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# Straightforward Access to Enantioenriched *cis*-3-Fluoro-Dihydroquinolin-4-ols Derivatives *via* Ru(II)-Catalyzed-Asymmetric Transfer Hydrogenation/Dynamic Kinetic Resolution

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**Abstract:** We herein report a practical method for the asymmetric transfer hydrogenation/dynamic kinetic resolution of *N*-Boc 3-fluoro-dihydro-tetrahydroquinolin-4-ones into the corresponding *cis*-fluoro alcohols in 70–96% yields, up to 99:1 dr and up to >99% ee by using the ruthenium complex Ts-DENEB and a formic acid/triethylamine (1:1) mixture as the hydrogen donor under mild conditions.

**Keywords:** Asymmetric Catalysis, Reduction, Fluorine, Hydrogen Transfer, Ruthenium

Dedicated to Professor H. B. Kagan for his outstanding contribution to asymmetric catalysis and organometallic chemistry

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## 1. Introduction

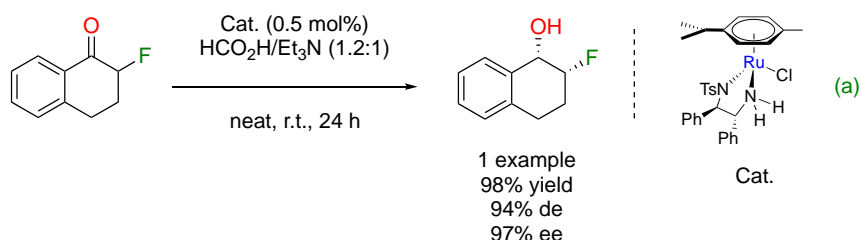
The number of bioactive molecules containing fluorine approved by the Food and Drug Administration FDA is in large increase as time passes by. The first decade of this century saw the introduction of 40 new compounds having a fluorine atom. This number represented at the time an important increase of 20% of the commercial drugs bearing at least one fluorine atom in their structure [1]. Compared to this data, exclusively in 2018, 17 fluorine-containing pharmaceuticals were approved by the FDA [2]. In a single year the number of new fluorinated drugs covered almost half of the increase of a whole decade.

This fluorine rush is easily explained by its ability to affect important parameters for a drug candidate such as permeability or pK<sub>a</sub> and to modify its pharmacokinetics and pharmacodynamics. Fluorine is thus an element that gives control to medicinal chemists over tailoring the properties of a molecule. Organic chemists must then work in consideration of this increasing demand and develop new methodologies to introduce fluorine into complex molecules and into versatile building blocks [3].

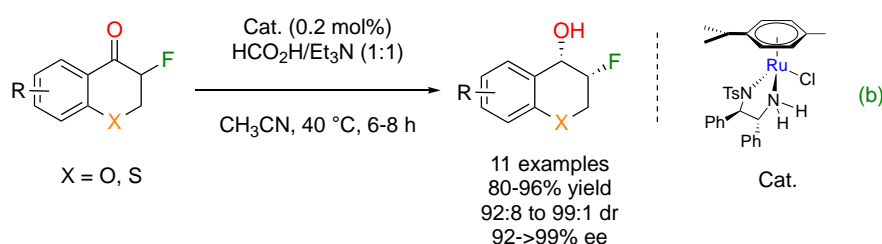
The tetrahydroquinoline core is a very common scaffold found in several biologically active molecules and therapeutic agents [4]. Introducing a fluorine atom in its structure would bring a novelty into the medicinal chemist toolbox and in theory open up its use in the screening and development of new biologically active molecules. Asymmetric transfer hydrogenation (ATH) has been reported by Lassaletta and co-workers [5] as a method to access enantiomerically enriched fluoro-tetralol *via* a dynamic kinetic resolution (DKR) process (Scheme 1; (a)). Following the premise of the fluorine need and the previous works in the laboratory including the ATH/DKR of 3-fluoro-chromanone derivatives [6] (Scheme 1; (b)), we herein report an asymmetric transfer hydrogenation/dy-

namic kinetic resolution [7] (ATH/DKR) of *tert*-butyl 3-fluoro-4-oxo-3,4-dihydroquinoline-1(2*H*)-carboxylates into the corresponding enantioenriched *cis*-fluorohydrins (Scheme 1; (c)).

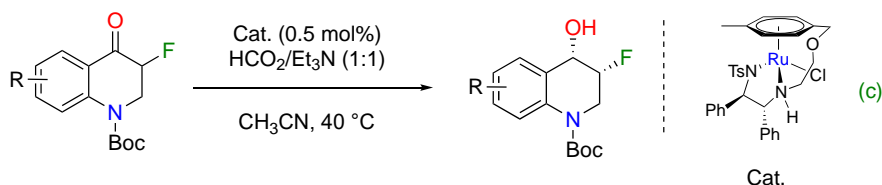
Lassaletta and co-workers:



Our previous work:



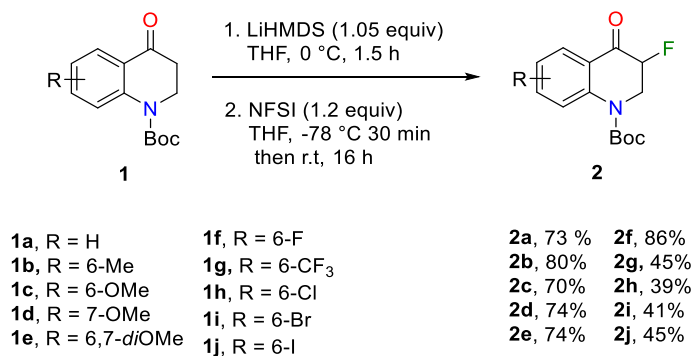
This work:



Scheme 1.

## 2. Results and Discussion

The study began with the synthesis of the fluorinated dihydroquinolin-4-ones **2a-j** via an electrophilic fluorination of the *N*-Boc-protected heterocyclic ketones **1a-j** by using lithium bis(trimethylsilyl)amide as a base and NFSI as the fluorine source (Scheme 2).

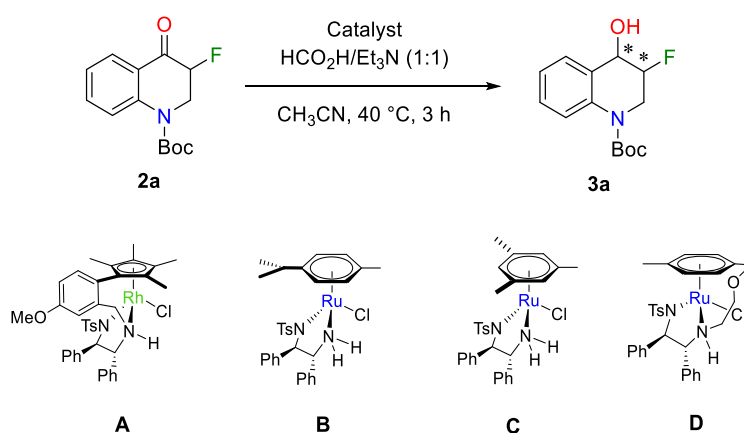


Scheme 2.

3-Fluoro-dihydroquinolin-4-one carboxylate **2a** was chosen as the standard substrate for the optimization of the asymmetric transfer hydrogenation. Based on previous studies, the ATH was set at 40 °C in acetonitrile with a (1:1) molar mixture of formic acid and

triethylamine as the hydrogen source. Under these conditions, a set of commercially available Ru(II) catalysts and a Rh(III) complex were screened (Table 1). After 3 h with a catalyst loading of 0.5 mol%, the Ru and Rh catalysts allowed a full conversion of the  $\alpha$ -fluoro ketone into the corresponding fluoro alcohol **3a**. All the complexes delivered excellent isolated yields and enantiomeric excesses for the desired fluoroalcohol **3a** (entries 1 to 4). The main difference appeared in the diastereoselectivity outcome. Whereas (*R,R*)-Ru(*p*-cymene)TsDPEN ((*R,R*)-**B**; entry 2) led to a moderate 79:21 dr, (*R,R*)-Rh-*teth*-TsDPEN ((*R,R*)-**A**; entry 1), (*R,R*)-Ru(mesitylene)TsDPEN ((*R,R*)-**C**; entry 3) and (*R,R*)-TsDENEb ((*R,R*)-**D**; entry 4) all gave an excellent diastereoisomeric ratio dr of 97:3 or 98:2 pointing out that the reaction was not affected by the nature of the arene ligand nor the tethered or untethered characteristic of the complex. From this series of results, the (*R,R*)-TsDENEb ((*R,R*)-**D**) catalyst was chosen for the optimization conditions.

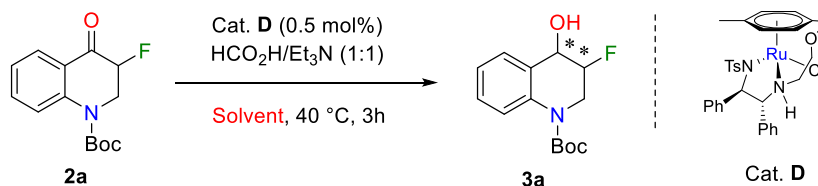
**Table 1.** Catalyst screening for the ATH/DKR<sup>a</sup>



Entry	Catalyst	Yield <sup>b</sup>	dr <sup>c</sup>	ee <sup>d</sup>
1	<b>A</b>	93%	97:3	99%
2	<b>B</b>	95%	79:21	95%
3	<b>C</b>	93%	97:3	98%
4	<b>D</b>	94%	98:2	>99%

<sup>a</sup>Conditions: **2a** (0.32 mmol), catalyst (0.5 mol%), HCO<sub>2</sub>H/Et<sub>3</sub>N (1:1) (6 equiv), CH<sub>3</sub>CN (1 mL), 40 °C, 3 h. <sup>b</sup>Isolated yield. <sup>c</sup>Determined by <sup>1</sup>H NMR analysis of the crude mixture. <sup>d</sup>Determined by SFC analysis.

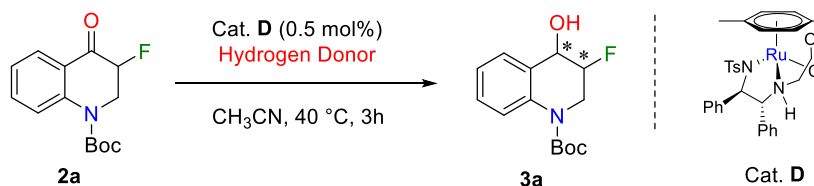
The solvent of the asymmetric transfer hydrogenation was then investigated. By using a catalyst loading of 0.5 mol% of (*R,R*)-**D**, formic acid/triethylamine (1:1) mixture and a reaction time of 3 h, we tested a series of different solvents (Table 2). Several polar aprotic (CH<sub>3</sub>CN, CH<sub>2</sub>Cl<sub>2</sub>, THF, Me-THF and EtOAc) and polar protic (MeOH, *i*PrOH and HFIP) as well as aromatic solvents were screened (toluene, chlorobenzene and trifluoromethylbenzene). The diastereo- and enantioselectivities remained mostly unchanged in all the tests. The conversion rate and the yield showed to be highly dependent on the solvent used. Isopropanol and acetonitrile led to 95% and 100% conversions respectively with excellent stereoselectivities in both solvents, acetonitrile slightly surpassing isopropanol in all of them. This prompted us to set acetonitrile as the solvent for the asymmetric transfer hydrogenation.

**Table 2.** Solvent optimization<sup>a</sup>

Entry	Solvent	Conversion <sup>b</sup>	Yield <sup>c</sup>	dr <sup>b</sup>	ee <sup>d</sup>
1	CH <sub>3</sub> CN	100%	93%	98:2	>99%
2	CH <sub>2</sub> Cl <sub>2</sub>	83%	78%	98:2	95%
3	EtOAc	48%	46%	98:2	98%
4	THF	38%	36%	98:2	>99%
5	Me-THF	18%	15%	96:4	98%
6	Toluene	26%	26%	97:3	96%
7	Cl-benzene	43%	42%	97:3	94%
8	CF <sub>3</sub> -benzene	60%	57%	98:2	96%
9	MeOH	76%	73%	98:2	96%
10	<i>i</i> PrOH	95%	94%	98:2	98%
11	HFIP	26%	24%	98:2	>99%

<sup>a</sup>Conditions: **2a** (0.32 mmol), catalyst (0.5 mol%), HCO<sub>2</sub>H/Et<sub>3</sub>N (1:1) (6 equiv), solvent (1 mL), 40 °C, 3 h. <sup>b</sup>Determined by <sup>1</sup>H NMR analysis of the crude mixture. <sup>c</sup>Isolated yield. <sup>d</sup>Determined by SFC analysis.

We continued the optimization by varying the hydrogen source of the reaction (Table 3). The acid to base ratio was examined (entries 1 to 4). Changing the formic acid to triethylamine ratio from 1:1 to 2:5 (entry 2), showed no impact on the reaction. However, switching to a HCO<sub>2</sub>H/Et<sub>3</sub>N ratio of 5:2 (entry 3), was detrimental to the diastereoselectivity which dropped to a 54:46 dr. Moreover, when this ratio was further increased to 12:1 (entry 4), no conversion was observed. This strong influence of the acid/base ratio of the hydrogen donor reflects the one that was observed in the ATH/DKR of 3-fluorochromanones [6]. Other organic bases associated to formic acid (DBU, entry 5 and DABCO, entry 6) as well as formate salts (ammonium formate and calcium formate, entries 7 and 8 respectively) were screened, all giving lower conversions and similar stereoselectivities than Et<sub>3</sub>N. Finally, the catalyst loading (0.2 mol%; entry 9) and the reaction temperature (room temperature; entry 10) were lowered. Both tests gave lower conversions in spite of longer reaction times so we set the optimized conditions for the ATH/DKR as 0.5 mol% of (*R,R*)-**D** as the catalyst, HCO<sub>2</sub>H/Et<sub>3</sub>N (1:1) mixture as the hydrogen donor, acetonitrile as the solvent and 40 °C as the reaction temperature.

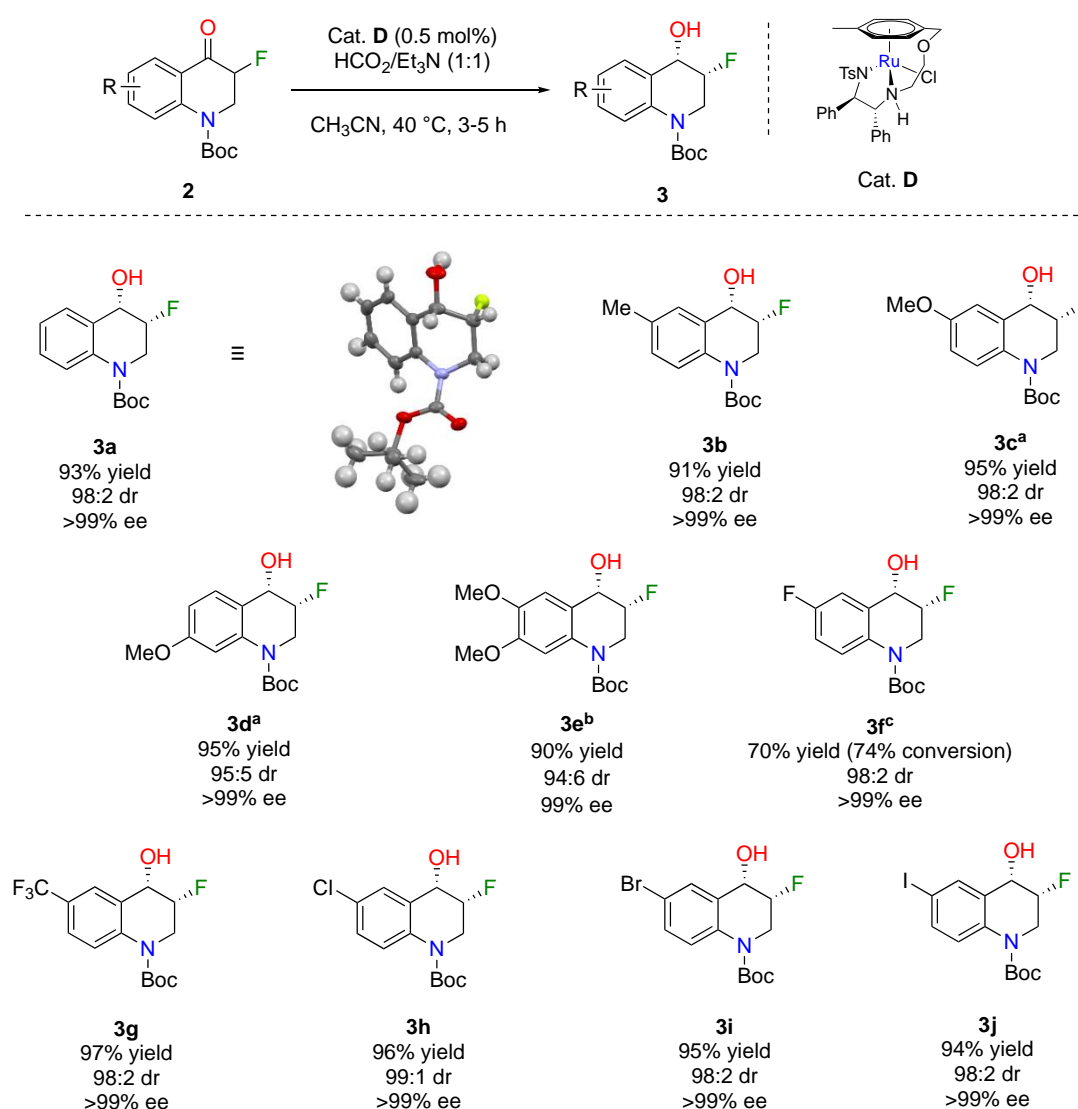
**Table 3.** Survey of the hydrogen donor<sup>a</sup>

Entry	Hydrogen donor	Conversion	Yield <sup>b</sup>	dr <sup>c</sup>	ee <sup>d</sup>
1	HCO <sub>2</sub> H/Et <sub>3</sub> N (1:1)	100%	93%	98:2	>99%
2	HCO <sub>2</sub> H/Et <sub>3</sub> N (2:5)	100%	95%	98:2	99%
3	HCO <sub>2</sub> H/Et <sub>3</sub> N (5:2)	100%	94%	54:46	92%
4	HCO <sub>2</sub> H/Et <sub>3</sub> N (12:1)	0%	-	-	-
5	HCO <sub>2</sub> H/DBU (1:1)	90%	88%	98:2	99%
6	HCO <sub>2</sub> H/DABCO (1:1)	75%	73%	98:2	98%
7	HCO <sub>2</sub> NH <sub>4</sub>	35%	34%	98:2	99%
8 <sup>e</sup>	(HCO <sub>2</sub> ) <sub>2</sub> Ca	20%	16%	98:2	98%
9 <sup>f</sup>	HCO <sub>2</sub> H/Et <sub>3</sub> N (1:1)	54%	53%	98:2	>99%
10 <sup>g</sup>	HCO <sub>2</sub> H/Et <sub>3</sub> N (1:1)	30%	23%	98:2	>99%

<sup>a</sup>Conditions: **2a** (0.32 mmol), catalyst (0.5 mol%), HCO<sub>2</sub>H/Et<sub>3</sub>N (1:1) (6 equiv), CH<sub>3</sub>CN (1 mL), 40 °C, 3 h. <sup>b</sup>Isolated yield. <sup>c</sup>Determined by <sup>1</sup>H NMR analysis of the crude mixture. <sup>d</sup>Determined by SFC analysis. <sup>e</sup>CH<sub>3</sub>CN/H<sub>2</sub>O (10:1) was used as solvent. <sup>f</sup>Catalyst loading of 0.2 mol% and reaction time of 24 h. <sup>g</sup>Reaction run at r. t. (22 °C) for 24 h.

Once the aforementioned conditions were identified, we subjected the family of substituted *tert*-butyl 3-fluoro dihydroquinolin-4-ones carboxylates **2a-j** previously synthesized to the asymmetric transfer hydrogenation. The scope (Scheme 3) showed that the fluoro dihydrotetrahydroquinolin-4-ones bearing electron-donating or electron-withdrawing groups all led in good yields (90–96%), diastereo- (94:6 to 99:1 dr) and enantioselectivities (99–>99% ee) to the corresponding fluorohydrins **3a-j**. Notably, the asymmetric transfer hydrogenation tolerated substituents such as chlorine, bromine or iodine along with other groups such as methyl, methoxy and trifluoromethyl. Concerning the outcome of the reaction, the two substrates bearing a methoxy- group in position 7 of the aromatic ring (**2d** and **2e**) gave a slightly lower diastereoselectivity compared to the rest of the dihydroquinolin-4-ones. The ATH of compound **2e**, possessing two methoxy groups in positions 6 and 7, also required a higher catalyst loading (1.0 mol%) and a longer reaction time (24 h) to achieve a full conversion.

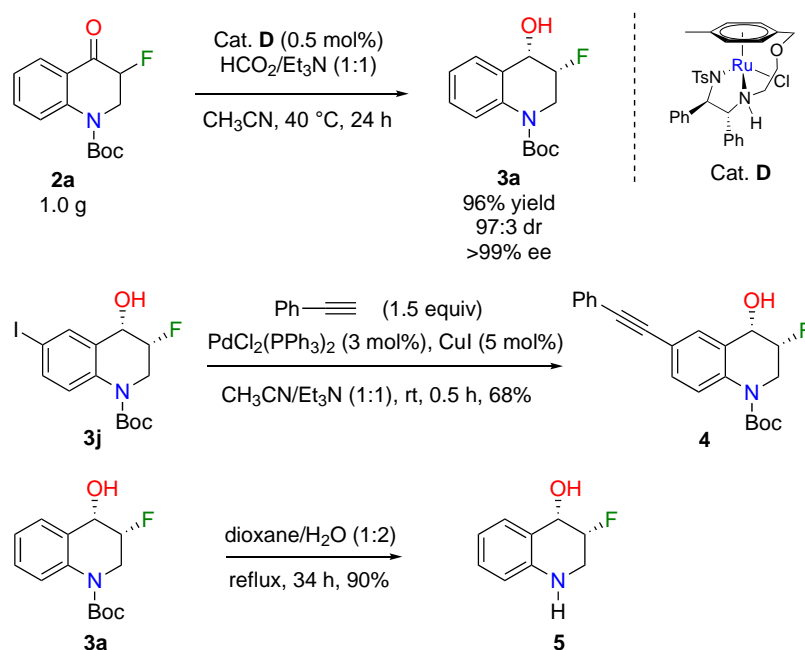
The absolute configuration of compound **3a** was unambiguously assigned as (3*R*, 4*S*) by the X-ray crystallographic analysis. We conjectured that the other products of the developed ATH/DKR reaction led to 3-fluoro-dihydrotetrahydroquinolin-4-ols derivatives with the same absolute configuration (Scheme 3).



Unless specified, the reaction time is of 3 h. <sup>a</sup>Reaction time of 5 h. <sup>b</sup>1.0 mol% of catalyst and reaction time of 24 h. <sup>c</sup>Reaction time of 24 h.

### Scheme 3. Scope of the reaction

The usefulness of the studied asymmetric transfer hydrogenation was finally evaluated by performing the reaction on a gram-scale with substrate **2a** (Scheme 4). 1.0 g of *tert*-butyl 3-fluoro-dihydro-2,3,4,5-tetrahydroquinolin-4-one carboxylate was subjected to the optimized reaction conditions which led to the *cis*-fluorohydrin **3a** in 96% yield, 97:3 dr and >99% ee. In addition, the post-functionalization of compound **3j** using a Sonogashira cross-coupling reaction with phenylacetylene was carried out in the presence of  $\text{PdCl}_2(\text{PPh}_3)_2$  and CuI to afford the corresponding alkyne **4** in 68% yield. Finally, *N*-Boc deprotection of compound **3a** was readily achieved by heating in dioxane/ $\text{H}_2\text{O}$  and the desired amine **5** was isolated in 90% yield.



Scheme 4. Scale-up, functionalization and deprotection experiments

### 3. Conclusion

In summary, a straightforward method to access enantiomerically and diastereomerically enriched *cis*-fluoro-dihydrotetrahydroquinolinols has been developed. Ru catalyst (*R,R*)-TsDENEb ensures an efficient asymmetric transfer hydrogenation/dynamic kinetic resolution (ATH/DKR) under mild conditions (acetonitrile as solvent and heating at 40 °C) and low catalyst loading (0.5 mol%). The reaction was performed successfully on a series of 3-fluoro-dihydrotetrahydroquinolin-4-ones substituted with several electron-donating or electron-withdrawing groups with excellent yields, diastereo- and enantioselectivities.

### 4. Materials and Methods

#### 4.1 General Information

All air and/or water sensitive reactions were carried out under an argon atmosphere. THF, CH<sub>2</sub>Cl<sub>2</sub> and toluene were dried over alumina columns in a solvent purification apparatus (Innovative technology). Methanol, isopropanol, chlorobenzene, trifluoromethylbenzene, ethyl acetate and acetonitrile from Sigma-Aldrich were used without further purification. Hexafluoroisopropanol (HFIP) was purchased from TCI and was used without further purification. Formic acid/triethylamine (1:1) mixture was purchased from Fluka or Alfa Aesar and was used without further purification. Reactions were monitored by thin layer chromatography carried out on precoated silica gel plates (Merck 60F254) and revealed with either an ultra-violet lamp ( $\lambda = 254\text{ nm}$ ) or a potassium permanganate solution. Proton nuclear magnetic resonance (<sup>1</sup>H NMR) spectra were recorded using a Bruker AC 400 (400 MHz). The chemical shifts are expressed in parts per million (ppm) referenced to residual chloroform (7.26 ppm). Data are reported as follows: chemical shifts ( $\delta$ ), multiplicity (recorded as s, singlet; d, doublet; t, triplet; q, quadruplet; quint, quintuplet; sext, sextuplet; hept, heptuplet; m, multiplet and br, broad), coupling constants and integration. Carbon-13 nuclear magnetic resonance (<sup>13</sup>C NMR) spectra were recorded using a Bruker AC 400 (101 MHz). The chemical shifts are expressed in parts per million (ppm) relative to the centre line of the triplet at 77.16 ppm for CDCl<sub>3</sub>. Melting points (m.



p.) were determined on a K f ler melting point apparatus. Optical rotations were measured on a Jasco P-1010 polarimeter. High resolution mass spectrometric (HRMS) analyses were measured on LTQ-Orbitrap (Thermo Fisher Scientific) at Sorbonne Universit .

#### 4.2 Method for the Synthesis of Compounds 2

Adapted from [8]

To a 0  C solution of LiHMDS (1.0 M in THF, 1.05 equiv) in a round-bottom tube set under argon, was slowly added a solution of the ketone (1.2 mmol, 1.0 equiv) in THF (2 mL) over 5 min. The mixture was stirred at 0  C for 1.5 h. The resulting solution was then added dropwise over 5 minutes *via* a cannula to a -78  C solution of NFSI (1.4 mmol, 1.2 equiv) in THF (4 mL). The reaction mixture was stirred at -78  C for 30 min then allowed to come to room temperature overnight. The reaction was diluted with 3 mL of CH    , quenched with 5 mL of saturated NH     aqueous solution and extracted with CH     (3 x 10 mL). The combined organic layers were washed with brine, dried over MgSO , filtered and concentrated under reduced pressure. The pure products were isolated by flash column chromatography on silica gel.

##### ***Tert*-butyl 3-fluoro-4-oxo-3,4-dihydroquinoline-1(2*H*)-carboxylate (2a):**

2.0 g of *tert*-butyl 4-oxo-3,4-dihydroquinoline-1(2*H*)-carboxylate (8.09 mmol, 1.0 equiv) were used following the described procedure. Purification via flash column chromatography on silica gel (95:5 petroleum ether/ethyl acetate) yielded 1.56 g of **2a** as a white solid (73%). m.p. 96-100  C. <sup>1</sup>H NMR (400 MHz, Chloroform-*d*)   8.03 (dd, *J* = 8.0, 1.6 Hz, 1H), 7.81 (dd, *J* = 8.5, 1.0 Hz, 1H), 7.54 (ddd, *J* = 8.7, 7.2, 1.7 Hz, 1H), 7.20 (td, *J* = 7.6, 1.0 Hz, 1H), 5.10 (ddd, *J* = 46.9, 10.5, 4.7 Hz, 1H), 4.63 (td, *J* = 13.0, 4.7 Hz, 1H), 4.04 (ddd, *J* = 13.3, 10.6, 4.8 Hz, 1H), 1.57 (s, 9H). <sup>19</sup>F NMR (376 MHz, Chloroform-*d*)   -198.77. <sup>13</sup>C NMR (101 MHz, Chloroform-*d*)   189.7 (C-F, <sup>2</sup>*J*<sub>CF</sub> = 14.1 Hz), 189.6 (C-F, <sup>2</sup>*J*<sub>CF</sub> = 14.1 Hz), 152.8, 144.1, 135.0, 128.1, 124.6, 123.8, 123.2, 88.1 (C-F, <sup>1</sup>*J*<sub>CF</sub> = 190.9 Hz), 86.2 (C-F, <sup>1</sup>*J*<sub>CF</sub> = 190.9 Hz), 83.2, 48.8 (C-F, <sup>2</sup>*J*<sub>CF</sub> = 27.3 Hz), 48.5 (C-F, <sup>2</sup>*J*<sub>CF</sub> = 27.3 Hz), 28.3. HRMS (APCI): *m/z* [M + H]<sup>+</sup> calcd for C<sub>14</sub>H<sub>17</sub>FNO<sub>3</sub> 266.1187, found 266.1186.

##### ***Tert*-butyl 3-fluoro-6-methyl-4-oxo-3,4-dihydroquinoline-1(2*H*)-carboxylate (2b):**

550 mg of *tert*-butyl 6-methyl-4-oxo-3,4-dihydroquinoline-1(2*H*)-carboxylate (2.10 mmol, 1.0 equiv) were used following the described procedure. Purification via flash column chromatography on silica gel (95:5 petroleum ether/ethyl acetate) yielded 470 mg of **2b** as a white solid (80%). m.p. 122-126  C. <sup>1</sup>H NMR (400 MHz, Chloroform-*d*)   7.75 (s, 1H), 7.62 (d, *J* = 8.5 Hz, 1H), 7.28 (dd, *J* = 9.8, 3.4 Hz, 1H), 5.01 (ddd, *J* = 46.9, 10.6, 4.7 Hz, 1H), 4.55 (tdd, *J* = 13.2, 4.7, 0.9 Hz, 1H), 3.93 (ddd, *J* = 13.5, 10.6, 4.8 Hz, 1H), 2.28 (s, 3H), 1.49 (s, 9H). <sup>19</sup>F NMR (376 MHz, Chloroform-*d*)   -198.75. <sup>13</sup>C NMR (101 MHz, Chloroform-*d*)   189.9 (C-F, <sup>2</sup>*J*<sub>CF</sub> = 14.1 Hz), 189.8 (C-F, <sup>2</sup>*J*<sub>CF</sub> = 14.1 Hz), 152.9, 141.8, 136.0, 134.5, 127.8, 123.7, 123.0, 88.2 (C-F, <sup>1</sup>*J*<sub>CF</sub> = 190.9 Hz), 86.3 (C-F, <sup>1</sup>*J*<sub>CF</sub> = 190.9 Hz), 83.0, 48.8 (C-F, <sup>2</sup>*J*<sub>CF</sub> = 27.3 Hz), 48.5 (C-F, <sup>2</sup>*J*<sub>CF</sub> = 27.3 Hz), 28.4, 20.7. HRMS (APCI): *m/z* [M - Boc + H]<sup>+</sup> calcd for C<sub>10</sub>H<sub>11</sub>FNO 180.0825, found 180.0819.

##### ***Tert*-butyl 3-fluoro-6-methoxy-4-oxo-3,4-dihydroquinoline-1(2*H*)-carboxylate (2c):**

700 mg of *tert*-butyl 6-methoxy-4-oxo-3,4-dihydroquinoline-1(2*H*)-carboxylate (2.52 mmol, 1.0 equiv) were used following the described procedure. Purification via flash column chromatography on silica gel (95:5 to 85:15 petroleum ether/ethyl acetate) yielded 487 mg of **2c** as a pale yellow solid (70%). m.p. 136-140  C. <sup>1</sup>H NMR (400 MHz, Chloroform-*d*)   7.69 (dd, *J* = 9.7, 2.9 Hz, 1H), 7.43 (dd, *J* = 7.0, 4.2 Hz, 1H), 7.11 (dt, *J* = 12.5, 8.2 Hz, 1H), 5.08 (ddd, *J* = 46.8, 17.6, 13.6 Hz, 1H), 4.60 (td, *J* = 13.2, 8.9 Hz, 1H), 4.01 (ddt, *J* =

15.3, 10.6, 3.2 Hz, 1H), 3.82 (s, 3H), 1.54 (s, 9H).  $^{19}\text{F}$  NMR (376 MHz, Chloroform-*d*)  $\delta$  –198.57.  $^{13}\text{C}$  NMR (101 MHz, Chloroform-*d*)  $\delta$  189.7 (C-F,  $^2J_{\text{CF}}$  = 15.2 Hz), 189.5 (C-F,  $^2J_{\text{CF}}$  = 15.2 Hz), 156.5, 152.9, 137.9, 125.5, 124.0, 123.5, 108.9, 88.7 (C-F,  $^1J_{\text{CF}}$  = 190.9 Hz), 86.4 (C-F,  $^1J_{\text{CF}}$  = 190.9 Hz), 82.9, 55.8, 48.9 (C-F,  $^2J_{\text{CF}}$  = 27.3 Hz), 48.7 (C-F,  $^2J_{\text{CF}}$  = 27.3 Hz), 28.3. HRMS (APCI):  $m/z$  [M – Boc + H] $^+$  calcd for  $\text{C}_{10}\text{H}_{11}\text{FNO}_2$  196.0774, found 196.0769.

***Tert*-butyl 3-fluoro-7-methoxy-4-oxo-3,4-dihydroquinoline-1(2H)-carboxylate (2d):**

313 mg of *tert*-butyl 7-methoxy-4-oxo-3,4-dihydroquinoline-1(2H)-carboxylate (1.13 mmol, 1.0 equiv) were used following the described procedure. Purification via flash column chromatography on silica gel (95:5 to 85:15 petroleum ether/ethyl acetate) yielded 247 mg of **2d** as a pale yellow solid (74%). m.p. 93–97 °C.  $^1\text{H}$  NMR (400 MHz, Chloroform-*d*)  $\delta$  7.98 (dd,  $J$  = 9.0, 1.3 Hz, 1H), 7.37 (d,  $J$  = 2.4 Hz, 1H), 6.74 (dd,  $J$  = 8.9, 2.4 Hz, 1H), 5.04 (ddd,  $J$  = 47.0, 10.3, 4.6 Hz, 1H), 4.58 (td,  $J$  = 13.4, 4.6 Hz, 1H), 4.03 (ddd,  $J$  = 13.2, 10.3, 5.2 Hz, 1H), 3.88 (s, 3H), 1.58 (s, 9H).  $^{19}\text{F}$  NMR (376 MHz, Chloroform-*d*)  $\delta$  –199.06.  $^{13}\text{C}$  NMR (101 MHz, Chloroform-*d*)  $\delta$  188.3 (C-F,  $^2J_{\text{CF}}$  = 15.2 Hz), 188.1 (C-F,  $^2J_{\text{CF}}$  = 15.2 Hz), 165.1, 152.7, 146.0, 130.2, 116.9, 112.0, 107.7, 87.8 (C-F,  $^1J_{\text{CF}}$  = 189.9 Hz), 85.9 (C-F,  $^1J_{\text{CF}}$  = 189.9 Hz), 83.2, 55.9, 48.9 (C-F,  $^2J_{\text{CF}}$  = 27.3 Hz), 48.7 (C-F,  $^2J_{\text{CF}}$  = 27.3 Hz), 28.4. HRMS (APCI):  $m/z$  [M + H] $^+$  calcd for  $\text{C}_{15}\text{H}_{19}\text{FNO}_4$  296.1293, found 296.1295.

***Tert*-butyl 3-fluoro-6,7-dimethoxy-4-oxo-3,4-dihydroquinoline-1(2H)-carboxylate (2e):**

330 mg of *tert*-butyl 6,7-dimethoxy-4-oxo-3,4-dihydroquinoline-1(2H)-carboxylate (1.07 mmol, 1.0 equiv) were used following the described procedure. Purification via flash column chromatography on silica gel (90:10 to 80:20 petroleum ether/ethyl acetate) yielded 257 mg of **2e** as a pale yellow solid (74%). m.p. 168–172 °C.  $^1\text{H}$  NMR (400 MHz, Chloroform-*d*)  $\delta$  7.46 – 7.36 (m, 2H), 5.05 (ddd,  $J$  = 47.0, 10.1, 4.5 Hz, 1H), 4.56 (ddd,  $J$  = 14.3, 13.2, 4.5 Hz, 1H), 4.06 (ddd,  $J$  = 13.3, 10.1, 5.4 Hz, 1H), 3.96 (s, 3H), 3.91 (s, 3H), 1.57 (s, 9H).  $^{19}\text{F}$  NMR (376 MHz, Chloroform-*d*)  $\delta$  –198.64.  $^{13}\text{C}$  NMR (101 MHz, Chloroform-*d*)  $\delta$  188.3 (C-F,  $^2J_{\text{CF}}$  = 15.2 Hz), 188.1 (C-F,  $^2J_{\text{CF}}$  = 15.2 Hz), 154.8, 152.8, 146.6, 140.0, 116.2, 108.0, 106.3, 87.9 (C-F,  $^1J_{\text{CF}}$  = 188.9 Hz), 86.0 (C-F,  $^1J_{\text{CF}}$  = 188.9 Hz), 83.0, 56.4, 56.3, 49.4 (C-F,  $^2J_{\text{CF}}$  = 27.3 Hz), 49.1 (C-F,  $^2J_{\text{CF}}$  = 27.3 Hz), 28.4. HRMS (APCI):  $m/z$  [M + H] $^+$  calcd for  $\text{C}_{16}\text{H}_{21}\text{FNO}_5$  326.1398, found 326.1399.

***Tert*-butyl 3,6-difluoro-4-oxo-3,4-dihydroquinoline-1(2H)-carboxylate (2f):**

500 mg of *tert*-butyl 6-fluoro-4-oxo-3,4-dihydroquinoline-1(2H)-carboxylate (1.88 mmol, 1.0 equiv) were used following the described procedure. Purification via flash column chromatography on silica gel (95:5 to 85:15 petroleum ether/ethyl acetate) yielded 474 mg of **2f** as a pale yellow solid (86%). m.p. 89–93 °C.  $^1\text{H}$  NMR (400 MHz, Chloroform-*d*)  $\delta$  7.81 (ddd,  $J$  = 9.7, 4.5, 2.3 Hz, 1H), 7.65 (dt,  $J$  = 8.3, 4.3 Hz, 1H), 7.31 – 7.20 (m, 1H), 5.10 (ddd,  $J$  = 46.7, 10.0, 2.7 Hz, 1H), 4.59 (td,  $J$  = 13.4, 3.8 Hz, 1H), 4.06 (ddd,  $J$  = 12.1, 5.1, 2.5 Hz, 1H), 1.55 (s, 9H). m.p. 89–93 °C.  $^{19}\text{F}$  NMR (376 MHz, Chloroform-*d*)  $\delta$  –116.65, –198.88.  $^{13}\text{C}$  NMR (101 MHz, Chloroform-*d*)  $\delta$  188.8 (C-F,  $^2J_{\text{CF}}$  = 15.2 Hz), 188.6 (C-F,  $^2J_{\text{CF}}$  = 15.2 Hz), 160.5 (C-F,  $^1J_{\text{CF}}$  = 248.5 Hz), 158.1 (C-F,  $^1J_{\text{CF}}$  = 248.5 Hz), 152.7, 140.4, 126.1 (C-F,  $^3J_{\text{CF}}$  = 7.1 Hz), 126.1 (C-F,  $^3J_{\text{CF}}$  = 7.1 Hz), 124.6 (C-F,  $^3J_{\text{CF}}$  = 6.1 Hz), 124.6 (C-F,  $^3J_{\text{CF}}$  = 6.1 Hz), 122.5 (C-F,  $^2J_{\text{CF}}$  = 23.2 Hz), 122.3 (C-F,  $^2J_{\text{CF}}$  = 23.2 Hz), 113.5 (C-F,  $^2J_{\text{CF}}$  = 23.2 Hz), 113.3 (C-F,  $^2J_{\text{CF}}$  = 23.2 Hz), 88.0 (C-F,  $^1J_{\text{CF}}$  = 191.9 Hz), 86.1 (C-F,  $^1J_{\text{CF}}$  = 191.9 Hz), 83.4, 48.8 (C-F,  $^2J_{\text{CF}}$  = 26.3 Hz), 48.5 (C-F,  $^2J_{\text{CF}}$  = 26.3 Hz), 28.3. HRMS (APCI):  $m/z$  [M – Boc + H] $^+$  calcd for  $\text{C}_9\text{H}_8\text{F}_2\text{NO}$  184.0574, found 184.0569.

***Tert*-butyl 3-fluoro-4-oxo-6-(trifluoromethyl)-3,4-dihydroquinoline-1(2H)-carboxylate (2g):**

700 mg of *tert*-butyl 4-oxo-6-(trifluoromethyl)-3,4-dihydroquinoline-1(2*H*)-carboxylate (2.22 mmol, 1.0 equiv) were used following the described procedure. Purification via flash column chromatography on silica gel (95:5 to 85:15 petroleum ether/ethyl acetate) yielded 335 mg of **2g** as a white solid (45%). m.p. 101–105 °C. <sup>1</sup>H NMR (400 MHz, Chloroform-*d*) δ 8.29 (d, *J* = 2.3 Hz, 1H), 8.03 (dd, *J* = 8.9, 0.6 Hz, 1H), 7.75 (dd, *J* = 7.1, 2.8 Hz, 1H), 5.13 (ddd, *J* = 46.7, 10.3, 4.6 Hz, 1H), 4.62 (ddd, *J* = 14.3, 13.6, 4.6 Hz, 1H), 4.12 (ddd, *J* = 13.5, 10.3, 5.1 Hz, 1H), 1.58 (s, 9H). <sup>19</sup>F NMR (376 MHz, Chloroform-*d*) δ −62.76, −199.13. <sup>13</sup>C NMR (101 MHz, Chloroform-*d*) δ 188.4 (C-F, <sup>2</sup>*J*<sub>CF</sub> = 15.2 Hz), 188.2 (C-F, <sup>2</sup>*J*<sub>CF</sub> = 15.2 Hz), 152.3, 146.6, 131.3 (C-F, <sup>3</sup>*J*<sub>CF</sub> = 4.0 Hz), 131.2 (C-F, <sup>3</sup>*J*<sub>CF</sub> = 4.0 Hz), 127.6 (C-F, <sup>1</sup>*J*<sub>CF3</sub> = 272.7 Hz), 127.2 (C-F, <sup>2</sup>*J*<sub>CF3</sub> = 33.3 Hz), 126.9 (C-F, <sup>2</sup>*J*<sub>CF3</sub> = 33.3 Hz), 126.6 (C-F, <sup>2</sup>*J*<sub>CF3</sub> = 33.3 Hz), 126.2 (C-F, <sup>2</sup>*J*<sub>CF3</sub> = 33.3 Hz), 125.6, 124.9 (C-F, <sup>1</sup>*J*<sub>CF3</sub> = 272.7 Hz), 124.2, 122.7, 122.2 (C-F, <sup>1</sup>*J*<sub>CF3</sub> = 272.7 Hz), 119.5 (C-F, <sup>1</sup>*J*<sub>CF3</sub> = 272.7 Hz), 87.6 (C-F, <sup>1</sup>*J*<sub>CF</sub> = 190.9 Hz), 85.7 (C-F, <sup>1</sup>*J*<sub>CF</sub> = 190.9 Hz), 84.1, 48.5 (C-F, <sup>2</sup>*J*<sub>CF</sub> = 27.3 Hz), 48.3 (C-F, <sup>2</sup>*J*<sub>CF</sub> = 27.3 Hz), 28.3. HRMS (APCI): *m/z* [M − Boc + H]<sup>+</sup> calcd for C<sub>10</sub>H<sub>8</sub>F<sub>4</sub>NO 234.0542, found 234.0537.

#### ***Tert*-butyl 6-chloro-3-fluoro-4-oxo-3,4-dihydroquinoline-1(2*H*)-carboxylate (2h):**

260 mg of *tert*-butyl 4-oxo-6-chloro-3,4-dihydroquinoline-1(2*H*)-carboxylate (0.92 mmol, 1.0 equiv) were used following the described procedure. Purification via flash column chromatography on silica gel (95:5 petroleum ether/ethyl acetate) yielded 107 mg of **2h** as a yellow solid (39%). m.p. 117–121 °C. <sup>1</sup>H NMR (400 MHz, Chloroform-*d*) δ 7.97 (d, *J* = 2.6 Hz, 1H), 7.81 (d, *J* = 9.0 Hz, 1H), 7.48 (dd, *J* = 9.0, 2.6 Hz, 1H), 5.09 (ddd, *J* = 46.8, 10.4, 4.7 Hz, 1H), 4.60 (td, *J* = 13.5, 4.6 Hz, 1H), 4.06 (ddd, *J* = 13.4, 10.4, 4.9 Hz, 1H), 1.56 (s, 9H). <sup>19</sup>F NMR (376 MHz, Chloroform-*d*) δ −198.92. <sup>13</sup>C NMR (101 MHz, Chloroform-*d*) δ 188.5 (C-F, <sup>2</sup>*J*<sub>CF</sub> = 15.2 Hz), 188.4 (C-F, <sup>2</sup>*J*<sub>CF</sub> = 15.2 Hz), 152.51, 142.5, 134.8, 130.4, 127.4, 125.3, 124.1, 87.8 (C-F, <sup>1</sup>*J*<sub>CF</sub> = 191.9 Hz), 85.9 (C-F, <sup>1</sup>*J*<sub>CF</sub> = 191.9 Hz), 83.6, 48.6 (C-F, <sup>2</sup>*J*<sub>CF</sub> = 26.3 Hz), 48.4 (C-F, <sup>2</sup>*J*<sub>CF</sub> = 26.3 Hz), 28.3. HRMS (APCI): *m/z* [M + H]<sup>+</sup> calcd for C<sub>14</sub>H<sub>16</sub>ClFNO<sub>3</sub> 300.0797, found 300.0799.

#### ***Tert*-butyl 6-bromo-3-fluoro-4-oxo-3,4-dihydroquinoline-1(2*H*)-carboxylate (2i):**

380 mg of *tert*-butyl 4-oxo-6-bromo-3,4-dihydroquinoline-1(2*H*)-carboxylate (1.16 mmol, 1.0 equiv) were used following the described procedure. Purification via flash column chromatography on silica gel (95:5 petroleum ether/ethyl acetate) yielded 165 mg of **2i** as a white solid (41%). m.p. 111–114 °C. <sup>1</sup>H NMR (400 MHz, Chloroform-*d*) δ 8.12 (d, *J* = 2.4 Hz, 1H), 7.75 (d, *J* = 9.0 Hz, 1H), 7.62 (dd, *J* = 9.0, 2.5 Hz, 1H), 5.09 (ddd, *J* = 46.8, 10.4, 4.7 Hz, 1H), 4.59 (td, *J* = 13.5, 4.6 Hz, 1H), 4.05 (ddd, *J* = 13.4, 10.4, 5.0 Hz, 1H), 1.56 (s, 9H). <sup>19</sup>F NMR (376 MHz, Chloroform-*d*) δ −198.90. <sup>13</sup>C NMR (101 MHz, Chloroform-*d*) δ 188.4 (C-F, <sup>2</sup>*J*<sub>CF</sub> = 15.2 Hz), 188.3 (C-F, <sup>2</sup>*J*<sub>CF</sub> = 15.2 Hz), 152.4, 143.0, 137.7, 130.5, 125.5, 124.4, 117.8, 87.8 (C-F, <sup>1</sup>*J*<sub>CF</sub> = 191.9 Hz), 85.9 (C-F, <sup>1</sup>*J*<sub>CF</sub> = 191.9 Hz), 83.7, 48.6 (C-F, <sup>2</sup>*J*<sub>CF</sub> = 27.3 Hz), 48.3 (C-F, <sup>2</sup>*J*<sub>CF</sub> = 27.3 Hz), 28.3. HRMS (APCI): *m/z* [M − Boc + H]<sup>+</sup> calcd for C<sub>9</sub>H<sub>8</sub>BrFNO 243.9773, found 243.9768.

#### ***Tert*-butyl 3-fluoro-6-iodo-4-oxo-3,4-dihydroquinoline-1(2*H*)-carboxylate (2j):**

339 mg of *tert*-butyl 4-oxo-6-iodo-3,4-dihydroquinoline-1(2*H*)-carboxylate (0.91 mmol, 1.0 equiv) were used following the described procedure. Purification via flash column chromatography on silica gel (97:3 petroleum ether/ethyl acetate) yielded 160 mg of **2j** as a white solid (45%). m.p. 113–117 °C. <sup>1</sup>H NMR (400 MHz, Chloroform-*d*) δ 8.31 (d, *J* = 2.2 Hz, 1H), 7.80 (dd, *J* = 8.9, 2.3 Hz, 1H), 7.61 (d, *J* = 8.9 Hz, 1H), 5.08 (ddd, *J* = 46.8, 10.4, 4.7 Hz, 1H), 4.59 (td, *J* = 13.5, 4.7 Hz, 1H), 4.04 (ddd, *J* = 13.4, 10.4, 5.0 Hz, 1H), 1.56 (s, 9H). <sup>19</sup>F NMR (376 MHz, Chloroform-*d*) δ −198.84. <sup>13</sup>C NMR (101 MHz, Chloroform-*d*) δ 188.3 (C-F, <sup>2</sup>*J*<sub>CF</sub> = 15.2 Hz), 188.2 (C-F, <sup>2</sup>*J*<sub>CF</sub> = 15.2 Hz), 152.4, 143.6, 143.4, 136.7, 125.6, 124.5, 88.0, 87.6 (C-F, <sup>1</sup>*J*<sub>CF</sub> = 190.9 Hz), 85.8 (C-F, <sup>1</sup>*J*<sub>CF</sub> = 190.9 Hz), 83.7, 48.5 (C-F, <sup>2</sup>*J*<sub>CF</sub> = 24.2 Hz), 48.3 (C-F,

$^2J_{\text{CF}} = 24.2$  Hz), 28.3. HRMS (APCI):  $m/z$   $[M + H]^+$  calcd for  $\text{C}_{14}\text{H}_{16}\text{FINO}_3$  392.0153, found 392.0155.

#### 4.3 General Procedure for the ATH of *tert*-butyl 3-fluoro-4-oxo-3,4-dihydroquinoline-1(2H)-carboxylates

In a round-bottom tube charged with the corresponding 3-fluoro-4-oxo-3,4-dihydroquinoline-1(2H)-carboxylate (1.0 equiv) set under argon, the necessary volume of a solution of (*R,R*)-**A** in acetonitrile (1.0 mL of a 1.24 mg/mL solution; 0.005 equiv) was added. The mixture was stirred for one minute before adding by syringe the necessary volume of formic acid/triethylamine (1:1) mixture (6.0 equiv). The reaction mixture was stirred at 40 °C for the time needed and then quenched with 3 mL of saturated  $\text{NaHCO}_3$  aqueous solution. The media was extracted with  $\text{CH}_2\text{Cl}_2$  (2 x 4 mL) and the organic layers dried over  $\text{MgSO}_4$ , filtered and concentrated under vacuum. The diastereoisomeric ratio was determined by  $^1\text{H}$  NMR analysis of the crude product. The product was purified with a flash column chromatography on silica gel (petroleum ether/EtOAc) and the enantiomeric excess was determined by SFC analysis (CHIRALPAK IE or IF column).

#### *Tert*-butyl (3*R*,4*S*)-3-fluoro-4-hydroxy-3,4-dihydroquinoline-1(2H)-carboxylate (**3a**):

100 mg of *tert*-butyl 3-fluoro-4-oxo-3,4-dihydroquinoline-1(2H)-carboxylate (**2a**) (0.38 mmol; 1.0 equiv) were used according to the described procedure (reaction time of 3 h). The product was purified by flash column chromatography on silica gel (petroleum ether/ethyl acetate 75:25) and yielded 93 mg of a beige/white solid (93%). m.p. 142–144 °C dr(*cis/trans*) = 98:2, ee<sub>cis</sub> = >99%.  $[\alpha]_{\text{D}}^{30} = -5$  (c 1.0,  $\text{CHCl}_3$ ).  $^1\text{H}$  NMR (400 MHz, Chloroform-*d*)  $\delta$  7.70 (d,  $J = 8.3$  Hz, 1H), 7.51 (dt,  $J = 7.6, 1.4$  Hz, 1H), 7.34–7.24 (m, 1H), 7.14 (td,  $J = 7.5, 1.2$  Hz, 1H), 5.05 (ddt,  $J = 50.8, 4.8, 3.5$  Hz, 1H), 4.78 (dd,  $J = 21.5, 5.9$  Hz, 1H), 4.10 (ddd,  $J = 14.9, 14.2, 4.8$  Hz, 1H), 3.94 (ddd,  $J = 29.9, 14.2, 3.7$  Hz, 1H), 2.38 (d,  $J = 8.3$  Hz, 1H), 1.53 (s, 9H).  $^{19}\text{F}$  NMR (376 MHz, Chloroform-*d*)  $\delta$  -200.34.  $^{13}\text{C}$  NMR (101 MHz, Chloroform-*d*)  $\delta$  153.8, 136.8, 128.2, 127.9, 127.3, 124.3, 123.5, 89.6 (C-F,  $^1J_{\text{CF}} = 176.8$  Hz), 87.9 (C-F,  $^1J_{\text{CF}} = 176.8$  Hz), 81.9, 68.0 (C-F,  $^2J_{\text{CF}} = 19.2$  Hz), 67.8 (C-F,  $^2J_{\text{CF}} = 19.2$  Hz), 46.3 (C-F,  $^2J_{\text{CF}} = 23.2$  Hz), 46.1 (C-F,  $^2J_{\text{CF}} = 23.2$  Hz), 28.4. HRMS (APCI):  $m/z$   $[M + H]^+$  calcd for  $\text{C}_{14}\text{H}_{19}\text{FNO}_3$  268.1343, found 268.1345. SFC: Chiralpak IE,  $\text{scCO}_2/\text{EtOH}$  96/4, 2.0 mL/min, P = 100 bar,  $\lambda = 215$  nm, tr [*trans*] = 11.04 min, tr [*trans*] = 12.85 min, tr [*cis*-(3*R*,4*S*)] = 14.54 min (major), tr [*cis*-(3*S*,4*R*)] = 15.43 min.

#### *Tert*-butyl (3*R*,4*S*)-3-fluoro-4-hydroxy-6-methyl-3,4-dihydroquinoline-1(2H)-carboxylate (**3b**):

100 mg of *tert*-butyl 3-fluoro-6-methyl-4-oxo-3,4-dihydroquinoline-1(2H)-carboxylate (**2b**) (0.36 mmol; 1.0 equiv) were used according to the described procedure (reaction time of 3 h). The product was purified by flash column chromatography on silica gel (petroleum ether/ethyl acetate 75:25) and yielded 91 mg of a colourless oil (91%). dr(*cis/trans*) = 98:2, ee<sub>cis</sub> = >99%.  $[\alpha]_{\text{D}}^{22} = -8.4$  (c 1.0,  $\text{CHCl}_3$ ).  $^1\text{H}$  NMR (400 MHz, Chloroform-*d*)  $\delta$  7.56 (d,  $J = 8.4$  Hz, 1H), 7.34–7.28 (m, 1H), 7.08 (dd,  $J = 8.4, 2.1$  Hz, 1H), 5.00 (dq,  $J = 50.9, 4.2$  Hz, 1H), 4.72 (dd,  $J = 22.0, 7.0$  Hz, 1H), 4.10–3.84 (m, 2H), 2.53 (s, 1H), 2.32 (s, 3H), 1.52 (s, 9H).  $^{19}\text{F}$  NMR (376 MHz, Chloroform-*d*)  $\delta$  -200.17.  $^{13}\text{C}$  NMR (101 MHz, Chloroform-*d*)  $\delta$  153.8, 134.2, 133.9, 128.9, 127.6, 123.4, 89.7, 88.0 (C-F,  $^1J_{\text{CF}} = 176.8$  Hz), 81.7 (C-F,  $^1J_{\text{CF}} = 176.8$  Hz), 67.9 (C-F,  $^2J_{\text{CF}} = 20.2$  Hz), 67.7 (C-F,  $^2J_{\text{CF}} = 20.2$  Hz), 46.2 (C-F,  $^2J_{\text{CF}} = 23.2$  Hz), 46.0 (C-F,  $^2J_{\text{CF}} = 23.2$  Hz), 28.4, 20.9. HRMS (APCI):  $m/z$   $[M + H]^+$  calcd for  $\text{C}_{15}\text{H}_{21}\text{FNO}_3$  282.1500, found 282.1502. SFC: Chiralpak IE,  $\text{scCO}_2/\text{EtOH}$  97.5/2.5, 2.0 mL/min, P = 100 bar,  $\lambda = 215$  nm, tr [*trans*] = 27.71 min, tr [*cis*-(3*R*,4*S*)] = 32.43 min (major), tr [*cis*-(3*S*,4*R*)] = 37.48 min, tr [*trans*] = 42.41 min.

***Tert*-butyl (3*R*,4*S*)-3-fluoro-4-hydroxy-6-methoxy-3,4-dihydroquinoline-1(2*H*)-carboxylate (3c):**

100 mg of *tert*-butyl 3-fluoro-6-methoxy-4-oxo-3,4-dihydroquinoline-1(2*H*)-carboxylate (2c) (0.34 mmol; 1.0 equiv) were used according to the described procedure (reaction time of 5 h). The product was purified by flash column chromatography on silica gel (petroleum ether/ethyl acetate 75:25) and yielded 95 mg of a white/red solid (95%). m.p. 114–118 °C dr(*cis*/*trans*) = 98:2, ee<sub>*cis*</sub> = >99%. [ $\alpha$ ]<sub>D</sub><sup>22</sup> = -13.3 (c 1.0, CHCl<sub>3</sub>). <sup>1</sup>H NMR (400 MHz, Chloroform-*d*)  $\delta$  7.57 (d, *J* = 9.1 Hz, 1H), 7.05 (dd, *J* = 3.0, 1.0 Hz, 1H), 6.83 (dd, *J* = 9.1, 3.0 Hz, 1H), 5.06 (dd, *J* = 51.1, 3.7 Hz, 1H), 4.74 (ddd, *J* = 22.7, 9.4, 3.2 Hz, 1H), 4.08 (ddd, *J* = 15.9, 14.4, 4.4 Hz, 1H), 3.92 (ddd, *J* = 31.4, 14.5, 3.7 Hz, 1H), 3.81 (s, 3H), 2.37 (d, *J* = 9.4 Hz, 1H), 1.52 (s, 9H). <sup>19</sup>F NMR (376 MHz, Chloroform-*d*)  $\delta$  -200.32. <sup>13</sup>C NMR (101 MHz, Chloroform-*d*)  $\delta$  156.5, 153.9, 129.8, 129.4, 125.0, 114.3, 111.4, 89.9 (C-F, <sup>1</sup>*J*<sub>CF</sub> = 176.8 Hz), 88.1 (C-F, <sup>1</sup>*J*<sub>CF</sub> = 176.8 Hz), 81.6, 68.2 (C-F, <sup>2</sup>*J*<sub>CF</sub> = 20.2 Hz), 68.0 (C-F, <sup>2</sup>*J*<sub>CF</sub> = 20.2 Hz), 55.7, 46.6 (C-F, <sup>2</sup>*J*<sub>CF</sub> = 23.2 Hz), 46.3 (C-F, <sup>2</sup>*J*<sub>CF</sub> = 23.2 Hz), 28.4. HRMS (APCI): *m/z* [M + Na]<sup>+</sup> calcd for C<sub>15</sub>H<sub>20</sub>FN<sub>2</sub>O<sub>4</sub>Na 320.1269, found 320.1273. SFC: Chiralpak IF, scCO<sub>2</sub>/EtOH 95.5/45, 2.0 mL/min, P = 100 bar,  $\lambda$  = 254 nm, tr [*trans*] = 15.71 min, tr [*cis*-(3*S*,4*R*)] = 17.11 min, tr [*trans*] = 18.69 min, tr [*cis*-(3*R*,4*S*)] = 22.79 min (major)

***Tert*-butyl (3*R*,4*S*)-3-fluoro-4-hydroxy-7-methoxy-3,4-dihydroquinoline-1(2*H*)-carboxylate (3d):**

100 mg of *tert*-butyl 3-fluoro-7-methoxy-4-oxo-3,4-dihydroquinoline-1(2*H*)-carboxylate (2d) (0.34 mmol; 1.0 equiv) were used according to the described procedure (reaction time of 5 h). The product was purified by flash column chromatography on silica gel (petroleum ether/ethyl acetate 75:25) and yielded 95 mg of a beige/white solid (95%). m.p. 92–96 °C dr(*cis*/*trans*) = 95:5, ee<sub>*cis*</sub> = >99%. [ $\alpha$ ]<sub>D</sub><sup>22</sup> = -9.2 (c 1.0, CHCl<sub>3</sub>). <sup>1</sup>H NMR (400 MHz, Chloroform-*d*)  $\delta$  7.42 – 7.31 (m, 2H), 6.70 (dt, *J* = 8.6, 2.3 Hz, 1H), 4.98 (dd, *J* = 50.0, 5.9 Hz, 1H), 4.76 (d, *J* = 19.1 Hz, 1H), 4.16 – 4.04 (m, 1H), 4.01 – 3.85 (m, 1H), 3.80 (s, 3H), 2.33 (s, 1H), 1.54 (s, 9H). <sup>19</sup>F NMR (376 MHz, Chloroform-*d*)  $\delta$  -201.08. <sup>13</sup>C NMR (101 MHz, Chloroform-*d*)  $\delta$  159.7, 153.6, 138.0, 128.9, 119.8, 110.8, 108.6, 89.1 (C-F, <sup>1</sup>*J*<sub>CF</sub> = 177.8 Hz), 87.3 (C-F, <sup>1</sup>*J*<sub>CF</sub> = 177.8 Hz), 82.0, 67.6 (C-F, <sup>2</sup>*J*<sub>CF</sub> = 19.2 Hz), 67.4 (C-F, <sup>2</sup>*J*<sub>CF</sub> = 19.2 Hz), 55.5, 45.7 (C-F, <sup>2</sup>*J*<sub>CF</sub> = 25.3 Hz), 45.5 (C-F, <sup>2</sup>*J*<sub>CF</sub> = 25.3 Hz), 28.5. HRMS (APCI): *m/z* [M + NH<sub>4</sub>]<sup>+</sup> calcd for C<sub>15</sub>H<sub>24</sub>FN<sub>2</sub>O<sub>4</sub> 315.1715, found 315.1718. SFC: Chiralpak IE, scCO<sub>2</sub>/EtOH 95/5, 2.0 mL/min, P = 100 bar,  $\lambda$  = 215 nm, tr [*trans*] = 11.87 min, tr [*trans*] = 13.19 min, tr [*cis*-(3*R*,4*S*)] = 16.63 min (major), tr [*cis*-(3*S*,4*R*)] = 18.62 min.

***Tert*-butyl (3*R*,4*S*)-3-fluoro-4-hydroxy-6,7-dimethoxy-3,4-dihydroquinoline-1(2*H*)-carboxylate (3e):**

100 mg of *tert*-butyl 3-fluoro-6,7-dimethoxy-4-oxo-3,4-dihydroquinoline-1(2*H*)-carboxylate (2e) (0.31 mmol; 1.0 equiv) were used according to the described procedure with 1.0 mol% of catalyst (*R,R*)-**D** (reaction time of 24 h). The product was purified by flash column chromatography on silica gel (petroleum ether/ethyl acetate 60:40) and yielded 90 mg of a colourless oil (90%). dr(*cis*/*trans*) = 94:6, ee<sub>*cis*</sub> = 99%. [ $\alpha$ ]<sub>D</sub><sup>22</sup> = -11.8 (c 1.0, CHCl<sub>3</sub>). <sup>1</sup>H NMR (400 MHz, Chloroform-*d*)  $\delta$  7.32 (s, 1H), 6.97 (d, *J* = 0.8 Hz, 1H), 4.99 (ddt, *J* = 50.2, 5.6, 3.4 Hz, 1H), 4.74 (dq, *J* = 19.9, 3.5 Hz, 1H), 4.20 – 4.11 (m, 1H), 3.94 – 3.80 (m, 7H), 2.38 (d, *J* = 8.4 Hz, 1H), 1.53 (s, 9H). <sup>19</sup>F NMR (376 MHz, Chloroform-*d*)  $\delta$  -202.03. <sup>13</sup>C NMR (101 MHz, Chloroform-*d*)  $\delta$  153.8, 148.5, 146.2, 130.4, 119.6, 110.0, 107.5, 89.1 (C-F, <sup>1</sup>*J*<sub>CF</sub> = 177.8 Hz), 87.3 (C-F, <sup>1</sup>*J*<sub>CF</sub> = 177.8 Hz), 81.7, 67.7 (C-F, <sup>2</sup>*J*<sub>CF</sub> = 20.2 Hz), 67.5 (C-F, <sup>2</sup>*J*<sub>CF</sub> = 20.2 Hz), 56.2, 56.1, 46.0 (C-F, <sup>2</sup>*J*<sub>CF</sub> = 24.2 Hz), 45.8 (C-F, <sup>2</sup>*J*<sub>CF</sub> = 24.2 Hz), 28.5. HRMS (APCI): *m/z* [M + Na]<sup>+</sup> calcd for C<sub>16</sub>H<sub>22</sub>FN<sub>2</sub>O<sub>5</sub>Na 350.1374, found 350.1377. SFC: Chiralpak IF, scCO<sub>2</sub>/EtOH 95/5, 2.0 mL/min, P = 100 bar,  $\lambda$  = 215 nm, tr [*trans*] = 14.75 min, tr [*trans*] = 15.89 min, tr [*cis*-(3*R*,4*S*)] = 17.45 min (major), tr [*cis*-(3*S*,4*R*)] = 19.07 min.

***Tert*-butyl (3*R*,4*S*)-3,6-difluoro-4-hydroxy-3,4-dihydroquinoline-1(2*H*)-carboxylate (3*f*):**

100 mg of *tert*-butyl 3,6-difluoro-4-oxo-3,4-dihydroquinoline-1(2*H*)-carboxylate (**2f**) (0.35 mmol; 1.0 equiv) were used according to the described procedure (reaction time of 24 h). The product was purified by flash column chromatography on silica gel (petroleum ether/ethyl acetate 75:25) and yielded 70 mg of a white solid (70%). m.p. 78–82 °C dr(*cis*/*trans*) = 98:2, ee<sub>*cis*</sub> = >99%. [ $\alpha$ ]<sub>D</sub><sup>22</sup> = −11.7 (c 1.0, CHCl<sub>3</sub>). <sup>1</sup>H NMR (400 MHz, Chloroform-*d*)  $\delta$  7.64 (dd, *J* = 9.3, 5.1 Hz, 1H), 7.23 (ddd, *J* = 8.8, 3.0, 1.0 Hz, 1H), 6.97 (td, *J* = 8.9, 8.1, 3.1 Hz, 1H), 5.08 (dq, *J* = 51.2, 3.6 Hz, 1H), 4.71 (ddd, *J* = 23.6, 9.5, 3.1 Hz, 1H), 4.09 (ddd, *J* = 16.6, 14.6, 3.9 Hz, 1H), 3.91 (ddd, *J* = 33.1, 14.6, 3.6 Hz, 1H), 2.48 (dd, *J* = 9.7, 2.0 Hz, 1H), 1.52 (s, 9H). <sup>19</sup>F NMR (376 MHz, Chloroform-*d*)  $\delta$  −118.05, −200.38. <sup>13</sup>C NMR (101 MHz, Chloroform-*d*)  $\delta$  160.8 (C-F, <sup>1</sup>*J*<sub>CF</sub> = 244.4 Hz), 158.4 (C-F, <sup>1</sup>*J*<sub>CF</sub> = 244.4 Hz), 153.7, 132.6 (C-F, <sup>4</sup>*J*<sub>CF</sub> = 2.0 Hz), 132.6 (C-F, <sup>4</sup>*J*<sub>CF</sub> = 2.0 Hz), 130.3 6 (C-F, <sup>3</sup>*J*<sub>CF</sub> = 6.1 Hz), 130.3 (C-F, <sup>3</sup>*J*<sub>CF</sub> = 6.1 Hz), 125.4 (C-F, <sup>3</sup>*J*<sub>CF</sub> = 7.1 Hz), 125.3 (C-F, <sup>3</sup>*J*<sub>CF</sub> = 7.1 Hz), 115.1 (C-F, <sup>2</sup>*J*<sub>CF</sub> = 22.2 Hz), 114.9 (C-F, <sup>2</sup>*J*<sub>CF</sub> = 22.2 Hz), 113.5 (C-F, <sup>2</sup>*J*<sub>CF</sub> = 22.2 Hz), 113.3 (C-F, <sup>2</sup>*J*<sub>CF</sub> = 22.2 Hz), 89.7 (C-F, <sup>1</sup>*J*<sub>CF</sub> = 176.8 Hz), 87.9 (C-F, <sup>1</sup>*J*<sub>CF</sub> = 176.8 Hz), 82.1, 67.9 (C-F, <sup>2</sup>*J*<sub>CF</sub> = 20.2 Hz), 67.7 (C-F, <sup>2</sup>*J*<sub>CF</sub> = 20.2 Hz), 46.8 (C-F, <sup>2</sup>*J*<sub>CF</sub> = 23.2 Hz), 46.6 (C-F, <sup>2</sup>*J*<sub>CF</sub> = 23.2 Hz), 28.4. HRMS (APCI): *m/z* [M + H]<sup>+</sup> calcd for C<sub>14</sub>H<sub>18</sub>F<sub>2</sub>NO<sub>3</sub> 286.1249, found 282.1248. SFC: Chiralpak IE, scCO<sub>2</sub>/MeOH 94/6, 2.0 mL/min, P = 100 bar,  $\lambda$  = 215 nm, tr [*trans*] = 4.50 min, tr [*trans*] = 4.96 min, tr [*cis*-(3*S*,4*R*)] = 6.44 min, tr [*cis*-(3*R*,4*S*)] = 7.00 min (major).

***Tert*-butyl (3*R*,4*S*)-3-fluoro-4-hydroxy-6-(trifluoromethyl)-3,4-dihydroquinoline-1(2*H*)-carboxylate (3*g*):**

100 mg of *tert*-butyl 3-fluoro-6-trifluoromethyl-4-oxo-3,4-dihydroquinoline-1(2*H*)-carboxylate (**2g**) (0.30 mmol; 1.0 equiv) were used according to the described procedure (reaction time of 3 h). The product was purified by flash column chromatography on silica gel (petroleum ether/ethyl acetate 75:25) and yielded 97 mg of a beige/white solid (97%). m.p. 152–156 °C dr(*cis*/*trans*) = 98:2, ee<sub>*cis*</sub> = >99%. [ $\alpha$ ]<sub>D</sub><sup>22</sup> = −0.9 (c 1.0, CHCl<sub>3</sub>). <sup>1</sup>H NMR (400 MHz, Chloroform-*d*)  $\delta$  7.88 (d, *J* = 8.7 Hz, 1H), 7.81 (d, *J* = 2.2 Hz, 1H), 7.51 (ddt, *J* = 8.7, 2.2, 0.7 Hz, 1H), 5.10 (dddd, *J* = 50.9, 7.7, 3.7, 1.4 Hz, 1H), 4.79 (dd, *J* = 23.1, 6.9 Hz, 1H), 4.17 (tdd, *J* = 15.5, 4.1, 1.1 Hz, 1H), 3.91 (dd, *J* = 33.7, 3.6 Hz, 1H), 2.45 (s, 1H), 1.54 (s, 9H). <sup>19</sup>F NMR (376 MHz, Chloroform-*d*)  $\delta$  −62.20, −200.84. <sup>13</sup>C NMR (101 MHz, Chloroform-*d*)  $\delta$  153.4, 139.7, 128.3 (C-F, <sup>1</sup>*J*<sub>CF3</sub> = 272.7 Hz), 128.2, 128.1, 126.5 (C-F, <sup>2</sup>*J*<sub>CF3</sub> = 33.3 Hz), 126.2 (C-F, <sup>2</sup>*J*<sub>CF3</sub> = 33.3 Hz), 125.9 (C-F, <sup>2</sup>*J*<sub>CF3</sub> = 33.3 Hz), 125.6 (C-F, <sup>1</sup>*J*<sub>CF3</sub> = 272.7 Hz + C-F, <sup>2</sup>*J*<sub>CF3</sub> = 33.3 Hz), 125.1 (C-F, <sup>3</sup>*J*<sub>CF3</sub> = 4.0 Hz), 125.1 (C-F, <sup>3</sup>*J*<sub>CF3</sub> = 4.0 Hz), 124.4 (C-F, <sup>3</sup>*J*<sub>CF3</sub> = 4.0 Hz), 124.4 (C-F, <sup>3</sup>*J*<sub>CF3</sub> = 4.0 Hz), 123.4, 122.9 (C-F, <sup>1</sup>*J*<sub>CF3</sub> = 272.7 Hz), 120.2 (C-F, <sup>1</sup>*J*<sub>CF3</sub> = 272.7 Hz), 89.1 (C-F, <sup>1</sup>*J*<sub>CF</sub> = 176.8 Hz), 87.4 (C-F, <sup>1</sup>*J*<sub>CF</sub> = 176.8 Hz), 82.7, 67.7 (C-F, <sup>2</sup>*J*<sub>CF</sub> = 20.2 Hz), 67.5 (C-F, <sup>2</sup>*J*<sub>CF</sub> = 20.2 Hz), 46.8 (C-F, <sup>2</sup>*J*<sub>CF</sub> = 22.2 Hz), 46.6 (C-F, <sup>2</sup>*J*<sub>CF</sub> = 22.2 Hz), 28.3. HRMS (APCI): *m/z* [M + NH<sub>4</sub>]<sup>+</sup> calcd for C<sub>15</sub>H<sub>21</sub>FN<sub>2</sub>O<sub>3</sub> 353.1483, found 353.1483. SFC: Chiralpak IF, scCO<sub>2</sub>/EtOH 96/4, 2.0 mL/min, P = 100 bar,  $\lambda$  = 254 nm, tr [*cis*-(3*R*,4*S*)] = 5.68 min (major), tr [*cis*-(3*S*,4*R*)] = 6.84 min, tr [*trans*] = 8.54 min.

***Tert*-butyl (3*R*,4*S*)-6-chloro-3-fluoro-4-hydroxy-3,4-dihydroquinoline-1(2*H*)-carboxylate (3*h*):**

60 mg of *tert*-butyl 3-fluoro-6-chloro-4-oxo-3,4-dihydroquinoline-1(2*H*)-carboxylate (**2h**) (0.20 mmol; 1.0 equiv) were used according to the described procedure (reaction time of 3 h). The product was purified by flash column chromatography on silica gel (petroleum ether/ethyl acetate 75:25) and yielded 97 mg of a beige/white solid (96%). m.p. 113–116 °C dr(*cis*/*trans*) = 99:1, ee<sub>*cis*</sub> = >99%. [ $\alpha$ ]<sub>D</sub><sup>22</sup> = −12.8 (c 1.0, CHCl<sub>3</sub>). <sup>1</sup>H NMR (400 MHz, Chloroform-*d*)  $\delta$  7.66 (d, *J* = 8.9 Hz, 1H), 7.51 (dd, *J* = 2.6, 1.1 Hz, 1H), 7.22 (d, *J* = 9.2 Hz, 1H), 5.07 (dt, *J* = 50.8, 3.9 Hz, 1H), 4.78 – 4.67 (m, 1H), 4.17 – 4.05 (m, 1H), 3.99 – 3.81 (m, 1H), 2.44 (br, 1H), 1.52 (s, 9H). <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>)  $\delta$  −200.57. <sup>13</sup>C NMR (101 MHz,

Chloroform-*d*)  $\delta$  153.5, 135.2, 129.6, 128.2, 126.9, 124.8, 89.4 (C-F,  $^1J_{CF}$  = 176.8 Hz), 87.7 (C-F,  $^1J_{CF}$  = 176.8 Hz), 82.3, 67.7 (C-F,  $^2J_{CF}$  = 20.2 Hz), 67.5 (C-F,  $^2J_{CF}$  = 20.2 Hz), 46.7 (C-F,  $^2J_{CF}$  = 23.2 Hz), 46.5 (C-F,  $^2J_{CF}$  = 23.2 Hz), 28.4. HRMS (APCI):  $m/z$  [M + H]<sup>+</sup> calcd for C<sub>14</sub>H<sub>18</sub>ClFNO<sub>3</sub> 302.0954, found 302.0955. SFC: Chiralpak IE, scCO<sub>2</sub>/MeOH 94/6, 2.0 mL/min, P = 100 bar,  $\lambda$  = 254 nm,  $t_R$  [*trans*] = 6.95 min,  $t_R$  [*trans*] = 7.49 min,  $t_R$  [*cis*-(3*S*,4*R*)] = 10.31 min,  $t_R$  [*cis*-(3*R*,4*S*)] = 11.09 min (major).

***Tert*-butyl (3*R*,4*S*)-6-bromo-3-fluoro-4-hydroxy-3,4-dihydroquinoline-1(2*H*)-carboxylate (3i):**

80 mg of *tert*-butyl 3-fluoro-6-bromo-4-oxo-3,4-dihydroquinoline-1(2*H*)-carboxylate (**2i**) (0.24 mmol; 1.0 equiv) were used according to the described procedure (reaction time of 3 h). The product was purified by flash column chromatography on silica gel (petroleum ether/ethyl acetate 75:25) and yielded 97 mg of a beige/white solid (95%). m.p. 128–131 °C dr(*cis/trans*) = 98:2, ee<sub>*cis*</sub> = >99%. [ $\alpha$ ]<sub>D</sub><sup>22</sup> = -15.6 (*c* 1.0, CHCl<sub>3</sub>). <sup>1</sup>H NMR (400 MHz, Chloroform-*d*)  $\delta$  7.68–7.65 (m, 1H), 7.59 (d, *J* = 8.9 Hz, 1H), 7.37 (dd, *J* = 8.9, 2.4 Hz, 1H), 5.05 (dq, *J* = 51.0, 3.6 Hz, 1H), 4.71 (ddd, *J* = 23.2, 9.3, 3.1 Hz, 1H), 4.10 (ddd, *J* = 15.7, 14.4, 4.1 Hz, 1H), 3.88 (ddd, *J* = 33.1, 14.5, 3.5 Hz, 1H), 2.52 (d, *J* = 9.4 Hz, 1H), 1.52 (s, 9H). <sup>19</sup>F NMR (376 MHz, Chloroform-*d*)  $\delta$  -200.49. <sup>13</sup>C NMR (101 MHz, Chloroform-*d*)  $\delta$  153.5, 135.7, 131.1, 130.0, 129.9, 125.1, 117.2, 89.4 (C-F,  $^1J_{CF}$  = 176.8 Hz), 87.6 (C-F,  $^1J_{CF}$  = 176.8 Hz), 82.3, 67.7 (C-F,  $^2J_{CF}$  = 20.2 Hz), 67.5 (C-F,  $^2J_{CF}$  = 20.2 Hz), 46.7 (C-F,  $^2J_{CF}$  = 23.2 Hz), 46.4 (C-F,  $^2J_{CF}$  = 23.2 Hz), 28.4. HRMS (APCI):  $m/z$  [M + NH<sub>4</sub>]<sup>+</sup> calcd for C<sub>14</sub>H<sub>21</sub>BrFN<sub>2</sub>O<sub>3</sub> 363.0714, found 363.0714. SFC: Chiralpak IE, scCO<sub>2</sub>/MeOH 93/7, 2.0 mL/min, P = 100 bar,  $\lambda$  = 254 nm,  $t_R$  [*trans*] = 7.72 min,  $t_R$  [*cis*-(3*S*,4*R*)] = 11.07 min,  $t_R$  [*cis*-(3*R*,4*S*)] = 11.96 min (major).

***Tert*-butyl (3*R*,4*S*)-3-fluoro-4-hydroxy-6-iodo-3,4-dihydroquinoline-1(2*H*)-carboxylate (3j):**

80 mg of *tert*-butyl 3-fluoro-6-iodo-4-oxo-3,4-dihydroquinoline-1(2*H*)-carboxylate (**2j**) (0.20 mmol; 1.0 equiv) were used according to the described procedure (reaction time of 3 h). The product was purified by flash column chromatography on silica gel (petroleum ether/ethyl acetate 75:25) and yielded 97 mg of a beige/white solid (94%). m.p. 125–128 °C dr(*cis/trans*) = 98:2, ee<sub>*cis*</sub> = >99%. [ $\alpha$ ]<sub>D</sub><sup>22</sup> = -19.6 (*c* 1.0, CHCl<sub>3</sub>). <sup>1</sup>H NMR (400 MHz, Chloroform-*d*)  $\delta$  7.84 (d, *J* = 0.8 Hz, 1H), 7.56 (dt, *J* = 8.7, 1.6 Hz, 1H), 7.47 (d, *J* = 8.8 Hz, 1H), 5.04 (dt, *J* = 50.9, 3.7 Hz, 1H), 4.71 (dd, *J* = 23.3, 8.8 Hz, 1H), 4.10 (ddt, *J* = 18.6, 15.1, 3.6 Hz, 1H), 3.97–3.78 (m, 1H), 2.46 (s, 1H), 1.52 (s, 9H). <sup>19</sup>F NMR (376 MHz, Chloroform-*d*)  $\delta$  -200.57. <sup>13</sup>C NMR (101 MHz, Chloroform-*d*)  $\delta$  153.4, 137.0, 136.6, 135.9, 130.1, 125.3, 89.3 (C-F,  $^1J_{CF}$  = 176.8 Hz), 87.8 (C-F,  $^1J_{CF}$  = 176.8 Hz), 87.5, 82.4, 67.5 (C-F,  $^2J_{CF}$  = 20.2 Hz), 67.3 (C-F,  $^2J_{CF}$  = 20.2 Hz), 46.6 (C-F,  $^2J_{CF}$  = 23.2 Hz), 46.3 (C-F,  $^2J_{CF}$  = 23.2 Hz), 28.4. HRMS (APCI):  $m/z$  [M + NH<sub>4</sub>]<sup>+</sup> calcd for C<sub>14</sub>H<sub>21</sub>FIN<sub>2</sub>O<sub>3</sub> 411.0575, found 411.0577. SFC: Chiralpak IE, scCO<sub>2</sub>/MeOH 93/7, 2.0 mL/min, P = 100 bar,  $\lambda$  = 254 nm,  $t_R$  [*trans*] = 10.92 min,  $t_R$  [*trans*] = 11.41 min,  $t_R$  [*cis*-(3*S*,4*R*)] = 15.73 min,  $t_R$  [*cis*-(3*R*,4*S*)] = 17.25 min (major).

#### 4.4 Gram-scale experiment

In a 30 mL-round-bottom tube charged with *tert*-butyl 3-fluoro-4-oxo-3,4-dihydroquinoline-1(2*H*)-carboxylate (**2a**, 1.0 g, 3.8 mmol; 1.0 equiv) and the catalyst (*R,R*)-D (12.4 mg, 12  $\mu$ mol; 0.005 equiv) set under argon were added 10 mL of acetonitrile. The mixture was stirred for one minute before adding by syringe a (1:1) mixture of formic acid and triethylamine (3.40 mL, 16 mmol; 6.0 equiv). The reaction mixture was stirred at 40 °C for 24 h. Then it was cooled down and quenched with 30 mL of NaHCO<sub>3</sub> aqueous solution. The mixture was extracted with CH<sub>2</sub>Cl<sub>2</sub> (2 x 40 mL) and the organic layers dried over

MgSO<sub>4</sub>, filtered and concentrated under vacuum. The diastereoisomeric ratio was determined by <sup>1</sup>H NMR analysis of the crude product. The product was purified with a flash column chromatography on silica gel (petroleum ether/EtOAc 70:30). 960 mg (96%) of (3*R*,4*R*)-3-fluorochroman-4-ol (**3a**) as a white solid were obtained. dr (*cis/trans*) 97:3, ee<sub>*cis*</sub> = >99%. <sup>1</sup>H NMR (400 MHz, Chloroform-*d*) δ 7.70 (d, *J* = 8.3 Hz, 1H), 7.51 (dt, *J* = 7.6, 1.4 Hz, 1H), 7.34–7.24 (m, 1H), 7.14 (td, *J* = 7.5, 1.2 Hz, 1H), 5.05 (ddt, *J* = 50.8, 4.8, 3.5 Hz, 1H), 4.78 (dd, *J* = 21.5, 5.9 Hz, 1H), 4.10 (ddd, *J* = 14.9, 14.2, 4.8 Hz, 1H), 3.94 (ddd, *J* = 29.9, 14.2, 3.7 Hz, 1H), 2.38 (d, *J* = 8.3 Hz, 1H), 1.53 (s, 9H). <sup>19</sup>F NMR (376 MHz, Chloroform-*d*) δ –200.34. <sup>13</sup>C NMR (101 MHz, Chloroform-*d*) δ 153.8, 136.8, 128.2, 127.9, 127.3, 124.3, 123.5, 89.6 (C-F, <sup>1</sup>*J*<sub>CF</sub> = 176.8 Hz), 87.9 (C-F, <sup>1</sup>*J*<sub>CF</sub> = 176.8 Hz), 81.9, 68.0 (C-F, <sup>2</sup>*J*<sub>CF</sub> = 19.2 Hz), 67.8 (C-F, <sup>2</sup>*J*<sub>CF</sub> = 19.2 Hz), 46.3 (C-F, <sup>2</sup>*J*<sub>CF</sub> = 23.2 Hz), 46.1 (C-F, <sup>2</sup>*J*<sub>CF</sub> = 23.2 Hz), 28.4. SFC: Chiralpak IE, scCO<sub>2</sub>/EtOH 96/4, 2.0 mL/min, P = 100 bar, λ = 215 nm, t<sub>R</sub> [*trans*] = 11.04 min, t<sub>R</sub> [*trans*] = 12.85 min, t<sub>R</sub> [*cis*-(3*R*,4*S*)] = 14.54 min (major), t<sub>R</sub> [*cis*-(3*S*,4*R*)] = 15.43 min.

#### 4.5 Sonogashira coupling reaction *tert*-Butyl (3*R*,4*S*)-3-fluoro-4-hydroxy-6-(phenylethynyl)-3,4-dihydroquinoline-1(2*H*)-carboxylate (**4**):

*tert*-Butyl (3*R*,4*S*)-3-fluoro-4-hydroxy-6-iodo-3,4-dihydroquinoline-1(2*H*)-carboxylate (**3j**) (77 mg ; 0.2 mmol ; 1.0 equiv) was introduced into a 10 mL round-bottom tube at open air with 1 mL of a 1 : 1 CH<sub>3</sub>CN/Et<sub>3</sub>N mixture. Next, phenylacetylene was added (31 µL ; 0.3 mmol ; 1.5 equiv) followed by CuI (2.8 mg ; 10 µmol ; 0.05 equiv) and PdCl<sub>2</sub>(PPh<sub>3</sub>)<sub>2</sub> (6.4 mg ; 6 µmol ; 0.03 equiv). The mixture was stirred for 30 min at room temperature. It was then quenched with 2 mL of NH<sub>4</sub>Cl saturated solution and extracted with EtOAc (3 × 10 mL). The combined organic phases were dried with MgSO<sub>4</sub>, filtered and concentrated under reduced pressure. The crude product was purified by flash column chromatography on silica gel, (petroleum ether/EtOAc/toluene 7:2:1) to afford **4** as a pale yellow solid (49 mg ; 68%). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.76–7.69 (m, 2H), 7.55–7.49 (m, 2H), 7.44 (ddd, *J* = 8.6, 2.1, 0.6 Hz, 1H), 7.37–7.30 (m, 3H), 5.06 (ddt, *J* = 50.8, 4.5, 3.4 Hz, 1H), 4.76 (dd, *J* = 23.3, 6.0 Hz, 1H), 4.13 (ddd, *J* = 15.3, 14.3, 4.4 Hz, 1H), 3.91 (ddd, *J* = 32.0, 14.3, 3.5 Hz, 1H), 2.52 (d, *J* = 9.0 Hz, 1H), 1.54 (s, 9H). <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>) δ –200.41. <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ 153.5, 136.7, 131.7, 131.3, 130.5, 128.5, 128.3, 127.9, 123.4, 123.2, 118.9, 89.4, 89.3 (C-F, <sup>1</sup>*J*<sub>CF</sub> = 176.8 Hz), 89.1, 87.6 (C-F, <sup>1</sup>*J*<sub>CF</sub> = 176.8 Hz), 82.3, 67.7 (C-F, <sup>2</sup>*J*<sub>CF</sub> = 20.2 Hz), 67.5 (C-F, <sup>2</sup>*J*<sub>CF</sub> = 20.2 Hz), 46.6 (C-F, <sup>2</sup>*J*<sub>CF</sub> = 23.2 Hz), 46.4 (C-F, <sup>2</sup>*J*<sub>CF</sub> = 23.2 Hz), 28.4. MS (ESI) [M – Boc + H]<sup>+</sup> = 267.

#### 4.6 N-Boc deprotection (3*R*,4*S*)-3-Fluoro-1,2,3,4-tetrahydroquinolin-4-ol (**5**):

*tert*-Butyl (3*R*,4*S*)-3-fluoro-4-hydroxy-3,4-dihydroquinoline-1(2*H*)-carboxylate **3a** (200 mg ; 0.75 mmol ; 1.0 equiv) was introduced in a 50 mL round bottom tube set under argon. 3.0 mL of 1,4-dioxane were added followed by 5.0 mL of HPLC-grade H<sub>2</sub>O. The mixture was heated to reflux for 34 h. It was then allowed to cool down to room temperature and it was extracted with MTBE (3 × 20 mL). The combined organic phases were dried with MgSO<sub>4</sub>, filtered and concentrated under reduced pressure. The crude product was purified by flash column chromatography on silica gel (petroleum ether/EtOAc 8:2 to 7:3) to afford **5** as a pale yellow solid (112 mg; 90%). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.35 (dt, *J* = 7.8, 1.0 Hz, 1H), 7.11 (ddd, *J* = 8.5, 7.5, 1.6 Hz, 1H), 6.75 (td, *J* = 7.4, 1.2 Hz, 1H), 6.55 (dd, *J* = 8.0, 1.1 Hz, 1H), 5.08–4.75 (m, 2H), 3.87 (s, 1H), 3.66 (dt, *J* = 11.9, 7.9 Hz, 1H), 3.45 (dddd, *J* = 22.2, 11.9, 3.3, 1.4 Hz, 1H), 2.26 (s, 1H). <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>) δ –203.21. <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ 143.4, 129.7, 129.6, 120.0, 118.2, 114.2, 88.8 (C-F, <sup>1</sup>*J*<sub>CF</sub> = 176.8 Hz), 87.1 (C-F, <sup>1</sup>*J*<sub>CF</sub> = 176.8 Hz), 67.3 (C-F, <sup>2</sup>*J*<sub>CF</sub> = 19.2 Hz), 67.1 (C-F, <sup>2</sup>*J*<sub>CF</sub> = 19.2 Hz), 41.8 (C-F, <sup>2</sup>*J*<sub>CF</sub> = 24.2 Hz), 41.6 (C-F, <sup>2</sup>*J*<sub>CF</sub> = 24.2 Hz). HRMS (ESI) m/z: [M+H]<sup>+</sup> Calcd for C<sub>9</sub>H<sub>11</sub>FNO 168.0819. Found 168.0819.



**Supplementary Materials:** The following are available online at [www.mdpi.com/xxx/s1](http://www.mdpi.com/xxx/s1), Figure S1: title, Table S1: title, Video S1: title.

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**Data Availability Statement:** CCDC 2132626 contains the supplementary crystallographic data for this paper. These data can be obtained free of charge via [www.ccdc.cam.ac.uk/data\\_request/cif](http://www.ccdc.cam.ac.uk/data_request/cif), or by emailing [data\\_request@ccdc.cam.ac.uk](mailto:data_request@ccdc.cam.ac.uk), or by contacting The Cambridge Crystallographic Data Centre, 12 Union Road, Cambridge CB2 1EZ, UK; fax: +44 1223 336033

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