

$\delta 180$ compound-specific stable isotope assessment: An advanced analytical strategy for sophisticated adulterations detection in essential oils - Application to spearmint, cinnamon, and bitter almond essential oils authentication

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- δ δ compound-specific stable isotope assessment : an advanced analytical
- 2 strategy for sophisticated adulterations detection in essential oils -
- 3 Application to spearmint, cinnamon, and bitter almond essential oils
- 4 authentication.

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<u>Abstract</u>

Natural plant extracts are primarily used as raw materials in the cosmetic and perfumery industry. However, adulterations with petrochemical products are occurring in the market, leading to non-100% natural products. Several analytical techniques such as impurity detection or enantioselective ratio assessments have been previously described as good indicators to detect any addition of synthetic products, but these techniques are ineffective with novel type of synthetic pathways such as semisynthesis. In order to improve authentication, development of advanced analytical strategies such as δ^{18} O stable isotopic ratios assessment was tested on spearmint, cinnamon and bitter almond essential oils major metabolites (carvone, (E)-cinnamaldehyde, and benzaldehyde). Natural δ^{18} O mean values (δ^{18} OCarvone = 18.4%; δ^{18} OCinnamaldehyde = 13.9%; δ^{18} OBenzaldehyde = 16.5%) were found to be 29 higher than semisynthetic origin for the 3 studied molecules (δ^{18} OCarvone = 9.2%; 30 δ^{18} OCinnamaldehyde = 8.8%; δ^{18} OBenzaldehyde = 10.9%). These measurements proved to be efficient to discriminate natural and semisynthetic origins of these components and therefore potentially lead to a novel way to authenticate natural products.

Keywords

- 36 Semisynthesis; Essential oil; Gas chromatography; Isotope ratio mass spectrometry;
- 37 Authentication

1. Introduction

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Natural aromatic raw materials market is growing exponentially nowadays to supply food industry, perfumery and cosmetics. To satisfy this increasing demand, researches have been focused on compounds issued from biotechnology processes used to produce aromas and fragrances from other plant origins. Unfortunately, these new technologies can lead to new sophisticated frauds. The adulterations found on the natural product market are constantly evolving and it is nowadays possible to observe additions of biotechnological origin molecules. These new processes have the capability to create seemingly natural molecules such as the vanillin, which has been regularly obtained from the ferulic acid contained in rice [1], or the frambinone obtained from 4-hydroxybenzaldehyde contained in Escherichia coli [2,3]. In the regulation field of essential oils, molecules must come from the raw material of the plant reference to be considered and labelled 100% pure and natural. Molecules produced from natural reagents, also called semisynthetic compounds, are suspected of being used to adulterate specific essential oils. Several examples are cited in the literature. Cinnamaldehyde molecules can be made from bitter almond benzaldehyde and used to adulterate an essential oil of cinnamon [4]. Greule et al. reported the use of carvacrol, produced from the carvone of caraway essential oil, to adulterate an essential oil of oregano [5]. Semisyntheses enable fraudsters to bypass many analytical controls. An essential oil that has undergone an addition of a semisynthetic molecule will be more difficult to detect than traditional synthetic molecule additions with usual analytical methods of authentication. One of the most common analysis is the determination of enantiomeric ratios to control chiral molecules such as linalool, because synthetic chiral

molecules are produced in racemic proportion, whereas natural molecules are generally stereospecific [6]. However, enantiomeric analyses will not be effective in the case of semisyntheses because the stereochemistry will be preserved. Another common analytical technique is the determination of radiocarbon activity, which determine the presence of fossil carbon in the sample [7]. However, a semisynthesis will eventually conduct to a 100% bio based carbon molecule because no petrochemicals would be used. Finally, stable isotope analyses of carbon and hydrogen will generally be inefficient to demonstrate this addition, because the transformations carried out will not generate significant isotopic variations of δ^{13} C and δ^{2} H ratios. Typical examples are the conversion of limonene into carvone or benzaldehyde into cinnamaldehyde. However, several analytical strategies are possible. A first possibility is the detection of residual impurities from semisyntheses. This approach has numerous flaws, as many synthetic pathways do not produce any impurities, and these residual molecules can be removed with a purification process [7]. A second analytical approach is the Site-Specific Natural Isotope Fractionation-Nuclear Magnetic Resonance (SNIF-NMR) technique. This system allows the measurement of isotopic fractionations of specific sites of a molecule, which can lead to the origin of the product [8]. Several authentication studies have been

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sample for analysis.

conducted using SNIF-NMR [4,9,10]. However, this analytical tool requires that the analysed molecules are almost entirely pure, which is difficult to achieve on complex matrices such as essential oils. In addition, there is a risk of isotopic fractionation during the purification step. Since SNIF-NMR is a relatively insensitive technique, it also requires using a large amount of

Analysis of oxygen stable isotopes can be an interesting approach. In many cases of semisynthesis of oxygenated terpenes or phenylpropanoids, the synthesis is characterized by the addition of an external oxygen atom from the original molecule. The δ^{18} O ratios can therefore be different, and it seems relevant to measure them. To determine this isotope ratio, the use of a GC-P-IRMS instrument is necessary to be used on most of the essential oils. This analytical system to operate is rarely used in authentication on natural products due to difficulties in the analytical process.

This article describes the development of the δ^{18} O analysis by GC-P-IRMS to highlight the additions of semisynthetic-origin compounds (carvone, (E)-cinnamaldehyde, and benzaldehyde) in essential oils of spearmint, cinnamon and bitter almond.

2. Material and methods

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2.1. Materials

2.1.1. Essential oils

Forty-eight samples of spearmint Mentha spicata var crispa, from India, USA, China, and France were analysed for this study. From these samples, 11 certified 100% pure and natural samples were provided by Albert Vieille SAS (Vallauris, France) and Young Living (Lehi, Utah, USA), and one sample was obtained from hydrodistillation process from fresh spearmint leaves at the Institute of Analytical Sciences (ISA Lyon, UMR5280 - CNRS, Villeurbanne, France) laboratory. Other essential oils (n=36) were commercial samples, obtained from various sources. Fifty-five samples of cinnamon essential oils were analysed for this study. These samples were obtained from various botanical and geographical origins: Cinnamomum verum essential oils were sourced from Sri Lanka and Madagascar, Cinnamomum cassia essential oils were sourced from China and Cinnamomum burmanii (Nees & T.Nees) Blume (Indonesia) came from Indonesia. Authentic samples (n=11) were provided by Albert Vieille SAS, Astérale (Madagascar), and Young Living. Moreover, homemade distillations of cinnamon bark were realized in the ISA Lyon laboratory to obtain cinnamon essential oils from several botanical origins. Other samples were commercial essential oils (n=44). In addition, 12 pure and natural samples of bitter almond essential oils, obtained from steam

distillation apricots almonds *Prunus armeniaca* L. have also been analysed. These essential oils were provided by Albert Vieille SAS and are originated from Morocco. For information, these samples were subjected to a cyanhydric acid disposal process prior to analysis.

2.1.2. Commercial molecules

Commercial molecules of carvone (R) (n=3), (E)-cinnamaldehyde (n=1), and benzaldehyde (n=3) were purchased from TCI (Tokyo, Japan), Alfa Aesar (Ward Hill, MA, USA) and Sigma Aldrich (Saint Louis, MO, USA).

Several carvone molecules were obtained from limonene conversion. The synthesis of the

2.1.3. Syntheses

is then obtained by steam distillation.

2.1.3.1. Synthesis of carvone (R) from limonene (S)

carvone has been carried out in 3 steps based on Rothenberger et al. protocol [11] (Figure 1). The first step leads to a first intermediate molecule: the limonene nitrosochloride (1). The second step leads to the synthesis of carvoxime (2). This compound is then used to produce the carvone (3).

In order to perform this synthesis, the limonene is firstly diluted in isopropanol at a temperature comprised between 0°C and 10°C. A solution of 37% chloridric acid and isopropanol is then gradually added to the mixture. The obtained solution is then mixed with NaNO₂ in deionised water. Limonene nitrosochloride product is then purified using Buchner vacuum filtration and traces of water were removed using a vacuum desiccator. This product is then dissolve in dimethylformamide and isopropanol. The mixture is heated to reflux. Carvoxime product is then extract using diethyl ether and the organic phase is washed with ultra-pure water until neutral pH. The solution is then dried using anhydrous sodium sulfate, then filtered. Diethyl ether is then evaporated and the carvoxime is dissolved in an aqueous solution of oxalic acid (5%), then heated at reflux for 30 minutes. The carvone final product

2.1.3.2. Aldol reaction of benzaldehyde

Ten samples of (E)-cinnamaldehyde were produced in laboratory using aldolisation of benzaldehyde from synthesis origin and natural origin from essential oils of bitter almond (Figure 2). The synthesis protocol is described below.

A volume of 600 μ L of benzaldehyde was diluted in 3 mL of ethanol 96%. The mixture was maintained at 0°C. 1 mL of NaOH 15% was ten added and the solution was stirred for 5 minutes. A volume of 200 μ L of acetaldehyde (TCI and Sigma Aldrich) was then added at 0°C. This step is renewed four times every 15 minutes to make sure that the acetaldehyde is maintained in excess during the reaction entire time. The mixture is then heated at 40°C during 10 minutes. Highly concentrated hydrochloric acid is then added to acid pH. Diethyl ether extraction was then carried out to collect the (E)-cinnamaldehyde final product.

2.1.3.3. Retro aldol reaction of cinnamaldehyde

Five samples of benzaldehyde have been produced in laboratory by retro aldol reaction of synthetic origin (E)-cinnamaldehyde (Fluka) and natural origin (E)-cinnamaldehyde from essential oils of *Cinnamomum verum*, *Cinnamomum cassia*, and *Cinnamomum burmanii*.

Ten grams of (E)-cinnamaldehyde are firstly diluted in an aqueous solution of sodium carbonate Na_2CO_3 (3%). This mixture was heated at reflux during 7 hours. A steam distillation was then carried out to extract the benzaldehyde.

2.2. Isotopic Analyses

2.2.1. Data Expression

165 Isotopic results are expressed in delta (δ) units according to the following formula [30]:

$$\delta^{i}E = \frac{({}^{i}R_{Sample} - {}^{i}R_{Reference})}{{}^{i}R_{Reference}}$$

where i is the mass number of the heavy isotope of element E, R_{Sample} is the isotope ratio of the sample, and $R_{Reference}$ is the isotope ratio of a relevant internationally recognized reference material. The δ values were multiplied by 1000 and are expressed in units "per mil" (‰).

High-purity gases of carbon dioxide (99.998%, 4.8 Messer), hydrogen (99.999%, 5.0 Messer), and carbon monoxide (99.997%, 4.7 Messer) were used as the working reference gases injected with each sample. Helium (99.999% purity) was used as a carrier gas.

All studied metabolites in the essential oils of interest were calibrated with commercial molecules (carvone, (E)-cinnamaldehyde, and benzaldehyde) previously calibrated against international standards in elemental analysis – IRMS (EA-IRMS).

International reference materials IAEA CH7 (polyethylene foil: δ^{13} C = -32.15‰, δ^2 H = -100‰), IAEA 601 (benzoic acid: δ^{18} O = 23.3‰), and USGS40 (L-glutamic acid: δ^{13} C = -26.39‰), supplied by the International Agency of Atomic Energy (IAEA, Vienna, Austria), were used to calibrate each molecule in the mixture, which allowed us to express our δ^{13} C results relative to the Vienna Pee Dee Belemnite (VPDB) scale and our δ^2 H and δ^{18} O results relative to the Standard Mean Ocean Water (VSMOW) scale. Each sample was analyzed versus a monitoring gas for which two pulses were admitted with each analyte. All sample values were analyzed in duplicate and verified by bracketing the analytical sequence with

international reference materials every six analyses. δ^{13} C isotopic analyses of the standards molecules were conducted using a Flash HT elemental analyzer connected to a Delta V Plus via a Conflo V interface. The δ^2 H and δ^{18} O measurements were carried out using a TC-EA elemental analyzer linked to a Delta V Plus via a Conflo V interface (all Thermo Fisher Scientific). The method is fully explained in a previous study [7].

2.2.2. Gas Chromatography - Mass Spectrometry - Combustion/Pyrolysis Isotope Ratio Mass Spectrometry (GC-MS-C/P-IRMS)

The δ^{13} C, δ^2 H, and δ^{18} O values of targeted spearmint, bitter almond, and cinnamon essential oils metabolites were determined by GC-IRMS using a GC Trace 1310 simultaneously connected to an ISQ MS quadrupole mass spectrometer and to a Delta V Plus isotope ratio mass spectrometer via a GC Isolink II interface. (Thermo Fisher, Bremen, Germany). This equipment enabled the identification of compounds via the ISQ MS instrument and specific compound isotopic ratio measurements of δ^{13} C, δ^2 H, and δ^{18} O via IRMS.

One microliter of diluted essential oil (1 μ L into 100 μ L of ethanol) was injected using a Tri Plus RSH autosampler in a Trace 1310 gas chromatograph equipped with an Agilent HP-1 capillary column (50 m * 0.320 mm * 0.52 μ m internal diameter film thickness) used for chromatographic separation. The injection port was held at 260°C, fitted with a split liner containing glass wool and operated in split mode (10:1). The oven temperature program started at 80°C for 2 min, increased to 150°C at a rate of 10°C/min, increased to 250°C at a rate of 20°C/min, increased to 325°C at a rate of 20°C/min. Components were separated at a flow rate of 1.1 mL/min and divided in 2 ways using a Sil Flow connector.

 δ^{13} C analyses requires to perform the combustion of carbon in carbon dioxide. It was achieved using an alumina tube containing nickel oxide maintained at 1000°C in a furnace.

Pyrolysis of the sample for δ^2 H measurement in H₂ was carried out at a high temperature (1400°C) using an alumina conversion tube (Thermo Fischer Scientific). Undecane, $C_{11}H_{24}$ (Sigma Aldrich), was regularly added to regenerate the alumina pyrolysis tube. For $\delta^{18}O$ measurements, it was necessary to ensure the conversion of oxygen atoms into carbon monoxide. The pyrolysis unit was designed by an inner platinum tub partly filled with nickel wires incorporated in an outer ceramic tube (Thermo Fisher) hold at 1280°C and work under a column flow 1.1 ml/min Helium added by 1.1 ml/min of a mix (98% Helium +2% H₂) necessary in this configuration. Pyrolysis tube was conditioned by injecting 1 μ l of high purity undecane paraffin (sigma Aldrich) in duplicate, allowing to create an inner homogenous coating into the platinum tube. This layer of carbon created would be necessary bring enough carbon reactant in the pyrolysis reaction and the production of carbon monoxide. This coating process has been monitored every week enabling to reconstitute the carbon layer.

2.3. Enantiomeric study and impurity detection

2.3.1. Carvone enantioselective analysis using GC-FID

Samples of spearmint essential oils have been analysed in Enantio GC-FID to control the enantioselective ratio of carvone. Enantioselective analyses were performed on a gas chromatograph 6890N GC System (Agilent Technologies) equipped with a flame ionization detector (FID) with the detector temperature set at 250°C. Separation of the carvone enantiomers were performed on a β -DEX 225 column (30 m * 0,250 mm * 0,25 μ m, Supelco Inc.) coated with cyclodextrin stationary phase. The GC conditions were programmed as follows: 0.2 μ L of diluted essential oil (1:100 in ethanol) was injected into the GC in split mode (100:1) with the injector temperature set at 250°C. The carrier gas was H₂ at a constant flow rate of 1.2 mL/min. The initial oven temperature was held at 80°C for 5 min,

ramped to 130°C at a rate of 2°C/min, then ramped to 180°C at a rate of 5°C/min and held for 2min.

2.3.2. Impurity detection using GC-MS analysis

GC–MS analyses were performed on a gas chromatograph 6890 N GC System (Agilent Technologies) interfaced with a 5975 inert XL MSD mass spectrometer with electron impact ionization (70 eV). 1 μL of pure cinnamon essential oil was injected into the GC in the split mode (100:1), with the injector temperature set at 250 °C. Helium was used as the carrier gas with a constant flow rate of 1.0 mL/min. Separation of the volatile compounds in cinnamon essential oils was achieved using a HP-5 capillary column (30 m*0.250 mm*0.25 μm, Agilent Technologies). The initial oven temperature was held at 60 °C for 3 min and then ramped at a rate of 5°C/min to reach 160°C, then ramped to 300°C at a rate of 10°C/min and held for 4 min. The SIM parameters were as follows: the targeted mass ions (m/z) were 128, 129 and 158 (molecular ion) to detect 5-phenylpenta-2,4-dienal. NIST library and lab-made library, combined with retention time, retention indices, literature references, and standards injections were used to identify the impurity produced during benzaldehyde aldolisation.

3. Results and discussion

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from spearmint carvone.

3.1. Spearmint Essential Oil

3.1.1. *Context*

Essential oils of spearmint Mentha spicata L. are mainly composed of carvone molecule. These extracts are generally processed with a fractional distillation to obtain more than 80% of carvone in the composition of the essential oil [12,13]. These products are usually originated from China, USA, and India. These essential oils are amongst the most important product in the essential oil market. Therefore, these extracts are highly subjects to be adulterated. Addition of synthetic carvone is a known adulteration and is easily highlighted with the control of the enantiomeric ratio [14,15]. An olfactory control is also very efficient, because enantiomers of carvone present different odours. Carvone (R) has a minty smell and is abundant in spearmint essential oils, whereas carvone (S) has a spicy aroma and is found in caraway essential oils. However, carvone (R) isomer can also be synthesized from natural origin limonene (S) from sweet orange essential oils [16]. This extract is currently the best-selling essential oil in the world and the cheapest in the market. Limonene represents over than 90% of the composition of sweet orange essential oil. The semisynthesis of carvone from limonene has the particularity to introduce an oxygen [11] (Figure 1). An isotopic monitoring of δ^{18} O may potentially be of interest to highlight the semisynthetic origin of carvone, because this newly introduced oxygen is from a different source and has not been impacted by the same isotopic fractionations as the original oxygen

3.1.2. Analysis of results

Enantiomeric and isotopic data are depicted in Table 1 and Figure 3.

The determination of the enantiomeric ratio has been carried out in order to control that carvone essential oils had not been adulterated with petrochemical origin carvone. The enantiomeric content of carvone (R) is measured between 97.3% and 100.0%. All the analysed samples present the carvone enantiomer (R) in excess, which is in line with the literature [14,15]. No significant differences were observed between authentic and commercial samples.

 δ^{13} C isotopic composition of carvone molecules derived from essential oils of genuine spearmint (-29.35 ± 1.54‰) is similar to the δ^{13} C of carvone molecules from commercial samples (-29.63 ± 0.87‰). These values are in line with the C3 origin [17] of the spearmint plant. According to the literature, C3 plants presents a δ^{13} C comprised between -20‰ and -30‰ [18]. The δ^{13} C of the carvone is not efficient to discriminate natural and synthetic origins of the carvone molecule according to the analyses. Indeed carvone molecules from petrochemical origins have δ^{13} C values comprised between -30.24‰ and -26.40‰, which coincide with δ^{13} C range value of authentic spearmint essential oils. Same result is observed with the carvone synthesized from limonene (δ^{13} C = -29.81‰).

As regards with δ^2 H isotopic values, significant isotopic fractionations are noticed between samples: authentic samples -262 \pm 31%; commercial samples -242 \pm 29%). These differences are probably explained by the different geographical origins, because δ^2 H stable isotope ratios are linked to the geographical location of the harvest [19]. USA spearmint essential oils have a mean value of -282 \pm 28%; China samples mean isotopic value is more enriched in deuterium (-235 \pm 18%); as well as India samples (-226 \pm 27%). These natural

values are significantly depleted by comparison with $\delta^2 H$ stable isotopic values of carvone molecules synthesized from petrochemical products (-190 \pm 24‰). The study of stable isotope ratios of hydrogen potentially highlights an origin of synthesis. However, this is unfortunately not possible in the case of a carvone obtained from limonene conversion, because the measured $\delta^2 H$ value (-204‰) is within the range of authentic sample values.

Finally, the study of δ^{18} O isotopic values highlights significant isotopic differences between authentic and commercial samples. The δ^{18} O value of 100% pure and natural carvone molecules are comprised between 14.1‰ and 22.1‰. These values are more positives than the δ^{18} O value measured on the carvone (R) obtained from limonene (9.2‰). From the panel of commercial samples, several essential oils presents δ^{18} O values similar to the measured range determined to authentic samples, but other samples present a lower δ^{18} O value, which potentially indicate that these samples have been adulterated with semisynthetic carvones.

3.2. Cinnamon Essential Oil & Bitter Almond Essential Oil

3.2.1. *Context*

Essential oils of cinnamon and bitter almond are amongst the most sold essential oils of the market. Bitter essential oils are obtained from the extraction of almond (*Prunus amygdalus* var. amara (DC.)) and apricot (*Prunus armeniaca* L.) kernels. This essential oil presents a composition with more than 95% of benzaldehyde. Cinnamon essential oil comes in two forms: oil cinnamon essential *Cinnamomum cassia* (L.) J. Presl, mainly from China and from distillation of wood and / or leaves, and cinnamon essential oil *Cinnamomun verum* J. Presl (syn. *Cinnamomum zeylanicum* Blume), mainly from Sri Lanka, India, and Madagascar, and

made from the distillation of cinnamon bark. These two essential oils are rich in (E) - cinnamaldehyde.

There are many frauds associated with the essential oils of bitter almond and cinnamon. Since the cassia origin of cinnamon is much cheaper than the Ceylon origin, the mixtures between these two essential oils are common. However, it is possible to highlight this type of addition by studying the differences in composition between the two essential oils [20], or by NIR and FTIR spectroscopic analysis [21,22]. Additions of (E) -cinnamaldehyde and benzaldehyde are also identifiable by radiocarbon analysis [23]. However, as shown in literature with the example of methyl salicylate or benzaldehyde [7,24], a labelling of synthetic molecule makes the measurement of 14 C activity inefficient. Isotopic methods by δ^2 H GC-P-IRMS are applicable to highlight the synthetic additions to benzaldehyde (Ruff et al., 2000) and (E) -cinnamaldehyde [25], as well as SNIF-NMR [4]. The synthesis of (E) -cinnamaldehyde from benzaldehyde produces a residual impurity, 5-phenylpenta-2,4-dienal [26], which is identifiable by GC, if the product has not been sufficiently purified.

In the same way as for mint, (E) -cinnamaldehyde can also be produced by semisynthesis, from benzaldehyde obtained from the essential oil of bitter almond. The scheme of reaction, presented in Figure 2, shows that (E)-cinnamaldehyde is obtained by aldolization of benzaldehyde (1) [4]. During the reaction, the oxygen in the benzaldehyde is removed and oxygen from acetaldehyde is introduced to form (E) -cinnamaldehyde. A stable isotopic assessment is therefore again of real interest. As prices fluctuate, essential oil bitter almond is sometimes cheaper than cinnamon bark essential oil (*Cinnamomum verum*). The fraud can be reversed. Benzaldehyde can also be hemi-synthesized from (E)-cinnamaldehyde [4], according to the retro-aldolization reaction (2). This time, the oxygen in the hemi-

synthesized benzaldehyde is obtained from water. However, the use of carbonates in the reaction suggests isotopic exchanges [27].

Therefore, adulteration can be realized in either ways, depending on market prices. The essential oil of cassia is ten times cheaper than the other two; it remains the main source of fraud.

3.2.2. Cinnamon Essential Oils

Some reference isotopic values of (E) -cinnamaldehyde have been published. The value $\delta^{13}C$ of (E) -cinnamaldehyde from natural cinnamon essential oil is between -28.20 and -28.60 ‰ according to Culp [23,24] and between -24.20‰ and -27.10‰ according to Sewenig [25], while the values $\delta^{13}C$ of synthetic (E) -cinnamaldehyde obtained from the oxidation of toluene, are between -24.61‰ and -25.70‰. According to the literature, the values $\delta^{2}H$ of natural (E) -cinnamaldehyde are given between -142‰ and -101‰ [23–25]. (E) – cinnamaldehyde of petrochemical origin are much more enriched in deuterium, which results in values $\delta^{2}H$ much higher, between 464‰ and 569‰ after Culp [23,24]. Finally, the naturally occurring (E)-cinnamaldehyde, a product of the shikimic pathway of biosynthesis [28], is expected to exhibit value $\delta^{18}O$ around 14.9‰ [29].

The isotopic values of (E) -cinnamaldehyde of the various essential oils of cinnamon, determined by GC-C / P-IRMS, are described in Table 2 and Figure 4.

The (E) -cinnamaldehydes of essential oils of cinnamon, which did not present impurities, have isotope ratios δ^{13} C of -28.37 \pm 1.14 ‰ and δ^{2} H of -142 \pm 10 ‰. These results are in agreement with the natural isotopic values presented by Culp and Sewenig[23–25]. δ^{2} H stable isotopic values are sometimes more negative however. The mean δ^{18} O value is

measured around 13.9 \pm 2.2‰, which is close to the value proposed by Schmidt [29]. All of these values are consistent with a natural origin.

Cinnamon essential oils that had impurities present various isotopic values. We can distinguish those which have $\delta^2 H$ indicators of one synthetic origin and others. The first ones have isotope ratios $\delta^{13} C$ -24.70 \pm 1.66‰ and $\delta^2 H$ between -42 and 844‰. These stable isotopic values demonstrate enrichment in $^2 H$ and $^{13} C$ compared to natural samples. The highest $\delta^2 H$ values are indicative of an origin of synthesis from toluene, while the lower values seem to correspond to benzyl chloride origin [25]. The measured $\delta^{18} O$ values are very variable, and fluctuate from 1.2 to 21.8‰. Isotope analyses confirm the detection of synthetic residues: these samples have undergone a partial or total addition of (E) - cinnamaldehyde of synthetic or semisynthetic origin.

However, other samples, which nevertheless had traces of 5-phenylpenta-2,4-dienal, exhibit naturally occurring (E) -cinnamaldehyde isotopic values (δ^{13} C -27.54 ± 0.65‰ and δ^{2} H -147 ± 12‰). However, we can see that the values in δ^{18} O are lower (4.0 ± 3.3‰). The oxygen values do not correspond to a natural origin from of the phenylpropanoid biosynthetic pathway from shikimic acid. The oxygen of (E) -cinnamaldehyde was probably altered, from a naturally occurring reagent: acetaldehyde from ethanol oxidation.

To verify this hypothesis, several semisyntheses of (E) -cinnamaldehyde were carried out, by aldolization of benzaldehydes, from different samples of almond essential oils bitter. Isotopic values $\delta^{13}C$ and δ^2H obtained correspond to a natural origin of (E) -cinnamaldehyde, but this is not the case with $\delta^{18}O$, which is measured between 7.9% and 9.5%. These values are lower than the values measured previously on essential oils of cinnamon natural, nor does it correspond to the value given by Schmidt [29].

The oxygen introduced during the aldolization of benzaldehyde comes from acetaldehyde used as reagent. It is the latter that will influence the final value of $\delta^{18}O$ of (E) - cinnamaldehyde produced by this semisynthesis. To verify this claim, the two commercial acetaldehyde samples used to produce the samples by aldolization, are analyzed by EA-P-IRMS, by direct introduction. The measure of their value $\delta^{18}O$ is 8.7 ± 0.1 %. By comparison, the mean value $\delta^{18}O$ measured on the 7 samples of (E) -cinnamaldehyde obtained by aldolization of benzaldehyde natural is 8.8 ± 0.6 %.

These results confirm the origin of the oxygen molecule introduced during the synthesis from benzaldehyde. If the $\delta^{18}O$ value of acetaldehyde is more negative, the (E) - cinnamaldehydes produced during aldolization will also have more negative $\delta^{18}O$ values, which may explain the values very low measured on some suspect samples. Indeed, acetaldehyde is produced industrially from agricultural ethanol, whose values $\delta^{18}O$ are low (between 3.0% and 8.1% after Schmidt [29]). It is therefore possible that syntheses can be carried out with acetaldehydes weakly enriched in ^{18}O .

3.2.3. Bitter Almond Essential Oil

For benzaldehyde, the isotopic values of natural origin indicated in the literature are between -27.06‰ and -29.58 ‰ for δ^{13} C, and between -145‰ and -84‰ for δ^{2} H from the study by Culp and Noakes [23,24], and between -113‰ and -148‰ for δ^{2} H from Ruff et al [30], for a bitter almond origin. Schmidt et al. [29] present a value δ^{18} O for one naturally occurring benzaldehyde from the shikimic route, around 19.3‰. Ruff's work and al. [30] also focused on samples of natural benzaldehyde ex-*Cinnamomum cassia*, the values δ^{2} H data is between -83 and -144 ‰, which merges with the range of values natural benzaldehyde from bitter almond essential oil. The isotopic values of Synthetic benzaldehyde are different

depending on the method of synthesis. A benzaldehyde from the acid or basic hydrolysis of (dichloromethyl) benzene has values between -28.52‰ and -29.99‰ for δ^{13} C [23,24], between -47 and -85‰ for δ^{2} H [23,24,30], and around 6.8‰ for δ^{18} O. While a benzaldehyde obtained by oxidation of toluene will have a value δ^{13} C between -24.70 and -26.50‰ according to Culp and Noakes [23,24], a strongly positive δ^{2} H isotopic value, between 420 ‰ and 720‰ approximately, and a value δ^{18} O around 14.1‰ according to the publication by Schmidt et al. [29].

- The isotopic values of benzaldehyde in various bitter almond essential oils analyzed,

 determined by GC-C / P-IRMS, are described in Table 3.
- Bitter almond essential oils have isotopic values corresponding to an origin natural benzaldehyde, isotopic values varying from -31.81% to -29.80% for δ^{13} C, from -131% to -97% for δ^{2} H, and from 13.4% to 19.4% for δ^{18} O.
- These values differ from those measured on benzaldehydes of petrochemical origin. Indeed,
 it is possible to observe more positive isotopic values on the three elements analyzed, which
 puts in evidence a greater enrichment in ¹³C, ²H, and ¹⁸O in the synthetic benzaldehydes.
 These values are characteristic of a synthesis from toluene.
 - Finally, various samples of benzaldehydes, obtained by retro-aldolization of (E) cinnamaldehydes from different origins were analyzed. Semisyntheses, carried out from (E) –cinnamaldehydes natural oils derived from cassia and verum cinnamon essential oils, have isotopic ratios stable δ^{13} C (-29.92 ± 0.54‰) and δ^{2} H (-108 ± 11‰) similar to the values obtained on oils bitter almond essentials. However, the isotopic values δ^{18} O are lower (10.9 ± 0.4‰) than those of natural essential oils.

The oxygen introduced during the retroaldolization reaction of (E) -cinnamaldehyde comes from water. However, isotopic exchanges occur between water and the carbonate used in the synthesis (Na₂CO₃), at 100°C [27]. The δ^{18} O value of benzaldehyde obtained by retroaldolization is therefore dependent on the isotopic value of the carbonate used during the synthesis, which explains why we find similar isotopic values on the different benzaldehydes produced by retroaldolization, despite the use as a starting reagent from different origins of (E) -cinnamaldehydes (synthetic or natural from essential oils of bitter almond).

The δ^{18} O of tap water, used for this synthesis, cannot be measured in EA-P-IRMS. To be able to carry out the measurement, we used an OA-ICOS system (Off Axis - Integrated cavity output spectroscopy) [31–33]. The δ^{18} O of water is measured at -11 ‰, while carbonates are generally highly enriched in 18 O (between 25‰ and 30‰, [27]). The isotopic exchange of oxygen between the carbonate ion CO₃²⁻ and H₂O lead to an intermediate value, which corresponds to the δ^{18} O values that we measure on benzaldehydes produced by retroaldolization.

Conclusion

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The results presented in this study are promising, as there are currently no analytical solutions to highlight evidence of semisynthetic product additions with $\delta^{13}C$ and $\delta^{2}H$ measurements, radiocarbon assessment, and enantiomeric ratio determination. In addition, biotechnologies will continue to grow with production of molecules from various origins using novel processes. δ^{18} O isotopic measurements will be of greater interest depending of technological advances. Other leads of investigation can be considered, such as the authentication of Oregano Origanum vulgare essential oils, primarily constituted of carvacrol. This molecule is easily synthesized from limonene (predominantly compound of sweet orange essential oil) and / or carvone (primarily found in spearmint and caraway essential oils)[34]. Another future prospect could be the study of the semisynthesis of (E)-anethol (Star anise Illicium verum, Anise Pimpinella anisum, and fennel Foeniculum vulgare essential oils) from methyl chavicol (basil Ocimum basilicum, tarragon Artemisia dracunculus, and chervil Anthriscus cerefolium essential oils). Furthermore, δ^{18} O isotopic study is not restricted to semisynthetic frauds. Many applications are potentially applicable, depending of the matrix of interest. δ^{18} O compound specific analyses using GC-P-IRMS can potentially be a new complementary tool for authentication, especially since many oxygenated constituents are present in the composition of essential oils, and that most of these metabolites have not yet been studied in δ^{18} O compound specific GC-P-IRMS analysis. As reported in previous studies [7], δ^{18} O metabolite specific values are efficient to highlight a petrochemical origin. In addition, this measure can provide

geographical origin authentication, as the isotopic fractionations of oxygen are linked to the atmospheric cycles of water, similar to those of hydrogen.

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Declaration of Competing Interests

The authors declare that there are no conflicts of interest.

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Table 1: Carvone compound-specific δ^{13} C, δ^{2} H, and δ^{18} O isotopic values and enantiomeric ratios measured in spearmint authentic and commercial essential oils, and carvone from petrochemical and from limonene conversion origins (n=number of samples).

Samples	Enantiomeric values of carvone				Compound-specific isotopic values of carvone (GC-C/P-IRMS)											
	% Carvone (R)			δ ¹³ C			δ^2 H					δ ¹⁸ Ο				
	Mean	Min	Max	SD	Mean	Min	Max	SD	Mean	Min	Max	SD	Mean	Min	Max	SD
Spearmint essential oils – Authentic samples (n=12)	99.7	98.7	100.0	0.4	-29.35	-33.34	-27.24	1.54	-262	-294	-211	31	18.4	14.1	22.1	2.6
Commercial spearmint essential oils (n=36)	99.1	97.3	100.0	0.7	-29.63	-30.96	-28.15	0.87	-242	-321	-201	29	9.0	-1.5	18.4	5.6
Carvone (R) – Petrochemical origin (n=3)	100.0	100.0	100.0	0.0	-27.94	-30.24	-26.40	2.03	-190	-217	-176	24	13.2	10.9	14.5	2.0
Carvone – Limonene conversion origin (n=1)	99.4	NA	NA	NA	-29.81	NA	NA	NA	-204	NA	NA	NA	9.2	NA	NA	NA
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Table 2: (E)-Cinnamaldehyde compound-specific δ^{13} C, δ^{2} H, and δ^{18} O isotopic values in cinnamon authentic and commercial essential oils, and (E)-Cinnamaldehyde from petrochemical and from benzaldehyde aldol origins (n=number of samples). Indication of the eventual detection of the 5-phenylpenta-2,4-dienal aldol reaction impurity using GC-MS and compliance of δ^{2} H value (maximal value of -101% based on [23–25]).

	δ^2 H	Impurity detected?	Compound-specific isotopic values of (E)-cinnamaldehyde (GC-C/P-IRMS)											
Samples	о п complied?		δ ¹³ C		δ^2 H				δ^{18} O					
	complied:		Mean	Min	Max	SD	Mean	Min	Max	SD	Mean	Min	Max	SD
Cinnamon essential oil – Authentic samples (n=11)	Yes	No	-28.37	-27.60	-26.13	1.14	-142	-156	-123	10	13.9	10.9	16.9	2.2
Cinnamon essential oil - Suspected hemi synthetic (E)- Cinnamaldehyde adulteration	Yes	Yes	-27.54	-28.68	-26.00	0.65	-147	-168	-125	12	4.0	-2.8	10.5	3.3
Cinnamon essential oil - Synthetic (E)-Cinnamaldehyde adulteration	No	Yes	-24.70	-27.57	-23.29	1.66	146	-42	844	391	11.7	1.2	21.8	7.6
(E)-Cinnamaldehyde – Petrochemical origin (n=1)	No	Yes	-25.96	NA	NA	NA	540	NA	NA	NA	16.1	NA	NA	NA
(E)-Cinnamaldehyde – natural benzaldehyde (bitter almond oil) aldol reaction origin (n=7)	Yes	Yes	-26.23	-28.51	-24.03	1.68	-125	-138	-109	9	8.8	7.9	9.5	0.6
(E)-Cinnamaldehyde – synthetic benzaldehyde aldol reaction origin (n=3)	Yes	Yes	-26.10	-26.69	-25.79	0.51	389	379	407	15	9.4	8.8	9.8	0.5

Table 3: Benzaldehyde compound-specific δ^{13} C, δ^{2} H, and δ^{18} O isotopic values in bitter almond authentic essential oils, and benzaldehyde from petrochemical and from (E)-cinnamaldehyde retro aldol origins (n=number of samples).

	Compound-specific isotopic values of benzaldehyde (GC-C/P-IRMS)											
Samples	$\delta^{13}C$	$\delta^2 H$	δ ¹⁸ Ο									
	Mean	Min	Max	SD	Mean	Min	Max	SD	Mean	Min	Max	SD
Bitter almond essential oil – Authentic samples (n=12)	-30.72	-31.81	-29.80	1.00	-117	-131	-97	12	16.5	13.4	19.4	2.1
Benzaldehyde – Petrochemical origin (n=3)	-26.17	-26.91	-25.26	0.84	494	389	546	91	20.9	19.2	23.7	2.4
Benzaldehyde – Natural (E)-cinnamaldehyde (cinnamon oil) retro-aldol reaction origin (n=4)	-29.92	-30.42	-29.18	0.54	-108	-120	-99	11	10.9	10.3	11.2	0.4
Benzaldehyde – Synthetic (E)-cinnamaldehyde retro-aldol reaction origin (n=1)	-26.57	NA	NA	NA	705	NA	NA	NA	11.2	NA	NA	NA

Figure 1: Synthesis of carvone from limonene in 3 steps: (1) Synthesis of limonene nitrosochloride from limonene. (2) Production of carvoxime.

607 (3) Synthesis of carvone.

Figure 2: Synthesis of (E)-cinnamaldehyde from aldolisation of benzaldehyde. This synthesis leads to the production of an impurity : 5-

phenylpenta-2,4-dienal

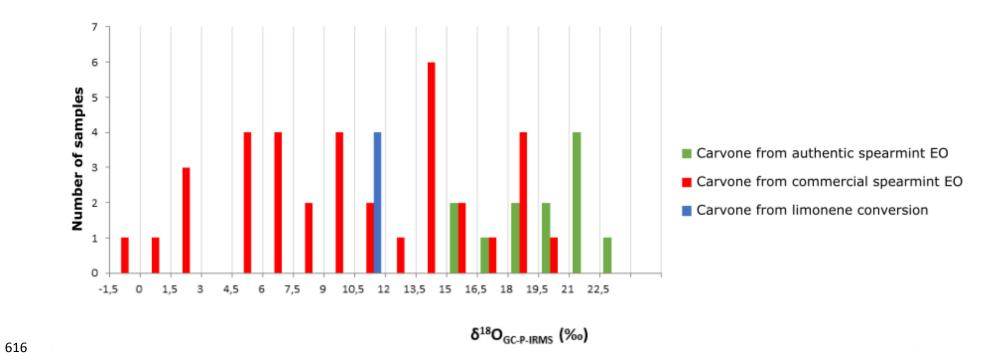


Figure 3: Distribution of $\delta^{18}O_{GC\text{-}P\text{-}IRMS}$ carvone isotopic values measured on 48 spearmint essential oils and 4 semisynthetic origin carvones from sweet orange limonene.

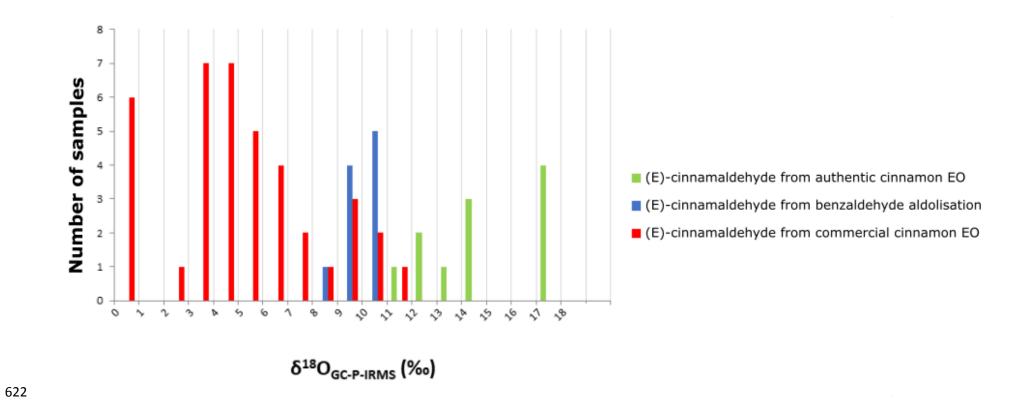


Figure 4: Distribution of $\delta^{18}O_{GC-P-IRMS}$ (E)-cinnamaldehyde isotopic values measured on 50 *Cassia* and *Verum* cinnamon essential oils and 10 (E)-cinnamaldehyde obtained from benzaldehyde aldolisation (Bitter almond and petrochemical origins).