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Adjusting the water-sensitivity of sugar/boronate-based organogels

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During the investigation of the water-sensitivity of (arylboronate alkylglucoside)-based organogels, we evaluated a series of twelve potential organogelators. They were synthesised in a single step from the corresponding arylboronic acids and alkylglucosides. Eight of them showed organogelation abilities of three solvents (toluene, cyclohexane, ethyl myristate). Conformational minimisations of the potential organogelators permitted to establish a clear relationship between the arylboronate orientation and the gelation effectiveness. Those gels were characterised by rheometry and SEM which revealed a gel-state originating from the self-assembly of the organogelators into long entangled fibres. SAXS confirmed the mode of packing in a hexagonal phase. Gels in toluene were found to be water-sensitive both after addition of a small amount of water and immersion into water. This study demonstrated that the main parameter impacting the water-sensitivity was the length of the alkyl chain at the anomeric position of the glucoside unit, much more than the functionalisation of the arylboronate moiety.

Introduction

Organogels refer to the immobilisation of an organic solvent in the presence of a suitable gelator. Polymeric networks usually provide chemical organogels whereas the hierarchical self-assembly of small molecules belongs to physical gels. Indeed, the latter results from the formation of a 3D network based on low energy interactions between organogelators.¹⁻⁵ The ability of low molecular weight compound to behave as organogelators is linked to their solubility/aggregation balance in a given solvent and therefore to their capacity to self-assemble through van der Waals, hydrogen bonding or/and π - π interactions.^{6,7} Several groups already rationalized the gelation ability of organogelators in function of solvent parameters, permitting to predict gelation in a given solvent.⁷⁻¹⁰ Sugar-based organogelators have played an important role in the development of organogels. They offer an interesting platform which can be further equipped with interacting groups (alkyl chains, aromatic rings, esters, amides, ureas, ..).¹¹⁻¹³ Recently, research groups have taken advantage of the self-assembling process to develop responsive organogels to various stimuli such as heat, light, sound, ions, redox conditions.¹⁴⁻²³ Within this context, sugar-boronate derivatives constitute a recent family of organogelators which already produced gels in

various solvents (Figure 1).²⁴ Starting from readily available glucoside, the selective reaction of boronic acids on position 4,6 of the glucoside represents an easy access to these molecules. Their chemical structures are very similar to benzylidene ones,²⁵⁻³⁵ but a boron atom instead of the acetalic carbon. Compared to the benzylidene function, the boronate one affords a slightly different geometry and a remarkable water sensitivity which deserve to be investigated in depth. Those water-sensitive gels might find applications in drug delivery for topical medicines or as appropriate ingredients for care lotions or cosmetic formulations. Furthermore, similar gelators exhibit important luminescent properties.^{36, 37} Using those boronate/sugar-based organogelators bearing water-sensitive properties could lead to water detection systems. Thus, with the aim to understand and to adjust the water-sensitivity of these organogels, we designed a novel series of sugar-boronate organogelators bearing structural variations both on the aromatic ring of the phenylboronate and at the anomeric position of the glucoside (i.e. aglycone) (Figure 1). Indeed, the length of the aglycone alkyl chain might impact the hydrophobicity of the gels and different substitutions on the aromatic ring of the boronate would modify the water-sensitive properties of this function. It is already known that increasing the steric hindrance around the boron atom (*o*-CH₃, *o*-CF₃) dramatically reduces the hydrolysis kinetics of such boronates. Shimada *et al.* took advantage of this feature to envisage them as protecting groups in synthetic schemes.^{38, 39} In addition, we increased the electron density on the phenyl ring (*p*-OCH₃) which should reduce the electrophilicity of the boron atom and therefore the water-sensitivity of the boronate function.

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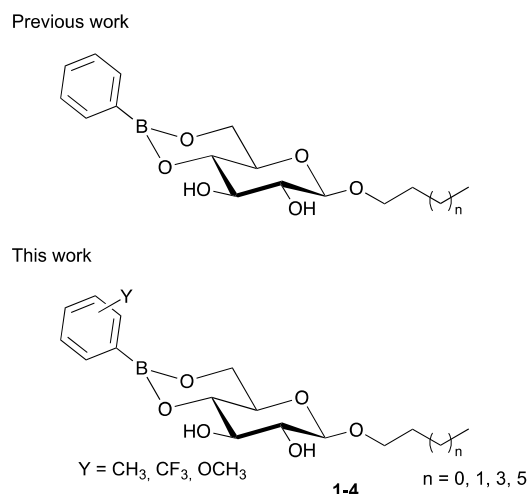


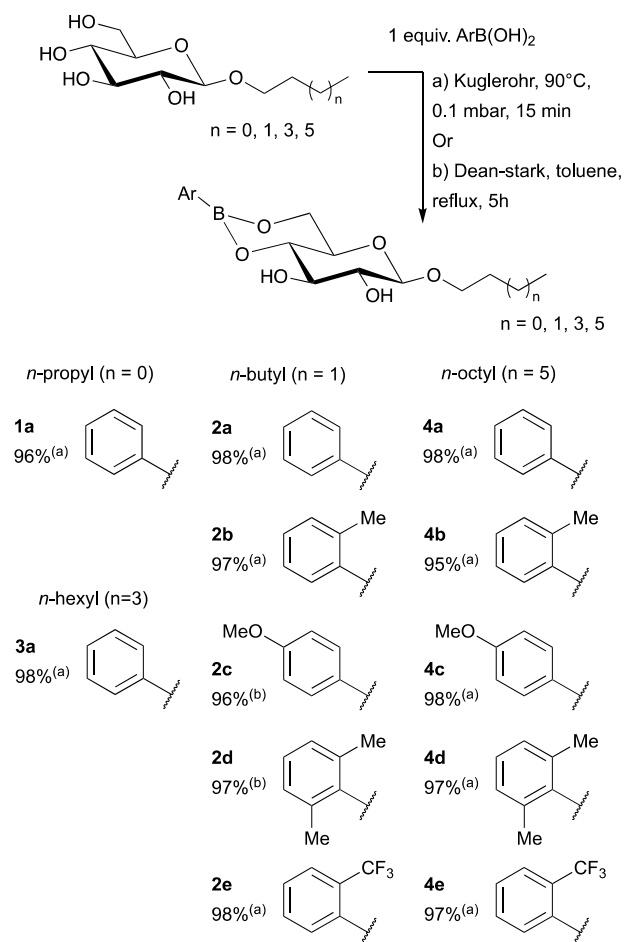
Figure 1 Sugar/boronate-based organogelators.

All compounds were obtained following a straightforward synthetic scheme, providing sufficient amount of each to determine their organogelation properties. First, we evaluated the effect of these relatively small structural changes on the gelation ability. Indeed, despite common structural features the influence of these small structural modifications remains difficult to predict and might enhance or prevent the gelation of a given solvent. Corresponding organogels were then characterised by rheometry, SEM and SAXS. Finally, their water-sensitivities were evaluated upon exposure to a small amount or to a large excess of water. Thus, this study offers to decipher the main molecular parameters favouring the gelation and/or responsible for the water-sensitivity of such organogelators.

Results and discussion

Synthesis

Arylboronate alkylglucosides **1-4** were prepared from the readily available alkyl glucosides in one single step. In fact, the formation of a boronate function requires only the presence of a boronic acid, a diol, and the removal of two water molecules (Scheme 1). Applied to alkyl glucosides, the reaction is regioselective towards the formation of the 4,6-boronate isomers only.⁴⁰ Two experimental procedures were used to perform this esterification, both involved the displacement of the equilibrium by removing the water produced during the transformation. Simple heating (90°C) in a Kugelrohr oven, under reduced pressure (0.1 mbar) and without solvent, provided most of the time successful reactions and excellent yields (95 to 98%) after only 15 min. When these solvent free conditions were not efficient enough due to incorrect mixing of the two powders, for **2c** and **2d** we used a Dean-Stark apparatus with toluene as solvent which led to complete reactions after few hours (5 h) with excellent yields (96-97%). As expected, the regioselectivity was fully in favour of the formation of the boronate function on the position 4,6 of the alkylglucoside as confirmed by NMR analysis. This series of 12 arylboronate alkylglucosides were then engaged in gelation assays.



Scheme 1 Synthesis of aromatic boronate alkylglucosides **1-4**

Gelation assays

The gelation ability of each compound was evaluated in several solvents including three representative ones: an aromatic solvent (toluene), an alkane (cyclohexane) and a biocompatible solvent (ethyl myristate)⁴¹. The goal of this evaluation was to determine the impact of small structural modifications (Scheme 1) on the gelation of organic solvents. In general, all compounds were soluble in ethyl acetate, dichloromethane, chloroform, THF and dioxane and insoluble in diisopropylether. Concerning toluene, cyclohexane and ethyl myristate, the results are shown in Table 1. The four phenylboronate derivatives (**1a-4a**, *n*-propyl, *n*-butyl, *n*-hexyl, *n*-octyl) gave gels in toluene at similar minimal gelation concentrations (MGC) around 10-12 mg ml⁻¹. These MGC are slightly dependent to the length of the alkyl chain, with a lowest MGC for the hexyl derivative (**3a**). The chain length also modified the optical properties of the gels which were transparent for propyl and butyl derivatives (**1a**, **2a**) but opaque for longer chains (**3a**, **4a**). This observation clearly means that different degrees of crystallinity appeared within these materials which can be related to solubility effects. We determined the gelation domain in the Hansen space for compound **2a** and **4a** following the methodology described by Bouteiller *et al.* (See ESI S7 for graphs).⁷ Even if our study was carried out with a limited number of solvents, it seems that longer alkyl chains shift the centre of gelation sphere and

reduce the gelation domain. Therefore, opaque gels for **3a** and **4a** could be linked to a reduced solubility compared to shorter chains (**1a**, **2a**). Also, during VT ^1H NMR experiments, gels prepared with **2a**, **3a** and **4a** in toluene showed broaden signals at 25°C which are characteristic of gel states. These broaden signals were maintained until 60°C, from which the signals became sharp enough to observe all scalar couplings. Interestingly, no significant differences were observed between those gels, demonstrating that the chain length has a limited impact on the gel/sol transition temperature. In cyclohexane, the butyl derivative **2a** provided the best gelation conditions, it furnished a transparent gel at a very low MGC (3 mg mL⁻¹). In ethyl myristate, MGC ranged from 8 to 12 mg mL⁻¹ with the lowest concentration for the octyl counterpart (**4a**). Therefore, one interesting finding that emerges from the first part of this work is that the gelation ability of those compounds can be adjusted to the solvent through a fine modulation of the length of the aglycone alkyl chain. Then, we evaluated the influence of phenyl ring substitutions, with the final aim to control the water-sensitivity of such gels.^{38, 39} However, additional substituents on the phenyl ring also induced a modification of the gelation ability of those molecules. Indeed, the addition of one methyl at the ortho position of the phenyl boronate (**2b**, **4b**) increased the MGC for the gelation of toluene to 15 mg mL⁻¹. Also, a gelation of cyclohexane at 10 mg mL⁻¹ (**4b**) was obtained while **4a** led to insoluble materials. Conversely **2b** gave insoluble material while **2a** was able to gel cyclohexane at a very low MGC. In ethyl myristate, **2b** provided a partial gel only and **4b** gelled this solvent but with an increased MGC compared to **4a**.

The addition of a methoxy group at the para position of the phenylboronate (**2c**, **4c**) had a very little influence on the gelation properties. **2c** and **4c** produced very similar results compared to the simpler phenyl boronate (**2a**, **4a**) both in toluene and in ethyl myristate. However, in cyclohexane, **2c** stayed insoluble and **4c** did not provide a complete gelation even at high concentration (**4c**, 20 mg mL⁻¹). It contrasted much with the results obtained with **2a** (3 mg mL⁻¹). To reduce even more the water-sensitivity, we envisaged the substitution of the phenyl ring with two *o*-methyl groups (**2d**, **4d**) or with one trifluoromethyl group (**2e**, **4e**).^{38, 39} These two modifications dramatically reduced the self-assembling properties of such compounds leading to solutions, even at concentrations higher than 20 mg mL⁻¹. Therefore, it demonstrates again that slight modulations of the molecular structure induce different or even opposite gelation capacities within this series of compounds.

Table 1 Gelation assays.^a

Compound	Toluene	Cyclohexane	Ethyl myristate
1a	G ^T 12	G ^P 2	G ^T 12
2a ²⁴	G ^T 11	G ^T 3	G ^T 10
3a ²⁴	G ^O 10	I	G ^O 10
4a ²⁴	G ^O 12	I	G ^O 8
2b	G ^O 15	I	G ^P 15
4b	G ^O 15	G ^T 10	G ^T 15
2c	G ^T 12	I	G ^O 12
4c	G ^T 10	G ^P 20	G ^T 10
2d	S	S	S
4d	S	S	S
2e	S	S	S
4e	S	S	S

a) I: Insoluble; S: Soluble; P: Precipitate; G^P: Partial gel; G^T: Translucent gel; G^O: Opaque gel. Values indicate minimum gelation concentrations (MGC) in mg mL⁻¹.

Computational studies and conformations

Intrigued by the impact of those relatively small structural variations on the gelation abilities, we investigated the conformation of this series of alkylglucoside arylboronates (Scheme 1) by Density Functional Theory methods. From previous published works on organoborate derivatives, several combinations of functional and basis set were compared to obtain accurate model of arylboronate molecular structure (See ESI Table S6.1). Geometric optimizations were developed at the selected level of theory ωB97XD/cc-pVTZ, as recently used for phenylboronate, to consider weak interactions involved particularly between the aromatic core and the six-membered ring of the boronate function.^{42, 43} Selected optimized geometries of **2a-e** are shown on Figure 2. As expected, the alkyl glucoside moieties have the same conformation for all compounds, but the aromatic ring ends with different orientations. Indeed, the aromatic core adopts a periplanar conformation for **2a**, **2b**, **2c** which are effectively organogelators. However, in the presence of a *o*-CF₃ (**2d**) or two *o*-Me (**2e**), those molecules, that did not provide gels, show increased torsion angles (55° and 52° respectively) with mainly a steric effect (See ESI Table S6.2). Therefore, the self-assembly leading to the formation of gels seems to require a specific molecular geometry in which the arylboronate must adopt a planar conformation. This conclusion is consistent with the mode of self-assembly that we demonstrated earlier for such compounds in which two aromatic rings are self-assembled through a herringbone packing.²⁴ Once the torsion angle increases, aromatic ring orientations modify the aromatic-aromatic interactions preventing an efficient self-assembly. Consequently, the gelation ability is dramatically reduced, explaining why **2d-e** and **4d-e** did not furnish gel.

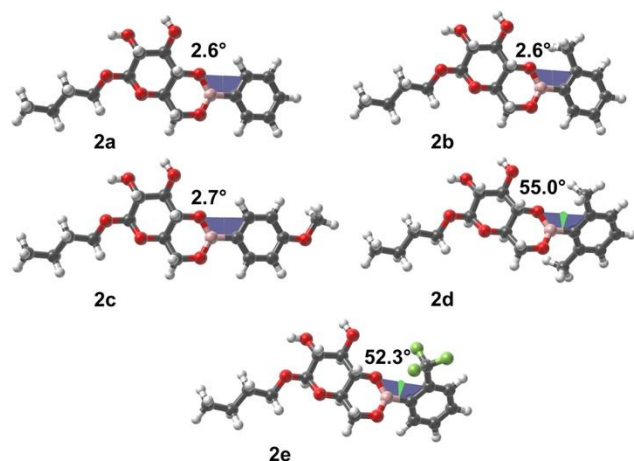


Figure 2 DFT optimized conformations of *n*-butylglucoside arylboronates **2a-e**.

Characterisation of the organogels

Rheometry. Mechanical properties of all gels were investigated by rheological amplitude-sweep experiments. Beforehand, we showed that the rheology of those gels was constant over a large range of frequency (see ESI S2.1. for frequency dependent rheology experiments). Figure 3 gathers graphs of G' and G'' against the applied strain obtained at a frequency of 1 Hz and at the MGC of each gelator in toluene (Graphs for other solvents are available in ESI, S2.2.). At low strain, all gels are characterized by storage moduli typically one order of magnitude higher than the loss moduli confirming the formation of gels that behave as elastic solids at small deformations (Table 2). However, the typical values of G' remains of the order of hundreds to a few thousand of Pa, meaning rather soft gels. Among those organogelators, **1a** led to a gel with the most interesting rheological properties in toluene with a high G' value and the highest G'/G'' ratio. In ethyl myristate, **1a** also gave a high G'/G'' ratio while gelation from **4b** provided the highest G' value (3508 Pa). Note also that some gels present a clear plateau of G' at low strains, which is a signature of a linear elastic regime extending on a large range of amplitude (Figure 3). Oppositely, others (from **2a**, **4a-b**) have elastic properties always reducing as the deformation increases, meaning some intrinsic microscopic rearrangements even at low amplitudes of deformation. Consistently, it is for these gels that the ratio G'/G'' is the smallest. As this amplitude is further increased, all the gels show a yielding behaviour, with G'' eventually dominating. The deduced yield strains vary from 0.02 to 0.2 and depend strongly on the organogelators. Above these yield strains, which corresponds here to low values for gels, the microstructure is irreversibly modified by the imposed shear. As shown in Figure 3, the more the gel is initially elastic (high elastic moduli G' , and a well-defined plateau of G') the lower is its yield strain. All together, these measurements show that one can adjust the macroscopic gel texture and its yielding properties by tuning the molecular chemical structure of the organogelator.

Table 2 Storage and loss moduli at low strain.^a

Compound	Solvent	G'	G''	G'/G''
1a	Toluene	1713	90	19.0
1a	Et myristate	1149	78	14.7
2a	Toluene	2724	213	12.8
2a	Cyclohexane	326	40	8.1
2a	Et myristate	1196	279	4.3
3a	Toluene	1124	111	10.1
3a	Et myristate	206	90	2.3
4a	Toluene	753	341	2.2
4a	Et myristate	1075	287	3.7
2b	Toluene	1827	439	4.2
4b	Toluene	1666	608	2.7
4b	Cyclohexane	552	56	9.9
4b	Et myristate	3508	298	11.8
2c	Toluene	1521	160	9.5
2c	Et myristate	763	229	3.3
4c	Toluene	1955	205	9.5
4c	Et myristate	1758	158	11.1

a) At strain < 1%, 1 Hz frequency.

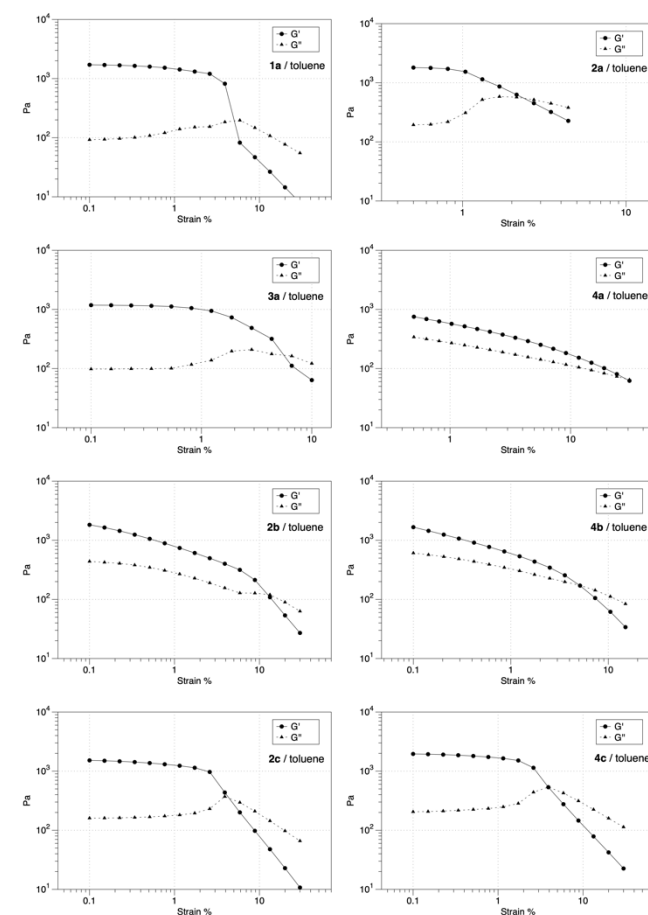


Figure 3 Storage modulus (G' , plain line) and loss modulus (G'' , dashed line) against strain % of organogels obtained in toluene with organogelators **1-4**.

Scanning Electron Microscopy. SEM images were recorded on xerogels obtained after removing the solvent of toluene-based gels (freeze-drying). Networks of fibres are easily observable in most of samples (Figure 4). Only **1a** and **2c** produced aggregated

structures in which thin fibres are visible at high magnification (See ESI, S3). Except these two, fibres are in general few hundred μm long and less than $1\ \mu\text{m}$ width. Keeping in mind that the drying process might cause collapse or other modifications of the fibrillar network, we are convinced that the fibres observed by SEM constitute a representative network responsible for the immobilisation of the solvent as gels. At least, it confirms that the gelation proceeded through the hierarchical self-assembly of the organogelator molecules.

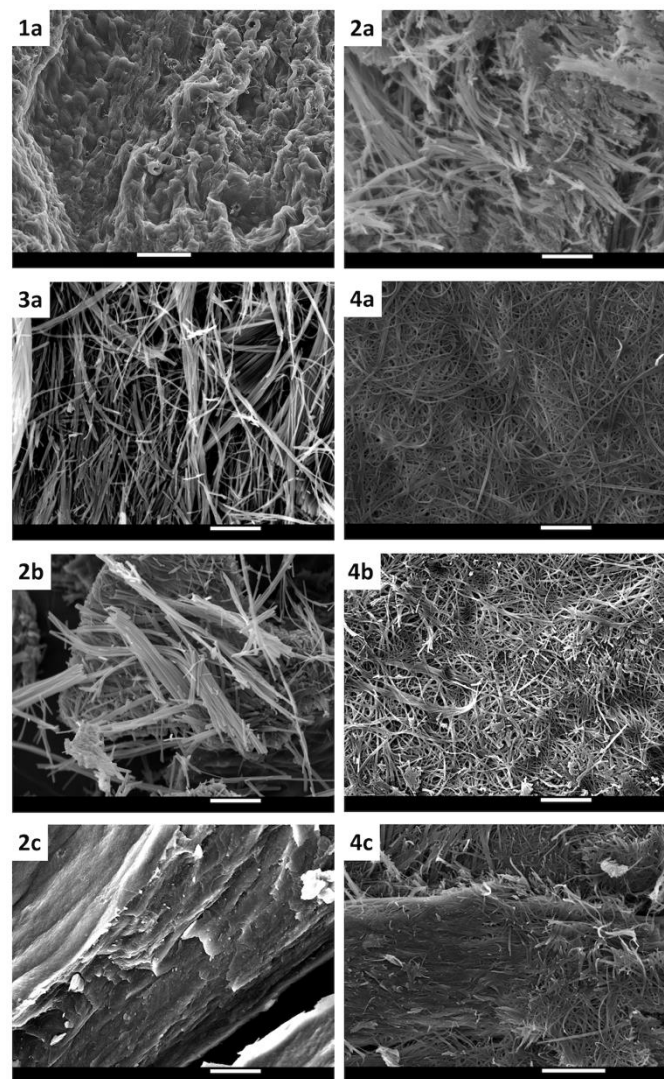


Figure 4 SEM images of xerogels obtained from toluene-based gels. Scale bar: $10\ \mu\text{m}$.

Small-Angle X-Ray Scattering. SAXS analysis can furnish insights in the mode of packing involved in the self-assembly. We were lucky to obtain a well-resolved SAXS diagram for xerogels prepared with **4b** originally coming from gels in toluene and in cyclohexane (Figure 5, See ESI S4 for cyclohexane). Both samples provided superposable data from which we extracted the Bragg peaks gathered in table 3. The corresponding indexation revealed the existence of a hexagonal phase which was assigned without any ambiguity. Consequently, tracing q_{hkl} in a hexagonal lattice, where h , k , and l refer to Miller indices,

gave access to repeating parameters of $a = b = 27.4\ \text{\AA}$ and $c = 5.1\ \text{\AA}$. These lattice parameters are compatible with the organisation of the aromatic ring into a herringbone packing and with the size of the organogelator (**4b**) (Figure 6). It is noteworthy that this type of self-assembly was already described for **4a** with slightly different lattice parameters ($a = b = 26.8\ \text{\AA}$; $c = 5.5\ \text{\AA}$).²⁴ It is noteworthy that the similar lattice parameters observed for **4a** and **4b** support comparable conformations and torsion angles as estimated by DFT. Also, the crystal packing of a close boronate derivative (methyl-2-deoxy- α -D-glucopyranoside 4,6-phenylboronate) involves the same molecular arrangement.⁴⁴

Interested by the mode of packing of those compounds, we performed SAXS analysis for all xerogels (see ESI). Despite less resolved data, all SAXS diagrams have a common pattern compatible with a similar self-assembly. Therefore, the herringbone packing constitutes a general feature involved in the self-assembling process of such molecules.

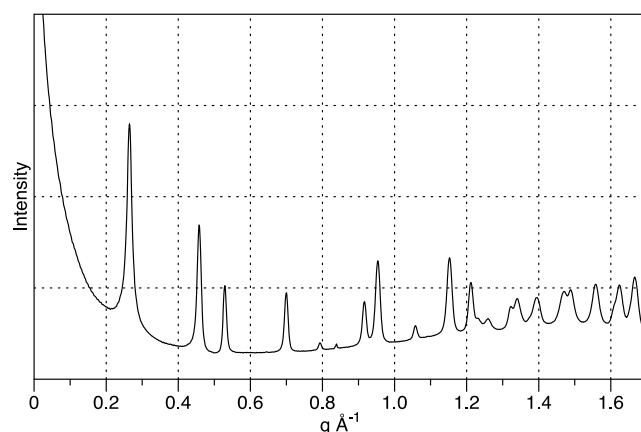


Figure 5 SAXS profiles of xerogel obtained with **4b** (toluene).

Table 3 Theoretical and observed indexations of the hexagonal phase shown in Figure 5 ($a = b = 27.4\ \text{\AA}$; $c = 5.1\ \text{\AA}$).

h	k	l	Q_{calc}^a	Q_{obs}^b
1	0	0	0.2642	0.2642
1	1	0	0.4577	0.4578
2	0	0	0.5285	0.5286
2	1	0	0.6991	0.6991
3	0	0	0.7927	0.7929
2	2	0	0.9153	0.9153
3	1	0	0.9527	0.9526
4	0	0	1.0569	1.0570
3	2	0	1.1517	1.1513
4	1	0	1.2108	1.2103
1	0	1	1.2576	1.2580
1	1	1	1.3120	1.3195
2	0	1	1.3383	1.3383
4	2	0	1.3982	1.3894
3	0	1	1.4629	1.4636
3	1	1	1.5554	1.5560
4	0	1	1.6214	1.6218

a) Theoretical indexation, b) Observed indexation.

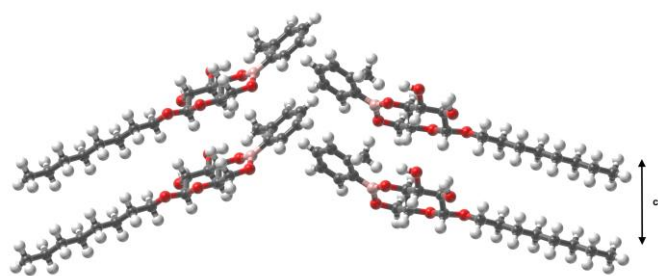
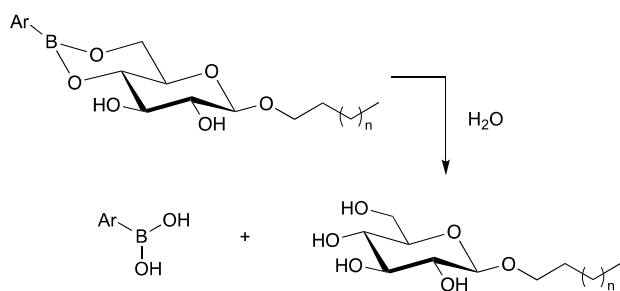


Figure 6 Representation of the self-assembly of **4b** following a herringbone packing between aromatic rings. (Octyl chain are shown as extended for clarity).

Water-sensitivity

In our preliminary study, we demonstrated that those organogels get disrupted upon contact with water.²⁴ At that time we showed that this water-sensitivity originated from the hydrolysis of the boronate function (Scheme 2). Then, it leads back to the corresponding arylboronic acid and alkylglucoside. These two compounds are not gelators themselves, thus it finally ends to a solution. Furthermore, within other contexts, arylboronate function hydrolysis has also been described.^{42, 45}



Scheme 2 Hydrolysis of arylboronate alkylglucoside

Here, we investigated more thoroughly, the water-sensitivity of those novel organogels with the aim to determine the molecular parameters modulating the gel disruption upon contact with water. This comparative study was performed with gels prepared with toluene, following two protocols: A) Addition of a small amount of water (5% v/v) on the top of the gels and visual follow-up until total disruption (over a maximum of one week); B) Video follow-up of a piece of gel (cylinder of \varnothing 2.4 x 1 cm) immersed in a large excess of water. Then image treatment permitted to plot graphs describing the gel disruptions against time.

Addition of 5% v/v of water permitted to establish that the length of the alkyl chain dramatically impacts the duration of the water-induced disruption of the toluene-based gels (Protocol A, Table 4). Indeed, the gel obtained with the propyl derivative (**1a**) disappeared in 1h30 followed by the one from the butyl derivative **2a** (3h00). Under the same conditions, corresponding gels from hexyl and octyl counterparts (**3a** and **4a**) were kept untouched even after one week of water exposure. The substitution of the aromatic ring of the boronate also modulated the water-sensitivity, increasing the disruption time from 3h00 (**2a**) to 8h00 (**2b**) and 4h00 (**2c**). Insertion of one bulky *o*-Me (**2b**) group induced a steric hindrance which contribute

to a protective effect on the boronate function in the self-assembly. It is noteworthy that the steric hindrance brought by the extra *o*-Me (**2b**) group decreased much more the water sensitivity of these gels than the electron donating *p*-OMe group (**2c**). Nevertheless, increasing the alkyl chain to octyl (**4c**, **4b**) avoid the hydrolytic disruption under these conditions, meaning that the aglycone chain length has a major influence here.

Table 4 Water-sensitivity in the presence of 5%v/v of water.^a

Compound	Time (h)
1a	1:30
2a	3:00
3a	> week ^b
4a	> week ^b
2b	8:00
4b	> week ^b
2c	4:00
4c	> week ^b

a) Gels were prepared with toluene at the MGC. b) No hydrolysis after one week

The second protocol (B) allowed to evaluate the behaviour of those gels in the presence of a large excess of water. A piece of gel was immersed in water and its disappearance was recorded by a camera placed aside of the gels (See ESI for GIF files). Therefore, the collected images were representative of the remaining gel and data treatment permitted to plot graphs describing the disruption progress against time (% of remaining gel against time (min)). Figure 7 reports the water-sensitivity of toluene-based organogels as a function of the chain length. For all experiments, we fixed the organogelator concentrations to 12 mg mL⁻¹ for comparison purpose. Here, the water, surrounding the bottom and the sides of the gel discs, induced gel disappearance in few minutes to few hours. In addition, the process turned out to be isotropic, as the shape of the gels remained similar with time. As for the previous study with 5% v/v of water, the propyl derivative (**1a**) was the fastest to break down (80% of gel in 20 minutes). The butyl derivative (**2a**) required more than double time (53 min) to lose 80% of gel state. At this stage, one can imagine that longer chains would lead to lower water sensitivity. However, octyl derivative (**4a**) appeared to disrupt faster than the hexyl counterpart (**3a**) (**4a**, 80% in 80 min; **3a**, 80% in 185 min).

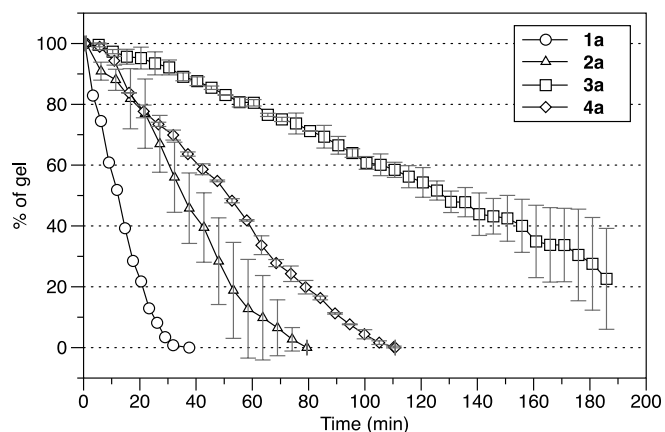


Figure 7 Percentage of remaining gel against time (min) after immersion in water. Gels prepared with **1a-4a** at 12 mg mL⁻¹ in toluene.

Then we compared organogelators bearing the same alkyl chain (*n*-butyl) but different substituents on the phenylboronate moiety (**2a,b,c**) (Figure 8). For comparison purpose, the concentration was fixed to 16 mg mL⁻¹. Note that the gel obtained with **2c** (*p*-OMe) behaved differently than the two others at the beginning of the water-sensitivity experiments. For some of the gels, in particular with **2c**, determining the accurate boundaries of the gel disc is optically complex. For such gels, a halo of solvent appears to be captured below and in the vicinity of the gel; this leads to an overestimation of the actual size of the gel as the boundary which is optically detected includes this halo. As a consequence, even though the gel continuously vanishes, an apparent flattening of the curve can be obtained (Figure 8, **2c**). Regarding the time to reach 80% of disruption, surprisingly the gel from **2b** showed the fastest disruption and those from **2a-2c** provided very similar results with disruption times longer by only 20 minutes.

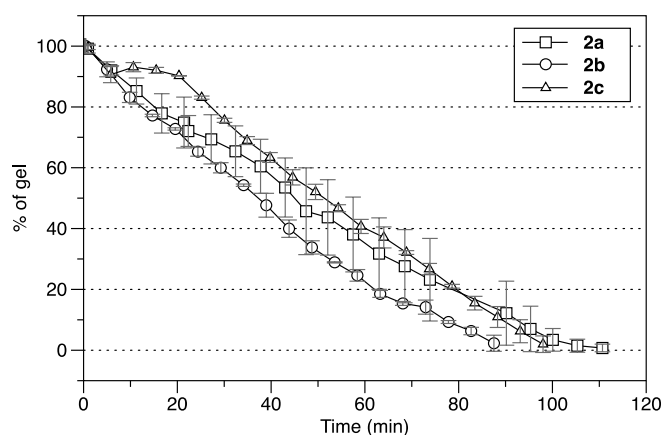


Figure 8 Percentage of remaining gel against time (min) after immersion in water. Gels prepared with **2a, 2b, 2c** at 16 mg mL⁻¹ in toluene.

To elucidate the main factors determining the water-sensitivity, we envisaged several plausible explanations: 1) the water-sensitivity would be related to the electrophilicity of the boronate function itself; 2) different modes of packing may conduce to different hydrolysis rates; 3) differences in

mechanical properties of the gels would be responsible for the water-sensitivity differences; 4) the fibre networks (size, density, ..) would facilitate or not the water entry into the gels. Shimida *et al.* already explained the stability of some arylboronates against water within an organic synthesis context, by comparing orbital energy levels.³⁹ Similarly, we established the energy levels of molecular orbitals for each compound **1a-4a** at the ω B97XD/cc-pVTZ level of theory (See ESI, Tables S6.3 and Figures S6.3). All compounds showed very close LUMO energies confirming that the different arylboronate functions should show similar reactivities that do not depend on the aglycone alkyl chain. Same conclusion has raised by considering the water-sensitivities of the gels from **2a, 2b** and **2c**. Indeed, **2c** should provide the least water-sensitive organogel when examining the relative LUMO energy levels (See ESI, Tables S6.3). However, gels with **2b** demonstrated a weaker water-sensitivity than with **2a** and **2c** (protocol A), and these three gels (in toluene) reached close disruption times after immersion in water (Protocol B). Therefore, these evolutions over time support influences of other factors than the electrophilicity of the boronate function. As showed here in earlier paragraphs, SAXS studies concluded in a very similar mode of packing for all organogelators. Thus, the different hydrolysis rates are not linked to different types of self-assembly. Very different rheological properties were also highlighted between gels obtained in toluene with **3a** and **4a** for instance. Indeed, the gel prepared with **3a** had stronger moduli than the ones for **4a**. Such a gap between both G' and G'' values can be associated to differences in dynamics of disruption which may find an origin in an easier diffusion of water molecule in the gels. However, this trend is far from being general, for example, gels obtained with **2a** which showed a higher water-sensitivity than with **3a**, has though stronger moduli. Then, the main structural feature governing the water-sensitivity is the chain length of the aglycone. One can conclude that increasing of the global hydrophobicity of the self-assembly reduces the water-sensitivity. In fact, hexyl and octyl derivatives have a stronger resistance upon water contact compared to propyl and butyl, whatever the structure of the aryl boronate. The specific inversion of this trend between hexyl (**3**) and octyl (**4**) is hard to explain. As the disruption dynamics probably start first on the outside of the fibres, an explanation can be based on the comparison of the size and the shape of the fibre networks. The SEM images of the corresponding xerogels displayed clear differences which can facilitate or not the hydrolysis step. Gel in toluene with **4a** is characterised by thinner fibres than the gel obtained with **3a** which reduces the penetration of water molecules for the latter. Note also that compounds **1a, 2a** and **2c** which led to the most water-sensitive gels present dense fibre networks constituted of thin fibres.

Finally, the water-sensitivity of those systems would be related to both the hydrophobicity of the self-assembly and the specific surfaces available for the water molecules to access the hydrolysable boronate function.

Conclusions

The synthesis of twelve arylboronate alkylglucosides was achieved in a single chemical step with excellent yields. It covered structural variations both at the anomeric position and on the aromatic ring. This series of potential or effective organogelators allowed to evaluate the impact of these structural variations at different scales both on the gelation ability and on the behaviour of the corresponding gels. One first conclusion raising from this work established that the conformation of the boronate function clearly modifies the gelation ability of such compounds. Rather planar conformations are required to obtain gelation of toluene, ethyl myristate and cyclohexane at low to very low minimal gelation concentrations. Interesting properties of these series of organogelators come from their capacity to disrupt upon contact with water. Even if the hydrolysis of the boronate function remains at the origin of the gel disruptions, the main structural feature that matters is the length of the alkyl chain aglycone. In general, the shorter the alkyl chain the higher is the water-sensitivity. In addition, careful analysis of the set of physicochemical data (Rheology, SEM, SAXS) allowed to identify the nature of the fibre network also as a determining parameter for the water-sensitivity of these gels (more than the mode of packing or the intrinsic reactivity of the boronate function). Therefore, this study provided important insights on the main structural features controlling the behaviour of this family of organogelators both for the gelation of apolar organic solvents and for their water-sensitivity. Additional structural modifications and gelation conditions would contribute in the future to reach a great control and a better modulation of both gelation and water-sensitivity properties of such organogels.

Experimental

General Information. ^1H NMR spectra (300 MHz, 400 MHz), ^{13}C NMR (75 MHz, 101 MHz), ^{19}F (282 MHz) and ^{11}B (96 MHz) were recorded on Bruker AC 300 and AC 400 spectrometers. Chemical shifts are given in ppm and coupling constants J in Hz. Multiplicities are presented as follows: s = singlet, d = doublet, t = triplet, q = quartet, quint = quintuplet, hex = hexuplet, m = multiplet, br = broad. The carbon bearing the boron atom was not observed due to multiplet structure: 1-1-1-1 quadruplet for ^{11}B -bearing ^{13}C (80%) and 1-1-1-1-1-1-1 septuplet for ^{10}B -bearing ^{13}C (20%) and to quadrupolar relaxation mechanism of both B isotopes.⁴⁶ Attribution numbers are related to the sugar nomenclature numbering. High-resolution mass spectra (HRMS) were recorded, either on a Bruker MaXis 4G, an Agilent 6510, or a Thermo Fisher Q-Exactive spectrometer (Centre Régional de Mesures Physiques de l'Ouest, Rennes) using positive ion Electron-Spray ionization techniques. Melting points were measured on a melting point apparatus Stuart SMP10 and are uncorrected. Specific rotation (in $\text{deg cm}^3 \text{g}^{-1}$) was measured on a Perkin Elmer-341 polarimeter.

Gel formation. The gels were prepared by mixing the appropriate amount of gelator into the chosen organic solvent

at various concentrations in capped tubes. The tubes were heated at 60°C (cyclohexane), 80°C (toluene) or 120°C (fatty esters) for 1 h or until clear solutions were obtained. Clear solutions were then cooled down to room temperature to allow the formation of a gel. MGC (Minimal Gelation Concentration) was determined as the lowest concentration leading to a complete gelation of the solvent (The concentration was reduced by 1 mg mL^{-1} from 20 mg mL^{-1} to the MGC).

Rheology. Organogel samples were presented under disc form (4 cm diameter and a few mm of thickness, depending on the organogelators). Rheological measurements were performed on an Anton-Paar MCR301 equipped with an upper plate of 75 mm diameter. Amplitude-sweep experiments were performed, where the frequency (ω) was fixed to 1 Hz and the amplitude deformation (γ) was gradually increased from 0 to 50% of shearing. Storage modulus G' and the loss modulus G'' were obtained at 25°C.

Complementary frequency-sweep experiments were also performed and showed that G' is always bigger than G'' over the whole range of frequencies (0,05 Hz to 10Hz).

Scanning Electron Microscopy (SEM). Metallisation by Au/Pd. Scanning Electron Microscopy (SEM) of xerogels were evaluated using the JEOL IT 300 Scanning Electron Microscope. Samples were collected and deposited on a Teflon plot. Each sample was examined using a voltage of 5 or 10 kV. Images were analysed by SMileView software.

Small Angle X-Ray Scattering (SAXS). Xerogel samples were prepared into capillaries. SAXS experiments were performed using X-ray patterns collected with a Pilatus 300k (Dectris, Grenoble, France), mounted on a microsource X-ray generator GeniX 3D (Xenocs, Sassenage, France) operating at 30 watts. The monochromatics $\text{CuK}\alpha$ radiation is of $\lambda = 1.541 \text{ \AA}$. The diffraction patterns were therefore recorded for reciprocal spacing $q = 4\pi \cdot \sin\theta/\lambda$ in a range of repetitive distances from 0.015 \AA^{-1} (418 \AA) and 1.77 \AA^{-1} (8 \AA). Images were transformed to graphics using the software program Fit2D (ESRF).

Hydrolysis experiments. Procedure A. 5% v/v of water was added on the top of gels formed in toluene (1 mL). Gelators were used at their minimal gelation concentrations. The disappearance of the gels was observed by eyes. **Procedure B.** Gels were prepared into 5 mL beakers, making possible to obtain gels approximately 10 mm in height and 24 mm in diameter. Gels were then unmoulded using a needle. Gels were immersed in water and maintained in place by a rod. A camera was installed to view the gel from the side and pictures were taken every 10 seconds. The images obtained were processed using ImageJ software permitting to plot the % of remaining gel against time.

Computational details.

Computational studies were carried out on 12-core Apple computing cluster using density functional theory (DFT) methods, as instituted in Gaussian 09 (version D.01) program,

tightening self-consistent field convergence thresholds (10^{-8} au).⁴⁷ Preliminary results from several hybrid functional/basis combinations have been compared (See ESI, Table S6.1). All arylboronate alkylglucosides were optimized at ω B97XD/cc-pVTZ and B3LYP/6-311+G(d,p) levels of theory and the first ω B97XD/cc-pVTZ was selected to consider dispersion forces.^{48, 49} So, torsion, distances, molecular orbital energies were given with this first combination as Figure 2 (See ESI, Tables S6.2). The vibrational analysis was performed at the same levels of theory to confirm that the optimized geometry is a true minimum (no imaginary frequency). Then evolution and convergence of geometric optimizations and energy minimization of frontier molecular orbitals were visualized and analysed with GaussView v5.2 in particular for distances and dihedral angles (ESI, Tables S6.2 and Figure S6.3).⁵⁰

General procedure for the synthesis of boronate glucosides.

Procedure A. In a 25 mL round bottom flask, alkylglucoside (0.3 mmol) and boronic acid (0.3 mmol) were added. The mixture was then stirred in a Kugelrohr distillation apparatus at 90°C; under 0.1 mm Hg for 15 min to give pure product. Compounds were washed by diisopropylether when necessary.

Procedure B. In a 25 mL round bottom flask, alkylglucoside (0.3 mmol) and boronic acid (0.3 mmol) were added. Toluene (15 mL) was added, and the reaction mixture was heated under stirring at reflux in a Dean-Stark Apparatus. After 5h, the toluene was evaporated under reduced pressure to afford pure products. Compounds were washed by diisopropylether when necessary.

Compounds **2a, 3a, 4a** were already described.²⁴

n-Propyl- β -D-glucopyranoside 4,6-phenylboronate **1a**.

89 mg (96%). White solid; mp = 184-188°C. $[\alpha]_D^{25} = -52$ (C 1, CH₂Cl₂). ¹H NMR (300 MHz, CDCl₃) δ 7.85-7.77 (m, 2H), 7.48-7.40 (m, 1H), 7.38-7.32 (m, 2H), 4.44 (d, $J = 7.7$ Hz, 1H, H1), 4.30 (dd, $J = 10.4, 5.4$ Hz, 1H, H6), 4.01 (t, $J = 10.4$ Hz, 1H, H6), 3.88 (dt, $J = 9.5, 6.8$ Hz, 1H, OCH₂), 3.81-3.71 (m, 2H, H3 H4), 3.62-3.48 (m, 3H, OCH₂, H2, H5), 2.81 (brs, 1H, OH), 2.53 (brs, 1H, OH), 1.68 (hex, $J = 7.3$ Hz, 2H), 0.96 (t, $J = 7.4$ Hz, 3H). ¹³C NMR (75 MHz, CDCl₃) δ 134.3, 131.4, 127.8, 103.4, 75.1 (C1), 74.7 (C3), 74.3 (C4), 72.3 (C5), 68.7 (OCH₂), 64.2 (C6), 23.0, 10.5. The carbon α to the boron atom was not observed. ¹¹B NMR (96 MHz, CDCl₃) δ 27.0. HRMS (ESI+) (M+Na)⁺ calculated for C₁₅H₂₁O₆¹¹BNa 331.1323, found 331.1324.

n-Butyl- β -D-glucopyranoside 4,6-*o*-methylphenylboronate **2b**.

98 mg (97%). White solid; mp = 157-159°C. $[\alpha]_D^{25} = -62$ (C 1, CH₂Cl₂). ¹H NMR (300 MHz, CDCl₃) δ 7.75 (d, $J = 7.0$ Hz, 1H), 7.30 (t, $J = 7.2$ Hz, 1H), 7.15 (t, $J = 7.0$ Hz, 1H), 7.14 (d, $J = 7.3$ Hz, 1H), 4.42 (d, $J = 7.7$ Hz, 1H, H1), 4.28 (dd, $J = 10.4, 5.3$ Hz, 1H, H6), 3.99 (t, $J = 10.3$ Hz, 1H, H6), 3.91 (dt, $J = 10.0, 6.8$ Hz, 1H, OCH₂), 3.83-3.67 (m, 2H, H4 H2), 3.65-3.44 (m, 3H, H3 OCH₂ H5), 3.22 (brs, 1H, OH), 3.06 (brs, 1H, OH), 2.51 (s, 3H, CH₃), 1.65 (quint, $J = 7.2$ Hz, 2H), 1.40 (hex, $J = 7.4$ Hz, 2H), 0.94 (t, $J = 7.3$ Hz, 3H). ¹³C NMR (75 MHz, CDCl₃) δ 143.9, 135.0, 130.3, 129.8, 124.5, 103.0 (C1), 74.7 (C4), 74.2 (C3), 73.8 (C2), 70.0 (OCH₂), 68.2 (C5), 63.7 (C6), 31.3, 22.3, 18.8, 13.5. The carbon α to the boron atom

was not observed. ¹¹B NMR (96 MHz, CDCl₃) δ 28.2. HRMS (ESI+) (M+Na)⁺ calculated for C₁₇H₂₅O₆¹¹BNa 359.1636, found 359.1637.

n-Butyl- β -D-glucopyranoside 4,6-*p*-methoxyphenylboronate **2c**. 101 mg (96%). White solid; mp = 178-180°C. $[\alpha]_D^{25} = -69$ (C 1, CH₂Cl₂). ¹H NMR (300 MHz, CDCl₃) δ 7.75 (d, $J = 8.6$ Hz, 2H), 6.87 (d, $J = 8.6$ Hz, 2H), 4.41 (d, $J = 7.7$ Hz, 1H, H1), 4.26 (dd, $J = 10.4, 5.4$ Hz, 1H, H6), 4.00-3.86 (m, 2H, H6 OCH₂), 3.80 (s, 3H, OCH₃), 3.76-3.73 (m, 2H, H4 H3), 3.46-3.61 (m, 3H, OCH₂ H2 H5), 3.31 (brs, 1H, OH), 3.06 (brs, 1H, OH), 1.62 (quint, $J = 7$ Hz, 2H), 1.39 (hex, $J = 7.3$ Hz, 2H), 0.93 (t, $J = 7.4$ Hz, 3H). ¹³C NMR (75 MHz, CDCl₃) δ 161.8, 135.6, 122.2 (br), 112.9, 102.9 (C1), 74.7 (C4), 74.1 (C3), 73.8 (C2), 70.0 (OCH₂), 68.3 (C5), 63.7 (C6), 54.8, 31.3, 18.8, 13.5. The carbon α to the boron atom was not observed. ¹¹B NMR (96 MHz, CDCl₃) δ 27.7. HRMS (ESI+) (M+Na)⁺ calculated for C₁₇H₂₅O₇¹¹BNa 375.1585, found 375.1587.

n-Butyl- β -D-glucopyranoside 4,6-*o*-dimethylphenylboronate **2d**.

102 mg (97%). Colorless oil. $[\alpha]_D^{25} = -62$ (C 1, CH₂Cl₂). ¹H NMR (300 MHz, CDCl₃) δ 7.12 (t, $J = 7.5$ Hz, 1H), 6.94 (d, $J = 7.6$ Hz, 2H), 4.42 (d, $J = 7.8$ Hz, 1H, H1), 4.28 (dd, $J = 10.4, 5.3$ Hz, 1H, H6), 4.01 (t, $J = 10.4$ Hz, 1H, H6), 3.89 (dd, $J = 9.4, 6.9$ Hz, 2H, H4 OCH₂), 3.78 (t, $J = 9.2$ Hz, 1H, H3), 3.73-3.44 (m, 3H, OCH₂ H2 H5), 3.20 (brs, 1H, OH), 2.95 (brs, 1H, OH), 2.34 (s, 6H, CH₃), 1.63 (quint, $J = 7.2$ Hz, 2H), 1.39 (hex, $J = 7.0$ Hz, 2H), 0.94 (t, $J = 7.4$ Hz, 3H). ¹³C NMR (75 MHz, CDCl₃) δ 140.4, 133.6 (br), 129.0, 126.5, 103.4 (C1), 75.0 (C4), 75.0 (C3), 74.0 (C2), 70.4 (OCH₂), 68.7 (C5), 64.4 (C6), 31.7, 22.3, 19.2, 13.9. The carbon α to the boron atom was not observed. ¹¹B NMR (96 MHz, CDCl₃) δ 31.2. HRMS (ESI+) (M+Na)⁺ calculated for C₁₈H₂₇O₆¹¹BNa 373.1792, found 373.1792.

n-Butyl- β -D-glucopyranoside 4,6-*o*-trifluoromethylphenylboronate **2e**.

114 mg (98%). Colorless oil. $[\alpha]_D^{25} = -62$ (C 1, CH₂Cl₂). ¹H NMR (300 MHz, CDCl₃) δ 7.71- 7.60 (m, 2H), 7.54-7.44 (m, 2H), 4.44 (d, $J = 7.7$ Hz, 1H, H1), 4.29 (dd, $J = 10.4, 5.3$ Hz, 1H, H6), 4.01 (t, $J = 10.4$ Hz, 1H, H6), 3.90 (dt, $J = 9.5, 6.8$ Hz, 1H, OCH₂), 3.80 (t, $J = 9.2$ Hz, 1H, H4), 3.70 (t, $J = 9.0$ Hz, 1H, H3), 3.63-3.48 (m, 3H, OCH₂ H2 H5), 3.06 (brs, 1H, OH), 2.88 (brs, 1H, OH), 1.63 (quint, $J = 6.9$ Hz, 2H), 1.40 (hex, $J = 7.2$ Hz, 2H), 0.93 (t, $J = 7.3$ Hz, 3H). ¹³C NMR (75 MHz, CDCl₃) δ 134.4, 133.85 (q, $J = 31.2$ Hz), 131.3, 130.25, 126.0 (q, $J = 5.0$ Hz), 125.2 (q, $J = 273.4$ Hz), 103.9 (C1), 75.6 (C4), 75.3 (C3), 74.5 (C2), 70.9 (OCH₂), 68.8 (C5), 65.0 (C6), 32.2, 19.7, 14.35. The carbon α to the boron atom was not observed. ¹⁹F NMR (282 MHz, CDCl₃) δ -59.4. ¹¹B NMR (96 MHz, CDCl₃) δ 28.7. HRMS (ESI+) (M+Na)⁺ calculated for C₁₇H₂₂¹¹BF₃O₆Na 413.1353, found 413.1358.

n-Octyl- β -D-glucopyranoside 4,6-*o*-methylphenylboronate **4b**.

112 mg (95%). White solid; mp = 142-144°C. $[\alpha]_D^{25} = -70$ (C 1, CH₂Cl₂). ¹H NMR (300 MHz, CDCl₃) δ 7.75 (dd, $J = 7.6, 1.6$ Hz, 1H), 7.30 (t, $J = 7.1$ Hz, 1H), 7.16 (t, $J = 6.9$ Hz, 1H), 7.14 (d, $J = 7.4$ Hz, 1H), 4.44 (d, $J = 7.7$ Hz, 1H, H1), 4.31 (dd, $J = 10.4, 5.4$ Hz, 1H, H6), 4.01 (t, $J = 10.4$ Hz, 1H, H6), 3.91 (dt, $J = 9.3, 6.8$ Hz, 1H,

OCH₂), 3.84-3.68 (m, 2H, H₃ H₄), 3.63-3.50 (m, 3H, OCH₂ H₂ H₅), 2.75 (brs, 1H, OH), 2.53 (brs, 1H, OH), 2.51 (s, 3H, CH₃), 1.64 (quint, *J* = 7.0 Hz, 2H), 1.20-1.43 (m, 10H), 0.88 (t, *J* = 6.7 Hz, 3H). ¹³C NMR (75 MHz, CDCl₃) δ 144.9, 135.9, 131.3, 130.8, 125.4, 103.8 (C₁), 75.6 (C₄), 75.2 (C₃), 74.7 (C₂), 71.3 (OCH₂), 69.1 (C₅), 64.6 (C₆), 32.4, 30.2, 29.9, 29.8, 26.5, 23.2, 23.1, 14.7. The carbon α to the boron atom was not observed. ¹¹B NMR (96 MHz, CDCl₃) δ 27.8. HRMS (ESI+) (M+Na)⁺ calculated for C₂₁H₃₃O₆¹¹BNa 415.2262, found 415.2263.

n-Octyl-β-D-glucopyranoside 4,6-*p*-methoxyphenylboronate **4c**. 120 mg (98%). White solid; mp = 156-158°C. [α]_D²⁰ = -51 (C 1, CH₂Cl₂). ¹H NMR (300 MHz, CDCl₃) δ 7.75 (d, *J* = 8.5 Hz, 2H), 6.87 (d, *J* = 9.0 Hz, 2H), 4.42 (d, *J* = 7.7 Hz, 1H, H₁), 4.26 (dd, *J* = 10.4, 5.3 Hz, 1H, H₆), 3.97 (t, *J* = 10.2 Hz, 1H, H₆), 3.88 (dd, *J* = 9.4, 6.9 Hz, 1H, OCH₂), 3.81 (s, 3H, OCH₃), 3.77-3.73 (m, 2H, H₃ H₄), 3.46-3.62 (m, 3H, H₂ OCH₂ H₅), 3.18 (brs, 1H, OH), 2.93 (brs, 1H, OH), 1.63 (quint, *J* = 7.0 Hz, 2H), 1.20-1.40 (m, 10H), 0.88 (t, *J* = 6.2 Hz, 3H). ¹³C NMR (75 MHz, CDCl₃) δ 162.2, 136.1, 113.4, 103.4 (C₁), 75.1 (C₄), 74.6 (C₃), 74.3 (C₂), 70.8 (OCH₂), 68.7 (C₅), 64.1 (C₆), 55.2, 31.9, 29.7, 29.5, 29.3, 26.0, 22.8, 14.2. The carbon α to the boron atom was not observed. ¹¹B NMR (96 MHz, CDCl₃) δ 21.4. HRMS (ESI+) (M+Na)⁺ calculated for C₂₁H₃₃O₇¹¹BNa 431.2212, found 431.2213.

n-Octyl-β-D-glucopyranoside 4,6-*o*-dimethylphenylboronate **4d**.

118 mg (97%). Colorless oil. [α]_D²⁰ = -53 (C 1, CH₂Cl₂). ¹H NMR (300 MHz, CDCl₃) δ 7.12 (t, *J* = 7.5 Hz, 1H), 6.95 (d, *J* = 7.6 Hz, 2H), 4.46 (d, *J* = 7.7 Hz, 1H, H₁), 4.31 (dd, *J* = 10.5, 5.4 Hz, 1H, H₆), 4.04 (t, *J* = 10.4 Hz, 1H, H₆), 3.91 (dt, *J* = 9.5, 6.2 Hz, 1H, OCH₂), 3.81 (t, *J* = 9.2 Hz, 1H, H₄), 3.71 (td, *J* = 9.2, 2.1 Hz, 1H, H₃), 3.66-3.49 (m, 3H, H₅ H₂ OCH₂), 2.82 (d, *J* = 2.2 Hz, 1H, OH), 2.56 (d, *J* = 2.4 Hz, 1H, OH), 2.35 (s, 6H, CH₃), 1.64 (quint, *J* = 7.1 Hz, 2H), 1.29 (m, 10H), 0.89 (t, *J* = 6.5 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 140.4, 133.6 (br), 129.0, 126.5, 103.4 (C₁), 75.0 (C₄), 75.0 (C₃), 74.0 (C₂), 70.8 (OCH₂), 68.7 (C₅), 64.4 (C₆), 31.0, 29.7, 29.5, 29.3, 26.0, 22.8, 22.3, 14.2. The carbon α to the boron atom was not observed. ¹¹B NMR (96 MHz, CDCl₃) δ 31.15. HRMS (ESI+) (M+Na)⁺ calculated for C₂₂H₃₅O₆¹¹BNa 429.2418, found 429.2421.

n-Octyl-β-D-glucopyranoside 4,6-*o*-trifluoromethylphenylboronate **4e**.

130 mg (97%). Colorless oil. [α]_D²⁰ = -55 (C 1, CH₂Cl₂). ¹H NMR (300 MHz, CDCl₃) δ 7.80 (d, *J* = 8.1 Hz, 2H), 7.59 (t, *J* = 7.9 Hz, 2H), 4.45 (d, *J* = 7.6 Hz, 1H, H₁), 4.29 (dd, *J* = 10.3, 5.4 Hz, 1H, H₆), 4.01 (t, *J* = 10.3 Hz, 1H, H₆), 3.89 (dt, *J* = 9.5, 6.9 Hz, 1H, OCH₂), 3.80 (t, *J* = 9.2 Hz, 1H, H₄), 3.71 (t, *J* = 9.2 Hz, 1H, H₃), 3.62-3.50 (m, 3H, OCH₂ H₂ H₅), 1.63 (quint, *J* = 7.0 Hz, 2H), 1.29 (m, 10H), 0.89 (t, *J* = 6.9 Hz, 3H). ¹³C NMR (75 MHz, CDCl₃) δ 134.0, 133.4 (q, *J* = 30.9 Hz), 130.9, 129.85, 125.6 (q, *J* = 5.1 Hz), 124.76 (q, *J* = 273.4 Hz), 103.5 (C₁), 75.2 (C₄), 74.9 (C₃), 74.1 (C₂), 70.85 (OCH₂), 68.4 (C₅), 64.6 (C₅), 31.9 (C₆), 29.7, 29.5, 29.3, 26.0, 22.8, 14.2. The carbon α to the boron atom was not observed. ¹⁹F NMR (282 MHz, CDCl₃) δ -59.4. ¹¹B NMR (96 MHz, CDCl₃) δ

28.3. HRMS (ESI+) (M+Na)⁺ calculated for C₂₁H₃₀¹¹BF₃O₆Na 469.1979, found 469.1982.

Author Contributions

We strongly encourage authors to include author contributions and recommend using [CRediT](#) for standardised contribution descriptions. Please refer to our general [author guidelines](#) for more information about authorship.

Conflicts of interest

There are no conflicts to declare.

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Notes and references

‡ Footnotes relating to the main text should appear here. These might include comments relevant not central to the matter under discussion, limited experimental and spectral data, and crystallographic data.

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etc.

- L. Zeng, X. Lin, P. Li, F.-Q. Liu, H. Guo and W.-H. Li, *Prog. Org. Coat.*, 2021, **159**, 106417.
- M. George and R. G. Weiss, *Acc. Chem. Res.*, 2006, **39**, 489-497.
- D. J. Abdallah and R. G. Weiss, *Adv. Mater.*, 2000, **12**, 1237-1247.
- P. R. A. Chivers and D. K. Smith, *Nat. Rev. Mater.*, 2019, **4**, 463-478.
- E. R. Draper and D. J. Adams, *Chem*, 2017, **3**, 390-410.
- S. S. Babu, V. K. Praveen and A. Ajayaghosh, *Chem. Rev.*, 2014, **114**, 1973-2129.
- D. Rosa Nunes, M. Raynal, B. Isare, P.-A. Albouy and L. Bouteiller, *Soft Matter*, 2018, **14**, 4805-4809.
- D. Rosa Nunes, M. Reche-Tamayo, E. Ressouche, M. Raynal, B. Isare, P. Foury-Leylekian, P.-A. Albouy, P. Brocorens, R. Lazzaroni and L. Bouteiller, *Langmuir*, 2019, **35**, 7970-7977.
- M. Raynal and L. Bouteiller, *Chemical Communications*, 2011, **47**, 8271-8273.
- Y. Lan, M. G. Corradini, R. G. Weiss, S. R. Raghavan and M. A. Rogers, *Chemical Society Reviews*, 2015, **44**, 6035-6058.
- J. Morris, J. Bietsch, K. Bashaw and G. Wang, *Gels*, 2021, **7**, 24.
- A. D. Ludwig, F. Berrée and L. Lemiègre, in *Carbohydrate Chemistry*, The Royal Society of Chemistry, 2022, vol. 45, pp. 379-415.
- A. Prathap and K. M. Sureshan, *Langmuir*, 2019, **35**, 6005-6014.
- T. Ishi-i and S. Shinkai, *Top. Curr. Chem.*, 2005, **258**, 119-160.
- Y. Kubo, W. Yoshizumi and T. Minami, *Chem. Lett.*, 2008, **37**, 1238-1239.

- 16 F. Xu, L. Pfeifer, S. Crespi, F. K.-C. Leung, M. C. A. Stuart, S. J. Wezenberg and B. L. Feringa, *J. Am. Chem. Soc.*, 2021, **143**, 5990-5997.
- 17 E. Suarez-Picado, M. Coste, J.-Y. Runser, M. Fossepre, A. Carvalho, M. Surin, L. Jierry and S. Ulrich, *Biomacromolecules*, 2022, **23**, 431-442.
- 18 S. J. Ren, H. Q. Liang, P. P. Sun, Y. N. Gao and L. Q. Zheng, *New J. Chem.*, 2020, **44**, 1609-1614.
- 19 A. Nuthanakanti, *New J. Chem.*, 2019, **43**, 13447-13456.
- 20 A. Nuthanakanti and S. G. Srivatsan, *ACS Appl. Mater. Interfaces*, 2017, **9**, 22864-22874.
- 21 J. A. Foster, R. M. Edkins, G. J. Cameron, N. Colgin, K. Fucke, S. Ridgeway, A. G. Crawford, T. B. Marder, A. Beeby, S. L. Cobb and J. W. Steed, *Chem. – Eur. J.*, 2014, **20**, 279-291.
- 22 C. D. Jones and J. W. Steed, *Chem. Soc. Rev.*, 2016, **45**, 6546-6596.
- 23 E. R. Draper and D. J. Adams, *Chem. Commun.*, 2016, **52**, 8196-8206.
- 24 A. D. Ludwig, A. Saint-Jalmes, C. Mériadec, F. Artzner, O. Tasseau, F. Berrée and L. Lemiègre, *Chem. – Eur. J.*, 2020, **26**, 13927-13934.
- 25 A. M. Vibhute, V. Muvvala and K. M. Sureshan, *Angew. Chem., Int. Ed.*, 2016, **55**, 7782-7785.
- 26 F. Ono, O. Hirata, K. Ichimaru, K. Saruhashi, H. Watanabe and S. Shinkai, *Eur. J. Org. Chem.*, 2015, 6439-6447.
- 27 J. Morris, P. Kozłowski and G. Wang, *Langmuir*, 2019, **35**, 14639-14650.
- 28 A. Chen, L. P. Samankumara, C. Garcia, K. Bashaw and G. Wang, *New J. Chem.*, 2019, **43**, 7950-7961.
- 29 A. Chen, I. S. Okafor, C. Garcia and G. Wang, *Carbohydr. Res.*, 2018, **461**, 60-75.
- 30 G. Wang, A. Chen, H. P. R. Mangunuru and J. R. Yerabolu, *RSC Adv.*, 2017, **7**, 40887-40895.
- 31 J. Kowalczyk, M. Bielejewski, A. Lapinski, R. Luboradzki and J. Tritt-Goc, *J. Phys. Chem. B*, 2014, **118**, 4005-4015.
- 32 H. Xu, J. Song, T. Tian and R. Feng, *Soft Matter*, 2012, **8**, 3478-3486.
- 33 J. Tritt-Goc and J. Kowalczyk, *Langmuir*, 2012, **28**, 14039-14044.
- 34 K. Sakurai, Y. Jeong, K. Koumoto, A. Friggeri, O. Gronwald, S. Sakurai, S. Okamoto, K. Inoue and S. Shinkai, *Langmuir*, 2003, **19**, 8211-8217.
- 35 O. Gronwald and S. Shinkai, *Chem. – Eur. J.*, 2001, **7**, 4329-4334.
- 36 N. Kameta, M. Masuda and T. Shimizu, *Chem. Commun.*, 2015, **51**, 11104-11107.
- 37 N. Kameta, K. Ishikawa, M. Masuda, M. Asakawa and T. Shimizu, *Chem. Mater.*, 2012, **24**, 209-214.
- 38 N. Shimada, K. Fukuhara, S. Urata and K. Makino, *Org. Biomol. Chem.*, 2019, **17**, 7325-7329.
- 39 N. Shimada, S. Urata, K. Fukuhara, T. Tsuneda and K. Makino, *Org. Lett.*, 2018, **20**, 6064-6068.
- 40 R. S. Mancini, J. B. Lee and M. S. Taylor, *J. Org. Chem.*, 2017, **82**, 8777-8791.
- 41 S. Uzan, D. Barış, M. Çolak, H. Aydın and H. Hoşgören, *Tetrahedron*, 2016, **72**, 7517-7525.
- 42 H. Li, H. Li, Q. Dai, H. Li and J.-L. Brédas, *Adv. Theory Simul.*, 2018, **1**, 1700015.
- 43 J.-D. Chai and M. Head-Gordon, *Phys. Chem. Chem. Phys.*, 2008, **10**, 6615-6620.
- 44 D. Heß and P. Klüfers, *Carbohydr. Res.*, 2011, **346**, 1752-1759.
- 45 M. Tanaka, A. Nakagawa, N. Nishi, K. Iijima, R. Sawa, D. Takahashi and K. Toshima, *J. Am. Chem. Soc.*, 2018, **140**, 3644-3651.
- 46 B. Wrackmeyer, *Prog. Nucl. Magn. Reson. Spectrosc.*, 1979, **12**, 227-259.
- 47 M. J. Frisch, G. W. Trucks, H. B. Schlegel, G. E. Scuseria, M. A. Robb, J. R. Cheeseman, G. Scalmani, V. Barone, G. A. Petersson, H. Nakatsuji, X. Li, M. Caricato, A. Marenich, J. Bloino, B. G. Janesko, R. Gomperts, B. Mennucci, H. P. Hratchian, J. V. Ortiz, A. F. Izmaylov, J. L. Sonnenberg, D. Williams-Young, F. Ding, F. Lipparini, F. Egidi, J. Goings, B. Peng, A. Petrone, T. Henderson, D. Ranasinghe, V. G. Zakrzewski, J. Gao, N. Rega, G. Zheng, W. Liang, M. Hada, M. Ehara, K. Toyota, R. Fukuda, J. Hasegawa, M. Ishida, T. Nakajima, Y. Honda, O. Kitao, H. Nakai, T. Vreven, K. Throssell, J. J. A. Montgomery, J. E. Peralta, F. Ogliaro, M. Bearpark, J. J. Heyd, E. Brothers, K. N. Kudin, V. N. Staroverov, T. Keith, R. Kobayashi, J. Normand, K. Raghavachari, A. Rendell, J. C. Burant, S. S. Iyengar, J. Tomasi, M. Cossi, J. M. Millam, M. Klene, C. Adamo, R. Cammi, J. W. Ochterski, R. L. Martin, K. Morokuma, O. Farkas, J. B. Foresman and D. J. Fox, *Gaussian, Inc., Wallingford CT*, 2016.
- 48 K. J. Donald, U. R. Gaillard and N. Walker, *J. Phys. Chem. A*, 2022, **126**, 5173-5185.
- 49 A. D. Becke, *J. Chem. Phys.*, 1993, **98**, 5648-5652.
- 50 R. Dennington, T. A. Keith and J. M. Millam, *Semichem Inc., Shawnee Mission, KS*, 2016.