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Gookbin Cho, Sawsen Azzouzi, Gaël Zucchi, Bérengère Lebental. Electrical and Electrochemical Sensors Based on Carbon Nanotubes for the Monitoring of Chemicals in Water-A Review. *Sensors*, 2022, 22 (1), pp.218. 10.3390/s22010218 . hal-03855860

HAL Id: hal-03855860

<https://hal.science/hal-03855860>

Submitted on 16 Nov 2022

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Review

Electrical and Electrochemical Sensors Based on Carbon Nanotubes for the Monitoring of Chemicals in Water—A Review

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Abstract: Carbon nanotubes (CNTs) combine high electrical conductivity with high surface area and chemical stability, which makes them very promising for chemical sensing. While water quality monitoring has particularly strong societal and environmental impacts, a lot of critical sensing needs remain unmet by commercial technologies. In the present review, we show across 20 water monitoring analytes and 90 references that carbon nanotube-based electrochemical sensors, chemistors and field-effect transistors (chemFET) can meet these needs. A set of 126 additional references provide context and supporting information. After introducing water quality monitoring challenges, the general operation and fabrication principles of CNT water quality sensors are summarized. They are sorted by target analytes (pH, micronutrients and metal ions, nitrogen, hardness, dissolved oxygen, disinfectants, sulfur and miscellaneous) and compared in terms of performances (limit of detection, sensitivity and detection range) and functionalization strategies. For each analyte, the references with best performances are discussed. Overall, the most frequently investigated analytes are H⁺ (pH) and lead (with 18% of references each), then cadmium (14%) and nitrite (11%). Micronutrients and toxic metals cover 40% of all references. Electrochemical sensors (73%) have been more investigated than chemistors (14%) or FETs (12%). Limits of detection in the ppt range have been reached, for instance Cu(II) detection with a liquid-gated chemFET using SWCNT functionalized with peptide-enhanced polyaniline or Pb(II) detection with stripping voltammetry using MWCNT functionalized with ionic liquid-dithizone based bucky-gel. The large majority of reports address functionalized CNTs (82%) instead of pristine or carboxyl-functionalized CNTs. For analytes where comparison is possible, FET-based and electrochemical transduction yield better performances than chemistors (Cu(II), Hg(II), Ca(II), H₂O₂); non-functionalized CNTs may yield better performances than functionalized ones (Zn(II), pH and chlorine).

Keywords: carbon nanotubes; nanomaterials; water quality; chemical sensor; chemistor; field effect transistor; electrochemical sensors



Citation: Cho, G.; Azzouzi, S.; Zucchi, G.; Lebental, B. Electrical and Electrochemical Sensors Based on Carbon Nanotubes for the Monitoring of Chemicals in Water—A Review. *Sensors* **2022**, *22*, 218. <https://doi.org/10.3390/s22010218>

Academic Editor: Abdelhafed Taleb

Received: 17 November 2021

Accepted: 24 December 2021

Published: 29 December 2021

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1. Introduction

While fresh water represents 3% of the total water on Earth, only 0.01% is available for human consumption [1]. Rapid population growth, unsustainable water use in agriculture and industry and climate changes are bringing about hydric stress worldwide. While drink water availability decreases, its quality also degrades: World Health Organization (WHO) reports that, in developing countries, 80% of human diseases, are water borne [2]. Drinking water quality in numerous countries does not meet WHO standards [3,4]. The presence of water contaminants critically impacts human beings and ecosystem. It is thus of vital importance to be able to analyze fresh water, whether it is groundwater, irrigation water or tap water.

Concerning water networks, water quality monitoring and control mainly takes place at the water supply intake or at the water treatment plant. However, this seems inadequate given the important variations in water quality observed throughout water distribution systems [5]. Online sensing—also called on-site sensing—is currently seen as the best solution to provide continuous, early warning systems for chemical contamination throughout the water network (from drink water production to waste water treatment). It designates the capability to monitor water quality accurately and in real-time, and it is expected to yield public health improvements via improved water safety [6]. To cover recent advances in the field, Kruse recently reviewed chemical sensors for water quality evaluation [7]. After detailing the parameters and contaminants that are currently relevant to water monitoring, the authors present exhaustively transduction methods for water quality sensors. The review shows that, despite worldwide efforts, there are still plenty of challenges to be met by online water quality sensors: reduction of costs and calibration frequency, increase in sensitivity and selectivity, reduction of power consumption and size and enhancement of lifetime [8].

These challenges have motivated a wide range of studies toward water quality sensors based on nanomaterials, for instance as described in references [9–11], as nanomaterials-based sensors are well-known to meet those specific challenges across all fields of research on sensors [12,13]. Among sensors fabricated with nanomaterials, those comprising carbon nanotubes (CNT) have been continuously proposed for chemical sensing since the early days of CNT research [14] taking advantage of their excellent chemical stability and their large surface area. Most recently, Schroeder et al. [15] reviewed CNT-based chemical sensors, with applications covering gas sensors, biosensors, food sensors or aqueous sensors.

However, there has been no review focusing specifically on water quality monitoring based on carbon nanotube sensors. The present paper endeavors to fill this gap. More specifically, we report on CNT-based electrical and electrochemical sensors, because they are particularly well-suited for online water monitoring applications [16–18]. The electrical transduction options for CNT-based chemical sensors are electrochemical, resistive, field-effect-based and electromechanical. The latter has only been reported for gas sensing, so is not discussed in the present paper [19]. The various CNT-based optical sensors are also beyond the scope of this paper. While they have been often proposed for water quality monitoring (see reference [20] for instance), their applicative use requires integration into microfluidic lab-on-chips, whose specifications are not suitable today for online use and much more appropriate for sampling-based (portable or off-site) water monitoring systems [21].

After a brief overview of the design, fabrication and operating principles of CNT-based chemical sensors, we present exhaustively the various electrical and electrochemical sensors reported in the literature from 2000 to mid-2021. We sort them by types of analytes and the various reports are analyzed in terms of sensing performances. Selected papers are highlighted in view of understanding the sensing mechanisms. Finally, the sensing performances of CNT-based sensors are compared to those of other upcoming nanomaterials.

2. Operating Principles of CNT-Based Chemical Sensors

2.1. Chemical Nanosensors

In general, a chemical sensor transforms chemical information (typically the presence or concentration of a target analyte in water) into an exploitable electrical signal. It consists of a chemical recognition layer (receptor) and a physicochemical transducer. The receptor interacts with target analytes, which affects the transducer then turns it into an exploitable signal [22]. When either the transducer or the recognition layer contains a nanomaterial or is nanostructured, the device is said to be a nanosensor.

The performance of a chemical sensor is notably characterized by its response curve, namely the relationship linking the sensor signal to the analyte concentration. The response is more often modeled as linear, though exponential and logarithmic responses are also

reported. The sensitivity of a sensor is defined as the slope of the response curve in its linear range. A chemical sensor is said to be selective if it can discriminate between a selected analyte and other species (said to be “interfering”) within a sample. Increasing sensitivity and selectivity is the main goal driving the use of nanomaterials in chemical sensors. Because of their high surface over volume ratio, nanomaterials are expected to have higher sensitivity. The capability to engineer their composition and crystalline structure at the atomic scale opens up the possibility to design more selective recognition layers.

2.2. Carbon Nanotubes (CNTs) Sensors

2.2.1. Carbon Nanotubes Structure

Carbon nanotubes composition, crystalline structure, fabrication and properties have been extensively reviewed over the years [23–26]. Briefly, carbon nanotubes are more easily described starting from graphene (Figure 1). Graphene consists of a single sheet of carbon atoms arranged in hexagonal cells with an in-plane structure. This form of coordination for carbon atoms is called the sp^2 hybridization of carbon: the 2s orbital and two of the 2p orbitals (the p_x and p_y) hybridize to form 3 covalent (σ) bonds per carbon atom spread at 120° from each other (trigonal structure), while one electron in the p_z orbital remains free to contribute to conduction. Carbon nanotubes (CNTs) can then be described as a rolled sheet of graphene with axial symmetry. They are sorted into two main categories, Single-Walled Carbon Nanotubes (SWCNTs) and Multi-Walled Carbon Nanotubes (MWCNTs) depending on the number of graphene layers rolled into a coaxial array (Figure 1). SWCNTs have typical diameters in the range 0.4 to 20 nm and typical lengths from 100 nm to 10 μm depending on the method used to synthesize them. In MWCNTs, each tube is separated from the next by 0.34 to 0.36 nm. MWCNTs may have diameter between 1 nm (double-walled carbon nanotubes (DWCNTs)) to 300 nm (about 100 coaxial tubes) and typical length from 1 μm to 150 μm . In both cases, the length-to-diameter ratio is very high, so they are regarded as 1D nanostructures. SWCNTs can be either semi-conducting or metallic depending on their chirality (the orientation of the lattice with respect to the tube axis) while MWCNT is metallic (except in rare DWCNTs cases).

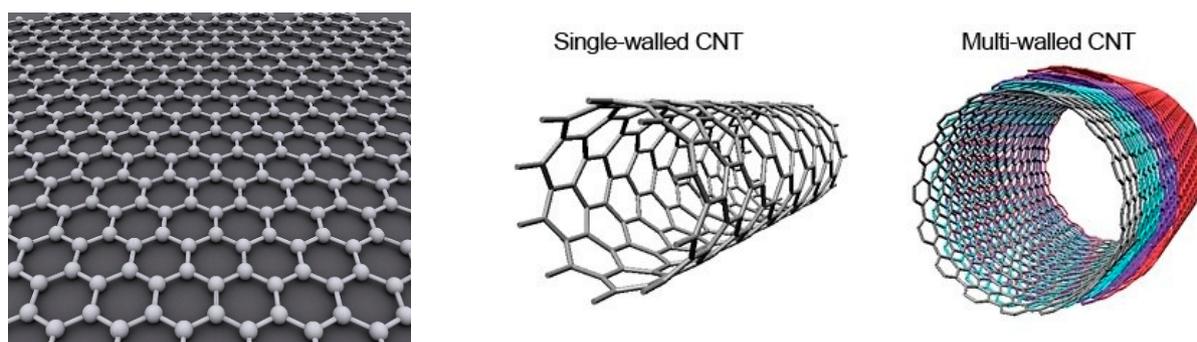


Figure 1. Basic structure of a graphene (left) [27], single-walled carbon nanotube (middle) and multi-walled carbon nanotube (right) [28]. CC BY-SA 3.0.

2.2.2. Functionalization of CNTs

CNTs are very attractive as active materials in chemical sensors as they have high adsorption capability to a wide range of species. However, various results show the limited selectivity of pristine CNT-based chemical sensors (for both gas-phase and liquid-phase sensing): they are often sensitive to different analytes with the same range of magnitude of sensitivity and response time [29,30].

Functionalizing CNT consists in hybridizing them with other molecules either by covalent [31,32] or non-covalent bonds [33,34]. Functionalization is advantageous for selective sensing because the functionalizing molecules can be selected for their affinity to

the target analyte. It has become the most popular approach to enhance the selectivity of CNT-based chemical sensors, though some studies report on modulating electrode material instead of functionalizing CNT as a mean to achieve selectivity [35].

One of the main challenges in using functionalization for selective sensing lies in ensuring that the changes occurring at the functionalizing molecules in the presence of the target analyte can be detected through the CNTs in the selected electronic device configuration. At the same time, the functionalization itself should not degrade dramatically the properties of the electronic device itself.

For instance, covalent functionalization is usually expected to allow for stronger charge transfer between CNT and functionalizing molecules, thus providing stronger sensitivity to the target analytes. However, covalent bonds degrade the crystalline structure of the CNTs, thus degrading their conduction properties and subsequently the transduction quality. As a consequence, the density of covalent functionalization that can be achieved in practice remains limited, which in turn may limit the gain in sensitivity and selectivity [36].

By contrast, using non-covalent functionalization, full coverage of the CNT surface may be achieved without degrading the intrinsic electronic properties of the CNTs; however, selecting functionalizing molecules that strongly impact the electronic properties of CNTs is challenging [37]. Usually, molecules that can functionalize CNTs by strong π -stacking are selected among aromatic molecules such as derivatives of benzene, fluorene, carbazole, or porphyrin, or conjugated polymers [38,39].

CNTs often carry some carboxyl groups (-COOH functions) on their sidewalls as a result of the synthesis process or of the post-synthesis purification (see Section 2.3.3). The density of these groups may for instance be evaluated by Raman spectroscopy [40,41], but is not systematically studied in the literature on CNT sensors. Hence CNTs reported on as non-functionalized CNTs may carry COOH groups. The COOH density may also be increased on purpose to enhance sensitivity to certain analytes, for instance by strong oxidative acidic treatments [42]. In this review, CNTs oxidized on purpose are labeled CNT-COOH. Articles reporting on those are classified jointly with the articles on pristine CNTs.

2.2.3. CNT-Based Electronic Devices

Using CNT in electrical sensors requires their integration into electronic devices. This topic has been extensively discussed in the literature (see reference [43] for instance). Briefly, one differentiates between devices based on as-grown CNTs or on prefabricated CNTs. Devices can be either based on a single CNT [44] or a CNT network [45]. In turn, this network may be either random or organized (for instance aligned).

In devices based on as-grown CNTs, CNTs are usually synthesized via chemical vapor deposition (CVD) directly onto pre-patterned electrodes within a temperature range from 550 °C to 1000 °C [46,47]. CVD CNT growth leads to robust electrode/CNT contacts and high CNT crystalline quality while avoiding bundling. However, the high-temperature CVD growth conditions usually prevent the use of flexible substrates. The need for metallic growth catalysts is often incompatible with the architecture of electronic devices (as they require well-defined insulating surfaces). For those reasons, as-grown CNT films are often transferred as a whole from the synthesis substrate onto more appropriate substrates via lift-off [46]. In addition, in-place synthesis does not allow for perfect control of CNT alignment, nor of their diameter, chirality or crystallinity, while these parameters have key impacts on device features. There are several purification and sorting techniques available to tune these parameters for CNTs on solid substrates. The most frequently reported post-growth processes are removal of the metallic CNTs by electrical breakdown [48] (application of a high current to a CNT network while the semiconducting CNTs are polarized in their OFF-state, which burns out metallic CNTs only) or degradation of the CNT crystalline quality by plasma etching [49], irradiation [50], or thermal oxidation [51].

By contrast, in devices relying on pre-fabricated CNTs, CNTs available in powder form are dispersed in a solvent and deposited onto the appropriate substrate via wet process. It is the most frequently reported approach to fabricate CNT-based sensors. It

is advantageous because it features little constraints regarding substrates and because it allows the use of a large panel of solution-based CNT pre-treatment protocols, such as purification, acidification, functionalization, sorting by chirality or by diameter [52]. A large variety of techniques is available to deposit CNTs from a dispersion onto a substrate: drop-casting [53], spin coating [54], dip-coating [55], inkjet printing [56], spray-coating [57], aerosol jet printing [58] or vacuum filtering [59]. After substrate deposition of a CNT dispersion, particularly following drop casting, dielectrophoresis may then be used to improve on the deposition quality, notably to control accurately CNT positioning or to achieve CNT alignment [60]. After vacuum filtering, the filter may be removed, forming a freestanding film often called buckypaper [61]. Let us note that, despite the advantages of using prefabricated CNTs compared to in-place growth CNTs, it also has a few drawbacks, such as: CNT placement on the substrate may not be as accurate; low network density is more difficult to achieve; CNT-substrate interaction may be less strong; CNT crystallinity may be degraded during liquid phase processing steps such as high-power sonication.

2.3. CNT-Based Electrochemical Sensors

2.3.1. Electrochemical Cells

An electrochemical sensor is a device that detects an electron exchange between sensor and analyte. It is usually composed of two basic components, a chemical recognition layer and a physicochemical transducer, the latter comprising several metal electrodes, the working electrode, the reference electrode and in most cases a counter electrode. Immersed into an electrolyte solution, they make up the electrochemical cell (or voltaic or galvanic cell) [62].

A two-electrode cell consists of only working and reference electrodes. It is used for low current operation (small-sized working electrodes, very low analyte concentrations) because at higher current, the potential of the working electrode becomes unstable. In most applications, a three-electrode cell is used; the reference electrode is maintained at a stable potential, while the current passes through working and counter electrodes. Two types of process may occur in electrochemical cells. In a Faradaic process, charge particles transfer from electrode to electrode through the electrolyte. In non-Faradaic, charge is progressively stored [63].

2.3.2. Electrochemical Transduction

There are various types of electrochemical transducers depending on how the electrochemical cell is operated. The most popular ones are briefly described in the following paragraphs.

In potentiometric sensors, the measured signal is the potential difference between the working electrode and the reference electrode in the absence of current. The working electrode potential depends on the concentration of the target analyte. A reference electrode is needed to provide a defined reference potential. The response of a potentiometric sensor is interpreted using the Nernst equation, which states that the activity of the species of interest is in a logarithmic relationship with the potential difference [64]. This approach works well when the activity of a given species can be approximated to the molar concentration, namely at low concentration.

In voltammetric sensors, the current response is measured as a function of the applied potential. It is directly correlated to the rate of electron transfer occurring via electrochemical reactions [65]. This approach differentiates well species with different redox potential (separated by more than ± 0.04 – 0.05 V). In turn, there are significant interfering effects if two or more species in the sample solution have similar redox potentials. There are different types of voltammetry depending on the way the voltage is applied, notably linear sweep or pulse-wise increase. The latter (usually called differential pulse voltammetry), is reported to be well suited for solid electrodes based on organic compound and more sensitive than the former (usually called cyclic voltammetry).

In electrochemical impedance spectroscopy (EIS), an alternating voltage is applied. The phase shift and amplitude of the current are measured over a range of frequencies. It provides information on the rate of the electrochemical reactions and on the ionic transport in the electrolyte [66].

Stripping voltammetry consists of two steps. First, target chemical species are electrolytically deposited on the surface of one of the electrodes using a constant potential, for instance by reduction of metal ions on the cathode. Second, a voltage scan is applied to the electrode, which progressively strips the target analytes from the electrode depending on their redox potential. At a given voltage, the resulting faradic current is proportional to the concentration of the target chemical ionic species [67]. If the different species are stripped at different voltages, selectivity is possible. The electrode deposition step has a pre-concentration effect on the target analyte, which yields this technique its considerable sensitivity (sub-nanomolar range for metal ions). Figure 2 shows example of measured responses by these three transduction methods.

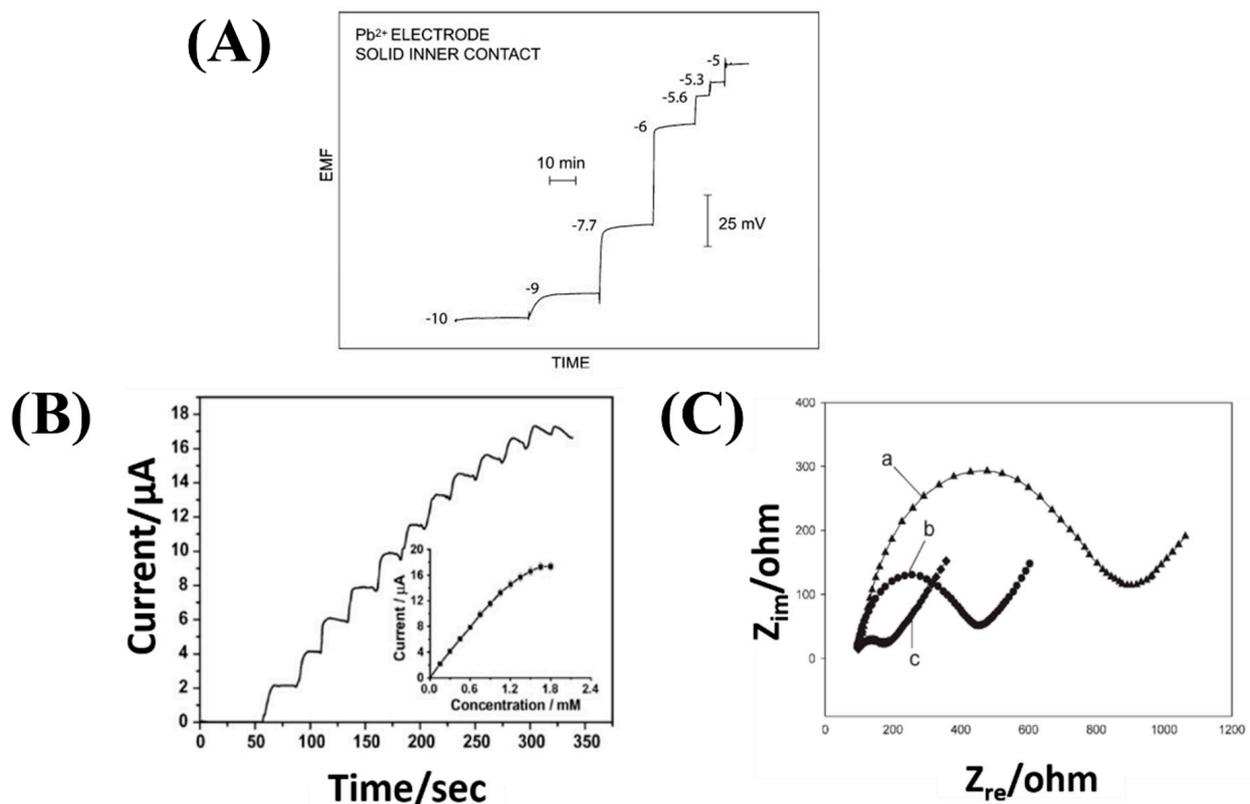


Figure 2. Measured response by three different transduction methods of electrochemical sensors: (A) Potentiometry (Potential at zero current—EMF electromotive force—vs. time under increasing volume of analyte—the numbers shown are logarithmic molar sample concentrations. Reproduced from [64] (B) Voltammetry (Current versus time under increasing volume of analyte) Reproduced from [68] (C) Electrochemical impedance spectroscopy (Nyquist plot: real impedance vs imaginary impedance for different of electrodes; a—Bare glassy carbon electrode (GCE) 1; b—CNTs/poly(1,2-diaminobenzene) prepared by cyclic voltammetry modified GCE; c—CNTs/poly(1,2-diaminobenzene) composite prepared by multipulse potentiostatic method modified GCE). Reproduced from [69].

2.3.3. Use of CNTs in Electrochemical Sensors

Because the performances of electrochemical sensors are driven by the specificities of the electrodes and of the electrolyte/electrode interfaces, improvements in sensor performances can be achieved by tuning either the electrode bulk material or the electrode surfaces, the latter using either dedicated coatings or by surface engineering (for instance,

roughness increase). CNTs are used both as coatings and as electrode material to leverage their high specific surface area. It allows for a large dynamic range and for a high loading in electrocatalysts (defined as the catalysts that participate in electrochemical reactions by increasing the rate of chemical reactions without being consumed in the process). Moreover, CNTs display resistance to fouling [70].

The details of the composition and fabrication process of the various CNT-based electrochemical sensors reported in this paper are provided later. As a summary, out of 66 reports, 31 (47%) reports address CNTs coated on glassy carbon electrodes, 10 (15%) are about electrodes directly made out of CNTs or CNT paste, the rest electrodes being made of miscellaneous metallic materials (gold, steel . . .). The most used method for coating electrodes with CNTs is drop casting (28 references, 42%): CNTs are first purified, then chemically activated (either oxidized or functionalized) and dispersed in a solvent with sonication. The dispersion is then dropped on the electrode surfaces and the solvent is evaporated rapidly [68,71–76].

The prevalence of drop-casting methods is due to their simple implementation. They are often used as a stepping stone on the path toward more reproducible, but often less straightforward, fabrication processes. One of the main shortcomings of techniques based on CNT dispersion (drop and spray casting, dip coating, dielectrophoresis, printing . . .) is that most solvents have low exfoliation efficiency for CNTs and the resulting solutions have low stability due to the rather weak interactions between these solvents and CNTs [77]. As a consequence, CNT-paste-based electrodes are a popular alternative to CNT-coated electrodes (13 references, 20%). The reported binders are often mineral oils, often mixed with graphite powder and/or ionic liquids ([78,79]).

Regarding electrochemical sensing mechanisms, the carbon atoms at the CNT ends have been shown to behave like the edge planes of highly orientated pyrolytic graphite (HOPG) and to feature rapid electron transfer kinetics: they contribute to the Faradaic processes and provide quick response time. By contrast, the carbon atoms of the sidewalls resemble the basal plane of HOPG and show slower electron transfer kinetics than end atoms [80] (though still higher than HOPG due to curvature [81]). In other words, they are much less involved in oxidoreduction reactions with the electrolyte. However, they contribute to non-Faradaic processes driven by adsorption and desorption mechanisms.

The processes enabling removal of the impurities from CNTs that remain from the synthesis process (carbon nanoparticles, nanocrystal metal catalysts, amorphous carbon . . .) play a strong role in the electrochemical properties of the CNTs. Raw CNTs are usually purified before use by thermal treatment at around 400 °C or by chemical oxidation via acidic treatment. It leads to shortened and partially oxidized CNT. In particular, the resulting CNTs feature functional oxygenated groups at the open ends and an increased defect density along the sidewalls [82]. In addition to CNT curvature, those defects also explain that CNT sidewalls contribute to the Faradaic process in electrochemical sensors. Luo et al. for instance detailed the oxidation–reduction reactions for carboxylic CNT sidewall defects in [83].

2.4. CNT-Based Chemistors

2.4.1. Chemistors

Chemi-resistors, or chemistors, are sensors operating by measuring the variation in the electrical resistance or the resistivity of a sensing (also called active) material as a consequence of its interaction with the target analyte. The target analyte has to be in direct contact or close proximity to the active material. The possible interactions are highly diversified: bulk or catalytic reactions, reversible or irreversible, chemi- or physisorption, surface or volume reactions or reactions at grain boundaries [84].

In most chemistors, resistance changes are measured in a two-terminal configuration (Figure 3a,b). A small constant current is applied between two electrodes separated by a short distance (μm to mm) and the resulting voltage is measured. Alternately, four-terminal configurations may also be used to reduce the influence of contact resistance on

the sensitivity, especially in the case of high resistance devices ($M\Omega$ range and higher). Four parallel electrodes are often used in those cases; the current is applied on the external electrodes and the voltage drop is measured across the two internal electrodes. In the case of arbitrary electrode disposition (for instance, anisotropic surface), the Van der Pauw method can be used to measure the bulk resistivity (ρ) and the Hall coefficient of the surface by using four different contact point [85].

2.4.2. Use of CNT in Chemistors

The use of CNT as active layer and/or electrode material in chemistor is prominent across various sensing applications (gas sensing, biological sensing) [86], as it is the most straightforward device structure available to assess sensitivity of CNTs to chemicals (in terms of design, fabrication, electronics, signal processing ...). The high surface area results in high adsorption rates for analytes leading to a rapid response time. Typically, only a fraction of μg or less of CNT material is needed, so the raw material cost is not a limiting factor [87]. Moreover, a small (1 cm^2) chip-based device can hold hundreds of sensor elements. Such miniaturization leads to a reduction in size and weight of the assembled systems.

Most reports on chemistors in this review (10 out of 13) use SWCNTs rather than MWCNTs. CNT chemistors are mostly often fabricated using electrode materials made of noble metals (platinum and gold), though occasionally (here 4 reports out of 12) the CNTs make up both electrode material and active layer. The electrode metal is usually thermally evaporated on the substrate and patterned with photolithography. After purification (eliminating synthesis residues), sorting (for instance by diameter) and dispersion in a solvent, the CNTs are deposited across the gap, bridging the electrodes or electrode fingers (Figure 3a,b), then the solvent is evaporated. Various methods can be used for this deposition step, either wet-processing techniques (such as drop-casting, inkjet printing, spraying ...—11 references) or dry-processing techniques (such as direct (in-place) chemical vapor deposition (CVD) growth or CVD-growth followed by solid-state transfer or by nanoimprint as nano-scale patterning process—2 references). The CNT networks are in most cases random (except in 3 references where there are aligned through dielectrophoresis [88] or threading of CVD-aligned CNTs [89]).

The baseline resistance level of a device and its sensitivity depends on the geometry of the electrodes, on the type and quality of CNTs as well as on their surface density. The latter actually depends on both the CNT concentration in the dispersion and the selected deposition process. The geometry of the electrodes is characterized primarily by their spacing—often called the gap—and the length of the gap. The gap ranges between $1\ \mu\text{m}$ and $100\ \mu\text{m}$, with the gap length of between $10\ \mu\text{m}$ and several mm. To optimize space occupation, the electrodes are often interdigitated (Figure 3c): instead of a straight gap, the gap is formed by a series of parallel fingers. The effective gap length is thus roughly equal to twice the finger length multiplied by the number of fingers. Finger widths are typically in the 1 to $10\ \mu\text{m}$ range, lengths in the 10 to $100\ \mu\text{m}$ range [90,91].

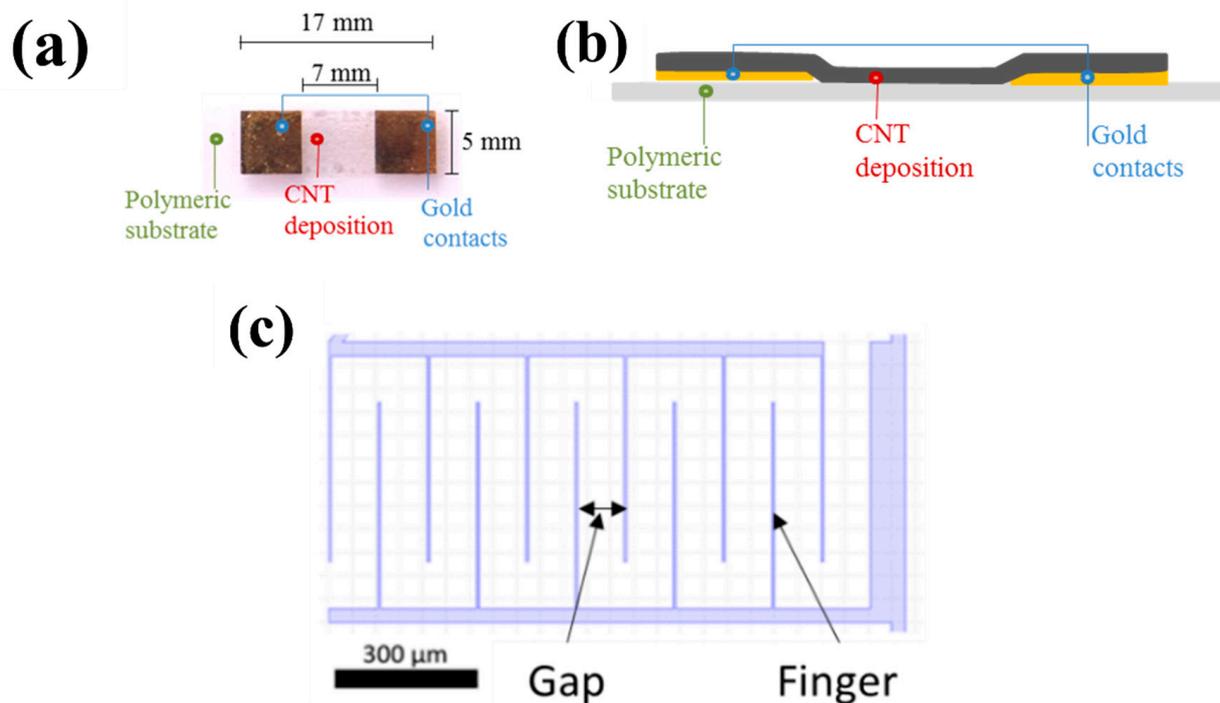


Figure 3. (a) Top view and (b) cross section of an example of two terminal resistive CNT sensors on ETFE (Ethylene Tetra Fluoro Ethylene). Reproduced from [92]. (c) Schematic of interdigitated electrodes.

2.5. CNT-Based ChemFET

2.5.1. ChemFET

A field-effect transistor (FET) is an electronic device consisting of a semiconducting layer, called channel, located between a source and a drain electrode; the density of electronic carriers flowing in the channel between source and drain electrodes is modulated by the potential of a third electrode, called gate electrode, close to the channel and insulated from it by a dielectric material. A chemical FET—chemFET—is a FET whose conduction characteristics are modulated by the presence and concentration of analytes around the device. The chemically sensitive layer is usually the semiconducting channel, though the electrodes and the dielectric layer have been reported to contribute to sensitivity. The device design allows for the semiconducting layer to be exposed to the target liquid. In an electrolyte-gated chemFET, the target liquid itself is used as both the gate and dielectric layer.

In broad terms, chemFETs are similar to chemistors in the sense that they can be described as chemistors whose baseline resistance is controlled by the gate voltage. Similarly to chemistors, chemical detection is enabled by short-range interactions between the target analyte and the active layer. However, while in chemistors only the resistance (and sometimes the resistivity and contact resistance) of the active layer may be exploited to derive the analyte concentration, there are much more varying parameters in chemFET architectures, which offers a finer understanding of the sensing mechanisms.

In more details, the electrical properties of chemFETs are derived from measuring its drain-source current I_{ds} as a function of gate V_g and drain voltage V_d . From these curves are extracted synthetic parameters such as: (i) in the linear regime of I_{ds} as a function of V_d , the (gate-voltage controlled chemFET resistance (slope of $I_{ds}(V_d)$); (ii) the ratio between ON and OFF current levels (values of I_{ds} when the semiconducting channel is respectively in its most conducting—ON—and most insulating—OFF—state); (iii) the transconductance (the maximum value of the first derivative of $I_{ds}(V_g)$, which is related to the mobility of the semiconducting channel); (iv) the threshold voltage (gate voltage value for which

the semiconducting channel transitions from insulating to conducting); (v) the hysteresis observed between threshold voltage values or transconductance values during upward and downward sweep of the gate voltage [93–95]. Regarding the latter, this hysteresis is attributed in large part to the adsorption of water molecules on the device surface creating charging effect [96]. Hence, it is expected to be a particularly relevant indicator in CNTFET-based water quality sensors but there has very been little study on it so far [97]. Because of this diversity of output parameters, chemFETs are usually considered to be more sensitive and more selective than chemistors. In turn, they usually require significantly more complex fabrication and characterization procedures as well as operating electronics and signal processing.

2.5.2. Use of CNT in ChemFET

CNT-FET is a chemFET with a CNT layer as channel. Because a semiconducting channel is required, only single-walled carbon nanotubes (SWCNTs) can be used [98] (MWCNTs being metallic). In general, the CNT-FET channel may be formed either by a single semi-conducting SWCNT or by a percolating network of SWCNTs with a semi-conducting behavior [99]. While devices based on a single SWCNT have remarkable electrical performances [43,100], devices based on a random-network of SWCNTs are more popular for sensing applications due to their higher effective sensing area, their simpler fabrication procedure as well as their better up-scalability (for mass production), even though their electrical performances are not as good as these of single SWCNT devices. In the field of water quality monitoring, only devices based on networks of CNT have been reported so far. Those networks are in most cases random except in 2 references where CNTs are aligned through dielectrophoresis [60,101].

As SWCNTs may be either semi-conducting or metallic depending on their chiral structure, percolating networks of SWCNTs are not normally semi-conducting because they contain a significant ratio of metallic SWCNTs. To achieve semiconducting SWCNTs networks, a variety of processes is available. For instance, semiconducting SWCNT may be sorted before deposition; electrical breakdown of metallic SWCNTs after their deposition or their in-place growth can also be used; almost systematically, low density networks are used to minimize the chance of forming a metallic path between electrodes [102–104].

There are four main types of device architecture for CNT-FET chemical sensors: top gate, bottom gate, liquid gate and hybrid structures; Among 11 reported CNT-FET-based chemical sensors, two are top-gated [105,106], three bottom-gated [107–109] and five liquid gated [60,110–113]. Finally, one is a hybrid dual-gate structure [54] (Figure 4). The original architecture is the bottom gate one, where the gate is embedded below the semiconducting layer with a separating dielectric layer [107]. In the context of water quality monitoring, it has the significant drawback of requiring a high gate voltage (usually several tens of Volts) for good electrical performances, which leads to hydrolysis of water (beyond 1 V). In top gate structures, the gate layer is located on top of the semiconducting channel instead, which makes it more straightforward to fabricate. It requires a lower operating gate voltage, but it is relatively little used for sensing applications as well because the top gate insulates the sensitive channel from the environment. A variation on the top gate structure, the liquid gate structure, consists in applying the gate voltage through the electrolyte surrounding the device [114]. It is particularly interesting for chemical sensing in water because it allows much lower-voltage operation (in the sub-volt range) compared to the usual bottom-gate structure (Figure 4a) [115]. It is also more straightforward to fabricate considering that it requires one less electrode by an embedded gate structure compared to the top gate structure [112]. Hybrid CNT-FET architectures consist in coupling in the same architecture several gating strategies. Notably, Pyo et al. [54] fabricated a double-gated CNT-FET with a separated extended gate. The extended gate concept consists on placing an ion-sensitive membrane on top of the top gate of the double-gated CNT-FET (Figure 4c).

SiO₂ is the most frequently used dielectric material used for the layer between the semiconducting channel and the substrate or gate electrode (Figure 4b) (6 papers out of

11). However, oxides with a higher dielectric constant such as Al_2O_3 or Si_3N_4 may also be used in order to have a thinner insulating layer with better homogeneity and durability compared to SiO_2 [116]. Takeda et al. used a sol-gel layer of 3-aminopropyltriethoxysilane (APS) as insulating layer, with the goal to better immobilize the SWCNTs in water [106]. Indeed, an electrostatic attraction occurs between the negatively-charged carboxyl groups of SWCNTs and the positively-charged APS.

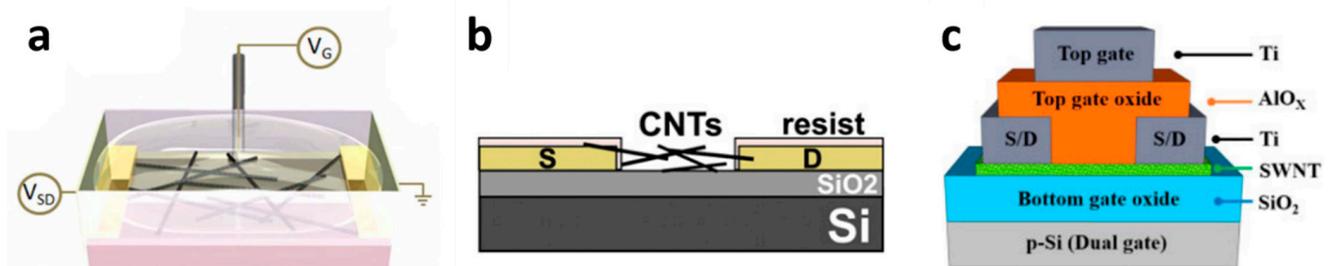


Figure 4. Topology of (a) liquid top gate, (b) bottom gate and (c) extended-dual gate CNT-FET. Reproduced from [54,60,107]. Reprinted with permission. CC BY 3.0.

2.6. Sensing Mechanisms in CNT-Based Chemistors and ChemFET

The mechanisms of sensitivity of CNT-based chemistors and chemFET are usually extrapolated from their mechanisms of sensitivity to gas [117], mechanisms that still remain somewhat debated. Overall, the response to analytes is attributed to a change in the conduction properties of either, or all, of the three following components of the devices, as shown in Figure 5 [15]: The conduction along the tube length (“intra-CNT”), the contact points between tubes behaving as tunnel junctions (“inter-CNT”) and the contact points between the tubes and the metal electrodes behaving as Schottky barriers. Sensitivity is attributed either to direct adsorption of the analytes on these sites, or to analytes not adsorbed, but at a distance small enough to these sites to perturb their electrical behavior.

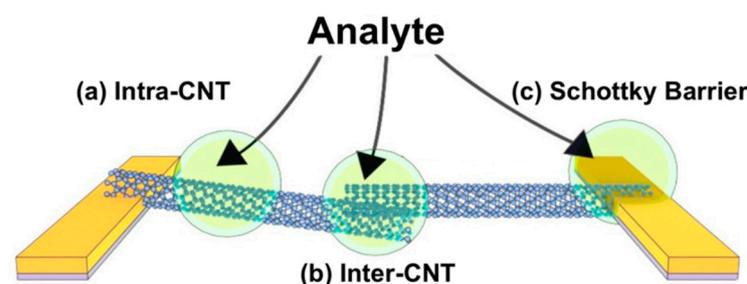


Figure 5. Schematic of possible sensitivity sites which affect the conductivity: (a) at the sidewall or along the length of the CNT itself, (b) interface between CNT-CNT (inter-CNT) and (c) at the interface between the metal electrodes and the CNT (Schottky barrier). Reprinted with permission from [15]. Copyright (2018) from American Chemical Society.

Modulation of the Schottky barrier is caused by a change in the work function of either the electrode metal or the CNTs in presence of the target analytes. The inter-CNT modulation corresponds to a change in the transmission coefficient of the inter-tube tunneling junction, which in turn can be attributed to either a change in the inter-tube distance, or a change in the work functions of the tubes. The intra-CNT conduction modulation is caused by a change either in the density of charge along the sidewalls (resulting in a doping effect) or in the carrier scattering properties of the sidewalls (impacting mobility).

Each of these three modulations may impact the global device response, with specificities depending on the type of transduction (ChemET or chemistor) and on the device morphology (particularly on the network density).

Regarding ChemFETs, Figure 6 provides an insight into the impact of the Schottky-barrier modulation and the intra-CNT modulation (doping or mobility variation) on the typical I-V characteristics of a chemFET [15]. Inter-CNT effects (contact resistance modulation) are usually neglected when analyzing sensing performances of chemFET as their electric performances are mostly driven by intra-CNT effects and Schottky barrier modulation.

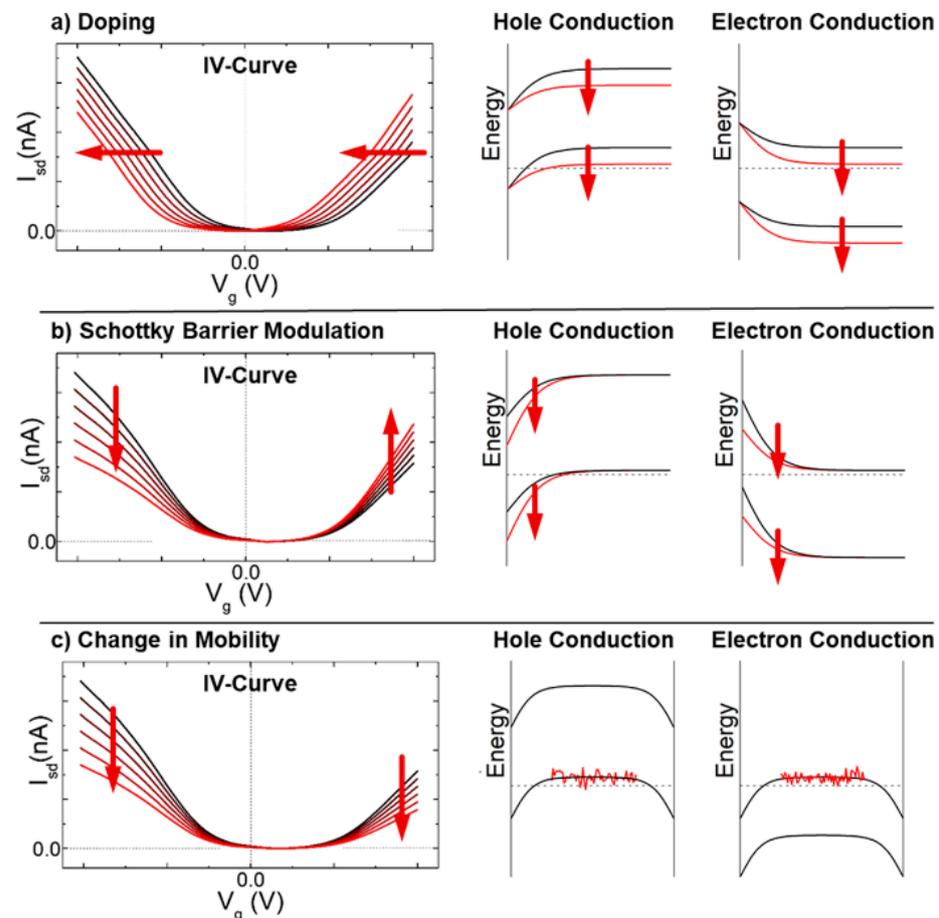


Figure 6. Theoretical I-V curves of a chemFET depending on (a) the doping (b) Schottky barrier modulation (c) change in mobility. Reprinted with permission from [15]. Copyright (2018) from American Chemical Society.

By contrast, in chemistors, the inter-CNT modulation is generally accepted to have the strongest impact on the device response, as the global baseline resistance of the network is mostly controlled by inter-CNT contacts. This is confirmed by modeling results for high density networks. In low density networks, modeling suggests that the variations of the electrode-CNT resistance and of the intra-CNT resistance may also contribute to the global relative resistance variation occurring upon exposure to chemicals [118].

3. CNT-Based Sensors with Different Analytes in Water

Table 1 summarizes the contents of the 90 reported references on CNT-based sensors considered in this review. The review includes references dealing with all the water-quality relevant analytes discussed until April 2021 except for pesticides, this specific and very large topic having been reviewed very recently elsewhere [119,120]. The most investigated analytes are H^+ (pH) and lead (with 18% of references each), then cadmium (14%) and nitrite (11%). Altogether, micronutrients and toxic metals cover 37 papers, so 40% of references, a lot of these references covering several analytes.

Table 1. Overview of reported CNT-based water quality monitoring sensors.

| Type of Analytes | Numbers of Refs. | Ref. SWCNT | Ref. MWCNT | Ref. CNTFET | Ref. Chemistors | Ref. EC | Ref. Functionalized (COOH Excluded) | |
|---|-------------------|-----------------|-----------------|-----------------|-----------------|-------------------|-------------------------------------|----|
| All Analytes | 90 | 26 (29%) | 64 (71%) | 11 (12%) | 13 (14%) | 66 (73%) | 74 (82%) | |
| pH | 16 (18%) | 12 | 4 | 6 | 7 | 5 (2 with CNTFET) | 8 | |
| Micronutrients and toxic metals (total) | All included | 36 (40%) | 5 | 32 | 2 | 2 | 33 | 31 |
| | Pb(II) | 16 (18%) | 0 | 16 | 0 | 0 | 16 | 12 |
| | Cd (II) | 13 (14%) | 0 | 13 | 0 | 0 | 13 | 11 |
| | Cu(II) | 9 (10%) | 1 | 9 | 1 | 0 | 8 | 8 |
| | Hg(II) | 8 (9%) | 3 | 5 | 1 | 1 | 6 | 6 |
| | As(III) | 5 (6%) | 0 | 5 | 0 | 0 | 5 | 5 |
| | Zn(II) | 4 (4%) | 0 | 4 | 0 | 0 | 4 | 2 |
| Miscellaneous | 2 (2%) | 2 | 0 | 1 | 1 | 0 | 2 | |
| Nitrite | 10 (11%) | 1 | 9 | 0 | 0 | 10 | 10 | |
| Water hardness | 2 (2%) | 1 | 1 | 1 | 0 | 1 | 2 | |
| DO | 2 (2%) | 0 | 2 | 0 | 0 | 2 | 2 | |
| Disinfectants | Free chlorine | 3 (3%) | 1 | 2 | 0 | 2 | 1 | 2 |
| | Hydrogen peroxide | 6 (7%) | 1 | 5 | 0 | 1 | 5 | 6 |
| Sulfur | Sulfide | 4 (4%) | 0 | 4 | 0 | 0 | 4 | 4 |
| | Sulfite | 2 (2%) | 0 | 2 | 0 | 0 | 2 | 2 |
| Miscellaneous | 9 (10%) | 4 | 5 | 2 | 1 | 5 | 8 | |

The large majority of reports addresses MWCNTs (71%), functionalized CNTs (82%) and electrochemical transduction—73%—(14% for chemistors and 12% for ChemFETs).

The following sections provide for each analyte a table with the highlights of the corresponding references. The tables include the following information:

- Materials: type of CNT (MWCNT or SWCNT), functional probe and type of functionalization (covalent or not),
- Device strategy: type of transduction (including type of electrochemical measurement and type of FET, gating), CNT deposition process, electrode material and configuration, choice of substrate
- Performances: limit of detection (LOD) (converted in the most used unit for the target analyte), sensitivity in the measured range of concentration (converted whenever possible in a common unit) and the results of interference study

For each analyte, the best results are figured out for discussions on the choice of functionalization and of the transduction strategies. Whenever possible, the impact of the fabrication strategy is discussed.

3.1. pH

pH is the physicochemical quantity defined as $\text{pH} = -\log[\text{H}_3\text{O}^+]$ [121]. Recommended levels of pH for drinking water are 6.5 to 8.5 according to World Health Organization [122]. Commercial pH sensors for water monitoring applications often cover larger ranges of pH (regularly from 1 to 14) to address abnormal situations regarding drink water (e.g., pollutant ingress) and to be applicable to industrial water processing (chlorination, waste water, aquaculture) as well.

Table 2 shows the 16 reported CNT-based pH sensors: seven are chemistors, six are chemFETs and five are electrochemical sensors (among which two are CNT-FET operated as EC sensors). Only four reports out of 16 use MWCNTs; eight reports address non-functionalized CNTs (including CNT-COOH). This is in contrast with other analytes (as summarized in Table 1), where functionalization is quasi-systematic and the use of chemistors is rare.

By contrast to other analytes also, the use of detection limits (LOD) expressed in M of H^+ as a mean of comparison between references is challenging because, due to the logarithmic pH scale, these LOD do not translate directly into pH detection limits. Moreover, it is only rarely provided (here seven papers out of 16 only). Because of the variety of reported response types (current, voltage, resistance, conductance, impedance, percentages) the absolute sensitivities (e.g., variation of the response by pH unit) cannot be compared either. As an alternative, we elected here to compare the relative sensitivity at pH 7 (e.g., the variation of the response by pH unit divided by the response at pH 7) which allows for comparison across transduction methods.

Using this indicator, we observed that the 3 non-functionalized and the COOH-based chemistors have considerably better performances than the functionalized ones, the best performance being achieved at 18%/pH unit ($63 \Omega/\text{pH unit}$) with MWCNTs sucked by vacuum force on filter paper [123].

For FET as well, the best performance (23%/pH unit) is achieved with spin coated non-functionalized SWCNT in a dual gate chemFET structure [54], the authors showing that double-gated operation performs better than single-gated.

The same performance (23%/pH unit) is achieved with impedance spectroscopy of COOH-functionalized MWCNT spin-coated on Kapton[®] with gold electrodes [59]. Using Aluminum electrode leads to a significant decrease in performance (14%/pH unit).

These results underline the very good sensitivity to pH of pristine CNTs and CNT-COOH for all three types of transduction methods, while the use of other functional probes degrades performances. This confirms the widespread theory mentioned in Section 2.2.2 that the sensitivity of pristine CNT and CNT-COOH to pH is due to the presence of carboxyl groups on the CNT sidewalls.

It is worth mentioning that the five references on potentiometry yield sensitivities in mV/pH unit very close to Nernst law irrespective of the functionalization (58 mV/pH unit) [54,60,124–126].

Finally, one observes that, while there are several reports with the same transduction mode (particularly resistive) and the same functionalization (pristine) [57,58,123,126], it is not possible to draw conclusions regarding to optimal design and fabrication of pH sensors as there is still too much variability in the choice of substrates, electrode materials and deposition method.

3.2. Micronutrients and Heavy Metals

Micronutrients (Iron, manganese, cobalt, copper, molybdenum, zinc, selenium, cadmium, iodine, boron, fluorine . . .) are mineral materials that play an important role in metabolic activities and tissue function maintenance in living beings. Subsequently, a suitable intake of micronutrients is necessary, often in trace amounts, and they should not be entirely removed from the water supply. However, they usually become harmful at higher doses and constitute a water quality concern.

Even though various heavy metals, notably cadmium, lead and mercury, are not micronutrients as they are very toxic even in trace amount, a lot of micronutrients actually are heavy metals (notably iron, copper, cobalt and zinc). Hence, micronutrients and heavy metals are often classified jointly in the water quality literature. We proceed in the same manner here.

Metal ions detection in water by CNT-based sensors has been heavily studied since 2005 with 37 papers reported out of 90 papers in total in this review, the large majority of these dealing with electrochemical transduction (33 papers) and functionalized CNTs (31 papers). In the following subsections, we summarize the results on the following ions by order of frequency of occurrence in the literature: Pb(II), Cd(II), Zn(II), Cu(II), Hg(II), As(III), Ni(II) and Co(II). We then discuss the multiplexing performances and the interference studies. The performances of the reported sensors are compared in terms of limit of detection which is the most frequently provided indicator.

Table 2. CNT-based pH sensors in water, sorted by transduction type then by relative sensitivity.

| Type of CNT | Functional Probe | Functionalization | Analyte | Detection Range | Detection Limit | Sensitivity Relative Sensitivity * | Transduction Method | CNT Deposition Method | Electrode Material Contact Configuration | Substrate | Comments | Ref. |
|-------------|-------------------------|--------------------|---------|-----------------|-----------------|--|----------------------------------|---|---|---|---|-------|
| SWCNT | Polyaniline | Non covalent | pH | pH 2.1~12.8 | 2.74 nM | N/A | Chemistor | Drop-casting | Ti/Au | Si/SiO ₂ | | [126] |
| SWCNT | Nafion | Non covalent | pH | pH 1~12 | N.P. | 3.5%/pH | Chemistor | Screen printing | SWCNT | Polymide | | [127] |
| MWCNT | Ni NP * | Non covalent | pH | pH 2~10 | N.P. | 5.0%/pH | Chemistor | Continuous pulling of super-aligned, CVD grown MWCNTs | MWCNT | PDMS | | [89] |
| SWCNT | Pristine | Non functionalized | pH | pH 1~11 | <10 pM | 34 nS/pH 3.4%/pH (pH 1~6) 163 nS/pH 9.3%/pH (pH 7~11) | Chemistor | Spray-casting | Cr | Si/SiO ₂ | | [57] |
| SWCNT | COOH | Covalent | pH | pH 5~9 | N.P. | 75 Ω/pH 11%/pH | Chemistor | Dielectrophoresis (aligned CNTs) | Cr/Au | Si/SiO ₂ | Response time: 2 s at pH 5, 24 s at pH 9 | [88] |
| SWCNT | Pristine | Non functionalized | pH | pH 4~10 | N.P. | 5.2 kΩ/pH 14%/pH | Chemistor | Aerosol jet printing | Ag | Kapton | | [58] |
| MWCNT | Pristine | Non functionalized | pH | pH 5~9 | N.P. | 63 Ω/pH 18%/pH | Chemistor | Sucked by vacuum force | MWCNT | Filter paper | | [123] |
| SWCNT | Malt extract agar | Non covalent | pH | pH 3~5 | 100 mM | N/A | FET (hybrid top gate) | Dip coating | Ti/Au (10/30 nm) contacts | Si/SiO ₂ (100 nm) | Multiplexed detection of Fungus (<i>A. niger</i> , <i>A. versicolor</i>) and Yeast (<i>S. cerevisiae</i>) * | [105] |
| SWCNT | ETH500 *, MDDA-Cl | Non covalent | pH | pH 2~7.5 | 10 mM | 71 nA/pH 7.5%/pH | FET (liquid gate) | Spray deposition | Aqueous electrolyte (gate) Cr/Au (5/50 nm) | Polymide (Kapton®) | Change from p-type to n-type transistor with the membrane layer | [112] |
| SWCNT | COOH | Covalent | pH | pH 3~8 | N.P. | 17 nA/pH 8.2%/pH | FET (top gate) | N.P. | Cr/Au (30/50 nm) source & drain electrodes, Ag/AgCl for reference electrode | Glass/APS(50~200 nm)/SWCNT /APS(500 nm)/TopGate | CNT placement controlled by location of APS (modified to immobilize the CNTs) | [106] |
| SWCNT | Pristine | Non functionalized | pH | pH 3.4~7.8 | 10 mM | 3.9 μA/pH 13%/pH | FET (bottom gate) | Spin coating | Cr/Au (5/40 nm) | Si/SiO ₂ (65 nm) | | [107] |
| SWCNT | Poly(1-aminoanthracene) | Non covalent | pH | pH 3~11 | 1 μM | FET 19 μS/pH 14%/pH potentiometry 55 mV/pH | FET, potentiometry (liquid gate) | Dielectrophoresis (aligned CNTs) | Au contacts, Pt wire (Auxillary), Ag/AgCl electrode (Reference) | Si/SiO ₂ (300 nm) | Multiplexed detection of Ca(II) and Na ⁺ | [60] |

Table 2. Cont.

| Type of CNT | Functional Probe | Functionalization | Analyte | Detection Range | Detection Limit | Sensitivity Relative Sensitivity * | Transduction Method | CNT Deposition Method | Electrode Material Contact Configuration | Substrate | Comments | Ref. |
|-------------|---------------------------|-----------------------|---------|-----------------|-----------------|---|-----------------------------------|--------------------------|---|--|--|-------|
| SWCNT | Pristine | Non functionalized | pH | pH 3~10 | 1 mM | 7600 mV/pH 23%/pH (Dual-gate mode) 59.5 mV/pH (single-gate mode potentiometry) | FET (double gate) | Spin coating | 100 nm Ti contacts for source, drain and top gate | p-Si (substrate acting as bottom gate) | | [54] |
| SWCNT | Polyaniline | Non covalent | pH | pH 1~13 | N.P. | 56 mV/pH | potentiometry | Spray casting | Polyvinyl chloride-coated steel wire | PVC | Highly selective against Li ⁺ , Na ⁺ , K ⁺ | [128] |
| MWCNT | COF _{THI-TFPB} * | Covalent | pH | pH 1~12 | N.P. | 54 mV/pH | Differential pulse voltammetry | Drop casting | Glassy carbon electrode | Glassy carbon | multiplexed detection of Ascorbic acid. | [125] |
| MWCNT | COOH | Covalent | pH | pH 4~9 | N.P. | 17 Ω/pH 23%/pH (Au), 16 Ω/pH 14%/pH (Al) | Impedance spectroscopy | Dip coating | Au and Al interdigitated electrodes | Kapton® | | [59] |

* The relative sensitivity is calculated using the formula $Relative\ Sensitivity = (x/x_0) * 100\ (%)$, with x the absolute sensitivity expressed (depending on the transduction) in units of resistance, voltage or current per pH unit and x_0 the baseline parameter (resistance, voltage or current) at pH 7. The relative sensitivity is not calculated for potentiometry-based transduction as it depends on the choice of reference voltages and the three references can be easily compared by their absolute sensitivity. N.P.: not provided Ni NP: Nickel nanoparticle, PDMS: Polydimethylsiloxane, APS: 3-aminopropyltriethoxysilane, ETH500: tetradodecylammonium tetrakis(4-chlorophenyl)borate, MDDA-Cl: methyltridodecylammonium chloride, *A. niger*: *Aspergillus niger*; *A. versicolor*: *Aspergillus versicolor*, *S. cerevisiae*: *Saccharomyces cerevisiae*, PVC: poly vinyl chloride, COF: Covalent organic framework, Thi: Thionine; TFPB: 1,3,5-tris(p-formylphenyl)benzene.

3.2.1. Detection of Pb(II)

Table 3 summarizes the different CNT-based Pb(II) sensors used for water quality monitoring. All references but three are based on functionalized MWCNTs sensors using stripping voltammetry, the other ones using stripping voltammetry with pristine MWCNTs [129,130] or potentiometry with functionalized MWCNTs [131,132]. Interestingly for comparison purposes, 6 out of 16 references discuss functionalized MWCNTs drop cast onto glassy carbon electrodes and operated through stripping voltammetry.

The reported ranges of detection cover a large scale, from 0.1 ppt to 100 ppb. By comparison, the maximum acceptable concentration (MAC) of lead worldwide ranges from 10 to 15 ppb [7]. The lowest limits of detection are 0.3 ppb, 0.04 ppb and 0.02 ppt with pristine [129], non-covalently functionalized (Nafion/Bismuth [73]) and covalently functionalized (Dithizone [132]) MWCNTs, respectively.

References [129,130] report the use of pristine CNTs and show that while the limits of detection (0.3 ppb and 1 ppb, respectively) of a pristine CNT electrode are theoretically sufficient for lead detection in the context of water quality monitoring, the ranges of detection are not compatible (lowest limit at respectively 210 ppb and 15 ppb). While ref [129] demonstrates that joint detection of several metal ions is possible (as expected from the principles of stripping voltammetry) both references remark on interferents (dissolved oxygen in [129] and cadmium and copper in [130]), as is expected from pristine nanotubes (non-selective sensing).

Concerning covalent functionalization, references [74,132,133] are based on the same architecture (drop casting of CNT dispersion on glassy carbon and stripping voltammetry), so the major differences in limits of detection can easily be linked to the functionalization strategy. Refs. [74,134] rely on grafting respectively cysteine and thiacalixarene (TCA) on MWCNT by exploiting their sidewall carboxyl groups as reaction sites. The major difference in performances is that TCA is much more favorable for lead complexation than cysteine. Notably, ref. [134] shows by computational method that Pb(II) ions can stably adsorb onto the TCA molecules and that there is a significant electron delocalization between Pb(II) and the sulfur atoms in the TCA molecule. To move beyond the performances of TCA-functionalized CNTs, ref. [132] relies not only on complexation of MWCNTs by dithizone (as thiols have strong interaction with metal ions) but also on the processing of MWCNTs into a bucky-gel, a porous MWCNT-based structure filled with ionic gel (here 1-butyl-3-methylimidazolium hexafluorophosphate). While the functionalization provides reactivity to the target metal ion, the bucky gel is thought to provide strong specific surface area enhancement [135].

In references reporting non-covalent functionalization strategies, bismuth or its derivatives are used in the majority of reports (8 out of 10), the other references citing mercury [131] and antimony oxide [136]. Bismuth is an environmentally friendly material often used as a replacement to mercury in electrochemical applications [137]. Its sensitivity to lead is attributed to its ability to form “fusible” alloys with heavy metals in general. It tends to facilitate their nucleation and subsequent reduction. However, this nucleation process of lead ions around bismuth could in theory limit the reusability of the sensing devices. In practice, Xu et al. (the reference with the best limit of detection among this category) report that the relative standard deviation (RSD) on sensitivity was lower than 5% for Pb(II) detection after 50 repetitive measurements [73].

It is worth noting that the three references about covalent functionalization did not identify any interferent materials, while among seven references reporting on non-covalent functionalization which discussed interferent, only one features no interferent [73], the others mentioned reduced or strong interferents. It suggests that, in the context of lead(II) monitoring, covalent functionalization yields not only better limits of detection but also better selectivity than non-covalent functionalization.

References [138,139] also give the opportunity to discuss the specificity of the stripping voltammetry protocol. Indeed, both references using the exact same materials and process (plating of Bismuth on screen printed CNT electrodes), Injang et al. [138] achieved an

improvement of almost one order of magnitude on lead(II) LOD and sensitivity compared to Hwang et al. [139] by complementing anodic stripping voltammetry (ASV) with sequential injection analysis [140]. It is a technique known to enhance sensitivity and selectivity in stripping voltammetry by better controlling reagent and sample volumes. In these two papers, the benefit for this technique occurs only for Pb(II), the two papers reporting similar LOD and sensitivity for Cd(II) and Zn(II) detection (see next sections). This suggests that the optimization of the electrochemical transduction scheme may have a very strong impact on sensitivity and selectivity, possibly stronger than the functionalization choice. However, this impact can rarely be assessed in the literature as it is seldom discussed and a reliable comparison between protocols requires references with very strong similarities in electrode design, fabrication process and material choices.

3.2.2. Detection of Cd(II)

Table 4 presents the 13 CNT sensors reported for Cd(II) detection in water. Similarly to the references reporting the detection of Pb(II), all references but one is based on functionalized MWCNTs sensors using stripping voltammetry, the other ones using stripping voltammetry with pristine MWCNTs [129] or stripping potentiometry with functionalized MWCNTs [131]. All except one paper [141] are common with the reports on Pb(II) sensors in the previous section. Unfortunately, there is no reference about covalently functionalized CNT for cadmium sensing.

The reported detection limits lie between 0.02 ppb and 17 ppb to be compared to the MAC of Cd(II) in water which stands between 3 and 5 ppb. Similarly to the case of lead, ref. [129] reporting the use of pristine CNTs shows an acceptable detection limit (0.23 ppb) but a range of detection not compatible with water quality monitoring (lowest limit at 170 ppb).

For the same reason as for Pb(II) detection, all but five papers out of 13 use functionalization compounds integrating bismuth. The best result in term of LOD (0.02 ppb) is achieved with non-covalent functionalization with Poly(sodium4-styrenesulfonate)-Bismuth (PSS-Bi) [142]. The remarkable LOD is attributed to the wrapping of the PSS polymer around the CNTs, providing a high density of adsorbing sites for metal binding without affecting the electronic properties of the CNTs.

From these references, and those about lead as well, one may note that paste-based approaches do not perform very well overall in terms of limits of detection compared to more traditional processing of MWCNTs.

Table 3. CNT-based sensors for detecting (II)Pb(II) ions in water, sorted by type of functionalization then detection limit.

| Type of CNT | Functional Probe | Functionalization | Analyte (Add. Analytes) | Detection Limit | Sensitivity (Detection Range) | Transduction Method | Deposition Method | Electrode Material Contact Configuration | Substrate | Interference Study | Ref. |
|-------------|--|--------------------|------------------------------------|-----------------|--|---|---|--|-----------------------|--|-------|
| MWCNT | Pristine | Non functionalized | Pb(II) (Cd(II), Zn(II), Cu(II)) | 0.3 ppb | 2.2 nA/ppb (210–830 ppb) | Stripping voltammetry | CNT thread | Metal wire and silver conductive epoxy | Glass capillary | Simultaneous determination of Cd(II), Cu(II), Pb(II) and Zn(II) demonstrated The presence of Dissolved Oxygen changes the calibration law for Cd(II) | [129] |
| MWCNT | Pristine | Non functionalized | Pb(II) | 1.0 ppb | 1.5 nA/ppb (15–40 ppb) 3.5 nA/ppb (40–70 ppb) | Stripping voltammetry | Inkjet printing | Inkjet-printed silver ink | PEN * | Effects of copper and cadmium are reported. | [130] |
| MWCNT | Ionic liquid—dithizone based bucky-gel | Covalent | Pb(II) | 0.02 ppt | 0.024 μ A/ppb (0.1ppt–210 ppb) | Stripping voltammetry | Drop-casting | Glassy carbon electrode | Glassy carbon | No interference of Cd(II) and Cu(II) ions with the detection of Pb(II) ion. | [132] |
| MWCNT | Thiacalixarene | Covalent | Pb(II) | 8 ppt | 3.8 μ A/ppb (0.04–2.07 ppb) | Differential pulse anodic stripping voltammetry | Drop casting | Glassy carbon electrode | Glassy carbon | Detection of Pb(II) was clearly not affected by Zn(II), Cd(II), Ni(II) (100-fold excess) | [134] |
| MWCNT | Cysteine | Covalent | Pb(II) (Cu(II)) | 1 ppb | 0.23 * μ A/ppb (25–750 ppb) | Differential pulse anodic stripping voltammetry | Drop casting | Glassy carbon electrode | Glassy carbon. | 40-fold Cl^- , 30-fold SO_4^{2-} and four fold CO_3^{2-} did not have any significant effect on the stripping peak current of Pb(II) and Cu(II) | [74] |
| MWCNT | Poly(o-toluidine) Ce(III)tungstate | Covalent | Pb(II) | 210 ppb | 27 mV/decade (0.1 ppt–100 ppb) | Potentiometry | Liquid mixing and membrane formation through drying | Calomel electrode | Glass tube (araldite) | Strong selectivity (from 50 to 500 times) against Zn(II), Sr(II), Hg(II), Ca(II), Pd(II), Cu(II), Mg(II) | [133] |
| MWCNT | Nafion/Bismuth | Non covalent | Pb(II), (Cd(II)) | 25 ppt | 0.22 μ A/ppb (0.05 to 5 ppb) 0.27 μ A/ppb (5–100 ppb) | Stripping voltammetry | Drop casting | Glassy carbon electrode | Glassy carbon | 500-fold of SCN^- , Cl^- , F^- , PO_4^{3-} , SO_4^{2-} , NO_3^- and various cations such as Na^+ , Ca(II), Mg(II), Al(III), K^+ , Zn(II), Co(II) and Ni(II) had no influences on the signals of Pb(II) and Cd(II). | [73] |
| MWCNT | PSS-Bi * | Non covalent | Pb(II) (Cd(II)) | 0.04 ppb | 0.079 μ A/ppb (0.5–90 ppb) | Stripping voltammetry | Drop casting | Glassy carbon electrode | Glassy carbon | 20-fold amounts of Zn(II), 5-fold amounts of Sn(II) and 1-fold amounts of Cu(II) have influence on the determination of Cd(II) and Pb(II) with deviation of 10%. | [142] |
| MWCNT | Bismuth | Non covalent | Pb(II) (Cd(II)) | ~0.04 ppb | N/A | Stripping voltammetry | Plasma-enhanced CVD (vertically aligned MWCNTs in epoxy matrix) | Cr | Silicon | N.P. * | [143] |
| MWCNT | Fe_3O_4 -LSG-CS-Bi * | Non covalent | Pb(II) (Cd(II)) | 0.07 ppb | 0.21 μ A/ppb (1–20 ppb) 0.24 μ A/ppb (20–200 ppb) | Stripping voltammetry | Drop casting | Glassy carbon electrode | Glassy carbon | Slight changes in peak currents of Pb(II) and Cd(II) were observed in presence of interfering ions Na^+ , Cl^- , SO_4^{2-} , PO_4^{3-} , Fe(II), Fe(III), Zn(II), As(III). Significant increase in response signals of Hg(II) was probably due to the formation of amalgam Dramatically decreased response signals of Cu(II) were ascribed to the formation of Pb-Cu inter-metallic compounds. | [144] |

Table 3. Cont.

| Type of CNT | Functional Probe | Functionalization | Analyte (Add. Analytes) | Detection Limit | Sensitivity (Detection Range) | Transduction Method | Deposition Method | Electrode Material Contact Configuration | Substrate | Interference Study | Ref. |
|-------------|----------------------------------|-------------------|----------------------------|-----------------|--|-------------------------|---|--|----------------------|--|-------|
| MWCNT | PPy-Bi NPs * | Non covalent | Pb(II) (Cd(II)) | 0.1 ppb | 1.1 $\mu\text{A/ppb}$ (0.11–120 ppb) | Stripping voltammetry | Paste mixture with MWCNT, paraffin oil and graphite powder | Stainless steel rod | Teflon (PTFE *) tube | Good selectivity towards Fe(II), Al(III), Zn(II), Mg(II), SO_4^{2-} , CO_3^{2-} , Ca(II), K^+ , Na^+ . The absolute relative change of signal varied from 0.40 to 4.88%. High interference from Cu(II) (1-fold mass ratio was found as the tolerance ratios for the detection of Pb and Cd ions) | [145] |
| MWCNT | rGO-Bi * | Non covalent | Pb(II) (Cd(II)) | 0.2 ppb | 930 nA/ppb cm^2 (20–200 ppb) | Stripping voltammetry | Spray coating | Cr(30 nm)/Au(200 nm) | Polymide (VTEC 1388) | 100-fold K^+ , Na^+ , Ca(II), Cl^- , NO_3^- and a 30-fold Fe(III) increase had no significant effect on the signals of Cd and Pb ions. Cu ions were found to reduce the response of target metal ions due to the competition between electroplating Bi and Cu on the electrode surface (close reduction potential of Cu and Bi.) | [146] |
| MWCNT | Bismuth | Non covalent | Pb(II) (Cd(II),Zn(II)) | 0.2 ppb | 0.39 $\mu\text{A/ppb}$ (2–18 ppb) 0.67 $\mu\text{A/ppb}$ (20–100 ppb) | Stripping voltammetry | Screen printing | Screen printed MWCNT based electrode | Ceramic substrates | N.P. | [138] |
| MWCNT | Bismuth | Non covalent | Pb(II) (Cd(II),Zn(II)) | 1.3 ppb | 1.2 $\mu\text{A/ppb}$ (2–100 ppb) | Stripping voltammetry | Screen printing | Screen printed MWCNT based electrode | Alumina plates | The addition of copper ions strongly influenced the stripping responses. Decrease of lead and cadmium pics by 65.5%. | [139] |
| MWCNT | Pristine | Non covalent | Pb(II) (Cd(II), Zn(II)) | 6.6 ppb | 0.47 * s/V/ppb (58–650 ppb) | Stripping potentiometry | Paste mixture of MWCNT and mineral oil | MWCNT paste electrode | Glass tube | Al(III), Mg(II), Fe(III), Ni(II), Co(II), Cr(III), Cu(II) and Sb(III) were investigated in the ratio analyte: Interferent 1:1 and 1:10. The interference was observed for the ratios analyte: interferent 1:1 and 1:10 for Co(II), 1:10 for Cr(III) and Cu(II). | [131] |
| MWCNT | Sb ₂ O ₃ * | Non covalent | Pb(II) (Cd(II)) | 24 ppb | 2.7 $\mu\text{A/ppb}$ (5–35 ppb) | Stripping voltammetry | Paste mixture of MWCNT, silicon oil, Sb ₂ O ₃ powder and ionic liquid | Copper wire | PTFE tube | N.P. | [136] |

* N.P.: Not provided. PSS-Bi: Poly(sodium 4-styrenesulfonate)-Bismuth. rGO-Bi: Reduced graphene oxide-Bismuth. PPy-Bi NPs: Polypyrrole-Bismuth NanoParticles. PTFE: poly tetra fluoro ethylene. LSG-Cs-Bi: laser-scribed graphene-chitosan-Bismuth. Sb₂O₃: antimony oxide. PEN: polyethylene naphthalate. 1 μM Pb(II) = 210 ppb Pb(II).

Table 4. CNT-based sensors for detecting Cd(II) ions in water, sorted by type of functionalization then by detection limit.

| Type of CNT | Functional Probe | Functionalization | Analyte (Add. Analytes) | Detection Limit | Sensitivity (Linear Range) | Transduction Method | Deposition Method | Electrode Material Contact Configuration | Substrate | Interference Study | Ref. |
|-------------|--------------------------------------|--------------------|--------------------------------------|-----------------|---|-----------------------|---|--|--------------------|---|-------|
| MWCNT | Pristine | Non functionalized | Cd(II) (Pb(II) Zn(II), Cu(II)) | 0.23 ppb | 3.9 nA/ppb (170–500 ppb) | Stripping voltammetry | CNT thread | Metal wire and silver conductive epoxy | Glass capillary | Simultaneous determination of Cd(II), Cu(II), Pb(II) and Zn(II) demonstrated The presence of Dissolved Oxygen changes the calibration law for Cd(II) | [129] |
| MWCNT | PSS-Bi * | Non covalent | Cd(II) (Pb(II)) | 0.02 ppb | 0.23 μ A/ppb (0.5–50 ppb) | Stripping voltammetry | Drop casting | Glassy carbon electrode | Glassy carbon | 20-fold amounts of Zn(II), 5-fold amounts of Sn(II) and 1-fold amounts of Cu(II) have influence on the determination of Cd(II) and Pb(II) with deviation of 10%. | [142] |
| MWCNT | Nafion/Bismuth | Non covalent | Cd(II) (Pb(II)) | 0.04 ppb | 0.18 μ A/ppb (0.08–5 ppb) 0.16 μ A/ppb (5–100 ppb) | Stripping voltammetry | Drop casting | Glassy carbon electrode | Glassy carbon | 500-fold of SCN^- , Cl^- , F^- , PO_4^{3-} , SO_4^{2-} , NO_3^- and various cations such as Na^+ , Ca(II) , Mg(II) , Al(III) , K^+ , Zn(II) , Co(II) and Ni(II) had no influences on the signals of Pb(II) and Cd(II). | [73] |
| MWCNT | Bismuth | Non covalent | Cd(II) (Pb(II)) | 0.04 ppb | 0.037 μ A/ppb (0.5–8 ppb) | Stripping voltammetry | Plasma-enhanced CVD (vertically aligned MWCNTs in epoxy matrix) | Cr | Silicon | N.P. | [143] |
| MWCNT | Fe_3O_4 -LSG-CS-Bi * | Non covalent | Cd(II) (Pb(II)) | 0.1 ppb | 0.097 μ A/ppb (1–20 ppb) 0.32 μ A/ppb (20–200 ppb) | Stripping voltammetry | Drop casting | Glassy carbon electrode | Glassy carbon | Slight changes in peak currents of Pb(II) and Cd(II) were observed in presence of interfering ions Na^+ , Cl^- , SO_4^{2-} , PO_4^{3-} , Fe(II), Fe(III), Zn(II), As(III). Significant increase in response signals of Hg(II) was probably due to the formation of amalgam Dramatically decreased response signals of Cu(II) was ascribed to the formation of Pb-Cu inter-metallic compounds. | [144] |
| MWCNT | PPy-Bi * | Non covalent | Cd(II) (Pb(II)) | 0.16 ppb | 0.47 μ A/ppb (0.16–120 ppb) | Stripping voltammetry | Paste mixture with MWCNT, paraffin oil and graphite powder | Stainless steel rod | Teflon (PTFE) tube | Good selectivity towards Fe(II), Al(III), Zn(II), Mg(II), SO_4^{2-} , CO_3^{2-} , Ca(II), K^+ , Na^+ . The absolute relative change of signal varied from 0.40 to 4.88%. High interference from Cu(II) (1-fold mass ratio was found as the tolerance ratios for the detection of Pb and Cd ions) | [145] |
| MWCNT | Poly(1,2-diaminobenzene) | Non covalent | Cd(II), (Cu(II)) | 0.25 ppb | 0.14 μ A/ppb (5–100 ppb) | Stripping voltammetry | Multipulse potentiostatic method | Glassy carbon electrode | Glassy carbon | N.P. | [69] |

Table 4. Cont.

| Type of CNT | Functional Probe | Functionalization | Analyte (Add. Analytes) | Detection Limit | Sensitivity (Linear Range) | Transduction Method | Deposition Method | Electrode Material Contact Configuration | Substrate | Interference Study | Ref. |
|-------------|--|-------------------|---------------------------|-----------------|--|-------------------------|--|--|----------------------|---|-------|
| MWCNT | rGO-Bi * | Non covalent | Cd(II) (Pb(II)) | 0.6 ppb | 26 nA/ppb cm ² (20–200 ppb) | Stripping voltammetry | Spray coating | Cr(30 nm)/Au(200 nm) | Polymide (VTEC 1388) | 100-fold K ⁺ , Na ⁺ , Ca(II), Cl ⁻ , NO ³⁻ and a 30-fold Fe(III) increase had no significant effect on the signals of Cd and Pb ions. Cu ions were found to reduce the response of target metal ions due to the competition between electroplating Bi and Cu on the electrode surface (close reduction potential of Cu and Bi.) | [146] |
| MWCNT | Bismuth | Non covalent | Cd(II) (Pb(II),Zn(II)) | 0.7 ppb | 0.22 μ A/ppb (2–18 ppb) 1.5 μ A/ppb (20–100 ppb) | Stripping voltammetry | Screen printing | Screen printed MWCNT based electrode | Alumina plates | The addition of copper ions strongly influenced the stripping responses. Decrease of lead and cadmium pics by 65.5%. | [139] |
| MWCNT | Bismuth | Non Covalent | Cd(II) (Pb(II),Zn(II)) | 0.8 ppb | 0.59 μ A/ppb (2–18 ppb) 0.80 μ A/ppb (20–100 ppb) | Stripping voltammetry | Screen printing | Screen printed MWCNT based electrode | Ceramic substrates | N.P. | [138] |
| MWCNT | Fe ₃ O ₄ /eggshell | Non covalent | Cd(II) | 2.4 ppb | 19 μ A/ppb (0.5–210 ppb) | Stripping voltammetry | Paste mixture of MWCNT, graphite powder, paraffin oil and Fe ₃ O ₄ -eggshell | Copper wire | Glass tube | 500-fold amounts of the following ions: Na ⁺ , Ca(II), Mg(II), Fe(III), Mn(II), Cr(III), Ba(II), Co(II), Hg(II), K ⁺ , NH ⁴⁺ , NO ₃ ⁻ , SO ₄ ²⁻ , PO ₄ ³⁻ made no alteration of the peak currents of Cd(II). 100-fold amounts of Sn(II) and Cu(II) with deviation of 9%, 50 fold amounts of Ni(II) and Zn(II) with deviations of 8% and 6% respectively had influence on the determination of Cd(II). | [141] |
| MWCNT | Pristine | Non covalent | Cd(II) (Pb(II),Zn(II)) | 8.4 ppb | 0.36 * s/V/ppb (58–646 ppb) | Stripping potentiometry | Paste mixture of MWCNT and mineral oil | MWCNT paste electrode | Glass tube | Al (III), Mg (II), Fe (III), Ni (II), Co (II), Cr (III), Cu (II) and Sb (III) were investigated in the ratio analyte: Interferent 1:1 and 1:10, the interference was observed for the ratios analyte: interferent 1:1 and 1:10 for Co (II), 1:10 for Cr (III) and Cu (II). | [131] |
| MWCNT | Sb ₂ O ₃ | Non covalent | Cd(II) (Pb(II)) | 17 ppb | 1.9 μ A/ppb (80–150 ppb) | Stripping voltammetry | Paste mixture of MWCNT, silicon oil, Sb ₂ O ₃ powder and ionic liquid | Copper wire | PTFE tube | N.P. | [136] |

* PSS-Bi: Poly(sodium 4-styrenesulfonate)-Bismuth. rGO-Bi: Reduced graphene oxide-Bismuth. PPy-BiNPs: Polypyrrole-Bismuth. LSG-Cs-Bi: laser scribed graphene-chitosan-Bismuth. Sb₂O₃: antimony oxide. 1 μ M Cd(II) = 112 ppb Cd(II).

3.2.3. Detection of Zn(II)

The maximum acceptable concentration of Zn(II) in water is 5 ppm. Detection of the Zn(II) ion in water using CNT sensors has only been investigated with electrochemical, MWCNT-based sensors (Table 5). The four references have been listed in the previous tables as they address also lead and cadmium. Unlike results on lead and cadmium where functionalized CNTs have much better performances than non-functionalized CNTs, one observes here that one of the two pristine MWCNTs references [129] has a remarkably better limit of detection (0.09 ppb—two orders of magnitude lower) than the two references with MWCNTs non covalently functionalized with bismuth. By contrast, the other reference with pristine MWCNT has worse detection limit than the Bi-functionalized devices. It points out once again the strong sensitivity of water quality sensor performances to CNT processing (CNT thread versus CNT paste) and to transduction mode (stripping voltammetry versus potentiometry).

3.2.4. Detection of Hg(II)

Table 6 presents CNT-based Hg(II) sensors that were reported to date. The literature is more varied in terms of transduction methods than for Pb(II), Cd(II) and Zn(II): one chemFET and one chemistor-based approaches (with pristine SWCNT) are proposed in addition to three references reporting stripping voltammetry and two others dealing with potentiometry (the last five reports being with functionalized MWCNT).

Only one paper reports the use of a chemistor. It uses pristine SWCNTs and it addresses the ppm range (LOD 0.6 ppm; range 1 to 30 ppm) which is not truly relevant for drink water monitoring as the maximum acceptable concentration of mercury allowed in water is 1 ppb [147]. By comparison, the pristine SWCNT-based chemFET structure provides widely improved performances (LOD of 2 ppb, range 0.2–200 ppm—still a bit high for drink water application). While an improvement on performances is expected when switching from chemistor to CNTFET, such a magnitude (two orders of magnitude of improvement) is usually not. It may be linked to the use of an octadecyl-trichlorosilane (OTS) self-assembled monolayer (SAM) in [110] to favor the adsorption of SWCNT on the SiO₂ substrate. This type of SAM is reported to improve FET electronic transport performances [148]. Interestingly, both references report very good selectivities despite the absence of functionalization, which suggests a strong natural affinity of pristine CNT to mercury. This is actually confirmed by studies that report water purification using CNTs and show remarkable adsorption capability of mercury by non-functionalized CNTs (containing CNT-COOH or -OH) [149].

It was observed that the LOD drops significantly and the range of detection shifts towards lower (and more relevant) detection limits when functionalization and electrochemical transduction are used. Covalent functionalization with thiophenol brings the LOD down to 0.6 ppb (range from 1 to 18 ppb) [150]. The lowest reported LOD is 2 ppt (range 2 ppt to 1000 ppm) [151]. It was reached (with differential pulse voltammetry) by functionalizing a 3D structure made of MWCNTs randomly arranged around graphene oxide sheets with bismuth-doped polyaniline chains (PANI). Once again, bismuth is used successfully for its ability to interact with heavy metals, while the 3D scaffold is thought to enhance the specific surface area.

Table 5. CNT-based sensors for detecting Zn(II) ions in water, sorted by detection limit.

| Type of CNT | Functional Probe | Functionalization | Analyte (Add. Analytes) | Detection Limit | Sensitivity (Linear Range) | Transduction Method | Deposition Method | Electrode Material Contact Configuration | Substrate | Interference Study | Ref. |
|-------------|------------------|--------------------|------------------------------------|-----------------|---|-------------------------|--|--|--------------------|---|-------|
| MWCNT | Pristine | Non functionalized | Zn(II) (Cd(II), Pb(II), Cu(II)) | 0.08 ppb | 3.4 pA/ppb (200–590 ppb) | Stripping voltammetry | CNT thread | Metal wire and silver conductive epoxy | Glass capillary | Simultaneous determination of Cd(II), Cu(II), Pb(II) and Zn(II) demonstrated The presence of Dissolved Oxygen changes the calibration law for Cd(II) | [129] |
| MWCNT | Bismuth | Non covalent | Zn(II) (Pb(II), Cd(II)) | 11 ppb | 0.18 μ A/ppb (12–18 ppb) 0.24 μ A/ppb (20–100 ppb) | Stripping voltammetry | Screen printing | Screen printed MWCNT based electrode | Ceramic substrates | N.P. | [138] |
| MWCNT | Bismuth | Non covalent | Zn(II) (Pb(II), Cd(II)) | 12 ppb | 0.38 μ A/ppb (20–100 ppb) | Stripping voltammetry | Screen printing | Screen printed MWCNT based electrode | Alumina plates | The addition of copper ions strongly influenced the stripping responses. Decrease of lead and cadmium pics by 65.5%. | [139] |
| MWCNT | Pristine | Non covalent | Zn(II) (Pb(II), Cd(II)) | 28 ppb | 0.11 * s/V/ppb (58–646 ppb) | Stripping potentiometry | Paste mixture of MWCNT and mineral oil | MWCNT paste electrode | Glass tube | Al(III), Mg(II), Fe(III), Ni(II), Co(II), Cr(III), Cu(II) and Sb(III) were investigated in the ratio analyte: Interferent 1:1 and 1:10, the interference was observed for the ratios analyte: interferent 1:1 and 1:10 for Co(II), 1:10 for Cr(III) and Cu(II). | [131] |

* 1 μ M Zn(II) = 65 ppb Zn(II).

Table 6. CNT-based sensors for detecting Hg(II) ions in water, sorted by type of functionalization, then detection limit.

| Type of CNT | Functional Probe | Functionalization | Analyte (Add. Analytes) | Detection Limit | Sensitivity (Linear Range) | Transduction Method | Deposition Method | Electrode Material Contact Configuration | Substrate | Interference Study | Ref. |
|-------------|-------------------------------|--------------------|-------------------------|-----------------|---|--------------------------------|---|--|-----------------------|---|-------|
| SWCNT | Pristine | Non functionalized | Hg(II) | 0.6 ppm | 12 mV/ppm (1–30 ppm) | Chemistor | CVD | SWCNT | Glass | 1000 fold excess of Fe(II), Fe(III), Ni(II), Cu(II), Zn(II), Cr(III) and 500 folds of As(III), Sb(III), Se(IV) and Pb(II) had no interfering effect in the analysis of mercury solution. | [147] |
| SWCNT | Pristine | Non functionalized | Hg(II) | 2 ppb | 0.22/decade 0.2 ppb–201 ppm | FET (Liquid gate) | Dip coating with selective CNT placement | Pd/Au (10/30 nm) | Glass | Good selectivity towards interferent ions (only Hg(II) causes conductance increase.) | [110] |
| SWCNT | Thiophenol | Covalent | Hg(II) | 0.6 ppb | 0.14 μ A/ppb (1–18 ppb) | Stripping voltammetry | Dip coating | Au | Au | The presence of 100-fold concentration of Cr(II), Mn(II), Co(II), Ni(II), Zn(II), 50-fold concentration of Fe(II) and 20-fold Cu(II), have no influence on the signals of 50 nM Hg(II) with deviation below 5%. | [150] |
| MWCNT | PANi-Bi NPs@GO * | Non covalent | Hg(II) (Cu(II)) | 2 ppt | 1.3 μ A/ppb (2 ppt–1000 ppm) | Differential pulse voltammetry | Screen printing | (commercial) Carbon ink | PET | Not provided | [151] |
| MWCNT | Au NPs | Non covalent | Hg(II) | 0.06 ppb | 0.59 μ A/ppb (0.1–1 ppb) 0.045 μ A/ppb (1–250 ppb) | Stripping voltammetry | Drop casting | Glassy carbon electrode | Glassy carbon | Not provided | [72] |
| MWCNT | * ENTZ | Non covalent | Hg(II) | 0.5 ppb | 29.3 mV/decade (1 ppb–20 ppm) | Potentiometry | Paste mixture of MWCNT, graphite powder, ENTZ ionophore and ionic liquid | Copper wire | Polypropylene syringe | The interfering ions (Ag^+ , Zn(II), Pb(II), Ni(II), Cd(II) and Cu(II)) do not have any effect on the response of proposed electrodes to Hg(II) | [79] |
| MWCNT | Thiol-functionalized chitosan | Non covalent | Hg(II) | 0.6 ppb | 1060 μ A/ppb (2–28 ppb) | Stripping voltammetry | Drop casting | Glassy carbon electrode | Glassy carbon | 100-fold Cd(II), 100-fold Pb(II), 50-fold Zn(II), 25-fold Cu(II), 10-fold Ag(II), 10-fold Fe(II) and 10-fold Mn(II) caused within \pm 5% changes of voltammetric signals for Hg(II). | [152] |
| MWCNT | Triazene (BEPT) | Non covalent | Hg(II) | 0.62 ppb | 29 mV/decade (0.8 ppb–440 ppm) | Potentiometry | Paste mixture of MWCNT, graphite powder, Triazene (BEPT) ionophore and paraffin oil | Copper wire | Polyethylene tube | The proposed electrode has a high performance to selective potentiometric assay of Hg(II) in aqueous samples containing some interfering ions (Cu(II), Ag(II), Cd(II), Co(II), Al(III), Pb(II), K^+). | [153] |

* ENTZ: 1-(2-ethoxyphenyl)-3-(3-nitrophenyl)triazene. PANI: polyaniline. GO: graphene oxide. NP: nanoparticles. 1 μ M Hg(II) = 200 ppb Hg(II).

3.2.5. Detection of As(III)

Table 7 presents the five stripping voltammetry sensors reported for the detection of As(III) in water using functionalized MWCNTs. No report was found on As(V) detection even though it is a relevant species for water quality monitoring. The maximum acceptable concentration of arsenic in drink water is 10 ppb.

COOH-functionalized CNTs [154] are sensitive to As(III) in the proper range of concentration (0.3 to 50 ppb), though the LOD is not provided. It suggests that pristine CNT (which naturally carry COOH groups on their sidewalls) are sensitive to As(III) as well, though the reference reports on interference with antimony.

Four references describe non-covalent functionalization, all but one [155] use metal nanoparticles (NP) as functional probes, as they have been reported to provide good performances for Arsenic detection in water [156]. More specifically, the best LOD is achieved with Au-NP with a value of 0.1 ppb [157]. The enhanced performances of gold NPs compared to others metal NPs for heavy metal detection are usually attributed to their high electrical conductivity, high surface area and catalytic activity. A comparable LOD (0.13 ppb) is achieved with a Leucine/Nafion functionalization [155]. The Leucine peptide is known for its capability to coordinate metal ions through hydrogen bonds occurring between its -NH(III) and -COOH groups.

Table 7. CNT-based sensors for detecting As(III) ions in water, sorted by detection limit.

| Type of CNT | Functional Probe | Functionalization | Analyte (Add. Analytes) | Detection Limit | Sensitivity (Linear Range) | Transduction Method | Deposition Method | Electrode Material Contact Configuration | Substrate | Interference Study | Ref. |
|-------------|------------------|-------------------|-------------------------|-----------------|---------------------------------|-----------------------|-------------------|--|---------------|--|-------|
| MWCNT | COOH | Covalent | As(III) | N.A | 0.24 μ A/ppb (0.3–50 ppb) | Stripping voltammetry | Dip coating | Au | Au electrode | Interference was significant when the Sb/As ratio is higher than 1. | [154] |
| MWCNT | Au-NP * | Non covalent | As(III) | 0.1 ppb | 26 μ A/ppb (75 ppt–5.3 ppm) | Stripping voltammetry | Drop casting | Glassy carbon | Glassy carbon | Not provided | [157] |
| MWCNT | Leucine/Nafion | Non covalent | As(III) | 0.13 ppb | 0.27 μ A/ppb (0.37–150 ppb) | Stripping voltammetry | Drop casting | Pt | Pt electrode | Zn(II) and Fe(II) could be tolerated up to at least 0.05 mM whereas commonly encountered matrix components such as Cd(II), Co(II), Mg(II), Ni(II) and Cu ⁺ did not show high percentage of interference. | [155] |
| MWCNT | Pt-Fe NP | Non covalent | As(III) | 0.75 ppb | 64 nA/ppb (0.75–22 ppb) | Stripping voltammetry | Drop casting | Glassy carbon | Glassy carbon | No interference from copper ion | [158] |
| MWCNT | Au-NPs | Non covalent | As(III) | 0.75 ppb | 2.6 Q/mL/ppb * (0.75–750 ppb) | Stripping voltammetry | Vacuum filtration | MWCNT membrane | PTFE membrane | The presence of copper at 10 μ M strongly affects the analytical response of As(III); The presence of Pb(II) caused a minor broadening of the peak of As(III) resulting in a slight reduction of the peak current; | [159] |

* NP: nanoparticles. Q/mL: charge at the peak current by mL of solution passing through (the conversion to A/ppb was not possible with provided information). 1 μ M As(III) = 75 ppb As(III).

3.2.6. Detection of Cu(II)

Table 8 shows the results of the nine reported papers for Cu(II) ions detection. All but one [129] (mentioned before regarding Zn(II), Cd(II) and Pb(II) detection) address functionalized CNTs-based sensors. All but one paper address stripping voltammetry with MWCNTs. The remaining one [113] achieves with a peptide-functionalized-SWCNT-FET structure a value of 3 ppt for the LOD over the range 0.6–600 ppt, which is the best one ever reported. The authors tested different combinations of peptides (of which there are in theory unlimited numbers) to identify the one with optimal sensitivity. The approach was also tested successfully for Ni(II) detection (see next section).

It should be noted that the MAC of Cu(II) in drink water is 1 ppm, so the other references targeting the ppb to ppm range with LOD in the ppb range are more relevant to drink water applications. Comparable LOD of 0.01 ppb, 0.02 ppb and 0.03 ppb are achieved respectively with Schiff base [160], pristine [129] and 2-amino-4-thiazoleacetic acid [161] functionalization and stripping voltammetry. While it suggests again a strong natural affinity of CNTs to Cu(II), both functional probes are found interesting as they carry amine groups which are well known to easily complex copper ions [162].

Table 8. CNT-based sensors for detecting Cu(II) ions in water, sorted by detection limit.

| Type of CNT | Functional Probe | Functionalization | Analyte (Add. Analytes) | Detection Limit | Sensitivity | Transduction Method | Deposition Method | Electrode Material Contact Configuration | Substrate | Interference Study | Ref. |
|-------------|--|--------------------|---------------------------------|-----------------|--------------------------------|---|---|--|-----------------------------|--|-------|
| SWCNT | PANI-GGHH * | Non covalent | Cu(II) | 3 ppt | N/A (3–29 ppt) | FET (liquid gate) | CVD | 300 nm Au | Si/SiO ₂ (120nm) | His ₆ shows higher chelation power for Ni(II) than to Cu(II). | [113] |
| MWCNT | C ₂₄ H ₃₀ N ₆ Schiff base | Non covalent | Cu(II) | 10 ppt | N/A (0.09–340 ppb) | Stripping voltammetry | Paste of MWCNT, Schiff base and mineral oil | Copper wire | Filter membrane | Not provided | [160] |
| MWCNT | Pristine | Non functionalized | Cu(II) (Cd(II), Zn(II), Pb(II)) | 17 ppt | 9.4 pA/ppb (32–220 ppb) | Stripping voltammetry | CNT thread aspirated into a glass capillary | Metal wire and silver conductive epoxy | Glass capillary | Simultaneous determination of Cd(II), Cu(II), Pb(II) and Zn(II) demonstrated The presence of Dissolved Oxygen changes the calibration law for Cd(II) | [129] |
| MWCNT | 2-amino-4-thiazoleacetic acid | Non covalent | Cu(II) | 30 ppt | 0.02 μA/ppb * (44 ppb–3.2 ppm) | Stripping voltammetry | Drop casting | Glassy carbon electrode | Glassy carbon | At a concentration ratio below 10, the presence of Zn(II), Mn(II), Ni(II), Co(II) has led to lower than 6% decreasing of DPSV currents of Cu(II) | [161] |
| MWCNT | PANI-Bi NPs@GO * | Non covalent | Cu(II) (Hg(II)) | 32 ppt | 0.23 uA/ppb (32 ppt–320 ppm) | Differential pulse voltammetry | Screen printing | (commercial) Carbon ink | PET | Not provided | [151] |
| MWCNT | N-doped carbon spheres | Non covalent | Cu(II) | 92 ppt | 0.28 μA/ppb (0.5–200 ppb) | Stripping voltammetry | Drop casting | Glassy carbon electrode | Glassy carbon | EDTA can seriously affect the stripping peak current of Cu(II) with a decrease of 79%. | [163] |
| MWCNT | Poly(1,2-diaminobenzene) | Non covalent | Cu(II) (Cd(II)) | 0.33 ppb | 0.11 μA/ppb (5–100 ppb) | Stripping voltammetry | Multipulse potentiostatic method | Glassy carbon electrode | Glassy carbon | Not provided | [69] |
| MWCNT | SSA/MoS ₂ * | Non covalent | Cu(II) | 3.6 ppb | 0.13 μA/ppb (6.4~700 ppb) | Stripping voltammetry | Drop casting | Glassy carbon electrode | Glassy carbon | 10-fold concentration of the metal ions (K ⁺ , Ca(II), Na ⁺ , Mg(II), Zn(II), Pb(II), Cd(II), Fe(III), Mn(II), Co(II), Cr(III), Cr ⁶⁺ , Ni(II) and Hg(II), has not any obvious effect on the Cu(II) peak current. | [164] |
| MWCNT | Cysteine | Covalent | Cu(II) (Pb(II)) | 15 ppb | 0.13 * μA/ppb (250–1500 ppb) | Differential pulse anodic stripping voltammetry | Drop casting | Glassy carbon electrode | Glassy carbon | 40-fold Cl ⁻ , 30-fold SO ₄ ²⁻ and four fold CO ₃ ²⁻ did not have any significant effect on the stripping peak current of Pb ²⁺ and Cu ²⁺ | [74] |

* SSA/MoS₂: 5-sulfosalicylic acid/MoS₂. PANI-GGHH: polyaniline functionalized with peptide chain glycine-glycine-histidine-histidine. PANI: polyaniline. GO: graphene oxide. NP: nanoparticles. 1 μM Cu(II) = 64 ppb Cu(II).

3.2.7. Detection of Other Metal Ions

As listed in Table 9, the detection of two additional metal ions, Ni(II) and Co(II), has been reported in the literature using SWCNT-FET and chemistor transduction, respectively.

The peptide-functionalized SWCNT-FET mentioned in the previous section for copper detection [113], was also applied to Ni(II) detection with a different peptide sequence. As for Cu(II), a remarkably low LOD was achieved (2.8 ppt) within the range 0.58 to 587 ppt. Such a low LOD is interesting for drink water monitoring as the MAC for Ni(II) is low (20 ppb).

Gou et al. [165] compared flexible polyazomethine -PAM- polymer and rigid (shape persistent macrocycle) functional probes on SWCNTs for chemiresistive Co(II) sensing. They indicate that the flexibility of the PAM allows for better performances as it rearranges over the SWCNT network when binding the metal ions, enabling strong electronic interaction with SWCNT. They report 0.04 ppt of LOD over an extremely large range (0.04 ppt~440 ppm), which is remarkable not only for chemistors (usually less sensitive than FET and electrochemical sensors) but also for electrochemical detection of heavy metals as discussed in the previous sections. It raises the question whether even better LOD could be achieved with alternative transduction modes.

3.2.8. Multiplexed Detection of Metal Ions

As reported above, studies on metal ions detection rely heavily on electrochemical transduction (32 papers out of 36 in total). Electrochemical detection, and more specifically stripping voltammetry, is particularly interesting for the simultaneous detection of different metals in water, as the current peaks for each metal appear at different voltage range, as can be shown in Figure 7 [129] (obtained with MWCNTs threads electrodes).

Naturally, among these 32 references reporting electrochemical transduction, 14 are reporting on multiplexed detection with stripping voltammetry while none of the four papers based on electrical transduction does. Investigated groups of metal ions are Cd(II)/Pb(II) (7), Cd(II)/Zn(II)/Pb(II) (3), Cd(II)/Zn(II)/Pb(II)/Cu(II) (1), Cd(II)/Cu(II) (1), Cu(II)/Pb(II) (1) and Hg(II)/Cu(II) (1). The simultaneous detection of lead and cadmium is particularly focused on (10 papers out of the 14), as these two heavy metals are commonly found together in soil and water supplies and are both severe environmental contaminants even at trace levels. Table 10 provides a comparison of the performances of the devices reported in these 14 papers as a function of the target species, with conversion from ppb to M (Molar concentration) unit to allow comparison between analytes. Overall, devices have slightly better detection limit to Pb(II) than to Cd(II) irrespective of the functionalization, except for the Bismuth-reduced graphene oxide functionalization reported in [146] with sensitivity to Pb(II) enhanced by a factor of 50 compared to Cd(II). By contrast, the limit of detection to Cd(II) is much lower than that to Zn(II) (by a factor of 6 to 30), except in [129] with non-functionalized MWCNT threads where it is 1.6 times higher. Finally, the limit of detection to Cu(II) is much higher than for other species by about one order of magnitude.

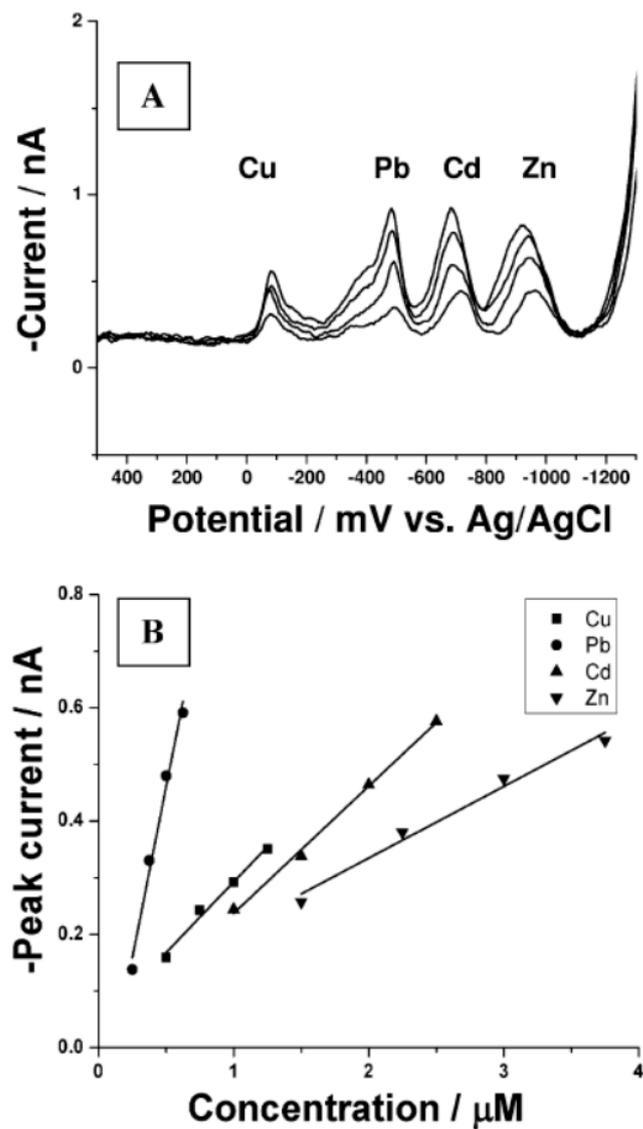


Figure 7. (A) Simultaneous detection of Cu(II), Pb(II), Cd(II) and Zn(II), ion concentrations 0.5, 0.25, 1.0, 1.5 μM for Cu(II), Pb(II), Cd(II), Zn(II), respectively; and 1.5, 2, 2.5 times of above concentrations for these metals ions. (B) Calibration Curve for Cu(II), Pb(II), Cd(II), Zn(II). Accumulation time: 120 s, deposit potential: -1.5 V. Reproduced from [129].

Table 9. CNT-based sensors for detecting Ni(II) and Co(II) ions in water.

| Type of CNT | Functional Probe | Functionalization | Analyte (Add. Analytes) | Detection Limit | Sensitivity | Transduction Method | Deposition Method | Electrode Material Contact Configuration | Substrate | Interference Study | Ref. |
|-------------|--------------------------------|-------------------|-------------------------|-----------------|--|---------------------|-------------------|--|------------------------------|---|-------|
| SWCNT | Polypyrrole-His _n * | Non covalent | Ni(II) | 2.8 ppt | 1.5 μ S/decade (5%/decade) (0.59 ppt–59 ppb) | FET (liquid gate) | CVD | 300 nm Au Pt wire (Counter electrode), Ag/AgCl (Reference electrode) | Si/SiO ₂ (120 nm) | His ₆ shows higher chelation power for Ni(II) than to Cu(II). | [113] |
| SWCNT | PAM * | Non covalent | Co(II) | 0.04 ppt | 0.014 */decade (0.04 ppt–440 ppm) | Chemistor | Spay-casting | Al tape Ag paint | Si/SiO ₂ | Selectivity to Co(II) was investigated in presence of Cu(II). The electrical response was higher with Co(II). | [165] |

* His: peptide histidine. PAM: polyazomethine.

Table 10. Comparison of the performances of sensors based on multiplexed detection as a function of the target species.

| Type of CNT | Functional Probe | Functionalization | Cd(II) LOD | Pb(II) LOD (LOD Pb/Cd) | Zn(II) LOD (LOD Zn/Cd) | Cu(II) LOD (LOD Cu/Cd) | Hg(II) LOD | Ref. |
|-------------|---|--------------------|---------------------|--------------------------|--------------------------|------------------------|------------|-------|
| MWCNT | Nafion/Bismuth | Non covalent | 0.04 ppb–0.4 nM | 0.025 ppb–0.12 nM 0.3 | | | | [73] |
| MWCNT | Bismuth | Non covalent | 0.04 ppb–0.4 nM | ~0.04 ppb–0.2 nM 0.5 | | | | [143] |
| MWCNT | PSS-Bi | Non covalent | 0.02 ppb–0.2 nM | 0.04 ppb–0.2 nM 1 | | | | [142] |
| MWCNT | rGO-Bi | Non covalent | 0.6 ppb–50 nM | 0.2 ppb–1 nM 0.02 | | | | [146] |
| MWCNT | PPy-Bi | Non covalent | 0.16 ppb–1.4 nM | 0.1 ppb–0.5 nM 0.4 | | | | [145] |
| MWCNT | Fe ₃ O ₄ -LSG-CS-Bi | Non covalent | 0.1 ppb–0.9 nM | 0.07 ppb–0.3 nM 0.3 | | | | [144] |
| MWCNT | Sb ₂ O ₃ | Non covalent | 17 ppb–0.15 μ M | 24 ppb–110 nM 0.7 | | | | [136] |
| MWCNT | Pristine | Non functionalized | 8.4 ppb–75 nM | 6.6 ppb–31 nM 0.4 | 28 ppb–0.43 μ M 6 | | | [131] |

Table 10. Cont.

| Type of CNT | Functional Probe | Functionalization | Cd(II) LOD | Pb(II) LOD (LOD Pb/Cd) | Zn(II) LOD (LOD Zn/Cd) | Cu(II) LOD (LOD Cu/Cd) | Hg(II) LOD | Ref. |
|-------------|--------------------------|--------------------|------------------|------------------------------|------------------------------|------------------------------|---------------|-------|
| MWCNT | Bismuth | Non covalent | 0.8 ppb–7 nM | 0.2 ppb–1 nM 0.14 | 11 ppb–0.17 μ M 24 | | | [138] |
| MWCNT | Bismuth | Non covalent | 0.7 ppb–6 nM | 1.3 ppb–6.2 nM 1 | 12 ppb–0.18 μ M 30 | | | [139] |
| MWCNT | Pristine | Non functionalized | 0.23 ppb–2 nM | 0.3 ppb–1 nM 0.5 | 0.08 ppb–1.2 nM 0.6 | 17 ppt–0.26 nM 0.13 | | [129] |
| MWCNT | Poly(1,2-diaminobenzene) | Non covalent | 0.25 ppb–0.22 nM | | | 0.33 ppb–5 nM 22 | | [69] |
| MWCNT | Cysteine | Covalent | | 1 ppb–4 nM | | 15 ppb–0.23 μ M | | [74] |
| MWCNT | PANi-Bi NPs@GO | Non covalent | | | | 32 ppt–0.5 nM | 2 ppt–0.01 nM | [151] |

PSS-Bi: Poly(sodium 4-styrenesulfonate)-Bismuth. rGO-Bi: Reduced graphene oxide-Bismuth. PPy-BiNPs: Polypyrrole-Bismuth. LSG-Cs-Bi: laser scribed graphene-chitosan-Bismuth. Sb₂O₃: antimony oxide. PANI: polyaniline. GO: graphene oxide. NP: nanoparticles. 1 μ M Cd(II) = 112 ppb Cd(II). 1 μ M Zn(II) = 65 ppb Zn(II). 1 μ M Pb(II) = 210 ppb Pb(II). 1 μ M Cu(II) = 64 ppb Cu(II). 1 μ M Hg(II) = 200 ppb Hg(II).

3.2.9. Interference Studies

Interferent studies are particularly significant regarding to toxic metal ions detection because in most water matrices, a wide range of ions are present at the same time, some of these at concentrations orders of magnitude larger than the target trace metals. For these reasons, most studies include interfering studies (30 out of 36 papers).

Most papers study interferences by other toxic ions typically present in the ppb range in water, such as Cu(II), Fe(II), Fe(III), Ni(II), Zn(II), Cr(III), As(III), Sb(III), Se(IV), Pb(II), Al(III), Fe(III), Ni(II), Co(II), F⁻ and SCN⁻. Among these, Cu(II) is the one reported most consistently as being an interferent for bismuth-functionalized CNT sensors due to the competition between bismuth ions and copper ions. It notably impacts performances for Zn(II), Cd(II), Pb(II) and As(III) detection [132,139,144–146,159].

Other papers rather focus on more ubiquitous ions usually present in the ppm range in water, such as Cl⁻, PO₄³⁻, SO₄²⁻, NO₃⁻, Na⁺, Ca(II), Mg(II), K⁺ or CO₃²⁻ [73,141]. For these ions, no interference to Pb(II), Cu(II) and Cd(II) detection was found.

These two types of interferents are mentioned in the literature on Hg(II) sensors. Out of eight papers, seven report on interferent studies. All these studies conclude toward a strong selectivity toward Hg(II) against the various interferent ions (see Table 10), irrespective of the types of functionalization (pristine, covalent, non-covalent) and of transduction (electrochemical, FET, chemistor). It suggests a strong selectivity of the CNTs themselves toward Hg(II).

Among other chemicals tested for interference, EDTA was found to particularly affect the detection of Cu(II) because EDTA forms complexes with every cation through its two amine and four carboxylate groups [163]. Benzene, xylene and some surfactants also interfere with metal ions detection by preventing the stripping of trace metals during stripping voltammetry measurements.

3.3. Nitrogen (Ammonia, Nitrite, Nitrate)

Ammonia (NH₃) is highly soluble in water and found under the form of dissolved gas or as the ammonium ion (NH₄⁺) depending on pH. Though it may be present in water as a result of normal biological degradations of proteins, it may also be brought by industrial water discharge. It is also sometimes used for drink water treatment (notably in the USA).

Nitrite ions (NO₂⁻) are widely used as fertilizing agents and food preservatives. They are in consequence among the pollutants most often identified in natural waters. They are highly toxic for human beings (fatal dose of nitrite ingestion is between 8.7 and 28.3 μM) [166]. Nitrate ions (NO₃⁻) are also widely found in groundwater and subsequently in drinking water. They primarily result from fertilizers, septic systems and manure storage or spreading operations. Although nitrite, nitrate and ammonia all have strong health and environmental impacts, only nitrite sensing has been reported with CNTs so far. Table 11 summarizes the reported performances. All papers rely on electrochemical transduction with non-covalently functionalized CNTs, and only one study reports the use of SWCNT.

The reported LODs vary from 0.016 μM to 25 μM (1 mM nitrite = 46 ppm), and the ranges of detection cover the scale from 0.1 μM to 10 mM. The MAC for nitrite in drinking water is around 1 ppm/20 μM, so nine papers out of 10 show acceptable limit of detection for nitrite monitoring in drink water with seven papers out of 10 reporting negligible interferences.

The best result is reported with a LOD of 0.016 μM with a functional probe based on a nanocomposite made of Co₃O₄ and rGO (reduced graphene oxide) [167]. With the same electrode and deposition process (drop casting on glassy carbon, using rGO only as the functional probe leads to a considerably higher LOD of 25 μM [76], underlying the role of the cobalt oxide functionalization in the sensitivity. Consistently, cobalt oxide on its own has been reported to be promising for nitrite sensing by its reduction process upon exposure to nitrite [168].

Table 11. CNT-based nitrogen sensors for water quality monitoring, sorted by detection limit.

| Type of CNT | Functional Probe | Functionalization | Analyte | DETECTION LIMIT | Sensitivity (Detection Range) | Transduction Method | Deposition Method | Electrode Material Contact Configuration | Substrate | Interference Study | Ref. |
|-------------|---|-------------------|---------|-----------------|---|---|--|--|---------------|--|-------|
| MWCNT | Co ₃ O ₄ ⁻ rGO * | Non covalent | Nitrite | 0.016 μM | 0.408 μA/μM/cm ² (0.1–8000 μM) | Voltammetry | Drop casting | Glassy carbon electrode | Glassy carbon | 100-fold of alcohol, Na ⁺ , K ⁺ , Cl ⁻ , NO ₃ ⁻ , N ₂ H ₄ , SO ₃ ²⁻ , SO ₄ ²⁻ , has no effect on sensor response. | [167] |
| MWCNT | PCMA * | Non covalent | Nitrite | 0.067 μM | -0.023 μA/μM (1–10 μM) -0.022 μA/μM (10–100 μM) -0.034 μA/μM (100–1000 μM) -0.026 μA/μM (1000–4000 μM) | Differential pulse voltammetry, Chronoamperometry | Drop cast of PCMA/MWCNT, then electrochemical crosslinking | Au | Au | Not provided | [169] |
| MWCNT | AuNPs/PEI*/MWCNT-COOH | Non covalent | Nitrite | 0.2 μM | -0.500 μA/μM* (1–2000 μM) -58 μA/mM (1–1400 μM) | Voltammetry | Drop casting | Au | Au | Na ⁺ , Mg(II), Ca(II), Zn(II), Fe(II), Cl ⁻ , I ⁻ and SO ₄ ²⁻ did not have significant interference in the detection of nitrite. | [170] |
| SWCNT | Pd | Non covalent | Nitrite | 0.25 μM | 420 μA mM ⁻¹ cm ⁻² (2–240 μM) 190 μA mM ⁻¹ cm ⁻² (280–1230 μM) | Differential pulse voltammetry | Vacuum filtration | SWCNT | PET | Negligible effect of K ⁺ , Na ⁺ , Cl ⁻ , PO ₄ ³⁻ , NH ₄ ⁺ , CH ₃ COO ⁻ and Zn(II) in concentration above 500 mM and concentrations of Mg(II), Ca(II), Cd(II), CO ₃ ²⁻ , NO ₃ ⁻ and SO ₄ ²⁻ above 200 mM | [171] |
| MWCNT | Ni ₇ S ₆ | Non covalent | Nitrite | 0.3 μM | 0.185 μA/μM (1–4200 μM) | Voltammetry | Drop casting | Glassy carbon electrode | Glassy carbon | Results comparable to high-performance liquid chromatography for lake water, tap water and pickle water | [172] |
| MWCNT | GO-MWCNT-PMA-Au | Non covalent | Nitrite | 0.67 μM | 0.484 μA/μM (2–10,000 μM) | Differential pulse voltammetry | Drop casting | Glassy carbon electrode | Glassy carbon | No obvious response was observed when injection of 0.4 Mm of Na ⁺ , Ca(II), NO ₃ ⁻ , CO ₃ ²⁻ , K ⁺ , Cl ⁻ , SO ₄ ²⁻ , IO ₃ ⁻ | [173] |
| MWCNT | Au/TiO ₂ | Non covalent | Nitrite | 3 μM | N/A (4–225 μM) | Differential pulse voltammetry | Pulsed electrodeposition | Glassy carbon electrode | Glassy carbon | The presence of arginine, serine, tyrosine, cysteine, glucose, alanine (each of 0.1 mM) causes less than 5% variation on sensor response. | [166] |
| MWCNT | Thionine | Non covalent | Nitrite | 4 μM | 0.002 μA/μM (6 μM–15,000 μM) | Voltammetry | Transfer via abrasion from filter paper to heated GC electrode | Glassy carbon electrode | Glassy carbon | Not provided | [174] |
| MWCNT | PANI * | Non covalent | Nitrite | 6.1 μM | 0.684 μA/μM/cm ² (N/A) | Voltammetry | Electrodeposition | Glassy carbon electrode | Glassy carbon | Not provided | [175] |
| MWCNT | rGO * | Non covalent | Nitrite | 25 μM | 0.01 μA/μM (75–6060 μM) | Differential pulse voltammetry | Drop casting | Glassy carbon electrode | Glassy carbon | 0.6 M Ca(II), Cu(II), K ⁺ , Na ⁺ , Zn(II), SO ₄ ²⁻ , L-cysteine, NO ₃ ⁻ and Cl ⁻ did not interfere with the pick signals of 0.15 mM HQ, 0.15 mM CC, 0.15 mM PC and 0.15 mM NO ₂ ⁻ . | [76] |

* In case of Nitrite ion (NO₂⁻), 1 mM = 46 ppm. PEI: polyethyleneimine. NP: nanoparticles. PCMA: poly(VMc-co-V Cz-coAA); VMc: 7-(4-vinylbenzyloxy)-4-methyl coumarin, VCz: 9-Vinylcarbazole, AA: Acrylic acid), GO: Graphene oxide; rGO: Reduced graphene oxide, GCE: Glassy carbon electrode, PANI: Polyaniline.

3.4. Water Hardness (Ca(II), Mg(II))

In general, the total hardness of water is defined as the sum of the concentrations of divalent calcium and magnesium, and of all the other alkaline earth ions in the water matrix. The concentration of calcium and magnesium ions is dominant to the other alkaline-earth metals, therefore water hardness is generally estimated from the concentration of these two ions [176]. Determination of water hardness is important as hard water can precipitate inside a water pipe and cause limescale. The sum of recommended Ca(II) and Mg(II) concentration in water ranges from 2 to 4 mM.

There have been relatively few studies on CNT sensors for Ca(II) and Mg(II) ions measurement, as reported in Table 12, all addressing functionalized CNT. To be noted, both sensors are tested to measure either Ca(II) or Mg(II) concentration in water, not the total water hardness. The best reported limit of detection is achieved with a chemFET approach, reaching down to 100 pM of Ca(II) (4 ppt). It is based on the functionalization of SWCNT by Fluo-4 AM (Fluorescent acetoxymethyl ester). It is a fluorescein derivative comprising amino carboxylate coordinating groups that has been widely used for calcium detection [177].

Table 12. CNT-based water hardness sensors in water.

| Type of CNT | Functional Probe | Functionalization | Analyte | Detection Limit | Sensitivity (Detection Range) | Transduction Method | Deposition Method | Electrode Material Contact Configuration | Substrate | Comments | Ref. |
|-------------|------------------|-------------------|-----------------|-----------------|------------------------------------|------------------------|-----------------------------------|---|--------------------------------------|---|-------|
| SWCNT | Fluo-4 AM * | Non-covalent | Ca(II) | 100 pM | 69 nA/decade (100 nM~1 mM) | CNT-FET | Dip coating | Ti (10 nm)/Au (30 nm) (liquid, floating gate) | Glass (borosilicate glass capillary) | FET at the end of a nanoneedle for intracell monitoring | [178] |
| MWCNT | PDMS * | Non-covalent | Ca(II) (Mg(II)) | 25 μ M | N/A (25 μ M~5 mM (Not linear)) | Capacitive measurement | Mold injection and thermal curing | MWCNT | PDMS | Measured at 2.4 kHz frequency | [179] |

* Fluo-4 AM: Fluorescent acetoxymethyl ester. PDMS: Polydimethylsiloxane. 1 ppm Ca(II) = 0.025 mM Ca(II).

3.5. Dissolved Oxygen (DO)

Dissolved oxygen (DO) refers to the amount of free oxygen present in water in gaseous form. It is measured in mg/L or in ppm. Algal biomass, dissolved organic matter, ammonia, volatile suspended solids and sediment oxygen demand can affect the variation of DO in water. Hence DO is widely used as indicator of the metabolism and pollution levels of waterbodies [180,181].

Several groups reported that molecular oxygen acts as dopant for CNTs and thus limits the selectivity and sensitivity of CNT-based sensors to other gases (in air) or chemicals (in water) [182,183]. In turn, this suggested the feasibility of CNT-based DO sensors. Table 13 shows the two reported DO sensors based on CNT reported so far. Both are based on cyclic voltammetry with non-covalently functionalized MWCNT coated on glassy carbon electrodes.

Regarding the first reported CNT-based dissolved oxygen sensor in 2004 [184], the functional probe is hemin. Hemin is an iron-containing porphyrin that can be found in red blood cells, and that efficiently binds dioxygen [185]. Hemin-functionalized MWCNTs show a better sensitivity to O₂ than non-functionalized ones in O₂-saturated phosphate buffer solution.

More recently, Tsai et al. (2013 [186]) used gold nanoparticles as functional probes, gold being selected as an effective catalyst for oxygen reduction. The electrodes showed a quasi-linear response to dissolved oxygen with a detection limit at 0.1 ppm (~3 μM). Such resolution is suitable to determine the spatial variation of DO concentration for oxygen profiling in water bodies [187].

3.6. Disinfectants (Hypochlorite, Hydrogen Peroxide, Chloramine, Peracetic Acid)

Free chlorine, hydrogen peroxide, peracetic acid, potassium permanganate and chloramine, are chemicals with an outstanding oxidation capacity. They are used either in the initial disinfection process of water or to keep the drinking water disinfected during distribution.

One of the most widely used drink water disinfectants is free chlorine. Its concentration in water should be in the range from 0.2 to 0.5 mg/L after disinfection (in which case it is called residual free chlorine). At lower concentrations, bacterial contamination may occur; at higher concentrations it is hazardous to human health.

Table 14 shows the reported CNT-based sensors for detecting disinfectants in water. The detection of hydrogen peroxide is reported in six articles, all but one by the means of electrochemical measurements with non-covalently functionalized CNT. The detection limit is about 3 ppm for the chemistor device (which is acceptable for the applicative range: conventional hydrogen peroxide sensors have a range from 0 to 2000 ppm and EPA (US) recommended levels in drink to range from 25 to 50 ppm). The use of electrochemical transduction with CNT functionalized by metallic materials lowers this threshold by several orders of magnitude, reaching down to 3.4 ppb with a 3D structure based on nitrogen doped Co-CNTs over graphene sheets [188]. An approach based on petal-like chromium hexacyanoferrate (Cr-hcf) crystallites yielded 17 ppb detection limit, this later material being specifically studied because of its electrocatalytic activity in the reduction of H₂O₂ [75]. As in the previous sections, the use of these two types of 3D structuration appeared to lead to large improvement (more than one order of magnitude) of performances compared to more traditional 2D architecture such as [189,190] (also metal based).

Regarding free chlorine (or hypochlorite detection for detection at pH higher than 7), the non-functionalized, aligned MWCNT-based chemistor device shows the lowest LOD below 5 ppb. The sensitivity in this reference is attributed to the oxidative properties of NaOCl leading to doping effect of the CNTs [191].

Table 13. CNT-based dissolved oxygen sensors in water.

| Type of CNT | Functional Probe | Functionalization | Analyte | Detection Limit | Sensitivity (Linear Range) | Transduction Method | Deposition Method | Electrode | Ref. |
|-------------|------------------|-------------------|----------------|-----------------|----------------------------|---------------------------------|--|-------------------------|-------|
| MWCNT | Hemin | Non-covalent | O ₂ | N/A | N/A (N/A) | Cyclic voltammetry, Amperometry | In-place CVD (densely-packed, vertically aligned CNTs) | Glassy carbon electrode | [184] |
| MWCNT | Au NP * | Non-covalent | O ₂ | 0.1 ppm | N/A (0~50 ppm) | Cyclic voltammetry | Not provided | Glassy carbon electrode | [186] |

* NP nanoparticles.

Table 14. CNT-based sensors for detecting disinfectants in water. References are sorted by type of analyte (hydrogen peroxide, free chlorine) then by limit of detection.

| Type of CNT | Functional Probe | Functionalization | Analyte | Detection Limit | Sensitivity (Detection Range) | Transduction | Deposition Method | Electrode Material Contact Configuration | Substrate | Interference | Ref. |
|-----------------------------|---|--------------------|--|-------------------------|---|--------------------------|--|--|-------------------------------|--|-------|
| MWCNT | PVC, DBE * | Non covalent | Hydrogen peroxide | N/A | Not Provided | Amperometry, voltammetry | Screen Printing | CNT electrodes | Alumina | Not provided | [192] |
| MWCNT | Nitrogen doped Co-CNTs over graphene sheets | Non covalent | Hydrogen peroxide | 100 nM 3.4 ppb | -0.85 μ A/ppm | Voltammetry, amperometry | Coating | Glassy carbon electrode | Glassy carbon electrode | No interference with uric acid, ascorbic acid and glucose | [188] |
| SWCNT | Cr-hcf * | Non covalent | Hydrogen peroxide | 0.5 μ M 17 ppb | 1 μ A/ppm (17 ppb-340 ppm) * | Amperometry, voltammetry | Drop casting | Glassy carbon electrode | Glassy carbon | No interference from ascorbic acid and uric acid | [75] |
| CNT (probably Multi-walled) | Fe-Ni | Non covalent | Hydrogen peroxide | 16 μ M 540 ppb | 1.2 μ A/ppm (34 ppm-510 ppm) | Voltammetry | Paste poured into electrode | Glassy carbon electrode | Glassy carbon electrode | Not provided | [189] |
| MWCNT | Chitosan/Cu/MWCNT-COOH | Non covalent | Hydrogen peroxide (pH) | <25 μ M <850 ppb | 0.97 nA/ppb (500 μ M-10 mM) | Amperometry | Potentiostatic polarization | Glassy carbon electrode | Chitosan-coated glassy carbon | No interference from ascorbic acid and uric acid | [190] |
| SWCNT | Phenyl capped aniline tetramer | Non covalent | Hydrogen peroxide | <3 ppm | 1%/ppm (3 ppm-8 ppm) Nonlinear <1%/100 ppm (48 ppm-1200 ppm) | Chemistor | Drop casting | Carbon ink | Glass | Not provided | [193] |
| MWCNT | Pristine | Non functionalized | Free chlorine in its hypochlorite ion form | <5 ppb | Logarithmic 39%/decade * (0.03-8 ppm) | Chemistor | Dielectrophoresis (aligned MWCNT) | Cr/Au | Glass | No information about selectivity, pH information not provided | [191] |
| MWCNT | Epoxy EpoTek H77A | Non covalent | Free chlorine under hypochlorous acid form (At pH 5.5) | 20 ppb | 0.15 μ A/ppb (0.02-4 ppm) | Voltammetry | Paste poured into tube and thermally cured | Epoxy/MWCNT composite | Not provided (tube) | Validated in real water matrices (tap water and swimming pool) | [194] |
| SWCNT | Phenyl capped aniline tetramer | Covalent | Free chlorine | <60 ppb | 92 nA/decade (0.06-60 ppm (linear up to 6 ppm)) | Chemistor | Drop casting | Au | Glass | Non selective to different oxidants list of oxidants not provided Regeneration possible | [53] |

* Cr hcf: Chromium hexacyanoferrate. PVC: Polyvinyl chloride. DBE: dibasic ester. 1 mM = 34 ppm H₂O₂.

3.7. Sulfur (Sulfide, Sulfite, Sulfate)

Sulfur can be found in aqueous environments in oxidized form as sulfite (SO_3^{2-}), sulfate (SO_4^{2-}), or in reduced form as sulfide (S^{2-}). The oxidized forms of sulfur play an important role within environmental systems [195]. In fact, they are detected in natural waters, waste waters and in boiler waters (those treated with sulfur for dissolved oxygen control). High concentrations of sulfite in boiler waters is harmful, since it decreases pH and subsequently, stimulates corrosion.

Table 15 compares the different CNT-based sulfur (sulfite and sulfide) sensors used for water quality monitoring. No sulfate sensor has been reported yet. All of these studies address electrochemical sensing with non-covalently functionalized CNTs.

Regarding sulfite detection, both reports [68,78], use a functional probe based on ferrocene. Zhou et al. (2008) used ferrocene-branched chitosan composites, while Hassan et al. (2011) used only ferrocene for GCE modification. Indeed, ferrocene and its derivatives have been reported as strong electrocatalysts for sulfite detection [196]. The LOD and sensitivity of the probe using ferrocene being directly in contact with MWCNTs are better by a factor of more than 20 than those of the probe using ferrocene-branched chitosan.

Regarding the detection of sulfide, all reports address electrochemical sensing with MWCNTs. Best LODs are in the range 0.2 to 0.3 μM (1 mM sulfide = 34 ppm). These LOD are too high compared to drink water quality requirements as sensitivity to sulfide in the ppt to sub-ppm range is required. The best limit of detection of 0.2 μM is reported with Hematoxylin [197], a compound reported to foster electrocatalytic oxidation of sulfide. Platinum [198] nanoparticles (also expected to oxidize sulfide) electrodeposited on vertically aligned CNT arrays perform also very well, comparably to non-functionalized CVD-grown MWCNTs [199].

Table 15. CNT-based sulfur sensors for water quality monitoring. References are sorted by analyte (sulfite and sulfide), then by limit of detection.

| Type of CNTs | Functional Probe | Functionalization | Analyte | Detection Limit | Sensitivity (Detection Range) | Transduction Method | Deposition Method | Electrode Material Contact Configuration | Substrate | Interference Study | Ref. |
|--------------|-----------------------------|--|---------|--|--|---|--|--|-----------------|---|-------|
| MWCNT | Ferrocene-branched chitosan | Non covalent | Sulfite | 2.8 μM | 0.013 $\mu\text{A}/\mu\text{M}$ (5 μM –1500 μM) | Amperometry | Drop casting | Glassy carbon electrode | Glassy carbon | 600-fold excess of Ca(II), Mg(II), Ba(II), PO_4^{3-} , NO_3^- , CO_3^{2-} and Cl^- did not interfere in the determination of sulfite. | [68] |
| MWCNT | Ferrocene | Non covalent (Physical immobilization) | Sulfite | 0.1 μM | 3.3 $\mu\text{A}/\mu\text{M}$ (0.4 μM –4 μM) 0.18 $\mu\text{A}/\mu\text{M}$ (4 μM –120 μM) | Differential Pulse Voltammetry | Paste mixture with graphite powder blended with paraffin oil | MWCNT paste, Copper wire | Glass tube | Not provided | [78] |
| MWCNT | Hematoxylin | Non covalent | Sulfide | 0.2 μM | 103 nA/ μM (0.5 μM –150 μM) | Amperometry | Paste mixture of MWCNT, mineral oil and graphite powder | Carbon paste | Teflon tube | No interference with Sn(II), Co(II), (II)Pb(II), (II)Zn(II), Cu(II), Ni(II), Mn(II), Fe(II) and Fe(III) | [197] |
| MWCNT | Platinum | Non covalent (plating) | Sulfide | 0.26 μM | 0.63 $\mu\text{A}/\mu\text{M}$ (0.26 μM –40 μM and 40 μM –100 μM) | Amperometry Differential pulse voltammetry | Thermal CVD (vertically aligned CNTs) | Stainless steel | Stainless steel | Not provided | [198] |
| MWCNT | Pristine | Not functionalized | Sulfide | 0.3 μM (CVD *), 12.5 μM (ARC *) | 0.12 $\mu\text{A}/\mu\text{M}$ (1.3 μM –113 μM) (CVD), 0.005 $\mu\text{A}/\mu\text{M}$ (12.5 μM –87.5 μM) (ARC) | Hydrodynamic voltammetry | Drop casting | Glassy carbon electrode | Glassy carbon | Not provided | [199] |
| MWCNT | Copper phenanthroline | Non covalent (Physical immobilization) | Sulfide | 1.2 μM | 34 nA/ μM (5 μM –400 μM) | Amperometry | Drop casting | Glassy carbon electrode | Glassy carbon | No interference with SO_3^{2-} , SO_4^{2-} , $\text{S}_2\text{O}_3^{2-}$, $\text{S}_4\text{O}_6^{2-}$, Cysteine. | [200] |

* ARC: Arc discharge method, CVD: Chemical vapor deposition method.

3.8. Other Contaminants

The detection of various additional analytes is also reported in the literature, as detailed in Table 16.

Zhao et al. reported that the threshold voltage of a CNT-FET with interdigitated electrodes using pristine, in-place grown SWCNT showed a response to glycerol in water [101]. This response is attributed to polar glycerol molecules adsorbing on the SWCNT sidewalls and acting as dopant for SWCNT. Glycerol is relevant to monitor in water as it is widely used in the food, beverage and e-cigarettes industry, and is also used in the formulation of numerous solvents. Thus, it ends up in the water cycle from human and industrial waste and may feature ecotoxicity [201].

Regarding to security applications (detection of explosive materials at extremely low concentration in water), Wei et al. demonstrated that a SWCNT-based chemistor functionalized with 1-pyrenemethylamine (PMA) could detect 2,4,6-trinitrotoluene in water, with a detection limit of 10 ppt and less than 1 min of response time [55]. The sensor showed high selectivity to several interfering molecules, for example, 2,6-dinitrotoluene (DNT) and 2,4-dinitrotoluene (2,4-DNT). The amino substituent in PMA was reported to interact selectively with TNT by forming negatively-charged complexes on the SWCNT sidewalls.

Regarding to the identification of dangerous toxins, Lee et al. reported that CNT-FET showed a response to botulinum neurotoxin (BoNT) in water with a detection limit up to 60 pM in case of peptide-modified CNT ((A)), and 52 fM in case of CNT modified with the anti-botulinum neurotoxin (B)) [109].

The detection of coliforms (notably *Escherichia coli*, but also other bacterial pathogens) is of major impact to drink water quality monitoring. However, standard assays take 24 h to 48h to determine presence or absence of coliforms, so reducing this detection time is of major interest. It relies on indirect detection of chemicals released by the bacteria, often upon addition of reagents. In [202], *p*-aminophenol is used as an indicator of the presence of coliform and detected through a glassy carbon electrode coated with Nafion/MWCNT. Coliform detection down to 10 cfu/mL was possible with 5 h response time.

Finally, references [76,203–205] address the topic of emerging contaminants with electrochemical sensors based on CNT modified carbon electrodes, through the angle of drugs and hormones [76,203] and of bisphenol A [204–206]. Emerging contaminants are compounds derived from manufactured chemicals and that despite being present only in $\mu\text{g/L}$ concentrations (or below) in water bodies are known to have strong impact on health and environment [207]. Among these, bisphenol A is notably acknowledged as endocrine disruptor and as toxic to reproduction. It is worth mentioning that regarding drug and health-care related chemicals, there are more references available beyond the field of drink water monitoring which are not included here, as detailed recently in [208].

Table 16. Reported chemical CNT sensors for water quality monitoring with different probes and analytes.

| Type of CNT | Functional Probe | Functionalization | Analyte | Detection Limit | Sensitivity (Detection Range) | Transduction Method | Deposition Method | Electrode Material Contact Configuration | Substrate | Interference Study | Ref. |
|-------------|----------------------------|--------------------|---|---|---|--|--|--|--------------------------------------|---|-------|
| SWCNT | Pristine | Not functionalized | Glycerol | N/A | ~10 Ω/Glycerol by weight % in water (10~50%) | CNT-FET | Dielectrophoresis | Cr/Au | Si/SiO ₂ | Not provided | [101] |
| SWCNT | 1-phyrenemethylamine | Non-covalent | Trinitrotoluene | ~ppt | N/A (>0.01 ppb) | Chemistor with interdigitated electrodes (IDEs) | Dip coating | Cr/Au | Si/SiO ₂ | Relatively selective to 2,6-DNT *, 2,4-DNT, 1,3-DNB *, 1-NB *, Response time~30 s | [55] |
| SWCNT | Peptides, anti-BoNT/E-Lc * | Non-covalent | BoNT* | 60 pM (Peptide probe), 52 fM (Anti-BoNT/E-Lc probe) | 27.95 nS/nM (Peptide), 313 nS/pM (Anti-BoNT) | CNT-FET | CVD (vertically aligned SWCNTs) | Au foils Bottom gate | 120 nm SiO ₂ on PDMS film | Not provided | [109] |
| MWCNT | Nafion | Non covalent | p-aminophenol (Coliforms) | 10 cfu/mL | 10 to 10 ⁴ cfu/mL | Cyclic voltammetry, amperometry | Drop casting | Glassy carbon electrode | Glassy carbon | Not provided | [202] |
| MWCNT | rGO* | Non covalent | Hydroquinone Catechol p-cresol (nitrite) | 2.6 μM 1.8 μM 1.6 μM | 0.19 μA/μM (8~391 μM) 0.07 μA/μM (5.5~540 μM) 0.04 μA/μM (5~430 μM) | Differential pulse voltammetry | Drop casting | Glassy carbon electrode | Glassy carbon | 0.6 M Ca(II), Cu(II), K ⁺ , Na ⁺ , Zn(II),SO ₄ ²⁻ , l-cysteine, NO ₃ ⁻ and Cl ⁻ did not interfere with the pic signals of 0.15 mM HQ, 0.15 mM CC, 0.15 mM PC | [76] |
| MWCNT | Fe-Co doped TNTs | Non-covalent | Sulpiride | 87 nM | 58.8 mV/decade (100 nM~10 mM) | Potentiometry | Paste mixture of graphite powder, MWCNT, Fe-CO-TNT, βCD ionophore, NaTPB anionic additive, DBP plasticizer | Carbon paste electrode | Syringe | No interference observed with K ⁺ , Na ⁺ , Ca(II), Mg(II), Cd(II), Co(II), Mn(II), Fe(II) | [203] |
| SWCNT | βCD * | Covalent | Bisphenol A | 1.0 nM | 1.3 mA/mM 11 nM~19 μM | Cyclic voltammetry | Drop casting | Glassy carbon electrode | Glassy carbon | No interference study, but tested on real plastic samples | [206] |
| MWCNT | βCD | Covalent | Bisphenol A | 14 nM | 7.2 μA/μM (125 nM~2 μM) 2.2 μA/μM (2 μM~30 μM) | Linear sweep voltammetry | Drop casting | Screen printed carbon electrode | Not provided | Selective to APAP *, BPA *, BPS | [204] |
| MWCNT | ZIF-67 * | Covalent | TBBPA * | 4.2 nM | 21.08 μA/μM (0.01~1.5 μM) | Differential pulse voltammetry, cyclic voltammetry | Paste mixture of paraffin oil, AB * and CNTs | Carbon paste electrode | Syringe | TBBME *, TBBDE *, BPAF *, BPA *, TCBPA *, TBBPS * did not show remarkable interference. | [205] |

* DNT: Dinitrotoluene. DNB: Dinitrobenzene. NB: Nitrobenzene. BoNT: Botulinum neurotoxin. rGO: reduced graphene oxide. TNT: titanate nanotube. E-Lc: E light chain. ZIF-67: Zeolitic imidazole framework-67. βCD: β-cyclodextrin. TBBPA: Tetrabromobisphenol A. AB: acetylene black. DBP: dibutyl phthalate. NaTPB: sodium tetraphenylborate. BPA: bisphenol A. TBBME: tetrabromobisphenol A-bis(dibromopropyl ether). TCBPA: tetrachlorobisphenol A. BPAF: hexa-fluorobisphenol A (BPAF, 98%), TBBPS: 4,4-sulphonyl-bis-(2,6-dibromophenol). TBBDE: tetrabromobisphenol A diallyl ether. BPS: bisphenol S. APAP: acetaminophen.

4. Discussion

4.1. Summary of Best Performances

The previous paragraphs show the various performances of the CNT sensors for all the reported analytes. To get an overview on the best device strategy to achieve the best performances, Table 17 summarizes the references for each type of functionalization and transduction for the 15 analytes that are addressed by more than one reference: pH, Pb(II), Cd(II), Zn(II), Hg(II), As(III), Cu(II), nitrite, calcium(II), dissolved oxygen, hydrogen peroxide, free chlorine, sulfite and sulfide.

4.2. Discussion on Sensor Design Choices

4.2.1. Choice of Transduction Mode

There are five analytes for which different transduction modes may be compared: pH, Hg(II), Cu(II), Ca(II) and H₂O₂:

- For pH, FET and impedance spectroscopy reach the same performance and are only slightly better than chemistor.
- For Cu(II), the LOD achieved with FET is three times better than voltammetry.
- For Hg(II), the LOD achieved with voltammetry is three orders of magnitude better than that obtained with FET, the latter being two orders of magnitude better than with chemistor.
- For H₂O₂, the LOD achieved with voltammetry is three orders of magnitude better than with a chemistor.
- For Ca(II), the LOD achieved with FET is five orders of magnitude better than capacitive measurements (which can be seen as a derivative of impedance spectroscopy).

While electrochemical measurements have been more widely used than FET-based approaches (probably due to easier manufacturing), the latter reach comparable or even widely improved performances for three out of four analytes. Testing FET architectures on a wider range of analytes would thus be valuable, as FETs are expected to be easier to operate than electrochemical sensors in field conditions.

Regarding chemistors, they feature larger limits of detection than the two other types, but the comparison is only possible on three analytes (out of 15). Moreover, for two out of three of these analytes (pH and H₂O₂), the detection limits are still acceptable for the monitoring of drink water. Finally, for several analytes (Co(II) and trinitrotoluene), ppt level detection limits are possible with chemistors. Considering that chemistors are easier to fabricate than FET and to operate than electrochemical sensors, their extensive testing against other types of analytes would be useful as well.

4.2.2. Functionalized versus Non-Functionalized CNT

Comparison of limits of detection between functionalized and non-functionalized CNTs (including COOH-CNTs) is possible for nine analytes. For Pb(II), Cd(II), Hg(II), As(III) and Cu(II), the use of functionalization improves significantly the limit of detection. Non-functionalized or COOH-functionalized CNT provide best performances for pH, free chlorine and Zn(II).

This suggests to systematically compare the performances of non-functionalized and functionalized CNTs (with the same device architecture), as they may be very sensitive without functionalization.

Table 17. Summary of best performances for all analytes addressed by more than one reference. When several transduction types or functionalization strategies are available for a given analyte, the table includes the best performing reference for each type.

| Analyte (Add. Analytes) | Type of CNT | Functional Probe | Functionalization | Detection Limit | Sensitivity (Detection Range) | Transduction Method | Deposition Method | Electrode Material Contact Configuration | Substrate | Interference Study | Ref. |
|-------------------------|-------------|--|--------------------|-----------------|--|----------------------------------|----------------------------------|---|--|---|-------|
| pH | MWCNT | Pristine | Non functionalized | N.P. | 63 Ω /pH 18%/pH pH 5–9 | Chemistor | Sucked by vacuum force | MWCNT | Filter paper | Not provided | [123] |
| | SWCNT | Pristine | Non functionalized | 1 mM | 7600 mV/pH 23%/pH (Dual-gate mode) pH 3–10 | FET, potentiometry (double gate) | Spin coating | 100 nm Ti contacts for source, drain and top gate | p-Si (substrate acting as bottom gate) | Not provided | [54] |
| | SWCNT | Poly(1-aminoanthracene) | Non covalent | 1 μ M | FET 19 μ S/pH 14%/pH | FET, potentiometry (liquid gate) | Dielectrophoresis (aligned CNTs) | Au contacts, Pt wire (Auxillary), Ag/AgCl electrode (Reference) | Si/SiO ₂ (300nm) | Multiplexed detection of Ca(II) and Na ⁺ | [60] |
| | MWCNT | COOH | Covalent | N.P. | 17 Ω /pH 23%/pH (Au) pH 4–9 | Impedance spectroscopy | Dip coating | Au and Al interdigitated electrodes | Kapton [®] | Not provided | [59] |
| Pb(II) | MWCNT | Pristine | Non functionalized | 0.3 ppb | 2.2 nA/ppb (210–830 ppb) | Stripping voltammetry | CNT thread | Metal wire and silver conductive epoxy | Glass capillary | Simultaneous determination of Cd(II), Cu(II), Pb(II) and Zn(II) demonstrated. The presence of Dissolved Oxygen changes the calibration law for Cd(II) | [129] |
| | MWCNT | Ionic liquid—dithizone based bucky-gel | Covalent | 0.02 ppt | 0.024 μ A/ppb (0.1ppt–210 ppb) | Stripping voltammetry | Drop-casting | Glassy carbon electrode | Glassy carbon | No interference of Cd(II) and Cu(II) ions with the detection of Pb(II) ion. | [132] |
| | MWCNT | Nafion/Bismuth | Non covalent | 25 ppt | 0.22 μ A/ppb (0.05 to 5 ppb) 0.27 μ A/ppb (5–100 ppb) | Stripping voltammetry | Drop casting | Glassy carbon electrode | Glassy carbon | 500-fold of SCN ⁻ , Cl ⁻ , F ⁻ , PO ₄ ³⁻ , SO ₄ ²⁻ , NO ₃ ⁻ and various cations such as Na ⁺ , Ca(II), Mg(II), Al(III), K ⁺ , Zn(II), Co(II) and Ni(II) had no influences on the signals of Pb(II) and Cd(II). | [73] |
| | MWCNT | PSS-Bi | Non covalent | 0.04 ppb | 0.079 μ A/ppb (0.5–90 ppb) | Stripping voltammetry | Drop casting | Glassy carbon electrode | Glassy carbon | 20-fold amounts of Zn(II), 5-fold amounts of Sn(II) and 1-fold amounts of Cu(II) have influence on the determination of Cd(II) and Pb(II) with deviation of 10%. | [142] |
| Cd(II) | MWCNT | Pristine | Non functionalized | 0.23 ppb | 3.9 nA/ppb (170–500 ppb) | Stripping voltammetry | CNT thread | Metal wire and silver conductive epoxy | Glass capillary | Simultaneous determination of Cd(II), Cu(II), Pb(II) and Zn(II) demonstrated. The presence of Dissolved Oxygen changes the calibration law for Cd(II) | [129] |
| | MWCNT | PSS-Bi | Non covalent | 0.02 ppb | 0.23 μ A/ppb (0.5–50 ppb) | Stripping voltammetry | Drop casting | Glassy carbon electrode | Glassy carbon | 20-fold amounts of Zn(II), 5-fold amounts of Sn(II) and 1-fold amounts of Cu(II) have influence on the determination of Cd(II) and Pb(II) with deviation of 10%. | [142] |

Table 17. Cont.

| Analyte (Add. Analytes) | Type of CNT | Functional Probe | Functionalization | Detection Limit | Sensitivity (Detection Range) | Transduction Method | Deposition Method | Electrode Material Contact Configuration | Substrate | Interference Study | Ref. |
|-------------------------|-------------|--|--------------------|-----------------|---|--------------------------------|---|--|-----------------------------|--|-------|
| Zn(II) | MWCNT | Pristine | Non functionalized | 0.08 ppb | 3.4 pA/ppb (200–590 ppb) | Stripping voltammetry | CNT thread | Metal wire and silver conductive epoxy | Glass capillary | Simultaneous determination of Cd(II), Cu(II), Pb(II) and Zn(II) demonstrated. The presence of Dissolved Oxygen changes the calibration law for Cd(II) | [129] |
| | MWCNT | Bismuth | Non covalent | 11 ppb | 0.18 μ A/ppb (12–18 ppb) 0.24 μ A/ppb (20–100 ppb) | Stripping voltammetry | Screen printing | Screen printed MWCNT based electrode | Ceramic substrates | N.P. | [138] |
| Hg(II) | SWCNT | Pristine | Non functionalized | 0.6 ppm | 12 mV/ppm (1–30 ppm) | Chemistor | CVD | SWCNT | Glass | 1000 fold excess of Fe(II), Fe(III), Ni(II), Cu(II), Zn(II), Cr(III) and 500 folds of As(III), Sb(III), Se(IV) and Pb(II) had no interfering effect in the analysis of mercury solution. | [147] |
| | SWCNT | Pristine | Non functionalized | 2 ppb | 0.22/decade 0.2 ppb–201 ppm | FET (Liquid gate) | Dip coating with selective CNT placement | Pd/Au (10/30 nm) | Glass | Good selectivity towards interferent ions | [110] |
| | SWCNT | Thiophenol | Covalent | 0.6 ppb | 0.14 μ A/ppb (1–18 ppb) | Stripping voltammetry | Dip coating | Au | Au | The presence of 100-fold concentration of Cr(II), Mn(II), Co(II), Ni(II), Zn(II), 50-fold concentration of Fe(II), and 20-fold Cu(II), have no influence on the signals of 50 nM Hg(II) with deviation below 5%. | [150] |
| As(III) | MWCNT | PANI-Bi NPs@GO | Non covalent | 2 ppt | 1.3 μ A/ppb (2 ppt–1000 ppm) | Differential pulse voltammetry | Screen printing | (commercial) Carbon ink | PET | Not provided | [151] |
| | MWCNT | COOH | Covalent | N.A | 0.24 μ A/ppb (0.3–50 ppb) | Stripping voltammetry | Dip coating | Au | Au electrode | Interference was significant when the Sb/As ratio is higher than 1. | [154] |
| | MWCNT | Au-NP | Non covalent | 0.1 ppb | 26 μ A/ppb (75 ppt–5.3 ppm) | Stripping voltammetry | Drop casting | Glassy carbon | Glassy carbon | Not provided | [157] |
| Cu(II) | SWCNT | PANI-GGHH | Non covalent | 3 ppt | N/A (3–29 ppt) | FET (liquid gate) | CVD | 300 nm Au | Si/SiO ₂ (120nm) | His ₆ shows higher chelation power for Ni(II) than to Cu(II). | [113] |
| | MWCNT | C ₂₄ H ₃₀ N ₆ Schiff base | Non covalent | 10 ppt | N/A (0.09–340 ppb) | Stripping voltammetry | Paste of MWCNT, Schiff base and mineral oil | Copper wire | Filter membrane | Not provided | [160] |
| | MWCNT | Pristine | Non functionalized | 17 ppt | 9.4 pA/ppb (32–220 ppb) | Stripping voltammetry | CNT thread aspirated into a glass capillary | Metal wire and silver conductive epoxy | Glass capillary | Simultaneous determination of Cd(II), Cu(II), Pb(II) and Zn(II) demonstrated. The presence of Dissolved Oxygen changes the calibration law for Cd(II) | [129] |
| Nitrite | MWCNT | Co ₃ O ₄ -rGO | Non covalent | 0.016 μ M | 0.408 μ A/ μ M/cm ² (0.1–8000 μ M) | Voltammetry | Drop casting | Glassy carbon electrode | Glassy carbon | 100-fold of alcohol, Na ⁺ , K ⁺ , Cl ⁻ , NO ₃ ⁻ , N ₂ H ₄ , SO ₃ ²⁻ , SO ₄ ²⁻ , has no effect on sensor response. | [167] |

Table 17. Cont.

| Analyte (Add. Analytes) | Type of CNT | Functional Probe | Functionalization | Detection Limit | Sensitivity (Detection Range) | Transduction Method | Deposition Method | Electrode Material Contact Configuration | Substrate | Interference Study | Ref. |
|-------------------------|-------------|---|--|--|--|--------------------------------|--|---|--------------------------------------|---|-------|
| Ca(II) | SWCNT | Fluo-4 AM | Non-covalent | 100 pM | 69 nA/decade (100 nM–1 mM) | FET | Dip coating | Ti (10 nm)/Au (30 nm) (liquid, floating gate) | Glass (borosilicate glass capillary) | FET at the end of a nanoneedle for intracell monitoring | [178] |
| | MWCNT | PDMS | Non-covalent | 25 μ M | N/A (25 μ M–5 mM (Not linear)) | Capacitive measurement | Mold injection and thermal curing | MWCNT | PDMS | Measured at 2.4 kHz frequency | [179] |
| O ₂ | MWCNT | Au NP | Non-covalent | 0.1 ppm | N/A (0–50 ppm) | Cyclic voltammetry | Not provided | Glassy carbon electrode | Glassy carbon | Not provided | [186] |
| Hydrogen peroxide | MWCNT | nitrogen doped Co-CNTs over graphene sheets | Non covalent | 100nM 3.4 ppb | –0.85 μ A/ppm | Voltammetry, amperometry | Coating | Glassy carbon electrode | Glassy carbon electrode | No interference with uric acid, ascorbic acid and glucose | [188] |
| | SWCNT | Phenyl capped aniline tetramer | Non covalent | <3 ppm | 1%/ppm (3 ppm–8 ppm) Nonlinear <1%/100 ppm (48 ppm–1200 ppm) | Chemistor | Drop casting | Carbon ink | Glass | Not provided | [193] |
| Free chlorine | MWCNT | Pristine | Non functionalized | <5 ppb (ClO [−]) | Logarithmic 39% /decade * (0.03–8 ppm) | Chemistor | Dielectrophoresis (aligned MWCNT) | Cr/Au | Glass | No information about selectivity, pH information not provided | [191] |
| | MWCNT | Epoxy EpoTek H77A | Non covalent | 20 ppb (HClO) | 0.15 μ A/ppb (0.02–4 ppm) | Voltammetry | Paste poured into tube and thermally cured | Epoxy/MWCNT composite | Not provided (tube) | Validated in real water matrices (tap water and swimming pool) | [194] |
| | SWCNT | Phenyl capped aniline tetramer | Covalent | <60 ppb | 92 nA/decade (0.06–60 ppm (linear up to 6 ppm)) | Chemistor | Drop casting | Au | Glass | Non selective to different oxidants—list of oxidants not provided Regeneration possible | [53] |
| Sulfite | MWCNT | Ferrocene | Non covalent (Physical immobilization) | 0.1 μ M | 3.3 μ A/ μ M (0.4 μ M–4 μ M) 0.18 μ A/ μ M (4 μ M–120 μ M) | Differential Pulse Voltammetry | Paste mixture with graphite powder blended with paraffin oil | MWCNT paste, Copper wire | Glass tube | Not provided | [78] |
| Sulfide | MWCNT | Hematoxylin | Non covalent | 0.2 μ M | 103 nA/ μ M(0.5 μ M–150 μ M) | Amperometry | Paste mixture of MWCNT, mineral oil and graphite powder | Carbon paste | Teflon tube | No interference with Sn(II), Co(II), Pb(II), Zn(II), Cu(II), Ni(II), Mn(II), Fe(II) and Fe(III) | [197] |
| | MWCNT | Pristine | Not functionalized | 0.3 μ M (CVD), 12.5 μ M (ARC) | 0.12 μ A/ μ M (1.3 μ M–113 μ M) (CVD), 0.005 μ A/ μ M (12.5 μ M–87.5 μ M) (ARC) | Hydrodynamic voltammetry | Drop casting | Glassy carbon electrode | Glassy carbon | Not provided | [199] |
| Bisphenol A | SWCNT | β CD | Covalent | 1.0 nM | 1.3 mA/mM 11 nM–19 μ M | Cyclic voltammetry | Drop casting | Glassy carbon electrode | Glassy carbon | No interference study, but tested on real plastic samples | [206] |

Moreover, while the literature often claims that pristine or COOH-CNTs do not have selectivity, one observes here that non-functionalized CNT sensors with excellent limit of detections may operate free from interferents as well [110,129,147].

One may wonder whether the overall remarkably good performances of non-functionalized CNTs could be explained by an “effective” functionalization during the fabrication process. To clarify, additional molecules may remain (intentionally or not) on the CNTs sidewalls during device fabrication. Similarly, CNT synthesized in place by CVD may still carry leftover catalysts particles. The role of these by-products of fabrication is not addressed in the papers. A systematic study of the role of solvents and catalysts in the sensitivity to analytes in water could be valuable.

4.2.3. Covalent versus Non-Covalent Functionalization

For pH, free chlorine, Cu(II), Hg(II) and Pb(II), covalent and non-covalent functionalization strategies are both reported. Except for Pb(II), non-covalent functionalization provides better performance than covalent functionalization. However, this conclusion should be tampered by the fact that it is never the same active compound being tested by both covalent or non-covalent functionalization. For instance, in the case of Pb(II) and Cd(II), bismuth is tested as an active compound of a lot of different functional probes, but all non-covalently functionalized. It would be very interesting to compare these results to a covalent functionalization strategy for bismuth or a bismuth derivative.

4.2.4. On the Diversity of Functional Probes

The functional probes inventoried in this review cover a wide range of size scale and feature different levels of complexity. The literature includes primarily a large number of “single-component” functional probes (e.g., that can be described by a single chemical formula): mono or bi-atomic dopants; small or macromolecules; nanoparticles; polymers.

These materials can be coupled together, forming “composite probes”. Two-component strategies are fairly standardized now: a primary functional probe such as a polymer or a macromolecule is itself functionalized by a secondary probe (for instance PSS-Bismuth in [142]). Three (or more)-component strategies are also reported. For instance, in [151] PANI is functionalized with bismuth, and the resulting two-component functional probe is used to functionalize graphene oxide sheets.

Multi-component functional probes are thought to enhance the 3D structuration of the CNT layer, hence its specific surface area, and adsorption capability and thus its sensitivity. It is worth mentioning that these three-component structures often include flagship bidimensional materials such as graphene oxide and graphene.

These observations are confirmed in Table 17: among the 12 analytes where the best performance is achieved through functionalization, half of these are achieved through a multi-component strategy (four papers on two-component probes, two papers on three-component probes). Moreover, three papers out of 12 include graphene or graphene oxide and three papers out of 12 include a polymer (PANI or PSS) functionalized by a secondary probe.

4.2.5. Type of CNT and CNT Alignment

The literature does not allow to compare between SWCNT against MWCNT as the choices between either is mostly guided by the transduction modes: MWCNTs are preferred for electrochemical and resistive sensing—as the CNT layer needs to be conducting; while the use of SWCNTs is mandatory with FETs. It would be very valuable to compare for the same analyte chemistors or electrodes with both types of MWCNTs (which is possible for electrochemical transduction and chemistors).

Similarly, there also appears to be no obviously optimal fabrication process. However, while definite comparison between references is not possible, it appears that alignment of CNT can provide outstanding performance, as is reported in [191] through dielectrophoresis or in [129] through threading.

4.3. Challenges and Perspectives

4.3.1. Optimal Sensing and Sensing Mechanisms—The Role of Modelling

With only 90 references over 20 years covering eight categories of analytes and dozens of different functional probes, the literature remains too limited to derive unequivocally the best functionalization, transduction and design strategies for a given target analyte, as well as to propose mechanisms of sensing that are valid across the whole literature (for instance regarding the impact of sensor footprint and choice of contact materials).

To support research on that front, the larger availability of fine-grained room temperature operation models for functionalized CNT networks would be highly valuable. As of now, the multi-scale nature of these structures and the complexity of their room-temperature electrical and chemical behavior makes this a challenging proposition. The model for ohmic CNT networks proposed by Benda et al. [118] is a first step in this direction, but covers neither chemical effects nor field effects. Regarding chemical effects, studying the sensitivity in water of a functional probe toward target analyte is possible [209], but reliably including carbon nanotubes in the modelled system and further predicting electronic transport remains an unanswered challenge.

4.3.2. Covering the Extreme Diversity of Analytes

This review details results on more than 25 target analytes. Despite this relatively large extent, and 20+ years of research, there are analytes of major impact that are not discussed by the literature, such as nitrate or the different forms of phosphate or iron in water. Furthermore, emerging contaminants such as drugs and hormones—which actually include dozens of relevant chemicals—will require monitoring tools in the near future. To cover more quickly the extreme diversity of analytes of interest, computer-aided design of functional probes should be promoted, as is reported in [209],

4.3.3. Managing the Complexity of the Water Matrices through e-Tongue Strategy

By nature, water monitoring requires sensors that are robust to very complex water matrices, e.g., water that contains a large variety of interfering ions. This explains why a large number of the study reported here includes interferent studies. While some level of selectivity is often reported, it is rarely perfect with CNT-based sensors. Moreover, for obvious practical reasons, reports can rarely cover all the species that should be tested to account for the complexity real water matrices.

In the field of gaz sensing with CNT, which features the same issues, the solution that is widely used is the electronic nose [210]: an array of CNT sensors is fabricated, each CNT sensor carrying a specific functional probe or having different contact metals, in order to make it selective to a different specie. The CNT sensor array thus aims at multiparameter sensing. As selectivity is usually not achieved perfectly, machine-learning algorithms are applied to determine the footprint of the target gaz in the gaz matrix out of the multiple sensors data.

In the field of sensing in liquids, the principle is called electronic tongue. Regularly used in CNT biosensors [211] and CNT sensors for food applications [212], it has so far remained mostly unexplored in the field of in-situ water monitoring. While recent work has shown the possibility to actually fabricate sensor arrays compatible with water monitoring application [213,214], a large amount of work remains to be done to prove the reliability of multiparameter measurements in complex water matrices.

4.3.4. Toward Real Applications: The Need for Ageing Studies

Besides the challenge of reliability measurements, water quality monitoring requires survival of the sensors in water over relatively long period of time, preferably without calibration. A chemical sensor with one year of operation in the field without recalibration would be a game changer. By contrast, ageing studies included in references so far discuss only short term cyclability (5 to 10 cycles of measurements) and short term sensor survival (usually a couple of days); the change in sensitivity over time is hardly studied. There is a

major discrepancy between what is currently done in laboratory in terms of ageing and what is required by end users for true applications. Major progress in marketability of CNT sensors for water applications could be expected if standardized protocols for ageing studies could be developed with reasonable scope and duration and then generalized among research teams.

5. Conclusions

This review identifies and compares 90 CNT-based water quality monitoring sensors reported from 2000 to April 2021. A set of 126 additional references provide context and supporting information. After reviewing the challenges of on-line drink water quality monitoring and presenting the highlights of CNT-based electronic devices and electrical transduction for chemical sensing, a quantitative comparison of the performances of reported sensors based on limit of detection, sensitivity and detection range was proposed. The target analytes are pH, micronutrients and heavy metals, nitrogen forms, sulfur forms, disinfectants and dissolved oxygen, as well as miscellaneous materials relevant to drink water quality.

Overall, there are so many parameters featuring in the design and operation of CNT based water quality sensors that a systematic comparison across all of these references was not possible with the current extent of the literature. However, some key conclusions can be drawn regarding the best transduction mode and functional probe considering a specific analyte.

Across all analytes, while electrochemical sensing with MWCNTs is the most frequently reported approach and allows to reach remarkable limit of detection (down to the ppt level), FET and chemistor approaches—which are much less frequently used—may also reach detection limits in the ppt range. In the rare instances where they are tested for the same analyte, FETs may perform as well or better than MWCNT electrodes, while chemistors usually perform worse than both FETs and electrochemical sensors. Overall, a more extensive evaluation of FET and chemistors for various analytes would be valuable.

A large variety of functional probes is reported. They cover the full-size scale from single atomic dopants to polymers and often couple two to three chemical building blocks. While these probes provide remarkable performances, especially the multi-component ones, there are—surprisingly—several analytes for which non-functionalized or COOH-functionalized CNTs provide better performances (pH, Zn(II), free chlorine). Non-functionalized or COOH-functionalized CNTs sensors are also reported to allow selectivity and to be resilient to interferents. These results suggest to systematically compare in new studies the performances of functionalized and non-functionalized CNTs.

Finally, despite the large numbers of analytes covered here, there are still a lot of highly environmentally relevant analytes that are not covered at all by the literature, for instance the various forms of iron (Fe(II), Fe(III)), ammonium, nitrate and phosphate ions or the various pesticides found at a ppb level in drink water. In addition, the literature does not address emerging contaminants such as drugs (e.g., antibiotics, hormones) and their degradation by-products or biological materials such as bacteria. While a few of these targets may be covered by the literature about optical CNT sensors, these results do not transfer easily to online monitoring applications, so that there are still a lot of research opportunities for electrical and electrochemical CNT sensors.

Author Contributions: Writing-original draft preparation, G.C. and S.A.; Writing-review and editing, B.L. and G.Z. All authors have read and agreed to the published version of the manuscript.

Funding: French National Research Agency (ANR): N° 17-CE04-0003-04; Horizon 2020 research and innovation program: N° 820881.

Acknowledgments: The authors acknowledge support from project 4-Water funded by French National Research Agency (ANR) and from project LOTUS co-funded by the European Commission under the Horizon 2020 research and by the Indian Government, Ministry of Science and Technology.

Conflicts of Interest: The authors declare the financial interests which may be considered as potential competing interests from the support by project 4-Water and LOTUS.

References

1. Hinrichsen, D.; Tacio, H. *The Coming Freshwater Crisis Is Already Here*; ESCP Publication: Washington, DC, USA, 2002; pp. 1–26.
2. Solomon, A.; Ahmed, Z.; Biruktawit, K.; Amare, D.; Solomon, A.; Endalew, Z.; Abera, S.; Zeyinudin, A.; Kebede, B.; Deribew, A.; et al. Bacteriological analysis of drinking water sources. *Afr. J. Microbiol. Res.* **2011**, *5*, 2638–2641. [[CrossRef](#)]
3. Aydin, A. The microbiological and physico-chemical quality of groundwater in West Thrace, Turkey. *Pol. J. Environ. Stud.* **2007**, *16*, 377–383.
4. Gyamfi, E.; Ackah, M.; Anim, A.; Hanson, J.; Kpattah, L.; Enti-Brown, S.; Adjei-Kyereme, Y.; Nyarko, E. Chemical analysis of potable water samples from selected suburbs of Accra, Ghana. *Proc. Int. Acad. Ecol. Environ. Sci.* **2012**, *2*, 118–127.
5. Clark, R.M.; Coyle, J.A. Measuring and Modeling Variations in Distribution System Water Quality. *J.-Am. Water Work. Assoc.* **1990**, *82*, 46–53. [[CrossRef](#)]
6. Raich, J. Review of sensors to monitor water quality. *Eur. Ref. Netw. Crit. Infrastruct. Prot.* **2013**. [[CrossRef](#)]
7. Kruse, P. Review on water quality sensors. *J. Phys. D Appl. Phys.* **2018**, *51*, 203002. [[CrossRef](#)]
8. Bhardwaj, J.; Gupta, K.K.; Gupta, R. A review of emerging trends on water quality measurement sensors. In Proceedings of the 2015 International Conference on Technologies for Sustainable Development (ICTSD), Mumbai, India, 4–6 February 2015; pp. 1–6. [[CrossRef](#)]
9. Mao, S.; Chang, J.; Zhou, G.; Chen, J. Nanomaterial-enabled Rapid Detection of Water Contaminants. *Small* **2015**, *11*, 5336–5359. [[CrossRef](#)]
10. Soni, R.; Soni, M.; Shukla, D.P. Emerging Techniques and Materials for Water Pollutants Detection. In *Sensors in Water Pollutants Monitoring: Role of Material*; Springer: Singapore, 2019; pp. 277–297. [[CrossRef](#)]
11. Willner, M.R.; Vikesland, P.J. Nanomaterial enabled sensors for environmental contaminants. *J. Nanobiotechnol.* **2018**, *16*, 95. [[CrossRef](#)] [[PubMed](#)]
12. Arduini, F.; Calvo, J.Q.; Palleschi, G.; Moscone, D.; Amine, A. Bismuth-modified electrodes for lead detection. *TrAC Trends Anal. Chem.* **2010**, *29*, 1295–1304. [[CrossRef](#)]
13. Yao, S.; Swetha, P.; Zhu, Y. Nanomaterial-Enabled Wearable Sensors for Healthcare. *Adv. Healthc. Mater.* **2017**, *7*, 1700889. [[CrossRef](#)]
14. Kong, J.; Franklin, N.R.; Zhou, C.; Chapline, M.G.; Peng, S.; Cho, K.; Dai, H. Nanotube Molecular Wires as Chemical Sensors. *Science* **2000**, *287*, 622–625. [[CrossRef](#)] [[PubMed](#)]
15. Schroeder, V.; Savagatrup, S.; He, M.; Lin, S.; Swager, T.M. Carbon nanotube chemical sensors. *Chem. Rev.* **2019**, *119*, 599–663. [[CrossRef](#)] [[PubMed](#)]
16. Khatri, P.; Gupta, K.K.; Gupta, R.K. Raspberry Pi-based smart sensing platform for drinking-water quality monitoring system: A Python framework approach. *Drink. Water Eng. Sci.* **2019**, *12*, 31–37. [[CrossRef](#)]
17. Subramanian, B.; Chen, S.S.; Reddy, K.R. *Emerging Technologies for Agriculture and Environment: Select Proceedings of ITsFEW 2018*; Springer: Singapore, 2020.
18. Luo, X.; Yang, J. A Survey on Pollution Monitoring Using Sensor Networks in Environment Protection. *J. Sens.* **2019**, *2019*, 6271206. [[CrossRef](#)]
19. Arash, B.; Wang, Q.; Varadan, V.K. Carbon Nanotube-Based Sensors for Detection of Gas Atoms. *J. Nanotechnol. Eng. Med.* **2011**, *2*, 021010. [[CrossRef](#)]
20. Li, C.; Shi, G. Carbon nanotube-based fluorescence sensors. *J. Photochem. Photobiol. C Photochem. Rev.* **2014**, *19*, 20–34. [[CrossRef](#)]
21. Jaywant, S.A.; Arif, K.M. A comprehensive review of micro fluidic water quality monitoring sensors. *Sensors* **2019**, *19*, 4781. [[CrossRef](#)]
22. Vikesland, P.J.; Wigginton, K.R. Nanomaterial Enabled Biosensors for Pathogen Monitoring—A Review. *Environ. Sci. Technol.* **2010**, *44*, 3656–3669. [[CrossRef](#)]
23. Iijima, S.; Ichihashi, T. Single-shell carbon nanotubes of 1-nm diameter. *Nature* **1993**, *363*, 603–605. [[CrossRef](#)]
24. Belin, T.; Epron, F. Characterization methods of carbon nanotubes: A review. *Mater. Sci. Eng. B* **2005**, *119*, 105–118. [[CrossRef](#)]
25. Qiu, H.; Yang, J. Chapter 2—Structure and Properties of Carbon Nanotubes. In *Industrial Applications of Carbon Nanotubes*; Peng, H., Li, Q., Chen, T., Eds.; Elsevier: Boston, MA, USA, 2017; pp. 47–69.
26. Soni, S.K.; Thomas, B.; Kar, V.R. A Comprehensive Review on CNTs and CNT-Reinforced Composites: Syntheses, Characteristics and Applications. *Mater. Today Commun.* **2020**, *25*, 101546. [[CrossRef](#)]
27. Aius, A. Graphene Honeycomb Lattice. CC BY-SA 3.0.
28. Zhang, Y.; Bai, Y.; Yan, B. Functionalized carbon nanotubes for potential medicinal applications. *Drug Discov. Today* **2010**, *15*, 428–435. [[CrossRef](#)] [[PubMed](#)]
29. Zhang, T.; Mubeen, S.; Myung, N.V.; Deshusses, M.A. Recent progress in carbon nanotube-based gas sensors. *Nanotechnology* **2008**, *19*, 332001. [[CrossRef](#)]
30. Andringa, A.-M.; Meijboom, J.R.; Smits, E.C.P.; Mathijssen, S.G.J.; Blom, P.W.M.; de Leeuw, D.M. Gate-Bias Controlled Charge Trapping as a Mechanism for NO₂ Detection with Field-Effect Transistors. *Adv. Funct. Mater.* **2010**, *21*, 100–107. [[CrossRef](#)]
31. Hirsch, A. Functionalization of single-walled carbon nanotubes. *Angew. Chem. Int. Ed.* **2002**, *41*, 1853–1859. [[CrossRef](#)]

32. Banerjee, S.; Hemraj-Benny, T.; Wong, S.S. Covalent Surface Chemistry of Single-Walled Carbon Nanotubes. *Adv. Mater.* **2005**, *17*, 17–29. [[CrossRef](#)]
33. Star, A.; Stoddart, J.F.; Steuerman, D.; Diehl, M.; Boukai, A.; Wong, E.W.; Yang, X.; Chung, S.-W.; Choi, H.; Heath, J.R. Preparation and Properties of Polymer-Wrapped Single-Walled Carbon Nanotubes. *Angew. Chem. Int. Ed.* **2001**, *40*, 1721–1725. [[CrossRef](#)]
34. Britz, D.A.; Khlobystov, A.N. Noncovalent interactions of molecules with single walled carbon nanotubes. *Chem. Soc. Rev.* **2006**, *35*, 637–659. [[CrossRef](#)] [[PubMed](#)]
35. Bondavalli, P.; Gorintin, L.; Feugnet, G.; Lehoucq, G.; Pribat, D. Selective gas detection using CNTFET arrays fabricated using air-brush technique, with different metal as electrodes. *Sens. Actuators B Chem.* **2014**, *202*, 1290–1297. [[CrossRef](#)]
36. Cardenas-Benitez, B.; Djordjevic, I.; Hosseini, S.; Madou, M.J.; Martinez-Chapa, S.O. Review—Covalent Functionalization of Carbon Nanomaterials for Biosensor Applications: An Update. *J. Electrochem. Soc.* **2018**, *165*, B103–B117. [[CrossRef](#)]
37. Zhou, Y.; Fang, Y.; Ramasamy, R.P. Non-Covalent Functionalization of Carbon Nanotubes for Electrochemical Biosensor Development. *Sensors* **2019**, *19*, 392. [[CrossRef](#)] [[PubMed](#)]
38. Toshimitsu, F.; Nakashima, N. Semiconducting single-walled carbon nanotubes sorting with a removable solubilizer based on dynamic supramolecular coordination chemistry. *Nat. Commun.* **2014**, *5*, 5041. [[CrossRef](#)] [[PubMed](#)]
39. Zucchi, G.; Lebental, B.; Loisel, L.; Ramachandran, S.; Gutiérrez, A.F.; Wang, X.; Godumala, M.; Bodeleot, L. Chemical Sensors Based on Carbon Nanotubes Functionalised by Conjugated Polymers for Analysis in Aqueous Medium. Patent 3609881A1, 10 April 2018.
40. Gómez, S.; Rendtorff, N.M.; Aglietti, E.F.; Sakka, Y.; Suárez, G. Surface modification of multiwall carbon nanotubes by sulfonitric treatment. *Appl. Surf. Sci.* **2016**, *379*, 264–269. [[CrossRef](#)]
41. Gong, H.; Kim, S.-T.; Lee, J.D.; Yim, S. Simple quantification of surface carboxylic acids on chemically oxidized multi-walled carbon nanotubes. *Appl. Surf. Sci.* **2013**, *266*, 219–224. [[CrossRef](#)]
42. Wepasnick, K.A.; Smith, B.A.; Schrote, K.E.; Wilson, H.K.; Diegelmann, S.R.; Fairbrother, D.H. Surface and structural characterization of multi-walled carbon nanotubes following different oxidative treatments. *Carbon* **2011**, *49*, 24–36. [[CrossRef](#)]
43. Anantram, M.P.; Léonard, F. Physics of carbon nanotube electronic devices. *Rep. Prog. Phys.* **2006**, *69*, 507–561. [[CrossRef](#)]
44. Tans, S.J.; Verschueren, A.R.M.; Dekker, C. Room-temperature transistor based on a single carbon nanotube. *Nature* **1998**, *393*, 49–52. [[CrossRef](#)]
45. Snow, E.S.; Novak, J.P.; Campbell, P.M.; Park, D. Random networks of carbon nanotubes as an electronic material. *Appl. Phys. Lett.* **2003**, *82*, 2145–2147. [[CrossRef](#)]
46. Zhou, X.; Boey, F.; Zhang, H. Controlled growth of single-walled carbon nanotubes on patterned substrates. *Chem. Soc. Rev.* **2011**, *40*, 5221–5231. [[CrossRef](#)]
47. Ahmad, M.; Silva, S.R.P. Low temperature growth of carbon nanotubes—A review. *Carbon* **2020**, *158*, 24–44. [[CrossRef](#)]
48. Seidel, R.; Graham, A.P.; Unger, E.; Duesberg, G.S.; Liebau, M.; Steinhögl, W.; Kreupl, F.; Hoenlein, W.; Pompe, W. High-Current Nanotube Transistors. *Nano Lett.* **2004**, *4*, 831–834. [[CrossRef](#)]
49. Zhang, G.; Qi, P.; Wang, X.; Lu, Y.; Li, X.; Tu, R.; Bangsaruntip, S.; Mann, D.; Zhang, L.; Dai, H. Selective Etching of Metallic Carbon Nanotubes by Gas-Phase Reaction. *Science* **2006**, *314*, 974–977. [[CrossRef](#)]
50. Zhang, Y.; Zhang, Y.; Xian, X.; Zhang, J.; Liu, Z. Sorting out Semiconducting Single-Walled Carbon Nanotube Arrays by Preferential Destruction of Metallic Tubes Using Xenon-Lamp Irradiation. *J. Phys. Chem. C* **2008**, *112*, 3849–3856. [[CrossRef](#)]
51. Zhao, Q.; Yao, F.; Wang, Z.; Deng, S.; Tong, L.; Liu, K.; Zhang, J. Real-Time Observation of Carbon Nanotube Etching Process Using Polarized Optical Microscope. *Adv. Mater.* **2017**, *29*, 1701959. [[CrossRef](#)]
52. Lei, T.; Pochorovski, I.; Bao, Z. Separation of Semiconducting Carbon Nanotubes for Flexible and Stretchable Electronics Using Polymer Removable Method. *Acc. Chem. Res.* **2017**, *50*, 1096–1104. [[CrossRef](#)]
53. Hsu, L.H.H.; Hoque, E.; Kruse, P.; Selvaganapathy, P.R. A carbon nanotube based resettable sensor for measuring free chlorine in drinking water. *Appl. Phys. Lett.* **2015**, *106*, 063102. [[CrossRef](#)]
54. Pyo, J.-Y.; Cho, W.-J. High-sensitivity pH sensor using separative extended-gate field-effect transistors with single-walled carbon-nanotube networks. *Jpn. J. Appl. Phys.* **2018**, *57*, 04FP02. [[CrossRef](#)]
55. Wei, L.; Lu, D.; Wang, J.; Wei, H.; Zhao, J.; Geng, H.; Zhang, Y. Highly sensitive detection of trinitrotoluene in water by chemiresistive sensor based on noncovalently amino functionalized single-walled carbon nanotube. *Sens. Actuators B Chem.* **2013**, *190*, 529–534. [[CrossRef](#)]
56. Eshkalak, S.K.; Chinnappan, A.; Jayathilaka, D.; Khatibzadeh, M.; Kowsari, E.; Ramakrishna, S. A review on inkjet printing of CNT composites for smart applications. *Appl. Mater. Today* **2017**, *9*, 372–386. [[CrossRef](#)]
57. Kwon, J.-H.; Lee, K.-S.; Lee, Y.-H.; Ju, B.-K. Single-Wall Carbon Nanotube-Based pH Sensor Fabricated by the Spray Method. *Electrochem. Solid-State Lett.* **2006**, *9*, H85–H87. [[CrossRef](#)]
58. Goh, G.L.; Agarwala, S.; Tan, Y.J.; Yeong, W.Y. A low cost and flexible carbon nanotube pH sensor fabricated using aerosol jet technology for live cell applications. *Sens. Actuators B Chem.* **2018**, *260*, 227–235. [[CrossRef](#)]
59. Stojanović, G.; Kojić, T.; Radovanović, M.; Vasiljević, D.; Panić, S.; Srdić, V.; Cvejić, J. Flexible sensors based on two conductive electrodes and MWCNTs coating for efficient pH value measurement. *J. Alloys Compd.* **2019**, *794*, 76–83. [[CrossRef](#)]
60. Gou, P.; Kraut, N.D.; Feigel, I.M.; Bai, H.; Morgan, G.J.; Chen, Y.; Tang, Y.; Bocan, K.; Stachel, J.; Berger, L.; et al. Carbon Nanotube Chemiresistor for Wireless pH Sensing. *Sci. Rep.* **2014**, *4*, 4468. [[CrossRef](#)]

61. Endo, M.; Muramatsu, H.; Hayashi, T.; Kim, Y.A.; Terrones, M.; Dresselhaus, M.S. 'Buckypaper' from coaxial nanotubes. *Nature* **2005**, *433*, 476. [[CrossRef](#)] [[PubMed](#)]
62. Bard, A.J.; Faulkner, R.L. *Electrochemical Methods: Fundamentals and Applications*, 2nd ed.; Wiley: Hoboken, NJ, USA, 2001; Volume 38, pp. 1364–1365.
63. Biesheuvel, P.M.; Porada, S.; Dykstra, J.E. The difference between Faradaic and non-Faradaic electrode processes. *arXiv* **2021**, arXiv:1809.02930.
64. Bakker, E.; Pretsch, E. Potentiometric sensors for trace-level analysis. *TrAC Trends Anal. Chem.* **2005**, *24*, 199–207. [[CrossRef](#)]
65. Hanrahan, G.; Patil, D.G.; Wang, J. Electrochemical sensors for environmental monitoring: Design, development and applications. *J. Environ. Monit.* **2004**, *6*, 657–664. [[CrossRef](#)] [[PubMed](#)]
66. Simões, F.R.; Xavier, M.G. 6—Electrochemical Sensors. In *Nanoscience and Its Applications*; Róz, A.L.D., Ferreira, M., Leite, F.L., Osvaldo, N., Oliveira, A., Eds.; William Andrew Publishing: Norwich, NY, USA, 2017; pp. 155–178.
67. March, G.; Nguyen, T.D.; Piro, B. Modified Electrodes Used for Electrochemical Detection of Metal Ions in Environmental Analysis. *Biosensors* **2015**, *5*, 241–275. [[CrossRef](#)]
68. Zhou, H.; Yang, W.; Sun, C. Amperometric sulfite sensor based on multiwalled carbon nanotubes/ferrocene-branched chitosan composites. *Talanta* **2008**, *77*, 366–371. [[CrossRef](#)]
69. Gao, X.; Wei, W.; Yang, L.; Guo, M. Carbon Nanotubes/Poly(1,2-diaminobenzene) Nanoporous Composite Film Electrode Prepared by Multipulse Potentiostatic Electropolymerisation and Its Application to Determination of Trace Heavy Metal Ions. *Electroanalysis* **2006**, *18*, 485–492. [[CrossRef](#)]
70. Farid, M.U.; Khanzada, N.K.; An, A.K. Understanding fouling dynamics on functionalized CNT-based membranes: Mechanisms and reversibility. *Desalination* **2019**, *456*, 74–84. [[CrossRef](#)]
71. Britto, P.; Santhanam, K.; Ajayan, P. Carbon nanotube electrode for oxidation of dopamine. *Bioelectrochem. Bioenerg.* **1996**, *41*, 121–125. [[CrossRef](#)]
72. Xu, H.; Zeng, L.; Xing, S.; Shi, G.; Xian, Y.; Jin, L. Microwave-radiated synthesis of gold nanoparticles/carbon nanotubes composites and its application to voltammetric detection of trace mercury(II). *Electrochem. Commun.* **2008**, *10*, 1839–1843. [[CrossRef](#)]
73. Xu, H.; Zeng, L.; Xing, S.; Xian, Y.; Shi, G.; Jin, L. Ultrasensitive Voltammetric Detection of Trace Lead(II) and Cadmium(II) Using MWCNTs-Nafion/Bismuth Composite Electrodes. *Electroanalysis* **2008**, *20*, 2655–2662. [[CrossRef](#)]
74. Morton, J.; Havens, N.; Mugweru, A.; Wanekaya, A.K. Detection of Trace Heavy Metal Ions Using Carbon Nanotube- Modified Electrodes. *Electroanalysis* **2009**, *21*, 1597–1603. [[CrossRef](#)]
75. Zhang, W.; Wang, G.; Zhang, X.; Fang, B. Amperometric Detection of Hydrogen Peroxide Using Glassy Carbon Electrodes Modified with Chromium Hexacyanoferrate/Single-Walled Carbon Nanotube Nanocomposites. *Electroanalysis* **2009**, *21*, 179–183. [[CrossRef](#)]
76. Hu, F.; Chen, S.; Wang, C.; Yuan, R.; Yuan, D.; Wang, C. Study on the application of reduced graphene oxide and multiwall carbon nanotubes hybrid materials for simultaneous determination of catechol, hydroquinone, p-cresol and nitrite. *Anal. Chim. Acta* **2012**, *724*, 40–46. [[CrossRef](#)]
77. Hu, C.; Hu, S. Carbon Nanotube-Based Electrochemical Sensors: Principles and Applications in Biomedical Systems. *J. Sens.* **2009**, *2009*, 187615. [[CrossRef](#)]
78. Karimi-Maleh, H.; Ensafi, A.A.; Beitollahi, H.; Nasiri, V.; Khalilzadeh, M.A.; Biparva, P. Electrocatalytic determination of sulfite using a modified carbon nanotubes paste electrode: Application for determination of sulfite in real samples. *Ionics* **2012**, *18*, 687–694. [[CrossRef](#)]
79. Khani, H.; Rofouei, M.K.; Arab, P.; Gupta, V.K.; Vafaei, Z. Multi-walled carbon nanotubes-ionic liquid-carbon paste electrode as a super selectivity sensor: Application to potentiometric monitoring of mercury ion(II). *J. Hazard. Mater.* **2010**, *183*, 402–409. [[CrossRef](#)]
80. Gong, K.; Yan, Y.; Zhang, M.; Su, L.; Xiong, S.; Mao, L. Electrochemistry and Electroanalytical Applications of Carbon Nanotubes: A Review. *Anal. Sci.* **2005**, *21*, 1383–1393. [[CrossRef](#)]
81. He, Y.; Chen, W.; Gao, C.; Zhou, J.; Li, X.; Xie, E. An overview of carbon materials for flexible electrochemical capacitors. *Nanoscale* **2013**, *5*, 8799–8820. [[CrossRef](#)]
82. Zhang, J.; Zou, H.; Qing, Q.; Yang, Y.; Li, Q.; Liu, Z.; Guo, A.X.; Du, Z. Effect of Chemical Oxidation on the Structure of Single-Walled Carbon Nanotubes. *J. Phys. Chem. B* **2003**, *107*, 3712–3718. [[CrossRef](#)]
83. Luo, H.; Shi, Z.; Li, N.; Gu, Z.; Zhuang, Q. Investigation of the Electrochemical and Electrocatalytic Behavior of Single-Wall Carbon Nanotube Film on a Glassy Carbon Electrode. *Anal. Chem.* **2001**, *73*, 915–920. [[CrossRef](#)] [[PubMed](#)]
84. Zahab, A.; Spina, L.; Poncharal, P.; Marlière, C. Water-vapor effect on the electrical conductivity of a single-walled carbon nanotube mat. *Phys. Rev. B* **2000**, *62*, 10000–10003. [[CrossRef](#)]
85. Van der Pauw, L.J. A method of measuring specific resistivity and hall effect of discs of arbitrary shape. *Philips Res. Rep.* **1958**, *13*, 1–9.
86. Tang, R.; Shi, Y.; Hou, Z.; Wei, L. Carbon nanotube-based chemiresistive sensors. *Sensors* **2017**, *17*, 882. [[CrossRef](#)] [[PubMed](#)]
87. Meyyappan, M. Carbon Nanotube-Based Chemical Sensors. *Small* **2016**, *12*, 2118–2129. [[CrossRef](#)]
88. Li, P.; Martin, C.M.; Yeung, K.K.; Xue, W. Dielectrophoresis aligned single-walled carbon nanotubes as pH sensors. *Biosensors* **2011**, *1*, 23–35. [[CrossRef](#)]

89. Jung, D.; Han, M.; Lee, G.S. pH-sensing characteristics of multi-walled carbon nanotube sheet. *Mater. Lett.* **2014**, *122*, 281–284. [[CrossRef](#)]
90. Li, J.; Lu, Y.; Ye, Q.; Delzeit, L.; Meyyappan, M. A Gas Sensor Array Using Carbon Nanotubes and Microfabrication Technology. *Electrochem. Solid-State Lett.* **2005**, *8*, H100–H102. [[CrossRef](#)]
91. Li, J.; Lu, Y.; Ye, Q.; Cinke, M.; Han, J.; Meyyappan, M. Carbon Nanotube Sensors for Gas and Organic Vapor Detection. *Nano Lett.* **2003**, *3*, 929–933. [[CrossRef](#)]
92. Michelis, F.; Bodelot, L.; Bonnassieux, Y.; Lebental, B. Highly reproducible, hysteresis-free, flexible strain sensors by inkjet printing of carbon nanotubes. *Carbon* **2015**, *95*, 1020–1026. [[CrossRef](#)]
93. Naderi, A.; Tahne, B.A. Review—Methods in Improving the Performance of Carbon Nanotube Field Effect Transistors. *ECS J. Solid State Sci. Technol.* **2016**, *5*, M131–M140. [[CrossRef](#)]
94. Imam, S.-A.; Kalam, N.; Abdullah, S. Comparative Analysis of Control Coefficients on the Performance of CNTFET Under Different Parameters. *Int. J. Nanosci.* **2016**, *15*, 1640005. [[CrossRef](#)]
95. Karimi, N.V.; Pourasad, Y. Investigating the effect of some parameters of the channel on the characteristics of tunneling carbon nanotube field-effect transistor. *Int. Nano Lett.* **2016**, *6*, 215–221. [[CrossRef](#)]
96. Kim, W.; Javey, A.; Vermesh, O.; Wang, Q.; Li, A.Y.; Dai, H. Hysteresis Caused by Water Molecules in Carbon Nanotube Field-Effect Transistors. *Nano Lett.* **2003**, *3*, 193–198. [[CrossRef](#)]
97. Joshi, S.; Bhatt, V.D.; Rani, H.; Becherer, M.; Lugli, P. Understanding the influence of in-plane gate electrode design on electrolyte gated transistor. *Microelectron. Eng.* **2018**, *199*, 87–91. [[CrossRef](#)]
98. Saito, R.; Fujita, M.; Dresselhaus, G.; Dresselhaus, M.S. Electronic structure of chiral graphene tubules. *Appl. Phys. Lett.* **1992**, *60*, 2204–2206. [[CrossRef](#)]
99. Di Bartolomeo, A.; Rinzan, M.; Boyd, A.K.; Yang, Y.; Guadagno, L.; Giubileo, F.; Barbara, P. Electrical properties and memory effects of field-effect transistors from networks of single- and double-walled carbon nanotubes. *Nanotechnology* **2010**, *21*, 115204. [[CrossRef](#)]
100. Derycke, V.; Martel, R.; Appenzeller, J.; Avouris, P. Carbon Nanotube Inter- and Intramolecular Logic Gates. *Nano Lett.* **2001**, *1*, 453–456. [[CrossRef](#)]
101. Zhao, J.; Hashmi, A.; Xu, J.; Xue, W. A compact lab-on-a-chip nanosensor for glycerol detection. *Appl. Phys. Lett.* **2012**, *100*, 243109. [[CrossRef](#)]
102. Pimparkar, N.; Kocabas, C.; Kang, S.J.; Rogers, J.; Alam, M. Limits of Performance Gain of Aligned CNT Over Randomized Network: Theoretical Predictions and Experimental Validation. *IEEE Electron Device Lett.* **2007**, *28*, 593–595. [[CrossRef](#)]
103. Barman, S.N.; Lemieux, M.C.; Baek, J.; Rivera, R.; Bao, Z. Effects of Dispersion Conditions of Single-Walled Carbon Nanotubes on the Electrical Characteristics of Thin Film Network Transistors. *ACS Appl. Mater. Interfaces* **2010**, *2*, 2672–2678. [[CrossRef](#)]
104. Ruiz-Soria, G.; Paz, A.P.; Sauer, M.; Mowbray, D.J.; Lacovig, P.; Dalmiglio, M.; Lizzit, S.; Yanagi, K.; Rubio, A.; Goldoni, A.; et al. Revealing the adsorption mechanisms of nitroxides on ultrapure, metallicity-sorted carbon nanotubes. *ACS Nano* **2014**, *8*, 1375–1383. [[CrossRef](#)] [[PubMed](#)]
105. Park, M.; Kim, H.S.; Kim, T.; Kim, J.; Seo, S.; Lee, B.Y. Real-time monitoring of microbial activity using hydrogel-hybridized carbon nanotube transistors. *Sens. Actuators B Chem.* **2018**, *263*, 486–492. [[CrossRef](#)]
106. Takeda, S.; Nakamura, M.; Ishii, A.; Subagyo, A.; Hosoi, H.; Sueoka, K.; Mukasa, K. A pH sensor based on electric properties of nanotubes on a glass substrate. *Nanoscale Res. Lett.* **2007**, *2*, 207–212. [[CrossRef](#)] [[PubMed](#)]
107. Münzer, A.; Melzer, K.; Heimgreiter, M.; Scarpa, G. Random CNT network and regioregular poly(3-hexylthiophen) FETs for pH sensing applications: A comparison. *Biochim. Biophys. Acta (BBA)-Gen. Subj.* **2013**, *1830*, 4353–4358. [[CrossRef](#)] [[PubMed](#)]
108. Chernov, A.I.; Eremina, V.A.; Shook, J.; Collins, A.; Walker, P.; Fedotov, P.V.; Zakhidov, A.A.; Obratsova, E.D. Field Effect Transistor Based on Solely Semiconducting Single-Walled Carbon Nanotubes for the Detection of 2-Chlorophenol. *Phys. Status Solidi B* **2017**, *255*, 1700139. [[CrossRef](#)]
109. Lee, N.H.; Nahm, S.-H.; Choi, I.S. Real-Time Monitoring of a Botulinum Neurotoxin Using All-Carbon Nanotube-Based Field-Effect Transistor Devices. *Sensors* **2018**, *18*, 4235. [[CrossRef](#)]
110. Kim, T.H.; Lee, J.; Hong, S. Highly Selective Environmental Nanosensors Based on Anomalous Response of Carbon Nanotube Conductance to Mercury Ions. *J. Phys. Chem. C* **2009**, *113*, 19393–19396. [[CrossRef](#)]
111. Lee, C.-S.; Ju, Y.; Kim, J.; Kim, T.H. Electrochemical functionalization of single-walled carbon nanotubes with amine-terminated dendrimers encapsulating Pt nanoparticles: Toward facile field-effect transistor-based sensing platforms. *Sens. Actuators B Chem.* **2018**, *275*, 367–372. [[CrossRef](#)]
112. Joshi, S.; Bhatt, V.D.; Jaworska, E.; Michalska, A.; Maksymiuk, K.; Becherer, M.; Gagliardi, A.; Lugli, P. Ambient Processed, Water-Stable, Aqueous-Gated sub 1 V n-type Carbon Nanotube Field Effect Transistor. *Sci. Rep.* **2018**, *8*, 11386. [[CrossRef](#)]
113. Forzani, E.S.; Li, X.; Zhang, P.; Tao, N.; Zhang, R.; Amlani, I.; Tsui, R.; Nagahara, L.A. Tuning the Chemical Selectivity of SWNT-FETs for Detection of Heavy-Metal Ions. *Small* **2006**, *2*, 1283–1291. [[CrossRef](#)]
114. Star, A.; Bradley, K.; Gabriel, J.-C.; Gruner, G. Nano-Electronic Sensors: Chemical Detection Using Carbon Nanotubes. *Polym. Mater. Sci. Eng.* **2003**, *89*, 204.
115. Rosenblatt, S.; Yaish, Y.; Park, J.; Gore, J.; Sazonova, V.; McEuen, P.L. High Performance Electrolyte Gated Carbon Nanotube Transistors. *Nano Lett.* **2002**, *2*, 869–872. [[CrossRef](#)]

116. Xu, X.; Tang, Z.; Shao, J.; Fan, Z. The study on the interface adhesion comparison of MgF_2 , Al_2O_3 , SiO_2 , and Ag thin films. *Appl. Surf. Sci.* **2005**, *245*, 11–15. [[CrossRef](#)]
117. Boyd, A.; Dube, I.; Fedorov, G.; Paranjape, M.; Barbara, P. Gas sensing mechanism of carbon nanotubes: From single tubes to high-density networks. *Carbon* **2014**, *69*, 417–423. [[CrossRef](#)]
118. Benda, R.; Cancès, E.; Lebental, B. Effective resistance of random percolating networks of stick nanowires: Functional dependence on elementary physical parameters. *J. Appl. Phys* **2019**, *126*, 44306. [[CrossRef](#)]
119. Noori, J.S.; Mortensen, J.; Geto, A. Recent Development on the Electrochemical Detection of Selected Pesticides: A Focused Review. *Sensors* **2020**, *20*, 2221. [[CrossRef](#)]
120. Wang, W.; Wang, X.; Cheng, N.; Luo, Y.; Lin, Y.; Xu, W.; Du, D. Recent advances in nanomaterials-based electrochemical (bio)sensors for pesticides detection. *TrAC Trends Anal. Chem.* **2020**, *132*, 116041. [[CrossRef](#)]
121. Covington, A.K.; Bates, R.G.; Durst, R.A. Definition of pH scales, standard reference values, measurement of pH and related terminology (Recommendations 1984). *Pure Appl. Chem.* **1985**, *57*, 531–542. [[CrossRef](#)]
122. World Health Organization. *pH in Drinking-Water Background Document for Development of WHO Guidelines for Drinking-Water Quality*; World Health Organization: Geneva, Switzerland, 1996.
123. Lei, K.F.; Lee, K.-F.; Yang, S.-I. Fabrication of carbon nanotube-based pH sensor for paper-based microfluidics. *Microelectron. Eng.* **2012**, *100*, 1–5. [[CrossRef](#)]
124. Jasti, R.; Bhattacharjee, J.; Neaton, J.B.; Bertozzi, C.R. Synthesis, Characterization, and Theory of [9]-, [12]-, and [18]-Cycloparaphenylene: Carbon NanoHoop Structures. *J. Am. Chem. Soc.* **2008**, *130*, 17646–17647. [[CrossRef](#)]
125. Wang, L.; Xie, Y.; Yang, Y.; Liang, H.; Wang, L.; Song, Y. Electroactive Covalent Organic Frameworks/Carbon Nanotubes Composites for Electrochemical Sensing. *ACS Appl. Nano Mater.* **2020**, *3*, 1412–1419. [[CrossRef](#)]
126. Liao, Y.; Zhang, C.; Zhang, Y.; Strong, V.; Tang, J.; Li, X.-G.; Kalantar-Zadeh, K.; Hoek, E.M.V.; Wang, K.L.; Kaner, R.B. Carbon Nanotube/Polyaniline Composite Nanofibers: Facile Synthesis and Chemosensors. *Nano Lett.* **2011**, *11*, 954–959. [[CrossRef](#)] [[PubMed](#)]
127. Jeon, J.-Y.; Kang, B.-C.; Ha, T.-J. Flexible pH sensors based on printed nanocomposites of single-wall carbon nanotubes and Nafion. *Appl. Surf. Sci.* **2020**, *514*, 145956. [[CrossRef](#)]
128. Kaempgen, M.; Roth, S. Transparent and Flexible Carbon Nanotube/Polyaniline pH Sensor. *J. Electroanal. Chem.* **2005**, *586*, 72–76. [[CrossRef](#)]
129. Zhao, D.; Guo, X.; Wang, T.; Alvarez, N.; Shanov, V.N.; Heineman, W.R. Simultaneous Detection of Heavy Metals by Anodic Stripping Voltammetry Using Carbon Nanotube Thread. *Electroanalysis* **2014**, *26*, 488–496. [[CrossRef](#)]
130. Rahm, C.E.; Torres-Canas, F.; Gupta, P.; Poulin, P.; Alvarez, N.T. Inkjet Printed Multi-walled Carbon Nanotube Sensor for the Detection of Lead in Drinking Water. *Electroanalysis* **2020**, *32*, 1533–1545. [[CrossRef](#)]
131. Tarley, C.R.T.; Santos, V.; Baeta, B.; Pereira, A.; Kubota, L.T. Simultaneous determination of zinc, cadmium and lead in environmental water samples by potentiometric stripping analysis (PSA) using multiwalled carbon nanotube electrode. *J. Hazard. Mater.* **2009**, *169*, 256–262. [[CrossRef](#)] [[PubMed](#)]
132. Wan, Q.; Yu, F.; Zhu, L.; Wang, X.; Yang, N. Bucky-gel coated glassy carbon electrodes, for voltammetric detection of femtomolar leveled lead ions. *Talanta* **2010**, *82*, 1820–1825. [[CrossRef](#)]
133. Wang, L.; Wang, X.; Shi, G.; Peng, C.; Ding, Y. Thiacalixarene Covalently Functionalized Multiwalled Carbon Nanotubes as Chemically Modified Electrode Material for Detection of Ultratrace Pb^{2+} Ions. *Anal. Chem.* **2012**, *84*, 10560–10567. [[CrossRef](#)] [[PubMed](#)]
134. Eltayeb, N.E.; Khan, A. Design and Preparation of a New and Novel Nanocomposite with CNTs and Its Sensor Applications. *J. Mater. Res. Technol.* **2019**, *8*, 2238–2246. [[CrossRef](#)]
135. Fukushima, T.; Kosaka, A.; Ishimura, Y.; Yamamoto, T.; Takigawa, T.; Ishii, N.; Aida, T. Molecular Ordering of Organic Molten Salts Triggered by Single-Walled Carbon Nanotubes. *Science* **2003**, *300*, 2072–2074. [[CrossRef](#)] [[PubMed](#)]
136. Hai, T.L.; Hung, L.C.; Phuong, T.T.B.; Ha, B.T.T.; Nguyen, B.-S.; Hai, T.D.; Nguyen, V.-H. Multiwall carbon nanotube modified by antimony oxide (Sb_2O_3 /MWCNTs) paste electrode for the simultaneous electrochemical detection of cadmium and lead ions. *Microchem. J.* **2020**, *153*, 104456. [[CrossRef](#)]
137. Wang, J. Stripping Analysis at Bismuth Electrodes: A Review. *Electroanalysis* **2005**, *17*, 1341–1346. [[CrossRef](#)]
138. Injang, U.; Noyrod, P.; Siangproh, W.; Dungchai, W.; Motomizu, S.; Chailapakul, O. Determination of trace heavy metals in herbs by sequential injection analysis-anodic stripping voltammetry using screen-printed carbon nanotubes electrodes. *Anal. Chim. Acta* **2010**, *668*, 54–60. [[CrossRef](#)]
139. Hwang, G.H.; Han, W.K.; Park, J.S.; Kang, S.G. Determination of trace metals by anodic stripping voltammetry using a bismuth-modified carbon nanotube electrode. *Talanta* **2008**, *76*, 301–308. [[CrossRef](#)]
140. Ivaska, A.; Kubiak, W.W. Application of sequential injection analysis to anodic stripping voltammetry. *Talanta* **1997**, *44*, 713–723. [[CrossRef](#)]
141. Mohammadi, S.; Taher, M.A.; Beitollahi, H.; Naghizadeh, M. Sensitive voltammetric determination of cadmium at a carbon nanotubes/ Fe_3O_4 /eggshell composites modified carbon paste electrode. *Environ. Nanotechnol. Monit. Manag.* **2019**, *12*, 100241. [[CrossRef](#)]
142. Jia, X.; Li, J.; Wang, E. High-Sensitivity Determination of Lead(II) and Cadmium(II) Based on the CNTs-PSS/Bi Composite Film Electrode. *Electroanalysis* **2010**, *22*, 1682–1687. [[CrossRef](#)]

143. Liu, G.; Lin, Y.; Tu, Y.; Ren, Z. Ultrasensitive voltammetric detection of trace heavy metal ions using carbon nanotube nanoelectrode array. *Analyst* **2005**, *130*, 1098–1101. [[CrossRef](#)]
144. Xu, Z.; Fan, X.; Ma, Q.; Tang, B.; Lu, Z.; Zhang, J.; Mo, G.; Ye, J.; Ye, J. A sensitive electrochemical sensor for simultaneous voltammetric sensing of cadmium and lead based on Fe₃O₄/multiwalled carbon nanotube/laser scribed graphene composites functionalized with chitosan modified electrode. *Mater. Chem. Phys.* **2019**, *238*, 121877. [[CrossRef](#)]
145. Oularbi, L.; Turmine, M.; El Rhazi, M. Preparation of novel nanocomposite consisting of bismuth particles, polypyrrole and multi-walled carbon nanotubes for simultaneous voltammetric determination of cadmium(II) and lead(II). *Synth. Met.* **2019**, *253*, 1–8. [[CrossRef](#)]
146. Xuan, X.; Park, J.Y. A miniaturized and flexible cadmium and lead ion detection sensor based on micro-patterned reduced graphene oxide/carbon nanotube/bismuth composite electrodes. *Sen. Actuators Chem.* **2018**, *255*, 1220–1227. [[CrossRef](#)]
147. Safavi, A.; Maleki, N.; Doroodmand, M.M. Fabrication of a selective mercury sensor based on the adsorption of cold vapor of mercury on carbon nanotubes: Determination of mercury in industrial wastewater. *J. Hazard. Mater.* **2010**, *173*, 622–629. [[CrossRef](#)] [[PubMed](#)]
148. Li, S.; Guerin, D.; Lmimouni, K. Improving performance of OFET by tuning occurrence of charge transport based on pentacene interaction with SAM functionalized contacts. *Microelectron. Eng.* **2018**, *195*, 62–67. [[CrossRef](#)]
149. Bassyouni, M.; Mansi, A.E.; ElGabry, A.; Ibrahim, B.A.; Kassem, O.A.; Alhebeshy, R. Utilization of carbon nanotubes in removal of heavy metals from wastewater: A review of the CNTs' potential and current challenges. *Appl. Phys. A* **2019**, *126*, 38. [[CrossRef](#)]
150. Wei, J.; Yang, D.; Chen, H.; Gao, Y.; Li, H. Stripping voltammetric determination of mercury(II) based on SWCNT-PhSH modified gold electrode. *Sens. Actuators B Chem.* **2014**, *190*, 968–974. [[CrossRef](#)]
151. Bao, Q.; Li, G.; Yang, Z.; Pan, P.; Liu, J.; Chang, J.; Wei, J.; Lin, L. Electrochemical performance of a three-layer electrode based on Bi nanoparticles, multi-walled carbon nanotube composites for simultaneous Hg(II) and Cu(II) detection. *Chin. Chem. Lett.* **2020**, *31*, 2752–2756. [[CrossRef](#)]
152. Deng, W.; Tan, Y.; Li, Y.; Wen, Y.; Su, Z.; Huang, Z.; Huang, S.; Meng, Y.; Xie, Q.; Luo, Y.; et al. Square wave voltammetric determination of Hg(II) using thiol functionalized chitosan-multiwalled carbon nanotubes nanocomposite film electrode. *Microchim. Acta* **2010**, *169*, 367–373. [[CrossRef](#)]
153. Mashhadizadeh, M.H.; Ramezani, S.; Rofouei, M.K. Development of a novel MWCNTs–triazene-modified carbon paste electrode for potentiometric assessment of Hg(II) in the aquatic environments. *Mater. Sci. Eng. C* **2015**, *47*, 273–280. [[CrossRef](#)] [[PubMed](#)]
154. Profumo, A.; Fagnoni, M.; Merli, D.; Quartarone, E.; Protti, S.; Dondi, D.; Albin, A. Multiwalled Carbon Nanotube Chemically Modified Gold Electrode for Inorganic As Speciation and Bi(III) Determination. *Anal. Chem.* **2006**, *78*, 4194–4199. [[CrossRef](#)]
155. Daud, N.; Yusof, N.A.; Tee, T.W.; Abdullah, A.H. Electrochemical sensor for As(III) utilizing CNTs/Leucine/Nafion modified electrode. *Int. J. Electrochem. Sci.* **2012**, *7*, 175–185.
156. Kempahanumakkagari, S.; Deep, A.; Kim, K.-H.; Kailasa, S.K.; Yoon, H.-O. Nanomaterial-based electrochemical sensors for arsenic—A review. *Biosens. Bioelectron.* **2017**, *95*, 106–116. [[CrossRef](#)]
157. Xiao, L.; Wildgoose, G.G.; Compton, R.G. Sensitive electrochemical detection of arsenic (III) using gold nanoparticle modified carbon nanotubes via anodic stripping voltammetry. *Anal. Chim. Acta* **2008**, *620*, 44–49. [[CrossRef](#)] [[PubMed](#)]
158. Shin, S.-H.; Hong, H.-G. Anodic Stripping Voltammetric Detection of Arsenic(III) at Platinum-Iron(III) Nanoparticle Modified Carbon Nanotube on Glassy Carbon Electrode. *Bull. Korean Chem. Soc.* **2010**, *31*, 3077–3083. [[CrossRef](#)]
159. Buffa, A.; Mandler, D. Arsenic(III) detection in water by flow-through carbon nanotube membrane decorated by gold nanoparticles. *Electrochim. Acta* **2019**, *318*, 496–503. [[CrossRef](#)]
160. Izadkhah, V.; Farmany, A.; Mortazavi, S. Voltammetric determination of copper in water samples using a Schiff base/carbon nanotube-modified carbon paste electrode. *J. Ind. Eng. Chem.* **2015**, *21*, 994–996. [[CrossRef](#)]
161. Zhao, H.; Jiang, Y.; Ma, Y.; Wu, Z.; Cao, Q.; He, Y.; Li, X.; Yuan, Z. Poly(2-amino-4-thiazoleacetic acid)/multiwalled carbon nanotubes modified glassy carbon electrodes for the electrochemical detection of copper(II). *Electrochim. Acta* **2010**, *55*, 2518–2521. [[CrossRef](#)]
162. Cabaniss, S.E. Forward Modeling of Metal Complexation by NOM: II. Prediction of Binding Site Properties. *Environ. Sci. Technol.* **2011**, *45*, 3202–3209. [[CrossRef](#)]
163. Qin, D.; Mamat, X.; Li, Y.; Hu, X.; Cheng, H.; Hu, G. A composite with botryoidal texture prepared from nitrogen-doped carbon spheres and carbon nanotubes for voltammetric sensing of copper(II). *Microchem. J.* **2019**, *153*, 104299. [[CrossRef](#)]
164. Wang, S.; Li, J.; Qiu, Y.; Zhuang, X.; Wu, X.; Jiang, J. Facile synthesis of oxidized multi-walled carbon nanotubes functionalized with 5-sulfosalicylic acid/MoS₂ nanosheets nanocomposites for electrochemical detection of copper ions. *Appl. Surf. Sci.* **2019**, *487*, 766–772. [[CrossRef](#)]
165. Gou, P.; Kraut, N.D.; Feigel, I.M.; Star, A. Rigid versus Flexible Ligands on Carbon Nanotubes for the Enhanced Sensitivity of Cobalt Ions. *Macromolecules* **2013**, *46*, 1376–1383. [[CrossRef](#)]
166. Li, P.; Sun, L.; Li, M.; Yang, B.; Li, H.; Li, C. Simultaneous Determination of Small Biomolecules and Nitrite Using an Au/TiO₂/Carbon Nanotube Composite-Modified Electrode. *J. Electrochem. Soc.* **2016**, *163*, 567–572.
167. Zhao, Z.; Zhang, J.; Wang, W.; Sun, Y.; Li, P.; Hu, J.; Chen, L.; Gong, W. Synthesis and electrochemical properties of Co₃O₄-rGO/CNTs composites towards highly sensitive nitrite detection. *Appl. Surf. Sci.* **2019**, *485*, 274–282. [[CrossRef](#)]
168. Thamae, M.; Nyokong, T. Cobalt(II) porphyrazine catalysed reduction of nitrite. *J. Electroanal. Chem.* **1999**, *470*, 126–135. [[CrossRef](#)]

169. Xu, M.; Zhu, X.; Hong, H.; Hu, K.; Yang, R.; Wang, X.; Li, X.; Liu, X. An electro-active amphiphilic copolymer to functionalize carbon nanotubes for highly sensitive determination of nitrite in water. *Colloids Surf. A Physicochem. Eng. Asp.* **2019**, *576*, 123–129. [[CrossRef](#)]
170. Azadbakht, A.; Abbasi, A.R.; Derikvand, Z.; Amraei, S. Immobilized organoruthenium(II) complexes onto polyethyleneimine-wrapped carbon nanotubes/in situ formed gold nanoparticles as a novel electrochemical sensing platform. *Mater. Sci. Eng. C* **2015**, *48*, 270–278. [[CrossRef](#)]
171. Pham, X.-H.; Li, C.A.; Han, K.N.; Huynh-Nguyen, B.-C.; Le, T.-H.; Ko, E.; Kim, J.H.; Seong, G.H. Electrochemical detection of nitrite using urchin-like palladium nanostructures on carbon nanotube thin film electrodes. *Sens. Actuators B Chem.* **2014**, *193*, 815–822. [[CrossRef](#)]
172. Wu, W.; Li, Y.; Jin, J.; Wu, H.; Wang, S.; Ding, Y.; Ou, J. Sensing nitrite with a glassy carbon electrode modified with a three-dimensional network consisting of Ni7S6 and multi-walled carbon nanotubes. *Microchim. Acta* **2016**, *183*, 3159–3166. [[CrossRef](#)]
173. Rao, D.; Sheng, Q.; Zheng, J. Self-assembly preparation of gold nanoparticle decorated 1-pyrenemethylamine functionalized graphene oxide-carbon nanotube composites for highly sensitive detection of nitrite. *Anal. Methods* **2016**, *8*, 4926–4933. [[CrossRef](#)]
174. Salimi, A.; Noorbakhash, A.; Karonian, F.S. Amperometric detection of nitrite, iodate and periodate on glassy carbon electrode modified with thionin and multi-wall carbon nanotubes. *Int. J. Electrochem. Sci.* **2006**, *1*, 435–446.
175. Hui, N.; Chai, F.; Lin, P.; Song, Z.; Sun, X.; Li, Y.; Niu, S.; Luo, X. Electrodeposited Conducting Polyaniline Nanowire Arrays Aligned on Carbon Nanotubes Network for High Performance Supercapacitors and Sensors. *Electrochim. Acta* **2016**, *199*, 234–241. [[CrossRef](#)]
176. Lerga, T.M.; O’Sullivan, C.K. Rapid determination of total hardness in water using fluorescent molecular aptamer beacon. *Anal. Chim. Acta* **2008**, *610*, 105–111. [[CrossRef](#)]
177. Gee, K.; Brown, K.; Chen, W.-N.; Bishop-Stewart, J.; Gray, D.; Johnson, I. Chemical and physiological characterization of fluo-4 Ca²⁺-indicator dyes. *Cell Calcium* **2000**, *27*, 97–106. [[CrossRef](#)]
178. Son, D.; Park, S.Y.; Kim, B.; Koh, J.T.; Kim, T.H.; An, S.; Jang, D.; Kim, G.T.; Jhe, W.; Hong, S. Nanoneedle Transistor-Based Sensors for the Selective Detection of Intracellular Calcium Ions. *ACS Nano* **2011**, *5*, 3888–3895. [[CrossRef](#)]
179. Akhter, F.; Nag, A.; Alahi, E.E.; Liu, H.; Mukhopadhyay, S. Electrochemical detection of calcium and magnesium in water bodies. *Sens. Actuators A Phys.* **2020**, *305*, 111949. [[CrossRef](#)]
180. Mulholland, P.J.; Houser, J.N.; Maloney, K.O. Stream diurnal dissolved oxygen profiles as indicators of in-stream metabolism and disturbance effects: Fort Benning as a case study. *Ecol. Indic.* **2005**, *5*, 243–252. [[CrossRef](#)]
181. Sánchez, E.; Colmenarejo, M.F.; Vicente, J.; Rubio, A.; García, M.G.; Travieso, L.; Borja, R. Use of the water quality index and dissolved oxygen deficit as simple indicators of watersheds pollution. *Ecol. Indic.* **2007**, *7*, 315–328. [[CrossRef](#)]
182. Collins, P.G.; Bradley, K.; Ishigami, M.; Zettl, A. Extreme Oxygen Sensitivity of Electronic Properties of Carbon Nanotubes. *Science* **2000**, *287*, 1801–1804. [[CrossRef](#)] [[PubMed](#)]
183. Moonosawmy, K.R.; Kruse, P. Cause and Consequence of Carbon Nanotube Doping in Water and Aqueous Media. *J. Am. Chem. Soc.* **2010**, *132*, 1572–1577. [[CrossRef](#)]
184. Ye, J.-S.; Wen, Y.; De Zhang, W.; Cui, H.-F.; Gan, L.M.; Xu, G.Q.; Sheu, F.-S. Application of multi-walled carbon nanotubes functionalized with hemin for oxygen detection in neutral solution. *J. Electroanal. Chem.* **2004**, *562*, 241–246. [[CrossRef](#)]
185. Shi, C.; Anson, F.C. Catalytic pathways for the electroreduction of oxygen by iron tetrakis(4-N-methylpyridyl)porphyrin or iron tetraphenylporphyrin adsorbed on edge plane pyrolytic graphite electrodes. *Inorg. Chem.* **1990**, *29*, 4298–4305. [[CrossRef](#)]
186. Tsai, T.H.; Yang, C.Y.; Chen, S.M. Development of a dissolved oxygen sensor for commercial applications. *Int. J. Electrochem. Sci.* **2013**, *8*, 5250–5261.
187. Sosna, M.; Denuault, G.; Pascal, R.W.; Prien, R.D.; Mowlem, M. Development of a reliable microelectrode dissolved oxygen sensor. *Sens. Actuators B Chem.* **2007**, *123*, 344–351. [[CrossRef](#)]
188. Balamurugan, J.; Thanh, T.D.; Karthikeyan, G.; Kim, N.H.; Lee, J.H. A novel hierarchical 3D N-Co-CNT@NG nanocomposite electrode for non-enzymatic glucose and hydrogen peroxide sensing applications. *Biosens. Bioelectron.* **2017**, *89*, 970–977. [[CrossRef](#)]
189. Naqvi, S.T.R.; Shirinfar, B.; Hussain, D.; Majeed, S.; Ashiq, M.N.; Aslam, Y.; Ahmed, N. Electrochemical Sensing of Ascorbic Acid, Hydrogen Peroxide and Glucose by Bimetallic (Fe, Ni)–CNTs Composite Modified Electrode. *Electroanalysis* **2019**, *31*, 851–857. [[CrossRef](#)]
190. Annamalai, S.K.; Palani, B.; Pillai, K.C. Highly stable and redox active nano copper species stabilized functionalized-multiwalled carbon nanotube/chitosan modified electrode for efficient hydrogen peroxide detection. *Colloids Surf. A Physicochem. Eng. Asp.* **2012**, *395*, 207–216. [[CrossRef](#)]
191. Yang, L.; Li, M.; Qu, Y.; Dong, Z.; Li, W.J. Carbon nanotube-sensor-integrated microfluidic platform for real-time chemical concentration detection. *Electrophoresis* **2009**, *30*, 3198–3205. [[CrossRef](#)]
192. Wang, J.; Musameh, M. Carbon nanotube screen-printed electrochemical sensors. *Analyst* **2003**, *129*, 1–2. [[CrossRef](#)]
193. Patel, V.; Kruse, P.; Selvaganapathy, P.R. Hydrogen peroxide chemiresistive detection platform with wide range of detection. In Proceedings of the 2019 IEEE SENSORS, Montreal, QC, Canada, 27–30 October 2019; pp. 1–4. [[CrossRef](#)]
194. Olivé-Monllau, R.; Pereira, A.; Bartrolí, J.; Baeza, M.; Céspedes, F. Highly sensitive CNT composite amperometric sensors integrated in an automated flow system for the determination of free chlorine in waters. *Talanta* **2010**, *81*, 1593–1598. [[CrossRef](#)]

195. Isaac, A.; Davis, J.; Livingstone, C.; Wain, A.J.; Compton, R.G. Electroanalytical methods for the determination of sulfite in food and beverages. *TrAC Trends Anal. Chem.* **2006**, *25*, 589–598. [[CrossRef](#)]
196. Raof, J.B.; Ojani, R.; Karimi-maleh, H. Electrocatalytic Determination of Sulfite at the Surface of a New Ferrocene Derivative-Modified Carbon Paste Electrode. *Int. J. Electrochem. Sci.* **2007**, *2*, 257–269.
197. Vu, L.; Cervenka, L. Determination of Sulfide by Hematoxylin Multiwalled Carbon Nanotubes Modified Carbon Paste Electrode. *Electroanalysis* **2013**, *25*, 1967–1973.
198. Mohajeri, S.; Dolati, A.; Salmani-Rezaie, S. Electrochemical sensors based on functionalized carbon nanotubes modified with platinum nanoparticles for the detection of sulfide ions in aqueous media. *J. Chem. Sci.* **2019**, *131*, 21. [[CrossRef](#)]
199. Lawrence, N.S.; Deo, R.P.; Wang, J. Electrochemical determination of hydrogen sulfide at carbon nanotube modified electrodes. *Anal. Chim. Acta* **2004**, *517*, 131–137. [[CrossRef](#)]
200. Shamsipur, M.; Roushani, M.; Pourmortazavi, S.M.; Shahabadi, N. Amperometric determination of sulfide ion by glassy carbon electrode modified with multiwall carbon nanotubes and copper (II) phenanthroline complex. *Open Chem.* **2014**, *12*, 1091–1099. [[CrossRef](#)]
201. Perales, E.; García, C.; Lomba, L.; García, J.L.; Pires, E.; Sancho, M.C.; Navarro, E.; Giner, B. Comparative ecotoxicity study of glycerol-biobased solvents. *Environ. Chem.* **2017**, *14*, 370–377. [[CrossRef](#)]
202. Cheng, Y.; Liu, Y.; Huang, J.; Xian, Y.; Zhang, W.; Zhang, Z.; Jin, L. Rapid amperometric detection of coliforms based on MWNTs/Nafion composite film modified glass carbon electrode. *Talanta* **2008**, *75*, 167–171. [[CrossRef](#)] [[PubMed](#)]
203. Khalil, M.M.; Farghali, A.A.; El Roubi, W.M.A.; Abd-Elgawad, I.H. Preparation and characterization of novel MWCNTs/Fe-Co doped TNTs nanocomposite for potentiometric determination of sulphuric acid in real water samples. *Sci. Rep.* **2020**, *10*, 8607. [[CrossRef](#)] [[PubMed](#)]
204. Ali, Y.; Alam, A.U.; Howlader, M.M. Fabrication of highly sensitive Bisphenol A electrochemical sensor amplified with chemically modified multiwall carbon nanotubes and β -cyclodextrin. *Sens. Actuators B Chem.* **2020**, *320*, 128319. [[CrossRef](#)]
205. Zhou, T.; Zhao, X.; Xu, Y.; Tao, Y.; Luo, D.; Hu, L.; Jing, T.; Zhou, Y.; Wang, P.; Mei, S. Electrochemical determination of tetrabromobisphenol A in water samples based on a carbon nanotubes@zeolitic imidazole framework-67 modified electrode. *RSC Adv.* **2020**, *10*, 2123–2132. [[CrossRef](#)]
206. Gao, Y.; Cao, Y.; Yang, D.; Luo, X.; Tang, Y.; Li, H. Sensitivity and selectivity determination of bisphenol A using SWCNT-CD conjugate modified glassy carbon electrode. *J. Hazard. Mater.* **2012**, *199–200*, 111–118. [[CrossRef](#)]
207. Dey, S.; Bano, F.; Malik, A. 1—Pharmaceuticals and personal care product (PPCP) contamination—A global discharge inventory. In *Pharmaceuticals and Personal Care Products: Waste Management and Treatment Technology*; Prasad, M.N.V., Vithanage, M., Kapley, A., Eds.; Butterworth-Heinemann: Oxford, UK; pp. 1–26.
208. Anzar, N.; Hasan, R.; Tyagi, M.; Yadav, N.; Narang, J. Carbon nanotube—A review on Synthesis, Properties and plethora of applications in the field of biomedical science. *Sens. Int.* **2020**, *1*, 100003. [[CrossRef](#)]
209. Benda, R.; Vezin, T.; Zucchi, G.; Cancès, E.; Lebental, B. Prediction of the interaction strength of an urea-based probe towards ions in water by means of DFT/PCM calculations. *J. Comput.-Aided Mol. Des.* **2021**. Preprint.
210. Star, A.; Joshi, V.; Skarupo, S.; Thomas, D.; Gabriel, J.-C.P. Gas Sensor Array Based on Metal-Decorated Carbon Nanotubes. *J. Phys. Chem. B* **2006**, *110*, 21014–21020. [[CrossRef](#)] [[PubMed](#)]
211. Fitzgerald, J.; Fenniri, H. Cutting Edge Methods for Non-Invasive Disease Diagnosis Using E-Tongue and E-Nose Devices. *Biosensors* **2017**, *7*, 59. [[CrossRef](#)]
212. Daikuzono, C.M.; Dantas, C.A.R.; Volpati, D.; Constantino, C.J.L.; Piazzetta, M.H.O.; Gobbi, A.L.; Taylor, D.M.; Oliveira, O.N.; Riul, A. Microfluidic electronic tongue. *Sens. Actuators B Chem.* **2015**, *207*, 1129–1135. [[CrossRef](#)]
213. Lebental, B.; Benda, R.; Bodelot, L.; Florea, I.; Godumala, M.; Gusarov, B.; Gutierrez, A.; Loisel, L.; Merliot, E.; Ramachandran, S.; et al. Carbon nanotube sensor array for water monitoring with conjugated polymers. In *Proceedings of the C’NANO 2017, The Nanoscience Meeting, Lyon, France, 5–7 December 2017*; p. 18.
214. Shaun, F.; Pellegrino, M.; Cesar, W.; Marty, F.; Xu, Z.; Capo-Chichi, M.; Basset, P.; Lebental, B.; Bourouina, T. Robust multi-parameter sensing probe for water monitoring based on ALD-coated metallic micro-patterns. In *Proceedings of the 2016 IEEE 11th Annual International Conference on Nano/Micro Engineered and Molecular Systems (NEMS), Sendai, Japan, 17–20 April 2016*; pp. 342–345. [[CrossRef](#)]