

Novel Chronic Anaplasmosis in Splenectomized Patient, Amazon Rainforest

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1 A novel agent of human anaplasmosis in Amazon rainforest

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27 Abstract

- We report a case of unusual human anaplasmosis from remote rainforest areas of French
- Guiana. Molecular typing showed that the pathogen is a novel Anaplasma species, different to
- 30 all known species, and more genetically related to recently described *Anaplasma* spp. causing
- 31 infections in rainforest wild fauna of Brazil.

Anaplasma genus. Eight *Anaplasma* species and several candidate species are described, and at least five species can be transmitted to humans (1,2). Of particular concern, the agent of human granulocytic anaplasmosis, *A. phagocytophilum*, has a specific tropism to polymorphonuclear neutrophils (1,3). Another species, *A. capra*, recently described from asymptomatic goats, is now recognized as an agent of human intraerythrocytic anaplasmosis in China (4). The three other species detected in humans are major veterinary agents sporadically identified in few patients worldwide: *A. ovis* and *A. bovis* in erythrocytes, and *A. platys* in platelets (1,5). Human anaplasmosis are consistently associated with people living in rural areas in habitats favorable to ticks or working closely with domestic animals (1,6). However, recent surveys report the presence of overlooked *Anaplasma* species of undetermined zoonotic potential in wild fauna (1,2).

The study

We assess the presence of *Anaplasma* in blood samples of clandestine gold miners working in rainforest of French Guiana, South America. This 83,000 km² territory, located between Suriname and Brazil, is one of the regions of highest biodiversity in the world, with over 280 species of wild mammals (7). Human population of French Guiana (284,000 inhabitants) is concentrated principally in a handful of towns spread along the coastline and the two main rivers (8). The interior is largely uninhabited and covered by dense rainforest where illegal gold mining camps are located (9,10).

We examined 363 archived DNA extracts obtained from human blood samples. We had primarily collected these samples in 2019 as part of a malaria survey in remote mining camps in French Guiana (Malakit) (See methodology in (11)). To characterize the whole bacterial

57 diversity, DNA blood samples were typed using a high-throughput bacterial 16S rDNA (rrs) 58 sequencing approach (bacterial barcoding, as described in (12)). Bacteria were characterized 59 as operational taxonomic unit (OTU) and amplicon sequence variants (ASV), and 60 taxonomically assigned using the Silva database (https://www.arb-silva.de/). 61 62 Examination of OTUs and ASVs revealed the presence of *Anaplasma* sequences in one DNA 63 sample. No OTU and ASV assigned to the Anaplasma genus or to the Anaplasmataceae 64 family was detected in the 362 other samples. We further conducted two independent 65 Anaplasma-specific PCR assays targeting a region of the 16S rDNA gene (544 bp) and the 23S-5S (ITS2) intergenic region (423 bp) following Calchi 2020 (13), and obtained 66 67 amplicons of correct sizes for the positive sample. The Sanger sequencing of amplicons obtained with each pair of primers each confirmed the presence of *Anaplasma*. These 68 sequences have been deposited on GenBank (accession numbers: ///pending///). 69 70 71 We used BLAST (http://www.ncbi.nlm.nih.gov/blast/Blast.cgi) to compare these 16S rDNA 72 and ITS2 nucleotide sequences with Anaplasma sequences available in GenBank. None of the 73 nucleotide sequences observed in this study is 100% identical to known Anaplasma 74 sequences. The 16S rRNA sequence showed highest identities with Anaplasma found in wild fauna of Brazil, including an Anaplasma sp. detected in Amblyomma coelebs ticks collected 75 76 on South American coatis, Nasua nasua (99.8%; GenBank MT019560), another Anaplasma 77 sp. of black rats, Rattus rattus (99.8%; GenBank KY391803), and Candidatus Anaplasma 78 amazonensis (13) of brown-throated sloths, *Bradypus variegatus*, and of two-toed sloths, 79 Choloepus didactylus (99.1%; GenBankMT199827). All other Anaplasma species showed 80 identities <99%. The ITS2 sequences showed highest nucleotide identity with Cand. 81 Anaplasma amazonensis of Brazilian brown-throated sloths and two-toed sloths (96.8%;

82 GenBankMT267354), and lower identities with other *Anaplasma* species or strains (<92%). 83 On account of these distinct genetic traits, we propose the designation Cand. Anaplasma 84 sparouinii for this novel bacterium. The specific name refers to the Sparouine River where the 85 infected patient lived. 86 87 We conducted phylogenetic analyses based on these 16S rDNA and ITS2 nucleotide sequences using the maximum-likelihood (ML) method. We obtained trees of similar 88 89 topologies with a robust clustering of Cand. Anaplasma sparouinii with some Anaplasma 90 associated with Brazilian wild fauna: Cand. Anaplasma sparouinii is phylogenetically more 91 related to Anaplasma sp. infections detected in ticks of coatis, black rats and, at a lesser 92 extent, to Cand. Anaplasma amazonensis of sloths (Figures 1A and B). Altogether, they 93 delineate a clade of neotropical Anaplasma divergent to all other Anaplasma species (Figures 94 1A and B). 95 96 The DNA sample positive for *Cand*. Anaplasma sparouinii was from a 58-years-old male who had a history of post-traumatic splenectomy and malaria attacks caused by *Plasmodium vivax*. 97 98 This patient originated from Maranhão, Brazil, but has been working exclusively in rainforest 99 of French Guiana from the past three years. The Sparouine anaplasmosis was retrospectively 100 diagnosed in September 2021 on the basis of PCR survey of previous blood samples (October 101 2019 and May 2021) and blood smears (October 2019) as further detailed. 102 103 We primarily detected the presence of *Cand*. Anaplasma sparouinii in a blood sample 104 collected in October 2019. At this time, the patient was asymptomatic, with no fever, normal

blood pressure, negative tests for agents of diseases usually tested in French Guiana

(serologies for yellow fever, Q fever, hepatitis B and C, HIV, syphilis, molecular tests for

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malaria and leptospirosis). He showed anemia with a hemoglobin level of 10.5 g/dL. The reexamination of Giemsa-stained thin blood film taken for malaria diagnosis at that time revealed the presence of intraerythrocytic bodies, which could be *Cand*. Anaplasma sparouinii. No infection was detected in granulocytes and platelets, but around one third of erythrocytes harbored one or two small, round dark purple inclusions located at their periphery and which could be *Anaplasma* (Figures 2A and B). We also detected the presence of Howell–Jolly bodies in erythrocytes (Figure 2B), which could be a consequence of splenectomy.

Eighteen months later (May 2021), the patient was admitted in the Cayenne Hospital Center with fever, myalgia, headache, epistaxis and severe anemia (Hb = 6.6 g/dL). A broad microbiological investigation ruled out covid-19, dengue, chikungunya, zika, influenza, malaria, HIV, HBV, HCV, and leptospirosis. The only positive test was a subnormal level of anti-*Coxiella burnetii* IgM and IgG leading to the introduction of antibiotic treatment (doxycycline 100 mg twice a day for 21 days and ceftriaxone 1 g per day for 5 days). The anemia was considered autoimmune hemolytic due to a positive Coombs test and was thus treated with prednisolone with decreasing doses from 60 mg/d to 10 mg/d for 3 months. The patient recovered within three weeks with resolution of symptoms and an improvement of the hemoglobin level to 9.4 g/dL at discharge. Our a posteriori *Anaplasma* PCR survey of May 2021 blood samples (prior to and at Day 7 of the antibiotic treatment) again revealed the presence of *Cand*. Anaplasma sparouinii. No further blood sample was preserved thus the disappearance of the *Anaplasma* infection at the end of the antibiotic treatment could not be ascertained.

Conclusions

We characterize here *Cand*. Anaplasma sparouinii as a novel human intraerythrocytic pathogen. The infection arose over at least 18 months in a patient living in rainforest and potentially more susceptible due to a previous splenectomy. The phylogenetic proximity of *Cand*. Anaplasma sparouinii with other *Anaplasma* associated with Amazonian wild mammals highlights that a genetic cluster of *Anaplasma* is circulating in French Guiana and Brazil. While these neotropical *Anaplasma* species are different to species currently recognized as human pathogens, they may represent a source of novel infections to humans in Amazon rainforest.

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Figure 1. *Anaplasma* phylogenies constructed using maximum-likelihood (ML) estimations based on (**A**) 16S rDNA (485 unambiguously aligned bp; best-fit approximation for the evolutionary model: HKY+I) and (**B**) ITS2 sequences (387unambiguously aligned bp; best-fit approximation for the evolutionary model: HKY+I) including *Cand*. Anaplasma sparouinii characterized in this study. GenBank accession numbers of sequences used in analyses are shown on the phylogenetic trees. Numbers at nodes indicate percentage support of 1,000 bootstrap replicates.

Figure 2. Thin blood films showing inclusions of *Cand*. Anaplasma sparouinii located at the periphery of the red blood cells as small round dots of $0.3-0.4 \,\mu m$ (arrow heads). Other red blood cells contain Howell-Joly bodies of various shapes and sizes >1 μm (arrows). Some Howell-Joly bodies are found in the background of the smear (Wright Giemsa stain, x100).