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CXCR2 intrinsically drives the maturation and function of neutrophils in mice

Pauline Delobel¹, Benjamin Ginter¹, Eliane Rubio¹, Karl Balabanian^{2,3}, Gwendal Lazenec^{1, 2*}

¹CNRS, SYS2DIAG-ALCEDIAG, Cap delta, 1682 rue de la Valsière, Montpellier, France.

²CNRS, GDR 3697 "Microenvironment of tumor niches", Micronit, France.

³Université Paris-Cité, Institut de Recherche Saint-Louis, INSERM U1160, Paris, France

* Correspondence:

Gwendal Lazenec

CNRS, SYS2DIAG, UMR9005, Cap delta, 1682 rue de la Valsière, Montpellier, F-34184, France.

Gwendal.lazenec@sys2diag.cnrs.fr

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Abstract

Neutrophils play a major role in the protection from infections but also in inflammation related to tumor microenvironment. However, cell-extrinsic and -intrinsic cues driving their function at steady state is still fragmentary. Using *Cxcr2* knock-out mice, we have evaluated the function of the chemokine receptor Cxcr2 in neutrophil physiology. We show here that Cxcr2 deficiency decreases the percentage of mature neutrophils in the spleen, but not in the bone marrow (BM). There is also an increase of aged CD62L^{lo} CXCR4^{hi} neutrophils in the spleen of KO animals. Spleen *Cxcr2*^{-/-} neutrophils display a reduced phagocytic ability, whereas BM neutrophils show an enhanced phagocytic ability compared to WT neutrophils. Spleen *Cxcr2*^{-/-} neutrophils show reduced reactive oxygen species production, F-actin and α -tubulin levels. Moreover, spleen *Cxcr2*^{-/-} neutrophils display an altered signaling with reduced phosphorylation of ERK1/2 and p38 MAPK, impaired PI3K-AKT, NF- κ B, TGF β and IFN γ pathways. Altogether, these results suggest that Cxcr2 is essential for neutrophil physiology.

Introduction

One of the first line of defense against pathogens such as bacteria, fungi, or parasites involves neutrophils, which are key mediator of innate immunity and inflammation. Neutrophils use different ways to clear the infection, including bacterial uptake (phagocytosis), phagolysosomal degradation of bacteria with a cocktail of antimicrobial factors and reactive oxygen species (ROS) (oxidative burst) or release of granules to neutralize extracellular pathogens (degranulation) (1). In addition, when intruders are too large or have escaped the other microbial killing processes, neutrophils can extrude a physical barrier to pathogen dissemination, called a neutrophil extracellular trap (NET), containing DNA, histones and granule proteases (2).

Neutrophils are the most abundant circulating leukocytes, representing 50% to 70% of all circulating leukocytes in humans and about 10 to 25% in mice (3). Neutrophils are relatively short lived cells (4), even if recent studies have questioned this aspect and it is now believed that human and murine neutrophils could have a half-life of 5 days or 18h, respectively (5).

Neutrophils arise from granulocyte–monocyte progenitors (GMPs), mostly within the bone marrow (BM) during hematopoiesis in response to several cytokines, principally granulocyte colony–stimulating factor (G-CSF) (6), but also from extramedullary tissues such as spleen (7-9). The first progenitor that is ‘neutrophil committed’ is the neutrophil promyelocyte (10), which then matures through granulocyte-committed precursors comprising myeloblasts, promyelocytes and myelocytes, to a post-mitotic or transition pool of metamyelocytes, band cells and segmented neutrophils (11). Mature post-mitotic neutrophils are released from the BM into the peripheral blood, extravasate from circulation into the tissues under the coordinate regulation of various adhesion molecules and chemokines (12). They are involved not only in the control of inflammation following infections or injuries but have also pro or anti-tumoral actions as tumor associated neutrophils (TANs) (13-15). In inflammatory sites, neutrophils fight the injury or infection and undergo apoptosis and are

59 phagocytosed by macrophages, once inflammation has been resolved (16). Senescent neutrophils can
60 also home back to the BM in a Cxcr4-dependent mechanism (17). The terminal differentiation of
61 neutrophils in the BM, before release into the bloodstream, is still a subject of debate, as some steps
62 of maturation could occur in other organs such as the spleen (9) and in the same line, neutrophil
63 progenitors have been found in the spleen (18). Maturation markers of neutrophils are also currently
64 discussed, but classically Ly6G, Cd101, Cxcr4 and Cxcr2 are used in mice (8, 12).

65 Cxcr2 appears as one of the key chemokine receptor expressed by neutrophils both in mice and
66 humans (19, 20). Cxcr2 binds the chemokines Cxcl1, 2, 3, 5, 6, 7 and 8 in human, which all have pro-
67 angiogenic properties and are located in a short cluster of chromosome 4 (21-23). Cxcr2 ligand action
68 is conditioned by its interaction with proteoglycans (24) and Cxcr2 signals through multiple
69 pathways including PI3K and Src (25). In addition, recent work has highlighted the role of Cxcr2 in
70 tumorigenesis, in particular through tumor-associated neutrophils (14, 23, 26, 27), but also in the
71 effects microbiota on pituitary function (28). *Cxcr2* knockout mice have been generated and are
72 characterized by a splenomegaly due to an increase of metamyelocytes, mature neutrophils and B
73 lymphocytes (29). These mice also display a defect in neutrophil recruitment after infection (30).

74 So far, the mechanism of Cxcr2 action in neutrophils remains poorly understood. In this study, we
75 have investigated the role of Cxcr2 in mouse neutrophils in the spleen and the BM, taking advantage
76 of *Cxcr2*^{-/-} mice. Our data show that Cxcr2 impairment in neutrophils affects differently spleen and
77 BM neutrophils in terms of maturation, phagocytosis and ROS production. Moreover, we analyzed at
78 the transcriptomic level the pathways that were altered by *Cxcr2* deletion in neutrophils. Altogether,
79 these results suggest that proper Cxcr2 expression and function is required for maturation and
80 effector functions of peripheral neutrophils.

84 Materials and methods**85 Animal Models and housing**

86 All animal experiments conformed to our animal protocols that were reviewed and approved by the
87 Institutional Animal Care and Use Committee. *Cxcr2*^{-/-} mice (29) were obtained from the Jackson
88 Laboratory. *Cxcr2*^{-/-} mice were backcrossed in FVB genetic background for more than 12 generations.
89 Control (WT) mice were also in a FVB background. All mice were housed in a SOPF (Specific and
90 Opportunistic Pathogen Free) animal facility.

91 Isolation of cells

92 Cells from the BM were isolated by centrifugation from the femurs and tibias of the animals, whereas
93 spleens were mashed on 100 µm nylon cell strainer. After centrifugation, red blood cells were
94 eliminated by treatment with ACK buffer (0.155 mM NH₄Cl, 1 mM KHCO₃, 0.1 mM EDTA) and
95 filtered on a 40 µm nylon cell strainer. After ACK treatment, cells were filtered on a 40 µm nylon cell
96 strainer. For neutrophil isolation, a first enrichment with EasySep™ Mouse CD11b Positive Selection
97 kit (StemCell technologies, Grenoble, France) was performed followed by cell sorting of CD45+
98 CD11b+ Ly6G+ F4/80- cells on an ARIA IIu FACS sorter (Becton Dickinson, Le Pont-de-Claix,
99 France).

100 Flow cytometry

101 Flow cytometry experiments were performed with the following conjugated antibodies from
102 Biolegend (London, United Kingdom): anti-mouse CD11b (clone M1/70), CD45 (clone 30-F11),
103 Cxcr2 (clone SA044G4), Ly6G (clone 1A8), CD62L (clone MEL-14) or BD Biosciences (BD
104 Biosciences, Le Pont-de-Claix, France): Cxcr4 (clone 2B11), or Ebiosciences (Fisher Scientific,
105 Illkirch, France): CD101 (clone Moushi101), Fixable viability dye (65-0866). Flow analysis was
106 performed on live singlets with a LSR II Fortessa flow cytometer (Becton Dickinson, Le Pont-de-
107 Claix, France). Data were analyzed using FlowJo (Tree Star).

108

109 Intracellular flow cytometry

110 Cells were first stained with antibodies directed against extracellular markers (CD45, CD11b, Ly6G)
111 and then permeabilized with Cytofix/Cytoperm and Permash buffer (BD Biosciences). Intracellular
112 staining of F-actin was performed with Phalloidin, Fluorescein Isothiocyanate Labeled (Sigma-
113 Aldrich, ref P5282), whereas α -tubulin was labeled with α -Tubulin antibody (clone 11H10, Cell
114 Signaling, ref 2125) followed by secondary an anti-rabbit IgG antibody coupled to Alexa Fluor 555
115 (ThermoFisher, ref A27039). Phospho-p42/44 MAPK ERK1/2 (Thr 202/ Tyr 204) (Cell signaling, ref
116 4370) followed by secondary an anti-rabbit IgG antibody coupled to Alexa Fluor 555 (ThermoFisher,
117 ref A27039) was used to detect Phospho-p42/44 MAPK ERK1/2. p-p38 MAPK (Thr 180/ Tyr 182)
118 coupled to PE (cell Signaling 6908), was used to detect phospho-p38 MAPK.

119 Annexin V staining and measure of mortality

120 To determine the proportion of apoptotic and dead cells, fresh neutrophils were stained with Annexin-
121 FITC and propidium iodide (PI) according to manufacturer instruction (Invitrogen, ref V13242).

122 Phagocytosis assay

123 CD45⁺ CD11b⁺ Ly6G⁺ neutrophils were incubated for various times at 37°C with with E. coli Red
124 Phrodo bioparticles (ThermoFisher, Illkirch, France) at a concentration of 15 μ g/ml and analyzed in
125 a kinetic manner by flow cytometry with a LSR II Fortessa flow cytometer (Becton Dickinson, Le
126 Pont-de-Claix, France). Data were analyzed using FlowJo (Tree Star). When opsonized particles were
127 used, they were opsonized for 1h at 37°C with opsonizing reagent (E2870, ThermoFisher, Illkirch,
128 France), according to manufacturer instructions.

129 Reactive oxygen species and mitochondrial superoxide quantification

130 Reactive oxygen species and mitochondrial quantification was performed by labelling fresh CD45⁺
131 CD11b⁺ Ly6G⁺ neutrophils for 20 min at 37°C, using CellRox Orange reagent (ref. C10443) and
132 Mitosox reagent (ref. M36008) respectively, following the manufacturer's instructions (Molecular
133 Probes, ThermoFisher Scientific).

RNA extraction and RNA-seq data processing

Total RNA was isolated using TRIzol reagent (Fisher Scientific, Illkirch, France), as described by the manufacturer. RNA integrity and quality were verified using RNA ScreenTape kit and Tapestation 2200 apparatus from AGILENT (Les Ulis, France). cDNA libraries were synthesized using NEBNext® rRNA Depletion and Ultra™ II Directional RNA Library Prep Kit (New England Biolabs, Evry-Courcouronnes, France). Library quality was checked on Tapestation 2200 apparatus from AGILENT (Les Ulis, France) with DNA 1000 ScreenTape. Samples were sequenced on Novaseq 6000 (Illumina) with an average sequencing depth of 30 million of paired-end reads. Length of the reads was 150 bp. Each 24 Plex Samples was sequenced on one Illumina SP FlowCell (2*800 million of 150bases reads). Raw sequencing data was quality-controlled with the FastQC program. Low quality reads were trimmed or removed using Trimmer (minimum length: 120 bp). Reads were aligned to the mouse reference genome (mm10 build) with the Star tool. Gene counts were obtained by read counting software Htseq. Normalization and differential analysis were performed with the DESeq2 package with Benjamini-Hochberg FDR multiple testing correction ($p < 0.05$; 1.5-fold or higher change) comparing WT and KO animals. The data discussed in this publication have been deposited in NCBI's Gene Expression Omnibus (31) and are accessible through GEO Series accession number GSE209860 (<https://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE209860>).

Bioinformatic analysis

To assess biological interpretation of the most differentially expressed genes, we used Gene ontology (GO) enrichment analysis. A gene set enrichment analysis (GSEA) was performed using signatures from GSEA collections. A normalized enrichment score (NES) was calculated for each gene set and only gene sets with an adjusted p value < 0.05 were selected.

Statistics

159 Statistical analyses were carried out using unpaired Mann-Whitney test.

160

Results

***Cxcr2* invalidation affects the maturation of neutrophils**

To evaluate the impact of *Cxcr2* knockout on neutrophil distribution and function, we first measured the presence of CD45⁺ CD11b⁺ Ly6G⁺ neutrophils in the BM and the spleen by flow cytometry (Fig.1A). Ly6G⁺ neutrophils were composed of two subpopulations with high (Ly6G^{hi}) or low (Ly6G^{lo}) levels of Ly6 (Fig. 1A). In BM, the percentage of total Ly6G⁺ (Ly6G^{hi} + Ly6G^{lo}) neutrophils was increased by about 30% in *Cxcr2*^{-/-} compared to WT animals, and this was much more pronounced in the spleen, with more than 8 - fold more neutrophils in *Cxcr2*^{-/-} animals (Fig. 1B). Of particular note, the percentage of CD45⁺ cells was not different in WT and KO spleens, whereas it slightly increased (88.7% vs 95.2%) in KO BM compared to WT BM (Supplemental Fig. 1). No major change in FSC and SSC distribution was observed for Ly6G⁺ neutrophils (Supplemental Fig. 2). In terms of absolute numbers, we observed a 5-fold increase of the total number of CD45⁺ cells and CD45⁺ CD11b⁺ Ly6G⁺ neutrophils in the spleen of *Cxcr2*^{-/-} animals, whereas minimal differences between *Cxcr2*^{-/-} and WT animals was seen in BM (no change of the number of CD45⁺ cells and less than 30 % increase for CD45⁺ CD11b⁺ Ly6G⁺) (Supplemental Fig. 3). This obviously creates a bias in the analysis of any type of subpopulation and for this reason, we decided to continue to look at percentage of subpopulations of neutrophils among neutrophils, to avoid this.

The level of Ly6G expression has been correlated with the degree of maturation of neutrophils with Ly6G^{hi} neutrophils being the most mature (32). In BM, there was a slight reduction of the percentage of immature Ly6G^{lo} neutrophils among total Ly6G⁺ in *Cxcr2*^{-/-} animals, whereas there was an increase by 2-fold of immature Ly6G^{lo} neutrophils in *Cxcr2*^{-/-} spleen (Fig. 1C). To strengthen these results, we used the CD101 marker, which characterizes mature neutrophils (8). Among neutrophils, we observed a reduction of CD101⁺ mature neutrophils in the spleen, but an increase of mature neutrophils in BM (Fig. 1D, F). Next, we also looked at *Cxcr4* expression in neutrophils, which is

186 correlated not only to immaturity of neutrophils (8), but also to aged neutrophils (33). The percentage
187 of Cxcr4⁺ neutrophils increased twice in the spleen, but was unaffected in the BM (Fig. 1E, G),
188 confirming the increase of the proportion of immature neutrophils in the neutrophil fraction of the
189 spleen. To assess more precisely whether these neutrophils corresponded to immature or aged
190 neutrophils in the spleen of KO animals, we used the marker CD62L (L-Selectin), in addition to
191 CXCR4 to identify "aged" CD62L^{lo}- CXCR4^{hi} neutrophils, as previously described (33). We
192 observed that the proportion of aged CD62L^{lo}- CXCR4^{hi} cells among neutrophils increased in KO
193 spleen compared to WT, but there was no significant change in BM (Supplemental Fig. 4).
194 To further assess the nature of neutrophils, we isolated by cell sorting CD45⁺ CD11b⁺ Ly6G⁺
195 neutrophils from WT and KO BM and spleen and colored them with Giemsa (Supplemental Fig. 9).
196 WT BM neutrophils displayed a condensed round nucleus, with a small cytoplasm, whereas KO BM
197 neutrophils seem to have a larger ring-shaped nucleus, which could suggest that they are more
198 mature. In the spleen, WT neutrophils had a strongly colored ring-shaped nucleus. Spleen KO
199 neutrophils exhibited a larger nucleus with a less pronounced staining and also a cytoplasmic center,
200 which could be reminiscent of band neutrophils (34). These morphological differences suggest that
201 WT and KO neutrophils might have distinct features.

202 ***Cxcr2*^{-/-} neutrophils display altered phagocytic ability**

203 Phagocytosis is one of the major function of neutrophils to eliminate infections. Thus, we analyzed
204 the phagocytic abilities of BM and spleen neutrophils. Kinetics of phagocytosis showed that BM
205 *Cxcr2*^{-/-} neutrophils had an enhanced phagocytic ability compared to WT neutrophils (Fig.2A),
206 whereas spleen *Cxcr2*^{-/-} neutrophils were less phagocytic than spleen WT neutrophils (Fig. 2B). We
207 have also performed an additional analysis, by separating Ly6G^{hi} and Ly6G^{lo} neutrophils
208 (Supplemental Fig. 5). Similar results were obtained for Ly6G^{hi} neutrophils as for Ly6G⁺
209 neutrophils, which is an increase of phagocytosis for BM KO neutrophils and a decrease of

phagocytosis for spleen KO neutrophils. Concerning Ly6G^{lo} neutrophils, they were less phagocytic than Ly6G^{hi} neutrophils, both in the spleen and BM. Moreover, BM KO Ly6G^{lo} were also more phagocytic than BM WT Ly6G^{lo} neutrophils. On the other hand, no difference was seen for spleen Ly6G^{lo} neutrophils between WT and KO animals. In addition, when using opsonized bioparticles (Supplemental Fig. 6), we obtained the same trend of difference between WT and KO neutrophils as for non-opsonized particles.

To better understand the mechanisms underlying these changes in phagocytic ability, we looked at ROS production and to actin-tubulin cytoskeleton organization of neutrophils, which are essential for phagocytosis (35). We observed a strong reduction in the production of cytoplasmic ROS (Fig 3A, B) and mitochondrial superoxide (Fig. 3C, D) of spleen *Cxcr2*^{-/-} neutrophils, whereas no difference could be measured between BM WT and *Cxcr2*^{-/-} neutrophils. Spleen *Cxcr2*^{-/-} neutrophils had also lower levels of F-Actin (Fig. 4A, B) as well as tubulin (Fig. 4C, D) compared to WT, but BM neutrophils were unaffected by *Cxcr2* impairment. These results of decreased levels of F-actin and tubulin were also confirmed, when comparing MFI values for F-actin and tubulin for the different types of neutrophils (Supplemental Fig. 7). Altogether, these results could account for the lower phagocytic ability of spleen *Cxcr2*^{-/-} neutrophils, which is in agreement with a lower maturity.

Spleen *Cxcr2*^{-/-} neutrophils have a higher viability

Neutrophil are short term living cells, so we wondered whether *Cxcr2* could modulate their survival ability. We observed that the percentages of apoptotic or dead cells were reduced for spleen *Cxcr2*^{-/-} neutrophils (Fig. 5B and 5C, respectively), whereas it was similar for WT and *Cxcr2*^{-/-} BM neutrophils. So, this suggests that the reduced phagocytic ability of spleen *Cxcr2*^{-/-} neutrophils is not a consequence of an impaired survival. We have also measured apoptosis and death after a 2h incubation at 37°C, and obtained similar results (Supplemental Fig. 8), which suggests that an

increased apoptosis or death after 2h incubation does not affect the phagocytosis ability of neutrophils.

Transcriptomic analysis confirms the impaired maturation of spleen *Cxcr2*^{-/-} neutrophils

We next focused on spleen neutrophils to decipher whether and how *Cxcr2* deficiency impacts their molecular identity. RNAseq analyses of WT and *Cxcr2*^{-/-} spleen neutrophils showed that more than 2,500 transcripts were up-regulated and about the same number down-regulated in *Cxcr2*^{-/-} spleen neutrophils (Fig. 6A, B). We then focused on their maturation using the neutrophil maturation signature reported by Xie and collaborators (36). GSEA analysis confirmed that spleen *Cxcr2*^{-/-} were less mature than their WT counterparts (Fig. 6D, E), with down regulation of genes encoding Cathepsin D (*Ctsd*), JunB or IL-1 β involved in Netosis (2), Arginase-2 (*Arg2*) controlling extra-urea cycle arginine metabolism and nitric oxide synthesis (12), C-type lectin receptor *Clec4d* crucial for bacteria elimination (37) or *Selp1g* (CD162), important for the rolling of neutrophils (38) . In addition, there was a cluster of GSEA Biological process (BP) signatures showing a down-regulation of the chemotaxis and migration of spleen *Cxcr2*^{-/-} neutrophils (Fig. 6F), suggesting a decrease in the migration ability of these neutrophils.

ERK and p38 MAPK pathways are down-regulated in spleen *Cxcr2*^{-/-} neutrophils

To look at the signaling that were affected in spleen *Cxcr2*^{-/-} neutrophils, we first focused on ERK and p38 MAPK signaling, which are known to modulate the migration and adhesion of neutrophils (39, 40). RNAseq data showed that both ERK1 and ERK2 cascade (Fig. 7A and C), as well as p38MAPK cascade (Fig. 7B and D), were down regulated in spleen *Cxcr2*^{-/-} neutrophils. Concerning ERK1/2 cascade, there is in particular a down-regulation of the AP-1 Transcription Factor Jun, of the Cyclic AMP-Dependent Transcription Factor Atf-3, IL1 β , growth factors such as TGF β 1 and IGF1, multiple chemokines such as Ccl3, 4, 15, 18, 23 and of a number of kinases such as Ptk2b (Protein Tyrosine Kinase 2 Beta), the tyrosine-protein kinase Syk, Csk (C-Terminal Src Kinase) or the

serine/threonine protein kinase BRAF (Fig. 7C). For p38MAPK cascade, we observed in particular a down regulation of Mitogen-Activated Protein Kinase 14 (Mapk14), Mitogen-Activated Protein Kinase Kinase Kinase 3 (Map3k3), Map3k5, and cytokines such as IL1 β , HGF.

To confirm these results, we measured by flow cytometry the intracellular content of Phospho-p42/44 MAPK ERK1/2 and phospho-p38 MAPK (Thr 180/ Tyr 182). The mean of fluorescence (MFI) of p-ERK1/2 and p-p38 were down-regulated by about 50% in spleen *Cxcr2*^{-/-} neutrophils compared to WT neutrophils (Fig. 7E and 7F, respectively) confirming the alterations of these pathways.

Several pathways are impaired in spleen *Cxcr2*^{-/-} neutrophils

To further investigate the mechanisms involved in the changes observed in spleen *Cxcr2*^{-/-} neutrophils, we explored the main pathways that were affected at the transcriptomic level. GSEA analysis showed a reduction of PI3K-AKT signaling (Fig. 8A), TNF α signaling via NF κ B (Fig. 8B), NIK-NF κ B signaling (Fig. 8C), as well of the cellular response to IL-1 (Fig. 8D) and TGF β signaling (Fig. 8E) and signaling by IFN γ (Fig. 8F) signatures in spleen *Cxcr2*^{-/-} neutrophils compared to WT neutrophils. Among the genes involved in PI3K-AKT pathway, we observed a down-regulation of Phosphatidylinositol-4,5-Bisphosphate 3-Kinase Catalytic Subunit Alpha, Delta and Gamma (Pik3ca, Pik3cd and Pik3cg), and Phosphoinositide-3-Kinase Regulatory Subunit 1, 2 and 5 (Pik3r1, Pik3r2, Pik3r5), Janus Kinase 1 (Jak1), Glycogen Synthase Kinase 3 Beta (GSK3b), and Kras, which are crucial for this pathway.

NF- κ B pathway was also impaired with in particular down-regulation of genes such as NF- κ B Subunits p52 (Nfkb2), Relb, C-Rel (Rel), TNF Alpha Induced Protein 2 and 3 (Tnfaip2 and Tnfaip3), Tnfaip3 Interacting Protein 1 (Tnfp1), TNF Receptor-Associated Factor 1, 3 and 5 (Traf1, Traf3, Traf5) or TNF Receptor Superfamily Member 10b (Tnfrsf10b).

Upstream of NF- κ B activation, cellular response to IL-1 was also altered, with a down-regulation of IL-1 itself, but also of Interleukin 1 Receptor Type 1 (Il1r1). Two other pathways were also linked to

282 PI3K/Akt, ERK, p38 and NF- κ B signaling, namely TGF β and IFN γ signaling. TGF β signaling was
283 impaired with a decrease in Transforming Growth Factor Beta 1 (Tgfb1), Transforming Growth
284 Factor Beta Receptor 1 and 2 (Tgfbr1 and Tgfbr2), and Smad2, 3, and 4. IFN γ signaling is also acting
285 through ERK and MAPK and PI3K signaling and exhibits in particular a down-regulation of Janus
286 kinase 2 (Jak2), suppressor of cytokine signaling protein 1n and 3 (Socs1 , Socs3), Signal transducer
287 and activator of transcription 4 (Stat4), Intercellular adhesion molecule-1 (ICAM-1) and Integrin
288 Subunit Beta 7 (ITGB7).

289

Discussion

Although the role of neutrophils in general inflammation but also tumor microenvironment inflammation is growing, the factors controlling their function are still not completely identified. In particular, the involvement of Cxcr2 in neutrophil physiology and features, as well as its impact on signalization in neutrophils is still a matter of debate. In this study, we report that impairment of *Cxcr2* leads to a modest increase of the percentage of neutrophils in the BM, but a strong one in the spleen, in agreement with previous studies (29). We also show that Cxcr2 differentially affects the maturation state of neutrophils in the spleen and the BM. More precisely, based on the maturation markers Ly6G (32) and Cd101 (8) expression, *Cxcr2* deletion led an increase of Ly6G^{lo} or CD101-immature neutrophils in the spleen, but to a decrease of immature neutrophils in the BM. The presence of resident immature neutrophils in the spleen is thought to serve as a reservoir of neutrophils, which will undergo a rapid proliferation and mobilization in case of infection to increase the number of active mature neutrophils (18, 32). Spleen is also a site of accumulation and destruction of neutrophils in human (41), so the presence of a high number of immature neutrophils could correspond to both resident and recruited neutrophils. On the other hand, considering that the BM is the main site of production of neutrophils, this would suggest that Cxcr2 impairment does not alter the maturation process on neutrophils in the BM, or that there could be a retention of mature neutrophils in BM. This latter hypothesis is unlikely, as we did not see any difference in the percentage Cxcr4⁺ neutrophils in the BM. Cxcr4 is indeed crucial for the retention of neutrophils in the BM (17), even though other studies have shown that the CXCR4 antagonist plerixafor did not mobilize neutrophils from the BM, but rather enhanced the release of neutrophils in the circulation from the marginated pool present in the lung (42). In contrast, the percentage of Cxcr4⁺ neutrophils was increased 2-fold in the spleen, which could account for several features: this could be the sign of more immature, since Cxcr4 expression is high in proliferating immature neutrophils (8). However, high levels of Cxcr4 can also be seen in senescent or aging neutrophils (17, 43). To clarify this point,

we analyzed the presence of aged CD62L^{lo} – CXCR4^{hi} neutrophils as defined previously (33). This shows that there is an increase of aged CD62L^{lo} – CXCR4^{hi} neutrophils in the spleen of KO animals, but no change in BM. In addition, we also observed that the percentage of apoptotic or dead neutrophils was reduced among spleen *Cxcr2*^{-/-} neutrophils compared to WT, whereas no difference was seen in BM. Overall, our data suggest that spleen *Cxcr2*^{-/-} neutrophils are distinct from BM neutrophils, as they are healthier and have more immature features (Ly6G^{lo} Cd101⁻ Cxcr4⁺) than their WT counterparts.

To go further in the understanding of the effects of *Cxcr2* impairment, we have observed that spleen *Cxcr2*^{-/-} neutrophils exhibited a reduced phagocytic ability than WT neutrophils. On the other hand, BM *Cxcr2*^{-/-} neutrophils had a higher phagocytic ability than the WT. This difference between spleen and BM neutrophils might be explained by their difference of maturation mentioned above. This is also true for basal phagocytic ability of WT neutrophils, which is higher in the spleen than in the BM. This might be due to a higher proportion of mature CD101⁺ neutrophils in WT spleen compared to WT BM (Fig. 1F), as we have also shown that Ly6G^{hi} neutrophils had a better phagocytic ability than Ly6G^{lo} neutrophils (Supplemental Fig. 5).

To explore the difference in phagocytic ability of WT and *Cxcr2*^{-/-} neutrophils more thoroughly, we looked at ROS production and actin and tubulin cytoskeleton, which are key elements in phagocytosis (35, 44). The percentage of cells with high cytoplasmic ROS, mitochondrial superoxide, F-actin and α -tubulin was reduced among spleen *Cxcr2*^{-/-} neutrophils compared to WT, but was not modified in BM neutrophils. This reduction could explain part of the reduced phagocytic ability of spleen *Cxcr2*^{-/-} neutrophils, although one can also notice that decreased phagocytic ability of WT BM neutrophils compared to WT spleen neutrophils is not correlated to their actin levels, suggesting that other parameters might be involved. It is interesting to notice that aged neutrophils are resistant to infections and display also a reduction of actin levels (43). Moreover, it has also been shown that in some cases, immature neutrophils could produce less ROS (8).

340 It was essential to analyze the mechanisms underlying the differences between spleen *Cxcr2*^{-/-} and
341 WT neutrophils. RNAseq analysis of both types of isolated neutrophils confirmed the defect in
342 maturation of spleen *Cxcr2*^{-/-} neutrophils, with in particular a down regulation of genes involved in
343 Netosis (Cathepsin D, JunB or IL-1 β) (45), of Arginase-2 controlling extra-urea cycle arginine
344 metabolism and nitric oxide synthesis, of Clec4d implicated in bacteria elimination (46) or Selp1g
345 (CD162), essential for the rolling of neutrophils (47). Several GO pathways of migration and
346 chemotaxis constituted a down-regulated cluster, suggesting also an impairment of spleen *Cxcr2*^{-/-}
347 chemotaxis. This could explain an accumulation of neutrophils in the spleen, with a weak ability to
348 migrate to other tissues. Earlier studies have shown that in inflammatory conditions, *Cxcr2* was
349 essential for the recruitment of neutrophils (48).

350 In terms of signaling, we report a down regulation of ERK and p38 MAPK pathways, both at the
351 transcriptomic level, but also when assessing the phosphorylation of ERK and p38 at the protein
352 level. The ERK and p38 MAP kinases are strongly stimulated in neutrophils upon activation by G-
353 protein coupled receptors agonists. Moreover, p38 MAPK is critical for the release of primary and
354 secondary granules, but not that of secretory vesicles by neutrophils (49). p38 inhibition has been
355 shown to delay apoptosis of neutrophils (50), which is in agreement of the reduced rate of apoptosis
356 that we observed in spleen *Cxcr2*^{-/-} neutrophils. Moreover, p38 MAPK promotes chemotaxis
357 towards fMLP by interfering with GRK2-mediated desensitization, whereas ERK MAPK is
358 inhibiting it (51, 52). However, the role of ERK MAPK pathway in neutrophil functions remains
359 unclear, due to contradictory studies (37).

360 In addition, PI3K-Akt, TNF α signaling and NF- κ B signaling, were also altered. TNF α is essential to
361 trigger neutrophil activation and phagocytic activity (53). It is interesting to note that in neutrophils,
362 the production of ROS is dependent on PI3K and ERK (54). Moreover, PI3K, ERK and p38 are
363 necessary for efficient phagocytosis (55). It has also been reported that PI3K γ ^{-/-} neutrophils display a

defect in migration towards fMLP, C5a, Cxcl8 or Ccl3 and respiratory burst upon activation by C5a or fMLP (56), but another study suggests that PI3K is not required for chemotaxis towards fMLP (51).

Cellular response to IL-1 was also reduced. IL1 α and IL1 β are very potent mediators of inflammation response, but their role in neutrophils remains poorly understood. Their main effect on neutrophils could be to increase their survival (37).

We also observed a down-regulation of TGF β and IFN γ pathways. In the steady state situation, the role of TGF β on neutrophil function is poorly understood. However, in cancer context, TGF β is responsible for promoting the generation of pro-tumoral type N2 neutrophils (57), whereas type IFN β and IFN γ might favor anti-tumoral type N2 neutrophils (58, 59). In non-cancerous situation, IFN γ has also been shown to enhance, or prime, increased ROS production in combination with a secondary stimulus and to promote phagocytosis (60), which could account for the decrease in ROS and phagocytosis that we observed in spleen *Cxcr2*^{-/-} neutrophils. As treatment of PMNs with IFN γ increases the production of TNF α and IL-1 β (61), this could also account for the down-regulation of TNF α and IL1 β signaling that we report.

In conclusion, this work highlights the multiple roles played by the chemokine receptor Cxcr2 in neutrophils and reinforces the importance of localization of neutrophils in terms of action and features. The identification of the pathways that are dependent on Cxcr2 and their further investigation will be essential to understand the roles of Cxcr2 in neutrophils in the steady state or inflammatory situation but also in the tumoral context.

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1 Conflict of Interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

2 Author Contributions

P.B., B.G., E.R. and have contributed to the investigation. K.B. participated to writing review and editing. GL was in charge of the conceptualization, investigation and supervision of the project, funding acquisition and writing original draft preparation. All authors have read, revised and agreed to the published version of the manuscript.

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Figure legends

Fig.1: *Cxcr2* knock-out decreases the percentage of mature neutrophils in the spleen

A. Representative dot plots of the gating strategy of CD45⁺ CD11b⁺ Ly6G^{hi} and CD45⁺ CD11b⁺ Ly6G^{lo} neutrophils among CD45⁺ cells in WT and *Cxcr2*^{-/-} bone marrow (BM) and spleen (SP). **B.** Quantification of the percentage of the total neutrophils (CD11b⁺ Ly6G⁺; sum of Ly6G^{hi} and Ly6G^{lo}) in the CD45⁺ fraction. **C.** Quantification of CD11b⁺ Ly6G^{hi} cells (left panel) and CD11b⁺ Ly6G^{lo} (right panel) in the CD11b⁺ Ly6G⁺ (Ly6G^{hi} and Ly6G^{lo} fractions) population. **D.** Gating strategy to identify CD101⁺ neutrophils in the CD45⁺ CD11b⁺ Ly6G⁺ fraction. **E.** Gating strategy to identify CXCR4⁺ neutrophils in the CD45⁺ CD11b⁺ Ly6G⁺ fraction. **F.** Percentage of mature CD101⁺ neutrophils in the CD11b⁺ Ly6G⁺ fraction. **G.** Percentage of CXCR4⁺ neutrophils in the CD11b⁺ Ly6G⁺ fraction. Data represent the mean $\pm$ SEM of at least 6 animals (Mann-Whitney test, NS: non-significant, * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$).

Fig. 2: spleen *Cxcr2*^{-/-} neutrophils have an impaired phagocytosis ability

A. To measure phagocytosis, BM CD11b⁺ Ly6G⁺ neutrophils were incubated with Red E. coli Phrodo bioparticles at 37°C for 0, 30 min, 1h or 2h and analyzed by flow cytometry. Upper panel: Gating strategy to identify phagocytic neutrophils (Phago⁺) in the CD11b⁺ Ly6G⁺ fraction at 2h time. Lower panel: Percentage of phagocytic Ly6G⁺ neutrophils. **B.** Same experiment with spleen neutrophils. Results are expressed as the percentage of phagocytic neutrophils in the CD11b⁺ Ly6G⁺ population and represent the mean $\pm$ SEM of at least 6 animals (Mann-Whitney test, NS: non-significant, * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$).

Fig. 3: spleen *Cxcr2*^{-/-} neutrophils display reduced ROS and superoxide levels

A. Gating strategy to measure cytoplasmic Ros levels using CellRox Orange in CD11b⁺ Ly6G⁺ neutrophils. **B.** ROS levels were quantified in spleen and BM CD11b⁺ Ly6G⁺ neutrophils using

CellRox Orange probe. Results are expressed as the percentage of ROS-positive neutrophils in the CD11b⁺ Ly6G⁺ population and represent the mean $\pm$ SEM of at least 6 animals (Mann-Whitney test, NS: non-significant, ** $p < 0.01$). **C.** Gating strategy to measure Mitochondrial superoxide levels using Mitosox in CD11b⁺ Ly6G⁺ neutrophils. **D.** Measure of the percentage of mitochondrial superoxide –positive CD11b⁺ Ly6G⁺ neutrophils using Mitosox reagent. Data represent the mean $\pm$ SEM of at least 6 animals (Mann-Whitney test, NS: non-significant, ** $p < 0.01$).

Fig. 4: Decrease of Actin⁺ and Tubulin⁺ levels in spleen *Cxcr2*^{-/-}

A. Gating strategy to measure F-Actin levels (Actin⁺) in CD11b⁺ Ly6G⁺ neutrophils. **B.** Percentage of CD11b⁺ Ly6G⁺ expressing Actin (mean $\pm$ SEM of at least 6 animals; Mann-Whitney test, NS: non-significant, ** $p < 0.01$). **C.** Gating strategy to measure α -Tubulin levels (Tubulin⁺) in CD11b⁺ Ly6G⁺ neutrophils. **D.** Percentage of CD11b⁺ Ly6G⁺ expressing Tubulin. (mean $\pm$ SEM of at least 6 animals; Mann-Whitney test, NS: non-significant, ** $p < 0.01$).

Fig. 5: spleen *Cxcr2*^{-/-} neutrophils exhibit a reduced apoptosis and mortality

A. Gating strategy to identify alive, dead and apoptotic cells based on Annexin V and PI staining. Control plot corresponds to cells labelled with cell surface markers, but not with Annexin V and PI. **B.** Measure of the percentage of apoptotic neutrophils in the spleen and BM of WT and *Cxcr2*^{-/-} animals by annexin V staining. **C.** Same measure of dead cells by PI staining. Results are expressed as the percentage of CD11b⁺ Ly6G⁺ neutrophils and represent the mean $\pm$ SEM of 6 animals; Mann-Whitney test, NS: non-significant, ** $p < 0.01$.

Fig. 6: *Cxcr2*^{-/-} neutrophils display decreased maturity at the transcriptomic level

A. Volcano plot showing the global changes in RNA expression patterns for spleen neutrophils isolated from *Cxcr2*^{-/-} (SP KO) versus WT (SP WT) animals. Data represent analysis of cpm

estimates with a log of fold change of more than 1.5 fold and $p < 0.05$ of 4 animals per group. Grey dots: NR: non-regulated genes; Green dots: genes with a log of fold change of more than 1.5 fold; blue dots: genes with a p -value < 0.05 ; red dots: genes with a log of fold change of more than 1.5 fold and $p < 0.05$. **B.** Number of differentially regulated genes for the spleen neutrophils. Up: genes up-regulated in spleen isolated from *Cxcr2*^{-/-} versus WT animals. Down: down-regulated genes. NR: non regulated genes. **C.** Simplified Heatmap of spleen *Cxcr2*^{-/-} versus WT neutrophils. **D.** Normalized enrichment score (NES) after GSEA analysis of the transcriptome of spleen neutrophils isolated from *Cxcr2*^{-/-} versus WT animals according to Neutrophil maturation of Xie et al. (36). **E.** Heatmap of the significantly regulated genes ($p < 0.05$) of Neutrophil maturation signature. **F.** Cluster of chemotaxis and migration GSEA analysis from *Cxcr2*^{-/-} versus WT animals according to Biologic process GO: Cell-chemotaxis (NES= -1.64; $q = 0.0002$), Leukocyte-chemotaxis (NES= -1.62; $q = 0.001$), Granulocyte chemotaxis (NES= -1.53; $q = 0.029$), Regulation of Chemotaxis (NES = -1.46; $q = 0.013$), Granulocyte migration (NES= -1.54; $q = 0.025$), Myeloid-leukocyte-migration (NES= -1.50; $q = 0.012$), leukocyte-migration (NES = -1.48; $q = 0.003$).

Fig. 7: ERK and p38 pathways are down-regulated in Spleen *Cxcr2*^{-/-} neutrophils.

A. NES after GSEA analysis of the transcriptome of spleen neutrophils isolated from *Cxcr2*^{-/-} versus WT animals according to ERK1 and ERK2 cascade signature. **B.** Same analysis for p38-MAPK signature. **C.** Heatmap of the significantly regulated genes ($p < 0.05$) of GO-BP ERK1 and ER2 cascade signature. **D.** Heatmap of the significantly regulated genes ($p < 0.05$) of GO-BP p38 MAPK cascade signature. **E.** Phospho-p42/44 MAPK ERK1/2 (Thr 202/ Tyr 204) was analyzed by flow cytometry in spleen CD11b⁺ Ly6G⁺ neutrophils. Left panel: representative Mean of Fluorescence (MFI) of WT (red line) and KO (blue line) Ly6G⁺ neutrophils. Results are expressed as MFI of phospho-ERK positive neutrophils in the CD11b⁺ Ly6G⁺ population and represent the mean $\pm$ SEM of at least 6

674 animals (Mann-Whitney test, * $p < 0.05$). **F.** Phosphorylation of p38MAPK on Thr180/Tyr182 was
675 analyzed by flow cytometry in the same conditions (Mann-Whitney test, *** $p < 0.001$).

676

677 **Fig. 8: Multiple pathways are down-regulated in Spleen *Cxcr2*^{-/-} neutrophils**

678 GSEA analysis of the following signatures: **A.** WIKI-PI3K-AKT signaling. **B.** Hallmark-TNF α
679 signaling via NF κ B. **C.** GO-BP NIK-NF κ B Signaling **D.** GO-BP Cellular response to IL-1 **E.** WP-
680 TGF β signaling pathway **F.** Hallmark IFN γ -response

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