



HAL
open science

Facile multipoint nanoholes array-based plasmonic platform: Development, characterization, and sensor evaluation

Bento Pereira Cabral Júnior, Jean Halison de Oliveira, Paulo Henrique Maciel Buzzetti, Maiara Mitiko Taniguchi, Johny Paulo Monteiro, Luís Henrique Cardozo Amorin, Alexandre Urbano, Eduardo Radovanovic, Emerson Marcelo Giroto

► To cite this version:

Bento Pereira Cabral Júnior, Jean Halison de Oliveira, Paulo Henrique Maciel Buzzetti, Maiara Mitiko Taniguchi, Johny Paulo Monteiro, et al.. Facile multipoint nanoholes array-based plasmonic platform: Development, characterization, and sensor evaluation. *Materials Letters*, 2022, 332, pp.133526. 10.1016/j.matlet.2022.133526 . hal-03916044

HAL Id: hal-03916044

<https://hal.science/hal-03916044v1>

Submitted on 6 Feb 2024

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

1 Facile Multipoint Nanoholes Array-Based Plasmonic Platform: Development,
2 Characterization, and Sensor Evaluation

3 Bento Pereira Cabral Júnior^a, Jean Halison de Oliveira^a, Paulo Henrique Maciel Buzzetti^a,
4 Maiara Mitiko Taniguchi^a, Johny Paulo Monteiro^b, Luís Henrique Cardozo Amorim^c,
5 Alexandre Urbano^c, Eduardo Radovanovic^a, Emerson Marcelo Giroto^{a*}

6 ^a *Department of Chemistry, State University of Maringá, Maringá, PR, Brazil.*

7 ^b *Department of Chemistry, Federal Technological University of Paraná, Apucarana, PR,*
8 *Brazil.*

9 ^c *Department of Physics, State University of Londrina, Londrina, PR, Brazil*

10 **Correspondent Author. E-mail address: emgirotto@uem.br.*

11

12 **Abstract**

13 This study demonstrates a novel nanoholes array-based plasmonic platform with potential for
14 application in multi-detection analysis. The nanostructured transducer was obtained by
15 nanosphere lithography (NSL), which was restricted to having a sensing multipoint. The
16 microscopic characterization showed sensing point (Sen-P) arrays with *ca.* 380 μm and
17 nanoholes with 234.1 ± 27.3 nm diameter. Nanoholes array transmission spectra presented a
18 plasmonic band in *ca.* 531 nm when the refractive index (RI) was 1.3397. The plasmonic
19 device showed a RI sensitivity of 271.92 ± 14.92 nm/RIU and could detect the bovine serum
20 albumin (BSA) with good feasibility. These results suggest the potential suitability of our
21 platform to develop multiple monitoring systems.

22 **Keywords:** biosensor, plasmonic holes, NSL, polymer resin, biodetection.

23 **1. Introduction**

24 Surface plasmon resonance (SPR)-based biosensors measure the RI changes on a
25 metallic-dielectric interface due to biomolecular bond (analyte-bioreceptor) events. These
26 devices have attracted great interest in label-free methods in the last decades [1]. In particular,
27 the devices that use nanohole arrays demonstrate high sensibility, and great potential for
28 miniaturized sensor production [2]. Another distinguishing feature is obtaining nanoholes with
29 high control in strategic points on the metallic film. This enables the integration with
30 microfluidic systems to get automatic and multipoint analysis, which is an important feature
31 for the future SPR biosensor technology [3]. Moreover, the nanoholes arrays can be projected
32 with a specific optic characteristic, controlling the hole diameter, geometry, and periodicity
33 [4].

34 The cost-effectivity of nanoholes production may make their use difficult on a large
35 scale due to the high-cost beam lithography techniques often used for nanoholes obtention,
36 such as focused ion beam (FIB) and electron beam lithography (EBL). As an alternative, many
37 authors have used nanospheres lithography (NSL). The accessible sphere self-assembly makes
38 possible the development of different structures [5]. For example, the sphere volume can be
39 adjusted by reactive ion etching (RIE) [6]. Thus, NSL has been used to prepare plasmonic
40 biosensors based on nanoholes [7], nanowires [8], and others [9].

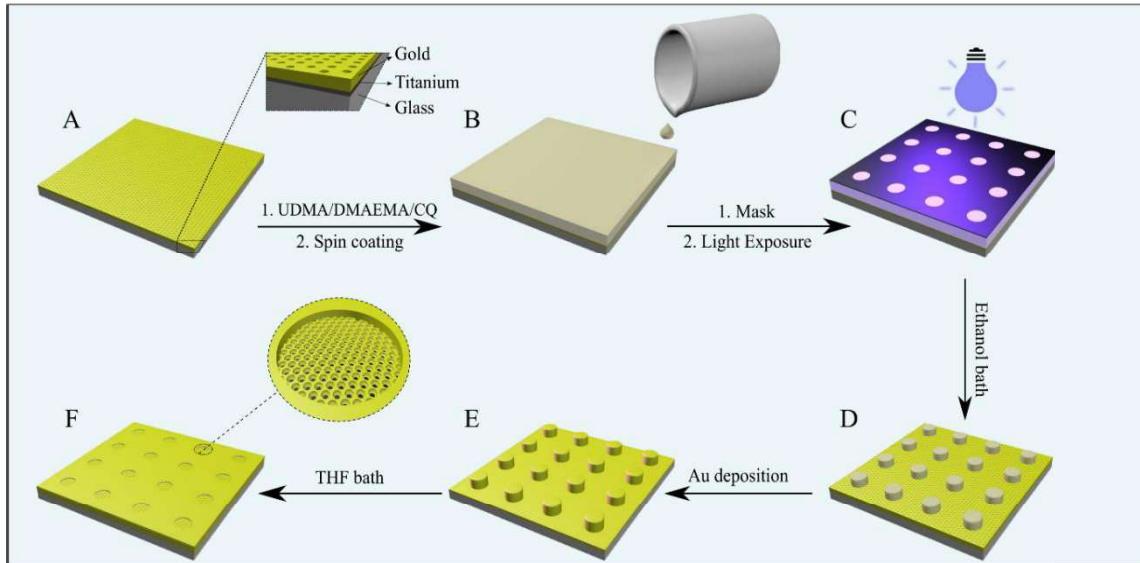
41 The NSL can thus provide a direct approach to overcoming the high cost of SPR
42 biosensors. Thus, in this work, we prepared an unprecedented plasmonic platform based on a
43 glass substrate covered with nanoholed gold thin film. The holes were obtained over the entire
44 metallic surface by NSL. This technique does not enable to have nanoholes arrays in specific
45 positions on the substrate. Thus, our main challenge was restricting the nanoholed substrate to
46 have a particular structure of Sen-PS on the substrate. The plasmonic platform had its sensitivity
47 characterized, and it was tested for BSA detection.

48 **2. Experimental section**

49 A glass substrate covered with an entire-nanoholed gold film (Figure 1A) fabricated by
50 NSL (support information) was used to develop a sensing multiple-point plasmonic device.
51 Firstly, the substrate was spin-coated at 500 rpm for 40 s and ramping at 1500 rpm for 15 s
52 (G3P-8) with a polymeric solution containing urethane dimethacrylate (UDMA, Aldrich,
53 100%), 2-(N, N-dimethylamino) ethyl methacrylate (DMAEMA, sigma-Aldrich, 100%), and

54 camphorquinone (CQ, Sigma-Aldrich, 97%) at the 85:10:5 wt% ratio (Figure 1B). The
55 polymeric solution was degassed before its use. Then, the Sen-Ps were obtained on the specific
56 regions of the substrate by photopolymerization. For that, a photomask containing transparent
57 circles of 150 μm diameter (printed in polyethylene terephthalate sheets with a high-resolution
58 printer) was positioned on the polymeric film and exposed to cure using LED curing light
59 (Radii-cal, 440-480 nm, 1200 mW/cm²) for 1 min (Figure 1C). The polymer on the regions
60 unexposed to light was removed using ethanol P.A. (synth) (Figure 1D). Then, using
61 a *sputtering* (Sputter coater SCD 050, Baltec, Canonsburg, USA), a second optically dense
62 gold layer was deposited onto the polymeric-recovered substrate using a current of 60 mA for
63 240 s (Figure 1E). Finally, the polymeric coating was removed by a THF bath (Neon, 99,0
64 wt%) for \sim 2 min (Figure 1F). Morphological characterizations were performed by scanning
65 electron microscopy (SEM, FEI Quanta 250), and the structure size was measured using Image-
66 Pro® PLUS (version 4.5.0.29) software. The sensibility was characterized by UV-Vis
67 spectroscopy (Ocean Optics, USB2000 with resolution of 0.1 nm), coupled to a metallographic
68 microscope (Quimis 9738 MIT with halogen lamp of 250 W) (Figure S1). The plasmonic
69 sensitivity to RI changes was conducted by exposing the substrate surface to glucose solutions
70 (Sigma-Aldrich, 99,5%) with different RIs (1.3397-1.3970). The substrate biodetection
71 capability was tested using BSA as a model biomolecule by the following procedure: 0.5
72 mmol/L of 11-mercaptoundecanoic acid (MUA, Sigma-Aldrich, \sim 95%) in ethanol covered the
73 plasmonic substrate for 6 h. Subsequently, the rinsed sensor was added to a 1:1 aqueous
74 solution containing N-ethyl-N-(3-diethylaminopropyl) carbodiimide (EDC, Sigma-Aldrich,
75 \geq 97%) 0.1 mol/L and N-hydroxysuccinimide (NHS, Sigma-Aldrich, \geq 99%) 0.1 mol/L for 4 h.
76 Finally, 0.5 $\mu\text{g/mL}$ of BSA (Sigma-Aldrich 96%) in PBS was passed through the device for
77 6 h.

78

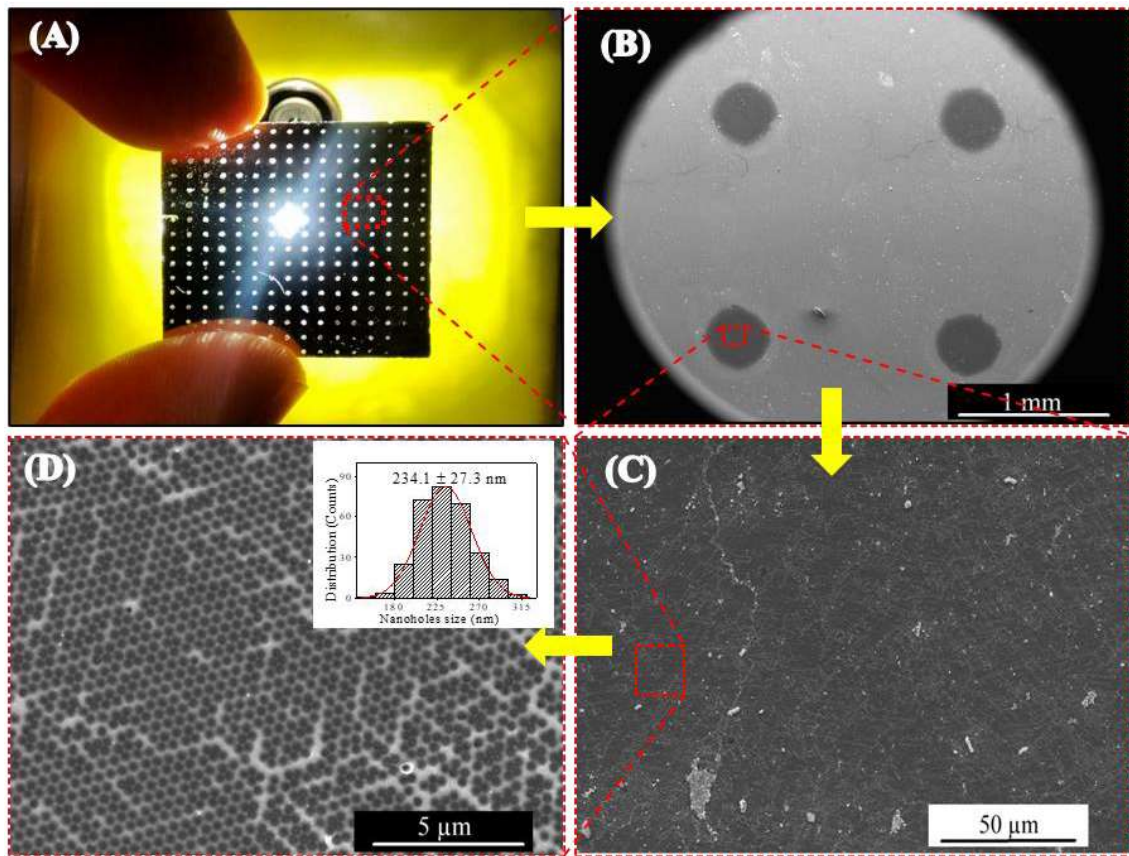


79

80 **Fig. 1.** Preparation process of the nanohole array-based plasmonic platform containing
 81 multiple Sen-Ps.

82 **3. Results and discussion**

83 Figure 2(A) shows a digital image of the demarcated substrate. The substrate had a 2.25
 84 cm² total area containing around 250 circular demarcations, called “sensing points (Sen-Ps)”,
 85 which are perfectly organized (bright spots in the image). The Sen-P arrays on the substrate
 86 can be customized just by changing the printed photomask design. Figure 2(B) shows an SEM
 87 image of a set of four Sen-Ps (highlighted in Fig. 1(A)) with a 380 μm diameter. The difference
 88 between the photomask transparent circular area (150 μm) and the actual area obtained for the
 89 Sen-Ps on the substrate (380 nm) can be attributed to the photopolymerization side-propagation
 90 [10]. Furthermore, light may enter beyond the edges of the transparent photomask due to a not
 91 fully collimated light beam. SEM images at different magnifications were obtained from a Sen-
 92 P and are shown in Figures 2(C)-2(D). The regions are filled with hexagonal-shaped nanoholes
 93 with an average size of 234,1 ± 27,3 nm (Inset in figure 2(D)).



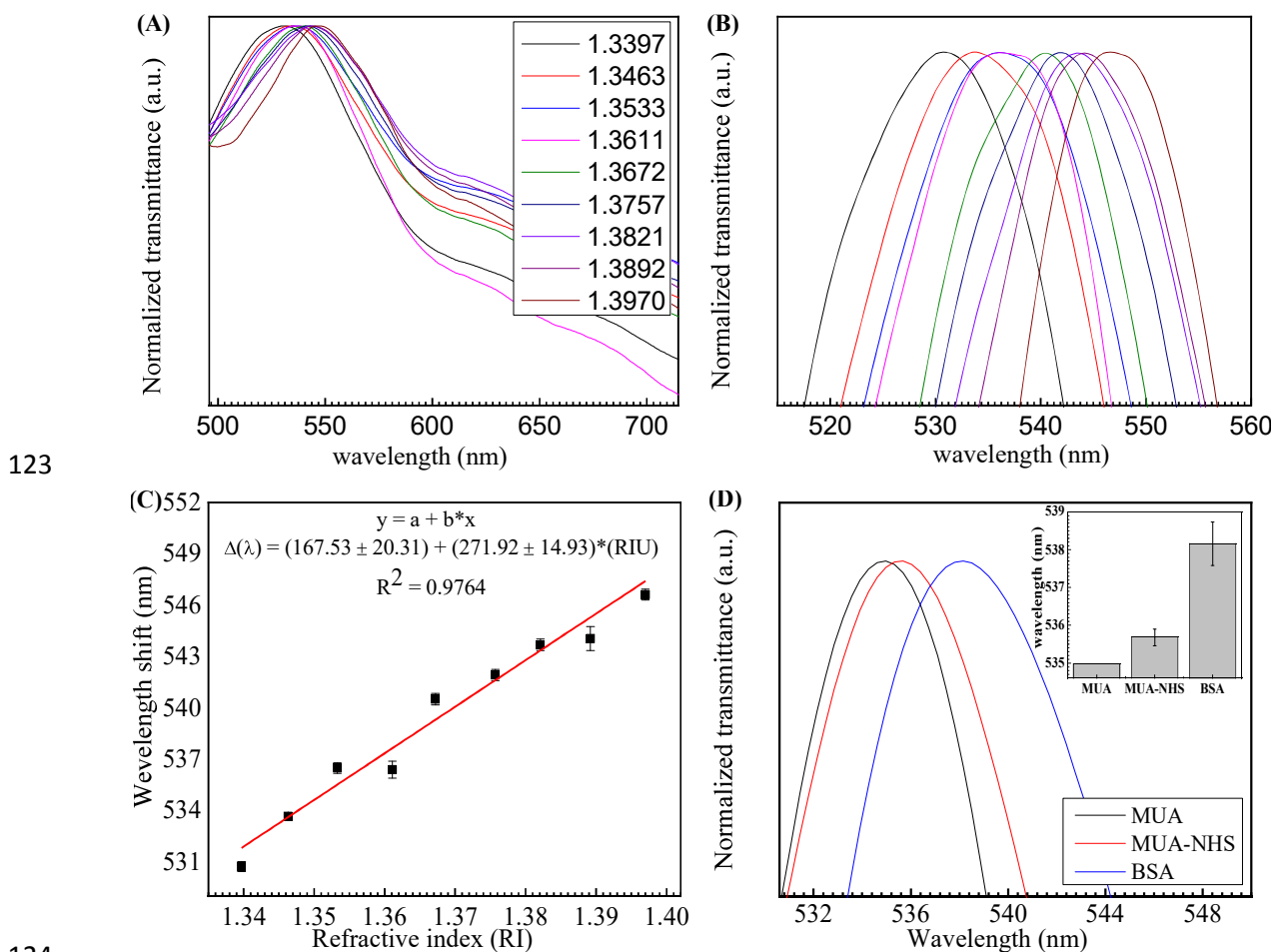
94

95 **Fig. 2.** (A) digital image of the demarcated substrate, (B) SEM image of the four Sen-Ps
 96 indicated in (A), and (C-D) show SEM images at different magnifications obtained within the
 97 Sen-P area. Inset in (D) shows a nanoholes size distribution obtained from the (D) image.

98 The evaluation of the Sen-P optical transmission revealed a plasmonic band at a 531
 99 nm maximum wavelength when a dilute glucose solution (RI=1,3397) was on it. This behavior
 100 was the same observed for other substrate Sen-Ps. The maximum transmission wavelength was
 101 close to the one observed in our previous work [11]. As observed in Fig. 3(A)-3(B), the
 102 transmission spectra were red-shifted as the RI increased. Then, the sensitivity (S) was
 103 determined using a linear fitting of the maximum transmission wavelength as a function of RI
 104 variation. The fitting curve inclination is defined as the Sen-P sensitivity (Fig. 3(C)), which
 105 was (271.92 ± 14.92) nm/RIU. Considering that the spectrophotometer precision was 0.1 nm,
 106 the calculated resolution was 3.7×10^{-4} RIU. Moreover, the R^2 for the linear fitting was
 107 acceptable (0.9764), indicating that the transmission maximum wavelength shift is proportional
 108 to RI changes.

109 The device response to the biomolecule immobilization was also investigated using the
 110 traditional model system of MUA/NHS-BSA [12]. The MUA-NHS layers are extensively used
 111 to provide a compact layer on the gold surface [13]. The BSA aminated groups quickly adsorb

112 on the MUA/NHS-modified gold surface and form a 4-7 nm thick monolayer [14] (see Figure
 113 S1 in the supplementary material). Thus, this molecular system can determine the substrate's
 114 ability to evaluate the sensor surface layer-by-layer modification. This is important to define if
 115 the substrate can work as a biosensor. The Sen-P transmission spectra were acquired for each
 116 modification step. The region of the spectra maximum absorbance can be observed in Figure
 117 3(D). The Fig. 3(D) inset shows the plasmonic band maximum wavelength for each
 118 modification. The redshift caused by the immobilization of the NHS layer over MUA was only
 119 (0.6 ± 0.22) nm. However, a significant shift of (2.48 ± 0.57) nm was observed when BSA
 120 molecules were immobilized on MUA-NHS layer. According to these results, and the literature
 121 support (supplementary information) the device has potential to detect surface biological
 122 interactions and also work as a plasmonic biosensor.



125 **Fig. 3.** (A) Transmission spectra for a Sen-P at different IRs (1.3397-1.3970). (B) Expansion
 126 of the maximum absorbance region observed in (A). (C) Maximum transmission wavelength
 127 as a function of RI variation (sensitivity curve). (D) Transmission spectra for each step of the
 128 gold surface modification on the Sen-P. Inset: column chart to plasmonic band redshift.

129 **4. Conclusion**

130 In this work, we presented a new nanoholes-based plasmonic substrate obtained by a
131 facile technique. The substrate was successfully prepared with several Sen-Ps with a circular
132 area filled with hexagonal-shaped nanoholes on gold thin film. The Sen-Ps were obtained using
133 a polymeric coating followed by remotion and then obtaining the plasmonic platform with a
134 multiplexing potential. The Sen-P arrays can be fully customized. It was shown that the device
135 was sensitive to RI changes and can respond to the biomolecules immobilization. The results
136 demonstrated that combining the techniques (nanohole development by NSL and demarcation
137 with an easy-to-remove polymeric system) enables the acquisition of plasmonic substrate with
138 the potential for multiple detections at a label-free single device.

139 **Acknowledgement**

140 The authors thank the Complex of Research Support Centers (COMCAP) for equipment
141 support, CNPq (process no. 305290/2020-7) and CAPES for fellowships.

142 **References**

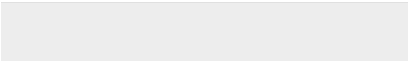
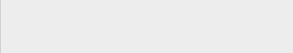
- 143 [1] J. Homola, Surface Plasmon Resonance Sensors for Detection of Chemical and
144 Biological Species, *Chem. Rev.* 108 (2008) 462–493.
145 <https://doi.org/10.1021/cr068107d>.
- 146 [2] C. Escobedo, On-chip nanohole array-based sensing: a review, *Lab Chip*. 13 (2013)
147 2445. <https://doi.org/10.1039/c3lc50107h>.
- 148 [3] P.H.M. Buzzetti, M.M. Taniguchi, N.S. Mendes, R.C. Vicentino, J.H. Oliveira, B.P.
149 Cabral Júnior, M. Souza, J.P. Monteiro, E.M. Giroto, Complete experimental and
150 theoretical characterization of nonlinear concentration gradient generator microfluidic
151 device for analytical purposes, *Microchim. Acta*. 11 (2022) 1–9.
152 <https://doi.org/10.1007/s00604-021-05110-7>.
- 153 [4] A. Shalabney, I. Abdulhalim, Sensitivity-enhancement methods for surface plasmon
154 sensors, *606* (2011) 571–606. <https://doi.org/10.1002/lpor.201000009>.
- 155 [5] P. Colson, C. Henrist, R. Cloots, Nanosphere Lithography: A Powerful Method for the
156 Controlled Manufacturing of Nanomaterials, *2013* (2013)
157 <https://doi.org/10.1155/2013/948510>.

- 158 [6] J.M. Chem, Elevated Ag nanohole arrays for high performance plasmonic sensors based
159 on extraordinary optical transmission, (2012) 8903–8910.
160 <https://doi.org/10.1039/c2jm30525a>.
- 161 [7] L. Niu, K. Cheng, Y. Wu, T. Wang, Q. Shi, D. Liu, Z. Du, Sensitivity improved
162 plasmonic gold nanoholes array biosensor by coupling quantum-dots for the detection
163 of specific biomolecular interactions, *Biosens. Bioelectron.* 50 (2013) 137–142.
164 <https://doi.org/10.1016/j.bios.2013.06.023>.
- 165 [8] S. Su, L. Lin, Z. Li, J. Feng, Z. Zhang, The fabrication of large-scale sub-10-nm core-
166 shell silicon nanowire arrays, *Nanoscale Res. Lett.* 8 (2013) 405.
167 <https://doi.org/10.1186/1556-276X-8-405>.
- 168 [9] Y. Cai, Y. Cao, P. Nordlander, P.S. Cremer, Fabrication of Split-Rings via Stretchable
169 Colloidal Lithography, *ACS Photonics.* 1 (2014) 127–134.
170 <https://doi.org/10.1021/ph400127g>.
- 171 [10] Z. Zhao, J. Wu, X. Mu, H. Chen, H.J. Qi, D. Fang, Origami by frontal
172 photopolymerization, *Sci. Adv.* 3 (2017) 1–8. <https://doi.org/10.1126/sciadv.1602326>.
- 173 [11] B.P. Cabral Júnior, J.H. Oliveira, P.H.M. Buzzetti, M.G.V. Fressatti, J.P. Monteiro,
174 L.H.C. Amorin, A. Urbano, E. Radovanovic, E.M. Girotto, Cost-effective plasmonic
175 device for label-free streptavidin detection, *Mater. Lett.* 227 (2018) 243–246.
176 <https://doi.org/10.1016/j.matlet.2018.05.064>.
- 177 [12] C.T.P. Silva, J.P. Monteiro, E. Radovanovic, E.M. Girotto, Unprecedented high
178 plasmonic sensitivity of substrates based on gold nanoparticles, *Sensors Actuators B*
179 *Chem.* 191 (2014) 152–157. <https://doi.org/10.1016/j.snb.2013.09.109>.
- 180 [13] J.P. Monteiro, J. Ferreira, R.G. Sabat, P. Rochon, M.J.L. Santos, E.M. Girotto, SPR
181 based biosensor using surface relief grating in transmission mode, *Sensors Actuators B*
182 *Chem.* 174 (2012) 270–273. <https://doi.org/10.1016/j.snb.2012.08.026>.
- 183 [14] H. Guner, E. Ozgur, G. Kokturk, M. Celik, E. Esen, A.E. Topal, S. Ayas, Y. Uludag, C.
184 Elbuken, A. Dana, A smartphone-based surface plasmon resonance imaging (SPRi)
185 platform for on-site biodetection, *Sensors Actuators B Chem.* 239 (2017) 571–577.
186 <https://doi.org/10.1016/j.snb.2016.08.061>.

Click here to access/download

Electronic Supplementary Material (online publication only)

Supplementary.docx





Click here to access/download

Electronic Supplementary Material (online publication only)

Supplementary materials_revised_.docx



CRedit authorship contribution statement

B. P. Cabral Júnior: Data curation, Methodology, Writing – original draft. **J. H. de Oliveira:** Formal analysis, Writing – review & editing, methodology. **J. P. Monteiro:** Conceptualization, Writing – review & editing. **P. H. M. Buzzetti:** Writing – review & editing, Investigation. **M. M. Taniguchi:** Writing – review & editing. **L. H. C. Amorin:** Methodology, Resources. **A. Urbano:** Resources. **E. Radovanovic:** Supervision, Methodology. **E. M. Giroto:** Supervision, Writing- Reviewing and Editing.

Declaration of interests

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

The authors declare the following financial interests/personal relationships which may be considered as potential competing interests:

1 **Facile** Multipoint Nanoholes Array-Based Plasmonic Platform: Development,
2 Characterization, and Sensor Evaluation

3 Bento Pereira Cabral Júnior^a, Jean Halison de Oliveira^a, Paulo Henrique Maciel Buzzetti^a,
4 Maiara Mitiko Taniguchi^a, Johny Paulo Monteiro^b, Luís Henrique Cardozo Amorim^c,
5 Alexandre Urbano^c, Eduardo Radovanovic^a, Emerson Marcelo Giroto^{a*}

6 ^a *Department of Chemistry, State University of Maringá, Maringá, PR, Brazil.*

7 ^b *Department of Chemistry, Federal Technological University of Paraná, Apucarana, PR,*
8 *Brazil.*

9 ^c *Department of Physics, State University of Londrina, Londrina, PR, Brazil*

10 **Correspondent Author. E-mail address: emgirotto@uem.br.*

11

12 **Abstract**

13 This study demonstrates a novel nanoholes array-based plasmonic platform with potential for
14 application in multi-detection analysis. The nanostructured transducer was obtained by
15 nanosphere lithography (NSL), which was restricted to having a sensing multipoint. The
16 microscopic characterization showed sensing point (Sen-P) arrays with *ca.* 380 μm and
17 nanoholes with 234.1 ± 27.3 nm diameter. Nanoholes array transmission spectra presented a
18 plasmonic band in *ca.* 531 nm when the refractive index (RI) was 1.3397. The plasmonic
19 device showed a RI sensitivity of 271.92 ± 14.92 nm/RIU and could detect the bovine serum
20 albumin (BSA) with good feasibility. These results suggest the potential suitability of our
21 platform to develop multiple monitoring systems.

22 **Keywords:** biosensor, plasmonic holes, NSL, polymer resin, biodetection.

23 **1. Introduction**

24 Surface plasmon resonance (SPR)-based biosensors measure the RI changes on a
25 metallic-dielectric interface due to biomolecular bond (analyte-bioreceptor) events. These
26 devices have attracted great interest in label-free methods in the last decades [1]. In particular,
27 the devices that use nanohole arrays demonstrate high sensibility, and great potential for
28 miniaturized sensor production [2]. Another distinguishing feature is obtaining nanoholes with
29 high control in strategic points on the metallic film. This enables the integration with
30 microfluidic systems to get automatic and multipoint analysis, which is an important feature
31 for the future SPR biosensor technology [3]. Moreover, the nanoholes arrays can be projected
32 with a specific optic characteristic, controlling the hole diameter, geometry, and periodicity
33 [4].

34 The cost-effectivity of nanoholes production may make their use difficult on a large
35 scale due to the high-cost beam lithography techniques often used for nanoholes obtention,
36 such as focused ion beam (FIB) and electron beam lithography (EBL). As an alternative, many
37 authors have used nanospheres lithography (NSL). The accessible sphere self-assembly makes
38 possible the development of different structures [5]. For example, the sphere volume can be
39 adjusted by reactive ion etching (RIE) [6]. Thus, NSL has been used to prepare plasmonic
40 biosensors based on nanoholes [7], nanowires [8], and others [9].

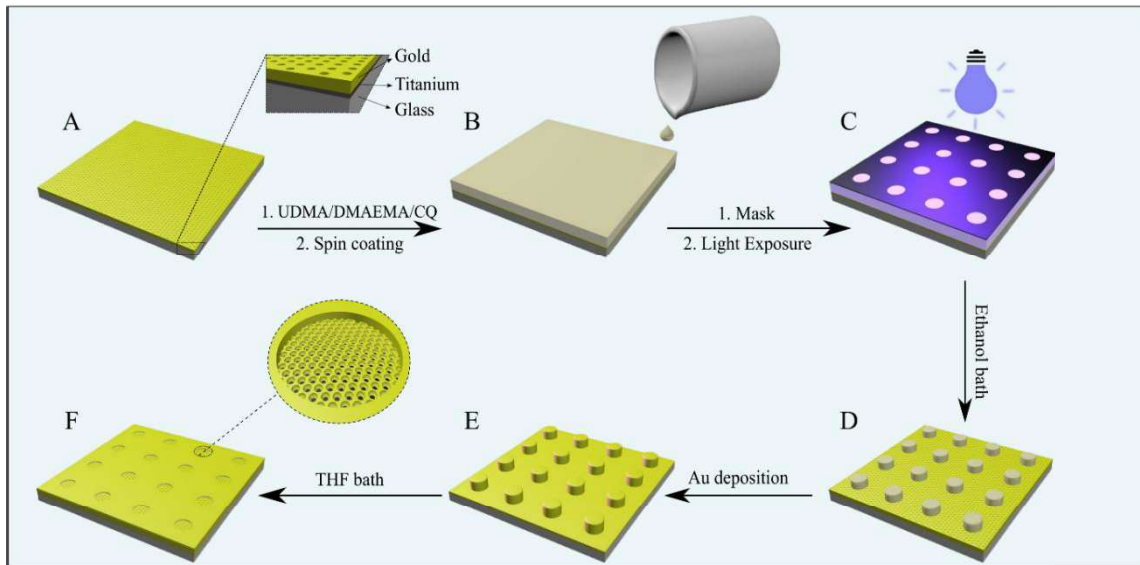
41 The NSL can thus provide a direct approach to overcoming the high cost of SPR
42 biosensors. Thus, in this work, we prepared an unprecedented plasmonic platform based on a
43 glass substrate covered with nanoholed gold thin film. The holes were obtained over the entire
44 metallic surface by NSL. This technique does not enable to have nanoholes arrays in specific
45 positions on the substrate. Thus, our main challenge was restricting the nanoholed substrate to
46 have a particular structure of Sen-PS on the substrate. The plasmonic platform had its sensitivity
47 characterized, and it was tested for BSA detection.

48 **2. Experimental section**

49 A glass substrate covered with an entire-nanoholed gold film (Figure 1A) fabricated by
50 NSL (support information) was used to develop a sensing multiple-point plasmonic device.
51 Firstly, the substrate was spin-coated at 500 rpm for 40 s and ramping at 1500 rpm for 15 s
52 (G3P-8) with a polymeric solution containing urethane dimethacrylate (UDMA, Aldrich,
53 100%), 2-(N, N-dimethylamino) ethyl methacrylate (DMAEMA, sigma-Aldrich, 100%), and

54 camphorquinone (CQ, Sigma-Aldrich, 97%) at the 85:10:5 wt% ratio (Figure 1B). The
55 polymeric solution was degassed before its use. Then, the Sen-Ps were obtained on the specific
56 regions of the substrate by photopolymerization. For that, a photomask containing transparent
57 circles of 150 μm diameter (printed in polyethylene terephthalate sheets with a high-resolution
58 printer) was positioned on the polymeric film and exposed to cure using LED curing light
59 (Radii-cal, 440-480 nm, 1200 mW/cm²) for 1 min (Figure 1C). The polymer on the regions
60 unexposed to light was removed using ethanol P.A. (synth) (Figure 1D). Then, using
61 a *sputtering* (Sputter coater SCD 050, Baltec, Canonsburg, USA), a second optically dense
62 gold layer was deposited onto the polymeric-recovered substrate using a current of 60 mA for
63 240 s (Figure 1E). Finally, the polymeric coating was removed by a THF bath (Neon, 99,0
64 wt%) for \sim 2 min (Figure 1F). Morphological characterizations were performed by scanning
65 electron microscopy (SEM, FEI Quanta 250), and the structure size was measured using Image-
66 Pro® PLUS (version 4.5.0.29) software. The sensibility was characterized by UV-Vis
67 spectroscopy (Ocean Optics, USB2000 with resolution of 0.1 nm), coupled to a metallographic
68 microscope (Quimis 9738 MIT with halogen lamp of 250 W) (Figure S1). The plasmonic
69 sensitivity to RI changes was conducted by exposing the substrate surface to glucose solutions
70 (Sigma-Aldrich, 99,5%) with different RIs (1.3397-1.3970). The substrate biodetection
71 capability was tested using BSA as a model biomolecule by the following procedure: 0.5
72 mmol/L of 11-mercaptoundecanoic acid (MUA, Sigma-Aldrich, \sim 95%) in ethanol covered the
73 plasmonic substrate for 6 h. Subsequently, the rinsed sensor was added to a 1:1 aqueous
74 solution containing N-ethyl-N-(3-diethylaminopropyl) carbodiimide (EDC, Sigma-Aldrich,
75 \geq 97%) 0.1 mol/L and N-hydroxysuccinimide (NHS, Sigma-Aldrich, \geq 99%) 0.1 mol/L for 4 h.
76 Finally, 0.5 $\mu\text{g}/\text{mL}$ of BSA (Sigma-Aldrich 96%) in PBS was passed through the device for
77 6 h.

78

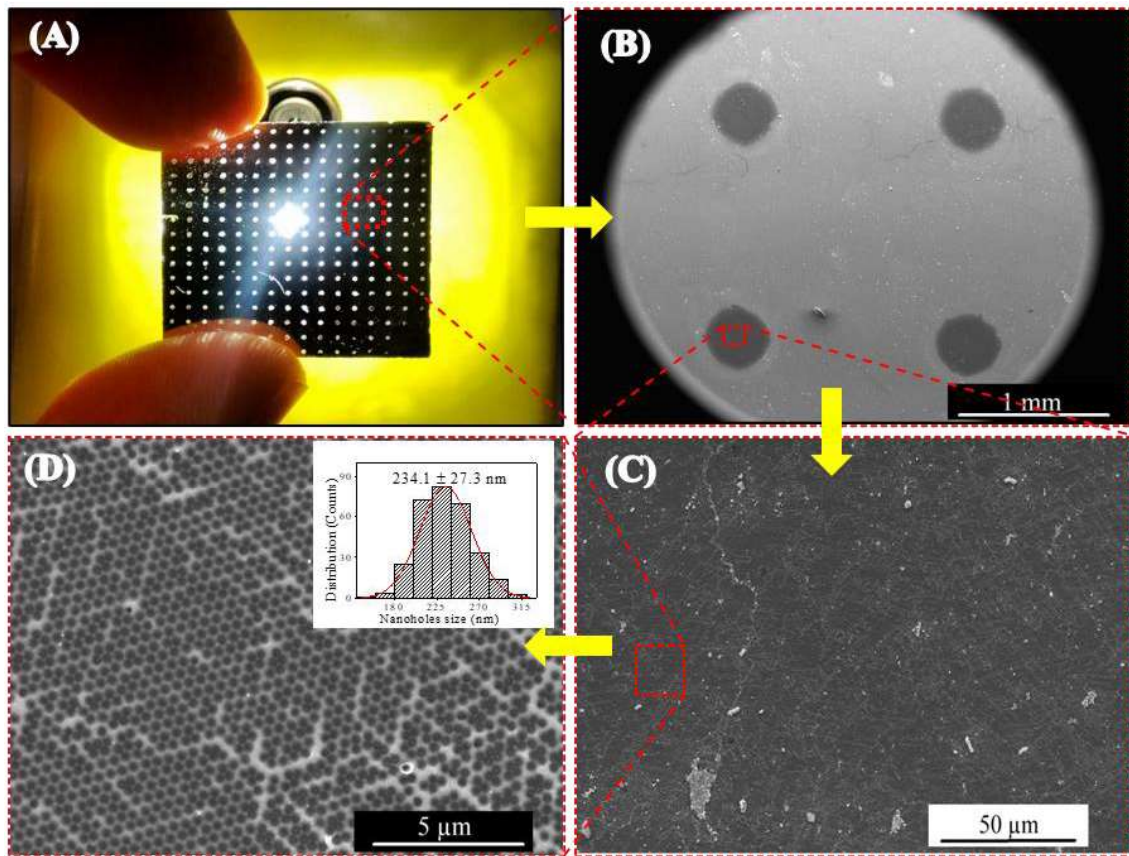


79

80 **Fig. 1.** Preparation process of the nanohole array-based plasmonic platform containing
 81 multiple Sen-Ps.

82 3. Results and discussion

83 Figure 2(A) shows a digital image of the demarcated substrate. The substrate had a 2.25
 84 cm² total area containing around 250 circular demarcations, called “sensing points (Sen-Ps)”,
 85 which are perfectly organized (bright spots in the image). The Sen-P arrays on the substrate
 86 can be customized just by changing the printed photomask design. Figure 2(B) shows an SEM
 87 image of a set of four Sen-Ps (highlighted in Fig. 1(A)) with a 380 μm diameter. The difference
 88 between the photomask transparent circular area (150 μm) and the actual area obtained for the
 89 Sen-Ps on the substrate (380 nm) can be attributed to the photopolymerization side-propagation
 90 [10]. Furthermore, light may enter beyond the edges of the transparent photomask due to a not
 91 fully collimated light beam. SEM images at different magnifications were obtained from a Sen-
 92 P and are shown in Figures 2(C)-2(D). The regions are filled with hexagonal-shaped nanoholes
 93 with an average size of 234,1 ± 27,3 nm (Inset in figure 2(D)).



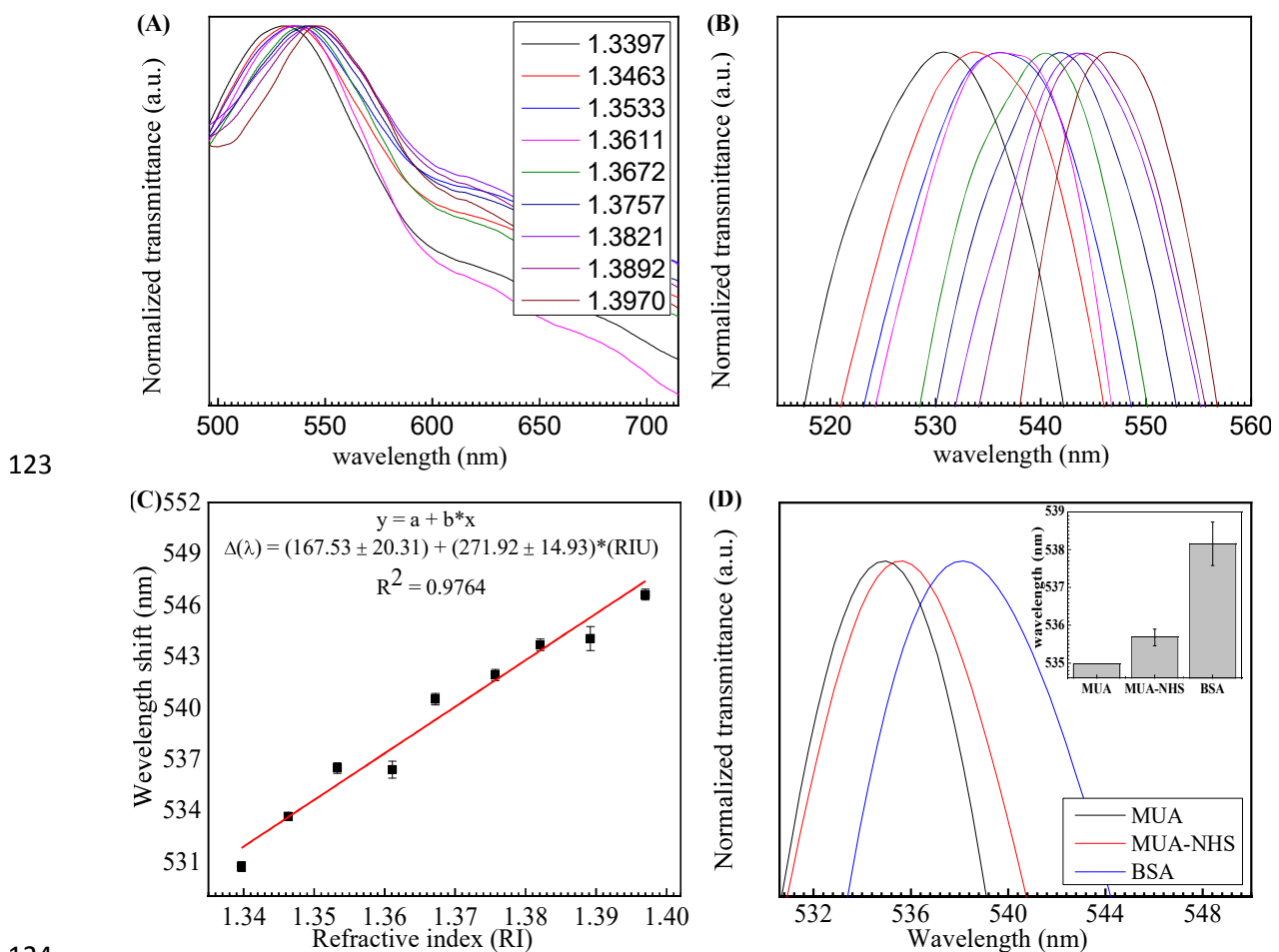
94

95 **Fig. 2.** (A) digital image of the demarcated substrate, (B) SEM image of the four Sen-Ps
 96 indicated in (A), and (C-D) show SEM images at different magnifications obtained within the
 97 Sen-P area. Inset in (D) shows a nanoholes size distribution obtained from the (D) image.

98 The evaluation of the Sen-P optical transmission revealed a plasmonic band at a 531
 99 nm maximum wavelength when a dilute glucose solution (RI=1,3397) was on it. This behavior
 100 was the same observed for other substrate Sen-Ps. The maximum transmission wavelength was
 101 close to the one observed in our previous work [11]. As observed in Fig. 3(A)-3(B), the
 102 transmission spectra were red-shifted as the RI increased. Then, the sensitivity (S) was
 103 determined using a linear fitting of the maximum transmission wavelength as a function of RI
 104 variation. The fitting curve inclination is defined as the Sen-P sensitivity (Fig. 3(C)), which
 105 was (271.92 ± 14.92) nm/RIU. Considering that the spectrophotometer precision was 0.1 nm,
 106 the calculated resolution was 3.7×10^{-4} RIU. Moreover, the R^2 for the linear fitting was
 107 acceptable (0.9764), indicating that the transmission maximum wavelength shift is proportional
 108 to RI changes.

109 The device response to the biomolecule immobilization was also investigated using the
 110 traditional model system of MUA/NHS-BSA [12]. The MUA-NHS layers are extensively used
 111 to provide a compact layer on the gold surface [13]. The BSA aminated groups quickly adsorb

112 on the MUA/NHS-modified gold surface and form a 4-7 nm thick monolayer [14] (see Figure
 113 S1 in the supplementary material). Thus, this molecular system can determine the substrate's
 114 ability to evaluate the sensor surface layer-by-layer modification. This is important to define if
 115 the substrate can work as a biosensor. The Sen-P transmission spectra were acquired for each
 116 modification step. The region of the spectra maximum absorbance can be observed in Figure
 117 3(D). The Fig. 3(D) inset shows the plasmonic band maximum wavelength for each
 118 modification. The redshift caused by the immobilization of the NHS layer over MUA was only
 119 (0.6 ± 0.22) nm. However, a significant shift of (2.48 ± 0.57) nm was observed when BSA
 120 molecules were immobilized on MUA-NHS layer. According to these results, and the literature
 121 support (supplementary information) the device has potential to detect surface biological
 122 interactions and also work as a plasmonic biosensor.



125 **Fig. 3.** (A) Transmission spectra for a Sen-P at different IRs (1.3397-1.3970). (B) Expansion
 126 of the maximum absorbance region observed in (A). (C) Maximum transmission wavelength
 127 as a function of RI variation (sensitivity curve). (D) Transmission spectra for each step of the
 128 gold surface modification on the Sen-P. Inset: column chart to plasmonic band redshift.

129 4. Conclusion

130 In this work, we presented a new nanoholes-based plasmonic substrate obtained by a
131 **facile** technique. The substrate was successfully prepared with several Sen-Ps with a circular
132 area filled with hexagonal-shaped nanoholes on gold thin film. The Sen-Ps were obtained using
133 a polymeric coating followed by remotion and then obtaining the plasmonic platform with a
134 multiplexing potential. The Sen-P arrays can be fully customized. It was shown that the device
135 was sensitive to RI changes and can respond to the biomolecules immobilization. The results
136 demonstrated that combining the techniques (nanohole development by NSL and demarcation
137 with an easy-to-remove polymeric system) enables the acquisition of plasmonic substrate with
138 the potential for multiple detections **at a label-free single** device.

139 Acknowledgement

140 The authors thank the Complex of Research Support Centers (COMCAP) for equipment
141 support, CNPq (process no. 305290/2020-7) and CAPES for fellowships.

142 References

- 143 [1] J. Homola, Surface Plasmon Resonance Sensors for Detection of Chemical and
144 Biological Species, *Chem. Rev.* 108 (2008) 462–493.
145 <https://doi.org/10.1021/cr068107d>.
- 146 [2] C. Escobedo, On-chip nanohole array-based sensing: a review, *Lab Chip*. 13 (2013)
147 2445. <https://doi.org/10.1039/c3lc50107h>.
- 148 [3] P.H.M. Buzzetti, M.M. Taniguchi, N.S. Mendes, R.C. Vicentino, J.H. Oliveira, B.P.
149 Cabral Júnior, M. Souza, J.P. Monteiro, E.M. Giroto, Complete experimental and
150 theoretical characterization of nonlinear concentration gradient generator microfluidic
151 device for analytical purposes, *Microchim. Acta*. 11 (2022) 1–9.
152 <https://doi.org/10.1007/s00604-021-05110-7>.
- 153 [4] A. Shalabney, I. Abdulhalim, Sensitivity-enhancement methods for surface plasmon
154 sensors, *606* (2011) 571–606. <https://doi.org/10.1002/lpor.201000009>.
- 155 [5] P. Colson, C. Henrist, R. Cloots, Nanosphere Lithography: A Powerful Method for the
156 Controlled Manufacturing of Nanomaterials, *2013* (2013)
157 <https://doi.org/10.1155/2013/948510>.

- 158 [6] J.M. Chem, Elevated Ag nanohole arrays for high performance plasmonic sensors based
159 on extraordinary optical transmission, (2012) 8903–8910.
160 <https://doi.org/10.1039/c2jm30525a>.
- 161 [7] L. Niu, K. Cheng, Y. Wu, T. Wang, Q. Shi, D. Liu, Z. Du, Sensitivity improved
162 plasmonic gold nanoholes array biosensor by coupling quantum-dots for the detection
163 of specific biomolecular interactions, *Biosens. Bioelectron.* 50 (2013) 137–142.
164 <https://doi.org/10.1016/j.bios.2013.06.023>.
- 165 [8] S. Su, L. Lin, Z. Li, J. Feng, Z. Zhang, The fabrication of large-scale sub-10-nm core-
166 shell silicon nanowire arrays, *Nanoscale Res. Lett.* 8 (2013) 405.
167 <https://doi.org/10.1186/1556-276X-8-405>.
- 168 [9] Y. Cai, Y. Cao, P. Nordlander, P.S. Cremer, Fabrication of Split-Rings via Stretchable
169 Colloidal Lithography, *ACS Photonics.* 1 (2014) 127–134.
170 <https://doi.org/10.1021/ph400127g>.
- 171 [10] Z. Zhao, J. Wu, X. Mu, H. Chen, H.J. Qi, D. Fang, Origami by frontal
172 photopolymerization, *Sci. Adv.* 3 (2017) 1–8. <https://doi.org/10.1126/sciadv.1602326>.
- 173 [11] B.P. Cabral Júnior, J.H. Oliveira, P.H.M. Buzzetti, M.G.V. Fressatti, J.P. Monteiro,
174 L.H.C. Amorin, A. Urbano, E. Radovanovic, E.M. Girotto, Cost-effective plasmonic
175 device for label-free streptavidin detection, *Mater. Lett.* 227 (2018) 243–246.
176 <https://doi.org/10.1016/j.matlet.2018.05.064>.
- 177 [12] C.T.P. Silva, J.P. Monteiro, E. Radovanovic, E.M. Girotto, Unprecedented high
178 plasmonic sensitivity of substrates based on gold nanoparticles, *Sensors Actuators B*
179 *Chem.* 191 (2014) 152–157. <https://doi.org/10.1016/j.snb.2013.09.109>.
- 180 [13] J.P. Monteiro, J. Ferreira, R.G. Sabat, P. Rochon, M.J.L. Santos, E.M. Girotto, SPR
181 based biosensor using surface relief grating in transmission mode, *Sensors Actuators B*
182 *Chem.* 174 (2012) 270–273. <https://doi.org/10.1016/j.snb.2012.08.026>.
- 183 [14] H. Guner, E. Ozgur, G. Kokturk, M. Celik, E. Esen, A.E. Topal, S. Ayas, Y. Uludag, C.
184 Elbuken, A. Dana, A smartphone-based surface plasmon resonance imaging (SPRi)
185 platform for on-site biodetection, *Sensors Actuators B Chem.* 239 (2017) 571–577.
186 <https://doi.org/10.1016/j.snb.2016.08.061>.