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Multistability and transitions between spatiotemporal patterns through versatile Notch-Hes signaling

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9 Abstract

The Delta-Notch-Hes signaling pathway is involved in various developmental processes ranging from the 10 formation of somites to the dynamic fine-grained patterns of cell types in developing or regenerating tissues. 11 Such broad patterning capabilities rely in part in the versatile and tunable dynamics of the Notch-Hes 12 feedback circuit eliciting both pulsatile and switching behaviors. This raises the theoretical issue of which 13 specific spatiotemporal features emerges from lateral inhibition between cells that can display and transit 14 between steady, oscillatory and bistable regimes. To address this issue, we consider a discrete cell lattice 15 model where intracellular dynamics is described by a phase-like variable and displays a typical cross-shaped 16 phase diagram. Model analysis identifies how the existence and stability of many spatially-inhomogeneous 17 and temporally-synchronized patterns depends on key intracellular and intercellular parameters, which high-18 lights an extensive multistability between those diverse spatiotemporal patterns as well as the existence of 19 multiple robust transition scenarios from temporal patterns to spatial patterns. Such broad repertoire and 20 multistability of spatiotemporal patterns is corroborated using a signaling network model of the Notch-Hes 21 pathway. 22

23 Keywords:

24 Multicellular development, Pattern formation, Oscillations, Cell-fate decision, Multistability

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25 Introduction

The interplay between Notch-mediated intercellular communications and Notch-driven intracellular ac-26 tivities is an important source of self-organized developmental patterns across metazoan tissues (Andersson 27 et al., 2011; Siebel and Lendahl, 2017; Henrique and Schweisguth, 2019; Boareto, 2020). The interaction 28 between Delta ligands and Notch receptors depends on many cell-specific features including the types and 29 the spatial distribution of ligands and receptors (Sprinzak et al., 2010; Shaya et al., 2017; Nandagopal et al., 30 2018). In turn, Notch is prone to inhibit the production of Delta ligands through diverse indirect and direct 31 signaling mechanisms involving the Hes family of proteins (Kageyama et al., 2007; Sjöqvist and Andersson, 32 2019). Hes proteins does not only mediate Notch-dependent repression of Delta or fate-inducing proteins, 33 but its autorepression is a source of intracellular oscillations that is prone to occur prior fate commitment 34 in developing and regenerating tissues (Kageyama et al., 2007, 2018). Notch-Hes pathway also contributes 35 to cell-fate decision programs by antagonizing some fate-promoting factors and reciprocally (Roese-Koerner 36 et al., 2016; Wahi et al., 2016; Sagner et al., 2018). This specific ability of Notch signaling pathway to elicit 37 both oscillatory and switching activities is prone to give rise to diverse and complex spatiotemporal patterns 38 of cell fates within tissues (Biga et al., 2021; Uriu et al., 2021), whose repertorie has yet to be fully explored 39 and characterized. 40

From a modeling viewpoint, a primary patterning role of Delta-Notch intercellular coupling is to destabi-41 lize homogeneous states and promote fine-grained patterning through a spatial symmetry-breaking process 42 triggered by the mutual inhibition between nearest-neighboring cells (Collier et al., 1996). This seminal 43 model has been further refined to illustrate how positive feedback, protrusions, signaling crosstalks or cell 44 division can modulate the relative stability or occurrence of diverse -periodic or aperiodic- spatial patterns 45 (Wearing et al., 2000; Hunter et al., 2016; Hadjivasiliou et al., 2016). In developmental contexts where 46 Notch-Hes signals contribute to cellular oscillations, various models, using a discrete or continuous descrip-47 tion of tissues and using a phase-like or biochemical description of oscillators, have been used to capture 48 the emergence of patterns such as traveling waves (Murray et al., 2011, 2013; Jörg et al., 2015; Tomka 49 et al., 2018), antiphase synchrony (Lewis, 2003; Wang et al., 2011) or dynamic clusters (Biga et al., 2021). 50 However, a consistent theoretical framework is still lacking to study the coexistence and transition between 51 synchronized patterns of oscillatory cells and spatial patterns of bistable cells in various developmentals 52 contexts. This issue of dynamical routes toward tissue patterning has been nevertheless investigated for 53 some specific developmental contexts and modeling settings (Owen, 2002; Plahte and Øyehaug, 2007; Fuji-54 moto et al., 2008; Koseska et al., 2010; Suzuki et al., 2011; Murray et al., 2011; Pfeuty and Kaneko, 2014; 55 Stanoev et al., 2021), including cell type-specific regulatory network models (Pfeuty, 2015; Keskin et al., 56 2018; Tiedemann et al., 2017). 57

In this study, we develop a modeling framework that combines minimal modeling and signaling network modeling in order to study developmental transitions between temporal to spatial patterns as those

driven by Notch-Hes pathway. First, we introduce an effective model of Notch-Hes intracellular dynamics 60 whose low-dimensional parameter and state spaces eases theoretical analysis of spatiotemporal dynamics 61 of cell population. Under some assumption about intercellular coupling and lattice topology, the stability 62 analysis of several archetypical classes of spatiotemporal patterns reveals the existence of signal-dependent 63 multistability between synchronization and inhomogeneous stationary states states. Biological implications 64 of multistable patterns are investigated at the level of developmental transition scenarios from temporal to 65 spatial patterns or by assessing the validity of the theoretical results in simulations of more realistic systems 66 biology models. 67

68 Results

⁶⁹ A minimal model of oscillatory-bistable cells coupled through lateral inhibition

Previous models of the intracellular Notch-Hes pathway frequently display both oscillatory and bistable 70 behaviors depending on some key parameters (Agrawal et al., 2009; Goodfellow et al., 2014; Pfeuty, 2015), 71 typically organized as a typical cross-shaped phase diagram where the transition between oscillation to 72 bistability can occur through diverse codimension-2 bifurcation (Boissonade and De Kepper, 1980; Pfeuty 73 and Kaneko, 2009). To further simplify the description of single-cell dynamics featured with oscillation 74 and bistability, we propose an effective one-dimensional model adapted from the so-called *theta* model 75 whose dynamics is described by a single angular variable. Initially introduced as the canonical model for a 76 neuron which undergoes a saddle-node on invariant circle (SNIC) bifurcation as external current increases 77 (Ermentrout, 1996; Laing, 2016), the model can be slightly modified to describe a transition from oscillation 78 to bistability through a two-SNIC bifurcation: 79

$$\frac{d\theta}{dt} = 1 - \cos\left(2\theta\right) - S_D\left(1 + \cos\left(2\theta\right)\right) + S_N\cos\left(\theta\right) \equiv f(\theta, S_D, S_N) \tag{1}$$

The phase diagram exhibits monostable, bistable and oscillatory domains which are symmetrically organized as function of S_D and S_N (Fig. 1A). The existence of and transition between these dynamical states are easily explained by the manner how the 2π periodic velocity field changes with S_D and S_N (Fig. 1(B)). Of note, the reduced form $\frac{d\theta}{dt} = 1 - \cos(2\theta)$ for $S_D = S_N = 0$ corresponds to the codimension-2 two-SNIC bifurcation for the transition between oscillations and bistability.

In the context of Notch-driven developmental patterning, the phase of the angular variable represents high/low value of Notch and Hes for $\theta \sim \pi/0$. While S_N represents Notch signaling cues, S_D represents other signaling cues (such as FGF or retinoic acid) or epigenetic cues (such as Id), which are known to interfere with intracellular oscillations and eventually promote stable cell-fate choice. The manner how S_N of the receiver cell *i* depends on the state θ of the sender cells *j* must be also 2π -periodic and is chosen as following:

$$S_N = S_{N0} + \gamma \cos\left(\theta_j (t - \tau_j)\right),\tag{2}$$



Figure 1: A low-dimensional model of oscillatory/bistable single-cell dynamics (A) Phase diagram in the signal parameter space S_D and S_N . (B) Vector fields associated with the regime of oscillations (left), bistability (right), monostability (up and down) and the co-dimension 2 saddle-node bifurcation (center). (C) Signal-driven transition from a single-cell oscillations to distinct steady states depending on signal timing

to implement the lateral inhibition property: the coupling term $\gamma \cos(\theta_i) \cos(\theta_j)$ becomes negative and stabilizing for different cell states ($\theta_j = \theta_i + \pi$), and positive and destabilizing for same cell states ($\theta_j = \theta_i$). The delay τ represents the signaling delays associated to this lateral inhibition.

⁹⁴ The population system can thus be written without loss of generality as

$$\frac{d\theta_i}{dt} = f(\theta_i, S_D, S_{N0}) + \Gamma/N\cos(\theta_i) \sum_{j=1,N} J_{ij}\cos\left(\theta_j(t-\tau)\right).$$
(3)

To account for the juxtacrine communication of Delta-Notch, we consider a coupling matrix $J_{ij} = 1$ for the nearest-neighbor cells and zero otherwise. The global coupling strength $\Gamma = N\gamma$ with a number of neighboring cells of N = 2 for 1D array, N = 8 for a square lattice or N = 6 for an hexagonal lattice.

98 Multistability between synchronization states and inhomogeneous steady states

A comprehensive analysis of the spatiotemporal patterning in the cell population model (Eq. (3)) is 99 made possible by the relatively low number of control parameters (Fig. 2(A)). The property of intercellular 100 coupling is monitored at the level of the coupling strength Γ and delay τ . In addition, the symmetric phase 101 diagram as function of the two parameters S_D and S_N (Fig. 1(A)) provides also the possibility to study 102 how spatiotemporal patterning depends on qualitative change in single-cell dynamics through continuous 103 parameter changes. To further simplify the analysis, we consider a ring lattice topology with homogeneous 104 signaling $(S_{D,i} = S_D \text{ and } S_{N0,i} = S_{N0} = 0)$, which left us with S_D , Γ and τ as the main control parameters 105 (Fig. 2(A,B)). 106

¹⁰⁷ A basic class of non-stationary pattern are phase-locked synchronization states (PSS) where neighboring ¹⁰⁸ oscillatory cells displays a constant phase shift ψ or time shift $\Delta = \psi \frac{T}{2\pi}$. $\Delta = 0$ and T/2 correspond to ¹⁰⁹ in-phase and anti-phase synchronized states while other values can be seen as uniform traveling (or rotating) ¹¹⁰ waves. With a one-dimensional ring topology, the phase-locked condition reads $\theta_{i+1}(t) = \theta_i(t + \Delta)$ which 111 can be replaced in Eq. (3) to obtain:

$$\frac{d\theta_i}{dt} = f(\theta, S_D, 0) + \Gamma \cos\left(\theta_i\right) \left[\cos\left(\theta_i(t - \tau - \Delta)\right) + \cos\left(\theta_i(t - \tau + \Delta)\right)\right]/2.$$
(4)

Defining the *T*-periodic delay function $H_u(\theta(t)) = \theta(t) - \theta(t-u)$ (that satisfies $H_0 = 0$ or $H_{u+v} = H_u + H_v$), trigonometric relations can then be used to obtain:

$$\frac{d\theta_i}{dt} = f(\theta_i, S_D, 0) + \Gamma \cos\left(\theta_i\right) \cos\left(H_\tau(\theta_i)\right) \left(\cos\left(\theta_i\right) \cos\left(H_\Delta\right) + \sin\left(\theta_i\right) \sin\left(H_\Delta\right)\right).$$
(5)

In the particular cases of in-phase synchronized solution with no coupling delay ($\Delta = \tau = 0$) and anti-phase synchronized solution with half-period delay ($\tau_{\Delta} = \tau = T/2$), Eq.(5) reduces to $\dot{\theta} = f(\theta, S_D - \Gamma/2, 0)$ for which a periodic solution exists only for $S_D < \Gamma/2$ (Fig. 1(A)) defining a maximal bound for the existence of PSS (Fig. 2(B)).

Stability analysis of phase-locked solutions $\theta_{i+1}(t) = \theta_i(t + \Delta)$ of identical oscillators could be done by 118 linearizing around the periodic solution manifold and compute Floquet multipliers or exponents. We use 119 a more heuristic approach that is to simulate the response of the full system to small spatially-periodic 120 transverse perturbation $\delta\theta$ ($\delta\theta_i(0) = -\delta\theta_{i+1}(0) = \delta\theta_{i+2}(0)$) and evaluate whether such perturbation is 121 relaxed or amplified after one period. Phase diagram shows that relative stability of different phase-locked 122 solutions (e.g., $\Delta = 0, T/2$ and T/25) essentially depends on coupling delays such that phase multistability 123 can naturally occur for some values of coupling strength and delays (Fig. 2(B)). Besides synchronization 124 states, incoherent state attractors are also observed (lower left panel of Fig. 2(C)) but are beyond the scope 125 of this study. 126

This system also displays stationary spatial patterns such as spatially periodic patterns in which cells settle in two states $\theta_{i=1,2}$ with a fixed proportion $k_{i=1,2}$ (i.e., order parameters) of neighbors in different states. In case of a square lattice, $k_{1,2} = 0.75$ for a 1-cell stripe pattern, $k_{1,2} = 0.5$ for a 1-cell or 2-cell checkerboard pattern, $k_{1,2} = \{0.125, 1\}$ for the spot/gap pattern and $k_{1,2} = \{0.375, 0.75\}$ for the 2-cell/1-cell stripe pattern. In case of a hexagonal lattice, $k_{1,2} = 2/3$ for a 1-cell stripe pattern and $k_{1,2} = \{1/3, 1\}$ for the spot/gap pattern. For a one-dimensional lattice, $k_{1,2} = 1$ for the 1-cell stripe pattern. For any of these patterns, steady-state solution of Eq. (3) can be casted into two equations associated to $d\theta_{1/2}/dt = 0$ as,

$$F(\theta_{1/2}, S_D, S_{N0}) + \Gamma \cos\left(\theta_{1/2}\right) \left((1 - k_{1/2}) \cos(\theta_{1/2}) + k_{1/2} \cos\left(\theta_{2/1}\right) \right) = 0.$$
(6)

A particular steady-state solution corresponds to a saddle-node instability for each cell type, which coincides with the appearance or disappearance of an inhomogeneous stationary state. Such instability necessarily occurs for $\theta_1 = 0$ and $\theta_2 = \pi$ which are replaced in Eq. (6) to finally obtain a set of relation between $k_{1/2}$, Γ and $S_{D/N0}$ as:

$$-2(S_D - \Gamma(1 - k_1)/2) - \Gamma k_1 + S_{N0} = 0$$
(7a)

$$-2(S_D - \Gamma(1 - k_2)/2) - \Gamma k_2 - S_{N0} = 0$$
(7b)

Summing these equations cancels out S_{N0} to finally derive the critical coupling strength for which the inhomogeneous state is destabilized:

$$\Gamma = \frac{2S_D}{1 - k_1 - k_2} \equiv \Gamma_c \tag{8a}$$

$$S_D = \frac{\Gamma}{2} (1 - k_1 - k_2)/2.$$
(8b)

The above condition for saddle-node instability boundary thus delimits the stability domain of an inhomo-134 geneous steady state (ISS) defined by the order parameter $k_1 + k_2$. On the one hand, spatial patterns for 135 which neighboring cells are in average more similar than different $(k_1 + k_2 < 1)$ are destabilized above some 136 inhibitory coupling strength $\Gamma > \Gamma_c > 0$ when cells are bistable $S_D > 0$, such as the fully homogeneous state 137 $(k_{1/2} = 0)$ destabilized for $\Gamma > 2S_D$ (Fig. 2(B)). On the other hand, spatial patterns for which neighboring 138 cells are in average more different than similar (i.e., $k_1 + k_2 > 1$) are always stable for $S_D > 0$ ($\Gamma_c < 0$) 139 and are stabilized above some inhibitory coupling strength $\Gamma > \Gamma_c = \frac{2S_D}{1-k_1-k_2}$ for oscillatory cells $(S_D < 0)$, 140 supporting a scenario of oscillation amplitude death. Numerical simulations show that lateral inhibition can 141 stabilize non-periodic spatial patterns for which neighboring cells are more different than similar, such as 142 labyrthin pattern for which $k_i = 5/8$ or 6/8 (lower right panel of Fig. 2(C)). 143

These stability properties of phase-locked synchronized states (PSS) and inhomogeneous steady states 144 (ISS) reveal two key complementary features of the spatiotemporal dynamics of oscillatory/bistable cells 145 coupled through delayed inhibition between nearest neighbors. Lateral inhibitory coupling stabilizes ISS 146 even when uncoupled cells oscillate and PSS even when uncoupled cells are in steady states. As a result, the 147 multicellular system exhibits a robust multistability between PSS and ISS especially when uncoupled cells 148 are operating close enough to the transition between oscillation and bistability (i.e., $|S_D| < \Gamma$). Accordingly, 149 multiple stationary or non-stationary attractors can coexist such as those satisfying spatial periodicity 150 $\theta_{i+2}(t) = \theta_i(t)$ (Fig. 2(D)), such that attractor selection would depend on the initial conditions. It is 151 important to note that this extensive multistability property does not occur for negative value of Γ and is 152 therefore mediated by the lateral inhibitory coupling. 153

¹⁵⁴ Pattern selection through transition from temporal to spatial patterns

The class of biological phenomena that motivates the present theoretical study concerns the developmental transition whereby a given population of oscillatory cells gives rise to two subpopulations of well-distinct cell states. In particular, a pending question is the ability of Notch pathways to underlie very diverse



Figure 2: Spatiotemporal dynamics of inhibitory-coupled oscillatory/bistable cells. (A) Cell population model organized as a 1D or 2D periodic lattice with first-neigbor inhibitory coupling. (B) Stability domains of typical phase-locked synchronization states and spatial stationary states as function of S_D and Γ (top: $\tau = 3$), and as function of S_D and τ (bottom: $\Gamma = 0.5$). (C) Example of stable temporal patterns for a 1D lattice with $\Gamma = 0.5$, $\tau = 3$ and $S_D = 0$ (up panels) and of stable spatial patterns for a 2D lattice with $\Gamma = 0.5$ and $S_D = 0$ (Down panels). (D) Example of multistability between phase-locked synchronization states (PSS) and inhomogeneous spatial states (ISS) in the 1D lattice plotted in the phase plane { $\theta_i, \theta_i + 1$ } for $S_D = -0.1$, $\Gamma = 0.5$ and $\tau = 3$.

symmetry-breaking scenarios from temporal to spatial patterns, involving stripe patterns as during somito-158 genesis or salt-and-pepper patterns as during neurogenesis. This issue is addressed in the population model 159 (Eq. (3)) by simulating the spatiotemporal dynamics in response to a temporal increase S_D corresponding 160 to a slow developmental change of signaling or epigenetic cues. According to the repertoire and stability 161 domains of states identified in Fig. 2, the population can be prepared and settled for $S_D(t=0) = S_{D1} < 0$ 162 into diverse spatiotemporal states such as in-phase synchronization, traveling waves and antiphase synchro-163 nization (Fig. 3). By stabilizing these states, intercellular coupling provides robustness to diverse sources 164 of noise, here in the initial conditions and in the signal dynamics $S_D(t)$. Following an increase of S_D , each 165 temporal pattern gives rise to a specific stationary pattern depending on the spatial pattern at the time of 166 the differentiation signal increase and the profile of such increase. First, a synchronous oscillatory pattern 167 translates into a homogeneous stationary pattern as far as $S_{D2} > \Gamma/2$ (Fig. 3(A)). Second, a traveling wave 168 pattern translates into a regular stripes (Fig. 3(B)) produces stripes of tunable size as far as $S_D(t = t_f)$ 169 is sufficiently high to stabilize these stripe patterns. Last, an antisynchronous pattern translates into an 170 inhomogeneous stationnary pattern stripe pattern of one-cell width in 1D (Fig. 3(C)). In these transition 171 processes, the role of lateral inhibition in stabilizing phase shifts between neighboring cells is very effec-172



Figure 3: Robust and tunable transition routes from temporal to spatial patterns. Transient spatiotemporal dynamics from a noisy initial spatial pattern $\theta_i(0)$ for $S_D(t) < 0$ (left panels of A-C) and driven by a noisy temporal pattern $S_D(t) = S_{D1} + (S_{D2} - S_{D1})\mathcal{H}(t) + \zeta(t)$ where $S_{D1} = -0.3$ and $s_{D2} = 0.2$ (uppest panel of A-C). The spatiotemporal dynamics $\cos(\theta_i(t))$ is compared with and without coupling ($\Gamma = 0.5$ for the upper-right panel and $\Gamma = 0$. for the lower-right panel). (A) From a noisy initial conditions, lateral inhibition promotes a robust transition from synchronous oscillations to an homogeneous spatial pattern. (B) From a noisey and spatially-periodic initial conditions, lateral inhibition promotes a robust transition from a traveling wave pattern to a regular stripe pattern. (C) From noisy checkerboard noisy initial conditions, lateral inhibition promotes a robust transition from antisynchronous oscillations to a checkerboard spatial patterns.

¹⁷³ tive to obtain regular and defect-free spatial patterns (compare dynamics with and without coupling in ¹⁷⁴ Fig. 3(A,B,C)).

¹⁷⁵ Multistable spatiotemporal patterns in a systems biology model of Delta-Notch-Hes circuit

To assess the biological relevance of results obtained with effective low-dimensional intracellular dynamics, we develop a systems biology model of Delta-Notch-Hes pathways whose feedback architecture enables a cross-shaped diagram similar to Fig.1(A). Previous single-cell models have first focused on the core autorepression of Hes1 protein (Lewis, 2003) before to incorporate additional set of factors mutually interacting with Notch or Hes, such as Rbpj (Agrawal et al., 2009), miR9 (Goodfellow et al., 2014), Cyclins-Cdks (Pfeuty, 2015) or Neurog (Tiedemann et al., 2017). Such interlocking between a core negative feedback mediated by Hes autorepression, and positive feedback mediated by mutual inhibition between Notch or Hes and other cell fate-specific factors is indeed a common feature in various tissues and cell types (Fig. 4(A)). For instance, members of Hes family proteins mutually antagonize with transcription factors including miR9 or Ascl1 in neural progenitors (Roese-Koerner et al., 2016; Vasconcelos et al., 2016), Olig2 in motoneuron progenitors (Sagner et al., 2018), Neurog3 in multipotent pancreatic progenitor cells (Ahnfelt-Rønne et al., 2012), Myod in myoblast (Lahmann et al., 2019) and Lfng or Mesp2 in presomitic stem cells (Wahi et al., 2016). Although the detailed set of regulatory mechanisms can differ depending on cell type, we can nevertheless develop a generic model where Notch, Hes and Delta concentration variables are supplemented with an antagonistic factor, named Y (Fig. 4(B)), based on the model of Delta-Notch-Hes-miR9 circuit developed in (Pfeuty, 2015):

$$\tau_N \frac{dN}{dt} = \frac{S_N}{1 + \mathbf{S}_N + k_{YN}Y^2} - N \tag{9a}$$

$$\tau_{H_m} \frac{dH_m}{dt} = \frac{k_{NH}N^2}{1 + k_{HH} \left(1 + k_{FH} S_{Dif}\right) H (t - \tau_1)^n + k_{NH}Y^2} - H_m$$
(9b)

$$\tau_H \frac{dH}{dt} = H_m (t - \tau_2) - d_H H \tag{9c}$$

$$\tau_Y \frac{dY}{dt} = \frac{1}{1 + k_{FY} S_{Dif}^2 + k_{HY} H^2} - Y$$
(9d)

$$\tau_D \frac{dD}{dt} = \frac{1}{1 + k_{HD}H^2} - D \tag{9e}$$

$$S_{N,i} = S_{N0,i} + \gamma \sum_{j} D_j(t-\tau) \tag{9f}$$

where N represents Notch activity, H_m and H represent concentrations of Hes mRNA and proteins, D represents Delta ligand and Y represents a fate-specific factor antagonizing with Notch and/or Hes proteins. Regarding signaling cues, S_N represents the extracellular juxtacrine signals associated with Delta-dependent activation of Notch receptor while S_{Dif} represents an extracellular morphogenetic fields associated with relative concentrations of extracellular signals (such as FGF and RA) assumed to promote expression of Y and inhibiting autorepression of H.

Degradation and time-delay parameters of the model (τ_i) are set with agreement with experimental 182 measurements and the approximate 2-hour period of Hes oscillation (Pfeuty, 2015). In the case where 183 the species Y is weakly expressed associated to S_{Dif} low (e.g., low RA and high FGF), the Notch-Hes 184 regulatory module displays oscillations for intermediate Notch signal S_N . In the case where the species 185 Y is highly expressed associated to S_{Dif} high (e.g., high RA and low FGF), the concomittant decrease 186 187 of Hes autorepression leads to a robust bistability due to mutual inhibition between Notch-Hes and Y. Given these two archetypical oscillation and bistable behaviors obtained for low and high S_{Dif} level, the 188 parameters for mutual inhibition between *Hes* and *Y* are then adjusted to obtain a phase diagram similar 189 with the low-dimensional model for the sake of comparison (Fig. 4(C) as compared to Figure 1(A)). Although 190 biochemical and low-dimensional models share a similar cross-shaped phase diagram, it is to mention that 191 the detailed model shows a more complex bifurcation scenario where the transition between oscillation and 192 bistability occurs typically through a concomittant Hopf and saddle-node bifurcation (instead of a two-193 SNIC bifurcation). These fine-grained dissimilarities do not impact the coarse-grained transition behavior 194 whereby increase of S_{Dif} leads to a cellular transition from an oscillatory state toward diverging steady states 195 depending on the relative timing between signal course and oscillatory phase (Figure 4(D) as compared to 196



Figure 4: Oscillation and bistability in a systems-biology model of Notch-Hes signaling. (A) Feedback regulatory circuits involved in Notch/Hes-driven fate decision in various tissue-specific cell types. (B) Schematic network model of the Notch-Hes-Delta circuit based on the common features of cell type-specific circuits in (A). (C) Phase diagram with a transition from oscillation to bistability obtained for the following parameter set: $d_N = 1$, $d_H = d_Y = 2$, $d_{mH} = 10$, $k_{NH} = 10$, $k_{HY} = 17$, $k_{YH} = 7$, $k_{YN} = 7$, $k_{FY} = 0.25$; $\tau_{1,2} = 0.5$ h. (D) Example of a transition from single-cell oscillations to distinc steady states function of the timing of S_{Dif} increase.

197 Figure 1(C)).

To address again the spatiotemporal dynamics of oscillatory/bistable cells coupled through Delta-Notch. 198 we consider a periodic 1D lattice with homogeneous signaling field, in order to define the same setting used 199 for the theoretical analysis of the simple model (Fig.5(A)). Multistability between diverse spatiotemporal 200 patterns are investigated through adiabatic increase of S_{Dif} starting from stable inphase or antiphase syn-201 chronization states for small S_{Dif} values and through adiabatic decrease of S_{Dif} starting from a typical 202 inhomogeneous stationary states of 1-cell period (Fig.5(B)) Simulation shows that Delta-Notch intercellular 203 communications can stabilize (i) inhomogeneous steady states (ISS) even for cells oscillating without cou-204 pling $(S_{Dif} < 0.5)$ and (ii) phase-locked synchronized states (PSS), both synchronous and antisynchronous 205 depending on coupling delay, even for stationary cells without coupling $(S_{Dif} > 0.5)$, indicating a robust 206 multistability between PSS and ISS. for intermediate differentiation signals. Such extensive multistability 207 between stationary and nonstationary patterns is exemplified for some intermediate value of coupling delay 208 $\tau = 0.5$ showing the phase-plane attractors associated with inhomogeneous stationary state and phase-locked 209 synchronization states of various phase shifts (Fig4(C,D)). 210

211 Discussion

The present modeling study investigates the transition properties between temporally-synchronized and spatially-inhomogeneous patterns driven by the Delta-Notch-Hes signaling axis. To address this issue, we introduced a particular class of discrete cell lattice model where single-cell dynamical repertoire comprises both autonomous oscillatory and switching behaviors. The model could recapitulate a wide spectrum of spatiotemporal behaviors that has been reported over last decades using distinct classes of theoretical models of lateral inhibition. Lateral inhibition itself is a notorious mechanism to generate diverse, and eventually



Figure 5: Multistability between temporally-synchronized and spatially-inhomogeneous states mediated through **Delta-Notch-Hes intercellular/intracellular signaling.** (A) One-dimensional ring lattice of Delta-Notch coupled cells. (B) Stability diagram of in-phase (up/blue) or anti-phase (down/turquoise) synchronization state and inhomogeneous stationary state (red) as function of S_{Dif} where $S_{N0,i} = 0.2$, $\Gamma = 1$ and $\tau = 0$ or 1h (i.e., T/2). (C) Multistability and coexistence of multiple archetypical (i.e., 2-cell periodic) attractors represented in a relevant section of the state space. Parameters are $S_{N0,i} = 0.2$, $S_{Dif} = 0.5$, $\Gamma = 1$ and $\tau = 0.5$ h. (D) Space-time representation of attractors in (C).

multistable, inhomogeneous fine-grained patterns (Collier et al., 1996; Owen et al., 2000; Hadjivasiliou et al., 218 2016; Hunter et al., 2016), but can also give rise to diverse coherent and synchronization states through time-219 delayed juxtacrine coupling between oscillatory cells (Lewis, 2003; Morelli et al., 2009; Murray et al., 2011; 220 Wang et al., 2011; Jörg et al., 2015; Tomka et al., 2018). Furthermore, repulsive coupling between synthetic 221 oscillators has been shown to generate a wide range of collective regimes including synchronization states and 222 inhomogeneous stationary states (Ullner et al., 2008; Koseska et al., 2010). By capturing those very diverse 223 collective behaviors as function of a very few parameters $(\tau, S_D, S_{N0}, \Gamma)$, the proposed low-dimensional 224 model proves well-suited to qualitatively study a wide range of developmental processes based on versatile 225 single-cell dynamics, in support or in parallel to more detailed signaling network models. 226

The extensive multistability reported in the present study builds on several mechanisms and has some 227 implications, notably for pattern selection in different contexts or between functional and spurious ones 228 (Morelli et al., 2009; Palau-Ortin et al., 2015; Pfeuty and Kaneko, 2016; Uriu et al., 2021). Mulistationarity 229 between spatial patterns typically arise from the combination of intrinsic bistability of cells and the in-230 tercellular positive feedback through Delta/Notch-mediated lateral inhibition (Collier et al., 1996; Wearing 231 et al., 2000). Multistability between synchronization states is also frequently observed in coupled oscillator 232 systems (Crowley and Epstein, 1989; Morelli et al., 2009; Williams et al., 2013). Less common is the here-233 characterized multistability between diverse temporally-synchronized and spatially-inhomogeneous states, 234 which entails the existence of many transition paths from a particular temporal pattern to a particular 235 spatial pattern. In addition, signal-driven transition from a stable spatiotemporal pattern toward a given 236 spatial pattern can operate as a both robust and flexible canalization process to the desired multicellular 237

outcome. This mechanism is complementary to pattern selection and fine-tuning mechanisms mediated 238 by the developmental modulation of Delta-Notch interactions and/or Notch-Delta intracellular signaling 239 (Formosa-Jordan and Ibañes, 2014; Palau-Ortin et al., 2015; Boareto et al., 2015; Sato et al., 2016; Luna-240 Escalante et al., 2018). This issue of pattern selection is illustrated by the differences observed during 241 neurogenesis and somitogenesis where a similar circuit gives rise either to dynamic salt-and-pepper patterns 242 or traveling waves (Liao and Oates, 2017; Kageyama et al., 2018). It is possible that the desynchronizing 243 action of asymmetric division in the neurogenic case and the synchronizing action of Wnt and Fringe in the 244 somitogenic case could explain diverging spatiotemporal trajectories and outcomes. Another set of differ-245 ential constraints would relate to the existence of pre-existing boundaries, axis and gradients, for instance 246 related to the layered structure of the developing neural systems or the antero-posterior axis in the somites. 247 In any case, the effective phase-like model could easily incorporate additional constraints at the levels of 248 intercellular coupling or tissue topology to investigate more specific developmental dynamics and reveals 249 how a similar signaling circuitry could give rise to such diversity of developmental patterning processes. 250

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