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LETTER TO THE EDITOR

Dear medical mycologists, it is time to look outside the box

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One sentence summary: Recent studies demonstrated that our environment can represent an alternative niche for several opportunistic fungal pathogens that can be a concern for human health.

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ABSTRACT

Opulente et al. (Opulente DA, Langdon QK, Buh KV et al. Pathogenic budding yeasts isolated outside of clinical settings. FEMS Yeast Res 2019;19:foz032) published early this year a study aiming to investigate the diversity of wild yeast species, by collecting 1000 environmental samples coming from different substrates across the United States of America. The main finding of this work is the recovery of 54 strains of budding yeasts of which several are having a pathogenic potential in the clinical setting, such as Candida albicans, C. parapsilosis, C. tropicalis, Nakaseomyces glabrata and Pichia kudriavzevii. These findings, discussed here in light of other recent studies highlighting the role of fungicides in the rise of antifungal resistance in the clinical setting or the emergence of Candida auris, demonstrate that our environment can represent an alternative niche for several opportunistic fungal pathogens that can be a concern for human health.

Keywords: yeast; antifungal resistance; environment; pesticides; opportunistic fungi; one health

I read with interest the manuscript of Opulente et al. (2019) published early this year. To gain further insights into the diversity of wild yeast species, the authors collected 1000 environmental samples (different substrates) across the United States of America. The main finding of this work is the recovery of 54 strains of budding yeasts of which several are having a pathogenic potential in the clinical setting, such as Candida albicans, C. parapsilosis, C. tropicalis, Nakaseomyces glabrata and Pichia kudriavzevii (the two latter known as Candida glabrata and Candida krusei, respectively in the clinical setting). The authors also highlighted that P. kudriavzevii is significantly associated with plants and C. tropicalis with soil.

Despite a relatively modest number of isolates of opportunistic yeasts (54 out of the 5000 isolates), these observations confirm earlier studies reporting human pathogenic yeasts

outside the clinical setting (Van Uden, De Faia and Assis-Lopes 1956; Yang et al. 2012; Wójcik, Kurnatowski and Błaszkowska 2013; Wójcik et al. 2016; Bensasson et al. 2019). For example, C. albicans, a major human pathogen and also a commensal of the human gut, has been recently reported in oaks in the United Kingdom (Bensasson et al. 2019). We also observed similar findings during a pilot study in western France, including 40 soil samples. Despite a different protocol and incubation at 30° C to expect a possible enrichment in medically relevant species, we identified, along with various basidio- and ascomycete yeasts, several opportunistic pathogens including C. tropicalis (n=2), Pichia guilliermondii (n=2), Pichia norvegensis (n=1), Clavispora lusitaniae (n=1), Candida pelliculosa (n=1), Cyberlindnera fabianii (n=1) and Candida sake (n=4) (unpublished data). Altogether this provides further evidence on the role of the environment as

a source of clinically relevant fungal pathogens. This is of major interest as, despite small variation between countries, *Candida* species rank in the top three or four pathogens responsible for health-care-associated bloodstream infections and are associated with a 10 to 20% attributable mortality (Pappas *et al.* 2018).

Despite the huge diversity of fungi evolving in our environment, their impact on human fungal diseases has been mainly highlighted for molds. One example is Aspergillus fumigatus, occurring in the environment as a saprophyte but which can cause a wide range of clinical manifestations, ranging from asthma to life-threatening infections, in susceptible hosts (van de Veerdonk et al. 2017). Since 10 years, physicians and researchers dealing with aspergillosis are facing antifungal resistance (Denning and Perlin 2011). Actually, environmental sampling has unveiled an unexpected environmental route of azole resistance that is turning into a public health issue (Verweij et al. 2009; Le Pape et al. 2016; Lavergne et al. 2017). Basically, when exposed to azole fungicides in the environment (fungicides are widely used for cereal or flower crop protection), A. fumigatus can develop acquired resistance mechanisms that also confer cross resistance to medical azoles. This is the scenario explaining the worrisome increase of azole resistance in A. fumigatus in the clinical setting. Actually, there are several other examples of environmental filamentous fungi impacting human health, such as dermatophytes, Alternaria spp., Fusarium spp. and Mucorales (Bongomin et al. 2017).

By comparison, this field of research has been less investigated and is almost neglected when dealing with yeast infections. In the clinical setting, as some Candida species belong to the natural flora (such as the gut for C. albicans and C. glabrata, and the skin for C. parapsilosis), the infective strain is usually considered to originate from the patient's endogenous flora (Neville et al. 2015). Hence, the occurrence of alternate environmental niches for these yeasts challenges this dogma. Be that as it may, a literature review of the last years provides additional arguments. One example is C. tropicalis, ranking first in patients with haematological malignancies with invasive candidiasis and being the second or third etiological agent of candidemia in Asia and Latin America (Horn et al. 2009; Nucci et al. 2013; Tan et al. 2016; Pappas et al. 2018). Since 2009, China is witnessing an increased prevalence of fluconazole-resistant C. tropicalis isolates in hospital wards (Fan et al. 2017). In a recent study, fluconazole non-susceptibility in clinical isolates of C. tropicalis reached 33% in Taiwan (Hii et al. 2019). Notably, molecular typing revealed the circulation of genetically related *C. tropicalis*-resistant clones across different hospitals and in soil samples (Yang et al. 2012; Chen et al. 2019). This explains why antifungal-naïve patients can develop invasive infections caused by de novo resistant isolates (Chen et al. 2019). This situation strongly reminds the Aspergillus fumigatus scenario described above. Therefore, in addition to act as niche for pathogenic yeasts, our environment is also a possible reservoir of de novo antifungal-resistant isolates resulting from cross-resistance with pesticides (Brilhante et al. 2019). An additional example is the elegant demonstration using population genetic analysis that P. kudriavzevii clinical isolates are not genetically distinct from environmental isolates suggesting that susceptible hosts can acquire P. kudriavzevii infections from the environment (Douglass et al. 2018). Last but not the least, Candida haemulonii, a multidrug-resistant species formerly described from fish guts and later isolated in cassava roots, has been responsible for invasive infections in the clinical setting (Van Uden and Kolipinsky 1962; Khan et al. 2007; Ferreira et al. 2010). Yet there are countless other examples, like

Diutina catenulata (known in the clinical setting as Candida catenulata), another pathogenic yeast whose draft genome has been recently published from an environmental isolate from Ireland (O'Brien et al. 2018). Susceptible hosts can also be occasionally infected by species associated with specific reservoirs or environments, such as the black yeast Exophiala xenobiotica, inhabiting environments rich in monoaromatic hydrocarbons and alkanes (Morio et al. 2012) or the neurotropic basidiomycetes Cryptococcus gattii, which causes ~225 000 new cases globally every year, and is associated with soil, pigeon droppings and tree hollows (Herkert et al. 2017). Finally, Candida auris, an emerging species in the clinical setting, has also been proposed to come from our environment, subjected to climate change and to spread worldwide with birds, as illustrated with C. glabrata (Al-Yasiri et al. 2016; Casadevall, Kontoyiannis and Robert 2019).

Besides the clinical interest, environmental sampling also has the potential to improve our knowledge about yeast biodiversity. For example, by bringing together public citizens and mycologists, the Citizen Science initiative conducted by the Westerdijk Fungal Biodiversity Institute and the Utrecht University Museum led to the discovery of at least six new ascomycetes yeast species in Dutch garden soil (Groenewald et al. 2018).

These findings also raise some questions: Do these observations only demonstrate that some spots are highly trafficked by humans or that the environment can serve as an alternate natural habitat for these yeasts? How environmental pressures exerted by climate change and fungicides could fade the boundaries between our environment and human health? Applying the 'One Health' concept, time has come to encourage any initiative aiming to extend our view about the role of the environment and its impact on human health. Such projects would require collaborative efforts at a global scale and the involvement of various specialists, including clinicians, medical mycologists, environmental specialists and veterinarians, along with support from governmental institutions. It is time to go ahead and look outside the box!

Conflicts of interest. None declared.

REFERENCES

- Al-Yasiri MH, Normand AC, L'Ollivier C et al. Opportunistic fungal pathogen Candida glabrata circulates between humans and yellow-legged gulls. Sci Rep 2016;6:36157.
- Bensasson D, Dicks J, Ludwig JM et al. Diverse lineages of Candida albicans live on Old Oaks. Genetics 2019;211:277–88.
- Bongomin F, Gago S, Oladele RO et al. Global and multi-national prevalence of fungal diseases estimate precision. *J Fungi* 2017;3:57.
- Brilhante RSN, Alencar LP, Bandeira SP et al. Exposure of Candida parapsilosis complex to agricultural azoles: an overview of the role of environmental determinants for the development of resistance. Sci Total Environ 2019;650:1231–8.
- Casadevall A, Kontoyiannis DP, Robert V. On the emergence of *Candida auris*: climate change, azoles, swamps, and birds. MBio 2019;**10**:e01397–19.
- Chen PY, Chuang YC, Wu UI et al. Clonality of fluconazolenonsusceptible Candida tropicalis in bloodstream infections, Taiwan, 2011–2017. Emerg Infect Dis 2019;25:1660–7.
- Denning DW, Perlin DS. Azole resistance in Aspergillus: a growing public health menace. Future Microbiol 2011;6:1229–32.

- Douglass AP, Offei B, Braun-Galleani S et al. Population genomics shows no distinction between pathogenic Candida krusei and environmental Pichia kudriavzevii: one species, four names. PLoS Pathog 2018;14:e1007138.
- Fan X, Xiao M, Liao K et al. Notable increasing trend in azole nonsusceptible Candida tropicalis causing invasive candidiasis in China (August 2009 to July 2014): molecular epidemiology and clinical azole consumption. Front Microbiol 2017;8:464.
- Ferreira N, Belloch C, Querol A *et al.* Yeast microflora isolated from Brazilian cassava roots: taxonomical classification based on molecular identification. *Curr Microbiol* 2010;**60**:287–93
- Groenewald M, Lombard L, de Vries M et al. Diversity of yeast species from Dutch garden soil and the description of six novel Ascomycetes. FEMS Yeast Res 2018;18:foy076.
- Herkert PF, Hagen F, Pinheiro RL et al. Ecoepidemiology of Cryptococcus gattii in developing countries. J Fungi 2017;3:62.
- Hii IM, Liu CE, Lee YL et al. Resistance rates of non-albicans Candida infections in Taiwan after the revision of 2012 Clinical and Laboratory Standards Institute breakpoints. Infect Drug Resist 2019;12:235–40.
- Horn DL, Neofytos D, Anaissie EJ et al. Epidemiology and outcomes of candidemia in 2019 patients: data from the prospective antifungal therapy alliance registry. Clin Infect Dis 2009;48:1695–703.
- Khan ZU, Al-Sweih NA, Ahmad S et al. Outbreak of fungemia among neonates caused by *Candida haemulonii* resistant to amphotericin B, itraconazole, and fluconazole. *J Clin Microbiol* 2007;45:2025–7.
- Lavergne RA, Chouaki T, Hagen F et al. Home environment as a source of life-threatening azole-resistant Aspergillus fumigatus in immunocompromised patients. Clin Infect Dis 2017;64:76–8.
- Le Pape P, Lavergne RA, Morio F et al. Multiple fungicide-driven alterations in azole-resistant Aspergillus fumigatus, Colombia, 2015. Emerg Infect Dis 2016;22:156–7.
- Morio F, Berre JY, Garcia-Hermoso D et al. Phaeohyphomycosis due to Exophiala xenobiotica as a cause of fungal arthritis in an HIV-infected patient. Med Mycol 2012;50:513–7.

- Neville BA, d'Enfert C, Bougnoux ME. Candida albicans commensalism in the gastrointestinal tract. FEMS Yeast Res 2015;15:fov081.
- Nucci M, Queiroz-Telles F, Alvarado-Matute T et al. Latin American Invasive Mycosis Network. Epidemiology of candidemia in Latin America: a laboratory-based survey. PLoS One 2013;8:e59373.
- O'Brien CE, McCarthy CGP, Walshe AE et al. Genome analysis of the yeast Diutina catenulata, a member of the Debaryomycetaceae/Metschnikowiaceae (CTG-Ser) clade. PLoS One 2018:13:e0198957.
- Opulente DA, Langdon QK, Buh KV et al. Pathogenic budding yeasts isolated outside of clinical settings. FEMS Yeast Res 2019:19:foz032.
- Pappas PG, Lionakis MS, Arendrup MC et al. Invasive candidiasis. Nat Rev Dis Primers 2018;4:18026.
- Tan TY, Hsu LY, Alejandria MM et al. Antifungal susceptibility of invasive Candida bloodstream isolates from the Asia-Pacific region. Med Mycol 2016;54:471–7.
- van Uden N, Kolipinsky MC. Torulopsis haemulonii nov. spec., a yeast from the Atlantic Ocean. Antonie Van Leeuwenhoek 1962;28:78–80.
- van de Veerdonk FL, Gresnigt MS, Romani L et al. Aspergillus fumigatus morphology and dynamic host interactions. Nat Rev Microbiol 2017;15:661–74.
- Van Uden N, De Faia MM, Assis-Lopes L. Isolation of Candida albicans from vegetable sources. J Gen Microbiol 1956;15:151–3.
- Verweij PE, Snelders E, Kema GH et al. Azole resistance in Aspergillus fumigatus: a side-effect of environmental fungicide use? Lancet Infect Dis 2009;9:789–95.
- Wójcik A, Błaszkowska J, Kurnatowski P et al. Sandpits as a reservoir of potentially pathogenic fungi for children. Ann Agric Environ Med 2016;23:542–8.
- Wójcik A, Kurnatowski P, Błaszkowska J. Potentially pathogenic yeasts from soil of children's recreational areas in the city of Lódź (Poland). Int J Occup Med Env 2013;26:477–87.
- Yang YL, Lin CC, Chang TP et al. Comparison of human and soil Candida tropicalis isolates with reduced susceptibility to fluconazole. PLoS One 2012;7:e34609.