

Sulfate groups position determines the ionic selectivity and syneresis properties of carrageenan systems

Said Elmarhoum, Sophie Mathieu, Komla Ako, William Helbert

▶ To cite this version:

Said Elmarhoum, Sophie Mathieu, Komla Ako, William Helbert. Sulfate groups position determines the ionic selectivity and syneresis properties of carrageenan systems. Carbohydrate Polymers, 2023, 10.1016/j.carbpol.2022.120166. hal-03812613

HAL Id: hal-03812613

https://hal.science/hal-03812613

Submitted on 12 Oct 2022

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

- 1 Sulfate groups position determines the ionic selectivity and syneresis
- 2 properties of carrageenan systems

3

- 4 Authors and affiliations:
- 5 Said ELMARHOUM ^a, Sophie MATHIEU ^b, Komla AKO * ^a, William HELBERT ^b
- 6 a) Univ. Grenoble Alpes, CNRS, Grenoble INP, LRP, 38000 Grenoble, France
- 7 b) Univ. Grenoble Alpes, CNRS, CERMAV, F-38000 Grenoble, France
- 8 *Corresponding author: komla.ako@univ-grenoble-alpes.fr; akokomla@hotmail.com

9

10

Abstract

- 11 The salt sensitivity and selectivity feature of α -carrageenan (α -Car) were investigated and
- 12 compared with κ -carrageenan (κ -Car) and iota-carrageenan (ι -Car). These carrageenans are
- identified by one sulfate group on the 3,6-anhydro-D-galactose (DA) for α -Car, D-galactose
- 14 (G) for κ -Car and on both carrabiose moieties (G and DA) for ι -Car. The viscosity and
- temperature, where order-disorder transition have been observed, were greater in presence of
- 16 CaCl $_2$ for α -Car and ι -Car compared with KCl and NaCl. Conversely, the reactivity of κ -Car
- 17 systems were greater in presence of KCl than $CaCl_2$. Unlike κ -Car systems, the gelation of α -
- 18 Car in presence of KCl was observed without syneresis. Thus, the position of sulfate group on
- 19 the carrabiose determines the importance of counterion valency too. The α -Car could be a
- 20 good alternative to κ -Car to reduce the syneresis effects.
- 21 **Keywords**: Alpha-carrageenan; Sulfate group position; Anhydro-ring; Viscosity; Salt
- 22 sensitivity; Syneresis

1 Introduction

23

24 Numerous applications of natural polymers are limited by the ability of the polymers to give a gel with exudation (syneresis) (Chen, Xu & Wang, 2007; Gonzalez, Loubes, Bertotto, Baeza 25 & Tolaba, 2021; Laplante, Turgeon & Paquin, 2006). Some of the applications are relevant 26 27 for human health management like cell cultures in hydrogels for disease diagnostics and tissue engineering (Divoux, Mao & Snabre, 2015; Hoffman, 2012; Jafari, Farahani, Sedighi, Rabiee 28 & Savoji, 2022; Peppas, Bures, Leobandung & Ichikawa, 2000; Zhang et al., 2020). To 29 understand the physicochemical causes of syneresis of natural polymers hydrogels, kappa-30 carrageenan polysaccharide is used as the material of this investigation, because this material 31 exhibits severe syneresis in relatively short experimental time with a dependence to many 32 physicochemical factors (Ako, 2015; Elmarhoum & Ako, 2021). In this study, the target is the 33 sulfate, -SO₃, ionic group position in the monomer of this polysaccharide. 34 Carrageenan (Car) defines a family of sulfated galactans extracted from marine algae 35 (Prechoux, Genicot, Rogniaux & Helbert, 2013; Souza, Hilliou, Bastos & Gonçalves, 2011; 36 Usov, 2011). The disaccharide repetition moieties (i.e. carrabiose) of the polysaccharide is 37 made with two galactose units identified as G and DA for respectively D-galactose and 3,6-38 39 anhydro-D-galactose, each of which can carry different number of sulfate functional group (Ueda, Saiki & Brady, 2001; Usov, 2011) (Fig.1). The number and position of the sulfate 40 group on the carrabiose identifies the type of carrageenan (Campo, Kawano, da Silva Jr & 41 Carvalho, 2009; Millane, Chandrasekaran & Arnott, 1988; Perez & Claudio, 2020; Souza, 42 Hilliou, Bastos & Gonçalves, 2011; Viebke, Borgström & Piculell, 1995). Kappa-carrageenan 43 (κ-Car) has one sulfate group at the position 4 of the G units and is coded as G4S-DA. Alpha-44 Carrageenan (α-Car) has one sulfate group at the position 2 of the DA units and is coded as G-45 DA2S accordingly (Bourgoin, Zablackis & Poli, 2008). Iota-carrageenan (1-Car) has two 46

sulfates on the carrabiose units on the same carbon position as for κ - and α -Car, and thus, is coded as G4S-DA2S. The number, the position and the distribution of ester sulfate along the carrageenans chain define the physicochemical properties of the polysaccharides (Millane, Chandrasekaran & Arnott, 1988; Souza, Hilliou, Bastos & Gonçalves, 2011). As an example, with the same concentration of positive charge counterions, the divalent counterions like calcium (Ca²⁺) influence strongly the flow behavior of 1-Car solution than the monovalent counterions like potassium (K⁺) (Bui, Nguyen, Renou & Nicolai, 2019; Elfaruk, Wen, Chi & Li, 2021). Conversely, K⁺ ions change drastically the flow behavior of κ-Car solution compared with 1-Car (Lai, Wong & Lii, 2000; Norton & Goodall, 1983b). The selectivity of 1-Car and κ-Car to respectively divalent and monovalent counterions and the fact that trivalent counterion induced gelation in λ -Car, which carries three sulfate groups on the carrabiose, lead to claim that the number of sulfate on the carrabiose is what determines the importance of counterion valency (Running, Falshaw & Janaswamy, 2012). The exact role of the sulfate position on the counterions binding and importance of the counterion valency in carrageenan systems are unclear (Klevens, 1950; Lindman, Karlstrom & Stigsson, 2010; Running, Falshaw & Janaswamy, 2012). The κ-Car hydrogels, for example, exhibit severe syneresis behavior in presence of K⁺ than 1-Car and other types of salts which are commonly used in food and biological domains (Thrimawithana, Young, Dunstan & Alany, 2010). Therefore, in order to understand the influence of the sulfate position on the rheological behavior of the carrageenans, we took advantage of the recent discovery of the endo-40-1-carrageenan sulfatase secreted by the marine bacteria Pseudoalteromonas atlantica T6c. This enzyme removes specifically the sulfate ester group located at the position 4 of the 1-carrabiose moiety and allows completely converting the 1-Car into α-Car (Prechoux, Genicot, Rogniaux & Helbert, 2013). We have investigated the rheological behavior of κ-Car

47

48

49

50

51

52

53

54

55

56

57

58

59

60

61

62

63

64

65

66

67

68

69

- and α -Car solution in presence of different types of salts to emphasize the role that the
- positions of -SO₃ may have in the flow properties of the polysaccharide.

2 Material and methods

74 2.1 Samples characteristics

- 75 Molecular weight. The samples were solubilized in 0.1 M NaNO₃ and filtered on 0.2 µm
- membrane. The samples were injected on two OHpak SB 806 M HQ Shodex columns
- mounted in series and eluted at 30°C in 0.1 M NaNO₃. Detection was monitored by a Optilab
- 78 rEX Wyatt refractometer and a mini DAWN TREOS Wyatt light scattering working at three
- angles. The dn/dC was 0.142 mL/g. Polydispersity indices was calculated Ip = Mw/Mn. All
- 80 measurements were done in triplicate.
- 81 Enzymatic preparation of alpha-carrageenan (α -Car). The α -carrageenan (Fig1) was
- prepared according to the method reported by (Prechoux, Genicot, Rogniaux & Helbert, 2013)
- using recombinant 40-iota-carrageenan sulfatase. Briefly, the genePatl 0889 (GenBank
- accession number ABG39415) encoding the sulfatase with a N-terminal poly-histidine tag
- was over expressed in BL21-DE3 Escherichia coli strain and purified on a Nickel affinity
- 86 column. Iota-carrageenan provided by Danisco (Brabrand, Denmark) was incubated with the
- purified sulfatase in 50 mM Tris-HCl pH 7.5. Incubation with additional fresh enzymes was
- 88 repeated until the complete conversion to α -carrageenan was achieved. The yield of
- desulfation was measured by ¹H NMR (Prechoux, Genicot, Rogniaux & Helbert, 2013). The
- 90 polysaccharide was ultra-filtered on 10 Da membrane in water and lyophilized. The average
- 91 molecular weight of the polysaccharide was $M_w = 3.8 \times 10^5$ g/mol, with an average
- polydispersity index of $M_w/M_n = 1.78$.

Kappa-carrageenan (k-Car). The κ-Car polysaccharide (Fig1) is a gift from Rhodia Food Switzerland, product name and reference: MEYPRO-GEL 01/2001 WG95-37 K-Car. The dialyzed product may contain a weak amount of glucose and was found in the pure potassium κ-Car form of total potassium at no more than 4.2 % (w/w); the other salts were considered to be only trace amounts. The measurement of its molecular weight was done in triplicate. The average molecular weight of the polysaccharide was $M_w = 3.3 \times 10^5$ g/mol, with an average polydispersity index of $M_w/M_n = 2.02$.

$$OSO_3^ CH_2OH$$
 $OSO_3^ CH_2OH$ $OSO_3^ OSO_3^ OSO_3^$

101 Figure 1: Structure of the carrabiose moieties composing kappa-, alpha- and iota-102 carrageenan.

2.2 Samples preparation

A 20 g/L solution was prepared by dissolving the κ -Car polysaccharides powder in demineralized water with 200 ppm sodium azide as the bacteriostatic agent under heat and stirring conditions. The resulting solution was dialyzed and filtered through 0.45 μ m pore size. A 9 g/L of solution was prepared by dilution of the 20 g/L solution and was stocked in fridge at 5 °C, with which, all the samples were prepared by dilution with demineralized water containing 200 ppm sodium azide with salt at the appropriate concentration. The dilution gives the desired final concentration of polysaccharide and salt concentration at the appropriate temperature. The α -Car polysaccharide in the lyophilized form were dissolved in demineralized water with 200 ppm sodium azide under heat and stirring conditions to 9 g/l stock solution. The samples were prepared in the same way like the κ -Car using the appropriate salt.

2.3 samples characterization

115

116

117

118

119

120

121

122

123

124

125

126

127

128

129

130

131

132

133

134

135

136

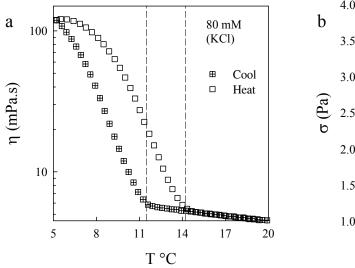
be specified in the text.

Viscosity measurements. The viscosity measurements were performed in Couette geometry using a DHR3 Rheometer (TA Instrument). The Couette geometry consisted of a concentric cylinder geometry of an inner rotor cylinder (bob) and outer stator cylinder (cup) of respectively a radius R₁ of 14 mm and R₂ of 15 mm defining a horizontal gap (R₂ - R₁) of 1 mm and an average radius R of 14.5 mm as $(R_1 + R_2)/2$. The height of the bob was 42 mm and the vertical gap between the bob and cup was set at 2 mm. The minimum and maximum torque of the instrument were 5 μ Nm and 5×10^3 Nm respectively. Both the torque (M) and the angular velocity (Ω) of the bob, which were proportional with respectively the stress (σ) and shear rate $(\dot{\gamma})$, were measured by the rheometer, from which the viscosity (η) was displayed as $\sigma/\dot{\gamma}$. The viscosity was determined as a function of the shear rate from 0.3 s⁻¹ to 30 s⁻¹ to define the Newtonian flow domain at a constant temperature in the range between 5 °C and 60 °C. The viscosity was also measured at a constant shear rate of 15 s⁻¹ while cooling or heating at 2 °C min⁻¹ or a lower rate 0.5 °C.min⁻¹. Viscoelastic modulus measurements. The rheological measurements were performed using a DHR3 Rheometer (TA Instrument) with etching conic geometry (Ø50 mm, 2.0°, TADC2°InR500) and etching plate. The conic geometry has a truncation gap of 110.0 µm. A portion of the prepared hot sample solution was loaded onto the Peltier of the rheometer and set at a temperature above the gelation temperature of the sample. After loading the sample, the surface was covered with a thin layer of mineral oil to avoid evaporation. Then, the temperature and the measurement program were run. The condition of each measurement will

3 Results and discussion

3.1 Flow behavior of α -carrageenan solution

The viscosity of α -Car in presence of 80 mM KCl is displayed in Fig. 2a for cooling and heating ramps. The viscosity increases when the temperature is decreased, and sharply from \approx 11.5 °C. The sharp increase of the viscosity was shown by Rochas and Rinaudo (1984) to be concomitant with the coil-helix transition. If the temperature is increased immediately from 5 °C, the viscosity decreases sharply and the helix-coil transition was found at \approx 14.3 °C above the coil-helix transition (Rochas & Rinaudo, 1984). The thermal hysteresis was reported as a result of inter-helical aggregation (Morris, Rees & Robinson, 1980; Piculell, Nilsson & Muhrbeck, 1992).



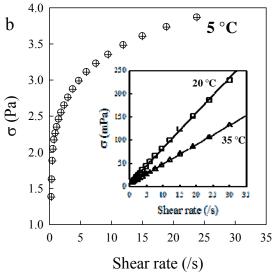


Figure 2: a) temperature dependence of α -Car solution viscosity for 3 g/L, 80 mM KCl, b) shear rate and stress relation displayed for the solution to show the viscosity behavior, at 5 °C in the helices domain the viscosity changes with the shear rate, in the coil domain the viscosity is constant (insert).

The thermal hysteresis defines three temperature domains which are identified in the Fig.2a as: (1) domain below the coil-helix transition temperature, < 11.5 °C, (2) domain between the coil-helix and helix-coil transition temperature, [11.5 °C, 14 °C] and (3) domain above the helix-coil transition temperature, > 14 °C. The conformational state of the carrageenan in temperature domain (2) depends on the temperature ramp (cooling or heating) but in the domain (1) or (3) the conformations of the polysaccharide chains were respectively either helices or coils. The shear rate dependence of the applied stress plots is shown in the Fig.2b. A non-linear relationship is observed, at 5 °C, in the domain below the coil-helix transition temperature and a linear relationship is observed at temperatures in the domain above the helix-coil transition temperature (insert of Fig.2b). We have observed that the flow behavior of κ-Car turns from Newtonian to non-Newtonian flow when the conformations of the polysaccharides turn from coils to helices (Elmarhoum & Ako, 2021). We think that similar conformational change could explain the temperature dependence of the flow behavior of α-Car. The impact of salt on the flow behavior of carrageenan solutions depends mainly on the kinetics of the helices aggregation and the kinetics increases with decreasing temperature below the coil-helix transition temperature (Meunier, Nicolai, Durand & Parker, 1999). We didn't observe the gelation for α-Car solutions in the interval of temperature explored in Fig.2a, when in the same KCl and polysaccharide concentration the κ -Car solutions gelled at temperature below 50 °C with syneresis (Ako, 2015, 2017). A lower helices aggregation kinetics is expected in relation with the weak coil-helix transition temperature in comparison with κ -Car. We didn't exclude the fact that the α -Car solutions at temperature below 12 °C behave like a suspension of aggregates (Garrec, Guthrie & Norton, 2013).

3.2 Coil-helix and helix-coil transition temperature: effect of salt

154

155

156

157

158

159

160

161

162

163

164

165

166

167

168

169

170

171

172

173

174

175

176

177

178

The transition from random coils to helices conformation, which is measured using optical rotation technique, have been observed to be concomitance with the steep rising of the

viscosity (Rochas & Rinaudo, 1984). Therefore, the onset of helix formation during cooling ramp is characterized by the temperature at which the viscosity starts to increase and is denoted as the coil-helix temperature (T_{ch}). Salt concentration dependence of the T_{ch} is shown in the Fig.3. The results for CaCl₂ salt are displayed for α -Car and the result for KCl salt is displayed in the insert of the Fig.3 for α -Car and κ -Car. The polysaccharides sensitivity to salt as a polyelectrolyte is expected (Ako, 2015; Ciancia, Milas & Rinaudo, 1997), but the fact that the coil-helix transition of α -Car is more sensitive to CaCl₂ than KCl was not obvious regarding the literature on κ -Car (Elmarhoum & Ako, 2021). Moreover, the sensitivity of κ -Car to KCl result in a greater coil-helix transition temperature than the sensitivity of α -Car to both KCl and CaCl₂. This finding emphasis the importance of the -SO₃⁻ position in the monomer as a physicochemical factor, which determines the polysaccharide reaction to salt.



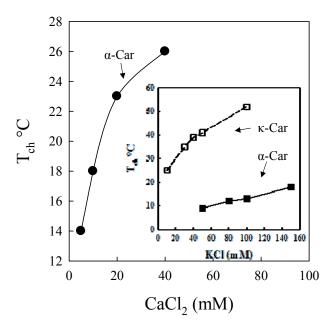


Figure 3: $CaCl_2$ dependence of the coil-helix transition temperature of α -Car and in the insert the KCl concentration dependence of the coil-helix transition of α -Car and κ -Car.

For α -Car, if CaCl₂ concentration is increased from 5 mM to 40 mM, the T_{ch} increases from \approx 14 °C to \approx 26 °C. The T_{ch} increases from \approx 10 °C to 20 °C, if the KCl concentration is increased from 45 mM to \approx 150 mM and the coil-helix transition was not observed for 20 mM KCl concentration, which is two time greater than 5 mM CaCl₂ in term of the concentration of positive charge counterions. Therefore, α -Car is more sensitive to divalent than the monovalent counterions. Based on this result, the behavior of α -Car with one sulfate anionic functional group on its monomer is rather similar to that of ι -Car with two sulfates anionic functional groups than κ -Car (Morris, Rees & Robinson, 1980). The κ -Car systems with one sulfate anionic functional group on its monomer react strongly in presence of KCl salt than the α -Car systems. The T_{ch} increases from \approx 25 °C to \approx 50 °C, if the KCl concentration is increased from 10 mM to 100 mM, with the hysteresis loop effect.

3.3 Monovalent salt sensitivity of α-Car

The fact that the conformational change of α -Car happens at a greater temperature in presence of calcium than potassium counterions, characterizes its greater sensitivity for the divalent salt than the monovalent salt (Morris, Rees & Robinson, 1980). At any rate, in the conditions where the coils conformations prevail, the viscosity of the solution doesn't depend on the cooling or heating ramps and the solution flow is Newtonian. In this section we have tried to bring some more results on the sensitivity of the polysaccharide for monovalent salts like; NaCl, KCl and KI.

In the Fig.4a, are shown the temperature dependence of the viscosity for the solution of 3 g/L as the reference (ref.) to show the effect of 10 mM of the selected monovalent salt. The results show a decrease of the viscosity in presence of the salts with no particular effect due to the type of salt for the concentration of 10 mM. Moreover, the KCl salt concentration is not enough to induce the coil-helix transition in the temperature range of the measurements as it

was observed for κ -Car. Based on the fact that, the temperature dependence of κ -Car solutions viscosity for 10 mM of NaCl, KI or KCl are different (Ciancia, Milas & Rinaudo, 1997), we didn't expect that the temperature dependence of α -Car solutions viscosity being similar. We cannot explain the difference between the behavior of α -Car and κ -Car, without considering the sulfate group position. If the NaCl concentration is increased to 100 mM, the solutions exhibited a coil-helix transition around 15 °C and a helix-coil transition around 17 °C. This may appear to be a small hysteresis but when scan rate dependence is accounted for these would be superimposable (Fig.4b). In presence of 100 mM of KCl, the coil-helix transition is observed around 12 °C and two transitional temperatures were observed for the heating ramp; the first one around 17 °C and the second one correspond to the helix-coil transition around 42 °C.



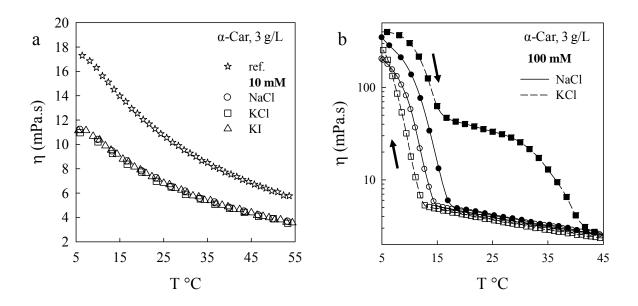


Figure 4: temperature dependence of the viscosity of α -Car solution,3 g/L in presence of a) 0 mM (ref.) and 10 mM of NaCl, KCl or KI, the cooling and heating ramps give similar results and b)100 mM of NaCl or KCl, the cooling (open symbols) and heating (full symbols) results are displayed.

The thermal hysteresis of the solution of α -Car with 100 mM KCl indicates an extensive aggregation of the helices which led the conserved solution to gel at 5 °C. The solution with 100 mM NaCl didn't gel after several days at 5 °C. The shift of the coil-helix transition temperature T_{ch} from 12 °C to 15 °C, when KCl is replaced by NaCl, leads to say that the conformational change of α -Car would be rather sensitive to NaCl than KCl. The role of monovalent salt to promote helix formation and then gelation for κ -Car systems was reported to follow the sequence RbCl > CsCl, KCl > NaCl > LiCl, HCl (Ciancia, Milas & Rinaudo, 1997; Rochas & Rinaudo, 1984). The superiority of the potassium on sodium to induce coilhelix conformational change of κ -Car is inversed in the case of α -Car. This inversion may indicate that the anhydro-ring promotes the accessibility to the sulfate group on the 3,6-anhydro-D-Galactose unit by small size counterions. However, extensive aggregation of the helices and gelation in presence of 100 mM KCl, indicate that the ion-pair formation of K⁺ with the -SO₃ is a time dependent process (Grasdalen & Smidsrod, 1981; Norton & Goodall, 1983a). The reduction of the sulfate charge by counterions would be fast for κ -Car, and thus, promotes immediate coil-helix transition and aggregation of the helices to gelation.

3.4 Influence of the degree of carrageenan sulfation

The gelation of different types of mono-composite (MC) of carrageenan polysaccharides made of α -carrabiose and ι -carrabiose on the same backbone are proposed in this section to show how the degree of conversion of ι -Car to α -Car can influence the coil-helix transition temperature. The gelation of a bi-composite (BC) solution, which is a blending of α -Car and ι -Car solution to the proportion of 67 wt.% and 33 wt.% respectively, is also shown. Given that the α -Car is obtained by conversion of ι -Car, the result of the bi-composite will indicate the influence that could have a few content of ι -Car in the α -Car solution on the coil-helix transition temperature (Piculell, Nilsson & Muhrbeck, 1992). The concentration of all the

solutions is 6 g/L and were prepared with 20 mM of $CaCl_2$ then cooled to give the results in Fig.5. The temperature dependence of the G' of the solutions were measured during the cooling ramp. The result of the BC solution is marked with a star in the legend of Fig5.b. The gelation of carrageenan systems is a time dependent process which is connected to the helices aggregation kinetic, thus the gelling point as the point where G' = G'' depends on the scan rate. The onset of elastic behavior of aqueous iota-carrageenan has been observed by Piculell et al., (1992) to closely follow the onset of helix formation, and the modulus of elasticity increases monotonically with an increasing helical content (Piculell, Nilsson & Muhrbeck, 1992). Therefore, if G' > G'' is observed, the temperature at which the helices are formed, is considered here as the gelation temperature. In this section, we have worked in the conditions where for all the samples, G' > G'' was observed during the cooling ramps.

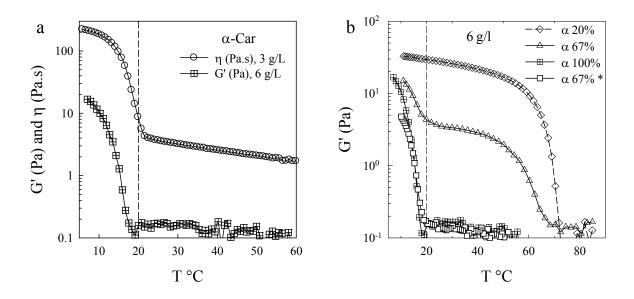


Figure 5: Temperature dependence of the G' and the viscosity of a) α -Car in presence of 20 mM $CaCl_2$ and b) different composition of α -Car and 1-Car carrabiose in the polysaccharide backbone. The composition α 67 %* is a solution made with 67 wt% of α -Car and 33 wt% of 1-Car. The solutions were prepared with 20 mM $CaCl_2$.

In Fig.5a, the gelation temperature is observed to be almost equal to the temperature at which the G' of the system increases sharply while cooling the solution. We can see that the domain of temperature where the viscosity increases coincides with that of G', which we said is 277 concomitant with the formation of helices (Belton, Chilvers, Morris & Tanner, 1984; Ciancia, 278 Milas & Rinaudo, 1997; Elmarhoum & Ako, 2021; Meunier, Nicolai, Durand & Parker, 1999; 279 Norton & Goodall, 1983b; Rochas & Rinaudo, 1984). The α-Car solution and the solution of 67 wt.% α -Car with 33 wt.% ι -Car, indicated in the Fig.5b respectively as α 100 % and α 67 %*, show the same gelation temperature. Therefore, presumably a few content of 1-Car in the α-Car solution would not affect the coil-helix transition temperature of α-Car and its 284 sensitivity to salt. The thermal behavior of α-Car solution seems to be very stable regarding a small variation of 1-Car concentration in wt.%. Considering the molecular weight of each composite of the blending solution, the 67 wt.% of α-Car may correspond to 90 mol%, which means that one molecules of ι-Car is surrounded by nine molecules of α-Car and this could explain the similarity between the thermal properties of the 100 mol% and 90 mol% α-Car 288 289 solution. In the MC polysaccharide, the proportion of α-carrabiose and ι-carrabiose are in mol% and 67 % of α-Car in the MC shows the gelation temperature of the part related to both 290 α-Car and ι-Car. This result, may indicate that the coil-helix transition would be rather an 291 intramolecular mechanism. In the condition where the α-carrabioses are randomly distributed 292 along the hybrid carrageenan chain (Prechoux, Genicot, Rogniaux & Helbert, 2013), we 293 should expect one gelation temperature. However, the probability to have a bloc of α-294 carrabioses along the hybrid chain which behave like a free chain of α -Car is increased by 295 296 increasing the degree of ι-Car conversion to α-Car and this fact seems to correspond to the results in Fig.5b. Below 50 % of α -Car in the MC, the gelation temperature of α -Car was not 297 clearly detectable. Supposed that the 20 mol% / 80 mol% of α -/ ι -Car were distributed like a 298 bloc copolymer, then the temperature dependence of G' would have shown distinctly the 299

275

276

280

281

282

283

285

286

gelation temperature of α -Car and ι -Car. Bui et al., (2019) have mixed 50 wt.% of κ -Car and 50 wt.% of ι -Car in 50 mM CaCl₂ aqueous solution and they have observed two distinct gelation temperatures of the BC solution indicating independent coil-helix transition (Bui, Nguyen, Renou & Nicolai, 2019). However the authors observe that κ -Car network is more homogeneous in the mixed gels than in the corresponding individual gels and the mixed gels were stiffer than the individual gels. The independent coil-helix transition of the polysaccharide which occurs in both BC and MC systems may indicate that the reduction of the polymers net charge is preferentially intramolecular by which the helices and aggregates are formed.

The claim of intramolecular mechanism to explain the reduction of the polysaccharide charge or ion-pair formation could also refer to the finding of Rochas and Rinaudo (1984). The authors found that the ionic selectivity for potassium counterions work also even if the κ -Car is in the coil conformation. If so, then the coil-helix conformational change may take place in concentration well below the critical concentration, where the polysaccharide are supposed to be individual. Hence, the polysaccharide conformation would be single helix. However, the authors have rejected the single-helix formation presumably because they didn't observe a specific role of the potassium ion in the crosslinking mechanism of the polysaccharide. It is not clear the way the intramolecular charge neutralization mechanism conciliates the hypothesis of double helices and the aggregation of the helices in BC and MC model, except if the double helices are considered as aggregate of two single-helices.

We can note the absence of peak or maximum point for the G' plots in Fig.5, which several authors reported for κ -Car gel systems as characteristic of slipping effect due to sever syneresis (release of liquid phenomena by the gels). The gelation of κ -Car solutions with K^+ , Cs^+ , Ca^{2+} gives gels with syneresis was reported by several authors (Ako, 2015; Lai, Wong & Lii, 2000; Piculell, Borgström, Chronakis, Quist & Viebke, 1997; Richardson & Goycoolea,

1994). The salt concentrations at which the syneresis was observed are relatively very weak compared with t-Car (Piculell, Nilsson & Muhrbeck, 1992; Thrimawithana, Young, Dunstan & Alany, 2010) and if the syneresis mechanism may exist for α -Car, then it should be caused by salt concentrations larger than those used for κ -Car and presumably over the experimental time conditions of this study. The gels coarsening and contraction cause syneresis in general, but in condition which is determined by the polysaccharide net charge reduction or neutralization, extensive helices aggregation induced syneresis reflects the salt ability to neutralize or reduce the intra and/or intermolecular repulsion forces. Therefore, sulfate group position on D-galactose unit is more favorable to neutralization by salt than the sulfate group position on 3,6-anhydro-D-galactose unit.

3.5 Discussion

Alpha-carrageenan and kappa-carrageenan are characterized respectively, by a mono sulfate group on the 3,6-anhydro-D-galactose unit and a mono sulfate group on the D-galactose unit. Zablackis et al., have studied the carrageenan extracted from *Catenella nipae Zanardini*, which according to the work of Falshaw et al., (1996) may correspond to hybrid α -/t-Car of variable proportion (Bourgoin, Zablackis & Poli, 2008; Falshaw, Furneaux, Wong, Liao, Bacic & Chandrkrachang, 1996), and the content of α -carrabiose in the native hybrid polysaccharide would not exceed 40 mol% (Prechoux, Genicot, Rogniaux & Helbert, 2013). The sodium form of this polysaccharide (in 100 mM NaCl) did not gel but exhibit a greater viscosity than κ -Car (Zablackis & Santos, 1986), while the sodium form of t-Car (in 100 mM NaCl) has been observed to form a gel (Piculell, Nilsson & Muhrbeck, 1992). In presence of salt the viscosity of the solutions of α -Car and κ -Car depends on the degree of their helices formation and aggregation. However, the decrease of the gelling capacity when the proportion of α -carrabiose increases in the hybrid polysaccharide indicates that the sulfate on the anhydro-D-galactose unit is less favorable for the inter-helical aggregation compared with the

sulfate on the D-galactose unit. Therefore, lower inter-helical aggregation activity and higher inter-helical aggregation activity, respectively, characterize the function of the sulfate on the anhydro-D-galactose unit and the function of the sulfate on the D-galactose unit, which explain, respectively, the weak gelling capacity of α -Car and the strong gelling capacity of κ -Car.

The viscosity of the solution of 3 g/L of α -Car and κ -Car at 20 °C were respectively 12.2 mPa.s and 7.8 mPa.s. The viscosity of the free salt solution of α -Car is observed to be greater than the viscosity of the free salt solution of κ -Car at a similar molecular weight, polysaccharide concentration and temperature conditions. Intrachain repulsive forces and water molecules binding capacity would be greater for α -Car than κ -Car (Ako, Elmarhoum & Munialo, 2022), and accordingly, the sulfate on the anhydro-D-galactose unit promotes more expansion of the polysaccharide than the sulfate on the D-galactose unit.

It is appropriate to emphasis that the α -Car system ionic selectivity for CaCl₂ rather than KCl was not expected, because the κ -Car system, which could derive from t-Car too, is known to be more sensitive to KCl than CaCl₂. Therefore, the sensitivity of carrageenan systems to counterions is not only a matter of number of sulfate groups on the carrabiose (Running, Falshaw & Janaswamy, 2012) but a result of many physicochemical factors. The size of hydrated counterions is quoted to play a role in the distance between the cation and the sulfate group (Piculell, Nilsson & Muhrbeck, 1992). The cation mobility can also affect the ion-pair formation stability or equilibrium constant, i.e., the small size cations could have a greater mobility, and thus, lead to a less stable or strong ion-pair formation (Ciancia, Milas & Rinaudo, 1997; Grasdalen & Smidsrod, 1981). Although α -Car and t-Car system are rather sensitive to CaCl₂, the conformational transition temperature of α -Car systems is found well below that of t-Car systems, which reflect the intramolecular characteristic of the charge reduction mechanism in carrageenan systems. The ability of salt to reduce the charge carried

by carrageenan polysaccharides could be pictured as an in-to-out process. The in-to-out neutralization process means that the system tries to neutralize first the charge carried by the carrabiose (intramolecular process), and if this fails like in the case of α -Car with calcium, the system tries to neutralize the charge carried by the carrageenan chain (intrachain process). In the intrachain process or interaction between carrabiose of same chain, if a block of carrabiose behave the same, like a block of α -Car in α -/t-Car hybrid polysaccharide, then the conformation transition characterizing the block could be detectable. The neutralization process could continue or stop by interchain/intermolecular process (aggregation), in this process the two sulfates or carrabiose which bind the multi valence counterions do not belong to the same chain.

4. Conclusion

The importance of counterions in the helical formation of carrageenan depends on many physicochemical factors. We show in this treatise that the influence of the counterion valency depends on the position and number of the sulfate group carried by the carrabiose. The sulfate on the 3,6-anhydro-D-galactose unit, which characterizes α -carrageenan (G-DA2S), is rather sensitive to divalent counterions and the sulfate group on the D-galactose unit, which characterizes κ -carrageenan (G4S-DA), is rather sensitive to monovalent counterions. The two types of sulfate, which characterize t-carrageenan (G4S-DA2S), is more sensitive to divalent counterions than the one type of sulfate carried by α -carrageenan (G-DA2S). The temperature dependence of the G' of α -/t-carrageenan, as a carrageenan block copolymer system in CaCl₂ solution, has shown distinctly the helical formation temperature of α -carrageenan and t-carrageenan. Based on the results of this study, the counterion influence, which we qualify as in-to-out process, would be preferentially and gradually intramolecular, then intrachain and finally interchain process.

399 **5 Acknowledgements**

- 400 We thank Vincent VERDOOT for his technical assistance with the rheometer instruments.
- 401 We thank Laurine Buon and Eric Bayma from Cermav for their kind assistance with the
- 402 products composition. This work was financially supported by I-MEP2 PhD graduate school
- and the PolyNatCarnotInstitut. The LaboratoireRhéologie et Procédés (LRP) is part of the
- 404 LabEx Tec 21 (Investissementsd'Avenir grant agreement n°ANR-11-LABX-0030) and of
- 405 the PolyNat Carnot InstitutInvestissementsd'Avenir grant agreement n°ANR-11-CARN-
- 406 030-01).

407

6 References

- Ako, K. (2015). Influence of elasticity on the syneresis properties of kappa-carrageenan gels.
- 409 *Carbohydrate Polymers*, *115*, 408-414.
- 410 Ako, K. (2017). Influence of osmotic and weight pressure on water release from
- 411 polysaccharide ionic gels. *Carbohydrate Polymers*, 169, 376-384.
- Ako, K., Elmarhoum, S., & Munialo, C. D. (2022). The determination of the lower critical
- concentration temperature and intrinsic viscosity: The syneresis reaction of polymeric gels.
- 414 *Food Hydrocolloids*, *124*, 107346.
- Belton, P. S., Chilvers, G. R., Morris, V. J., & Tanner, S. F. (1984). Effects of group I cations
- on the gelation of iota carrageenan. *International Journal of Biological Macromolecules*, 6,
- 417 303-308.
- Bourgoin, A., Zablackis, E., & Poli, J. B. (2008). Characterization of a-carrageenan solution
- behavior by field-flow fractionation and multiangle light scattering. Food Hydrocolloids, 22,
- 420 1607-1611.
- 421 Bui, V. T. N. T., Nguyen, B. T., Renou, F., & Nicolai, T. (2019). Rheology and
- microstructure of mixtures of iota and kappa-carrageenan. Food Hydrocolloids, 89, 180-187.

- 423 Campo, V. L., Kawano, D. F., da Silva Jr, D. B., & Carvalho, I. (2009). Carrageenans:
- 424 Biological properties, chemical modifications and structural analysis A review.
- 425 Carbohydrate Polymers, 77, 167-180.
- 426 Chen, H.-H., Xu, S.-Y., & Wang, Z. (2007). Interaction between flaxseed gum and meat
- protein. *Journal of Food Engineering*, 80, 1051-1059.
- 428 Ciancia, M., Milas, M., & Rinaudo, M. (1997). On the specific role of coions and counterions
- on kappa-carrageenan conformation. *International Journal of Biological Macromolecules*, 20,
- 430 35-41.
- Divoux, T., Mao, B., & Snabre, P. (2015). Syneresis and delayed detachment in agar plates.
- 432 *Soft Matter, 11,* 3677-3685.
- Elfaruk, M. S., Wen, C., Chi, C., & Li, X. (2021). Effect of salt addition on iota-carrageenan
- 434 solution properties. *Food Hydrocolloids*, 113, 106491.
- 435 Elmarhoum, S., & Ako, K. (2021). Lower critical concentration temperature as
- 436 thermodynamic origin of syneresis: Case of kappa-carrageenan solution. Carbohydrate
- 437 *Polymers, 267*, 118191.
- 438 Falshaw, R., Furneaux, R. H., Wong, H., Liao, M.-L., Bacic, A., & Chandrkrachang, S.
- 439 (1996). Structural analysis of carrageenans from Burmese and Thai samples of Catenella
- nipae Zanardini. Carbohydrate Research, 285, 81-98.
- 441 Garrec, D. A., Guthrie, B., & Norton, I. T. (2013). Kappa carrageenan fluid gel material
- properties. Part 1: Rheology. *Food Hydrocolloids*, *33*, 151-159.
- Gonzalez, L. C., Loubes, M. A., Bertotto, M. M., Baeza, R. I., & Tolaba, M. (2021). Flow
- behavior and syneresis of ball milled rice starch and their correlations with starch structure.
- 445 *Carbohydrate Polymer Technologies and Applications*, 2, 100168.

- Grasdalen, H., & Smidsrod, O. (1981). (133)Cs NMR in the Sol-Gel States of Aqueous
- Carrageenan. Selective Site Binding of Cesium and Potassium Ions in k-Carrageenan Gels.
- 448 *Macromolecules, 14*, 229-231.
- Hoffman, A. S. (2012). Hydrogels for biomedical applications. Advanced Drug Delivery
- 450 *Reviews*, *64*, 18-23.
- Jafari, A., Farahani, M., Sedighi, M., Rabiee, N., & Savoji, H. (2022). Carrageenans for tissue
- engineering and regenerative medicine applications: A review. Carbohydrate Polymers, 281,
- 453 119045.
- Klevens, H. B. (1950). Solubilization. *Chemical Reviews*, 47, 1-74.
- Lai, V. M. F., Wong, P. A.-L., & Lii, C.-Y. (2000). Effects of Cation Properties on Sol-gel
- 456 Transition and Gel Properties of k-carrageenan. *Journal of Food Science*, 65(8), 1332-1337.
- Laplante, S., Turgeon, S. L., & Paquin, P. (2006). Emulsion-stabilizing properties of chitosan
- in the presence of whey protein isolate: Effect of the mixture ratio, ionic strength and pH.
- 459 *Carbohydrate Polymers*, *65*, 479-487.
- Lindman, B., Karlstrom, G., & Stigsson, L. (2010). On the mechanism of dissolution of
- 461 cellulose. *Journal of Molecular Liquids*, 156, 76-81.
- 462 Meunier, V., Nicolai, T., Durand, D., & Parker, A. (1999). Light Scattering and
- Viscoelasticity of Aggregating and Gelling kappa-Carrageenan. Macromolecules, 32, 2610-
- 464 2616.
- 465 Millane, R. P., Chandrasekaran, R., & Arnott, S. (1988). The molecular structure of kappa-
- carrageenan and comparison with iota-carrageenan. Carbohydrate Research, 182, 1-17.
- 467 Morris, E. R., Rees, D. A., & Robinson, G. (1980). Cation-specific Aggregation of
- 468 Carrageenan Helices: Domain Model of Polymer Gel Structure. Journal of Molecular
- 469 Biology, 138, 349-362.

- Norton, I. T., & Goodall, D. M. (1983a). Equilibrium and Dynamic Studies of the Disorder-
- Order Transition of Kappa Carrageenan. J. Chem. Soc., Faraday Trans. 1, 79, 2489-2500.
- Norton, I. T., & Goodall, D. M. (1983b). Role of Cations in the Conformation of Iota and
- Kappa Carrageenan. Journal of the Chemical Society, Faraday Transactions 1, 79, 2475-
- 474 2488.
- Peppas, N. A., Bures, P., Leobandung, W., & Ichikawa, H. (2000). Hydrogels in
- 476 pharmaceutical formulations. European Journal of Pharmaceutics and Biopharmaceutics, 50,
- 477 27-46.
- 478 Perez, S. J. L. P., & Claudio, G. C. (2020). Molecular dynamics simulations of two double-
- 479 helical hexamer fragments of iota-carrageenan in aqueous solution. Journal of Molecular
- 480 Graphics and Modelling, 98, 107588.
- Piculell, L., Borgström, J., Chronakis, I. S., Quist, P.-O., & Viebke, C. (1997). Organisation
- and association of k-carrageenan helices under different salt conditions. *International Journal*
- 483 *of Biological Macromolecules, 21*, 141-153.
- 484 Piculell, L., Nilsson, S., & Muhrbeck, P. (1992). Effects of small amounts of kappa-
- carrageenan on the rheology of aqueous iota-carrageenan. Carbohydrate Polymers, 18, 199-
- 486 208.
- Prechoux, A., Genicot, S., Rogniaux, H., & Helbert, W. (2013). Controlling Carrageenan
- 488 Structure Using a Novel Formylglycine-Dependent Sulfatase, an Endo-4S-iota-Carrageenan
- 489 Sulfatase. *Marine Biotechnology, 15*, 265-274.
- 490 Richardson, R. K., & Goycoolea, F. M. (1994). Rheological measurement of kappa-
- 491 carrageenan during gelation. *Carbohydrate Polymers*, 24, 223-225.
- 492 Rochas, C., & Rinaudo, M. (1984). Mechanism of gel formation in κ-Carrageenan.
- 493 *Biopolymers, 23*(4), 735-745.

- Running, C. A., Falshaw, R., & Janaswamy, S. (2012). Trivalent iron induced gelation in
- lambda-carrageenan. *Carbohydrate Polymers*, 87, 2735–2739.
- 496 Souza, H. K. S., Hilliou, L., Bastos, M., & Gonçalves, M. P. (2011). Effect of molecular
- 497 weight and chemical structure on thermal and rheological properties of gelling k/i-hybrid
- 498 carrageenan solutions. *Carbohydrate Polymers*, 85, 429-438.
- Thrimawithana, T. R., Young, S., Dunstan, D. E., & Alany, R. G. (2010). Texture and
- 500 rheological characterization of kappa and iota carrageenan in the presence of counter ions.
- 501 *Carbohydrate Polymers*, 82, 69-77.
- 502 Ueda, K., Saiki, M., & Brady, J. W. (2001). Molecular Dynamics Simulation and NMR Study
- of Aqueous Neocarrabiose 4(1)-Sulfate, a Building Block of K-Carrageenan. Journal of
- 504 *Physical Chemistry B, 105*, 8629-8638.
- 505 Usov, A. I. (2011). Polysaccharides of the red algae. Advances in Carbohydrate Chemistry
- 506 *and Biochemistry, 65*, 115-217.

514

- Viebke, C., Borgström, J., & Piculell, L. (1995). Characterisation of kappa- and iota-
- carrageenan coils and helices by MALLS/GPC. Carbohydrate Polymers, 27, 145-154.
- Zablackis, E., & Santos, G. A. (1986). The Carrageenan of Catenella nipae Zanard, a Marine
- 510 Red Alga. Botanica Marina, 29, 319-322.
- 511 Zhang, Y., Yang, N., Zhang, Y., Hou, J., Han, H., Jin, Z., Shen, Y., & Guo, S. (2020). Effects
- of κ-carrageenan on pullulan's rheological and texture properties as well as pullulan hard
- capsule performances. *Carbohydrate Polymers*, 238, 116190.