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#### Genetic causes of functional adrenocortical adenomas

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**Abbreviations**: PA, primary aldosteronism; CS, Cushing syndrome; sCS, subclinical Cushing syndrome; APA, aldosterone producing adenomas; CPA, cortisol producing adenomas; A/CPA, aldosterone and cortisol producing adenoma; ACA, adrenocortical adenoma; BAH: bilateral adrenal hyperplasia; PBMAH, primary bilateral macronodular adrenal hyperplasia; FH, familial hyperaldosteronism

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#### Abstract

Aldosterone and cortisol, the main mineralocorticoid and glucocorticoid hormones in humans, are produced in the adrenal cortex, which is composed of three concentric zone with specific functional characteristics. Adrenocortical adenomas (ACA) may lead to the autonomous secretion of aldosterone responsible for primary aldosteronism, the most frequent form of secondary arterial hypertension. In the case of cortisol production, ACA lead to overt or subclinical Cushing syndrome. Genetic analysis driven by next generation sequencing technology has enabled the discovery, during the last seven years, of the genetic causes of a large subset of ACA. In particular, somatic mutations in genes regulating intracellular ionic homeostasis and membrane potential have been identified in aldosterone producing adenoma. These mutations all promote increased intracellular calcium concentrations, with activation of calcium signaling, the main trigger for aldosterone production. In cortisol producing adenomas, recurrent somatic mutations in PRKACA (coding for the cAMP-dependent protein kinase catalytic subunit alpha) affect cyclic AMP-dependent protein kinase A signaling, leading to activation of cortisol biosynthesis. In addition to these specific pathways, the Wnt/ $\beta$ -catenin pathway appears to play an important role in adrenal tumorigenesis, as  $\beta$ catenin mutations have been identified in both aldosterone as well as cortisol producing adenomas. This, together with different intermediate states of aldosterone and cortisol cosecretion, raises the possibility that the two conditions share a certain degree of genetic susceptibility. Alternatively, different hits may be responsible for the diseases, with one hit leading to adrenocortical cell proliferation and nodule formation, with the second one specifying the hormonal secretory pattern.

#### 1 **1. Introduction**

Adrenal masses are quite common being found in  $\approx 3\%$  of the general population with the 2 prevalence increasing with age (1). Functional adrenocortical adenomas are a group of benign 3 tumors of the adrenal cortex autonomously secreting excess hormones. Aldosterone producing 4 adenomas (APA) are responsible for primary aldosteronism (PA), the most frequent form of 5 6 secondary hypertension. Cortisol producing adenomas (CPA) are responsible for overt or 7 subclinical Cushing syndrome (CS). In addition, both diseases may also result from different forms of adrenal hyperplasia: bilateral adrenal hyperplasia (BAH) in PA and primary bilateral 8 macronodular adrenal hyperplasia (PBMAH) responsible for overt or subclinical CS. Primary 9 10 pigmented nodular adrenal hyperplasia is a rare condition responsible for overt CS. Unilateral adrenal hyperplasia leading to autonomous aldosterone or cortisol production has also been 11 described. Importantly, in between the two conditions are histological forms of multinodular 12 13 adrenals where hormone production may be localized to one large nodule or to multiple nodules; sometimes co-secretion of aldosterone and cortisol is observed in one adrenal, either 14 15 from one adenoma or from distinct adenomas.

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#### 2. Aldosterone and cortisol biosynthesis in the adrenal cortex

Aldosterone and cortisol are produced in the adrenal cortex, which is composed in humans of 18 three concentric functionally distinct zones. Steroid hormones are produced from cholesterol 19 20 through sequential enzymatic steps involving different cytochrome P450 enzymes and hydroxysteroid dehydrogenases and the functional properties of each zone results from the 21 expression of specific steroidogenic enzymes and the ability to respond to unique regulatory 22 stimuli. In particular, the outer zona glomerulosa produces aldosterone through the activity of 23 aldosterone synthase (encoded by CYP11B2), which catalyzes the three terminal reactions of 24 aldosterone biosynthesis: hydroxylation at position C11 of the steroid intermediate 25 deoxycorticosterone (DOC) yielding corticosterone, followed by a hydroxylation at position 26

27	C18 yielding 18-hydroxy-corticosterone and a subsequent oxidation of the hydroxyl group at
28	C18, which results in the formation of aldosterone. Cortisol biosynthesis occurs in the zona
29	fasciculata, where the expression of $17\alpha$ hydroxylase mediates conversion of 17-deoxy-21-
30	carbon steroids into 17-hydroxy-21-carbon steroids, precursors of cortisol. As 17a
31	hydroxylase is not expressed in rats and mice, the main glucocorticoid in these species is
32	corticosterone (2). Cortisol is synthesized from 17OH-progesterone through 21-hydroxylation
33	to form 11-deoxycortisol, followed by hydroxylation at position C11 by 11 $\beta$ -hydroxylase
34	(encoded by CYP11B1). The two highly homologous genes CYP11B2 and CYP11B1 are
35	located in tandem on human chromosome 8q21-22. Unequal crossing over of the two genes
36	during meiosis results in a chimeric gene composed of the regulatory regions of CYP11B1
37	fused to the coding sequence of CYP11B2, which is responsible for familial
38	hyperaldosteronism (FH) type 1 (also known as glucocorticoid remediable aldosteronism) (3).
39	Under these conditions, aldosterone synthase expression and aldosterone biosynthesis are
40	regulated by ACTH in the zona fasciculata of the adrenal cortex, rather than by angiotensin II
41	(AngII), with a circadian rhythmicity of aldosterone production following that of cortisol (4)
42	and production of the hybrid steroids 18-hydroxycortisol and 18-oxocortisol (5,6).
43	The main regulators of aldosterone biosynthesis are the renin-angiotensin system, the
44	concentration of extracellular potassium and ACTH (7). The renin-angiotensin system
45	translates changes in renal perfusion or sodium concentration sensed by the distal tubule into
46	changes in aldosterone production from the adrenal cortex, in order to maintain volume and
47	electrolyte homeostasis. AngII acts via the AT1 receptor expressed in zona glomerulosa cells,
48	activating a Gaq-phospholipase C mediated pathway leading to increased inositol 1,4,5-
49	triphosphhate (IP3) and 1,2-diacylglycerol concentrations. IP3 increases intracellular calcium
50	concentrations by calcium release from intracellular stores.

AngII also inhibits the background potassium channel TASK (TWIK-related acid-sensitive 51 potassium channel) and the Na<sup>+</sup>,K<sup>+</sup>-ATPase, leading to cell membrane depolarisation and 52 activation of voltage-gated Ca<sup>2+</sup> channels, again increasing intracellular calcium 53 concentrations. Extracellular potassium is the other major regulator of aldosterone production, 54 exerting a large amplitude of control over a wide range of potassium concentrations. This is 55 possible because the main ionic conductance of zona glomerulosa cells is that of K<sup>+</sup> and the 56 cell membrane potential closely follows the equilibrium potential of K<sup>+</sup> over a large range of 57 extracellular K<sup>+</sup> concentrations (8). Elevation of the extracellular K<sup>+</sup> concentration, a decrease 58 in the intracellular K<sup>+</sup> concentration, inhibition of Na<sup>+</sup>, K<sup>+</sup>-ATPase or closure of potassium 59 channels all lead to cell membrane depolarization, opening of voltage-dependent Ca<sup>2+</sup> 60 channels and an increase in intracellular calcium concentrations resulting in activation of 61 calcium signaling. 62

63 Increased calcium signalling affects aldosterone production in several ways. Calcium signalling increases activity of cholesterol ester hydrolase with subsequent release of 64 65 deesterified cholesterol from cytoplasmic stores, promotes cholesterol delivery to the outer mitochondrial membrane and its transfer to the inner mitochondrial membrane by increasing 66 expression of the steroid acute regulatory protein (StAR), and increases the synthesis of 67 cofactors required for p450 cytochrome enzymes (9). Stimulation of CYP11B2 transcription is 68 mediated by calcium/calmodulin binding and activation of protein kinases regulating 69 phosphorylation of several transcription factors, in particular nuclear receptor subfamily 4 70 group A (NR4A) members 1 and 2 (NURR77 or NGF1B and NURR1), the cyclic AMP-71 72 dependent transcription factor ATF-1 and the cyclic AMP-responsive element-binding protein (CREB) (10) (Figure 1A). 73

Cortisol production is regulated by the hypothalamus-pituitary-adrenal (HPA) axis, whereby
 circadian input and stress stimuli induce release of cortisol via corticotrophin releasing

hormone (CRH) -dependent ACTH secretion and cortisol signals back to exert negative 76 feedback on the hypothalamus and the pituitary. In the zona fasciculata (and zona reticularis) 77 of the adrenal cortex, ACTH binds to its receptor MC2R (Melanocortin receptor 2), which 78 79 signals through a Gas mediated signaling to increase intracellular cAMP levels and to activate protein kinase A (PKA) (Figure 1B) (11). Acute stimulation by ACTH elicits effects on 80 steroidogenesis by regulating cholesterol availability, mobilizing cholesterol from lipid stores 81 82 and increasing cholesterol transport into mitochondria and by stimulating transcription of steroidogenic genes. Rapid effects occurring within minutes mainly affect StAR activity 83 through phosphorylation, while longer stimulation (days) by ACTH results in transcriptional 84 regulation of StAR as well as steroidogenic enzymes. In particular, PKA signaling activates 85 transcription factors regulating expression of steroidogenic genes, including steroidogenic 86 factor 1 (SF-1), CREB, CRE modulator (CREM), CCAAT/enhancer-binding proteins 87 88 (C/EBPs), and activator protein 1 (AP-1) (11). ACTH also regulates aldosterone production acutely via cAMP mediated pathways, whereas chronically, ACTH suppresses plasma 89 90 aldosterone both in human and animal models and downregulates CYP11B2, inducing 91 *CYP11B1* expression and promoting a zona fasciculata phenotype (7,12). Long-term exposure to ACTH (weeks or months) stimulates adrenal growth and promotes adrenal cell hypertrophy 92 and hyperplasia. In extreme cases of glucocorticoid deficiency or ACTH excess, as well as in 93 experimental models of CRH or ACTH deficient mice, the trophic effects of the hormone lead 94 to major hyperplasia or atrophy respectively of the inner zones of the adrenal cortex (12). In 95 addition to acting through cAMP, ACTH also stimulates calcium influx in glomerulosa cells 96 by activation of L-type  $Ca^{2+}$  channels (9). Many other factors have been shown to modulate 97 aldosterone and cortisol production, including neurotransmitters, cytokines, vascular products, 98 99 and adipokines, as well as locally produced ACTH acting through a paracrine mechanism (12, 13).100

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# 102 **3. Somatic mutations in APA**

PA is due to autonomous aldosterone production from the adrenal cortex and is diagnosed on 103 104 the basis of hyperaldosteronism associated with suppressed renin and often hypokalemia (14). It is the most common form of secondary hypertension (HT), with a prevalence of 5-10% in 105 patients with arterial hypertension (15,16) and up to 20% in patients with resistant HT (17). 106 107 PA is associated with increased cardiovascular risk, in particular coronary artery disease, 108 nonfatal myocardial infarction, heart failure, atrial fibrillation, renal damage and stroke (18-20), due to blood-pressure independent effects of aldosterone on cardiac and vascular 109 110 remodeling (21,22), The majority of cases of PA are due to an APA or to bilateral adrenal hyperplasia (BAH, also 111 112 called idiopathic hyperaldosteronism). Only in rare cases has unilateral adrenal hyperplasia 113 been described, and aldosterone producing adrenocortical cancers are extremely rare. APA are generally small (< 2 cm in diameter) and are diagnosed by adrenal CT and adrenal venous 114 115 sampling, which identifies lateralized aldosterone secretion in the presence or absence of a 116 visible adrenal nodule at imaging (14,23). Recent advances in genome technology have

allowed identification, during the last five years, of several somatic mutations in APA,

118 providing a pathogenic model for PA development. Whole exome sequencing of paired

tumour (somatic) and germline DNA performed by different groups on a total of more than

120 100 APA samples has identified recurrent somatic mutations in genes coding for ion channels

121 (*KCNJ5* (24) and *CACNA1D* (25,26)) and ATPases (*ATP1A1* and *ATP2B3*, (25,27))

regulating intracellular ionic homeostasis and cell membrane potential, thus defining APA as

a channelopathy. These mutations all promote increased intracellular calcium concentrations,

124 leading to activation of calcium signaling, the main trigger for aldosterone biosynthesis

125 (Figure 1). A small fraction of APA also carries somatic mutations in *CTNNB1*, coding for  $\beta$ -

catenin, similar to CPA (see below) and adrenocortical cancer (28,29) (Table 1). In the largest 126 127 multicenter study published, exploring the prevalence of somatic mutations in KCNJ5, CACNA1D, ATP1A1 and ATP2B3 in 474 patients from seven centers belonging to the 128 129 ENS@T network (European Network for the Study of Adrenal Tumors), somatic mutations were detected in 54% of APA, ranging from 27.2% to 56.8% across different centers (30). In 130 this study, KCNJ5 mutations were the most prevalent genetic abnormality and were found in 131 132 38% of APA. Mutations affecting CACNA1D were the second most prevalent genetic abnormalities being present in 9.3% of cases, while ATP1A1 and ATP2B3 mutations were 133 identified in 5.3% and 1.7% of cases respectively. There was no difference in cellular 134 135 composition (percentage of cells resembling to zona fasciculata (ZF) or zona glomerulosa (ZG)) of APA or in CYP11B2, KCNJ5, CACNA1D or ATP1A1 gene expression in APA across 136 genotypes. Patients with KCNJ5 mutations were more frequently female, diagnosed younger 137 138 and with higher plasma potassium concentrations when compared to CACNA1D mutation carriers or non-carriers. CACNA1D mutations were associated with smaller adenomas. There 139 140 was no association between the mutation status and preoperative plasma aldosterone or renin 141 levels, the aldosterone to renin ratio or the number of medications taken before surgery. There was also no association with post-operative blood pressure outcomes, treatment scores at 142 follow-up, cure or improvement of hypertension. These associations were largely center-143 dependent, which might be due to the heterogeneity of APA that has to be taken into account 144 when considering genotype-phenotype correlations (30). 145 Furthermore, less frequent mutations have been identified in a few genes in patients with 146 peculiar phenotypes of PA, including early-onset PA and tumors cosecreting aldosterone and 147 cortisol (31,32) (Table 1). Recently, somatic mutations in PRKACA (coding for the cAMP-148 dependent protein kinase catalytic subunit alpha) have been reported in patients with APA 149 (32). Using whole exome and targeted sequencing in 122 patients, Rhayem et al identified 150

two somatic heterozygous mutations in *PRKACA*, one of them being the p.Leu206Arg
mutation previously reported in CPA (see below). The carrier of this variant had biochemical
Cushing syndrome and high expression of 11β-hydroxylase in the tumor. The second
p.His88Asp variant was not associated with cortisol excess, nor with a gain of function of the
PKA catalytic activity, leaving open the question of its role in the development of PA.

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## 3.1. Pathogenic mechanisms of APA development

KCNJ5 codes for the G protein-activated inward rectifier potassium channel GIRK4 158 (alternative protein name Kir3.4), which is composed of two membrane-spanning domains 159 160 (M1 and M2), one pore forming region (H5) and cytoplasmic N- and C-termini that contribute to the pore structure. Recurrent somatic mutations identified in APA cluster within or near the 161 selectivity filter of the channel pore, between amino acids Thr149 to Gly153 (Table 2). They 162 163 affect its ionic selectivity, rendering the channel permeable to sodium, with increased sodium influx into the cell leading to chronic cell membrane depolarization and opening of voltage-164 gated calcium channels (24). This in turn, leads to increased intracellular calcium 165 concentrations and activation of calcium signaling (Figure 1A). Transient transfection of 166 167 KCNJ5 mutants into adrenocortical cells increase aldosterone production and expression of 168 CYP11B2 in a calcium-dependent manner (33); in some cases, mutations lead to reduced membrane and total abundance of GIRK4 (34). Surprisingly, transiently transfected KCNJ5 169 mutants did not lead to enhanced cell proliferation ex vivo, but rather to reduced cell viability 170 171 or sodium dependent cell death, which does not explain the cell proliferation and APA formation (33,35). Whether these results are due to experimental artefacts, due to high 172 expression of transfected channels leading to excessively increased sodium influx into the 173 cell, which does not reflect the in vivo situation, or whether additional hits are required for 174 cell proliferation, remains a matter of debate. 175

176	<i>CACNA1D</i> codes for the $\alpha$ 1 subunit of the L-type voltage gated Cav1.3 calcium channel
177	(calcium channel, voltage-dependent, L type, alpha-1d subunit), whereas ATP1A1 codes for
178	the $\alpha$ 1 subunit of the Na <sup>+</sup> , K <sup>+</sup> -ATPase and ATP2B3 for the plasma membrane calcium-
179	transporting ATPase 3 (PMCA3). Na <sup>+</sup> ,K <sup>+</sup> -ATPase and PMCA3 are members of the P-type
180	family of ATPases and are composed of ten transmembrane domains $(M1 - M10)$ with
181	intracellular N- and C-tails. ATP1A1 mutations affect amino acids in transmembrane helices
182	M1, M4 and M9 of the $\alpha$ 1 subunit of the Na <sup>+</sup> , K <sup>+</sup> -ATPase (Table 2). They lead to a loss of
183	pump activity and a reduced affinity for K <sup>+</sup> , as well as to an inward proton or sodium leak,
184	which is proposed to increase aldosterone production through cell membrane depolarization
185	and increased calcium influx, as is observed in the presence of KCNJ5 mutations (25,27).
186	However, transient transfections of Na <sup>+</sup> , K <sup>+</sup> -ATPase $\alpha$ 1 subunits carrying the p.Leu104Arg,
187	p.Val332Gly or p.Gly99Arg mutations in adrenocortical NCI-H295R cells did not modify
188	basal cytosolic calcium levels and hardly increased potassium-stimulated calcium
189	concentrations, despite depolarizing adrenal cells and stimulating aldosterone secretion (36).
190	Rather they induced increased intracellular acidification which is suggested to play a role in
191	regulating aldosterone biosynthesis in this context (Figure 1A) (36).
192	Mutations of PMCA3 are located in the transmembrane domain M4 and are all deletions of
193	multiple amino acids in the region between Leu425 and Leu433 (Table 2).
194	Electrophysiological and functional studies have shown that the p.Leu425_Val426del
195	mutation leads to reduced $Ca^{2+}$ export due to loss of the physiological pump function and
196	increased $Ca^{2+}$ influx due to opening of depolarization-activated $Ca^{2+}$ channels. A $Ca^{2+}$ leak
197	through the mutated pump has also been observed, inducing CYP11B2 expression and
198	aldosterone biosynthesis (Figure 1A) (37).
199	Finally, CACNA1D mutations affect the function of the L-type $Ca^{2+}$ channel $\alpha$ -subunit $Ca_v 1.3$ ,

200 which is composed of four repeat domains (I to IV), each one consisting of six

transmembrane segments S1-S6 and a membrane associated loop between S5 and S6 (Table
2) (38). Whereas the S4 segments are involved in voltage sensing, the S5 and S6 domains as
well as the loop in between line the pore of the channel (39). 18 different *CACNA1D*mutations have been identified (25,26,30); they are gain-of-function mutations leading to a
shift of voltage-dependent channel activation towards more negative voltages, or to reduced
inactivation of the channel, followed by increased intracellular calcium concentrations with
induction of aldosterone biosynthesis (Figure 1A) (25,26).

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#### **3.2.** Genotype-phenotype correlations in APA

210 KCNJ5 mutations are the most frequent type of somatic mutation within APA (Table 1). Their

reported frequency ranges from 13-77%, with higher frequency in studies from Japan, China,

Korea and Taiwan (40-46). CACNA1D mutations are the second most frequent mutations (up

to 10%), followed by ATP1A1 and ATP2B3. Three studies have also explored CTNNB1

mutations, which are found in 2.1 - 5.1% of cases (26,28,29).

215 The correlations of clinical and molecular parameters with the mutation status are

216 heterogeneous among studies (Table 1). In a recent meta-analysis, Lenzini et al analyzed the

clinical correlates of *KCNJ5* mutations in 1636 patients from 13 studies (47). The overall

218 prevalence of *KCNJ5* mutations was 43%, ranging from 12-80% among different studies,

219 with a higher prevalence in studies from Japan and China compared to studies on populations

from Europe, US and Australia. Carriers of *KCNJ5* mutations were younger, more often

221 females, had larger tumors and had higher plasma aldosterone levels when compared to non-

222 carriers. No association was observed between the mutation status and systolic or diastolic

blood pressure and serum potassium levels overall, confirming previous results from the study

by Fernandes-Rosa et al (30). An association between the presence of KCNJ5 mutations and

the lateralization index at adrenal vein sampling was described in two studies ((42,48)

, but not replicated in another study (49). In four studies, *KCNJ5* mutations were associated
with tumors composed mainly of large, lipid-laden cells resembling zona fasciculata cells
(Table 1). The other mutations were associated mainly with male sex and smaller tumors (2528,30,45,50), and the cellular phenotype of APA carrying *CACNA1D*, *ATP1A1* or *ATP2B*mutations was more heterogeneous than *KCNJ5* mutated tumors. Although few studies have
investigated *CTNNB1* mutations in large cohorts, they seem to be more prevalent in females
(26,28,29,51).

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#### 4 **3.3. Germline mutations in APA**

In addition to somatic mutations, the occurrence of germline CACNA1H (coding for the 235 voltage-dependent T-type calcium channel subunit alpha-1H) mutations, and rarely, germline 236 KCNJ5 and ARMC5 variants has been reported in PA (31,52,53). Daniil et al performed 237 238 whole exome sequencing in patients with different phenotypic presentations of PA, including 23 subjects with APA, 10 patients with a family history of PA and one subject with early-239 240 onset PA. CACNA1H mutations were identified in early onset PA and patients with a family 241 history, and a germline CACNA1H p.Val1951Glu mutation was identified in one patient with an APA (31). CACNA1H encodes the pore-forming al subunit of the T-type voltage-242 dependent calcium channel Cav3.2, which has been implicated in membrane potential 243 oscillations in ZG cells (54). The variant is located in the C-terminal cytoplasmic domain of 244 Cav3.2, in a region possibly implicated in fast channel activation. Electrophysiological 245 investigation revealed a gain-of-function phenotype, whereby p.Val1951Glu induced a faster 246 recovery from inactivation and marked increase in voltage-dependent facilitation, two 247 properties which could favor larger calcium entry during repetitive electrical activity. In 248 H295R adrenocortical cells, transient transfection of the mutant channel led to an increase in 249 aldosterone biosynthesis and expression of CYP11B2 (31). 250

Genetic abnormalities in APA driver genes are present as germline mutations in some familial 251 252 forms of PA, which account for 1 to 5% of cases and are transmitted as an autosomal dominant trait. Although the terminology for FH is still evolving, to date four different forms 253 254 of familial hyperaldosteronism, FH-I to FH-IV, have been described. FH-I, resulting from the formation of a chimeric gene between CYP11B2 and CYP11B1 (see above), is found in 0.5% 255 to 1.0% of cases of PA (55-57), but may represent up to 3% of cases in the paediatric 256 257 hypertensive population (58). FH-I is associated with BAH, but in rare cases adrenocortical 258 adenomas have been described. Heterozygous germline mutations in KCNJ5 are found in FH-III, a severe form of PA presenting in typical cases in childhood with severe hypertension and 259 260 profound hypokalemia, due to massive bilateral adrenal hyperplasia which requires bilateral adrenalectomy to control the hypertension (24). In other cases, the disease is milder and with 261 normal appearing adrenals on imaging studies (59). Germline mutations in CACNA1H have 262 263 recently been identified as the cause of a new form of early onset PA associated or not with a developmental disorder (31,60). In familial cases, the disease (named FH-IV) is transmitted as 264 an autosomal dominant trait. Similar mutations are also found in patients diagnosed as FH-II 265 with or without an adrenal adenoma as well as in APA (31). Finally, de novo germline 266 CACNA1D mutations have been described in a rare disease called PASNA (Primary 267 268 aldosteronism, seizures, neurological abnormalities), with no adrenal abnormality reported on imaging (26). 269

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# **4.** Pathological and genetic heterogeneity and the origin of APA

While the functional link between mutations in driver genes in APA and aldosterone
production has been clearly demonstrated, the origin of APA in terms of increased cell
proliferation and nodule formation is not yet well understood, and work from different groups
has provided contradictory results on the sequence of events leading to aldosterone

overproduction and adenoma formation. Although the fact that familial forms of PA are due 276 277 to germline mutations in driver genes supports a role for these mutations in adrenal cortex cell proliferation and nodulation, adrenal morphology is highly variable among these subjects. 278 279 Patients with FH-III harboring germline *KCNJ5* mutations may exhibit adrenal phenotypes ranging from a normal adrenal to massive bilateral adrenal hyperplasia or adrenal nodules 280 (24,61,62). Similarly, patients carrying germline CACNA1H mutations show different adrenal 281 282 phenotypes including normal, hyperplastic and adenomatous adrenals (31,60). No abnormality in adrenal morphology was observed in patients with PASNA 283 harboring germline CACNA1D mutations (26). Ex vivo, adrenocortical cells overexpressing 284 285 mutant GIRK4 channels show increased intracellular Na<sup>+</sup> concentration leading, counterintuitively, to reduced cell proliferation and sodium-dependent apoptosis (33,35). 286 It has been previously shown that adrenals with APA exhibit adrenal cortex remodeling, 287 288 decreased vascularization, increased nodulation and ZG hyperplasia (63,64). These findings suggest that adrenal cortex remodeling may precede APA development and that somatic 289 290 mutations in driver genes are secondary events specifying the pattern of hormone secretion. The 291 recent development of an aldosterone-synthase antibody (65) has allowed assessment of the genetic and histological characteristics of adrenals with APA and has reinforced the hypothesis 292 293 of independent events leading to nodulation and aldosterone overproduction. Two studies in particular have identified different genetic abnormalities in different aldosterone producing 294 nodules from the same adrenal. In one study, Dekkers et al analyzed 28 multinodular adrenals 295 and identified five adrenals with more than one nodule staining positively for aldosterone 296 synthase; in one adrenal two aldosterone-synthase positive nodules each harbored a different 297 KCNJ5 mutation (66). In the second study, assessment of the KCNJ5, ATP1A1, ATP2B3 and 298 CACNA1D mutation status in aldosterone synthase positive nodules from 27 multinodular 299 adrenals with APA identified 7 adrenals harboring different mutations between the principal 300

nodule and the secondary nodule within the same adrenal, including two adrenals with an APA 301 302 harboring a CACNA1D mutation and in one secondary nodule, a KCNJ5 mutation (67). Further studies extended the heterogeneity of aldosterone synthase expression to the APA itself, with 303 304 KCNJ5 mutations only identified in APA regions positive for aldosterone synthase (68). Finally, in a young PA patient with a germline APC mutation and a multinodular adrenal with 305 somatic biallelic inactivation of APC, we identified a somatic KCNJ5 mutation in the nodule 306 expressing aldosterone synthase only (see below) (69). These data support a two-hit model for 307 308 APA formation whereby a specific genetic or epigenetic event is responsible for adrenal cortex remodeling and nodule formation and somatic mutations in driver genes leading to excessive 309 310 aldosterone secretion represent second hits specifying the pattern of hormonal secretion (Figure 2). 311

A second model for the origin of APA has emerged from genetic studies performed on 312 aldosterone-producing cell clusters (APCC). APCC, which are found in normal adult adrenals 313 314 and in adrenals with APA, are composed by ZG-like cells expressing disabled 2 (Dab2, a 315 marker of ZG) in contact with the capsule and inner columnar Dab2-negative ZF-like cells, which do not express CYP11B1, forming cords along sinusoids (70,71). APCC cells are 316 positive for CYP11B2 and 3BHSD staining, but not for CYP17, consistent with their capacity 317 to synthesize aldosterone (70). Targeted next generation sequencing was performed on the 318 DNA of 23 APCC microdissected from adrenals from kidney transplant donors; somatic 319 320 mutations in CACNA1D, ATP1A1 and ATP2B3, similar to those already described in APA, were 321 identified in 35% of APCC (72). The authors proposed that these findings are consistent with 322 the development of APA from APCC (Figure 2) and suggest different hypotheses for this progression: single somatic mutations responsible for aldosterone production and cell 323 proliferation or second-hit mutations within the APCC leading to cell proliferation. In some 324 325 cases APCCs may represent terminal lesions not having the capacity to progress towards an

adenoma (72). Interestingly, no KCNJ5 mutations, the most frequent genetic alteration 326 327 observed in APA, were identified in APCC. The authors suggest that this could be explained by a rapid progression of KCNJ5 mutated APCC to APA (72). Subsequently, CACNA1D 328 329 mutations were identified in 17 of 26 APCC from patients with lateralized PA negative for detectable masses at imaging, suggesting that these lesions may contribute to PA in this 330 331 situation (73). Recently, a novel aldosterone-producing structure named a possible APCC-to-APA transitional lesion (pAATL) was described in multinodular adrenals from two patients 332 with lateralized PA, which was suggested to be an intermediate lesion between APCC and APA 333 (74). While the APCC-like portion consists of a subcapsular ZG-like region and an inner ZF-334 335 like region expressing CYP11B2 but not CYP11B1, the micro-APA-like portion is composed of a heterogeneous mixture of CYP11B1-positive and CYP11B2-positive cells. Sequencing of 336 the known APA-associated mutations in three pAATL identified a KCNJ5 mutation and an 337 338 ATP1A1 mutation in the micro-APA-like portion of two pAATL, but not in their corresponding APCC-like portions. The authors suggest that these data support the hypothesis that the micro-339 340 APA derived from the APCC following the appearance of an APA driver mutation (74). However, another pAATL carried two novel ATP1A1 mutations in both the mAPA-like and 341 APCC-like portions, in this case supporting a clonal origin of the two portions (74). New 342 343 genetic studies in a larger number of normal and pathological adrenals with APCC are necessary to clarify the relationship between these aldosterone-producing lesions and the origin of APA. 344

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### 346 5. Genetic abnormalities in Cortisol Producing Adenomas

The genetics of CPA was unraveled in 2014 by the work of four independent groups
identifying somatic mutations of *PRKACA* as a cause of CS. Beuschlein and coworkers
performed WES in 10 paired adrenal tumor and germline DNA samples from patients with
CS, which revealed the presence of a recurrent somatic p.Leu206Arg mutation in the

PRKACA gene in 7 cases, with an additional p.Leu199\_Cys200insTrp mutation in one case 351 (75). Sequencing of 129 additional samples identified the p.Leu206Arg mutation in 22 of 59 352 CPA (37%) from patients with overt CS, but not in tumors from 40 patients with subclinical 353 354 hypercortisolism (sCS), 20 APA, 20 non-functioning adenomas or 42 adrenocortical carcinomas. Interestingly, among patients with different forms of bilateral adrenal hyperplasia 355 leading to CS, including bilateral micro- or macronodular hyperplasia, a germline copy-356 357 number gain (duplication) of the genomic region on chromosome 19 that includes PRKACA 358 was identified in 5 patients from four kindreds. In one family with bilateral macronodular hyperplasia, the genetic defect was transmitted in an autosomal dominant manner from the 359 360 mother to her affected child. The p.Leu206Arg mutation is located in the highly conserved core of the interaction between the regulatory (RII $\beta$ ) and catalytic (C $\alpha$ ) subunits of PKA. In 361 vitro studies demonstrated gain-of-function of the PKA catalytic subunit mutants with 362 363 impaired inhibition by the PKA regulatory subunit, and increased protein expression of the PKA catalytic subunit in cells from patients with germline chromosomal gains, with both 364 365 mechanisms leading to increased PKA activity. Basal PKA activity in cells transfected with mutant Ca subunits was increased and was not reduced by excess nonmutant RIIB subunit 366 expression, nor was it responsive to cAMP analogues, indicating that the mutations made the 367 368 catalytic subunit resistant to the physiological suppression. The presence of *PRKACA* mutations was associated with a more severe phenotype in patients with overt CS and 369 PRKACA mutation carriers had significantly increased expression of MC2R, StAR, CYP21A1, 370 HSD3B2 and CYP11A1 mRNA in adrenal tumors. These results were confirmed in the three 371 372 other studies. By performing WES in 25 tumor samples producing excess cortisol, including 22 CPA and 3 ACC, Goh et al identified six cases carrying the recurrent p.Leu206Arg 373 374 substitution (76). Six cases had gain-of-function mutations in CTNNB1 and two cases in GNAS (coding for Gas), with PRKACA, GNAS and CTNNB1 mutations being mutually 375

exclusive. Additional targeted sequencing in 38 patients identified the p.Leu206Arg 376 substitution collectively in 13 out of 63 tumors representing 24% of all ACA and 35% of all 377 ACA associated with overt CS. In this study, adenomas carrying PRKACA or GNAS 378 379 mutations were significantly smaller than those without these mutations, and the patients were of younger age at diagnosis with overt CS. WES also revealed a group of tumors harboring 380 many somatic copy number variants with frequent deletion of CDC42 and CDKN2A, 381 382 amplification of 5q31.2 and protein-altering mutations in TP53 and RB1. This mutational spectrum is more similar to that of malignant tumors, indicating that they may have greater 383 malignant potential or represent early steps towards the development of ACC. 384 385 A higher rate of somatic *PRKACA* mutations seems to occur in subjects with an Asian ancestry. Cao et al (77) reported the occurrence of the p.Leu205Arg mutation (equivalent to 386 387 the p.Leu206Arg mutation in the other reports) in 65.5% of a total of 87 CPA from China 388 screened by WES and targeted sequencing. In this study, there was no significant correlation between the mutation status and clinical or biological parameters. The authors also identified 389 390 one somatic PRKAR1A mutation, one CTNNB1 mutation, a truncating APC mutation and two 391 activating GNAS mutations in CPA; an additional CTNNB1 mutation was found in an adrenal oncocytoma (a very rare form of cortisol producing adrenal tumor). The authors also 392 identified the recurrently mutated gene *DOT1L* coding for a histone H3 methyltransferase as 393 well as one HDAC9 mutation in ACTH-independent macronodular adrenocortical hyperplasia 394 (AIMAH), suggesting involvement of chromatin deregulation, and CLASP2 mutations in 2 of 395 three adrenocortical oncocytomas. RNA-sequencing data comparing PRKACA mutated and 396 397 non-mutated tumors identified 232 differentially expressed genes, with significant enrichment of gene ontology terms "biosynthesis and metabolism of steroid and cholesterol" and 398 "response to chemical stimulus", with increased expression of the genes StAR, MC2R, 399 GSTA1, CXCL2, and S100A8/9. 400

Finally, in a Japanese cohort, 50% of cases (4/8) with adrenocortical adenomas associated 401 with corticotropin-independent Cushing's syndrome were carriers of the PRKACA 402 p.Leu206Arg mutation (78). An additional patient carried a recurrent p.Arg201Cys GNAS 403 404 mutation. Follow-up sequencing in an additional 57 cases identified the PRKACA p.Leu206Arg mutation in 30 cases and GNAS mutation in 10 cases, representing a total of 405 52.3% and 16.9% of mutational events and which were mutually exclusive. These mutations 406 407 were found in 76% of patients with overt CS; interestingly, two patients with sCS were also carriers of a PRKACA and a GNAS mutation. In this study, carriers of the recurrent PRKACA 408 p.Leu206Arg mutation, but also those with GNAS mutations had higher cortisol levels after an 409 410 overnight dexamethasone suppression test and smaller adenoma size, suggesting a higher cortisol output compared to non-mutated tumors. 411

412 After these seminal works, a similar prevalence of *PRKACA* mutations was found in different 413 cohorts of CS patients from different countries (Table 3). The largest study published to date was performed within the ENS@T and included 149 samples from nine different European 414 415 centers, among which were 64 CPA from patients with overt CS, 36 CPA samples from 416 patients with sCS, 32 nonsecreting adenomas, 4 androgen producing tumors, 5 ACC and 8 samples from PBMAH. Heterozygous somatic PRKACA mutations were found in 22/64 417 samples from patients with overt CS (34%). 18 patients carried the recurrent p.Leu206Arg 418 mutation, while a new mutation p.Cys200\_Gly201insVal was identified in three patients and a 419 p.Ser213Arg+p.Leu212\_Lys214insIle-Ile-Leu-Arg mutation was identified in one patient. 420 These newly identified mutations involved a region implicated in the interaction between the 421 422 PKA regulatory and catalytic subunits. As in previous studies, patients with PRKACA mutations showed more severe cortisol abnormalities (higher levels of cortisol after an 423 overnight dexamethasone suppression test) and smaller adenoma size compared with non-424 carriers (75,76). In a study exploring 60 patients with overt (n=36) or subclinical (n=24) CS, 425

Thiel et al (79) identified the recurrent somatic PRKACA p.Leu206Arg mutation in 23.1% of 426 tumors, somatic CTNNB1 mutations (p.Ser45Pro, p.Ser45Phe) in 23.1%, the recurrent GNAS 427 p.Arg201Cys mutation in 5.8%, and a double mutation of CTNNB1 and GNAS in one case. 428 429 *PRKACA* and *GNAS* mutations were again mutually exclusive. In one patient with unilateral hyperplasia, a somatic PRKACA p.Leu206Arg mutation was identified, while patients with 430 bilateral adenomas did not have known somatic mutations. The presence of *PRKACA* 431 432 mutations was associated with younger age, overt Cushing's syndrome, and higher cortisol levels after an overnight dexamethasone suppression test compared with non-PRKACA-433 mutant or CTNNB1-mutant lesions (79). Lastly, targeted genetic screening for GNAS, 434 PRKACA, and KCNJ5 mutations in Japanese patients, 15 with CS, 9 with sCS, and 33 with 435 an APA, found mutations in the PRKACA gene in 4 (26%) patients with CS, while mutations 436 in the GNAS gene were detected in 2 (13%). A mutation in the GNAS gene was detected in 1 437 438 patient with sCS (80,81). Altogether, more than 580 patients with CPA or hyperplasia have been investigated. Somatic 439 440 PRKACA mutations were found in 28%, CTNNB1 mutations in 4%, and GNAS mutations in 4% of cases (Table 3). PRKACA mutations were associated with overt CS, more severe 441

442 cortisol abnormalities and smaller tumors. The question as to whether other recurrent

443 mutations or genetic abnormalities are found in CPA has been addressed by Ronchi et al, who

444 performed WES in 99 patients with ACA, including 74 CPA and 25 non-functioning

adenomas negative for *PRKACA* mutations (82). Different mutations affecting genes of the

446 cAMP/PKA and Wnt/ $\beta$ -catenin signalling pathway were identified (Figure 1B and 1C), the

447 latter are more frequently associated with larger tumors and endocrine inactivity. Genetic

448 variants were also identified in genes belonging to the calcium signalling pathway. To what

449 extent these single variants are causative for cortisol production and/or adenoma formation

450 remains to be established.

452	6. Genetics of aldosterone and cortisol co-secretion
453	Co-secretion of aldosterone and cortisol has been described in several reports and biochemical
454	abnormalities of cortisol excess in PA have been associated with worse cardiovascular
455	outcomes compared to APA without cortisol secretion (83). Recent studies suggest that
456	cortisol excess may be more frequent in patients with APA than previously thought.
457	Histological examination of adrenals with APA has consistently shown co-expression of
458	CYP11B2 and CYP11B1 in a number of cases (84).
459	A Japanese study explored somatic GNAS, PRKACA, and KCNJ5 mutations in 15 patients
460	with CS, 9 patients with sCS, and 33 patients with APA (80,81) (Table 4). From 33 APA
461	tested, 10 had autonomous cortisol secretion based on an overnight dexamethasone
462	suppression test, increased midnight cortisol and/or decreased plasma ACTH levels.
463	Mutations in the PRKACA gene were found in 4 (26%) patients with CS, while mutations in
464	the GNAS gene were detected in 2 (13%). Mutations in the GNAS gene were also detected in 1
465	patient (11%) with sCS. Twenty-four patients with APA (72%) harboured KCNJ5 mutations.
466	Among APA patients with autonomous cortisol secretion, six carried a somatic KCNJ5
467	mutation, while two had a somatic GNAS mutation. KCNJ5 and GNAS mutations in APA
468	were mutually exclusive. No PRKACA mutations were identified in patients with APA (80).
469	In another study, the same authors identified a KCNJ5 mutation in 2 out of 3 patients with
470	APA and sCS (85).
471	In a study exploring 60 patients with cortisol excess, Thiel et al identified 4 cases with
472	aldosterone co-secretion, 2 with overt and 2 with subclinical CS. Among those, 2 were
473	carriers of mutations in KCNJ5 (79). Finally, Rhayem et al identified somatic PRKACA
474	mutations in two patients diagnosed with PA, a newly identified p.His88Asp variant and the

recurrent p.Leu206Arg mutation (32). The patient with the recurrent p.Leu206Arg had 475 476 biochemical CS, while the carrier of the p.His88Asp variant had normal cortisol levels. A distinct histopathological entity is the adrenal harboring two nodules each one secreting a 477 478 different hormone. Fallo and coworkers have investigated the occurrence of concurrent PA and subclinical cortisol hypersecretion in a prospective series of 76 consecutive patients with 479 PA (86). Three out of 76 patients failed to appropriately suppress cortisol after an overnight 480 481 dexamethasone suppression test; one of them also showed suppressed ACTH levels and mildly elevated urinary cortisol excretion. The resected adrenal showed a 4-cm nodule 482 expressing CYP11B1 by in-situ hybridization, whereas CYP11B2 expression was restricted to 483 484 the peritumoral region composed of zona glomerulosa-like cells, suggesting the co-existence of a cortisol-producing adenoma and an aldosterone-producing hyperplasia in the same 485 adrenal. Namba et al described a patient with lateralized PA and CS, in whom examination of 486 487 the adrenal revealed two distinct nodules, one expressing CYP11B1 and carrying a p.Leu206Arg PRKACA mutation, while the other nodule expressed CYP11B2 and carried a 488 489 somatic KCNJ5 mutation (87). 490 Altogether these studies raise a certain number of questions about the histological, cellular and genetic nature of adrenal lesions producing aldosterone and cortisol. In particular, they 491

emphasize the need to explore the common pathways leading to APA and CPA in terms of

493 signals triggering cell proliferation and/or defining hormonal output.

494

# 495 **7. Common hits for APA and CPA**

### 496 **7.1.** The role of the Wnt/β-catenin pathway in adrenal tumorigenesis

497 The role of the Wnt/ $\beta$ -catenin pathway has emerged as crucial in adrenal development and

498 disease.  $\beta$ -catenin, as well as ARMC5 (Armadillo repeat-containing protein 5, see below), are

499 both members of the Armadillo repeat containing (ARMC) gene family. Armadillo repeats are

approximately 40 amino acid long tandemly repeated domains which usually fold together to 500 form a single, rigid protein domain called the armadillo (ARM) domain. β-catenin is a key 501 downstream component of the canonical Wnt signaling pathway. In the absence of Wnt 502 503 ligand, β-catenin is in complex with AXIN1, AXIN2, APC, CSNK1A1 and GSK3B, which promotes its phosphorylation on N-terminal Ser and Thr residues, ubiquitination and 504 subsequent degradation by the proteasome. In the presence of Wnt ligand, the complex is 505 506 disrupted and  $\beta$ -catenin accumulates in the nucleus, where it acts as a coactivator for 507 transcription factors of the TCF/LEF family, leading to transcriptional regulation of Wnt responsive genes. 508

509 Modulation of different components of the Wnt/β-catenin pathway in mice leads to adrenal abnormalities. In the absence of Wnt4, one of the main ligands of the Wnt/ $\beta$ -catenin pathway, 510 mice die immediately after birth due to organ defects such as kidney dysfunction or poor 511 512 pituitary gland development. Despite normal adrenal morphology, Wnt4 knockout animals produce less aldosterone due to decreased expression of *cyp11b2* and show reduced 513 514 expression of other markers of ZG cells, suggesting a defect in ZG development in the 515 absence of Wnt4 (88). As observed in mice, loss of function mutations of WNT4 in humans lead to dysgenesis of the kidneys and to female and male sex reversal; adrenal development 516 and external genitalia are also affected in human fetuses (89). Along the same lines, the 517 518 targeted disruption of  $\beta$ -catenin restricted to sf-1 expressing cells (adrenal cortex, somatic cells of the gonads, spleen, pituitary gonadotropes and ventromedial hypothalamic nucleus 519 (90) leads to complete adrenal aplasia when  $\beta$ -catenin is totally absent during development 520 521 and to a defect in the maintenance of adrenocortical tissue due to increased apoptosis in adult mice when the level of expressed  $\beta$ -catenin is low (91). In H295R cells, a human 522 523 adrenocortical cell line with activating *CTNNB1* mutations, a decrease in  $\beta$ -catenin expression induces cell cycle arrest, changes in cell morphology, modification of the expression of 524

epithelial to mesenchymal transition markers and a modification of cell motility (92). 525 526 Whereas these results highlight the central role of  $\beta$ -catenin in adrenal development, tight regulation of its activation is also crucial. Mice carrying a heterozygous mutation of APC, 527 528 leading to the formation of a truncated protein and constitutive  $\beta$ -catenin activation, exhibit multiple intestinal tumors. In addition these mice also develop hyperaldosteronism, 529 hypervolemia and increased blood pressure, but their adrenal phenotype has not been explored 530 (93,94). Mice expressing a constitutively active  $\beta$ -catenin in the adrenal cortex exhibit 531 progressive adrenal hyperplasia and dysplasia leading, with time, to the formation of nodules 532 in both cortex and medulla; abnormal vascularization was also observed, and in later stages in 533 534 rare cases, development of adrenal carcinoma (95). In humans, activation of  $\beta$ -catenin, demonstrated by its cytoplasmic and/or nuclear accumulation, was found in a large number of 535 adrenocortical tumors, including adrenocortical adenomas (71,96-98). Mutations in the 536 *CTNNB1* gene, leading to constitutive activation of  $\beta$ -catenin were found in both malignant 537 and benign tumors (Figure 1C) (96,97). In benign tumors, the presence of  $\beta$ -catenin mutations 538 539 was found to be associated with larger adenomas and absence of hormonal secretion (97). In 540 hormonally active ACA, the prevalence of CTNNB1 mutations is estimated between 2 to 5% in APA (28,29,99) and  $\approx 4\%$  in CPA (see previous sections). The presence of CTNNB1 541 542 mutations in APA was not associated with a particular histological or morphological phenotype, but seems to be more prevalent in females (26,28,29,51). In addition, Teo and co-543 workers reported three women with PA, harboring CTNNB1 mutations in their adenoma that 544 were associated with aberrant expression of G-protein coupled receptors (GPCR), two during 545 546 pregnancy and one after menopause (51), suggesting a possible acceleration of the disease during pregnancy and menopause due to high gonadotropin levels acting on aberrantly 547 expressed gonadotropin receptors. Since CTNNB1 mutations were also reported in non-548 pregnant women (28,29) and men, the relation between *CTNNB1* mutations and pregnancy 549

and the possible interaction with aberrant gonadotropin receptors remain to be furtherclarified (100,101).

The role of the Wnt  $\beta$ -catenin pathway in the development of APA is further supported by the

553 case of a young patient who was diagnosed with severe arterial hypertension due to primary aldosteronism at age 26, followed by hemorrhagic stroke four years later, due to lateralized 554 aldosterone secretion from an APA in the context of bilateral multinodular adrenal glands 555 556 (69). The patient, in whom additional exploration identified asymptomatic Familial 557 Adenomatous Polyposis (FAP) associated with a heterozygous germline APC mutation, showed biallelic APC inactivation due to loss of heterozygosity in two adrenal nodules. The 558 559 aldosterone producing nodule carried an additional somatic KCNJ5 mutation supporting a two-hit model for APA development (69). This patient also had incomplete cortisol 560 suppression after an overnight dexamethasone suppression test, but normal 24-h urinary free 561 562 cortisol, maintained circadian rhythm of cortisol secretion and normal ACTH levels; CYP11B1 expression was detected in all adrenal nodules, including the one carrying the 563 564 KCNJ5 mutation (unpublished). 565 Remarkably, a link between the Wnt/ $\beta$ -catenin and the PKA pathway has recently been established, showing that activation of PKA prevents zona glomerulosa differentiation 566

through WNT4 repression and Wnt pathway inhibition, suggesting that PKA activation in the

zona fasciculata is a key driver of Wnt inhibition and lineage conversion. (102).

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# 570 **7.2. ARMC5 mutations in adrenocortical tumors**

571 Evidence that ARMC5 is involved in adrenal tumorigenesis came from recent studies

572 exploring PBMAH (also called ACTH-independent macronodular adrenal hyperplasia or

573 AIMAH). PBMAH is a form of macronodular adrenal hyperplasia characterized by multiple,

bilateral, non-pigmented, benign, adrenocortical nodules. It results in excessive production of
cortisol leading to ACTH-independent overt or subclinical Cushing syndrome (103).

576 Germline *ARMC5* mutations were identified in familial cases and are present in 50% of

577 apparently sporadic cases. Somatic events occur independently in adrenal nodules, suggesting

that *ARMC5* acts as a tumor suppressor gene (103).

A recent study has shown that PBMAH is not truly ACTH-independent since 579 proopiomelanocortin expression was described in clusters of steroidogenic cells in PBMAH 580 581 tissue, suggesting local ACTH production with paracrine regulation of cortisol biosynthesis (104). The existence of heterogeneous phenotypes (105), such as that observed in a Brazilian 582 583 family with adrenal macronodular disease showing bilateral and unilateral adrenal nodules, has highlighted the complexity of the condition, leading some authors to prefer the name primary 584 macronodular adrenal hyperplasia (PMAH) (106). PBMAH is usually diagnosed after the 585 586 investigation of an adrenal incidentaloma or by the presence of overt CS, although insidious or mild CS is more frequent (107,108). Aldosterone secretion was also described in patients with 587 588 PBMAH (69,109,110). Previous studies have demonstrated that cortisol secretion in PBMAH 589 may be regulated by various hormones via aberrant adrenal expression of several G-proteincoupled receptors (107,111,112). Illegitimate G-protein-coupled receptors expressed in 590 PBMAH include the gastric inhibitory polypeptide (GIP) receptor, vasopressin receptors,  $\beta$ -591 592 adrenergic receptors, LH/hCG receptors, and the serotonin 5-HT4 and 5-HT7 receptors ((107) and references therein). 593

Although sporadic forms appear to be more frequent, the description of familial forms and the occurrence of bilateral adrenal lesions support a genetic cause for PBMAH. Recently, combined SNP array and microsatellite marker analysis have shown a somatic 16p11 loss of heterozygosity (LOH) in 8 of 33 tumor samples from patients with PBMAH. In parallel, WES analysis followed by direct sequencing of blood and tumor samples has identified somatic mutations in *ARMC5*, the armadillo repeat containing 5 gene mapped to 16p11.2, in 18 of 33 PBMAH patients (113). All patients carried a germline *ARMC5* mutation and a second somatic *ARMC5* event in the tumor DNA, including LOH, a second mutation or microdeletion, suggesting a 2-hit model for the development of PBMAH (113). Interestingly, distinct somatic *ARMC5* alterations are observed in different nodules from the same adrenal (113), exemplified by an emblematic case of 16 different somatic *ARMC5* mutations in 20 adrenal nodules from the same patient (114).

606 The function of ARMC5 is unknown and no other diseases have been associated with mutations in this gene. ARMC5 encodes a 935 amino acid protein of the armadillo repeat family containing 607 an armadillo repeat domain in the N-terminal region and a BTB/POZ in the C-terminal region 608 (115). Analysis of the expression of four ARMC5 isoforms in 46 human tissues has shown that 609 at least one ARMC5 isoform is ubiquitously expressed throughout the body, but only seven 610 611 tissues expressed all four isoforms, including the adrenal gland (116). Armc5 knockout mice have defective development and immune responses, with adrenal hyperplasia in old animals 612 613 (117). Transfection of wild-type ARMC5 in H295R cells induces apoptosis, which is not 614 observed when mutant ARMC5 are transfected. Furthermore, silencing of ARMC5 in H295R cells reduced the expression of the steroid enzymes CYP17A1 and CYP21A2; NR5A1 and 615 MC2R, as well as cortisol production (113). PBMAH patients with ARMC5 mutations show 616 more overt CS than patients without ARMC5 mutations (113,118). In a cohort of 98 unrelated 617 PBMAH subjects, ARMC5 mutations were found in 24 patients (26%). Patients with ARMC5 618 mutations again showed more frequently overt CS and had larger adrenals with a higher number 619 620 of nodules than non-mutated patients (119). These results were confirmed in an Italian cohort of PBMAH patients, in which ARMC5 mutated patients exhibited higher cortisol levels, more 621 severe hypertension and diabetes, and larger adrenal lesions (120). ARMC5 mutations have also 622 been identified in familial forms of PBMAH (105,121-123). In one Brazilian family with 623

PBMAH, a germline ARMC5 mutation was found which segregated with the disease, although 624 625 the clinical phenotype and adrenal involvement (unilateral or bilateral) were variable among carriers (105). In five families with PBMAH, Gagliardi et al have found germline ARMC5 626 627 mutations in four families with CS and no ARMC5 mutation in one family with PA and sCS (121). Interestingly, in a large French-Canadian PBMAH family, ARMC5 mutations were found 628 in affected subjects together with aberrant expression of  $\beta$ -adrenergic and V1-vasopressin 629 630 receptors in adrenal nodules (123). In this case, the aberrant expression of GPCR may represent a secondary event in the development of PBMAH, but the link between ARMC5 mutations and 631 this pattern of receptor expression is not yet clarified. 632

Zilbermint et al have sequenced ARMC5 in germline DNA from 56 subjects with PA and have 633 identified 12 heterozygous variants, 9 missense variants and three variants leading to alternative 634 splicing (124). In silico analysis of the ARMC5 variants has shown that six variants were 635 636 predicted to be damaging, including the ARMC5 variant p.R898W, which was previously described in one patient with PBMAH (118), and a novel missense mutation ARMC5 p.P826H 637 (124). All carriers of these variants were African-American, two had bilateral macronodular 638 639 hyperplasia, and tumor DNA sequencing showed neither additional somatic ARMC5 mutations nor ARMC5 loss of heterozygosity. APA with predicted damaging ARMC5 variants exhibit 640 641 lower ARMC5 expression than the adjacent adrenal cortex. The functional consequences of these ARMC5 variants are not completely understood. Silencing of ARMC5 in H295R cells 642 leads to decreased CYP11B2 expression, and therefore a reduction of the aldosterone-secretory 643 capacity. The authors suggest that the higher aldosterone secretion in PA patients carrying 644 645 ARMC5 variants may be explained by the increased adrenocortical mass, similar to what was observed in patients with PBMAH and hypercortisolism (124). An association of ARMC5 646 647 mutations with PA was not replicated in an Italian study assessing the presence of ARMC5 mutations in 39 PA patients with bilateral computed tomography-detectable adrenal alterations, 648

from which 8 had unilateral and 27 bilateral aldosterone secretion (125). Different *ARMC5* variants were identified in 18 subjects, among them 11 common variants, two rare ones and two previously unreported variants were found, but *in silico* analysis did not identify possibly damaging *ARMC5* variants (125). These contradictory data do not allow confirmation of the role of *ARMC5* mutations in PA and further studies are required to better understand the role of *ARMC5* in the development of adrenal nodules and aldosterone excess.

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#### 7.3. Germline mutations associated with ACA and macronodular hyperplasia

Genetic alterations leading to multiple tumor syndromes and mutations affecting genes of the 657 cAMP/PKA signaling pathway have been associated with PBMAH (review in (106,108)). 658 Familial adenomatous polyposis (FAP) is a disease characterized by multiple colonic polyps 659 with an increased risk of colon carcinomas, due to an inactivating germline mutation of APC 660 661 and a somatic second hit (mutation or LOH), resulting in constitutive activation of the Wnt/βcatenin pathway (126). Adrenal masses are more frequently found in patients with FAP than in 662 the general population (127), including non-functioning adrenal adenomas, adrenocortical 663 cancers, and PBMAH (128,129). Multiple endocrine neoplasia type 1 (MEN1) is an autosomal 664 dominant syndrome resulting from inactivating mutations in the MEN1 gene, which is 665 666 characterized by the development of multiple endocrine and non-endocrine tumors, most frequently hyperparathyroidism, pituitary adenomas, and pancreatic neuroendocrine tumors 667 (130). MEN1 encodes the protein menin which has a role in the regulation of cell cycle, cell 668 growth and proliferation (131). The analysis of adrenal involvement in 715 patients with MEN1 669 670 has shown adrenal tumors (>10mm in size) in 10% of the cohort; in 19.4% of these cases the tumors were bilateral, and in 12.5% of the cases the tumors were >40 mm in size (132). A MEN1 671 mutation was also identified in one patient with PBMAH and hyperparathyroidism, without a 672 family history or other endocrine abnormalities (129). In addition, LOH at the MEN1 locus was 673

also reported in APA (133). Another hereditary disease associated with PBMAH is the 674 675 hereditary leiomyomatosis and renal cell carcinoma syndrome (HLRCC), a disease caused by germline mutations in the fumarate hydratase (FH) gene (134). Adrenal lesions are found in 676 677 7.8% of patients with HLRCC, including bilateral lesions associated with ACTH-independent hypercortisolism (135), and germline FH mutations were identified in patients with PBMAH 678 associated with HLRCC (129,134). Genetic alterations leading to activation of cAMP/PKA 679 signaling were also described in patients with PBMAH. Post-zygotic somatic mutations in the 680 gene encoding the alpha subunit of stimulatory G protein (GNAS), leading to constitutive 681 activation of adenylate cyclase, result in McCune Albright syndrome, which is characterized 682 683 by a triad constituted by polyostotic fibrous dysplasia, cafe' au lait skin spots, and gonadotropinindependent precocious puberty, associated with other endocrine and non-endocrine diseases, 684 including CS (136). Somatic GNAS mutations were described in a few cases of PBMAH 685 686 without other typical signs of McCune Albright syndrome and might represent variants of this syndrome (129,137). In isolated cases, variants in PDE11A, encoding the isoform 11A of the 687 phosphodiesterase enzyme, and in MC2R were associated with PBMAH (138-140). Finally, 688 689 duplication of a genomic region of chromosome 19 including PRKACA was also identified in patients with PBMAH (75,141). 690

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#### 692 8. Concluding remarks

Over the past few years, extraordinary progress has been made in our understanding of the genetic abnormalities leading to APA and CPA. While genetic testing in inherited forms of the diseases has been translated into clinical practice, and family screening is useful for management of affected family members, the clinical utility of diagnosing somatic mutations is difficult to anticipate from available studies on genotype-phenotype correlations and outcome. Ideally, identification of mutations prior to surgery, through the application of

surrogate biomarkers or genetic testing on circulating cell-free DNA, could guide treatments 699 to specifically target mutated proteins, as shown for APA (142,143). There seems to be an 700 overlap of genetic abnormalities leading to different endocrine abnormalities in ACA. In 701 702 particular, CTNNB1 mutations are found at relatively high frequency (at least comparable to 703 what observed for ATP1A1 or ATP2B3 mutations in APA) in CPA but also usually in APA. Furthermore, genetic variants affecting genes of the calcium signaling pathway are enriched 704 in WES data from CPA, suggesting that calcium, in addition to PKA signaling, may lead to 705 706 autonomous cortisol overproduction. This is not surprising, as a subset of APA show some cortisol co-secretion, or biochemical glucocorticoid abnormalities (144), and in some cases 707 708 those tumors harbor somatic KCNJ5 mutations. In addition, overexpression of mutated 709 GIRK4 channels in HAC15 cells not only increases CYP11B2 expression but also CYP11B1 (33). Future studies need to address the sequence of events leading to adrenal cell 710 711 proliferation, nodule formation and specification of the hormonal secretory pattern. It will be 712 of particular interest to explore whether common susceptibility alleles or rare variant may 713 predispose to the development of APA and CPA. 714 715

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1326 Figure legends

1327

# Figure 1. Specific and common mechanism of aldosterone and cortisol producing 1328 adenoma development. A. Specific mechanisms of APA formation. In absence of 1329 stimulation (AngII or K<sup>+</sup>), ZG cells are hyperpolarized due to the presence of a large number 1330 of K<sup>+</sup> channels. In response to an increase of extracellular K<sup>+</sup> concentration or to the binding 1331 of AngII to its receptor (AT1R), inhibition of K<sup>+</sup> channels leads to cell membrane 1332 depolarization, opening of voltage gated Ca<sup>2+</sup> channels, increased intracellular calcium 1333 concentrations and activation of $Ca^{2+}$ signaling, the major trigger for aldosterone biosynthesis. 1334 Moreover, binding of AngII to AT1R leads to the inhibition of the Na<sup>+</sup>/K<sup>+</sup>-ATPase, increasing 1335 cell depolarization, whereas signaling through $G\alpha q$ promotes the release of $Ca^{2+}$ from the 1336 endoplasmatic reticulum through binding of IP3 to its receptor, again stimulating aldosterone 1337 biosynthesis. Mutations in KCNJ5, encoding the GIRK4 potassium channel, induce changes 1338 in channel selectivity, allowing Na<sup>+</sup> entry into the cell instead of K<sup>+</sup> efflux, resulting in cell 1339 depolarization, opening of voltage gated $Ca^{2+}$ channel, stimulation of $Ca^{2+}$ signaling and 1340 increase in aldosterone production. Mutations in ATP1A1, coding for the $\alpha$ 1 subunit of the 1341 Na<sup>+</sup>/K<sup>+</sup>-ATPase, lead to increased aldosterone production by increasing intracellular Na<sup>+</sup> and 1342 H<sup>+</sup> concentration, resulting in cell membrane depolarization and decrease of intracellular pH. 1343 Mutations in CACNA1D, encoding the Cav1.3 calcium channel, and ATP2B3, coding for the 1344 $Ca^{2+}$ pump PMCA3 directly affect intracellular $Ca^{2+}$ concentrations and thus aldosterone 1345 biosynthesis. B. Specific mechanisms of CPA formation. In absence of stimulation, the 1346 catalytic and regulatory subunits of the PKA form an inactive complex. Binding of ACTH to 1347 its receptor, the melanocortin 2 receptor (MC2R), leads to activation of adenylate cyclase 1348 resulting in an increase of cAMP. cAMP binds to the regulatory subunits of the PKA 1349 complex, inducing conformational changes and release of the catalytic subunits from the 1350 1351 complex. Active PKA phosphorylates specific target proteins such as the transcription factor

CREB that binds to the promoter regions of specific target genes, ie CYP11B1 leading to 1352 1353 increased cortisol biosynthesis. In cells, the cAMP is converted into AMP through the action of specific PDE. Mutations in PRKACA or in GNAS lead to the constitutive activation of 1354 1355 PKA, phosphorylation or CREB and increased expression of CYP11B1 and autonomous cortisol biosynthesis. C. Common mechanisms of APA and CPA formation. In 1356 1357 unstimulated cells,  $\beta$ -catenin is phosphorylated, ubiquitinated by a protein complex formed by 1358 APC, GSK3, Axin and CK1 $\alpha$  and then degraded by the proteasome. Binding of Wnt ligand to Frizzled receptor and LRP5/6 co-receptor leads to the dissociation of the β-catenin 1359 degradation complex. β-catenin can translocate into the nucleus where it acts as a 1360 1361 transcriptional coactivator for transcription factors of the TCF/LEF family inducing the expression of specific target genes. In APA as well as in CPA, CTNNB1 mutations affecting 1362 specific residues of  $\beta$ -catenin lead to constitutive activation of the protein. In APA, mutations 1363 1364 in APC also lead to constitutive  $\beta$ -catenin activation.

1365

### 1366 Figure 2. The two current models for APA development. Left panel: Two hit model of **APA development.** In ZG cells, activation of specific signaling pathways, such as $\beta$ -catenin, 1367 lead to abnormal cell proliferation. Abnormal cell proliferation creates a propitious 1368 1369 environment for occurrence of recurrent somatic mutations specifying the pattern of hormonal secretion. Right panel: APCC hypothesis. In ZG cells, occurrence of somatic mutations in a 1370 certain number of APA driver genes, lead to the formation of aldosterone producing cell 1371 clusters (APCC) with autonomous aldosterone production. In a certain number of cases these 1372 structures would evolve towards an aldosterone producing adenoma a possible APCC-to-APA 1373 1374 transitional lesion (pAATL).

1375

Total n of	KCNJ5	CACNA1D	ATP1A1	ATP2B3	CTNNB1	Other	Correlations of mutation status	Reference
patients screened*	N (%)	N (%)	N (%)	N (%)	N (%)			
22	8 (36%)	NA	NA	NA	NA	NA	7/8 patients with KCNJ5 mutations were female	(24)
380	129 (34%)	NA	NA	NA	NA	NA	<i>KCNJ5</i> : Higher prevalence in females and younger patients; higher plasma aldosterone at diagnosis; no specific pangenomic trascriptomic profile; no association with pre-operative blood pressure, serum potassium and adenoma size or cure after adrenalectomy	(145,146)
							Lower relative GIRK4 protein expression in <i>KCNJ5</i> mutated tumors	
348	180 (45%)	NA	NA	NA	NA	NA	<i>KCNJ5</i> : Higher prevalence in females, males younger at diagnosis and with larger tumors; no correlation with <i>KCNJ5</i> protein expression	(147)
23	15 (62.5%)	NA	NA	NA	NA	NA	<i>KCNJ5:</i> higher <i>KCNJ5</i> mRNA expression compared to non-mutated tumors	(40)
73	30 (41%)	NA	NA	NA	NA	NA	<i>KCNJ5</i> : larger APA, predominant ZF-like cells, younger patients, higher <i>CYP17A1</i> expression;	(148,149)
91	29 (32%)	NA	NA	NA	NA	NA	<i>KCNJ5</i> : Higher aldosterone levels, lower plasma renin activity, higher ARR higher lateralization index; more frequent contralateral suppression Higher <i>CYP11B2</i> mRNA expression	(48)

 Table 1. Clinical and molecular characteristics of patients with APA.

308	118 (38.3)	NA	16 (5.2%)	5 (1.6%)	NA	NA	<i>ATP1A1</i> and <i>ATP2B3</i> : more frequent in males, higher preoperative aldosterone concentrations, lower serum potassium concentrations	(27)
152	20 (13.2%)	12 (7.9%)	12 (7.9%)	0/10 exomes	1/10 exomes	NA	<i>ATP1A1</i> and <i>CACNA1D</i> : more frequent in older males; smaller APAs with more compact, zona glomerulosa–like cells;	(25)
							<i>KCNJ5</i> : more frequent ZF-like cells, higher <i>CYP11B1</i> mRNA expression:	
							No difference in <i>ATP1A1</i> and <i>CACNA1D</i> expression among genotypes	
64	21 (32.8%)	5 (7.8%)	1 (1.6%)	2 (3.1%)	2 (3.1%)	NA	CACNA1D: smaller APA, trend towards older age	(26)
59	19 (32%)	NA	5 (8.5%)	3 (5%)	NA	NA	No impact of mutation status on lateralization index and contralateral suppression index	(49)
474	180 (38%)	44 (9.3%)	25 (5.3%)	8 (1.7%)	NA	NA	<i>KCNJ5</i> : Higher prevalence in females and younger patients; higher minimal plasma potassium at diagnosis;	(30)
							CACNA1D: smaller adenomas**	
							All mutations: no association with pre-operative blood pressure, serum potassium and adenoma size or cure after adrenalectomy; no correlation with cellular composition nor <i>CYP11B2</i> or <i>CYP11B2</i> expression; no correlation with <i>KCNJ5</i> , <i>CACNA1D</i> , and <i>ATP1A1</i> mRNA expression	
112****	44 (39.3%)	NA	7 (6.3%)	1 (0.9%)	NA	NA	<i>ATP1A1</i> , <i>ATP2B3</i> : higher <i>CYP11B2</i> mRNA expression compared to <i>KCNJ5</i> mutated APA	(150)

108	75 (69.4%)	2 (1.9)	3 (2	2.8)	NA	NA	<i>KCNJ5</i> : younger patients, higher plasma and urine aldosterone levels; significant improvement of left ventricular mass index after surgery; lower post- op SBP and aldosterone levels	
195	48 (24.6%)	NA	NA	NA	NA	NA	Not investigated	(151)
165	90 (54.5%)	5 (3.0%)	10 (6.1%)	5 (3.0%)	NA	NA	<i>KCNJ5</i> : Higher prevalence in females and younger patients;	(50)
							<i>CACNA1D</i> , <i>ATP1A1</i> and <i>ATP2B3</i> : older at diagnosis, more often males with smaller tumors; higher <i>CYP11B2</i> and <i>NPNT</i> mRNA expression compared to APA with <i>KCNJ5</i> mutation;	
							All mutations: No correlation with <i>ATP1A1</i> , <i>ATP2B3</i> or <i>KCNJ5</i> mRNA expression	
69	26 (37.7%)	NA	NA	NA	NA	NA	<i>KCNJ5</i> : Lower relative GIRK4 protein expression in <i>KCNJ5</i> mutated APA; Higher prevalence in females and younger patients; larger tumors and lower plasma potassium	(34)
168	129 (76.8%)	1 (0.6%)	4 (2.4%)	1 (0.6%)	NA	NA	<i>KCNJ5</i> : Higher prevalence in females, larger tumors, higher aldosterone levels and lower minimal serum potassium, higher lateralization index, lower plasma renin activity, higher ARR	(42)
71***	27 (38%)	3 (4.2%)	5 (7%)	1 (1.4%)	NA	NA	<i>KCNJ5</i> : more frequently zona fasciculata-like cells, high <i>CYP11B1</i> protein expression;	(152)
							<i>ATP1A1</i> , <i>ATP2B3</i> and <i>CACNA1D</i> : more frequently ZG-like cells with high <i>CYP11B2</i> protein expression	

90	36 (37.1%)	10 (10.3%)	8 (8.2%)	3 (3.1%)	2 (2.1%)	NA	<i>KCNJ5</i> : Higher prevalence in females, larger tumors, lower pre-contrast Hounsfield units on CT; more frequently ZF-like cells	(28)
							ATP1A1, ATP2B3 and CACNA1D: more frequent in males, heterogenous cell composition	
148	88 (59.5%)	0	2 (1.4%)	1 (0.7%)	NA	NA	Mutation carriers: younger, higher preoperative aldosterone and ARR, lower serum potassium, higher glomerular filtration rate, lower serum CRP levels, greater chance of recovery from hypertension post-adrenalectomy	(44)
114	86 (75.4%)	1 (0.09%)	0	0	NA	NA	Mutation carriers: younger, lower preoperative serum potassium level, higher LV mass index and significant improvement after surgery; higher mRNA expression of <i>KCNJ5</i> , <i>CYP11B2</i> , and <i>ATP2B3</i>	(43)
							male (but not female) mutation carriers: higher preoperative plasma aldosterone and blood pressure;	
198	92 (46.5%)	3 (1.5%)	6 (3.0%)	3 (1.5%)	10 (5.1%)	NA	<i>CTNNB1</i> : more frequent in females; larger tumors compared to APA without mutation; Higher <i>CYP11B2</i> mRNA expression compared to APA with <i>KCNJ5</i> mutation;	(29)
159	117 (73.6%)	4 (2.5%)	1 (0.6%)	4 (2.5%)	NA	NA	<i>ATPase</i> : tendency to smaller tumors, mainly ZG-like cells	(45)
							<i>CACNA1D</i> : tendency to smaller tumors, predominant ZF-like cells	
							KCNJ5: not analyzed in this report	

66	47 (71.2%)	0	0	0	NA	NA	<i>KCNJ5</i> : Higher prevalence in females and younger patients, no association with pre- operative blood pressure, plasma aldosterone, serum potassium, lateralization index, and adenoma size; better cure following adrenalectomy	(46)
219	116 (52.9%)	0	3 (1.4%)	1 (0.5%)	8 (3.7%)		CTNNB1: more frequent in females, older patients, heterogeneous staining of $\beta$ -catenin and variable expression of gonadal receptors and both CYP11B1 and CYP11B2	(99)

\*by WES and/or targeted sequencing; \*\*frequency of mutations and associations dependent on center; \*\*\*patients included in (30) ;\*\*\*\* included 32 samples from (27). ZF, zona fasciculate; ZG, zona glomerulosa.

Gene	Mutations	<b>References first description</b>
KCNJ5	p.Gly151Arg, pLeu168Arg, pThr158Ala	(24)
	p.Gly151Glu	(62)
	p.Ile157del	(153)
	p.Tyr152Cys	(154)
	p.Trp126Arg	(150)
	p.Glu145Gln, p.Arg115Trp, p.Glu246Gly	(34)
	p.Phe154Cys, p.Ile150_Gly151del_InsMet,	(28)
	p.Ser143_ile144del_InsAla	
	p.Ala139_Phe142dup	(155)
ATP1A1	p.Leu104Arg, p.Val332Gly, pPhe100_Leu104del	(27)
	p.Glu960_Ala963del	(25)
	p.Gly99Arg	(150)
	p.Met102_Leu103del	(42)

# Table 2. List of the different mutations identified in APA driver genes.

	p.Met102_Ile106del, p.leu103_Leu104del, p.Phe959_Glu961del,	(50)
	p.Glu960_Leu694del, p.Phe956_Glu961del	
ATP2B3	p.Leu425_426del, p.Val426_427del	(27)
	p.Val424_Leu425del	(30)
	p.Ala428_Val429del	(156)
	p.Val426_Val429del	(50)
	p.Val422_Val426del_InsSerThrLeu	(42)
	p.Ala428_Leu433del_InsGlyGln	(28)
CACNA1D	p.Val259Asp, p.Gly403Argp.Phe747Leu, p.Ile750Met,	(25)
	p.Arg990His	
	p.Gly403Arg, p.Ile750Met, p.Phe767Val, p.Val1373Met	(26)
	p.Ser652Leu, p.Leu655Pro, p.Tyr741Cys, p.Val949Asp,	(30)
	p.Lys981Asnp.Ala998Ile, p.Ala998Valp.Val1151Phe,	
	p.Ile1152Asnp.Val1338Met	
	p.Val401Leu	(50)
	p.Phe747Val	(68)

Total n of patients screened*	Clinical phenotype	<i>PRKACA</i> N (phenotype)	<i>GNAS</i> N (phenotype)	<i>CTNNB1</i> N (phenotype)	<i>APC</i> N (phenotype)	Reference
139	59 CS, 40 sCS,	22 (CS)	0 in WES	0 in WES	0 in WES	(75)
	20 APA, 20 NSA					
63 (55 CPA + 8 ACC)	36 CS, 27 sCS	13 (10 CS, 3 sCS)	3 CS	9 (3 CS, 6 sCS)	0 in WES	(76)
87	87 (CS)	57 (CS)	2 (CS)	1 (CS)	1 (CS)	(77)
64	55 CS, 9 sCS	34 (33 CS, 1 sCS)	11 (10 CS, 1 sCS)	0 in WES	0 in WES	(78)
144	64 CS, 36 sCS, 32 NSA, 4 androgen producing tumors, 8 PBMAH	22 (CS)	ND	ND	ND	(157)
60	36 CS, 22 sCS	12 (11 CS, 1 sCS)	4 (3 CS, 1 sCS <sup>\$</sup> )	13 (5 CS, 8 sCS <sup>\$</sup> )	ND	(79)
57	15 CS, 9 sCS, 33 APA	4 CS	5 (2 CS, 1 sCS, 2 APA+sCS	ND	ND	(81,158)

Table 3. Clinical and genetic characteristics of patients with CPA.

\*by whole exome sequencing and/or targeted sequencing, excluding ACC; CS, overt Cushing syndrome; sCS, subclinical Cushing syndrome; APA, aldosterone producing adenoma; NSA, non-secreting adenoma; ACA, adrenocortical adenoma; ACC, adrenocortical carcinoma; <sup>\$</sup>including one patient with a double *CTNNB1* + *GNAS* mutation; ND, not determined.

Characteristics	(81)	(85)	(79)	(32)	(69)
Total n of APA patients screened	33	unknown	4	122	1
N of patients with cortisol co-secretion (F:M)	10 (x:x)	3 (3:0)	4	?	1 (0:1)
Clinical phenotype	PA/sCS	PA/sCS	PA/2CS-2sCS	1 case with PA/sCS, 1 with PA only	PA/sCS
KCNJ5 mutations	6	2	2	0	1 (somatic)
GNAS mutations	2	Not described	0	Not described	Not described
<b>PRKACA</b> mutations	0*	Not described	0	2	Not described
CTNNB1 mutations	Not described	Not described	0	Not described	0
APC mutations	Not described	Not described	Not described	Not described	1 (germline)

Table 4. Clinical and genetic characteristics of patients with A/CPA

\*in another paper, the same authors report about the analysis of 33 APA, 11 of them with sCS, with none of them carrying a PRKACA mutation (80). It is unclear, what the overlap is between these patients and those of this reference.





Figure 2