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Interactions Between Severe Acute Respiratory Syndrome Coronavirus 2 Replication and Major Respiratory Viruses in Human Nasal Epithelium

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1 Major Article

2 **Interactions between Severe Acute Respiratory Syndrome Coronavirus 2**

3 **Replication and Major Respiratory Viruses in Human nasal Epithelium**

4

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Abstract

The emergence of severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2), along with extensive non-pharmacological interventions, have profoundly altered the epidemiology of major respiratory viruses. Some studies have described virus-virus interactions, particularly manifested by viral interference mechanisms at different scales. Still, our knowledge of the mutual interactions between SARS-CoV-2 and other respiratory viruses remains incomplete. Here, we studied the interactions between SARS-CoV-2 and several respiratory viruses (influenza, RSV, hMPV, and hRV) in a reconstituted human epithelial airway model, exploring different scenarios affecting the sequence and timing of co-infections. We show that the virus type and the sequence of infections are key parameters of virus-virus interactions, having the impact of primary infections on the regulation of the immune response a determinant role in the outcome of secondary infections.

Keywords

SARS-CoV-2; virus-virus interactions; influenza virus; Respiratory syncytial virus; Human Rhinovirus; Human Metapneumovirus

Running Title

SARS-CoV-2 respiratory viral coinfections

1 INTRODUCTION

2

3 The emergence of SARS-CoV-2 in late 2019 and the consequent COVID-19 pandemic had,
4 and still have, a massive impact on our societies. Beyond the more than 526 million
5 confirmed cases and over 6.28 million deaths [1], multivariate estimates point to over 40% of
6 the global population having been infected at least once before the recent rising of the SARS-
7 CoV-2 omicron variant [2]. The magnitude of the health crisis has somewhat overshadowed
8 the contribution of acute respiratory infections to global mortality, which are responsible for 3
9 to 5 million deaths per year in non-pandemic periods [3]. Several studies have shown that the
10 global circulation of SARS-CoV-2 has profoundly altered the epidemiological pattern of the
11 main respiratory viruses, such as influenza viruses and Respiratory Syncytial Virus (RSV) [4].
12 Early in 2020, many countries implemented non-pharmaceutical interventions (e.g., face
13 masks, social distancing, lockdown, school closures, or teleworking) and travel restrictions to
14 mitigate the transmission of SARS-CoV-2 and subsequently saw a rapid decline in influenza
15 infections, with influenza circulation in the northern hemisphere at an all-time low despite
16 higher numbers of specimens tested [4–7]. At the same time, a shift on RSV outbreaks,
17 occurring at unusual times of the year, has been observed [8–10]. Although other mechanisms
18 for the predominance of SARS-CoV-2 cannot be excluded (e.g.,
19 underestimation/detection/reporting of other respiratory viruses, more limited access to
20 treatment of other respiratory infections, competitive interference), health restrictions seem to
21 have played a major role, as suggested by recent reports showing significant increases in the
22 circulation of influenza and other respiratory viruses concomitantly with the ease of
23 restrictions [11,12].

24 There is relatively little information on the severity of COVID-19 in the context of co-
25 infection with other viruses, apart from a few studies involving a small number of patients,

1 some of which suggest a possible deleterious effect [13–16]. The simultaneous circulation of
2 different viruses can give rise to synergistic, neutral, or antagonistic interactions, ranging
3 from the local environment of the respiratory epithelial cell to a population epidemic
4 dynamics level [17]. Competitive interference has been described in the case of co-circulation
5 between respiratory viruses, notably involving influenza viruses. However, there is still no
6 clear consensus about the consequences of these co-infections in terms of disease severity
7 [18,19]. *In vitro* approaches have shown, for example, that RSV growth is blocked by
8 competitive infection with influenza A virus [20] or that primary viral infection with a
9 rhinovirus reduces replication of subsequent influenza infection [21,22]. The underlying
10 mechanisms are not yet fully understood and would involve viral competition for host
11 resources but also the induction of temporary non-specific host immunity [19,21,22].

12 Several teams have tried to address these questions concerning the potential interactions
13 between SARS-CoV-2 and other respiratory viruses, with relatively similar experimental
14 designs and observations. They have notably shown that primary infection with rhinovirus
15 prevents secondary infection with SARS-CoV-2 [23–26]. Fage and colleagues recently
16 showed that prior infection of human epithelial cells with SARS-CoV-2 interferes with the
17 replication kinetics of H1N1 influenza virus and RSV, while only H1N1 infection reduces
18 subsequent SARS-CoV-2 infection [27]. Cheemarla and collaborators have notably shown
19 that the activity of Interferon-Stimulated Genes-mediated response at the time of SARS-CoV-
20 2 exposure impacts infection progression [24]. Altogether, these studies suggest a prominent
21 role for the innate immune response in these virus-virus interactions, at least when co-
22 infections are very close in time. Nevertheless, our knowledge of the mutual interactions
23 between SARS-COV-2 and respiratory viruses remains incomplete.

24 In this study, we aimed to further explore the interactions between SARS-CoV-2 and several
25 respiratory viruses, such as influenza viruses, RSV, hMPV, or hRV using a reconstituted

1 human airway epithelium model (HAE). We studied and compared the mutual impacts on
2 viral replication, epithelial integrity, but also innate immune signatures in different infection
3 and superinfection scenarios. Our results suggest that the timing, sequence, and specific
4 nature of the induced immune response are key elements in the interactions between SARS-
5 CoV-2 and other viruses.

6 7 METHODS

8 9 *Viral strains*

10 All experiments involving clinical samples and the manipulation of infectious SARS-CoV-2
11 were performed in biosafety level 3 (BSL-3) facilities, using appropriate protocols. The
12 SARS-CoV-2 strain used in this study (BetaCoV/France/IDF0571/2020; GISAID accession
13 ID EPI_ISL_411218) is a Wuhan-Like B strain isolated from a nasal swab sample collected
14 from a one of the first COVID-19 cases confirmed in France [28]. Rhinovirus A16 (hRV-A16
15 ATCC VR-283) was a kind gift from Samuel Constant (Epithelix SARL). Influenza
16 A/Lyon/969/2009(H1N1), Respiratory syncytial virus (hRSV-A Long strain ATCC-VR26)
17 and human metapneumovirus (hMPV-B strain CAN97-82 (B1)) were previously described
18 [29]. All these viruses were produced and titrated in dedicated cell lines, and the replication
19 kinetics of each strain in the HAE model were previously performed [28,29].

20 21 *Viral infections in reconstituted human airway epithelia (HAE)*

22 Infections protocols in HAE were previously described [28,29]. Briefly, MucilAirTM HAE
23 reconstituted from human primary cells obtained from nasal biopsies, were provided by
24 Epithelix SARL (Geneva, Switzerland) and maintained in air-liquid interphase with specific

1 culture medium in Costar Transwell inserts (Corning, NY, USA) according to the
2 manufacturer's instructions. For infection experiments, apical poles were gently washed twice
3 with warm OptiMEM medium (Gibco, ThermoFisher Scientific) and then infected directly
4 with 150 μ l of calibrated viral suspension in OptiMEM medium, at a specific multiplicity of
5 infection (MOI). For mock infection, the same procedure was followed using OptiMEM as
6 inoculum. Transepithelial electrical resistance (TEER) was measured using a dedicated volt-
7 ohm meter (EVOM2, Epithelial Volt/Ohm Meter for TEER) and expressed as Ohm.cm^2 .

8

9 *Transmission electron microscopy*

10 Infected nasal HAE were fixed with 2% glutaraldehyde (EMS) in 0.1 M sodium cacodylate
11 (pH 7.4) buffer at room temperature for 30 min. After washing three times in 0.2 M sodium
12 cacodylate buffer, cell cultures were post-fixed with 2% osmium tetroxide (EMS) at room
13 temperature for 1 h and dehydrated in a graded series of ethanol at room temperature and
14 embedded in Epon. After polymerization, ultrathin sections (100 nm) were cut on a UCT
15 (Leica) ultramicrotome and collected on 200 mesh grids. Sections were stained with uranyl
16 acetate and lead citrate before observations on a Jeol 1400JEM (Tokyo, Japan) transmission
17 electron microscope, equipped with an Orius 600 camera and Digital Micrograph Software
18 (Gatan).

19

20 *Real-time quantitative PCR*

21 Relative quantification of viral genome was performed by one-step real-time quantitative
22 reverse transcriptase and polymerase chain reaction (RT-qPCR) from viral or total RNA
23 extracted using QiAmp viral RNA or RNeasy Mini Kit (Qiagen) in the case of
24 supernatants/apical washings or cell lysates, respectively. The relative quantification of the

1 viral genes was performed by RTqPCR or SYBR-Green using the StepOnePlus™ Real-Time
2 PCR System (Applied Biosystems) in 96-well plates. Each sample was analyzed in triplicate,
3 and the cycle threshold (Ct) values were normalized against the endogenous GAPDH
4 reference, when necessary. The primers and probes used are described in Table 1.

5

6 *Nanostring gene expression analysis*

7 The NanoString nCounter® technology, a hybridization-based multiplex assay characterized
8 by its amplification-free step, was used for mRNA detection of two customized panels:
9 “immune response” (96 genes) and “type III IFNs” (12 genes) [28]. Data treatment and
10 normalization were next performed using nSolver analysis software (version 4.0, NanoString
11 technologies). A first step of normalization using the internal positive controls allowed
12 correction of a potential source of variation associated with the technical platform. To
13 normalize for differences in RNA input we used the same method as in the positive control
14 normalization, except that geometric means were calculated over several housekeeping genes.
15 After normalization steps, the ratio between infected/co-infected condition and the mock
16 condition were calculated to compare gene expression. Results were expressed as fold change
17 induction. Heatmaps were generated using Morpheus (Broad Institute, Morpheus,
18 <https://software.broadinstitute.org/morpheus>).

19 RESULTS

20

21 *Simultaneous influenza virus and SARS-CoV-2 infections*

22 Based on protocols previously described [28,29], we initially performed single infections or
23 co-infection with H1N1 influenza virus (H1N1pdm09) and/or SARS-CoV-2 (B, Wuhan-like
24 strain) in a HAE model of nasal origin. Infections/co-infections were performed at a MOI of 1
25 and monitored over a 48-hour period (Figure 1A). As expected, single infections strongly

1 impact epithelial integrity at 48 hpi, with a 40-80% decrease in trans-epithelial resistance
2 (TEER ohms.cm²) values for H1N1 and SARS-CoV-2, respectively. Such deleterious impact
3 of co-infection on epithelium integrity was comparable to that observed in the case of single
4 infection with SARS-CoV-2 (Figure 1B). In terms of viral replication, we found a very
5 modest decrease in normalized intracellular SARS-CoV-2 genome levels between single
6 infection and co-infection (18%, nsp14/GAPDH, Figure 1C), whereas this difference was
7 much greater in the case of H1N1 (74%, M/GADPH, Figure 1D). Observation of the different
8 experimental conditions by electron microscopy (Figure 1E) shows characteristic signs of
9 infection with SARS-CoV-2 (Figure 1E, panel b) or H1N1 (Figure 1E, panel c). In the case of
10 co-infection, these same characteristics are found on neighboring cells (Figure 1E, panel D),
11 but rarely within the same cell. Altogether, our initial observations suggest a mutual
12 antagonism of the two viruses yet with differential amplitude in HAE model.

14 *Influenza and SARS-CoV-2 mutual superinfections*

15 We then performed mutual superinfection experiments in HAE, in which a primary viral
16 infection with one of the viruses (MOI 0.1) was followed by a secondary infection with the
17 other virus (MOI 0.1) 48h later. The final readout was fixed at 96hpi, namely 48h post
18 superinfection (Figure 2A). Under these conditions, single infection with influenza virus
19 induces a marked (>50%) drop in TEER values at 48hpi and to a lesser extent at 96hpi. Single
20 SARS-CoV-2 infection induced a comparable decrease at 48hpi though TEER values
21 remained low at 96hpi (Figure 2B). In the case of superinfections, TEER values followed the
22 same pattern observed for single infections with the primary virus, suggesting no or minor
23 supplementary impact of the viral superinfection on the integrity of the respiratory epithelium
24 at 96 hpi (Figure 2B). We then quantified for each condition the viral genome of both viruses
25 at the apical region of the HAE (Figure 2C) and also at the intracellular level (Figure 2D).

1 While SARS-CoV-2 superinfection 48h after a primary influenza infection does not
2 significantly alter influenza M gene copy levels either at the apical or intracellular level, the
3 opposite sequence of superinfection (i.e. H1N1 superinfection following primary SARS-CoV-
4 2 infection) appears to marginally reduce apical SARS-CoV-2 nsp14 gene copy levels (Figure
5 2C/D). These results are in line with previous TEER observations suggesting a minimal level
6 of interference during viral superinfection in HAE model.

7 To complete our investigations, we used the nanostring technology to study the expression of
8 a wide panel of genes involved in the host innate immune response at 96hpi (Figure 2E).
9 Interestingly, the comparison of the immune signatures of superinfections with those of the
10 corresponding single infections reflect quite different profiles for influenza virus or SARS-
11 CoV-2. Indeed, compared to the condition of simple influenza infection, we observe a
12 significant upregulation of a large proportion of the genes in the panel during superinfection
13 with SARS-CoV-2, with the notable exception of several genes (IL1A, IL1B, IL1R2,
14 FAM89A, PTGS2, IL7R, MIP2 α , IL18, and IL6) which are mainly involved in the
15 inflammatory response (orange box, Figure 2E). In contrast, several genes involved in the
16 early interferon-stimulated innate immune response (OAS1, OAS2, SOCS1, DDX58,
17 CXCL10) are upregulated following influenza superinfection when compared to
18 corresponding SARS-CoV-2 simple infection (green boxes, Figure 2E). Overall, these
19 observations suggest that the sequential order of superinfection is crucial for the differential
20 regulation of early host immune and inflammatory responses.

21
22 *SARS-CoV-2 delayed superinfection following an influenza, RSV, hMPV or hRV primo-*
23 *infection*

24 As the experimental co-infection and superinfection protocols described above do not
25 necessarily fully reflect the different interaction scenarios found in patients, we finally sought

1 to mimic a situation of delayed superinfection by SARS-CoV-2 in the context of a primary
2 infection by influenza but also by other major respiratory viruses, such as Respiratory
3 Syncytial Virus (RSV), Human Metapneumovirus (hMPV) or Human Rhinoviruses (hRV).
4 For this, we performed primary infections with H1N1 (MOI 0.01), RSV (MOI 0.1), hMPV
5 (MOI 1) and hRV (MOI 5) in HAE and followed by superinfection with SARS-CoV-2 (MOI
6 0.1) at 7dpi, with final readout 48h later (9dpi) (Figure 3A). As expected, primary influenza,
7 RSV and hMPV infections had a deleterious impact on epithelial integrity, with the greatest
8 decreases on TEER values observed for H1N1 and RSV at 7 and 9dpi. No significant effect of
9 Rhinovirus infection on TEER was observed (Figure 3B). Similar to our previous
10 observations, we did not observe any additional effect of SARS-CoV-2 superinfection on
11 epithelium integrity at 9dpi (Figure 3B). Furthermore, superinfection with SARS-CoV-2 had
12 no significant impact on the intracellular genome levels of influenza, hMPV and hRV, with
13 only a 2-fold reduction in the context of primary RSV infection (Figure 3C).
14 On the other hand, SARS-CoV-2 nsp14 gene copy levels at 9dpi (48h post SARS-CoV-2
15 infection or superinfection) were significantly lower in the context of primary influenza, RSV,
16 and hMPV viral infections than in the mock primo-infected condition, hence suggesting very
17 strong viral interference in HAE model. This effect was much less pronounced in the context
18 of hRV primoinfection (Figure 3D). These observations are in line with the immune
19 signatures determined by nanostring (Figure 4). We have plotted the deregulation of gene
20 expression during SARS-CoV-2 superinfection against the level of expression measured
21 during primary infection (Superinf. H1N1, hRSV, hMPV & hRV, Figure 4 & Supplementary
22 Figure 1). This heatmap visualization reflects quite well the very strong interference of
23 primary viral infection, despite a long infection time, and illustrates the similarities and
24 differences between each viral model. Indeed, there is a clear clustering between H1N1/hRSV
25 and hMPV/hRV SARS-CoV-2 superinfections, with the regulation of genes such as IL18,

1 IL1A, IRF3, JAK2 being particularly discriminating, with down- or up-regulation,
2 respectively (yellow box, Figure 4). Some gene clusters are deregulated in a way that is very
3 specific to the nature of the primary infection, for example the downregulation of ZBTB16,
4 TBP, EI2AK4, DECTIN-1, POU2F2, IL7R is very specific to the RSV+SARS-CoV-2
5 superinfection (green box, Figure 4). Similarly, the upregulation of a very large number of
6 genes, including CXCL10, OAS1/OAS2, IRAK2, STAT2, IFI44L, MX1, IFITM1 or IFI35,
7 seems characteristic of hRV+SARS-COV-2 superinfection (orange boxes, Figure 4) and
8 suggests a more pronounced level of interference than for the other viruses studied, under
9 these experimental conditions. Altogether, these observations show that the nature of the
10 primary infection is an important driver in the host response.

11

12 DISCUSSION

13

14 After two years of an ongoing pandemic and the gradual lifting of sanitary restrictions in
15 many countries, we now find ourselves in a rather unprecedented situation, with the co-
16 circulation of SARS-CoV-2 with other main respiratory viruses. This situation raises many
17 questions about the mechanisms of interaction between these different viruses, but also about
18 the consequences of these co-infections/superinfection and their clinical management. In this
19 study, we aimed to further explore the interactions between SARS-CoV-2 and several
20 respiratory viruses, such as influenza viruses, RSV, hMPV, or hRV using a reconstituted
21 HAE model. This model, consisting of human airway primary cells grown at the air/liquid
22 interface, is a truly relevant model for studying respiratory viruses and within-host
23 interactions. For example, our group has used this model several times to characterize and
24 compare different respiratory viruses such as influenza viruses, RSV, hMPV [29,34] or

1 SARS-CoV-2 [28], but also to experimentally simulate scenarios of interactions between
2 pathogens, including bacterial/fungal superinfections [35–37].
3
4 One of our first observations showed a reciprocal deleterious impact between influenza
5 viruses and SARS-CoV-2 in the context of simultaneous infection (Figure 1). Although this
6 experimental scheme might be uncommon in the context of natural infections, it allowed us to
7 show that simultaneous infection of the HAE model by viruses using distinct receptors and
8 entry pathways was possible, but also that the reciprocal impact between viruses was not
9 necessarily of the same order. Our results do indeed show a much greater impact of SARS-
10 CoV-2 on influenza than vice versa, but it is very difficult to know whether this reflects a
11 characteristic of SARS-CoV-2 or simply a difference due to a distinct infectivity or greater
12 replication. We then chose to explore models of superinfection, prioritizing sufficiently large
13 time gaps between primary and secondary infection to allow for multi-cycle replication and
14 significant host response to the infections. While many studies have examined the effect of a
15 pre-existing infection on the outcome of the second infection, in one direction only, it is
16 extremely instructive to be able to compare the mirror-image superinfection scenarios. In a
17 context of successive infections with a 48-hour delay, we showed that SARS-CoV-2 infection
18 is slightly reduced by superinfection with H1N1, which is not the case in the reverse situation
19 (Figure 2). In a context of successive infections with a much longer delay (7 days), we
20 showed that superinfection with SARS-CoV-2 had no significant impact on primary
21 infections, except for RSV. Under the same conditions, we also found that primary viral
22 infections induced an unfavorable state for SARS-CoV-2 replication, with significant
23 differences for some viruses – hRV being the less impactful (Figure 3). These results are in a
24 relatively good agreement with the work previously published by Essaidi-Laziozi &

1 colleagues [25] and only partially with those obtained by Fage & colleagues [27], which can
2 probably be explained by differences in protocols and experimental parameters.

3

4 The comparison of immune signatures between simple infections and superinfections allowed
5 us to highlight specificities linked to the sequential order of the infections. While a
6 superinfection with influenza virus in the context of a primary SARS-COV-2 infection
7 manifests itself by a rather characteristic antiviral signature (Figure 2E), the opposite scenario
8 results in an inflammatory signature which suggests a more severe context when SARS-CoV-
9 2 is the second pathogen. Although primary viral infections provide varying degrees of
10 control of secondary infection with SARS-CoV-2, the nature of the resulting immune
11 response may be distinct for different combinations of pathogens and may constitute a more
12 or less favorable terrain for disease progression. Our results in HAE model are in line with *in*
13 *vivo* studies that showed that greater severity was associated with SARS-CoV-2
14 superinfection. For example, Kim and colleagues showed that SARS-CoV-2/influenza virus
15 coinfections in K18-ACE2 mice caused an unbalanced immune response in the lungs and
16 peripheral blood, with more pronounced lung damage and longer secondary infections [38]. In
17 another model, (golden Syrian hamster), it was recently shown that simultaneous or sequential
18 co-infection with SARS-CoV-2 and H1N1 induced more severe disease than single infection
19 with either virus. Interestingly, prior infection with H1N1 virus reduced the pulmonary viral
20 load of SARS-CoV-2 but increased lung damage [39], in line with the enhanced inflammatory
21 expression profile observed in the context of H1N1+SARS-CoV-2 superinfections in our
22 study (Figure 2).

23

24 Although our results and those available in the literature indicate quite clearly the importance
25 of the IFN response in SARS-CoV-2/respiratory virus interactions, with differences specific

1 to each virus, it remains very difficult to really determine whether this is related to intra-
2 specific, virus-specific characteristics or rather to experimental biases. Indeed, as is the case
3 in many studies, particularly the study of *in vitro* interactions between pathogens, the
4 comparison between different viruses as different as influenza, RSV or even SARS-CoV-2, in
5 a given model, is very complex, as very large differences can exist notably in terms of
6 infectivity, delay of the induction of the IFN response, replication kinetics and within-host
7 interaction. Therefore, it is important to multiply the different possible scenarios of co-
8 infection, by playing with the parameters of infection, sequence, and timing of infection when
9 possible. Although the data generated in our experimental models are not the perfect proxy
10 for simulating clinical situations of co-infections, they are nevertheless a very good starting
11 point, complementary to *in vivo* studies, but also mathematical modeling approaches [18] to
12 improve our understanding of the underlying virus-virus interactions. In order to better
13 understand these interactions at the scale of the respiratory epithelium, it will also be
14 necessary to explore the various models set up with much more global approaches, in
15 particular by calling upon "omics" approaches (e.g., transcriptional profiling, metabolomics)
16 in order to have a more complete vision of within-host interactions, beyond the responses
17 involving the immune response.

18 In this situation of co-circulation of respiratory viruses and SARS-CoV-2, it is essential to
19 continue our efforts to better understand the interactions between these different viruses, at
20 different scales. This knowledge is essential to guide prophylactic and therapeutic approaches
21 in such clinical situations. From this point of view, the development of host-targeted antiviral
22 strategies with broad-spectrum efficacy could strengthen our arsenal to fight coinfections.

23

1 NOTES

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7

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12

13 *Potential conflicts of interests*

14 The authors declare no conflicts of interest in relation to this study

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7

8

ACCEPTED MANUSCRIPT

1 FIGURES

2

3 **Figure 1. Simultaneous infection with influenza virus and SARS-CoV-2.** **A.** Nasal HAE
4 were mock-infected or infected by influenza (H1N1) and/or SARS-CoV-2 at a MOI of 1 and
5 incubated for 48h (n=3). Viral loads of SARS-CoV-2 and H1N1 were determined by RT-
6 qPCR from total RNA extracted at 48hpi. **B.** Trans-Epithelial resistance (TEER ohms.cm²)
7 was measured for all experimental conditions. **C/D.** Intracellular viral genome relative
8 quantification by RTqPCR for SARS-CoV-2 (nsp14 gene) and influenza (M gene). **E.**
9 Remodeling of the HAE ultrastructure during infection and co-infection by H1N1 and SARS-
10 CoV-2. a) Regular ultrastructure of mock-infected nasal reconstituted human airway epithelia;
11 b-d) MucilAir™ HAE were infected on the apical surface with SARS-CoV-2 (b), or H1N1 (c)
12 or co-infected with both viruses (d). At 48 hpi, HAE were fixed and processed for
13 transmission electron microscopy analysis, as described in Materials and Methods. Section of
14 SARS-CoV-2 infected cells (*) showing numerous viral vesicles (DMVs) and H1N1-infected
15 cells (#) showing numerous M1-associated rod-like structures (arrowhead) accumulated
16 within the nucleoplasm. N: nucleus, Nu: nucleolus, V: viral particles.

17

18 **Figure 2. Influenza/SARS-CoV-2 mutual superinfections.** **A.** Nasal HAE were mock-
19 infected or infected by influenza (H1N1) or SARS-CoV-2 at a MOI of 0.1 and were
20 superinfected at 48hpi by the other virus (MOI 0.1) and incubated for another 48h. **B.** Trans-
21 Epithelial resistance (TEER ohms.cm²) was measured for all experimental conditions. **C.**
22 Apical or **D.** Intracellular viral genome relative quantification by RTqPCR for SARS-CoV-2
23 (nsp14 gene) and influenza (M gene). **E.** Differential expression of both immune response (96
24 genes) and type III IFNs (12 genes) panels were evaluated in infected nasal HAE using the
25 Nanostring technology. Heatmap and hierarchical clustering (one minus pearson correlation)

1 of differentially expressed genes compared to mock condition. Genes of interest in the text are
2 highlighted with orange and green boxes.

3 **Figure 3. SARS-CoV-2 superinfection in the context of influenza, RSV, hMPV, or hRV**
4 **prior infections.** A. Nasal HAE were mock-infected or infected by influenza (H1N1) or RSV,
5 hMPV or hRV at a MOI of 0.01, 0.1, 1 and 5, respectively and incubated for seven days
6 (T7dpi), and then superinfected by SARS-CoV-2 (MOI 0.1) for 2 additional days. B. Trans-
7 Epithelial resistance (TEER ohms.cm²) was measured for all experimental conditions. C.
8 Intracellular viral genome relative quantification by RTqPCR for influenza (M gene), RSV (F
9 gene), hMPV (N gene) or hRV (N gene) D. Intracellular viral genome relative quantification
10 by RTqPCR for SARS-CoV-2 (nsp14 gene).

11 **Figure 4. Immune response of SARS-CoV-2 superinfection in the context of influenza,**
12 **RSV, hMPV or hRV prior infections.** Differential expression of both immune response (96
13 genes) and type III IFNs (12 genes) panels were evaluated in infected nasal HAE using the
14 Nanostring technology. Heatmap and hierarchical clustering (one minus pearson correlation)
15 of differentially expressed genes in the context of superinfection compared to primary
16 infection. Genes of interest in the text are highlighted with yellow, orange and green boxes.

17
18 **Supplementary Figure 1. Immune response of SARS-CoV-2 superinfection and**
19 **influenza, RSV, hMPV or hRV primary infections.** Differential expression of both immune
20 response (96 genes) and type III IFNs (12 genes) panels were evaluated in infected nasal HAE
21 using the Nanostring technology. Heatmap and hierarchical clustering of differentially
22 expressed genes compared to mock condition.

23

24

Virus	Viral strain	Target	Primers/probes
SARS-CoV-2	BetaCoV/France/IDF0571/2020	Nsp14 [28]	Forward 5'-TGGGGYTTTACRGGTAACCT-3'
			Reverse 5'-AACRCGCTTAACAAAGCACTC-3'
			Probe 5'-FAM-TAGTTGTGATGCWATCATGACTAG-TAMRA-3'
Influenza	A/Lyon/969/2009(H1N1)	M [30]	Forward 5'-CTTCTAACCGAGGTCGAAACGTA-3'
			Reverse 5'-GGTGACAGGATTGGTCTTGTCTTTA-3'
			Probe 5'-TCAGGCCCCCTCAAAGCCGAG-3'
RSV	hRSV-A Long strain ATCC-VR26	F [31]	Forward 5'-CTGTGATAGARTTCCAACAAAAGAACA-3'
			Reverse 5'-AGTTACACCTGCATTAACACTAAATCC-3'
			Probe 5' FAM-CAGACTACTAGAGATTAC-3'
hMPV	hMPV-B strain CAN97-82 (B1)	N [32]	Forward 5'-AGAGTCTCAGTACACAATAAAAAGAGATGTGGG-3'
			Reverse 5'-CCTATTTCTGCAGCATATTTGTAATCAG-3'
hRV	hRV-A16 ATCC VR-283	5'UTR [33]	Forward 5' AGCCTGCGTGCCCKGCC 3'
			Reverse 5' GAAACACGGACACCCAAAGTAGT 3'
			Probe 5' FAM-CTCCGGCCCCTGAATGYGGCTAA-TAMRA 3'

1 **Table 1.** List of primer & probes used in this study

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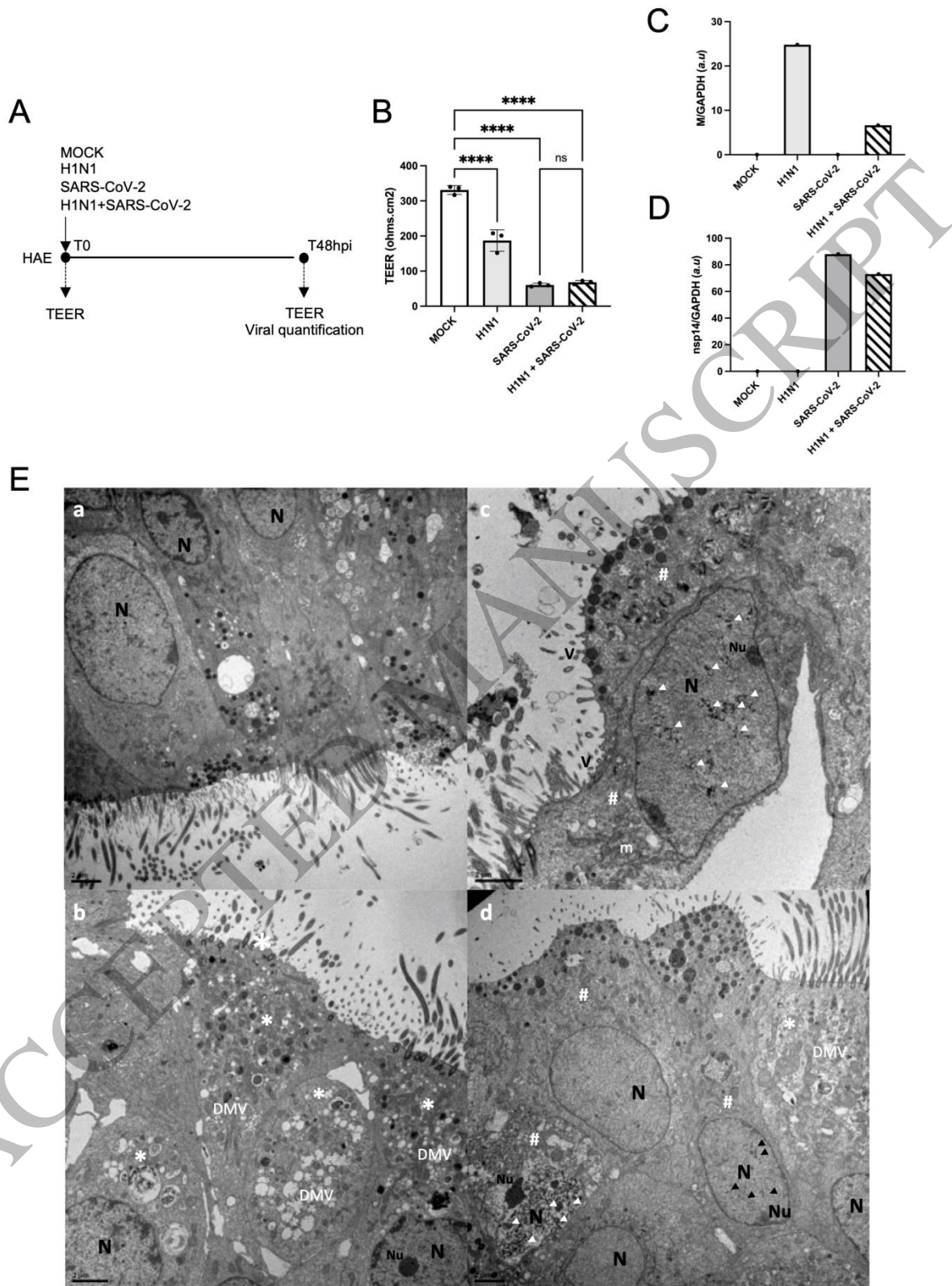


Figure 1

Figure 1
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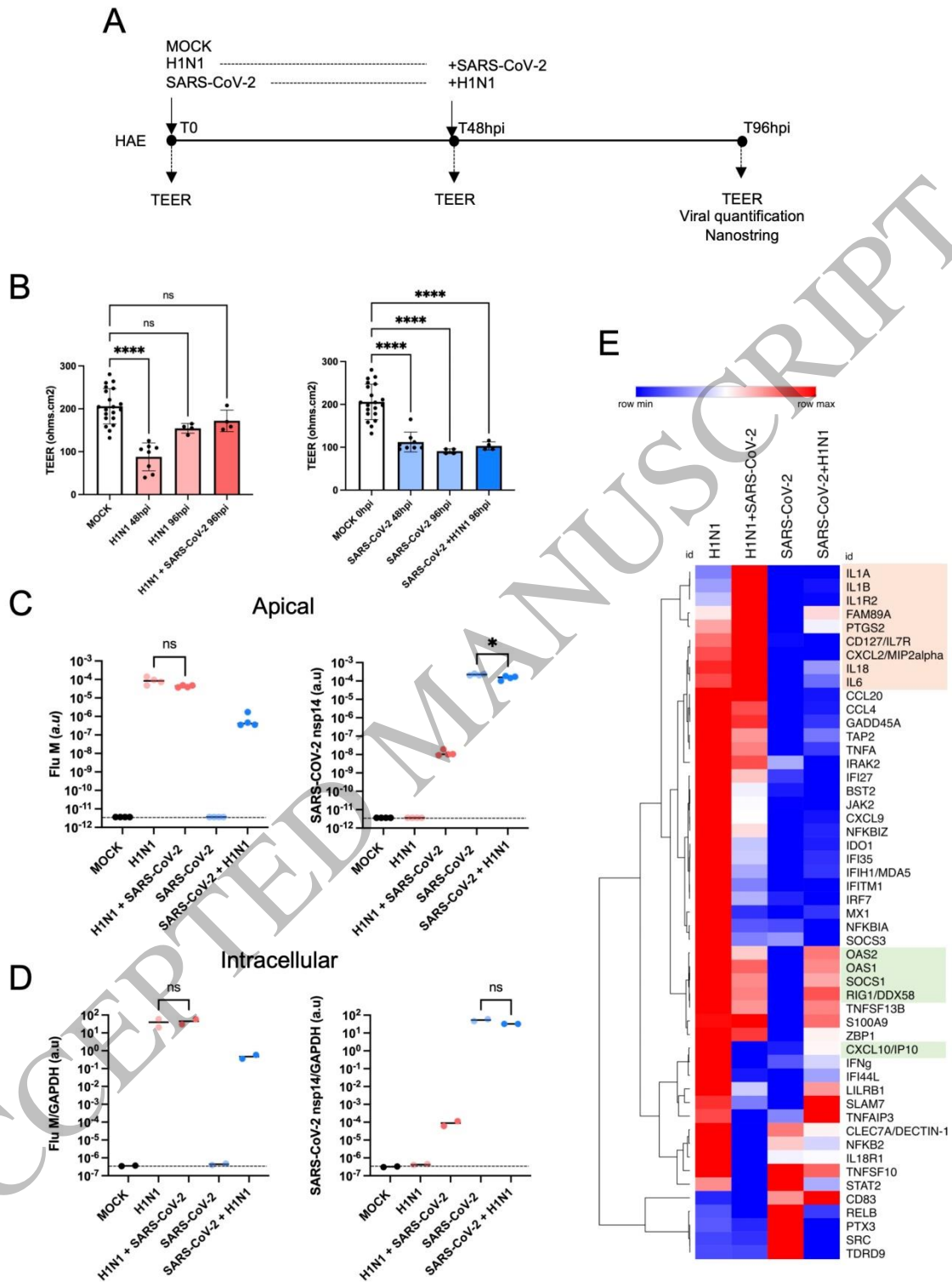


Figure 2

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Figure 2
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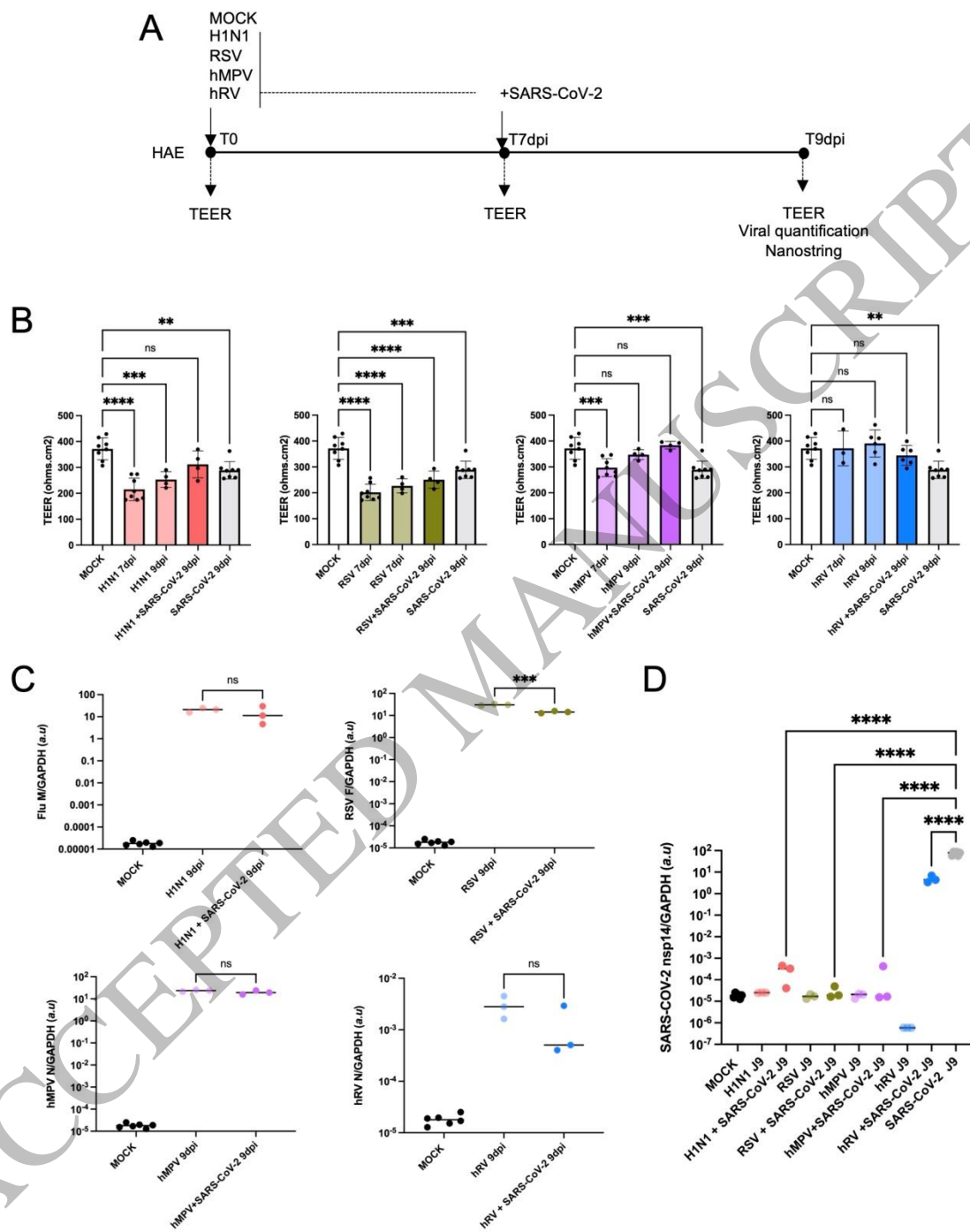


Figure 3

Figure 3
190x275 mm (79 x DPI)

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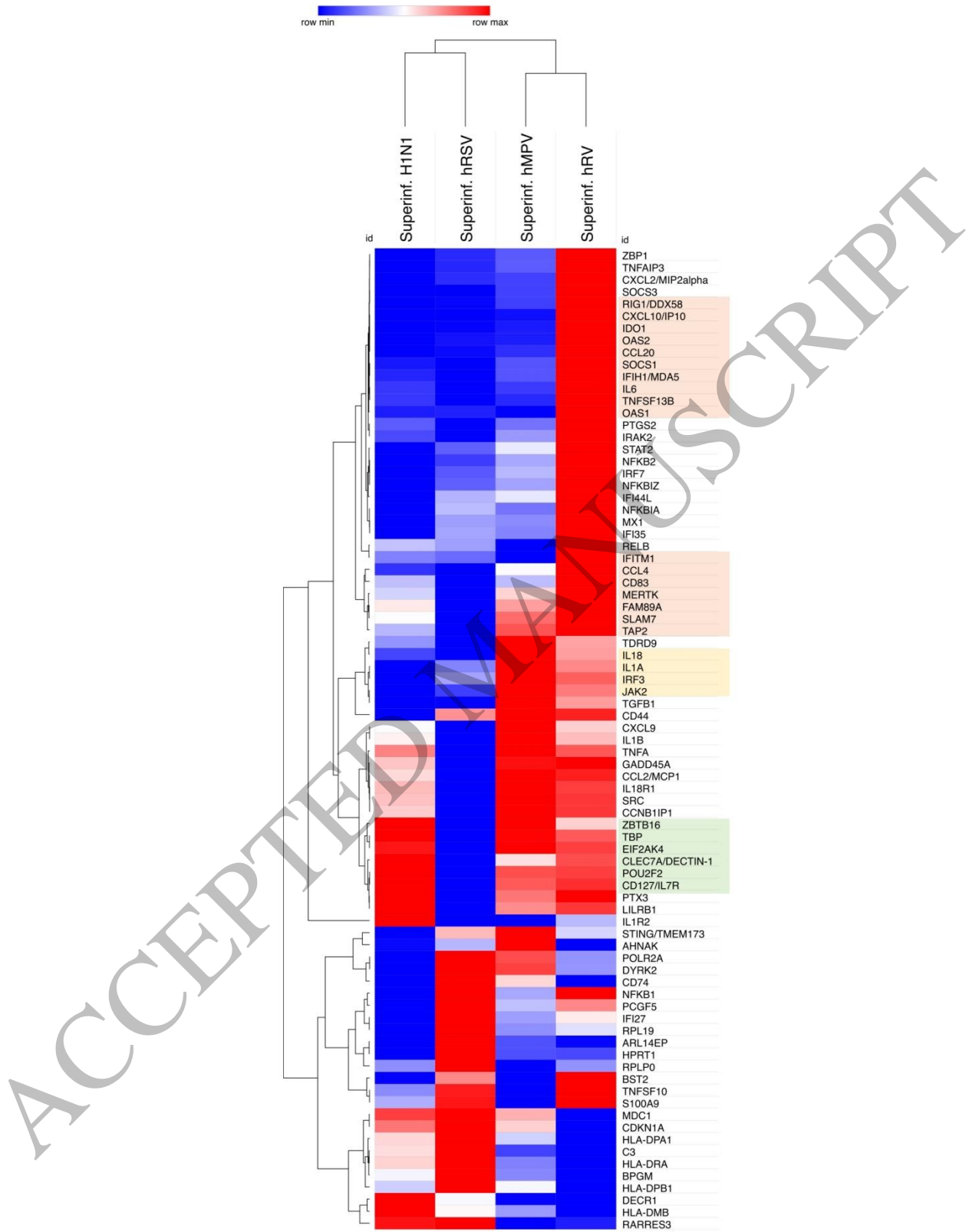


Figure 4

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Figure 4
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