

Direct [11 C]Methylation of Amines from [11C]CO 2 for the Synthesis of PET Radiotracers

François Liger, Ture Eijsbouts, Florence Cadarossanesaib, Christian Tourvieille, Didier Le Bars, Thierry Billard

► To cite this version:

François Liger, Ture Eijsbouts, Florence Cadarossanesaib, Christian Tourvieille, Didier Le Bars, et al.. Direct [11 C]Methylation of Amines from [11C]CO 2 for the Synthesis of PET Radiotracers. European Journal of Organic Chemistry, 2015, 2015 (29), pp.6434-6438. 10.1002/ejoc.201500924 . hal-03719209

HAL Id: hal-03719209 https://hal.science/hal-03719209v1

Submitted on 11 Jul2022

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

Direct ¹¹C-Methylation of Amines from [¹¹C]CO₂ for the Synthesis of PET Radiotracers.

François Liger,^[b] Ture Eijsbouts,^{[a],[b]} Florence Cadarossanesaib,^[b] Christian Tourvieille,^[b] Didier Le Bars,^{[a],[b]} Thierry Billard^{*[a],[b]}

Abstract: ¹¹C-Methylation ($t_{1/2} = 20.4$ min) is a main labeling strategy in the development of PET radiotracers. A straight radiomethylation of amines by use of cyclotron-produced ¹¹CO₂ was developed to obtain various radiolabeled compounds with good radiochemical yields. This strategy constitutes a new approach to simplify the radiolabeling processes and transpositions onto automatic synthesizers. This method was applied to the syntheses of radiolabeled drugs and a PET radiotracer used in Alzheimer's disease studies.

Introduction

PET (positron emission tomography) imaging is a nuclear medicine technology increasingly used not only in routine clinical diagnosis^[1] but also in biomedical research and drug development.^[2] These growing applications of PET imaging require the development of more and more various radiotracers. However, the short-lived positron-emitters impose specific synthetic methods, easily transferable onto automatic synthesizers.^[16, 3]

In general, [¹⁸F] labeled molecules are preferred for production and use in routine clinical imaging due to the higher half-life of fluorine-18, as illustrated by the recent development of various methods for radiofluorination.^[4] Nevertheless, despite the ¹¹C short half-life (t_{1/2} = 20.4 min), [¹¹C] labeled molecules still represent valuable targets for research studies. Indeed, easy-to-introduce carbon-substituents are present in a large majority of drugs and bioactive compounds. Consequently, a direct radiolabeled equivalent of molecule is easier to envisage.

The radioisotope carbon-11 is usually produced from a cyclotron by the ¹⁴N(p, α)¹¹C nuclear reaction in the form of either of two precursors: ¹¹CH₄ or ¹¹CO₂. Direct use of these two compounds is quite limited and they are, usually, converted into more reactive species. Some direct synthetic applications of

[a]	T. Eijsbouts, Dr D. Le Bars, Dr T. Billard Institute of Chemistry and Biochemistry (ICBMS – UMR CNRS 5246)
	Université de Lyon, Université Lyon 1, CNRS
	43 Bd du 11 novembre 1918 – 69622 Lyon (France)
	E-mail: Thierry.billard@univ-lyon1.fr www.FMI-Lyon.fr
[b]	F. Liger, T. Eijsbouts, F. Cadarossanesaib, C. Tourvieille, Dr D. Le Bars, Dr T. Billard
	CERMEP - IN VIVO IMAGING Groupement Hospitalier Est
	59 Bd Pinel – 69677 I von (France)
	www.cermep.fr

¹¹CO₂ have been previously described^[5] to obtain [¹¹C] labeled carboxylic acids by reaction with Grignard and organolithium reagents^[3a, 6] or, more recently, by cross-coupling reactions with boronic acids.^[7] The syntheses of [¹¹C] labeled ureas and carbamates have been also described.^[8]

Methylation reactions of heteroatoms, in particular nitrogen, constitute one of the more classic approaches for ¹¹C-labeling of molecules,^[3a, 6] as illustrated by the radiosynthesis of the β amyloid imaging radioligand [11C]PIB.[9] These radiomethylations require preliminary transformation of ¹¹CO₂ into very reactive species,¹¹CH₃I or ¹¹CH₃OTf.^[3a, 6, 10] These reagents are often used under basic conditions in a strictly anhydrous environment. Such conditions can require additional protection and deprotection steps of incompatible functional groups that substantially prolong and complicate the radiolabeling process. Furthermore, from a technical point of view the simplest radiosynthesis, with a minimum of steps, will lead to an easier automation.^[10-11] Consequently, a direct radiomethylation of amines using ¹¹CO₂, generated from cyclotron, appears very attractive. Some previous works have been described,^[12] however, these strategies require a multi-steps procedure: ¹¹CO₂ is first fixed with silvlated amines (preliminary prepared) to give O-silyl carbamates, which are subsequently reduced in situ by LiAlH₄ (a harsh reducer). Therefore, a more direct and soft procedure could be beneficial.

Results and Discussion

Recently, some groups have described the application of CO₂ as a C1-building block for catalytic methylation of amines.^[13] Such elegant works appeared in perfect adequacy with our effort to develop a simpler and more direct way to perform ¹¹C-methylation of amines. This approach would eliminate the preliminary time-consuming, and sometimes sources of failure during the automatic radiolabeling, steps of preparation of reactive methylating reagents. We have focused our interest onto the strategy developed by Cantat et al. using standard and simple catalytic system (ZnCl₂, IPr), without use of additional hydrogen gas.^[13b] As within this article, N-methyl aniline (**1a**) has been selected as model substrate (Table 1).

With a similar catalytic system, in the same solvent and at the same temperature than the optimal described conditions, only 3% of expected methylation was observed in 20 min (entry 1). Such disappointing results can be rationalized by the short time of reaction, imposed by ¹¹C half-life, compared to the 20 hours required in original publication. To increase the reaction kinetic, a solvent with similar properties and polarity but with a higher boiling point was selected to increase reaction temperature. Thus, in diglyme, at 150°C, a satisfactory radiochemical yield of 40% has been obtained (entry 2). Because of the small molar amount of ¹¹CO₂ generated by cyclotron (e.g. 1 Ci, i.e. 37 GBq, correspond to 108 pmol), the precursor (1a) quantity was decreased to match standard amount (around 1-2 mg) used in classic radiolabeling without any change in yield (entry 3). It should be noticed that, for practical reasons (weighing), the same amount of catalyst was preserved, bringing the catalyst amount to stoichiometry (related to 1a). With a shortened reaction time of 10 min, the radiochemical yield was halved (entry 4), even with a doubled catalyst amount (entry 5). The presence of both ZnCl₂ and IPr is proving to be essential (entries 6-8). Preliminary complex formation [ZnCl₂;IPr]^[14] appears to be deleterious for radiolabeling, in contrast with the work of Cantat et al. (entry 9).^[13b] In order to increase ¹¹CO₂ concentration in solution to potentially accelerate the reaction, TMEDA (N,N,N',N'tetramethylethylenediamine) as CO2 trap was added, as described by Pike et al.[7] However, no reaction was then observed: TMEDA certainly competes with 1a to catch CO2 (entry 10). Finally, microwave irradiation appeared to be also detrimental for the reaction; the increased molecular agitation contributes to break the amine-CO2 interaction, which seems to be essential to initiate the methylation (entry 11). Furthermore, Dyson et al. have described similar methylation in metal-free conditions with another carbene (NHC, 1,3-bis(2,4,6trimethylphenyl)-1,3-dihydro-2H-imidazol-2-ylidene).[13e] Unfortunately, in our conditions, no expected radiolabeling was

Moreover, the trapping efficiency of cyclotron-produced ${}^{11}CO_2$ was measured using the optimal conditions (Table 1, entry 3) at a trapping flow rate of 10 mL/min. Untrapped ${}^{11}CO_2$ was confined within an ascarite[®] cartridge attached to the reactor outlet. Up to 80% of the delivered ${}^{11}CO_2$ was kept inside the reactor at the end of entrapment (trapping time = 60s). More precisely, the trapping efficiency depends on the amines basicity: 80% with **1d**, 70% with **1a** and 65% with **1e**.

observed (entry 12).

Table 1. Direct ¹¹C-methylation of 1a with ¹¹CO₂.^[a]



Entry	[1a] (M)	Solvent	T (°C)	t (min)	RCY ^[b] (%)
1	0.231	THF	80	20	3
2	0.231	Diglyme	150	20	40
3	0.023	Diglyme	150	20	43
4	0.023	Diglyme	150	10	23
5 ^[c]	0.023	Diglyme	150	10	27
6 ^[d]	0.023	Diglyme	150	20	0
7 ^[e]	0.023	Diglyme	150	20	0
8 ^[f]	0.023	Diglyme	150	20	3
ð [a]	0.023	Diglyme	150	20	6
10 ^[h]	0.023	Diglyme	150	20	0
11 ^[i]	0.023	Diglyme	150	5	4
120	0.023	Diglyme	150	20	0

[a] Conditions: Solvent (400µL), ZnCl₂ (1,3 mg), **IPr** (3,5 mg), PhSiH₃ (23 µL). [b] Radiochemical yields (RCY) are estimated from trapped ¹¹CO₂ within the reactor and are decay-corrected from the end of ¹¹CO₂ trapping inside reactor. [c] ZnCl₂ (3,5 mg), **IPr** (7 mg). [d] Without ZnCl₂ and **IPr**. [e] Without **IPr**. [f] Without ZnCl₂. [g] With preformed complex [IPr;ZnCl₂] (6 mg). [h] With addition of TMEDA (70µL). [i] Under microwave irradiation (140 W). [j] Without ZnCl₂, with **NHC** (2 mg) instead of **IPr** and Ph₂SiH₂ (23 µL) instead of PhSiH₃.

With the optimal reaction conditions in hand (Table 1, entry 3), the scope of this new radiolabeling strategy was investigated using various amines (scheme 1).



Scheme 1. Direct ¹¹C-methylation of amines with ¹¹CO₂. RCY values are mean (n=3). They are estimated from trapped ¹¹CO₂ within the reactor and are decay-corrected from the end of ¹¹CO₂ trapping inside reactor. [a] ratio **3h/3b** = 60:40. [b] with 2 equiv. of **1h** vs ZnCl₂.

These ¹¹C-methylations gave generally satisfactorv radiochemical yields with aromatic or aliphatic amines. Primary amines are selectively mono-methylated and, generally, only a small amount of bis-methylation was observed. With anilines, presence of electron-withdrawing or electron-donor the substituents seems to have no relevant influence (3b-3h). In accordance with the supposed mechanism (see scheme 2), electron-withdrawing groups should favor the first step of CO2 trapping by amine whereas electron-donor substituents should more favor the second and third reduction steps. This reaction is also compatible with ester function (3f). In the case of cyano group, a more moderate yield is observed, maybe due to partial chelation of ZnCl₂ by the nitrogen atom partially guenching the reaction. Starting from p-bromoaniline (1h), a satisfactory yield of expected product was obtained, despite the formation of 3b, arising from the partial reduction of the C-Br bond. This radiolabeling appears also to be chemoselective since in case of amino-alcohol, only N-methylation is observed (3k). However, to achieve similar radiochemical yields compared to other amines, an excess of amino-alcohol (vs ZnCl₂) was required. This could be explained by chelate formation between amino-alcohol and zinc which could deactivate the nitrogen atom and thus would disfavor the CO₂ trapping. Finally, this methodology could be applied to the synthesis of radiolabeled drugs, such as ephedrine (sympathomimetic amine, 3k), imipramine (tricyclic antidepressant, **3**j) and the β -amyloid radiotracer [¹¹C]PIB (Alzheimer disease diagnostic, 3I).

From a mechanistic point of view, Cantat et al. have proposed the over reduction of an intermediate formamide (Scheme 2, steps 2-3).^[13b] Furthermore, radiomethylation by reduction of a formamide has been previously reported.^[15] In our conditions the CO₂-amine interaction appeared to be determining. Indeed, an increase of molecular agitation by micro-wave^[16] disfavored the reaction by breaking this interaction. In addition, starting from imipramine hydrochloride (**3g**), no labeling was observed due to the impossible CO₂-amine interaction. During the radiomethylation of **1a**, the corresponding labeled formamide, could be identified by HPLC. These observations led us to proposed the following pathway (Scheme 2): 1) formation of CO₂-amine complex; 2) 1st reduction in formamide; 3) 2nd reduction in expected methyl amine.



Scheme 2. Hypothesized mechanism for the ¹¹C-methylation with ¹¹CO₂.

This direct ¹¹C-labeling strategy was applied to effective production of the β -amyloid radiotracer [¹¹C]PIB^[17] (**3i**) with HPLC purification and final formulation step (Scheme 3). [¹¹C]PIB was produced in sufficient radioactive amount (57 mCi – RCY = 38%), with a high radiochemical and chemical purity. Such result is very encouraging and demonstrates the possibility to extrapolate this new strategy to a "productive scale" of radiotracers. Specific radioactivity remains however weak (SA = 15 GBq.µmol⁻¹), certainly due to excess of environmental CO₂ which dilutes ¹¹CO₂. Nevertheless, in a classical production of [¹¹C]PIB ([¹¹C]methyl triflate method) with the same synthesizer and a similar production of ¹¹CO₂ by the same cyclotron, [¹¹C]PIB is obtained with a similar radiochemical yield of 45% and a specific activity of around 50 GBq.µmol⁻¹.



Scheme 3. Production of β -amyloid radiotracer [¹¹C]PIB. RCY value is estimated from trapped ¹¹CO₂ within the reactor and are decay-corrected from the end of ¹¹CO₂ trapping inside reactor

Conclusions

In conclusion, we have brought the proof of concepts of direct and selective amine ¹¹C-methylation by direct use of cyclotronproduced ¹¹CO₂. Even if some improvements are still required, these preliminary data should open the way for new developments of direct and simple radiolabeling methods.

Experimental Section

Synthesis of [¹¹C]PIB (**3i**). Cyclotron-produced ¹¹CO₂, trapped (953 mCi) on a column of molecular sieves (4 Å), was released by purging the heated column (350°C) with He gas and bubbled, through Teflon lines, into the reactor containing ZnCl₂ (1.3mg), **IPr** (3.5 mg), **1i** (1.7 mg) in 400 μ L of diglyme and PhSiH₃ (23 μ L) cooled at 0°C (¹¹CO₂ traps: 711 mCi). Teflon lines were removed when radioactivity content reached a maximum and the reacting mixture was heated at 150°C for 20 min. After cooling, 2.2 mL of HPLC mobile phase were added to the mixture and the resulting solution was purified by HPLC on a Waters Symmetry-Prep C18 column (7 μ m, 7.8 x 300 mm) at a flow rate of 4 mL/min (H₂O/MeCN 60:40 v/v). [¹¹C]PIB fraction (t_R = 11 min) was collected, diluted in 40 mL of water and formulated by SPE. After rinsing with 10 mL of water, the purified product was released from the Sep-Pak (Waters Plus tC18) with 1 mL of ethanol and 2 mL of water in a sterile vial (57 mCi).

Acknowledgements

This work has been supported by the RADIOMI project (EU FP7-PEOPLE-2012-ITN-RADIOMI). Dr Anis Tlili (ICBMS – UMR CNRS 5246) is thanked for fruitful discussions. Frédéric Bonnefoi and Thibaut lecker (CERMEP - in vivo imaging) are acknowledged for their technical assistance. The authors wish to thank CNRS for financial supports.

Keywords: alkylation • carbon-11 • positron emission tomography • radiochemistry • isotopic labeling

- a) W. Cai, Mol. Pharm. 2014, 11, 3773-3776; b) L. Zimmer, A. Luxen, Neuroimage 2012, 61, 363-370; c) G. B. Saha, Basics of PET Imaging: Physics, Chemistry, and Regulations, Springer New York, 2010.
- a) C. C. Wagner, O. Langer, *Adv. Drug Del. Rev.* 2011, 63, 539-546; b)
 R. Chakravarty, H. Hong, W. Cai, *Mol. Pharm.* 2014, *11*, 3777-3797; c)
 J. K. Willmann, N. van Bruggen, L. M. Dinkelborg, S. S. Gambhir, *Nat. Rev. Drug Discov.* 2008, *7*, 591-607.
- a) P. W. Miller, N. J. Long, R. Vilar, A. D. Gee, Angew. Chem. 2008, 120, 9136-9172; Angew. Chem. Int. Ed. 2008, 47, 8998-9033; b) Z. Li,
 P. S. Conti, Adv. Drug Del. Rev. 2010, 62, 1031-1051; c) P. A. Schubiger, L. Lehmann, PET Chemistry: The Driving Force in Molecular Imaging, Springer, 2007.

- [4] a) L. Li, M. N. Hopkinson, R. L. Yona, R. Bejot, A. D. Gee, V. Gouverneur, *Chem. Sci.* 2011, *2*, 123; b) E. L. Cole, M. N. Stewart, R. Littich, R. Hoareau, P. J. H. Scott, *Curr. Top. Med. Chem.* 2014, *14*, 875-900.
- B. H. Rotstein, S. H. Liang, J. P. Holland, T. L. Collier, J. M. Hooker, A. A. Wilson, N. Vasdev, *Chem. Commun.* **2013**, *49*, 5621-5629.
- [6] a) M. Allard, E. Fouquet, D. James, M. Szlosek-Pinaud, *Curr. Med. Chem.* 2008, *15*, 235-277; b) P. J. H. Scott, *Angew. Chem.* 2009, *121*, 6115-6118; *Angew. Chem. Int. Ed.* 2009, *48*, 6001-6004.
- P. J. Riss, S. Lu, S. Telu, F. I. Aigbirhio, V. W. Pike, Angew. Chem. 2012, 124, 2752-2756; Angew. Chem. Int. Ed. 2012, 51, 2698-2702.
- [8] a) J. M. Hooker, A. T. Reibel, S. M. Hill, M. J. Schueller, J. S. Fowler, Angew. Chem. 2009, 121, 3534-3537; Angew. Chem. Int. Ed. 2009, 48, 3482-3485; b) J. M. Hooker, M. Schönberger, H. Schieferstein, J. S. Fowler, Angew. Chem. 2008, 120, 6078-6081; Angew. Chem. Int. Ed. 2008, 47, 5989-5992; c) A. A. Wilson, A. Garcia, S. Houle, O. Sadovski, N. Vasdev, Chem. Eur. J. 2011, 17, 259-264.
- a) C. A. Mathis, Y. Wang, D. P. Holt, G.-F. Huang, M. L. Debnath, W. E. Klunk, *J. Med. Chem.* 2003, *46*, 2740-2754; b) A. A. Wilson, A. Garcia, A. Chestakova, H. Kung, S. Houle, *J. Labelled Compd. Radiopharm.* 2004, *47*, 679-682.
- [10] G. Antoni, J. Labelled Compd. Radiopharm. 2015, 58, 65-72.
- a) V. Gomez-Vallejo, J. Llop, *Nucl. Med. Commun.* 2011, *32*, 1011-1017; b) C. Philippe, D. Haeusler, M. Mitterhauser, J. Ungersboeck, H. Viernstein, R. Dudczak, W. Wadsak, *Appl. Radiat. Isot.* 2011, *69*, 1212-1217; c) M. Verdurand, G. Bort, V. Tadino, F. Bonnefoi, D. Le Bars, L. Zimmer, *Nucl. Med. Commun.* 2008, *29*, 920-926.
- a) S. Ram, R. E. Ehrenkaufer, D. M. Jewett, *Appl. Radiat. Isot.* **1986**, *37*, 391-395; b) S. Ram, L. D. Spicer, *Appl. Radiat. Isot.* **1989**, *40*, 413-416; c) S. Ram, R. E. Ehrenkaufer, L. D. Spicer, *Appl. Radiat. Isot.* **1989**, *40*, 425-427.
- [13] a) O. Jacquet, C. Das Neves Gomes, M. Ephritikhine, T. Cantat, J. Am. Chem. Soc. 2012, 134, 2934-2937; b) O. Jacquet, X. Frogneux, C. Das Neves Gomes, T. Cantat, Chem. Sci. 2013, 4, 2127-2131; c) Y. Li, I. Sorribes, T. Yan, K. Junge, M. Beller, Angew. Chem. 2013, 125, 12378-12382; Angew. Chem. Int. Ed. 2013, 52, 12156-12160; d) Y. Li, X. Fang, K. Junge, M. Beller, Angew. Chem. 2013, 125, 9747-9750; Angew. Chem. Int. Ed. 2013, 52, 9568-9571; e) S. Das, F. D. Bobbink, G. Laurenczy, P. J. Dyson, Angew. Chem. 2014, 126, 13090-13093; Angew. Chem. Int. Ed. 2014, 53, 12876-12879.
- [14] D. Wang, K. Wurst, M. R. Buchmeiser, J. Organomet. Chem. 2004, 689, 2123-2130.
- [15] G. Berger, M. Maziere, R. Knipper, C. Prenant, D. Comar, Int. J. Appl. Radiat. Isot. 1979, 30, 393-399.
- a) M. B. Gawande, S. N. Shelke, R. Zboril, R. S. Varma, *Acc. Chem. Res.* 2014, *47*, 1338-1348; b) P. Lidström, J. Tierney, B. Wathey, J. Westman, *Tetrahedron* 2001, *57*, 9225-9283.
- [17] a) J. P. Holland, S. H. Liang, B. H. Rotstein, T. L. Collier, N. A. Stephenson, I. Greguric, N. Vasdev, *J. Labelled Compd. Radiopharm.* 2014, *57*, 323-331; b) N. S. Mason, C. A. Mathis, W. E. Klunk, *J. Labelled Compd. Radiopharm.* 2013, *56*, 89-95.