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A one-pot synthesis and X-Ray structural characterization of

new highly substituted-allyl carbamates

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Research highlights

- One-pot synthesis of new variously substituted allylic carbamates starting from β -chloro- α . β -unsaturated aldehydes in high yields.
- Full characterization of the new compounds and the corresponding intermediates by NMR and IR spectroscopic techniques and HRMS.
- X-ray crystallography structural determination of two allyl carbamates.

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ABSTRACT

A one-pot synthesis of new variously substituted γ -chloro- γ , β -allylic carbamates have been carried out starting from β-chloro-α,β-unsaturated aldehydes. The new carbamates and the corresponding intermediates, the γ -chloro- γ , β -allylic alcohols, were fully characterized by (¹H and ¹³C) NMR, IR and HRMS spectroscopic techniques. The crystal structures of two allylic carbamates, were established by single-crystal X-ray diffraction.

Keywords: β-chlorovinylaldehydes; one-pot synthesis ; allylic carbamates; X-ray diffraction.

1. Introduction

Carbamates and allylic carbamates in particular, are of considerable interest due to their important role in various areas. Indeed, they belong to structural or functional part of many approved drugs [1-5] and prodrugs [6], which are used for the treatment of various diseases such as Alzheimer's disease [7]**,** cancer [8], hepatitis C [1, 9], HIV infection [10], anxiety [11] and epilepsy [11]. They are found to be important raw materials in the synthesis of polyurethanes [12-14], paints [15], wood preservatives [15], pesticides, herbicides, insecticides and fungicides [16]. They are also used as intermediates in numerous organic synthesis [17, 18] as protecting amino groups in peptide synthesis [19, 20], polymers synthesis [21] and as linkers in combinatorial chemistry [22] as well as in the cyclic [23] and heterocyclic compounds synthesis [24-27]. Therefore continuous efforts have been

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made to obtain this type of compounds, particularly the allylic carbamates, through simple and efficient methods. The conventional method for preparation is the reaction of an isocyanate with the corresponding allyl alcohol [28, 29] or through a *trans*-carbamoylation reaction of the hydroxyl group with the phenyl carbamate [30]. Coupling reaction of aryloxyallenes with $CO₂$ and amines using a silver-catalyzed is also reported [31] as well as the reaction of carbonyl compounds with N,N-dialkyl carbamoyl chloride [32] . However, the structural variation of allyl carbamates have shown serious limitations due to either the limited availability of various allyl alcohols bearing tri- or tetra-substituted double bond [28] or by the use of palladium complexes as catalysts strong bases [33] and strong oxidants [34] which are not suitable for allyl carbamates since they can promote unwanted cyclization reactions[35].

In our previous works, we have explored the reactivity of the β-chlorovinyl aldehydes as precursors of highly substituted β–fluoroalkoxyvinyl aldehydes and corresponding alcohols [36], benzenes [37], allylic alcohols [38], phosphoramidates [38], phosphonates and phosphates [39]. As an extension of our work, we report here the onepot synthesis of a new series of variously substituted allylic carbamates. The new compounds were obtained in mild conditions with good to excellent yields and were fully confirmed by $(^1H$ and $^{13}C)$ NMR, IR spectroscopies and HRMS. The structures of two allyl carbamates were determined by X-Ray crystallography.

2. Results and discussion

2.1. Synthesis

The starting substrates **1, 2** and **4-9** were synthesized from the corresponding α-methylene ketones using the Vilsmeier-Haack reaction according to literature procedures [40]. The synthesis of the new γ-chloro-β,γunsaturated alcohols (11, 12 and 14-18) was performed via carbonyl reduction of the β -chloro- α , β -unsaturated aldehydes. The chlorovinylaldehydes and cinnamaldehyde **(1-10)** reacted with sodium borohydride as reducing agent in dry THF and gave the products (**11-20)** in good yields (Table1).

Table1: Optimized synthesis of γ-chloro-β,γ-unsaturated alcoohols (11-20).

^a: *Z/E* isomer ratios measured from ¹H NMR aldehydic signals ^b yields measured after purification. °: : *Z/E* Isomer ratios measured in crude products from¹H-NMR CH₂OH signals.

Sodium borohydride reacted regioselectively with the vinylic aldehydes **1-10** in dry THF in good to excellent yields. The obtained pale yellow or colorless oils of allylic alcohols **11-20** are stable at room temperature with *Z/E* proportions identical to those of the starting aldehydes.

Different reaction conditions were attempted to optimize the synthesis of the target carbamates. When the reaction of N-phenyl isocyanate with the allylic alcohol **12** was carried out without any base in dry THF at 25°C, the desired product **27** was observed after 24 h in moderate conversion percentage (Table 2, entry 1). The use of Et₃N with slight excess gave total conversion (>99%) with excellent yield (entry 3). In CH₂Cl₂ or Et₂O, the reaction led to lower conversion percentages (entries 4 and 5). The various explored conditions are displayed in Table 2.

Entry	$Et3N$ (eq.)	Solvent	Time (h)	Conversion $\lceil % \rceil^a$
		THF	24	
		THF		96
	1.5	THF	n	>99
	1.5	DCM		74
		Et ₂ O		

Table 2: Optimization of the reaction conditions of the allylic carbamate **27** at 25°C

^a: The conversion % was monitored by ¹H NMR spectroscopy in crude products.

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With the optimal reaction conditions outlined above, the synthesized γ -chloro allylic alcoohls **11-20** were subjected to carbamoylation reaction with commercial N-alkyl and N-aryl-isocyanates in dry THF in mild basic medium at room temperature affording successfully new series of enol carbamates **20-45** in moderate to excellent yields (52-95%). Moreover, both carbonyl reduction and carbamoylation were succesfuly realized in one-pot procedure, starting from the aldehydes. The resulting variously substituted carbamates from the cyclic aldehydes **21-28** (Table 3), of the cinnamaldehydes **29-37** (Table 4) or from the acyclic α , β -unsaturated aldehydes (Table 5), were obtained as viscous oils or solids, soluble in both CHCl₃ and DMSO. All the reactions were monitored by TLC and ¹H-NMR. All final products were stable at 0°C and were characterized by IR, (¹H and ¹³C) NMR spectroscopies and HRMS.

Table 3: one-pot synthesis of carbamates from σ , β -unsaturated cyclic aldehydes (21-28)

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'Yan wasan ƙafa ta ƙasar Ingila. ^a: yields after purification.

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Table 4: One-pot synthesis of carbamates from cinnamaldehyde derivatives (**29-37**)

R_{4}		R٠ R,
	1) NaBH ₄ (0.25eq), dry THF, reflux, $(4h)$	
1eq	${}^{\mathcal{W}\!{\mathcal{R}}}_2$ 2) Et $_3$ N (1.5eq), R_3 NCO (1.3eq), rt, 12h	
$3 - 5$		29-37

Table 5: One-pot synthesis of carbamates from alkylated α **,** β **- unsaturated aldehydes (38-45)**

R_1	1) NaBH ₄ (0.25eq), dry THF, $T(^{\circ}C)/t(h)$		
	$R_2^{\mathcal{U}}$ CI 2) Et ₃ N (1.5eq), R ₃ NCO (1.3eq), rt, 15 h		
1eg		$38 - 45$	
6, 7 and 8			

^a: yields after purification ; ^b: *Z/E* Isomer percentages measured in crude products from ¹H NMR spectra.

Analysis of the ¹H-NMR spectra showed the formation of the target carbamates by the presence of signals assigned to both N=C*H* and CON*H* protons. However, for the carbamate **42**, these signals appear to be duplicated due to *cis* and *trans* conformers. In particular, the compound **36**, after a long term storing or recording the corresponding spectrum in DMSO as solvent instead of CDCl3, the additional signals of the second conformer are observed. In all spectra, The N*H*-Ph signals had moved substantially from chemical shifts 6.6-7.58 ppm in CDCl³ to 9.7-9.9 ppm in DMSO without significantly affecting the *cis* and *trans* ratios. This is obviously due to the favored formation of the H-bond between the carbamate and DMSO. This is in accordance with our previous work [41].

Table 6: (δ _{NH} ppm) *cis/trans*-amide conformers and ratios in CDCl3 and DMSO-d₆

A.

a: *cis/trans*-amide ratio measured at room temperature from ¹H-NMR NH signals integration in CDCl₃ and DMSO-d₆.

The same behavior was observed in ¹³C-NMR spectra of compounds **36** and **42**, which revealed duplication not only around the CH-N*H* bonds, but also in more remote parts of the molecules, such as the carbonyl and the substituents in γ and β positions of the carbamates. The ¹³C-NMR spectra of carbamates 23 and 27 in CDCl₃ show only one set of peaks corresponding to a single rotamer. The 2D-NOESY spectrum exhibited a cross correlation between NH and CH2O in compound **23** and **27** only indicating the *trans* conformation of OR group with respect of NH. However, no indication on the *trans* NH-CO conformation was collected from the spectrum.

On the other hand, it has been reported that, in IR spectroscopy of compounds bearing NH-CO group, the NHstretching vibration v_{NH} at 3200 cm⁻¹ is assigned to the *cis*-amide whereas the v_{NH} above 3200 cm⁻¹ is attributed to the *trans*-amide with in addition, a characteristic band at 1540 cm⁻¹ [42]. In most cases, the infrared spectra of the compounds **20-45** in pure state (Table 6) showed two v_{NH} stretching bands; a large vibrational band ranging from 3330 to 3350 cm⁻¹ related to bound NH and a thin vibrational band ranging from 3420 to 3460 cm⁻¹ related to free NH. Furthermore, the presence of a thin and intense bending band at 1513-1590 cm−1 indicate reliably the *trans* conformation of NH-CO. All the spectra show also intense bands related to the carbonyl C=O at about 1620–1700 cm⁻¹ and to the double bond C=C at 1625–1685 cm⁻¹. All the characteristic bands are summarized in Table 7.

Table 7: IR data of allylic carbamates **21-45** in cm-1

Crystallographic data collection, structure determination and refinement

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The proposed structures of allylic carbamates **24** and **36** were confirmed by single crystal X-ray diffraction (SCXRD) analysis. The suitable crystals for both compounds were prepared from their solution in cyclohexane/diethylether (1:4) by slow evaporation of the solvent. The corresponding intensities datasets were measured at room temperature using a RIGAKU *XtaLabPro* diffractometer equipped with a Mo microfocus sealed tube *MM003* generator coupled to a double-bounce confocal Max-Flux® multilayer optic and a HPAD *PILATUS3R 200K* detector. *CrysAlisPro 1.171.39.46* [43] was employed for the data processing, with *SCALE3 ABSPACK* scaling algorithm implemented for the empirical absorption correction using spherical harmonics. The structures were readily solved by intrinsic phasing methods (SHELX-T program) [44]: one molecule of **24** was placed in the asymmetric unit (asu) of the orthorhombic unit cell, space group Pna2₁ and four independent conformers of **36** in that of the monoclinic unit cell, space group P21. All the non-hydrogen atoms were refined with anisotropic displacement parameters by full-matrix least-squares methods on $F²$ using SHELX-L. [45] The crystal studied of **36** turned out to be an inversion twin whose component mass ratio refined to *ca* 0.52:0.48. H atoms could be identified in difference maps and therefore the methyl H atoms borne by C11 were allowed as rigid groups to rotate but not tip, so as to match the electron-density maxima. The other H atoms were positioned geometrically and refined with U_{iso} set to $xU_{eq}(C)$, $x=1.2$ whereas $x = 1.5$ for the methyl carbons. The positions of the hydrogen atoms in the amide groups were refined with N-H distance restrained to target 0.87(2) Å in **24** or to be similar in **36**, with Uiso set to 1.2Ueq(N) (only for **36**, Uiso being freely refined in **24**). Figure 1 shows the molecular structure of **24** whose correct orientation relative to the polar axis direction from the crystal picked up for data collection was established by means of the Flack *x* parameter; [46] $x = -0.05$ (4), calculated [46] using

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1334 quotients of the type $[(I^+)-(I^-)]/[(I^+)+(I^-)]$ and one copy of the highly flexible E-isomer molecular structure of **36**. The conformational differences between the four copies are highlighted in an overlay diagram shown in Figure 2. In its asu, these molecules are elongated along the [010] direction and those relatively oriented in parallel way are linked by N–H … O hydrogen bonds [graph set *C* 1 ¹(4) commonly found in structures made of simple amides, as well in the **24** one] developing along the [0 0 1] direction molecular chains (Figure 3), stacking on top of each other in antiparallel way in the *a* direction and segregated at the x, $\frac{1}{4}$, z and x, $\frac{3}{4}$, z interfaces of 2₁ symmetry-related (chloromethyl)benzene moieties through edge-to-face and Cl...Cl contacts $(d_{Cl-Cl} = 3.79Å ≥ van der Waals radii sum). In 24, the N–H ... O chains propagate infinitely along the [1 0 0]$ direction developing paddlewheel structure motifs centered around x, 0.25, 0 and x, 0.75, 0.5 positions, involving molecules related by the glide plane perpendicular to $[0 1 0 1 0]$ with glide component 0.5, 0, 0 (Figure 4). No halogen contacts are noticed in the **24** crystal packing. Crystal data, data collection and structure refinement details are summarized in Table 8. CCDC 2084634 (**24**) - 2084224 (**36**) contain the supplementary crystallographic data for this paper. These data can be obtained free of charge from the Cambridge Crystallographic Data Centre via www.ccdc.cam.ac.uk/data_request/cif.

Compound	24	36
	CI. O Ĥ	Ò CI.
	(2-chlorocyclohex-1-en-1-	(E) -3-chloro-2-methyl-3-
	yl)methyl(3-chlorophenyl)carbamate	phenylallyl phenylcarbamate
Empirical formula	$C_{14} H_{15} Cl_2 N O_2$	C_{17} H_{16} Cl N O_2
Formula weight	300.17	301.76
Temperature (K)	293(2)	293(2)
Wavelength (Å)	0.71073	0.71073
Crystal system,	Orthorhombic,	Monoclinic,
space group	P na 21	P_1
Unit cell dimensions $ (\AA)$	9.9172(7) 10.2917(8) 14.0882(10) 90	9.2313(5) 34.2963(12) 10.0818(5) 90
\vert (°)	90 90	105.979(6) 90
Volume (\AA^3)	1437.91(18)	3068.5(3)
Z.	4,	8,
Calculated density $(Mg/m3)$	1.387	1.306
Absorption coefficient (mm ⁻ $\mathbf{1}$	0.448	0.252
F(000)	1264	1264
Crystal size (mm)	$0.44 \times 0.20 \times 0.17$	0.45 x 0.23 x 0.05
θ range for data collection (°)	4.063 to 27.102	3.639 to 25.350
Limiting indices	$-12 \le h \le 12$, $-13 \le k \le 13$, $-18 \le l \le 18$	$-10 \le h \le 11$, $-41 \le k \le 40$, $-11 \le l \le 12$
Reflections collected unique	14153 / 3127 0.0749	22490 / 10718 0.0434

Table 8: Crystal data and structure refinement for compounds **24** and **36**

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Figure 1 Ortep view of **24** (left), and one **36** conformer (right) in their asu with their respective atom numbering scheme. Displacement ellipsoids are drawn at the 50% probability level. The H atoms are shown as small spheres of arbitrary radii.

Figure 2 Overlay of the four independent molecules of **36**.

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Figure 3. Two independent hydrogen-bonded (N-H…O) chains (underscored by different colors applied to carbon atoms) of molecules in the central part of the unit cell of 36 running along the *c* axis and framed by $2₁$ symmetry-related molecules. Cyan dashed lines indicate N-H…O intermolecular hydrogen bonds (see table 9).

Figure 4. One hydrogen-bonded (N-H…O) chain of the molecules in the unit cell of **24** running along the *a* axis. Cyan dashed lines indicate N-H…O intermolecular hydrogen bonds (see table 9).

 Symmetry transformations used to generate equivalent atoms: (**24**) #1 x+1/2,-y+3/2,z ; (**36**) #2 x,y,z-1 #3 x,y,z+1

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3. Conclusion

In summary, new series of $γ$ -chloro-γ,β-unsaturated alcohols and corresponding enol carbamates have been synthesized from β -chloro- α,β -unsaturated aldehydes using a facile and efficient method in most cases. The major advantage of this transformation is the non-use of metal catalysts or toxic reagents. The desired compounds were obtained in high yields and characterized by different spectroscopic techniques and X-Ray crystallography.

Experimental Section Instruments and chemicals

All solvents were dried, distilled and stored over molecular sieves (4 Å) using standard techniques. [47] The starting aldehydes were prepared using Vilsmeir–Haack reaction. [40] TLC was performed on silica gel, 60F-250 (0.26 mm thickness) plates. The plates were visualized using a UV lamp light (254 nm) or by use of potassium permanganate and $5 \text{ g } K_2 \text{CO}_3/100 \text{ mL}$ water. NMR spectra were performed on Bruker AVANCE 300 MHz spectrometer at the Faculty of Pharmacy at University Paris-Saclay in France or an Ultrafield AVANCE 300 MHz spectrometer at High Institute of Environmental Sciences and Technology at University of Carthage in Tunisia. All NMR spectra were recorded at ambient temperature (290K) unless otherwise noted. ¹H NMR spectra are reported as follows: chemical shift (multiplicity, coupling constant, integration). The following abbreviations are used to indicate multiplicities: s, singlet; d, doublet; t, triplet; q, quartet; m, multiplet. Chemical shifts (δ) were given in parts per million (ppm) from tetramethylsilane with the solvent resonance as the internal standard in CDCl₃ solution. High resolution masse spectra were obtained using a Bruker Esquire electrospray ionization apparatus at the Faculty of Pharmacy at University Paris-Saclay in France.

FTIR spectra were recorded on Nicolet IR200 spectrometer within the wave number range of 4000-400cm−1 at 25°C at High Institute of Environmental Sciences and Technology at University of Carthage in Tunisia.

Unless otherwise noted, commercially available NaBH4, isocyanates and triethylamine were purchased from various commercial sources (Acros, Aldrich) and used without further purification.

General Procedure for preparation of γ-chloro-γ,β-allylic alcohols (11-19).

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To a stirred solution of β -chloro- α, β -vinylic aldehyde (1 mmol) in dry THF (10 mL) was added NaBH₄ (0.25 mmol) at 0° C. The reaction mixture was stirred at adequate temperature. The progress of the reaction was monitored by TLC until the conversion of β-chloro-α,β- unsaturated aldehydes was complete. Then, cold distilled water (100 mL) was added to the mixture. The product was extracted with diethyl ether (3*30mL) and washed by 10 mL of a saturated solution of NaCl. The solvent (diethyl ether) was evaporated, and the crude product was purified by short-column chromatography using (cyclohexane:diethylether) (8:2) as eluent. The same procedure will be used for the reduction of cinnamaldehyde.

(2-chlorocyclohex-1-en-1-yl) methanol (11)

Light Yellow oil, **FTIR: neat (cm-1)**: 3317 (OH), 1663 (C=C); **¹H NMR (300 MHz, CDCl3) δ:** 4.16 (s, 2H, CH₂OH), 3.37 (s, 1H, O<u>H), 2.27 (m, 2H, CH₂cyc), 2.18-2.16 (m, 2H, CH₂cyc), 1.66-1.56 (m, 4H, CH₂CH₂cyc);</u> **¹³C NMR (75 MHz, CDCl3) δ:** 132.52 (C-Cl), 128.35 (C=CCH2OH), 62.68 (CH2OH), 33.81 (CH2cyc), 27.78 (CH₂cyc), 23.79 (CH₂cyc), 22.13 (CH₂cyc).

(2-chlorocyclohept-1-en-1-yl) methanol (12)

Light Yellow oil, **FTIR: neat (cm-1)**: 3307 (OH), 1651 (C=C); **¹H NMR (300 MHz, CDCl3) δ:** 4.21 (s, 2H, CH₂OH), 2.58-2.55 (m, 2H, CH₂cyc), 2.50 (s, 1H, OH), 2.34-2.30 (m, 2H, CH₂cyc), 1.73-1.48 (m, 6H, **CH₂CH₂CH₂cyc**); ¹³**C** NM**R** (75 MHz, CDCl₃) δ: 137.51 (<u>C</u>-Cl), 132.56 (<u>C</u>=CCl), 64.75 (CH₂OH), 38.87 (CH_{2cyc}) , 31.24 (CH_{2cyc}), 30.37 (CH_{2cyc}), 26.10 (CH_{2cyc}), 25.23 (CH_{2cyc}).

3-Phenylprop-2-en-1-ol (13)

Colorless oil **, FTIR: neat (cm-1)**: 3308 (OH), 1657 (C=C); **¹H NMR (300 MHz, CDCl3) δ** : 7.42-7.25 (m, 5H, H_{arm}), 6.64 (d, ³J_{H-H}= 15.9Hz, 1H, C<u>H</u>Ph=CH), 6.39 (dt, ³J_{H-H}= 15.9Hz, ³J_{H-H}= 5.66Hz, 1H, CHPh=CH), 4.33 (m, 2H, CH2O), 2.28 (t, ³ JH-H= 5.59Hz, 1H,OH); **¹³C NMR (75 MHz, CDCl3) δ :** 136.76 (C=CPh) 131.04- 127.66 (C_{arm}), 126.48 (HC=CPh), 63.56 (CH₂OH).

3-chloro-3-phenylprop-2-en-1-ol (14)

Light Yellow oil, **FTIR: neat (cm-1)**: 3307 (OH), 1636 (C=C) ; **¹H NMR (300 MHz, CDCl3) δ :** 7.63-7.35 (m, 5H , H_{arm}), 6.37 (Z, t, ³J_{H-H}= 5.90Hz, 1H, C<u>H</u>), 6.22 (E, t, ³J_{H-H}= 7.25Hz, 1H, C<u>H</u>), 4.54 (Z, d, ³J_{H-H}= 5.90Hz, 2H, CH2OH), 4.16 (*E*, d, ³ JH-H= 7.23Hz, 2H, CH2OH), 3.45 (*Z+E*, s, 1H, CH2OH); **¹³C NMR (75 MHz, CDCl3) δ:** 137.26 (*Z*, C-Cl), 136.33(*E*, C-Cl), 133.70-126.44 (Carm), 128.92 (*Z*, HC=CCl), 128.68 *(E*, HC=CCl), 60.50 (*Z*, CH2OH), 59.76 (*E,* CH2OH).

3-chloro-2-methyl-3-phenylprop-2-en-1-ol (15)

Light Yellow oil, **FTIR: neat (cm-1)**: 3317 (OH), 1644 (C=C); **¹H NMR (300 MHz, CDCl3) δ**: 7.30 (m, 5H, Harm), 4.40 (*Z*, s, 2H, CH2OH), 3.99 (*E*, s, 2H, CH2OH), 2.04 (*E*, s, 3H, CH3), 1.79 (*Z*, s,3H, CH3); **¹³C NMR (75 MHz, CDCl3) δ:** 138.24 (*E*, C-Cl), 133.47-128.30 (Carm), 128.54 (*E*, C=C-CH3), 63.73 (*E*, CH2OH), 22.72 (*Z*, CH3), 17.93 (*E*, CH3).

3-chloro-4,4-dimethylpent-2-en-1-ol (16)

Light Yellow oil, **FTIR: neat (cm-1)**: 3307 (OH), 2969-2872 (CH³ and CH2), 1649 (C=C); **¹H NMR (200 MHz, CDCl₃) δ:** 5.65 (t, ³J_{H-H}= 5.80Hz, 1H, C<u>H</u>), 4.21 (d, ³J_{H-H}= 5.80Hz, 2H, C<u>H</u>₂OH), 3.89 (s, 1H, O<u>H</u>), 1.11 (s, 3H, 'Bu); ¹³**C NMR (75 MHz, CDCl3) δ:** 145.62 (<u>C</u>-Cl), 121.99 (<u>C</u>=CCl), 60.03 (<u>C</u>H₂OH), 38.40 (C(CH₃)₂), 28.59 (C(CH₃)₂).

Ethyl 2-chloro-4-hydroxybut-2-enoate (17)

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Light Yellow oil; ¹H NMR (300 MHz, CDCl₃) δ : 7.19 (t, 1H, CH, ³J_{H-H}= 5.45Hz), 4.50 (d, 2H, CH₂, ³J_{H-H}= 5.45Hz), 4.30 (q, 2H, CH₂CH₃, ³J_{H-H}= 7Hz), 2.11 (s, 1H, OH), 1.34 (t, 3H, CH3, ³J_{H-H}= 7Hz)**; ¹³C NMR (75 MHz, CDCl₃) δ:** 161.94 (C=O), 140.84 (C-Cl), 124.10 (C=CH), 62.55 (CH₃CH₂O), 60.30 (CH₂OH), 14.09 $(CH₃CH₂O).$

3-chloro-2-methylpent-2-en-1-ol (18)

Yellow oil, **FTIR: neat (cm-1)**: 3408 (OH), 1741(C=C) cm-1 ; **¹H NMR (300 MHz, CDCl3)δ :** 4.22 (*Z*, s, 2H, CH₂OH), 4.09 (*E*, s, 2H, CH₂OH), 2.92 (Z+*E*, s, 1H, O<u>H</u>), 2.43 (*E*, q, ³J_{H-H}= 7.38Hz, 2H, CH₂CH₃), 2.37 (Z, q, ³ JH-H= 7.44Hz, 2H,CH2CH3), 1.87 (*E*, s, 3H, CH3), 1.79 (*Z*, s, 3H, CH3), 1.11-1.05 (*Z+E*, t, 3H, CH2CH3); **¹³C NMR (75 MHz, CDCl3) δ:** 135.47 (*E*, CCl), 132.15 (*Z*, CCl), 129.82 (*E*, CH3C=CCl), 129.38 (*E*, CH3C=CCl), 64.06 (*Z*, CH2OH), 62.44 (*E*, CH2OH), 28.94 (*Z*, CH2CH3), 28.58 (*E*, CH2CH3), 17.85 (*E*, CH3), 15.85 (*Z*, CH3), 13.13 (*E*, CH₂CH₃), 12.01 (*Z*, CH₂CH₃).

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3-chloro-2-phenylbut-2-en-1-ol (19)

Yellow oil, **FTIR: neat (cm-1)**: 3326 (OH), 1651(C=C) cm-1 ; **¹H NMR (300 MHz, CDCl3) δ:** 7.40-7.23 (m, 5H, Harm), 4.57 (*Z*, s, 2H, CH2OH), 4.42 (*E*, s, 2H, CH2OH), 2.37 (*E*, s, 3H, CH3), 2.08 (*Z*, s, 3H, CH3), 2.08 (s, 1H, OH); **¹³C NMR (75 MHz, CDCl3) δ:** 139.28 (*E*, C-Cl), 138.47 (*Z*, C-Cl), 136.65-128.32 (Carm), 127.58 (*Z*, CCH3), 127.45 (*E,* CCH3), 64.39 (*Z*, CH2OH), 63.30 (*E,* CH2OH), 23.99 (*E***,** CH3), 22.88 (*Z,* CH3).

General Procedure for preparation of *γ*−chloro-*γ*,β-allylic alcohols (20).

0.25 mmol of NaBH4 was added to the ethyl 3-chloro-4,4,4-trifluoro-2-formylbut-2-enoate aldehyde (1mmol) stirred at -10°C in absolute ethanol. The progress of the reaction was monitored by TLC (by dissolving the sample in ether) using cyclohexane : diethylether (8:2) until the conversion of the aldehyde was complete. Then, water (100 mL) was added to the mixture. The product was extracted with diethyl ether (3x30mL) and washed 2 times with 10 mL of a saturated solution of NaCl. The solvent (diethylether) was evaporated, and the crude product was purified by column chromatography using (cyclohexane:diethylether) as eluent (8:2).

Ethyl 3-chloro-4,4,4-trifluoro-2-(hydroxymethyl)but-2-enoate (20)

coloress oil, **FTIR: neat** (cm⁻¹): \vee 3442 (OH), 11727(C=O) cm⁻¹; 1652(C=C) cm⁻¹; ¹H NMR (300 MHz, **CDCl3) δ :** 4.59 (*Z+E*, s, 2H, CH2O), 4.42-4.29 (*Z+E*, m, 2H, CH2CH3), 2.68 (*Z*, s, 1H, OH), 2.25 (*E*, s, 1H, OH), 1.40-1.32 (**Z+E,** m, 3H, CH2CH3); **¹³C NMR (75 MHz, CDCl3) δ :** 165.01 (*E,* CO2Et), 164.44 (*Z,* CO2Et), 139.73 (Z, CCl), 132.69 (E, C=CCl), 119.46 (Z+E, q, CF₃, ³J_{C-F}=271.68Hz), 62.71 (Z, CH₂CH₃), 62.51(E,

CH2CH3), 61.27 (*Z*, CH2O), 59.05 (*E,* CH2O), 13.92 (*E*, CH2CH3), 13.72 *(Z*, CH2CH3); **¹⁹F NMR (188 MHz, CDCl3**) δ : -61.55 (*E*, s, CF₃), -65.03 (*Z*, s, CF₃).

General Procedure for preparation of allylic carbamates (21-45).

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To a stirred solution of 3-chloro allyl alcohols (1 mmol) in dry THF (10 mL) with triethyle amine (1.5mmol) was added commercial isocyanates (1.3 mmol) at 0°C. The reaction mixture was stirred at room temperature for 2−12 h. the progress of the reaction was monitored by TLC. After, the reaction mixture was evaporated and the crude product was purified by column chromatography on silica gel using cyclohexane: diethyl ether (8:2) as the eluent to give the allylic carbamates products.

(2-chlorocyclohex-1-en-1-yl) methylpropylcarbamate (21)

FTIR: neat (cm⁻¹): v 3346 (NH), 1711 (C=O), 1695 (C=C), 1043 (C-O), 734 (C-Cl) cm⁻¹; ¹**H NMR (300 MHz, CDCl₃**</sub> δ 5.05 (s, 1H, N<u>H)</u>, 4.65 (s, 2H, CH₂O), 3.08-3.02 (m, 2H, NHCH₂CH₂CH₂), 2.27 (m, 2H, CH₂cyc), 2.07 (m, 2H, CH₂cyc), 1.67-1.50 (m, 4H, CH₂CH₂cyc), 1.48-1.41 (m, 2H, NHCH₂CH₂CH₃), 0.88-0.81 (*Cis+Trans*, m, 3H, NHCH2CH2CH3); **¹³C NMR (75 MHz, CDCl3) δ:** 156.59 (C=O), 130.17 (C=CCl), 128.99 (C=CCl), 64.46 (CH2O), 44.29/42.71 (*cis/trans*, NHCH2CH2CH3), 33.79 (CH2cyc), 27.68 (CH2cyc), 23.56 (CH2cyc), 23.08/21.00 (*trans/cis*, NHCH2CH2CH3), 21.90 (CH2cyc), 11.06/10.93 (*trans/cis*, NHCH2CH2CH3); **HRMS** (ESI), m/z: calcd for $[C_{11}H_{18}CINO_2Na]^+$ 254.0924 $[M + Na]$ +; found 254.0926.

(2-chlorocyclohex-1-en-1-yl)methylbutylcarbamate (22)

FTIR: neat (cm⁻¹): v 3452 (NH), 1712 (C=O), 1515 (C=C), 1137 (C-O), 667 (C-Cl) cm⁻¹; ¹**H NMR (300 MHz, CDCl3**) δ **:** 5.11 (s, 1H, NH), 4.63 (s, 2H, CH₂O), 3.06 (*Cis+Trans*, q, ³J_{H-H}= 6.41Hz, 2H, NHCH₂CH₂CH₂CH₂CH₃), 2.24 (m, 2H, CH₂cyc), 2.04 (m, 2H, CH₂cyc), 1.62-1.53 (m, 4H, CH₂CH₂cyc), 1.43-1.12 (*Cis+Trans*, q, ³J_{H-H}= 6.33Hz, 4H, NHCH2CH2CH2CH3), 0.81 (*Cis+trans*, t, ³ JH-H= 7.30Hz, 3H, NHCH2CH2CH2CH3); **¹³C NMR (75 MHz, CDCl₃**)</sub> **δ :** 156.57 (C=O), 130.04 (C=CCl), 129.00 (C=CCl), 64.40 (CH₂O), 40.67 (NHCH₂CH₂CH₂CH₃), 33.76 (CH2cyc), 31.93/29.52 (*trans/cis*, NHCH2CH2CH2CH3), 27.63 (CH2cyc), 23.55 (CH2cyc), 21.88 (CH2cyc), 20.75/19.76 (*cis/trans*, NHCH2CH2CH2CH3), 14.01/13.55 (cis/*Trans*, NHCH2CH2CH2CH2CH3); **HRMS** (ESI), m/z: calcd for $|C_{12}H_{21}CINO_2Na|$ ⁺ 268.1080 [M+Na]+; found 268.1077.

(2-chlorocyclohex-1-en-1-yl)methyl phenylcarbamate (23)

FTIR: neat (cm-1): 3425 (NH), 1732 (C=O), 1601 (C=C), 1050 (C-O) cm-1 ; **¹H NMR (300 MHz, CDCl3) δ** : 7.40-7.03(m, 5H, Harm), 7.01(s,1H,NH), 4.84 (s, 2H, CH2O), 2.34-2.32 (m, 2H, CH2cyc), 2.18- 2.14 (m, 2H, CH2cyc), 1.70-1.59 (m, 4H, CH2CH2cyc); **¹³C NMR (75 MHz, CDCl3) δ:** 153.67(C=O), 137.90(C-Cl), 128.97 (CCl=C), 131.15-118.72(C_{arm}), 65.12(CH₂O), 33.89(CH₂cyc), 27.93(CH₂cyc), 23.58(CH₂cyc), 21.94(CH₂cyc); **HRMS** (ESI), m/z: calcd for $[C_{14}H_{16}CNO_2Na]^+$ 288.0767 [M+Na]+; found 288.0769.

(2-chlorocyclohex-1-en-1-yl)methyl (3-chlorophenyl)carbamate (24)

FTIR: neat (cm⁻¹): v 3432 (NH), 1732 (C=O), 1598(C=C), 1049 (C-O), 667 (C-Cl) cm⁻¹; ¹**H NMR (300 MHz, CDCl3) δ:** 7.54 (s, 1H, NH), 7.26-7.02 (m, 4H, Harm), 4.86 (s, 2H, CH2O), 2.37 (m, 2H, CH2cyc), 2.18 (m, 2H, CH2cyc), 1.73-1.62 (m, 4H, CH2cyc); **¹³C NMR (75 MHz, CDCl3) δ:** 153.43 (C=O), 139.16 (C=CCl), 134.69-

118.78 (Carm), 116.72 (C=CCl), 65.39 (CH₂O), 33.90 (CH₂cyc), 27.99 (CH₂cyc), 23.58 (CH₂cyc), 21.95 $(CH_2$ cyc); **HRMS** (ESI), m/z: calcd for $[C_{14}H_{15}Cl_2NO_2Na]^+322.0378$ [M + Na]+; found 322.0374.

(2-chlorocyclohept-1-en-1-yl)methylpropylcarbamate (25)

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FTIR: neat (cm⁻¹): \vee 3345/3337 (NH), 1710 (C=O), 1695 (C=C), 1135 (C-O) cm⁻¹; ¹**H NMR (300 MHz, CDCl₃**) δ: 4.80 (s, 1H, N<u>H)</u>, 4.65(s, 2H, C<u>H</u>₂O), 3.08-3.04 (m, 2H, NHCH₂CH₂CH₂CH₃), 2.55-2.52(m, 2H, CH₂cyc), 2.20-2.17(m, 2H, CH₂cyc), 1.68-1.65 (m, 2H, NHCH₂CH₂CH₃), 1.56-1.41 (m, 6H, CH₂cycCH₂cycH₂cyc), 0.87- 0.82 (*Cis+Trans*, m, 3H, NHCH2CH2CH3); **¹³C NMR (75 MHz, CDCl3) δ:** 156.60(C=O), 134.74(C=CCl), 133.78(C=CCl), 66.37(CH₂O), 44.38/42.77 (*cis/trans*, NHCH₂CH₂CH₃), 38.90 (CH₂cyc), 31.24 (CH₂cyc), 30.28 (CH₂cyc), 25.75 (CH₂cyc), 25.07 (CH₂cyc), 23.16/21.07 (*Trans/cis*, NHCH₂CH₂CH₃), 11.12/11.01 (*Trans/cis*, NHCH₂CH₂CH₃); **HRMS** (ESI), m/z: calcd for [C₁₂H₂₀ClNO₂Na]⁺ 268.1080[M + Na]+; found 268.1080.

(2-chlorocyclohept-1-en-1-yl)methyl butylcarbamate (26)

FTIR: neat (cm⁻¹): v 3452/3334 (NH), 1704 (C=O), 1516 (C=C), 1135(C-O), 666 (C-Cl) cm⁻¹; ¹**H NMR (300 MHz, CDCl₃**)</sub> **δ :** 5.20 (s, 1H, N<u>H</u>), 4.63 (s, 2H, CH₂O), 3.10-3.05 (m, 2H, NHCH₂CH₂CH₂CH₃), 2.54-2.50 (m, 2H, CH₂cyc), 2.19-2.16 (m, 2H, CH₂cyc), 1.66-162 (m, 2H, NHCH₂CH₂CH₂CH₂CH₃), 1.55-1.50 (m, 2H, NHCH2CH2CH2CH3), 1.47-1.20 (m, 6H, C<u>H2CH2CH2</u>cyc), 0.84 (t, ³J_{H-H}= 7.27Hz, 3H, NHCH2CH2CH2C<u>H</u>3); **¹³C NMR (75 MHz, CDCl3) δ:** 156.58 (C=O), 134.33 (C-Cl), 133.80 (ClC=C), 66.06 (CH2O), 40.64 (NHCH₂CH₂CH₂CH₃), 38.79 (CH₂cyc), 31.92 (NHCH₂CH₂CH₂CH₂CH₃), 31.16 (CH₂cyc), 30.05 (CH₂cyc), 25.65 (CH₂cyc), 24.98 (CH₂cyc), 19.75 (NHCH₂CH₂CH₂CH₃), 13.55 (NHCH₂CH₂CH₂CH₃); **HRMS** (ESI), m/z: calcd for $[C_{13}H_{22}CINO_2Na]^+$ 282.1237 $[M + Na]^+$; found 282.1239.

(2-chlorocyclohept-1-en-1-yl)methyl phenylcarbamate (27)

FTIR: neat (cm⁻¹): v 3435 (NH), 1732 (C=O), 1602 (C=C), 1051 (C-O), 667 (C-Cl) cm⁻¹; ¹**H NMR (300 MHz, CDCl3) δ :** 7.44-7.07 (m, 5H, Harm), 6.82 (s, 1H, NH), 4.87 (s, 2H, CH2O), 2.67-2.63 (m, 2H, CH2cyc), 2.35- 2.31 (m, 2H, CH2cyc), 1.81-1.53 (m, 6H, CH2CH2CH2cyc)**; ¹³C NMR (75 MHz, CDCl3) δ:** 153.26 (C=O), 139.08 (C-Cl), 116.64 (ClC=C), 135.94-118.76(Carm), 67.30 (CH2O), 38.97 (CH2cyc), 31.20 (CH2cyc), 30.60 (CH₂cyc), 25.81 (CH₂cyc), 25.04 (CH₂cyc); **HRMS** (ESI), m/z: calcd for [C₁₅H₁₈ClNO₂Na]⁺ 302.0924 [M+Na]⁺; found 302.0923.

(2-chlorocyclohept-1-en-1-yl)methyl (3-chlorophenyl)carbamate (28)

FTIR: neat (cm⁻¹): v 3428/3322 (NH), 1711 (C=O), 1596(C=C), 1051 (C-O), 680 (C-Cl) cm⁻¹;¹**H NMR (300 MHz, CDCl₃**) δ: 7.43 (s, 1H, NH), 7.13-6.78 (m, 4H, H_{arm}), 4.75 (s, 2H, CH₂O), 2.56-2.52 (m, 2H, CH₂cyc), 2.23-2.19 (m, 2H, CH2cyc), 1.68-1.44 (m, 6H, CH2CH2CH2cyc)**; ¹³C NMR (75 MHz, CDCl3) δ:** 153.32 (C=O), 134.75 (C-Cl), 123.45 (ClC=C), 139.09-116.68 (Carm), 67.29 (CH₂O), 38.96 (CH₂cyc), 31.19 (CH₂cyc), 30.57 (CH2cyc), 25.80 (CH2cyc), 25.04 (CH2cyc); **HRMS** (ESI), m/z: calcd for [C15H17Cl2NO2Na]⁺336.0534 [M+Na]⁺; found 336.0532.

Cinnamylpropylcarbamate **(29)**

FTIR: neat (cm-1): 3448 (NH), 1720 (C=O), 1695 (C=C), 1037 (C-O) cm-1 ; **¹H NMR (300 MHz, CDCl3) δ** 7.40-7.20 (m, 5H, Harm), 6.61 (d, ${}^{3}J_{H-H}= 15.90$ Hz, 1H, PhC<u>H</u>=CH), 6.30 (td, ${}^{3}J_{H-H}= 6Hz$, ${}^{3}J_{H-H}= 15.90$ Hz, 1H,

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CH=CHPh), 5.06 (s, 1H, NH), 4.74 (d, ³J_{H-H}= 6Hz, 2H, C<u>H</u>₂OH), 3.20-3.13 (m, 2H, NHC<u>H</u>₂CH₂CH₃), 1.60-1.48 (m, 2H, NHCH2CH2CH3), 0.93 (t, ³ JH-H= 7.37Hz, 3H, NHCH2CH2CH3)**; ¹³C NMR (75 MHz, CDCl3) δ:** 156.4853 (C=O), 136.43 (C=CPh), 133.41-126.57 (Carm), 124.19 (CH=CPh), 65.22 (CH2O), 42.81 (NHCH2CH2CH3), 23.19 (NHCH2CH2CH3), 11.18 (NHCH2CH2CH3); **HRMS** (ESI), m/z: calcd for $[C_{13}H_{17}NO_2Na]^+$ 242.1157 [M + Na]⁺; found 242.1157.

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Cinnamyl butylcarbamate (30)

FTIR: neat (cm-1): 3340 (NH), 1695 (C=O), 1519 (C=C), 1137 (C-O) cm-1 ; **¹H NMR (300 MHz, CDCl3) δ :** 7.26-7.11 (m, 5H, H_{arm}), 6.50 (d, ³J_{H-H}=15.91Hz, 1H, PhC<u>H</u>=CH), 6.16 (dt, ³J_{H-H}=15.88Hz, ³J_{H-H}=6.28Hz, 1H,PhCH=CH), 4.98 (s, 1H, NH), 4.60 (d, ³ JH-H= 6.03 Hz, 2H, CH2O), 3.16-3.03 *(cis+trans*, m, 2H, NHC<u>H</u>₂CH₂CH₂CH₃), 1.39-1.18 (m, 4H, NHCH₂C<u>H₂CH₂CH₃), 0.80 (*trans*, t, ³J_{H-H}=7.25, 3H,</u> NHCH2CH2CH2CH3); **¹³C NMR (75 MHz, CDCl3) δ:** 156.48/156.41 (*Trans/cis*, C=O), 133.36-126.54 (Carm), 136.42 (CH=CHPh), 124.18 (PhHC=CH), 65.20 (CH₂O), 40.79/40.39 (*Trans/Cis*, NHCH₂CH₂CH₂CH₃), 32.05/31.76 (*Trans/Cis*, NHCH2CH2CH2CH3), 20.14/19.88 (*Cis/Trans*, CH2CH2CH2CH3), 13.70 (*Trans*, $CH_2CH_2CH_2CH_3$; **HRMS** (ESI), m/z: calcd for $[C_{14}H_{19}ClNO_2Na]^+$ 256.1313 [M+Na]⁺; found 256.1310.

Cinnamylphenylcarbamate (31)

FTIR: neat (cm-1): 3435 (NH), 1728 (C=O), 1601(C=C), 1050 (C-O) cm-1 ; **¹H NMR (300MHz, CDCl3) δ:** 7.50-7.11 (m, 11H, $H_{arm} + NH_{}$), 6.74 (d, ${}^{3}J_{H-H}$ = 15.89Hz, 1H, CH), 6.39 (dt, $({}^{3}J_{H-H})$ doublet = 15.87Hz, $({}^{3}J_{H}$ ^H)triplet= 6.38 Hz, 1H, CH=CHPh), 4.90 (d, ³ JH-H= 6.35Hz, 2H, CH2O); **¹³C NMR (75 MHz, CDCl3) δ:** 153.70 (C=O), 138.05-123.52 (Carm), 134.23 (PhHC=CH), 119.05 (PhHC=CH), 65.85 (CH2O); **HRMS** (ESI), m/z: calcd for $[C_{16}H_{15}NO_2Na]^+$ 276.1000 $[M+Na]^+$; found 276.1003.

Cinnamyl (3-chlorophenyl) carbamate (32)

FTIR: neat (cm⁻¹): v 3322 (NH), 1713 (C=O), 1596(C=C), 1048 (C-O), 692 (C-Cl) cm⁻¹; ¹**H NMR (300 MHz, CDCl3**) **ô:** 7.58 (s, 1H, N<u>H</u>), 7.43-7.06 (m, 9H, H_{arm}), 6.71 (*d*, ³J_{H-H}= 15.90Hz, 1H, PhC<u>H</u>=CH), 6.34 (td, ³J_{H-H}= 6.40Hz, ³ JH-H= 15.90Hz, 1H, PhCH=CH), 4.87 (d, ³ JH-H= 6.39Hz, 2H, CH2OH); **¹³C NMR (75 MHz, CDCl3) δ:** 153.65 (C=O), 139.37-117.08 (Carm), 134.46 (PhC=CH), 126.67 (PhC=CH), 66.16 (CH2O); **HRMS** (ESI), m/z: calcd for $[C_{16}H]_4CINO_2Na]$ ⁺ 310.0611 $[M+Na]$ ⁺; found 310.0603.

3-Chloro-3-phenylallyl propylcarbamate (33)

FTIR: neat (cm⁻¹): v 3346/3353 (NH), 1716 (C=O), - (C=C), 1037 (C-O) cm⁻¹; ¹H NMR (300 MHz, CDCl3) δ **:** (Z) 7.62-7.34 (m, 5H, H_{arm}), 6.30 (t, ³J_{H-H}= 6.04Hz, 1H, ClC=C<u>H</u>), 4.94 (s, 1H, N<u>H</u>), 4.95 (d, ³J_{H-H}= 6.04Hz, 2H, CH₂OH), 3.18-3.16 (m, 2H, NHCH₂CH₂CH₃), 1.61-1.49 (m, 2H, NHCH₂CH₂CH₃), 1.74-1.63 (m, 2H, CH₂CH₂CH₃), 0.93 (t, ³J_{H-H}= 6.64Hz, 3H, NHCH₂CH₂C<u>H</u>₃). *(E)*: 6.17 (t, ³J_{H-H}= 5.83Hz, 1H, C<u>H</u>), 4.56 (d, ³J_{H-H}= 5.80Hz, 2H, CH2OH); **¹³C NMR (75 MHz, CDCl3) δ:** 156.33 (*Z*, C=O), 137.13 (*Z*, C-Cl), 135.46-126.48 (Carm), 124.36 (*E*, ClC=CH), 122.47 (*Z*, ClC=CH), 62.31 *(Z*, CH2O), 42.85 (NHCH2CH2CH32), 23.18 $(NHCH_2CH_2CH_3)$, 11.16 (NHCH₂CH₂CH₃); **HRMS** (ESI), m/z: calcd for $[C_{13}H_{16}ClNO_2Na]^+$ 276.076 [M+Na]⁺; found 276.0767.

3-chloro-3-phenylallyl (3-chlorophenyl) carbamate (34)

FTIR: neat (cm⁻¹): \vee 3420/3336 (NH), 1732 (C=O), 1597 (C=C), 1062 (C-O) cm⁻¹; ¹**H NMR (300 MHz, CDCl3**) **ô:** 7.64-7.05 (m, 10H, H_{arm}), 6.95 (s, 1H, N<u>H</u>), 6.35 (Z, t, ³J_{H-H}= 6.23Hz, 1H, C<u>H</u>), 6.20 (*E*, t, ³J_{H-H}= 7.5Hz, 1H, CH), 5.06 (*Z*, d, ³ JH-H= 6.23Hz, 2H, CH2O), 4.69 (*E*, d, ³ JH-H= 7.5Hz, 2H, CH2O)**; ¹³C NMR (75 MHz, CDCl3) δ:** 153.14 (C=O), 138.96-116.76 (Carm), 136.91 (*Z*, C=CCl), 121.34 (*Z*, C=CCl), 120.84 (*E*, C=CCl), 62.88 (*Z*, CH2O), 62.29 (*E*, CH2O); **HRMS** (ESI), m/z: calcd for [C16H13Cl2NO2Na] ⁺344.0221 $[M+Na]^{+}$; found 344.0247.

3-chloro-2-methyl-3-phenylallyl propylcarbamate (35)

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FTIR: neat (cm⁻¹): v 3445/3351 (NH), 1715 (C=O), 1699 (C=C), 1135 (C-O), 667 (C-Cl) cm⁻¹; ¹H NMR **(300 MHz, CDCl₃)** δ **:** 7.34 (m, 5H, H_{arm}), 4.92 (s, 1H, N<u>H</u>), 4.54 (*E*, s, 2H, C<u>H</u>₂O), 3.11 (*m*, ³J_{H-H}= 6.69Hz, 2H, NHC<u>H</u>2CH2CH3), 2.05 (E, s, 3H, C<u>H</u>3), 1.78 (Z, s, 3H, C<u>H3), 1.57-1.45 (m, 2H, NHCH2CH2</u>CH3), 0.91 (t, ³J_{H-H}= 7.37Hz, 2H, NHCH2CH2CH3); **¹³C NMR (75 MHz, CDCl3) δ:** 156.30 (C=O), 138.46 (*Z*, CCl), 137.96 (*E*, CCl), 132.27-128.27 (Carm), 128.62 (C=CCl), 66.96 (*Z*, CH2O), 65.36 (*E*, CH2O), 44.42 (Z, NHCH2CH2CH3), 43.42/42.79 (cis/Trans *E*, NHCH2CH2CH3), 23.14/21.12 (*trans/Cis, E*, NHCH2CH2CH3), 17.95 (*E*, CH3), 17.52 (Z, CH_3) , 11.15 (*Cis+Trans, E*, NHCH₂CH₂CH₃); **HRMS** (ESI), m/z: calcd for $[C_{14}H_{18}CINO_2Na]$ ⁺ 290.0924 [M+Na]+; found 290.0930.

3-chloro-2-methyl-3-phenylallyl phenylcarbamate (36)

FTIR: neat (cm⁻¹): v 3423/3338 (NH), 1732 (C=O), 1599 (C=C), 1047 (C-O), 895 (C-Cl) cm⁻¹; ¹**H NMR (300 MHz, CDCl₃**)</sub> δ **:** 7.41-7.08 (m, 10H, H_{arm}), 6.73 (s, 1H, NH), 5.10 (**Z**, s, 2H, CH₂O), 4.66 (**E**, s, 2H, CH₂O), 2.12 (*E*, s, 3H, CH3), 1.85 (*Z*, s, 3H, CH3)**; ¹³C NMR (75 MHz, CDCl3) δ:** 153.17 (C=O), 137.86 (*E*, C=CCl), 137.68(*Z*, C=CCl), 133.03-123.61 (Carm), 119.79 (*Z*, C=CCl), 118.75 (*E*, C=CCl), 65.93 (*E*, CH2O), 22.29 (*Z*, CH₃), 18.11 (*E*, CH₃); **HRMS** (ESI), m/z: calcd for $[C_{17}H_{16}CINO_{2}Na]$ ⁺ 324.0767 [M+Na]+; found 324.0765. *3-chloro-2-methyl-3-phenylallyl (3-chlorophenyl) carbamate (37)*

FTIR: neat (cm⁻¹): v 3406/3320 (NH), 1708 (C=O), 1594(C=C), 1043 (C-O), 696 (C-Cl) cm⁻¹; ¹**H NMR (300 MHz, CDCl3) δ:** 7.52 (s, 1H, NH), 7.38-6.96 (m, 9H, Harm), 5.10 (*Z*, s, 2H, CH2O), 4.67 (*E*, s, 2H, CH2O), 2.11 (*E*, s, 3H, CH3), 1.90 (*Z,* s, 3H, CH3); **¹³C NMR (75 MHz, CDCl3) δ:** 153.07 (C=O), 138.99-116.79 (Carm), 137.80 **(***E***,** CH3C=CCl), 123.63 (*E***,** CH3C=CCl), 68.34 (*Z*, CH2O), 66.17 (*E*, CH2O), 18.10 (*E*, CH3), 17.82 (*Z*, CH_3 ; HRMS (ESI), m/z : calcd for $[C_{17}H_{15}Cl_2NO_2Na]$ ⁺ 258.0378 [M+Na]⁺; found 258.0381.

(Z)-3-chloro-4,4-dimethylpent-2-en-1-yl phenylcarbamate (38)

FTIR: neat (cm⁻¹): \vee 3423/3336 (NH), 1731 (C=O), 1600 (C=C), 1061 (C-O) cm⁻¹; ¹**H NMR (300 MHz, CDCl₃**) δ **:** 7.43-7.06 (m, 5H, Harm), 6.62 (s, 1H, N<u>H</u>), 5.76 (t, ³J_{H-H}= 5.92Hz, 1H, ClC=C<u>H</u>), 4.87 (d, 2³J_{H-H}= 5.92Hz, H, CH2O), 1.22 (s, 9H, (CH3)3)**; ¹³C NMR (75 MHz, CDCl3) δ:** 153.60/153.54 (cis/trans, C=O), 148.65 (CH=CCl), 137.83-118.74 (*cis+trans,* Carm), 117.40 (CH=CCl), 62.77 (CH2O), 38.81 (C(CH3)3), 28.65 $(C(\underline{CH}_3)_3)$; **HRMS** (ESI), m/z: calcd for $[C_{14}H_{18}CINO_2Na]^+$ 290.0924 [M+Na]⁺; found 290.0921.

3-chloro-2-ethylbut-2-en-1-yl propylcarbamate (39)

FTIR: neat (cm-1): 3335 (NH), 1692 (C=O), 1660 (C=C), 1042 (C-O)cm-1 ; **¹H NMR (300 MHz, CDCl3) δ** 4.86 (s, 1H, NH), 4.68 (*Z*, s, 2H, CH2O), 4.52 (*E*, s, 2H, CH2O), 3.07-3.05 (m, 2H, NHCH2CH2CH3), 2.43 (*E*, q,

3 JH-H= 7.38Hz, 2H, CH2CH3), 2.34 (*E*, q, ³ JH-H= 7.47Hz, 2H, CH2CH3), 1.79 (*E*, s, CH3), 1.69 (*Z*, s, CH3), 1.51- 1.39 (m, 2H, NHCH₂CH₂CH₃), 1.04 (*E*, t, ³J_{H-H}= 7.4Hz, 3H, CH₂CH₃), 1.03(*Z*, t, ³J_{H-H}= 7.38Hz, 3H, CH₂CH₃), 0.84 (t, ³ JH-H= 7.39Hz, 3H, NHCH2CH2CH3) ; **¹³C NMR (75 MHz, CDCl3) δ:** 156.58/156.40 (*cis/trans*, C=O), 137.39 (*Z+E*, C=CCl), 126.20 (*E*, C=CCl), 120.82 (*Z*, C=CCl), 65.72 (*Z*, CH2O), 64.18 (*E*, CH2O), 44.37 (*Z*, CH2CH3) ,42.76 (*E*, CH2CH3), 28.95 (*Z*, CH3), 28.82 (*E*, CH3), 23.11/21.05 (*Trans/ Cis*, NHCH2CH2CH3), 18.10/15.70 (*Trans Cis/,* NHCH2CH2CH3), 13.01 (*E*, CH2CH3), 11.98 (*Z*,CH2CH3), 11.08/10.98 (*Trans/Cis*, $NHCH_2CH_2CH_3$); **HRMS** (ESI), m/z: calcd for $[C_{10}H_{18}ClNO_2Na]^+$ 242.0924 $[M+Na]^+$; found 242.0917.

3-chloro-2-ethylbut-2-en-1-yl phenylcarbamate (40)

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FTIR: neat (cm-1): 3328 (NH), 1711 (C=O), 1600(C=C), 1048 (C-O) cm-1 ; **¹H NMR (300 MHz, CDCl3) δ:** 7.48-7.07 (m, 6H, H_{arm}+N<u>H</u>), 4.94 (Z, s, 2H, C<u>H</u>₂O), 4.75 (E, s, 2H, C<u>H</u>₂O), 2.56 (E, q, ³J_{H-H}= 7.38Hz, 2H, CH₂CH₃), 2.46 (Z, s, ³J_{H-H}= 7.43Hz, 3H, CH₂CH₃), 1.95 (E, s, 3H, C<u>H</u>₃), 1.84 (Z, s, 3H, C<u>H</u>₃), 1.18 (E, t, 3³J_H ^H= 7.38Hz, H, CH2CH3), 1.17 (*Z*, t, ³ JH-H= 7.43Hz, 3H, CH2CH3) ; **¹³C NMR (75 MHz, CDCl3) δ:** 153.87/153.71 (*cis/trans,* C=O), 138.05 *(E*, C=CCl), 137.93 *(Z*, C=CCl), 134.50-119.02 (Carm), 123.57 *(E*, C=CCl), 123.50 *(Z*, C=CCl), 66.31 *(Z*, CH2O), 64.68 *(E*, CH2O), 29.08 (*Z*, CH2CH3), 28.95 (*E*, CH2CH3), 18.28 (*E*, CH3), 15.91 (*Z*, CH3), 13.15 (*E*, CH2CH3), 12.09 (*Z*, CH2CH3); **HRMS** (ESI), m/z: calcd for $[C_{13}H_{16}CINO_2Na]^+$ 276.0767 [M+ Na]+; found 276.0769.

3-chloro-2-ethylbut-2-en-1-yl (3-chlorophenyl) carbamate (41)

FTIR: neat (cm⁻¹): v 3432 (NH), 1731 (C=O), 1598(C=C), 1050 (C-O), 668 (C-Cl) cm⁻¹;¹**H NMR (300 MHz, CDCl3) δ:** 7.55/7.52 (cis/trans, s, 1H, NH), 7.23-6.95 (m, 4H, Harm), 4.89 (*Z*, s, 2H, CH2O), 4.73 (*E*, s, 2H, CH₂O), 2.54 (*E*, q, ³J_{H-H}= 7.39Hz, 2H, CH₂CH₃), 2.44 (**Z**, q, ³J_{H-H}= 7.41Hz, 2H, CH₂CH₃), 1.92 (*E*, s, 3H, C<u>H</u>₃), 1.83 (**Z**, s, 3H, CH₂), 1.15 (**E**, t, ³J_{H-H}= 7.39Hz, 3H, CH₂CH₂), 1.14 (**Z**, t, ³J_{H-H}= 7.41Hz, 3H, CH₂CH₂); ¹³C **NMR (75 MHz, CDCl3) δ:** 153.17/152.86 (*trans/cis*, C=O), 138.37 (*E*, CCl), 138.05 (*Z*, CCl), 138.99-116.74 (Carm), 123.56 (*E*, C=CCH3), 123.48 (*Z*, C=CCH3), 66.57 (*Z*, CH2O), 64.93 (*E*, CH2O), 29.06 *(Z,* CH2CH3) , 28.93*(E,* CH2CH3), 18.28 *(E,* CH3), 16.00 *(Z,* CH3), 13.11 *(E,* CH2CH3), 12.04 *(Z,* CH2CH3) ; **HRMS** (ESI), m/z: calcd for $[C_{13}H_{15}Cl2NO_2Na]^+$ 310.0378 $[M+Na]+$; found 310.0386.

3-chloro-2-phenylbut-2-en-1-yl phenylcarbamate *(42)*

FTIR: neat (cm⁻¹): v 3431/3301(NH), 1727 (C=O), 1600 (C=C), 1053 (C-O), 692 (C-Cl) cm⁻¹; ¹**H NMR (300 MHz, CDCl3) δ:** 7.58-6.85 (m, Harm), 6.68/6.61 *(Trans/cis*, s, 1H, N*H*), 5.20/5.16 (*cis/Trans, Z*, s, 2H, CH2O), 5.02/4.97 (trans/*Cis E*, s, 2H, CH2O), 2.46/2.34 (trans/*Cis E*, s, 3H, CH3), 2.12/2.02 (trans/*Cis Z*, s, 3H, CH3) ; **¹³C NMR (75 MHz, CDCl3) δ:** 155.56/153.16 (*trans/Cis,*C=O), 132.54/132.07 (*Trans /cis Z* C=CCl), 131.53/131.16 (*trans/Cis E*, C=CCl), 124.11/124.04 *(Cis/trans E*, C=CCl), 123.59/123.42 (*Cis/trans Z*, C=CCl), 138.00-118.08 (*Cis+Trans Z+E*, Carm), 67.69/66.76 *(Trans/cis E*, CH2O), 66.06/65.09 (*Trans/cis Z*, CH2O), 24.13/23.94 *(Trans/cis Z*, CH3), 23.31/23.10 *(trans/Cis E,* CH3) ; **HRMS** (*ESI*), m/z: calcd for $[C_{17}H_{16}CINO_{2}Na]^{+}$ 324.0767 [M+Na]⁺; found 324.0763.

3-chloro-2-phenylbut-2-en-1-yl (3-chlorophenyl)carbamate **(43)**

FTIR: neat (cm⁻¹): \vee 3432 (NH), 1733 (C=O), 1598 (C=C), 1214 (C-O), 667 (C-Cl) cm⁻¹; ¹**H NMR (300 MHz, CDCl3) δ:** 7.35- 6.91 (m, 5H, Harm), 6.54 (*E*, s, 1H, NH), 6.47 (*Z*, s, 1H, NH), 5.07 (*Z*, s, 2H, CH2O), 4.90 (*E*, s, 2H, CH2O), 2.34 (*E*, s, 3H, CH3), 1.99 (*Z*, s, 3H, CH3); **¹³C NMR (75 MHz, CDCl3) δ**: 152.83 (C=O), 139.10-116.68 (Carm), 129.99 (*E*, PhC=CCl), 129.92 (*Z*, PhC=CCl), 123.64 (*E*, PhC=CCl), 123.49 (*Z*, PhC=CCl), 66.28 (*Z*, CH2O), 65.30 (*E*, CH2O), 24.11 (*Z*, CH3), 23.28 (*E*, CH3); **HRMS** (*ESI*), m/z: calcd for $[C_{17}H_{15}Cl_2NO_2Na]^+$ 358.0378 [M+Na]⁺; found 358.0383.

Ethyl 3-chloro-4,4,4-trifluoro-2-(((propylcarbamoyl)oxy)methyl)but-2-enoate (44)

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FTIR: neat (cm⁻¹): v 3450 (NH), 1735 (C=O), 1651(C=C), 1215 (C-O), 667 (C-Cl) cm⁻¹; ¹H NMR (200 MHz, **CDCl3) δ :** 4.97 (*Z+E*, s, 2H, CH2O), 4.23 (*Z+E*, m, 2H, CH2CH3), 3.11 (m, 2H, NHCH2CH2CH3), 1.42 (m, 2H, NHCH₂CH₂CH₃), 1.27 (Z+*E*, m, 3H, CH₂C<u>H</u>₃), 0.87 (m, 2H, NHCH₂CH₂CH₃); ¹³**C NMR (75 MHz, CDCl**3) **δ :** 163.92 (*E*, CO2Et), 163.35 (*Z*, CO2Et), 155.08 (C=O), 137.49 (*E*, CCl), 136.29 (*Z*, CCl), 121.21 (*Z*, C=CCF3), 119.40 *(Z+E,* q, ³ JC-F= 271.87Mhz, CF3), 62.50 (*E*, CH2O), 62.34 (*Z*, CH2O), 61.71 (*Z*, CH2CH3), 59.56 (*E*, CH2CH3), 43.39/42.86 (*Cis/trans*, NHCH2CH2CH3), 22.93/22.29 (*trans/Cis*, NHCH2CH2CH3), 13.81 (*E*, CH₂CH₃), 13.64 (*Z*, CH₂CH₃), 10.92/10.14 (*trans/Cis*, NHCH₂CH₂CH₃); ¹⁹**F** NM**R** (188 MHz, CDCl₃) δ : -61.83 (*E*, s, CF₃), -65.15 (*Z*, s, CF₃); **HRMS** (*ESI*), m/z: calcd for $[C_{11}H_{15}CINO_4F_3Na]^+$ 340.0539 [M+Na]⁺; found 340.0534.

Ethyl 3-chloro-4,4,4-trifluoro-2-(((phenylcarbamoyl)oxy)methyl)but-2-enoate (45)

FTIR: neat (cm⁻¹): v 3432 (NH), 1740 (C=O), 1601(C=C), 1214 (C-O), 666 (C-Cl) cm⁻¹; ¹**H NMR (200 MHz, CDCl3) δ :** 7.35-7.06 (m, 5H, Harm), 6.95 (s, 1H, NH), 5.12 (*Z+E*, s, 2H, CH2O), 4.31 (*Z+E*, m, 2H, CH2CH3), 1.29 (*Z+E*, m, 3H, CH2CH3); **¹³C NMR (75 MHz, CDCl3) δ** : 164.00 (*E*, CO2Et), 163.44 (*Z*, CO2Et), 152.22 (C=O), 137.22 (Z, CCl), 135.75-117.62 (Carm), 119.43 (Z, q, ³ JC-F= 272.43Hz, CF3), 62.82 (*Z*, CH2O), 62.65 (*E*, CH2O), 62.07 (*Z*, CH2CH3), 59.61(*E*, CH2CH3), 13.88 (*E*, CH2CH3), 13.71 (*Z*, CH2CH3); **¹⁹F NMR (188 MHz, CDCl3**) δ **:** -61.62 (*E*, s, CF₃), -64.98 (*Z*, s, CF₃); **HRMS** (*ESI*), m/z: calcd for $[C_{14}H_{13}CINO_4F_3Na]^+374.0383$ [M+Na]⁺; found 374.0394.

CRediT author statement

- **S. G.** performed and validated the experiments,
- **P. R.** XRay and structural investigation
- **B. C.** provided expertise and feedback.
- **T. B. S.** conceived, supervised, wrote the original manuscript and revised it.

conflict of interest

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