

Arctic methylmercury cycling

Sofi Jonsson, Michelle Nerentorp Mastromonaco, Feiyue Wang, Andrea G Bravo, Warren R.L. Cairns, John Chételat, Thomas A. Douglas, Gretchen Lescord, Liisa Ukonmaanaho, Lars-Eric Heimbürger-Boavida

To cite this version:

Sofi Jonsson, Michelle Nerentorp Mastromonaco, Feiyue Wang, Andrea G Bravo, Warren R.L. Cairns, et al.. Arctic methylmercury cycling. Science of the Total Environment, 2022, pp.157445. $10.1016/j.scitotenv.2022.157445$. hal-03737703

HAL Id: hal-03737703 <https://hal.science/hal-03737703>

Submitted on 15 Nov 2022

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

Arctic Methylmercury Cycling

pool of MeHg available for bioaccumulation. Methylation of Hg occurs in diverse Arctic

1. Introduction

 Mercury (Hg) is a bioaccumulative neurotoxin that is released from the earth's crust through natural processes and anthropogenic activities (Streets 2019). Since the industrial revolution, direct anthropogenic emissions have exceeded natural emissions by an order of magnitude and concentration of Hg in the atmosphere has increased 3-5 times (Driscoll et al. 2013, Li et al 2020). Since Hg is well mixed globally in Earth's atmosphere these emissions have also reached the Arctic environment. Due to a variety of physical, chemical, and biological processes, some of which are unique to environments in cold regions, high levels of Hg exposure have been detected among Arctic communities and wildlife (AMAP 2021).

 The biogeochemical cycle that connects Hg emissions to Hg in Arctic food webs involves inorganic and organic forms of Hg that undergo a wide range of transformation processes in air, water, and land. There are three main chemical forms of Hg in Arctic environmental compartments and biota: gaseous 66 elemental mercury $(Hg(0))$, inorganic divalent mercury $(Hg(II))$ and MeHg. Figure 1 shows the partition of each in the major Arctic environmental compartments and biota.

 Industrial activities (e.g., artisanal and small-scale gold mining, coal-fired power stations, waste 69 incineration, cement manufacture, and others) release both $Hg(0)$ and $Hg(II)$. The main natural Hg sources to the atmosphere are volcanoes and forest fires. While Hg(II) is typically deposited closer to the emission source, Hg(0) has a residence time ranging from several months to over a year in the atmosphere and can be transported from the emission source to remote polar regions within a few days to weeks (Durnford et al, 2010, Dastoor et al. 2015). Atmospheric mercury depletion events, during polar sunrise occur in both Arctic and Antarctic regions and lead to the rapid oxidation of Hg(0) to the 75 more easily deposited Hg(II) species. A large fraction of $Hg(0)$ also dissolves in natural waters where it is referred to as dissolved gaseous mercury (DGM) (Jiskra et al. 2021). On land, foliar (plant) uptake of Hg(0) drives the atmospheric deposition of Hg in the Arctic tundra (Obrist et al., 2017; Jiskra et al. 2018; Douglas and Blum, 2019). Through this process, large amounts of Hg have accumulated in northern permafrost soils from the atmosphere over thousands of years. Today, concerns are raised

 about the potential remobilization of this permafrost Hg pool as climate warming leads to increased soil temperatures, thawing permafrost, and changes in the hydrological cycle (Schaefer et al., 2020, Lim et al. 2020). In natural waters, soils and sediments, Hg is primarily found as Hg(II) (Skyllberg, 83 2012). Due to a high affinity for particles (e.g. mineral surfaces and organic matter), most $Hg(II)$ in sediments and soils is found in the solid phase and these systems commonly act as important sinks for Hg. At the same time, these systems are known to be important sites for Hg methylation. The main form of MeHg existing in natural environments is monomethylmercury (MMHg), although, in some environments, dimethylmercury (DMHg) may become the dominant species, especially in seawater. Since most analytical data on methylated Hg do not differentiate MMHg and DMHg, we will use MeHg to refer to methylated Hg in general from hereon, unless otherwise specified. While both MMHg and DMHg are toxic and lipophilic, only MMHg has been shown to biomagnify in aquatic food webs to concentrations of concern for wildlife and human health (Lavoie et al., 2013). Because of this, the potential risk Hg pollution poses to Arctic ecosystems is not only controlled by the amount of inorganic Hg transported into the system and cycling within the Arctic environment, but also by the extent of the Hg pool that is methylated and accumulated in aquatic food webs. In this review and contribution to the Virtual Special Issue on Arctic mercury, we present a synthesis of work conducted for the AMAP Assessment 2021: Mercury in the Arctic on the processes affecting mercury transformation and biotic uptake in the Arctic (AMAP 2021, Chapter 4). Given the high complexity of the Hg biogeochemical cycle, it is not surprising that significant advances are still being

made despite over 50 years of active research. Here, we focus on scientific progress made in the last

decade on processes leading to environmental MeHg exposure by presenting (1) a new mass balance

for MeHg in the Arctic, and new information on (2) our mechanistic understanding of Hg

(de)methylation processes, (3) where hotspots of MeHg are found in the Arctic and, finally, (4)

processes affecting Hg accumulating in Arctic biota.

 budgets were calculated for land areas north of 60°N and the Arctic Ocean, including the central basin, and the Barents, Kara, Laptev, East Siberian, Chuckhi and Beauford Seas.

 To estimate the stock for the 0-20 m, 20-200 m, 200-500 m and <500 m water masses (Table 2) average concentrations of THg and MeHg in seawater calculated from available data (Wang et al., 2012b, 2018; Heimbürger et al., 2015; Agather et al., 2019; Charette et al., 2020; Kim et al., 2020b; Petrova et al., 2020; Jonsson et al., 2022, Kohler et al., 2022) were used. For biota, we used the estimate previously provided by Soerensen et al., 2016b. Remaining reservoirs (soil, snow, ice and air) were estimated from the stocks of Hg (Dastoor et al. 2022; Table 3.8.1 in AMAP (2021), key data for this review are also shown in Table S1) and the best estimates of %MeHg for each environmental compartment (Table 1 and S1). The MeHg pool in surface soils was estimated by multiplying the total Hg pool by the median %MeHg in surface soils (St. Pierre et al., 2015: 0.82%) and wetlands (Tarbier et al., 2021: 1.7%, not impacted by recent permafrost thaw) and assuming a 25% coverage of wetlands for the Arctic Region (Kåresdotter et al., 2021). Methylmercury reservoirs in snowpack and glaciers were estimated using %MeHg measurements for snow (6.1%, range: 0.11-11%; St. Pierre et al., 2019, Varty et al., 2021 and Jonsson et al., 2022) and median %MeHg (5.5 %) reported by Zdanowicz et al., (2016) for glaciers, respectively. For sea ice, the mean %MeHg reported by Dimento et al., (2019) was used, resulting in an estimated MeHg pool of 0.32 Mg. This value is close to the estimate of MeHg (0.33 Mg) previously estimated by Schartup et al. (2020). For the atmosphere, %MeHg were calculated from the concentrations of MMHg and DMHg (Table 1) previously reported by Baya et al. (2015) and total Hg concentration reported by Dastoor et al. (2022). Using geochemical modelling, Soerensen et al., (2016b) estimated MeHg reservoirs in the Arctic Ocean (also including the Canadian Arctic Archipelago and the Baffin Bay), sediments and plankton

resulting in a total of 576 Mg. In the updated MeHg budget presented by Petrova et al. (2020), the total

pool of MeHg in the Arctic Ocean was estimated to 800 Mg MeHg. Our budget for the Arctic Ocean

143 water sums up to 590 \pm 341 Mg MeHg and is thus in the same range as earlier estimates.

Petrova et al. (2020) found a MeHg maxima in the subsurface waters (100–300 m) of the Arctic Ocean

(in line with observations from Wang et al. (2012b, 2018), Heimbürger et al. (2015), Agather et al.

 (2019), Jonsson et al. (2022)), leading to a net export of MeHg from the Arctic Ocean to the Nordic Seas and the North Atlantic of 7.5 Mg/yr. A part of the MeHg produced in the subsurface ocean layer (20–200 m; 17 Mg/yr) is converted to DMHg, which diffuses up to the polar mixed layer (PML; 0–20 149 m), where some of it evades to the atmosphere. Here DMHg is proposed to rapidly photochemically 150 degrade back to MMHg and deposit back to the PML. For the AMAP 2021 report, MeHg (MMHg + 151 DMHg) evasion was estimated to 4.5 (2.8-6.3) Mg/yr using a DMHg to Hg^0 evasion ratio of 0.16 152 (Soerensen et al, 2016b) and the new Hg^0 evasion estimate (32 (20-45) Mg/yr (Table 3.8.1 in AMAP (2021)). The MeHg deposition was then calculated to 2.6 Mg/yr assuming the evasion to deposition ratio (0.57) presented by Soerensen et al. (2016b). In addition, DMHg may be photochemically degraded in surface waters to MMHg (West et al. 2022). However, as our budget deals with the total stock of MeHg, transformation reactions between methylated Hg species within compartments are not considered here. Riverine MeHg inputs (0.8 Mg/yr) were estimated from Sonke et al., (2018) and equal inputs by coastal erosion was assumed (as done for Hg, Table 3.8.1 in AMAP (2021)). The benthic flux of MeHg to subsurface waters was estimated to 0.9-4.6 Mg/yr (Soerensen et al., 2016b, Kim et al., 2020b). The *in situ* net water column methylation (5.1 Mg/yr) was estimated by calculating the difference between estimated inputs (total input: 18.4 Mg/yr; rivers: 0.8 Mg/yr; coastal erosion: 0.8 Mg/yr; atmospheric deposition: 2.6 Mg/yr; oceans: 9 Mg/yr; sediment: 4.7 Mg/yr, sea ice: 0.5 Mg/yr) and outputs (total output: 23.5 Mg/yr; atmospheric evasion: 4.5 Mg/yr; ocean inputs: 16.5 Mg/yr; sediment: 1.4 Mg/yr, sea ice: 0.4 Mg/yr; plankton uptake 0.7 Mg/yr). Our *in situ* net water column methylation for the Arctic Ocean is close to the *in situ* net water column methylation estimates provided from earlier models (Kim et al., 2020: 10 Mg/yr; Soerensen et al., 2022: 15 Mg/yr for the upper 200 m). Our estimated *in situ* production corresponds to a daily net Hg methylation potential of 168 7.5 10⁻⁶ d⁻¹ (calculated using the water column total Hg budget estimate (1,870 Mg Hg) from Dastoor et al., 2022). This calculated net Hg methylation potential (net result of *in situ* Hg methylation and *in situ* MeHg demethylation processes) is three orders of magnitude lower than the potential Hg 171 methylation rate constants $(0.6-13 \times 10^{-3} \text{ d}^{-1})$ reported from incubation studies by Lehnherr et al., (2011).

 The Arctic MeHg budget highlights the importance of several processes that control MeHg reservoirs. Specifically, MeHg production in the water column and in shelf sediments are dominant inputs to the Arctic Ocean (Figure 2). Atmospheric deposition and marine evasion also represent large fluxes of MeHg, with a net deposition flux. Similarly, fluxes of MeHg via oceanic currents are large, with an estimated net export from the Arctic Ocean to the North Atlantic. The modelling of estimates for pools and fluxes is based on existing data, which have varying degrees of uncertainty. In particular, our *in situ* net Hg methylation potential is associated with high uncertainty, although the much lower *in situ* 179 net methylation used in our model (in comparison to potential methylation rates (k_m) values available) suggests a rapid turnover of MeHg in Arctic waters (Lehnherr et al., 2011 reported potential 181 demethylation rates of 0.2-0.6 d^{-1}). Some budget estimates are based on a "best estimate" approach (when no data was available) or a very limited set of observations. Therefore, improved spatial and temporal coverage of measurements is important to improve estimates of MeHg fluxes, including from meltwater and erosion.

186 **Table 1.** Total MeHg concentrations, fluxes and pools in the Arctic environment (excluding aquatic biota larger than plankton). The total MeHg 187 concentrations reported cover both freshwater and the marine compartments, while the fluxes focus on those relevant for the Arctic Ocean.

0.046±0.028 ng/L (n=177) 32% 200-500 m

0.040±0.026 ng/L (n=439) 21% 20-200 m Wang et al., 2012, 2018;

Heimbürger et al., 2015;

189 **Table 2.** Total MeHg fluxes and pools in the Arctic environment (excluding aquatic biota larger than

190 plankton). While the MeHg concentrations reported (Table 1) cover both freshwater and the marine

191 compartments, the fluxes focus on those relevant for the Arctic Ocean. Details on how the MeHg

192 pools (and their uncertainty) were calculated is provided in Table S1.

193

194

195

196 **Figure 2.** MeHg fluxes and budget in the Arctic environment. Updated from Soerensen et al. (2016b) 197 and Petrova et al. (2020) with data shown in Table 1 and Table 2.

198

¹⁹⁹ 3. Mechanistic understanding of Hg (de)methylation processes

3.1. Formation and degradation pathways of MeHg

 Given that MeHg is the bioaccumulative form of Hg, understanding the rates of both methylation and demethylation as well as Hg(II) reduction, which might limit the availability of this species for methylation, have implications for biotic uptake and exposure of Hg. While photochemical processes might govern Hg chemistry at the surface of the waters, biological processes are the key to understand Hg cycling in freshwater, seawater and in sediments.

 Methylation of Hg is primarily thought to be a biotic process carried out by various microorganisms, though evidence for abiotic methylation exists. In 2013, the gene pair involved in biological Hg methylation (*hgcAB* genes) were discovered (Parks et al., 2013). This opened up new possibilities to search for Hg methylators (Gilmour et al., 2013, Jones et al., 2019; Capo et al., 2020; Gionfriddo et al., 2016; McDaniel et al., 2020) and uncover the intracellular processes involved in Hg methylation (Smith et al., 2015). Carriers of the *hgcAB* genes that are known to methylate Hg include a wide range of sulfate reducing bacteria, iron reducing bacteria, methanogens, and some fermentative microorganisms (Gilmour et al., 2013, 2018). A global analysis of metagenomes was first to reveal the presence of *hgcAB* in the Arctic environment by identifying the genes in *Deltaproteobacteria*, *Firmicutes*, and particularly in methanogenic archaea from thawing permafrost soil (Podar et al., 2015). Fahnestock et al. (2019) also showed that concentrations of MeHg in porewater increased across a thaw gradient along with the abundance and diversity of potential Hg methylators. It is important to note that not all Hg methylating microorganisms have the same capacity to methylate Hg and that this appears to depend on individual strains rather than taxonomic ranking (Ma et al., 2019). Marine microorganisms with a greater oxygen tolerance and broader habitat range might be important Hg methylators in the seawater (Villar et al. 2020; Lin et al., 2021). In the last decade, several studies have highlighted the importance of Hg methylation in suboxic and

even oxic waters for MeHg bioaccumulation and biomagnification in aquatic food webs (Sunderland

et al., 2009; Heimbürger et al. 2010; Lehnher et al., 2011; Mason et al., 2012; Wang et al., 2012b,

2018; Blum et al., 2013; Heimbürger et al., 2015; Schartup et al., 2015a; Soerensen et al., 2016a;

Gascón Díez et al., 2016; Rosati et al. 2018; Agather et al., 2019; Petrova et al., 2020). While

 anaerobic biological Hg methylation is mediated by the *hgcAB* genes that are abundant in marine sediments (Podar et al., 2015), the aerobic pathway, if it exists, remains unclear. *HgcA* or *hgcAB*-like genes have recently been identified in global ocean oxic seawater (Villar et al., 2020) as well as in Antarctic sea ice (Gionfriddo et al. 2020), though not in the Arctic Ocean (Bowman et al., 2020a). Further studies are needed to identify key players for Hg methylation in polar marine waters and sea ice.

 It is generally agreed that primarily dissolved complexes of Hg(II) and MMHg are taken up by Hg- methylating bacteria at the base of aquatic food webs (Drott et al., 2007; Schaefer and Morel, 2009; Chiasson-Gould et al., 2014; Mangal et al., 2019). Adsorption of Hg(II) and MMHg to the solid phase are thus important processes limiting their biological uptake (Schartup et al., 2015a; Mazrui et al., 2016). However, as shown in previous work from other marine or brackish water systems, different dissolved and solid chemical forms of Hg differ in their bioavailability (Jonsson et al., 2012, 2014; 240 Mazrui et al., 2016). Although recent terrestrial inputs of $Hg(II)$ and MeHg did not contribute much to the stock of MeHg in the estuarine system, 40 to 70% of the bioaccumulated Hg pool originated from these recent terrestrial Hg inputs. In these studies, refractory pools of Hg(II) and MMHg were not fully accounted for.

 MeHg demethylation can be biotically or abiotically mediated. Abiotic demethylation is mostly photochemical degradation in surface waters by different mechanisms including direct photolysis and indirect photolysis via reactions with reactive oxygen species such as hydroxyl radicals. Research in the Arctic, subarctic, and temperate areas has shown photochemical demethylation from ultraviolet (UV) radiation is an important aspect of freshwater Hg budgets (Seller et al., 1996; Lehnherr et al., 2012a; Poste et al., 2015; Klapstein and O'Driscoll, 2018). Because organic matter compounds in surface waters readily absorb UV radiation, they mediate the photodemethylation of aqueous MeHg (Klapstein et al., 2017). A recent study on ponds with high DOC concentrations resulting from permafrost thaw slumps found higher rates of photodemethylation when compared to an oligotrophic lake in the polar desert with low DOC concentrations (Girard et al., 2016). Photolytically produced reactive oxygen species from organic matter have been shown to mediate MeHg demethylation in

 Alaskan lacustrine systems (Hammerschmidt and Fitzgerald, 2010). In the Arctic, photodemethylation rates are likely regulated by prolonged ice coverage (Poste et al., 2015; Klapstein and O'Driscoll 2018). Non-photochemical abiotic demethylation may also occur via reactions with selenoamino acids (Khan and Wang, 2010), but the importance of such reactions remains to be further examined outside of the laboratory.

 Bacterial resistance to MeHg (and Hg(II)) is one of the most widely observed phenotypes in eubacteria (Barkay et al., 2003) and is mediated by the *mer* operon, which has been found in a diverse range of habitats including soils, sediment and water, both freshwater and marine systems (Barkay et al., 2003 and references therein). Within the operon (which contains different types of genes), attention has been focused on the *merA* gene, which catalyzes reaction transforming Hg(II) into Hg(0) and *merB*, which codifies for the alkaline mercury lyase, a protein that demethylates MeHg into Hg(II). Little is known about the occurrence, diversity and distribution of *these genes* in the Arctic region. Of potential importance for Hg demethylation, a recent study, however, unveiled the presence of *merB* in deep waters of the Arctic Ocean (Bowman et al., 2020a). Also, a study performed in lake sediment cores showed the presence of hgcA in Arctic lakes (Ruuskanen et al., 2020). The diversity of both Hg methylators and MeHg demethylators in the Arctic needs to be further explored.

3.2. Formation and degradation pathways of DMHg

 The main formation and degradation pathways of the DMHg remain unknown in marine waters, including the Arctic Ocean (Agather et al., 2019; Petrova et al., 2020; Bowman et al., 2020b, Jonsson et al. 2022). Although early studies identified synergies between the activity of organisms and production of DMHg (Baldi et al.,1993; Sommar et al., 1999; Pongratz and Heumann, 1998), there is currently no experimental data to support that it is a biological product. Baldi et al. (1993) suggested the formation of DMHg in bacterial cultures was the result of an abiotic reaction between dissolved sulfide and MeHg (via the formation of bismethylmercurysulfide). Recent work has also shown abiotic formation of DMHg from MMHg when MMHg is adsorbed onto organic or inorganic surfaces

 containing reduced sulfur sites (Jonsson et al., 2016). Other known pathways of DMHg formation of potential relevance in natural environments include the reaction of MeHg with selenoamino acids (Khan and Wang, 2010) and methylation with methylcobalamin (Imura et al., 1971). Volatilization of DMHg from the marine boundary layer and the subsequent atmospheric demethylation of DMHg to MMHg has been suggested as a source of MeHg in surface snow (Kirk et al., 2008; St. Louis et al., 2007), marine waters (Baya et al., 2015) and coastal terrestrial environments (St. Pierre et al., 2015). Baya et al. (2015), who were first to quantify DMHg in ambient air, also suggested the evasion of DMHg from marine waters in Hudson Bay and the Canadian Arctic Archipelago was enough to sustain the atmospheric concentrations of MMHg observed. In addition to degradation of DMHg in air, DMHg could potentially also be degraded in marine waters, although there have been contradictory claims in the literature on whether the rates are high enough for the process to play a role in the cycling of DMHg (Mason and Sullivan, 1999; Black et al., 2009). West et al. (2022), however, recently demonstrated that DMHg in arctic seawater readily degrades under natural sunlight. The photodecomposition rates observed in tested waters were in the same range as the rate observed for MMHg, suggesting that a significant fraction of the DMHg that is transported to surface waters may be degraded to MMHg directly in surface waters rather than in the atmosphere. It is also possible that DMHg is degraded through other pathways which do not involve photochemical reactions. DMHg has recently been shown to degrade in aqueous solutions to MeHg in the presence of reduced sulfur as dissolved sulfide or as the metastable iron sulfide mackinawite (West et al., 2020).

4. Where are methylmercury hotspots in the Arctic?

4.1. Terrestrial Environments

4.1.1 Arctic Tundra

 Mercury has a strong affinity to organic substances, and organic soils therefore play an important role in the mobilization and transportation of Hg from soils to water bodies. As such, surface vegetation, soils, and organic matter are likely locations where elevated Hg is present and available for transformation. St. Pierre et al. (2015) reported that soil MeHg concentrations varied by two orders of magnitude (0.02–2.11 ng/g) across several sites in the Canadian Arctic Archipelago, with a median of 0.12 ng/g. A tundra study (Olson et al., 2018) reported concentrations of MeHg between 0.5 and 3.4 ng/g in organic horizons and 0.1 to 1.0 ng/g in B horizons, with MeHg on average accounting for 1% of THg (THg with a maximum of 3% in the lower B horizon). A study from Fennoscandia further 314 found MeHg accounting for $0.48 \pm 1.7\%$ of THg in peat plateau soils, respectively (Tarbier et al. 2021). As discussed further below, higher %MeHg values has been reported from subarctic and Arctic 316 soils saturated with water (Tarbier et al. 2021: $6.1 \pm 6.5\%$, Varty et al. 2021: $>10\%$). In addition to driving the deposition of Hg to Arctic tundra by foliar uptake (Obrist et al., 2017), plants also bioaccumulate Hg from the soil via the roots (Zhou and Obrist, 2021). Generally, very few data are available on MeHg concentrations in Arctic vegetation. Lichens in some coastal areas of the Canadian High Arctic (MeHg: 1.41–17.1 ng/g) were more enriched in MeHg (and THg) relative to underlying soils than at inland sites. This suggests greater uptake/deposition of MeHg may originate from the evasion of DMHg and MMHg from open marine waters in coastal areas (St. Pierre et al., 2015). Further research is needed to identify sources and pathways of MeHg uptake in terrestrial vegetation.

 A major consequence of climate warming in the Arctic and Subarctic has been the thawing and degradation of permafrost (Jorgenson et al., 2001, 2006; Lantz and Kokelj, 2008; Walvoord et al., 2019). Given the large amounts of Hg that potentially could be released from Arctic soils in a warmer climate (Schaefer et al., 2020, Lim et al. 2020), methylation rates in the Arctic terrestrial environment have gained more attention. Permafrost degrades through two dominant mechanisms; first is the top down thaw of near-surface permafrost through the downward expansion of the seasonally thawed active layer (Douglas et al., 2021). This mechanism, which typically encompasses a broad areal extent but is not vertically extensive, generates wetter and more anoxic conditions that are potentially favorable for Hg methylating bacteria (Yang et al., 2016). While increased Hg methylation under such conditions has been demonstrated in temperate and tropical zones (Hsu-Kim et al., 2018), few studies have covered Hg methylation in zones with thawing permafrost (Fahnestock et al., 2019; Tarbier et al., 2021; Varty et al., 2021).

 The second mechanism for permafrost degradation is the development of thermokarst features through thaw subsidence of ice-rich frozen ground (Vonk et al., 2015; Olefeldt et al., 2016; Olefeldt et al., 2021). This typically leads to creation of a variety of features including scarps, pits, thaw slumps, and ponding of surface water that do not encompass large regions but are hotspots for geomorphologic processes. Thermokarst wetlands, ponds and lakes have been repeatedly identified as Hg methylation hotspots across the Arctic due to the concurrent mobilization of carbon and nutrients, creating conditions suitable for methylation. In previously formed thaw ponds, Lehnherr et al. (2012a, 2012b) 344 measured potential Hg methylation rates (1.8 to 40 ng/m²/day) similar to rates in already identified Hg 345 methylation hotspots. High concentrations of dissolved MeHg (ranging up to 22 ng L^{-1}), likely from either increased export of MeHg from peat soils, or *in situ* methylation in pond sediments, have also been found in eastern Canadian thaw ponds (MacMillan et al., 2015). Gordon et al. (2016) observed up to an order of magnitude higher MeHg concentrations in the poor fens associated with recent 349 permafrost thaw $(0.43 \text{ ng/L}, \text{filtered})$ than in the hydrologically isolated bogs $(0.04 \text{ to } 0.1 \text{ ng/L},$ filtered) in the Scotty Creek catchment of the discontinuous permafrost zone (in Canada's Northwest Territories). In the Stordalen Mire in Subarctic Sweden, MeHg concentrations increased by two orders

 of magnitude along the permafrost thaw gradient, from 0.01–0.05 ng/L in palsas underlain by permafrost to 1.0–2.0 ng/L in thawed fens (Fahnestock et al., 2019). The thawed fens were associated with heightened Hg accumulation, reducing conditions, higher sulfur concentrations and a greater proportional representation of putative Hg methylators (i.e. microbial strains carrying the Hg methylation gene pair but have not yet have been confirmed to methylate Hg in laboratory studies) within the microbial communities (from 0.5% to 7.8%), compared to the dry frozen palsas and bogs at intermediate stages of thaw (Fahnestock et al., 2019). In another Fennoscandian permafrost peatland study, an order of magnitude higher concentration of MeHg (and fraction of THg as MeHg) was found in topsoil (upper 50 cm) from collapse fens (edge of the fen receiving peat from the collapsing peat 361 plateau; 1.8 ± 4.7 ng g⁻¹; $6.1 \pm 6.5\%$ of THg) compared to the peat plateau soils $(0.17 \pm 0.55$ ng g⁻¹; 0.48 \pm 1.7% of THg; Tarbier et al. 2021). Median MeHg and %MeHg in the soil from the collapse fen $(1.8 \pm 4.7 \text{ ng g}^{-1}; 1.7 \pm 2.4\% \text{ of THg})$ was also 3.6 times higher than in the nearby fens assumed to have no recent history of permafrost, suggesting higher methylation of Hg in the recently formed fens. In line with these studies, Varty et al. (2021) observed higher %MeHg in soils saturated with water in comparison to drier soils collected on Ellesmere Island (Canada).

 Permafrost thaw has also been shown to influence microbial processes (Mackelprang et al., 2011) and biogeochemical cycling (Frey and McClelland, 2009) in downstream freshwater systems. Several recent studies have reported sites with permafrost slumps (ì.e. slope failures or mass-wasting features resulting from the thawing of ice-rich permafrost) are abundant with both organic matter compounds and Hg. In fact, the highest concentrations of THg and MeHg (1200 and 7 ng/L unfiltered, respectively) ever recorded in natural flowing freshwater were measured downstream of a permafrost thaw slump in the Northwest Territories of Canada (St. Pierre et al., 2018). Other studies show similarly elevated levels of both THg and MeHg in water and sediments from areas influenced by permafrost degradation (MacMillan et al., 2015; Gordon et al., 2016; Houben et al., 2016). In stratified thaw ponds, elevated MeHg concentrations and percentages in bottom waters suggest *in situ* methylation is occurring in anoxic waters (MacMillan et al. 2015). In addition to slump age, many factors may be contributing to the reported differences in organic matter and Hg levels downstream of

 permafrost slumps, highlighting the need for further study of how permafrost degradation will affect interactions between organic matter and Hg across various Arctic and Subarctic landscapes. Factors such as the depth of the permafrost active layer, peatland buffering and interferences as well as ground water influences may affect organic matter-Hg dynamics downstream of thermokarst processes (Frey and McClelland, 2009; Lim et al., 2019). Furthermore, thaw slumps are likely affecting a wide range of other biogeochemical processes that will either directly or indirectly alter Hg methylation or bioaccumulation in receiving water bodies (Pokrovsky et al., 2018).

4.1.2. Freshwater systems

 Arctic freshwater systems support several species of fish, which can be important in the traditional diet of some Indigenous peoples (Lockhart et al, 2005). Mercury enters lakes and streams primarily via terrestrial runoff and atmospheric deposition, but it can also originate from groundwater passing through unfrozen parts of permafrost below the water body (i.e. taliks). Inorganic Hg(II) entering 391 Arctic freshwater systems can either be photolytically reduced to $Hg(0)$ and evade back to the atmosphere, be methylated to MeHg or bind to particles and sink to the bottom sediments. Where methylation of Hg dominates in freshwater systems has been shown to vary between different Arctic freshwater bodies (see Table S2; Chételat et al., 2015).

 Mercury and MeHg dynamics in freshwater systems change with the seasons, having low or no mobilization during freezing conditions (November–May) which increases with higher river flows and flooding during the melting season from May to June. Methylmercury in terrestrial snow, for example, enters High Arctic lakes during snowmelt via melt streams, having higher concentrations during spring melt than during summer (Loseto et al., 2004). Methylmercury concentrations in Canadian Arctic freshwater systems are generally low ranging from 0.04 to 0.18 ng/L in unfiltered samples in rivers, 0.02 to 0.10 ng/L in lakes and greater than 0.02 to 0.53 ng/L in ponds (see Table 1). Compared to deeper lakes, shallow water bodies generally have higher MeHg due to warmer temperatures and higher organic matter available for microbial metabolism and are probable methylation hotspots (Lehnherr et al., 2012a; Lehnherr, 2014; MacMillan et al., 2015). Varty et al. (2021) showed that small, shallow Arctic lakes can reach higher concentrations of water column MeHg (up to 0.72 ng/L)

 in winter when ice covered. In addition, snow MeHg concentrations in spring (0.19-0.67 ng/L) suggest that snowpack may be an important MeHg source during the melt period. Water residence time may also play a role, with less within-lake processing of aqueous MeHg in smaller lakes (Richardson et al., 2021). In Alaskan lakes, sediment methylation has been shown to be the major source of MeHg to those ecosystems (Hammerschmidt and Fitzgerald, 2006). Varty et. al. (2021) also suggested sediment MeHg production is important for the annual MeHg budget of Skeleton Lake and can explain the elevated concentrations of MeHg observed during the ice-covered season.

 Generally, in freshwater ecosystems, the main carrier for MeHg is DOC (Tsui and Finlay, 2011), which has the double role of promoting the solubility and transport of MeHg in surface waters and attenuating dissolved MeHg uptake by aquatic food webs by limiting transport across cell membranes. High Arctic lakes typically have low biological productivity, limiting Hg uptake by algae and suspended detrital organic matter. At the same time, many Arctic lakes have low DOC concentrations, often below the suggested threshold of 8 to 11 mg/L DOC, above which DOC is hypothesized in some studies to counteract MeHg bioaccumulation (French et al., 2014; Braaten et al., 2018). For MeHg production, methylation of Hg in High Arctic wetland ponds was suggested to be negatively correlated 421 with DOC levels by decreasing Hg(II) availability to methylating microorganisms (Lehnherr et al., 2012a). While some studies suggest DOC concentrations may be declining in Arctic lakes (Saros et al., 2015), others show increases in DOC because of climate change, land use practices and other water chemistry interactions (Macdonald et al., 2005; Isles et al., 2020).

 During melting, most transported Hg is bound to particles. A total flux of (dissolved) MeHg flux from all Arctic rivers was estimated by Sonke et al. (2018) to be 584 kg/y. The larger variability

among different flux estimates for rivers, e.g. the Mackenzie River in various studies (72 kg/yr of

dissolved MeHg (Sonke et al. 2018), 14 kg/yr MeHg (Mu et al. 2019) and 13 and 15 kg/y MeHg

(unfiltered) (Emmerton et al, 2013)) stresses the need for additional studies to better constrain these

fluxes. The total Hg flux from larger Arctic rivers estimated by Sonke et al. (2018) was 27.6 Mg/yr

(13.4 Mg/yr as dissolved Hg and 13.9 as particulate Hg). The fate of Hg exported to the Arctic Ocean

is less clear. Coastal sediments and oligotrophic lake sediments in a High Arctic region of the

 Canadian Arctic Archipelago were found to methylate Hg, though at a low rate (St. Pierre et al., 2014; Hudelson et al., 2020).

4.1.3 Glaciers

 Methylmercury has been shown to be released in glacier-fed streams of the Canadian High Arctic (Ellesmere Island; St. Louis et al., 2005; St. Pierre et al., 2019), Yukon (Halm and Dornblaser, 2007) and Alaska (Nagorski et al., 2014; Vermilyea et al., 2017). Reported MeHg concentrations in unfiltered glacial meltwater vary from <0.010 to 0.26 ng/L and account for 1 to 26% of THg, while concentrations of MeHg in filtered meltwater vary from <0.010 to 0.075 ng/L and account for 4 to 32% of THg (see Table S3). Estimates of unfiltered MeHg fluxes in glacier-fed streams of Ellesmere Island are between 0.03 and 14.9 g/y, which translate to annual yields ranging from 0.49 to 11.7 443 mg/km²/y in basins with 21 to 82% glacier coverage (St. Pierre et al., 2019; see Table S3). An increase in the glacial sediment Hg flux in response to enhanced glacier melt rates might lead to higher exposure risks to inorganic Hg or MeHg in downstream aquatic ecosystems in some catchments. On land, changes might occur with the development of seasonally wet areas in newly deglaciated forelands where methylation of sediment-bound Hg could occur. In southeastern Alaska, for example, watersheds with greater wetland abundance have higher Hg concentrations, in both water and aquatic fauna, than glacierized basins (Nagorski et al., 2014). An increased supply of labile and bioavailable DOM from terrestrial sources such as wetlands or newly formed organic soils (e.g., Wietrzyk-Pełka et al., 2020) might also increase the risk of riverine Hg uptake by biota in these watersheds. It could also promote Hg methylation by sulfur-reducing bacteria in catchments where subglacial oxidation of sulfides causes enhanced sulfidization of ponds, wetlands and lakes (Graham et al., 2012). Cryoconite holes (dirt-filled melt cavities on glacier surfaces) are abundant over glacier margins or stagnant ice and can accumulate atmospherically deposited impurities released by ice melt, as well as microbes (Cook et al., 2016). These micro-environments could potentially act as local hotspots of Hg methylation in glacial environments, but direct evidence supporting this is presently limited to Tibetan glaciers (Zhang et al., 2020).

 In the marine environment, meltwater plumes from tidewater glaciers carrying both dissolved and particulate Hg may be injected into zones of locally high primary productivity in coastal surface waters, where subsequent methylation and biological uptake can occur (Schartup et al., 2015a). Some 462 glaciers of southern Alaska are sources of old (up to \sim 3900 y) and highly labile DOM, of possible subglacial bacterial origin, which is discharged into coastal waters (Hood et al., 2009, 2015). Likewise, retreating glaciers in fjords in southern Spitsbergen, Svalbard, discharge sediments containing up to 2.8% weight of OC (Kim et al., 2020a). Seafloor sediments in these fjords have THg concentrations of up to 84 ng/g. Hawking et al. (2021) recently reported dissolved MeHg concentrations from Greenland glacial catchments that were similar to values previously reported from wetlands. As discussed elsewhere (Dastoor et al., 2022) for THg, further studies are warranted to confirm the high levels of MeHg in these glacial catchments, particularly since the source(s) and processes leading to high MeHg yield remain unknown. Hg concentrations correlate closely with OC and are thought to be largely derived from glacially eroded sedimentary rocks (e.g., shales) in the glacierized basin. The discharge of Hg-bearing glacial sediments containing organic matter might promote Hg methylation and uptake by marine biota within the water column, which has not been well studied. However, as a recent and exhaustive review highlighted, the local effect of glacial runoff on coastal marine primary productivity is likely limited to a few kilometers beyond glacier termini, and whether runoff promotes or limits productivity varies between land- or marine-terminating glaciers (Hopwood et al., 2020). For example, observations in fjords in northwestern Svalbard suggest that glacial meltwater plumes could reduce the phytoplankton biomass (Piquet et al., 2014), and this might effectively limit MeHg production and uptake in the marine food chain. Furthermore, the high deposition rates of inorganic matter near tidewater glaciers, or at the mouth of proglacial channels, dilute the organic matter content in the seafloor sediments of Svalbard fjords (Liu et al., 2015), and this might limit benthic Hg methylation. On the whole, therefore, the environmental impact of Hg releases from melting glaciers and ice caps is likely to be local, rather than widespread across the Arctic. A study on the East Greenland shelf found no enhanced Hg or MeHg levels near the 79°N glacier, precluding large inputs or locally enhanced MeHg production (Petrova et al. 2020).

4.2. Marine systems

4.2.1 Seawater

MeHg concentrations in the western Arctic Ocean have been reported to be 0.011±0.010 ng/L in

- filtered seawater (Agather et al., 2019, Figure 3) and 0.046±0.024 ng/L in unfiltered seawater (Wang
- et al., 2018). Heimbürger et al. (2015) found similar levels of 0.045±0.017 ng/L and 0.034±0.022 ng/L
- in unfiltered seawater in the Eastern Arctic and Central Ocean. Methylmercury (MeHg) inflow to the
- Arctic Ocean totalled 9±2 Mg/yr, most of which enters with deep and mid-depth waters of the Atlantic
- Ocean (7.5±2 Mg/yr; Petrova et al., 2020). Similar to THg, more MeHg flows out of the Arctic Ocean
- 494 into the North Atlantic (16.5±3 Mg/yr, 14±2 Mg/yr via the Fram Strait, 2.5±1 Mg/yr via the Davis
- Strait; Petrova et al. 2020) than flows in. The MeHg outflow via the Fram Strait conserves the shallow
- MeHg peak found throughout the Arctic Ocean, and an average DMHg:MMHg ratio of 2.5:1 (Petrova
- et al., 2020). *In situ* MeHg formation in the Arctic Ocean water column (Lehnherr et al., 2011; Wang
- et al., 2012b, 2018; Heimbürger et al., 2015; Soerensen et al., 2016b; Petrova et al., 2020) results in a

net MeHg outflow of 7.5± 2Mg/yr (Petrova et al. 2020).

A subsurface enrichment of MeHg in seawater has been observed in almost all major ocean basins

(AMAP/UNEP 2019), including the Atlantic Ocean (Bowman et al., 2015; Bratkič et al., 2016),

Pacific Ocean (Sunderland et al., 2009; Hammerschmidt and Bowman, 2012; Munson et al., 2015;

Bowman et al., 2016; Kim et al., 2017), Southern Ocean (Cossa et al., 2011; Gionfriddo et al., 2016);

Mediterranean Sea (Cossa et al., 2009, 2012, 2017; Heimbürger et al., 2010), Baltic Sea (Soerensen et

- al., 2016b) and the Black Sea (Rosati et al., 2018) basins. However, the Arctic Ocean basin differs
- from the others because the MeHg enrichment occurs at a much shallower depth (100–300 m depth)
- where dissolved oxygen saturation is well above 75% (Wang et al., 2012b, 2018; Heimbürger et al.,
- 2015; see Figure 4). The Arctic subsurface enrichment of MMHg and DMHg is also detected in water
- exiting the Arctic Ocean to the North Atlantic Ocean (Petrova et al. 2020). Heimbürger et al. (2015)
- first hypothesized that the characteristic shallow MeHg peaks lead to enhanced biomagnification and
- may be an alternative explanation for high arctic biota Hg levels, rather than anthropogenic Hg inputs.
- As this subsurface MeHg maximum lies within the habitat of zooplankton and other lower trophic

 level biota it has been suggested that biological uptake of the subsurface MeHg and subsequent biomagnification explain a long-standing mystery in the Canadian Arctic on why marine animals in the western Arctic have higher Hg levels than those in the east (see Figure 4; Wang et al., 2018). Increasing amounts of bioavailable Hg in surface waters could have an effect on Hg accumulation at the bottom of Arctic marine food webs. THg levels of up to 242 ng/g have been reported in zooplankton from Hudson Bay (Foster et al., 2012). As discussed further below, regional differences in food web length (and thus biomagnification) have also been provided as a contributing factor explaining the higher Hg levels marine animals in the western Arctic (St. Louis et al., 2011; Brown et al., 2016).

 The mechanisms of subsurface MeHg formation in the Arctic Ocean and other ocean basins in the world is under debate. Most of the studies have attributed subsurface MeHg to *in situ* Hg methylation in the water column associated with organic matter remineralization (Sunderland et al., 2009; Cossa et al., 2009; Wang et al., 2012b; Heimbürger et al., 2010, 2015), yet the biotic or abiotic processes that are responsible for methylation remain unidentified (Bowman et al., 2020a). Recent work in the Canadian Arctic (Wang et al., 2018) and the Siberian Shelf (Kim et al., 2020) suggest that advection of MeHg produced in shelf sediments also could contribute to the subsurface peak of MeHg found in the water column. This debate calls for further studies on MeHg sources and dynamics in the Arctic and global oceans.

 Figure 3. Monomethylmercury (MMHg) and dimethylmercury (DMHg) and dissolved oxygen (O2) in 534 filtered seawater along a looped latitudinal section (\sim 75°N to the North Pole and then back to \sim 73°N) in the western Arctic Ocean. The top panel shows sampling station locations along the section.

Adapted from Agather et al., 2019.

 Figure 4. Methylmercury (MeHg) concentrations in unfiltered seawater (top panel) and the marine food web (bottom panel) along a longitudinal (west-to-east) section (the red line in the bottom panel) across the Canadian Arctic and Labrador Sea. Bar charts in the bottom panel show mean concentrations of monomethylmercury (MMHg) in *Calanus* spp. and *Themisto* spp. collected from 1998 to 2012 (±1SD), THg in muscle of adult ringed seals collected in 2007 and 2011 and THg in liver of polar bears (*Ursus maritimus*) collected from 2005 to 2008. Adapted from Wang et al., 2018.

4.2.2. Marine sediment

 Methylmercury concentration measurements in deep Arctic Ocean sediments are sparse. Based on a range of means, MeHg concentrations in Beaufort Sea and Chukchi Sea shelf sediments were estimated to range from 0.15 to 0.37 ng/g (dry weight, Fox et al., 2014). A recent study in the East Siberian Sea shows that MeHg concentrations range from 0.09 to 0.17 ng/g in shelf sediments, with much lower concentrations (~0.003 ng/g dry weight) in the slope sediments (Kim et al., 2020b). Liem- Nguyen et al (2022) reported MeHg concentrations ranging from 0.19 to 2.6 ng/g in Laptev Sea sediments along a 330 km long cross-shelf transect. The higher concentrations of MeHg (and MeHg/OC ratio) observed close to the Lena river delta than further out on the shelf was explained by trapping of terrestrial MeHg close to the river delta. Based on estimated MeHg concentrations of 0.2 ng/g (dry weight) for shelf sediments and 0.05 ng/g (dry weight) for the deep basin, Soerensen et al. 571 (2016b) estimated a MeHg sedimentation flux of 1.3 Mg/yr and 0.1 Mg/yr to shelf and deep basin, respectively. Resuspension was not estimated due to an absence of data. Soerensen et al., (2016b) 573 estimated MeHg diffusion to the overlying water column at 0.9 Mg/yr and 0.1 Mg/yr from shelf and deep basin sediments, respectively, for a total flux of 1.0 Mg/yr. However, a recent study in the East Siberian Sea suggests that the MeHg diffusion flux from sediments to the water column could be much larger, amounting to 4.6±0.2 Mg/yr for the Arctic shelf system alone (Kim et al., 2020b). There is a need to better constrain the benthic MeHg flux.

4.2.3. Sea ice and snow

The sea ice environment, including first-year ice, multi-year ice and the overlying snow cover,

provides a spatially and temporally dynamic interface between the ocean and the atmosphere in the

Arctic Ocean where physical transport, chemical reactions, and biological uptake occur (Wang et al.,

- 2017). While sea ice is semi-permeable (e.g., Loose et al. 2010), elevated Hg(0) concentrations under
- sea ice compared with ice-free waters suggest that ice is a barrier for gas exchange (Dimento et al.
- 2019). The sea ice environment is also an important habitat and feeding ground for many organisms,

 including bacteria, algae, zooplankton and larger mammals such as polar bears and whales. Beattie et al. (2014) found elevated MeHg concentrations in multi-year Arctic sea ice of up to 0.5 ng/L (unfiltered) in the lower part of the ice (see Figure 5). Recent work has shown concentrations of both MMHg and DMHg in sea ice, especially within first-year ice (4.4% MMHg vs. 1.5% in multi-year ice) from across the central Arctic Ocean (Schartup et al., 2020). Snow, which covers large parts of the Arctic landscape as well as sea icescape has a large interstitial surface area for air-snow exchange of Hg and plays a major role in atmospheric mercury depletion events (Douglas et al., 2012, 2017; Steffen et al., 2014). However, Douglas et al. (2012) report that the small amounts of MeHg found in snowpack before spring snowmelt does not add significantly to MeHg levels in aquatic systems. As the high concentrations of MeHg in the bottom layer of sea ice (Beattie et al., 2014) cannot be explained by downward transport of MeHg from the overlying snow (Gionfriddo et al., 2016; Soerensen et al., 2016b), it has been postulated that Hg methylation could take place in sea ice. Discrete peaks in MMHg and DMHg concentrations within sea ice cores (Schartup et al., 2020) also suggest that they may be produced by microorganisms within the sea ice. This is supported by the fact that microbes present in snow and ice are metabolically active even under freezing temperatures (Douglas et al., 2012). Methylation of Hg has earlier been suggested to be strictly anaerobic and to occur in the anaerobic brine environment (droplets, channels or pockets of salt solution in sea ice), or in the lower part of the ice, due to the involvement of sulfate-reducing bacteria (Chaulk et al., 2011; Gilmour et al., 2013; Beattie et al., 2014). During spring melt, heterotrophic activity could lead to oxygen depletion in the bottom of the ice column and denitrification processes that favor anoxic environments could then be important in the lower parts of the ice (Rysgaard and Glud, 2004). A study in the Antarctic has suggested methylation of Hg in sea ice might be carried out by aerophilic bacteria, such as *Nitrospina*, or by sea ice phytoplankton (Gionfriddo et al., 2016), but this has yet to be confirmed in the Arctic. Gionfriddo et al. (2016) suggest that sea ice phytoplankton play a role in the stimulation of methylation of oxidized Hg that may occur within brine pockets and periphytic biofilms where organic matter is trapped and decaying. Due to the seasonality of primary production, MeHg formation could show seasonal variations. Chlorophyll a (Chla) showed a covariation with MeHg,

 which also has been shown in other studies (Antarctica: Cossa et al., 2011; Arctic: Beattie et al., 2014), suggesting a microbial mercury methylation and the involvement of phytoplankton and phototrophs. Gionfriddo et al. (2016) also identify Proteobacteria, which possesses the *mer* operon, as a potentially important Hg reductive phylum. It should be noted that direct support for the production of MeHg in Arctic snow and ice (from e.g. Hg methylation assays) is still limited. From terrestrial sites, where Hg methylation and demethylation assays have been conducted on intact snow or melted snow, low or non-detectable potential Hg methylation rates were reported (Varty et al. 2021; Willis et al., 2018). In addition, both Varty et al. (2021) and Willis et al. (2018) found low methylation to demethylation ratios and concluded that *in situ* MeHg production likely did not contribute significantly to the pool of MeHg observed in the snowpack.

 $120°W$

115°W

 110° W

105°W

125°W

624 **Figure 5** Profiles of total Hg (Hg_T) methylmercury (MeHg_T) concentrations and the ratio of MeHg to THg (MeHg:THg) in multi-year sea ice sampled at two sites (MY2 and MY3) in the eastern Beaufort Sea and the M'Clure Strait. Adapted from Beattie et al., 2014.

 The uptake and transport of Hg and subsequent formation of MeHg in sea ice is determined by its thickness and permeability, which varies dramatically with temperature and season. During winter, the ice is cold and less permeable, which slows the transport of Hg species through the ice. However, exchange continues with the seawater as the bottom of the ice is warmer (greater permeability) and with the atmosphere as brine exclusion expels Hg to the overlying snowpack. In spring, the ice warms up and the brine volume increases. Contaminants such as Hg then percolate from snow to ice and could lead to their flushing into the underlying seawater. Summer sea ice melt leads not only to dissolved Hg being released from the ice but also particle bound Hg and MeHg being incorporated in the bulk ice (Wang et al., 2017). The presence of sea ice also decreases photodegradation of MeHg in underlying water (Ebinghaus et al., 2002, Point et al. 2012). The dynamic motion of sea ice further makes it possible to transport Hg and MeHg over time and space as the ice can drift over large areas in the Arctic Ocean, depending on winds and underwater currents. The export of MeHg via drifting sea ice has been calculated by Wang et al. (2017) at less than 4% of the total Hg export from the Arctic Ocean. Petrova et al. (2020) estimated a net export of MeHg in sea ice of 0.4 Mg/yr, considering 10% of the total Hg export is in the form of MeHg.

 The total MeHg pool in sea ice in the Arctic Ocean was estimated by Schartup et al. (2020) to be 0.33 Mg, which is consistent with estimates of 0.3 to 2.6 Mg, based on studies performed in the M'Clure Strait and Beaufort Sea (Beattie et al., 2014; Soerensen et al., 2016b).

5. Processes affecting mercury accumulation in Arctic biota

 Methylmercury concentrations in fish and wildlife vary within and across geographic regions in the circumpolar Arctic, reflecting the variability in Hg loading to the ecosystem and environmental conditions that control *in situ* production of MeHg, both of which are discussed above. It also reflects on differences in biological uptake at the base of the food web, and ecological characteristics that control trophic transfer of MeHg within the food web (see Dietz et al., this issue). All of these conditions can change over time resulting in temporal variation of MeHg concentrations in animals (see Morris et al., this issue). The delineation of specific conditions promoting MeHg bioaccumulation is important for identifying Arctic ecosystems at greater risk of MeHg exposure (Munthe et al., 2007). Recent research has identified key environmental characteristics that enhance MeHg bioaccumulation in the Arctic.

Mercury speciation determines (a) how Hg is transported between different environmental

compartments, (b) organismal exposure, (c) the potential toxicity of the exposure to the organism, (d)

the extent to which Hg is distributed between tissues within the organism, (e) the toxicity of the Hg

661 compound (e.g., MMHg, DMHg, Hg(II)), (f) organism assimilation and excretion, and (g) its

propensity to bioaccumulate and biomagnify through the food web. In this section, the uptake of Hg at

 the base of Arctic aquatic and terrestrial food webs is described, with an emphasis on the biomagnified chemical form MeHg.

5.1 Mercury uptake at the base of Arctic food chains

 The bioconcentration of aqueous MeHg in bacteria and algae, either suspended in the water column (plankton) or growing in biofilms on surfaces such as sediment, ice and rocks, is understood to be a critical step in the transfer of Hg to top predators in freshwater and marine environments. Aquatic food webs with greater MeHg uptake by microorganisms also have higher concentrations in subsequent trophic levels (Wu et al., 2019), and greater uptake at the base of terrestrial food webs likely has the same consequence (Tsz-Ki Tsui et al., 2019).

 Despite the critical role of phytoplankton as the dominant energy source for food webs in the ocean, few measurements of MeHg have been made of Arctic phytoplankton. In the Subarctic estuary of Lake Melville, phytoplankton contained 0.03 ng/g dw of MeHg (Schartup et al., 2015a), which is low relative to concentrations in marine phytoplankton at more temperate latitudes (Schartup et al., 2018). Phytoplankton of the Chukchi Sea had MeHg concentrations below the study analytical detection limit of 1.5 ng/g dw (Fox et al., 2017). Cellular uptake of MeHg by phytoplankton may occur by passive or active transport across the cell membrane (Lee and Fisher, 2016; Skrobonja et al., 2019). Smaller phytoplankton cells have larger surface to volume ratios and, consequently, bioconcentrate more MeHg than larger cells (Lee and Fisher, 2016; Schartup et al., 2018). Smaller phytoplankton are also known to stimulate MeHg production, because they have a higher surface to volume ratio and are more effectively remineralized (Heimbürger et al., 2010). In the Arctic Ocean, water column MeHg 684 concentrations are often highest at subsurface depths of \sim 100–200 m, where the proximity to phytoplankton may enhance uptake into pelagic food webs (Heimbürger et al., 2015; Wang et al., 2018). Arctic estuaries may also be important zones for MeHg uptake by phytoplankton because of salinity-driven water-column stratification and the stimulation of MeHg production by inputs of terrestrial organic matter (Schartup et al., 2015b). Seasonal changes also affect Hg uptake by marine phytoplankton, such as in the Baltic Sea, where reduced sea ice cover and eutrophication enhanced bioaccumulation during warmers months (Bełdowska and Kobos, 2016; Soerensen et al., 2016a). Sea ice is another habitat where microbial uptake of MeHg may be critical for basal uptake into food webs. As stated before, MeHg production can occur in polar sea ice (Beattie et al., 2014; Gionfriddo et al., 2016; Shartup et al., 2020) and in the marginal sea ice zone (Heimbürger et al., 2015). Algal growth on the under-ice surface is a significant energy source for pelagic food webs in the Arctic (Kohlbach et al., 2016). Burt et al. (2013) characterized uptake of inorganic Hg by ice algae during a

spring bloom associated with first-year ice and suggested that this could represent an important entry

point for marine invertebrates grazing either directly on the ice or on settling particulate matter

released from melting ice.

 Methylmercury within the sea ice can be taken up by ice-associated biota, algae, under-ice amphipods and ice-dependent animals such as fishes, polar bears and birds and can bioaccumulate in marine food chains throughout the year. When sea ice melts, Hg and MeHg in the ice are released to the surface ocean. Fast settling of sea ice algae upon melt (Boetius et al. 2013) may also be an important transport route for MeHg into the deep ocean, but this needs further research.

 With greater light penetration into the water column, the melting period is also a period of high primary production in surface water, and the concurrent release of contaminants during this period could lead to additional uptake of contaminants within the marine food web. Mercury attached to particles is released from the ice during complete ice melt. Sinking particles may be remineralized in the water column, releasing Hg for uptake by pelagic microbes, or reach the sediment and become a source of Hg available for uptake by benthic organisms. Hg uptake by sea ice algae is only limited by the amount of Hg available for uptake when spring blooms commence (Burt et al., 2013). This is important since ice algae represent 10 to 60% of annual primary production in the Arctic, and they may become even more abundant under mild climate conditions. Burt et al. (2013) estimate that with the replacement of multi-year sea ice with first-year ice due to climate change, an extra 48 kg/y of particle-bound Hg could enter the Beaufort Sea in the coming years.

 Benthic algae and biofilms are a dominant primary energy source for food webs in many Arctic lakes (Vadeboncoeur et al., 2003; Chételat et al., 2010; Rautio et al., 2011). In the eastern Canadian Arctic, MeHg concentrations ranged from 2 to 12 ng/g dw for rocky shoreline biofilms (Chételat et al., 2018), 0.3 to 1.3 ng/g dw for microbial mats (Camacho et al., 2015), and 2 to 56 ng/g dw for periphyton and filamentous algae (van der Velden et al., 2013a). In comparison, MeHg concentrations of water- column seston measured in some of those same lakes ranged from less than 0.4 to 30 ng/g dw, and bioconcentration factors of MeHg in biofilms and seston were higher in lakes with low aqueous DOC (Chételat et al., 2018).

Invertebrates play a key role in the trophic transfer of MeHg entering food webs via microorganisms.

In the last decade, large amounts of data were compiled on MeHg concentrations in marine

zooplankton from across the circumpolar Arctic (Foster et al., 2012; Pućko et al., 2014; Pomerleau et

 al., 2016). Zooplankton MeHg concentrations ranged from 8 to 96 ng/g dw and were highest in the southern Beaufort Sea region compared to the Chukchi Sea, Canadian Arctic Archipelago, Hudson Bay and northern Baffin Bay (Pomerleau et al., 2016; Wang et al., 2018). Larger taxa of marine zooplankton such as *Themisto* and *Paraeuchaeta* predate on smaller zooplankton, and biomagnification of MeHg was found between omnivorous and herbivorous taxa (Foster et al., 2012; Pomerleau et al., 2016). Recent measurements of MeHg concentrations in zooplankton from Arctic lakes (Lescord et al., 2015; Chételat et al., 2018, Varty et al., 2021) add to data from earlier studies (Chételat and Amyot, 2009; Gantner et al., 2010), which showed that zooplankton in lakes can have 734 higher concentrations (>100 ng/g dw) than those in the marine environment. In a freshwater system on Ellesmere Island (Canada), zooplankton MeHg concentrations were lowest under ice, highest at the onset of ice-free conditions and then subsequently declined, indicating considerable seasonality in bioaccumulation (Varty et al., 2021). While copepods tend to dominate in lacustrine zooplankton communities in the Arctic, large-bodied grazers (Cladocera, *Daphnia*) are also present in some systems and can have elevated MeHg concentrations (Chételat and Amyot, 2009; Lehnherr et al., 2012a; Lescord et al. 2015; MacMillan, 2018). Modelling shows a potential for increase distribution of daphnia across Arctic lakes due to climate-induced changes in lake productivity, a trend that could enhance MeHg bioaccumulation patterns in pelagic food chains (Chételat and Amyot 2009). The developmental stage can also influence the MeHg concentration of Arctic freshwater copepods, with dilution occurring as adults accumulate lipid stores (Chételat et al., 2012).

 A considerable number of studies of Arctic food webs have also characterized Hg concentrations in a variety of benthic invertebrate taxa in marine environments (Bidleman et al., 2013; van der Velden et al., 2013a; Fox et al., 2014; Jörundsdóttir et al., 2014; Clayden et al., 2015; McMeans et al., 2015; Góngora et al., 2018; Pedro et al., 2019) as well as in lakes (van der Velden et al., 2013a; Chételat et al., 2015, 2018; Kahilainen et al., 2016, 2017; Keva et al., 2017; Rohonczy et al., 2020). These studies showed variation in Hg bioaccumulation among benthic invertebrate taxa, which was partly related to

trophic position.

5.2. Processes affecting mercury bioaccumulation at higher trophic levels

 Mercury concentrations in fish and wildlife reflect ecological processes that influence dietary exposure as well as animal physiology that affects internal accumulation of Hg (Bradley et al., 2017, Chételat et al., 2020). Trophic position, shifts in diet, habitat-specific feeding, cross-ecosystem subsidies and movement or migration can be important determinants of dietary Hg consumed by fish and wildlife. Differences in the extent of assimilation and elimination of MeHg related to sex, age, body condition and species also affect animal Hg burdens. Recent advances have identified Arctic-specific processes affecting Hg bioaccumulation in fish and wildlife.

 Arctic marine ecosystems have long food chains (~4–5 levels), and top predators (such as polar bear, ringed seal, beluga whale, and Greenland shark) have elevated Hg concentrations due to feeding at a higher trophic position (St. Louis et al., 2011; McMeans et al., 2015; Brown et al., 2016, 2018). These slow-growing, long-lived top predators are prone to long-term Hg bioaccumulation with age (Masbou et al., 2018; Ewald et al., 2019). Recent quantification of food-chain length using nitrogen stable isotopes demonstrated that regional differences in Hg concentrations of top predators, such as between western and eastern regions of the Canadian Arctic (St. Louis et al., 2011; Brown et al., 2016) is, in part, due to food-chain length. There can also be spatial variation in the carbon sources which support Arctic marine food webs, with potential consequences for food web bioaccumulation of Hg (Cyr et al., 2019). Recent variation of Hg concentrations in beluga whales reflect complex influences of diet, foraging habitat (e.g., benthic versus pelagic feeding), and climate drivers (Desforges et al., 2022; Loseto et al., 2015, Loseto et al., 2008). Considerable research effort has also focused on measuring the rate that Hg biomagnifies through food webs in Arctic ecosystems. A global meta-analysis showed that biomagnification rates were higher at northern latitudes compared with the tropics (Lavoie et al., 2013). Within the Arctic, Hg biomagnification rates can vary among marine and freshwater food webs (Clayden et al., 2015; Lescord et al., 2015; McMeans et al., 2015; Ruus et al., 2015; Fox et al., 2017); although in a study of co-located marine and lacustrine food webs of Arctic char, no difference was found in biomagnification rates (van der Velden et al., 2013b). While some evidence suggests that

 cold, low productivity ecosystems have higher Hg biomagnification (Lavoie et al., 2013), factors controlling the biomagnification rate of Hg through Arctic food webs remain unclear. Marine subsidies to the diet of terrestrial carnivores and freshwater fish can strongly affect Hg bioaccumulation in the Arctic. Wolves and Arctic foxes that scavenged on carcasses of marine mammals had higher Hg concentrations than those that had an exclusively terrestrial diet (Bocharova et al., 2013; McGrew et al., 2014; Hallanger et al., 2019). In the case of Arctic fox at Svalbard, interannual variation in their Hg concentration was positively related to the extent of sea ice cover, which provides a platform to access marine carcasses (Hallanger et al., 2019). Sea-run Arctic char tend to be larger and have faster growth rates than their lacustrine conspecifics, and therefore they have lower muscle Hg concentrations (Swanson et al., 2011; van der Velden et al., 2013b). The lower Hg bioaccumulation in sea-run char was also attributed to lower Hg exposure at the base of marine food webs relative to lakes in the Arctic (van der Velden et al., 2013a).

 Migration by wildlife such as seabirds and marine mammals can influence their dietary Hg exposure. Little auks (*Alle alle*) that breed in Greenland during summer and overwinter off the coast of eastern Canada were found to accumulate more Hg during the wintering period (Fort et al., 2014). Similarly, yellow-billed loons (*Gavia adamsii*), which breed in lakes of Arctic North America and overwinter in marine areas of eastern Asia, showed evidence of higher winter exposure (Evers et al., 2014). An evaluation of seasonal variation in Hg contamination of nine species of Arctic seabirds showed, on average, three times higher Hg concentrations in feathers grown at lower latitudes during the non- breeding period than those grown while breeding in the Arctic (Albert et al., 2021). Greater Hg bioaccumulation was found for seabirds overwintering in the western Atlantic relative to the eastern Atlantic or the Pacific region (Albert et al., 2021). Thus, consideration of overwinter exposure at lower latitudes is necessary for interpreting Hg bioaccumulation in migratory Arctic seabirds. Bowhead whales (*Balaena mysticetus*) undergo long seasonal migrations within the Arctic in response to sea ice conditions and prey availability, and examination of their baleen plates provided a record of temporal 804 variation in Hg exposure (Pomerleau et al., 2018). Annual cycles of Hg and δ^{13} C values (an isotopic

 indicator of diet) in the baleen plates showed seasonal shifts in diet and Hg accumulation by bowhead whales in both the eastern and western Arctic.

 Biological transport is also a pathway for the mobility of contaminants in the Arctic environment such as from migratory spawning fish and bird colonies (Michelutti et al. 2010, Choy et al. 2010, Kristiansen et al., 2019). Migrating salmonids that return to remote freshwater ecosystems to spawn act as important transport (bio)vectors of Hg from ocean to rivers (Blais et al., 2007). Similarly, high- density nesting of seabirds and their deposition of guano can lead to inputs of marine-derived nutrients and Hg to nearby ponds and tundra (Michelluti et al., 2010; Kristiansen et al., 2019, Choy et al., 2010). The body condition of fish and wildlife is sometimes correlated with their Hg concentration, although reported observations have been both positive and negative. Trudel and Rasmussen (2006) showed that Hg concentration may be either positively or negatively correlated to changes in growth and growth efficiency, depending on how fish allocate energy between activity costs and growth rates. Mercury concentrations of northern pike were negatively correlated with growth rate though also influenced by water chemistry and characteristics of the subarctic catchments (Moslemi-Aqdam et al., 2022). Tissue Hg concentrations were negatively correlated with body condition in sea-run char (Evans et al., 2015), caribou (*Rangifer tarandus groenlandicus*; Gamberg et al., 2016) and little auks (Amélineau et al., 2019), while positive correlations were observed for eider (Provencher et al., 822 2016). For polar bear, both negative and positive correlations were found between tissue Hg concentrations and body condition (Bechshoft et al., 2016; McKinney et al., 2017). Those contrasting observations reflect several potential underlying physiological processes driving the effect of body condition. Fasting or lower food availability and the subsequent loss of body mass can concentrate Hg already in tissues (Keva et al., 2017; Seewagen et al., 2016; Peterson et al., 2018; Martyniuk et al. 827 2020). The reverse has also been suggested, that poorer body condition may be a negative effect of a higher Hg burden (Provencher et al., 2016; Amélineau et al., 2019), although this has been discounted for northern pike (*Esox lucius*; Sandheinrich and Drevnick, 2016). A positive association between body condition and Hg concentration could result from greater Hg intake associated with increased feeding (Bechshoft et al., 2016). Given these bioenergetic influences, seasonal and interannual

changes in food resource availability in the Arctic can have important consequences for Hg

concentrations in animals (Amélineau et al., 2019; Eckbo et al., 2019; Hallanger et al., 2019).

5.3. Organic matter influences on mercury bioaccumulation

 While numerous water chemistry parameters have been shown to alter MeHg bioaccumulation in Arctic systems, organic matter has complex linkages with the Hg cycle that can both dampen or promote MeHg bioaccumulation. The complexity of these interactions is reflected in the varying relationships reported between organic matter and Hg in the literature (Lavoie et al., 2019). A recent global review found these relationships to be broadly influenced by geographic location and temporal 841 scale, as well by as human activities (Lavoie et al., 2019). The chemical composition of DOC also plays a critical role in its effects on Hg biogeochemical cycling (Schartup et al., 2015b; Lescord et al., 2018; Jiang et al., 2020), including bioaccumulation (see Section 4.4). For example, OC of terrestrial origin is involved in the transport of MeHg to downstream freshwater and marine environments. In the Subarctic Baltic Sea, interannual variation of seawater MeHg concentrations were related to DOC inputs and composition (Soerensen et al., 2017). In Arctic fresh waters, DOC can promote or inhibit the photodegradation of MeHg depending on the amount of DOC present (Girard et al., 2016) and alter Hg(II) bioavailability to microbes (Chiasson-Gould et al., 2014) because of the multiple roles that organic matter plays in those processes. Dissolved organic carbon can play a key role in promoting MeHg bioaccumulation, although those interactions are complex, and consideration of site-specific conditions are likely important.

 Numerous studies on the effects of organic matter on Hg accumulation in Arctic biota have been conducted in lakes and rivers across the Arctic over the past ten years. For example, Hudelson et al. (2019) recently reported strong negative correlations between DOC and POC concentrations versus total Hg (THg) levels in landlocked Arctic char (*Salvelinus alpinus*) in lakes around Resolute Bay in Nunavut, Canada. In the same set of lakes, Lescord et al. (2015) reported negative correlations between DOC (in combination with total nutrient levels) and MeHg concentrations in zooplankton. In High Arctic lakes, waters with very low concentrations of DOC were associated with greater bioaccumulation of MeHg at the base of the food web (e.g., chironomid larvae and zooplankton) even when concentrations of MeHg in the water were low (Chételat et al., 2018). Overall, these results suggest that Arctic aquatic systems with lower DOC concentrations, in combination with other physicochemical variables, may have higher rates of Hg bioaccumulation in biota. However, variation in the influence of DOC likely occurs. Furthermore, climate change is rapidly altering both carbon and Hg cycling, resulting in a general increase in concentrations of organic matter in aquatic environments across the Arctic (MacDonald et al., 2005; Isles et al., 2020; Chételat et al. 2022).

 Some research in Arctic and Subarctic systems has also suggested that the relationship between biotic Hg and organic matter concentrations may not be linear and may, in fact, be dependent on DOC concentrations. More specifically, several studies have reported a curvilinear relationship between 869 DOC concentrations and THg or MeHg concentrations in aquatic biota, with an inflection point of ~8– 11 mg/L of DOC (French et al., 2014; Braaten et al., 2018). This relationship was first suggested based on data from the Adirondack lakes and is dependent on system pH (Driscoll et al., 1994). However, given that concentrations of DOC are often strongly correlated with aqueous concentrations of both THg and MeHg, the curvilinear relationship between DOC and biotic Hg concentrations warrants further investigation with consideration of other factors such as water chemistry and trophic position (Chételat et al., 2018; Broadley et al., 2019). For example, Chételat et al. (2018) found that the ratio of aqueous MeHg concentrations to DOC concentrations was a strong and linear predictor of invertebrate MeHg concentrations across an array of Arctic lakes.

 Recent studies, some related to this curvilinear Hg-DOC relationship, also consider the effect of various measures of DOM quality rather than quantity (i.e., DOC concentrations) have on Hg bioaccumulation. French et al. (2014) suggest that changes in DOM compounds and bioaccessibility may account for changes in Hg bioavailability and bioaccumulation in Arctic freshwater invertebrates; at lower DOC concentrations, Hg was found to be bound with smaller fulvic acids, while at higher DOC concentrations Hg was bound with larger and less bioaccessible humic acids. In the Subarctic, biotic Hg concentrations were similarly higher in systems with higher proportions of more

 bioaccessible DOM compounds and lower proportions of large and aromatic humic substances (Lescord et al., 2018). Conversely, other studies have reported that humic lakes are associated with greater MeHg bioaccumulation in Subarctic boreal ecoregions (Braaten et al., 2018; Poste et al., 2019). Future studies on Arctic Hg and DOM interactions may benefit from incorporating simple structural measurements, such as SUVA254, which provides an estimate of DOM aromaticity (e.g., Skyllberg et al., 2009; Burns et al., 2013).

 While several studies have shown changes in aqueous Hg levels in relation to permafrost degradation and changing organic matter dynamics, none to our knowledge have examined their combined effect on biotic Hg levels, Hg bioaccumulation potentials, or rates of biomagnification. In the retrogressive thaw slump in the western Canadian Arctic, over 95% of the Hg was released in the particulate phase (St. Pierre et al. 2018) and may not be readily bioavailable. Furthermore, a recent review by Tank et al. (2020) found considerable variation in DOC concentrations and DOM quality among studies examining the impact of permafrost thawing on biogeochemical cycling across aquatic networks. Therefore, future studies assessing the impact of thawing permafrost on downstream bioaccumulation 899 patterns would benefit from considering the effect of DOM, including the structural or quality measurements discussed above.

 Given the biogeochemical cycling of Hg in the Arctic environment is sensitive to climate change, its effects on MeHg production and breakdown in the Arctic, including the connection between climate

 Future mapping of microbial communities carrying the genes involved in methylation and demethylation of Hg should be carried out as such mapping will provide critical information on these processes across the Arctic, as well as reveal key linkages to the biogeochemical

 cycle of other elements such as carbon (e.g. by understanding the involvement of methanogens in Hg methylation).

The thawing of permafrost represents a potential source of both DOM and Hg to Arctic and

- *Subarctic aquatic environments; future work should examine the effects of permafrost-derived*
- *organic matter on Hg bioaccumulation in Arctic biota, which may add to our understanding*
- *of the complex Hg-DOM relationship as permafrost thawing increases in a changing climate.*
- *Future research should also be directed towards a better understanding of the formation and environmental fate of DMHg in the Arctic Ocean, and its role in controlling the pool of Hg available for bioaccumulation in marine and coastal food webs.*

Where are the hotspots of methylmercury in the Arctic?

- 8. We previously knew that the production of MeHg in natural systems was controlled by substrate availability (e.g., inorganic Hg and organic matter), the presence and abundance of microbial Hg methylators, and MeHg degradation processes. Thus, anoxic systems, such as organic-rich sediments and flooded soils, are understood to be typical hotspots for MeHg production in the Arctic as elsewhere.
- 9. In the terrestrial environment, foliar uptake of Hg from the atmosphere, a process that could be enhanced in recently or soon-to-be deglaciated foreland, is a key driver for sequestering inorganic Hg that may be available for methylation.
- 10. Permafrost thaw in low-relief regions with ice-rich soils and sediments may result in the
- formation of thermokarst wetlands, ponds and lakes that have been repeatedly identified as Hg
- methylation hotspots across the Arctic due to the concurrent mobilization of carbon and nutrients, creating conditions suitable for methylation.
- 11. Locally significant MeHg production may occur in newly-deglaciated forelands (especially where wetlands develop), in cryoconite holes on the margins of glacier surfaces or margins,

Acknowledgements

This review is a result of efforts led by the Arctic Monitoring and Assessment Programme (AMAP)

- and the AMAP Mercury Expert Group to produce the 2021 AMAP Mercury Assessment. We thank
- Rune Dietz (Aarhus University) and Simon Wilson (AMAP) for provided guidance and support.
- Graphics production and technical editing were supported by AMAP. We also acknowledge Adam
- Kirkwood (Laurentian University), Daniel Obrist (University of Massachusetts), Peter Outridge
- (Geological Survey of Canada/ University of Manitoba), Kyra A. St Pierre (University of British
- Columbia) and Christian Zdanowicz (Uppsala University) for their contributions to chapter 4 of the
- 2021 AMAP Mercury Assessment. This work was supported by the Swedish Research Council (2017-
- 05275 to SJ), the Swedish Research Council for Environment, Agricultural Sciences and Spatial
- Planning (2020-01868 to S.J.), Environment and Climate Change Canada (to J.C.), Chantier Arctique
- Francais (Pollution in the Arctic System to L.E.H.B) and the AXA Research Fund (to L.E.H.B).
-
-

References

 Adam D. Morris, Frank R. Rigét, Simon Wilson, Rob Fryer, Philippe J. Thomas, et al. 2022. Temporal trends of mercury in Arctic biota: 10 more years of progress in Arctic monitoring. Science of the Total Enviroment, in review.

- Agather, A.M., Bowman, K.L., Lamborg, C.H. and Hammerschmidt, C.R., 2019. Distribution of
- mercury species in the Western Arctic Ocean (U.S. GEOTRACES GN01). Marine Chemistry,
- 216:103686.
- Albert, C., Helgason, H.H., Brault-Favrou, M., Robertson, G.J., Descamps, S., Amélineau, F., et al. 2021. Seasonal variation of mercury contamination in Arctic seabirds: A pan-Arctic assessment. Science of the Total Environment 750, 142201.
- AMAP, 2011. AMAP Assessment 2011: Mercury in the Arctic. Arctic Monitoring and Assessment Programme (AMAP), Oslo, Norway. xiv + 193 pp.
- AMAP/UNEP, 2013. Technical Background Report to the Global Atmospheric Mercury Assessment 2013. Arctic Monitoring and Assessment Programme (AMAP), Oslo, Norway/United Nations Environment Programme (UNEP), Chemicals Branch, Geneva, Switzerland. vi + 263pp.
- AMAP, 2021. Assessment 2021: Mercury in the Arctic. Arctic Monitoring and Assessment Programme
- (AMAP), Oslo, Norway.
- Amélineau, F., Grémillet, D., Harding, A.M.A., Walkusz, W., Choquet, R. and Fort, J., 2019. Arctic climate change and pollution impact little auk foraging and fitness across a decade. Scientific Reports, 9 (1):1014.
- Baldi, F., Pepi, M. and Filippelli, M.,1993. Methylmercury resistance in *Desulfovibrio desulfuricans* strains in relation to methylmercury degradation. Applied and Environmental Microbiology, 59 (8):2479-2485.
- Barkay, T., Miller, S.M. and Summers, A.O., 2003. Bacterial mercury resistance from atoms to ecosystems. FEMS Microbiology Reviews, 27 (2-3):355-384.
- Baya, P.A., Gosselin, M., Lehnherr, I., St. Louis, V.L. and Hintelmann, H., 2015. Determination of monomethylmercury and dimethylmercury in the Arctic marine boundary layer. Environmental Science & Technology, 49 (1):223-232
- Beattie, S.A., Armstrong, D., Chaulk, A., Comte, J., Gosselin, M. and Wang, F., 2014. Total and methylated mercury in Arctic multiyear sea ice. Environmental Science & Technology, 48 (10):5575- 5582.
- Bechshoft, T., Derocher, A.E., Richardson, E., Lunn, N.J. and St. Louis, V.L., 2016. Hair mercury concentrations in Western Hudson Bay polar bear family groups. Environmental Science & Technology, 50 (10):5313-5319.
- Bełdowska, M. and Kobos, J., 2016. Mercury concentration in phytoplankton in response to warming of an autumn - winter season. Environmental Pollution, 215:38-47.
- Bidleman, T.F., Stern, G.A., Tomy, G.T., Hargrave, B.T., Jantunen, L.M. and MacDonald, R.W.,
- 2013. Scavenging amphipods: sentinels for penetration of mercury and persistent organic chemicals into food webs of the deep Arctic Ocean. Environmental Science & Technology, 47 (11):5553-5561.
- Black, F.J., Conaway, C.H. and Flegal, A.R., 2009. Stability of dimethyl mercury in seawater and its Conversion to monomethyl mercury. Environmental Science & Technology, 43 (11):4056-4062.
- Blais, J. M., Macdonald, R.W., Mackay, D., Webster, E., Harvey, C. and Smol, J. P., 2007. Biologically mediated transport of contaminants to aquatic systems. Environmental Science & Technology, 41 (4):1075-1084.
- Blum, J.D., Popp, B.N., Drazen, J.C., Anela Choy, C. and Johnson, M.W., 2013. Methylmercury production below the mixed layer in the North Pacific Ocean. Nature Geoscience, 6 (10):879-884.
- 1073 Bocharova, N., Treu, G., Czirják, G.Á., Krone, O., Stefanski, V., Wibbelt, G., Unnsteinsdóttir, E.R., 1074 Hersteinsson, P., Schares, G., Doronina, L., Goltsman, M. and Greenwood, A.D., 2013. Correlates Hersteinsson, P., Schares, G., Doronina, L., Goltsman, M. and Greenwood, A.D., 2013. Correlates between feeding ecology and mercury levels in historical and modern Arctic foxes (*Vulpes lagopus*). PLOS ONE, 8 (5):e60879.
- Bowman, K.L., Hammerschmidt, C.R., Lamborg, C.H. and Swarr, G., 2015. Mercury in the North Atlantic Ocean: the U.S. GEOTRACES zonal and meridional sections. Deep Sea Research Part II: Topical Studies in Oceanography, 116:251-26.
- Bowman, K.L., Hammerschmidt, C.R., Lamborg, C.H., Swarr, G.J. and Agather, A.M., 2016.
- Distribution of mercury species across a zonal section of the eastern tropical South Pacific Ocean
- (U.S. GEOTRACES GP16). Marine Chemistry, 186:156-166.
- Bowman, K.L., Collins, R.E., Agather, A.M., Lamborg, C.H., Hammerschmidt, C.R., Kaul, D.,
- Dupont, C.L., Christensen, G.A. and Elias, D.A., 2020a. Distribution of mercury-cycling genes in the
- Arctic and equatorial Pacific Oceans and their relationship to mercury speciation. Limnology and Oceanography, 65 (S1):S310-S320.
- Boetius, A., Albrecht, S., Bakker, K., Bienhold, C., Felden, J., Fernández-Méndez, M., Hendricks, S.,
- Katlein, C., Lalande, C., Krumpen, T., Nicolaus, M., Peeken, I., Rabe, B., Rogacheva, A., Rybakova,
- E., Somavilla, R., Wenzhöfer, F., and RV Polarstern ARK27-3-Shipboard Science Party: Export of
- algal biomass from the melting Arctic sea ice, Science, 339, 1430–1432,
- doi:10.1126/science.1231346, 2013.
- Bowman, K.L., Lamborg, C.H. and Agather, A.M., 2020b. A global perspective on mercury cycling in 1093 the ocean. Science of the Total Environment, 710:136166.
- Braaten, H.F.V., de Wit, H.A., Larssen, T. and Poste, A.E., 2018. Mercury in fish from Norwegian lakes: the complex influence of aqueous organic carbon. Science of the Total Environment, 627:341-348.
- Bradley MA, Barst BD, Basu N. A Review of Mercury Bioavailability in Humans and Fish. Int J Environ Res Public Health. 2017 Feb 10;14(2):169. doi: 10.3390/ijerph14020169. PMID: 28208586; PMCID: PMC5334723.
- Bratkič, A., Vahčič, M., Kotnik, J., Obu Vazner, K., Begu, E., Woodward, E.M.S. and Horvat, M., 2016. Mercury presence and speciation in the South Atlantic Ocean along the 40°S transect. Global Biogeochemical Cycles, 30 (2):105-119.
- Broadley, H.J., Cottingham, K.L., Baer, N.A., Weathers, K.C., Ewing, H.A., Chaves-Ulloa, R.,
- Chickering, J., Wilson, A.M., Shrestha, J. and Chen, C.Y., 2019. Factors affecting MeHg
- bioaccumulation in stream biota: the role of dissolved organic carbon and diet. Ecotoxicology, 28 (8):949-963.
- Brown, T.M., Fisk, A.T., Wang, X., Ferguson, S.H., Young, B.G., Reimer, K.J. and Muir, D.C.G.,
- 2016. Mercury and cadmium in ringed seals in the Canadian Arctic: influence of location and diet. Science of the Total Environment, 545-546:503-511.
- Brown, T.M., Macdonald, R.W., Muir, D.C.G. and Letcher, R.J., 2018. The distribution and trends of persistent organic pollutants and mercury in marine mammals from Canada's Eastern Arctic. Science
- 1111 of the Total Environment, 618:500-517.
- Burns, D.A., Aiken, G.R., Bradley, P.M., Journey, C.A. and Schelker, J., 2013. Specific ultra-violet
- absorbance as an indicator measurement of merucry sources in an Adirondack River basin.
- Biogeochemistry, 113 (1-3):451-466.
- Burt, A., Wang, F., Pućko, M., Mundy, C.-J., Gosselin, M., Philippe, B., Poulin, M., Tremblay, J.-É.
- and Stern, G.A., 2013. Mercury uptake within an ice algal community during the spring bloom in first-year Arctic sea ice. Journal of Geophysical Research: Oceans, 118 (9):4746-4754.
- Camacho, A., Rochera, C., Hennebelle, R., Ferrari, C. and Quesada, A., 2015. Total mercury and
- methyl-mercury contents and accumulation in polar microbial mats. Science of the Total Environment,
- 509-510:145-153.
- Capo, E., Bravo, A.G., Soerensen, A.L., Bertilsson, S., Pinhassi, J., Feng, C., Andersson, A.F., Buck,
- M. and Björn, E., 2020. Deltaproteobacteria and spirochaetes-like bacteria are abundant putative
- mercury methylators in oxygen-deficient water and marine particles in the Baltic Sea. Frontiers in
- Microbiology, 11:2277.
- Charette, M.A., Kipp, L.E., Jensen, L.T., Dabrowski, J.S., Whitmore, L.M., Fitzsimmons, J.N.,
- Williford, T., Ulfsbo, A., Jones, E., Bundy, R.M., Vivancos, S.M., Pahnke, K., John, S.G., Xiang, Y.,
- Hatta, M., Petrova, M.V., Heimbürger-Boavida, L.-E., Bauch, D., Newton, R., Pasqualini, A., Agather,
- A.M., Amon, R.M.W., Anderson, R.F., Andersson, P.S., Benner, R., Bowman, K.L., Edwards, R.L., Gdaniec, S., Gerringa, L.J.A., González, A.G., Granskog, M., Haley, B., Hammerschmidt, C.R., Hansell, D.A., Henderson, P.B., Kadko, D.C., Kaiser, K., Laan, P., Lam, P.J., Lamborg, C.H., Levier, M., Li, X., Margolin, A.R., Measures, C., Middag, R., Millero, F J., Moore, W S., Paffrath, R., Planquette, H., Rabe, B., Reader, H., Rember, R., Rijkenberg, M.J.A., Roy-Barman, M., Rutgers van der Loeff, M., Saito, M., Schauer, U., Schlosser, P., Sherrell, R M., Shiller, A.M., Slagter, H., Sonke, J.E., Stedmon, C., Woosley, R.J., Valk, O., van Ooijen, J. and Zhang, R., 2020. The Transpolar Drift as a source of riverine and shelf‐derived trace elements to the central Arctic Ocean. Journal of Geophysical
- Research: Oceans, 125 (5):e2019JC015920.
- Chaulk, A.; Stern, G. A.; Armstrong, D.; Barber, D. G.; Wang, F. Mercury distribution and transport
- across the ocean-sea-ice-atmosphere interface in the Arctic Ocean. Environ. Sci. Technol. 2011,
- 45:1866−1872.
- Chételat, J., Ackerman, J.T., Eagles-Smith, C.A. and Hebert, C.E., 2020. Methylmercury exposure in
- wildlife: a review of the ecological and physiological processes affecting contaminant concentrations
- and their interpretation. Science of the Total Environment, 711:135117.
- Chételat, J. and Amyot, M., 2009. Elevated methylmercury in High Arctic *Daphnia* and the role of productivity in controlling their distribution. Global Change Biology, 15 (3):706-718.
- Chételat, J., Amyot, M. and Cloutier, L., 2012. Shifts in elemental composition, methylmercury content and δ15N ratio during growth of a High Arctic copepod. Freshwater Biology, 57 (6):1228-1240.
- Chételat, J., Cloutier, L. and Amyot, M., 2010. Carbon sources for lake food webs in the Canadian High Arctic and other regions of Arctic North America. Polar Biology, 33 (8):1111–1123.
- Chételat, J., Richardson, M.C., MacMillan, G.A., Amyot, M. and Poulain, A.J., 2018. Ratio of
- methylmercury to dissolved organic carbon in water explains methylmercury bioaccumulation across a
- latitudinal gradient from north-temperate to Arctic lakes. Environmental Science & Technology, 52 (1):79-88.
- Chételat, J., Amyot, M., Arp, P., Blais, J.M., Depew, D., Emmerton, C.A., Evans, M., Gamberg, M.,
- Gantner, N., Girard, C., Graydon, J., Kirk, J., Lean, D., Lehnherr, I., Muir, D., Nasr, M., Poulain, A.J.,
- Power, M., Roach, P., Stern, G., Swanson, H. and van der Velden, S., 2015. Mercury in freshwater
- ecosystems of the Canadian Arctic: recent advances on its cycling and fate. Science of the Total
- Environment, 509-510:41-66.
- Chételat, J., McKinney, M.A., Amyot, M., Dastoor, A., Douglas, T.A., Heimbürger-Boavida, L.-E.,
- Kirk, J., Kahilainen, K.K., Outridge, P.M., Pelletier, N., Skov, H., St Pierre, K., Vuorenmaa, J., Wang, F., 2022. Climate change and mercury in the Arctic: Abiotic interactions. Sci. Total Environ. 824,
- 153715
	- Chiasson-Gould, S.A., Blais, J.M. and Poulain, A.J., 2014. Dissolved Organic Matter Kinetically Controls Mercury Bioavailability to Bacteria. Environmental Science & Technology, 48 (6):3153-3161.
	- Choy ES, Gauthier M, Mallory ML, Smol JP, Douglas MSV, Lean D, et al. An isotopic investigation
	- of mercury accumulation in terrestrial food webs adjacent to an Arctic seabird colony. Science of the
	- Total Environment 2010; 408: 1858-1867.
	- Clayden, M.G., Arsenault, L.M., Kidd, K.A., O'Driscoll, N.J. and Mallory, M.L., 2015. Mercury
	- bioaccumulation and biomagnification in a small Arctic polynya ecosystem. Science of the Total Environment, 509-510:206-215.
	- Cook, J., Edwards, A., Takeuchi, N. and Irvine-Fynn, T., 2016. Cryoconite: the dark biological secret of the cryosphere. Progress in Physical Geography: Earth and Environment, 40 (1):66-111.
- Cossa, D., Heimbürger, L.-E., Lannuzel, D., Rintoul, S.R., Butler, E.C.V., Bowie, A.R., Averty, B.,
- Watson, R.J. and Remenyi, T., 2011. Mercury in the Southern Ocean. Geochimica et Cosmochimica Acta, 75 (14):4037-4052.
- Cossa, D., Averty, B. and Pirrone, N., 2009. The origin of methylmercury in open Mediterranean waters. Limnology and Oceanography, 54 (3):837-844.
- Cossa, D., Madron, X.D. de, Schäfer, J., Lanceleur, L., Guédron, S., Buscail, R., Thomas, B., Castelle, S. and Naudin, J.-J., 2017. The open sea as the main source of methylmercury in the water column of the Gulf of Lions (Northwestern Mediterranean margin). Geochimica et Cosmochimica Acta, 199:222- 237.
- Cossa, D., Harmelin-Vivien, M., Mellon-Duval, C., Loizeau, V., Averty, B., Crochet, S., Chou, L. and
- Cadiou, J.-F., 2012. Influences of bioavailability, trophic Position, and growth on methylmercury in
- hakes (*Merluccius merluccius*) from northwestern Mediterranean and northeastern Atlantic.
- Environmental Science & Technology, 46 (9):4885-4893.
- 1185 Cossa, D., Garnier, C., Buscail, R., Elbaz-Poulichet, F., Mikac, N., Patel-Sorrentino, N., Tessier, E., 1186 Rigaud, S., Lenoble, V. and Gobeil, C., 2014. A Michaelis-Menten type equation for describing
- Rigaud, S., Lenoble, V. and Gobeil, C., 2014. A Michaelis–Menten type equation for describing
- methylmercury dependence on inorganic mercury in aquatic sediments. Biogeochemistry, 119 (1):35- 43
- Cyr, A., López, J.A., Rea, L., Wooller, M.J., Loomis, T., Mcdermott, S. and O'Hara, T.M., 2019. Mercury concentrations in marine species from the Aleutian Islands: spatial and biological determinants.
- Science of the Total Environment, 664:761-770.
- 1192 Dastoor A, Ryzhkov A, Durnford D, Lehnherr I, Steffen A, Morrison H. 2015, Atmospheric mercury
1193 in the Canadian Arctic. Part II: Insight from modeling. Science of The Total Environment, 509-510:
- in the Canadian Arctic. Part II: Insight from modeling. Science of The Total Environment, 509-510: 16-27.
- Dastoor A., Angot H., Bieser J., Christensen J.H., Douglas T.A., Heimbürger-Boavida L-E., Jiskra M.,
- Mason R.P., McLagan D.S., Obrist D., Outridge P.M, Petrova M.V, Ryjkov A., St. Pierre K.A.,
- Schartup A.T., Soerensen A.L., Toyota K., Travnikov O., Wilson S.J., Zdanowicz C., 2022, Arctic
- mercury cycling, Nature Reviews Earth & Environment volume 3, pages270–286 (2022)
- Desforges J-P, Outridge P, Hobson KA, Heide-Jørgensen MP, Dietz R. Anthropogenic and Climatic
- Drivers of Long-Term Changes of Mercury and Feeding Ecology in Arctic Beluga (Delphinapterus
- leucas) Populations. Environmental Science & Technology 2022; 56: 271-281.
- DiMento BP, Mason RP, Brooks S, Moore C. The impact of sea ice on the air-sea exchange of mercury in the Arctic Ocean. Deep Sea Research Part I: Oceanographic Research Papers 2019; 144: 28- 38.Loseto, L.L., Lean, D.R. and Siciliano, S.D., 2004. Snowmelt sources of methylmercury to High Arctic ecosystems. Environmental Science & Technology 38 (11):3004-3010.
- Douglas, T.A., Loseto, L.L., Macdonald, R.W., Outridge, P., Dommergue, A., Poulain, A., Amyot, M., Barkay, T., Berg, T., Chételat, J., Constant, P., Evans, M., Ferrari, C., Gantner, N., Johnson, M.S., Kirk, J., Kroer, N., Larose, C., Lean, D., Nielsen, T.G., Poissant, L., Rognerud, S., Skov, H., Sørensen, S., Wang, F.Y., Wilson, S. and Zdanowicz, C.M., 2012. The fate of mercury in Arctic terrestrial and aquatic ecosystems, a review. Environmental Chemistry, 9 (4):321-355.
- Douglas, T.A., Sturm, M., Blum, J.D., Polashenski, C., Stuefer, S., Hiemstra, C., Steffen, A., Filhol, S.
- and Prevost, R., 2017. A pulse of mercury and major ions in snowmelt runoff from a small Arctic Alaska watershed. Environmental. Science & Technology, 51 (19):11145-11155.
-
- Douglas, T.A. and Blum, J.D., 2019. Mercury isotopes reveal atmospheric gaseous mercury deposition directly to the Arctic coastal snowpack. Environmental Science & Technology Letters, 6 (4):235-242.
- Douglas TA, Hiemstra CA, Anderson JE, Barbato RA, Bjella KL, Deeb EJ, Gelvin AB, Nelsen PE,
- Newman SD, Saari SP, Wagner AM. 2021, Recent degradation of Interior Alaska permafrost mapped
- with ground surveys, geophysics, deep drilling, and repeat airborne LiDAR. The Cryosphere
- Discussions. Feb 22:1-39.

 Driscoll, C.T., Yan, C., Schofield, C.L., Munson, R. and Holsapple, J., 1994. The mercury cycle and fish in the Adirondack lakes. Environmental Science & Technology, 28 (3):136A-143A.

 Driscoll CT, Mason RP, Chan HM, Jacob DJ, Pirrone N. 2013. Mercury as a Global Pollutant: Sources, Pathways, and Effects. Environmental Science & Technology, 47: 4967-4983

- Drott, A., Lambertsson, L., Björn, E. and Skyllberg, U., 2007. Importance of dissolved neutral mercury sulfides for methyl mercury production in contaminated sediments. Environmental Science & Technology, 41 (7):2270-2276.
- Durnford, D., Dastoor, A., Figueras-Nieto, D., and Ryjkov, A. 2010. Long range transport of mercury to the Arctic and across Canada, Atmos. Chem. Phys., 10, 6063–6086, https://doi.org/10.5194/acp-10- 6063-2010.
- Dietz R, Letcher R.J., Aars J., Andersen M., Andriashek et al. 2022 What are the toxicological effects of mercury in Arctic marine and terrestrial mammals? Science of the Total Environment, in review.
- Ebinghaus, R., Kock, H.H., Temme, C., Einax, J.W., Löwe, A.G., Richter, A., Burrows, J.P. and
- Schroeder, W.H., 2002. Antarctic springtime depletion of atmospheric mercury. Environmental
- 1234 Science & Technology, 36 (6):1238-1244.
- Eckbo, N., Le Bohec, C., Planas-Bielsa, V., Warner, N.A., Schull, Q., Herzke, D., Zahn, S., Haarr, A.,
- Gabrielsen, G.W. and Borgå, K., 2019. Individual variability in contaminants and physiological status in a resident Arctic seabird species. Environmental Pollution, 249:191-199.
- Emmerton, C.A., Graydon, J.A., Gareis, J.A.L., St. Louis, V.L., Lesack, L.F.W., Banack, J.K.A.,
- Hicks, F. and Nafziger, J., 2013. Mercury export to the Arctic Ocean from the Mackenzie River,
- Canada. Environmental Science & Technology, 47 (14):7644-7654.
- Evans, M.S., Muir, D.C.G., Keating, J. and Wang, X., 2015. Anadromous char as an alternate food
- choice to marine animals: A synthesis of Hg concentrations, population features and other influencing factors. Science of the Total Environment, 509-510:175-194.
- Evers, D.C., Schmutz, J.A., Basu, N., DeSorbo, C.R., Fair, J., Gray, C.E.,. Paruk, J.D., Perkins, M.,
- Regan, K., Uher-Koch, B.D. and Wright, K.G., 2014. Historic and contemporary mercury exposure
- and potential risk to yellow-billed loons (*Gavia adamsii*) breeding in Alaska and Canada. Waterbirds, 1247 37 (sp1):147-159.
- Ewald JD, Kirk JL, Li M, Sunderland EM. Organ-specific differences in mercury speciation and accumulation across ringed seal (Phoca hispida) life stages. Science of the Total Environment 2019; 650: 2013-2020.
- 1251 Fahnestock, M.F., Bryce, J.G., McCalley, C.K., Montesdeoca, M., Bai, S., Li, Y., Driscoll, C.T., Crill, 1252 P.M., Rich, V.I. and Varner, R.K., 2019. Mercury reallocation in thawing Subarctic peatlands. P.M., Rich, V.I. and Varner, R.K., 2019. Mercury reallocation in thawing Subarctic peatlands. Geochemical Perspectives Letters, 11:33-38.
- Fort, J., Robertson, G.J., Grémillet, D., Traisnel, G. and Bustamante, P., 2014. Spatial ecotoxicology:
- migratory Arctic seabirds are exposed to mercury contamination while overwintering in the Northwest Atlantic. Environmental Science & Technology, 48 (19):11560-11567.
- Foster, K.L., Stern, G.A., Pazerniuk, M.A., Hickie, B., Walkusz, W., Wang, F. and Macdonald, R.W.,
- 2012. Mercury biomagnification in marine zooplankton food webs in Hudson Bay. Environmental
- 1259 Science & Technology, 46 (23):12952-12959.

 Fox, A.L., Hughes, E.A., Trocine, R.P., Trefry, J.H., Schonberg, S.V., McTigue, N.D., Lasorsa, B.K., Konar, B. and Cooper, L.W., 2014. Mercury in the northeastern Chukchi Sea: distribution patterns in seawater and sediments and biomagnification in the benthic food web. Deep Sea Research Part II: Topical Studies in Oceanography, 102:56-67.

- Fox, A.L., Trefry, J.H., Trocine, R.P., Dunton, K.H., Lasorsa, B.K., Konar, B., Ashjian, C.J., Cooper,
- L.W., 2017. Mercury biomagnification in food webs of the northeastern Chukchi Sea, Alaskan Arctic. Deep Sea Research Part II: Topical Studies in Oceanography, 144:63-77.
-
- French, T.D., Houben, A.J., Desforges, J.P.W., Kimpe, L.E., Kokelj, S.V., Poulain, A.J., Smol, J.P., Wang, X. and Blais, J.M., Dissolved organic carbon thresholds affect mercury bioaccumulation in Arctic lakes. Environmental Science & Technology, 48 (6):3162-3168.
- 1270 Frey, K.E. and McClelland, J.W., 2009. Impacts of permafrost degradation on Arctic river 1271 biogeochemistry. Hydrological Processes. 23 (1):169-182. biogeochemistry. Hydrological Processes, (1):169-182.
- Gamberg, M., Cuyler, C. and Wang, X., 2016. Contaminants in two West Greenland caribou populations. Science of the Total Environment, 554-555:329-336.
- Gantner, N., Muir, D.C., Power, M., Iqaluk, D., Reist, J.D., Babaluk, J.A., Meili, M., Borg, H.,
- Hammar, J., Michaud, W., Dempson, B. and Solomon, K.R., 2010. Mercury concentrations in
- landlocked Arctic char (*Salvelinus alpinus*) from the Canadian Arctic. Part II: Influence of lake biotic
- and abiotic characteristics on geographic trends in 27 populations. Environmental Toxicology and Chemistry, 29 (3):633-643.
- Gascón Díez, E., Loizeau, J.-L., Cosio, C., Bouchet, S., Adatte, T., Amouroux, D. and Bravo, A.G.,
- 1280 2016. Role of settling particles on mercury methylation in the oxic water column of freshwater 1281 systems. Environmental Science & Technology, 50 (21):11672-11679.
- systems. Environmental Science & Technology, 50 (21):11672-11679.
- Gilmour, C.C., Podar, M., Bullock, A.L., Graham, A.M., Brown, S.D., Somenahally, A.C., Johs, A., Hurt, R.A., Bailey, K.L. and Elias, D.A., 2013. Mercury methylation by novel microorganisms from
- new environments. Environmental Science & Technology, 47 (20):11810-11820.
- Gilmour, C.C., Bullock, A.L., McBurney, A., Podar, M., Elias, D.A., 2018, Robust Mercury Methylation across Diverse Methanogenic Archaea, mBio, 9(2), 10.1128/mBio.02403-17
- Gionfriddo, C.M., Tate, M.T., Wick, R.R., Schultz, M.B., Zemla, A., Thelen, M.P., Schofield, R.,
- Krabbenhoft, D.P., Holt, K.E. and Moreau, J.W., 2016. Microbial mercury methylation in Antarctic sea ice. Nature Microbiology, 1 (10):16127.
- Gionfriddo C.M., Stott, M.B., Power, J.F., Ogorek, J.M., Krabbenhoft, D.P., Wick, R., Holt, K., Chen,
- L-.,X., Thomas, B.C., Banfield, J.F., Moreau, J.W. and Cann, I., 2020. Genome-resolved
- metagenomics and detailed geochemical speciation analyses yield new insights into microbial mercury
- cycling in geothermal springs. Applied and Environmental Microbiology, 86 (15):e00176-20.
- Girard, C., Leclerc, M. and Amyot M., 2016. Photodemethylation of methylmercury in Eastern Canadian Arctic thaw pond and lake ecosystems. Environmental Science & Technology, 50 (7):3511- 3520.
- Gordon, J., Quinton, W., Branfireun, B.A. and Olefeldt, D., 2016. Mercury and methylmercury biogeochemistry in a thawing permafrost wetland complex, Northwest Territories, Canada. Hydrological Processes, 30 (20):3627-3638.
- Góngora, E., Braune, B.M. and Elliott, K.H., 2018. Nitrogen and sulfur isotopes predict variation in mercury levels in Arctic seabird prey. Marine Pollution Bulletin, 135:907-914.
- Graham, A.M., Aiken, G.R. and Gilmour, C.C., 2012. Dissolved organic matter enhances microbial mercury methylation under sulfidic conditions. Environmental Science & Technology, 46 (5):2715- 2723.
- Hallanger, I.G., Fuglei, E., Yoccoz, N.G., Pedersen, Å.Ø., König, M. and Routti, H., 2019. Temporal trend of mercury in relation to feeding habits and food availability in Arctic foxes (*Vulpes lagopus*) from Svalbard, Norway. Science of the Total Environment, 670:1125-1132.
- Halm, D.R. and Dornblaser, M.M., 2007. Water and sediment quality in the Yukon River and its tributaries between Atlin, British Columbia, Canada, and Eagle, Alaska, USA, 2004 (Open-File Report 2007-1197). U.S. Geological Survey. Available at: **https://doi.org/10.3133/ofr20071197**.
- Hammerschmidt, C.R. and Fitzgerald, W.F., 2006. Photodecomposition of methylmercury in an Arctic Alaskan Lake. Environmental Science & Technology, 40 (4):1212-1216.
- Hammerschmidt, C.R. and Bowman, K.L., 2012. Vertical methylmercury distribution in the subtropical North Pacific Ocean. Marine Chemistry, 132-133:77-82.
- Hammerschmidt, C.R. and Fitzgerald, W.F., 2010. Iron-mediated photochemical decomposition of methylmercury in an Arctic Alaskan Lake. Environmental Science & Technology, 44 (16):6138-6143.
- Hawkings, J. R. Linhoff B.S., Wadham J.L., Stibal M., Lamborg C.H., Carling G. T., Lamarche-
- Gagnon G., Kohler T.J., Ward R., Hendry K.R., Falteisek L., Kellerman A. M., Cameron K. A.,
- Hatton J. E., Tingey S., Holt A.D., Vinšová P., Hofer S., Bulínová M., Větrovský T., Meire L. &
- Spencer R.G. M., 2021, Large subglacial source of mercury from the southwestern margin of the
- Greenland Ice Sheet. Nat. Geosci. 14, 496–502
- Heimbürger, L.-E., Sonke, J.E, Cossa, D., Point, D., Lagane, C., Laffont, L, Galfond, B., Nicolaus, M., Rabe, B. and Rutgers van der Loeff, M., 2015. Shallow methylmercury production in the marginal sea ice zone of the central Arctic Ocean. Scientific Report, 5 (1):10318.
- Heimbürger, L.-E., Cossa, D., Marty, J.-C., Migon, C., Averty, B., Dufour, A. and Ras, J., 2010.
- Methyl mercury distributions in relation to the presence of nano- and picophytoplankton in an oceanic water column (Ligurian Sea, North-western Mediterranean). Geochimica et Cosmochimica Acta, 74 (19):5549-5559.
- Hollweg, T.A., Gilmour, C.C. and Mason, R.P., 2010. Mercury and methylmercury cycling in
- sediments of the mid-Atlantic continental shelf and slope. Limnology and Oceanography, 55 (6):2703- 2722.
- Hood, E., Fellman, J., Spencer, R.G.M., Hernes, P.J., Edwards, R., D'Amore, D. and Scott, D., 2009. Glaciers as a source of ancient and labile organic matter to the marine environment. Nature, 462
- (7276):1044-1047.
- 1335 Hood, E., Battin, T.J., Fellman, J., O'Neel, S. and Spencer, R.G.M., 2015. Storage and release of organic carbon from glaciers and ice sheets. Nature Geoscience, 8 (2):91-96. carbon from glaciers and ice sheets. Nature Geoscience, 8 (2):91-96.
- Hopwood, M.J., Carroll, D., Dunse, T., Hodson, A., Holding, J.M., Iriarte, J.L., Ribeiro, S., Achterberg,
- E.P., Cantoni, C., Carlson, D.F., Chierici, M., Clarke, J.S., Cozzi, S., Fransson, A., Juul-Pedersen, T.,
- Winding, M.H.S. and Meire, L., 2020. How does glacier discharge affect marine biogeochemistry and
- primary production in the Arctic? The Cryosphere, 14 (4):1347-1383.
- Houben, A.J., French, T.D., Kokelj, S. V., Wang, X., Smol, J.P. and Blais, J.M., 2016. The impacts of permafrost thaw slump events on limnological variables in upland tundra lakes, Mackenzie Delta region. Fundamental and Applied Limnology, 189:11-35.
-
- Hsu-Kim, H., Eckley, C.S., Achá, D., Feng, X., Gilmour, C.C., Jonsson, S. and Mitchell, C.P.J., 2018. Challenges and opportunities for managing aquatic mercury pollution in altered landscapes. Ambio, 47 (2):141-169.
- Hudelson, K.E., Drevnick, P.E., Wang, F., Armstrong, D. and Fisk, A.T., 2020. Mercury methylation and demethylation potentials in Arctic lake sediments. Chemosphere, 248:126001.
- Hudelson, K.E., Muir, D.C.G., Drevnick, P.E., Köck, G., Iqaluk, D., Wang, X., Kirk, J.L., Barst, B.D., Grgicak-Mannion, A., Shearon, R. and Fisk, A.T., 2019. Temporal trends, lake-to-lake variation, and climate effects on Arctic char (*Salvelinus alpinus*) mercury concentrations from six High Arctic lakes in Nunavut, Canada. Science of the Total Environment, 678:801-812.
- Imura, N., Sukegawa, E., Pan, S.-K., Nagao, K., Kim, J.-Y., Kwan, T. and Ukita, T., 1971. Chemical 1354 methylation of inorganic mercury with methylcobalamin, a vitamin B_{12} analog. Science, 172 (3989):1248-1249.
- Isles, P.D.F., Jonsson, A., Creed, I.F. and Bergström, A.-K., 2020. Does browning affect the identity of limiting nutrients in lakes? Aquatic Sciences, 82 (2):45.
- Jiang, T., Wang, D., Meng, B., Chi, B., Laudon, H. and Liu, J., 2020. The concentrations and characteristics of dissolved organic matter in high-latitude lakes determine its ambient reducing capacity. Water Research, 169:115217.
- Jiskra, M., Sonke, J.E., Obrist, D., Bieser, J., Ebinghaus, R., Myhre, C.L., Pfaffhuber, K.A.,
- Wängberg, I., Kyllönen, K., Worthy, D., Martin, L.G., Labuschagne, C., Mkololo, T., Ramonet, M.,
- Magand, O., Dommergue, A., 2018. A vegetation control on seasonal variations in global atmospheric mercury concentrations. Nat. Geosci. 11, 244–250
- Jiskra, M., Heimbürger-Boavida, L.-E., Desgranges, M.-M., Petrova, M.V., Dufour, A., Ferreira-
- Araujo, B., Masbou, J., Chmeleff, J., Thyssen, M., Point, D., Sonke, J.E., 2021. Mercury stable
- isotopes constrain atmospheric sources to the ocean. Nature 597, 678–682.
- Jones, D.S., Walker, G.M., Johnson, N.W., Mitchell, C.P.J., Coleman Wasik, J.K. and Bailey, J.V.,
- 2019. Molecular evidence for novel mercury methylating microorganisms in sulfate-impacted lakes. The ISME Journal, 13 (7):1659-1675.
- Jonsson, S., Skyllberg, U., Nilsson, M.B., Westlund, P.-O., Shchukarev, A., Lundberg, E. and Björn,
- E., 2012. Mercury methylation rates for geochemically relevant HgII species in sediments.
- Environmental Science & Technology, 46 (21):11653-11659.
- Jonsson, S., Skyllberg, U., Nilsson, M.B., Lundberg, E., Andersson, A. and Björn, E., 2014.
- Differentiated availability of geochemical mercury pools controls methylmercury levels in estuarine sediment and biota. Nature Communications, 5 (1):4624.
-
- Jonsson, S., Mazrui, N.M. and Mason, R.P., 2016. Dimethylmercury formation mediated by inorganic and organic reduced sulfur surfaces. Scientific Reports, 6 (1):27958.
- Jonsson, S., Nerentorp Mastromonaco, M.G, Gårdfeldt, K., Mason, R.P., Distribution of Total
- Mercury and Methylated Mercury Species in Central Arctic Ocean Water and Ice, 2022, Marine Chemistry, 242, 104105.
- Jorgenson, M.T., Racine, C.H., Walters, J.C. and Osterkamp, T.E., 2001. Permafrost degradation and
- ecological changes associated with a warming climate in central Alaska. Climatic Change, 48 (4):551-
- 579.
- Jorgenson, M.T., Shur, Y.L. and Pullman, E.R., 2006. Abrupt increase in permafrost degradation in Arctic Alaska. Geophysical Research Letters, 33 (2): L02503.
- Jörundsdóttir, H.Ó., Jensen, S., Hylland, K., Holth, T.F., Gunnlaugsdóttir, H., Svavarsson, J.,
- Ólafsdóttir, Á., El-Taliawy, H., Rigét, F., Strand, J., Nyberg, E., Bignert, A., Hoydal, K.S. and
- Halldórsson, H.P., 2014. Pristine Arctic: background mapping of PAHs, PAH metabolites and
- inorganic trace elements in the North-Atlantic Arctic and Sub-Arctic coastal environment. Science of
- the Total Environment, 493:719-728.
- Kahilainen, K.K., Thomas, S.M., Keva, O., Hayden, B., Knudsen, R., Eloranta, A.P., Tuohiluoto, K.,
- Amundsen, P.,-A., Malinen, T. and Järvinen, A., 2016. Seasonal dietary shift to zooplankton
- influences stable isotope ratios and total mercury concentrations in Arctic charr (*Salvelinus alpinus* (L.)). Hydrobiologia, 783 (1):47-63.
- Kahilainen, K.K., Thomas, S.M., Nystedt, E.K.M., Keva, O., Malinen, T. and Hayden, B., 2017. Ecomorphological divergence drives differential mercury bioaccumulation of polymorphic European whitefish (*Coregonus lavaretus*) populations of Subarctic lakes. Science of the Total Environment, 599- 600:1768-1778.
- Keva, O., Hayden, B., Harrod, C. and Kahilainen, K.K., 2017. Total mercury concentrations in liver and
- muscle of European whitefish (*Coregonus lavaretus* (L.)) in a Subarctic lake assessing the factors
- driving year-round variation. Environmental Pollution, 231 (Part 2):1518-1528.
- Khan, M.A. and Wang, F., 2010. Chemical demethylation of methylmercury by selenoamino acids. Chemical Research in Toxicology, 23 (7):1202-1206.
- 1405 Kim, H., Soerensen, A.L., Hur, J., Heimbürger, L.-E., Hahm, D., Rhee, T.S., Noh, S. and Han, S., 1406 2017. Methylmercury mass budgets and distribution characteristics in the western Pacific Ocean.
- 2017. Methylmercury mass budgets and distribution characteristics in the western Pacific Ocean. Environmental Science & Technology, 51 (3):1186-1194.
- Kim, H., Kwon, S.Y., Lee, K., Lim, D., Han, S., Kim, T.-W., Joo, Y.J., Lim, J., Kang, M.-H. and Nam, S.-I., 2020a. Input of terrestrial organic matter linked to deglaciation increased mercury transport to the Svalbard fjords. Scientific Reports, 10 (1):3446.
- Kim, J., Soerensen, A.L., Kim, M.S., Eom, S., Rhee, T.S., Jin, Y K. and Han, S., 2020b. Mass budget of methylmercury in the east Siberian Sea: the importance of sediment sources. Environmental Science
- and Technology, 54 (16):9949–9957.
- Kirk, J.L., St. Louis, V.L., Hintelmann, H., Lehnherr, I., Else, B. and Poissant, L., 2008. Methylated mercury species in marine waters of the Canadian High and Sub Arctic. Environmental Science & Technology, 42 (22):8367-8373.
- Klapstein, S.J. and O'Driscoll, N.J., 2018. Methylmercury biogeochemistry in freshwater ecosystems:
- a review focusing on DOM and photodemethylation. Bulletin of Environmental Contamination and Toxicology, 100 (1):14-25.
- Klapstein, S.J., Ziegler, S.E. and O'Driscoll, N.J., 2017. Methylmercury photodemethylation is inhibited in lakes with high dissolved organic matter. Environmental Pollution, 232:392-401.
- Kohlbach, D., Graeve, M., Lange, B.A., David, C., Peeken, I. and Flores, H., 2016. The importance of
- ice algae-produced carbon in the central Arctic Ocean ecosystem: food web relationships revealed by
- lipid and stable isotope analyses. Limnology and Oceanography, 61 (6):2027-2044.
- Kohler, Stephen G., Heimbürger-Boavida, L.E., Petrova, M.V., Digernes, M.G., Sanchez, N., Dufour,
- A., Simić, A., Ndungu, K., Ardelan, M.V., 2022. Arctic Ocean's wintertime mercury concentrations limited by seasonal loss on the shelf. in press
- Kristiansen SM, Leinaas HP, Herzke D, Hylland K, Gabrielsen GW, Harju M, et al. Seabird-
- Transported Contaminants Are Reflected in the Arctic Tundra, but Not in Its Soil-Dwelling Springtails
- (Collembola). Environmental Science and Technology 2019; 53: 12835-12845.

 Kåresdotter, E., Destouni, G., Ghajarnia, N., Hugelius, G., & Kalantari, Z., 2021. Mapping the vulnerability of Arctic wetlands to global warming. Earth's Future, 9, e2020EF001858.

- Lantz, T.C. and Kokelj, S.V., 2008. Increasing rates of retrogressive thaw slump activity in the Mackenzie Delta region, N.W.T., Canada. Geophysical Research Letters, 35 (6):L06502.
- Lavoie, R.A., Jardine, T.D., Chumchal, M.M., Kidd, K.A. and Campbell, L.M., 2013.
- Biomagnification of mercury in aquatic food webs: a worldwide meta-analysis. Environmental 1437 Science & Technology, 47 (23):13385-13394.
- Lavoie, R.A., Amyot, M., Lapierre, J.-F., 2019. Global meta-analysis on the relationship between
- mercury and dissolved organic carbon in freshwater environments. Journal of Geophysical Research: Biogeosciences, 124 (6):1508-1523.
- Lee, C.-S. and Fisher, N.S., 2016. Methylmercury uptake by diverse marine phytoplankton.
- Limnology and Oceanography, 61 (5):1626-1639.
- Lehnherr, I., St. Louis, V.L., Hintelmann, H. and Kirk, J.L., 2011. Methylation of inorganic mercury in polar marine waters. Nature Geoscience, 4 (5):298-302.
- Lehnherr, I., St. Louis, V.L. and Kirk, J.L., 2012a. Methylmercury cycling in High Arctic wetland ponds: sources and sinks. Environmental Science & Technology, 46 (19):10514-10522.
- Lehnherr, I., St. Louis, V.L. and Kirk, J.L., 2012b Methylmercury cycling in High Arctic wetland ponds: controls on sedimentary production. Environmental Science & Technolog, 46 (19):10523-10531.
- Lehnherr, I., 2014. Methylmercury biogeochemistry: a review with special reference to Arctic aquatic ecosystems. Environmental Reviews, 22 (3):229-243.
- Lescord, G.L., Emilson, E.J.S., Johnston, T.A., Branfireun, B.A. and Gunn, J.M., 2018. Optical
- properties of dissolved organic matter and their relation to mercury concentration in water and biota across a remote freshwater drainage basin. Environmental Science & Technology, 52 (6): 3344-3353.
- Lescord, G.L., Kidd, K.A., Kirk, J.L., O'Driscoll, N.J., Wang, X. and Muir, D.C.G., 2015. Factors affecting biotic mercury concentrations and biomagnification through lake food webs in the Canadian
- High Arctic. Science of the Total Environment, 509-510:195-205.
- Li, C., Sonke, J.E., Le Roux, G., Piotrowska, N., Van der Putten, N., Roberts, S.J., Daley, T.,
- Rice, E., Gehrels, R., Enrico, M., Mauquoy, D., Roland, T.P., De Vleeschouwer, F., 2020.
- Unequal Anthropogenic Enrichment of Mercury in Earth's Northern and Southern
- Hemispheres. ACS Earth Space Chem. 4, 2073–2081
- Liem-Nguyen, V., Wild, B., Gustafsson, Ö., Semiletov, I., Dudarev, O. and Jonsson, S., 2022. Spatial
- patterns and distributional controls of total and methylated mercury off the Lena River in the Laptev Sea
- sediments. Marine Chemistry, 238, 104052
- Lim, A.G., Sonke, J.E.; Krickov, I.V.; Manasypov, R.M., Loiko, S.V. and Pokrovsky, O.S., 2019.
- Enhanced particulate Hg export at the permafrost boundary, western Siberia. Environmental Pollution,
- 254, (Part B):113083.
- Lim, A. G., Jiskra, M., Sonke, J. E., Loiko, S. V., Kosykh, N., and Pokrovsky, O. S., 2020, A revised pan-Arctic permafrost soil Hg pool based on Western Siberian peat Hg and carbon observations, Biogeosciences, 17, 3083–3097
- Lin H.., Ascher D.B., Myung Y., Lamborg C.H., Hallam S.J., Gionfriddo C.M., Holt K..E., Moreau
- J.W., 2021. Mercury methylation by metabolically versatile and cosmopolitan marine bacteria. The
- ISME Journal 15:1810–1825
- Liu, Y., Chai, X., Hao, Y., Gao, X., Lu, Z., Zhao, Y., Zhang, J. and Cai, M., 2015. Total mercury and methylmercury distributions in surface sediments from Kongsfjorden, Svalbard, Norwegian Arctic.
- Environmental Science and Pollution Research , 22 (11):8603-8610.
- Lockhart WL, Stern GA, Low G, Hendzel M, Boila G, Roach P, et al. A history of total mercury in edible muscle of fish from lakes in northern Canada. Science of The Total Environment 2005; 351-352: 427-463.
- Loose, B., SchlosseR, P., Perovich, D., Ringelberg, D., HO, D., Takahashi, T., Richter-Menge, J.,
- Reynolds, C., McGillis, W. and Tison, J.-L. (2011), Gas diffusion through columnar laboratory sea ice: implications for mixed-layer ventilation of CO2 in the seasonal ice zone. Tellus B, 63: 23-39.
- Loseto LL, Stern GA, Deibel D, Connelly TL, Prokopowicz A, Lean DRS, et al. Linking mercury exposure to habitat and feeding behaviour in Beaufort Sea beluga whales. Journal of Marine Systems 2008; 74: 1012-1024.
- Loseto LL, Stern GA, Macdonald RW. Distant drivers or local signals: Where do mercury trends in western Arctic belugas originate? Science of The Total Environment 2015; 509-510: 226-236.
- Ma, M., Du, H. and Wang, D., 2019. Mercury methylation by anaerobic microorganisms: a review. Critical Reviews in Environmental Science and Technology, 49 (20):1893-1936.
- Macdonald, R.W., Harner, T. and Fyfe, J., 2005. Recent climate change in the Arctic and its impact on contaminant pathways and interpretation of temporal trend data. Science of the Total Environment, 342 (1-3):5-86.
- Mackelprang, R., Waldrop, M.P., DeAngelis, K.M., David, M.M., Chavarria, K.L., Blazewicz, S.J.,
- Rubin, E.M. and Jansson, J.K., 2011. Metagenomic analysis of a permafrost microbial community reveals a rapid response to thaw. Nature, 480 (7377):368-371.
- MacMillan, G.A., 2018. Hunting for trace metals in the rapidly changing North: using limnological,
- ecological and collaborative approaches. Doctoral thesis, Department of Biological Sciences,
- University of Montreal, Quebec, Canada. Available at:
- https://papyrus.bib.umontreal.ca/xmlui/handle/1866/21777.
- MacMillan, G.A., Girard, C., Chételat, J., Laurion, I., and Amyot, M., 2015. High methylmercury in
- Arctic and Subarctic ponds is related to nutrient levels in the warming eastern Canadian Arctic. Environmental Science & Technology, 49 (13):7743-7753.
- Mangal, V., Stenzler, B.R., Poulain, A.J. and Guéguen, C., 2019. Aerobic and anaerobic bacterial
- mercury uptake is driven by algal organic matter composition and molecular weight. Environmental
- Science & Technology, 53 (1):157-165.
- Masbou, J., Sonke, J.E., Amouroux, D., Guillou, G., Becker, P.R. and Point, D., 2018. Hg-stable isotope variations in marine top predators of the western Arctic Ocean. ACS Earth and Space Chemistry 2
- (5):479-490.
- Mazrui, N.M., Jonsson, S., Thota, S., Zhao, J. and Mason, R.P., 2016. Enhanced availability of
- mercury bound to dissolved organic matter for methylation in marine sediments. Geochimica et Cosmochimica Acta, 194:153-162.
- Martyniuk, M.A.C., Couture, P., Tran, L., Beaupré, L. and Power, M., 2020. Seasonal variation of total
- 1512 mercury and condition indices of Arctic charr (Salvelinus alpinus) in Northern Quebec, Canada. Science
1513 of the Total Environment. 738:139450.
- of the Total Environment, 738:139450.
- Mason, R.P., Choi, A.L., Fitzgerald, W.F., Hammerschmidt, C.R., Lamborg, C.H., Soerensen, A.L. and
- Sunderland, E.M., 2012. Mercury biogeochemical cycling in the ocean and policy implications. Environmental Research, 119:101-117.
- Mason, R.P. and Sullivan, K.A., 1999. The distribution and speciation of mercury in the South and equatorial Atlantic. Deep Sea Research Part II: Topical Studies in Oceanography 46 (5):937-956.
- McDaniel, E.A., Peterson, B.D., Stevens, S.L.R., Tran, P.Q., Anantharaman, K., McMahon, K.D. and
- Kent, A.D., 2020. Expanded phylogenetic diversity and metabolic flexibility of mercury-methylating
- microorganisms. mSystems, 5 (4):e00299-20.
- McGrew, A.K., Ballweber, L.R., Moses, S.K., Stricker, C.A., Beckmen, K.B., Salman, M.D. and O'Hara, T.M., 2014. Mercury in gray wolves (*Canis lupus*) in Alaska: increased exposure through consumption of marine prey. Science of the Total Environment, 468-469:609-613.
- McKinney, M.A., Atwood, T.C., Pedro, S. and Peacock, E., 2017. Ecological change drives a decline in mercury concentrations in southern Beaufort Sea polar bears. Environmental Science & Technology, 51 (14):7814-7822.
- McMeans, B.C., Arts, M.T. and Fisk, A.T., 2015. Impacts of food web structure and feeding behavior on mercury exposure in Greenland Sharks (*Somniosus microcephalus*). Science of the Total Environment, 509-510:216-225.
- Michelutti N, Blais JM, Mallory ML, Brash J, Thienpont J, Kimpe LE, et al. Trophic position
- influences the efficacy of seabirds as metal biovectors. Proceedings of the National Academy of Sciences of the United States of America 2010; 107: 10543-10548.
- Moslemi-Aqdam M, Baker LF, Baltzer JL, Branfireun BA, Evans MS, Laird BD, et al. Understanding among-lake variability of mercury concentrations in Northern Pike (Esox lucius): A whole-ecosystem
- study in subarctic lakes. Science of The Total Environment 2022; 822: 153430.
- Munson, K.M., Lamborg, C.H., Swarr, G.J. and Saito, M.A., 2015. Mercury species concentrations and fluxes in the central tropical Pacific Ocean. Global Biogeochemical Cycles, 29 (5):656-676.
- Munthe, J., Bodaly, R.A., Branfireun, B.A., Driscoll, C.T., Gilmour, C.C., Harris, R., Horvat, M., Lucotte, M. and Malm, O., 2007. Recovery of mercury-contaminated fisheries. Ambio, 36 (1):33-44.
- Nagorski, S.A., Engstrom, D.R., Hudson, J.P., Krabbenhoft, D.P., Hood, E., DeWild, J.F. and Aiken, G.R., 2014. Spatial distribution of mercury in southeastern Alaskan streams influenced by glaciers,
- wetlands, and salmon. Environmental Pollution, 184:62-72.
- Obrist, D., Agnan, Y., Jiskra, M., Olson, C.L., Colegrove, D.P., Hueber, J., Moore, C.W., Sonke, J.E.
- and Helmig, D., 2017. Tundra uptake of atmospheric elemental mercury drives Arctic mercury pollution.
- Nature, 547 (7662):201-204.

 Olefeldt, D., Goswami, S., Grosse, G., Hayes, D., Hugelius, G., Kuhry, P., McGuire, A.D., Romanovsky, V.E., Sannel, A.B.K., Schuur, E.A.G. and Turetsky, M.R., 2016. Circumpolar distribution and carbon storage of thermokarst landscapes. Nature Communications, 7 (1):13043.

 Olefeldt D, Heffernan L, Jones MC, Sannel AB, Treat CC, Turetsky MR. 2021. Permafrost thaw in northern peatlands: rapid changes in ecosystem and landscape functions. Ecosystem Collapse and Climate Change.:27-67.

- Olson, C., Jiskra, M., Biester, H., Chow, J. and Obrist, D., 2018. Mercury in active-layer tundra soils of
- Alaska: concentrations, pools, origins, and spatial distribution. Global Biogeochemical Cycles, 32 (7):1058-1073.
- Parks, J.M., Johs, A., Podar, M., Bridou, R., Hurt, R.A., Jr., Smith, S.D., Tomanicek, S.J., Qian, Y.,
- Brown, S.D., Brandt. C.C., Palumbo, A.V., Smith, J.C., Wall, J.D., Elias, D.A. and Liang, L., 2013. The
- genetic basis for bacterial mercury methylation. Science, 339 (6125):1332-1335.
- Pedro, S., Fisk, A.T., Ferguson, S.H., Hussey, N.E., Kessel, S.T. and McKinney, M.A., 2019. Limited 1560 effects of changing prey fish communities on food quality for aquatic predators in the eastern Canadian
1561 Arctic in terms of essential fatty acids, methylmercury and selenium. Chemosphere, 214:855-865. Arctic in terms of essential fatty acids, methylmercury and selenium. Chemosphere, 214:855-865.
- Peterson, S.H., Ackerman, J.T., Crocker, D.E. and Costa, D.P., 2018. Foraging and fasting can
- influence contaminant concentrations in animals: an example with mercury contamination in a free-
- ranging marine mammal. Proceedings of the Royal Society B: Biological Sciences, 285
- (1872):20172782.
- Petrova, M.V., Krisch, S., Lodeiro, P., Valk, O., Dufour, A., Rijkenberg, M.J.A., Achterberg, E.P., Rabe,
- B., Rutgers van der Loeff, M., Hamelin, B., Sonke, J.E., Garnier, C., Heimbürger-Boavida, L.-E., 2020.
- Mercury species export from the Arctic to the Atlantic Ocean. Marine Chemistry, 225:103855.
- Piquet, A.M.-T., van de Poll, W.H., Visser, R.J.W., Wiencke, C., Bolhuis, H., Buma, A.G.J., 2014.
- 1570 Springtime phytoplankton dynamics in Arctic Krossfjorden and Kongsfjorden (Spitsbergen) as a 1571 function of glacier proximity. Biogeosciences, 11 (8):2263–2279.
- function of glacier proximity. Biogeosciences, 11 (8):2263–2279.
- Point, D., Sonke, J. E., Day, R. D., Roseneau, D. G., Hobson, K. A., Vander Pol, S. S., Moors, A. J.,
- Pugh, R. S., Donard, O. F. X., Becker P. R., 2011, Methylmercury photodegradation influenced by sea-ice cover in Arctic marine ecosystems, Nature Geoscience, 4, pages188–194
- Podar, M., Gilmour, C.C., Brandt, C.C., Soren, A., Brown, S.D., Crable, B.R., Palumbo, A.V.,
- Somenahally, A.C. and Elias, D.A., 2015. Global prevalence and distribution of genes and
- microorganisms involved in mercury methylation. Science Advances, 1 (9):e1500675.
- Pomerleau, C., Matthews, C.J.D., Gobeil, C., Stern, G.A., Ferguson, S.H. and Macdonald, R.W., 2018. Mercury and stable isotope cycles in baleen plates are consistent with year-round feeding in two bowhead whale (*Balaena mysticetus*) populations. Polar Biology, 41 (9):1881-1893.
- Pomerleau, C., Stern, G.A., Pućko, M., Foster, K.L., Macdonald, R.W. and Fortier, L., 2016. Pan-1582 Arctic concentrations of mercury and stable isotope ratios of carbon (δ^{13} C) and nitrogen (δ^{15} N) in
- marine zooplankton. Science of the Total Environment, 551-552:92-100.
- Pongratz, R. and Heumann, K.G., 1998. Production of methylated mercury and lead by polar macroalgae — a significant natural source for atmospheric heavy metals in clean room compartments. Chemosphere, 36 (9):1935-1946.
- Pokrovsky, O.S., Bueno, M., Manasypov, R.M., Shirokova, L.S., Karlsson, J. and Amouroux, D.,
- 2018. Dissolved organic matter controls seasonal and spatial selenium concentration variability in
- thaw lakes across a permafrost gradient. Environmental Science & Technology, 52 (18):10254-10262.
- Poste, A.E., Braaten, H.F.V., de Wit, H.A., Sørensen, K. and Larssen, T., 2015. Effects of photodemethylation on the methylmercury budget of boreal Norwegian lakes. Environmental Toxicology and Chemistry, 34 (6):1213-1223.
- Poste, A.E., Hoel, C.S., Andersen, T., Arts, M.T., Frærøvig, P-J. and Borgå, K., 2019. Terrestrial organic matter increases zooplankton methylmercury accumulation in a brown-water boreal lake. Science of the Total Environment, 674:9-18.
- Provencher, J.F., Forbes, M.R., Hennin, H.L., Love, O.P., Braune, B.M., Mallory, M.L., and Gilchrist, 2016. Implications of mercury and lead concentrations on breeding physiology and phenology in an Arctic bird. Environmental Pollution, 218:1014-1022.
- Pućko, M., Burt, A., Walkusz, W., Wang, F., Macdonald, R.W., Rysgaard, S., Barber, D.G., Tremblay, J.-É. and Stern, G.A., 2014. Transformation of mercury at the bottom of the Arctic food web: an overlooked puzzle in the mercury exposure narrative. Environmental Science & Technology, 48 (13):7280-7288.
- 1603 Rautio, M., Dufresne, F., Laurion, I., Bonilla, S., Vincent, W.F. and Christoffersen, K.S., 2011. Shallow
1604 freshwater ecosystems of the circumpolar Arctic. Écoscience. 18 (3):204-222. freshwater ecosystems of the circumpolar Arctic. Écoscience, $18 (3):204-222$.
- Richardson M, Chételat J, MacMillan GA, Amyot M. Mercury concentrations and associations with dissolved organic matter are modified by water residence time in eastern Canadian lakes along a 30° latitudinal gradient. Limnology and Oceanography 2021; 66: S64-S80.
- Rohonczy, J., Cott, P.A., Benwell, A., Forbes, M.R., Robinson, S.A., Rosabal, M., Amyot, M. and Chételat, 2020. Trophic structure and mercury transfer in the Subarctic fish community of Great Slave Lake, Northwest Territories, Canada. Journal of Great Lakes Research 46 (2):402-413.
- Rosati, G., Heimbürger, L.E., Melaku Canu, D., Lagane, C., Laffont, L., Rijkenberg, M.J.A., Gerringa,
- L.J.A., Solidoro, C., Gencarelli, C.N., Hedgecock, I.M., De Baar, H.J.W. and Sonke, J.E., 2018.
- 1613 Mercury in the Black Sea: new insights from measurements and numerical modeling. Global 1614 Biogeochemical Cycles, 32 (4):529-550.
- Biogeochemical Cycles, (4):529-550.
- Ruus, A., Øverjordet, I.B., Braaten, H.F.V., Evenset, A., Christensen, G., Heimstad, E.S., Gabrielsen, G.W. and Borgå, K., 2015. Methylmercury biomagnification in an Arctic pelagic food web.
- Environmental Toxicology & Chemistry, 34 (11):2636-2643.
- Ruuskanen M.O., Colby G., St. Pierre K.A., St.Louis V.AL., Aris-Brosou S., Poulain A.J. 2020.,
- Microbial genomes retrieved from High Arctic lake sediments encode for adaptation to cold and oligotrophic environments, Limnol. Oceanogr.65, S233–S247
- Rysgaard, S. and Glud, R.N., 2004. Anaerobic N2 production in Arctic sea ice. Limnology and Oceanography, 49 (1):86-94.
- Sandheinrich, M.B. and Drevnick, P.E., 2016. Relationship among mercury concentration, growth rate, and condition of northern pike: A tautology resolved? Environmental Toxicology and Chemistry, 35 (12):2910-2915.
- Saros, J.E., Osburn, C.L., Northington, R.M., Birkel, S.D., Auger, J.D., Stedmon, C.A. and Anderson, N.J., 2015. Recent decrease in DOC concentrations in Arctic lakes of southwest Greenland. Geophysical
- Research Letters, 42 (16):6703-6709.
- Schaefer, J.K. and Morel, F.M.M., 2009. High methylation rates of mercury bound to cysteine by
- Geobacter sulfurreducens. Nature Geoscience, 2 (2):123-126.
- Schaefer, K., Elshorbany, Y., Jafarov, E., Schuster, P.F., Striegl, R.G., Wickland, K.P. and Sunderland,
- E.M., 2020*.* Potential impacts of mercury released from thawing permafrost. Nature Communications,
- 11 (1):4650.
- Schartup, A.T., Balcom, P.H., Soerensen, A.L., Gosnell, K.J., Calder, R.S.D., Mason, R.P. and Sunderland, E.M., 2015a. Freshwater discharges drive high levels of methylmercury in Arctic marine biota. Proceedings of the National Academy of Sciences, 112 (38):11789-11794.
- Schartup, A.T., Ndu, U., Balcom, P.H., Mason, R.P. and Sunderland, E.M., 2015b. Contrasting effects of marine and terrestrially derived dissolved organic matter on mercury speciation and bioavailability in seawater. Environmental Science & Technology, 49 (10):5965-5972.
- Schartup, A.T., Qureshi, A., Dassuncao, C., Thackray, C.P., Harding, G. and Sunderland, E.M., 2018.
- 1641 A model for methylmercury uptake and trophic transfer by marine plankton. Environmental Science & Technology, 52 (2):654-662.
- Schartup, A.T., Soerensen, A.L. and Heimbürger-Boavida, L.-E., 2020. Influence of the Arctic sea-ice regime shift on sea-ice methylated mercury trends. Environmental Science & Technology Letters, 7 (10):708-713.
- Seewagen, C.L., Cristol, D.A. and Gerson, A.R., 2016. Mobilization of mercury from lean tissues during simulated migratory fasting in a model songbird. Scientific Reports, 6 (1):25762.
- Seller, P., Kelly, C.A., Rudd, J.W.M. and MacHutchon, A.R., 1996. Photodegradation of methylmercury in lakes. Nature, 380 (6576):694-697.
- Skrobonja, A., Gojkovic, Z., Soerensen, A.L., Westlund, P.-O., Funk, C. and Björn, E., 2019. Uptake
- kinetics of methylmercury in a freshwater alga exposed to methylmercury complexes with
- environmentally relevant thiols. Environmental Science & Technology, 53 (23):13757-13766.
- Skyllberg, U., Westin, M.B., Meili, M. and Björn, E., 2009. Elevated concentrations of methyl
- 1654 mercury in streams after forest clear-cut: a consequence of mobilization from soil or new methylation?
1655 Environmental Science & Technology, 43 (22):8535-8541.
- Environmental Science & Technology, 43 (22):8535-8541.
- Skyllberg U., 2012, Chemical Speciation of Mercury in Soil and Sediment, in: Liu, G., Cai, Y.,
- O'Driscoll, N.J. (Eds.), Environmental Chemistry and Toxicology of Mercury, pp. 219-258.
- Smith, S.D., Bridou, R., Johs, A., Parks, J.M., Elias, D.A., Hurt, R.A., Brown, S.D., Podar, M., Wall,
- J.D. and Kivisaar, M., 2015. Site-directed mutagenesis of *hgcA* and *hgcB* reveals amino acid residues
- important for mercury methylation. Applied and Environmental Microbiology, 81 (9):3205-3217.
- Sommar, J., Feng, X. and Lindqvist, O., 1999. Speciation of volatile mercury species present in digester and deposit gases. Applied Organometallic Chemistry, 13 (6):441-445.
- Soerensen, A.L., Schartup, A.T., Gustafsson, E., Gustafsson, B.G., Undeman, E. and Björn, E., 2016a.
- Eutrophication increases phytoplankton methylmercury concentrations in a coastal sea—a Baltic Sea case study. Environmental Science & Technology, 50 (21):11787-11796.
- Soerensen, A.L., Jacob, D.J., Schartup, A.T., Fisher, J.A., Lehnherr, I., St. Louis, V.L., Heimbürger, L.- E., Sonke, J.E., Krabbenhoft, D.P. and Sunderland, E.M., 2016b. A mass budget for mercury and methylmercury in the Arctic Ocean. Global Biogeochemical Cycles, 30 (4):560-575.
- Soerensen, A.L., Schartup, A.T., Skrobonja, A. and Björn, E., 2017. Organic matter drives high interannual variability in methylmercury concentrations in a Subarctic coastal sea. Environmental
- Pollution, 229:531-538.
- Sonke, J.E., Teisserenc, R., Heimbürger-Boavida, L.-E., Petrova, M.V., Marusczak, N., Le Dantec, T.,
- Chupakov, A.V., Li, C., Thackray, C.P., Sunderland, E.M., Tananaev, N. and Pokrovsky, O.S., 2018.
- Eurasian river spring flood observations support net Arctic Ocean mercury export to the atmosphere and
- Atlantic Ocean. Proceedings of the National Academy of Sciences, 115 (50):E11586-E11594.
- Steffen, A., Bottenheim, J., Cole, A., Ebinghaus, R., Lawson, G. and Leaitch, W.R., 2014. Atmospheric mercury speciation and mercury in snow over time at Alert, Canada. Atmospheric Chemistry and Physics, 14 (5):2219-2231.
- Streets, D.G, Horowitz, H.M., Lu, Z., Levin, L., Thackray, C.P., Sunderland, E.M., 2019.Five hundred years of anthropogenic mercury: spatial and temporal release profiles, Environ. Res. Lett. 14 084004
- St. Louis, V.L., Derocher, A.E., Stirling, I., Graydon, J.A., Lee, C., Jocksch, E., Richardson, E.,
- Ghorpade, S., Kwan, A.K., Kirk, J.L., Lehnherr, I. and Swanson, H.K., 2011. Differences in mercury
- bioaccumulation between polar bears (*Ursus maritimus*) from the Canadian High- and Sub-Arctic.
- Environmental Science & Technology, 45 (14):5922-5928.
- St. Louis, V.L., Sharp, M.J., Steffen, A., May, A., Barker, J., Kirk, J.L., Kelly, D.J.A., Arnott, S.E.,
- Keatley, B. and Smol, J.P., 2005. Some sources and sinks of monomethyl and inorganic mercury on
- Ellesmere Island in the Canadian High Arctic. Environmental Science & Technology, 39 (8):2686-2701.
- St. Louis, V.L., Hintelmann, H., Graydon, J.A., Kirk, J.L., Barker, J., Dimock, B., Sharp, M.J. and Lehnherr, I., 2007. Methylated mercury species in Canadian High Arctic marine surface waters and snowpacks. Environmental Science & Technology, 41 (18):6433-6441.
- St. Pierre, K.A., St. Louis, V.L., Kirk, J.L., Lehnherr, I., Wang, S. and La Farge, C., 2015. Importance of open marine waters to the enrichment of total mercury and monomethylmercury in lichens in the
- Canadian High Arctic. Environmental Science & Technology, 49 (10):5930-5938.
- St. Pierre, K.A., Zolkos, S., Shakil, S., Tank, S.E., St. Louis, V.L. and Kokelj, S.V., 2018. 1695 Unprecedented increases in total and methyl mercury concentrations downstream of retrogressive thaw
1696 slumps in the western Canadian Arctic. Environmental Science & Technology, 52 (24):14099-14109. slumps in the western Canadian Arctic. Environmental Science & Technology, 52 (24):14099-14109.
- St. Pierre, K.A., St. Louis, V.L., Lehnherr, I., Gardner, A.S., Serbu, J.A., Mortimer, C.A., Muir, D.C.G., Wiklund, J.A., Lemire, D., Szostek, L. and Talbot, C., 2019. Drivers of mercury cycling in the rapidly changing glacierized watershed of the High Arctic's largest lake by volume (Lake Hazen, Nunavut, Canada). Environmental Science & Technology, 53 (3):1175-1185.
- St. Pierre, K.A., Chétélat, J., Yumvihoze, E. and Poulain, A. J., 2014. Temperature and the sulfur cycle 1702 control monomethylmercury cycling in High Arctic coastal marine sediments from Allen Bay, Nunavut,
1703 Canada. Environmental Science & Technology, 48 (5):2680-2687. Canada. Environmental Science & Technology, 48 (5):2680-2687.
- Sunderland, E.M., Gobas, F.A.P.C., Branfireun, B.A. and Heyes, A., 2006. Environmental controls on the speciation and distribution of mercury in coastal sediments. Marine Chemistry, 102 (1-2):111-123.
- Sunderland, E.M., Krabbenhoft, D.P., Moreau, J.W., Strode, S.A. and Landing, W.M., 2009. Mercury sources, distribution, and bioavailability in the North Pacific Ocean: insights from data and models. Global Biogeochemical Cycles, 23 (2):GB2010.
- Swanson, H., Gantner, N., Kidd, K.A., Muir, D.C.G. and Reist, J.D., 2011. Comparison of mercury
- concentrations in landlocked, resident, and sea-run fish (*Salvelinus* spp.) from Nunavut, Canada. Environmental Toxicology and Chemistry, 30 (6):1459-1467.
	- Tank, S.E., Vonk, J.E., Walvoord, M.A., McClelland, J.W., Laurion, I. and Abbott, B.W., 2020. Landscape matters: predicting the biogeochemical effects of permafrost thaw on aquatic networks with
- a state factor approach. Permafrost and Periglacial Processes, 31 (3):358-370.
- Tarbier, B., Hugelius, G., Sannel, A.B.K., Baptista-Salazar, C. and Jonsson, S., 2021. Permafrost thaw
- increases methylmercury formation in Subarctic Fennoscandia. Environmental Science & Technology,
- 55 (10):6710–6717.
- Trudel, M. and Rasmussen, J.B., 2006. Bioenergetics and mercury dynamics in fish: a modelling perspective. Canadian Journal of Fisheries and Aquatic Sciences, 63 (8):1890-1902.
- Tsui, M.T.K. and Finlay, J.C., 2011. Influence of dissolved organic carbon on methylmercury
- bioavailability across Minnesota stream ecosystems. Environmental Science & Technology, 45
- (14):5981-5987.
- Tsz-Ki Tsui, M., Liu, S., Brasso, R.L., Blum, J.D., Kwon, S.Y., Ulus, Y., Nollet, Y.H., Balogh, S.J., Eggert, S.L. and Finlay, J.C., 2019. Controls of methylmercury bioaccumulation in forest floor food webs. Environmental Science & Technology, 53 (5):2434-2440.
- Vadeboncoeur, Y., Jeppesen, E., Zanden, M.J.V., Schierup, H.-H., Christoffersen, K. and Lodge,
- D.M., 2003. From Greenland to green lakes: cultural eutrophication and the loss of benthic pathways in lakes. Limnology and Oceanography, 48 (4):1408-1418.
- van der Velden, S., Evans, M.S., Dempson, J.B., Muir, D.C.G. and Power, M., 2013a. Comparative
- analysis of total mercury concentrations in anadromous and non-anadromous Arctic charr (*Salvelinus*
- *alpinus*) from eastern Canada. Science of the Total Environment, 447:438-449.
- van der Velden, S., Dempson, J.B., Evans, M.S., Muir, D.C.G. and Power, M. 2013b. Basal mercury concentrations and biomagnification rates in freshwater and marine food webs: effects on Arctic charr
- (*Salvelinus alpinus*) from eastern Canada. Science of the Total Environment, 444: 531-542.
- Varty S, Lehnherr I, St. Pierre K, Kirk J, Wisniewski V. Methylmercury Transport and Fate Shows Strong Seasonal and Spatial Variability along a High Arctic Freshwater Hydrologic Continuum. Environmental Science and Technology 2021; 55: 331-340.
- Vermilyea, A.W., Nagorski, S.A., Lamborg, C.H., Hood, E.W., Scott, D. and Swarr, G.J., 2017. Continuous proxy measurements reveal large mercury fluxes from glacial and forested watersheds in Alaska. Science of the Total Environment, 599-600:145-155.
- Villar, E., Cabrol, L. and Heimbürger-Boavida, L.-E., 2020. Widespread microbial mercury methylation genes in the global ocean. Environmental Microbiology Reports, 12 (3):277-287.
- Vonk, J.E., Tank, S.E., Bowden, W.B., Laurion, I., Vincent, W.F., Alekseychik, P., Amyot, M., Billet,
- M.F., Canário, J., Cory, R.M., Deshpande, B.N., Helbig, M., Jammet, M., Karlsson, J., Larouche, J.,
- MacMillan, G., Rautio, M., Walter Anthony, K.M. and Wickland, K.P., 2015. Reviews and syntheses:
- effects of permafrost thaw on Arctic aquatic ecosystems. Biogeosciences, 12 (23):7129-7167.
- Walvoord, M.A., Voss, C.I., Ebel, B.A. and Minsley, B.J., 2019. Development of perennial thaw
- zones in boreal hillslopes enhances potential mobilization of permafrost carbon. Environmental Research Letters, 14:015003.
- - Wang, F., Macdonald, R.W., Armstrong, D.A. and Stern, G.A., 2012b. Total and methylated mercury in the Beaufort Sea: the role of local and recent organic remineralization. Environmental Science &
	- Technology, 46 (21):11821-11828.
	- Wang F., Lemes M. and Khan M.A.K., 2012a. Metallomics of mercury: role of thiol- and selenol-
	- containing biomolecules. In: Cai, Y. and O'Driscoll, N. (Eds.). Environmental Chemistry and Toxicology of Mercury, pp. 517-544. John Wiley & Sons, Ltd.
- Wang, F., Pućko, M. and Stern, G., 2017. Chapter 19: Transport and transformation of contaminants in sea ice. In: Thomas, D.N. (Ed.). Sea Ice, Third Edition, pp. 472-491 John Wiley & Sons, Ltd.
- Wang, K., Munson, K.M., Beaupré-Laperrière, A., Mucci, A., Macdonald, R.W. and Wang, F., 2018. Subsurface seawater methylmercury maximum explains biotic mercury concentrations in the Canadian Arctic. Scientific Reports, 8 (1):14465.
- West, J., Graham, A.M., Liem-Nguyen, V. and Jonsson, S., 2020. Dimethylmercury degradation by dissolved sulfide and mackinawite. Environmental Science & Technology, 54 (21):13731-13738.
- West J., Gindorf S., Jonsson S., 2022, Photochemical Degradation of Dimethylmercury in Natural Waters, Environ. Sci. Technol. 56, 9, 5920–5928
- Wietrzyk-Pełka, P., Rola, K., Szymański, W. and Węgrzyn, M.H., 2020. Organic carbon accumulation in the glacier forelands with regard to variability of environmental conditions in different ecogenesis
- stages of High Arctic ecosystems. Science of the Total Environment, 717:135151.
- Willis C.E., Kirk J., St. Louis V.L., Lehnherr I., Ariya P.A., Rangel-Alvarado R.B., 2018. Sources of
- Methylmercury to Snowpacks of the Alberta Oil Sands Region: A Study of In Situ Methylation and
- Particulates. Environmental Science & Technology 52,2,531-540
- Wu, P., Kainz, M.J., Bravo, A.G., Åkerblom, S., Sonesten, L. and Bishop, K., 2019. The importance
- of bioconcentration into the pelagic food web base for methylmercury biomagnification: a meta-
- analysis. Science of the Total Environment, 646:357-367.
- Yang, Z., Fang, W., Lu, X., Sheng, G.-P., Graham, D.E., Liang, L., Wullschleger, S.D. and Gu, B., 2016. Warming increases methylmercury production in an Arctic soil. Environmental Pollution, 214:504-509.
- Zdanowicz, C. M., E. M. Krümmel, A. J. Poulain, E. Yumvihoze, J. Chen, M. Štrok, M. Scheer, and H. Hintelmann., 2016, Historical variations of mercury stable isotope ratios in Arctic glacier firn and ice cores, Global Biogeochem. Cycles, 30, 1324–1347
- Zhang, B., Chen, T., Guo, J., Wu, M., Yang, R., Chen, X., Wu, X., Zhang, W., Kang, S., Liu, G. and
- Dyson, P., 2020. Microbial mercury methylation profile in terminus of a high-elevation glacier on the
- northern boundary of the Tibetan Plateau. Science of the Total Environment, 708: 135226.
- 1783 Zhou J, Obrist D. 2021. Global Mercury Assimilation by Vegetation. Environmental Science & 1784 Technology: 55: 14245-14257. Technology; 55: 14245-14257.