

Non-covalent interactions reveal the protein chain δ conformation in a flexible single-residue model

Zeynab Imani, V. Rao Mundlapati, Valérie Brenner, Eric Gloaguen, Katia Le Barbu-Debus, Anne Zehnacker, Sylvie Robin, David J Aitken, Michel Mons

▶ To cite this version:

Zeynab Imani, V. Rao Mundlapati, Valérie Brenner, Eric Gloaguen, Katia Le Barbu-Debus, et al.. Non-covalent interactions reveal the protein chain δ conformation in a flexible single-residue model. Chemical Communications, 2023, 10.1039/D2CC06658K. hal-03923111

HAL Id: hal-03923111

https://hal.science/hal-03923111

Submitted on 4 Jan 2023

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



View Article Online
View Journal

ChemComm

Chemical Communications

Accepted Manuscript

This article can be cited before page numbers have been issued, to do this please use: Z. Imani, V. R. Mundlapati, V. Brenner, E. Gloaguen, K. Le Barbu-Debus, A. Zehnacker, S. Robin, D. J. Aitken and M. Mons, *Chem. Commun.*, 2023, DOI: 10.1039/D2CC06658K.



This is an Accepted Manuscript, which has been through the Royal Society of Chemistry peer review process and has been accepted for publication.

Accepted Manuscripts are published online shortly after acceptance, before technical editing, formatting and proof reading. Using this free service, authors can make their results available to the community, in citable form, before we publish the edited article. We will replace this Accepted Manuscript with the edited and formatted Advance Article as soon as it is available.

You can find more information about Accepted Manuscripts in the <u>Information for Authors</u>.

Please note that technical editing may introduce minor changes to the text and/or graphics, which may alter content. The journal's standard <u>Terms & Conditions</u> and the <u>Ethical guidelines</u> still apply. In no event shall the Royal Society of Chemistry be held responsible for any errors or omissions in this Accepted Manuscript or any consequences arising from the use of any information it contains.



Open Access Article. Published on 04 January 2023. Downloaded on 1/4/2023 2:58:10 PM.

This article is licensed under a Creative Commons Attribution-NonCommercial 3:0 Unported Licence

View Article Online DOI: 10.1039/D2CC06658K

COMMUNICATION

Non-covalent interactions reveal the protein chain δ conformation in a flexible single-residue model

Received 00th January 20xx, Accepted 00th January 20xx

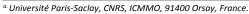
DOI: 10.1039/x0xx00000x

Zeynab Imani, [‡]a* Venkateswara Rao Mundlapati, [‡]b* Valérie Brenner, ^b Eric Gloaguen, ^b Katia Le Barbu-Debus, ^c Anne Zehnacker-Rentien, ^c Sylvie Robin, ^{a,d} David J. Aitken, ^{*a} Michel Mons *b

The δ conformation is a local secondary structural feature in proteins that implicates a π_{amide} N-H···N interaction between a backbone N atom and the NH of the following residue. Small-molecule models of this conformation have been limited so far to rigid proline-type models that may over-emphasize the significance of the interaction. We show here that in derivatives of a cyclic amino acid with a sulphur atom in the γ -position, specific sidechain/backbone N-H···S interactions stabilize the δ conformation sufficiently to allow it to compete with classical C5 and C7 H-bonding conformers.

Nature makes use of non-covalent interactions to stabilize well-defined secondary and tertiary structures in peptides and proteins. Backbone N-H···O=C hydrogen bonding is preeminent in the repertoire and provides the basis for the well-known topologies of turns, helices and sheets. The appreciation of other types of interaction, often found in cooperative combination with classical backbone hydrogen bonding, is still emerging; examples of such phenomena are intra-residue C5 H-bonding, C-H···O interactions, hoperative interactions, C-bonds, cation— π interactions, amongst others. The cooperative interactions, amongst others.

An intriguing feature of folded proteins is a local backbone geometry referred to as the δ conformation, 11 that appears on the Ramachandran (φ,ψ) map as an extension of the helix-promoting α region stretching towards the β region, centered at $\psi=0^\circ$ and $\varphi=-90^\circ$, known as the bridge (or δ) region (Fig. 1). It has been suggested that the δ conformation may be stabilized in proteins by a favorable interaction between H(i+1) of a given residue and the amide π electron density on N(i), $^{12-15}$ and this tenet has been consolidated by statistical, theoretical and experimental studies. $^{16-19}$ A role for such an NH(i+1)···N(i) interaction, also referred to as a $\pi_{\rm amide}$ ($\pi_{\rm am}$) interaction, has been suggested in the enzymatic isomerization of proline, 20,21 and it has recently been evoked in the stabilization of synthetic azapeptides 22 and as an accompaniment to β turn conformers in small peptides. 23,24 While the $\pi_{\rm am}$ interaction is



^{b.} Université Paris-Saclay, CEA, CNRS, LIDYL, 91191 Gif-sur-Yvette, France.

Université de Toulouse. France

Electronic Supplementary Information (ESI) available. See DOI: 10.1039/x0xx00000

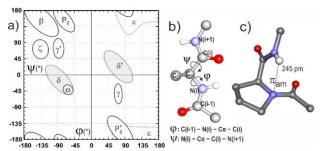


Figure 1 (a) The classical Ramachandran map, highlighting the δ region and its δ' mirror image. (b) Definitions of (φ, ψ) protein backbone angles. (c) An example of a π_{am} interaction implicating a proline residue in a protein; NH(i+1) is located perpendicular to the plane of the preceding amide bond to interact with the π electron density on N(i). In this example, $\varphi = -69^{\circ}$ and $\psi = -23^{\circ}$.

considered to be very weak, the prevalence of local δ conformations in proteins vindicates further investigation and characterization, in order to better understand its role in protein folding. ²⁵

Small-molecule models (comprising of a single capped α -amino acid) that facilitate experimental spectroscopic characterization of the δ conformation and an assessment of its energetics have remained elusive in the literature. Early pioneering studies on single-residue derivatives of glycine (Gly) and α -aminoisobutyric acid (Aib) suggested that short range C5 and C7 H-bonding interactions preclude the adoption of δ conformations in solution; only with imino acid derivatives could the $\pi_{\rm am}$ interaction be detected using IR spectroscopy. 26 The dihedral angle ϕ of proline is advantageously fixed around -60° and the limited panel of small-molecule model systems studied so far has retained a proline-inspired structural constraint in order to impose a δ conformation. 16,20,21,27 To date, no small-molecule model has yet been described for which a δ conformation is shown to compete with accessible C7 and C5 conformers without imposition of covalent constraints.

In order to procure a non-proline δ conformation model, we considered that an α,α -disubstituted amino acid backbone would be auspicious, since a δ -folded conformer has been detected in a gas phase conformational landscape of Cbz-(Aib)₂-OMe,²⁸ and in theoretical²⁹ and experimental³⁰ gas phase studies of 1-aminocyclobutane-1-carboxylic acid (Ac4c), a constrained cyclic analog of Aib (Fig. 2). Local δ conformations implicating Aib residues have been observed in peptide crystal structures on several occasions.³¹

^{c.} Université Paris-Saclay, CNRS, ISMO, 91400 Orsay, France.

d. Université de Paris, Faculté de Pharmacie, 75006 Paris, France.

[‡] These authors contributed equally to this work.

[❖] Current address: Département de Chimie, Université de Montréal, Canada ℋ Current address: Institut de Recherche en Astrophysique et Planétologie,

ChemComm

Communication

A further conceptual premise was that a local backbone conformation could be influenced through finely-controlled side-chain/backbone interactions, particularly those induced by a sulphur atom in the γ -position of the side-chain (Fig. 2). In this regard, both cysteine (Cys) 32 and S-methylcysteine (Cys(Me)) 33 can benefit from intra-residue (C5 γ) or vicinal (C6 γ) NH···S H-bonding interactions that stabilize extended or folded forms, respectively, while in the cyclic thioether analog, 3-aminothietane-3-carboxylic acid (Attc), the combination of C6 γ (side-chain) and C5 (backbone) H-bonds stabilizes a predominant extended C5-C6 γ conformer. We reasoned that tailoring the covalent constraints in the cyclic side chain should have a significant impact on the relative strengths of C6 γ and C5 γ H-bonds as well as their compatibility with canonical backbone C7 and C5 H-bonding regimes.

To substantiate this hypothesis, taking Attc derivative ${\bf 1}$ as the starting point, we have made a comparative study of the behavior of compounds ${\bf 2}$ and ${\bf 3}$, which are derivatives of the corresponding 5-and 6-membered cyclic α,α -disubstituted amino acids, 3-aminothiolane-3-carboxylic acid (Atlc) and 3-aminothiane-3-carboxylic acid (Atc), respectively (Fig. 2).

Ring-size constraints lead to NH···S geometries that favor 5γ H-bonds \underline{and} stabilize δ conformations.

This article is licensed under a Creative Commons Attribution-NonCommercial 3.0 Unported Licence

pen Access Article. Published on 04 January 2023. Downloaded on 1/4/2023 2:58:10 PM

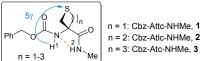


Figure 2 Rationale for this study and molecules **1-3** investigated. Racemic materials were used experimentally; theoretical calculations were carried out on molecules with the *S* configuration. For clarity, the C label is dropped in all figures when designating the C5 and C7 H-bonds.

In the process, we have discovered the ability of the sulphur atom to enable C5 γ H-bonds that stabilize δ -folded conformations, both in the gas phase and in weakly polar solution, providing new IR spectroscopic data for a π_{am} interaction.

We began with a quantum chemistry structural modelling study of molecules **1-3** both isolated in the gas phase (DFT-D level) and in chloroform solution, modelled using a polarizable continuum (see details in the ESI). Three families of low energy backbone structures characterized by distinct Ramachandran regions were obtained (Fig. 3a-c), with only minor differences between gas and solution phase geometries (Fig. S2.1). The classical extended and γ -folded forms, stabilized by C5 and C7 interactions respectively, were accompanied by the quested δ -folded conformations. These latter were characterized by quasi-perpendicular peptide bonds and a weak π_{am} interaction with a noticeably short NH(2)···N(1) distance (ca. 230 pm). A striking feature was the juxtaposition of the S atom with the neighboring NH moiety, featuring short NH···S distances and H-bonding interactions that varied significantly with the molecule and the backbone type being considered.

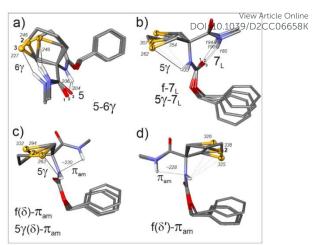


Figure 3 Most stable theoretical structures of each backbone family of species **1-3** in chloroform solution, illustrated with the *gauche*—rotamer of the *trans* benzyl carbamate (a-c). The illustrations show the more stable ring puckering conformers, all of which have the sulphur atom pointing towards N2. For structural comparison, the C1, C α , and N2 atoms of each conformation have been overlapped. The H-bond NH···O/S/N distances are given in pm. The nomenclature indicates the H-bonding status of each NH along the backbone. Within each family, less stable orientations of the ring relative to the backbone also occur, as do backbone mirror structures, *e.g.* with a δ'-fold (d) or a 7_D H-bond (see Fig. S2.2). Strong interactions are represented with full lines, weaker ones with dotted lines.

The C5 extended forms of each of the three compounds (Fig. 3a) featured an inter-residue NH(2)···S C6 γ interaction with a short NH···S distance (within the range 225-250 pm). The C7 γ -folded (Fig. 3b) and the δ -folded (Fig. 3c) forms of each compound both featured an intra-residue NH(1)···S C5 γ interaction, for which the H-bonding distance (and thus strength) varied between the three molecules depending on a number of factors: the chirality of the backbone fold (7_L or 7_D , δ or δ'); the ring puckering (depending on whether the S atom is closer to O(1) or N(2)); the local conformation (gauche +/- or trans) of the benzyl carbamate group as well as its configuration (trans or cis). Each type of backbone described above therefore gave rise to a conformational family, wherein all these structural features can vary (see Figs. S2.1 & S2.2).

The significance of the stabilizing effects of NH···S bonds is reflected in the relative energetics of the three backbone conformation families (Fig. 4 and Fig. S2.4). In the series $\mathbf{1}\rightarrow\mathbf{2}\rightarrow\mathbf{3}$, the energetics suggest that the δ conformer family becomes increasingly favorable and that the accompaniment of an NH···S C5 γ interaction plays a primordial role (Fig. 3c). The stabilizing effect of the NH···S interaction is commensurate with the shortening of the NH···S distances highlighted by comparison of the energies of the "mirrorimage" δ and δ ' conformer families of $\mathbf{2}$ and $\mathbf{3}$ (Fig. 3c and 3d); the former incorporate the C5 γ interaction and are 3-4 kJ/ mol more stable than the latter, which have no such interaction.

Despite the presence of a strong C6 γ H-bond in the extended form of all three molecules, a progressive destabilization (by ca. 10 kJ/mol) of the extended (C5/C6 γ) form relative to folded (C7 or δ) forms is observed in the series $1 \rightarrow 2 \rightarrow 3$, indicating that side-chain/backbone H-bonds do not alone control the energetics and that geometric distortions of both backbone and side-chain are also contributing factors.

Journal Name ARTICLE

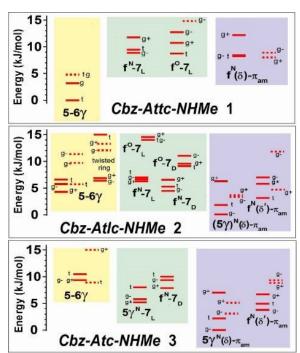


Figure 4 Chloroform solution (300 K) energetic conformational landscape of compounds 1-3 obtained from quantum chemistry calculations (DFT-D level and polarizable continuum; details in ESI, section S2), illustrating the three backbone families present in these species - C5/C6 γ (yellow), C7 (green) and δ (violet) - and the stabilizing effect of the C5 γ interaction on the δ conformer. See Fig. 3 and Fig. S2.2 for the conformational nomenclature. The labels g+, g– and t designate the orientation of the Cbz.

A gas phase conformation-selective laser spectroscopic study was carried out using UV and IR/UV double-resonance spectroscopies³⁶ and the data were compared to theoretical calculations (see ESI, sections S3 and S4). Compounds 1 and 2 behaved similarly, each exhibiting a single conformation assigned to an extended backbone family featuring two red-shifted H-bonds (close to 3360 and 3400 cm⁻¹), consistent with the calculated spectrum for the most stable C5-C6y structure in the gas phase. In contrast, the numerous UV features of 3 provided five different NH stretch IR spectra (designated A-E; see ESI, section S4.2 for details). Comparison with calculated spectra for the most stable forms of each family led us to assign A to the C5-C6 γ family and B to the C5 γ -C7 family. More interestingly, spectra C-E (Fig. 5) were assigned to conformations belonging to the δ -folded C5 γ - π _{am} family, characterized by a significantly red-shifted band (3345-3405 cm⁻¹ range) and a blue band (3465-3475 cm $^{-1}$), corresponding respectively to the C5 γ Hbond and the π_{am} features. This important observation demonstrates the existence of and provides spectral characterization for the δ folded family of compound 3 in the gas phase, albeit in competition with the classical extended and folded forms.

The search for evidence of δ conformations of compounds **1-3** in solution phase was rewarding. IR absorption spectra were recorded in chloroform (5 mM), without concentration-related effects (Fig S5.2), indicating the intramolecular nature of any non-covalent interactions. Considerable differences were observed in the amide NH stretch region in the series: one main band was observed for **1**, four were observed for **2** and two were observed for **3** (Fig. 6).

The standout feature in the spectrum of **3** was the strong blueshifted band at 3450 cm $^{-1}$ (yellow zone in Fig. 16). This cand assigned to NH(2) of δ -folded forms (Fig. 3c), in agreement with the solution state energetics study (Fig. 4). Concomitantly, the H-bonded NH(1) in a C5 γ (δ)- π _{am} conformer of **3** is red-shifted and gives rise to the broad band at 3365 cm $^{-1}$ (cyan zone in Fig. 6). These assignments are supported by the theoretical solution state spectra (Fig. 6) and demonstrate that the δ -folded conformer family is indeed predominant for compound **3** in solution.

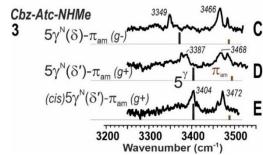


Figure 5 Gas-phase conformation-selective IR spectra (black) of compound **3**, recorded by IR/UV double resonance method: the letter labels (right) refer to the UV band used for conformer selection. For comparison, theoretical IR spectra (sticks) are given for selected representative low energy gas phase conformations. The π_{am} bands (around 3470 cm⁻¹) appear as intense doublets due to a Fermi resonance with one of the CO stretches (see details in ESI).

The case of **2** was more complex since several conformer families appear to populate the solution state landscape. While the δ -folded $5\gamma(\delta)$ - π_{am} conformer was less prominent than for compound **3** it was still in evidence, giving rise to bands at around 3415 cm $^{-1}$ for NH(1) (with a weaker C5 γ H-bond than in compound **3**) and at 3450 cm $^{-1}$ for NH(2) involved in a π_{am} interaction. The presence of a C5-C6 γ conformer was indicated by the band at 3385 cm $^{-1}$, typical for N(1) in a C5 interaction (violet zone in Fig. 6), 30 with NH(2) appearing in the broad red-shifted region around 3340 cm $^{-1}$. Furthermore, a f-7 conformer may contribute to the bands observed at 3430 and 3340 $^{-1}$.

The δ conformation was not in evidence in compound **1**, whose IR spectrum showed one broad band at 3375 cm⁻¹, assigned to the overlapping NH(1) and NH(2) absorptions of a predominant C5-C6 γ conformer. Only previously noted in passing,³⁴ the two minor bands at higher frequencies (3430 and 3450 cm⁻¹) can now be attributed to diminutive contributions from higher energy f(δ)- π _{am} and/or f-7 conformers of **1**.

The prevalence of a δ -folded conformer of **3** in chloroform solution was supported by 1 H NMR experiments. Titration with DMSO- d_6 induced small downfield shifts ($\Delta\delta$ < 0.3 ppm for 10% added DMSO- d_6) indicating limited solvent exposure, while a NOESY experiment showed correlations between the NHs and nearby (within 3.5 Å) CH atoms in a δ -folded conformer (see ESI, section S.6).

In summary, these studies provide spectroscopic characterization, supported by theory, of the first example of a single-residue model of the δ conformation of protein secondary structure without recourse to the backbone restriction deployed in previous proline-derived models. Stabilized by a propitious interplay of non-covalent

Communication ChemComm

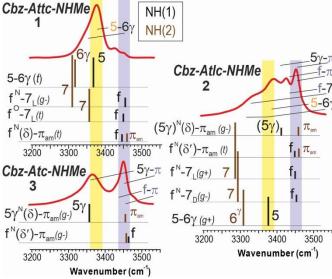


Figure 6. Solution state IR spectra of compounds **1-3** (5 mM in chloroform) and the calculated IR spectra of their most energetically relevant conformations (a more complete set is shown Fig. S5.3). The colored zones indicate the typical locations of diagnostic spectral features: nearly free (π_{am}) NH in a δ conformation (violet) and NH engaged in a C5 H-bond (yellow).

interactions including a C5 γ intra-residue NH···S interaction, the δ -folded form of derivative **3** competes successfully with the alternative C5-C6 γ and C7 conformers. In solution, the NH vibration in δ -folded **3** (and also in δ -folded **2**) appears at ca. 3450 cm⁻¹. This compares with an absorption in the 3420-3430 cm⁻¹ range for constrained proline-derived models, 20,21 suggesting that the backbone restrictions in the latter may over-emphasize the red-shift. The present study reiterates that the N-H···N interaction should be considered as very weak. In the conformational analysis of many small peptides over the years, NH vibrations around 3450 cm⁻¹ have been designated as "free"; the present findings suggest that such absorbance bands may also include contributions from δ -folded conformations.

Support from the French National Research Agency (Grant ANR-17-CE29-0008) and from "Investissements d'Avenir" programmes (LabEx PALM; grant ANR-10-LABX-0039-PALM; DIRCOS) is acknowledged. We thank the Grand Equipement National de Calcul Intensif at TGCC/CINES/IDRIS (Grant 2020-A0070807540), the CCRT High Performance Computing at CEA (Grant CCRT2020-p606bren) and MésoLUM (U. Paris-Saclay) for computing facilities.

Conflicts of interest

There are no conflicts to declare

Notes and references

- 1 A. Karshikoff, *Non-covalent Interactions in Proteins*, Imperial College Press, London, UK, 2006.
- 2 G. A. Jeffrey and W. Sänger, Hydrogen Bonding in Biological Structures, Springer-Verlag, Berlin, Germany, 1991.
- R. W. Newberry and R. T. Raines, *Nat. Chem. Biol.*, 2016, **12**, 1084.

- Z. S. Derewenda, L. Lee and U. Derewenda, J. Mol. Biol. 1995
 252, 248. DOI: 10.1039/D2CC06658K
- 5 S. Horowitz and R. C. Trievel, J. Biol. Chem., 2012, 287, 41576.
- G. B. McGaughey, M. Gagne and A. K. Rappe, J. Biol. Chem., 1998, 273. 15458.
- 7 G. J. Bartlett, A. Choudhary, R. T. Raines and D. N. Woolfson, Nat. Chem. Biol., 2010, 6, 615.
- 8 V. R. Mundlapati, D. K. Sahoo, S. Bhaumik, S. Jena, A. Chandrakar and H. S. Biswal, *Angew. Chem. Int. Ed.*, 2018, **57**, 16496.
- 9 D. A. Dougherty, Science, 1996, **271**, 163.
- 10 R. W. Newberry and R. T. Raines, ACS Chem. Biol., 2019, 14, 1677.
- 11 S. Hollingsworth and P. Karplus, BioMol. Concepts, 2010, 1, 271.
- 12 F. M. Pohl, Nature-New Biology, 1971, 234, 277.
- A. Gieren, B. Dederer and F. Schanda, Z. Naturforsch. C, 1980, 35, 741.
- 14 J. N. Scarsdale, C. Vanalsenoy, V. J. Klimkowski, L. Schafer and F. A. Momany, J. Am. Chem. Soc., 1983, 105, 3438.
- 15 P. A. Karplus, *Protein Sci.*, 1996, **5**, 1406.
- 16 R. Adhikary, J. Zimmermann, J. Liu, R. P. Forrest, T. D. Janicki, P. E. Dawson, S. A. Corcelli and F. E. Romesberg, J. Am. Chem. Soc., 2014, 136, 13474.
- 17 M. Holcomb, R. Adhikary, J. Zimmermann and F. E. Romesberg, J. Phys. Chem. A, 2018, 122, 446.
- 18 J. Dhar, R. Kishore and P. Chakrabarti, Sci. Rep., 2016, **6**, 31483.
- 19 R. Deepak and R. Sankararamakrishnan, *Biophys. J.*, 2016, **110**, 1967.
- C. Cox, V. G. Young and T. Lectka, J. Am. Chem. Soc., 1997, 119, 2307.
- 21 C. Cox and T. Lectka, J. Am. Chem. Soc., 1998, 120, 10660.
- 22 K. Baruah, B. Sahariah, S. S. Sakpal, J. K. R. Deka, A. K. Bar, S. Bagchi and B. K. Sarma, *Org. Lett.*, 2021, 23, 4949.
- 23 K. Scholten and C. Merten, Phys. Chem. Chem. Phys., 2022, 24, 3611.
- 24 V. C. D'mello, G. Goldsztejn, V. R. Mundlapati, V. Brenner, E. Gloaguen, F. Charnay-Pouget, D. J. Aitken and M. Mons, *Chem. Eur. J.*, 2022, **28**, e202104328.
- 25 A native protein fold is only 20-50 kJ/mol more stable than misfolded or unfolded forms; see: G.D. Rose, P.J. Fleming, J.R. Banavar, A. Maritan, *Proc. Natl. Acad. Sci. U. S. A.* 2006, 103, 16623.
- 26 M. Marraud, J. Néel and B. Maigret, J. Chim. Phys. Phys.- Chim. Biol., 1975, 72, 1173.
- 27 T. Vermeyen and C. Merten, Phys. Chem. Chem. Phys., 2020, 22, 15640.
- 28 J. R. Gord, D. M. Hewett, A. O. Hernandez-Castillo, K. N. Blodgett, M. C. Rotondaro, A. Varuolo, M. A. Kubasik and T. S. Zwier, *Phys. Chem. Chem. Phys.*, 2016, **18**, 25512.
- 29 J. Casanovas, D. Zanuy, R. Nussinov and C. Alemán, *Chem. Phys. Lett.*, 2006, **429**, 558.
- 30 V. R. Mundlapati, Z. Imani, V. C. D'mello, V. Brenner, E. Gloaguen, J.-P. Baltaze, S. Robin, M. Mons and D. J. Aitken, *Chem. Sci.*, 2021, **12**. 14826.
- 31 S. Aravinda, N. Shamala and P. Balaram, *Chem. Biodiversity*, 2008, **5**, 1238.
- 32 G. Goldsztejn, V. R. Mundlapati, V. Brenner, E. Gloaguen, M. Mons, C. Cabezas, I. Leon and J. L. Alonso, *Phys. Chem. Chem. Phys.*, 2020, 22, 20284.
- 33 V. R. Mundlapati, Z. Imani, G. Goldsztejn, E. Gloaguen, V. Brenner, K. Le Barbu-Debus, A. Zehnacker-Rentien, J.-P. Baltaze, S. Robin, M. Mons and D. J. Aitken, *Amino Acids*, 2021, 53, 621.
- 34 Z. Imani, V. R. Mundlapati, G. Goldsztejn, V. Brenner, E. Gloaguen, R. Guillot, J.-P. Baltaze, K. Le Barbu-Debus, S. Robin, A. Zehnacker, M. Mons and D. J. Aitken, *Chem. Sci.*, 2020, 11, 9191.
- 35 The synthesis of Cbz-Attc-NHMe **1** has been described before (ref 34); related methods were used to prepare Cbz-Atlc-NHMe **2** and Cbz-Atc-NHMe **3**. Full details are provided in the ESI, section S1.
- 36 E. Gloaguen, M. Mons, K. Schwing and M. Gerhards, Chem. Rev., 2020, 120, 12490.