



HAL
open science

Microbial ecology of the atmosphere

Tina Šantl-Temkiv, Pierre Amato, Emilio Casamayor, Patrick Lee, Stephen Pointing

► **To cite this version:**

Tina Šantl-Temkiv, Pierre Amato, Emilio Casamayor, Patrick Lee, Stephen Pointing. Microbial ecology of the atmosphere. *FEMS Microbiology Reviews*, 2022, 46 (4), 10.1093/femsre/fuac009 . hal-03806309

HAL Id: hal-03806309

<https://hal.science/hal-03806309>

Submitted on 7 Oct 2022

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

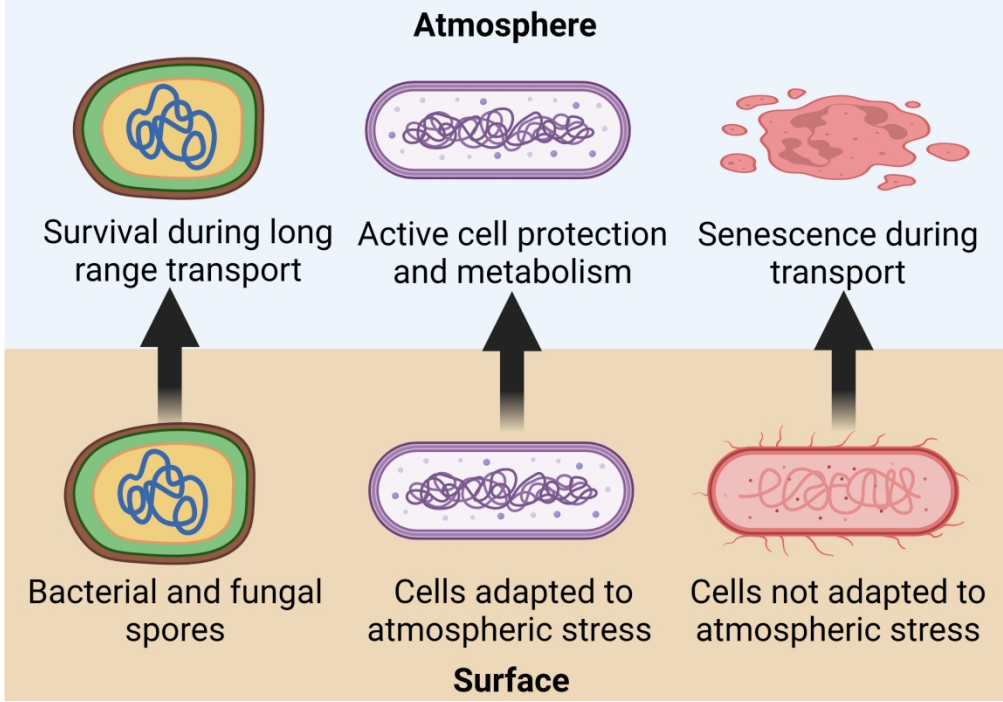
<http://mc.manuscriptcentral.com/fems>

Microbial ecology of the atmosphere

Journal:	<i>FEMS Microbiology Reviews</i>
Manuscript ID	FEMSRE-21-10-0057
Manuscript Type:	Review Article
Date Submitted by the Author:	31-Oct-2021
Complete List of Authors:	Santl-Temkiv, Tina; Aarhus University, Department of Biology Amato, Pierre; Université Clermont Auvergne, ICCF Casamayor, Emilio; Centre Estudis Avancats-CSIC, Integrative Freshwater Ecology Lee, Patrick; City University of Hong Kong, School of Energy and Environment Pointing, Stephen; National University of Singapore, Yale-NUS College
Keywords:	Aeromicrobiology, Ecosystems ecology, Microbial biogeography, Microbial ecology, Microbial dispersal, Atmospheric microbiology

SCHOLARONE™
Manuscripts

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60



645x452mm (118 x 118 DPI)

1
2
3 **1 Microbial ecology of the atmosphere**
4

5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

3 Tina Šantl-Temkiv^{1,2}, Pierre Amato³, Emilio O. Casamayor⁴, Patrick K.H. Lee^{5,6}, Stephen

4 B. Pointing^{7,8*}

6 ¹ *Department of Biology, Aarhus University, Aarhus, Denmark*

7 ² *Stellar Astrophysics Centre, Department of Physics and Astronomy, Aarhus University,*
8 *Aarhus, Denmark*

9 ³ *Institut de Chimie de Clermont-Ferrand, SIGMA Clermont, CNRS, Université Clermont*
10 *Auvergne, Clermont-Ferrand, France*

11 ⁴ *Centre for Advanced Studies of Blanes, Spanish Council for Research (CSIC), Blanes, Spain*

12 ⁵ *School of Energy and Environment, City University of Hong Kong, Hong Kong, China*

13 ⁶ *State Key Laboratory of Marine Pollution, City University of Hong Kong, Hong Kong,*
14 *China*

15 ⁷ *Yale-NUS College, National University of Singapore, Singapore*

16 ⁸ *Department of Biological Sciences, National University of Singapore, Singapore*

17 *e-mail: stephen.pointing@yale-nus.edu.sg

19 **Summary**

20 This review highlights the ecological importance of atmospheric microorganisms with
21 particular focus on recent advances in environmental metagenomics, a critical
22 assessment of the evidence for microbially-mediated atmospheric biochemical and
23 biophysical transformations and consideration of potential impacts due to
24 anthropogenic forcing.

26 **Keywords**

27 Aeromicrobiology, Atmospheric microbiology, Ecosystems ecology, Microbial
28 biogeography, Microbial ecology, Microbial dispersal

30 **Abstract**

31 The atmosphere facilitates connectivity between microbial habitats across multiple
32 spatial scales via airborne transport of cells, propagules and biomolecules. Airborne
33 microorganisms are also implicated in biochemical transformation of atmospheric
34 compounds and biophysical ice nucleation that influence climate processes. Recent
35 advances in methodology for interrogating ultra-low biomass habitats such as air have
36 led to significant advances in understanding. In this review, the contribution of
37 environmental metagenomics to atmospheric diversity estimation, evidence for diverse
38 metabolic transformations, and elucidation of ice nucleation mechanisms are critically
39 assessed, and knowledge gaps identified. The central importance of the atmospheric
40 microbiota to macroecological outcomes including long-distance pathogen dispersal is
41 highlighted. The potential for a metabolically active atmospheric microbiome is
42 evaluated and microbial influence on the water cycle is highlighted. Finally, possible

1
2
3 43 impacts of anthropogenic forcing on the atmospheric microbiota and ecological
4
5 44 outcomes are identified.
6
7
8 45

10 46 **Introduction**

11
12
13 47 Microorganisms have been extensively studied in terrestrial and aquatic natural
14
15 48 environments where an implicit and persistent assumption for observed ecological
16
17 49 patterns has been that atmospheric transport between habitats is essentially a neutral
18
19 50 process due to the extensive aerosolization and long-range transport of cells. This was
20
21 51 first embodied in the classic “everything is everywhere, but, the environment selects”
22
23 52 statement by Lourens Baas Becking and has driven hypotheses in microbial ecology
24
25 53 research for decades (de Wit and Bouvier 2006). Evidence for the distribution of
26
27 54 microorganisms in the atmosphere (aeromicrobiology) has been reported since the
28
29 55 earliest days of modern life science, with visionaries including Louis Pasteur and
30
31 56 Charles Darwin identifying airborne microorganisms (Schwartz 2001; Gorbushina *et al.*
32
33 57 2007).
34
35
36
37
38

39 58 A major challenge for aeromicrobiology has been that traditional cultivation
40
41 59 based enumeration is strongly biased towards cultivable taxa (Šantl-Temkiv *et al.*
42
43 60 2020), and even the most modern high-throughput sequencing approaches that more
44
45 61 fully capture environmental diversity are subject to bias from contaminant signal due to
46
47 62 the ultra-low biomass of microorganisms in air (Salter *et al.* 2014). Recent advances in
48
49 63 high-volume air mass recovery and application of bioinformatics approaches to
50
51 64 decontamination of environmental sequence data have led to significant novel insight,
52
53 65 e.g. (Cáliz *et al.* 2018; Gusareva *et al.* 2019; Tignat-Perrier *et al.* 2019; Archer *et al.*
54
55 66 2021), thus theoretical frameworks in ecology that challenge the traditional notion of
56
57
58
59
60

1
2
3 67 ubiquitous microbial distribution in the atmosphere, e.g. (Dumbrell *et al.* 2010; Lowe
4
5 68 and McPeck 2014) are being validated and revised.
6
7

8 69 It is now apparent that the atmosphere supports vast and highly variable
9
10 70 numbers of microbial cells, pollen, propagules, cell fragments and biomolecules
11
12 71 (Després *et al.* 2012), which comprise the primary biological aerosol particles (PBAP)
13
14 72 estimated to account for 25% of all particles larger than 200 nm (Jaenicke 2005).
15
16 73 Overall, the majority of cells in the atmosphere are likely metabolically inactive and are
17
18 74 undergoing transport between source and sink destinations with a high turnover
19
20 75 (Burrows *et al.* 2009). The fate of the viable fraction of PBAP is important to dispersal
21
22 76 outcomes in terrestrial and marine ecosystems globally (Hanson *et al.* 2012), where the
23
24 77 heterogenous air microbiota may strongly influence ecological outcomes, e.g. affecting
25
26 78 resilience through acting as microbial “seed banks” (Lennon and Jones 2011). In
27
28 79 addition, the recognition that some microorganisms mediate biochemical (Deguillaume
29
30 80 *et al.* 2008) and biophysical transformations (Morris *et al.* 2008) during brief residence
31
32 81 periods in the atmosphere offers the prospect of an active microbial contribution to
33
34 82 atmospheric processes. Finally, PBAP have important health-related effects, through the
35
36 83 dispersal of animal (Lai, Emberlin and Colbeck 2009) and plant (Brown and Hovmøller
37
38 84 2002) pathogens, gene-flow of antibiotic resistance genes (Pal *et al.* 2016), and
39
40 85 microbially derived allergens (Woo *et al.* 2013).
41
42
43
44
45
46
47

48 86 In this review, the unique complexity of the atmospheric environment and its
49
50 87 microbiota is identified and the extent and limitations in our current understanding of
51
52 88 its ecological relevance to other biomes is considered. The scope encompasses
53
54 89 microorganisms that occur in outdoor atmospheric air for extended periods or travel
55
56 90 across regional or inter-continental distances. The short-range dispersal of human
57
58 91 pathogens facilitated by exhaled respiratory droplets in air (Zhang *et al.* 2020) and
59
60

1
2
3 92 airborne transmission within the indoor built environment (Gilbert and Stephens 2018)
4
5 93 are excluded. Specific focus is given to how insight from environmental metagenomics
6
7 94 has identified atmospheric microbiology as key to understanding the interdependence
8
9 95 of ecosystems via flux of diverse taxa between atmosphere and surface habitats, and
10
11 96 critical examination of the genetic and physiological evidence for microbial activity in
12
13 97 the atmosphere. Furthermore, given the atmosphere is facing profound change due to
14
15 98 anthropogenic emissions, consideration is given to how this may impact its
16
17 99 microbiology in future.
18
19
20
21
22
23

24 101 **Boundaries for habitability in the atmosphere**

25
26 102 The atmosphere comprises a series of vertically delineated concentric layers whose
27
28 103 altitude and depth are primarily defined by thermal properties (FIG. 1a). The
29
30 104 troposphere is the lowermost atmospheric layer that contains 75% of its molecular and
31
32 105 gaseous mass as well as most water vapour and atmospheric particles (Barry and
33
34 106 Chorley 2010), including the majority of microbial cells (FIG. 1b). This is where water
35
36 107 cycles through clouds, precipitation and surface environments and it includes the
37
38 108 atmospheric boundary layer (a.k.a. planetary boundary layer), which is in direct contact
39
40
41 109 with the surface, and the free troposphere, which is of great relevance for regional and
42
43 110 global dispersal due to long-range movement of air. Vertical mixing above the
44
45 111 troposphere is limited due to thermal inversion. The bulk of atmospheric ozone is
46
47
48 112 located in the lower stratosphere and this is thought to represent the altitudinal limit to
49
50 113 survival for the majority of microbial cells due to lethal UV exposure at higher
51
52
53 114 altitudes as a function of reduced attenuation by ozone and extended residence time for
54
55
56 115 suspended biological particles at higher altitudes (Bryan *et al.* 2019).
57
58
59
60

1
2
3 116 The troposphere presents an extremely challenging but nonetheless potentially
4
5 117 habitable environmental gradient for microorganisms. The presence of sufficient liquid
6
7
8 118 water in cells – a fundamental pre-requisite for any microbial activity – is subject to
9
10 119 atmospheric relative humidity that is determined by physical processes and
11
12 120 evapotranspiration. In the troposphere, relative humidity is generally bimodal, with
13
14
15 121 higher values in the lower and upper troposphere and a drier mid-troposphere (FIG. 1b)
16
17 122 (Gettelman *et al.* 2006). The lifted condensation level (LCL) is the altitude at which
18
19 123 vapour pressure of an air parcel becomes saturated and above the LCL, at slight
20
21
22 124 supersaturations, the water vapour will condense on aerosol particles and form clouds.
23
24 125 LCL varies widely depending on temperature and water vapour content, which fluctuate
25
26 126 with latitude, location and time, determining the cloud patterns (FIG. 1b). Global
27
28 127 differences reflect atmospheric circulation, surface climate and cloud cover, and range
29
30 128 from saturation at equatorial latitudes to near-zero above some polar locations.
31
32
33 129 Microbial exploitation of low moisture in xeric environments (Pointing and Belnap
34
35 130 2012; Lebre, De Maayer and Cowan 2017) suggests that sub-saturated relative humidity
36
37 131 levels throughout the troposphere are not inhibitory to adapted microbial taxa although
38
39 132 the lower limit of relative humidity that supports active metabolism has not been
40
41
42 133 determined. In contrast, only regions of high relative humidity and clouds are likely to
43
44 134 support the majority of microorganisms not specifically adapted to low water activity.
45
46
47 135 Cloud droplets provide aqueous microenvironments (Joly *et al.* 2015), also the thin
48
49 136 layer of liquid water that occurs at the surface of deliquescent aerosol particles may be
50
51
52 137 sufficient to sustain microbial activity (Stevenson *et al.* 2015a), but future research will
53
54 138 be key to understand the relationship between hygroscopicity and activity of airborne
55
56 139 cells.
57
58
59
60

1
2
3 140 Among other major challenges to microbial survival and proliferation in the
4
5 141 troposphere are the level of photo-oxidative and thermal stress (FIG. 1b). A strong
6
7 142 altitude-dependent gradient occurs in the troposphere due to the attenuation by ozone,
8
9 143 oxygen, water vapour and particulates, and overall UV-A and UV-B radiation increases
10
11 144 approximately 10-20% with every km in altitude (Barry and Chorley 2010). Whilst UV
12
13 145 radiation exposure *per se* is a strong environmental filter to microbial survival and
14
15 146 influences diurnal bacterial occurrence in air (Tong and Lighthart 2006), it is unlikely to
16
17 147 be a limiting factor to radiation-tolerant taxa. Air temperature decreases with altitude in
18
19 148 the troposphere at the rate of approximately 6.5°C per km from ambient near-surface
20
21 149 temperatures to -50°C or lower at the tropopause (NOAA, NASA and USAirForce 1976)
22
23 150 in the absence of inversions. The freezing isotherm occurs at variable altitudes between
24
25 151 ground level and about 5km according to season, latitude, terrain, and atmospheric
26
27 152 conditions (FIG. 1b). This delineates the mechanism of precipitation formation and the
28
29 153 altitudinal limit for non-psychrophilic microbial metabolism. Microbial metabolism has
30
31 154 been demonstrated at temperatures below those encountered even in the uppermost
32
33 155 region of the troposphere (Amato and Christner 2009), and so whilst the freezing
34
35 156 isotherm undoubtedly represents a physiological boundary, temperature *per se* is
36
37 157 unlikely to inhibit potential microbial metabolism or survival in the troposphere.

38
39 158 A peculiarity of the atmosphere contrasting with other environments such as
40
41 159 soils and oceans is its highly dynamic nature, with high cell turnover rates (Burrows *et*
42
43 160 *al.* 2009). Temperature, humidity, and liquid water content can change within minutes,
44
45 161 exposing cells to repeated thermal and osmotic shock, and freeze-thaw cycles during
46
47 162 residence in air. Additionally, as air density decreases with altitude so does the spatial
48
49 163 proximity of cells and their potential substrates. The limited availability of surfaces in
50
51 164 the atmosphere is also a challenge to microbial community development as it limits
52
53
54
55
56
57
58
59
60

1
2
3 165 conventional biofilm formation and the possibility for microbial interactions, e.g. via
4
5 166 quorum sensing (Monier and Lindow 2003). Overall, the atmosphere has been
6
7
8 167 described as possessing qualities of a chaotic system in terms of its physicochemical
9
10 168 complexity (Hochman *et al.* 2019), and this underlines its heterogeneity as a microbial
11
12 169 environment.
13
14

15 170

17 171 **Diversity and biogeography of atmospheric microorganisms**

19 172 The physical volume of the troposphere dwarfs that of the equivalent microbially
20
21
22 173 inhabited layers of the ocean (i.e. the photic zone) or land (i.e. topsoil) although
23
24 174 microbial abundance is several orders of magnitude lower (FIG. 2). On a global scale,
25
26 175 modelled atmospheric cell concentrations fall within the range of 10^0 - 10^5 per m^3 for
27
28 176 bacteria and fungi (Burrows *et al.* 2009; Hoose, Kristjánsson and Burrows 2010;
29
30 177 Spracklen and Heald 2014), and observed geographic patterns in relative abundance
31
32 178 are broadly congruent with modelled estimates of biogeography for cell numbers
33
34 179 (Mayol *et al.* 2017; Tignat-Perrier *et al.* 2019; Archer *et al.* 2021) (FIG. 2a,b). Estimates
35
36 180 from fresh snowfall and from cloud water suggest $4 \times 10^2 - 4 \times 10^3$ bacterial cells per m^3
37
38 181 and $4 \times 10^0 - 4 \times 10^2$ eukaryotic cells per m^3 assuming condensed water content of 0.4 g
39
40 182 per m^3 (Christner *et al.* 2008b; Tesson, Skjøth and Šantl-temkiv 2016; Amato *et al.* 2017;
41
42 183 Hu *et al.* 2018; Šantl-Temkiv *et al.* 2019). Cells associated with aerosolized desert dust
43
44 184 may reach 10^7 per m^3 (Maki *et al.* 2016). Aggregations of cells may create localised
45
46 185 elevated concentrations of free biomass or on surfaces of airborne particulates.
47
48
49

51
52 186 Localised concentrations of microorganisms within clouds and particulate dust
53
54 187 plumes are likely to be significant: The global total mass of clouds has been estimated at
55
56 188 194,000 Tg (Pruppacher and Jaenicke 1995), and the fraction of the atmosphere under
57
58 189 cloud cover has been estimated at 67% globally (King *et al.* 2013). Up to two billion
59
60

1
2
3 190 tonnes of desert dust is transported in the atmosphere annually (Shepherd *et al.* 2016),
4
5 191 and a comparable volume of soil particulates are also emitted to the atmosphere by
6
7
8 192 arable lands every year (Borrelli *et al.* 2017). Attempts to extrapolate these numbers to
9
10 193 yield global atmospheric cell abundance have a high level of uncertainty and have not
11
12 194 been able to adequately encompass variability between boundary layer, troposphere,
13
14 195 stratosphere, clouds and dust. Nonetheless, a global scale meta-analysis of bacterial
15
16 196 abundance across biomes indicated approximately 5×10^{19} cells may occur globally in
17
18 197 the atmospheric boundary layer (Whitman, Coleman and Wiebe 1998). This estimate
19
20 198 was several orders of magnitude lower than those for ocean, soil and subsurface
21
22 199 biomes, and reflects the extremely low density of cells in the atmosphere.
23
24
25

26
27 200 Direct cell counting, e.g. (Mayol *et al.* 2017), estimates from environmental
28
29 201 metagenomics, e.g. (Bryan *et al.* 2019; Gusareva *et al.* 2019; Tignat-Perrier *et al.* 2019;
30
31 202 Archer *et al.* 2021), and historical cultivation approaches (Després *et al.* 2012),
32
33 203 collectively indicate that the most abundant microorganisms within the atmospheric
34
35 204 boundary layer globally are bacteria and fungi. Other microbial components such as
36
37 205 archaea and protists appear to comprise a relatively minor component in air (Gusareva
38
39 206 *et al.* 2019; Archer *et al.* 2021). Few studies have focused on viruses although there is
40
41 207 evidence that a significant volume of free and attached viral particles occurs in air
42
43 208 within and above the atmospheric boundary layer (Whon *et al.* 2012; Reche *et al.* 2018).
44
45 209 Cell-free microbial metabolites and cell fragments constitute a significant portion of
46
47 210 suspended particulate organic matter in air, above both terrestrial and ocean surfaces
48
49 211 where they are important for cloud ice formation (Wilson *et al.* 2015; Tobo *et al.* 2019).
50
51
52
53

54
55 212 Microbiological diversity estimations for the atmosphere have been strongly
56
57 213 influenced by methodological biases (Šantl-Temkiv *et al.* 2020). Direct comparisons of
58
59 214 cultivation versus molecular ecological approaches have indicated very few
60

1
2
3 215 atmospheric microorganisms are cultivable, e.g. (Temkiv *et al.* 2012), and this is
4
5 216 supported by order of magnitude higher diversity estimates from environmental
6
7 217 metagenomics surveys, e.g. (Cáliz *et al.* 2018; Tignat-Perrier *et al.* 2019). The recent
8
9 218 emergence of portable high-volume air sampling devices has greatly helped in recovery
10
11 219 of sufficient biomass for environmental metagenomics studies with sample sizes of
12
13 220 several cubic metres of air now possible during short timeframes (Šantl-Temkiv *et al.*
14
15 221 2020). A persistent issue for diversity estimation has been in distinguishing genuine
16
17 222 microbial signatures from the unavoidable contaminants that occur from equipment
18
19 223 and reagents when interrogating ultra-low biomass samples. Assessment of
20
21 224 contamination during sequence-based diversity estimation indicates that at sample
22
23 225 DNA template concentrations encountered in air contaminants may form a significant
24
25 226 component of sequences (Salter *et al.* 2014). For this reason a rigorous approach to
26
27 227 mitigation is required during experimental and bioinformatic workflows (Eisenhofer *et*
28
29 228 *al.* 2019). Therefore, it is also necessary to strongly nuance findings where
30
31 229 decontamination effort and outcome were not fully reported. Nonetheless, a growing
32
33 230 body of recent research indicates a growing consensus in the understanding of
34
35 231 microorganisms in bulk air (i.e. including free cells, aggregates and particulate-
36
37 232 associated cells). Taxa adapted to survival under high UV irradiance, moisture limitation
38
39 233 and other challenges are over-represented in atmospheric communities which is
40
41 234 indicative of strong environmental filtering. This is supported by evidence that
42
43 235 atmospheric microbial communities exhibit distinct phylogenetic structuring within the
44
45 236 boundary layer and at higher altitudes (Tignat-Perrier *et al.* 2019; Archer *et al.* 2021).
46
47 237 Across broad geographic, altitudinal and temporal scales in the troposphere taxonomic
48
49 238 diversity broadly reflects that of underlying surface source habitats, and commonly
50
51 239 encountered bacterial taxa belong to the Actinobacteria, Alphaproteobacteria,
52
53
54
55
56
57
58
59
60

1
2
3 240 Firmicutes and Gammaproteobacteria (FIG. 2c); and commonly encountered fungi in air
4
5 241 largely comprise spores of Agaricomycetes and Dothidiomycetes (Cáliz *et al.* 2018; Els
6
7 242 *et al.* 2019b; Gusareva *et al.* 2019; Tignat-Perrier *et al.* 2019; Archer *et al.* 2021) (FIG.
8
9
10 243 2c).

11
12 244 Global air circulation patterns have been proposed to define boundaries of
13
14 245 biogeographic regions for atmospheric microorganisms (FIG. 1a) (Womack, Bohannan
15
16 246 and Green 2010). This is supported by recent evidence for regionalisation (Uetake *et al.*
17
18 247 2020) and biogeographic patterns in atmospheric microbial diversity on a global scale
19
20 248 (Archer *et al.* 2021). A high microbial species richness within a given air mass in the
21
22 249 atmospheric boundary layer reflects recruitment from a variety of terrestrial and
23
24 250 marine sources, and source tracking has indicated combined influence of local and
25
26 251 distantly recruited taxa to observed diversity (Archer *et al.* 2021). Diversity estimates
27
28 252 for near-ground air within the atmospheric boundary layer at various locales have
29
30 253 indicated bacterial and fungal communities that were correlated with local abiotic
31
32 254 variables such as temperature, precipitation and humidity or land use/surface cover,
33
34 255 reflecting the role of local cell emissions (Bowers *et al.* 2011; Fröhlich-Nowoisky *et al.*
35
36 256 2012; Tignat-Perrier *et al.* 2019; Archer *et al.* 2021; Spring *et al.* 2021). Thus, diversity
37
38 257 of fungi is relatively higher above regions with a well-developed phyllosphere
39
40 258 (Lymperopoulou, Adams and Lindow 2016; Archer *et al.* 2021; Zhou *et al.* 2021), and
41
42 259 lowest above oceans (Mayol *et al.* 2017; Archer *et al.* 2020; Uetake *et al.* 2020) and low
43
44 260 productivity terrestrial systems such as drylands and polar/alpine regions where
45
46 261 underlying surface communities are less diverse and microbially dominated by bacteria
47
48 262 (Archer *et al.* 2019, 2021). Several studies have also related diversity to factors
49
50 263 associated with dispersal such as history of air masses and UV radiation, suggesting
51
52 264 combined influence of predominant remote sources and conditions during transit
53
54
55
56
57
58
59
60

1
2
3 265 (*Bowers et al.* 2012; *Cáliz et al.* 2018; *Archer et al.* 2019, 2020; *Els et al.* 2019b; *Uetake*
4
5 266 *et al.* 2019; *Tignat-Perrier et al.* 2020).

7
8 267 Temporal patterns in local atmospheric microbiota reflect seasonal changes in
9
10 268 underlying surface cover and source of incoming air mass. A pronounced diel variation
11
12 269 of specific bacterial and fungal groups was found in the equatorial tropics using
13
14 270 metagenomics (*Gusareva et al.* 2019). Inter-seasonal variation in the atmospheric
15
16 271 boundary layer was found to be absent (*Gusareva et al.* 2019), weak (*Tignat-Perrier et*
17
18 272 *al.* 2020), pronounced for some taxa (*Bowers et al.* 2012) or stochastic (*Els et al.*
19
20 273 2019b), depending on the location. Still, a large number of studies have identified
21
22 274 differences in bacterial and/or fungal communities in summer versus winter at
23
24 275 temperate and sub-tropical terrestrial locations (*Bowers et al.* 2012, 2013; *Woo et al.*
25
26 276 2013; *Barberán et al.* 2014; *Cáliz et al.* 2018; *Els et al.* 2019b; *Uetake et al.* 2019). One of
27
28 277 the scarce continuous long-term studies demonstrated a clear and well-delineated
29
30 278 seasonal pattern in bacterial and fungal communities (*Woo et al.* 2013). At polar
31
32 279 latitudes elevated cell abundance in air corresponded to summer and the growing
33
34 280 season (*Šantl-Temkiv et al.* 2019). These studies strengthen the view that the
35
36 281 underlying phyllosphere is a major determinant of atmospheric microbial diversity. As
37
38 282 the elevation above ground increases, the source area of influence widens (*Schmid*
39
40 283 2002; *Hsieh and Katul* 2009), and the small-scale spatial and temporal heterogeneities
41
42 284 observed near the ground blur (*Archer et al.* 2019, 2021; *Els et al.* 2019a). Above the
43
44 285 atmospheric boundary layer long-range air movements progressively decouple
45
46 286 atmospheric diversity from its emission sources (*Burrows et al.* 2009) (*Smith et al.*
47
48 287 2018).

49
50 288 Whilst most research has focused on the atmospheric boundary layer and higher
51
52 289 altitudes within the troposphere, understanding the biogeographic limit of microbial

1
2
3 290 occurrence and survival at higher altitudes is also of interest. It has been postulated that
4
5 291 limited microbial survival and transport may occur in the stratosphere (DasSarma and
6
7
8 292 DasSarma 2018), but the role of the stratosphere in microbial dispersal may be limited
9
10 293 due to the combined effects of extreme environmental stress and extended residence
11
12 294 times (Bryan *et al.* 2019). Several studies have employed balloon and aircraft platforms
13
14 295 to recover stratospheric air samples and they have described a limited diversity and
15
16 296 viability of bacterial and fungal taxa (Wainwright *et al.* 2003; Smith, Griffin and
17
18 297 Schuergler 2010; Smith *et al.* 2011, 2018; Bryan *et al.* 2019), with taxa not markedly
19
20 298 differentiated from those occurring in the troposphere below (Smith *et al.* 2018).
21
22 299 Estimates of abundance vary greatly but an upper estimate concluded that bacterial
23
24 300 abundance in the troposphere and lower stratosphere may be comparable (Bryan *et al.*
25
26 301 2019). However, extreme caution is required during interpretation of results from
27
28 302 stratospheric air samples due to the major uncertainty surrounding effective exclusion
29
30 303 of the unavoidable contaminants that occur when interrogating the ultra-low biomass
31
32 304 system. There is currently insufficient evidence to draw robust conclusions about the
33
34 305 occurrence, transport, or viability of microorganisms in the stratosphere. The discipline
35
36 306 of astrobiology has considered the potential role of the stratosphere and upper
37
38 307 atmosphere for interplanetary dispersal of microorganisms (panspermia) as free cells
39
40 308 or via mineral vectors (Yang, Yokobori and Yamagishi 2009) but this will require major
41
42 309 investigative effort to be further considered.
43
44
45
46
47
48
49
50
51

52 311 **What are the determinants of microbial flux between the atmosphere and surface**
53
54 312 **habitats?**

55
56
57 313 The atmosphere has long been identified as a major conduit for microbial transport
58
59 314 between surface habitats, e.g. (Womack, Bohannan and Green 2010), and this has a
60

1
2
3 315 profound impact on the function and resilience of terrestrial and marine surface
4
5 316 communities due to its contribution to gene flow, community assembly, and dispersal of
6
7
8 317 pathogens. Recent developments in ecological theory have begun to challenge the view
9
10 318 that microbial transport is a neutral process (Lowe and McPeck 2014), and recent
11
12 319 evidence from large scale biogeographic (Mayol *et al.* 2017; Archer *et al.* 2021) and
13
14 320 temporal (Cáliz *et al.* 2018; Tignat-Perrier *et al.* 2020) studies of atmospheric
15
16 321 microbiota increasingly support this view. Deterministic influence due to
17
18 322 environmental filtering and microbial stress tolerance both play a significant role in
19
20 323 shaping atmospheric microbiota (Archer *et al.* 2021), and hence the composition of
21
22 324 assemblages that are exchanged between atmosphere and surface habitats.

26 325 The major determinants of microbial fluxes in the atmosphere between source
27
28 326 and sink habitats are emission, survival during transport, and deposition (FIG. 3).
29
30 327 Emission from the surface, known as aerosolization, is dependent upon the substrate
31
32 328 and a major factor is the nature of the interface with air, such as the composition of
33
34 329 plant cover (Zhou *et al.* 2021), cohesiveness of soil (Joung, Ge and Buie 2017; Archer *et*
35
36 330 *al.* 2021), or chemistry of the water-surface microlayer (Michaud *et al.* 2018) (FIG. 3a).
37
38 331 Biotic microbial traits that affect aerosolization are also important, including cell
39
40 332 hydrophobicity and membrane composition (Michaud *et al.* 2018), extracellular
41
42 333 polymeric substance (EPS) production (Morris and Monier 2003), spore production and
43
44 334 release (Lagomarsino Oneto *et al.* 2020), and allometry (Norros *et al.* 2014). Bacterial
45
46 335 cells occur as free cells or are attached to particulate matter whereas the fungal spores
47
48 336 likely occur mostly as freely suspended particles. Modelling emissions for particulates
49
50 337 1-3µm diameter as a proxy for single and particle-associated bacterial cells revealed
51
52 338 that the total annual emissions were in the range of 7.6×10^{23} - 3.5×10^{24} cells largely
53
54 339 originating from terrestrial environments, i.e. crops, grasslands and shrubs (Burrows *et*

1
2
3 340 *al.* 2009, 2013). This equated to an estimated annual emission of 470–1100 Gg bacterial
4
5 341 biomass (Burrows *et al.* 2013). Fungal emissions occur predominantly as free spores
6
7 342 and were estimated at 9.5×10^{23} total spores from terrestrial surfaces per year which
8
9 343 amounts to ~50 Tg of fungal biomass annually (Elbert *et al.* 2007).

12 344 The major marine aerosolization process is initiated by breaking waves which
13
14 345 produces small air bubbles that scavenge microorganisms and organic compounds from
15
16 346 the sea column and transport them to the sea surface (Aller *et al.* 2005)(Carlucci and
17
18 347 Williams 1965), where they burst to eject small droplets containing salts, biogenic
19
20 348 material and microorganisms (Blanchard and Syzdek 1970), into the atmosphere (Afeti
21
22 349 and Resch 1990)(Blanchard 1989). Even though oceans are thought to represent weak
23
24 350 sources of airborne microorganisms on a global per surface area basis and bioaerosols
25
26 351 largely comprise bacteria and viruses (Wilson *et al.* 2015), selective transfer and
27
28 352 enrichment of organic compounds and microbial cells has been shown experimentally
29
30 353 (Rastelli *et al.* 2017). In addition, large spatial and temporal variability in marine
31
32 354 emissions and effects on atmospheric processes may occur due to biological processes
33
34 355 and meteorological factors (Wang *et al.* 2015; Wilson *et al.* 2015; DeMott *et al.* 2016).

37 356 Survival during transit in air is influenced by biotic traits related to stress
38
39 357 tolerance, and particularly with regard to mitigating the effects of low water activity
40
41 358 (Stevenson *et al.* 2015b), desiccation (Potts 1994), photo-oxidative UV (Daly 2009;
42
43 359 Sharma *et al.* 2017) and general oxidative stress (Ziegelhoffer and Donohue 2009;
44
45 360 Ezraty *et al.* 2017), and low temperatures/freezing (Amato 2013) (FIG. 4). Dormancy
46
47 361 and sporulation are strategies that provide protection against major stressors and
48
49 362 functional metagenomics of air has indicated genes associated with these traits are
50
51 363 over-represented in atmospheric communities compared to underlying surface habitats
52
53 364 (Aalismail *et al.* 2019; Archer *et al.* 2021). Many spores as well as dormant and active
54
55
56
57
58
59
60

1
2
3 365 cells possess pigment compounds such as melanins, mycosprines and scytonemins that
4
5 366 protect against oxidative and desiccation stress largely due to scavenging of reactive
6
7 367 species (Gao and Garcia-Pichel 2011). A number of biodiversity surveys in air have
8
9 368 revealed taxa known to produce protective pigments are over-represented in air
10
11 369 compared to underlying terrestrial and marine habitats (Fröhlich-Nowoisky *et al.* 2012;
12
13 370 Tignat-Perrier *et al.* 2019; Archer *et al.* 2021), and this implies a survival advantage.
14
15 371 Additional strategies to counter stress may include the secretion of extracellular
16
17 372 polymeric substances to mitigating both low water activity via hygroscopic
18
19 373 carbohydrate polymers as well as extracellular protection from oxidative stress
20
21 374 (Matulová *et al.* 2014). Aerosolization of cells as aggregations, biofilms or attached to
22
23 375 particulates may also confer survival advantages during dispersal (Monier and Lindow
24
25 376 2003; Morris and Monier 2003) (FIG. 3), but may also reduce residence time in air due
26
27 377 to increased size and density (Burrows *et al.* 2009). Metagenomic profiling of
28
29 378 atmospheric communities has begun to reveal prevalence of genes encoding cold shock,
30
31 379 oxidative stress, and UV repair enzymes (Aalismail *et al.* 2019; Archer *et al.* 2021), and
32
33 380 metatranscriptomics of cloud water has revealed active metabolic responses to counter
34
35 381 oxidative, osmotic and thermal stresses (Amato *et al.* 2019). Physiological responses to
36
37 382 counter simulated atmospheric stress have also been observed for taxa isolated from
38
39 383 the atmosphere. Strains of *Pseudomonas syringae* isolated from rain and cloud water
40
41 384 withstood aerosolization, freezing, osmotic shock, short-term exposure to UVA and UVB
42
43 385 radiation (Alsved *et al.* 2018; de Araujo *et al.* 2019; Ling *et al.* 2021), and oxidative
44
45 386 stress imposed by hydrogen peroxide (H₂O₂) (Wirgot *et al.* 2019). Metabolomic studies
46
47 387 on *P.syringae* also revealed a suite of responses to cold shock under simulated
48
49 388 atmospheric conditions (Jousse *et al.* 2017), including production of cryoprotectants,
50
51 389 antioxidants, alkaloids and metabolites involved in energy metabolism.
52
53
54
55
56
57
58
59
60

1
2
3 390 The deposition of microorganisms occurs through a combination of wet and dry
4
5 391 deposition (Després *et al.* 2012). The wet deposition is affected by a combination of in-
6
7
8 392 cloud partitioning during cloud droplet / ice particle formation and below-cloud aerosol
9
10 393 scavenging processes, which can both contribute to segregation of taxa between the
11
12 394 wet and dry phases (Bauer *et al.* 2003; Moore *et al.* 2020; Woo and Yamamoto 2020).
13
14
15 395 In particular, cloud condensation ability was modelled to shorten the average bacterial
16
17 396 residence time in air (and hence dispersal distance) from ~8d to 3d (Burrows *et al.*
18
19 397 2009) and ice nucleating microbial cells were shown to be efficiently precipitated
20
21 398 (Amato *et al.* 2015; Stopelli *et al.* 2015) Wet deposition rates of $10^7 - 5 \times 10^9$ bacteria
22
23 399 cells $m^{-2} d^{-1}$ have been reported (Reche *et al.* 2018; Woo and Yamamoto 2020). In
24
25 400 contrast, dry deposition velocity is largely determined by allometric considerations,
26
27 401 such as PBAP size, shape, and density (FIG. 3d). Dry deposition rates of $\sim 10^7 - 5 \times 10^8$
28
29 402 bacteria cells $m^{-2} d^{-1}$ have been reported but with no significant difference in estimated
30
31 403 overall wet and dry deposition (Reche *et al.* 2018; Woo and Yamamoto 2020). Patterns
32
33 404 in deposition of fungal spores and pollen are thought to follow the same pattern where
34
35 405 overall wet and dry deposition contributions are similar (Aylor 2017; Woo and
36
37 406 Yamamoto 2020), although taxon-specific variation is likely.

38
39 407 Aside from the emission and deposition processes, cell fluxes are also affected by
40
41 408 the altitude that the cells reach, which is a deterministic factor related to stress
42
43 409 exposure (FIG. 3b). A very large fraction (97%) of depositing bacterial cells have been
44
45 410 found to be particle associated (Reche *et al.* 2018), and with a large mean diameter (~ 9
46
47 411 μm) (Woo and Yamamoto 2020), thus supporting the idea that the mixing state of cells
48
49 412 in air, i.e. free cells, aggregates or cells vectored on particulate matter, has a significant
50
51 413 effect on cell residence time (FIG. 3c). It should also be noted that processes affecting
52
53 414 atmospheric cell fluxes are highly variable and subject to stochastic influence from
54
55
56
57
58
59
60

1
2
3 415 events such as dust storms (González-Toril *et al.* 2020) and wildfires (Moore *et al.*
4
5 416 2021).

6
7
8 417 A pertinent question for atmospheric communities is the extent to which the
9
10 418 complex variation in emission and deposition flux, together with environmental filtering
11
12 419 during transit, influence the assembly of terrestrial and marine surface communities.
13
14 420 The flux of diversity and biomass from air to soil or oceans is likely smaller by
15
16 421 magnitudes than that of standing communities but given the ability of microorganisms
17
18 422 to rapidly exploit favourable ecological niches, atmospheric flux should be viewed as a
19
20 423 critical control point in aspects of connectivity and ecological resilience across biomes
21
22 424 (Dumbrell *et al.* 2010; Lowe and McPeck 2014; Zhou and Ning 2017). This occurs by
23
24 425 facilitating microbial dispersal between spatially separated natural systems (Kellogg
25
26 426 and Griffin 2006), contributing to maintenance of diversity via recruitment (Jones and
27
28 427 Lennon 2010), and in promoting dispersal of dormant propagules that may act as
29
30 428 microbial seed banks (Lennon and Jones 2011).
31
32
33
34
35

36 429

37 38 430 **What is the evidence for long distance atmospheric pathogen dispersal?**

39
40 431 Some evidence exists for regional dispersal of pathogens without any obvious
41
42 432 adaptation to the environmental stress of atmospheric transport, e.g. foot and mouth
43
44 433 diseases virus (Sorensen *et al.* 2000; Björnham, Sigg and Burman 2020). However, long-
45
46 434 range atmospheric dispersal of pathogens (i.e. beyond short-range inter-organismal
47
48 435 transmission in air) typically requires protection from environmental stress, whether
49
50 436 facilitated by pre-adaptation as spores, or due to association with particulate or
51
52 437 biological vectors. Correlations have been made between atmospheric dust vectoring
53
54 438 and regional trans-continental scale pathogen dispersal (Tobias *et al.* 2019), and
55
56 439 consistent inter-annual dynamics for specific taxa may exist, which make them
57
58
59
60

1
2
3 440 foreseeable over time with dominance of plant pathogens over those of animal and
4
5 441 humans (Triado-Margarit, Caliz and Casamayor 2021).
6
7

8 442 Fungal spores are particularly well suited to long-range dispersal and are
9
10 443 problematic as a threat to agriculture and global food security (Brown and Hovmøller
11
12 444 2002; Savary *et al.* 2019). Many plant pathogens have historically displayed distinct
13
14 445 regionalisation but globalisation of agriculture has exacerbated range shifts (Bebber,
15
16 446 Holmes and Gurr 2014). Airborne fungal pathogens have been detected above land with
17
18 447 agricultural land use (Nicolaisen *et al.* 2017; Archer *et al.* 2021), and some fungal
19
20 448 pathogens are able to disperse across inter-continental distances and re-establish
21
22 449 infections even when host plants are seasonally absent (Brown and Hovmøller 2002). A
23
24 450 striking example is the re-emergence of wheat stem rust as a threat to global food
25
26 451 supply. The disease caused by the basidiomycete fungus *Puccinia graminis* f.sp. *tritici* is
27
28 452 dispersed readily in air by basidiospores. It was virtually eradicated in the twentieth
29
30 453 century, however, a highly pathogenic new strain, Ug99 and its variants, emerged in the
31
32 454 last decade and have threatened to devastate global wheat cultivation (Singh *et al.*
33
34 455 2011). Combination of disease surveillance and dispersal modelling has resulted in
35
36 456 strong evidence for atmospheric transmission as a major determinant of wheat rust
37
38 457 epidemiological zones (Meyer *et al.* 2017).
39
40
41
42
43
44

45 458 Evidence for long-range atmospheric dispersal of human pathogens also exists.
46
47 459 Seasonal outbreaks of meningococcal meningitis in sub-Saharan Africa have been
48
49 460 correlated with bacterial vectoring on dust (Jusot *et al.* 2017), and this also appears to
50
51 461 occur for fungal diseases such as Valley Fever caused by *Coccidioides* spores transmitted
52
53 462 on desert dust across the south western United States and northern Mexico (Kollath,
54
55 463 Miller and Barker 2019). Bacterial plant pathogens may have significantly higher
56
57 464 relative abundance than fungi during intercontinental dust intrusions from Africa to
58
59
60

1
2
3 465 Europe, although predominant western air mass may also bring *Coccidioides* spp., a
4
5 466 potentially severe and understudied fungal pathogen endemic from arid and semiarid
6
7 467 regions of North and Central America, to Europe (Triado-Margarit, Caliz and Casamayor
8
9 468 2021). Vectoring on biological surfaces also provides a protected means of long-
10
11 469 distance dispersal. Wind-borne transport of *Anopheles* mosquitoes carrying the
12
13 470 *Plasmodium* parasite was recorded over several hundred kilometres in the Sahel region
14
15 471 of Africa (Huestis *et al.* 2019), and pollen has been demonstrated as a vector for
16
17 472 bacteria and bacterial allergens (Oteros *et al.* 2019).
18
19
20
21
22 473

23 24 474 **What is the evidence for metabolically active atmospheric microorganisms?**

25
26 475 The majority of cells in the atmosphere are unlikely to be metabolically active but are
27
28 476 simply undergoing transport between source and sink destinations with a high turnover
29
30 477 (Burrows *et al.* 2009). Given that most fungi in air occur as dormant spores it is likely
31
32 478 that the active fraction of atmospheric microbiota is comprised almost exclusively of
33
34 479 bacteria that possess adaptation to atmospheric environmental stress (Archer *et al.*
35
36 480 2021) (FIG. 4). Putative generation times for bacteria in cloud water have been
37
38 481 estimated at 3.6-19.5 days (Sattler, Puxbaum and Psenner 2001), which fit within their
39
40 482 modelled airborne residence times (Burrows *et al.* 2009). However, whether metabolic
41
42 483 transformations *in situ* under the highly fluctuating atmospheric conditions are
43
44 484 sufficient to sustain only quasi-dormancy, or if cell homeostasis and reproduction are
45
46 485 energetically feasible within the short timeframe of favourable conditions offered
47
48 486 during atmospheric transport remains unexplored.

49
50
51
52
53
54 487 Evidence from environmental samples has demonstrated that atmospheric
55
56 488 microorganisms possess the potential to transform substrates encountered in the
57
58 489 atmosphere (FIG. 4). Metagenomic surveys of cloud water (Amato *et al.* 2019), free air

1
2
3 490 (Gusareva *et al.* 2019; Archer *et al.* 2021), and airborne desert dust plumes (Aalismail *et*
4
5 491 *al.* 2019) indicate diverse autotrophic and heterotrophic metabolic pathways in
6
7
8 492 atmospheric assemblages that broadly reflect source environments. Photosynthesis
9
10 493 may be limited in atmospheric microorganisms due to light-inhibition (Barber and
11
12 494 Andersson 1992), but clouds and desert dust provide shading against damaging levels
13
14
15 495 of direct UV radiation and diverse viable and stress-tolerant microalgae and
16
17 496 cyanobacteria have been reported from clouds (Tesson, Skjøth and Šantl-temkiv 2016).
18
19 497 A range of pathways indicating atmospheric trace gas metabolism including hydrogen,
20
21 498 methane and carbon monoxide were widespread among atmospheric metagenomes
22
23
24 499 (Archer *et al.* 2021) and methane oxidation has been demonstrated by atmospheric
25
26 500 isolates of *Methylocystis* and *Methylosinus* (Šantl-Temkiv *et al.* 2013), although it is
27
28 501 unclear if this is energetically feasible under atmospheric conditions. Most research has
29
30 502 focused on the potential for heterotrophic metabolism of simple organic compounds in
31
32 503 clouds as these are envisaged as “hotspots” for atmospheric microbial activity.

33
34
35 504 Microbial assemblages in cloud water have revealed general markers of
36
37 505 metabolic activity under simulated atmospheric conditions including ATP, high
38
39 506 ribosomal content, transcribed RNA, enzymatic and respiratory activities (Klein *et al.*
40
41 507 2016; Šantl-Temkiv *et al.* 2018; Amato *et al.* 2019). Under conditions of favourable
42
43 508 water activity, temperature, substrate availability and reduced stress exposure,
44
45 509 atmospheric microbial assemblages appear capable of active heterotrophic metabolism
46
47 510 as evidenced by chemical speciation in recovered cloud water (Vaitilingom *et al.* 2013;
48
49 511 Bianco *et al.* 2019). A comparative metagenomic and metatranscriptomic study of cloud
50
51 512 water assemblages highlighted that active metabolism in clouds is closely coupled with
52
53 513 the energetic demands of active stress response metabolism and particularly with
54
55 514 regard to mitigating oxidative stress (Amato *et al.* 2019). A complex metabolism was

1
2
3 515 envisaged that also included synthesis of cryoprotectant and extracellular polymeric
4
5 516 substances implicated in a protective role. The study identified C1 and C2 compounds as
6
7
8 517 important substrates and ammonium as the major nitrogen source (Amato *et al.* 2019).
9
10 518 Another combined metatranscriptomic and metabolic screening of cloud water
11
12 519 provided strong evidence for phenol metabolism by bacteria, and this was envisaged as
13
14
15 520 an important atmospheric substrate derived from anthropogenic emissions (Lallement
16
17 521 *et al.* 2018).

19 522 Several studies have demonstrated the ability of atmospheric isolates to
20
21 523 metabolise compounds encountered in clouds: A screening of sixty atmospheric isolates
22
23 524 including the bacterial genera *Bacillus*, *Pseudomonas*, *Micrococcus* and *Sphingomonas*
24
25 525 plus fungal yeasts from cloud water revealed that under laboratory conditions all were
26
27 526 capable of degrading diverse carboxylic acids and volatile organic compounds (Amato
28
29 527 *et al.* 2007). Atmospheric bacterial isolates of *Bacillus* sp, *Frigoribacterium* sp,
30
31 528 *Pseudomonas* sp. and the methanotrophic genera *Methylocystis* sp. and *Methylosinus* sp.
32
33 529 have been demonstrated to oxidize common atmospheric volatile organic compounds
34
35 530 including formaldehyde and methanol (Husárová *et al.* 2011; Šantl-Temkiv *et al.* 2013).
36
37 531 *Rhodococcus enclensis* isolated from cloud water was shown to metabolise catechol and
38
39 532 phenol under simulated atmospheric conditions at rates comparable to that of chemical
40
41 533 transformations in the atmosphere (Jaber *et al.* 2020). Larger molecules may also serve
42
43 534 as substrates, for example a *Bacillus* strain from cloud water was demonstrated to
44
45 535 metabolise a range of pentose and hexose sugars a range of mono- and disaccharides
46
47 536 (Matulová *et al.* 2014).

48 537 Based on bulk cloud water measurements, which were unable to account for the
49
50 538 multiphase structure and high dynamic nature of clouds, it has been proposed that
51
52 539 heterotrophic microbial activity could be the main driver for the oxidation of common
53
54
55
56
57
58
59
60

1
2
3 540 atmospheric organic compounds in air during night and could compete with
4
5 541 photochemistry during the day (Vaithilingom *et al.* 2010). Modeling has indicated that in
6
7 542 clouds where only a small fraction ($\sim 10^{-4}$) of spatially separated microdroplets contains
8
9 543 microbial cells, the quantitative contribution of microbial activity to the transformation
10
11 544 of atmospheric compounds is likely to be low (Fankhauser *et al.* 2019), although this
12
13 545 needs to be verified with experimental approaches simulating *in situ* atmospheric
14
15 546 conditions. Modelling of biodegradation potential for organic compounds by bacteria
16
17 547 in clouds has indicated that acetate and formate are the most utilised substrates and
18
19 548 that overall bacterial biodegradation may represent an underestimated sink for
20
21 549 atmospheric organics (Khaled *et al.* 2020).
22
23
24
25
26
27
28

29 551 **What is the microbial role in atmospheric biophysical processes?**

30
31 552 Microbial involvement in atmospheric biophysical transformations occurs
32
33 553 independently from the metabolic activity that is thought to be rare during short
34
35 554 residence times in the extreme environment. Biological ice nucleation contributes to
36
37 555 cloud formation, precipitation, and the hydrological cycle, as well as impacting
38
39 556 microbial flux between atmospheric and surface habitats, and recent advances have
40
41 557 shed new insight on the mechanistic basis for this. Diverse microorganisms and
42
43 558 biomolecules, including bacteria (particularly Gammaproteobacteria) (Joly *et al.* 2013),
44
45 559 fungi (Fröhlich-Nowoisky *et al.* 2014), microalgae (Tesson and Šantl-Temkiv 2018) and
46
47 560 microbially-derived cell-free ice nucleation proteins (INpro) (O'Sullivan *et al.* 2015) can
48
49 561 nucleate ice in super-cooled cloud droplets at temperatures much warmer than other
50
51 562 atmospheric particles (between -13 and -1°C) and have been implicated in cloud
52
53 563 processes. This property has been linked with specific surface- or excreted proteins and
54
55 564 is essentially a biophysical process that occurs independently of cell viability or
56
57
58
59
60

1
2
3 565 association (FIG. 5). Whilst terrestrial sources are dominant on a global level, marine
4
5 566 sources associated with INpro excreted by marine microalgae (Wilson *et al.* 2015) may
6
7
8 567 be important in remote regions unaffected by continents (e.g. Southern Ocean) (DeMott
9
10 568 *et al.* 2016).

11
12 569 Among identified biological ice nucleation active molecules, the best understood
13
14 570 are bacterial INpro, which are large membrane-associated proteins (>120 kDa) that
15
16 571 structure interfacial water molecules at low temperature promoting ice formation
17
18 572 (Roeters *et al.* 2021) (FIG. 5). Bacterial INpro are long repetitive proteins that present
19
20 573 on the surface of the outer membrane and nucleate ice through a yet to be fully resolved
21
22 574 mechanism (Pandey *et al.* 2016; Hudait *et al.* 2018; Lukas *et al.* 2020). Monomers or
23
24 575 small oligomers nucleate ice at temperatures from -7 to -10 °C and are known as Type C
25
26 576 INpro; and higher order oligomers of INpro form at well-defined sizes that nucleate ice
27
28 577 between -2 to -5 °C and are described to as Type A INpro (Wex *et al.* 2015; Ling *et al.*
29
30 578 2018). Laboratory studies using cloud simulation chambers have shown that the
31
32 579 activity of bacterial INpro is independent of cell viability (Hartmann *et al.* 2013; Amato
33
34 580 *et al.* 2015). Fungal spores and hyphae also carry INpro that exhibit different molecular
35
36 581 structure and properties to those in bacteria and they are yet to be fully characterised.
37
38 582 Fungal spores are the dominant state for fungi in the atmosphere and support surface-
39
40 583 associated INpro and ice nucleating activity (Pouleur *et al.* 1992; Morris *et al.* 2013)
41
42 584 (FIG. 5). Fungal hyphae occur very rarely in the atmosphere and so the major
43
44 585 contribution arises from dissociation of fungal INpro from cell walls and adhesion to soil
45
46 586 particulates that subsequently become aerosolized (O'Sullivan *et al.* 2015) (FIG. 5). The
47
48 587 ubiquitous soil fungal genera *Fusarium* and *Mortierella* have been demonstrated to
49
50 588 excrete INpro into the environment and soil particles with associated INpro were
51
52 589 capable of ice nucleation (Fröhlich-Nowoisky *et al.* 2014; O'Sullivan *et al.* 2015).
53
54
55
56
57
58
59
60

1
2
3 590 Once they induce freezing, Microbial INpro can also contribute to the formation
4
5
6 591 of secondary ice particles in clouds which enhances the impact of low abundance
7
8 592 biological INPs through ice-multiplication, particularly at high sub-zero temperatures
9
10 593 (Korolev and Leisner 2020) (Fig. 5). Rain that is produced via the ice phase through the
11
12 594 Hallett–Mossop process dominates over the midlatitude oceans and continents
13
14 595 (Mülmenstädt *et al.* 2015). This process occurs in mixed phase clouds, in which some of
15
16 596 the supercooled cloud droplets freeze due to the presence of ice nucleating particles,
17
18 597 promoting the deposition of water vapor on thus-formed ice particles and allowing
19
20 598 them to grow to precipitation sizes (Fig. 5). Precipitation induced by ice-nucleation
21
22 599 active bacteria, whose natural habitats are plant leaf surfaces, contributes to the
23
24 600 bioprecipitation cycle by enhancing the growth of vegetation and epiphytic
25
26 601 microorganisms, which in turn leads to enhanced emissions of ice-nucleation active
27
28 602 bacteria resulting in a positive feedback (Fig. 3d) (Morris *et al.* 2014). Bioprecipitation
29
30 603 may be important above regions with a well-developed phyllosphere where emissions
31
32 604 of bacterial INpro are highest, and accompanied by relatively low levels of abiotic INPs
33
34 605 (Huffman *et al.* 2013; Healy *et al.* 2014). Other precipitation-related feedbacks that
35
36 606 increase bioaerosol generation and thus enhance the atmospheric INpro include
37
38 607 immediate and delayed interactions of rain with soil and plants (Huffman *et al.* 2013;
39
40 608 Joung, Ge and Buie 2017). These positive feedbacks occur alongside non-biological ice
41
42 609 nucleation and precipitation and the significance of these processes for meteorology
43
44 610 and climate is yet to be quantified.
45
46
47
48
49
50
51

611

612 **How might anthropogenic activities affect atmospheric microbiota?**

56
57 613 The atmosphere is a primary sink for anthropogenic emissions of greenhouse
58
59 614 gases, chemical pollutants and particulates from fossil fuel combustion (Archer and
60

1
2
3 615 Pointing 2020). Additional particulate emissions occur as a result of biomass burning
4
5 616 due to land clearance and wildfires, soil destabilization due to land use change and
6
7
8 617 expansion of intensive agriculture, and climate-change induced desertification (Archer
9
10 618 and Pointing 2020). Most of these emissions are concentrated in the troposphere where
11
12 619 long-range microbial transport, metabolism and biophysical transformations occur.
13
14
15 620 Atmospheric forcing due to emissions has been acute in the industrial age and this has
16
17 621 the potential to create positive and negative impacts on the dispersal of microorganisms
18
19 622 in air (Archer and Pointing 2020) (FIG. 6). A recent modelling study suggests that
20
21 623 overall microbial diversity in the atmosphere may decline under future climate
22
23 624 scenarios and this may severely impact ecosystem connectivity and function (Ontiveros
24
25 625 *et al.* 2021).

26
27
28
29 626 Gaseous emissions from fossil fuel combustion including SO_x and NO_x have been
30
31 627 shown to inhibit cell function and survival of *Pseudomonas* strains isolated from clouds
32
33 628 (Kondakova *et al.* 2016). Greenhouse gaseous, e.g. methane (Šantl-Temkiv *et al.* 2013),
34
35 629 can be metabolised by atmospheric bacteria. However, as *in situ* microbial activity and
36
37 630 its influence on atmospheric chemistry is still not well constrained, it also remains
38
39 631 unclear if elevated substrate levels in the atmosphere have the potential to significantly
40
41 632 affect microbial activity and its outcomes (Archer and Pointing 2020). The ice
42
43 633 nucleation activity of PBAP may also become more important to precipitation patterns
44
45 634 in a warming world because they are more efficient at warmer temperatures from -10°C
46
47 635 to 0°C, compared to mineral particulates (Christner *et al.* 2008a). The potential impacts
48
49 636 from direct and indirect anthropogenic particulate emissions are more apparent and
50
51 637 widespread. Anthropogenic particulates including brown carbon from biomass
52
53 638 combustion and black carbon from fossil fuel combustion account for approximately
54
55 639 20% of global emissions (Chen *et al.* 2018), and climate change increases mineral dust
56
57
58
59
60

1
2
3 640 aerosolization due to desertification (Pointing and Belnap 2014). Elevated
4
5 641 concentrations of anthropogenic and desert dust particulates have been shown to
6
7 642 influence atmospheric microbial diversity (Maki *et al.* 2019), and this may have
8
9 643 implications for dispersal outcomes at deposition sites. The transport of
10
11 644 microorganisms in plumes of airborne particulates is envisaged to provide additional
12
13 645 protection from UV damage during atmospheric transport (González-Toril *et al.* 2020).
14
15 646 Particulates arising from combustion are more hygroscopic than natural mineral
16
17 647 particles and whilst this may be viewed as facilitating greater survival and even
18
19 648 potentially enhanced microbial activity, it must be balanced against the often toxic
20
21 649 levels of PAH associated with these particulates (Hayakawa *et al.* 2018). Anthropogenic
22
23 650 forcing through particulate emissions may also affect ice nucleation patterns in clouds
24
25 651 and thus impact microbial flux and deposition in the atmosphere more generally
26
27 652 through changes in precipitation. Finally, airborne microorganisms themselves may
28
29 653 become increasingly prevalent due to climate-change forcing or pollution, e.g. marine
30
31 654 phytoplankton blooms (Lewis, van Dijken and Arrigo 2020). This includes strong
32
33 655 emission from point sources for pathogenic microorganisms, antibiotic resistance genes
34
35 656 and allergens, e.g. wastewater treatment plants (Han *et al.* 2020) and urban centres
36
37 657 (Brodie *et al.* 2007; Woo *et al.* 2013), and these have direct public health relevance
38
39 658 although dispersal appears to be localised. Assessing the extent of anthropogenic
40
41 659 impacts remains somewhat speculative as more dedicated studies are needed both to
42
43 660 provide a better constrained understanding of current processes and to investigate
44
45 661 impacts of highly complex anthropogenic changes to the atmospheric gaseous and
46
47 662 particulate composition.
48
49
50
51
52
53
54
55
56
57
58
59
60

664 **Concluding remarks**

1
2
3 665 The discipline of aeromicrobiology has benefitted enormously from recent advances in
4
5 666 methodology that overcome the limitations associated with the study of ultra-low
6
7
8 667 biomass habitats, and we view this is beneficial in a wider sense to improved ecological
9
10 668 understanding of linkages between the atmosphere and surface terrestrial and marine
11
12 669 biomes. Specifically, the ability to accurately estimate microbial diversity in air through
13
14 670 rigorous decontamination of environmental metagenomic data has revealed that the
15
16 671 atmospheric boundary layer supports a diverse microbiota. Atmospheric microbial
17
18 672 assemblages display taxonomic structuring that reflects strong environmental filtering,
19
20 673 as well as distinct spatio-temporal biogeographic signals that reflect a complex
21
22 674 interplay of local and regional recruitment to the atmosphere. The long-held ecological
23
24 675 view of atmospheric transport as neutral to dispersal outcomes is therefore no longer
25
26 676 supportable, and a revision of ecological assumptions about the influence of
27
28 677 atmospheric microbial transport is warranted. Knowledge gaps that impede greater
29
30 678 understanding in this regard include observational data on flux to and from surface
31
32 679 environments and the atmosphere, and a focus on accurate quantitative methods should
33
34 680 be a priority for future research.

40 681 The biophysical role of microorganisms in ice nucleation and precipitation
41
42 682 formation is supported by convincing evidence and the molecular mechanisms that
43
44 683 underpin biogenic ice nucleation are becoming better resolved. Given the temperature
45
46 684 range at which biogenic ice nucleation occurs is relevant to a warming climate we urge
47
48 685 greater focus and inclusion of microbial ice nucleation in research to predict future
49
50 686 impacts on the water cycle. In the absence of evidence for widespread metabolic
51
52 687 activity that influences biogeochemical outcomes, lack of microbial interactions and
53
54 688 food webs or evidence for cell proliferation, we conclude that microbial biochemical
55
56 689 transformations make a very limited contribution to overall atmospheric chemistry
57
58
59
60

1
2
3 690 although they may be important in localised habitable areas such as clouds or plumes of
4
5 691 hydrated particles. The notion of a metabolically active atmospheric microbiome must
6
7
8 692 therefore be contemplated with a high degree of nuance. Development of improved
9
10 693 laboratory simulations for atmospheric conditions and *in situ* measurements will assist
11
12 694 with further understanding of atmospheric microbial metabolism and its boundaries in
13
14
15 695 air.

16
17 696 In a broader sense we advocate that estimates of the pan-global microbiota and
18
19 697 its ecological relevance require greater consideration of biomass in the gas-phase
20
21 698 atmospheric microbial environment alongside solid-phase terrestrial and liquid-phase
22
23 699 marine habitats. The increasing threats to environmental, organismal and human health
24
25 700 due to changes in atmospheric transport of microorganisms will be more clearly
26
27 701 understood when empirical considerations of the atmospheric microbiota are
28
29 702 incorporated into analytical frameworks. We therefore urge expansion of atmospheric
30
31 703 microbiology monitoring and inclusion as part of global scale atmospheric data
32
33 704 networks.
34
35
36
37
38
39
40

41 706 **References**

- 42
43 707 Aalismail NA, Ngugi DK, Díaz-Rúa R *et al.* Functional metagenomic analysis of dust-
44
45 708 associated microbiomes above the Red Sea. *Sci Rep* 2019;**9**:1–12.
46
47 709 Afeti GM, Resch FJ. Distribution of the liquid aerosol produced from bursting bubbles in
48
49 710 sea and distilled water. *Tellus B Chem Phys Meteorol* 1990;**42**:378–84.
50
51 711 Aller JY, Kuznetsova MR, Jahns CJ *et al.* The sea surface microlayer as a source of viral
52
53 712 and bacterial enrichment in marine aerosols. *J Aerosol Sci* 2005;**36**:801–12.
54
55 713 Alsved M, Holm S, Christiansen S *et al.* Effect of Aerosolization and Drying on the
56
57 714 Viability of *Pseudomonas syringae* Cells. *Front Microbiol* 2018;**9**:1–11.
58
59
60

- 1
2
3 715 Amato P. Energy metabolism in low temperature and frozen conditions. In: Yumoto I
4
5 716 (ed.). *Cold-Adapted Microorganisms*. 1st ed. Poole: Caister Academic Press, 2013.
6
7
8 717 Amato P, Besaury L, Joly M *et al*. Metatranscriptomic exploration of microbial
9
10 718 functioning in clouds. *Sci Rep* 2019;**9**:4383.
11
12 719 Amato P, Christner BC. Energy Metabolism Response to Low-Temperature and Frozen
13
14 720 Conditions in *Psychrobacter cryohalolentis*. *Appl Environ Microbiol* 2009;**75**:711–8.
15
16
17 721 Amato P, Demeer F, Melaouhi A *et al*. A fate for organic acids, formaldehyde and
18
19 722 methanol in cloud water: Their biotransformation by micro-organisms. *Atmos*
20
21 723 *Chem Phys* 2007;**7**:4159–69.
22
23
24 724 Amato P, Joly M, Besaury L *et al*. Active microorganisms thrive among extremely diverse
25
26 725 communities in cloud water. *PLoS One* 2017;**12**:e0182869.
27
28
29 726 Amato P, Joly M, Schaupp C *et al*. Survival and ice nucleation activity of bacteria as
30
31 727 aerosols in a cloud simulation chamber. *Atmos Chem Phys* 2015;**15**:6455–65.
32
33
34 728 de Araujo GG, Rodrigues F, Gonçalves FLT *et al*. Survival and ice nucleation activity of
35
36 729 *Pseudomonas syringae* strains exposed to simulated high-altitude atmospheric
37
38 730 conditions. *Sci Rep* 2019;**9**:7768.
39
40
41 731 Archer S, Lee K, Caruso T *et al*. Diverse recruitment to a geographically structured
42
43 732 atmospheric microbiome. *Nat Ecol Evol* 2021;**5**:[https://doi.org/10.21203/rs.3.rs-](https://doi.org/10.21203/rs.3.rs-244923/v2)
44
45 733 [244923/v2](https://doi.org/10.21203/rs.3.rs-244923/v2).
46
47
48 734 Archer SDJ, Lee KC, Caruso T *et al*. Airborne microbial transport limitation to isolated
49
50 735 Antarctic soil habitats. *Nat Microbiol* 2019;**4**:925–32.
51
52
53 736 Archer SDJ, Lee KC, Caruso T *et al*. Air mass source determines airborne microbial
54
55 737 diversity at the ocean–atmosphere interface of the Great Barrier Reef marine
56
57 738 ecosystem. *ISME J* 2020;**14**:871–6.
58
59
60 739 Archer SDJJ, Pointing SB. Anthropogenic impact on the atmospheric microbiome. *Nat*

- 1
2
3 740 *Microbiol* 2020;**5**:229–31.
- 4
5 741 Aylor D. *Aerial Dispersal of Pollen and Spores*. St Paul: APS Press, 2017.
- 6
7 742 Barber J, Andersson B. Too much of a good thing: light can be bad for photosynthesis.
- 8
9 743 *Trends Biochem Sci* 1992;**17**:61–6.
- 10
11 744 Barberán A, Henley J, Fierer N *et al*. Structure, inter-annual recurrence, and global-scale
- 12
13 745 connectivity of airborne microbial communities. *Sci Total Environ* 2014;**487**:187–
- 14
15 746 95.
- 16
17 747 Barry RG, Chorley RJ. *Atmosphere, Weather and Climate*. 9th Editio. New York:
- 18
19 748 Routledge, 2010.
- 20
21 749 Bauer H, Giebl H, Hitzenberger R *et al*. Airborne bacteria as cloud condensation nuclei. *J*
- 22
23 750 *Geophys Res Atmos* 2003;**108**:4658.
- 24
25 751 Bebbler DP, Holmes T, Gurr SJ. The global spread of crop pests and pathogens. *Glob Ecol*
- 26
27 752 *Biogeogr* 2014;**23**:1398–407.
- 28
29 753 Bianco A, Deguillaume L, Chaumerliac N *et al*. Effect of endogenous microbiota on the
- 30
31 754 molecular composition of cloud water: a study by Fourier-transform ion cyclotron
- 32
33 755 resonance mass spectrometry (FT-ICR MS). *Sci Rep* 2019;**9**:7663.
- 34
35 756 Björnham O, Sigg R, Burman J. Multilevel model for airborne transmission of foot-and-
- 36
37 757 mouth disease applied to Swedish livestock. *PLoS One* 2020;**15**:1–31.
- 38
39 758 Blanchard DC. The Ejection of Drops from the Sea and Their Enrichment with Bacteria
- 40
41 759 and Other Materials: A Review. *Estuaries* 1989;**12**:127.
- 42
43 760 Blanchard DC, Syzdek L. Mechanism for the Water-to-Air Transfer and Concentration of
- 44
45 761 Bacteria. *Science (80-)* 1970;**170**:626–8.
- 46
47 762 Borrelli P, Lugato E, Montanarella L *et al*. A New Assessment of Soil Loss Due to Wind
- 48
49 763 Erosion in European Agricultural Soils Using a Quantitative Spatially Distributed
- 50
51 764 Modelling Approach. *L Degrad Dev* 2017;**28**:335–44.
- 52
53
54
55
56
57
58
59
60

- 1
2
3 765 Bowers RM, Clements N, Emerson JB *et al.* Seasonal variability in bacterial and fungal
4
5 766 diversity of the near-surface atmosphere. *Environ Sci Technol* 2013;**47**:12097–106.
6
7
8 767 Bowers RM, McCubbin IB, Hallar AG *et al.* Seasonal variability in airborne bacterial
9
10 768 communities at a high-elevation site. *Atmos Environ* 2012;**50**:41–9.
11
12
13 769 Bowers RM, McLetchie S, Knight R *et al.* Spatial variability in airborne bacterial
14
15 770 communities across land-use types and their relationship to the bacterial
16
17 771 communities of potential source environments. *ISME J* 2011;**5**:601–12.
18
19
20 772 Brodie EL, DeSantis TZ, Moberg Parker JP *et al.* Urban aerosols harbor diverse and
21
22 773 dynamic bacterial populations. *Proc Natl Acad Sci U S A* 2007;**104**:299–304.
23
24
25 774 Brown JKM, Hovmøller MS. Aerial Dispersal of Pathogens on the Global and Continental
26
27 775 Scales and Its Impact on Plant Disease. *Science (80-)* 2002;**297**:537–41.
28
29 776 Bryan NC, Christner BC, Guzik TG *et al.* Abundance and survival of microbial aerosols in
30
31 777 the troposphere and stratosphere. *ISME J* 2019;**13**:2789–99.
32
33
34 778 Burrows SM, Butler T, Jöckel P *et al.* Bacteria in the global atmosphere - Part 2: Modeling
35
36 779 of emissions and transport between different ecosystems. *Atmos Chem Phys*
37
38 780 2009;**9**:9281–97.
39
40
41 781 Burrows SM, Rayner PJ, Butler T *et al.* Estimating bacteria emissions from inversion of
42
43 782 atmospheric transport: sensitivity to modelled particle characteristics. *Atmos Chem*
44
45 783 *Phys* 2013;**13**:5473–88.
46
47
48 784 Cáliz J, Triadó-Margarit X, Camarero L *et al.* A long-term survey unveils strong seasonal
49
50 785 patterns in the airborne microbiome coupled to general and regional atmospheric
51
52 786 circulations. *Proc Natl Acad Sci U S A* 2018;**115**:12229–34.
53
54
55 787 Carlucci AF, Williams PM. Concentration of bacteria from sea water by bubble
56
57 788 scavenging. *ICES J Mar Sci* 1965;**30**:28–33.
58
59
60 789 Chen S, Jiang N, Huang J *et al.* Quantifying contributions of natural and anthropogenic

- 1
2
3 790 dust emission from different climatic regions. *Atmos Environ* 2018;**191**:94–104.
4
5 791 Christner BC, Cai R, Morris CE *et al.* Geographic, seasonal, and precipitation chemistry
6
7 792 influence on the abundance and activity of biological ice nucleators in rain and
8
9 793 snow. *Proc Natl Acad Sci U S A* 2008a;**105**:18854–9.
10
11 794 Christner BC, Morris CE, Foreman CM *et al.* Ubiquity of biological ice nucleators in
12
13 795 snowfall. *Science (80-)* 2008b;**319**:1214.
14
15 796 Cifuentes JO, Comino N, Trastoy B *et al.* Structural basis of glycogen metabolism in
16
17 797 bacteria. *Biochem J* 2019;**476**:2059–92.
18
19 798 Daly MJ. A new perspective on radiation resistance based on *Deinococcus radiodurans*.
20
21 799 *Nat Rev Micro* 2009;**7**:237–45.
22
23 800 DasSarma P, DasSarma S. Survival of microbes in Earth's stratosphere. *Curr Opin*
24
25 801 *Microbiol* 2018;**43**:24–30.
26
27 802 Deguillaume L, Leriche M, Amato P *et al.* Microbiology and atmospheric processes:
28
29 803 Chemical interactions of primary biological aerosols. *Biogeosciences* 2008;**5**:1073–
30
31 804 84.
32
33 805 DeMott PJ, Hill TCJ, McCluskey CS *et al.* Sea spray aerosol as a unique source of ice
34
35 806 nucleating particles. *Proc Natl Acad Sci* 2016;**113**:5797–803.
36
37 807 Després VR, Alex Huffman J, Burrows SM *et al.* Primary biological aerosol particles in the
38
39 808 atmosphere: A review. *Tellus, Ser B Chem Phys Meteorol* 2012;**64**:doi:
40
41 809 10.3402/tellusb.v64i0.15598.
42
43 810 Dumbrell AJ, Nelson M, Helgason T *et al.* Relative roles of niche and neutral processes in
44
45 811 structuring a soil microbial community. *ISME J* 2010;**4**:337–45.
46
47 812 Eisenhofer R, Minich JJ, Marotz C *et al.* Contamination in Low Microbial Biomass
48
49 813 Microbiome Studies: Issues and Recommendations. *Trends Microbiol* 2019;**27**:105–
50
51 814 17.
52
53
54
55
56
57
58
59
60

- 1
2
3 815 Elbert W, Taylor PE, Andreae MO *et al.* Contribution of fungi to primary biogenic
4
5 816 aerosols in the atmosphere: wet and dry discharged spores, carbohydrates, and
6
7 817 inorganic ions. *Atmos Chem Phys* 2007;**7**:4569–88.
8
9
10 818 Els N, Baumann-Stanzer K, Larose C *et al.* Beyond the planetary boundary layer:
11
12 819 Bacterial and fungal vertical biogeography at Mount Sonnblick, Austria. *Geo Geogr*
13
14 820 *Environ* 2019a;**6**:e00069.
15
16
17 821 Els N, Larose C, Baumann-Stanzer K *et al.* Microbial composition in seasonal time series
18
19 822 of free tropospheric air and precipitation reveals community separation.
20
21 823 *Aerobiologia (Bologna)* 2019b;**35**:671–701.
22
23
24 824 Ezraty B, Gennaris A, Barras F *et al.* Oxidative stress, protein damage and repair in
25
26 825 bacteria. *Nat Rev Microbiol* 2017;**15**:385–96.
27
28
29 826 Fankhauser AM, Antonio DD, Krell A *et al.* Constraining the Impact of Bacteria on the
30
31 827 Aqueous Atmospheric Chemistry of Small Organic Compounds. *ACS Earth Sp Chem*
32
33 828 2019;**3**:1485–91.
34
35
36 829 Fröhlich-Nowoisky J, Burrows SM, Xie Z *et al.* Biogeography in the air: Fungal diversity
37
38 830 over land and oceans. *Biogeosciences* 2012;**9**:1125–36.
39
40
41 831 Fröhlich-Nowoisky J, Hill TCJ, Pummer BG *et al.* Ice Nucleation Activity in the
42
43 832 Widespread Soil Fungus *Mortierella alpina*. *Biogeosciences Discuss* 2014;**11**:12697–
44
45 833 731.
46
47
48 834 Gao Q, Garcia-Pichel F. Microbial ultraviolet sunscreen. *Nat Rev Microbiol* 2011;**9**:791–
49
50 835 802.
51
52
53 836 Gettelman A, Collins WD, Fetzer EJ *et al.* Climatology of Upper-Tropospheric Relative
54
55 837 Humidity from the Atmospheric Infrared Sounder and Implications for Climate. *J*
56
57 838 *Clim* 2006;**19**:6104–21.
58
59
60 839 Gilbert JA, Stephens B. Microbiology of the built environment. *Nat Rev Microbiol*

1
2
3 840 2018;**16**:661–70.
4

5 841 González-Toril E, Osuna S, Viúdez-Moreiras D *et al.* Impacts of Saharan Dust Intrusions
6
7
8 842 on Bacterial Communities of the Low Troposphere. *Sci Rep* 2020;**10**:6837.
9

10 843 Gorbushina AA, Kort R, Schulte A *et al.* Life in Darwin's dust: intercontinental transport
11
12 844 and survival of microbes in the nineteenth century. *Env Microbiol* 2007;**9**:2911–22.
13

14
15 845 Gusareva ES, Acerbi E, Lau KJX *et al.* Microbial communities in the tropical air ecosystem
16
17 846 follow a precise diel cycle. *Proc Natl Acad Sci U S A* 2019;**116**:23299–308.
18

19 847 Han Y, Li L, Wang Y *et al.* Composition, dispersion, and health risks of bioaerosols in
20
21 848 wastewater treatment plants: A review. *Front Environ Sci Eng* 2020;**15**:38.
22

23
24 849 Hanson CA, Fuhrman JA, Horner-Devine MC *et al.* Beyond biogeographic patterns:
25
26 850 Processes shaping the microbial landscape. *Nat Rev Microbiol* 2012;**10**:497–506.
27

28
29 851 Hartmann S, Augustin S, Clauss T *et al.* Immersion freezing of ice nucleation active
30
31 852 protein complexes. *Atmos Chem Phys* 2013;**13**:5751–66.
32

33
34 853 Hayakawa K, Tang N, Nagato EG *et al.* Long term trends in atmospheric concentrations
35
36 854 of polycyclic aromatic hydrocarbons and nitropolycyclic aromatic hydrocarbons: A
37
38 855 study of Japanese cities from 1997 to 2014. *Environ Pollut* 2018;**233**:474–82.
39

40
41 856 Heald CL, Spracklen D V. Atmospheric budget of primary biological aerosol particles
42
43 857 from fungal spores. *Geophys Res Lett* 2009;**36**, DOI:

44
45 858 <https://doi.org/10.1029/2009GL037493>.
46

47
48 859 Healy DA, Huffman JA, O'Connor DJ *et al.* Ambient measurements of biological aerosol
49
50 860 particles near Killarney, Ireland: A comparison between real-time fluorescence and
51
52 861 microscopy techniques. *Atmos Chem Phys* 2014;**14**:8055–69.
53

54
55 862 Hochman A, Alpert P, Harpaz T *et al.* A new dynamical systems perspective on
56
57 863 atmospheric predictability: Eastern Mediterranean weather regimes as a case
58
59 864 study. *Sci Adv* 2019;**5**:eaau0936.
60

- 1
2
3 865 Hoose C, Kristjánsson JE, Burrows SM. How important is biological ice nucleation in
4
5 866 clouds on a global scale? *Environ Res Lett* 2010;**5**:24009.
6
7
8 867 Hsieh CI, Katul G. The Lagrangian stochastic model for estimating footprint and water
9
10 868 vapor fluxes over inhomogeneous surfaces. *Int J Biometeorol* 2009;**53**:87–100.
11
12 869 Hu W, Niu H, Murata K *et al.* Bacteria in atmospheric waters: Detection, characteristics
13
14 870 and implications. *Atmos Environ* 2018;**179**:201–21.
15
16
17 871 Hudait A, Odendahl N, Qiu Y *et al.* Ice-Nucleating and Antifreeze Proteins Recognize Ice
18
19 872 through a Diversity of Anchored Clathrate and Ice-like Motifs. *J Am Chem Soc*
20
21 873 2018;**140**:4905–12.
22
23
24 874 Huestis DL, Dao A, Diallo M *et al.* Windborne long-distance migration of malaria
25
26 875 mosquitoes in the Sahel. *Nature* 2019;**574**:404–8.
27
28
29 876 Huffman JA, Prenni AJ, DeMott PJ *et al.* High concentrations of biological aerosol
30
31 877 particles and ice nuclei during and after rain. *Atmos Chem Phys* 2013;**13**:6151–64.
32
33
34 878 Husárová S, Vařtilingom M, Deguillaume L *et al.* Biotransformation of methanol and
35
36 879 formaldehyde by bacteria isolated from clouds. Comparison with radical chemistry.
37
38 880 *Atmos Environ* 2011;**45**:6093–102.
39
40
41 881 Jaber S, Lallement A, Sancelme M *et al.* Biodegradation of phenol and catechol in cloud
42
43 882 water: comparison to chemical oxidation in the atmospheric multiphase system.
44
45 883 *Atmos Chem Phys* 2020;**20**:4987–97.
46
47
48 884 Jaenicke R. Abundance of Cellular Material and Proteins in the Atmosphere. *Science (80-*
49
50 885 *)* 2005;**308**:73.
51
52
53 886 Joly M, Amato P, Sancelme M *et al.* Survival of microbial isolates from clouds toward
54
55 887 simulated atmospheric stress factors. *Atmos Environ* 2015;**117**:92–8.
56
57
58 888 Joly M, Attard E, Sancelme M *et al.* Ice nucleation activity of bacteria isolated from cloud
59
60 889 water. *Atmos Environ* 2013;**70**:392–400.

- 1
2
3 890 Jones SE, Lennon JT. Dormancy contributes to the maintenance of microbial diversity.
4
5 891 *Proc Natl Acad Sci U S A* 2010;**107**:5881–6.
6
7
8 892 Joung YS, Ge Z, Buie CR. Bioaerosol generation by raindrops on soil. *Nat Commun*
9
10 893 2017;**8**:14668.
11
12 894 Jousse C, Dalle C, Canet I *et al.* Metabolomic study of the response to cold shock in a
13
14 strain of *Pseudomonas syringae* isolated from cloud water. *Metabolomics*
15 895 2017;**14**:11.
16
17 896
18
19 897 Jusot J-F, Neill DR, Waters EM *et al.* Airborne dust and high temperatures are risk factors
20
21 for invasive bacterial disease. *J Allergy Clin Immunol* 2017;**139**:977-986.e2.
22 898
23
24 899 Kellogg CA, Griffin DW. Aerobiology and the global transport of desert dust. *Trends Ecol*
25
26 900 *Evol* 2006;**21**:638–44.
27
28
29 901 Khaled A, Zhang M, Amato P *et al.* Biodegradation by bacteria in clouds: An
30
31 902 underestimated sink for some organics in the atmospheric multiphase system.
32
33 903 *Atmos Chem Phys Discuss* 2020;**2020**:1–32.
34
35
36 904 King MD, Platnick S, Menzel WP *et al.* Spatial and Temporal Distribution of Clouds
37
38 905 Observed by MODIS Onboard the Terra and Aqua Satellites. *IEEE Trans Geosci*
39
40 906 *Remote Sens* 2013;**51**:3826–52.
41
42
43 907 Klein AM, Bohannan BJM, Jaffe DA *et al.* Molecular evidence for metabolically active
44
45 908 bacteria in the atmosphere. *Front Microbiol* 2016;**7**:1–11.
46
47
48 909 Kollath DR, Miller KJ, Barker BM. The mysterious desert dwellers: *Coccidioides immitis*
49
50 910 and *Coccidioides posadasii*, causative fungal agents of coccidioidomycosis.
51
52 911 *Virulence* 2019;**10**:222–33.
53
54
55 912 Kondakova T, Catovic C, Barreau M *et al.* Response to Gaseous NO₂ Air Pollutant of *P.*
56
57 913 *fluorescens* Airborne Strain MFAF76a and Clinical Strain MFN1032. *Front Microbiol*
58
59 914 2016;**7**:379.
60

- 1
2
3 915 Korolev A, Leisner T. Review of experimental studies of secondary ice production. *Atmos*
4
5 916 *Chem Phys* 2020;**20**:11767–97.
6
7
8 917 Krumins V, Mainelis G, Kerkhof LJ *et al.* Substrate-Dependent rRNA Production in an
9
10 918 Airborne Bacterium. *Environ Sci Technol Lett* 2014;**1**:376–81.
11
12 919 Lagomarsino Oneto D, Golan J, Mazzino A *et al.* Timing of fungal spore release dictates
13
14 survival during atmospheric transport. *Proc Natl Acad Sci* 2020;**117**:5134–43.
15
16
17 921 Lai KM, Emberlin J, Colbeck I. Outdoor environments and human pathogens in air.
18
19 922 *Environ Heal A Glob Access Sci Source* 2009;**8**:1–5.
20
21
22 923 Lallement A, Besaury L, Tixier E *et al.* Potential for phenol biodegradation in cloud
23
24 924 waters. *Biogeosciences* 2018;**15**:5733–44.
25
26
27 925 Lebre PH, De Maayer P, Cowan DA. Xerotolerant bacteria: surviving through a dry spell.
28
29 926 *Nat Rev Microbiol* 2017;**15**:285–96.
30
31
32 927 Lennon JT, Jones SE. Microbial seed banks: the ecological and evolutionary implications
33
34 928 of dormancy. *Nat Rev Microbiol* 2011;**9**:119–30.
35
36 929 Lewis KM, van Dijken GL, Arrigo KR. Changes in phytoplankton concentration now drive
37
38 930 increased Arctic Ocean primary production. *Science (80-)* 2020;**369**:198–202.
39
40
41 931 Ling M, Marshall IPG, Rosati B *et al.* Properties relevant to atmospheric dispersal of the
42
43 932 ice-nucleation active *Pseudomonas syringae* strain R10.79 isolated from rain
44
45 933 water. *Aerobiologia (Bologna)* 2021;**37**:225–41.
46
47
48 934 Ling ML, Wex H, Grawe S *et al.* Effects of Ice Nucleation Protein Repeat Number and
49
50 935 Oligomerization Level on Ice Nucleation Activity. *J Geophys Res Atmos*
51
52 936 2018;**123**:1802–10.
53
54
55 937 Lowe WH, McPeck MA. Is dispersal neutral? *Trends Ecol Evol* 2014;**29**:444–50.
56
57 938 Lukas M, Schwidetzky R, Kunert AT *et al.* Electrostatic Interactions Control the
58
59 939 Functionality of Bacterial Ice Nucleators. *J Am Chem Soc* 2020;**142**:6842–6.
60

- 1
2
3 940 Lympelopoulou DS, Adams RI, Lindow SE. Contribution of vegetation to the microbial
4
5 941 composition of nearby outdoor air. *Appl Environ Microbiol* 2016;**82**:3822–33.
6
7
8 942 Maki T, Kurosaki Y, Onishi K *et al.* Variations in the structure of airborne bacterial
9
10 943 communities in Tsogt-Ovoo of Gobi desert area during dust events. *Air Qual Atmos*
11
12 944 *Heal* 2016:1–12.
13
14
15 945 Maki T, Lee KC, Kawai K *et al.* Aeolian Dispersal of Bacteria Associated With Desert Dust
16
17 946 and Anthropogenic Particles Over Continental and Oceanic Surfaces. *J Geophys Res*
18
19 947 *Atmos* 2019;**124**:5579–88.
20
21
22 948 Matulová M, Husárová S, Capek P *et al.* Biotransformation of various saccharides and
23
24 949 production of exopolymeric substances by cloud-borne *Bacillus* sp. 3B6. *Environ Sci*
25
26 950 *Technol* 2014;**48**:14238–47.
27
28
29 951 Mayol E, Arrieta JM, Jiménez MA *et al.* Long-range transport of airborne microbes over
30
31 952 the global tropical and subtropical ocean. *Nat Commun* 2017;**8**:201.
32
33
34 953 Meyer M, Cox JA, Hitchings MDT *et al.* Quantifying airborne dispersal routes of
35
36 954 pathogens over continents to safeguard global wheat supply. *Nat Plants*
37
38 955 2017;**3**:780–6.
39
40
41 956 Michaud JM, Thompson LR, Kaul D *et al.* Taxon-specific aerosolization of bacteria and
42
43 957 viruses in an experimental ocean-atmosphere mesocosm. *Nat Commun*
44
45 958 2018;**9**:2017.
46
47
48 959 Monier J-M, Lindow SE. Differential survival of solitary and aggregated bacterial cells
49
50 960 promotes aggregate formation on leaf surfaces. *Proc Natl Acad Sci*
51
52 961 2003;**100**:15977–82.
53
54
55 962 Moore RA, Bomar C, Kobziar LN *et al.* Wildland fire as an atmospheric source of viable
56
57 963 microbial aerosols and biological ice nucleating particles. *ISME J* 2021;**15**:461–72.
58
59 964 Moore RA, Hanlon R, Powers C *et al.* Scavenging of Sub-Micron to Micron-Sized
60

- 1
2
3 965 Microbial Aerosols during Simulated Rainfall. *Atmosphere (Basel)*
4
5 966 2020;**11**:doi.org/10.3390/atmos11010080.
6
7
8 967 Morris CE, Conen F, Alex Huffman J *et al.* Bioprecipitation: a feedback cycle linking Earth
9
10 968 history, ecosystem dynamics and land use through biological ice nucleators in the
11
12 969 atmosphere. *Glob Chang Biol* 2014;**20**:341–51.
13
14
15 970 Morris CE, Monier J-M. The ecological significance of biofilm formation by plant-
16
17 971 associated bacteria. *Annu Rev Phytopathol* 2003;**41**:429–53.
18
19
20 972 Morris CE, Sands DC, Bardin M *et al.* Microbiology and atmospheric processes: an
21
22 973 upcoming era of research on bio-meteorology. *Biogeosciences Discuss* 2008;**5**:191–
23
24 974 212.
25
26
27 975 Morris CE, Sands DC, Glaux C *et al.* Urediospores of rust fungi are ice nucleation active at
28
29 976 > –10 °C and harbor ice nucleation active bacteria. *Atmos Chem Phys*
30
31 977 2013;**13**:4223–33.
32
33
34 978 Mülmenstädt J, Sourdeval O, Delanoë J *et al.* Frequency of occurrence of rain from liquid-
35
36 979 , mixed-, and ice-phase clouds derived from A-Train satellite retrievals. *Geophys Res*
37
38 980 *Lett* 2015;**42**:6502–9.
39
40
41 981 Nicolaisen M, West JS, Sapkota R *et al.* Fungal Communities Including Plant Pathogens in
42
43 982 Near Surface Air Are Similar across Northwestern Europe. *Front Microbiol*
44
45 983 2017;**8**:1729.
46
47
48 984 NOAA, NASA, USAirForce. *U.S. Standard Atmosphere*. Washington D.C., 1976.
49
50 985 Norros V, Rannik Ü, Hussein T *et al.* Do small spores disperse further than large spores?
51
52 986 *Ecology* 2014;**95**:1612–21.
53
54
55 987 O’Sullivan D, Murray BJ, Ross JF *et al.* The relevance of nanoscale biological fragments
56
57 988 for ice nucleation in clouds. *Sci Rep* 2015;**5**:8082.
58
59
60 989 Ontiveros VJ, Cáliz J, Triadó-Margarit X *et al.* General decline in the diversity of the

- 1
2
3 990 airborne microbiota under future climatic scenarios. *Sci Rep* 2021;**11**:20223.
4
5 991 Oteros J, Bartusel E, Alessandrini F *et al.* Artemisia pollen is the main vector for airborne
6
7 992 endotoxin. *J Allergy Clin Immunol* 2019;**143**:369–77.
8
9
10 993 Pal C, Bengtsson-Palme J, Kristiansson E *et al.* The structure and diversity of human,
11
12 994 animal and environmental resistomes. *Microbiome* 2016;**4**:54.
13
14 995 Pandey R, Usui K, Livingstone RA *et al.* Ice-nucleating bacteria control the order and
15
16 996 dynamics of interfacial water. *Sci Adv* 2016;**2**:e1501630.
17
18 997 Pointing SB, Belnap J. Microbial colonization and controls in dryland systems. *Nat Rev*
19
20 998 *Microbiol* 2012;**10**:551–62.
21
22 999 Pointing SB, Belnap J. Disturbance to desert soil ecosystems contributes to dust-
23
24 1000 mediated impacts at regional scales. *Biodivers Conserv* 2014;**23**:1659–67.
25
26 1001 Potts M. Desiccation tolerance of prokaryotes. *Microbiol Mol Biol Rev* 1994;**58**:755–805.
27
28 1002 Pouleur S, Richard C, Martin J *et al.* Ice nucleation activity in *Fusarium acuminatum* and
29
30 1003 *Fusarium avenaceum*. *Appl Environ Microbiol* 1992;**58**:2960–4.
31
32 1004 Pruppacher HR, Jaenicke R. The processing of water vapor and aerosols by atmospheric
33
34 1005 clouds, a global estimate. *Atmos Res* 1995;**38**:283–95.
35
36 1006 Rastelli E, Corinaldesi C, Dell'anno A *et al.* Transfer of labile organic matter and
37
38 1007 microbes from the ocean surface to the marine aerosol: An experimental approach.
39
40 1008 *Sci Rep* 2017;**7**:1–10.
41
42 1009 Reche I, D'Orta G, Mladenov N *et al.* Deposition rates of viruses and bacteria above the
43
44 1010 atmospheric boundary layer. *ISME J* 2018;**12**:1154–62.
45
46 1011 Roeters SJ, Golbek TW, Bregnhøj M *et al.* Ice-nucleating proteins are activated by low
47
48 1012 temperatures to control the structure of interfacial water. *Nat Commun*
49
50 1013 2021;**12**:1183.
51
52 1014 Salter SJ, Cox MJ, Turek EM *et al.* Reagent and laboratory contamination can critically

- 1
2
3 1015 impact sequence-based microbiome analyses. *BMC Biol* 2014;**12**:87.
4
5 1016 Šantl-Temkiv T, Finster K, Hansen BM *et al.* Viable methanotrophic bacteria enriched
6
7
8 1017 from air and rain can oxidize methane at cloud-like conditions. *Aerobiologia*
9
10 1018 (*Bologna*) 2013;**29**:373–84.
11
12 1019 Šantl-Temkiv T, Gosewinkel U, Starnawski P *et al.* Aeolian dispersal of bacteria in
13
14
15 1020 southwest Greenland: Their sources, abundance, diversity and physiological states.
16
17 1021 *FEMS Microbiol Ecol* 2018;**94**:fiy031.
18
19 1022 Šantl-Temkiv T, Lange R, Beddows D *et al.* Biogenic Sources of Ice Nucleating Particles
20
21
22 1023 at the High Arctic Site Villum Research Station. *Environ Sci Technol*
23
24 1024 2019;**53**:10580–90.
25
26 1025 Šantl-Temkiv T, Sikoparija B, Maki T *et al.* Bioaerosol field measurements: Challenges
27
28
29 1026 and perspectives in outdoor studies. *Aerosol Sci Technol* 2020;**54**:520–46.
30
31 1027 Sattler B, Puxbaum H, Psenner R. Bacterial growth in supercooled cloud droplets.
32
33 1028 *Geophys Res Lett* 2001;**28**:239–42.
34
35 1029 Savary S, Willocquet L, Pethybridge SJ *et al.* The global burden of pathogens and pests
36
37
38 1030 on major food crops. *Nat Ecol Evol* 2019;**3**:430–9.
39
40 1031 Schmid HP. Footprint modeling for vegetation atmosphere exchange studies: A review
41
42
43 1032 and perspective. *Agric For Meteorol* 2002;**113**:159–83.
44
45 1033 Schwartz M. The life and works of Louis Pasteur. *J Appl Microbiol* 2001;**91**:597–601.
46
47 1034 Sharma A, Gaidamakova EK, Grichenko O *et al.* Across the tree of life, radiation
48
49
50 1035 resistance is governed by antioxidant Mn²⁺, gauged by paramagnetic resonance.
51
52 1036 *Proc Natl Acad Sci U S A* 2017;**114**:9253–60.
53
54 1037 Shepherd G, Terradellas E, Baklanov A *et al.* *Global Assessment of Sand and Dust Storms*.
55
56
57 1038 United Nations Environment Program (UNEP), 2016.
58
59 1039 Singh RP, Hodson DP, Huerta-Espino J *et al.* The Emergence of Ug99 Races of the Stem
60

- 1
2
3 1040 Rust Fungus is a Threat to World Wheat Production. *Annu Rev Phytopathol*
4
5 1041 2011;**49**:465–81.
6
7
8 1042 Smith DJ, Griffin DW, McPeters RD *et al.* Microbial survival in the stratosphere and
9
10 1043 implications for global dispersal. *Aerobiologia (Bologna)* 2011;**27**:319–32.
11
12 1044 Smith DJ, Griffin DW, Schuerger AC. Stratospheric microbiology at 20 km over the Pacific
13
14 1045 Ocean. *Aerobiologia (Bologna)* 2010;**26**:35–46.
15
16
17 1046 Smith DJ, Ravichandar JD, Jain S *et al.* Airborne bacteria in earth's lower stratosphere
18
19 1047 resemble taxa detected in the troposphere: Results from a new NASA Aircraft
20
21 1048 Bioaerosol Collector (ABC). *Front Microbiol* 2018;**9**:1752.
22
23
24 1049 Sorensen JH, Mackay DKJ, Jensen C *et al.* An integrated model to predict the atmospheric
25
26 1050 spread of foot-and-mouth disease virus. *Epidemiol Infect* 2000;**124**:577–90.
27
28
29 1051 Spracklen D V., Heald CL. The contribution of fungal spores and bacteria to regional and
30
31 1052 global aerosol number and ice nucleation immersion freezing rates. *Atmos Chem*
32
33 1053 *Phys* 2014;**14**, DOI: 10.5194/acp-14-9051-2014.
34
35
36 1054 Spring AM, Domingue KD, Kerber T V. *et al.* Land use effects on airborne bacterial
37
38 1055 communities are evident in both near-surface and higher-altitude air. *Diversity*
39
40 1056 2021;**13**:1–19.
41
42
43 1057 Stevenson A, Burkhardt J, Cockell CS *et al.* Multiplication of microbes below 0.690 water
44
45 1058 activity: implications for terrestrial and extraterrestrial life. *Environ Microbiol*
46
47 1059 2015a;**17**:257–77.
48
49
50 1060 Stevenson A, Cray JA, Williams JP *et al.* Is there a common water-activity limit for the
51
52 1061 three domains of life? *ISME J* 2015b;**9**:1333–51.
53
54
55 1062 Stopelli E, Conen F, Morris CE *et al.* Ice nucleation active particles are efficiently
56
57 1063 removed by precipitating clouds. *Sci Rep* 2015;**5**:16433.
58
59
60 1064 Temkiv TŠ, Finster K, Hansen BM *et al.* The microbial diversity of a storm cloud as

- 1
2
3 1065 assessed by hailstones. *FEMS Microbiol Ecol* 2012, DOI: 10.1111/j.1574-
4
5 1066 6941.2012.01402.x.
6
7
8 1067 Tesson SVM, Šantl-Temkiv T. Ice Nucleation Activity and Aeolian Dispersal Success in
9
10 1068 Airborne and Aquatic Microalgae. 2018;**9**:1–14.
11
12 1069 Tesson SVM, Skjøth A, Šantl-temkiv T. Airborne Microalgae: Insights, Opportunities, and
13
14 1070 Challenges. *Appl Environ Microbiol* 2016;**82**:1978–91.
15
16
17 1071 Tignat-Perrier R, Dommergue A, Thollot A *et al.* Global airborne microbial communities
18
19 1072 controlled by surrounding landscapes and wind conditions. *Sci Rep* 2019;**9**:1–11.
20
21 1073 Tignat-Perrier R, Dommergue A, Thollot A *et al.* Seasonal shift in airborne microbial
22
23 1074 communities. *Sci Total Environ* 2020;**716**:137129.
24
25
26 1075 Tobias A, Karanasiou A, Amato F *et al.* Health effects of desert dust and sand storms: a
27
28 1076 systematic review and meta-analysis. *Environ Epidemiol* 2019;**3**:doi:
29
30 1077 10.1097/01.EE9.0000610424.75648.58.
31
32
33 1078 Tobo Y, Adachi K, DeMott PJ *et al.* Glacially sourced dust as a potentially significant
34
35 1079 source of ice nucleating particles. *Nat Geosci* 2019;**12**:253–8.
36
37
38 1080 Tong Y, Lighthart B. Effect of simulated solar radiation on mixed outdoor atmospheric
39
40 1081 bacterial populations. *FEMS Microbiol Ecol* 2006;**26**:311–6.
41
42
43 1082 Triado-Margarit X, Caliz J, Casamayor E. A long-term atmospheric baseline for
44
45 1083 intercontinental exchange of airborne pathogens. *Environ Int* 2021:in press.
46
47
48 1084 Uetake J, Hill TCJ, Moore KA *et al.* Airborne bacteria confirm the pristine nature of the
49
50 1085 Southern Ocean boundary layer. *Proc Natl Acad Sci U S A* 2020;**117**:13275–82.
51
52
53 1086 Uetake J, Tobo Y, Uji Y *et al.* Seasonal changes of airborne bacterial communities over
54
55 1087 Tokyo and influence of local meteorology. *Front Microbiol* 2019;**10**:1572.
56
57
58 1088 Vaitilingom M, Amato P, Sancelme M *et al.* Contribution of microbial activity to carbon
59
60 1089 chemistry in clouds. *Appl Environ Microbiol* 2010;**76**:23–9.

- 1
2
3 1090 Vaitilingom M, Deguillaume L, Vinatier V *et al.* Potential impact of microbial activity on
4
5 1091 the oxidant capacity and organic carbon budget in clouds. *Proc Natl Acad Sci*
6
7 1092 2013;**110**:559–64.
8
9
10 1093 Wainwright M, Wickramasinghe NC, Narlikar J V *et al.* Microorganisms cultured from
11
12 1094 stratospheric air samples obtained at 41 km. *FEMS Microbiol Lett* 2003;**218**:161–5.
13
14
15 1095 Wang X, Sultana CM, Trueblood J *et al.* Microbial Control of Sea Spray Aerosol
16
17 1096 Composition: A Tale of Two Blooms. *ACS Cent Sci* 2015;**1**:124–31.
18
19
20 1097 Wex H, Augustin-Bauditz S, Boose Y *et al.* Intercomparing different devices for the
21
22 1098 investigation of ice nucleating particles using Snomax® as test substance. *Atmos*
23
24 1099 *Chem Phys* 2015;**15**:1463–85.
25
26
27 1100 Whitman WB, Coleman DC, Wiebe WJ. Prokaryotes: The unseen majority. *Proc Natl Acad*
28
29 1101 *Sci U S A* 1998;**95**:6578–83.
30
31
32 1102 Whon TW, Kim M-S, Roh SW *et al.* Metagenomic Characterization of Airborne Viral DNA
33
34 1103 Diversity in the Near-Surface Atmosphere. *J Virol* 2012;**86**:8221–31.
35
36 1104 Wilson TW, Ladino LA, Alpert PA *et al.* A marine biogenic source of atmospheric ice-
37
38 1105 nucleating particles. *Nature* 2015;**525**:234–8.
39
40
41 1106 Wirgot N, Lagrée M, Traïkia M *et al.* Metabolic modulations of *Pseudomonas graminis* in
42
43 1107 response to H₂O₂ in cloud water. *Sci Rep* 2019;**9**, DOI: 10.1038/s41598-019-
44
45 1108 49319-2.
46
47
48 1109 de Wit R, Bouvier T. “Everything is everywhere, but, the environment selects”; what did
49
50 1110 Baas Becking and Beijerinck really say? *Environ Microbiol* 2006;**8**:755–8.
51
52
53 1111 Womack AM, Bohannan BJM, Green JL. Biodiversity and biogeography of the
54
55 1112 atmosphere. *Philos Trans R Soc Lond B Biol Sci* 2010;**365**:3645–53.
56
57
58 1113 Woo AC, Brar MS, Chan Y *et al.* Temporal variation in airborne microbial populations
59
60 1114 and microbially-derived allergens in a tropical urban landscape. *Atmos Environ*

- 1
2
3 1115 2013;**74**:291–300.
4
5
6 1116 Woo C, Yamamoto N. Falling bacterial communities from the atmosphere. *Environ*
7
8 1117 *Microbiome* 2020;**15**:22.
9
10 1118 Yang Y, Yokobori S, Yamagishi A. Assessing Panspermia Hypothesis by Microorganisms
11
12 1119 Collected from The High Altitude Atmosphere. *Biol Sci Sp* 2009;**23**:151–63.
13
14
15 1120 Zhang R, Li Y, Zhang AL *et al.* Identifying airborne transmission as the dominant route
16
17 1121 for the spread of COVID-19. *Proc Natl Acad Sci* 2020;**117**:14857–63.
18
19
20 1122 Zhou J, Ning D. Stochastic Community Assembly: Does It Matter in Microbial Ecology?
21
22 1123 *Microbiol Mol Biol Rev* 2017;**81**:e00002–17.
23
24 1124 Zhou S-Y-D, Li H, Giles M *et al.* Microbial Flow Within an Air-Phyllosphere-Soil
25
26 1125 Continuum. *Front Microbiol* 2021;**11**:3325.
27
28
29 1126 Ziegelhoffer EC, Donohue TJ. Bacterial responses to photo-oxidative stress. *Nat Rev*
30
31 1127 *Microbiol* 2009;**7**:856–63.
32
33
34 1128
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

Microbial ecology of the atmosphere

Figure legends

FIG. 1 | Defining the atmosphere as an ecological niche.

a) The atmosphere comprises a series of vertically delineated concentric layers that are primarily defined by thermal properties. The lowermost layer, the troposphere contains 75% of the molecular and gaseous mass of the atmosphere and almost all the water vapour, clouds and aerosolised particles (Barry and Chorley 2010) and including microorganisms. b) Four boundaries that occur at varying altitude within the troposphere are important in terms of atmospheric microbial ecology: The atmospheric boundary layer delineates the region of air closest to the surface where the bulk of surface-atmosphere interactions occur, and this includes exchange of microorganisms with terrestrial and marine ecosystems. The lifted condensation level (LCL) delineates the layer above which cloud formation occurs. It is variable with temperature and humidity and delineates the thermal limit for condensation of saturated water vapour on particulate surfaces. The freezing isotherm delineates the layer above which freezing occurs and is also the limit for active non-psychrophilic microbial metabolism. We note that the relationship between the atmospheric boundary layer, the LCL and the freezing isotherm, which are highly dynamic, is significantly more complex than presented in the figure, e.g., both the LCL and the freezing isotherm can sit at ground level well within the boundary layer. The boundary between the LCL and freezing isotherm represent the theoretical range for mesophilic microbial metabolic activity, with potential for psychrophilic and xerophilic activity to extend beyond this zone. The tropopause marks the upper limit of the troposphere and mixing with upper layers is limited by the thermal inversion in the stratosphere. The bulk of atmospheric ozone is in the lower stratosphere and this likely delineates the altitudinal limit of survival for many atmospheric microorganisms (Bryan et al. 2019).

FIG. 2 | Patterns of microbial diversity and abundance in the atmosphere.

a) Simulated concentration of bacterial cells and fungal spores in the near surface atmospheric boundary layer (adapted from ref (Hoose, Kristjánsson and Burrows 2010)). Models assumed a uniform bacterial cell diameter of 1 μm and fungal spore diameter of 5 μm (Burrows et al. 2009), and emission rates were estimated from observed and modelled data (Burrows et al. 2009; Heald and Spracklen 2009). b) Predicted global relative abundance of bacteria and fungi in the near surface atmospheric boundary layer from observed quantitative PCR of rRNA genes in air above locations within different climatically defined biomes (estimated using data from ref (Archer et al. 2021)). Estimates are relative and cannot be used to directly infer absolute cell abundance, they are also domain-specific and cannot be directly compared between bacteria and fungi. c) Taxonomic composition of the ten most abundant classes of bacteria and fungi in the near surface atmospheric

1
2
3 boundary layer inferred from rigorously decontaminated environmental rRNA gene
4 sequence data from globally distributed locations (adapted from (Archer et al. 2021)).
5 Classes are ranked in order of relative abundance in soil to emphasize those that were over-
6 represented in air.
7
8
9

10
11 FIG. 3 | Factors affecting atmospheric microbial transport and macroecological outcomes.

12
13 a) Emission and recruitment of microorganisms from continental areas to the atmosphere is
14 strongly dependent on the source habitat and growing season. For example, the
15 phyllosphere is the major contributor in high productivity environments, whereas in low
16 productivity environments with reduced plant cover and in areas affected by intense
17 agricultural practices, the mobilisation of soil particulates becomes an increasingly
18 important emission source. A range of biotic, e.g., passive versus active spore release; and
19 abiotic factors, e.g., rainfall-induced aerosolization from soil, can also influence emissions.
20 Marine surfaces emit microorganisms through bubble-bursting processes driven by wind
21 speed, but the extent of marine emissions on a per surface area basis is low compared to
22 terrestrial sources. The effect of their spatiotemporal determinants and patterns on PBAP
23 emissions remain largely unknown and may include areas of relative high emission in coastal
24 and highly productive marine locations (Wang et al. 2015). b) Above terrestrial and marine
25 surfaces at local scales transport occurs largely within the atmospheric boundary layer over
26 relatively short distances. At larger spatial scales microbial transport involves transit above
27 the boundary layer and this results in longer residence times and greater environmental
28 filtering (Mayol et al. 2017; Archer et al. 2021). c) The association of cells with clouds or
29 particulates directly impacts survival and transport. Combustion particulates may be less
30 efficient vectors than mineral particles from desert dust and soil because they co-aerosolize
31 with lower numbers of microorganisms, and they are associated with toxic combustion
32 products (Archer and Pointing 2020). d) Deposition of cells from the atmosphere occurs via
33 wet deposition as rainfall, hail, and snow, and via dry deposition that relies on
34 sedimentation of cells due to gravity and occurs at lower velocities than wet deposition
35 (Woo and Yamamoto 2020). Wet deposition over continents is typically induced by ice
36 nucleation, which leads to preferential deposition of ice-nucleation active cells and
37 contributes to bioprecipitation and other feedbacks (Huffman et al. 2013; Morris et al. 2014;
38 Jung, Ge and Buie 2017).
39
40
41
42
43
44
45
46
47
48
49

50 FIG. 4 | Stress response and potential substrate utilization by atmospheric microorganisms.

51
52 Microbial spores are pre-adapted to survival during atmospheric transport and are over-
53 represented in the atmosphere compared to underlying surface habitats (Mayol et al. 2017;
54 Tignat-Perrier et al. 2019; Archer et al. 2021). Cells that are not adapted to environmental
55 stress undergo strong environmental filtering upon emission to the atmosphere. Cells
56 adapted to environmental stress possess a suite of stress response mechanisms which may
57 reflect source environment stress (Archer et al. 2021). A range of intracellular protective
58 and repair strategies to reactive oxygen species (ROS) and other stressors may be
59
60

1
2
3 complemented by extracellular protection due to extracellular polymeric substances (EPS)
4 that also confer protection against xeric, thermal, and ultraviolet radiation (UVR) stress
5 (Lebre, De Maayer and Cowan 2017). Bacterial isolates from the atmosphere have been
6 demonstrated to oxidize volatile organic compounds (VOC) such as methane and a range of
7 small organic compound carboxylic acids, pentose and hexose sugars, amino acids, and
8 phenolic compounds (Bianco et al. 2019). It has been speculated that oxidation of other
9 atmospheric trace gases and carbon fixation may also be potential pathways for activity in
10 the atmosphere although supporting data is currently lacking (Archer and Pointing 2020).
11 Finally, storage compounds accumulated in source environments could support basal
12 activity in airborne bacteria, providing the energy required to preserve cell integrity
13 (Cifuentes et al. 2019).
14
15
16
17
18
19

20
21 FIG. 5 | Biophysical role of atmospheric microorganisms in ice nucleation.

22 An important biophysical interaction of atmospheric microorganisms with cloud formation is
23 ice nucleation by specific microbial proteins that contributes to precipitation and the
24 hydrological cycle, as well as impacting microbial negative flux, i.e., deposition from the
25 atmospheric to surface habitats. a) Bacterial ice nucleating proteins (INpro) are long
26 repetitive proteins that are anchored on the surface of the outer membrane and nucleate
27 ice through a yet to be fully resolved mechanism (Pandey et al. 2016; Hudait et al. 2018;
28 Lukas et al. 2020). They may be cell-associated or retain activity in cell fragments.
29 Monomers or small oligomers nucleate ice at temperatures from -7 to -10 oC and higher-
30 order oligomers of INpro form at well-defined sizes that nucleate ice between -2 to -5 oC
31 (Wex et al. 2015; Ling et al. 2018). Fungal spores and hyphae are associated with INpro that
32 exhibit different molecular structure and properties to those in bacteria, but they are yet to
33 be fully described. b) Fungal spores are the dominant state for fungi in the atmosphere and
34 support surface associated INpro and ice nucleating activity (Pouleur et al. 1992; Morris et
35 al. 2013). c) Fungal hyphae occur very rarely in the atmosphere and so the major
36 contribution arises from fungal INpro that are easily dissociated from cell walls and adhere
37 to soil particulates that subsequently become aerosolized (O'Sullivan et al. 2015). d) Low
38 concentrations of atmospheric INpro nucleate cloud ice and may induce subsequent
39 secondary ice formation via different mechanisms, of which the best understood is riming
40 and splintering (Hallet-Mossop process) (Korolev and Leisner 2020). This process involves
41 freezing of supercooled cloud droplets that encounter primary ice particles (riming) and
42 subsequent ejection of frozen droplets that form independent ice particles (splintering).
43 Primary and secondary ice particles form precipitation through the Bergeron-Findeisen
44 mechanism where ice growth occurs at the expense of surrounding water droplets. Images
45 in panels a-d are not to scale, in a)-c) hexagonal shapes represent ice embryos forming on
46 INpro and in d) hexagonal shapes represent ice particles in clouds.
47
48
49
50
51
52
53
54
55
56
57

58 FIG. 6 | Anthropogenic risks to the atmospheric microbiology.
59
60

1
2
3 Direct and indirect anthropogenic forcing of the atmosphere will potentially impact the
4 emission and deposition flux, atmospheric burden, biodiversity, biophysical, and metabolic
5 activity of microorganisms in the atmosphere (Archer and Pointing 2020). We identify three
6 main outcomes of anthropogenic forcing that are of potential concern, although these are
7 likely inter-linked and overlap. They are: Altered atmospheric transport arising from changes
8 to particulate matter (PM) burden in the atmosphere; shifts in atmospheric microbial
9 emission and deposition flux due to change in surface microbial communities; and impacts
10 on human systems due to agricultural and human pathogen dispersal. We advocate for
11 future research focus to address these impacts.
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

For Peer Review

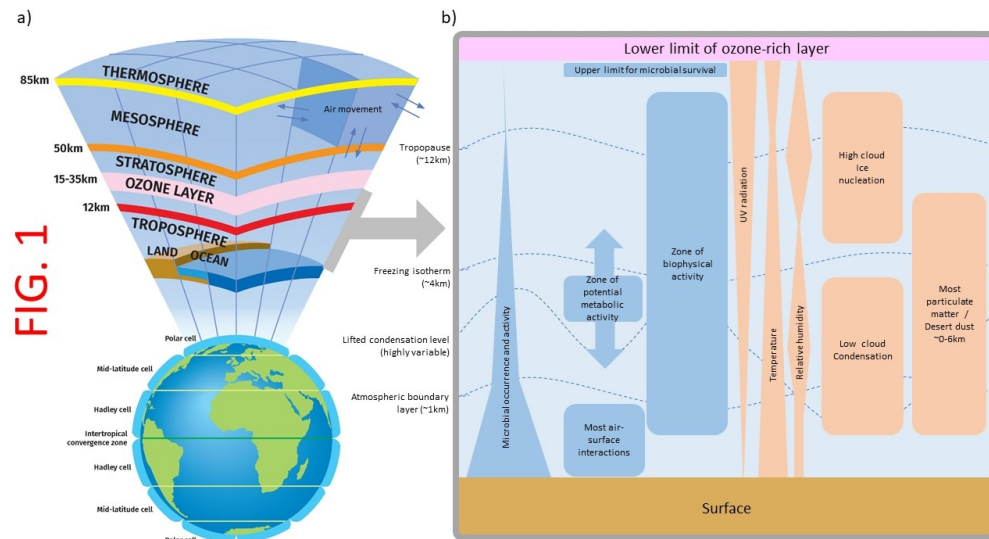


FIG. 1 | Defining the atmosphere as an ecological niche.

a) The atmosphere comprises a series of vertically delineated concentric layers that are primarily defined by thermal properties. The lowermost layer, the troposphere contains 75% of the molecular and gaseous mass of the atmosphere and almost all the water vapour, clouds and aerosolised particles (Barry and Chorley 2010) and including microorganisms. b) Four boundaries that occur at varying altitude within the troposphere are important in terms of atmospheric microbial ecology: The atmospheric boundary layer delineates the region of air closest to the surface where the bulk of surface-atmosphere interactions occur, and this includes exchange of microorganisms with terrestrial and marine ecosystems. The lifted condensation level (LCL) delineates the layer above which cloud formation occurs. It is variable with temperature and humidity and delineates the thermal limit for condensation of saturated water vapour on particulate surfaces. The freezing isotherm delineates the layer above which freezing occurs and is also the limit for active non-psychrophilic microbial metabolism. We note that the relationship between the atmospheric boundary layer, the LCL and the freezing isotherm, which are highly dynamic, is significantly more complex than presented in the figure, e.g., both the LCL and the freezing isotherm can sit at ground level well within the boundary layer. The boundary between the LCL and freezing isotherm represent the theoretical range for mesophilic microbial metabolic activity, with potential for psychrophilic and xerophilic activity to extend beyond this zone. The tropopause marks the upper limit of the troposphere and mixing with upper layers is limited by the thermal inversion in the stratosphere. The bulk of atmospheric ozone is in the lower stratosphere and this likely delineates the altitudinal limit of survival for many atmospheric microorganisms (Bryan et al. 2019).

338x190mm (96 x 96 DPI)

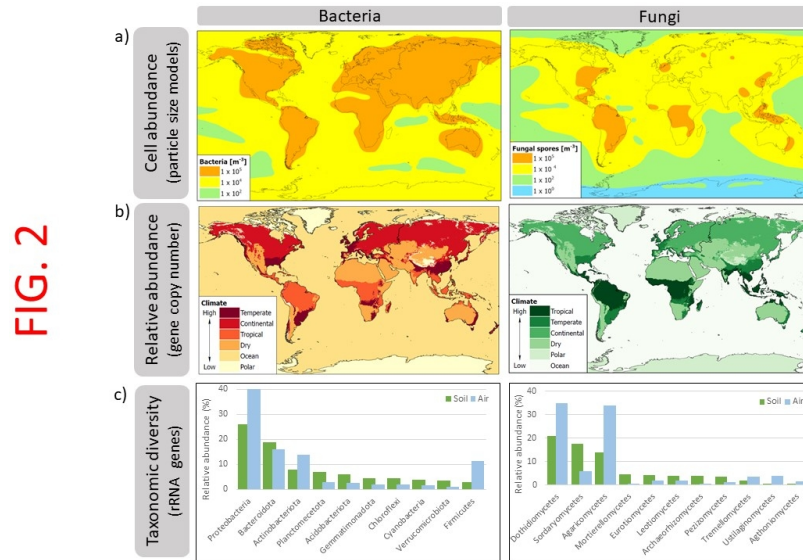


FIG. 2 | Patterns of microbial diversity and abundance in the atmosphere.

a) Simulated concentration of bacterial cells and fungal spores in the near surface atmospheric boundary layer (adapted from ref (Hoose, Kristjánsson and Burrows 2010)). Models assumed a uniform bacterial cell diameter of 1 μm and fungal spore diameter of 5 μm (Burrows et al. 2009), and emission rates were estimated from observed and modelled data (Burrows et al. 2009; Heald and Spracklen 2009). b) Predicted global relative abundance of bacteria and fungi in the near surface atmospheric boundary layer from observed quantitative PCR of rRNA genes in air above locations within different climatically defined biomes (estimated using data from ref (Archer et al. 2021)). Estimates are relative and cannot be used to directly infer absolute cell abundance, they are also domain-specific and cannot be directly compared between bacteria and fungi. c) Taxonomic composition of the ten most abundant classes of bacteria and fungi in the near surface atmospheric boundary layer inferred from rigorously decontaminated environmental rRNA gene sequence data from globally distributed locations (adapted from (Archer et al. 2021)). Classes are ranked in order of relative abundance in soil to emphasize those that were over-represented in air.

338x190mm (96 x 96 DPI)

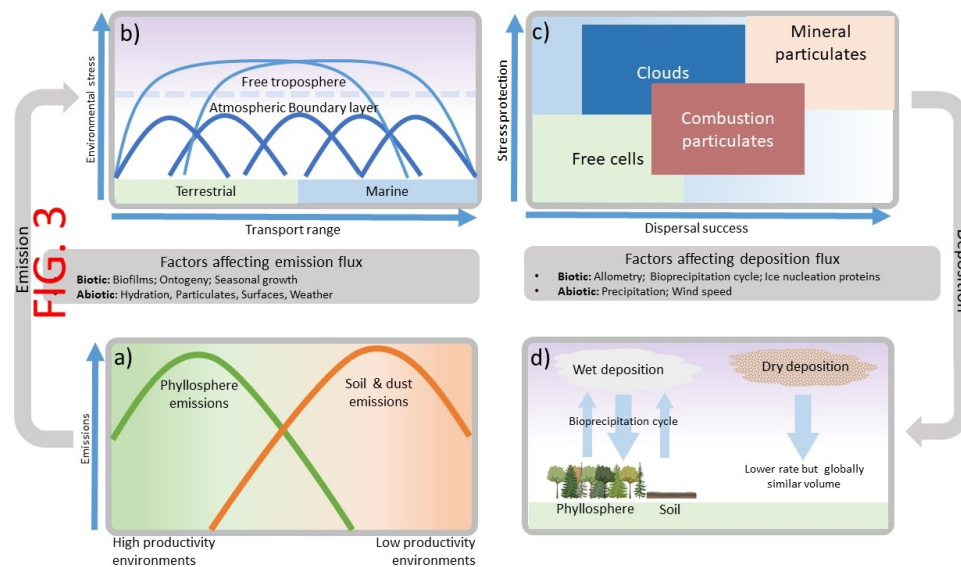


FIG. 3 | Factors affecting atmospheric microbial transport and macroecological outcomes.

a) Emission and recruitment of microorganisms from continental areas to the atmosphere is strongly dependent on the source habitat and growing season. For example, the phyllosphere is the major contributor in high productivity environments, whereas in low productivity environments with reduced plant cover and in areas affected by intense agricultural practices, the mobilisation of soil particulates becomes an increasingly important emission source. A range of biotic, e.g., passive versus active spore release; and abiotic factors, e.g., rainfall-induced aerosolization from soil, can also influence emissions. Marine surfaces emit microorganisms through bubble-bursting processes driven by wind speed, but the extent of marine emissions on a per surface area basis is low compared to terrestrial sources. The effect of their spatiotemporal determinants and patterns on PBAP emissions remain largely unknown and may include areas of relative high emission in coastal and highly productive marine locations (Wang et al. 2015). b) Above terrestrial and marine surfaces at local scales transport occurs largely within the atmospheric boundary layer over relatively short distances. At larger spatial scales microbial transport involves transit above the boundary layer and this results in longer residence times and greater environmental filtering (Mayol et al. 2017; Archer et al. 2021). c) The association of cells with clouds or particulates directly impacts survival and transport. Combustion particulates may be less efficient vectors than mineral particles from desert dust and soil because they co-aerosolize with lower numbers of microorganisms, and they are associated with toxic combustion products (Archer and Pointing 2020). d) Deposition of cells from the atmosphere occurs via wet deposition as rainfall, hail, and snow, and via dry deposition that relies on sedimentation of cells due to gravity and occurs at lower velocities than wet deposition (Woo and Yamamoto 2020). Wet deposition over continents is typically induced by ice nucleation, which leads to preferential deposition of ice-nucleation active cells and contributes to bioprecipitation and other feedbacks (Huffman et al. 2013; Morris et al. 2014; Joung, Ge and Buie 2017).

338x190mm (96 x 96 DPI)

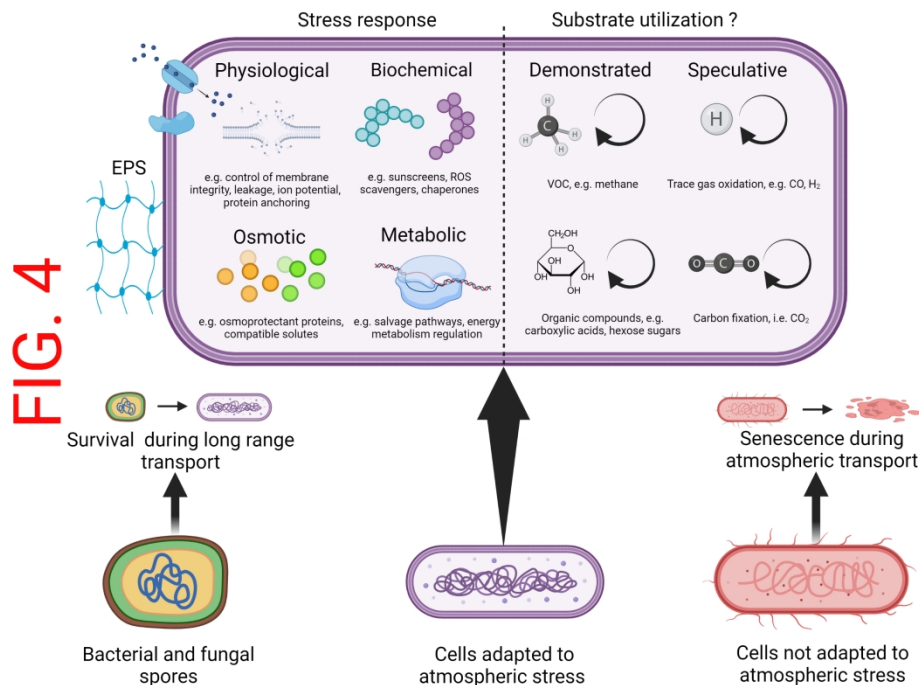


FIG. 4 | Stress response and potential substrate utilization by atmospheric microorganisms. Microbial spores are pre-adapted to survival during atmospheric transport and are over-represented in the atmosphere compared to underlying surface habitats (Mayol et al. 2017; Tignat-Perrier et al. 2019; Archer et al. 2021). Cells that are not adapted to environmental stress undergo strong environmental filtering upon emission to the atmosphere. Cells adapted to environmental stress possess a suite of stress response mechanisms which may reflect source environment stress (Archer et al. 2021). A range of intracellular protective and repair strategies to reactive oxygen species (ROS) and other stressors may be complemented by extracellular protection due to extracellular polymeric substances (EPS) that also confer protection against xeric, thermal, and ultraviolet radiation (UVR) stress (Lebre, De Maayer and Cowan 2017). Bacterial isolates from the atmosphere have been demonstrated to oxidize volatile organic compounds (VOC) such as methane and a range of small organic compound carboxylic acids, pentose and hexose sugars, amino acids, and phenolic compounds (Bianco et al. 2019). It has been speculated that oxidation of other atmospheric trace gases and carbon fixation may also be potential pathways for activity in the atmosphere although supporting data is currently lacking (Archer and Pointing 2020). Finally, storage compounds accumulated in source environments could support basal activity in airborne bacteria, providing the energy required to preserve cell integrity (Cifuentes et al. 2019).

645x452mm (118 x 118 DPI)

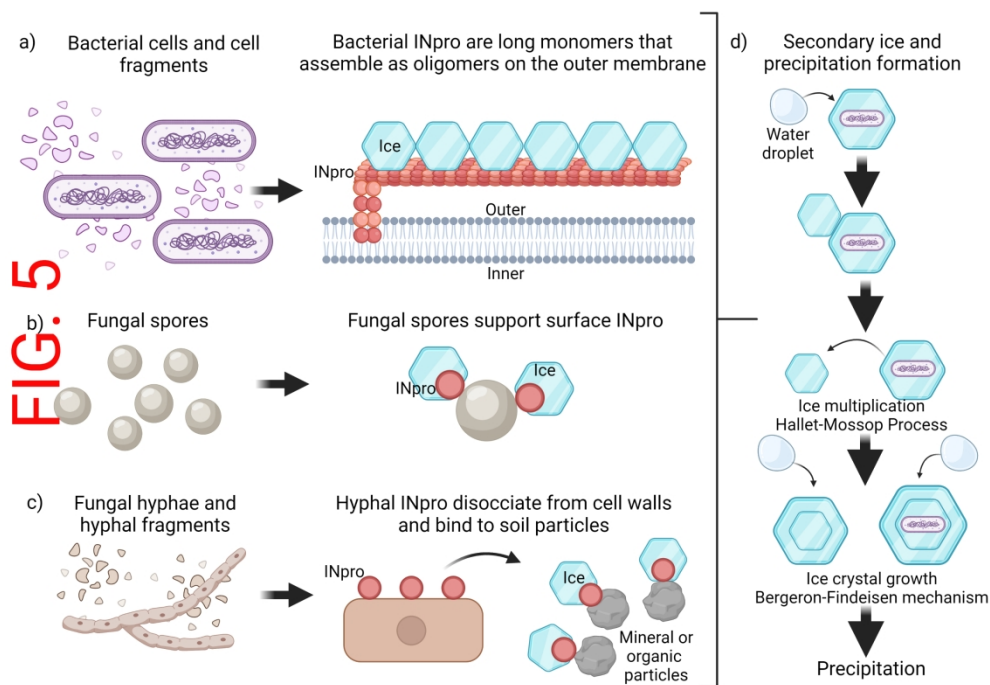


FIG. 5 | Biophysical role of atmospheric microorganisms in ice nucleation.

An important biophysical interaction of atmospheric microorganisms with cloud formation is ice nucleation by specific microbial proteins that contributes to precipitation and the hydrological cycle, as well as impacting microbial negative flux, i.e., deposition from the atmospheric to surface habitats. a) Bacterial ice nucleating proteins (INpro) are long repetitive proteins that are anchored on the surface of the outer membrane and nucleate ice through a yet to be fully resolved mechanism (Pandey et al. 2016; Hudait et al. 2018; Lukas et al. 2020). They may be cell-associated or retain activity in cell fragments. Monomers or small oligomers nucleate ice at temperatures from -7 to -10 oC and higher-order oligomers of INpro form at well-defined sizes that nucleate ice between -2 to -5 oC (Wex et al. 2015; Ling et al. 2018). Fungal spores and hyphae are associated with INpro that exhibit different molecular structure and properties to those in bacteria, but they are yet to be fully described. b) Fungal spores are the dominant state for fungi in the atmosphere and support surface associated INpro and ice nucleating activity (Pouleur et al. 1992; Morris et al. 2013). c) Fungal hyphae occur very rarely in the atmosphere and so the major contribution arises from fungal INpro that are easily dissociated from cell walls and adhere to soil particulates that subsequently become aerosolized (O'Sullivan et al. 2015). d) Low concentrations of atmospheric INpro nucleate cloud ice and may induce subsequent secondary ice formation via different mechanisms, of which the best understood is riming and splintering (Hallett-Mossop process) (Korolev and Leisner 2020). This process involves freezing of supercooled cloud droplets that encounter primary ice particles (riming) and subsequent ejection of frozen droplets that form independent ice particles (splintering). Primary and secondary ice particles form precipitation through the Bergeron-Findeisen mechanism where ice growth occurs at the expense of surrounding water droplets. Images in panels a-d are not to scale, in a)-c) hexagonal shapes represent ice embryos forming on INpro and in d) hexagonal shapes represent ice particles in clouds.

645x452mm (118 x 118 DPI)

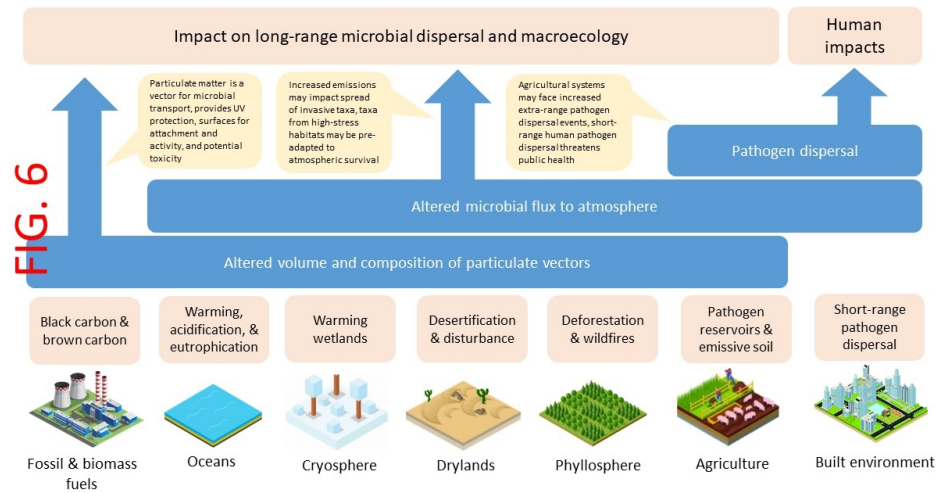


FIG. 6 | Anthropogenic risks to the atmospheric microbiology.

Direct and indirect anthropogenic forcing of the atmosphere will potentially impact the emission and deposition flux, atmospheric burden, biodiversity, biophysical, and metabolic activity of microorganisms in the atmosphere (Archer and Pointing 2020). We identify three main outcomes of anthropogenic forcing that are of potential concern, although these are likely inter-linked and overlap. They are: Altered atmospheric transport arising from changes to particulate matter (PM) burden in the atmosphere; shifts in atmospheric microbial emission and deposition flux due to change in surface microbial communities; and impacts on human systems due to agricultural and human pathogen dispersal. We advocate for future research focus to address these impacts.

338x190mm (96 x 96 DPI)