



HAL
open science

Morph-dependent nematode infection and its association with host movement in the land snail *Cepaea nemoralis* (Mollusca, Gastropoda)

M Dahirel, M. Proux, C. Gerard, Armelle Ansart

► To cite this version:

M Dahirel, M. Proux, C. Gerard, Armelle Ansart. Morph-dependent nematode infection and its association with host movement in the land snail *Cepaea nemoralis* (Mollusca, Gastropoda). *Journal of Zoology*, 2022, 318 (3), pp.181-192. 10.1111/jzo.13012 . hal-03796331

HAL Id: hal-03796331

<https://hal.science/hal-03796331v1>

Submitted on 14 Feb 2023

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

1 **Morph-dependent nematode infection and its association with**
2 **host movement in the land snail *Cepaea nemoralis* (Mollusca,**
3 **Gastropoda)**

4 Maxime Dahirel^{1,2*}, Marine Proux³, Claudia Gérard³, Armelle Ansart³

5 ¹Université Côte d'Azur, INRAE, CNRS, ISA, F-06903, Sophia-Antipolis, France

6 ²Department of Biology, Ghent University, B-9000, Ghent, Belgium

7 ³Univ Rennes, UR1, CNRS, ECOBIO (Ecosystèmes, Biodiversité, évolution), UMR 6553, F-35000,
8 Rennes, France

9 * corresponding author: maxime.dahirel@yahoo.fr

10

11 **Abstract**

12 Host behaviour can be influenced by parasitic risk and infection through a variety of direct and
13 indirect mechanisms. We can expect individuals expressing different phenotypes to also differ in
14 the ways their behaviour is altered by parasites. We used the land snail *Cepaea nemoralis*, a
15 species with various shell colour morphs differing in behaviour and physiology, as a model to
16 study the link between parasite response and individual behaviour variation. We analysed
17 metazoan parasite abundance and its relation to behaviour (movement and food intake) in
18 snails from three shell morphs (from light unbanded to darker five-banded) and from two
19 neighbouring populations from contrasted environments. Snails were parasitized by mites,
20 trematodes and nematodes, from rarest to most frequent. We confirm that terrestrial
21 gastropods can defend against infection by trapping parasitic nematodes in their shell. We show
22 that nematode encapsulated in shells can uncover past infections even when a snail population
23 is currently nematode-free, possibly due to parasite seasonality. We present new observations
24 suggesting that not only nematodes but also mites might be encapsulated in shells. Infection
25 levels varied between morphs, with darker snails harbouring fewer nematodes. Behaviour
26 (movement and food intake) was linked to nematode, but not trematode infection. Individuals
27 with higher nematode load ate less, irrespective of morph and population. The most-infected
28 morph (unbanded snails) showed reduced activity in the nematode-infected population
29 compared to the one that was nematode-free at sampling time. As a result, parasites may
30 cancel previously described behavioural differences between morphs. We discuss the possible
31 mechanisms explaining morph-dependent responses to parasites, and how parasite risk may be
32 an unseen force shaping *C. nemoralis* morph variation in space and time. We conclude on the
33 possible ecological consequences of the link, mediated by shell colour, between thermal and
34 immune responses.

35 **Keywords:** activity; *Brachylaima*; food intake; *Riccardoella*; shell colour polymorphism; snail
36 host-parasite interactions

37 Introduction

38 Parasites and the behaviour of their hosts are interlinked in complex ways: parasites change the
39 environmental context in which potential hosts express their behaviour, and directly alter actual
40 hosts' internal phenotypic state; host behaviour may favour or hinder parasite spread. Focusing
41 on movement-related behaviours (Binning, Shaw & Roche, 2017 for a review), potential hosts
42 may react to the perceived risk of infection by moving to avoid contact with parasites (e.g.
43 Davies & Knowles, 2001). Infection costs may force infected individuals to limit or delay
44 movements (e.g. Debeffe *et al.*, 2014; O'Dwyer, Kamiya & Poulin, 2014), possibly reshaping the
45 distribution of infection risk in space. All these responses may be shaped by pre-existing
46 behavioural variation, leading to feedbacks between parasite dynamics and the behaviour of
47 uninfected and infected individuals (Ezenwa *et al.*, 2016). To untangle these dynamics, and
48 understand their consequences, we need to map the existing relationships between parasite
49 infection and behavioural variation.

50 Land snails and slugs host a diverse array of metazoan parasites and parasitoids, including flies,
51 mites, cestodes, trematodes and nematodes (Barker, 2004; Morley & Lewis, 2008; Gracenea &
52 Gállego, 2017; Pieterse, Malan & Ross, 2017; Gérard *et al.*, 2020). Depending on species,
53 parasites use terrestrial gastropods as their only hosts, or as intermediate hosts with definitive
54 hosts often being birds and mammals (Barker, 2004). Some of these parasites can cause
55 diseases (e.g., strongylosis) in domesticated animals and humans (Meerburg, Singleton &
56 Kijlstra, 2009; Butcher, 2016; Giannelli *et al.*, 2016). Parasite infection is often highly variable
57 between populations (Baur & Baur, 2005; Żbikowska *et al.*, 2020).

58 Terrestrial gastropods can defend against metazoan parasites in several ways. They may alter
59 their behaviour to avoid parasite-infected microhabitats (Wynne, Morris & Rae, 2016). If
60 parasites manage to enter snails, they can be neutralized through several physiological
61 mechanisms, including cellular and non-cellular immune responses (Furuta & Yamaguchi, 2001;
62 Coaglio *et al.*, 2018). Snails can also trap nematodes directly in the inner layer of their shell (Rae,
63 2017). As in insects (González-Santoyo & Córdoba-Aguilar, 2012; San-Jose & Roulin, 2018),
64 melanin and the phenoloxidase pathway involved in melanisation play a key role in snail

65 defences against parasites (Coaglio *et al.*, 2018). This association between immunity and
66 pigment-related pathways could be especially relevant in land snails. Indeed, many species
67 exhibit shell colour polymorphism (Williams, 2017), which affects behaviour, thermal responses
68 and vulnerability to predators (e.g. Jones, Leith & Rawlings, 1977; Rosin *et al.*, 2011; Surmacki,
69 Ożarowska-Nowicka & Rosin, 2013; Köhler *et al.*, 2021).

70 The grove snail *Cepaea nemoralis* has a long history as an iconic model in evolutionary biology
71 (Jones *et al.*, 1977; Ożgo, 2009). Individuals can differ in background shell color (yellow, pink or
72 brown) as well as in the number of dark bands (zero to five). Decades of research have aimed to
73 disentangle the roles of thermal selection, predation risk and neutral processes in driving the
74 widely observed, across multiple scales, among-population differences in morph frequencies
75 (Cain & Sheppard, 1954; Lamotte, 1959; Jones *et al.*, 1977; Schilthuizen, 2013; Ożgo *et al.*,
76 2017). These color differences are associated with behavioural and physiological differences
77 (e.g. Kavaliers, 1992; Dahirel, Gaudu & Ansart, 2021), leading to potentially adaptive phenotypic
78 syndromes.

79 If susceptibility and response to parasite infection are shaped by host behaviour and physiology,
80 this means that (i) *C. nemoralis* morphs might consistently differ in their infection levels, (ii)
81 variation in infection between populations may alter host behaviour in a morph-dependent
82 way, (iii) infection and behaviour may also covary independently of morph. In this context, we
83 compared natural infections by metazoan parasites across two habitats differing in microclimate
84 conditions. We examined whether infections varied with morph and potentially morph-
85 dependent behaviours, namely food intake and activity.

86 Based on the link between pigments and immune defence outlined above, we expected snails
87 with more dark bands hosted fewer parasites. Since darker snails are less mobile (Dahirel *et al.*,
88 2021), this is in line with some predictions from the pace-of-life syndrome hypothesis, namely
89 that fast active individuals have lower immune responses (Réale *et al.*, 2010; but see Royauté *et*
90 *al.*, 2018). We also hypothesized that metazoan parasites would have notable negative effects
91 on snail performance (McElroy & de Buron, 2014), leading to slower behaviours and changed

92 food intake in more infected snails (both at the morph level and within morphs). We expected
93 these effects on behaviour to be stronger in morph(s) more vulnerable to parasite infection.

94 **Methods**

95 **Snail collection and maintenance**

96 We collected snails in early May 2019 (i.e. at most \approx 1.5 month after hibernation) from two
97 contrasted sites in and near the village of Arçais, France. From previous studies, we know that
98 snail parasites are present, albeit in another species (*Cornu aspersum*, Gérard *et al.*, 2020), and
99 that environmental differences between the sites are linked to differences in the frequency of
100 darker *Cepaea nemoralis* morphs (Dahirel *et al.*, 2021).

- 101 • The “open habitat” population is in a garden with little to no trees and thus exposed to
102 the sun throughout the year (\approx 46°17'50" N, 0°41'30" W).
- 103 • The “shaded habitat” population (\approx 46°18'01" N, 0°42'56" W) is in a small humid
104 woodlot which is not directly exposed to the sun during snail activity months, and
105 occasionally experiences partial winter flooding. The substrate is poorer in calcite than in
106 the open habitat (Dupuis *et al.*, 1983; BRGM, 2005). Five-banded snails are more
107 frequent there, in agreement with hypotheses on shell colour and thermal selection
108 (22.3% of individuals, compared to 13.5% in the open habitat; Dahirel *et al.*, 2021).

109 We sampled 90 adult individuals per population, divided equally into unbanded, three-banded,
110 and five-banded snails. Adults were recognizable by the presence of a reflected “lip” around the
111 shell opening. We only sampled snails with yellow background shells, leaving aside the rarer
112 brown and pink-shelled morphs. Sampling was random with respect to other shell features
113 (e.g. band fusion).

114 We kept snails under standard conditions ($20 \pm 1^\circ\text{C}$, L:D 16:8) in $30 \times 45 \times 8 \text{ cm}^3$ plastic crates
115 lined with 1-2 cm of soil kept humid and regularly renewed, with a lid made of a fine plastic
116 mesh. Snails were grouped by morph \times population of origin. Food (vegetal flour supplemented
117 with calcium, Hélinove, Saint Paul en Pareds, France) was available *ad libitum* in Petri dishes.

118 Four days before the experiments, we marked snails with a unique identifier on the side of their
119 shell (using Posca pens, uni Mitsubishi Pencil, Tokyo, Japan). At the same time, we measured
120 snail size (greater shell diameter) to the nearest 0.1 mm.

121 **Movement behaviour and food intake**

122 Behavioural experiments began two weeks after snail sampling. We tested each snail twice for
123 both movement and food intake, to separate repeatable among-individual variation from
124 within-individual variability. To ensure a manageable workload, we split the 180 snails in three
125 groups of 60, each starting the experimental sequence on a different day, with each group
126 containing snails from all band number × population combinations in equal proportions.

127 We first tested snails for movement behaviour. We stimulated snail activity by placing them in a
128 Petri dish containing ≈ 1 mm of water until they were out of their shell. We then placed each
129 snail individually on a clean lab bench, and used the time snails took to move more than 10 cm
130 in a straight line from their starting point as a measure of movement (longer times implying
131 slower individuals). Importantly, we only counted time after snails moved more than 2 cm
132 (i.e. about one shell length) from their starting point, to ensure all observed snails were truly
133 active. Some snails stayed inactive and the relevant observations were thus excluded (11
134 individuals missed one of the two tests, 6 individuals missed both). We stopped observation 20
135 min after placing snails on the lab bench (not after the start of activity).

136 Movement tests were immediately followed by food intake tests. We kept all snails individually
137 in 10 × 7 × 6 cm³ boxes for 18 hours (including 8 hours of darkness) and provided them with 1.5
138 g of dry food (a non-limiting quantity, large enough to feed one snail for at least three days,
139 based on preliminary tests). Afterwards, we recovered, dried (60°C for 60 hours) and weighed
140 the remaining food (to the nearest 0.1 mg) to estimate food intake.

141 Snails were then placed back in their original box. The second test for each behaviour was done
142 one week later.

143 **Parasitological research**

144 After freezing at -20°C and thawing, we dissected each snail under a binocular microscope. After
145 removing the body from the shell, we looked for and counted metazoan parasites in various
146 locations: space between shell and soft body, lung, heart, kidney, body cavity and digestive
147 gland, as in Gérard et al. (2020). We found nematodes, larval trematodes and parasitic mites
148 (Acari). Trematodes were identified to the genus level as in Gérard et al. (2020), and mites using
149 Fain (2004). That level of taxonomic precision could not be reached for nematodes. However,
150 nematodes from different families consistently use/infect different organs and reach different
151 body sizes within snails (Morand, Wilson & Glen, 2004; Wilson, 2012). We assigned nematodes
152 to size classes (<1 mm, 1-3 mm, 3-5 mm, >5 mm) using graph paper, and used nematode
153 location and size to infer the most likely families infecting our snails. We additionally observed
154 shells and shell fragments under brighter light to detect encapsulated parasites (Rae, 2017). By
155 simplicity, we use “live” as a shorthand to designate parasites found free within the body or
156 between the body and the shell during dissection, and thus presumably still living before snails
157 were sacrificed, by opposition with “trapped” or “encapsulated” parasites found within shells.

158 **Statistical analyses**

159 We analysed our data in a Bayesian framework, using R (version 4.2.0, R Core Team, 2022) and
160 the brms R package (Bürkner, 2017) as frontends for the Stan language (Carpenter *et al.*, 2017).
161 We used the tidybayes, bayesplot, patchwork packages for data preparation, model evaluation
162 and plotting, as well as the tidyverse suite of packages (Gabry *et al.*, 2019; Kay, 2019; Wickham
163 *et al.*, 2019; Pedersen, 2020). One snail (out of 180) died during the experiments and was
164 excluded from statistical analyses, but was used for natural history information (see **Results**).

165 We ran two models, which both included at least effects of population of origin, number of shell
166 bands, as well as population \times banding interactions as categorical explanatory variables
167 (hereafter “morph and population effects”).

168 We first used a Gaussian linear model to analyse how snail shell diameter (mean centred and
169 scaled to unit 1 SD), varied with morph and population effects.

170 We then ran a multivariate generalized mixed/multilevel model to examine how behaviour and
171 parasite infections varied between morph and populations, and whether these traits were
172 correlated at the individual level. Trematodes and nematodes could not be identified to species,
173 so we used the total numbers of trematodes and nematodes per host snail in our analyses (as in
174 e.g. Debeffe *et al.*, 2014). Mite infections were too rare to be analysed statistically (**Results**).
175 This multivariate model included four submodels:

- 176 • live trematode abundance was modelled as a function of morph and population effects,
177 as well as shell size, using a Poisson model. This model included random effects of test
178 group and individual identity. The latter acts as an observation-level random effect and
179 accounts for overdispersion (Harrison, 2014); it also allows us to build an individual-level
180 correlation matrix connecting all four traits.
- 181 • live nematode abundance was modeled as trematode abundance, with one change. As
182 no live nematodes were found in the shaded population, we only fit this submodel to
183 data from the open population and removed the population fixed effect. Importantly,
184 keeping all data instead does not change our conclusions (see **Data availability**).
- 185 • food consumption (mean-centred and scaled to unit 1SD) was analysed using a Gaussian
186 model including the same fixed and random effects as trematode abundance. Although
187 food consumption data are proportions (of total food), and so are well-suited to Beta
188 models (Douma & Weedon, 2019), the Gaussian model behaved much better regarding
189 posterior predictive checks (**Data availability**). As individuals were measured twice, the
190 individual random effect here allows separation between within- and among individual
191 variation.
- 192 • movement behaviour (latency to move more than 10 cm from the starting point) was
193 inverse-transformed, mean-centred and scaled to unit 1SD, and then analysed using a
194 Gaussian model including the same fixed and random effects as above. We initially
195 attempted to analyse untransformed latencies using lognormal or Gamma models, but
196 the Gaussian model performed better regarding posterior predictive checks (**Data**
197 **availability**).

198 In both models we used weakly informative priors (McElreath, 2020): Normal(0,1) priors for
199 fixed effects, half – Normal(0,1) priors for standard deviations (random effect and residual),
200 and a LKJ(2) prior for the correlation matrix in the multivariate model. See **Supporting**
201 **Information S1** for detailed model descriptions. We ran four chains per model for 4000
202 iterations each, with the first half of each chain used as warmup. We checked convergence and
203 mixing following Vehtari *et al.* (2021). All parameters had satisfactory effective sample sizes
204 (Vehtari *et al.*, 2021). Posteriors are summarised in text and figures as means [95% Highest
205 Posterior Density Intervals].

206 We also looked at the proportion of snails with encapsulated nematodes as a function of shell
207 morph and population. Here we did not run statistical models, and simply present these
208 proportions as qualitative information about past parasite exposure in both populations. We
209 explicitly refrained from analysing the prevalence or abundance of encapsulated nematodes at
210 the individual level within populations, as it is likely linked to cumulative exposure time/age,
211 and snail age is not known (and cannot be approximated by e.g. size, given *Cepaea nemoralis*
212 exhibits determinate growth).

213 **Results**

214 **Snail size**

215 Snails from the shaded population were on average smaller than those from the open habitat if
216 they were three-banded ($\Delta_{[\text{shaded}-\text{open}]}$ = -0.70 mm [-1.38; -0.04], **Fig. 1**) or five-banded
217 ($\Delta_{[\text{shaded}-\text{open}]}$ = -1.37mm [-2.07; -0.71], **Fig. 1**). For unbanded snails, there was no clear
218 difference in size between habitats ($\Delta_{[\text{shaded}-\text{open}]}$ = -0.39 mm [-1.03; 0.33], **Fig. 1**).

219

220 **Parasitological information**

221 Snails from both populations and all morphs were infected by metazoan parasites (**Figs. 2-3**, see
222 **Supporting Information S2** for detailed descriptive statistics about prevalence and infection
223 intensity). All trematodes found were live *Brachylaima* sp. metacercariae in snails' kidneys; no

224 live sporocysts were recorded and no trematodes of any stage were found trapped in shells.
225 Among the organs we examined, we detected live nematodes only within the lung and in the
226 space between the body and the shell; most of these nematodes were in the 1-3 mm size class
227 (297/371; 211/269 excluding the snail that died during the experiments). Based on this, we can
228 tentatively assume that most nematodes found belonged to the Rhabditidae family (Morand *et*
229 *al.*, 2004; Wilson, 2012). In addition we frequently (**Fig. 3**) found nematodes encapsulated in
230 shells. No live nematodes were found in individuals from the shaded population (**Fig.3**), by
231 contrast to the open population where 65.6% of snails were infected (59/90, including the dead
232 snail). However, a high number of snails contained encapsulated nematodes in their shells, in
233 both populations (**Figs 2, 3**), indicating past exposure to nematodes even in the shaded
234 population. Only one snail (unbanded from the shaded habitat) harboured live mites ($N = 93$
235 mites), identified as *Riccardoella* sp. Three other snails (all five-banded from the shaded habitat)
236 had (unidentified) mites trapped in their shell ($N = 1, 2$ and 15 mites, **Fig. 2**). As mentioned in
237 **Methods**, one snail among 180 died during the experiments; it harboured 102 live nematodes,
238 more than 4 times the number of the most infected surviving snail (**Fig. 3**).

239
240 In the open population where live nematodes were found, nematode abundance was morph-
241 dependent: five-banded snails were on average infected by about a third of the nematodes
242 infecting other snails ($N_{[5bands]}/N_{[0bands]} = 0.29$ [0.11; 0.52], $N_{[5bands]}/N_{[3bands]} = 0.34$ [0.12;
243 0.62]; **Table 1, Fig. 3**), mostly due to differences in prevalence between morphs (**Supporting**
244 **Information S2**). Five-banded snails also contained fewer trematodes, but only when compared
245 to three-banded snails and only clearly in the shaded habitat ($N_{[5bands]}/N_{[3bands]} = 0.19$ [0.00;
246 0.57]; **Fig. 3, Supporting Information S3**). Parasite abundance (trematode or nematode) was not
247 linked to shell size (**Table 1**).

248 249 **Snail behaviour**

250 Both movement and food intake were repeatable at the individual level (see individual-level SDs
251 **Table 1**), although adjusted repeatability (sensu Nakagawa & Schielzeth, 2010) was weaker and

252 closer to 0 for movement ($r_{[\text{movement}]} = 0.11 [0.00; 0.23]$; $r_{[\text{food intake}]} = 0.29 [0.15; 0.42]$;
253 **Supporting Information S4**).

254 Movement depended on both population of origin and shell band number (**Table 1, Fig. 4,**
255 **Supporting Information S3**). In the shaded population, five-banded snails were on average less
256 mobile than unbanded snails ($\Delta_{[5\text{bands}-0\text{bands}]} = -0.61$ SDs $[-1.00; -0.21]$). In snails from the
257 open habitat however, there was no such difference between morphs ($\Delta_{[5\text{bands}-0\text{bands}]} = 0.17$
258 SDs $[-0.19; 0.59]$). This was due to unbanded snails being less mobile when coming from the
259 open habitat than when coming from the shaded population ($\Delta_{[\text{shaded}-\text{open}]} = 0.71$ SDs $[0.31;$
260 $1.08]$), while there was no clear between-population difference for five-banded snails
261 ($\Delta_{[\text{shaded}-\text{open}]} = -0.08$ SDs $[-0.47; 0.32]$)(**Table 1, Fig. 4**). In any case, there was no detectable
262 link between movement and shell size (**Table 1**).

263 Larger snails tended to eat more (**Table 1**). Source population had no effect on food intake if
264 snails were three- ($\Delta_{[\text{shaded}-\text{open}]} = 0.03$ g $[-0.02; 0.08]$, **Fig. 4**) or five-banded ($\Delta_{[\text{shaded}-\text{open}]} =$
265 0.02 g $[-0.03; 0.07]$, **Fig. 4**), but similarly to movement, unbanded snails coming from the shaded
266 population ate more on average than their counterparts from the open habitat ($\Delta_{[\text{shaded}-\text{open}]} =$
267 0.05 g $[0.00; 0.10]$, **Fig. 4**).

268
269 Looking at the individual-level correlation matrix between traits (i.e. once fixed effects of shell
270 morph, size and population of origin are accounted for), food intake and nematode abundance
271 are negatively correlated, while all other correlations (including between nematode and
272 trematode abundance) are not markedly different from zero (**Table 2**). The food-nematode
273 correlation persists if we refit the nematode model to include data from the shaded population
274 (**-0.58 [-0.87; -0.29]**).

275

276 **Discussion**

277 To understand how individual variation in behaviour and infection interact, we studied
278 movement, food intake and natural parasite infection across two populations and three morphs
279 of the land snail *Cepaea nemoralis*. We found evidence of morph-dependent responses to
280 parasites, as well as links between parasite infection and behaviour. We additionally present
281 evidence of land snails encapsulating parasitic arthropods (mites) in their shell.

282 Snails of the darker morphs were smaller in the shaded population (**Fig. 1**). While the two
283 habitats differ along various abiotic dimensions (calcium availability in substrate,
284 microclimate..., see **Methods**), their documented effect on shell size are highly species- and
285 study-dependent, and there is no consistent evidence for or against their role in driving the
286 differences we observe (Goodfriend, 1986). Differences in parasite infection between the two
287 habitats might also influence body size, for instance if high parasite risk drives resource
288 allocation away from growth and towards immune defence (Rantala & Roff, 2005). In any case,
289 our models conditioned on size, meaning that any effect of morph/population we discuss below
290 is “size being equal”. In addition, we did not find any effect of size itself on parasites or
291 movement (**Table 1**), meaning that the effects of morph/population we found are likely not
292 mediated by size differences.

293 One snail was found with live *Riccardoella* mites, hemolymph feeders with negative effects on
294 mollusc life-history and behavior (e.g. Schüpbach & Baur, 2008a). They are transmitted
295 horizontally during mating, but may find new hosts by leaving their host and following snail
296 mucus trails (Schüpbach & Baur, 2008b). However, their limited prevalence in our sample
297 prevented us from investigating their effects. Intriguingly, we found encapsulated mites in three
298 other snails (**Fig. 2**). Many land snails can defend themselves against nematode infection by
299 trapping them in their shell (Rae, 2017), including *Cepaea nemoralis* (this study, Williams & Rae,
300 2016; Rae, 2017). Based on extensive surveys, Rae (2017) wrote that encapsulation in the shell
301 is a specific response against nematode infection; our observations might contradict this

302 affirmation. *Riccardoella* infection is highly variable across populations or species, with reported
303 prevalences of 0% or near 0% being frequently observed (this study; Russo & Madec, 2013; Baur
304 & Baur, 2005; Segade *et al.*, 2013), which may explain why encapsulated mites were not
305 previously found. We need further studies to determine with certainty whether shell
306 encapsulation is a more general response to metazoan parasites or if it is a nematode-specific
307 response, as posited by Rae (2017), but occasionally and anecdotally catches other foreign
308 bodies.

309 A majority of snails in the open population were hosting nematodes, most of which were likely
310 Rhabditidae based on size and organ location (Morand *et al.*, 2004; Wilson, 2012). Interactions
311 between Rhabditidae and terrestrial molluscs are complex, including phoresy/endozoochory,
312 facultative parasitism, necromeny (feeding and reproducing on the host's body after its death),
313 and obligate larval parasites that actively infect snails through e.g. the lung cavity (Morand *et al.*,
314 2004; Pieterse *et al.*, 2017; Sudhaus, 2018). Delimiting these interaction types is complicated
315 because even when they have no effects on adults, these nematodes can have dramatically
316 negative effects on juveniles (Grannell, Cutler & Rae, 2021). By contrast to the open population,
317 we found no live nematodes in any snail from the shaded population (**Fig. 3**; note that our
318 methods were not designed to detect nematodes embedded in e.g. snail foot tissue). Thanks to
319 nematodes trapped in shells (**Fig. 3**), which integrate infections over time, we know parasitic
320 nematodes are not truly absent from the shaded population; they were simply not infecting
321 snails at the time of sampling. The "shaded" woodlot habitat is sheltered from the sun, more
322 humid and likely colder than the open one. Infections are often dependent on abiotic
323 parameters, including temperature and humidity, leading to seasonality (Altizer *et al.*, 2006).
324 Differences in nematode infection between the habitats may then be explained, at least partly,
325 by differences in microclimate. First, the different microclimates experienced by the two
326 populations may favour different nematode species within the soil community, differing in their
327 tendency to parasitism. Second, even if nematode communities are similar, conditions
328 favourable to nematode activity, development or infection may be reached only later in the
329 year in the shaded habitat. For instance, *Deroceras reticulatum* slugs were more negatively
330 impacted by *Phasmarhabditis hermaphrodita* infection when conditions were artificially

331 warmed during winter (El-Danasoury & Iglesias-Piñeiro, 2017), indicating that warmer
332 conditions may favour nematode activity. In any case, these results mean that any between-
333 population differences in snail behaviour in the present study should be interpreted not only in
334 relation with microhabitat and microclimate differences, but also in terms of average infection
335 levels.

336 In the open population where active nematodes were actually found, morph differences in
337 infection seem to validate our initial hypothesis of increased immune defence in darker snails
338 (**Fig. 3**). By contrast, no significant morph difference in immune defence was found in the
339 related *Cepaea hortensis* (Scheil *et al.*, 2013, 2014), but their endpoints were different
340 (phenoloxidase activity and mortality). In addition, our behavioural results (discussed below) are
341 also consistent with the hypothesis of morph-dependent immunity. These results are interesting
342 in the light of recent evidence that melanins may not be the pigments forming the dark bands in
343 *C. nemoralis* (Affenzeller *et al.*, 2020). Indeed, our initial hypothesis when designing this
344 experiment (in 2019) was based on the oft-seen association between immunity and melanin
345 (San-Jose & Roulin, 2018), including in snails (Coaglio *et al.*, 2018), although other pigments may
346 also have links to immunity (Tan *et al.*, 2020). While it is still likely that melanin-associated
347 pathways play a role in immune defence in *Cepaea* (Scheil *et al.*, 2014), it is probably not as
348 simple as “banded snails constitutively produce more melanin, and thus are better protected”.
349 More studies are needed to understand the mechanisms linking shell morph to nematode
350 resistance.

351 Snails from all morphs and both populations were infected by *Brachylaima* trematodes (**Fig. 3**).
352 We previously documented *Brachylaima* infecting *Cornu aspersum* in the same area (Gérard *et al.*,
353 2020); finding these parasites a year later and in *C. nemoralis* may indicate that they are
354 well-implanted in local communities. Contrary to nematodes, we found no evidence that morph
355 or population influenced trematode abundance (**Fig. 3**). This is consistent with another study
356 analyzing both *C. nemoralis* and *C. hortensis* (Žbikowska *et al.*, 2020). We must acknowledge
357 however that snail infections by *Brachylaima* vary seasonally, and that spring may not be the
358 infection peak (Gracenea & Gállego, 2017). Longitudinal studies, possibly combined with

359 experimental infections or cures, may help confirm whether morphs respond differently to
360 trematode risk.

361 Nematode and trematode numbers were uncorrelated (**Table 2**). Terrestrial gastropods are
362 commonly infected by both nematodes and trematodes at the same time (e.g. Segade *et al.*,
363 2013; Antzée-Hyllseth *et al.*, 2020). Our results extend these previous studies, and suggest that
364 despite sharing a host, nematode and trematode niches are sufficiently distinct within snails to
365 limit spatial and trophic competition. We reiterate however that studies more expansive in
366 space and time, including seasons and/or populations with higher prevalence and abundance of
367 trematodes and nematodes, are needed to confirm this. Combined with finer-grained studies
368 (e.g. at the parasite species level; Morand *et al.*, 2004), this would allow us to understand better
369 how parasites with similar or different niches within snails interact.

370 We found that movement behaviour in *C. nemoralis* was morph-dependent, but that this
371 dependency varied between populations: five-banded morphs were less mobile, but contrary to
372 Dahirel *et al.* (2021), this was only the case in the shaded population (**Fig. 4**). In the open
373 population, snail morphs are mobile at similarly low levels: five-banded snails do not change
374 behaviour between the two populations, while unbanded snails go down to five-banded snail
375 levels of mobility (**Fig. 4**). One key difference between the present study and Dahirel *et al.*
376 (2021) is the collection date. While here snails were collected several weeks after the onset of
377 spring activity and kept active throughout, in Dahirel *et al.* (2021) they were collected in late fall
378 and stored at cold temperatures inducing dormancy for months before being tested. This
379 difference may be especially relevant: winter or summer dormancy can lead to sharp decreases
380 in parasite infection in several parasite-snail systems (Solomon, Paperna & Matkovics, 1996;
381 Haeussler *et al.*, 2012; Segade *et al.*, 2013). We can thus reconcile the behavioral results from
382 Dahirel *et al.* (2021) from those of the present study if we assume that in the former
383 experiment, hibernation reduced parasite load across all morphs and populations, and that in
384 the present study, lower temperatures in the shaded habitat led to longer hibernation. We can
385 then interpret population differences in the present study in the light of their different infection
386 patterns. Five-banded snails' behaviour is almost unchanged between the two populations,
387 while unbanded snails from the "infected" population are slower than in the nematode-free

388 population (**Fig. 4**). This is again consistent with the idea that unbanded snails are more
389 vulnerable to parasites than banded ones. The question then becomes: is this reduction in
390 activity in infected or at-risk unbanded snails simply the direct consequence of the costs of
391 infection, or does it reflect something more? For instance, infective nematodes may be able to
392 follow mucus trails from active snails (Andrus *et al.*, 2020). Reducing activity and the associated
393 mucus production may then be protective for snails, by limiting the risk of further infections.
394 Experimental manipulations of both snail infection and parasite risk in the environment may
395 help disentangle these two responses. Future studies including species-level identification of
396 nematodes will also be needed, to determine whether nematodes with more negative impacts
397 also trigger stronger behavioural responses, or if snails respond indiscriminately to nematode
398 abundance.

399 Food intake was influenced by nematode infection: highly parasitized snails ate less (**Table 2**).
400 Feeding reduction in nematode-infected individuals has been documented in other terrestrial
401 molluscs (Antzée-Hyllseth *et al.*, 2020; Saeedizadeh & Niasti, 2020) and is an indication that
402 parasite infection is indeed costly for snails even if they are non-lethal, at least in adults (Wilson
403 *et al.*, 2000; Williams & Rae, 2016; Grannell *et al.*, 2021). By contrast, trematode infection had
404 no effect on behaviour, including feeding behaviour (**Table 2**). We however only found snails
405 infected by *Brachylaima* metacercariae. At the sporocyst stage, *Brachylaima* parasitizes the
406 digestive gland (e.g. Gérard *et al.*, 2020), which would likely have a stronger effect on snail
407 behaviour. Indeed, sporocysts can cause high damage, direct and indirect, to that key organ for
408 digestion and for energy storage (Segade *et al.*, 2011; Żbikowska *et al.*, 2020).

409 The fact that snail response to parasites seems morph-dependent for nematodes, but not
410 trematodes, has interesting implications for host-parasite and snail-predator interactions in a
411 changing world. Trematodes infecting land snails are trophically transmitted to vertebrate
412 definitive hosts including rodents and birds (Gérard *et al.*, 2020; Sitko & Heneberg, 2020). Visual
413 selection by thrushes (*Turdus philomelos*) is held as one of the key explanations for the
414 maintenance of *Cepaea* shell polymorphism (Jones *et al.*, 1977), and predation by rodents may
415 also be morph-dependent (Rosin *et al.*, 2011; Rosin *et al.*, 2013). If confirmed, our results
416 suggest that morph-dependent predation confers no additional trematode-avoiding advantage.

417 In addition, snail morphs differ in their thermal responses: lighter-coloured morphs are better
418 adapted to heat, becoming more frequent with global warming or urbanization (Jones *et al.*,
419 1977; Silvertown *et al.*, 2011; Özgo *et al.*, 2017; Kerstes *et al.*, 2019). If lighter morphs are more
420 vulnerable to infection by nematodes, then we need to understand how snail immunity will
421 interact with thermal tolerance to predict how snail and nematode dynamics will be altered in
422 response to climate change (El-Danasoury & Iglesias-Piñeiro, 2017; Aleuy & Kutz, 2020).

423 **Acknowledgements**

424 We thank participants to the 4th French Conference on Animal Ecophysiology (CEPA4, Rennes,
425 2019) for interesting discussions. Funding for MD's position was obtained by Elodie Vercken (see
426 Funding).

427 **Data availability**

428 Data and R scripts to reproduce all analyses presented here are on Github
429 (<https://github.com/mdahirel/cepaea-parasite-behaviour-2019>) and archived in Zenodo (DOI:
430 <https://doi.org/10.5281/zenodo.6327265>). Representative images of parasites trapped in shells
431 are also archived in Zenodo (DOI: <https://doi.org/10.5281/zenodo.6326305>).

432 **Funding**

433 MD was a postdoctoral researcher funded by the French Agence Nationale de la Recherche
434 (PushToiDeLa, ANR-18-CE32-0008), during the course of this research.

435 **References**

436 Affenzeller, S., Wolkenstein, K., Frauendorf, H. & Jackson, D.J. (2020). Challenging the concept
437 that eumelanin is the polymorphic brown banded pigment in *Cepaea nemoralis*. *Sci. Rep.* **10**,
438 2442.

439 Aleuy, O.A. & Kutz, S. (2020). Adaptations, life-history traits and ecological mechanisms of
440 parasites to survive extremes and environmental unpredictability in the face of climate change.
441 *Int. J. Parasitol. Parasites Wildl.* **12**, 308–317.

442 Altizer, S., Dobson, A., Hosseini, P., Hudson, P., Pascual, M. & Rohani, P. (2006). Seasonality and
443 the dynamics of infectious diseases. *Ecol. Lett.* **9**, 467–484.

444 Andrus, P., Ingle, O., Coleman, T. & Rae, R. (2020). Gastropod parasitic nematodes
445 (*Phasmarhabditis* sp.) are attracted to hyaluronic acid in snail mucus by cGMP signalling. *J.*
446 *Helminthol.* **94**, e9.

447 Antzée-Hyllseth, H., Trandem, N., Torp, T. & Haukeland, S. (2020). Prevalence and parasite load
448 of nematodes and trematodes in an invasive slug and its susceptibility to a slug parasitic
449 nematode compared to native gastropods. *J. Invertebr. Pathol.* **173**, 107372.

450 Barker, G.M. (Ed.). (2004). *Natural enemies of terrestrial molluscs*. Wallingford, UK: CABI.

451 Baur, A. & Baur, B. (2005). Interpopulation variation in the prevalence and intensity of parasitic
452 mite infection in the land snail *Arianta arbustorum*. *Invertebr. Biol.* **124**, 194–201.

453 Binning, S.A., Shaw, A.K. & Roche, D.G. (2017). Parasites and host performance: incorporating
454 infection into our understanding of animal movement. *Integr. Comp. Biol.* **57**, 267–280.

455 BRGM. (2005). Geological map at 1:50 000 (BD-CHARM-50), department: Deux-Sèvres (79,
456 France).

457 Bürkner, P.-C. (2017). brms: an R package for Bayesian multilevel models using Stan. *J. Stat.*
458 *Softw.* **80**, 1–28.

459 Butcher, A.R. (2016). Children, snails and worms: the *Brachylaima cribbi* story. *Microbiol. Aust.*
460 **37**, 30–33.

461 Cain, A.J. & Sheppard, P.M. (1954). Natural selection in *Cepaea*. *Genetics* **39**, 89–116.

462 Carpenter, B., Gelman, A., Hoffman, M.D., Lee, D., Goodrich, B., Betancourt, M., Brubaker, M.,
463 Guo, J., Li, P. & Riddell, A. (2017). Stan: a probabilistic programming language. *J. Stat. Softw.* **76**,
464 1–32.

465 Coaglio, A.L., Ferreira, M.A.N.D., dos Santos Lima, W. & de Jesus Pereira, C.A. (2018).
466 Identification of a phenoloxidase- and melanin-dependent defence mechanism in *Achatina*
467 *fulica* infected with *Angiostrongylus vasorum*. *Parasites Vectors* **11**, 113.

468 Dahirel, M., Gaudu, V. & Ansart, A. (2021). Boldness and exploration vary between shell morphs
469 but not environmental contexts in the snail *Cepaea nemoralis*. *Ethology* **13**.

470 Davies, M.S. & Knowles, A.J. (2001). Effects of trematode parasitism on the behaviour and
471 ecology of a common marine snail (*Littorina littorea* (L.)). *J. Exp. Mar. Biol. Ecol.* **260**, 155–167.

472 Debeffe, L., Morellet, N., Verheyden-Tixier, H., Hoste, H., Gaillard, J.-M., Cargnelutti, B., Picot,
473 D., Sevila, J. & Hewison, A.J.M. (2014). Parasite abundance contributes to condition-dependent
474 dispersal in a wild population of large herbivore. *Oikos* **123**, 1121–1125.

475 Douma, J.C. & Weedon, J.T. (2019). Analysing continuous proportions in ecology and evolution:
476 a practical introduction to beta and Dirichlet regression. *Methods Ecol. Evol.* **10**, 1412–1430.

477 Dupuis, J., Cariou, E., Coirier, B. & Ducloux, J. (1983). *Notice explicative, Carte géologique de la*
478 *France à 1/50 000, feuille Niort (610)*. Orléans, France: BRGM.

479 El-Danasoury, H. & Iglesias-Piñeiro, J. (2017). Performance of the slug parasitic nematode
480 *Phasmarhabditis hermaphrodita* under predicted conditions of winter warming. *J. Pestic.*
481 *Science* **42**, 62–66.

482 Ezenwa, V.O., Archie, E.A., Craft, M.E., Hawley, D.M., Martin, L.B., Moore, J. & White, L. (2016).
483 Host behaviour–parasite feedback: an essential link between animal behaviour and disease
484 ecology. *Proc. Royal Soc. B* **283**, 20153078.

485 Fain, A. (2004). Mites (Acari) parasitic and predaceous on terrestrial gastropods. In *Natural*
486 *enemies of terrestrial molluscs*: 505–524. Barker, G.M. (Ed.). Wallingford, UK: CABI.

487 Furuta, E. & Yamaguchi, K. (2001). Haemolymph: blood cell morphology and function. In *The*
488 *biology of terrestrial molluscs*: 289–306. Barker, G.M. (Ed.). Wallingford, UK: CABI.

- 489 Gabry, J., Simpson, D., Vehtari, A., Betancourt, M. & Gelman, A. (2019). Visualization in Bayesian
490 workflow. *J. Royal Stat. Soc. A* **182**, 389–402.
- 491 Gérard, C., Ansart, A., Decanter, N., Martin, M.-C. & Dahirel, M. (2020). *Brachylaima* spp.
492 (Trematoda) parasitizing *Cornu aspersum* (Gastropoda) in France with potential risk of human
493 consumption. *Parasite* **27**, 15.
- 494 Giannelli, A., Cantacessi, C., Colella, V., Dantas-Torres, F. & Otranto, D. (2016). Gastropod-borne
495 helminths: a look at the snail–parasite interplay. *Trends Parasitol.* **32**, 255–264.
- 496 González-Santoyo, I. & Córdoba-Aguilar, A. (2012). Phenoloxidase: a key component of the
497 insect immune system. *Entomol. Exp. Appl.* **142**, 1–16.
- 498 Goodfriend, G.A. (1986). Variation in land-snail shell form and size and its causes: a review. *Syst.*
499 *Zool.* **35**, 204–223.
- 500 Gracenea, M. & Gállego, L. (2017). Brachylaimiasis: *Brachylaima* spp. (Digenea: Brachylaimidae)
501 metacercariae parasitizing the edible snail *Cornu aspersum* (Helicidae) in Spanish public
502 marketplaces and health-associated risk factors. *J. Parasitol.* **103**, 440–450.
- 503 Grannell, A., Cutler, J. & Rae, R. (2021). Size-susceptibility of *Cornu aspersum* exposed to the
504 malacopathogenic nematodes *Phasmarhabditis hermaphrodita* and *P. californica*. *Biocontrol Sci.*
505 *Technol.* **31**, 1149–1160.
- 506 Haeussler, E.M., Pizá, J., Schmera, D. & Baur, B. (2012). Intensity of parasitic mite infection
507 decreases with hibernation duration of the host snail. *Parasitology* **139**, 1038–1044.
- 508 Harrison, X.A. (2014). Using observation-level random effects to model overdispersion in count
509 data in ecology and evolution. *PeerJ* **2**, e616.
- 510 Jones, J.S., Leith, B.H. & Rawlings, P. (1977). Polymorphism in *Cepaea*: a problem with too many
511 solutions? *Annu. Rev. Ecol. Evol. Syst.* **8**, 109–143.
- 512 Kavaliers, M. (1992). Opioid systems, behavioral thermoregulation and shell polymorphism in
513 the land snail, *Cepaea nemoralis*. *J. Comp. Physiol. B* **162**, 172–178.

514 Kay, M. (2019). *tidybayes: tidy data and geoms for Bayesian models*.
515 <https://doi.org/10.5281/zenodo.1308151>

516 Kerstes, N.A.G., Breeschoten, T., Kalkman, V.J. & Schilthuizen, M. (2019). Snail shell colour
517 evolution in urban heat islands detected via citizen science. *Commun. Biol.* **2**, 264.

518 Köhler, H.-R., Capowiez, Y., Mazzia, C., Eckstein, H., Kaczmarek, N., Bilton, M.C., Burmester,
519 J.K.Y., Capowiez, L., Chueca, L.J., Favilli, L., Gomila, J.F., Manganelli, G., Mazzuca, S.,
520 Moreno-Rueda, G., Peschke, K., Piro, A., Cardona, J.Q., Sawallich, L., Staikou, A.E., Thomassen,
521 H.A. & Triebkorn, R. (2021). Experimental simulation of environmental warming selects against
522 pigmented morphs of land snails. *Ecol. Evol.* **11**, 1111–1130.

523 Lamotte, M. (1959). Polymorphism of natural populations of *Cepaea nemoralis*. *Cold Spring*
524 *Harb. Symp. Quant. Biol.* **24**, 65–86.

525 McElreath, R. (2020). *Statistical rethinking: a Bayesian course with examples in R and Stan*. 2nd
526 edition. Boca Raton, USA: Chapman and Hall/CRC.

527 McElroy, E.J. & de Buron, I. (2014). Host performance as a target of manipulation by parasites: a
528 meta-analysis. *J. Parasitol.* **100**, 399–410.

529 Meerburg, B.G., Singleton, G.R. & Kijlstra, A. (2009). Rodent-borne diseases and their risks for
530 public health. *Crit. Rev. Microbiol.* **35**, 221–270.

531 Morand, S., Wilson, M.J. & Glen, D.M. (2004). Nematodes (Nematoda) parasitic in terrestrial
532 molluscs. In *Natural enemies of terrestrial molluscs*: 525–557. Barker, G.M. (Ed.). Wallingford,
533 UK: CABI.

534 Morley, N.J. & Lewis, J.W. (2008). The influence of climatic conditions on long-term changes in
535 the helminth fauna of terrestrial molluscs and the implications for parasite transmission in
536 southern England. *J. Helminthol.* **82**, 325–335.

537 Nakagawa, S. & Schielzeth, H. (2010). Repeatability for Gaussian and non-Gaussian data: a
538 practical guide for biologists. *Biol. Rev.* **85**, 935–956.

539 O'Dwyer, K., Kamiya, T. & Poulin, R. (2014). Altered microhabitat use and movement of littorinid
540 gastropods: the effects of parasites. *Mar. Biol.* **161**, 437–445.

541 Özgo, M. (2009). Current problems in the research of *Cepaea* polymorphism. *Folia Malacol.* **16**,
542 55–60.

543 Özgo, M., Liew, T.-S., Webster, N.B. & Schilthuizen, M. (2017). Inferring microevolution from
544 museum collections and resampling: lessons learned from *Cepaea*. *PeerJ* **5**, e3938.

545 Pedersen, T.L. (2020). *patchwork: the composer of plots*. [https://CRAN.R-](https://CRAN.R-project.org/package=patchwork)
546 [project.org/package=patchwork](https://CRAN.R-project.org/package=patchwork)

547 Pieterse, A., Malan, A.P. & Ross, J.L. (2017). Nematodes that associate with terrestrial molluscs
548 as definitive hosts, including *Phasmarhabditis hermaphrodita* (Rhabditida: Rhabditidae) and its
549 development as a biological molluscicide. *J. Helminthol.* **91**, 517–527.

550 R Core Team. (2021). *R: a language and environment for statistical computing*. Vienna, Austria:
551 R Foundation for Statistical Computing.

552 Rae, R. (2017). The gastropod shell has been co-opted to kill parasitic nematodes. *Sci. Rep.* **7**,
553 4745.

554 Rantala, M.J. & Roff, D.A. (2005). An analysis of trade-offs in immune function, body size and
555 development time in the Mediterranean Field Cricket, *Gryllus bimaculatus*. *Funct. Ecol.* **19**, 323–
556 330.

557 Réale, D., Garant, D., Humphries, M.M., Bergeron, P., Careau, V. & Montiglio, P.-O. (2010).
558 Personality and the emergence of the pace-of-life syndrome concept at the population level.
559 *Phil. Trans. R. Soc. B* **365**, 4051–4063.

560 Rosin, Z.M., Kobak, J., Lesicki, A. & Tryjanowski, P. (2013). Differential shell strength of *Cepaea*
561 *nemoralis* colour morphs—implications for their anti-predator defence. *Naturwissenschaften*
562 **100**, 843–851.

- 563 Rosin, Z.M., Olborska, P., Surmacki, A. & Tryjanowski, P. (2011). Differences in predatory
564 pressure on terrestrial snails by birds and mammals. *J. Biosci.* **36**, 691–699.
- 565 Royauté, R., Berdal, M.A., Garrison, C.R. & Dochtermann, N.A. (2018). PACELESS life? A meta-
566 analysis of the pace-of-life syndrome hypothesis. *Behav. Ecol. Sociobiol.* **72**, 64.
- 567 Russo, J. & Madec, L. (2013). Linking immune patterns and life history shows two distinct
568 defense strategies in land snails (Gastropoda, Pulmonata). *Physiol. Biochem. Zool.* **86**, 193–204.
- 569 Saeedizadeh, A. & Niasti, F. (2020). Response of grey slug to entomopathogenic nematodes.
570 *Bragantia* **79**, 447–456.
- 571 San-Jose, L.M. & Roulin, A. (2018). Toward understanding the repeated occurrence of
572 associations between melanin-based coloration and multiple phenotypes. *Am. Nat.* **192**, 111–
573 130.
- 574 Scheil, A.E., Hilsmann, S., Tribskorn, R. & Köhler, H.-R. (2013). Shell colour polymorphism,
575 injuries and immune defense in three helicid snail species, *Cepaea hortensis*, *Theba pisana* and
576 *Cornu aspersum maximum*. *Results Immunol.* **3**, 73–78.
- 577 Scheil, A.E., Hilsmann, S., Tribskorn, R. & Köhler, H.-R. (2014). Shell colouration and parasite
578 tolerance in two helicoid snail species. *J. Invertebr. Pathol.* **117**, 1–8.
- 579 Schilthuizen, M. (2013). Rapid, habitat-related evolution of land snail colour morphs on
580 reclaimed land. *Heredity* **110**, 247–252.
- 581 Schüpbach, H.U. & Baur, B. (2008a). Parasitic mites influence fitness components of their host,
582 the land snail *Arianta arbustorum*. *Invertebr. Biol.* **127**, 350–356.
- 583 Schüpbach, H.U. & Baur, B. (2008b). Experimental evidence for a new transmission route in a
584 parasitic mite and its mucus-dependent orientation towards the host snail. *Parasitology* **135**,
585 1679–1684.
- 586 Segade, P., Crespo, C., García, N., García-Estévez, J.M., Arias, C. & Iglesias, R. (2011).
587 *Brachylaima aspersae* n. sp. (Digenea: Brachylaimidae) infecting farmed snails in NW Spain:

588 Morphology, life cycle, pathology, and implications for heliculture. *Vet. Parasitol.* **175**, 273–
589 286.

590 Segade, P., García-Estévez, J., Arias, C. & Iglesias, R. (2013). Parasitic infections in mixed system-
591 based heliculture farms: dynamics and key epidemiological factors. *Parasitology* **140**, 482–497.

592 Silvertown, J., Cook, L., Cameron, R., Dodd, M., McConway, K., Worthington, J., Skelton, P.,
593 Anton, C., Bossdorf, O., Baur, B., Schilthuizen, M., Fontaine, B., Sattmann, H., Bertorelle, G.,
594 Correia, M., Oliveira, C., Pokryszko, B., Ożgo, M., Stalažs, A., Gill, E., Rammul, Ü., Sólymos, P.,
595 Féher, Z. & Juan, X. (2011). Citizen science reveals unexpected continental-scale evolutionary
596 change in a model organism. *PLOS ONE* **6**, e18927.

597 Sitko, J. & Heneberg, P. (2020). Emerging helminthiasis of song thrush (*Turdus philomelos*) in
598 Central Europe. *Parasitol Res.* **119**, 4123–4134.

599 Solomon, A., Paperna, I. & Matkovic, A. (1996). The influence of aestivation in land snails on
600 the larval development of *Muellerius cf. capillaris* (Metastrongyloidea: Protostrongylidae). *Int. J.*
601 *Parasitol.* **26**, 363–367.

602 Sudhaus, W. (2018). Dispersion of nematodes (Rhabditida) in the guts of slugs and snails. *Soil*
603 *Org.* **90**, 101–114.

604 Surmacki, A., Ożarowska-Nowicka, A. & Rosin, Z.M. (2013). Color polymorphism in a land snail
605 *Cepaea nemoralis* (Pulmonata: Helicidae) as viewed by potential avian predators.
606 *Naturwissenschaften* **100**, 533–540.

607 Tan, K., Zhang, H., Lim, L.-S., Ma, H., Li, S. & Zheng, H. (2020). Roles of carotenoids in
608 invertebrate immunology. *Front. Immunol.* **10**.

609 Vehtari, A., Gelman, A., Simpson, D., Carpenter, B. & Bürkner, P.-C. (2021). Rank-normalization,
610 folding, and localization: an improved \hat{R} for assessing convergence of MCMC (with discussion).
611 *Bayesian Anal.* **16**, 667–718.

612 Wickham, H., Averick, M., Bryan, J., Chang, W., McGowan, L., François, R., Grolemund, G.,
613 Hayes, A., Henry, L., Hester, J., Kuhn, M., Pedersen, T., Miller, E., Bache, S., Müller, K., Ooms, J.,
614 Robinson, D., Seidel, D., Spinu, V., Takahashi, K., Vaughan, D., Wilke, C., Woo, K. & Yutani, H.
615 (2019). Welcome to the Tidyverse. *J. Open Source Softw.* **4**, 1686.

616 Williams, A. & Rae, R. (2016). *Cepaea nemoralis* (Linnaeus, 1758) uses its shell as a defence
617 mechanism to trap and kill parasitic nematodes. *J. Molluscan Stud.* **82**, 349–350.

618 Williams, S.T. (2017). Molluscan shell colour. *Biol. Rev.* **92**, 1039–1058.

619 Wilson, M.J. (2012). Pathogens and parasites of terrestrial molluscs. In *Manual of techniques in*
620 *invertebrate pathology*: 427–439. Lacey, L.A. (Ed.). Oxford; New York: Academic Press.

621 Wilson, M.J., Hughes, L.A., Hamacher, G.M. & Glen, D.M. (2000). Effects of *Phasmarhabditis*
622 *hermaphrodita* on non-target molluscs. *Pest. Manag. Sci.* **56**, 711–716.

623 Wynne, R., Morris, A. & Rae, R. (2016). Behavioural avoidance by slugs and snails of the parasitic
624 nematode *Phasmarhabditis hermaphrodita*. *Biocontrol Sci. Technol.* **26**, 1129–1138.

625 Żbikowska, E., Marszewska, A., Cichy, A., Templin, J., Smorağ, A. & Strzała, T. (2020). *Cepaea*
626 *spp.* as a source of *Brachylaima mesostoma* (Digenea: Brachylaimidae) and *Brachylecithum sp.*
627 (Digenea: Dicrocoeliidae) larvae in Poland. *Parasitol. Res.* **119**, 145–152.

628

629 **Figure Legends**

630 **Figure 1.** Snail shell diameter as a function of band number and population of origin. Grey dots
631 are observed values, displayed along the posterior distributions of the average sizes (black dots
632 and segments are the means and 95% CI of the posteriors).

633 **Figure 2.** Examples of encapsulated parasites in *Cepaea nemoralis* shells. (A) mite; (B)
634 nematode. Original images, including other images of encapsulated parasites, are made
635 available with the rest of the data (see **Data availability**).

636 **Figure 3.** Parasite-related variables as a function of morph and population of origin ($N = 179$,
637 one dead snail was excluded). (A) Proportion of snails having at least one nematode trapped in
638 their shells; (B) Live nematode abundance; (C) Live *Brachylaima* sp. trematode abundance
639 (metacercariae). For (B) and (C): grey dots are observed values, displayed along the posterior
640 distributions of the average abundances based on the model described **Table 1** (black dots and
641 segments are the means and 95% CI of the posteriors). Posterior predictions are based on fixed
642 effects only, and the effects of shell size are averaged out. See **Supporting Information S3** for
643 pairwise comparisons.

644 **Figure 4.** Behaviours as a function of morph and population of origin. (A) Movement metric
645 ($1/\text{latency to move beyond 10 cm}$, centered and scaled to unit 1SD); (B) Food intake. Grey dots
646 are observed values, displayed along the posterior distributions of the average abundances
647 based on the model described in **Table 1** (black dots and segments are the means and 95% CI of
648 the posteriors). Observed values from the same individual are connected by a line. Posterior
649 predictions are based on fixed effects only, and the effects of shell size are averaged out. See
650 **Supporting Information S3** for pairwise comparisons.

651

652

653 **Tables**

654 **Table 1.** Summary of posterior model coefficients for the parasite and behaviour multivariate model.
 655 Individual-level correlations between traits are presented in **Table 2**. See **Methods** for model details, and
 656 **Supporting Information S3** for pairwise comparisons among morph × population groups. Fixed effects of
 657 population and size that are different from 0, based on 95% intervals are in bold.

	Nematode abundance*	Trematode abundance	Movement	Food intake
Fixed effects				
Intercept _[0 bands]	0.84 [0.09; 1.52]	-0.94 [-2.44; 0.48]	-0.26 [-0.66; 0.16]	-0.22 [-0.61; 0.19]
Intercept _[3 bands]	0.67 [-0.08; 1.38]	-0.10 [-1.54; 1.33]	0.16 [-0.24; 0.56]	-0.09 [-0.50; 0.31]
Intercept _[5 bands]	-0.47 [-1.31; 0.32]	-1.26 [-2.72; 0.23]	-0.09 [-0.50; 0.31]	-0.04 [-0.45; 0.37]
Population = “shaded” _[0 bands]	—	-0.02 [-1.42; 1.39]	0.71 [0.31; 1.08]	0.42 [0.04; 0.83]
Population = “shaded” _[3 bands]	—	0.06 [-1.31; 1.34]	-0.23 [-0.61; 0.13]	0.22 [-0.18; 0.63]
Population = “shaded” _[5 bands]	—	-0.88 [-2.44; 0.66]	-0.08 [-0.47; 0.32]	0.13 [-0.26; 0.55]
Shell size (standardised)	-0.19 [-0.53; 0.13]	0.06 [-0.57; 0.72]	0.09 [-0.03; 0.21]	0.23 [0.10; 0.35]
Random effects SDs				
Test group SD	0.53 [0.00; 1.23]	1.95 [0.94; 3.19]	0.23 [0.00; 0.70]	0.21 [0.00; 0.67]
Individual-level SD	1.14 [0.85; 1.46]	2.57 [1.99; 3.21]	0.32 [0.04; 0.54]	0.56 [0.43; 0.70]
Residual SD	—	—	0.92 [0.82; 1.02]	0.8 [0.72; 0.88]

658 * The nematode model is only estimated for the “sun-exposed” population, since no live nematodes
 659 were recorded in the “shaded” population

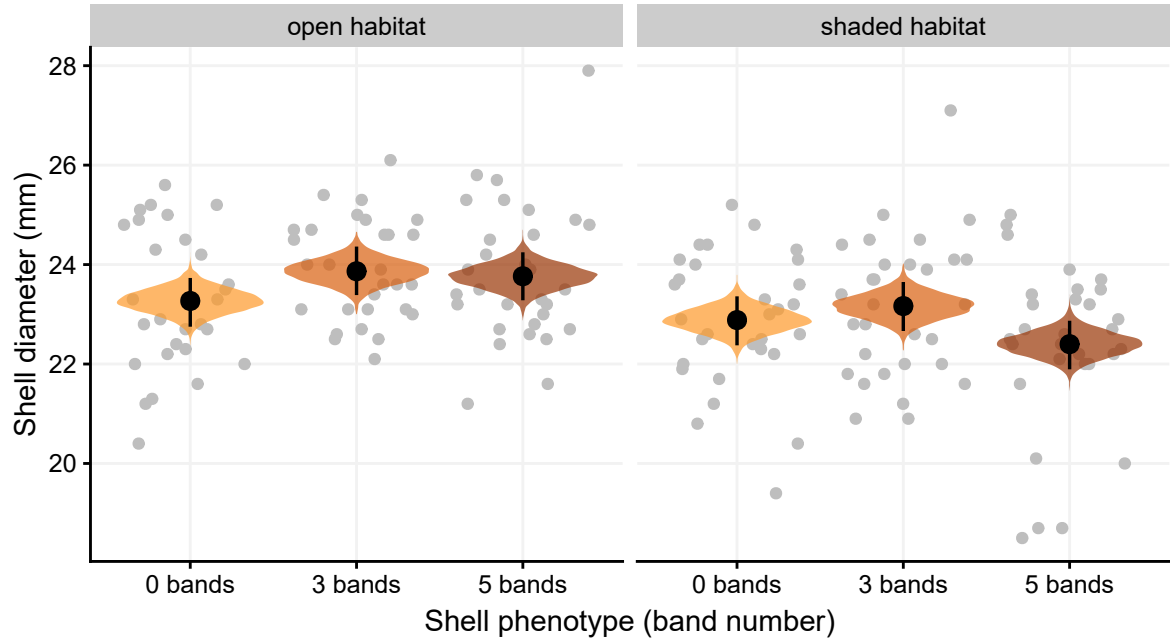
660

661

662 **Table 2.** Individual-level correlations between parasite abundances and behavioural traits. Correlations
663 involving nematode abundance are only based on individuals from the open population (see Methods);
664 results are not qualitatively different if the model is refitted to include all observations (see **Data**
665 **availability**). Correlations that are different from 0 (based on 95% intervals) are in bold.

	Nematode abundance	Trematode abundance	Movement
Trematode abundance	0.01 [-0.35; 0.37]	--	--
Movement	-0.24 [-0.73; 0.25]	-0.17 [-0.68; 0.28]	--
Food intake	-0.60 [-0.87; -0.30]	-0.12 [-0.46; 0.19]	0.24 [-0.23; 0.71]

666



A



200 μm

B



200 μm

0.25 mm

