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Clarisse Dumont[1], Guangping Li[1], Julien Castel[1], Serge Luquet[1], Giuseppe Gangarossa[1,*]

[1] Université Paris Cité, CNRS, Unité de Biologie Fonctionnelle et Adaptative, F-75013 Paris, France

*Correspondence to Giuseppe Gangarossa (giuseppe.gangarossa@u-paris.fr, @PeppeGanga)

Running title: Hindbrain→thalamus paths scale feeding and homeostasis

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Key points

1. The paraventricular thalamus (PVT) is known to receive projections from the hindbrain. Here, we confirm and further extend current knowledge on the existence of hindbrainTH→PVT catecholaminergic (CA) inputs, notably from the locus coeruleus (LC) and the nucleus tractus solitarius (NTS), with the NTS representing the main source.

2. Disruption of hindbrainTH→PVT inputs contribute to the modulation of PVT-neurons activity.

3. HindbrainTH→PVT inputs scale feeding strategies in environmental, behavioral, physiological and metabolic stress-like contexts.

4. HindbrainTH→PVT inputs participate in regulating metabolic efficiency and nutrient partitioning in stress-like contexts.

5. HindbrainTH→PVT, directly and/or indirectly, contribute in modulating the downstream activity of lateral (LH) and dorsomedial (DMH) hypothalamic neurons.

Abstract

The regulation of food intake and energy balance relies on the dynamic integration of exteroceptive and interoceptive signals monitoring nutritional, metabolic, cognitive and emotional states. The paraventricular thalamus (PVT) is a central hub that, by integrating sensory, metabolic and emotional states, may contribute to the regulation of feeding and homeostatic/allostatic processes. However, the underlying PVT circuits remain still elusive. Here, we aimed at unraveling the role of catecholaminergic (CA) inputs to the PVT in scaling feeding and metabolic efficiency. First, using region-specific retrograde disruption of CA projections, we show that PVT CA inputs mainly arise from the hindbrain, notably the locus coeruleus (LC) and the nucleus tractus solitarius (NTS). Second, taking advantage of integrative calorimetric measurements of metabolic efficiency, we reveal that CA inputs to the PVT scale adaptive feeding and metabolic responses in environmental, behavioral, physiological and metabolic stress-like contexts. Third, we show that hindbrainTH→PVT inputs contribute in modulating the activity of PVT as well as lateral (LH) and dorsomedial (DMH) hypothalamic neurons.

In conclusion, this study, by assessing the key role of CA inputs to the PVT in scaling homeostatic/allostatic regulations of feeding patterns, reveals the integrative and converging hindbrainTH→PVT paths that contribute to whole-body metabolic adaptations in stress-like contexts.

Introduction

In mammals, the regulation of food intake intricately relies on the orchestration of several signals mirroring the dynamic integration of interoceptive and exteroceptive (environment) states (Sweeney & Yang, 2017). Indeed, emotional states (stress, anxiety, motivation), by modulating the activity of central neuronal hubs, thoroughly scale the regulation of food intake and the establishment of feeding habits (Ulrich-Lai *et al.*, 2015). This is particularly evident in stress-related contexts where emotional states compete with the homeostatic regulation of feeding (Maniam & Morris, 2012; Herzog, 2020), thereby leading to feed-forward allostatic adaptations (*stability through changes*) which may culminate in psychiatric and metabolic disorders. These observations support the idea that emotional and homeostatic states may share similar, although not identical, neuronal networks (Sweeney & Yang, 2017). Nonetheless, the systems underlying the functional connection between these states remain poorly understood.

Emerging evidence strongly suggests that the paraventricular nucleus of the thalamus (PVT), a dorsal midline thalamic relay, may represent a functional *hot-spot* where interoceptive and exteroceptive stimuli may converge to orchestrate the selection of adaptive and appropriate behavioral responses aimed at regulating homeostatic and cognitive processes (Penzo & Gao, 2021). Given their elaborated connectivity (Kirouac, 2015), PVT excitatory (glutamate) neurons are well positioned to serve as functional integrators of orexigenic and anorexigenic stimuli (Ong *et al.*, 2017; Meffre *et al.*, 2019), glucose fluctuations (Labouèbe *et al.*, 2016; Kessler *et al.*, 2021), learning and memory processes (Do-Monte *et al.*, 2015; Penzo *et al.*, 2015) as well as emotional states (Heydendael *et al.*, 2011; Barson *et al.*, 2020; Pliota *et al.*, 2020). This plethora of PVT-related brain functions highly relies on different afferent projections which, by carrying distinct neurochemical information, synergistically scale the activity of PVT-neurons and their downstream connected regions. Among the different afferents, the PVT also harbors a dense plexus of catecholaminergic (CA) fibers mainly arising from the hindbrain (Schroeter *et al.*, 2000; Beas *et al.*, 2018; Sofia Beas *et al.*, 2020; Wang *et al.*, 2021*b*) and only few scattered CA fibers from the hypothalamus (Li *et al.*, 2014). In addition, recent reports have indicated that brain CA (norepinephrine, NE, and/or dopamine, DA), by modulating different homeostatic functions (*i.e.* wakefulness, arousal, glucoprivation-

induced food seeking), may represent key neuromodulators of the PVT (Beas *et al.*, 2018; Sofia Beas *et al.*, 2020; Wang *et al.*, 2021*b*). Moreover, brain CA are also important contributors to the regulation of stress-like responses (Valentino & Van Bockstaele, 2008; Kvetnansky *et al.*, 2009) which ultimately impact, directly and/or indirectly, on feeding patterns and behaviors (Xu *et al.*, 2019; Qu *et al.*, 2020).

Indeed, the PVT has already been involved in food-seeking behaviors mostly associated to positive (motivation, reward, reinforcement) or negative valance (Labouèbe *et al.*, 2016; Otis *et al.*, 2017, 2019; Do-Monte *et al.*, 2017; Wang *et al.*, 2021*a*; Engelke *et al.*, 2021) as well as in stress and emotional arousal (Hsu *et al.*, 2014). However, whether and how the PVT and its afferent CA inputs may contribute to the regulation of food intake and metabolic efficiency in stress-related contexts remain to be fully established.

In order to dissect the contribution of PVT CA inputs in the regulation of food intake, we suppressed local CA inputs by microinjecting the neurotoxin 6-OHDA into the PVT and performed several experiments aimed at revealing the adaptive strategies associated to the regulation of feeding and energy balance. Here, we demonstrate that CA inputs to the PVT exerted a modulatory role on food intake and metabolic efficiency specifically in stress-related contexts since no major alterations were detected in basal conditions. Notably, we found that 6-OHDA^{PVT}-lesioned mice, following both exteroceptive (environmental) and interoceptive (body energy) stressors, showed enhanced food intake and metabolic efficiency.

Altogether, our results reveal a novel neuronal network by which stressors impinge on the regulatory allostatic processes underlying feeding behaviors, metabolic efficiency and nutrient partitioning to scale stress-associated adaptive responses.

Materials and methods

Ethics statement and animals

All experiments were approved by the Animal Care Committee of the Université Paris Cité (APAFiS #35585 and #11003) and carried out following the 2010/63/EU directive. 8-14 weeks old male C57Bl/6J mice (Janvier, Le Genest St Isle, France) were used and housed in a room maintained at 22 +/-1 °C, with a light period from 7h00 to 19h00. Regular chow diet (3.24 kcal/g, reference SAFE® A04, Augy, France) and water were provided *ad libitum* unless otherwise stated.

Stereotaxic microinjections for viral tracing studies and 6-OHDA-induced catecholaminergic denervation

Mice were anaesthetized with isoflurane (3.5% for induction, 1.5% for maintenance), administered with Buprécare® (buprenorphine 0.3 mg) and Ketofen® (ketoprofen 100 mg), and placed on a stereotactic frame (Model 940, David Kopf Instruments). During surgery, body temperature was maintained at 37°C.

6-OHDA-HCl (Sigma-Aldrich, #H4381) was dissolved in a saline solution (NaCl 0.9% w/v) containing 0.02% of ascorbic acid at a final concentration of 6 µg/µl. Animals were randomly assigned to either 6-OHDA or vehicle microinjections. A volume of 0.35 µl of 6-OHDA or vehicle (0.02% ascorbic acid) was injected into the PVT (L= 0.0; AP= -1.46; V= -2.8, mm) at an infusion rate of 0.05 µl/min. The injection needle was carefully removed after waiting 5 minutes at the injection site. 24-hrs after, animals were re-administered with Buprécare® and Ketofen®, and had facilitated access to jelly food (DietGel Boost #72-04-5022, Clear H_2O) for 2 consecutive days. Recovery from surgery was monitored during 3-5 days post-surgery. Animals were allowed to recover for 3-4 weeks before any experimental evaluation.

pAAV-CAG-tdTomato (titer ≥ 1×10^{13} vg/mL) was a gift from Edward Boyden (Addgene viral prep #59462-AAV9; http://n2t.net/addgene:59462; RRID:Addgene_59462). A volume of 0.20 µl of pAAV-CAG-tdTomato was injected into the PVT (L= 0.0; AP= -1.46; V= -2.8, mm) at an infusion rate of 0.05 µl/min. The injection needle was carefully removed after waiting 5 minutes at the injection site. Viral expression was evaluated 4 weeks after microinjection.

158 **Indirect calorimetry and metabolic efficiency analysis**

159 Indirect calorimetry was performed as previously described (Berland *et al.*, 2022).
160 Mice were monitored for whole energy expenditure (EE), O_2 consumption and CO_2
161 production, respiratory exchange rate (RER=VCO_2/VO_2), fatty acid oxidation (FAO),
162 and locomotor activity using calorimetric cages with bedding, food and water
163 (Labmaster, TSE Systems GmbH, Bad Homburg, Germany). Ratio of gases was
164 determined through an indirect open circuit calorimeter (Arch *et al.*, 2006; Even &
165 Nadkarni, 2012). This system monitors O_2 and CO_2 concentration by volume at the
166 inlet ports of a tide cage through which a known flow of air is being ventilated (0.4
167 L/min) and compared regularly to a reference empty cage. For optimal analysis, the
168 flow rate was adjusted according to the animal body weights to set the differential in
169 the composition of the expired gases between 0.4-0.9% (Labmaster, TSE Systems
170 GmbH, Bad Homburg, Germany). The flow was previously calibrated with O_2 and
171 CO_2 mixture of known concentrations (Air Liquide, S.A. France). O_2 consumption and
172 CO_2 production were recorded every 15 min for each animal during the entire
173 experiment. Whole energy expenditure (EE) was calculated using the Weir equation
174 for respiratory gas exchange measurements. Food consumption was measured as
175 the instrument combines a set of highly sensitive feeding sensors for automated
176 online measurements. Mice had access to food and water *ad libitum*. To allow
177 measurement of every ambulatory movement, each cage was embedded in a frame
178 with an infrared light beam-based activity monitoring system with online
179 measurement at 100 Hz. The sensors for gases and detection of movements
180 operated efficiently in both light and dark phases, allowing continuous recording.
181 When required, the inversion of circadian light/dark cycles was programmed using
182 the Labmaster software.
183 Body mass composition was analyzed using an Echo Medical systems' EchoMRI
184 (Whole Body Composition Analyzers, EchoMRI, Houston, USA), according to
185 manufacturer's instructions. Readings of body composition were given within 1 min.
186 Data analysis was performed on Excel XP using extracted raw values of VO_2
187 consumed (expressed in ml/h), VCO_2 production (expressed in ml/h), and energy
188 expenditure (kcal/h).
189

190 **Novelty-suppressed feeding (NSF)**

After an overnight fasting, mice were placed in a cage (40×40×40 cm) with a single regular chow pellet in the center. The latency (time in seconds) to eat was scored. To measure food intake, food consumption was evaluated 60 minutes after the beginning of the test.

Open field (OF)

Spontaneous exploratory behavior was monitored in an open field (40×40×40 cm, BIOSEB) for 20 min, video-tracked and analyzed using the SMART software (BIOSEB). The open field was wiped with 70% ethanol between sessions.

Infrared temperature measurements

Thermogenesis was visualized using a high-resolution infrared camera (FLIR E8; FLIR Systems, Portland, OR, USA). To measure the temperature (°C) of the brown adipose tissue (BAT, interscapular regions), lower back and tail, images were captured before and after the open field (OF) test. Infrared thermography images were analyzed using the FLIR TOOLS software.

Food intake following food or water deprivation

In two distinct experiments we measured food intake following either an overnight fasting (food deprivation) or water deprivation.

Overnight fasting. Mice were first weighted in the morning following an overnight fasting to ensure the loss of body weight. Then, they were exposed to pre-weighted chow pellets. Food intake was measured at the following time points: 30 min, 1h, 2h, 3h, 4h.

Overnight water deprivation. Mice were first weighted in the morning following an overnight water deprivation to ensure the loss of body weight. Then, they were exposed to a calibrated drinking bottle and pre-weighted chow pellets. Water consumption and food intake were measured at the following time points: 30 min, 1h, 2h, 3h, 4h.

Food intake induced by 2-DG and ghrelin

Mice were handled and injected with vehicle during 3 consecutive days before drugs administration. After this habituation period, they were administered with ghrelin (#1465, Tocris, 0.5 mg/kg, i.p.) or 2-DG (#14325, Cayman Chemical, 500 mg/kg, i.p.)

and exposed to chow pellets 30 min after the injections. Food intake was measured during 3 hours. For 2-DG treated mice, blood glucose (counterregulatory response) was measured from the vein tail using a glucometer (Menarini Diagnotics, Rungis, France) at 0 and 30 min.

Glucose dynamics

Oral glucose tolerance test (OGTT). Animals were fasted 5 hours before receiving a bolus of glucose solution (2 g/kg) by gavage. Blood glucose (hyperglycemia) was measured from the vein tail using a glucometer (Menarini Diagnotics, Rungis, France) at 0, 5, 10, 15, 30, 60, 90, and 120 min.

Insulin tolerance test (ITT). Animals were fasted 5 hours before receiving an injection of insulin (0.5 U/kg, Novo Nordisk, i.p.). Blood glucose (hypoglycemia) was measured from the vein tail at 0, 5, 10, 15, 20, 30, 60, 90, and 120 min.

Tissue preparation and immunofluorescence

Animals were injected with pentobarbital (500 mg/kg, i.p., Sanofi-Aventis, France). Once anaesthetized, they were transcardially perfused with cold (4°C) PFA 4% for 5 minutes. Brains were collected, put overnight in PFA 4% and then stored in PBS (4°C). 40 µm-thick sections were sliced with a vibratome (Leica VT1000S, France) and stored in a cryoprotective solution at -20 °C until immunofluorescence investigations. Immunofluorescence on brain slices was performed as previously described (Gangarossa *et al.*, 2019; Berland *et al.*, 2020).

Briefly, sections were processed as it follows. Day 1: free-floating sections were rinsed in Tris-buffered saline (TBS; 0.25 M Tris and 0.5 M NaCl, pH 7.5), incubated for 5 min in TBS containing 3% H_2O_2 and 10% methanol, and then rinsed three times for 10 min each in TBS. After 15 min incubation in 0.2% Triton X-100 in TBS, sections were rinsed three times in TBS again. Slices were then incubated 48 hrs at 4°C with the following primary antibodies: rabbit anti-cFos (1:1000, Synaptic Systems, #226 003), rabbit anti-TH (1:1000, Millipore, #AB153) or rat anti-DAT (1:500, Millipore, #MAB369). Sections were rinsed three times for 10 min in TBS and incubated for 60 min with secondary donkey anti-rabbit Cy3 AffiniPure (1:1000, Jackson ImmunoResearch, #711-165-152) or donkey anti-rat Cy3 AffiniPure (1:1000, Jackson ImmunoResearch, #712-165-153). Sections were rinsed for 10 min twice in TBS, stained with DAPI (10 min) and rinsed in TB (0.25 M Tris) before mounting.

Acquisitions were performed with a confocal microscope (Zeiss LSM 510). The objective (10X) and the pinhole setting remained unchanged during the acquisition of a series for all images. Depending on the extension of the region of interest, either single or mosaic acquisitions were conducted. Quantification of immunoreactive cells (cFos- or TH-positive neurons) was performed using the cell counter plugin of the ImageJ software taking as standard reference a fixed threshold of fluorescence. For cell counting, three (TH) or two (cFos) rostro-caudal levels for each brain region were used. Quantifications of immunopositive neurons were averaged between hemispheres and then summed for consecutive brain slices.

Statistics

All data are presented as mean ± SD. Statistical tests were performed with Prism 7 (GraphPad Software, La Jolla, CA, USA). Detailed statistical analyses are listed in the **Statistical Summary Table**. Normality was assessed by the D'Agostino-Pearson test. Depending on the experimental design, data were analyzed using either Student t-test (paired or unpaired) with equal variances, One-way ANOVA or Two-way ANOVA. The significance threshold was automatically set at $p<0.05$. ANOVA analyses were followed by Bonferroni *post hoc* test for specific comparisons only when overall ANOVA revealed a significant difference (at least $p<0.05$).

Results

Catecholaminergic inputs modulate the activity of PVT-neurons

To study whether catecholaminergic (TH-positive) inputs to the PVT participate/contribute to the regulation of food intake, we decided to ablate catecholamine (CA) fibers projecting to the PVT by locally microinjecting 6-OHDA (**Figure 1A**), a neurotoxin reuptaken by DAT- and/or NET-expressing terminals. Since the PVT extends throughout the midline thalamic axis, we decided to mainly focus on the mid-posterior PVT as this region has been shown to be involved in stress and stress-induced hypophagia (Heydendael *et al.*, 2011; Barson *et al.*, 2020; Barrett *et al.*, 2021). Indeed, 6-OHDA was able to strongly reduce TH immunostaining in the PVT (**Figure 1B**), with few remaining terminals most likely corresponding to NET-negative TH fibers (terminals releasing adrenaline and/or devoid of monoamine transporters) since the PVT does not seem to contain DAT-positive terminals as compared to DAT-rich regions such as the tail of the striatum [TS, (Gangarossa *et al.*, 2013; Valjent & Gangarossa, 2021)] and the central amygdala (**Suppl. Figure 1A**, https://doi.org/10.6084/m9.figshare.19683228.v1).

We therefore examined the regional sources of our 6-OHDA-induced degeneration of TH-afferents. We focused on putative noradrenergic TH-positive projecting neurons since the PVT do not receive dopaminergic inputs from the midbrain (SNc and VTA) (Li *et al.*, 2014) and it harbors only minor, if any, scattered DAT-fibers (García-Cabezas *et al.*, 2009; Clark *et al.*, 2017). Immunofluorescence analysis revealed a significant reduction of TH-neurons in the nucleus tractus solitarius (NTS) and the locus coeruleus (LC), in line with the presence of *Slc6a2* (NET)-cathecolaminergic neurons in these nuclei (Schroeter *et al.*, 2000; Zhang *et al.*, 2021) and the sensitivity of these neurons to 6-OHDA (Szot *et al.*, 2012*b*; Lin *et al.*, 2013). However, we observed a more robust reduction of PVT-projecting TH-neurons in the NTS (-34.2%) compared to the LC (-15.1%) (**Figure 1C, D**). No differences were observed in the A1 area of the hindbrain (**Figure 1C, D**) as well as in the hypothalamus (**Suppl. Figure 1B**, https://doi.org/10.6084/m9.figshare.19683228.v1).

Then we investigated whether the reduction in local TH-afferents was followed by the modulation of PVT-neurons activity. Since the PVT shows higher activity during wakefulness (Ren *et al.*, 2018), mice were perfused 1h before the onset of the dark phase (spontaneous feeding period) and at basal conditions. Using cFos as a

molecular proxy of cellular activity, we observed a significant increase in cFos-positive cells in the PVT of 6-OHDA^{PVT} compared to Sham^{PVT} mice (**Figure 1E, F**). This set of results suggests that a subset of hindbrain CA inputs (hindbrainTH→PVT projections) may serve as modulators of PVT-neurons activity.

Catecholaminergic inputs to the PVT contribute to novelty-induced hypophagia

Following 3-4 weeks from 6-OHDA microinjections, no major differences in body weight were detected in 6-OHDA^{PVT} compared to Sham^{PVT} mice (group-housed animals, **Figure 2A**). In order to study the role of PVT catecholaminergic inputs in the regulation of feeding patterns, 6-OHDA^{PVT} and Sham^{PVT} mice were single-housed (**Figure 2B**). Environmental and social isolation, as triggered by single housing, represent behavioral/environmental stress-like allostatic stimuli (Lee *et al.*, 2020, 2021) which can trigger a transient reduction of food intake (Takatsu-Coleman *et al.*, 2013; Benfato *et al.*, 2022). Interestingly, we observed that 6-OHDA^{PVT} mice were less sensitive to single housing-induced hypophagia compared to Sham^{PVT} mice (**Figure 2C**). This phenotype prompted us to investigate whether catecholaminergic inputs to the PVT were important in driving feeding patterns and metabolic adaptations to exposure to other behavioral/environmental challenges.

After 2 weeks of habituation to single-housing, we analyzed the metabolic efficiency of 6-OHDA^{PVT} and Sham^{PVT} mice exposed to a novel environment during two consecutive exposures during the dark period (spontaneous feeding period, **Figure 2D**). While Sham^{PVT} mice transiently (Exp.1) showed a reduction in food intake during the dark period (**Figure 2E, E^{1}**), 6-OHDA^{PVT} mice were again less sensitive to novelty-induced hypophagia. No differences in food intake were measured during Exp.2 period between the two groups, indicating a rapid restoration of homeostatic regulations associated to environmental habitation (**Figure 2E, E^{1}**). This phenotype was not associated to changes in locomotor activity as indicated by the similar patterns of exploration (Exp.1, novelty) and habituation (Exp.2) (**Figure 2F, F^{1}**). We also measured key whole-body metabolic parameters such as the respiratory exchange ratio (RER, indicative of the energy substrates used, RER=~1 for carbohydrates, RER=~0.7 for lipids), fatty acid oxidation (FAO) and energy expenditure (EE) during the exposure to the novel environment (Exp.1). Compared to Sham^{PVT} mice, 6-OHDA^{PVT} animals showed increased RER (**Figure 2G, G^{1}**) and decreased FAO (**Figure 2H, H^{1}**), mirroring the changes in food intake and indicating

a shift of energy substrates (carbohydrates and lipids) use during exposure to a novel environment. However, these adaptations did not impact on energy expenditure (**Figure 2I, I[1]**), suggesting that nutrient partitioning (Joly-Amado *et al.*, 2012), rather than total energy balance, was affected by the loss of hindbrainTH→PVT fibers.

One may wonder whether the absence of novelty-induced hypophagia in 6-OHDA^{PVT} mice may be associated to enhanced perception and/or reward value of food. To investigate this aspect, Sham^{PVT} and 6-OHDA^{PVT} mice were intermittently (1h/day) exposed to high-fat high-sugar (HFHS) diet during two consecutive days. No differences in HFHS food intake were observed between groups (**Suppl. Figure 2A**, https://doi.org/10.6084/m9.figshare.19683228.v1), suggesting intact food palatability, perception and preference. These results suggest that hindbrainTH→PVT fibers represent an important node for the integration of homeostatic regulations.

Exposure to novel environments can lead to the occurrence of anxiogenic traits and novelty-triggered thermogenic adaptations (Lecorps *et al.*, 2016) which may impact on, and therefore confound, the mechanisms underlying feeding strategies. Thus, we performed an open field test (OF, **Figure 3A**) to evaluate both anxiety and thermogenic adaptations. We observed no significant differences in anxiety-like parameters between Sham^{PVT} and 6-OHDA^{PVT} animals (**Figure 3B-F**). Moreover, both groups showed similar thermogenic enhancements in the brown adipose tissue (BAT), the lower back and the tail (**Figure 3G-J**). These results indicate that PVT-projecting TH-afferents modulate feeding patterns and metabolic efficiency independently from affective (anxiety) and thermogenic adaptations.

Feeding patterns and metabolic efficiency highly depend on circadian rhythms and strong functional interactions exist between circadian rhythms, feeding and energy balance (Challet, 2019). Moreover, the PVT, which is bidirectionally connected with the suprachiasmatic nucleus (SCN) (Peng & Bentivoglio, 2004; Colavito *et al.*, 2015), has been pointed as a contributor to circadian cycles and its activity varies depending on active/inactive phases (Colavito *et al.*, 2015; Kirouac, 2015). Thus, we decided to investigate whether PVT catecholaminergic inputs participated to the adaptive metabolic strategies occurring during circadian challenges by inverting the light/dark cycle (**Figure 4A**). First, before inverting the light/dark cycle, we confirmed (**Figure 2**) that exposure to the new environment was associated to reduced novelty-induced hypophagia in 6-OHDA^{PVT} compared to Sham^{PVT} mice (**Suppl. Figure 3A, B**, https://doi.org/10.6084/m9.figshare.19683228.v1). Then, after habituation, light/dark

cycles were inverted. When measuring food intake, we observed that both experimental groups rapidly shifted and adapted toward the new light/dark schedule (average of first 2 days of standard and inverted cycles) (**Figure 4B, B^{1}**). In fact, despite the adaptive shift (**Figure 4B, B^{1}**), no differences were observed in the cumulative food intake (**Figure 4C**). Moreover, we also detected a similar pattern of circadian adaptation in the EE profile of both groups (**Figure 4D, D^{1}**), with an increased EE in the 7h-19h inverted period (iDark) and a decreased EE in the 19h-7h inverted period (iLight) (**Figure 4E, E^{1}**). Interestingly, we found a significant difference in FAO and RER. In particular, while Sham^{PVT} animals rapidly adapted to the inverted light/dark cycle (**Figure 4F**), 6-OHDA^{PVT} mice did not show significant changes in FAO during the 7h-19h inverted period (iDark) (**Figure 4F^{1}, G**), whereas both groups showed increased FAO during the 19h-7h inverted period (iLight) (**Figure 4G^{1}**). In line with this shift in energy substrates partitioning, measurements of RER indicated an impaired adjustment of metabolic efficiency during the iDark period (7h-19h) in 6-OHDA^{PVT} mice (**Figure 4H, H^{1}, I, I^{1}**).

These results suggest that hindbrainTH→PVT inputs, although not involved in the adaptation of food intake to circadian challenges, are important in adjusting peripheral energy substrates utilization (*i.e* lipids and carbohydrates as revealed by RER and FAO) and overall metabolic efficiency.

Catecholaminergic inputs to the PVT contribute to feeding under physiological and metabolic challenges

Next, we wondered whether hindbrainTH→PVT inputs guided feeding following physiological and metabolic stressors. First, to mimic a conflict between hunger and environmental stress, we decided to study food intake in overnight fasted mice in a novelty-induced hypophagia test. After an overnight fasting, both grouped showed a similar reduction in body weight and plasma glucose levels (**Suppl. Figure 4A, B**, https://doi.org/10.6084/m9.figshare.19683228.v1). Then mice underwent the novelty-suppressed feeding (NSF) test (**Figure 5A**). Sham^{PVT} and 6-OHDA^{PVT} mice showed similar latency to eat (**Figure 5B**), with a significant difference in food intake which was higher in 6-OHDA^{PVT} mice (**Figure 5C, D**). The NSF test (latency to eat) is mainly used to assess anxiety- and depressive-like phenotypes. Therefore, our results suggest that the increased food intake observed following PVT catecholaminergic ablation may not be confounded by potential alterations triggered

by anxiety. This is in line with our above-mentioned observations using the open field test (**Figure 3A-F**).

Then, we used an acute restraint (immobilization) paradigm which is known to alter metabolism and food intake (Rybkin *et al.*, 1997; Vallès *et al.*, 2000; Rabasa & Dickson, 2016). Sham^{PVT} and 6-OHDA^{PVT} mice underwent a 30 min acute restraint and food intake was measured during the dark period. Although both groups showed a significant reduction in food intake (**Figure 5E**), stress-induced hypophagia was significantly more pronounced in Sham^{PVT} compared to 6-OHDA^{PVT} mice (**Figure 5E**).

To further investigate the role of hindbrainTH→PVT inputs in scaling feeding, we used metabolic stressors to modulate food intake. First, Sham^{PVT} and 6-OHDA^{PVT} mice were administered with 2-deoxy-d-glucose (2-DG) which, in virtue of its glucoprivic effects (neuroglucopenia), elicits food consumption as well as the typical glucose counterregulatory response (CRR) (Pénicaud *et al.*, 1986; Lewis *et al.*, 2006). Moreover, PVT-neurons are highly sensitive to glucoprivation (Labouèbe *et al.*, 2016). In this conditions, 6-OHDA^{PVT} mice consumed more chow food than Sham^{PVT} mice (**Figure 6A**), even though the magnitude of the glucose excursion as counterregulatory response was similar between groups (**Figure 6B**). This result suggests that hindbrainTH→PVT projections are required to fully express feeding adaptions to glucoprivic conditions but are dispensable for the autonomic control of glycogen breakdown and glucose production in CRR. In the same line, glucose clearance dynamics during an oral glucose tolerance test (OGTT, **Figure 6C**) or an insulin tolerance test (ITT, **Figure 6D**) were similar between sham and 6-OHDA^{PVT} mice, indicating that glucose metabolism and insulin sensitivity remained unaltered following the loss of hindbrainTH→PVT inputs.

Second, we performed a fasting-refeeding test to mimic a negative energy balance (food deprivation). In line with the NSF test (**Figure 5C, D**), after an overnight fasting and a similar loss of body weight (**Suppl. Figure 4C**, https://doi.org/10.6084/m9.figshare.19683228.v1), both experimental groups showed an enhanced food intake with 6-OHDA^{PVT} mice consuming more food than Sham^{PVT} mice (**Figure 6E**). Third, we also decided to measure drinking and food intake in overnight water-deprived animals. Both groups showed again a similar decrease in body weight (**Suppl. Figure 4D**, https://doi.org/10.6084/m9.figshare.19683228.v1). After deprivation, mice were exposed to water. While no differences were observed

in drinking behavior (**Figure 6F**), 6-OHDA^{PVT} mice again consumed more food than Sham^{PVT} mice (**Figure 6G**). These results suggest that hindbrainTH→PVT inputs scale food intake also following physiological and metabolic stressors.

In order to assess whether orexigenic signals without metabolic challenges also required an intact PVT catecholaminergic transmission, we administered ghrelin in fed mice. As shown in **Figure 6H**, ghrelin similarly induced an increase in food intake in both groups, thereby indicating that canonical orexinergic circuits are not altered.

Reduced catecholaminergic transmission in the PVT promotes the activation of hypothalamic regions

The above-mentioned results point to hindbrainTH→PVT afferents as major actors in scaling food intake and metabolic efficiency. Since these homeostatic functions highly depend on the hypothalamus, classically described as the master regulator of energy balance (Dietrich & Horvath, 2013; Timper & Brüning, 2017), we decided to study whether 6-OHDA^{PVT} mice were characterized by an altered basal activity (cFos-positive cells) of key hypothalamic regions such as the dorsomedial hypothalamus (DMH), the ventromedial hypothalamus (VMH), the lateral hypothalamus (LH) and the arcuate nucleus (Arc). As for **Figure 1**, mice were perfused 1h before the onset of the dark phase. Interestingly, in 6-OHDA^{PVT} mice we observed an increase in cFos-positive cells specifically in the DMH and LH (**Figure 7A-C**) compared to Sham^{PVT} mice, whereas no differences were detected in the VMH and Arc (**Figure 7A, D, E**).

In order to see whether PVT-neurons may potentially modulate hypothalamic functions by directly projecting to the DMH and LH, we microinjected an AAV9-CAG-TdTomato virus in the PVT (**Figure 7F, G**). As shown in **Figure 7G**, we observed direct PVT→DMH and PVT→LH projections. These results suggest that the increased activity of PVT-neurons following catecholamines depletion (**Figure 1**) may impact, directly (**Figure 7G**) and/or indirectly (polysynaptic circuits), on the regulatory activity of the hypothalamus which may ultimately result in the modulation of food intake and metabolic efficiency.

Discussion

The regulation of food intake represents one of the most complex biological functions. Pivotal for adaptation and survival, the regulatory processes underlying feeding are constantly shaped by signals reflecting/sensing physiological adjustments. In virtue of their heterogeneity (exteroceptive *vs* interoceptive sources), stress-like allostatic stimuli are indeed powerful modulators of food intake, feeding habits and metabolic adaptations.

In this study, we report that hindbrain catecholaminergic (putative noradrenergic) inputs to the PVT play a key role in modulating food intake and metabolic efficiency in stress-related contexts. In fact, permanent ablation of TH-afferents to the PVT resulted in enhanced food intake, adjusted metabolic efficiency and nutrient partitioning whenever environmental, behavioral, physiological and/or metabolic (acute/transient) stressors were introduced as dependent variables of feeding behaviors. In particular, 6-OHDA^{PVT} mice were resistant or less sensitive to environmental and behavioral stress-induced hypophagia and showed enhanced feeding patterns following physiological and metabolic challenges. The different nature of stressors used in this study highlights the highly conserved role of CA inputs to the PVT in readily scaling feeding and metabolic adaptations. Moreover, beyond the impact on feeding and metabolic efficiency, it is important to mention that stressors-elicited homeostatic adaptations such as energy expenditure and thermogenesis did not depend on PVT CA inputs, suggesting a functional tropism of hindbrainTH→PVT circuits toward food intake on one hand and peripheral nutrient partitioning on the other. This is relevant since previous studies have suggested that PVT-neurons, by facilitating hypothalamic-pituitary-adrenal (HPA) responses (Bhatnagar *et al.*, 2000), may contribute to the regulation of core temperature rhythms as well as body weight gain in chronically stressed rats (Bhatnagar & Dallman, 1999). Whether distinct PVT networks (inputs/outputs) are differently engaged by acute *vs* chronic stressors on the regulation of body hemostasis will deserve in-depth investigations. Overall, these results indicate that the PVT, a key node of the limbic circuitry (Barson *et al.*, 2020), contributes to the elaboration of food-related decisions and strategies by integrating, among others, also catecholaminergic information. In addition, our results provide new evidence for the existence of distinct, but converging, hindbrain CA inputs (a subset of NTSTH- and

LCTH-neurons) capable of gating food-related homeostatic adaptations under transient stress-like allostatic stimuli.

Surprisingly, we found that the NTS represented one of the major sources of TH-positive projections to the PVT. In fact, local microinjection of 6-OHDA resulted in a significant reduction of TH-expressing neurons in the NTS and to a lesser extend in the LC, sparing CA neurons in the medulla (A1 area) and the hypothalamus, the latter known to send only minor scattered projections to the PVT (Wang *et al.*, 2021*b*). Although to our knowledge no other studies have functionally assessed this NTSTH→PVT connection, the impact of 6-OHDA^{PVT} on NTSTH-neurons is in line with the presence of a dense plexus of PVT-reaching TH fibers when fluorescent recombinant markers are directly microinjected in the NTS of *Th*-Cre animals (Aklan *et al.*, 2020) as well as with a recent retrograde viral study identifying a subset of NTSTH-neurons projecting to the PVT (Kirouac *et al.*, 2022). This evidence is of paramount important since NTS and LC catecholaminergic neurons, by converging onto the PVT, may synergistically modulate feeding patterns and metabolic efficiency in stressogenic contexts. In fact, NTSTH- and LCTH-neurons are well-known to modulate food intake and stress/novelty, respectively (McCall *et al.*, 2015; Roman *et al.*, 2016; Takeuchi *et al.*, 2016).

Recent studies have shown that activation of CA-releasing NTS$^{DBH/TH}$-neurons may result in a reduction (Roman *et al.*, 2016; Chen *et al.*, 2020) as well as in an increase (Aklan *et al.*, 2020; Chen *et al.*, 2020) of food intake depending on CA cell types and/or projection sites [parabrachial nucleus (PBN) *vs* arcuate nucleus (Arc)]. Although not directly assessed in our study, our results suggest that NTSTH→PVT projecting neurons may serve as anorexigenic stimuli since their ablation enhances food intake under stress-related contexts. Indeed, it may be legitimately argued that the use of local microinjections of 6-OHDA may result in the degeneration of LCTH- and NTSTH-neurons projecting to the PVT but eventually to also other brain regions. However, loss of hindbrainTH neurons projecting to the medial hypothalamus resulted in a loss of glucoprivation-induced feeding (Fraley & Ritter, 2003; Hudson & Ritter, 2004), while in our study we show an increased food consumption under glucoprivic conditions when hindbrainTH→PVT projections were ablated. Moreover, opto-activation of NTSTH→Arc projections leads to an increase in food intake (Aklan *et al.*, 2020), whereas in our case enhanced food intake was elicited by the absence of hindbrainTH→PVT projections. These effects may substantiate the functional

546 selectivity of hindbrainTH→PVT projections in the adaptive responses of feeding,
547 metabolic efficiency and nutrient partitioning to stress-related contexts. Indeed, future
548 investigations using projection-specific optogenetics and/or chemogenetics may
549 definitely help in dissecting out the distinct and, most importantly, synergistic roles of
550 NTSTH→PVT and LCTH→PVT transmissions in guiding the tight interaction between
551 food intake, metabolism and stress-like allostatic stimuli.
552 Moreover, we observed that deletion of hindbrainTH→PVT inputs led to an increase in
553 PVT-cells activity (cFos), suggesting that catecholamines may act, directly and/or
554 indirectly, as negative modulators onto PVT-neurons. At first, this is surprising and
555 counterintuitive since *ex vivo* bath-applications of DA (precursor of NE) or NE, which
556 can be synaptically co-released from CA terminals (Smith & Greene, 2012;
557 Kempadoo *et al.*, 2016), lead to disinhibition [DA, (Beas *et al.*, 2018)] or activation
558 [NE, (Wang *et al.*, 2021*b*)] of PVT-neurons. However, it should be mentioned that
559 mid-posterior PVT-neurons express DA D2 and D3 receptors (Rieck *et al.*, 2004;
560 Clark *et al.*, 2017; Beas *et al.*, 2018; Gao *et al.*, 2020) as well as NE receptors such
561 as the α1, α2, β1 and β2 receptors (Rainbow *et al.*, 1984; Pieribone *et al.*, 1994;
562 Rosin *et al.*, 1996). Indeed, (*i*) how this variety of G-protein-coupled receptors (G_i-,
563 G_s-, G_q- and β-arrestin-coupled receptors) mechanistically contribute to the overall
564 modulation of PVT-neurons and (*ii*) whether 6-OHDA-induced TH deletion
565 reorganizes the expression of the above-mentioned CA receptors in the PVT require
566 future investigations. Although our results cannot distinguish between the functional
567 roles of distinct catecholamines onto their associated multiple receptors located onto
568 PVT-neurons, it is worth to mention that the PVT does not receive pure DA-fibers
569 from the midbrain (SNc and VTA) (Li *et al.*, 2014; Papathanou *et al.*, 2019) and that
570 direct 6-OHDA-induced LCTH-neurons loss was associated to an increased
571 expression of Gq-coupled α1 receptor in the thalamus (Szot *et al.*, 2012*a*) which may
572 explain, at least in part, the increase in PVT cFos-neurons.
573 The enhanced activation of PVT-neurons in 6-OHDA^{PVT} mice and the associated
574 feeding behaviors are in line with reports showing that stressors as well as hunger
575 are able to activate PVT-neurons (Bubser & Deutch, 1999; Beas *et al.*, 2018; Hua *et*
576 *al.*, 2018). In addition, our results are also in line with a recent report showing that
577 activation of PVT-neurons by oxytocin was efficient in suppressing stress-induced
578 hypophagia (Barrett *et al.*, 2021). However, it is worth to mention that satietogenic

signals are also able to activate PVT-neurons (Ong *et al.*, 2017), therefore indicating that PVT excitatory (glutamate) neurons may be actually segregated into several cell types with distinct neurochemical, cellular and functional features. This is already supported by the existence of at least two neuronal populations [galanin- and dopamine 2 receptor (D2R)-positive neurons, (Gao *et al.*, 2020)] and, as already suggested by the presence of several neuropeptides in PVT-neurons (Curtis *et al.*, 2020), it may not be hazardous to hypothesize that future cell type-specific transcriptomic analyses will reveal new sub-families and clusters.

The homeostatic processes underlying food intake, energy balance and metabolic efficiency strongly depend on the activity of the hypothalamus (Dietrich & Horvath, 2013; Timper & Brüning, 2017). We observed that depletion of TH-afferents to the PVT resulted not only in the activation of PVT-neurons but also in the concomitant activation of hypothalamic regions, notably the lateral (LH) and the dorsomedial (DMH) hypothalamus. Indeed, activation of LH and DMH cell types has been shown to promote feeding (Jennings *et al.*, 2015; Navarro *et al.*, 2016; Otgon-Uul *et al.*, 2016; Jeong *et al.*, 2017) even following anxiogenic environmental cues (Cassidy *et al.*, 2019). Although we cannot rule out yet whether and how the adaptive activation of PVT-neurons following TH deletion may be responsible for the direct activation of LH and DMH regions, it is interesting to note that PVT excitatory (glutamate) neurons also project to the hypothalamus (Engelke *et al.*, 2021; Li *et al.*, 2021), therefore potentially modulating feeding and energy homeostasis. This is also supported by our viral tracing strategy which revealed direct PVT→DMH/LH projections. However, we cannot formally exclude that the partial loss of hindbrain TH-neurons may impact on the hypothalamic activity in virtue of other circuits (hindbrainTH→hypothalamus and/or hindbrainTH→PBN→hypothalamus paths). Indeed, while the existence of a hypothalamus→PVT→accumbal circuit seems critical for behavioral adaptations (Betley *et al.*, 2013; Zhang & van den Pol, 2017; Otis *et al.*, 2019; Meffre *et al.*, 2019; Zhang *et al.*, 2020; Iglesias & Flagel, 2021; Engelke *et al.*, 2021), our results, together with previous and recent literature (Otake *et al.*, 1994; Ong *et al.*, 2017; Beas *et al.*, 2018; Sofia Beas *et al.*, 2020; Li *et al.*, 2021), also suggest a hindbrain→PVT→hypothalamus path that may regulate homeostatic functions requiring the integration of exteroceptive and interoceptive signals.

In conclusion, the PVT has been classically positioned as a functional node of the limbic circuit (Barson *et al.*, 2020). Only recently the hypothesis of the PVT as a

homeostatic relay has been proposed (Penzo & Gao, 2021). Altogether, our results support the working hypothesis according to which the PVT, through its afferent connections with NTS^{TH}- and LC^{TH}-neurons, may represent a functional interface between homeostatic and emotional states, thereby leading to allostatic adaptations. This study, besides highlighting the existence of a dual hindbrain-to-thalamus connection, (*i*) provides new evidence to better understand the dynamic processes underlying the regulation of food intake and energy metabolism, and (*ii*) may serve as starting step to explore the functional relationships and comorbidities between psychiatric (stress) and metabolic (anorexia, obesity, binge eating) disorders.

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Author Contributions

C.D. performed and analyzed most of the experiments. G.L. performed immunofluorescence studies. J.C. performed surgeries. S.L. provided critical feedback. G.G. conceived and supervised the whole project, and wrote the manuscript with contribution from all coauthors.

Data availability statement

All data are presented in the manuscript or supplementary information. For Suppl. Figures see https://doi.org/10.6084/m9.figshare.19683228.v1.

Competing interests

The authors declare no competing interests.

References

Aklan I, Sayar Atasoy N, Yavuz Y, Ates T, Coban I, Koksalar F, Filiz G, Topcu IC, Oncul M, Dilsiz P, Cebecioglu U, Alp MI, Yilmaz B, Davis DR, Hajdukiewicz K, Saito K, Konopka W, Cui H & Atasoy D (2020). NTS Catecholamine Neurons Mediate Hypoglycemic Hunger via Medial Hypothalamic Feeding Pathways. *Cell Metab* **31,** 313-326.e5.

Arch JRS, Hislop D, Wang SJY & Speakman JR (2006). Some mathematical and technical issues in the measurement and interpretation of open-circuit indirect calorimetry in small animals. *Int J Obes (Lond)* **30,** 1322–1331.

Barrett LR, Nunez J & Zhang X (2021). Oxytocin activation of paraventricular thalamic neurons promotes feeding motivation to attenuate stress-induced hypophagia. *Neuropsychopharmacology* **46,** 1045–1056.

Barson JR, Mack NR & Gao W-J (2020). The Paraventricular Nucleus of the Thalamus Is an Important Node in the Emotional Processing Network. *Front Behav Neurosci* **14,** 598469.

Beas BS, Wright BJ, Skirzewski M, Leng Y, Hyun JH, Koita O, Ringelberg N, Kwon H-B, Buonanno A & Penzo MA (2018). The locus coeruleus drives disinhibition in the midline thalamus via a dopaminergic mechanism. *Nat Neurosci* **21,** 963–973.

Benfato ID, Quintanilha ACS, Henrique JS, Souza MA, Rosário BDA, Beserra Filho JIA, Santos RLO, Ribeiro AM, Le Sueur Maluf L & de Oliveira CAM (2022). Effects of long-term social isolation on central, behavioural and metabolic parameters in middle-aged mice. *Behav Brain Res* **417,** 113630.

Berland C et al. (2020). Circulating Triglycerides Gate Dopamine-Associated Behaviors through DRD2-Expressing Neurons. *Cell Metab* **31,** 773-790.e11.

Berland C, Castel J, Terrasi R, Montalban E, Foppen E, Martin C, Muccioli GG, Luquet S & Gangarossa G (2022). Identification of an endocannabinoid gut-brain vagal mechanism controlling food reward and energy homeostasis. *Mol Psychiatry*; DOI: 10.1038/s41380-021-01428-z.

Betley JN, Cao ZFH, Ritola KD & Sternson SM (2013). Parallel, redundant circuit organization for homeostatic control of feeding behavior. *Cell* **155,** 1337–1350.

Bhatnagar S & Dallman MF (1999). The paraventricular nucleus of the thalamus alters rhythms in core temperature and energy balance in a state-dependent manner. *Brain Res* **851,** 66–75.

Bhatnagar S, Viau V, Chu A, Soriano L, Meijer OC & Dallman MF (2000). A

cholecystokinin-mediated pathway to the paraventricular thalamus is recruited in chronically stressed rats and regulates hypothalamic-pituitary-adrenal function. *J Neurosci* **20,** 5564–5573.

Bubser M & Deutch AY (1999). Stress induces Fos expression in neurons of the thalamic paraventricular nucleus that innervate limbic forebrain sites. *Synapse* **32,** 13–22.

Cassidy RM, Lu Y, Jere M, Tian J-B, Xu Y, Mangieri LR, Felix-Okoroji B, Selever J, Xu Y, Arenkiel BR & Tong Q (2019). A lateral hypothalamus to basal forebrain neurocircuit promotes feeding by suppressing responses to anxiogenic environmental cues. *Sci Adv* **5,** eaav1640.

Challet E (2019). The circadian regulation of food intake. *Nat Rev Endocrinol* **15,** 393–405.

Chen J, Cheng M, Wang L, Zhang L, Xu D, Cao P, Wang F, Herzog H, Song S & Zhan C (2020). A Vagal-NTS Neural Pathway that Stimulates Feeding. *Curr Biol* **30,** 3986-3998.e5.

Clark AM, Leroy F, Martyniuk KM, Feng W, McManus E, Bailey MR, Javitch JA, Balsam PD & Kellendonk C (2017). Dopamine D2 Receptors in the Paraventricular Thalamus Attenuate Cocaine Locomotor Sensitization. *eNeuro*; DOI: 10.1523/ENEURO.0227-17.2017.

Colavito V, Tesoriero C, Wirtu AT, Grassi-Zucconi G & Bentivoglio M (2015). Limbic thalamus and state-dependent behavior: The paraventricular nucleus of the thalamic midline as a node in circadian timing and sleep/wake-regulatory networks. *Neurosci Biobehav Rev* **54,** 3–17.

Curtis GR, Oakes K & Barson JR (2020). Expression and Distribution of Neuropeptide-Expressing Cells Throughout the Rodent Paraventricular Nucleus of the Thalamus. *Front Behav Neurosci* **14,** 634163.

Dietrich MO & Horvath TL (2013). Hypothalamic control of energy balance: insights into the role of synaptic plasticity. *Trends Neurosci* **36,** 65–73.

Do-Monte FH, Minier-Toribio A, Quiñones-Laracuente K, Medina-Colón EM & Quirk GJ (2017). Thalamic Regulation of Sucrose Seeking during Unexpected Reward Omission. *Neuron* **94,** 388-400.e4.

Do-Monte FH, Quiñones-Laracuente K & Quirk GJ (2015). A temporal shift in the circuits mediating retrieval of fear memory. *Nature* **519,** 460–463.

Engelke DS, Zhang XO, O'Malley JJ, Fernandez-Leon JA, Li S, Kirouac GJ, Beierlein

M & Do-Monte FH (2021). A hypothalamic-thalamostriatal circuit that controls approach-avoidance conflict in rats. *Nat Commun* **12,** 2517.

Even PC & Nadkarni NA (2012). Indirect calorimetry in laboratory mice and rats: principles, practical considerations, interpretation and perspectives. *Am J Physiol Regul Integr Comp Physiol* **303,** R459-476.

Fraley GS & Ritter S (2003). Immunolesion of norepinephrine and epinephrine afferents to medial hypothalamus alters basal and 2-deoxy-D-glucose-induced neuropeptide Y and agouti gene-related protein messenger ribonucleic acid expression in the arcuate nucleus. *Endocrinology* **144,** 75–83.

Gangarossa G, Castell L, Castro L, Tarot P, Veyrunes F, Vincent P, Bertaso F & Valjent E (2019). Contrasting patterns of ERK activation in the tail of the striatum in response to aversive and rewarding signals. *J Neurochem* **151,** 204–226.

Gangarossa G, Espallergues J, Mailly P, De Bundel D, de Kerchove d'Exaerde A, Hervé D, Girault J-A, Valjent E & Krieger P (2013). Spatial distribution of D1R- and D2R-expressing medium-sized spiny neurons differs along the rostro-caudal axis of the mouse dorsal striatum. *Front Neural Circuits* **7,** 124.

Gao C, Leng Y, Ma J, Rooke V, Rodriguez-Gonzalez S, Ramakrishnan C, Deisseroth K & Penzo MA (2020). Two genetically, anatomically and functionally distinct cell types segregate across anteroposterior axis of paraventricular thalamus. *Nat Neurosci* **23,** 217–228.

García-Cabezas MA, Martínez-Sánchez P, Sánchez-González MA, Garzón M & Cavada C (2009). Dopamine innervation in the thalamus: monkey versus rat. *Cereb Cortex* **19,** 424–434.

Herzog H (2020). Integrated pathways that control stress and energy homeostasis. *Nat Rev Endocrinol* **16,** 75–76.

Heydendael W, Sharma K, Iyer V, Luz S, Piel D, Beck S & Bhatnagar S (2011). Orexins/hypocretins act in the posterior paraventricular thalamic nucleus during repeated stress to regulate facilitation to novel stress. *Endocrinology* **152,** 4738–4752.

Hsu DT, Kirouac GJ, Zubieta J-K & Bhatnagar S (2014). Contributions of the paraventricular thalamic nucleus in the regulation of stress, motivation, and mood. *Front Behav Neurosci* **8,** 73.

Hua R, Wang X, Chen X, Wang X, Huang P, Li P, Mei W & Li H (2018). Calretinin Neurons in the Midline Thalamus Modulate Starvation-Induced Arousal. *Curr Biol* **28,**

3948-3959.e4.

Hudson B & Ritter S (2004). Hindbrain catecholamine neurons mediate consummatory responses to glucoprivation. *Physiol Behav* **82,** 241–250.

Iglesias AG & Flagel SB (2021). The Paraventricular Thalamus as a Critical Node of Motivated Behavior via the Hypothalamic-Thalamic-Striatal Circuit. *Front Integr Neurosci* **15,** 706713.

Jennings JH, Ung RL, Resendez SL, Stamatakis AM, Taylor JG, Huang J, Veleta K, Kantak PA, Aita M, Shilling-Scrivo K, Ramakrishnan C, Deisseroth K, Otte S & Stuber GD (2015). Visualizing hypothalamic network dynamics for appetitive and consummatory behaviors. *Cell* **160,** 516–527.

Jeong JH, Lee DK & Jo Y-H (2017). Cholinergic neurons in the dorsomedial hypothalamus regulate food intake. *Mol Metab* **6,** 306–312.

Joly-Amado A, Denis RGP, Castel J, Lacombe A, Cansell C, Rouch C, Kassis N, Dairou J, Cani PD, Ventura-Clapier R, Prola A, Flamment M, Foufelle F, Magnan C & Luquet S (2012). Hypothalamic AgRP-neurons control peripheral substrate utilization and nutrient partitioning. *EMBO J* **31,** 4276–4288.

Kempadoo KA, Mosharov EV, Choi SJ, Sulzer D & Kandel ER (2016). Dopamine release from the locus coeruleus to the dorsal hippocampus promotes spatial learning and memory. *Proc Natl Acad Sci U S A* **113,** 14835–14840.

Kessler S, Labouèbe G, Croizier S, Gaspari S, Tarussio D & Thorens B (2021). Glucokinase neurons of the paraventricular nucleus of the thalamus sense glucose and decrease food consumption. *iScience* **24,** 103122.

Kirouac GJ (2015). Placing the paraventricular nucleus of the thalamus within the brain circuits that control behavior. *Neurosci Biobehav Rev* **56,** 315–329.

Kirouac GJ, Li S & Li S (2022). Convergence of monosynaptic inputs from neurons in the brainstem and forebrain on parabrachial neurons that project to the paraventricular nucleus of the thalamus. 2022.02.23.481069. Available at: https://www.biorxiv.org/content/10.1101/2022.02.23.481069v1 [Accessed April 18, 2022].

Kvetnansky R, Sabban EL & Palkovits M (2009). Catecholaminergic systems in stress: structural and molecular genetic approaches. *Physiol Rev* **89,** 535–606.

Labouèbe G, Boutrel B, Tarussio D & Thorens B (2016). Glucose-responsive neurons of the paraventricular thalamus control sucrose-seeking behavior. *Nat Neurosci* **19,** 999–1002.

783 Lecorps B, Rödel HG & Féron C (2016). Assessment of anxiety in open field and
784 elevated plus maze using infrared thermography. *Physiol Behav* **157,** 209–216.
785 Lee J-S, Kang J-Y & Son C-G (2020). A Comparison of Isolation Stress and
786 Unpredictable Chronic Mild Stress for the Establishment of Mouse Models of
787 Depressive Disorder. *Front Behav Neurosci* **14,** 616389.
788 Lee J-S, Lee S-B, Kim D-W, Shin N, Jeong S-J, Yang C-H & Son C-G (2021). Social
789 isolation-related depression accelerates ethanol intake via microglia-derived
790 neuroinflammation. *Sci Adv* **7,** eabj3400.
791 Lewis SR, Ahmed S, Khaimova E, Israel Y, Singh A, Kandov Y, Kest B & Bodnar RJ
792 (2006). Genetic variance contributes to ingestive processes: a survey of 2-deoxy-D-
793 glucose-induced feeding in eleven inbred mouse strains. *Physiol Behav* **87,** 595–601.
794 Li S, Dong X & Kirouac GJ (2021). Extensive divergence of projections to the
795 forebrain from neurons in the paraventricular nucleus of the thalamus. *Brain Struct*
796 *Funct* **226,** 1779–1802.
797 Li S, Shi Y & Kirouac GJ (2014). The hypothalamus and periaqueductal gray are the
798 sources of dopamine fibers in the paraventricular nucleus of the thalamus in the rat.
799 *Front Neuroanat* **8,** 136.
800 Lin L-H, Moore SA, Jones SY, McGlashon J & Talman WT (2013). Astrocytes in the
801 rat nucleus tractus solitarii are critical for cardiovascular reflex control. *J Neurosci* **33,**
802 18608–18617.
803 Maniam J & Morris MJ (2012). The link between stress and feeding behaviour.
804 *Neuropharmacology* **63,** 97–110.
805 McCall JG, Al-Hasani R, Siuda ER, Hong DY, Norris AJ, Ford CP & Bruchas MR
806 (2015). CRH Engagement of the Locus Coeruleus Noradrenergic System Mediates
807 Stress-Induced Anxiety. *Neuron* **87,** 605–620.
808 Meffre J, Sicre M, Diarra M, Marchessaux F, Paleressompoulle D & Ambroggi F
809 (2019). Orexin in the Posterior Paraventricular Thalamus Mediates Hunger-Related
810 Signals in the Nucleus Accumbens Core. *Curr Biol* **29,** 3298-3306.e4.
811 Navarro M, Olney JJ, Burnham NW, Mazzone CM, Lowery-Gionta EG, Pleil KE,
812 Kash TL & Thiele TE (2016). Lateral Hypothalamus GABAergic Neurons Modulate
813 Consummatory Behaviors Regardless of the Caloric Content or Biological Relevance
814 of the Consumed Stimuli. *Neuropsychopharmacology* **41,** 1505–1512.
815 Ong ZY, Liu J-J, Pang ZP & Grill HJ (2017). Paraventricular Thalamic Control of
816 Food Intake and Reward: Role of Glucagon-Like Peptide-1 Receptor Signaling.

Neuropsychopharmacology **42,** 2387–2397.

Otake K, Reis DJ & Ruggiero DA (1994). Afferents to the midline thalamus issue collaterals to the nucleus tractus solitarii: an anatomical basis for thalamic and visceral reflex integration. *J Neurosci* **14,** 5694–5707.

Otgon-Uul Z, Suyama S, Onodera H & Yada T (2016). Optogenetic activation of leptin- and glucose-regulated GABAergic neurons in dorsomedial hypothalamus promotes food intake via inhibitory synaptic transmission to paraventricular nucleus of hypothalamus. *Mol Metab* **5,** 709–715.

Otis JM, Namboodiri VMK, Matan AM, Voets ES, Mohorn EP, Kosyk O, McHenry JA, Robinson JE, Resendez SL, Rossi MA & Stuber GD (2017). Prefrontal cortex output circuits guide reward seeking through divergent cue encoding. *Nature* **543,** 103–107.

Otis JM, Zhu M, Namboodiri VMK, Cook CA, Kosyk O, Matan AM, Ying R, Hashikawa Y, Hashikawa K, Trujillo-Pisanty I, Guo J, Ung RL, Rodriguez-Romaguera J, Anton ES & Stuber GD (2019). Paraventricular Thalamus Projection Neurons Integrate Cortical and Hypothalamic Signals for Cue-Reward Processing. *Neuron* **103,** 423-431.e4.

Papathanou M, Dumas S, Pettersson H, Olson L & Wallén-Mackenzie Å (2019). Off-Target Effects in Transgenic Mice: Characterization of Dopamine Transporter (DAT)-Cre Transgenic Mouse Lines Exposes Multiple Non-Dopaminergic Neuronal Clusters Available for Selective Targeting within Limbic Neurocircuitry. *eNeuro* **6,** ENEURO.0198-19.2019.

Peng Z-C & Bentivoglio M (2004). The thalamic paraventricular nucleus relays information from the suprachiasmatic nucleus to the amygdala: a combined anterograde and retrograde tracing study in the rat at the light and electron microscopic levels. *J Neurocytol* **33,** 101–116.

Pénicaud L, Thompson DA & Le Magnen J (1986). Effects of 2-deoxy-D-glucose on food and water intake and body temperature in rats. *Physiol Behav* **36,** 431–435.

Penzo MA & Gao C (2021). The paraventricular nucleus of the thalamus: an integrative node underlying homeostatic behavior. *Trends Neurosci* **44,** 538–549.

Penzo MA, Robert V, Tucciarone J, De Bundel D, Wang M, Van Aelst L, Darvas M, Parada LF, Palmiter RD, He M, Huang ZJ & Li B (2015). The paraventricular thalamus controls a central amygdala fear circuit. *Nature* **519,** 455–459.

Pieribone VA, Nicholas AP, Dagerlind A & Hökfelt T (1994). Distribution of alpha 1 adrenoceptors in rat brain revealed by in situ hybridization experiments utilizing

subtype-specific probes. *J Neurosci* **14,** 4252–4268.

Pliota P, Böhm V, Grössl F, Griessner J, Valenti O, Kraitsy K, Kaczanowska J, Pasieka M, Lendl T, Deussing JM & Haubensak W (2020). Stress peptides sensitize fear circuitry to promote passive coping. *Mol Psychiatry* **25,** 428–441.

Qu N, He Y, Wang C, Xu P, Yang Y, Cai X, Liu H, Yu K, Pei Z, Hyseni I, Sun Z, Fukuda M, Li Y, Tian Q & Xu Y (2020). A POMC-originated circuit regulates stress-induced hypophagia, depression, and anhedonia. *Mol Psychiatry* **25,** 1006–1021.

Rabasa C & Dickson SL (2016). Impact of stress on metabolism and energy balance. *Current Opinion in Behavioral Sciences* **9,** 71–77.

Rainbow TC, Parsons B & Wolfe BB (1984). Quantitative autoradiography of beta 1- and beta 2-adrenergic receptors in rat brain. *Proc Natl Acad Sci U S A* **81,** 1585–1589.

Ren S et al. (2018). The paraventricular thalamus is a critical thalamic area for wakefulness. *Science* **362,** 429–434.

Rieck RW, Ansari MS, Whetsell WO, Deutch AY & Kessler RM (2004). Distribution of dopamine D2-like receptors in the human thalamus: autoradiographic and PET studies. *Neuropsychopharmacology* **29,** 362–372.

Roman CW, Derkach VA & Palmiter RD (2016). Genetically and functionally defined NTS to PBN brain circuits mediating anorexia. *Nat Commun* **7,** 11905.

Rosin DL, Talley EM, Lee A, Stornetta RL, Gaylinn BD, Guyenet PG & Lynch KR (1996). Distribution of alpha 2C-adrenergic receptor-like immunoreactivity in the rat central nervous system. *J Comp Neurol* **372,** 135–165.

Rybkin II, Zhou Y, Volaufova J, Smagin GN, Ryan DH & Harris RB (1997). Effect of restraint stress on food intake and body weight is determined by time of day. *Am J Physiol* **273,** R1612-1622.

Schroeter S, Apparsundaram S, Wiley RG, Miner LH, Sesack SR & Blakely RD (2000). Immunolocalization of the cocaine- and antidepressant-sensitive l-norepinephrine transporter. *J Comp Neurol* **420,** 211–232.

Smith CC & Greene RW (2012). CNS dopamine transmission mediated by noradrenergic innervation. *J Neurosci* **32,** 6072–6080.

Sofia Beas B, Gu X, Leng Y, Koita O, Rodriguez-Gonzalez S, Kindel M, Matikainen-Ankney BA, Larsen RS, Kravitz AV, Hoon MA & Penzo MA (2020). A ventrolateral medulla-midline thalamic circuit for hypoglycemic feeding. *Nat Commun* **11,** 6218.

Sweeney P & Yang Y (2017). Neural Circuit Mechanisms Underlying Emotional

885 Regulation of Homeostatic Feeding. *Trends Endocrinol Metab* **28,** 437–448.
886 Szot P, Franklin A, Sikkema C, Wilkinson CW & Raskind MA (2012*a*). Sequential
887 Loss of LC Noradrenergic and Dopaminergic Neurons Results in a Correlation of
888 Dopaminergic Neuronal Number to Striatal Dopamine Concentration. *Front*
889 *Pharmacol* **3,** 184.
890 Szot P, Knight L, Franklin A, Sikkema C, Foster S, Wilkinson CW, White SS &
891 Raskind MA (2012*b*). Lesioning noradrenergic neurons of the locus coeruleus in
892 C57Bl/6 mice with unilateral 6-hydroxydopamine injection, to assess molecular,
893 electrophysiological and biochemical changes in noradrenergic signaling.
894 *Neuroscience* **216,** 143–157.
895 Takatsu-Coleman AL, Patti CL, Zanin KA, Zager A, Carvalho RC, Borçoi AR, Ceccon
896 LMB, Berro LF, Tufik S, Andersen ML & Frussa-Filho R (2013). Short-term social
897 isolation induces depressive-like behaviour and reinstates the retrieval of an aversive
898 task: mood-congruent memory in male mice? *J Psychiatry Neurosci* **38,** 259–268.
899 Takeuchi T, Duszkiewicz AJ, Sonneborn A, Spooner PA, Yamasaki M, Watanabe M,
900 Smith CC, Fernández G, Deisseroth K, Greene RW & Morris RGM (2016). Locus
901 coeruleus and dopaminergic consolidation of everyday memory. *Nature* **537,** 357–
902 362.
903 Timper K & Brüning JC (2017). Hypothalamic circuits regulating appetite and energy
904 homeostasis: pathways to obesity. *Dis Model Mech* **10,** 679–689.
905 Ulrich-Lai YM, Fulton S, Wilson M, Petrovich G & Rinaman L (2015). Stress
906 exposure, food intake and emotional state. *Stress* **18,** 381–399.
907 Valentino RJ & Van Bockstaele E (2008). Convergent regulation of locus coeruleus
908 activity as an adaptive response to stress. *Eur J Pharmacol* **583,** 194–203.
909 Valjent E & Gangarossa G (2021). The Tail of the Striatum: From Anatomy to
910 Connectivity and Function. *Trends Neurosci* **44,** 203–214.
911 Vallès A, Martí O, García A & Armario A (2000). Single exposure to stressors causes
912 long-lasting, stress-dependent reduction of food intake in rats. *Am J Physiol Regul*
913 *Integr Comp Physiol* **279,** R1138-1144.
914 Wang C, Zhou W, He Y, Yang T, Xu P, Yang Y, Cai X, Wang J, Liu H, Yu M, Liang
915 C, Yang T, Liu H, Fukuda M, Tong Q, Wu Q, Sun Z, He Y & Xu Y (2021*a*). AgRP
916 neurons trigger long-term potentiation and facilitate food seeking. *Transl Psychiatry*
917 **11,** 11.
918 Wang Y, Xu L, Liu M-Z, Hu D-D, Fang F, Xu D-J, Zhang R, Hua X-X, Li J-B, Zhang L,

Huang L-N & Mu D (2021*b*). Norepinephrine modulates wakefulness via α1 adrenoceptors in paraventricular thalamic nucleus. *iScience* **24,** 103015.

Xu Y, Lu Y, Cassidy RM, Mangieri LR, Zhu C, Huang X, Jiang Z, Justice NJ, Xu Y, Arenkiel BR & Tong Q (2019). Identification of a neurocircuit underlying regulation of feeding by stress-related emotional responses. *Nat Commun* **10,** 3446.

Zhang C, Kaye JA, Cai Z, Wang Y, Prescott SL & Liberles SD (2021). Area Postrema Cell Types that Mediate Nausea-Associated Behaviors. *Neuron* **109,** 461-472.e5.

Zhang J, Chen D, Sweeney P & Yang Y (2020). An excitatory ventromedial hypothalamus to paraventricular thalamus circuit that suppresses food intake. *Nat Commun* **11,** 6326.

Zhang X & van den Pol AN (2017). Rapid binge-like eating and body weight gain driven by zona incerta GABA neuron activation. *Science* **356,** 853–859.

Figure legends

Figure 1: Retrograde ablation of hindbrain catecholaminergic inputs contribute to PVT activity. (**A**) Drawing represents the microinjection of 6-OHDA or Vehicle (Veh) in the mid-posterior PVT. (**B**) Immunofluorescence detection of tyrosine hydroxylase (TH) within the PVT in Sham^{PVT} and 6-OHDA^{PVT} mice. Scale bar: 150 µm. (**C**) Immunofluorescence detection of TH in PVT-projecting hindbrain regions [nucleus tractus solitarius (NTS), area A1 and locus coeruleus (LC)] in Sham^{PVT} and 6-OHDA^{PVT} mice. Scale bars: 150 µm. (**D**) Quantification of TH-positive neurons in the NTS, A1 and LC. Statistics: * $p<0.05$, ** $p<0.01$, 6-OHDA^{PVT} *vs* Sham^{PVT} mice. (**E**) Immunofluorescence detection of cFos-positive neurons in the mid-posterior PVT (mPVT and pPVT). Scale bars: 150 µm. (**F**) Quantification of cFos-positive cells in the mid-posterior PVT. Statistics: *** $p<0.001$, 6-OHDA^{PVT} *vs* Sham^{PVT} mice. Data are presented as mean ± SD. For statistical details see **Statistical Summary Table**.

Figure 2: HindbrainTH→PVT inputs participate to novelty-induced hypophagia. (**A**) Body weight of Sham^{PVT} and 6-OHDA^{PVT} mice following 3-4 weeks from 6-OHDA or Veh microinjection in the PVT. (**B**) Experimental design indicating the transition from grouped to singled housing. (**C**) Food intake (g/day and kcal/day) during the first three days of isolation (D1 to D3). Statistics: * $p<0.05$, ** $p<0.01$, 6-OHDA^{PVT} *vs* Sham^{PVT} mice. (**D**) Investigation of food intake and metabolic efficiency using metabolic cages during two consecutive exposures to a novel environment. (**E**) Food intake during the dark phase (spontaneous eating) in Sham^{PVT} and 6-OHDA^{PVT} mice during two consecutive exposures to a novel environment. (**E^{1}**) Cumulative food intake. Statistics: *** $p<0.01$, Sham^{E2} *vs* Sham^{E1}, $^{\#\#}$ $p<0.01$, 6-OHDA^{E1} *vs* Sham^{E1} groups. (**F**) Spontaneous locomotor activity (beam breaks, bb) during the dark phase in Sham^{PVT} and 6-OHDA^{PVT} mice during two consecutive exposures to a novel environment. (**F^{1}**) Cumulative locomotor activity. Statistics: ** $p<0.01$, Sham^{E2} *vs* Sham^{E1}; $^{\#\#\#}$ $p<0.001$, 6-OHDA^{E2} *vs* 6-OHDA^{E1} groups. Note that both experimental groups showed similar degrees of habituation (reduced locomotor activity during the dark period). Measurements of the respiratory exchange ratio (RER, **G**), fatty acid oxidation (FAO, **H**), and energy expenditure (EE, **I**) in Sham^{PVT} and 6-OHDA^{PVT} mice during the first exposure (Exp.1) to a novel environment. (**G^{1}-1^{1}**) Averaged RER,

FAO and EE during the dark phase. Statistics: ** p<0.01, 6-OHDA^{E1} *vs* Sham^{E1}. Data are presented as mean ± SD. For statistical details see **Statistical Summary Table**.

Figure 3: HindbrainTH→PVT inputs do not alter novelty-induced anxiety and thermogenesis. (**A**) Drawing represents the open field (OF) test. (**B-F**) Parameters measured during a 20 min OF test: total distance, number of entries in the center, % of time in the center, mean exploration visits to the center, distance in the center. (**G**) Infrared thermographic images of animals after the OF test (20 min). (**H-J**) Temperature (°C) of the brown adipose tissue (BAT), lower back and tail in Sham^{PVT} and 6-OHDA^{PVT} mice before and after the OF test. Statistics: *** p<0.001, After *vs* Before OF test (Sham^{PVT} mice); $^{###}$ p<0.001, After *vs* Before OF test (6-OHDA^{PVT} mice). No differences were detected between Sham^{PVT} and 6-OHDA^{PVT} mice. Data are presented as mean ± SD. For statistical details see **Statistical Summary Table**.

Figure 4: HindbrainTH→PVT inputs contribute to the circadian adaptation of metabolic efficiency. (**A**) Drawing indicates the experimental procedure used to study feeding and metabolic adaptations during an inverted cycle (transition from Light-to-Dark to Dark-to-Light). (**B, B^{1}**) Food intake in Sham^{PVT} (**B**) and 6-OHDA^{PVT} (**B^{1}**) mice during the standard and inverted cycles. Note how the temporal dynamics of feeding change during the inverted cycle. (**C**) Cumulative food intake in Sham^{PVT} and 6-OHDA^{PVT} mice during the standard and inverted cycles. (**D, D^{1}**) Temporal dynamics of EE adaptations in Sham^{PVT} (**D**) and 6-OHDA^{PVT} (**D^{1}**) mice during the standard and inverted cycles. (**E, E^{1}**) Averaged EE in Sham^{PVT} and 6-OHDA^{PVT} mice according to matched inverted phases (7h-19h, **E**, and 19h-7h, **E^{1}**). Statistics: *** p<0.001, Inverted *vs* Standard cycle (Sham^{PVT} mice); $^{###}$ p<0.001, Inverted *vs* Standard cycle (6-OHDA^{PVT} mice). (**F, F^{1}**) Temporal dynamics of FAO adaptations in Sham^{PVT} (**F**) and 6-OHDA^{PVT} (**F^{1}**) mice during the standard and inverted cycles. (**G, G^{1}**) Averaged FAO in Sham^{PVT} and 6-OHDA^{PVT} mice according to matched inverted phases (7h-19h, **G**, and 19h-7h, **G^{1}**). Statistics: *** p<0.001, ** p<0.01, Inverted *vs* Standard cycle (Sham^{PVT} mice); $^{##}$ p<0.01, Inverted *vs* Standard cycle (6-OHDA^{PVT} mice). (**H, H^{1}**) Temporal dynamics of RER adaptations in Sham^{PVT} (**H**) and 6-OHDA^{PVT} (**H^{1}**) mice during the standard and inverted cycles. (**I, I^{1}**) Averaged RER in Sham^{PVT} and 6-OHDA^{PVT} mice according to matched inverted phases (7h-19h, **I**, and

19h-7h, **I**[1]). Statistics: *** p<0.001, ** p<0.01, Inverted *vs* Standard cycle (Sham^{PVT} mice); ## p<0.01, Inverted *vs* Standard cycle (6-OHDA^{PVT} mice). Data are presented as mean ± SD. For statistical details see **Statistical Summary Table**.

Figure 5: HindbrainTH→PVT inputs scale feeding following physiological stressors. (**A**) Drawing represents the novelty-suppressed feeding (NSF) test in overnight fasted Sham^{PVT} and 6-OHDA^{PVT} mice. (**B**) Latency to eat (first bite) during the NSF test. (**C, D**) Food intake, total kcal (**C**) and normalized kcal/BW (**D**) during the NSF test. Statistics: ** p<0.01, 6-OHDA^{PVT} *vs* Sham^{PVT} mice. (**E**) Food intake in Sham^{PVT} and 6-OHDA^{PVT} mice after a restraint stress (immobilization). NoR: No Restraint (control conditions). R: Restraint. Statistics: *** p<0.001, Restraint *vs* No Restraint (Sham^{PVT}); ## p<0.01, Restraint *vs* No Restraint (6-OHDA^{PVT}); $$$ p<0.001, 6-OHDA^{PVT} *vs* Sham^{PVT} mice (Restraint). Data are presented as mean ± SD. For statistical details see **Statistical Summary Table**.

Figure 6: HindbrainTH→PVT inputs scale feeding following metabolic stressors. (**A**) Food intake in Sham^{PVT} and 6-OHDA^{PVT} mice administered with 2-DG (500 mg/kg, i.p., neurogluopenia-induced feeding). (**B**) Glucose variation (%) following administration of 2-DG (0 *vs* 30 min post administration). Note: neuroglucopenia-induced hyperglycemia is the typical readout of the glucose counterregulatory response (CRR). (**C, D**) Glucose dynamics during the oral glucose tolerance test (OGTT, **C**) and the insulin tolerance test (ITT, **D**). No differences were observed between groups. (**E**) Food intake in overnight fasted Sham^{PVT} and 6-OHDA^{PVT} mice (refeeding). (**F**) Water intake in overnight water-deprived Sham^{PVT} and 6-OHDA^{PVT} mice. (**G**) Food intake in overnight water-deprived Sham^{PVT} and 6-OHDA^{PVT} mice. (**H**) Food intake in Sham^{PVT} and 6-OHDA^{PVT} mice administered with ghrelin (0.5 mg/kg, i.p., orexigenic response). Statistics: * p<0.05, ** p<0.01, *** p<0.001, 6-OHDA^{PVT} *vs* Sham^{PVT} mice. Data are presented as mean ± SD. For statistical details see **Statistical Summary Table**.

Figure 7: Deletion of hindbrainTH→PVT inputs increases cFos expression in the hypothalamus. (**A**) Immunofluorescence detection of cFos in hypothalamic regions of Sham^{PVT} and 6-OHDA^{PVT} mice, notably the dorsomedial hypothalamus (DMH), the

lateral hypothalamus (LH), the ventromedial hypothalamus (VMH) and the arcuate nucleus (Arc). Regions of interest are delineated by white dotted lines. Insets represent higher magnifications of DMH and LH regions. Scale bars: 500 µm. Quantification of cFos-positive cells in the DMH (**B**), LH (**C**), VMH (**D**) and Arc (**E**) regions. Statistics: ** $p<0.01$, 6-OHDA^{PVT} *vs* Sham^{PVT} mice. Abbreviations: f (fornix). (**F**) Drawing indicates the microinjection of AAV9-CAG-TdTomato in the PVT. (**G**) Immunofluorescence detection of TdTomato in the PVT (injection site), DMH and LH (projections). Scale bars: 150 µm. Data are presented as mean ± SD. For statistical details see **Statistical Summary Table**.

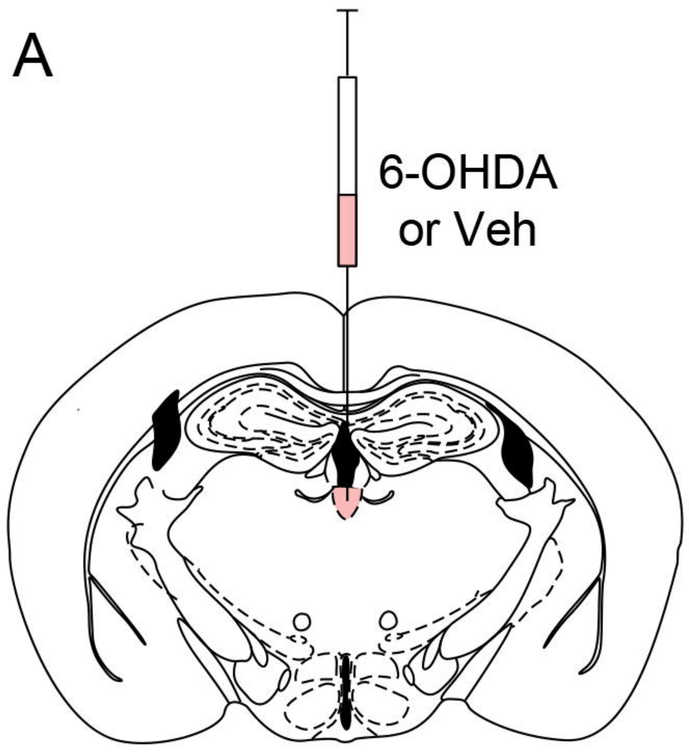
Suppl. Figure 1: Detection of DAT-positive fibers in the PVT and TH-positive neurons in the hypothalamus following PVT 6-OHDA microinjections. (**A**) Immunofluorescence detection of the dopamine transporter (DAT, red) and DAPI (blue) in the PVT, tail of the striatum (TS) and central amygdala (CeA). The lack of DAT-positive fibers in the PVT suggests no direct projections from dopamine-(DAT)-containing midbrain regions. Scale bar: 150 µm. (**B**) Immunofluorescence detection of TH (red) and DAPI (blue) in the hypothalamus of Sham^{PVT} and 6-OHDA^{PVT} mice. Scale bar: 500 µm.

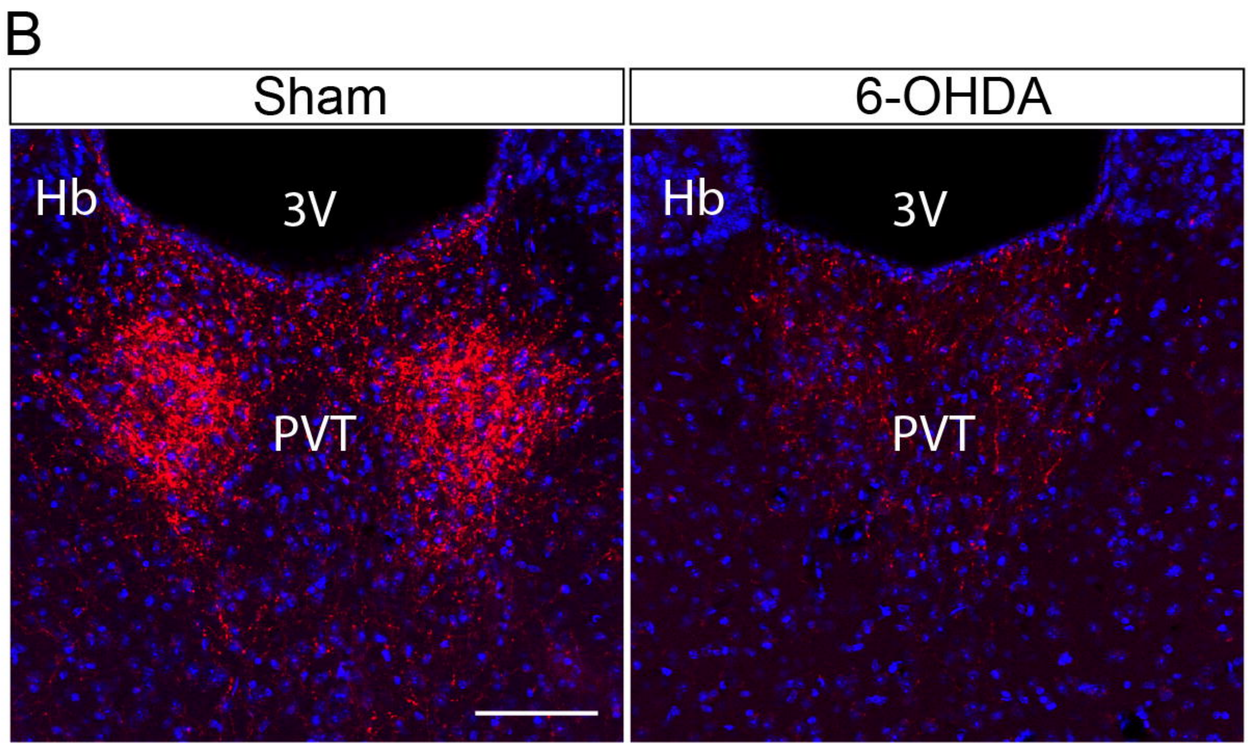
Suppl. Figure 2: HindbrainTH→PVT inputs dos not alter palatability for HFHS diet. (**A**) Food intake in Sham^{PVT} and 6-OHDA^{PVT} mice following time-locked feeding (1h) of high-fat high-sugar (HFHS) diet during two consecutive days. Statistics: ** $p<0.01$, Day2 *vs* Day1 (Sham^{PVT} mice); $^{\#\#}$ $p<0.01$, Day2 *vs* Day1 (6-OHDA^{PVT} mice). No differences between experimental groups. Data are presented as mean ± SD. For statistical details see **Statistical Summary Table**.

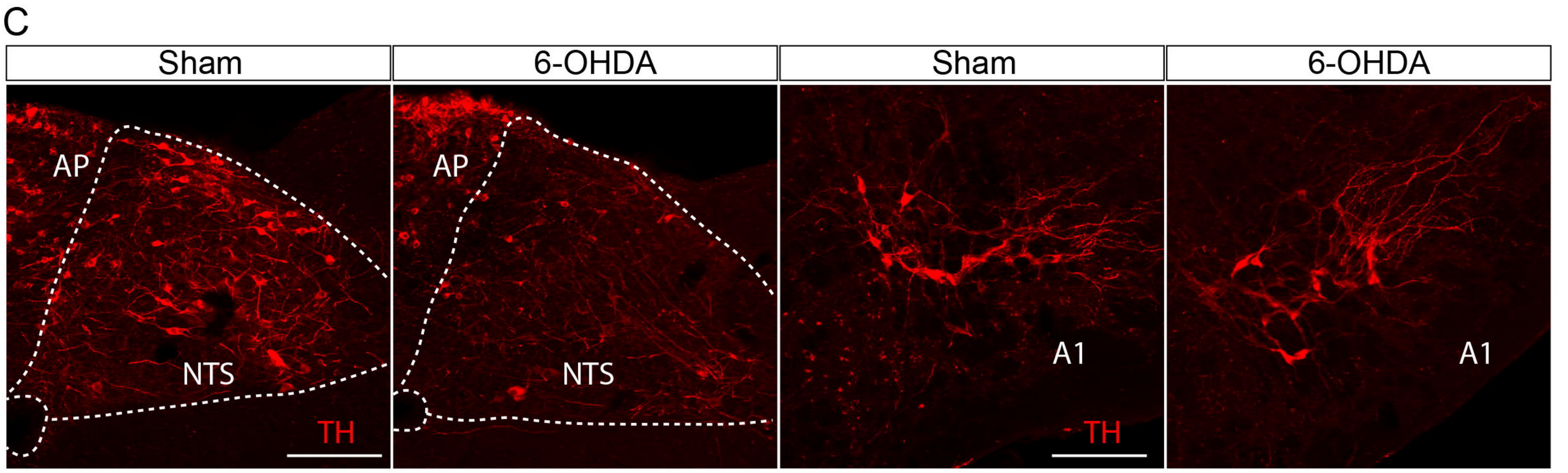
Suppl. Figure 3: Confirmation of sensitivity to novelty-induced hypophagia in Sham^{PVT} and 6-OHDA^{PVT} mice before the inverted cycle. (**A**) Cumulative food intake during the dark phase (spontaneous eating) in Sham^{PVT} and 6-OHDA^{PVT} mice during the first exposure to a novel environment (calorimetric chambers). (**B**) Total food intake during the dark phase. Statistics: *** $p<0.001$, ** $p<0.01$, 6-OHDA^{PVT} *vs* Sham^{PVT} mice. Data are presented as mean ± SD. For statistical details see **Statistical Summary Table**.

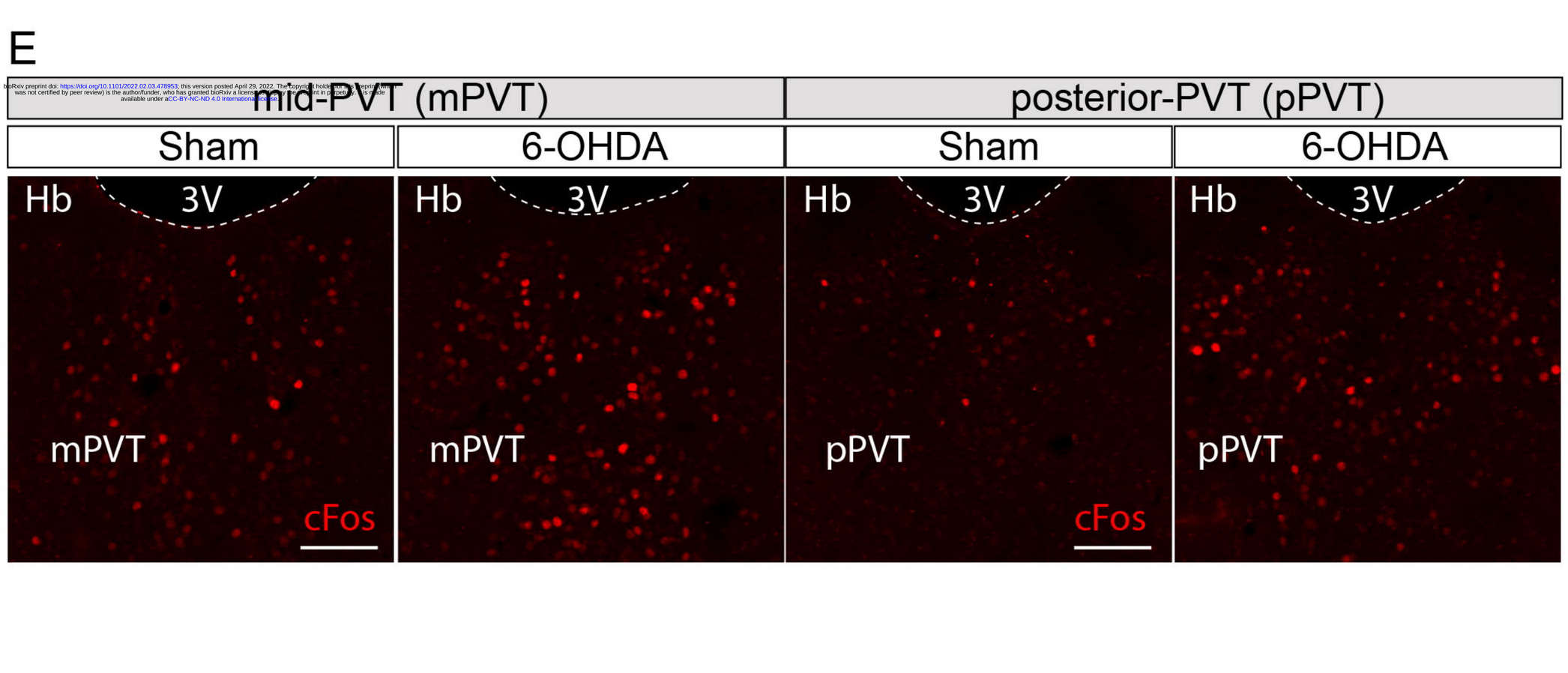
Suppl. Figure 4: Effect of fasting and water deprivation in Sham^{PVT} and 6-OHDA^{PVT} mice. (**A**) Loss of body weight and (**B**) glucose variations in overnight fasted animals used for the novelty-suppressed feeding (NSF) test. (**C**) Loss of body weight in overnight fasted animals before the refeeding schedule. (**D**) Loss of body weight in overnight water-deprived animals before having access to water and chow pellets. No differences between experimental groups. Data are presented as mean ± SD. For statistical details see **Statistical Summary Table**.

Figure 1









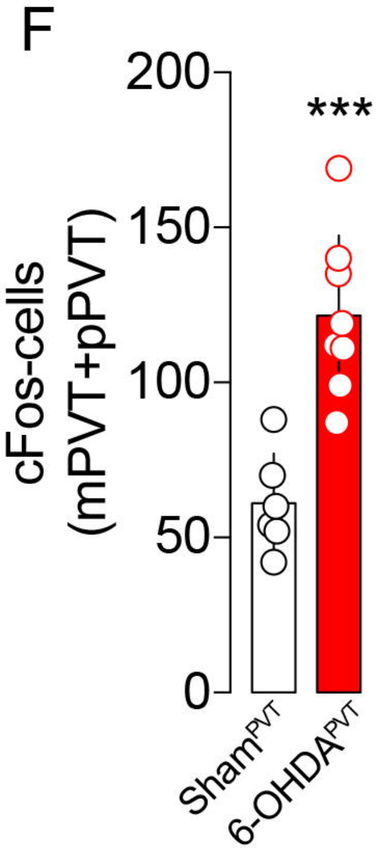


Figure 2

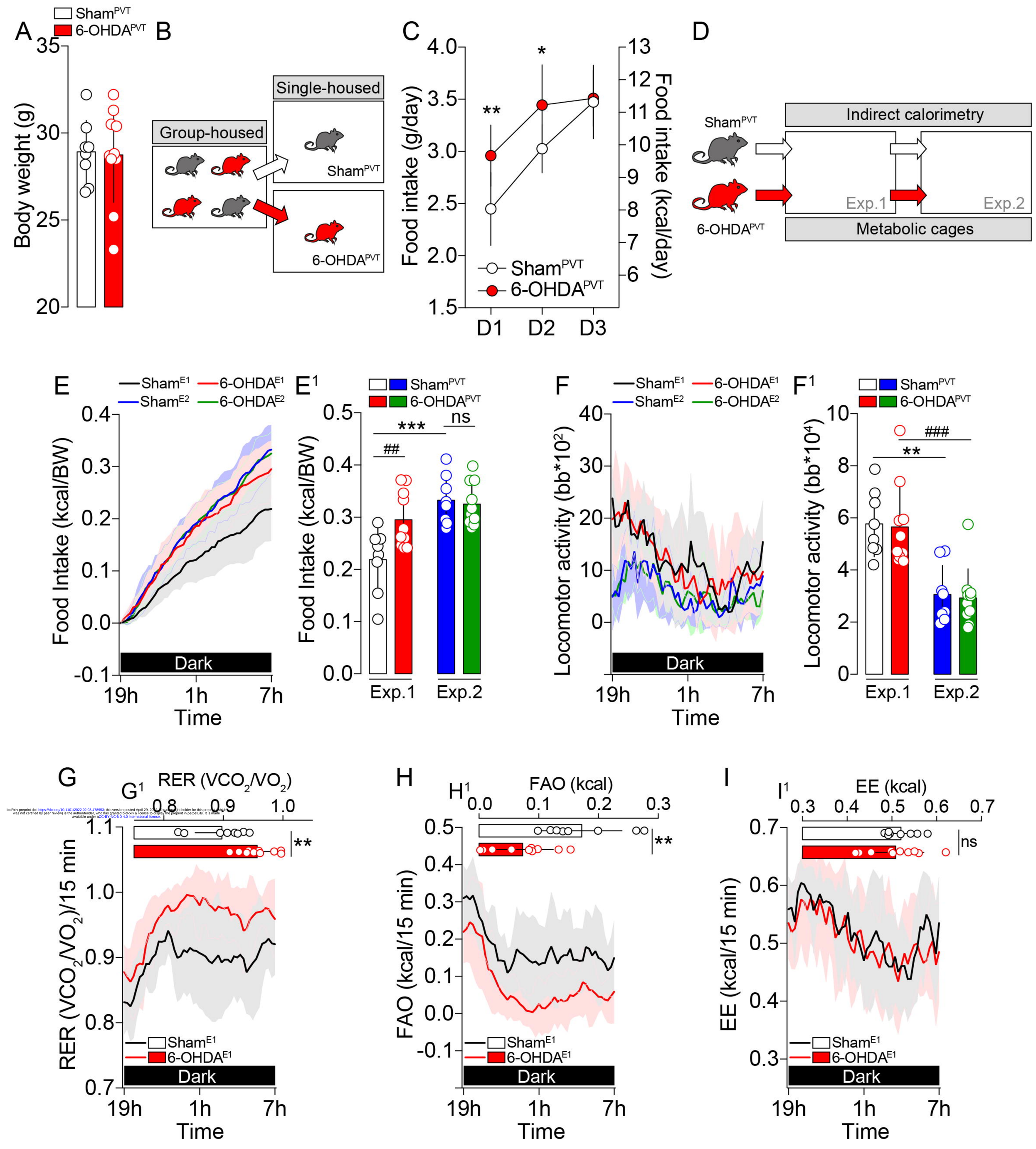
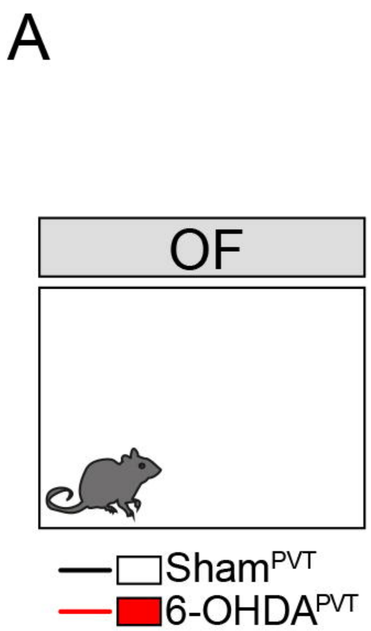
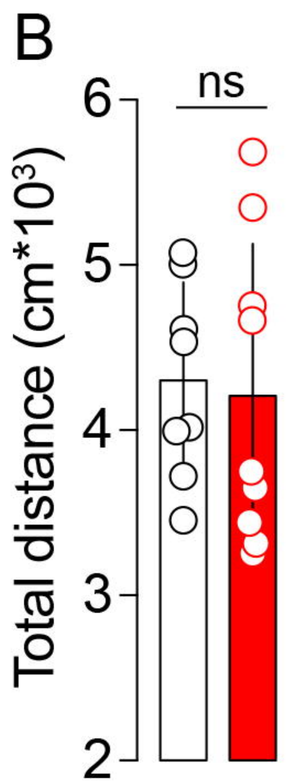
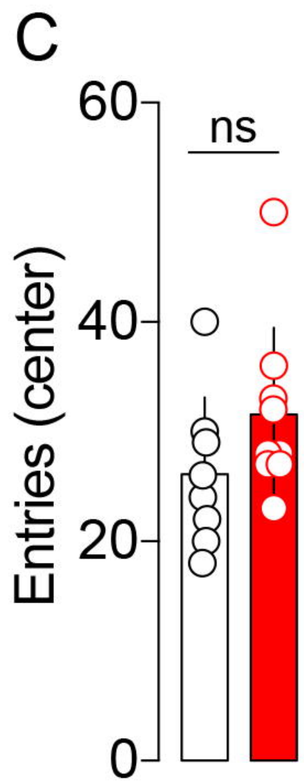
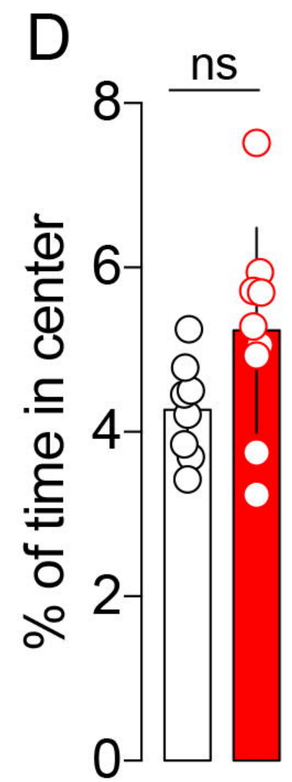


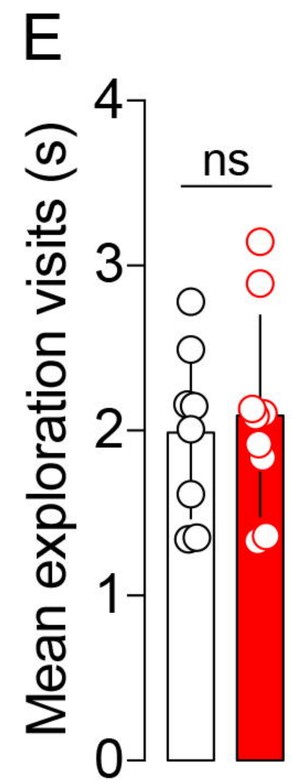
Figure 3

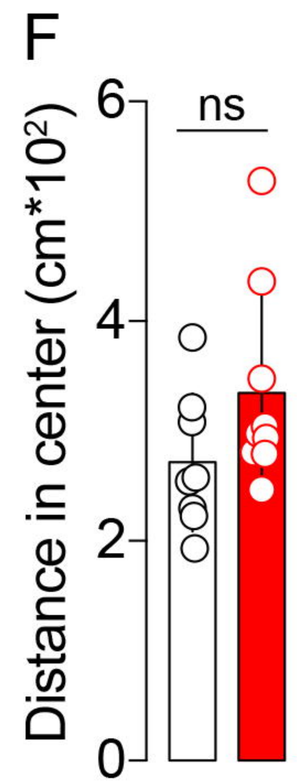


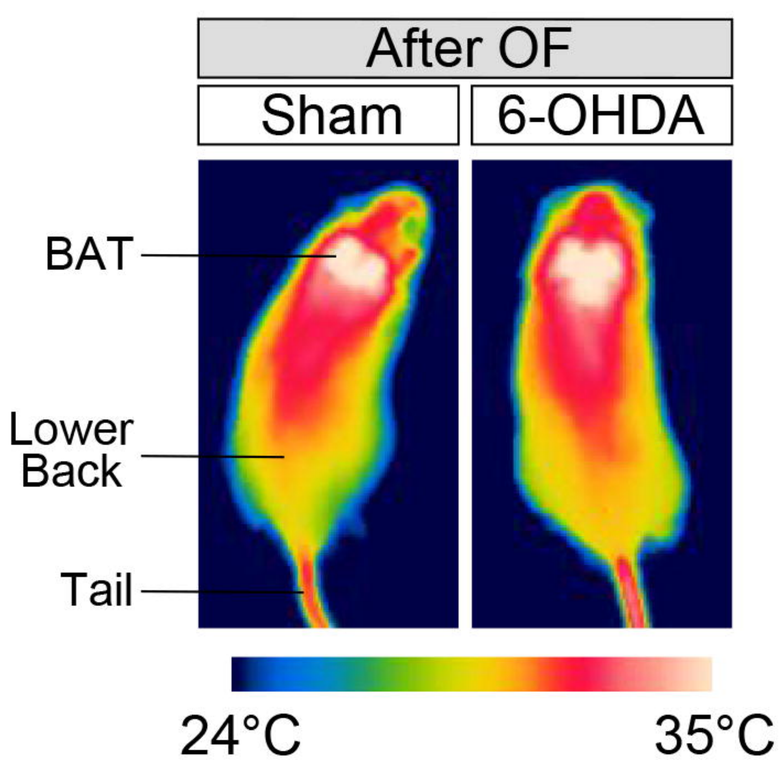


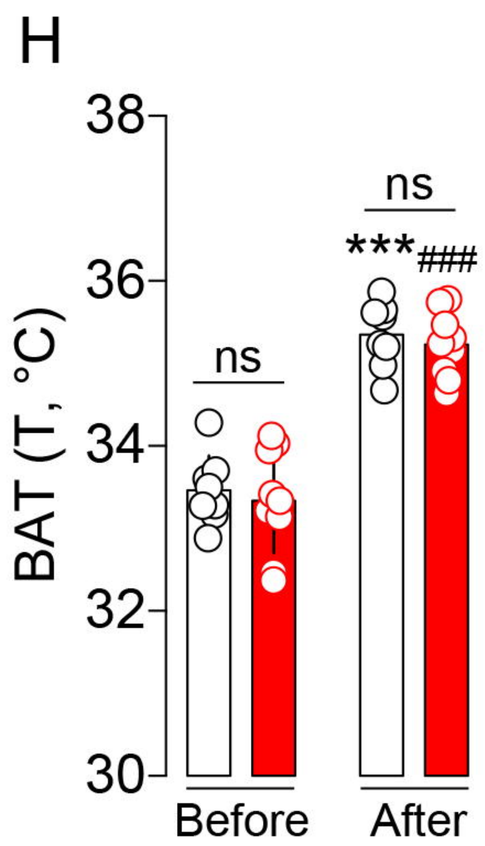


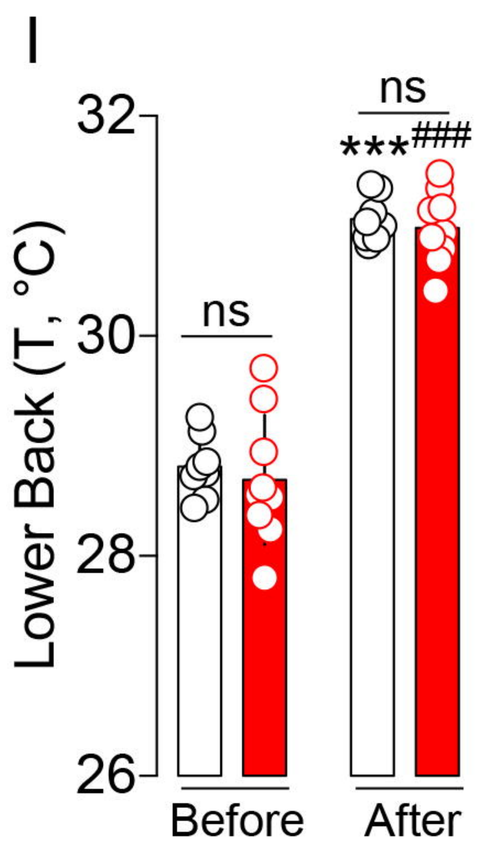












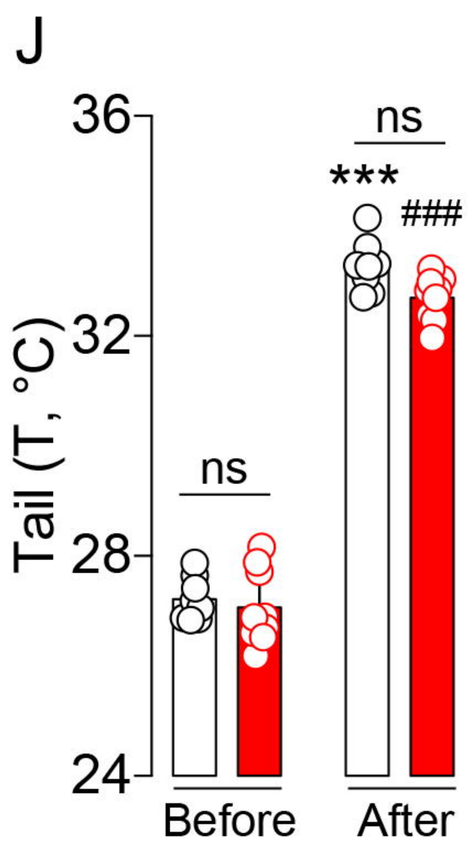
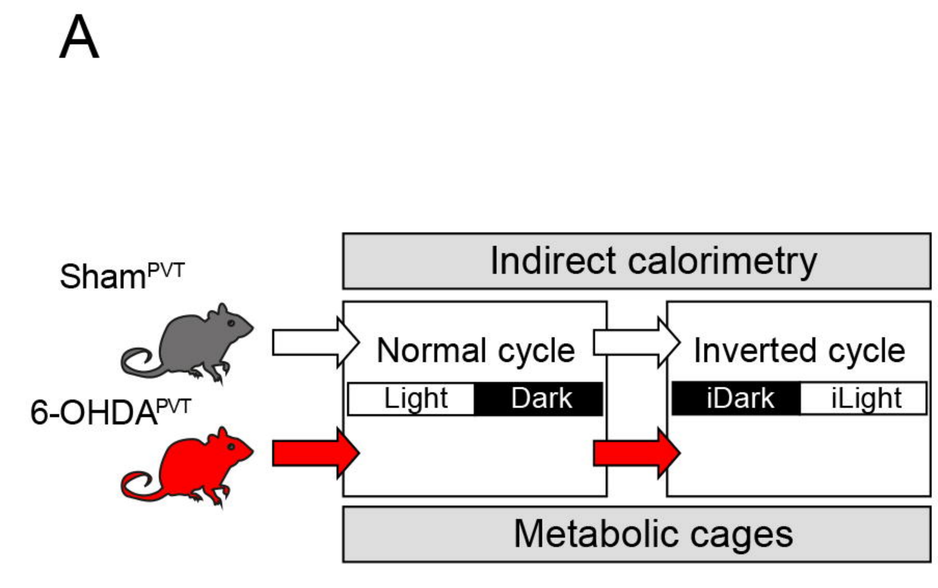
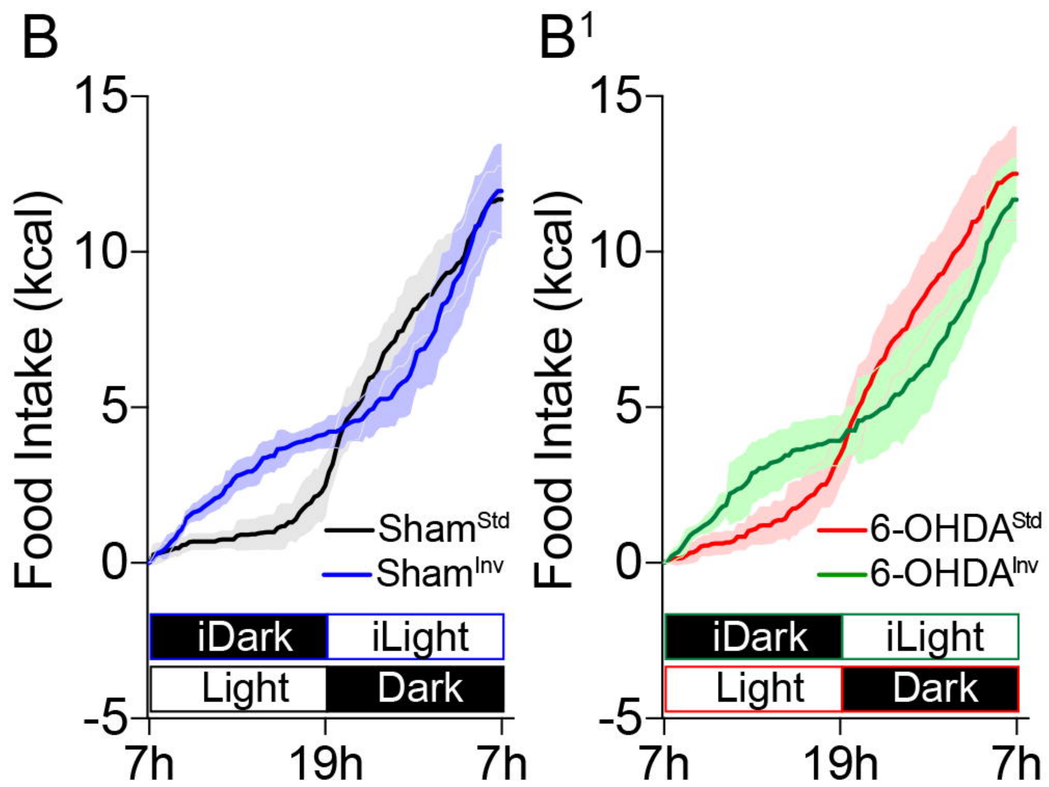
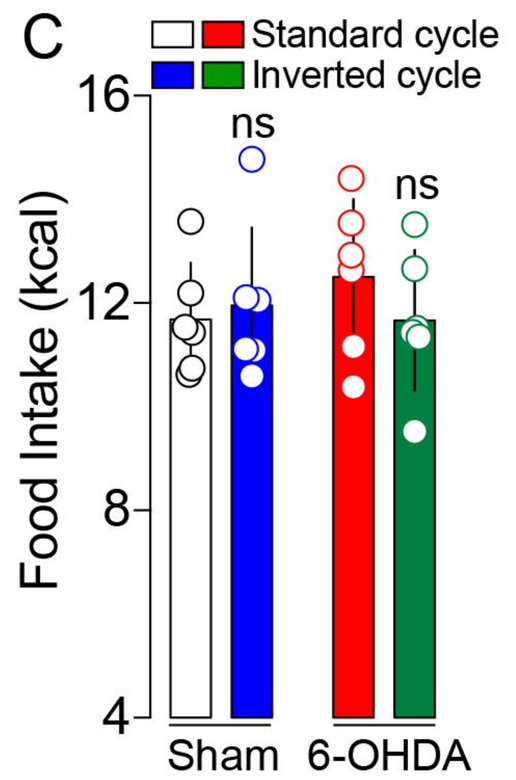
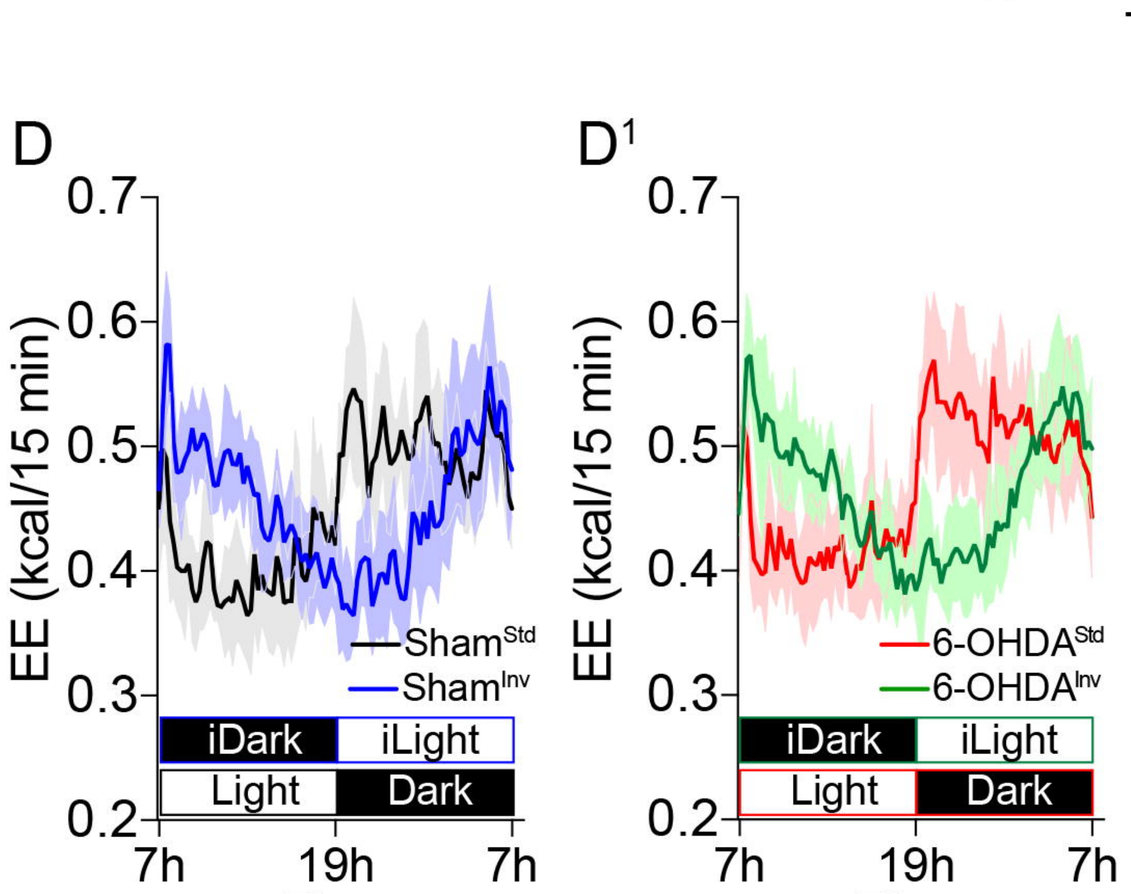


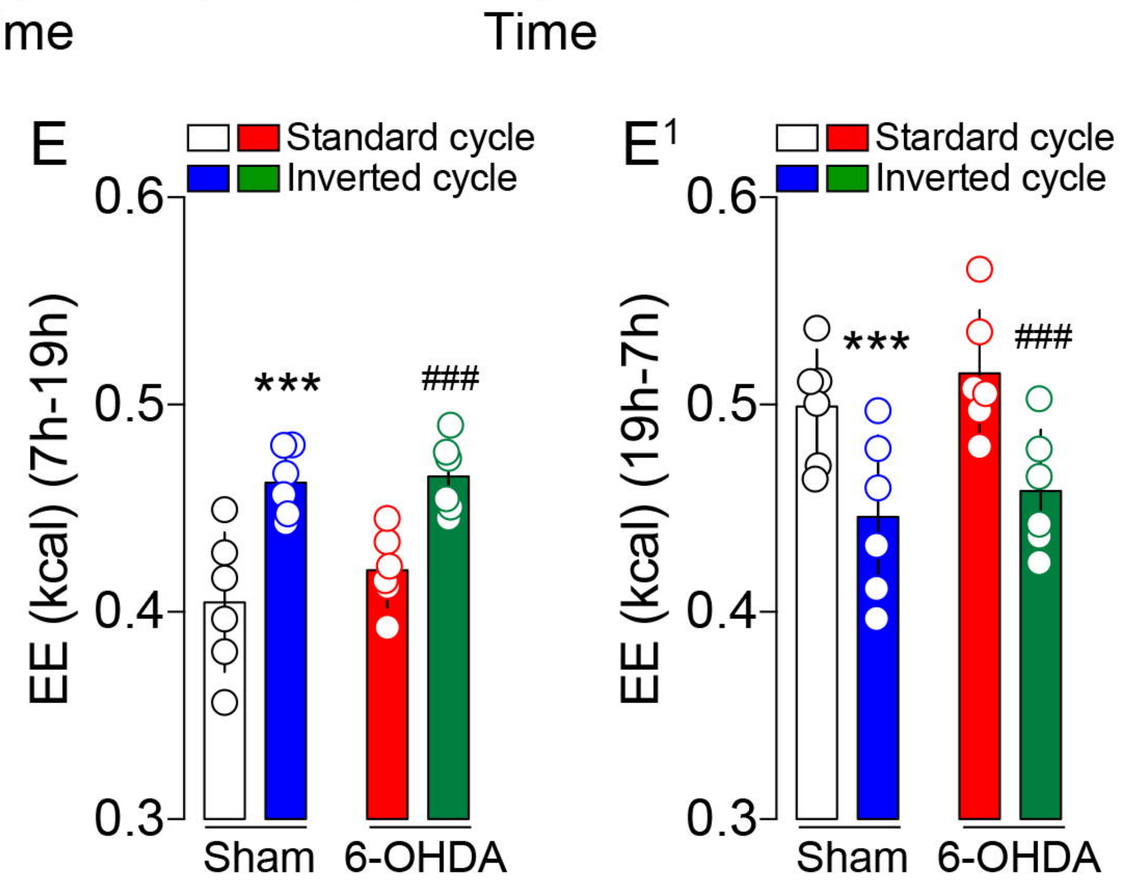
Figure 4

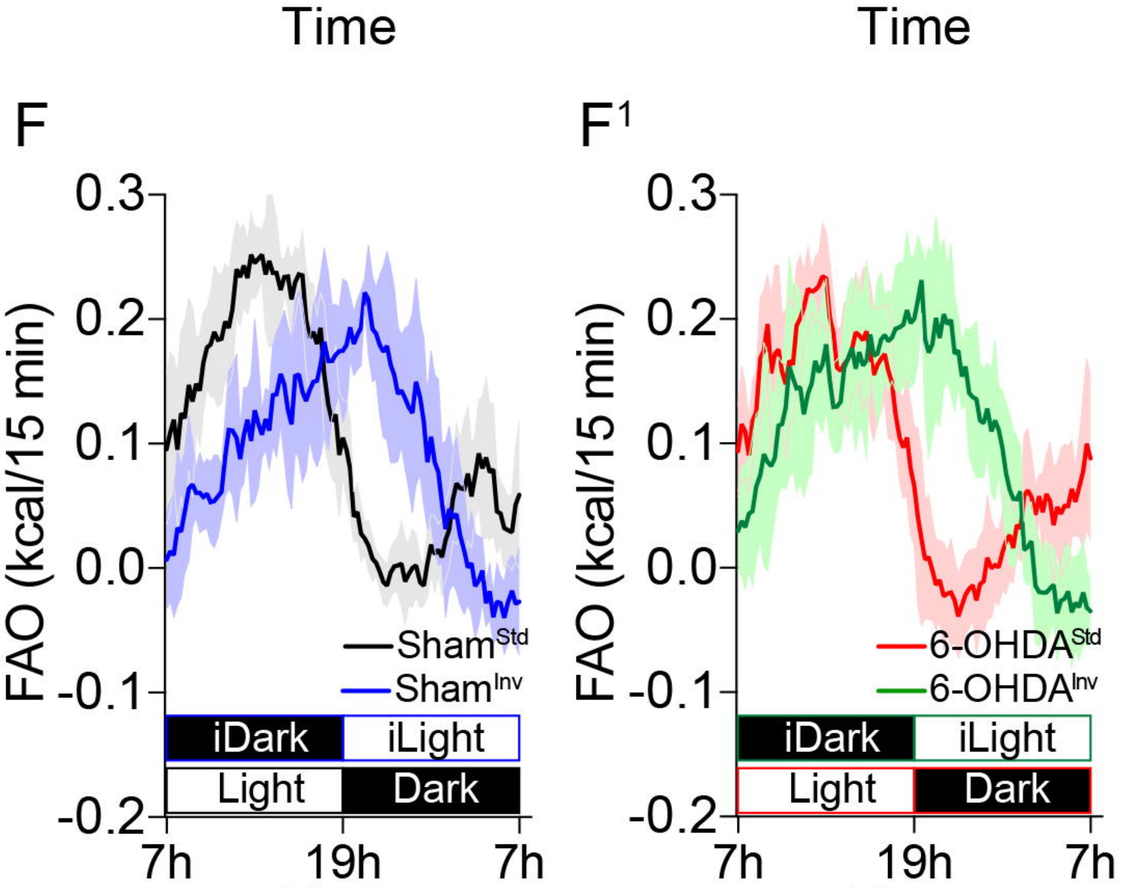


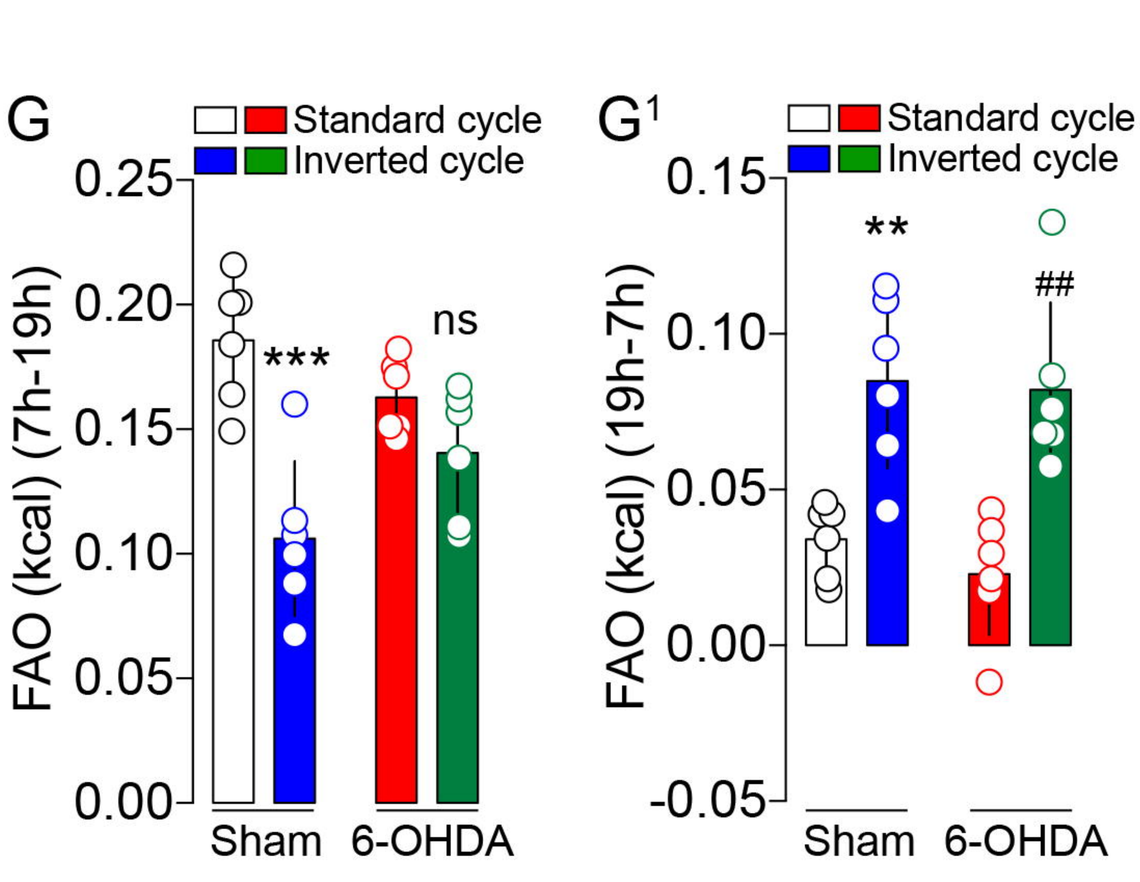


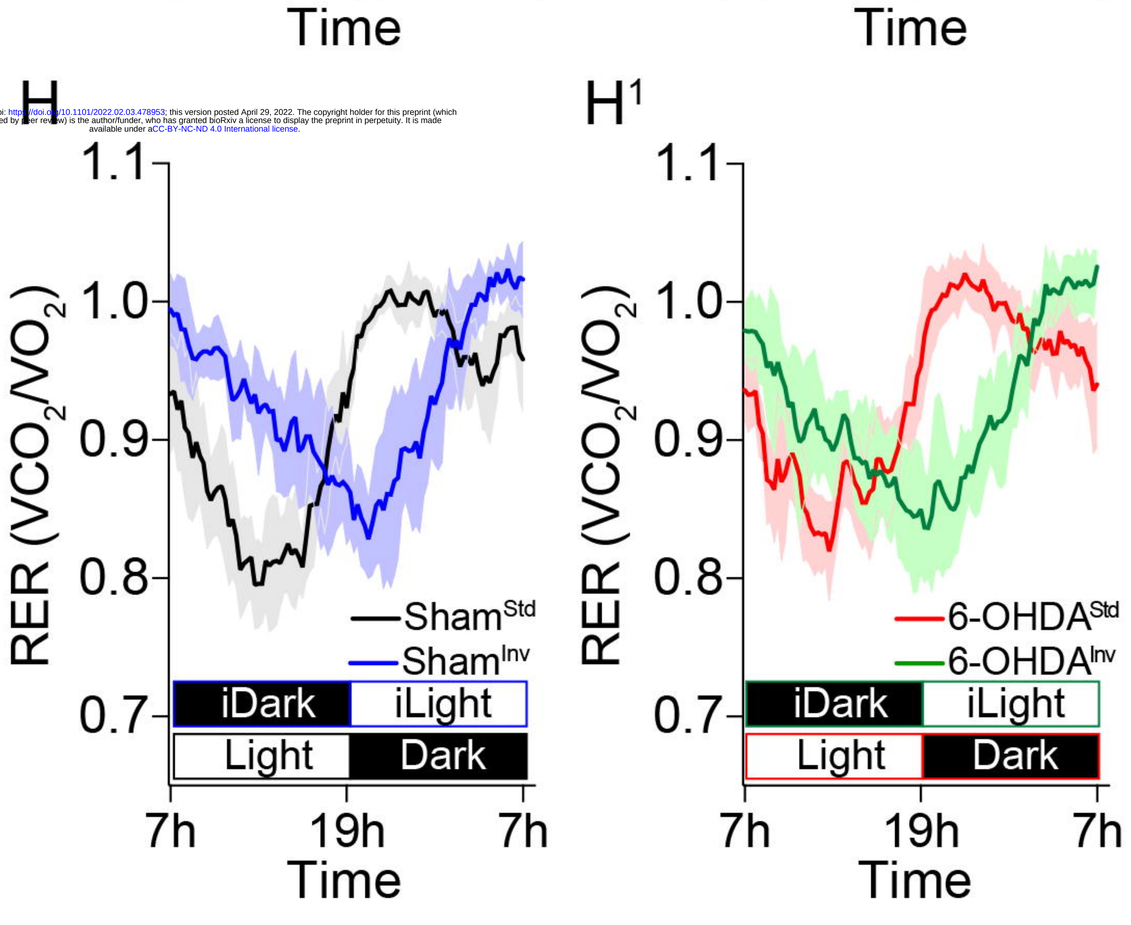












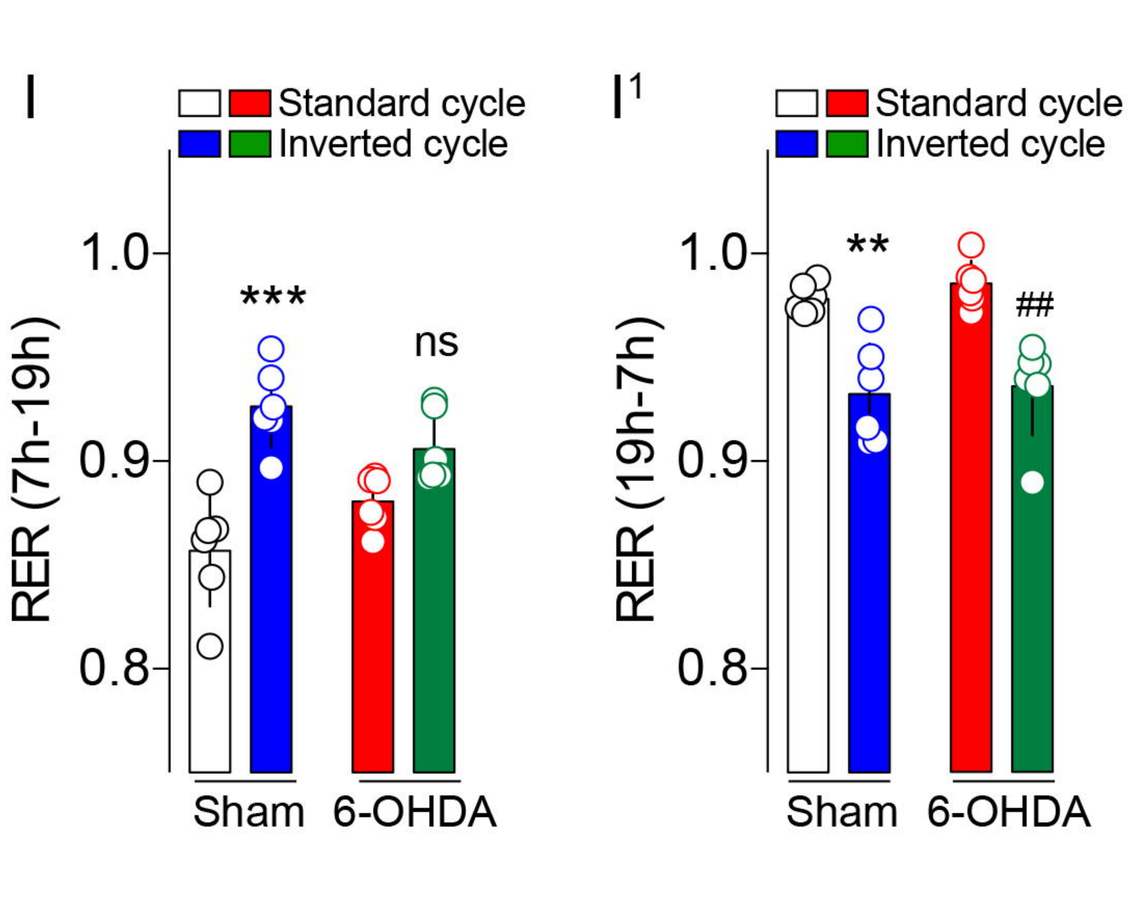
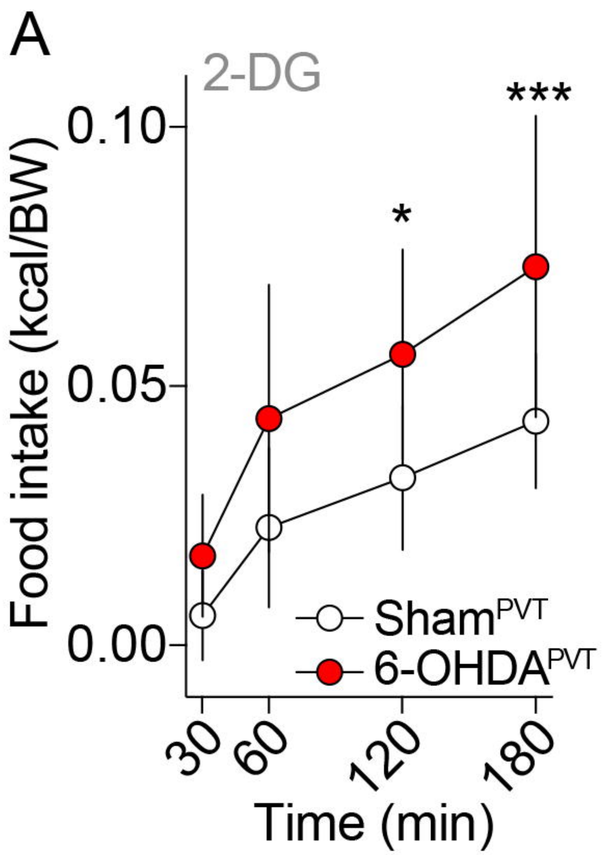
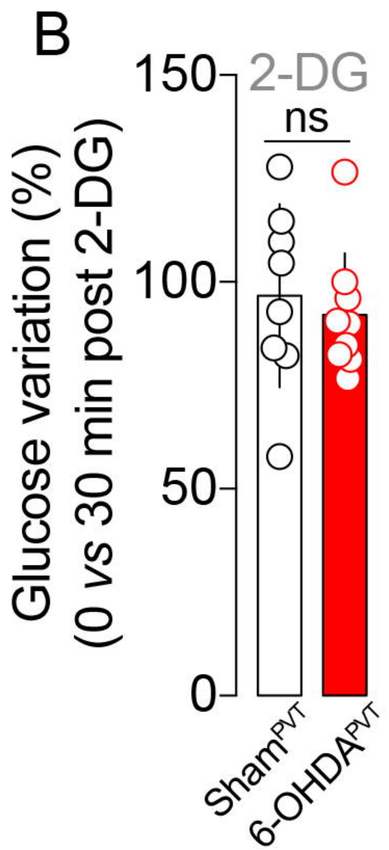
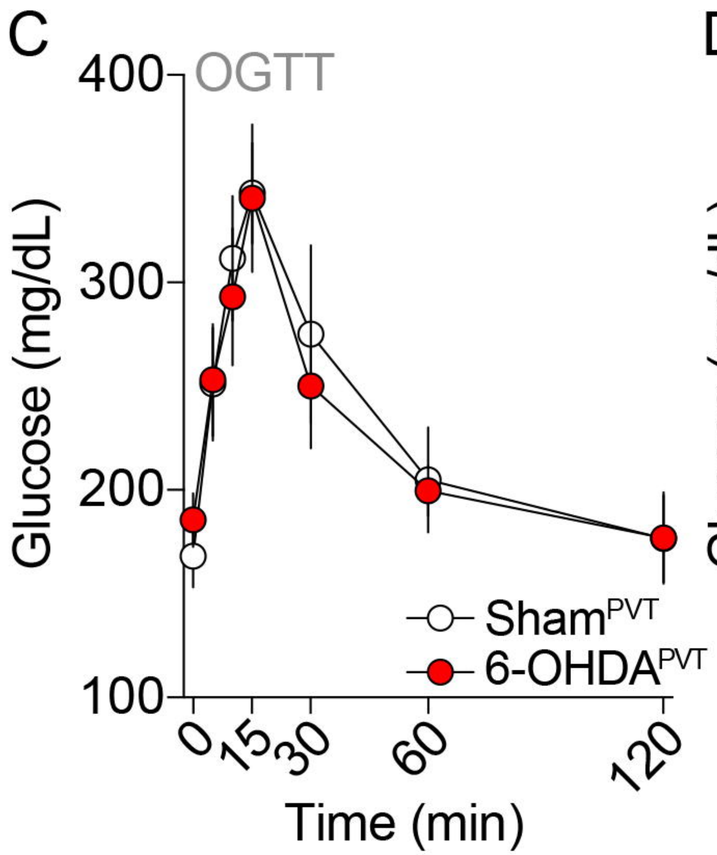


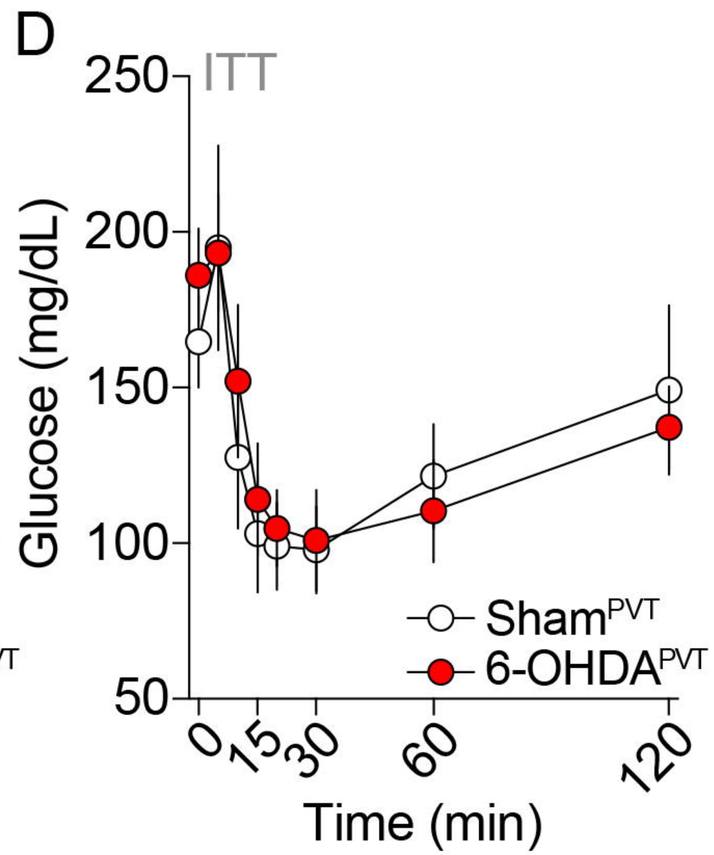
Figure 5

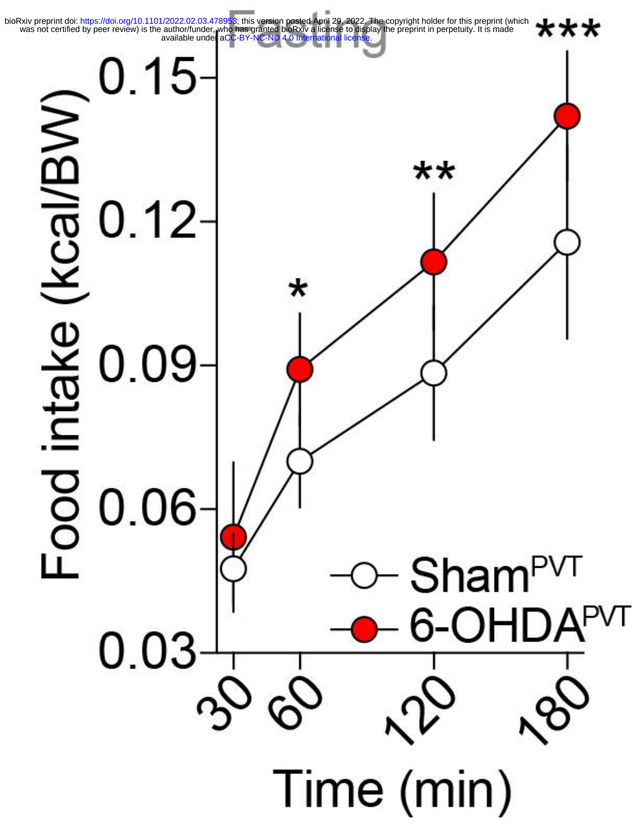
Figure 6

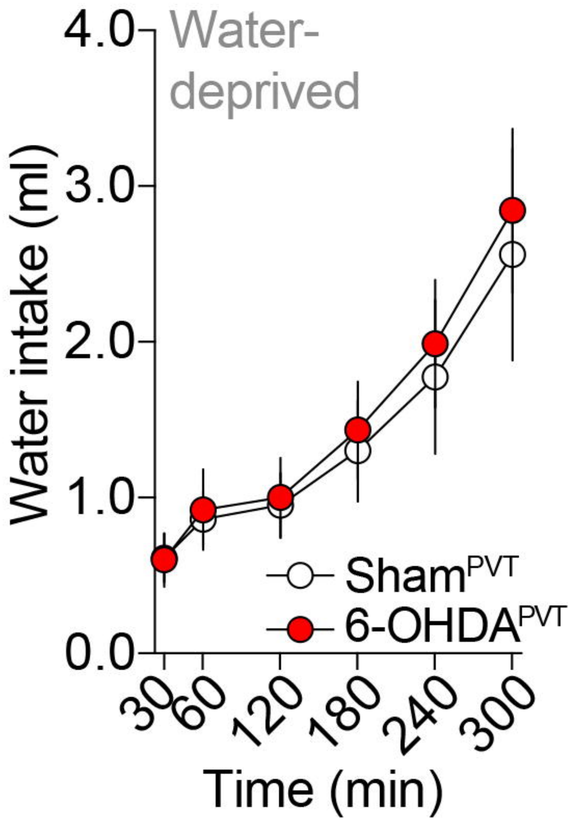


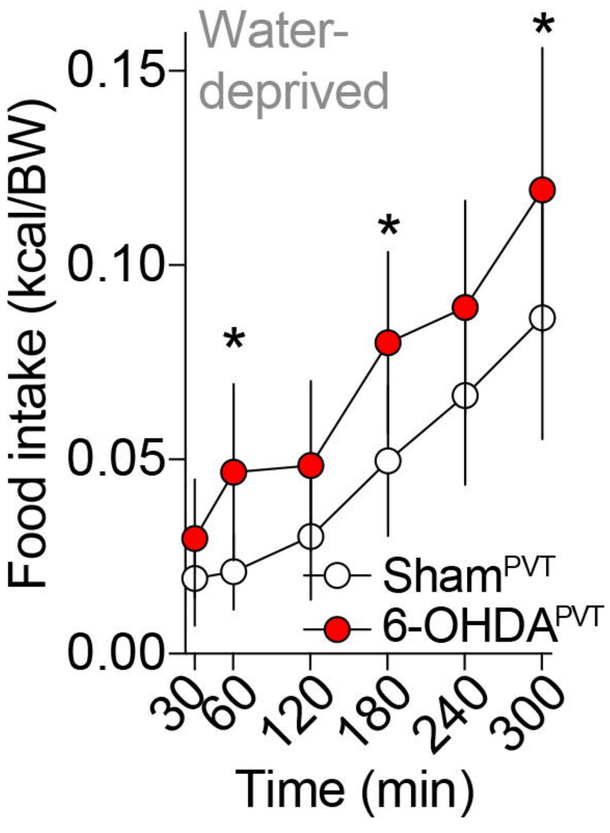












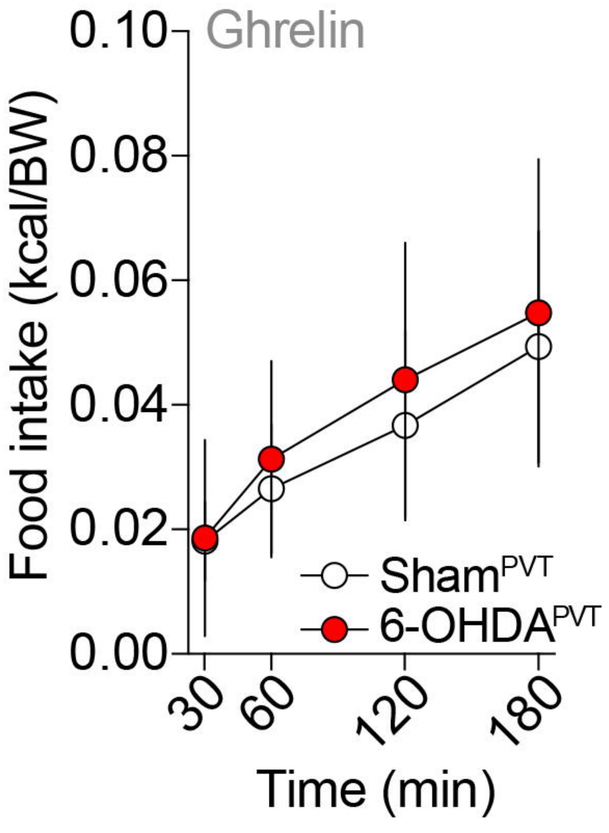


Figure 7