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Régine Verlaque, Laurent Hardion, Carla Lambertini, Kim Canavan, Marc Verlaque, Bruno Vila

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New highlights on Old World giant *Phragmites* (Poaceae) using leaf and floral bract 1 microscopic characters 2 R. Verlaque^a, L. Hardion^b, C. Lambertini^c, K. Canavan^d, M. Verlaque^e, B. Vila^a. 3 ^a Aix Marseille University, IRD, LPED, Marseille, France 4 ^b University of Strasbourg, CNRS, Laboratoire Image Ville Environnement, UMR 7362, 5 6 Strasbourg, France 7 ^c Department of Bioscience, University of Milan, via Celoria 26, 20133 Milan ^d Centre for Biological Control, Department of Zoology and Entomology, Rhodes University, 8 Makhanda, South Africa 9 ^e Aix Marseille University and Université de Toulon, CNRS, IRD, Mediterranean Institute of 10 Oceanography (MIO), UM 110, Marseille, France 11 12 13 14 Keywords: Systematics, Epidermis, Anatomy, Reed, Micromorphology 15

16 **1. Introduction**

The genus *Phragmites* Adans. (Poaceae) has a worldwide distribution from the tropics to the 17 cold-temperate regions. Its positive attributes as an ecosystem engineer in wetlands provide 18 19 essential ecological and economic services, such as wildlife habitat, erosion control, and biomass production (Saltonstall and Meyerson, 2016). All species form dense stands, playing a 20 crucial role as carbon sinks over a long-time scale (Brix et al., 2001), and they are also known 21 22 for their phytoremediation abilities. Like other tall grasses, *Phragmites* spp. have followed 23 humans since the Paleolithic period due to their countless uses, e.g. energy source, construction, fodder, weapons, music, fiber and medicine (Täckholm et al., 1973; Oyen, 2011). Due to their 24 25 usefulness, they were introduced into many adventive ranges, like the reed genus Arundo L. (Hardion et al., 2014). The cosmopolitan P. australis (Cav.) Trin. ex Steud. has become a 26 27 "model organism for plant invasions" driven by its competitive abilities, particularly in North America (Pyšek et al., 2019). 28

Although *Phragmites* are the subject of an extensive number of articles, their taxonomy remains unclear. Thereby, the species number is widely debated, from only one species *P*. *australis* (Täckholm et al., 1973, under the name *P. communis* Trin.) to seven only for Corsica Island (Mabille, 1869) and for Russia (Tsvelev and Probatova, 2019), and up to 17 in the world (Steudel, 1854). In recent studies (Clevering and Lissner, 1999; Scholz and Böhling, 2000; Lambertini et al., 2006), the most cited taxa are *P. australis* (cosmopolitan), *P. karka* (Retz.)

- 35 Trin. ex Steud. (pantropical, except the American continent), *P. mauritianus* Kunth (tropical
- 36 Africa, Madagascar, Mascarene Islands), *P. japonicus* Steudel (N-Japan, E-Russia, NE-China),
- and *P. frutescens* Scholz (Crete, Greece, SW-Turkey).

The poor taxonomic resolution of *Phragmites* results from many overlaps of quantitative 38 macroscopic traits, and the ambiguity of descriptive terms. For example according to Conert 39 (1961) or Clayton (1970), the upper glume is either lanceolate or acute in *P. karka*, and either 40 ovoid or acuminate in P. mauritianus. Long considered as distinct species, Clayton (1970) 41 42 suggests a subspecific status for P. mauritianus within P. karka, while some floras simply include P. mauritianus within P. karka, without any infraspecific rank (e.g. Phillips, 1995). 43 Moreover for Veldkamp (1992), P. karka should be named P. vallatoria (Plunk. ex L.) Veldk., 44 a name now considered as invalid (Kirkbride and Wiersema, 2007). 45

Surprisingly, despite its morphological differentiation, online taxonomic databases (*e.g.* Kew,
Paris) place *P. frutescens* in synonymy within *P. australis*. Described in 1996, *P. frutescens* is
a rare species with infrequent flowering, but morphologically more related to *P. mauritianus*than to *P. australis* (Scholz and Böhling, 2000), as confirmed by AFLP analyses (Lambertini
et al., 2006: Fig. A1). In fact, the three taxa, *P. karka, P. mauritianus, P. frutescens*, form a
distinct genetic unit, with two unique cpDNA haplotypes (U, I:Table 1).

In his monograph, Conert (1961) lists 11 varieties within *P. australis*, while Clayton (1970) 52 recognizes only two taxa: the temperate subsp. *australis* (< 3 m) and the Mediterranean subsp. 53 altissimus (Benth.) Clayton (height and panicles taller). These two subspecies are adopted by 54 Feinbrun-Dothan (1986), but many floras include subsp. altissimus within P. australis (e.g. 55 Tutin, 1980; Fish et al., 2015), or place it under various names and ranks, such as var. isiaca 56 (Del.) Coss. (Täckholm et al., 1973; Hultén and Fries, 1986), but rarely as a species [Coste, 57 1937, as Arundo maxima Forsk.; Tsvelev and Probatova, 2019, as P. altissimus (Benth.) Mab.]. 58 All genetic studies separate this taxon from the subsp. *australis*, but the individual groupings 59 differ according to methods and sampling, (AFLPs, haplotypes, SSRs: Lambertini et al., 2006, 60 61 2012; RAD-sequencing: Wang et al., 2021).

Finally, according to genetic studies, *P. japonicus* is separate from the *P. australis* subsp. *australis* (hereafter *P. australis s.str.*), which includes two distinct phylogeographic groups: the Afro-European unit (AE) and the Australian-E. Asian unit (AA) (Lambertini et al., 2006, 2012, 2020). In America, there are three native genetic units that highly differ from all Old World taxa, based on AFLPs, haplotypes, microsatellites (Lambertini et al., 2006: Fig. A1; Saltonstall and Meyerson, 2016) and RAD-sequencing (Wang et al., 2021).

In the absence of reliable macroscopic characters in Phragmites, we investigated the 68 microscopic traits of leaves and floral bracts. The high phylogenetic value of leaf epidermis and 69 anatomy traits is known since the founding work of Prat (1932) and has proven to be valuable 70 for the systematics of Poaceae (Watson et al., 1992; Kellogg, 2015), and for the identification 71 of South African Phragmites (Gordon-Gray and Ward, 1971). Such traits can be useful when 72 phylogenetic markers fail to resolve complex grass groups. The traits of floral bracts were also 73 found to have high discriminant value, phylogenetic significance (Liu et al., 2010; Hardion et 74 al., 2021), and are comparatively easy to use. Based on extensive sampling, including plants 75 76 previously analyzed in molecular studies, our aim here was to test the value of micromorphologic characters to distinguish Old World Phragmites, especially the problematic giant 77 78 reeds.

79

80 **2. Materials and Methods**

For a representative cover of the global distribution of taxa in the Old World, specimens from 81 several international herbaria, Aix-Marseille University collections, and twenty samples 82 83 previously studied in molecular analyses were investigated. The study was performed on 160 specimens of Phragmites [35 P. karka, 30 P. mauritianus, 11 P. frutescens, 44 P. australis 84 subsp. altissimus, 31 Afro-European (AE) P. australis subsp. australis, 5 Australian-E. Asian 85 (AA) P. australis, 4 P. japonicus], and one Molinia caerulea (L.) Moench., the sister group of 86 Phragmites (Hardion et al., 2021). All information on specimens are reported in the table A1. 87 Prior identifications were based on macroscopic characters observed on herbarium specimens, 88 except for the genetic unit AA P. australis for which no distinctive macro-morphological 89 character is yet known. These traits were chosen from original diagnosis, from some floras 90 (Coste, 1937; Clayton, 1970; Feinbrun-Dothan, 1986), and from the synthesis of Scholz and 91 Böhling (2000). 92

Courtesy of herbarium curators, we collected four mature spikelets on each sample, and a 93 small piece from the middle part of the two terminal leaves of the primary axis, because they 94 possess the most differentiated epidermis (Prat, 1932). To avoid damages on type specimens, 95 two spikelets (taken in the bag of fragments) and an imprint of leaf epidermis with clear nail 96 polish were taken. The material was re-hydrated in distilled water for ca. one week. For each 97 spikelet, glumes, lemmas and paleas of the two first basal flowers were separated by cutting at 98 99 their base. For each leaf, the adaxial epidermis was prepared by scraping away the abaxial face and mesophyll cells with a razor blade. Floral bracts, leaf margin and epidermis were stained 100

with iodine green solution for 3 min and mounted in glycerol. For anatomy, about 10 transversal
sections per leaf were performed in lateral part (out of the central rib), cleared with sodium
hypochlorite, washed in distilled water, stained with carmine green and mounted in glycerol.

104 In the spikelet, only lemmas and paleas, which are more differentiated than glumes in reeds, 105 were studied in detail. Classical discriminant traits were studied, as well as several new characters, such as marginal teeth of leaves and paleas, and floral bract coloring. A total of 56 106 characters were retained: 23, 12 and 21 for the leaf epidermis, anatomy, and spikelet, 107 respectively (Table A2). The micromorphological terminology follows Prat (1932) and its 108 revisions (Metcalfe, 1960; Watson et al., 1992; ICPT, 2019). All specimens were studied using 109 a light microscope (LM; ca.10 observations for each trait), and some samples were observed 110 under scanning electron microscope (SEM). All traits were coded as discrete states (Table A2a-111 c). Before each analysis, datasets were converted into binary variables, with one new variable 112 113 for each state of initial variables, using the dummies R-package in the R environment (R Core Team, 2022). Data were then analyzed in correspondence and hierarchical clustering analyses 114 115 with Jaccard binary dissimilarities using the *ade4* R-package (Thioulouse et al., 2018).

116

3. Results

Based on 21 epidermal characters of lemmas and paleas, the hierarchical clustering analysis 118 (HCA) highlights two main clusters (Fig. 1a). The clade A strictly includes the African and 119 Mediterranean P. mauritianus, P. frutescens and P. australis subsp. altissimus, while the clade 120 B includes the remaining taxa: P. karka, P. japonicus and P. australis s.str. The clade A mainly 121 differs from clade B by glumes and lemmas being green and spread out, vs brownish and rolled 122 up (Fig. 2a vs 2b), and lemma hooks larger and pointed to all directions, vs small (<30 µm) and 123 pointed to the top (Fig. 3c-d vs 3a-b). In addition, within each clade, palea characters distinguish 124 the different taxa (Fig. 3g-h and e-f). The correspondence analysis (CA) clearly confirms these 125 two clades, with the first three axes representing 59.8% of the total variance (26.4%, 12.5% and 126 127 10.9%, respectively; Fig. A2a). The main distinctive traits of the two clades A and B, and taxa are summarized in a key (Box 1). 128

The HCA of the 23 leaf epidermal characters also highlights two main clusters that mainly match with previous ones, excepted for the inverted position of *P. australis* subsp. *altissimus* and *P. karka* between the two clades (Fig. 1b). The clade I includes the three highest (up to 8 m) and thermophilic species, *P. karka, P. mauritianus,* and *P. frutescens*, while the clade II includes the more temperate and smaller taxa, *P. australis* subsp. *altissimus,* subsp. *australis,*

and P. japonica. The clade I differs from clade II in its simpler epidermal structure: ribs with 134 large base and long-pointed prickles (vs small in clade II), intercostal zones with bicellular 135 microhairs (vs absent), many cork-cells (vs rare), rare hooks (vs frequent) (Fig. 3i-j vs 3k-m), 136 and weakly ornate leaf margin (vs more ornate) (Fig. 2c-e vs 2f-j). In our preparations, the 137 iodine green staining of floral bracts lasts several years, but that of leaf teeth fades after a few 138 weeks. The CA confirms the two clades I and II, with the three first axes representing 78% of 139 the total variance (54.1%, 14.9% and 9%, respectively; Fig. A2b). The main traits are reported 140 in the Box 2. 141

The coupled analysis of floral and leaf epidermis shows a better discrimination of taxa (Fig.
4), notably for *P. frutescens*, the Afro-European (AE) and Australian-E. Asian (AA) *P. australis*. The three first axes of the CA represent 50.6% of the total variance (24.2%, 15.6% and 10.8%, respectively). The two S-African hybrids genetically attested (Canavan et al., 2018),
KC2 (P. *mauritianus* x AE *P. australis*), and KC3 (*P. mauritianus* x *P. altissimus*), have a mix of parental traits, and the CA placed them far from their parents, especially from *P. mauritianus*.

Leaf anatomy shows the main structural characteristics of *Phragmites*, especially the shape and size of bulliform cells (BC), with the round bottom forming a gutter of regular adaxial groups between the vascular bundles (Fig. 5). The CA separates the giant reed taxa (> 3m) from the smaller ones along the axis 1, as well as the different taxa of each group (Fig. 6) and shows the polymorphism of AA *P. australis*. The three first axes represent 66% of the total variance (32%, 22% and 12%, respectively). The anatomical key based on the HCA distinguishes all the Old World taxa (Box 3).

155 **4. Discussion**

156 *4.1. Taxonomy*

The objective of this study was to delimit well supported Old World Phragmites taxa. As 157 expected, microscopic characters have more discriminant resolution than macroscopic ones, 158 and shed a new light on this problematic genus. Both leaf and spikelet traits show a strong 159 individualization of four giant taxa: P. mauritianus, P. karka, and more surprisingly P. 160 frutescens and P. australis subsp. altissimus. Given the stability and the high taxonomic value 161 of these micro-characters, we consider them as four distinct species. For the subsp. *altissimus*, 162 which possesses highly specific traits, the most appropriate name should be *P. altissimus* 163 164 (Benth.) Mabille (1869).

The floras of S-Africa only report *P. australis* and ignore *P. altissimus* (Fish et al., 2015), 165 while both taxa co-exist. Our results show that the S-African "P. australis" illustrated by 166 Gordon-Gray and Ward (1971), and genetically studied (Canavan et al. 2018) is in fact P. 167 altissimus. Our sampling does not show obvious micro-characters differences between S-168 African and Mediterranean P. altissimus, in agreement with DNA microsatellites analyses 169 (Lambertini et al., 2012). Within *P. australis s.l.* of the Danube Delta, Hanganu et al. (1999) 170 found two distinct taxa called "giant reeds" (2n = 96), and "fine reeds" (2n = 48), which likely 171 correspond to P. altissimus and P. australis s.str., respectively. As suggested by the map of 172 Hultén and Fries (1986) and the presence of the exclusive 200-bp waxy band (Lambertini et al., 173 2012), the distribution of P. altissimus is larger than expected, including African, 174 Mediterranean, and Middle East regions up to Armenia and Kashmir. Phragmites frutescens is 175 sometimes sympatric with P. altissimus and morphologically close to P. mauritianus, but it is 176 restricted to the Mediterranean Basin, and it presents distinct specific traits, especially in leaf 177 anatomy. 178

Unlike the giant reeds, the taxonomy of Old World small reeds (< 3 m), *P. japonicus* and *P. australis s.str.* (AE, AA), appears more challenging. Except leaf anatomy, the other characters
distinguish these three taxa less clearly than genetic studies (Lambertini et al., 2006, 2020; Liu
et al., 2020; Wang et al., 2021). Further investigations are needed, based on a broader sampling
in Australia and Asia, to clarify their morphological distinction and geographic distribution.

184 *4.2. Anatomy*

Leaf anatomy shows the most discriminant traits, particularly the bulliform cells (BC). The 185 huge flabellate reed BC are unique among Poaceae (Chen et al., 2020), and called "Phragmites-186 type" as wetland markers in paleoecology. These are the only traits that easily separate P. 187 188 frutescens (BC protruding) and P. japonicus (BC wider than deep) from all others. Anatomical analyses produce original groupings, and clearly distinguish three taxa within Old World P. 189 australis s.str. (Box 3). The two temperate to cold small reeds (AE P. australis, and P. 190 japonicus) possess thin leaves, and small fan-shaped BC, penetrating ca. 1/2 of the blade. In 191 contrast, thick leaves and deep BC, penetrating ca. 2/3 of the blade, separate the thermophilic 192 taxa (the giant reeds and AA P. australis) from the first two taxa. This group includes the 193 tropical pair P. karka - P. mauritianus, and the three other subtropical taxa: the Mediterranean 194 pair P. altissimus - P. frutescens, and the humid subtropical AA P. australis. 195

In Poaceae, Cyperaceae and Juncaceae, BC have a crucial role in leaf rolling, and water
storage, in response to drought and salt stress. All *Phragmites* live in wetlands, but AE *P. australis* has a better salt tolerance than AA *P. australis* (Lambertini et al., 2020), and *P. altissimus* (Hanganu et al., 1999). Therefore, the strong anatomical differentiation of *Phragmites*taxa could be an adaptation to eco-climatic conditions.

201 *4.3. Staining and waxy genes*

In iodine green solution, floral bracts, and leaf margin teeth of *P. frutescens, P. mauritianus* and *P. altissimus* turned green, while those of other taxa remain unstained. These results are consistent with the banding pattern of the waxy genes: green colour corresponds to the presence of 100 bp band in *P. mauritianus* and *P. frutescens*, 200 bp band in *P. altissimus*, and unstained parts match to the lack of waxy bands in the other taxa (Lambertini et al., 2012; Canavan et al., 2018).

Waxy plants, notably cereals, are of great interest for their countless industrial uses. Iodine 208 solutions are used to detect waxy genes, they colour the starch of pollen and endosperm 209 differently: waxy plants rich in amylopectin turn reddish, whereas non-waxy plants with only 210 amylose turn blue (Bates et al., 1943). If in herbarium specimens, these organs are rarely in 211 good condition, our results show that the spikelet and leaf staining can easily detect waxy plants. 212 Similar observations were cited for waxy rice leaves (Abé, 1985). This phenomenon might 213 result from the relationship between the presence of amylopectin and a better permeability of 214 cell walls (Michailova et al., 2001). The spreading out or rolling up of glumes and lemmas in 215 216 glycerol can also be explained by starch types: water soluble amylopectin, hydrophilic with alcohol, would cause spreading out bracts, while water insoluble amylose, hydrophobic with 217 218 alcohol, would induce rolling up bracts. Waxy rice (glutinous) has been used for millennia in Asia, due to its better digestibility, early ripening, and tolerance to temperature variation and 219 220 drought (Abé, 1985). Waxy corn also has a high resistance to pathogenic fungi (Walden, 1978), which can explain our leaf observations: taxa without waxy genes are often infected with rusts, 221 222 while waxy taxa are almost always healthy. Biochemical differences could also explain the distinct uses of species, e.g. in African traditional medicine (Oyen, 2011). 223

4.4. Ploidy levels and hybrids

In karyological studies of *Phragmites*, the authors considered 2n = 48 as the tetraploid level, with a basic chromosome number n = 12, even if n = 6 is more likely for the Arundinoideae, particularly for *Molinia* spp. with 2n = 36 (Conert, 1961; Hardion et al., 2011). According to the main syntheses (Gorenflot, 1976; Connor et al., 1998; Clevering and Lissner, 1999; Ishii and Kadono, 2001; Hansen et al., 2007), the *Phragmites* include: (1) four stable species with 2n = 48: *P. karka, P. mauritianus, P. frutescens* and *P. japonicus;* (2) the AE *P. australis* with 2n = 48 (rarely 2n = 96); and (3) two greatly variable taxa: AA *P. australis* with 2n = 96 (120, 264), and *P. altissimus* with 2n = 48 (72) in Senegal and S. Europe, 96 (144) in S. Africa, S. Mediterranean, Danube Delta, and Middle East.

Some herbarium samples were difficult to identify, as evidenced by the various names 234 attributed by different botanists (Table A1). Notably, two plants previously named "P. karka" 235 showed mixed characters: (i) the Cameroonian P2 with spikelet traits of P. mauritianus and, 236 leaf epidermis and anatomy of P. karka, and (ii) the Japanese P11 with spikelet traits and 237 anatomy of AA P. australis and leaf epidermis of P. japonicus. Both could be hybrids of these 238 239 close related taxa, or rather the expression of strong polymorphism, because the true S-African hybrids KC2 (P. mauritianus x AE P. australis: 100 bp band), and KC3 (P. mauritianus x P. 240 altissimus: 100-200 bp bands), get away from their parent taxa in CA. In hybridizations between 241 waxy and non-waxy taxa (P2: P. mauritianus x P. karka, or KC2), floral bracts and leaf margins 242 243 always turned green. This could explain some hybrids with 200 bp waxy band (S-Africa, Danube Delta), but related to AE P. australis in some genetic traits (Lambertini et al., 2012; 244 Canavan et al., 2018). 245

The proportion of natural hybrids is challenging, given the strong polymorphism of reeds due 246 247 to the high polyploidy resulting from a reticulated evolution (Lambertini et al., 2012; Liu et al., 2020; Wang et al., 2021). Like most polyploids (Richard, 1997), Phragmites spp. have weak 248 249 genetic barriers that explain the occurrence of some hybrids. Based on a large P. japonicus (2n 250 = 48) and AA P. australis (2n = 96, 120) sampling, Ishii and Kadono (2001) argue that hybrids 251 are very rare due to: (i) mismatched flowering periods between species, (ii) temporal 252 reproductive isolation related to the short flowering period, (iii) short distance of anemophilous pollination, and (iv) distinct ploidy levels without intermediate numbers. According to Scholz 253 and Böhling (2000) and many floras, clear asynchronous flowering of sympatric taxa applies to 254 all Phragmites, e.g. the early flowering of P. mauritianus vs P. karka, AE P. australis vs P. 255 altissimus, and AA P. australis vs P. japonicus. In addition, the pollen of Mediterranean "P. 256 *australis*" hybrids (2n = 72) is entirely sterile (Gorenflot et al., 1990). Lastly, in the wild, hybrid 257 formation is also strongly limited by the almost exclusive vegetative growth of dense long-lived 258 clones (Clevering and Lissner, 1999), often distinct geographic areas and eco-climatic 259 preferences (geo-climatic barriers; Lambertini et al., 2012). 260

261 *4.5. Evolution and biogeography*

Micromorphological studies are not only descriptive, but they can also be used in phylogeny 262 (Liu et al., 2010). Floral bract and leaf epidermis characters define different clades, implying 263 that each category of traits has a distinct evolutionary significance. The main leaf traits of the 264 clade I (P. karka, P. mauritianus, P. frutescens) are (i) ribs with huge prickles and oblong silica-265 266 bodies, and (ii) intercostal zone without hooks, but with straight wall long cells, many bicellular microhairs and cork-cells. Comparisons of descriptions and illustrations (Metcalfe, 1960; 267 Watson et al., 1992) show that these traits are also present in the earliest groups of the BEP 268 clade in the phylogenetic classification of Poaceae (Soreng et al., 2017), *i.e.* the first three basal 269 tribes of the Oryzoideae. The Phragmites share other characters with ancestral states, since this 270 genus is the only one of the PACMAD clade having arm cells in the mesophyll, like the basal 271 272 Oryzoideae and Bambusoideae (Kellogg, 2015).

The species of the clade I (P. karka, P. mauritianus, P. frutescens) form a morphological unit 273 characterized by their height (8-10 m), large panicles, deciduous leaf sheaths, and leaf blades 274 275 that are rough to the touch. Epidermis comparison between Phragmites and its sister group 276 Molinia (Hardion et al., 2021) shows that M. caerulea shares a maximal similarity with P. karka: 85 and 78% for flower and leaf traits, respectively. Thus, P. karka retains the most 277 278 number of ancestral traits across *Phragmites* spp., in accordance with its basal position in the AFLP tree (Lambertini et al., 2006: Fig. A1). In addition, P. karka and P. mauritianus have a 279 280 large distribution in tropical warm riparian habitats, like the earliest Poaceae (Kellogg, 2015). Opposingly, the taxa of clade II (P. altissimus, P. australis s.str., and P. japonica) exhibit 281 282 derived traits and are distributed in subtropical and temperate to cold zones. On the other hand, spikelet traits draw on two other evolutionary lineages, divergent from P. karka with an 283 284 increasing differentiation. The clade B includes the non-waxy taxa (brownish bracts), with lightly ornamented lemma and palea: P. karka, AE P. australis, P. japonicus (2n = 48), and 285 lastly AA P. australis (2n = 96). The clade A includes the waxy taxa (green bracts), with 286 increasingly ornate paleas: P. mauritianus, P. frutescens (2n = 48), and P. altissimus (2n = 48), 287 96). 288

In conclusion, we have highlighted several valuable micro-characters allowing for the identification of four Old World giant *Phragmites*, which match well with the previous genetic clustering. Future investigations will focus on the small reeds, particularly those from Asia and Australia, and the macroscopic characters of Old World *Phragmites*.

293 CRediT authorship contribution statement

R. Verlaque: Conceptualization, species identification, morphological investigation, and writing. L.
Hardion: Data analyses, figures, and writing. K. Canavan, C. Lambertini and B. Vila: Field work,
documents, and writing. M. Verlaque: tables, figures, and writing.

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303 Appendix A. Supporting information

304 Supplementary data associated with this article can be found in the online version at

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423 Legends of figures

- 424 Figure 1: Hierarchical clustering analyses: 1a. Floral bract epidermis; 1b. Leaf epidermis; 1c.
- 425 Leaf anatomy. Clade A: Floral bracts green and spread out; Clade B: Floral bracts brownish
- 426 and rolled up; Clade I: ribs with large prickles and bicellular microhairs; Clade II: ribs with
- 427 small prickles, without bicellular microhairs.
- Figure 2: Lemmas. (a) Clade A, green and spread out: *P. mauritianus* (B24); (b) Clade B,
 brownish and rolled up: *Phragmites karka* (P9). Leaf margin: teeth rounded tipped, similar,
 low: (c) *P. mauritianus* (L1); (d-i): teeth large and small, pointed, erect, and more spaced: (d)
- 431 P. karka (B11); (e) P. frutescens (C196); (f) P. japonicus (C177); (g) Afro-European P.
- 432 *australis* (MS24); (h) Australian-E-Asian *P. australis* (MP7); (i) *P. australis* subsp. *altissimus*:
- 433 (MS26); (j) detail of basal internal ornament of large teeth (MS20). Scale = $100 \mu m$.
- 434 Figure 3: Lemma epidermis, Clade B, hooks pointed to the top: (a) *Phragmites karka* (B13),
- (b) AE *P. australis* (MS27). Clade A, hooks pointed in all directions: (c) *P. mauritianus* (B24),
- 436 (d) P. australis subsp. altissimus (MS1). Palea apex, Clade B, few ornamented: (e) P. karka

- (B13); (f) *P. japonicus* (MP10, similar in AE *P. australis*). Clade A, well ornamented, large
 teeth pointed in all directions: (g) *P. mauritianus* (B24); (h) *P. australis* subsp. *altissimus*(MS2): entire palea with marginal long hairs. Leaf characters (abaxial face): Clade I, ribs with
 large prickles and bicellular microhairs: (i) *P. karka* (B39, similar in *P. mauritianus*); (j) *P.*
- 441 frutescens (MS8) smaller prickles. Clade II, ribs with small prickles, without bicellular
- 442 microhairs: (k) AE *P. australis* (MS29, similar in AA); (l) *P. japonicus* (C177) many obvious
- 443 hooks; (m) P. australis subsp. altissimus (C174) blunt prickles. R = rib; p = prickle; b =
- bicellular microhair; h = hook. Scales: $(a-g) = 40 \ \mu m$; $(h) = 250 \ \mu m$; $(i-m) = 20 \ \mu m$.
- Figure 4: Hierarchical clustering analysis of floral bracts and leaves epidermal characters.
 Hybrids: KC2 (*P. mauritianus* x AE *P. australis*) and KC3 (*P. mauritianus* x *P. altissimus*).
- 447 Figure 5: Leaf anatomy (transversal section). (a) *Phragmites karka* (P6); (b) *P. mauritianus*
- 448 (P14); (c) P. frutescens (MS6); (d) P. australis subsp. altissimus (MP16); (e) Australian-E.
- Asian P. australis (MS16); (f) Afro-European P. australis (MS13); (g) P. japonicus (C177).
- 450 BC = bulliform cells, ad = adaxial face, ab = abaxial face, arrow = obvious underlying colorless
- 451 cells, I = primary vascular bundles (elliptic or circular), separated by 3-4 secondary bundles
- 452 (=II, always circular). Scale = $50 \mu m$.
- **Figure 6:** Hierarchical clustering analysis of leaf anatomic characters.
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455

- **Box 1**: Key of spikelet epidermal characters.
- **Box 2**: Key of leaf epidermal characters.
- **Box 3**: Key of leaf anatomy characters.

460	Box 1: Key of spikelet epidermal characters (specific traits in italics)			
461	1. Glumes and lemmas green and spread out*; lemma hooks: many large (> 30µm), pointed in all directions: clade A			
462 463	2. Paleas densely ornate: margins with large teeth (70-140µm), pointed in all directions, and some long hairs (up to 1.5 mm); apex retuse with many hooks: P. australis subsp. altissimus			
464	2'. Paleas less ornate: margins with smaller teeth (<100 μ m), and without long hairs			
465	3. Hooks of lemmas and paleas large (30-150µm); palea apex bifid: <i>P. mauritianus</i>			
466	3'. Hooks and margin teeth of lemmas and paleas smaller (<40µm); palea apex retuse-truncated: P. frutescens			
467	1'. Glumes and lemmas brownish and rolled up*; lemma hooks: smaller (< 30µm) and pointed to the top: clade B			
468 469	4. Lemmas with straight long-cell walls and many hooks; palea: margins with small teeth (< 50 μm), veins stopped long before the top, apex hyaline generally rounded: P. karka			
470 471	4'. Lemmas with sinuous long-cell walls and <i>rare hooks</i> ; palea margins with larger teeth (> 50µm), veins going to the top, apex not hyaline, generally truncated: small reeds including <i>P. australis</i> subsp. <i>australis</i> (AE, AA) and <i>P. japonicus</i> .			
472	* After staining with iodine green and mounting in glycerin.			
473				
474	Box 2: Key of leaf epidermal characters (specific traits in italics)			
475 476	1. Ribs with large (> 65 μm) long-pointed prickles; intercostal zone (IZ) with bicellular microhairs, cork-cells and rare hooks clade l			
477 478	2. Ribs with oblong silica bodies; IZ with superposed cork-cells; marginal teeth: rounded tip, quite similar, low, close together, green-colored: <i>P. mauritianus</i>			
479 480	2'. Ribs with saddle-shaped silica bodies; IZ with solitary cork-cells; marginal teeth: pointed tip, higher, irregular, the largest (64-120 μm) separated by several small teeth:			
481	3 . Ribs with many large prickles; marginal teeth few colored, the largest (< 290 μm) few spaced: <i>P. karka</i>			
482	3'. Ribs with rare smaller prickles; marginal teeth green, irregular, the largest (290-450 μm) well-spaced: P. frutescen			
483	1'. Ribs with smaller (< 60µm) short-pointed prickles; IZ without microhairs, rare cork-cells and many hooks: clade II			
484 485	4 . Rib with blunt tip prickles, flat silica bodies; IZ with many short-cell pairs; marginal teeth green, the largest (0.5-1 mm) with obvious basal internal cells, well-spaced: P. australis subsp. altissimus			
486 487	4'. Rib prickles with a small terminal tip (< 8µm), thick silica bodies; IZ with rare short-cell pairs; marginal teeth not or few colored, the largest shorter, without obvious basal internal cells, less spaced: small reeds			
488	5. IZ with small hooks: P. australis subsp. australis (AA and AE)			
489	5'. IZ with many large hooks: <i>P. japonicus</i> and P11 (AA).			
490				

491	Box 3: Key of leaf anatomy characters (specific traits in italics) (Figure 5)
492 493	1 . Blade thin (< 190 μm, on primary ribs); bulliform cells (BC) small, penetrating ca. 1/2 of the blade, without underlying colorless cells; all bundles circular:
494	2. Blade undulated, <i>BC semicircular</i> (ø < 70 μm), straight sided; small hooks: Afro-European <i>P. australis</i> (AE)
495	2'. Blade nearly flat, BC wider than deep (90 x 70 μ m); many obvious hooks: P. japonicus
496	1'. Blade mostly thicker (> 190 μ m); BC deeper than wide, penetrating ca. 2/3 of the blade:
497	3 . BC (45-60 x 70-95 μ m); blade undulated, abaxial face without hooks.
498	4. BC with obvious underlying colorless cells, straight sides; primary bundles elliptic: P. karka
499	4'. BC with inconspicuous underlying colorless cells, sides flared toward the top; bundles circular: P. mauritianus
500	3'. BC bigger, sides flared toward the top, with inconspicuous colorless cells:
501	5. Blade flat, without hooks; BC (75-96 x 100-140 µm) protruding in upper part; all bundles circular: P. frutescens
502	5'. Blade undulated, with small hooks; BC not protruding; primary bundles elliptic:
503	6 . Blade thick (220-280μm), well undulated; BC big (70-96 x 85-110 μm): <i>P. australis</i> subsp<i>. altissimus</i>
504	6', Blade thinner (170-230um), few undulated: BC smaller (50-65 x 65-90 um); Australian-E, Asian P. australis (AA)

The genus *Phragmites* includes taxa that are very difficult to distinguish macroscopically, and even genetically. To better discriminate these taxa, microscopic traits, such as the epidermis of floral bracts and leaves, and leaf anatomy, were investigated. In the Old World, four distinct giant reeds species (> 3 m) were delineated: two tropical species, *P. karka* and *P.* mauritianus, and two Mediterranean species, P. altissimus and P. frutescens. Previously included within P. australis sensu lato, the rehabilitation of P. altissimus and P. frutescens is supported by relevant distinctive characteristics. Conversely, the small reeds (< 3 m) from temperate to cold regions form a distinct group. Although this group is still difficult to subdivide with epidermis traits, three taxa can be separated using leaf anatomy: both the Afro-European and the Australian-E. Asian P. australis sensu stricto, and P. japonicus. Using iodine green solution, we link our morphological analyses with previous genetic clustering, the green coloration of floral bracts and margin leaf teeth matches with the presence of nuclear waxy gene bands in P. frutescens and P. mauritianus (100 bp), and P. altissimus (200 bp), while P. karka, P. australis subsp. australis and P. japonicus have no coloration nor waxy band. While leaf anatomy features seem to be correlated with eco-climatic conditions, the epidermal traits delimited other clades. For leaf epidermis P. karka, P. mauritianus, and P. frutescens present the more ancestral characters, whereas P. australis, P. japonicus, and P. altissimus possess derived traits. Lastly, floral bract epidermis traits separate two divergent evolutionary lineages from the basal P. karka.







Figure 1



P. frutescens

P. mauritianus

P. japonicus

P. a. subsp. *australis* Afro-European

P. karka +P2

P. austra@#ick here to access/download;Figure;Fig1.eps **±** subsp. *altissimus*





Taxa	AFLPs groups ^a	cpDNA groups ^b	cpDNA haplo-types ^{b, c}	Waxy bands ^b
Afro-European P. australis	P. australis core group	European Phragmites	L ¹ , M, O ^{1,2} , AD, AH, AI, AJ, AK, K	
Australian-E.Asian P. australis	P. australis 8x Australia-Asia	Far East - Australian Phragmites	P ^{1,2}	
P. karka	P. vallatoria	Tropical Phragmites (P. karka)	U^1	
P. mauritianus	P. mauritianus	Tropical Phragmites (P. mauritianus)	\mathbf{I}^1	100 bp
P. frutescens	P. frutescens	Tropical Phragmites (P. frutescens)	I^1 , U^1	100 bp
P. altissimus	P. australis Africa	Mediterranean Phragmites	M ¹ , AL	200 bp
P. japonicus	P. japonicus	P. japonicus	AM	

Table 1. Summary of genetic and phylogeogra	aphic groups of the Old World <i>Phragmites</i> taxa.
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^a Lambertini et al., 2006; ^b Lambertini et al., 2012; ^c Lambertini et al., 2020
¹ includes cpDNA microsatellite variants (ex. M1, M2, M3, M4 Lambertini et al., 2012)
² includes Haplotype O-related and Haplotype P-related haplotypes (Lambertini et al., 2020)