

## Ageing of copper, zinc and synthetic pesticides in particle-size and chemical fractions of agricultural soils

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1	Ageing of copper, zinc and synthetic pesticides
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#### 26 **Abstract**

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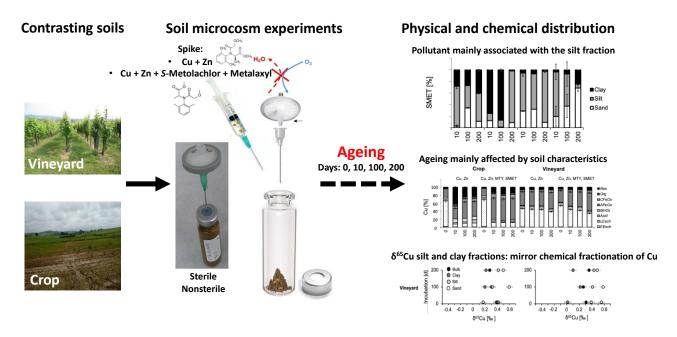
The transformation and mobility of heavy metals and synthetic pesticides in soil depend on ageing, involving their chemical and physical distributions among soil fractions over time. Heavy metals and synthetic pesticides often co-occur in soil, although their ageing is usually evaluated separately and in bulk soil. Here, contrasting vineyard and crop soils were spiked with copper (Cu; 700 mg kg<sup>-1</sup>) and zinc (Zn; 200 mg kg<sup>-1</sup>) and/or synthetic pesticides (5 mg kg<sup>-1</sup>), i.e., the fungicide metalaxyl (MTY) and herbicide S-metolachlor (SMET), to evaluate within 200 days their distribution among soil physical and chemical fractions. More than 90% of MTY and SMET in soil was released into water, even 200 days after spiking. MTY, SMET, Cu and Zn were associated mainly with the silt fraction and to a lesser extent 36 (<30%) with the sand and clay fractions. Overall, the ageing of MTY, SMET, Cu and Zn in agricultural soil was affected by the soil type and sterilization and only to a minor extent by their co-occurrence. Sorption controlled the dissipation of MTY and SMET in soil, while biodegradation contributed to <10%. Cu was mainly (37  $\pm$  2%) associated with Fe oxides after 200 days, while Zn was found (33  $\pm$  2%) in the residual fraction. The silt fraction of the nonsterile vineyard soil became enriched in  $^{65}$ Cu over time ( $\Delta\delta^{65}$ Cu = 0.25 ± 0.07 %), whereas the clay fraction was depleted in  $^{65}$ Cu ( $\Delta\delta^{65}$ Cu =  $-0.20 \pm 0.07$  ‰). Cu isotope fractionation mirrored the Cu distribution in soil chemical fractions, suggesting that Cu stable isotopes may help to follow-up Cu ageing. In contrast, no significant Zn isotope fractionation was observed among experiments or over time. Overall, our study emphasizes the variability in ageing of synthetic pesticides and heavy metals co-occurring in agricultural soils and their interplay in soil fractions. **Keywords:** agricultural soils; heavy metals; pesticides; ageing; particle-size fractions;

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sequential chemical extraction; stable isotope analysis.

# **Highlights:** MTY, SMET, Cu and Zn were associated (>40%) with the silt fraction of agricultural soils MTY, SMET, Cu and Zn distribution among soil fractions reflected ageing within 200 days Sorption is a major process of MTY and SMET dissipation in crop and vineyard soils 55 • 56 • MTY, SMET, Cu and Zn ageing was mainly affected by soil texture and organic content $\delta^{65}\text{Cu}$ values in the silt and clay fractions of the soil mirrored chemical fractionation of Cu 57 •

### **Graphical abstract**



#### 1. Introduction

The use of fertilizers, pesticides and sewage sludge has led to mixtures of synthetic pesticides and heavy metals in agricultural soils. Synthetic pesticides and heavy metals often co-occur in soil and can adversely affect nontarget organisms (Imfeld and Vuilleumier, 2012; Muturi et al., 2017; Wyszkowska et al., 2012). Vineyard soils are of particular concern because synthetic pesticides, inorganic copper (Cu) fungicides and zinc (Zn) amendments are regularly applied (Komárek et al., 2010a; Imfeld et al., 2020). The physical and chemical distribution and diffusion of synthetic pesticides and heavy metals over time in soil porewater, minerals and organic matter are referred to as ageing (Komárek et al., 2010b; Ma et al., 2006). Ageing depends on both the physicochemical characteristics of the soil and pollutant properties (Komárek et al., 2010b; Swarcewicz and Gregorczyk, 2012; Zhang et al., 2014). Ageing increases pollutant sequestration and persistence, and reduces pollutant availability and toxicity (Hatzinger and Alexander, 1995). Ageing also controls pollutant export from agricultural soil since rainfall patterns influence the preferential mobilization of specific soil fractions (Babcsányi et al., 2016; Meite et al., 2018). However, the concomitant ageing of synthetic pesticides and heavy metals in agricultural soils remains poorly understood.

Dissolution, precipitation, complexation, sorption, diffusion and entrapment control the sorption and ageing of synthetic pesticides and heavy metals in soil. The sorption of synthetic pesticides involves hydrophobic binding to soil organic matter (SOM) and/or oxides and clay minerals (Clausen et al., 2001; Cornejo et al., 2008; Đurović et al., 2009; Helal et al., 2006). During ageing, Cu and Zn sorb onto oxides, mainly retained by clay minerals (Komárek et al., 2010b; Ma et al., 2006). The slow diffusion of Cu and Zn in Fe oxides (Bruemmer et al., 1988), Al oxides and Mn oxides, clay minerals (Ma and Uren, 1998) and carbonate precipitation (Nakhone and Young, 1993; Ma et al., 2006) governs their

sequestration in soil. Cu and Zn also present high affinity for organic matter (Duplay et al., 2014; Néel et al., 2007; Quenea et al., 2009). It is worth noting that the ageing of synthetic pesticides and heavy metals is generally examined separately and in bulk soil. However, their concomitant occurrence may affect their distribution among soil fractions due to competition for sorption sites, the formation of complexes between pollutants (e.g., El Azzi et al., 2018), or the alteration of soil biogeochemistry and microbial activities impacting pollutant sorption.

Compared with the physicochemical evaluation of soil fractions, the natural abundance signatures of metal stable isotopes (Wiederhold, 2015) can reflect processes that metals undergo in soil (Wang et al., 2021). In particular, transition metals such as Cu and Zn may exhibit distinct isotopic signatures in soil fractions. Cu isotope ratios (65Cu/63Cu) change during Cu sorption onto Fe and Al oxy(hydr)oxides (Balistrieri et al., 2008a; Pokrovsky et al., 2008) or clay minerals (Li et al., 2015a), oxidation-reduction (Ehrlich et al., 2004; Zhu et al., 2002), uptake by plants (Jouvin et al., 2012; Ryan et al., 2013) or microorganisms (Navarrete et al., 2011), precipitation as Cu mineral phases (Ehrlich et al., 2004; Maréchal and Albarède, 2002), and complexation with organic matter (Bigalke et al., 2010; Ryan et al., 2014).  $\delta^{65}$ Cu values significantly differ between particle-size soil fractions, including both SOM and mineral phases (El Azzi et al., 2013; Vance et al., 2008; Babcsányi et al., 2016). Similarly, Zn stable isotope fractionation has been observed following Zn sorption on carbonates (Schott et al., 2014) and several oxides, including goethite and ferrihydrite Fe oxides (Lv et al., 2020; Juillot et al., 2008), birnessite and pyrolusite Mn oxides, and Al oxides (Pokrovsky et al., 2005).  $\delta^{66}$ Zn also varied during Zn complexation with humic acids (Jouvin et al., 2009) and Zn sorption and bacterial incorporation (Kafantaris and Borrok, 2014). Although signals of isotope fractionating processes of transition metals in soil may overlap, reference experiments on Cu and Zn isotope fractionation during long-term ageing (>100 days) in agricultural soils are missing thus far.

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Here, we designed a laboratory microcosm study on contrasting vineyard and crop soils to investigate the ageing of the widely used fungicides metalaxyl (MTY) and herbicides S-metolachlor (SMET), as well as Cu and Zn. The objective of this study was to examine (i) the influence of soil characteristics on the independent and concomitant ageing of synthetic pesticides and heavy metals and (ii) the stable isotope fractionation of Cu and Zn during the ageing of Cu and Zn in agricultural soils. Experiments with sterile (abiotic) and nonsterile (biotic) soil were incubated separately for 1, 10, 100 and 200 days to recover soil particle-size fractions for sequential chemical extraction and stable isotope analyses of Cu and Zn. We hypothesized that (i) the ageing of synthetic pesticides and heavy metals in soil differs if they co-occur or not, and (ii) the distribution of Cu and Zn among soil fractions during ageing is associated with specific stable isotope fractionating processes in agricultural soil.

#### 2. Material and methods

#### 2.1. Chemicals

Metalaxyl ( $C_{15}H_{21}NO_4$ , methyl N-(methoxyacetyl)-N-(2,6-xylyl)-DL-alaninate) and S-metolachlor ( $C_{15}H_{22}ClNO_2$ , (S)-2-chloro-N-(2-ethyl-6-methyl-phenyl)-N-(1-methoxypropan-2-yl)acetamide) were purchased from Sigma–Aldrich (St. Louis, MO, USA), with purities of 99.8% and 98.2%, respectively. The physicochemical characteristics of MTY and SMET are provided in Table S1. Copper (CuCl<sub>2</sub>) and zinc (ZnCl<sub>2</sub>) chlorides for soil spiking and salts for preparing artificial rainwater were purchased from Sigma–Aldrich (St. Louis, MO, USA) with a purity  $\geq$ 97%. Ultrapure water and dichloromethane (DCM), BaCl<sub>2</sub> (0.1 M), Pb(NO<sub>3</sub>)<sub>2</sub> (0.05 M), CH<sub>3</sub>COONa (1 M), NH<sub>2</sub>OH·HCl (0.01 M), H<sub>2</sub>C<sub>2</sub>O<sub>4</sub> (0.1 M), Na<sub>2</sub>C<sub>2</sub>O<sub>4</sub> (0.175 M) and Na<sub>4</sub>P<sub>2</sub>O<sub>7</sub> (0.1 M) for the sequential extraction of soil chemical fractions were purchased from Sigma–Aldrich.

#### 2.2. Experimental soils

A vineyard and crop soil with contrasting physicochemical properties (Table S2) were collected with a distilled water- and ethanol-cleaned shovel on August 6 and 7, 2015, from well-characterized sites. The vineyard soil was collected from a plot in an experimental catchment in Rouffach (Alsace, France) (Babcsányi et al., 2016; Duplay et al., 2014; Imfeld et al., 2020). Fifty kg of topsoil (0 to 5 cm, Hor. A) were collected by random subsampling (>50) at each site, since this soil layer is directly impacted by pesticides. Subsamples of calcareous vineyard topsoil were collected from a 1486 m² plot managed under conventional wine-growing practices across seven vine inter-rows at ten locations in each inter-row. Subsamples of arable crop soil were obtained from three transects in a field site cultivated with sugar beet, corn and wheat at Alteckendorf (Alsace, France) (Lefrancq et al., 2017). Subsamples of the crop soil were collected from a 47-ha crop catchment (corn and wheat) across three transects.

Soil samples were transported in coolers to the laboratory, dried at 30 °C with an airforced oven for soil analysis or at room temperature for further soil incubation, and sieved using a 2-mm mesh. After complete homogenization by mixing four times in quarters, an aliquot of each dried soil was weighed and dried at 105 °C to quantify the soil moisture content. Soil physicochemical characteristics (Table S2) were measured following ISO methods and procedures (Meite et al., 2018; Pérez-Rodríguez et al., 2021). Briefly, the clay-loamy vineyard soil was alkaline (pH 8.1), with a low content of organic matter (1.7%) and a high content of carbonates (27.1%). The crop soil was neutral (pH 7.0), characterized by higher contents of organic matter (5.5%), sand (10.3%) and silt (61.5%) than the vineyard soil.

#### 2.3. Experimental set-up

Seven series of soil microcosm experiments were set up in parallel for both the vineyard and crop soils (Table S3): (i) nonsterile (biotic) soils spiked with MTY and SMET;

(ii) nonsterile soils spiked with Cu and Zn; (iii) nonsterile soils spiked with MTY, SMET, Cu and Zn; (iv) sterile (autoclaved) soils spiked with MTY and SMET; (v) sterile soils spiked with Cu and Zn; (vi) sterile soils spiked with MTY, SMET, Cu and Zn; and (vii) nonsterile and sterile soils adjusted with sterile distilled water only as controls.

The soil microcosms consisted of 60 g of air-dried soil in sterile 100 mL HDPE flasks. A 0.2 µm syringe filter (Rotilabo®, Carl Roth®, France) was mounted on a syringe tip stuck through the vial cap to allow oxic conditions in the vials through permanent soil-atmosphere gas exchange while limiting water loss and avoiding contamination (Torabi et al., 2020). The soil for sterile experiments was autoclaved three times at 12 h intervals. The soil microcosms were prepared under laminar flow with sterile material, as described earlier (Torabi et al., 2020). Briefly, pesticide stock solutions of individual MTY and SMET in DCM were added to distilled water and stirred until complete DCM evaporation. The aqueous solutions of MTY, SMET, CuCl<sub>2</sub>, and ZnCl<sub>2</sub> were filter-sterilized separately for soil spiking.

Experiments with nonsterile and sterile soils were spiked with MTY and SMET at 5 mg kg<sup>-1</sup> and/or Cu at 700 mg kg<sup>-1</sup> and Zn at 200 mg kg<sup>-1</sup>, equivalent to high but environmentally relevant concentrations in agricultural soil (Imfeld et al., 2020; Komárek et al., 2010b). Solution spiking was adjusted to reach a final water content of 20% (w/w). To homogenize the soil microcosms, aqueous pesticide solution, or deionized water as a control, was added between soil layers, i.e., 4 layers of soil and 5 layers of aqueous solution. The microcosms were then vortexed for 30 s.

All experiments were incubated in the dark at 20 °C. Sampling was carried out using a sacrificial approach on days 1 (following an equilibration of 24 h), 10, 100, and 200. In total, 16 experiments were set up, each in independent triplicate microcosms, and for each of the four sampling days, yielding a total of 192 individual soil microcosms (Table S3).

2.17 2.4. Physical separation of soil particle-size fractions

To separate the water and soil fractions for MTY and SMET analysis, soil suspensions with 1:5 bulk soil:ultrapure sterile water were agitated for 24 h at 60 rpm and centrifuged at 750 rpm for 5 min. The obtained supernatant constituted the water fraction. The sand (>63  $\mu$ m), silt (2 and 63  $\mu$ m) and clay (<2  $\mu$ m) soil fractions were separated as described previously (Wang and Keller, 2008). The sand fraction was wet-sieved (63  $\mu$ m) and carefully rinsed with 18.2 M $\Omega$  water. The silt and clay fractions were separated by centrifugation at 750 rpm for 5 min. The obtained supernatant was eventually centrifuged (4000 rpm, 35 min) to collect the clay fraction. The sand, silt and clay fractions were weighed, and the recovery ( $\geq$ 95  $\pm$ 5%) for Cu or Zn was verified. The sand, silt and clay fractions were stored at -4 °C prior to further treatments and quantification of Cu, Zn, MTY and SMET.

#### 2.4.MTY and SMET quantification

MTY and SMET were extracted from water using SPE with SolEx C18 cartridges (1 g phase, Dionex-Thermo Fischer) method described previously (Masbou et al. 2018). Extraction from the particle-size fractions was carried out with ethyl-acetate using a modified solid—liquid extraction protocol (Ivra et al. 2014). Extraction recoveries from water and sediment samples were highly reproducible (SE<10%). MTY and SMET were quantified by gas chromatography (GC, Trace 1300, Thermo Fisher Scientific) coupled with a mass spectrometer (MS, ISQ<sup>TM</sup>, Thermo Fisher Scientific). Recoveries, limits of detection (LODs), limits of quantification (LOQs) and accuracy of GC-MS measurements are provided in Table S4 (Supporting Information).

### 2.5. Sequential chemical extraction of Cu and Zn

The distribution of Cu and Zn among soil chemical fractions was quantified with a sequential extraction procedure adapted from Rao et al. (2008) and summarized in Table S5. Eight fractions were extracted sequentially: (i) easily exchangeable (*EExch*); (ii) less easily exchangeable (*LExch*); (iii) acid-soluble, carbonate-bound (*Acid*); (iv, v and vi) reducible,

Mn- (MnOx), amorphous Fe- (AFeOx) and crystallized Fe-oxide bound (CFeOx); (vii) oxidizable, organic matter-bound (Org); and (viii) residual (Res), associated with the silicate matrix.

Homogenized soil samples (1 g) were extracted with 25 mL of extractant. A solution of BaCl<sub>2</sub> extracted the *EExch*, followed by PbNO<sub>3</sub> and sodium acetate in acetic acid (pH 5) to extract the *LExch* and *Acid* fractions, respectively. The *MnOx*, *AFeOx* and *CFeOx* fractions were extracted with hydroxylamine in hydrochloric acid NH<sub>2</sub>OH·HCl, oxalate H<sub>2</sub>C<sub>2</sub>O<sub>4</sub> and ammonium oxalate (NH<sub>4</sub>)<sub>2</sub>C<sub>2</sub>O<sub>4</sub>, respectively. The *Org* fraction was extracted with sodium pyrophosphate (Na<sub>4</sub>P<sub>2</sub>O<sub>7</sub>). Extraction of amorphous and crystallized Fe oxide-bound metals was carried out in the dark and under UV light, respectively. The *Res* fraction was dissolved twice in 10 mL of bidistilled HNO<sub>3</sub> heated at 200 °C on a hot plate, followed by dissolution in 10 mL of bidistilled HCl and 10 mL of supra-pure HF heated at 100 °C for 24 h. Acids were evaporated at 120 °C and dissolved in 5 mL of HCl 7 M in an ultrasonic bath for 5 min and heated at 70 °C for 12 h.

The fraction obtained after each extraction stage was washed with 20 mL of 18.2 M $\Omega$  cm water with an end-over-end shaker and centrifuged for 30 min at 5000 rpm. The solution containing the extracted elements was acidified with distilled HNO<sub>3</sub> (0.5 M) and stored in polyethylene vials at 4 °C until analysis. The analytical uncertainty for the entire sequential extraction procedure was <10% based on separate triplicate extractions and measurements (Semhi et al., 2014).

### 2.6.Quantification of Cu and Zn

For Cu and Zn quantification, soil samples were first oven-dried (60 °C) and powdered using an agate disk mill (<100 µm) prior to total dissolution in acids under a laminar fume hood in a clean room, as described previously (Babcsányi et al., 2016). Measurements were carried out by ICP-AES (ICAP6500, Thermo Fisher Scientific) with geological standards,

BCR-2 (US Geological Survey, Reston, VA, USA) and SCL-7003 (Analytika, Prague, Czech Republic), for quality control (Babcsányi et al., 2014). The total analytical uncertainty of measurements was  $\pm$  5%, and the detection limit was <3 ppb.

2.7.Cu and Zn isotope analysis and notations

For Cu and Zn isotope analysis, 100 mg of bulk soil, sand, silt or clay powder (<63  $\mu$ m) was weighed in PFA vials. Liquid samples were evaporated to reach at least 200 ng of Cu and Zn. All samples were dissolved in successive acid baths: HF-HNO<sub>3</sub> acid (v:v = 4:1) at 70 °C, HClO<sub>4</sub> acid at 150 °C and a mixture of HCl and H<sub>3</sub>BO<sub>3</sub> at 110 °C.

Protocols for double Cu and Zn purifications were adapted from Maréchal et al. (1999) and Babcsányi et al. (2014) for Cu and from Moynier et al. (2009) for Zn (Table S6). Each Cu purification step was followed by sample digestion in 1 mL of HNO<sub>3</sub> to mineralize the organic matter recovered from the AGMP1 resin. Purified samples and standards were dissolved in HNO<sub>3</sub> 0.5 M to reach 100  $\mu$ g L<sup>-1</sup> of Cu and 200  $\mu$ g L<sup>-1</sup> of Zn. The total procedure contributed to  $\leq$ 1% of the total Cu and Zn content in the soil samples (data not shown).

Cu and Zn isotope ratios were measured with a Finnigan-Neptune MC-ICP-MS (Thermo Scientific, Waltham, MA, USA) using a Ni standard (NIST SRM 986) for Cu and a Cu standard (NIST SRM 976) for Zn. For Cu and Zn isotope ratio measurements, the Cu standard (NIST SRM 976) and Zn standard (IRMM 3702) were used for correction of instrumental mass biases as previously described (Li et al., 2009). Each measurement of stable isotope ratios consisted of 40 (Cu) and 30 (Zn) cycles of 8 s of integration.

The total analytical uncertainty for digestion, purification and isotope analysis was  $\leq 0.05 \% (2\sigma, n=15)$  for  $\delta^{65}$ Cu values and  $\leq 0.09 \% (2\sigma, n=15)$  for  $\delta^{66}$ Zn values. The total analytical uncertainty was determined using the soil reference material CRM-7003 ( $\delta^{65}$ Cu =  $0.18 \pm 0.05\%$  and  $\delta^{66}$ Zn =  $-0.25 \pm 0.09\%$ , including 3 samples digested separately and 15 isotope measurements), in agreement with the literature (Bigalke et al., 2010). Cu and Zn

isotope ratios were expressed as  $\delta^{65}$ Cu and  $\delta^{66}$ Zn values [‰] relative to the NIST 976 and IRMM 3702 reference materials, respectively.

2.8.Data analysis

#### 2.8.1. Pesticide dissipation and biodegradation

The contribution of abiotic and biotic dissipation of MTY and SMET in soil was estimated by comparing dissipation in nonsterile (biotic) soils and in sterile (abiotic) soils. The dissipated fractions [%] of pesticides after 200 days in sterile soils ( $D_{abiotic, 200}$ ) define the contribution of ageing and abiotic degradation, while dissipation in nonsterile experiments ( $D_{biotic, 200}$ ) additionally features biotic degradation. Thus, the extent of biodegradation ( $B_{diss}$ ) [%] of MTY or SMET at 200 days was estimated as the difference between  $D_{abiotic, 200}$  and  $D_{biotic, 200}$ .

#### 2.8.2. Statistical analyses

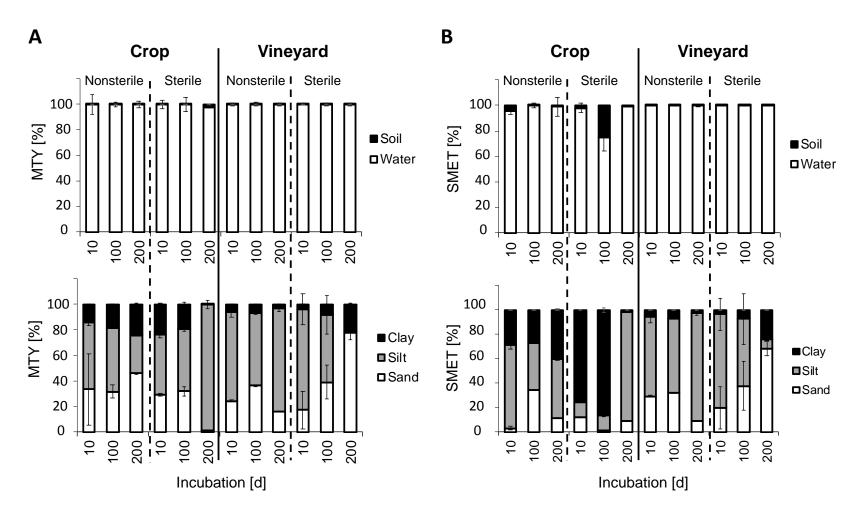
Statistical tests were performed using the R program (Copyright 2005, The R Foundation for Statistical Computing, Version 2.15.1). Nonparametric Wilcoxon and Kruskal–Wallis tests were applied to compare the ageing of MTY, SMET, Cu and Zn according to the soil type, sterilization or distribution in soil fractions.

Cluster analysis was used to visualize dissimilarities between the distribution patterns of MTY, SMET, Cu and Zn in the particle-size fractions of soil and of Cu and Zn in the chemical fractions. The distance between distribution patterns was calculated with the Bray-Curtis dissimilarity index. A hierarchical cluster analysis was performed on the resulting dissimilarity matrix using Ward's method (Ward, 1963).

#### 3. Results and discussion

3.1. Distribution of MTY and SMET among particle-size fractions during ageing

More than 96% of residual MTY and SMET was released from the soil into the water phase in all experiments and after all incubation times (Figure 1). The high solubility in water and low sorption propensity of MTY and SMET (Table S1) favoured partitioning into the water phase at all incubation times. Only a minor and constant portion of MTY and SMET remained associated with soil over time. Since the water phase did not include soil particles, MTY and SMET were mainly freely dissolved and associated to DOC. This emphasizes the potential of MTY and SMET mobilization from agricultural soils into runoff water and further export to aquatic ecosystems, even 200 days after pesticide application. However, the distribution of pesticides among particle-size fractions of the soil may also affect pesticide export into runoff since the rainfall pattern determines the preferential export of specific soil fractions (Martínez-Casasnovas et al., 2002; Mohamadi and Kavian, 2015; Meite et al., 2018; Imfeld et al., 2020).



**Figure 1**. Distribution of (A) metalaxyl (MTY) and (B) *S*-metolachlor (SMET) among fractions of crop and vineyard soils, including water, sand ( $x>50 \mu m$ ), silt ( $2< x<50 \mu m$ ) and clay ( $x<2 \mu m$ ), in nonsterile and sterile experiments. The distribution of MTY and SMET among soil fractions does not reflect their dissipation extent (see section 3.2). Error bars denote 95% confidence intervals.

The MTY and SMET distribution among particle-size fractions largely varied among conditions but did not change significantly (*p*>0.1) with ageing or soil type in nonsterile experiments (Figure 1). However, both MTY and SMET dissipated in soil over time (see section 3.2). Overall, MTY and SMET were mainly associated with the silt fraction in both soil types, although the silt content was approximately three times lower in the vineyard soil (23.1%) than in the crop (61.5%) (Table S2). In the silt fraction, both MTY and SMET slightly increased over time in nonsterile experiments with vineyard soil. Differences in both the content and nature of SOM and minerals among particle-size fractions can alter the distribution of MTY and SMET. The SOM content generally increases with decreasing particle size (Guo et al., 2010; Quenea et al., 2009; Schmidt and Kögel-Knabner, 2002), which may favour MTY and SMET sorption in the silt and clay fractions. In addition, the proportion of particulate organic matter (POM, >50 μm) and mineral-associated organic matter (MOM, <50 μm) constituting SOM is expected to differ among particle-size fractions in crop and vineyard soils (Benoit et al., 2008; Guo et al., 2010; Moyo et al., 2014; Tirloni et al., 2012).

We thus hypothesize that an intermediate degree of SOM alteration and the specific POM/MOM ratio in the silt fraction favoured the sorption of nonionic pesticides, such as MTY and SMET (Figure 1). In particular, less degraded SOM with a medium-density organic fraction and a lower POM/MOM ratio in the silt than in the sand fraction may favour MTY and SMET sorption in the silt fraction of the vineyard soil (Besnard et al., 2001; Parat et al., 2002; Quenea et al., 2009). Interactions with minerals in the silt and clay fractions can further stabilize SOM in the form of organomineral complexes (Mertz et al., 2005; Poirier et al., 2005; Schmidt and Kögel- Knabner, 2002) and pesticide-SOM complexes. These results are in agreement with previous studies showing fast incorporation of metalaxyl in silt- and clay-

organic matter (Kalathoor et al. 2015). The binding mechanism may include hydrogen and ionic bonds between the oxygen-rich metalaxyl residues and the soil humic surface. In addition, nonionic pesticides can sorb onto soil oxides, especially on clay minerals (Clausen et al., 2001; Cornejo et al., 2008; Moyo et al., 2014; Remucal and Ginder-Vogel, 2014). This may explain the slight increase in MTY and SMET in the clay fraction of the crop soil after 200 days of ageing (Figure 1).

In comparison with the nonsterile experiments, both MTY and SMET significantly increased over time in the silt and sand fractions of sterile crop and vineyard soils, respectively. Sterilization by autoclaving affected the distribution of MTY and SMET among soil fractions, presumably due to changes in the availability of soil-binding sites and SOM alteration. Autoclaving results in physical aggregate fractionation, decreasing soil aggregation and increasing the clay fraction (Berns et al., 2008). The lysis of microorganisms and subsequent transformation of microbial carbon also alter the SOM structure. Thus, alteration of SOM and lower soil aggregation following autoclaving likely favoured ageing of both MTY and SMET in the sand fraction in sterile experiments with vineyard soil (Figure 1). Indeed, the sand fraction generally contains higher POM, which is known as a light-density organic fraction and the most accessible SOM fraction for pesticides (Quenea et al., 2009). Overall, higher aggregate fractionation and SOM alteration, together with changes in POM/MOM ratios across soil fractions, led to distinct distributions of MTY and SMET among soil fractions in the nonsterile and sterile experiments.

#### 3.2. Dissipation of MTY and SMET in crop and vineyard soils

Ageing also controlled the dissipation of MTY and SMT in agricultural soils. The extent of dissipation by irreversible sorption and biodegradation was estimated based on differences between the dissipated fraction of MTY and SMET in nonsterile and sterile

experiments (Table 1). Overall, soil type, sterilization and ageing did not significantly (p>0.1) affect MTY and SMET dissipation in soil. For all experiments and based on concentration decrease, more than 90% of MTY and SMET dissipated within 200 days (Table 1). The large uncertainties associated with dissipation estimates in the crop than in the vineyard soil may reflect higher heterogeneity of biological, physical and chemical characteristics. The contribution of MTY and SMT volatilization, dilution, plant uptake, export or photodegradation can be safely discarded in our setup. Hence, biodegradation in the nonsterile experiments ranged from <8 to <72 given the uncertainty associated with dissipation estimates and generally accounted for dissipation of less than 10% (Table 1). Sorption was thus identified as the major process responsible for MTY and SMET dissipation in all experiments.

**Table 1**. Dissipated fractions of pesticides in nonsterile ( $D_{biotic,t}$ ) and sterile soils ( $D_{abiotic,t}$ ), along with the extent of biodegradation ( $B_{diss}$ ) at 200 days.

	Стор			Vineyard		
	$D_{biotic}$ [%] $\pm { m SE}$	$egin{aligned} D_{abiotic}  [\%] \ \pm \mathrm{SE} \end{aligned}$	<i>B<sub>diss</sub></i> [%] ±SE	$D_{biotic}$ [%] $\pm { m SE}$	$D_{abiotic} [\%] \ \pm { m SE}$	<i>B<sub>diss</sub></i> [%] ±SE
MTY	$92 \pm 68$	$96 \pm 23$	<72	$97 \pm 7$	96 ± 7	<10
SMET	$94 \pm 23$	$95 \pm 4$	<23	$99 \pm 4$	$93 \pm 7$	<8

The contribution of sorption to the overall MTY and SMET dissipation is expected to increase with lower water content in soil. The low (20%) water content in crop and vineyard soils limited sorption-desorption, MTY and SMET bioavailability and microbial activity, thus decreasing the contribution from biodegradation. In contrast, with a lower water content, the fraction of nonextractable residues (NER) may have increased, resulting in an apparent dissipation of MTY and SMET (Rodríguez-Liébana and Peña, 2020). It can be hypothesized that a lower water content enhanced intraparticle diffusion over time through inner soil pores of more soluble and less hydrophobic pesticides, such as MTY and SMET (solubility >400

mg L<sup>-1</sup> and K<sub>dsoil</sub><2.5 mL g<sup>-1</sup>; Table S1). In turn, this reduced MTY and SMET bioavailability and biodegradation while increasing their half-lives in the soil. Sorption was thus the main dissipation process controlling MTY and SMET extraction and dissipation during ageing in soils with low water content. Our results also emphasize the persistence and strong binding of pesticide NER in dryer soils (Odukkathil and Vasudevan, 2013) after 200 days, even for soluble and nonionic pesticides, such as MTY and SMET.

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#### 3.3. Distribution of Cu and Zn among particle-size fractions during ageing

Cu partitioning between soil and water was mainly affected by soil sterilization and ageing, whereas Zn partitioning only slightly varied during ageing. The fraction of Cu released into the water phase (36  $\pm$  2%) was three times higher than that of Zn (12  $\pm$  1%) in nonsterile experiments (Figure 2). Partitioning of both Cu and Zn into water only slightly decreased between 10 and 200 days of ageing. Overall, this emphasizes the potential release of Cu- and Zn-DOM complexes from agricultural soils into the aqueous phase during Cu and Zn ageing. The formation and dominance of Cu- and Zn-DOM complexes at pH>7 in aqueous solution was supported by speciation modelling using Minteq with Nica-Donnan and Stockholm Humic models (data not shown) (Vance et al., 2008). The amount of Cu in the aqueous phase was generally approximately twice as high in sterile than in nonsterile experiments (Figure 2). The higher release of Cu- and Zn-DOM complexes into the water phase in sterile experiments was due to the physical aggregate fractionation of the soil during autoclaving, favouring DOM release. In addition, the quality of DOM has profound and different effects on Cu and Zn binding (e.g., Baken et al., 2011). Different DOM qualities among experiments may have resulted in preferential Cu-DOM binding and release compared with Zn-DOM complexes and distinct patterns of water-soil partitioning in sterile and nonsterile experiments with crop and vineyard soils.

The Cu and Zn distribution among soil particle-size fractions in nonsterile and sterile experiments did not significantly change during ageing (p>0.1; Figure 2). Cu and Zn distribution in both crop and vineyard soils followed a silt ( $54 \pm 3\%$  of Cu and  $51 \pm 3\%$  of Zn) > clay ( $24 \pm 1\%$  of Cu and  $34 \pm 2\%$  of Zn) > sand pattern ( $23 \pm 1\%$  of Cu and  $15 \pm 1\%$  of Zn) (Figure 2). This emphasizes that Cu and Zn were mainly associated with silt, despite the larger proportion (68%; Table S2) of clay in the vineyard than in the crop soil. Less degraded SOM and a lower POM/MOM ratio in the silt fraction possibly favoured Cu and Zn sorption, especially in the vineyard soil. Similarly, POM associated with the sand fraction likely favoured Cu and Zn binding (Quenea et al., 2009), whereas sorption to sand minerals, mainly composed of quartz, remained limited (Nenadović et al., 2010).

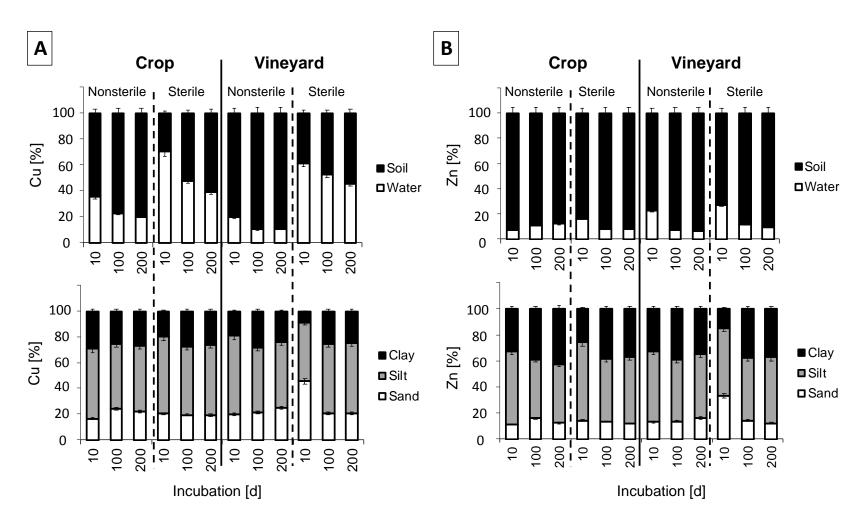


Figure 2. Distribution of (A) Cu and (B) Zn among crop and vineyard soils, including water, sand (x>50 μm), silt (2 <x<50 μm) and clay (x<2 442 fractions in sterile 95% confidence intervals. 443 μm) nonsterile and experiments. Error bars denote

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3.4. Distribution of Cu and Zn among chemical fractions

The Cu and Zn distribution among chemical fractions mainly differed with soil type and more subtly over time (Figure 3). The cluster analysis highlighted that the distribution of both Cu and Zn among chemical fractions differed between the crop and vineyard soils and different distribution patterns between Cu and Zn (Figure 4). In particular, the LExch-Cu and Zn fractions were significantly higher (p<0.001), whereas the Res-Cu and Zn fractions were lower (p < 0.001) in the vineyard than in the crop soil. In both soils, Cu and Zn were mainly associated with *LExch* ( $\overline{X} \pm SD$  across conditions: 34 ± 2% for Cu and 32 ± 2% for Zn), AFeOx (37  $\pm$  2% for Cu and 15  $\pm$  1% for Zn) and Res fractions (11  $\pm$  1% for Cu and 33  $\pm$  2% for Zn) (Figure 3). This distribution agrees with previous studies, showing that Cu and Zn in the LExch fraction were mainly covalently bound to organic and inorganic sites (Rao et al., 2008). Clay minerals and organic matter controlling the cation exchange capacity (CEC) of soil (Liu et al., 2017) may have favoured Cu and Zn release from vineyard soil (CEC=15.4 cmol<sup>+</sup> kg<sup>-1</sup>, Table S2) compared with crop soil (CEC=9.1 cmol<sup>+</sup> kg<sup>-1</sup>). In the AFeOx fraction, Cu and Zn were generally associated with amorphous Fe oxides following coprecipitation, adsorption, surface complex formation, ion exchange and/or penetration into the mineral. In contrast, Cu and Zn in the Res fraction were strongly bound or entrapped in minerals, especially clays and oxides (Clausen et al., 2001; Đurović et al., 2009; Remucal and Ginder-Vogel, 2014; Ruggiero et al., 2002). Higher proportions of both Al<sub>2</sub>O<sub>3</sub> and Fe<sub>2</sub>O<sub>3</sub> in the crop soil (Table S2) may explain the higher Res-Cu and Zn in the crop than in the vineyard soil.

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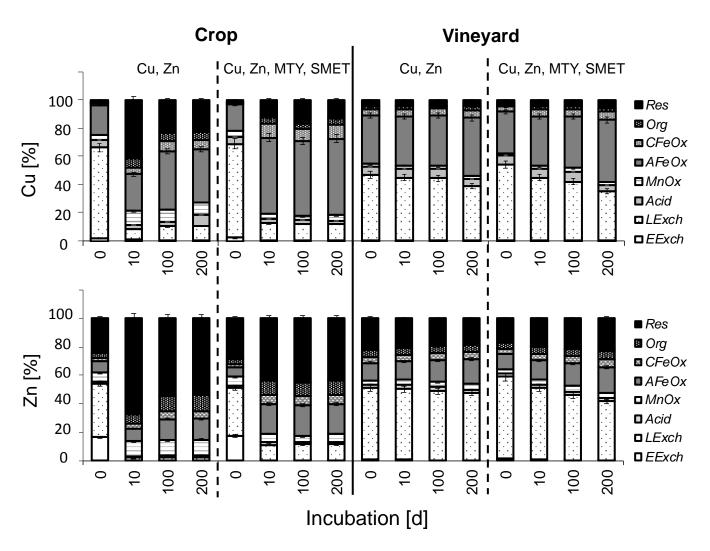
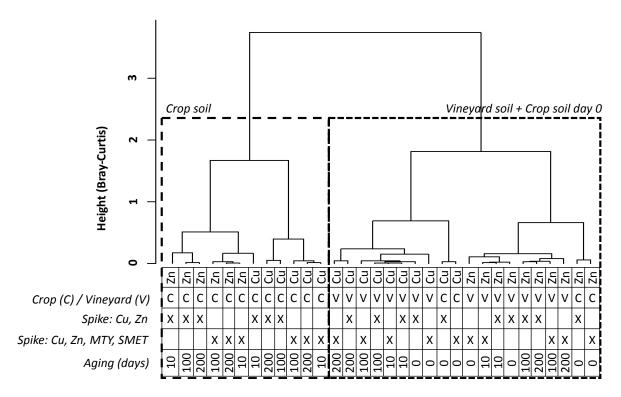


Figure 3. Distribution of Cu and Zn among chemical fractions of nonsterile crop and vineyard soils spiked with Cu and Zn only or with Cu, Zn, metalaxyl (MTY) and S-metolachlor (SMET). Error bars denote 95% confidence intervals.



**Figure 4**. Hierarchical cluster analysis (Ward's method) of Cu and Zn distribution in chemical soil fractions (*EExch*, *LExch*, *Acid*, *MnOx*, *AFeOx*, *CFeOx*, *Org* and *Res*) in nonsterile crop (C) and vineyard (V) soils spiked with Cu and Zn or with Cu, Zn, metalaxyl (MTY) and S-metolachlor (SMET) for 0, 10, 100 or 200 days of ageing.

The distribution of Cu and Zn among chemical fractions also varied over time, especially in the crop soil. For both Cu and Zn, *EExch* and *LExch* fractions and, to a lesser extent, *Org*- and *CFeOx* fractions significantly differed (*p*<0.05) between Days 0 and 10 in the crop soil. This suggests a redistribution of Cu and Zn from *LExch* and *EExch* to *Org* and *CFeOx* fractions in the early ageing stage, in agreement with previous studies (Ma et al., 2006; Jalali and Khanlari, 2008). Notably, the uncertainty associated with sequential chemical extraction of Cu and Zn from the *EExch* and *LExch* fractions may be high since MgCl used to extract exchangeable elements partly dissolves carbonates and releases Mg.

3.5. Effect of co-contamination on the distribution of Cu and Zn among chemical fractions

The distribution of Cu and Zn among chemical fractions slightly differed in experiments spiked with MTY, MET, Cu and Zn and with Cu and Zn only (Figure 3), particularly in the crop soil between Days 10 and 200 (Figure 4). Concomitant ageing of MTY, MET, Cu and Zn significantly (p<0.01) increased the *LExch* fraction and decreased the *MnOx* fraction for both Cu and Zn compared with experiments spiked with Cu and Zn only.

These results suggest that the co-occurrence of MTY, MET, Cu and Zn in soil affects their complexation and competitive sorption to soil-binding sites and thus modifies their sorption rates and distribution, mainly in the LExch and MnOx fractions. The less degraded SOM and a lower POM/MOM ratio in the silt fraction is likely to control the sorption of nonionic MTY and SMET and heavy metals in the studied soils (see section 3.1 and 3.3). First, competitive sorption on SOM and minerals can affect the sorption rates of both organic pollutants and metals. This has been observed previously for Cu and p-nitrophenol in soil (Pei et al., 2006) or Cu and chlordimeform sorption on montmorillonite (Maqueda et al., 1998). Although no experiments on competitive sorption with MTY and SMET and heavy metals have been reported, Cu and Zn are expected to form stronger bounds at pH>6 and competition between metals and non-ionic pesticides is expected to be limited (e.g., Jacobson et al. 2005). In addition, the formation of pesticide-metal complexes may alter the sorption of MTY and SMET in the presence of trace metals and vice versa. Metals can enhance the linkage between synthetic pesticides and organic matter, with Cu linking SOM and soluble pesticides thus facilitating their sorption (Zhou et al., 2004; Morillo et al., 2002). The introduction of synthetic pesticides in soil can also alter the physicochemical characteristics of the solution, thereby modifying the sorption rates. For instance, reductive dissolution of oxides by synthetic pesticides may enhance the release of Cu and Zn associated with oxides in the silt and clay fractions in experiments with MTY, MET, Cu and Zn, as previously observed with

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Mn oxides (Remucal and Ginder-Vogel, 2014). Finally, Cu or Zn ions in soil may also affect the activity of microorganisms, thus decreasing MTY and SMET degradation (Liu et al., 2007) and altering ageing patterns.

Overall, our results show that the co-occurrence of synthetic pesticides and metals can slightly affect their sorption behaviour and ageing, depending on the soil characteristics. However, the order of pollutant addition in soil can also influence sorption and competition for binding sites (Maqueda et al., 1998), although the effect of consecutive addition of pollutants has not yet been explored.

3.6. Cu and Zn stable isotope fractionation during Cu and Zn ageing in agricultural soils

Stable isotope signatures of Cu ( $\delta^{65}$ Cu) and Zn ( $\delta^{66}$ Zn) were analysed from Day 10 onwards to evaluate the isotope fractionation and associated processes during long-term ageing (>100 days) of Cu and Zn in soil fractions. Cu and Zn isotope fractionation was analysed in experiments spiked with Cu, Zn, MTY and SMET, which are more representative of multicontaminated agricultural soils. It is worth noting that Cu and Zn isotope fractionation among soil fractions mainly resulted from ageing of fresh Cu and Zn since the Cu and Zn spikes represented >90% and >70% of the initial Cu and Zn mass in soil, respectively. As expected,  $\delta^{65}$ Cu and  $\delta^{66}$ Zn values of bulk soils on Day 0 (data not shown) did not change significantly over time or among experiments. The isotopic mass balance of Cu and Zn in soil physical fractions fitted with the isotopic composition of the bulk soil, emphasizing the accuracy of Cu and Zn isotopic analysis in soil physical fractions.

The  $\delta^{65}$ Cu values differed mainly among soil fractions, particularly in the crop soil experiments ( $\Delta\delta^{65}$ Cu up to 1 ‰), and slightly over time in the clay fraction of the vineyard soil (Figure 5). In contrast,  $\delta^{66}$ Zn values did not vary significantly among soil types, granulometric fractions or over time ( $\Delta\delta^{66}$ Zn<0.4 ‰) (Figure S1). Both binding to organic

complexes and sorption to secondary minerals, such as clays and Fe-Mn oxyhydroxides, may compete to control Cu and Zn behaviour in soils (Grybos et al., 2007) and is certainly key for isotope fractionation in soil. Nevertheless, different chemical distributions of Cu and Zn (Figures 3 and 4), and redox transitions in the case of Cu, and specific values of the stability constant for each complex (Ryan et al., 2014) resulted in distinct Cu and Zn isotope fractionation in soil. Similar patterns of  $\delta^{65}$ Cu and  $\delta^{66}$ Zn values in nonsterile and sterile experiments emphasize that slight differences in the physical and chemical distribution of Cu and Zn and biotic processes did not significantly affect Cu and Zn stable isotope fractionation.



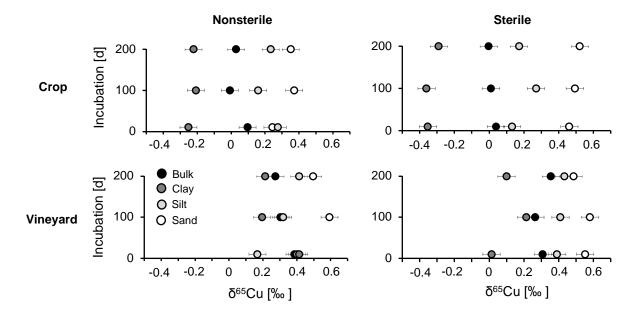


Figure 5.  $\delta^{65}$ Cu values in bulk soil and clay, silt and sand fractions of nonsterile and sterile crop and vineyard soils spiked with Cu, Zn, MTY and SMET. Error bars denote analytical uncertainty ( $\pm$  0.05).

The difference in  $\delta^{65}$ Cu in the soil fractions of the crop and vineyard soils suggests that different processes affected Cu isotope fractionation. In both soils, the clay fraction was depleted in  $^{65}$ Cu, whereas the silt and sand fractions were enriched in  $^{65}$ Cu compared with the bulk soil. The formation of SOM-Cu complexes and Fe-oxide and organo-clay complexes

generally drives Cu sorption in soil, although Cu precipitation as carbonate or hydroxide may prevail in calcareous vineyard soil (El Azzi et al., 2013; Komárek et al., 2010a). Our results are in agreement with previous results insofar as Cu sorption to Fe oxides (Balistrieri et al., 2008b) and SOM (Bigalke et al., 2010) is expected to enrich Cu in <sup>65</sup>Cu, whereas Cu sorption to clay minerals may deplete Cu in <sup>65</sup>Cu (Li et al., 2015b). In particular, the formation of Cu-SOM complexes at pH above 7 may affect  $\delta^{65}$ Cu among soil fractions (Quenea et al., 2009). However, the SOM quality varies in grain-size fractions, with more altered (oxidized) SOM and stabilized organomineral complexes generally prevailing in the clay fraction (Poirier et al., 2005). We thus hypothesize that the larger Res-Cu fraction in the crop than in the vineyard soil (Figure 3) is characterized by strongly bound or entrapped Cu in clays, explaining the higher Cu enrichment in <sup>65</sup>Cu in the clay fraction of crop soil. In addition, Cu precipitation in carbonates is also expected to deplete the clay fraction in <sup>65</sup>Cu (Maréchal and Sheppard, 2002) in the vineyard soil since CaCO<sub>3</sub> contributed to 27.1% of the soil content (Table S2). However, XRD data showed that carbonate phases prevailed in the silt fraction of the vineyard soil that was enriched in <sup>65</sup>Cu, whereas only traces of calcite were found in clays (Babcsányi et al., 2016), and <1% of carbonates were found in the crop soil (Table S2). Hence, it is unlikely that Cu precipitation in carbonate enriched the silt fraction and depleted the clay fraction in <sup>65</sup>Cu. The  $\delta^{65}$ Cu values of the soil fractions did not change much over time, especially after 100 days of ageing. This suggests that the Cu distribution among soil fractions was stable

The  $\delta^{65}$ Cu values of the soil fractions did not change much over time, especially after 100 days of ageing. This suggests that the Cu distribution among soil fractions was stable from Day 10 onwards (Figures 2 and 3), reflecting limited Cu redistribution among chemical fractions during longer-term ageing in both soil types. However, a redistribution of Cu associated with Cu isotope fractionation in the early ageing stage, i.e., from Day 0 to 10, cannot be excluded, and the signature may be acquired in the early stages of ageing. Moreover, the silt fraction was gradually enriched in  $^{65}$ Cu from Day 10 to 200 ( $\Delta\delta^{65}$ Cu = 0.25

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 $\pm$  0.10 %) in nonsterile experiments with vineyard soil, whereas the clay fraction was depleted in  $^{65}$ Cu ( $\Delta\delta^{65}$ Cu =  $-0.20 \pm 0.10$  %) (Figure 5). Interestingly, the enrichment in  $^{65}$ Cu of the silt fraction mirrored the increasing contribution of the AFeOx fraction of nonsterile vineyard soil over time (Figure 3). Since the AFeOx fraction involved Cu association with amorphous Fe oxides, it may be inferred that Cu sorption to Fe oxides gradually increased during ageing in the silt fraction of vineyard soil to reach  $\delta^{65}$ Cu typically ranging between 0.24 0.98 ‰ (Balistrieri et al., 2008a; Pokrovsky and et al., 2008).

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#### 4. Conclusion

Current research issues in the field of pollutant dissipation in agricultural soils include (i) how soil characteristics affect the ageing of synthetic pesticides and heavy metals and (ii) how the co-occurrence of synthetic pesticides and heavy metals affects their distribution among soil fractions during ageing. It is our contention that studies such as ours may help to address these issues and enable more systematic examinations of concomitant ageing of synthetic pesticides and heavy metals in soil fractions rather than in bulk soil. Specifically, our study emphasised that the distribution of MTY and SMET among particle-size fractions varied during ageing, thereby controlling the sorption of MTY and SMT and the potential release of MTY and SMET into water. In contrast, the physical and chemical distributions of Cu and Zn differed with soil type and more subtly over time.

Most importantly, we showed that the co-occurrence of synthetic pesticides and heavy metals may slightly alter metal distribution in soil fractions, depending on soil characteristics. Thus, the simultaneous evaluation of synthetic pesticides and heavy metals in soil physical and chemical fractions can reinforce the evaluation of ageing in agricultural fields and help identify risks associated with pollutant distribution among soil fractions in multiple-contaminated areas. Here, we could identify field plots where preferential release into water and export of aged pollutants associated with specific soil fractions occurred, thereby allowing us to focus remediation efforts in these areas. Our results also provide new insights into Cu isotope fractionation during ageing in soils, emphasizing that  $\delta^{65}$ Cu partly mirrored the Cu distribution among chemical fractions of the soils, potentially involving Cu binding in SOM and Fe oxides in specific soil fractions. Overall, our study illustrates the usefulness of integrating approaches to jointly evaluate the ageing of micropollutant molecules and heavy metals in soil for more robust assessment and remediation of multiple polluted sites.

512	Credit authorship contribution statement					
514	Fatima Meite: Conceptualization, Investigation, Methodology, Data curation, Writin					
515	- review & editing. Mathieu Granet: Investigation, Methodology, Data curation, Writing					
516	review & editing. Gwenaël Imfeld: Conceptualization, Methodology, Funding acquisition					
517	Project administration, Supervision, Formal analysis, Writing - original draft.					
518						
519	Declaration of competing interest					
520	The authors declare that they have no known competing financial interests or personal					
521	relationships that could have appeared to influence the work reported in this paper.					
522						
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528						
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