

# Impact of Ionic Liquids on the (bio)degradability of Poly(butylene succinate)/Poly(lactic acid) blends

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#### CORVEIGHT

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# Impact of Ionic Liquids on the (bio)degradability of Poly(butylene succinate)/Poly(lactic acid) blends

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Bio-based and (bio)degradable polymers constitute an important material innovation because they reduce the amount of waste materials inducing persistent microplastics and can offer similar benefits to conventional polymer materials. Poly(butylene succinate) and poly(lactic acid) blends exhibit interesting properties and can be possible alternatives to some traditional polymers. Some of their properties can be tailored by adding small proportions of ionic liquids (IL) that can act as interfacial agents between PBS and PLA. In our study, samples formulated with ionic liquids display a broader morphology with thermal properties close to the PBS/PLA reference, whereas Young's modulus is lowered in the presence of one of the IL studied. Nevertheless, the blends have a rather different ability to (bio) degrade. Indeed, disintegration experiments show that PBS/PLA/IL exhibit higher weight losses and faster fragmentation. DSC thermograms display an important decrease of PLA melting temperature after composting experiment, indicating that PLA phases are affected the most by degradation at 58°C. Our study shows that elaborating polymer materials, for which degradation processes are preferentially located in a "predegraded" dispersed phase, can be considered as a way to speed up macroscopic (bio)degradation. In the present work, morphologies, mechanical properties as well as (bio)degradability can be tailored by adding a small amount of ionic liquids.

KEYWORDS

PLA, PBS, ionic liquids, thermal properties, morphologies, (bio)degradation

#### Introduction

The development of polymer blends has been attracting the attention of academic and industrial research for years, and represents both important scientific and economic challenges (Wu, 1985; Wu, 1987; Yu et al., 2006). Indeed, binary and ternary mixtures are promising routes for the production of high performance polymers with cost-effectiveness. In the field of polymer materials, many authors have highlighted the

advantages of combining the properties of neat polymers. There are many examples of commercial polymer blends made of 1) polyolefins such as polypropylene (PP), polyethylene (PE) and its copolymers (Fel et al., 2016); 2) engineering polymers such as polyamide (PA), polyethylene terephthalate (PET) (Utracki, 1998; Lins et al., 2015; Quitadamo et al., 2017).

Concerning biodegradable polymers, many authors have developed blends containing poly(lactic acid) (PLA), which is well-known to have excellent properties, such as a high stiffness and good water vapor barrier properties (Ma, 2004; Bordes et al., 2009; Okamoto and John, 2013) that make it suitable for packaging and medical applications. However, its major drawback, i.e. its low stretchability, limits its use in packaging applications (Kopinke et al., 1996; Taubner and Shishoo, 2001). To circumvent this problem, PLA is widely blended with more ductile biodegradable polymers such as poly(butylene-adipateco-terephtalate) (PBAT), poly(butylene-succinate) (PBS) or polycaprolactone (Na et al., 2002; Furukawa et al., 2005; Jiang et al., 2006). In such a context, poly(butylene succinate) (PBS) and poly(lactic acid) (PLA), two bio-based polyesters, combine satisfying mechanical and thermal properties, as well as an ability for (bio)degradation (Bhatia et al., 2007; Su et al., 2019).

PBS, obtained by polycondensation of fossil or bio-based succinic acid and 1,4 butanediol, can be a relevant alternative to polyethylene for various uses, such as packaging, films. PBS and its copolymers are able to biodegrade in diverse environments (Xu et al., 2010a), with potential regulation of biodegradation as a function of the formulations. In an objective of reinforcement, PBS can be blended with PLA, with which it is not miscible, at proportions higher than 20%wt of PBS in PLA (Shibata et al., 2006; Bhatia et al., 2007; Deng and Thomas, 2015). It was found that PBS particles in PLA acted as a nucleating agent for PLA (Yokohara and Yamaguchi, 2008; Wu et al., 2012; Deng and Thomas, 2015; Ostrowska et al., 2019). Qiu et al. characterized and modelled the mechanical behavior of PLA/PBS blends with different proportions (Qiu et al., 2016). More recently, Deng et al. blended PBS and PLA as well, and found that the addition of PBS in PLA matrix increases the ductility due to a continuous phase morphology (Deng and Thomas, 2015). Morphology, as well as thermal and mechanical properties, can be adjusted by selecting appropriate formulations (Livi et al., 2015). Moreover, parameters such as crystallinity, molecular weight, hydrophobicity of polymer materials, influence their behavior. For example, crystalline regions biodegrade less easily, and small molecular weights facilitate biodegradation. Hydrophilicity is preferred for improving affinity with bacteria and enzymes able to catalyze chain scissions. Weak phases of multiphase materials, leading to voids after degradation, can facilitate diffusion of bacteria and enzymes. Biodegradation is also easier at temperatures close or higher than the glass transition temperature (Tg) (Agarwal et al., 1998; Andersson et al., 2010; Delamarche et al., 2020a). Studies mentioning biodegradation of PBS/PLA blends showed that adding PBS in a PLA matrix leads

to a slower degradation rate in compost at 58°C due to PBS phases being highly crystalline (Luzi et al., 2016). On the contrary, at ambient temperature in soil, degradability of PBS/PLA blends increases with increasing the PBS content (Zhang et al., 2013).

In addition, from a thermodynamical point of view, it is wellknown that mixtures of polymers lead to immiscible blends, with a high interfacial tension, making melt mixing very difficult, and inducing the formation of unstable morphologies with weak interfacial adhesion, which are the main causes of poor final properties, such as mechanical performances. In fact, for polymer blends composed of two immiscible polymers denoted A and B, two types of macromolecular chains tend to minimize their contact interface, due to their mutual repulsion. This phenomenon results in a macro-separated phase leading to their bad mutual adhesion and therefore to the poor quality of the blends formed. Depending on the proportion of each polymer phase, two types of morphologies can be generated: 1) nodular morphology where the minor phase forms the dispersed nodular phase in the continuous phase of the second polymer and 2) co-continuous morphology where an increase of the minor phase induces a percolation phenomenon of this phase. For these reasons, many authors have used ionic liquids to produce polymer blends with enhanced final properties. For example, Fortunati et al. studied the influence of tributyl acetate citrate and isosorbide diester as plasticizers in PLA/ PBS blends (Fortunati et al., 2017). Compatibilization agents used in PLA/PBS blends were listed by Su et al. (Su et al., 2019). The two polyesters being immiscible, hydrolysis mechanisms can take place faster than for neat polymers, due to the presence of voids at the interfaces between PBS and PLA phases (Wang et al.,

In 2015, Lins et al. have investigated the effect of ILs based on trihexyltetradecyl phosphonium cation with different counter anions, on blends composed of poly(butyleneadipate-co-terephtalate) (PBAT) and poly(lactid acid) (PLA) (Lins et al., 2015). Phosphonium ILs combined with chloride (IL-Cl), phosphinate (IL-TMP), bistriflimide (IL-TFSI), hexanoate (IL-EHT) and phosphate (IL-EHP) have been introduced in PBAT/PLA blends by using a DSM microextruder with co-rotating screws leading to the formation of PLA fibrils or small droplets associated with the generation of gradient interfacial zones between PLA and PBAT polymers, depending on the interactions between both polymers and the counter anion, in agreement with Luzi et al. (Luzi et al., 2016). More recently, Megevand et al. have revealed the formation of interfacial mixture between PBAT and PLA compatibilized by ILs, with Atomic Force Microscopy. The authors have demonstrated that IL-Cl led to the formation of a thick interphase generating a good local miscibility of PBAT and PLA phases while IL-TMP induced the formation of an interface layer ensuring interactions with the two polymers (Megevand et al., 2016).

The first part of this work has consisted in investigating the influence of ILs based on ammonium or phosphonium cations on the final properties of PBS/PLA blends. In particular, an important loss of PLA molecular weight in presence of phosphonium ionic liquid was attained during melt processing, impacting its (bio)degradability.

In order to tailor use properties and (bio)degradation, we have chosen a PLA 005 grade suitable for good mixing with our PBS and IL, a ratio PBS/PLA of 60/40 to attain Young's modulus, convenient for applications such as packaging. 1% by weight of IL was selected as it is enough for IL to act as an interfacial agent confined in the material. For amounts in the range of 2–5%, the IL could migrate out of the material and to the PLA phase with increased plasticization and degradation of the PLA.

The second part deals with the study of both degradation and biodegradation of PLA/PBS/ionic liquids blends, with the objective of attaining a compromise between thermal, mechanical properties and ability for (bio)degradation. Indeed, physical, chemical degradations can be associated with biodegradation (Krzan et al., 2006).

Polyesters, polyamides, polysaccharides and polycarbonates, and in general polymers synthesized by polycondensation, are particularly subject to hydrolysis, in neutral, acid, and also in alkaline conditions (Hawkins, 1984).

Abiotic hydrolysis can be performed at neutral or basic pH when it is known that this promotes degradation (Cho et al., 2001; Zhou and Xanthos, 2008). They then allow the study of abiotic hydrolysis of polymers, which may be the limiting mechanism in the degradation of biodegradable polymers in complex media. In abiotic environments, PLA degrades faster in basic solutions than in neutral or acidic solutions (Makino et al., 1986; Schliecker et al., 2003; Xu et al., 2010). Hydrolysis of PBS is also favoured in basic media (Xu and Guo, 2010b).

As a consequence, in this study, 3-months-abiotic hydrolysis was conducted in deionized water and in alkaline solution. We have chosen to perform the abiotic tests under the same temperature conditions (58°C) as composting experiment. 3-months-composting was conducted in open containers, under the temperature of 58°C as ISO (2012). These identical temperature conditions allow us to determine whether abiotic hydrolysis is the limiting mechanism for biodegradation of the tested polymers.

Moreover, 58°C is not only a temperature required for ISO experiments but is also very close to the glass transition temperature of PLA and superior to the one of PBS.

To summarize, the present work aims at studying the relations between polymer materials properties and (bio) degradation. In fact, characteristics such as molecular weights, crystallinities, morphologies, surface properties are well known parameters that impact the biodegradability (Delamarche et al., 2020b). 3-months degradation essays were conducted on polyester samples to evaluate their degradability 1) under composting conditions, 2) in water at 58°C, 3) under soil

burial conditions, and 4) in humid atmosphere at room temperature. Weight loss, <sup>1</sup>H NMR spectra, and thermal properties measurements were used to follow degradation.

#### Materials and methods

#### **Products**

Products used in this study are listed in Table 1. PBS and PLA pellets were supplied by Natureplast (PBE 003 and PLA 005 grades, respectively). L-lactic acid/D-lactic acid content for PLA is 94:6 mol% (Dorigato et al., 2012). Trihexyl(tetradecyl) phosphonium chloride (denoted in the text as P-Cl) was purchased from Cytec. Tris(2-hydroxyethyl)methylammonium methylsulfate (denoted in the text as A-M) was purchased from Sigma Aldrich.

#### Sample preparation

Polymer pellets were dried in an oven at 70°C for 12 h. Pellets and additives were extruded using a 15g-capacity DSM microextruder (Midi 2000 Heerlen, Netherlands) with co-rotating screws (L/D ratio of 18) at 190°C, with a 100 rpm speed, for 3 min. PBS/PLA (60:40 wt. ratio), PBS/PLA/P-Cl (60:40:1 wt. ratio) and PBS/PLA/A-M (60:40:1 wt. ratio) were extruded. Extruded samples were injected in a 10 cm³ mould at 30°C to obtain 2 mm thick and 4 mm wide dumbbell-shaped specimens. 0.2 mm thin films were processed under compression at 210°C and cut for ageing experiments.

#### Characterization

#### Morphologies

Transmission electron microscopy (TEM) was carried out at the Technical Center of Microstructures of Lyon on a Philips CM 120 microscope with an accelerating voltage of 80 kV. 80–100 nm thin samples, which were cut using an ultramicrotome equipped with a diamond knife, were set on copper grids.

#### <sup>1</sup>H NMR

One dimensional <sup>1</sup>H NMR spectroscopy was used to determine the PBS and PLA contents of the blends. Samples, cut from films, were dissolved in CDCl<sub>3</sub> and analyzed at 25°C using a Bruker Advance III spectrometer (400 MHz), equipped with a 5 mm multinuclear broadband probe (BBFO+). In order to evaluate the weight percentage of PBS and PLA in the blends, peak resonance (a) of PLA (CH) at 5.1 ppm and resonance peak (a) of PBS (CH2) at 2.6 ppm were integrated to calculate the weight content of PBS:

TABLE 1 Formula of studied polymers and Ionic Liquids.

Abbreviation	Full name	Formula	$\overline{Mn}$ (g/mol)	Tm (°C)	Structural formula
PBS	Poly(butylene succinate)	$(C_8H_{12}O_4)_n$	50 × 10 <sup>3</sup>	113	
PLA	Poly(lactic acid)	$(C_3H_4O_2)_n$	34 × 10 <sup>3</sup>	170	
P-Cl	Trihexyl(tetradecyl)phosphonium chloride	C <sub>32</sub> H <sub>68</sub> ClP	519.3	-	OH O 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0
A-M	Tris(2-hydroxyethyl) methylammoniummethylsulfate	C <sub>8</sub> H <sub>21</sub> NO <sub>7</sub> S	275.3	-	P+ CI-

$$f_{(PBS)} = \frac{\frac{I_{a(PBS)}}{4}}{\frac{I_{a(PBS)}}{4} + I_{a(PLA)}}$$
(1)

$$wt\%_{(PBS)} = \frac{f_{(PBS)} \times M_{PBS}}{f_{(PBS)} \times M_{PBS} + \left(1 - f_{(PBS)}\right) \times M_{PLA}} \times 100 \quad (2)$$

with  $f_{(PBS)}$  the molar fraction of PBS,  $I_{a(PBS)}$  the integral of peak (a) of PBS,  $I_{a(PLA)}$  the integral of peak (a) of PLA, and  $wt\%_{(PBS)}$  the weight content of PBS.

End chain group analysis was carried out to calculate mean molecular weights of PBS. It was assumed that each polyester chain exhibits one hydroxyl and one carboxylic end groups. Resonance peak of hydroxyl end-group (CH<sub>2</sub>-OH) of PBS, appearing at 3.7 ppm (Labruyère et al., 2014), was integrated as well as CH<sub>2</sub> (a) one in the monomer repeating unit at 2.6 ppm  $\overline{M_n}$  was determined as follows:

$$\overline{M_n} = \frac{I_a}{4} \times \frac{2}{I_{CH_2-OH}} \times M_{PBS}$$
 (3)

with  $M_{PBS} = 172.2 \ g/mol$ , the molecular weight of PBS repeating unit,  $I_a$  the integral of resonance peak  $CH_2$  in the polymer

repeating unit, and  $I_{CH_2-OH}$  the integral of resonance peak of hydroxyl end group (CH<sub>2</sub>-OH).

In order to evaluate the molecular weight of PLA phase after processing, hydroxyl end-group of PLA (CH-OH) signed at 4.34 ppm and CH (a) of the monomer at 5.1 ppm were analyzed to calculate the molecular weights. However, this peak is close to peak (b) of PBS. Hence, PLA end-chain titration was carried out after extracting PLA from the PBS/PLA blend with the following protocol. First, samples were dissolved in CHCl<sub>3</sub>. Then, tetrahydrofuran was added to induce precipitation of PBS. After filtration and solvent evaporation, the remaining PLA and PBS oligomers (soluble in THF) were analyzed by NMR in CDCl<sub>3</sub>.  $\overline{M_n}$  was determined as follows:

$$\overline{M_n} = \frac{I_a}{1} \times \frac{1}{I_{CH OH}} \times M_{PLA} \tag{4}$$

with  $M_{PLA}=72.1~g/mol$ , the molecular weight of PLA repeating unit,  $I_a$  the CH peak integral in the polymer repeat unit,  $I_{CH-OH}$  the integral of peak of hydroxyl end group (CH-OH). Spectra with labels have been published in a previous work (Delamarche et al., 2020a).

#### Thermal properties

Differential scanning calorimetry analyses were carried out using a model Q20 TA Instruments equipment (TA Instruments, New Castle, DE, United States). Samples were subjected twice to a cycle of heating and cooling ramps of  $10^{\circ}$ C/min, from -70 to  $200^{\circ}$ C and from  $200^{\circ}$ C to  $-70^{\circ}$ C.

Thermogravimetric analyses were carried out using a model Q500 TA Instruments equipment. Samples were subjected to a heating rate of  $20^{\circ}\text{C}$  min $^{-1}$  under nitrogen atmosphere from 25 to 600°C. Temperatures at which 1% of the initial weight was lost  $T_{1\%~\text{deg}}$  (°C), and degradation temperatures  $T_{\text{degPLA}}$  and  $T_{\text{degPBS}}$  (obtained from the maxima of the derivative curves of the weight loss as a function of temperature) were determined.

#### Mechanical properties

Uniaxial tensile tests were conducted using an INSTRON 33R4469 (Instron, Norwood, MA, United States) tensile machine at 25°C. Dumbbell specimens were tested for a 50 mm/min elongation speed. Young's modulus, maximum and average strain at break were determined.

#### Dynamic mechanical analysis

Dynamic mechanical measurements were carried out using an ARES G2 rheometer (TA Instruments, New Castle, DE, United States). The heating rate was 3°C min<sup>-1</sup> from -60–90°C at a frequency of 1Hz, (0.1% oscillation strain). The change of shear storage modulus, G', and shear loss modulus, G'', were determined.  $tan(\delta)$  was considered to determine the alpharelaxation temperatures,  $T\alpha$ , related to Tg of PBS and PLA.  $\Delta T\alpha$  was defined as the difference between  $T(\alpha(PBS))$  and  $T(\alpha(PLA))$ .

#### Molecular weights

Molecular weights were determined using a steric exclusion chromatography (SEC) equipment, comprising Agilent Technologies columns (Argilent Technologies, Palo Alto, CA, United States), a light scattering detector, and a Shimadzu RID-10A detector. 3 mg samples were cut and dissolved in chloroform (1 mg/ml). Analysis took place at 30°C under a 1 ml/min flow.  $\frac{dn}{dc}$  used was 0.06 ml/g for PBS and in PBS/PLA blends, and 0.0237 for neat PLA only (Malmgren et al., 2006).

#### Degradatation experiments

Degradation experiments for 3 months were performed considering films exposed to various environments. Every month, two samples of each formulation were recovered, washed with deionized water, and gently dried on absorbent paper. Then, they were dried in a vacuum oven at 30°C for 12 h and kept in a closed desiccator. Weight losses were calculated as

follows ( $W_0$  being the weight before degradation experiment and  $W_t$  the weight after degradation experiment):

weight loss (%) = 
$$\frac{W_0 - W_t}{W_0} * 100$$
 (5)

#### Abiotic hydrolysis

3-months-abiotic hydrolysis was conducted at  $58^{\circ}$ C in deionized water and in alkaline solution (NaOH 0.01 M). 1.25\*1.5\*0.02 cm films were disposed in 10 ml of aqueous solution in individual closed flasks.

#### Composting experiment

3-months-composting was conducted at 58°C in open containers. Compost was from composting facility of Racine - Ecopole la Rize (Décines-Charpieu, France). Organic matter percentage was obtained by calcination of dry samples and was found to be  $58.2 \pm 0.3$  wt%. The water content and ability to retain water were analyzed. 2.5\*1.5\*0.02 cm polymer samples were buried 15 cm beneath the surface. Moisture content was regularly adjusted to be 90% of the maximum capacity of water retention.

#### Soil burial

3-months-soil burial experiment was conducted at room temperature in open containers. Soil was from LyonTech Campus La Doua, Villeurbanne, France. Organic matter percentage was obtained by calcination of dry samples and was found to be  $21\pm3$  wt%. Water composition and ability to retain water of the soil were analyzed as well. 2.5\*1.5\*0.02 cm polymer samples were buried  $10\,\mathrm{cm}$  beneath the surface. Moisture content was regularly adjusted to 90% of the maximum capacity of water retention.

#### Humid atmosphere

2.5\*1.5\*0.02 cm polymer samples were disposed in a closed transparent chamber with humidity-saturated air at room temperature for 3 months.

#### Results and Discussion

## Tailoring PBS/PLA blends properties with ILs

TEM micrographs (Figure 1) show examples of the dispersion of PLA phases (white domains) in the PBS matrix, confirming that PBS and PLA are immiscible (Bhatia et al., 2007). PLA phases in PBS/PLA blend without IL are irregularly dispersed (Figure 1A), and their maximum surface area is about 0.8 μm². The non-uniform dispersion of PLA domains has been observed by Deng et al., and it was assumed that, since PBS exhibits a lower melt viscosity than PLA, it is not able to

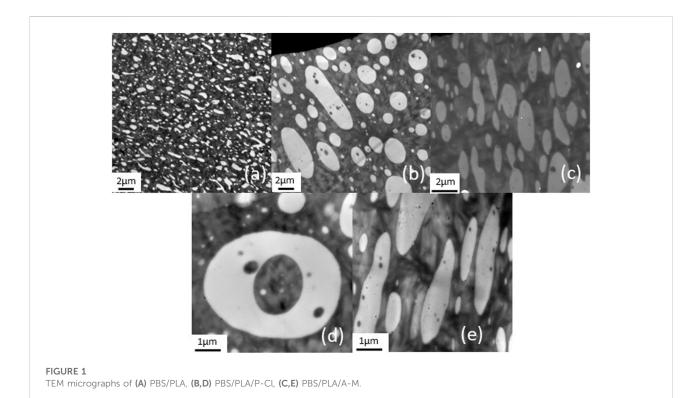


TABLE 2 Mechanical properties of neat PLA and PBS and PBS/PLA blends. \*Data provided in product datasheet.

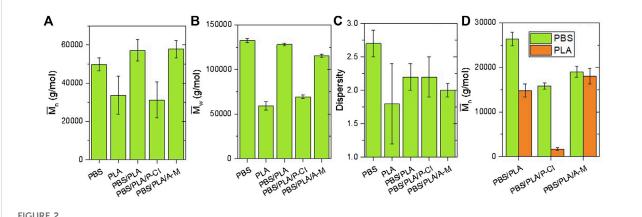
Formulation	Young's modulus (MPa)	Mean strain at break (%)	Maximum strain at break (%)
PBS	$320 \pm 27$	310 ± 90	385
PLA	3500*	5*	-
PBS/PLA	603 ± 57	$391 \pm 84$	418
PBS/PLA/P-Cl	$312 \pm 24$	$500 \pm 73$	600
PBS/PLA/A-M	665 ± 104	$413 \pm 43$	470

break down PLA droplets (Deng and Thomas, 2015). Similar compatible blends have been prepared in previous studies, including PBAT/PLA/LI and PBAT/PLA/lignin/LI blends (Lins et al., 2015; Livi et al., 2015) where it was clearly demonstrated that ionic liquids act as compatibilizing agents. Indeed, ionic liquids were located at the interface of the PLA domains and when increasing the amount of ionic liquids, a migration of ionic liquids in the minority phase was observed (Megevand et al., 2016). In the present study, the addition of ionic liquids induces the presence of larger PLA domains (up to  $27\,\mu\text{m}^2$  with P-Cl). Furthermore, PLA domains have ovoid shapes. Then, inclusions of PBS are observed in PLA domains. Since the PBS/PLA weight proportion is 60/40, phase inversion is likely to occur. Indeed, Wu et al. reported that, if the weight proportion of PBS is 60% or more, it becomes the continuous

phase, and the inversion point occurs at around 50/50 (Wu et al., 2012), which can explain the increase of PLA phase. At molecular scale, ionic liquids are observed at the interfaces.

The fact that adding ionic liquids induces larger PLA domains in the PBS matrix was not expected regarding our earlier studies. Lins et al. added 1% of phosphonium ILs in PBAT/PLA blends and showed that ILs generated interfacial zones due to a strong attraction to ester groups, leading to a decrease of the interfacial tension (Lins et al., 2015).

Tensile properties of PBS/PLA blends, reported in Table 2, show that neat PBS/PLA exhibits a rather high Young's modulus due to brittle PLA phases, acting as reinforcing agent, and a high strain at break due to PBS being ductile (Qiu et al., 2016). In presence of IL, complex morphologies, at ratio PBS/PLA 60/40 close to the inversion phase make the discussion rather



 $\overline{M_n}$ ,  $\overline{M_w}$  and dispersity of PBS, PLA and PBS/PLA blends determined in CHCl<sub>3</sub> SEC (A–C), and calculated with <sup>1</sup>H NMR titration (D). Results are given with experimental uncertainty.

difficult to establish relationship between tensile properties and morphology of the blends. Young's modulus and strain at break are slightly improved with ammonium IL, which could suggest the IL acting as an interfacial agent. The increase of modulus can also be explained by the fact that PLA domains are larger, and hence constitute more effective reinforcing agents. However, regarding PBS/PLA blend with phosphonium IL, the stiffness is reduced while elongation at break is significantly higher. PLA domains do not behave as a reinforcing agent in this blend. Elongation at break is probably enhanced due to a plasticization process, which could be induced by small PLA chains and phosphonium IL. It has been seen in the literature that phosphonium IL can plasticize PLA, as well as catalyze its degradation during processing (Park and Xanthos, 2009). By using Phosphonium IL when processing the PBS/PLA blend, it is not necessary to add a preliminary step aiming at cutting commercially available PLA chains in order to facilitate biodegradation. Hence, the increase of strain at break could be a consequence of low molecular weight PLA chains  $(\overline{M_n})$  of 1.7 kDa, see SEC measurements on Figure 2) and/or of phosphonium IL plasticizing PLA domains. The hypothesis of short PLA chains could explain the loss of stiffness as well, since PLA domains provide stiffness in the PBS/PLA blends. It was seen by Deng et al. that PBS/PLA blends exhibit a low ductility from 40 wt% to 80 wt% of PBS, attributed to a poor adhesion between the two polyesters at such proportions (Deng and Thomas, 2015). Hence, a better adhesion between PBS and PLA, provided by the ILs is another way to explain the gain in elongation at break with ILs. Nevertheless, Young's modulus decreases in presence of IL P-Cl, due to the depolymerization of PLA, while PBS is much less affected (Figure 2D). In addition, molecular weight being a parameter affecting the degradability of materials, faster degradation is expected for PBS/PLA blends containing IL P-Cl and a "predegraded" PLA phase, with a compromise between mechanical properties and degradability.

TABLE 3 Dynamic mechanical analysis of PBS/PLA blends with and without ionic liquids (1 phr).

Formulation	Ta, PBS (°C)	Ta, PLA (°C)	ΔTα (°C)
PBS/PLA	-25.6	61.1	86.7
PBS/PLA/P-Cl	-18.9	56.5	75.4
PBS/PLA/A-M	-25.5	61.7	87.2

Considering environmental and safety issues, it is well known that ILs, which catalyze the glycolysis of PET, can be used to depolymerize many polymers such as polyethylene terephthalate (Kamber et al., 2010). In our study, only a small amount of ILs is used (1 wt%), acting as interfacial agents and migrating into PLA matrix, which considerably limits environmental and safety issues. In our work, in presence of ionic liquid, the degradation products of PLA with the lowest molecular weights, might be lactic acid, that is neither considered dangerous nor very volatile (Song et al., 2014; Elsawy et al., 2017).

Data acquired with dynamic mechanical analyses are displayed in Table 3. Two distinct alpha-relaxation temperatures confirm that PBS and PLA are not fully miscible at the studied proportions (Deng and Thomas, 2015). A slight merging of the alpha-relaxation temperatures of PLA and PBS with phosphonium ionic liquid is observed. This small shift may be due to partial miscibility of low molecular weight chains (Lins et al., 2015). The decrease of the T $\alpha$  of PLA is also consistent with a migration of P-Cl ionic liquid in the PLA phase, leading to reduced molecular weights (Figure 2) and lower T $\alpha$ . Ammonium IL does not have a significant impact on the alpha-relaxation temperatures. These results are in agreement with partial miscibility and/or perhaps transesterification in presence of phosphonium IL.

Number average molecular weight  $(\overline{M_n})$ , weight average molecular weight  $(\overline{M_w})$ , and dispersity  $(\overline{M_w}/\overline{M_n})$  are

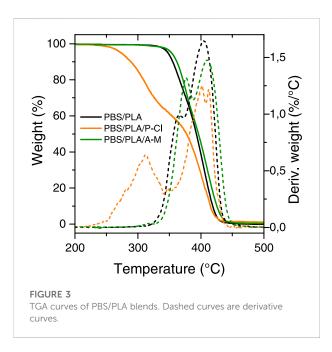


TABLE 4 Degradation temperatures of PLA and PBS phases and degradation temperature for 1% wt. loss from thermogravimetric analyses.

Formulation	$T_{1\% \ deg.}$ (°C)	$T_{degPLA}$ (°C)	T <sub>degPBS</sub> (°C)
PBS	304.3	-	409.1
PLA	319.0	379.0	-
PBS/PLA	323.6	365.2	404.7
PBS/PLA/P-Cl	240.6	311.5	402.3
PBS/PLA/A-M	314.3	377.1	409.7

summarized in Figures 2A–C. PLA and PBS are not distinguishable on SEC chromatograms. Hence, to assess the average molecular weight  $\overline{M_n}$  of specifically PBS or PLA phases, <sup>1</sup>H NMR spectroscopy (end group analysis) is used as well (Figure 2D). Molecular weights measured with SEC (Figure 2A) are higher than the ones calculated with end group analysis (Figure 2D), where it is assumed that the studied polyesters are not branched. Hence, values obtained with end group analysis may be underestimated. For this reason, trends, rather than absolute molecular weights, are discussed in this work.

Regarding SEC measurements on injected samples, PBS/PLA blends with phosphonium IL exhibit significantly lower molecular weights than PBS/PLA without additive. Using NMR titration, it is observed that the  $\overline{M_n}$  of PLA drastically decreases from 14.8 kg.mol<sup>-1</sup> to 1.7 kg.mol<sup>-1</sup> with phosphonium IL, while it slightly increases to 18.0 kg mol<sup>-1</sup> with ammonium IL.  $\overline{M_n}$  of PBS decreases with both ILs (from 26.3 kg mol<sup>-1</sup> to 15.8 with P-Cl, 19.0 with A-M). This suggests the phosphonium IL degrades the polymers during processing, as

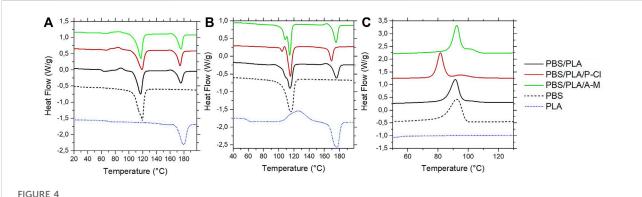
observed by Park et al. with neat PLA (Park and Xanthos, 2009). This could be due to transesterification reactions, causing random chain scissions at first (Lins et al., 2015). It was observed by Freyermouth that no or few transesterification reactions take place between PBS and PLA in PBS/PLA blends, at 180°C for 105 min (Freyermouth, 2014), however, these conclusions could differ with the addition of ionic liquids. Nonetheless, no additional peak, corresponding to a possible transesterification phenomenon, was detected using <sup>1</sup>H NMR spectroscopy. It is yet clear that P-Cl induces thermal degradation during processing, leading to lower molecular weight chains (especially PLA), which can explain partial miscibility, in agreement with a slight merging of alpha relaxation temperatures in DMA (Table 3). These results are consistent with thermogravimetric analyses (TGA) (Figure 3; Table 4), that show that PLA phases in PBS/PLA/P-Cl blends degrade at lower temperature than in PBS/PLA and PBS/PLA/ A-M blends. Regarding PBS phases, degradation temperatures do not change considerably.

As reported in our previous article (Delamarche et al., 2020a), temperature at 1% degradation (T1% deg.) and at the maxima of the derivative curves of the weight loss as a function of temperature, were 304 and 409°C for PBS; 319 and 379°C for PLA. This is consistent with the fact that PLA phases degraded to a greater extent during processing with P-Cl. Results are also in agreement with differential scanning calorimetry (DSC) results (Table 5), where it can be seen that melting temperature of PLA, during the second heating ramp, decreases with phosphonium IL (from 174.4°C on the first heating ramp to 169.5°C), while for neat PBS/PLA and PBS/PLA with ammonium IL, the melting temperature remains similar between the first and second heating ramp. This can be explained by a degradation of PLA phases taking place during the measurement, at the end of the first heating ramp, leading to smaller molecular weight PLA chains. Melting temperature indeed depends on the molecular weights: under a critical value of molecular weight, the melting temperature drops, whereas above this critical value, the melting temperature is constant (Fatou and Mandelkern, 1965; Sánchez-Soto et al., 2002). Xiang et al. observed that melting temperature of PLA diminishes as the molecular weight diminishes (Xiang et al., 2016).

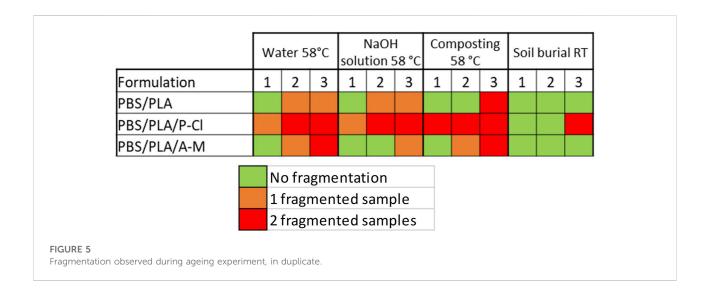
For thermogravimetric analyses, as the IL contents are only 1%, we have assumed their own thermal stability is not influencing significantly the blends behavior. PBS/PLA blend with ammonium IL exhibits slightly higher thermal degradation temperatures (+11.9°C for PLA, +5°C for PBS), which is consistent with a higher molecular weight of PLA measured earlier, and a possible transesterification phenomenon. This could also be due to good interfacial interactions between the ammonium IL and the polyesters, which increase the activation energy necessary to cause degradation (Lins et al., 2015). Therefore, thermogravimetric analysis, like molecular weight measurements, show that the phosphonium IL degrades more

TABLE 5 Glass transition, melting, and crystallization temperatures of PBS, PLA, PBS/PLA (60:40 phr), PBS/PLA/P-Cl and PBS/PLA/A-M (60:40:1 phr) films determined using DSC (second heating/cooling ramp) before exposure. \*\*Hardly visible peak.

Formulation	Tg, PBS (°C)	Tg, PLA (°C)	Tm, PBS (°C)	Tm, PLA (°C)	Tc (°C)
PBS	-29.2	x	116.0	x	92.4
PLA	x	60.2	X	177.1	95.1**
PBS/PLA	-29.4	62.2	114.7	176.0	91.4
PBS/PLA/P-Cl	-25.4	62.7	115.4	169.5	81.6-96.1
PBS/PLA/A-M	-28.4	62.4	114.4	175.4	92.4

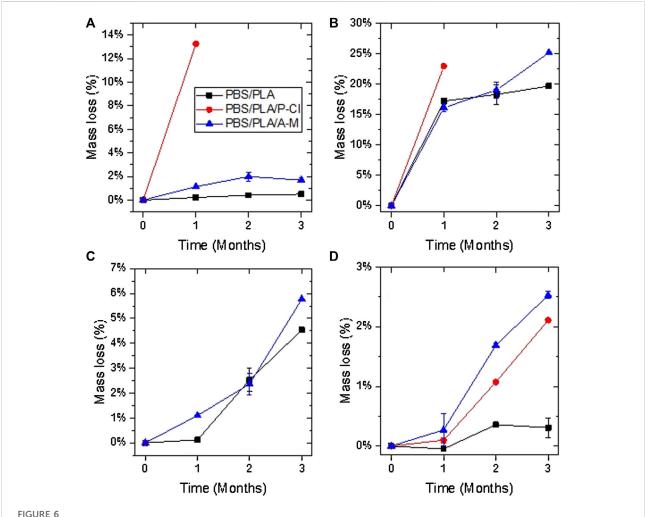


Melting (first ramp (A); second ramp (B)) and crystallization (C) curves of PBS, PLA, PBS/PLA (60:40 phr), PBS/PLA/P-Cl and PBS/PLA/A-M (60:40 phr) films determined using DSC (heating/cooling rate of 10°C.min<sup>-1</sup> under nitrogen atmosphere) before exposure.



specifically PLA phases during processing, which can no more play the role of a reinforcing agent: it can be assumed that PLA chains were degraded to an extent where PLA domains could not provide stiffness to the blend. Ammonium ionic liquid, however, leads to a better stability of the blend.

In PBS/PLA blends, glass transition temperatures appear to be difficult to determine using DSC, because, in the case of PBS, they are wide transitions, and in the case of PLA, they are close to a low intensity melting endotherm of PBS, known in the literature as an annealing peak (Wang et al., 2007). However,



Weight loss in deionized water at 58°C (A), in NaOH solution at 58°C (B), under composting conditions at 58°C (C), and under soil burial conditions at room temperature (D). Results are given with standard deviation. If not shown, only one sample could be recovered and weighted.

we can still observe similar tendencies as the ones observed for alpha relaxation temperatures measured using dynamic mechanical analysis: the Tg of PBS increases by 4°C with the addition of ILs P-Cl, by 1.8°C with IL M-A, whereas, the Tg of PLA phases do not show significant changes (Table 5). As discussed earlier, the shift of glass transition temperature of PBS can be due to a partial miscibility of PLA oligomers in the PBS matrix. Indeed, it has been seen in the literature, that PBS and PLA can exhibit partial miscibility in the amorphous phases to a small extent (Deng and Thomas, 2015; Ostrowska et al., 2019).

Melting and crystallization curves (second ramp) are plotted in Figure 4. Both polymers lead to two melting peaks looking like those of each neat polymer (Bhatia et al., 2007). Melting peak appearing at Tm(PBS) exhibits a more pronounced separation with ILs at second heating ramp. This peak division is typical of PBS melting behavior, and usually appears after crystallization at

temperatures above 80°C, with the generation of crystals that have different thermal stability (Wang et al., 2007). Here, adding IL in PBS/PLA blends seems to have the same effect on the crystallinity of PBS as an annealing treatment. This is in agreement with a certain affinity of ILs with PBS. Regarding PLA, melting peak of PLA in PBS/PLA/P-Cl blend appears at lower temperature than for neat PBS/PLA blend on the second heating ramp, suggesting the presence of small crystallites, probably due to low molecular weight PLA chains that were formed during processing and first heating ramp. The melting curve of PBS/PLA blend with ammonium IL is very similar to PBS/PLA without additive, except for a slight separation of melting peak of PBS.

It is well-known that the use of IL-modified nanoparticles may induce a heteregenous nucleation effect as well as a lamellar ordering effect in the polymer matrix (Du et al., 2007; Xie et al., 2016). But in this case, we have used only 1 wt% of ILs (without

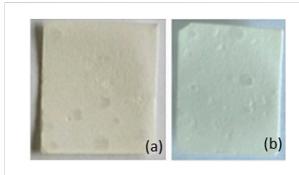


FIGURE 7 PBS/PLA/A-M after 1 (A) and 3 months (B) in alkaline medium at 58  $^{\circ}\text{C}.$ 

nanoparticles) and we have demonstrated in a previous paper that a small amount has no significant effect on the crystallization temperature of PBAT matrix (Livi et al., 2014).

Neat PBS/PLA blend shows two crystallization peaks, a broad one looking similar to the melting peak of PBS alone, and a small peak that merges and appears at higher temperature. PBS/PLA blend with ammonium IL shows a similar curve, although the small peak appearing at higher temperature is more pronounced, yet not completely separated. Phosphonium IL induces a crystallization peak division. The larger peak, appearing at lower temperature might be PBS crystallization, while the small peak could be associated with PLA crystallization. Another explanation can be that PLA crystallites act as nucleating agents for PBS once they are formed.

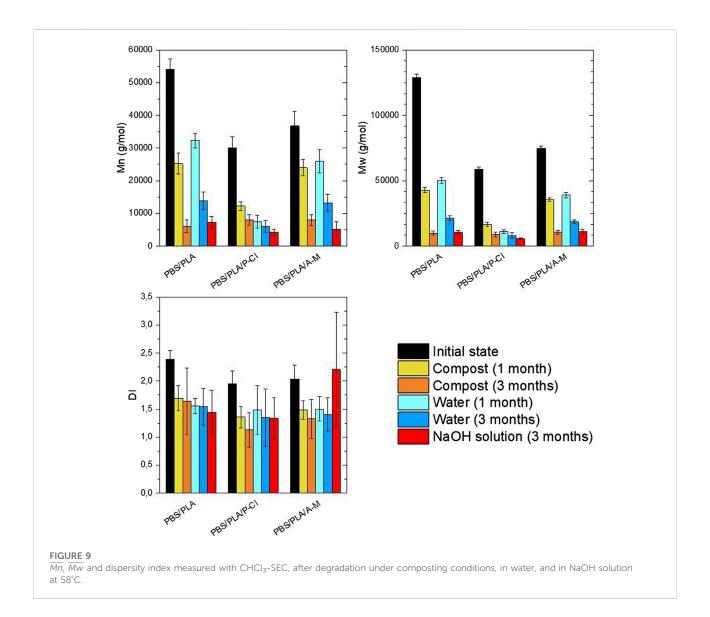
# Degradation in water, soda solution, compost and soil

#### Visual observations and weight loss

Fragmentation of samples is reported in Figure 5. After 1 month in deionized water and in NaOH solution at 58°C,

	PBS	PLA	PBS/PLA	PBS/PLA/P-Cl	PBS/PLA/A-M
Initial time to					
Compost 58°C 2 months					
Compost 58°C 3 months	V)				
Soil burial, RT, 3 months				S. Comments	

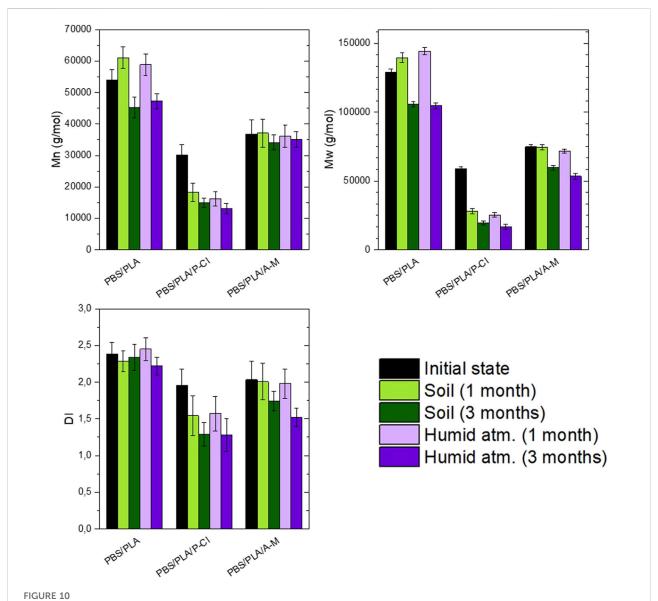
FIGURE 8
Photographs of the samples at initial time, in compost at 58°C after 2 and 3 months, in soil at RT after 3 months.



one of the two samples of PBS/PLA blends with phosphonium IL was fragmented, whereas both samples of neat blend and PBS/ PLA with ammonium IL maintained their integrity. In alkaline medium, PBS/PLA blend and PBS/PLA/A-M exhibit similar weight loss (Figure 6B) except on the third month, where the blend with ammonium IL shows a higher weight loss, while blends with P-Cl exhibit a more important weight loss in the first month (more than 20% weight loss). Data could not be collected for blend PBS/PLA/P-Cl after 2 months because not all the fragments could be recovered. In deionized water, it is observed that PBS/PLA blends containing ionic liquids lose more weight than neat PBS/PLA blend (Figure 6A). This weight loss dramatically increases with phosphonium IL, and is slightly higher with ionic liquid A-M. Regarding the surface aspect, circles are observed on the surface of samples of PBS/ PLA/A-M after degradation in NaOH solution, which is less

pronounced on neat PBS/PLA blend (Figure 7). This can indicate a preferential degradation at weak points. Figure 8 shows that samples containing P-Cl undergo enhanced visual degradation, both in compost at 58°C for 2 & 3 months as well as in soil at RT, for 3 months.

Under composting conditions at 58°C, all samples with phosphonium IL were fragmented after 1 month. After 2 months, one of the two samples with A-M was fragmented. After 3 months, all blends showed fragmentation. Weight loss (Figure 6C) of the blends with ammonium ionic liquid is similar to the reference without ionic liquid. Samples with P-Cl could not be weighed because of their advanced stage of fragmentation, making it impossible to recover all the fragments. Hence, PBS/PLA blend with phosphonium IL degrade faster than PBS/PLA and PBS/PLA/A-M at 58°C. This blend undergoes the most important degradation during processing due to the addition



Mn, Mw and dispersity index measured with CHCl<sub>3</sub>-SEC, after degradation under soil burial conditions, and in humid atmosphere at room temperature.

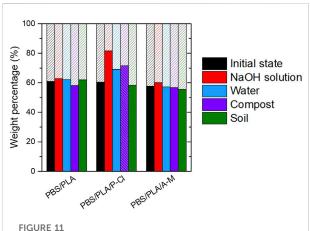
of IL, as discussed earlier. Since the initial molecular weight of blends with P-Cl is lower, integrity is lost more rapidly during random chain scission caused by hydrolysis. Furthermore, it is well known that PLA degrades well at 58°C (Agarwal et al., 1998; Itävaara et al., 2002; Yagi et al., 2009). Since it was observed, here, that PLA domains were degraded the most by the ionic liquid during processing, shorter chains can constitute weak points in the matrix.

Under soil burial conditions at room temperature, only PBS/PLA blend with phosphonium IL undergoes fragmentation after 3 months. However, PBS/PLA/A-M shows a slightly higher weight loss (Figure 6D). It is not clear yet if exudation of

ionic liquids occurs, and if so, at which extent. In humid atmosphere, cracks appeared on PBS/PLA/P-Cl blend after 3 months, indicating that samples became brittle.

#### Evolution of molecular weights

Molecular weights analyzed using size exclusion chromatography are reported in Figures 9, 10. Since it is not possible to distinguish PLA and PBS phases on SEC chromatograms, the dn/dc of neat PBS (0.06) was used to calculate the average molecular weight of both polymers. The resulting  $\overline{M_n}$  are used to assess the trends, since these ones are semi-quantitative values.



Weight proportions of PBS and PLA determined by 1H NMR titration in CDCl<sub>3</sub> after 3 months-degradation experiments (\*except PBS/PLA/P-Cl after 2 months-composting experiment). Plain columns: PBS wt%, hatched columns: PLA wt%.

Number average molecular weight of PBS phases was determined by end group analysis using <sup>1</sup>H NMR spectroscopy. As said previously, these molecular weights might be underestimated, since it is assumed that PBS chains are not branched.

After experiments at 58°C, under composting conditions and in deionized water, molecular weight loss is significant from the first month and does not importantly decrease afterwards for PBS/PLA blends with phosphonium IL, suggesting that  $\overline{M_n}$  reaches a limit of about 10 000 g/mol. Beyond this limit, polymer chains might diffuse to the medium. For PBS/PLA and PBS/PLA/A-M, the average molecular weight decreases significantly in the first month and in the third month as well. Polydispersity decreases during degradation. The molecular weight loss under composting conditions is similar to its loss in deionized water, suggesting that abiotic hydrolysis is the predominant degradation mechanism under composting conditions. If biotic degradation occurs, it does not affect the heart of the sample (surface etching), or it induces ester bond breaking at chain ends, rather than random chain scission. A decrease of dispersity index is observed, which is due to long chains undergoing chain scission.

Under soil burial conditions and in humid atmosphere (Figure 10), at room temperature, PBS/PLA blends with phosphonium IL exhibit a molecular weight loss in the first month, whereas PBS/PLA and PBS/PLA/A-M show no sign of degradation during the first month. In the third month, all blends exhibit molecular weight loss. Molecular weight loss in soil is similar to its loss in humid atmosphere, suggesting that the predominant degradation mechanism under soil burial conditions is abiotic hydrolysis. The more molecular weights decrease, the more the dispersity

index decreases, which is a consequence of bond breaking in long chains, as said earlier.

#### Molecular scale analysis

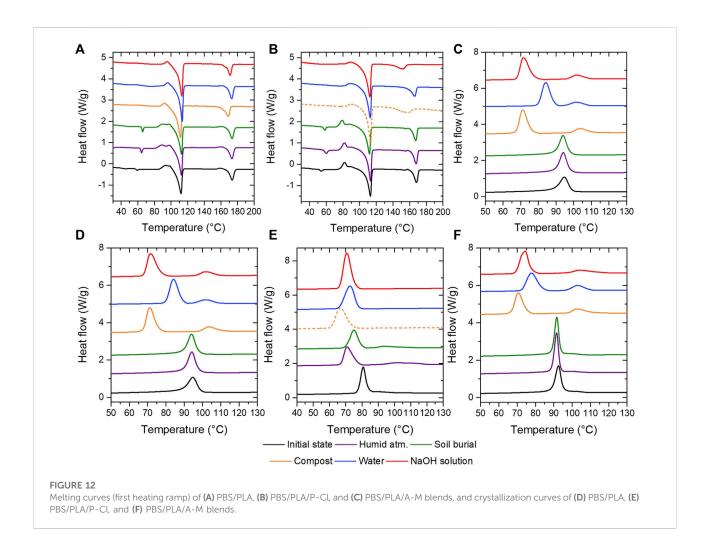
After 3 months, samples were analyzed in CDCl<sub>3</sub> using <sup>1</sup>H NMR spectroscopy, to observe the evolution of PBS/PLA proportions (Figure 11). It can be seen that, after 3 months in deionized water and under composting conditions at 58°C, weight proportions of PBS and PLA in PBS/PLA/P-Cl changed from 60:40 to about 70:30, and to about 80:20 in alkaline conditions, and remained unchanged under soil burial conditions. PBS/PLA and PBS/PLA/A-M did not undergo a change in weight proportions. This result is consistent with the assumption that PLA domains degraded faster than PBS, and diffused out of the matrix at 58°C.

Melting and crystallization behavior of PBS and PLA based blends were studied by using differential scanning calorimetry (Figure 12). Melting temperature of PLA in blend PBS/PLA/P-Cl considerably decreases after degradation at 58°C in water (loss of roughly 30°C) and is absent in alkaline conditions. This drop has been observed by Pantani et al. and is certainly attributed to the low molecular weight chains (Pantani and Sorrentino, 2013). The evolution of melting temperatures of PLA and PBS in blend PBS/PLA/A-M is similar to the reference without additive, i.e. a slight decrease of melting temperature of PLA is observed after degradation under composting conditions and in alkaline medium. This confirms that PLA phases are importantly degraded in PBS/PLA/P-Cl blends during degradation experiments at 58°C, which can be observed through the decrease of melting temperature of PLA.

Crystallization of PBS/PLA blends was studied using a 10°C/min cooling ramp. PBS/PLA without additive showed no change of crystallization behavior after degradation at room temperature (under soil burial conditions and in humid atmosphere). The crystallization temperature remains unchanged and is about 94°C. However, after degradation at 58°C, a peak separation is observed. A first peak, seen at about 103°C, probably corresponds to PLA crystallization, and a second peak at lower temperature, corresponds to PBS crystallization. A second peak reaches a maximum at about 71°C after degradation experiment in alkaline water and under composting conditions, while it is at about 84°C after degradation in deionized water. A similar behavior is observed with PBS/PLA/A-M formulation.

PBS/PLA/P-Cl shows a more pronounced crystallization peak separation after degradation in humid atmosphere and under soil burial conditions (Figure 12). PLA melting peak appears to be broad and flat. After degradation at 58°C, this peak is hardly observable. As seen earlier, this is due to PLA degrading and diffusing out of the PBS matrix.

Finally, evolutions of weight losses, molecular weights show that abiotic degradation can be associated with possible biodegradation in presence of microrganisms and that both are enhanced for blends containing P-Cl.



#### Conclusion

In this study, PBS/PLA blends were processed with ionic liquids P-Cl and A-M, that could act as interfacial agents between PBS and PLA phases. Surprisingly, it was seen that PLA phases in the PBS matrix were considerably larger in PBS/PLA blends with additives than in neat PBS/PLA blend. Furthermore, it was seen, through molecular weight measurements, that phosphonium ionic liquid induced a significant decrease of molecular weight, leading to the formation of PLA oligomers, which induces a lower thermal stability. It had already been observed earlier that phosphonium ionic liquid catalyzes PLA degradation during processing (Park and Xanthos, 2009; Lins et al., 2015). Consequently, PLA oligomers do not provide stiffness to the PBS matrix in PBS/PLA/P-Cl blend. Then, the slight merging of alpha relaxation temperatures, observed in DMA, suggests that small chains of PLA might be miscible in PBS phases, and vice versa. Blend with ammonium IL exhibits a higher thermal stability, and improved mechanical properties, assessing the role of interfacial agent of A-M. The mechanism is not established yet, although the

increase of  $\overline{M_n}$  of PLA after processing could suggest a possible transesterification reaction.

Degradation experiments conducted at  $58^{\circ}\text{C}$  indicate that the blends degrading faster are the blends undergoing the most important degradation during processing (PBS/PLA/P-Cl>>>PBS/PLA/A-M  $\geq$  PBS/PLA). It was also observed that PLA domains were affected the most during degradation, through the decrease of melting temperature and PLA weight percentage, which, macroscopically, leads to a significant weight loss and to fragmentation. It can easily be assumed that PLA phases play the role of weak points in the matrix, leading to a rapid loss of integrity. This shows that "predegraded" phases in a polymer matrix can accelerate macroscopic degradation, without compromising the whole material's uses.

However, PLA phases were not affected to the same extent after degradation at room temperature, which was expected since it is well known that PLA do not easily degrade under mesophilic conditions (Agarwal et al., 1998; Itävaara et al., 2002; Yagi et al., 2009). At room temperature, no significant change was observed using DSC, hence it is not clear yet whether any polymer

constitutes a weak point. <sup>1</sup>H NMR results show no serious change in PBS/PLA proportions. Nevertheless, under soil burial conditions, all blends containing either IL show a more important weight loss than neat PBS/PLA blend.

Regarding SEC measurements, it is observed that the molecular weight loss is very similar in abiotic and biotic conditions at same temperatures, which suggests that hydrolysis is the main degradation mechanism in the studied polyester blends.

This study shows that ionic liquids can be used to tune mechanical properties of polymer blends as well as their (bio) degradability. However, further investigation is needed to understand degradation mechanisms at room temperature. Moreover, it is necessary to identify degradation products and to study their eco-toxicity. Moreover, potential toxicity of ILs has been raising interest in the literature indeed (Zhao et al., 2007; Petkovic et al., 2010; Thuy Pham et al., 2010; Ventura et al., 2012; Hou et al., 2013; El-Harbawi et al., 2014; Ventura et al., 2014; Radošević et al., 2015). Affinity with microorganisms of the tested IL is under study. It can be assumed, through a change in morphology, that ILs induce surface modification, and hence affect interactions of the material with microorganisms.

#### Data availability statement

The original contributions presented in the study are included in the article/Supplementary Material, further inquiries can be directed to the corresponding author.

#### **Author contributions**

Conceptualization, SL and VM; Methodology, SL, AM, ED, VM, RB, and J-FG; Formal Analysis, AM, ED, VM, SL, and J-FG;

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#### Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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