

Automated keratinocyte identification on in vivo reflectance confocal microscopy images of the epidermis (Poster)

Imane Lboukili, Xavier Descombes, Georgios N Stamatas

► To cite this version:

Imane Lboukili, Xavier Descombes, Georgios N Stamatas. Automated keratinocyte identification on in vivo reflectance confocal microscopy images of the epidermis (Poster). YSN 2022 - 3rd Imabio Young Scientist Network Conference, Jul 2022, Lyon, France. hal-03760766

HAL Id: hal-03760766 https://hal.science/hal-03760766

Submitted on 25 Aug 2022

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

Automated keratinocyte identification on *in vivo* reflectance confocal microscopy images of the epidermis

Imane Lboukili^{a,b}, Xavier Descombes^b, Georgios Stamatas^a ^aJohnson & Johnson Santé Beauté France; ^bUCA – INRIA – I3S/CNRS

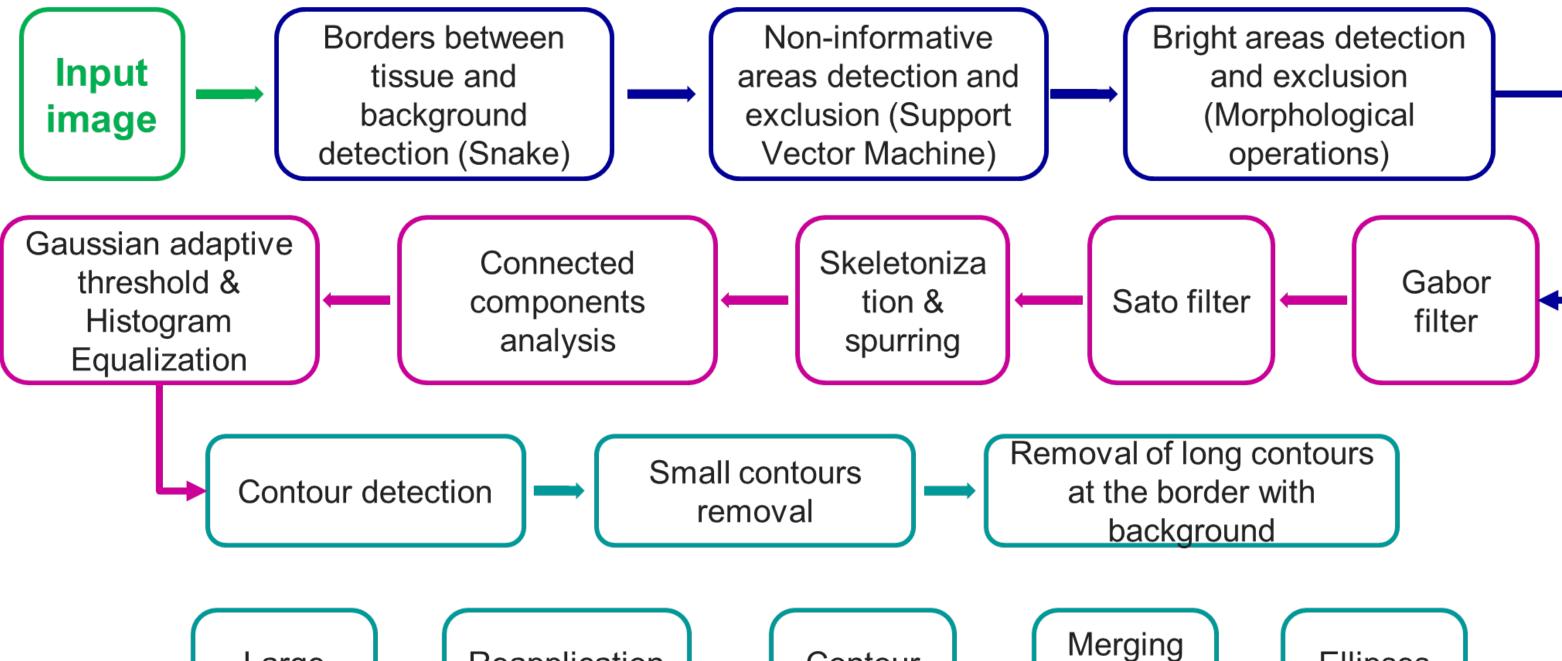
INTRODUCTION

(Johnson Johnson CONSUMER HEALTH

Reflectance Confocal Microscopy^{1,2} (RCM) is a real-time non-invasive *in vivo* technology that allows the visualization of the epidermis and upper layers of the dermis at cellular level. It provides information on the geometrical and topological properties of the skin, which play important roles in the makeup of the skin barrier. In most cases, analysis of RCM stacks is done manually, providing qualitative observations. However, manual analysis is time-consuming and subject to human

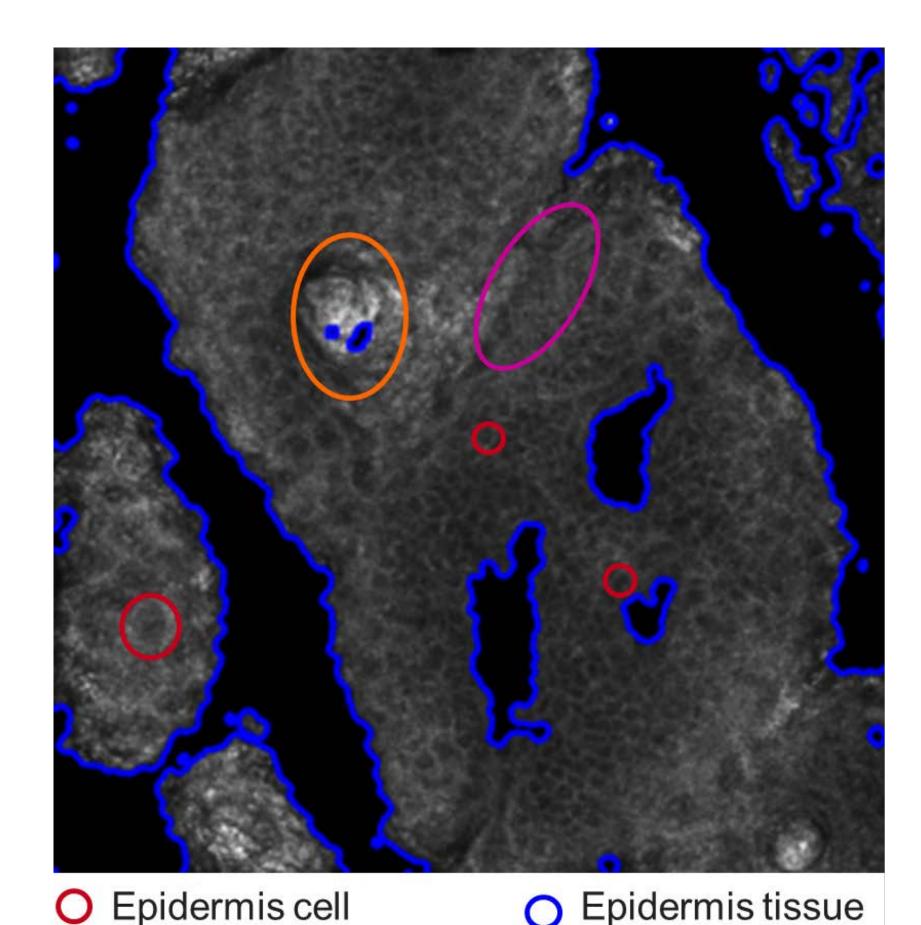
interpretation and inter-expert differences. Thus, we could benefit from automated methods to quantitatively analyze RCM images.

Our approach focuses on RCM images of the *stratum granulosum* (SG) and *stratum spinosum* (SS) where viable cells, called keratinocytes, appear in a honeycomb pattern. They are both characterized by a dark center and a grainy cytoplasm due to micro-structures, surrounded by bright grainy membranes.



UNIVERSITÉ Côte d'Azur

Attempts at the automated identification of epidermal cells on RCM images have been made^{4,5}. Unfortunately, the amount of noise and heterogeneity of RCM images hinders the development of accurate segmentation methods.



Reapplication Contour Ellipses Large of small of filter locally fitting contours detection contours Two groups of contours Combination of all Small Accuracy contours & centers **Evaluation** contours detection

Fig. 2 Steps of automated cell identification **RESULTS**

Image	Precision	Recall
Image 1	86%	72%
Image 2	71%	82%
Image 3	81%	86%
Image 4	72%	91%

We have shown that the automated detection of keratinocytes on RCM images of

the SG and SS is achievable and comparable to expert's manual identification.

HEALTHY SKIN MATURATION

O Non-informative areas O Bright areas

Fig. 1 RCM image of the *stratum spinosum* of light-pigmented skin. In blue, the border between tissue and background. In pink, non-informative areas. In orange, bright spots. And in red, epidermal cells.

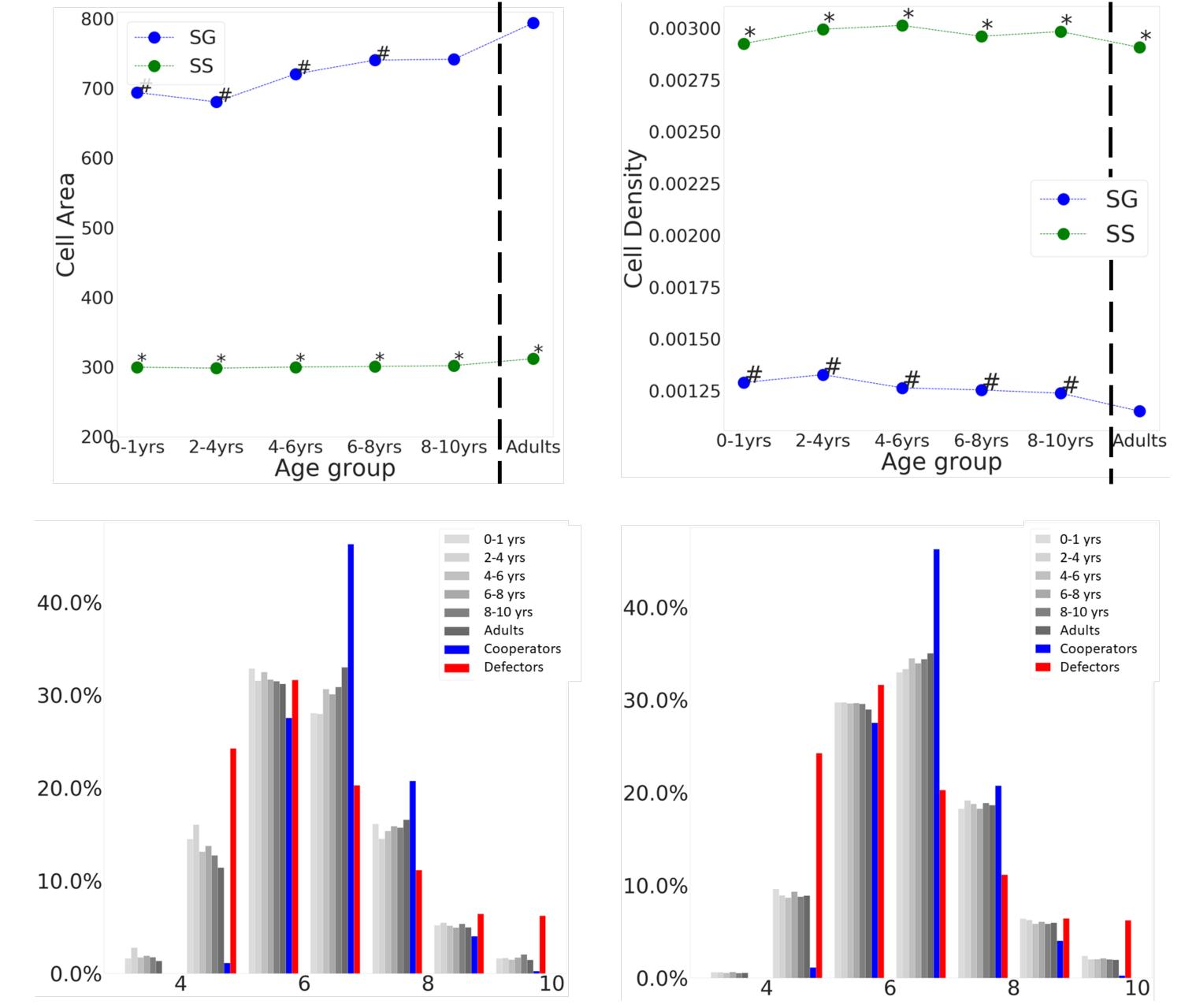
We propose a method to automatically detect keratinocytes centers on RCM images

of the stratum granulosum and stratum spinosum.

IMAGE ANALYSIS WORKFLOW

The first step is to identify the region-of-interest (ROI) containing cells. We start by identifying the tissue using the Morphological-Geodesic-Active-Contour method. We further refine the ROI by removing non-informative bright areas sometimes present in RCM images by applying an alternate sequential filter on the binarized blurred image. Other spurious areas are removed using a Support Vector Machine algorithm trained on Grey-level-Co-occurrence-Matrix features.

The second step is to identify individual cells in the segmented tissue area. To perform this task, we use successive application of Sato and Gabor filters. The outputs of these filters are then locally normalized, binarized, and skeletonized. The obtained skeletons (representing cell membranes) are pruned to remove spurious branches.



The third step is a post-processing to improve the detection and remove size outliers (detected areas too small or too large to be accurate detections of cells).

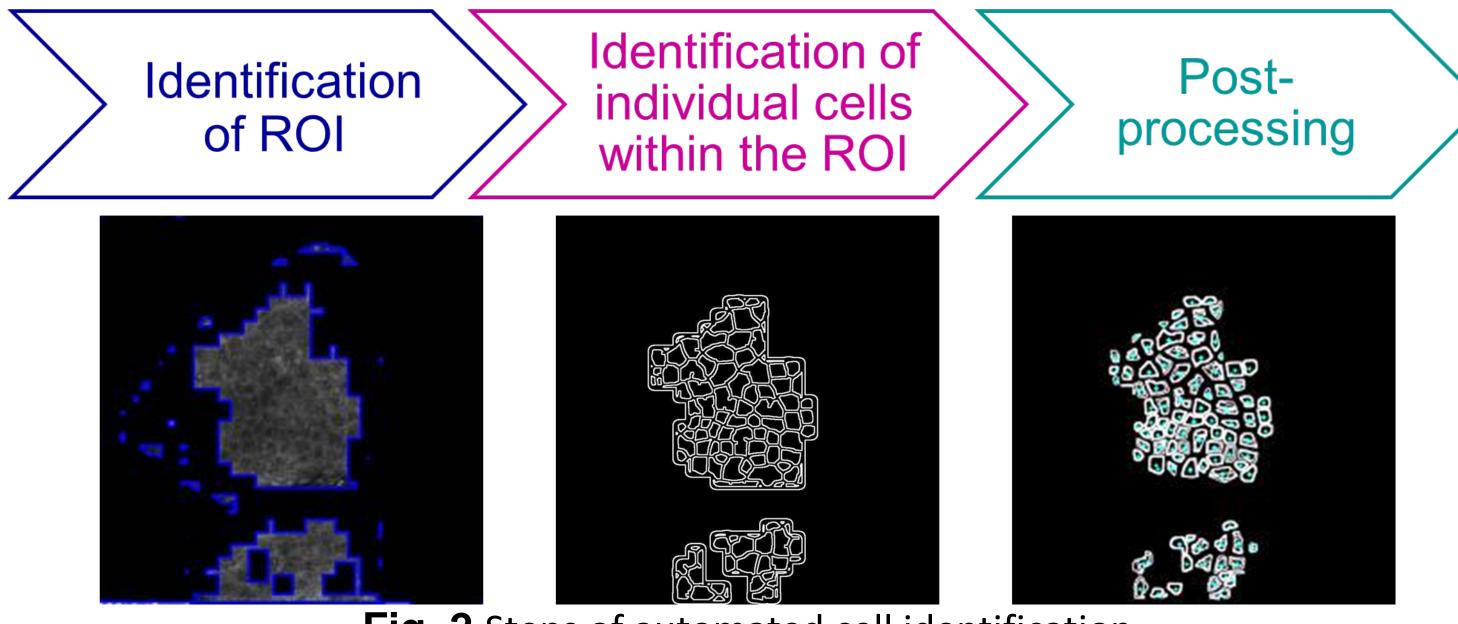


Fig. 2 Steps of automated cell identification

* indicates that the SS value is significantly different from the SG value;
indicates that group value is significantly different from the adult group value.

The epidermis continuously changes between 0 and 10 years-old with important inter-person variability. Cell geometry changes with age and per epidermal layer but topology remains the

REFERENCES

1. M. Minsky, "Microscopy apparatus," US3013467A (1961).

same.

- 2. M. Minsky, "Memoir on inventing the confocal scanning microscope," Scanning, (1988).
- 3. J. Bensaci et al., "Geometrical and topological analysis of in vivo confocal microscopy images reveals dynamic maturation of epidermal structures during the first years of life," J. Biomed. Opt., (2015).
- 4. M. A. Harris et al., "A Pulse Coupled Neural Network Segmentation Algorithm for Reflectance Confocal Images of Epithelial Tissue," PLoS ONE, (2015).
- 5. D. Gareau, "Automated identification of epidermal keratinocytes in reflectance confocal microscopy," J. Biomed. Opt., (2011).
- 6. F. MEYER et al., "Morphological segmentation," J. Vis. Commun. Image Represent., (1990).