

# New highlights on Old World giant Phragmites (Poaceae) using leaf and floral bract microscopic characters

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- 2 microscopic characters
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- Oceanography (MIO), UM 110, Marseille, France
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### 1. Introduction

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- 17 The genus *Phragmites* Adans. (Poaceae) has a worldwide distribution from the tropics to the
- cold-temperate regions. Its positive attributes as an ecosystem engineer in wetlands provide
- 19 essential ecological and economic services, such as wildlife habitat, erosion control, and
- biomass production (Saltonstall and Meyerson, 2016). All species form dense stands, playing a
- crucial role as carbon sinks over a long-time scale (Brix et al., 2001), and they are also known
- for their phytoremediation abilities. Like other tall grasses, *Phragmites* spp. have followed
- humans since the Paleolithic period due to their countless uses, e.g. energy source, construction,
- fodder, weapons, music, fiber and medicine (Täckholm et al., 1973; Oyen, 2011). Due to their
- usefulness, they were introduced into many adventive ranges, like the reed genus Arundo L.
- 26 (Hardion et al., 2014). The cosmopolitan *P. australis* (Cav.) Trin. ex Steud. has become a
- 27 "model organism for plant invasions" driven by its competitive abilities, particularly in North
- 28 America (Pyšek et al., 2019).
- 29 Although *Phragmites* are the subject of an extensive number of articles, their taxonomy
- remains unclear. Thereby, the species number is widely debated, from only one species P.
- 31 australis (Täckholm et al., 1973, under the name P. communis Trin.) to seven only for Corsica
- 32 Island (Mabille, 1869) and for Russia (Tsvelev and Probatova, 2019), and up to 17 in the world
- 33 (Steudel, 1854). In recent studies (Clevering and Lissner, 1999; Scholz and Böhling, 2000;

- Lambertini et al., 2006), the most cited taxa are *P. australis* (cosmopolitan), *P. karka* (Retz.)
- 35 Trin. ex Steud. (pantropical, except the American continent), P. mauritianus Kunth (tropical
- Africa, Madagascar, Mascarene Islands), *P. japonicus* Steudel (N-Japan, E-Russia, NE-China),
- and *P. frutescens* Scholz (Crete, Greece, SW-Turkey).
- The poor taxonomic resolution of *Phragmites* results from many overlaps of quantitative
- macroscopic traits, and the ambiguity of descriptive terms. For example according to Conert
- 40 (1961) or Clayton (1970), the upper glume is either lanceolate or acute in *P. karka*, and either
- 41 ovoid or acuminate in *P. mauritianus*. Long considered as distinct species, Clayton (1970)
- suggests a subspecific status for *P. mauritianus* within *P. karka*, while some floras simply
- include *P. mauritianus* within *P. karka*, without any infraspecific rank (e.g. Phillips, 1995).
- Moreover for Veldkamp (1992), P. karka should be named P. vallatoria (Plunk. ex L.) Veldk.,
- a name now considered as invalid (Kirkbride and Wiersema, 2007).
- Surprisingly, despite its morphological differentiation, online taxonomic databases (e.g. Kew,
- 47 Paris) place *P. frutescens* in synonymy within *P. australis*. Described in 1996, *P. frutescens* is
- a rare species with infrequent flowering, but morphologically more related to *P. mauritianus*
- 49 than to *P. australis* (Scholz and Böhling, 2000), as confirmed by AFLP analyses (Lambertini
- et al., 2006: Fig. A1). In fact, the three taxa, P. karka, P. mauritianus, P. frutescens, form a
- distinct genetic unit, with two unique cpDNA haplotypes (U, I:Table 1).
- In his monograph, Conert (1961) lists 11 varieties within *P. australis*, while Clayton (1970)
- recognizes only two taxa: the temperate subsp. *australis* (< 3 m) and the Mediterranean subsp.
- 54 altissimus (Benth.) Clayton (height and panicles taller). These two subspecies are adopted by
- Feinbrun-Dothan (1986), but many floras include subsp. altissimus within P. australis (e.g.
- Tutin, 1980; Fish et al., 2015), or place it under various names and ranks, such as var. isiaca
- 57 (Del.) Coss. (Täckholm et al., 1973; Hultén and Fries, 1986), but rarely as a species [Coste,
- 58 1937, as *Arundo maxima* Forsk.; Tsvelev and Probatova, 2019, as *P. altissimus* (Benth.) Mab.].
- All genetic studies separate this taxon from the subsp. *australis*, but the individual groupings
- differ according to methods and sampling, (AFLPs, haplotypes, SSRs: Lambertini et al., 2006,
- 61 2012; RAD-sequencing: Wang et al., 2021).
- Finally, according to genetic studies, *P. japonicus* is separate from the *P. australis* subsp.
- 63 australis (hereafter P. australis s.str.), which includes two distinct phylogeographic groups: the
- Afro-European unit (AE) and the Australian-E. Asian unit (AA) (Lambertini et al., 2006, 2012,
- 65 2020). In America, there are three native genetic units that highly differ from all Old World
- taxa, based on AFLPs, haplotypes, microsatellites (Lambertini et al., 2006: Fig. A1; Saltonstall
- and Meyerson, 2016) and RAD-sequencing (Wang et al., 2021).

In the absence of reliable macroscopic characters in *Phragmites*, we investigated the microscopic traits of leaves and floral bracts. The high phylogenetic value of leaf epidermis and anatomy traits is known since the founding work of Prat (1932) and has proven to be valuable for the systematics of Poaceae (Watson et al., 1992; Kellogg, 2015), and for the identification of South African *Phragmites* (Gordon-Gray and Ward, 1971). Such traits can be useful when phylogenetic markers fail to resolve complex grass groups. The traits of floral bracts were also found to have high discriminant value, phylogenetic significance (Liu et al., 2010; Hardion et al., 2021), and are comparatively easy to use. Based on extensive sampling, including plants previously analyzed in molecular studies, our aim here was to test the value of micromorphologic characters to distinguish Old World *Phragmites*, especially the problematic giant reeds.

### 2. Materials and Methods

For a representative cover of the global distribution of taxa in the Old World, specimens from several international herbaria, Aix-Marseille University collections, and twenty samples previously studied in molecular analyses were investigated. The study was performed on 160 specimens of *Phragmites* [35 *P. karka*, 30 *P. mauritianus*, 11 *P. frutescens*, 44 *P. australis* subsp. *altissimus*, 31 Afro-European (AE) *P. australis* subsp. *australis*, 5 Australian-E. Asian (AA) *P. australis*, 4 *P. japonicus*], and one *Molinia caerulea* (L.) Moench., the sister group of *Phragmites* (Hardion et al., 2021). All information on specimens are reported in the table A1. Prior identifications were based on macroscopic characters observed on herbarium specimens, except for the genetic unit AA *P. australis* for which no distinctive macro-morphological character is yet known. These traits were chosen from original diagnosis, from some floras (Coste, 1937; Clayton, 1970; Feinbrun-Dothan, 1986), and from the synthesis of Scholz and Böhling (2000).

Courtesy of herbarium curators, we collected four mature spikelets on each sample, and a small piece from the middle part of the two terminal leaves of the primary axis, because they possess the most differentiated epidermis (Prat, 1932). To avoid damages on type specimens, two spikelets (taken in the bag of fragments) and an imprint of leaf epidermis with clear nail polish were taken. The material was re-hydrated in distilled water for ca. one week. For each spikelet, glumes, lemmas and paleas of the two first basal flowers were separated by cutting at their base. For each leaf, the adaxial epidermis was prepared by scraping away the abaxial face and mesophyll cells with a razor blade. Floral bracts, leaf margin and epidermis were stained

with iodine green solution for 3 min and mounted in glycerol. For anatomy, about 10 transversal sections per leaf were performed in lateral part (out of the central rib), cleared with sodium hypochlorite, washed in distilled water, stained with carmine green and mounted in glycerol. In the spikelet, only lemmas and paleas, which are more differentiated than glumes in reeds, were studied in detail. Classical discriminant traits were studied, as well as several new characters, such as marginal teeth of leaves and paleas, and floral bract coloring. A total of 56 characters were retained: 23, 12 and 21 for the leaf epidermis, anatomy, and spikelet, respectively (Table A2). The micromorphological terminology follows Prat (1932) and its revisions (Metcalfe, 1960; Watson et al., 1992; ICPT, 2019). All specimens were studied using a light microscope (LM; ca.10 observations for each trait), and some samples were observed under scanning electron microscope (SEM). All traits were coded as discrete states (Table A2a-c). Before each analysis, datasets were converted into binary variables, with one new variable for each state of initial variables, using the *dummies* R-package in the R environment (R Core Team, 2022). Data were then analyzed in correspondence and hierarchical clustering analyses with Jaccard binary dissimilarities using the *ade4* R-package (Thioulouse et al., 2018).

### 3. Results

Based on 21 epidermal characters of lemmas and paleas, the hierarchical clustering analysis (HCA) highlights two main clusters (Fig. 1a). The clade A strictly includes the African and Mediterranean *P. mauritianus*, *P. frutescens* and *P. australis* subsp. *altissimus*, while the clade B includes the remaining taxa: *P. karka*, *P. japonicus* and *P. australis s.str*. The clade A mainly differs from clade B by glumes and lemmas being green and spread out, *vs* brownish and rolled up (Fig. 2a *vs* 2b), and lemma hooks larger and pointed to all directions, *vs* small (<30 μm) and pointed to the top (Fig. 3c-d *vs* 3a-b). In addition, within each clade, palea characters distinguish the different taxa (Fig. 3g-h and e-f). The correspondence analysis (CA) clearly confirms these two clades, with the first three axes representing 59.8% of the total variance (26.4%, 12.5% and 10.9%, respectively; Fig. A2a). The main distinctive traits of the two clades A and B, and taxa are summarized in a key (Box 1).

The HCA of the 23 leaf epidermal characters also highlights two main clusters that mainly match with previous ones, excepted for the inverted position of *P. australis* subsp. *altissimus* and *P. karka* between the two clades (Fig. 1b). The clade I includes the three highest (up to 8 m) and thermophilic species, *P. karka*, *P. mauritianus*, and *P. frutescens*, while the clade II includes the more temperate and smaller taxa, *P. australis* subsp. *altissimus*, subsp. *australis*,

and *P. japonica*. The clade I differs from clade II in its simpler epidermal structure: ribs with large base and long-pointed prickles (*vs* small in clade II), intercostal zones with bicellular microhairs (*vs* absent), many cork-cells (*vs* rare), rare hooks (*vs* frequent) (Fig. 3i-j *vs* 3k-m), and weakly ornate leaf margin (*vs* more ornate) (Fig. 2c-e *vs* 2f-j). In our preparations, the iodine green staining of floral bracts lasts several years, but that of leaf teeth fades after a few weeks. The CA confirms the two clades I and II, with the three first axes representing 78% of the total variance (54.1%, 14.9% and 9%, respectively; Fig. A2b). The main traits are reported in the Box 2.

The coupled analysis of floral and leaf epidermis shows a better discrimination of taxa (Fig. 4), notably for *P. frutescens*, the Afro-European (AE) and Australian-E. Asian (AA) *P. australis*. The three first axes of the CA represent 50.6% of the total variance (24.2%, 15.6% and 10.8%, respectively). The two S-African hybrids genetically attested (Canavan et al., 2018), KC2 (P. *mauritianus* x AE *P. australis*), and KC3 (*P. mauritianus* x *P. altissimus*), have a mix of parental traits, and the CA placed them far from their parents, especially from *P. mauritianus*.

Leaf anatomy shows the main structural characteristics of *Phragmites*, especially the shape and size of bulliform cells (BC), with the round bottom forming a gutter of regular adaxial groups between the vascular bundles (Fig. 5). The CA separates the giant reed taxa (> 3m) from the smaller ones along the axis 1, as well as the different taxa of each group (Fig. 6) and shows the polymorphism of AA *P. australis*. The three first axes represent 66% of the total variance (32%, 22% and 12%, respectively). The anatomical key based on the HCA distinguishes all the Old World taxa (Box 3).

### 4. Discussion

### 4.1. Taxonomy

The objective of this study was to delimit well supported Old World *Phragmites* taxa. As expected, microscopic characters have more discriminant resolution than macroscopic ones, and shed a new light on this problematic genus. Both leaf and spikelet traits show a strong individualization of four giant taxa: *P. mauritianus*, *P. karka*, and more surprisingly *P. frutescens* and *P. australis* subsp. *altissimus*. Given the stability and the high taxonomic value of these micro-characters, we consider them as four distinct species. For the subsp. *altissimus*, which possesses highly specific traits, the most appropriate name should be *P. altissimus* (Benth.) Mabille (1869).

The floras of S-Africa only report P. australis and ignore P. altissimus (Fish et al., 2015), while both taxa co-exist. Our results show that the S-African "P. australis" illustrated by Gordon-Gray and Ward (1971), and genetically studied (Canavan et al. 2018) is in fact P. altissimus. Our sampling does not show obvious micro-characters differences between S-African and Mediterranean P. altissimus, in agreement with DNA microsatellites analyses (Lambertini et al., 2012). Within *P. australis s.l.* of the Danube Delta, Hanganu et al. (1999) found two distinct taxa called "giant reeds" (2n = 96), and "fine reeds" (2n = 48), which likely correspond to P. altissimus and P. australis s.str., respectively. As suggested by the map of Hultén and Fries (1986) and the presence of the exclusive 200-bp waxy band (Lambertini et al., 2012), the distribution of P. altissimus is larger than expected, including African, Mediterranean, and Middle East regions up to Armenia and Kashmir. Phragmites frutescens is sometimes sympatric with P. altissimus and morphologically close to P. mauritianus, but it is restricted to the Mediterranean Basin, and it presents distinct specific traits, especially in leaf anatomy. 

Unlike the giant reeds, the taxonomy of Old World small reeds (< 3 m), *P. japonicus* and *P. australis s.str.* (AE, AA), appears more challenging. Except leaf anatomy, the other characters distinguish these three taxa less clearly than genetic studies (Lambertini et al., 2006, 2020; Liu et al., 2020; Wang et al., 2021). Further investigations are needed, based on a broader sampling in Australia and Asia, to clarify their morphological distinction and geographic distribution.

# 4.2. Anatomy

Leaf anatomy shows the most discriminant traits, particularly the bulliform cells (BC). The huge flabellate reed BC are unique among Poaceae (Chen et al., 2020), and called "Phragmitestype" as wetland markers in paleoecology. These are the only traits that easily separate P. frutescens (BC protruding) and P. japonicus (BC wider than deep) from all others. Anatomical analyses produce original groupings, and clearly distinguish three taxa within Old World P. australis s.str. (Box 3). The two temperate to cold small reeds (AE P. australis, and P. japonicus) possess thin leaves, and small fan-shaped BC, penetrating ca. 1/2 of the blade. In contrast, thick leaves and deep BC, penetrating ca. 2/3 of the blade, separate the thermophilic taxa (the giant reeds and AA P. australis) from the first two taxa. This group includes the tropical pair P. karka - P. mauritianus, and the three other subtropical taxa: the Mediterranean pair P. altissimus - P. frutescens, and the humid subtropical AA P. australis.

In Poaceae, Cyperaceae and Juncaceae, BC have a crucial role in leaf rolling, and water storage, in response to drought and salt stress. All *Phragmites* live in wetlands, but AE *P. australis* has a better salt tolerance than AA *P. australis* (Lambertini et al., 2020), and *P. altissimus* (Hanganu et al., 1999). Therefore, the strong anatomical differentiation of *Phragmites* taxa could be an adaptation to eco-climatic conditions.

### 4.3. Staining and waxy genes

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In iodine green solution, floral bracts, and leaf margin teeth of *P. frutescens, P. mauritianus* and *P. altissimus* turned green, while those of other taxa remain unstained. These results are consistent with the banding pattern of the waxy genes: green colour corresponds to the presence of 100 bp band in *P. mauritianus* and *P. frutescens*, 200 bp band in *P. altissimus*, and unstained parts match to the lack of waxy bands in the other taxa (Lambertini et al., 2012; Canavan et al., 2018).

Waxy plants, notably cereals, are of great interest for their countless industrial uses. Iodine solutions are used to detect waxy genes, they colour the starch of pollen and endosperm differently: waxy plants rich in amylopectin turn reddish, whereas non-waxy plants with only amylose turn blue (Bates et al., 1943). If in herbarium specimens, these organs are rarely in good condition, our results show that the spikelet and leaf staining can easily detect waxy plants. Similar observations were cited for waxy rice leaves (Abé, 1985). This phenomenon might result from the relationship between the presence of amylopectin and a better permeability of cell walls (Michailova et al., 2001). The spreading out or rolling up of glumes and lemmas in glycerol can also be explained by starch types: water soluble amylopectin, hydrophilic with alcohol, would cause spreading out bracts, while water insoluble amylose, hydrophobic with alcohol, would induce rolling up bracts. Waxy rice (glutinous) has been used for millennia in Asia, due to its better digestibility, early ripening, and tolerance to temperature variation and drought (Abé, 1985). Waxy corn also has a high resistance to pathogenic fungi (Walden, 1978), which can explain our leaf observations: taxa without waxy genes are often infected with rusts, while waxy taxa are almost always healthy. Biochemical differences could also explain the distinct uses of species, e.g. in African traditional medicine (Oyen, 2011).

### 4.4. Ploidy levels and hybrids

In karyological studies of *Phragmites*, the authors considered 2n = 48 as the tetraploid level, with a basic chromosome number n = 12, even if n = 6 is more likely for the Arundinoideae, particularly for *Molinia* spp. with 2n = 36 (Conert, 1961; Hardion et al., 2011). According to

- the main syntheses (Gorenflot, 1976; Connor et al., 1998; Clevering and Lissner, 1999; Ishii
- and Kadono, 2001; Hansen et al., 2007), the *Phragmites* include: (1) four stable species with
- 230 2n = 48: P. karka, P. mauritianus, P. frutescens and P. japonicus; (2) the AE P. australis with
- 231 2n = 48 (rarely 2n = 96); and (3) two greatly variable taxa: AA P. australis with 2n = 96 (120,
- 232 264), and P. altissimus with 2n = 48 (72) in Senegal and S. Europe, 96 (144) in S. Africa, S.
- 233 Mediterranean, Danube Delta, and Middle East.
- Some herbarium samples were difficult to identify, as evidenced by the various names
- attributed by different botanists (Table A1). Notably, two plants previously named "P. karka"
- showed mixed characters: (i) the Cameroonian P2 with spikelet traits of P. mauritianus and,
- leaf epidermis and anatomy of *P. karka*, and (ii) the Japanese P11 with spikelet traits and
- anatomy of AA *P. australis* and leaf epidermis of *P. japonicus*. Both could be hybrids of these
- close related taxa, or rather the expression of strong polymorphism, because the true S-African
- 240 hybrids KC2 (*P. mauritianus* x AE *P. australis:* 100 bp band), and KC3 (*P. mauritianus* x *P.*
- 241 *altissimus*: 100-200 bp bands), get away from their parent taxa in CA. In hybridizations between
- waxy and non-waxy taxa (P2: *P. mauritianus* x *P. karka*, or KC2), floral bracts and leaf margins
- 243 always turned green. This could explain some hybrids with 200 bp waxy band (S-Africa,
- Danube Delta), but related to AE *P. australis* in some genetic traits (Lambertini et al., 2012;
- 245 Canavan et al., 2018).
- The proportion of natural hybrids is challenging, given the strong polymorphism of reeds due
- 247 to the high polyploidy resulting from a reticulated evolution (Lambertini et al., 2012; Liu et al.,
- 248 2020; Wang et al., 2021). Like most polyploids (Richard, 1997), *Phragmites* spp. have weak
- genetic barriers that explain the occurrence of some hybrids. Based on a large *P. japonicus* (2n
- = 48) and AA *P. australis* (2n = 96, 120) sampling, Ishii and Kadono (2001) argue that hybrids
- are very rare due to: (i) mismatched flowering periods between species, (ii) temporal
- reproductive isolation related to the short flowering period, (iii) short distance of anemophilous
- pollination, and (iv) distinct ploidy levels without intermediate numbers. According to Scholz
- and Böhling (2000) and many floras, clear asynchronous flowering of sympatric taxa applies to
- all Phragmites, e.g. the early flowering of P. mauritianus vs P. karka, AE P. australis vs P.
- altissimus, and AA P. australis vs P. japonicus. In addition, the pollen of Mediterranean "P.
- australis" hybrids (2n = 72) is entirely sterile (Gorenflot et al., 1990). Lastly, in the wild, hybrid
- formation is also strongly limited by the almost exclusive vegetative growth of dense long-lived
- clones (Clevering and Lissner, 1999), often distinct geographic areas and eco-climatic
- preferences (geo-climatic barriers; Lambertini et al., 2012).

Micromorphological studies are not only descriptive, but they can also be used in phylogeny (Liu et al., 2010). Floral bract and leaf epidermis characters define different clades, implying that each category of traits has a distinct evolutionary significance. The main leaf traits of the clade I (*P. karka, P. mauritianus, P. frutescens*) are (i) ribs with huge prickles and oblong silicabodies, and (ii) intercostal zone without hooks, but with straight wall long cells, many bicellular microhairs and cork-cells. Comparisons of descriptions and illustrations (Metcalfe, 1960; Watson et al., 1992) show that these traits are also present in the earliest groups of the BEP clade in the phylogenetic classification of Poaceae (Soreng et al., 2017), *i.e.* the first three basal tribes of the Oryzoideae. The *Phragmites* share other characters with ancestral states, since this genus is the only one of the PACMAD clade having arm cells in the mesophyll, like the basal Oryzoideae and Bambusoideae (Kellogg, 2015).

The species of the clade I (P. karka, P. mauritianus, P. frutescens) form a morphological unit characterized by their height (8-10 m), large panicles, deciduous leaf sheaths, and leaf blades that are rough to the touch. Epidermis comparison between *Phragmites* and its sister group Molinia (Hardion et al., 2021) shows that M. caerulea shares a maximal similarity with P. karka: 85 and 78% for flower and leaf traits, respectively. Thus, P. karka retains the most number of ancestral traits across *Phragmites* spp., in accordance with its basal position in the AFLP tree (Lambertini et al., 2006: Fig. A1). In addition, P. karka and P. mauritianus have a large distribution in tropical warm riparian habitats, like the earliest Poaceae (Kellogg, 2015). Opposingly, the taxa of clade II (P. altissimus, P. australis s.str., and P. japonica) exhibit derived traits and are distributed in subtropical and temperate to cold zones. On the other hand, spikelet traits draw on two other evolutionary lineages, divergent from P. karka with an increasing differentiation. The clade B includes the non-waxy taxa (brownish bracts), with lightly ornamented lemma and palea: P. karka, AE P. australis, P. japonicus (2n = 48), and lastly AA P. australis (2n = 96). The clade A includes the waxy taxa (green bracts), with increasingly ornate paleas: P. mauritianus, P. frutescens (2n = 48), and P. altissimus (2n = 48)96).

In conclusion, we have highlighted several valuable micro-characters allowing for the identification of four Old World giant *Phragmites*, which match well with the previous genetic clustering. Future investigations will focus on the small reeds, particularly those from Asia and Australia, and the macroscopic characters of Old World *Phragmites*.

### CRediT authorship contribution statement

- 294 R. Verlaque: Conceptualization, species identification, morphological investigation, and writing. L.
- 295 Hardion: Data analyses, figures, and writing. K. Canavan, C. Lambertini and B. Vila: Field work,
- documents, and writing. M. Verlaque: tables, figures, and writing.

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### Appendix A. Supporting information

304 Supplementary data associated with this article can be found in the online version at

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### Legends of figures

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- Figure 1: Hierarchical clustering analyses: 1a. Floral bract epidermis; 1b. Leaf epidermis; 1c.
- Leaf anatomy. Clade A: Floral bracts green and spread out; Clade B: Floral bracts brownish
- and rolled up; Clade I: ribs with large prickles and bicellular microhairs; Clade II: ribs with
- small prickles, without bicellular microhairs.
- Figure 2: Lemmas. (a) Clade A, green and spread out: P. mauritianus (B24); (b) Clade B,
- brownish and rolled up: *Phragmites karka* (P9). **Leaf margin**: teeth rounded tipped, similar,
- low: (c) P. mauritianus (L1); (d-i): teeth large and small, pointed, erect, and more spaced: (d)
- 431 *P. karka* (B11); **(e)** *P. frutescens* (C196); **(f)** *P. japonicus* (C177); **(g)** Afro-European *P.*
- australis (MS24); (h) Australian-E-Asian P. australis (MP7); (i) P. australis subsp. altissimus:
- 433 (MS26); (j) detail of basal internal ornament of large teeth (MS20). Scale =  $100 \mu m$ .
- Figure 3: Lemma epidermis, Clade B, hooks pointed to the top: (a) Phragmites karka (B13),
- (b) AE *P. australis* (MS27). Clade A, hooks pointed in all directions: (c) *P. mauritianus* (B24),
- 436 (d) P. australis subsp. altissimus (MS1). Palea apex, Clade B, few ornamented: (e) P. karka

- 437 (B13); (f) P. japonicus (MP10, similar in AE P. australis). Clade A, well ornamented, large
- 438 teeth pointed in all directions: (g) P. mauritianus (B24); (h) P. australis subsp. altissimus
- 439 (MS2): entire palea with marginal long hairs. Leaf characters (abaxial face): Clade I, ribs with
- large prickles and bicellular microhairs: (i) P. karka (B39, similar in P. mauritianus); (j) P.
- 441 frutescens (MS8) smaller prickles. Clade II, ribs with small prickles, without bicellular
- microhairs: (k) AE P. australis (MS29, similar in AA); (l) P. japonicus (C177) many obvious
- hooks; (m) P. australis subsp. altissimus (C174) blunt prickles. R = rib; p = prickle; b =
- bicellular microhair; h = hook. Scales: (a-g) =  $40 \mu m$ ; (h) =  $250 \mu m$ ; (i-m) =  $20 \mu m$ .
- Figure 4: Hierarchical clustering analysis of floral bracts and leaves epidermal characters.
- 446 Hybrids: KC2 (*P. mauritianus* x AE *P. australis*) and KC3 (*P. mauritianus* x *P. altissimus*).
- Figure 5: Leaf anatomy (transversal section). (a) Phragmites karka (P6); (b) P. mauritianus
- 448 (P14); (c) P. frutescens (MS6); (d) P. australis subsp. altissimus (MP16); (e) Australian-E.
- Asian P. australis (MS16); (f) Afro-European P. australis (MS13); (g) P. japonicus (C177).
- BC = bulliform cells, ad = adaxial face, ab = abaxial face, arrow = obvious underlying colorless
- cells, I = primary vascular bundles (elliptic or circular), separated by 3-4 secondary bundles
- 452 (=II, always circular). Scale =  $50 \mu m$ .

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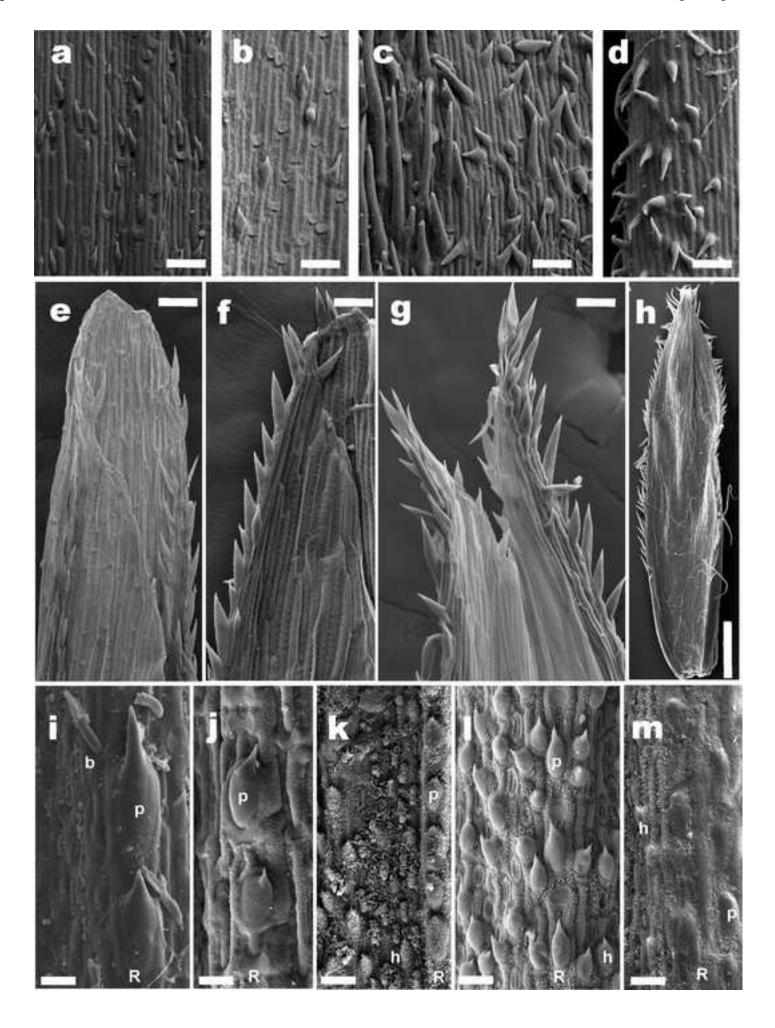
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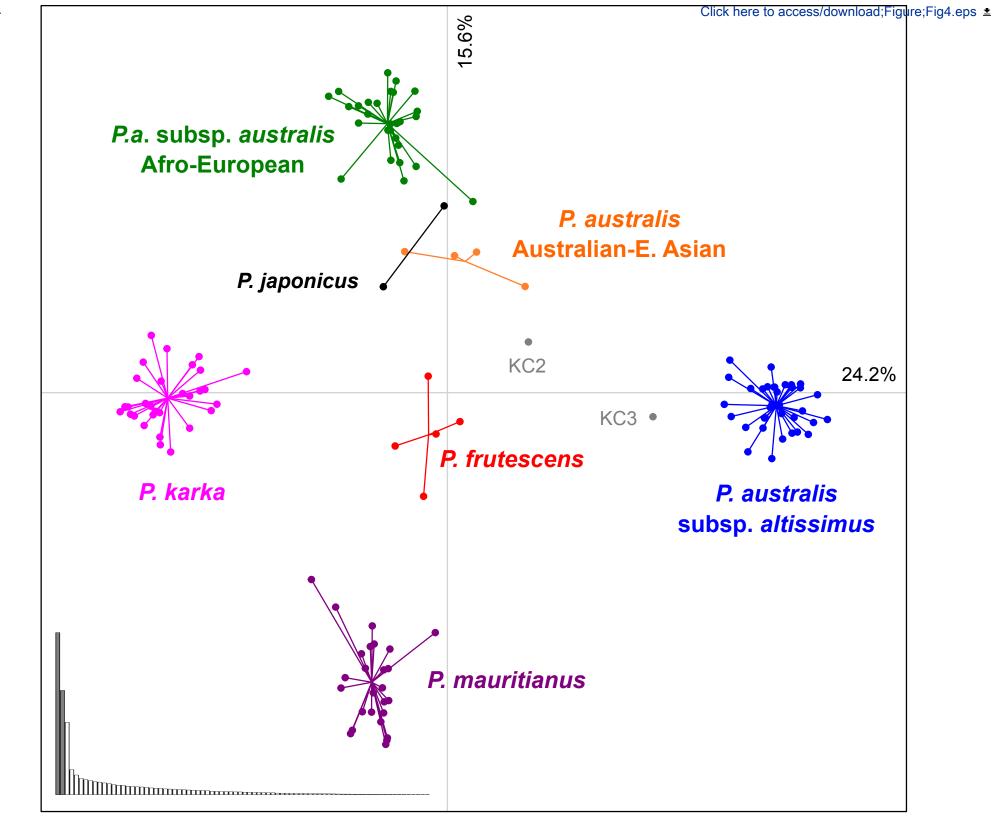
Figure 6: Hierarchical clustering analysis of leaf anatomic characters.

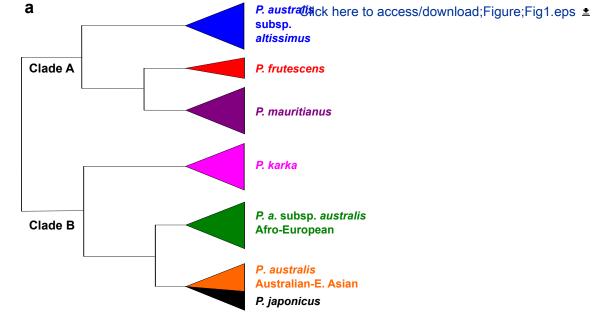
**Box 1**: Key of spikelet epidermal characters. 456 **Box 2**: Key of leaf epidermal characters. 457 **Box 3**: Key of leaf anatomy characters. 458 459 Box 1: Key of spikelet epidermal characters (specific traits in italics) 460 461 1. Glumes and lemmas green and spread out\*; lemma hooks: many large (> 30µm), pointed in all directions: clade A 462 2. Paleas densely ornate: margins with large teeth (70-140µm), pointed in all directions, and some long hairs (up to 1.5 mm); apex retuse with many hooks: P. australis subsp. altissimus 463 464 2'. Paleas less ornate: margins with smaller teeth (<100µm), and without long hairs 465 3. Hooks of lemmas and paleas large (30-150µm); palea apex bifid: P. mauritianus 3'. Hooks and margin teeth of lemmas and paleas smaller (<40µm); palea apex retuse-truncated: **P. frutescens** 466 467 1'. Glumes and lemmas brownish and rolled up\*; lemma hooks: smaller (< 30µm) and pointed to the top: clade B 4. Lemmas with straight long-cell walls and many hooks; palea: margins with small teeth (< 50 µm), veins stopped long 468 469 before the top, apex hyaline generally rounded: P. karka 470 4'. Lemmas with sinuous long-cell walls and rare hooks; palea margins with larger teeth (> 50µm), veins going to the top, apex not hyaline, generally truncated: small reeds including P. australis subsp. australis (AE, AA) and P. japonicus. 471 472 \* After staining with iodine green and mounting in glycerin. 473 474 Box 2: Key of leaf epidermal characters (specific traits in italics) 475 1. Ribs with large (> 65 μm) long-pointed prickles; intercostal zone (IZ) with bicellular microhairs, cork-cells and rare hooks: 476 clade I 477 2. Ribs with oblong silica bodies; IZ with superposed cork-cells; marginal teeth: rounded tip, quite similar, low, close 478 together, green-colored: P. mauritianus 2'. Ribs with saddle-shaped silica bodies; IZ with solitary cork-cells; marginal teeth: pointed tip, higher, irregular, the 479 largest (64-120 µm) separated by several small teeth: 480 481 3. Ribs with many large prickles; marginal teeth few colored, the largest (< 290 µm) few spaced: P. karka 482 3'. Ribs with rare smaller prickles; marginal teeth green, irregular, the largest (290-450 µm) well-spaced: P. frutescens 483 1'. Ribs with smaller (< 60µm) short-pointed prickles; IZ without microhairs, rare cork-cells and many hooks; clade II 484 4. Rib with blunt tip prickles, flat silica bodies; IZ with many short-cell pairs; marginal teeth green, the largest (0.5-1 mm). 485 with obvious basal internal cells, well-spaced: P. australis subsp. altissimus 486 4'. Rib prickles with a small terminal tip (< 8µm), thick silica bodies; IZ with rare short-cell pairs; marginal teeth not or few colored, the largest shorter, without obvious basal internal cells, less spaced: small reeds 487 488 5. IZ with small hooks: P. australis subsp. australis (AA and AE) 489 5'. IZ with many large hooks: P. japonicus and P11 (AA).

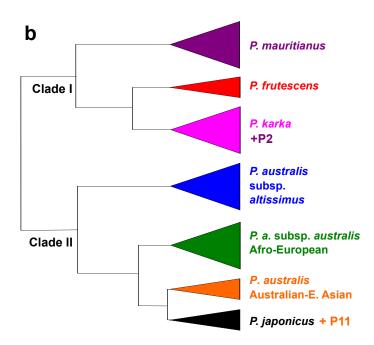
491	Box 3: Key of leaf anatomy characters (specific traits in italics) (Figure 5)
492 493	1. Blade thin (< 190 μm, on primary ribs); bulliform cells (BC) small, penetrating ca. 1/2 of the blade, without underlying colorless cells; all bundles circular:
494	2. Blade undulated, BC semicircular (ø < 70 µm), straight sided; small hooks: Afro-European P. australis (AE)
495	2'. Blade nearly flat, BC wider than deep (90 x 70 μm); many obvious hooks: P. japonicus
496	1'. Blade mostly thicker (> 190 µm); BC deeper than wide, penetrating ca. 2/3 of the blade:
497	3. BC (45-60 x 70-95 μm); blade undulated, abaxial face without hooks.
498	4. BC with obvious underlying colorless cells, straight sides; primary bundles elliptic: P. karka
499	4'. BC with inconspicuous underlying colorless cells, sides flared toward the top; bundles circular: P. mauritianus
500	3'. BC bigger, sides flared toward the top, with inconspicuous colorless cells:
501	5. Blade flat, without hooks; BC (75-96 x 100-140 µm) protruding in upper part; all bundles circular: P. frutescens
502	5'. Blade undulated, with small hooks; BC not protruding; primary bundles elliptic:
503	6. Blade thick (220-280μm), well undulated; BC big (70-96 x 85-110 μm): <i>P. australis</i> subsp. <i>altissimus</i>
504	6'. Blade thinner (170-230μm), few undulated; BC smaller (50-65 x 65-90 μm): Australian-E. Asian <i>P. australis</i> (AA)

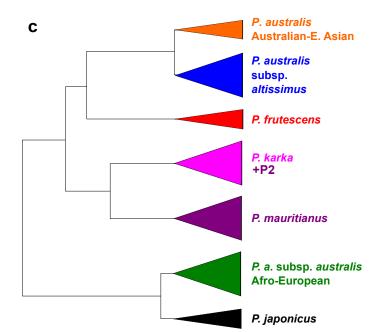
The genus *Phragmites* includes taxa that are very difficult to distinguish macroscopically, and even genetically. To better discriminate these taxa, microscopic traits, such as the epidermis of floral bracts and leaves, and leaf anatomy, were investigated. In the Old World, four distinct giant reeds species (> 3 m) were delineated: two tropical species, P. karka and P. mauritianus, and two Mediterranean species, P. altissimus and P. frutescens. Previously included within P. australis sensu lato, the rehabilitation of P. altissimus and P. frutescens is supported by relevant distinctive characteristics. Conversely, the small reeds (< 3 m) from temperate to cold regions form a distinct group. Although this group is still difficult to subdivide with epidermis traits, three taxa can be separated using leaf anatomy: both the Afro-European and the Australian-E. Asian P. australis sensu stricto, and P. japonicus. Using iodine green solution, we link our morphological analyses with previous genetic clustering, the green coloration of floral bracts and margin leaf teeth matches with the presence of nuclear waxy gene bands in P. frutescens and P. mauritianus (100 bp), and P. altissimus (200 bp), while P. karka, P. australis subsp. australis and P. japonicus have no coloration nor waxy band. While leaf anatomy features seem to be correlated with eco-climatic conditions, the epidermal traits delimited other clades. For leaf epidermis P. karka, P. mauritianus, and P. frutescens present the more ancestral characters, whereas P. australis, P. japonicus, and P. altissimus possess derived traits. Lastly, floral bract epidermis traits separate two divergent evolutionary lineages from the basal *P. karka*.

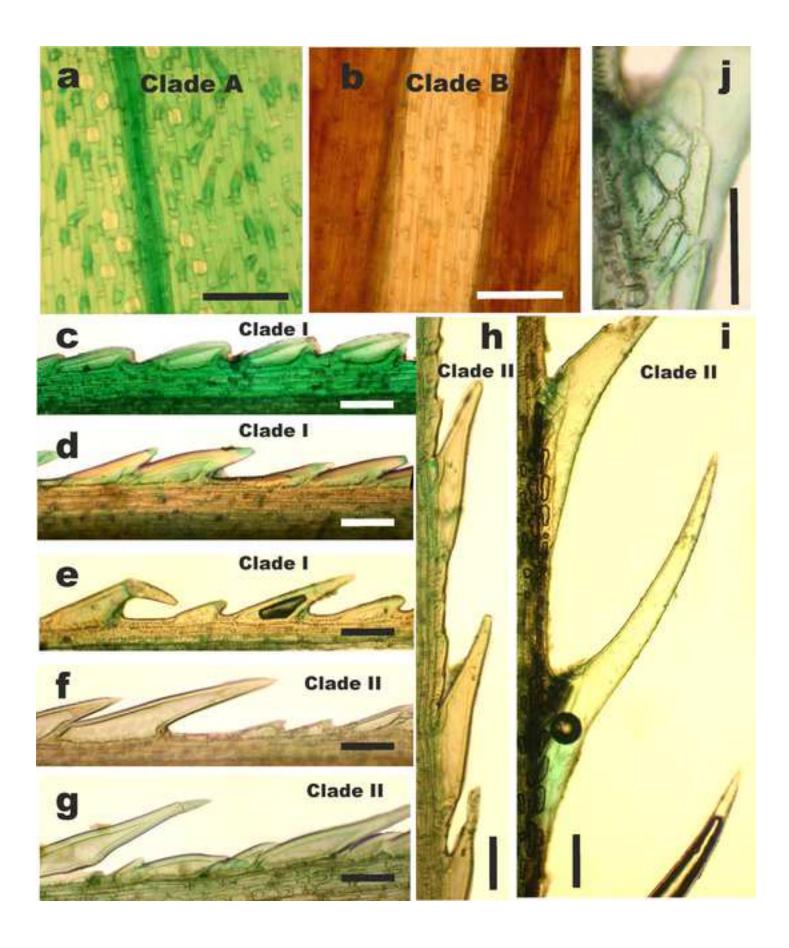












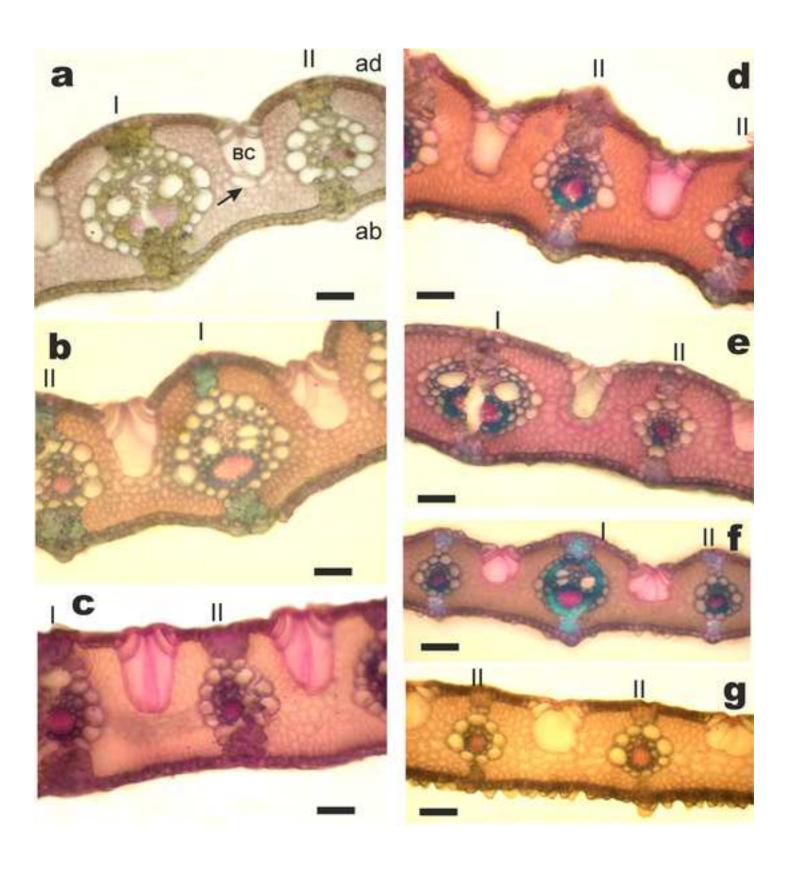


Table 1. Summary of genetic and phylogeographic groups of the Old World Phragmites taxa.

Taxa	AFLPs groups <sup>a</sup>	cpDNA groups <sup>b</sup>	cpDNA haplo-types b, c	Waxy bands b
Afro-European P. australis	P. australis core group	European Phragmites	L1, M, O1,2, AD, AH, AI, AJ, AK, K	
Australian-E. Asian P. australis	P. australis 8x Australia-Asia	Far East - Australian Phragmites	P <sup>1,2</sup>	
P. karka	P. vallatoria	Tropical Phragmites (P. karka)	$\mathrm{U}^1$	
P. mauritianus	P. mauritianus	Tropical Phragmites (P. mauritianus)	$\mathbf{I}^1$	100 bp
P. frutescens	P. frutescens	Tropical Phragmites (P. frutescens)	$\mathrm{I}^{1},\mathrm{U}^{1}$	100 bp
P. altissimus	P. australis Africa	Mediterranean Phragmites	$M^1$ , $AL$	200 bp
P. japonicus	P. japonicus	P. japonicus	AM	

 <sup>&</sup>lt;sup>a</sup> Lambertini et al., 2006;
 <sup>b</sup> Lambertini et al., 2012;
 <sup>c</sup> Lambertini et al., 2020
 <sup>l</sup> includes cpDNA microsatellite variants (ex. M1, M2, M3, M4 Lambertini et al., 2012)
 <sup>e</sup> includes Haplotype O-related and Haplotype P-related haplotypes (Lambertini et al., 2020)