



## Brackish diatom species observed at the bituminous Poix spring: an island of curiosity

Lory-Anne Baker, Aude Beauger, Carlos E Wetzel, Olivier Voldoire, Christelle Blavignac, Elisabeth Allain, Luc Ector, David Biron

### ► To cite this version:

Lory-Anne Baker, Aude Beauger, Carlos E Wetzel, Olivier Voldoire, Christelle Blavignac, et al.. Brackish diatom species observed at the bituminous Poix spring: an island of curiosity. BIOM - Revue scientifique pour la biodiversité du Massif central, 2022, 3 (1), pp.40 - 51. 10.52497/biom.v3i1.304 . hal-03703802

**HAL Id: hal-03703802**

**<https://hal.science/hal-03703802>**

Submitted on 28 Jun 2022

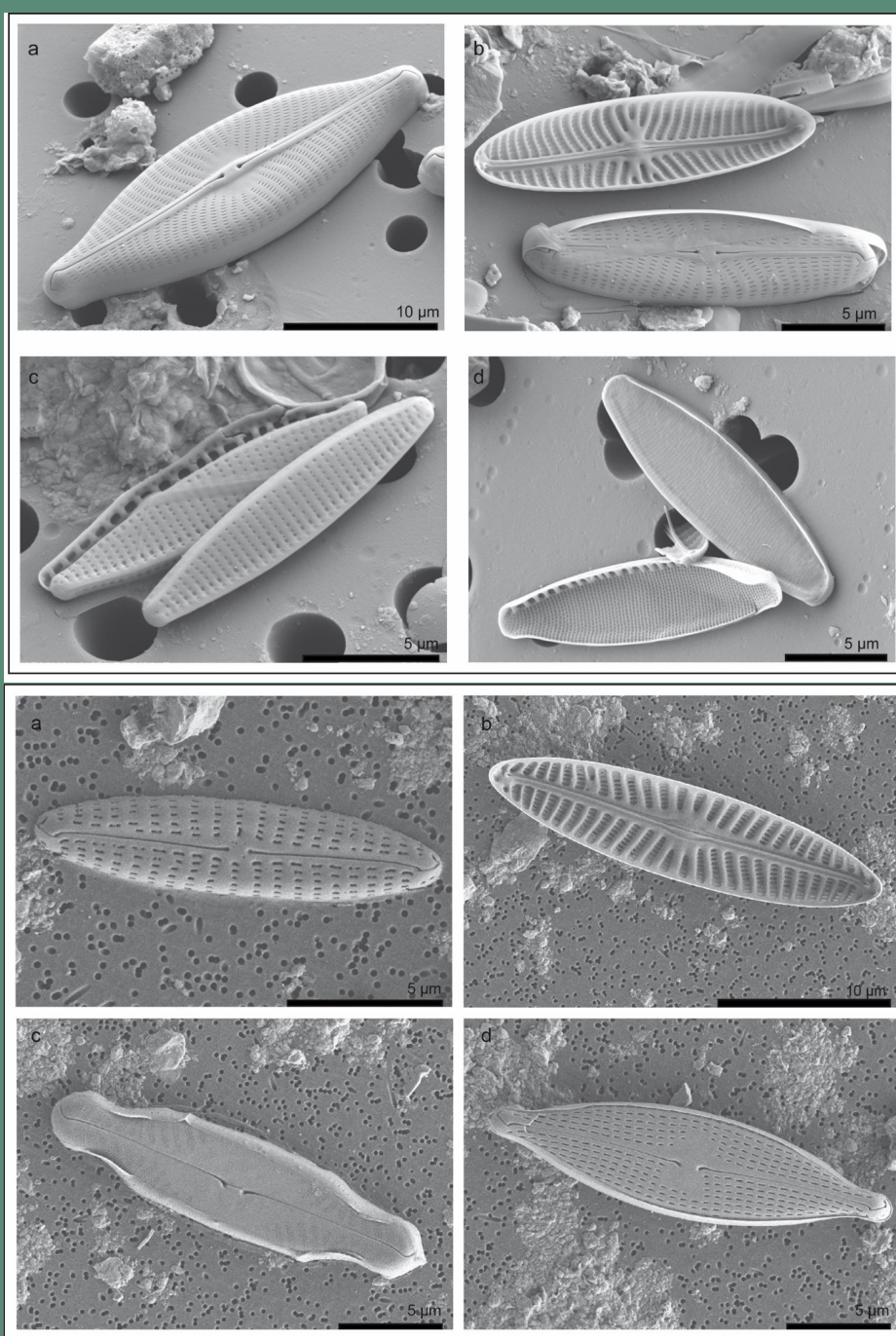
**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



# BIOM

Revue scientifique pour la biodiversité  
du Massif central



**Brackish diatom species observed at the bituminous Poix spring:  
an island of curiosity**

*Baker et al. / BIOM 3 (2022) : 40-51*

## Brackish diatom species observed at the bituminous Poix spring: an island of curiosity

Lory-Anne Baker<sup>1,2</sup>, Aude Beauger<sup>2,3</sup>, Carlos E. Wetzel<sup>4</sup>, Olivier Voldoire<sup>3</sup>, Christelle Blavignac<sup>5</sup>,  
Elisabeth Allain<sup>3</sup>, Luc Ector<sup>4</sup>† & David Biron<sup>1,2</sup>†

<sup>1</sup>Université Clermont Auvergne, CNRS, LMGE, F-63000 Clermont-Ferrand, France ;

<sup>2</sup>LTSER "Zone Atelier Territoires Uranifères", 63000 Clermont-Ferrand, 63178 Aubière Cedex, France ;

<sup>3</sup>Université Clermont Auvergne, CNRS, GEOLAB, F-63000 Clermont-Ferrand, France ;

<sup>4</sup>Luxembourg Institute of Science and Technology (LIST), Environmental Research and Innovation department (ERIN), Environment and Biodiversity (OCEB), 4422 Belvaux, Luxembourg ;

<sup>5</sup>Centre Imagerie Cellulaire Santé, UCA PARTNER, F-63000 Clermont-Ferrand, France.

Corresponding author : Lory-Anne Baker, l-anne.baker@doctorant.uca.fr, Université Clermont Auvergne, CNRS, LMGE, F-63000 Clermont-Ferrand, France.

Soumis le 22 mars 2022

Accepté le 30 mai 2022

Publié le 20 juin 2022

### Résumé

La source de la Poix est une particularité géologique du Massif central français car elle est connue pour être bitumineuse associée à de l'eau salée, du méthane et des traces d'hydrogène sulfuré. Entre mars 2017 et juin 2020, trois prélèvements ont été effectués en raclant les diatomées sur la couche de bitume. De plus, les conditions physiques et chimiques ont été examinées. Il apparaît que la conductivité de l'eau salée est supérieure à celle de l'océan, entraînant le développement de certaines espèces de diatomées saumâtres, observées pour certaines d'entre elles sur les sédiments intertidaux ou dans les estuaires. Même si les conditions abiotiques étaient extrêmes pour ces algues, la biodiversité était élevée et révèle des espèces ayant une grande capacité à supporter des concentrations et des fluctuations de salinité élevées.

### Abstract

The Poix spring is a geological particularity of the French Massif central as it is known to be bitumen associated with salt water, methane and traces of hydrogen sulphide. Between March 2017 and June 2020, three samples were taken by scraping the diatoms on the layer of bitumen. Moreover, the physical and chemical conditions were examined. It appears that the conductivity of the saline water is higher than the one of the ocean leading to the development of some brackish diatom species observed for some of them on the intertidal sediments or estuaries. Even if the abiotic conditions were extreme for these algae, the biodiversity was high and reveals species with high ability to withstand in high salinity concentrations and fluctuations.

#### Mots-clés

Eau saumâtre  
Biodiversité  
Communautés de diatomées  
Sources minérales

#### Keywords

Brackish water  
Biodiversity  
Diatoms communities  
Mineral springs

## Introduction

Brackish water is defined as having a chloride content more than  $400 \text{ mg.L}^{-1}$  or an electrical conductivity greater than  $1.5 \text{ dS.m}^{-1}$  (Shevah 2014). Brackish environments are classically observed on coastal areas, estuary but also in inland habitats. When considering inland areas, these are habitats under the influence of saline waters that accompany the salt deposits, as well as areas exposed to water deficit in dry climate (Reimold & Queen 1974; Poljakoff-Mayber & Gale 1975). These brackish environments are relatively widespread throughout the world and cover ca. 10% of the land (O'Leary & Glenn 1994; Piernik 2012). Inland salt environments occur in limited areas, mostly on "fossil salt deposits and around salty springs" (Waisel 1972; Piernik 2012).

Among the springs, there a wide range of physical and chemical conditions with conductivity ranging from 10 to  $70,000 \text{ }\mu\text{S.cm}^{-1}$  (Sabater & Roca 1992; Angeli et al. 2010; Želazna-Wieczorek 2011; Wojtal 2013; Beauger et al. 2015, 2016, 2017, 2019). When the conductivity is the highest with high sodium concentration, springs are associated with halophyte vegetation or micro-organisms adapted to high osmotic pressure. Among these micro-organisms, diatoms are present and distributed in this particular habitat. These microscopic algae are known to

have ecological preferences for most of them. So, brackish diatom species are associated and grow in salty springs (Rakowska 1997; Wojtal 2013; Lai et al. 2016).

In the French Massif central, there are many salt marshes and among the 500 inventoried mineral springs (Lecoq 1864), some are salty springs that induced this brackish ecosystem. Indeed, an on-going survey on diatoms that began in 2015, reveals that the conductivity of the mineral springs ranges between 37 and  $123,200 \text{ }\mu\text{S.cm}^{-1}$  with an average of  $4800 \text{ }\mu\text{S.cm}^{-1}$  (Beauger, personal communication). The Poix spring at Clermont-Ferrand is the mineral spring with the highest conductivity and salinity of this region. Since at least antiquity, this spring has drawn attention particularly for the occurrence of bitumen with water. In 1575, the French author François de Belle-Forêt described the area as a hill where bitumen and spring water flow in the book "La cosmologie universelle de tout le monde". Later, this spring was also mentioned by the doctor Jean Banc in 1604, the chemist Joseph de Tournefort in 1734, the geologist Jean-Etienne Guettard in 1759 and the famous naturalists Antoine Delarbre in 1749, Georges-Louis Leclerc de Buffon in 1799 and Henri Lecoq in 1864 (Nivet 1846; Truchot 1878; Legrand 2019). Focusing on the mineral Poix spring, the main aim of the study was to estimate the biodiversity associated with these harsh conditions by studying diatom communities during different seasons.

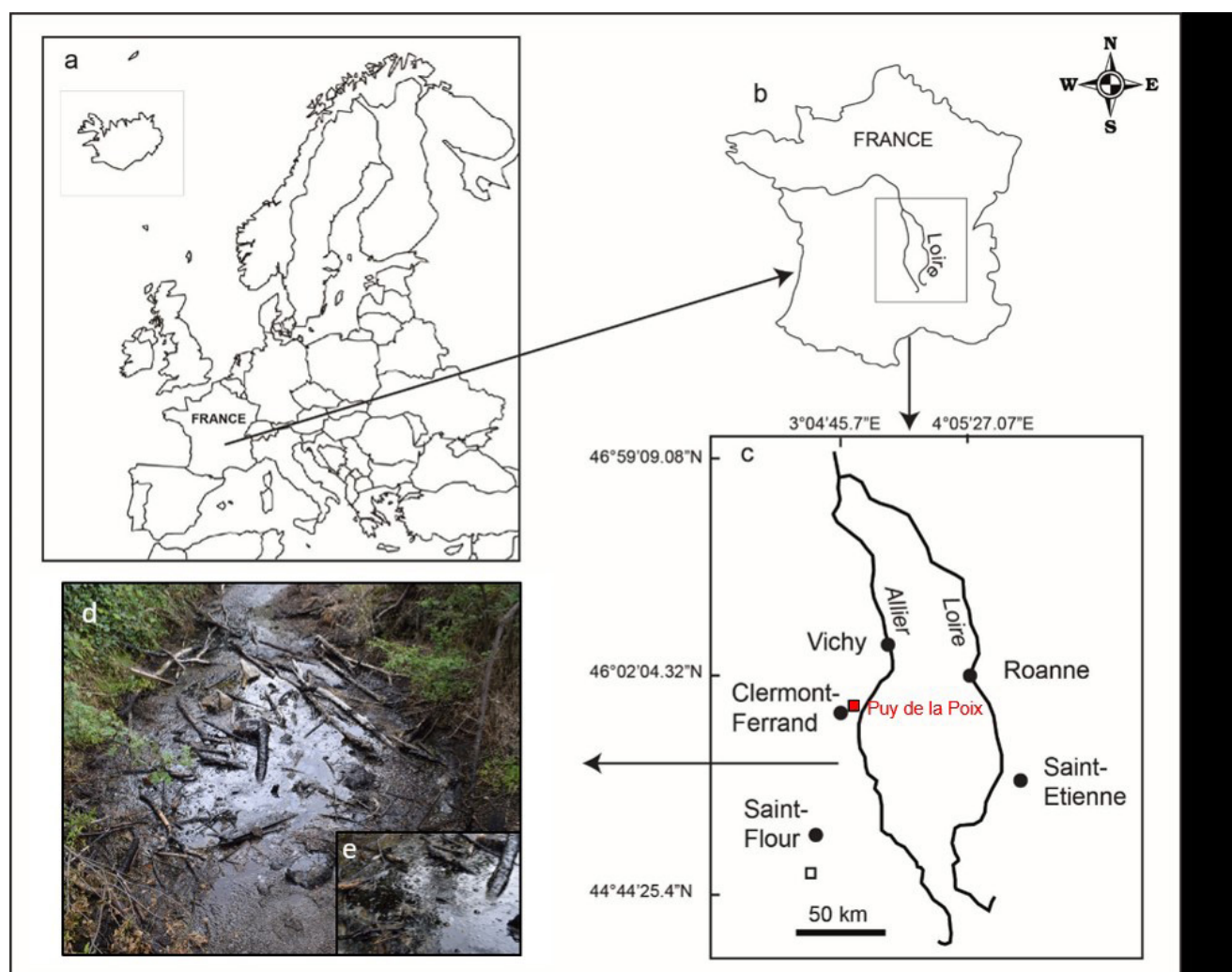


Figure 1 – Map of the studied mineral spring. a: map of Europe; b and c: position location of the Poix spring; d: photo of the Poix spring and the bitumen.



## Materials and methods

### Study site

The Poix spring is situated in the French Massif central and more particularly in the plain of Limagne near Clermont-Ferrand (Fig. 1) (E Lambert 93: 711392.04; N Lambert 93: 6520283.61; altitude: 337m). In the Limagne plain, the Oligocene sediments are naturally rich in hydrocarbons. Indeed, the former lake of Limagne was very rich in life, and the conditions of sedimentation preserved a large part of the organic matter. This organic matter has undergone diagenesis and has partly become bitumen. These hydrocarbons (less dense than the water and the surrounding rocks) rise through the rocks of the Limagne, by following the volcanic vents of the peperitic mini-volcano situated in the area (Thomas 2008). Moreover, this bitumen is associated with salt water, methane and traces of hydrogen sulphide (Gonnard 1906). Lecoq (1864) underlined that out of 82 g of material there is 70 g of sodium chloride, 7 g of de sodium sulphate, 0.1 g of bitumen and organic matter, a small amount of other salts, carbonate of lime, carbonate of magnesia, etc. and finally only traces of sulfur and silica. The place where the spring emerges is not shaded and takes full advantage of the sunshine, which is favourable for the development of diatoms.

### Physical and chemical analyses and diatom sampling

The sampling survey was carried out on the 16<sup>th</sup> March 2017, on the 22<sup>nd</sup> October 2019 and on the 8<sup>th</sup> June 2020 (table 1) in order to make an inventory of the diatom flora of the site. The geographical position of the sampling site was georeferenced using a DGPS Trimble Geo7x, in Lambert 93. *In situ*, conductivity ( $\mu\text{S}\cdot\text{cm}^{-1}$ ), pH (pH units), water temperature ( $^{\circ}\text{C}$ ) and the dissolved oxygen (%) were measured using respectively a multi-parameter WTW FC probe 340i and an oximeter Ysi ProODO. A water sample was collected for further chemical analysis in the laboratory and was analysed using the high-pressure ion chromatography technique. First, these samples were filtered using Whatmann GF/C filters. For the cation analysis, a Thermo Scientific Dionex ICS1100 system was used, whereas for the anions, a Thermo Scientific Dionex Aquion system was used. The concentrations ( $\text{mg L}^{-1}$ ) in lithium ( $\text{Li}^{+}$ ), sodium ( $\text{Na}^{+}$ ), ammonium ( $\text{NH}_4^{+}$ ), potassium ( $\text{K}^{+}$ ), magnesium ( $\text{Mg}^{2+}$ ), calcium ( $\text{Ca}^{2+}$ ), fluorine ( $\text{F}^{-}$ ), chloride ( $\text{Cl}^{-}$ ), nitrite ( $\text{NO}_2^{-}$ ), nitrate ( $\text{NO}_3^{-}$ ), phosphate ( $\text{PO}_4^{3-}$ ) and sulphate ( $\text{SO}_4^{2-}$ ) were measured in the laboratory. In addition, carbonate concentration ( $\text{HCO}_3^{-}$ ) was measured using a Hach Carbonate AL-DT kit (Hach, Loveland, CO, USA).

Diatom sampling was possible at certain locations where the bitumen was submerged at the emergence of the spring. The sampling area was equivalent to a 30x30 cm area. Diatoms were collected by scraping the bituminous deposit at the bottom of the spring using a toothbrush and a pipette (Fig. 1d). Then, vial was named and dated.

### Slide preparation, microscopy and analysis

For each sample collected at the Poix spring, a small fraction of the sample (2 ml) was prepared for light microscopy (LM) observation following the method described in Prygiel & Coste

(2000). Samples were cleaned using hydrogen peroxide ( $\text{H}_2\text{O}_2$ , 35%) and hydrochloric acid (HCl, 37%), and rinsed several times with distilled water. A drop of the diluted cleaned material was dried on coverslips and mounted in Naphrax. LM observations and morphometric measurements were done using a Leica® DM2700M microscope with a 100x oil immersion objective using a differential interference contrast. For each slide, 400 valves were counted. Once the counting was complete, the relative abundances of diatom species were calculated.

Diatom species were identified according to Krammer & Lange-Bertalot (1997a, 1997b, 2000a, 2000b, 2004), Krammer (2000, 2002, 2003), Lange-Bertalot (2001), Werum & Lange-Bertalot (2004), Żelazna-Wieczorek (2011), Wojtal (2013), Levkov et al. (2016), Lange-Bertalot et al. (2017), associated to further articles (Spaulding & Stoermer 1997; Spaulding et al. 1999; Beauger et al. 2015, 2016, 2017, 2019; Wetzel et al. 2015).

For the scanning electron microscopy (SEM), parts of the oxidized suspensions were filtered with additional deionized water through a 3 or 0.2  $\mu\text{m}$  Isopore polycarbonate membrane filter (Merck Millipore). Filters were mounted on stubs using adhesive carbon tabs and coated with carbon (Quorum Q150 TES). An ultrahigh-resolution analytical field emission (FE) scanning electron microscope, Hitachi SU-70 for the sample taken in 2017 and Hitachi Regulus 8230 (Hitachi High-Technologies Corporation, Japan) for the two other samples, was operated at 2 or 5 kV and 10 mm distance for image analysis. SEM images were taken using the lower (SE-L) and upper (SE-U) detector signal.

Shannon index and evenness were calculated using “vegan” package version 2.6-2 (Oksanen et al. 2022) in R version 4.1.2. Ecological traits were calculated according to Van Dam et al. (1994) using OMNIDIA 6.0.8 software (Lecointe et al. 1993) and Lange-Bertalot et al. (2017).

## Results

### Physical and chemical environment

The physico-chemical data of the Poix spring measured during the three campaigns are presented in table 1. On the whole the conductivity and all ionic concentrations were very high contrary to the dissolved oxygen. In March 2017, the conductivity was the highest with  $123,200 \mu\text{S}\cdot\text{cm}^{-1}$  and at this period, the concentration in sodium and chloride was the double compared to 2019 and 2020 with 44,295.2 and 60,477.2  $\text{mg}\cdot\text{L}^{-1}$ . In 2019 and 2020, the conductivity was 46,260 and 53,300  $\mu\text{S}\cdot\text{cm}^{-1}$  respectively and other ionic concentrations were comparable except magnesium and calcium.

### Diatom communities

On the whole, thirty-one different species were observed at the Poix spring (Table 2) with many species of *Navicula* and *Nitzschia*. The *Navicula* species sampled were *N. cincta* (Ehrenberg) Ralfs in Pritchard, *N. gregaria* Donkin, *N. meulemansii* A.Mertens, Witkowski & Lange-Bertalot (Figs 2a-e, 3b), *N. salinarum* Grunow in Cleve & Grunow (Figs 2f-i, 3a), *N. salinicola* Hustedt (Figs 2j-o, 4a) and *N. veneta* Kützinger. The *Nitzschia* species observed

were *Nitzschia inconspicua* Grunow (Fig. 3c), *Nitzschia* aff. *lacuum* Lange-Bertalot, *Nitzschia pusilla* Grunow (Fig. 3d), *Nitzschia* cf. *epithemioides* Grunow in Cleve & Grunow and *Nitzschia* aff. *liebethruthii* Rabenhorst.

As it is shown in Table 3, biodiversity was the lowest in 2017 and 2019 with 11 species and there were 26 species in 2020. Shannon index was 1.41 in 2017, 0.75 in 2019 and 2.51 in 2020. Evenness varied between 0.39 and 0.77. In March 2017, the dominant species were *Navicula salinarum* (54.4%) (Fig. 3a), *Nitzschia pusilla* (Fig. 3d) (20.5%) and *Navicula veneta* (9.7%). In October 2019, it was *Navicula salinicola* (84.0%) (Fig. 4a). At last, in June 2020, *Navicula veneta* (22.5%), *Navicula cincta* (Fig. 4b) (16.7%), *Nitzschia* cf. *liebethruthii* (11.4%), *Pinnularia jocolata* (Manguin) Krammer (10.7%) (Fig. 4c), *Navicula meulemansii* (6.8%) (Fig. 3a) and *Crenotia thermalis* (Rabenhorst) Wojtal (4.7%). Some individual of *Muelleria terrestris* (J.B.Petersen) S.A.Spaulding & Stoermer were observed (Fig. 5). Some unknown species were also observed such as *Halamphora* sp. and two species of *Pinnularia* undetermined.

Related to the ecological trait “salinity”, the community was dominated by brackish water species such as *Navicula salinarum* in 2017 (Fig. 6). In 2019, marine-brackish taxa dominated such as *Navicula salinicola*. At last, in 2020, it was a mix of halophilous, brackish and marine-brackish water species such as *Navicula veneta*.

Table 1 – Ionic concentrations ( $\text{mg.L}^{-1}$ ), physical and chemical results of the measures done in-situ at the Poix spring.

Sampling date	3/16/2017	10/22/2019	6/8/2020
$\text{Li}^+$	171.3	56.4	59.5
$\text{Na}^+$	44,295.2	13,748.7	10,583.7
$\text{NH}_4^+$	2.9	5.4	17.1
$\text{K}^+$	228.0	146.8	107.8
$\text{Mg}^{2+}$	658.0	548.7	294.0
$\text{Ca}^{2+}$	622.1	1,072.9	504.4
$\text{F}^-$	2.5	1.2	0.2
$\text{Cl}^-$	60,477.2	22,929.4	17,417.5
$\text{NO}_2^-$	0.0	0.0	0.0
$\text{Br}^-$	230.5	73.8	56.4
$\text{NO}_3^-$	1.3	0.1	0.1
$\text{PO}_4^{3-}$	0.1	0.1	1.3
$\text{SO}_4^{2-}$	5,417.8	2,570.0	2,121.3
$\text{HCO}_3^-$	4,290.0	2,320.0	1,360.0
Conductivity ( $\mu\text{S.cm}^{-1}$ )	123.2	46.26	53.3
pH (pH units)	7.2	6.8	7.5
$\text{O}_2$ (%)	6.3	2.8	7.8
Water temperature ( $^\circ\text{C}$ )	10.6	12.9	13.6

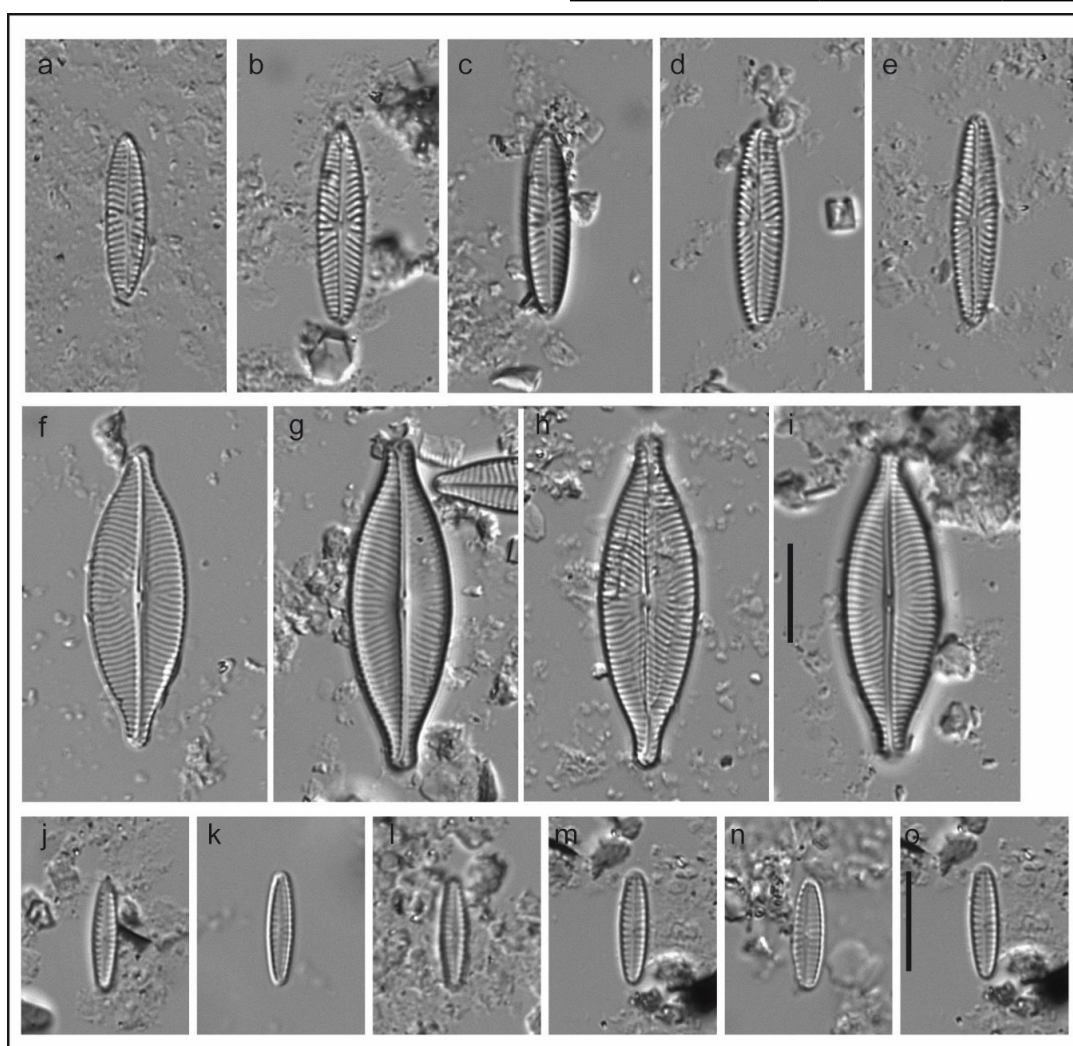


Figure 2 – Some *Navicula* species observed at the Poix spring. *Navicula meulemansii* (a-e), *Navicula salinarum* (f-i), *Navicula salinicola* (j-o). Scale bar = 10  $\mu\text{m}$ .

Table 2 – Relative abundances of the diatom species observed at each sampling period.

Sampling date	3/16/2017	10/22/2019	6/8/2020
<i>Caloneis fontinalis</i> (Grunow in Van Heurck) A.Cleve	0	0	1.1
<i>Crenotia thermalis</i> (Rabenhorst) Wojtal	0	0	4.7
<i>Gomphonema parvulum</i> Kützing	0	0	0.21
<i>Halamphora</i> sp.	0.71	0	0
<i>Hantzschia amphioxys</i> (Ehrenberg) Grunow in Cleve & Grunow	0	2.73	2.5
<i>Hippodonta subtilissima</i> Lange-Bertalot. Metzeltin & Witkowski	0	1.74	0
<i>Humidophila brekkaensis</i> (J.B.Petersen) R.L.Lowe. Kociolek. J.R.Johansen.Van deVijver. Lange-Bertalot & Kopalová	0	0	0.21
<i>Humidophila contenta</i> (Grunow) R.L.Lowe. Kociolek. J.R.Johansen. Van de Vijver. Lange-Bertalot & Kopalová	0.23	0.75	1.31
<i>Luticola</i> aff. <i>frequentissima</i> Levkov. Metzeltin & A.Pavlov	0	0	0.7
<i>Luticola ventricosa</i> (Kützing) D.G.Mann in Round et al.	0	0	0.6
<i>Muelleria terrestris</i> (J.B.Petersen) S.A.Spaulding & Stoermer	0	0	0.65
<i>Navicula cincta</i> (Ehrenberg) Ralfs in Pritchard	0.24	0.25	17.13
<i>Navicula gregaria</i> Donkin	5.25	0	0.21
<i>Navicula meulemansii</i> A.Mertens. Witkowski & Lange-Bertalot	4.53	0	7.01
<i>Navicula salinarum</i> Grunow in Cleve & Grunow	54.42	0	1.94
<i>Navicula salinicola</i> Hustedt	0	84.08	5.7
<i>Navicula veneta</i> Kützing	9.79	5.22	23.07
<i>Nitzschia</i> aff. <i>lacuum</i> Lange-Bertalot	0	0	1.97
<i>Nitzschia</i> aff. <i>liebethuthii</i> Rabenhorst	1.67	0	11.5
<i>Nitzschia</i> cf. <i>epithemioides</i> Grunow in Cleve & Grunow	0	0	1.1
<i>Nitzschia inconspicua</i> Grunow	2.39	0	0
<i>Nitzschia pusilla</i> Grunow	20.53	1.99	2.14
<i>Pinnularia joculara</i> (Manguin) Krammer	0	1.74	10
<i>Pinnularia</i> sp.1	0	0	2.2
<i>Pinnularia</i> sp.2	0	0	1.2
<i>Planothidium frequentissimum</i> (Lange-Bertalot) Lange-Bertalot	0.24	0	0
<i>Planothidium lanceolatum</i> (Brébisson ex Kützing) Lange-Bertalot	0	0.75	0.21
<i>Pseudofallacia monoculata</i> (Hustedt) Y.Liu. Kociolek & Q.Wang in Liu et al.	0	0	1.32
<i>Pseudofallacia</i> sp.	0	0	0.88
<i>Sellaphora bacillum</i> (Ehrenberg) D.G.Mann	0	0.5	0
<i>Stauroneis tackei</i> (Hustedt) Krammer & Lange-Bertalot in Kusber et al.	0	0.25	0.44

Table 3 –  $\alpha$ -diversity of samples showing species richness, Shannon's index diversity and evenness.

	Richness	Shannon	Evenness
2017	11	1.41	0.59
2019	11	0.75	0.31
2020	26	2.51	0.77



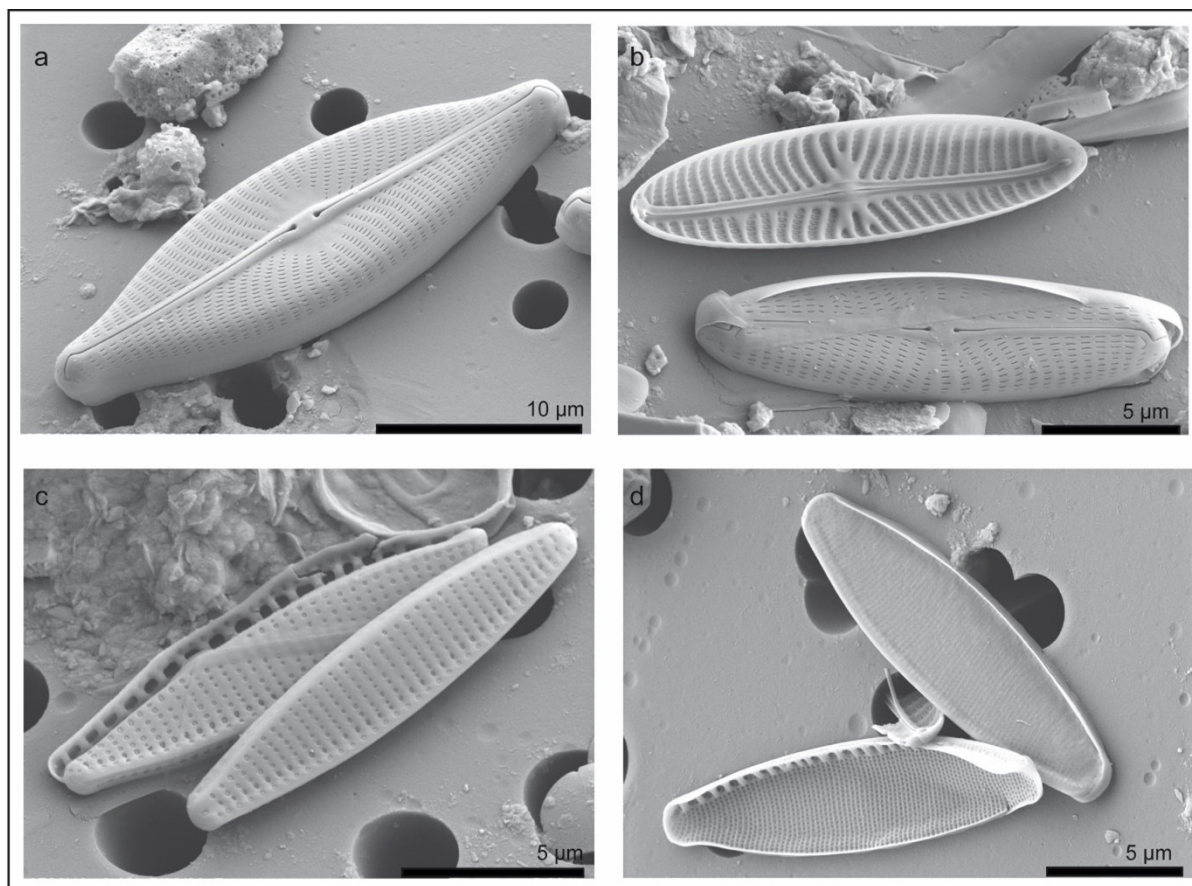


Figure 3 – Photographs at the scanning electron microscopy of some species: *Navicula salinarum* (a), *Navicula meulemansii* (b), *Nitzschia inconspicua* (c), *Nitzschia pusilla* (d).

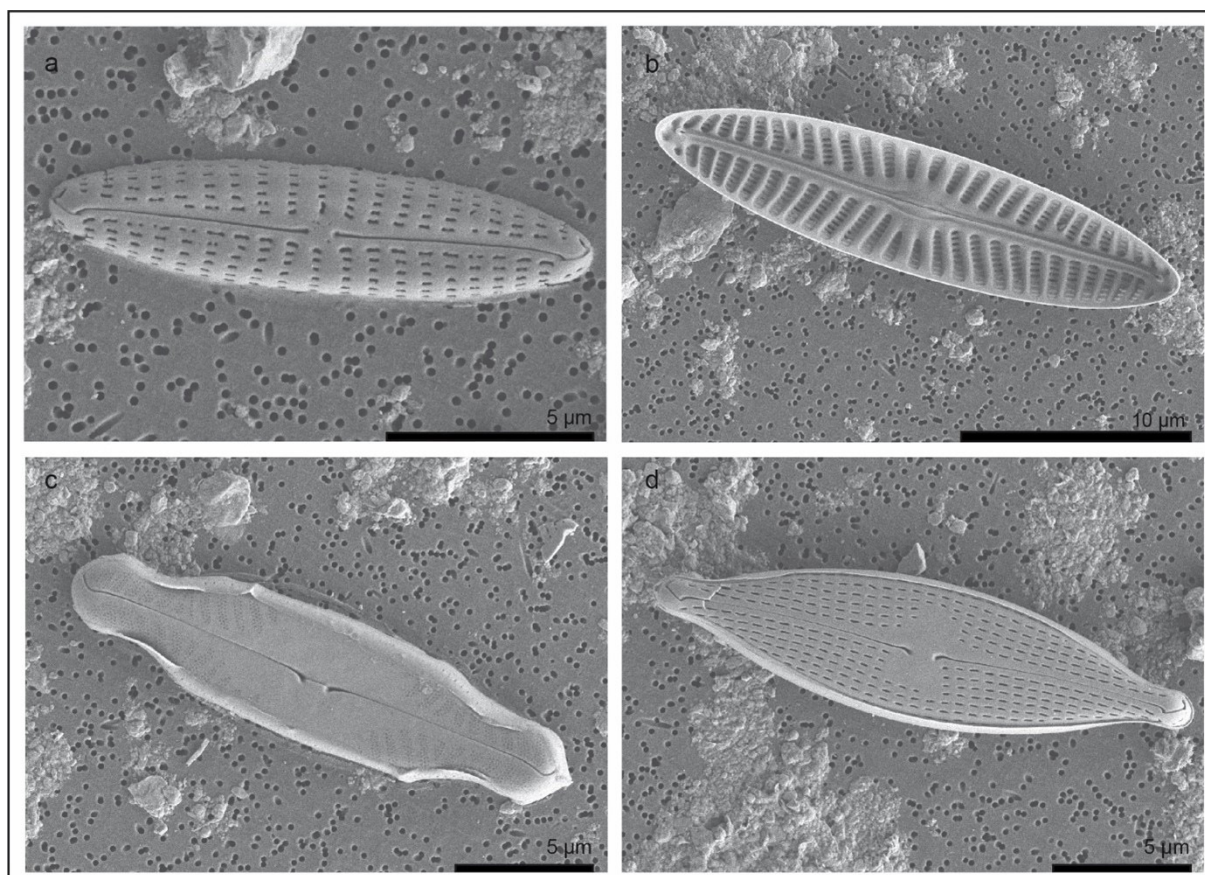


Figure 4 – Photographs at the scanning electron microscopy of some species: *Navicula salinicola* (a), *Navicula cincta* (b), *Pinnularia jocolata* (c), *Navicula gregaria* (d).



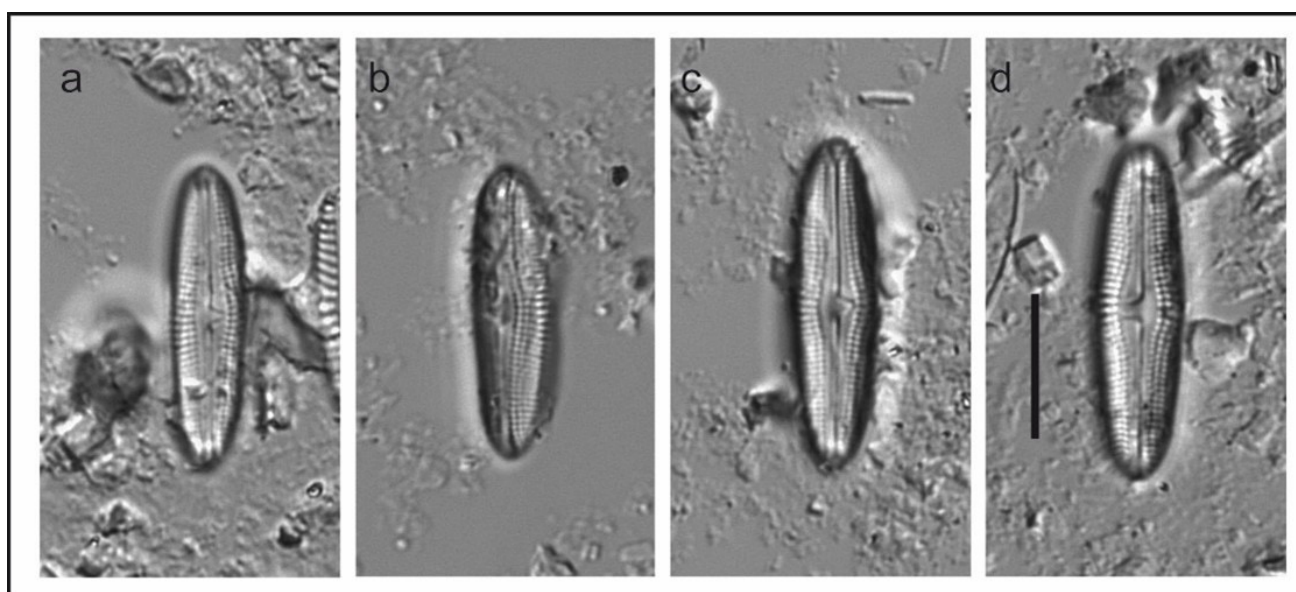


Figure 5 (a-d) – *Muelleria terrestris* observed at the Poix spring. Scale bar = 10  $\mu\text{m}$ .

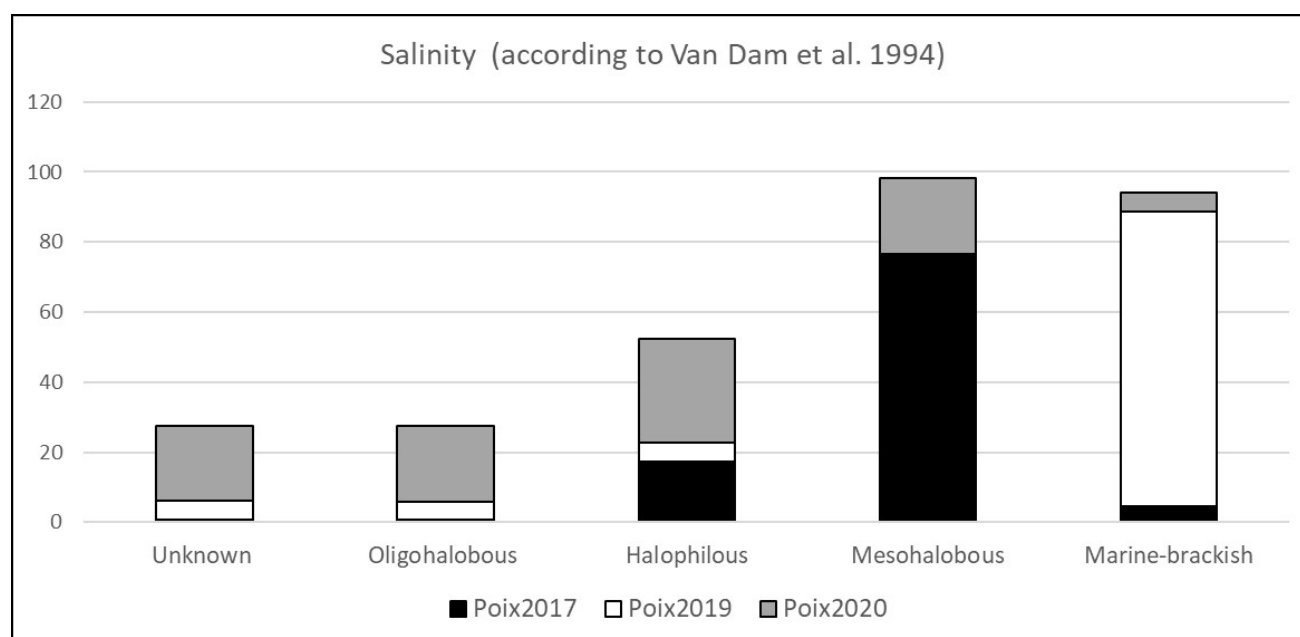


Figure 6 – Ecological traits retained: salinity at the Poix spring for the three samples.

## Discussion

In the Poix spring, characterized by a high mineralization as underlined the high conductivity and salinity, the pH is about 7 and the oxygenation is low. Moreover, the concentration in Lithium and Bromine underlined the deep origin of the water (Michard 1990). The conductivity ranged between 46,260 and 123,200  $\mu\text{S}\cdot\text{cm}^{-1}$  and is consequently higher than ocean that was evaluated as 32,000  $\mu\text{S}\cdot\text{cm}^{-1}$  by some authors (Schnepf et al. 2014; Sabaka et al. 2015). Thus, the abiotic conditions lead to an extreme environment where diatoms were living. In such environment, only species with ecological preferences in adequation with the properties of the water could be encountered. Species richness in 2017 and 2019 was 11 and reached 26 in 2020. The estimated richness varies greatly depending on the

spring. In Lai et al. (2019a), the species richness of mineral springs sampled in Auvergne and Sardinia varied between 2 and 14. Also in Beauger et al. (2020), 17 springs of the Auvergne region were sampled with a richness ranging between 2 and 21 (mean: 9). Lai et al. (2019c) sampled several Mediterranean karst springs, the richest spring was Sa Vena Manna-Sedini with 26 species detected while the mean of richness of the 8 springs sampled was 8. When Cantonati et al. (2012) collected diatoms in 110 springs of the Alps during summer, species richness ranged between 3 and 36 (mean: 13). These observations suggest that diatoms richness is high during 2020 in the mineral spring of the Poix. Shannon diversity was higher in 2020 and 2017 when evenness was higher also.

Diatom species richness is about twice as high in 2020 as in 2017 and 2019. The first hypothesis made to explain these

observations is that it is a seasonal effect, as species richness is classically higher in the warmer seasons (Lavoie et al. 2003). Indeed, in 2020 diatoms were sampled in spring whereas in 2017 and 2019 they were sampled in winter and autumn respectively. The second hypothesis is that the stress conditions in the spring have decreased because the conductivity in the spring has decreased from 123,200 to 53,300  $\mu\text{S}\cdot\text{cm}^{-1}$ . However, in 2019, a decrease in conductivity did not lead to an increase in species richness.

The community was mainly dominated by different species of *Navicula* such as *N. cincta*, *N. gregaria*, *N. meulemansii*, *N. salinarum*, *N. salinicola* and *N. veneta* and also by different species of *Nitzschia* such as *N. inconspicua*, *N. aff. lacuum*, *N. pusilla*, *Nitzschia cf. epithemioides* and *Nitzschia aff. liebethuthii*. Moreover, *Crenotia thermalis* in 2020 and *Pinnularia jocolata* at the three campaigns, were also well represented in the diatom community.

The physical and chemical conditions encountered by diatoms in this spring induced the presence of several halophilous to brackish species such as *Crenotia thermalis* and the cosmopolitan *Navicula salinarum*, *N. salinicola* and *N. veneta* (Van Dam et al. 1994; Lange-Bertalot 2001). Indeed, the synthetic ecological spectra highlighted the dominance of species that prefer high concentrations of dissolved salts. *Crenotia thermalis* occurred in electrolyte-rich inland habitats, particularly thermal and mineral springs (Wojtal 2013; Lange-Bertalot et al. 2017). *Navicula salinarum* was abundant on the coasts, in inland brackish waters, including inland brackish lakes where salts are supplied mainly by rocks and soil weathering (athalassic ecosystems) such as in Slovenia, where it occurred in the only brackish lake (Krivograd Klemenčič et al. 2006). Even if this taxon is not classically encountered in springs, it was observed in some salt springs in Poland (Lange-Bertalot 2001; Wojtal 2013; Źelazna-Wieczorek et al. 2015). In athalassic habitats in Poland, *N. salinarum* was observed associated with *N. veneta* and *N. meulemansii* (Źelazna-Wieczorek et al. 2015). *Navicula veneta* is common in electrolyte rich to brackish waters (Lange-Bertalot 2001). In Poland, it was observed in the sulphide spring of slightly alkaline water of high specific conductivity but very low dissolved oxygen (Wojtal 2013). Moreover, *N. meulemansii* has a wide geographical distribution in coastal brackish waters (The Netherlands and Germany), European estuaries (Portugal), and the San Francisco Bay (U.S.A.) (Mertens et al. 2014). Indeed, it has been identified in samples collected from intertidal sediments from the Jadebusen near Dangast (Witkowski et al. 2004), the Tagus estuary (Ribeiro 2010), the San Francisco Bay area, and from inland brackish waters in the Netherlands (Mertens et al. 2014).

Among the other dominant species, *N. salinicola* was collected along the coast in brackish waters, in river estuaries, in inland salt springs and in freshwaters with a high electrolyte content (Targett et al. 1983; Lange-Bertalot 2001; Guiry & Guiry 2022). *Navicula gregaria* was also observed at the Poix spring (Fig. 4d). It is a species that was found on coasts and inland salt springs of different countries (Lange-Bertalot 2001; Owen et al. 2008; Źelazna-Wieczorek et al. 2015; Leira et al. 2017; Angel et al. 2018; Lai et al. 2019a, 2019b).

*Navicula cincta* was also present in the samples. It is considered

as oligohalobous in Van Dam et al. (1994) whereas this species lives in electrolyte rich to brackish waters (Lange-Bertalot 2001). *Navicula cincta* was observed in athalassic habitats in Poland and also in mineral springs of a very high specific conductivity, in thermo-mineral springs in Sardinia and in Slovenia (Krivograd Klemenčič et al. 2006; Wojtal 2013; Źelazna-Wieczorek et al. 2015; Lai et al. 2019a, 2019b).

*Nitzschia pusilla* was observed in brackish waters (Krammer & Lange-Bertalot 1997b), saline ponds (Trobajo et al. 2011), a hypersaline coastal lagoon (Sylvestre et al. 2001), and a saline lake (Taukulis & John 2006).

At last, some taxa living in a wide amplitude of ecological conditions were present at the Poix spring, such as *Planothidium frequentissimum* (Lange-Bertalot) Lange-Bertalot, *P. lanceolatum* (Brébisson ex Kützing) Lange-Bertalot and *Nitzschia inconspicua* (Lange-Bertalot et al. 2017) (Fig. 3c) that is also known to extend into brackish waters (Kelly et al. 2015). *Hantzschia amphioxys* (Ehrenberg) Grunow in Cleve & Grunow that lives in prolongedly dry and only temporarily wetted (aerial) biotopes, such as soils and rock crevices, underlined that the level of water was low at this site (Lange-Bertalot et al. 2017). *Muelleria terrestris* was observed in soils (Petersen 1915; Spaulding et al. 1999; Foets et al. 2021), and on wet mosses in an intermittent spring of a village in Macedonia (Levkov et al. 2019). *Muelleria terrestris* was also present in a petrifying *Cratoneurion* spring in Belgium (Denys & Oosterlynck 2015). *Pinnularia jocolata* was described in freshwater (Bourrelly & Manguin 1952; Krammer 2000) and was also observed in mineral springs (Quintela et al. 2013; Lai et al. 2019b).

Eight species *Humidophila contenta* (Grunow) R.L.Lowe, Kocielek, J.R.Johansen, Van de Vijver, Lange-Bertalot & Kopalová, *Navicula cincta*, *N. gregaria*, *N. meulemansii*, *N. salinarum*, *N. veneta*, *Nitzschia aff. liebethuthii*, and *Pinnularia jocolata* were present during the three periods of sampling. These species seem to have great capacity to maintain in extreme environments with high fluctuations of conductivity and ion concentrations suggesting the presence of high salinity tolerance mechanisms named osmoregulation. These mechanisms allow cells to compensate osmotic and ionic stress.

Furthermore, the presence of the eight species and the majority of brackish species in the Poix spring as an isolated environment is remarkable and allows the emergence of a question on the origin of these species. This question was already addressed in the literature. Since 1940s, Remane proposed the idea that brackish species are freshwater or marine species adapted to brackish conditions or descendants of these two categories of species (Remane 1940). He also suggested that there are identifiable brackish assemblages (Cognetti & Maltagliati 2000; Potapova 2011). In contrast, Barnes (1989) suggested the inexistence of specific brackish assemblages but the presence of some populations of thalassogenous species in brackish environments (Cognetti & Maltagliati 2000). Each argument has its value depending on the environment studied. To go further, Round & Sims (1981) support the marine origin of diatoms and the colonisation of freshwaters by some lineages because of the high number of fossils and genera of marine diatoms found

in the early Pliocene (Sims et al. 2006). The reconstruction of phylogeny of Thalassiosirales by Alverson et al. (2007) revealed more complicated schema and no unidirectional colonization. Indeed, in their study three lineages of Thalassiosirales seems to have colonized freshwater habitats independently. Another interesting result of this study is that there were at least three recolonizations of marine habitats by these lineages that led to speciation events resulting in the presence of these lineages in marine and freshwater habitats. The genus *Navicula* sensu stricto and *Nitzschia* found in the Poix spring are among the species of raphid diatoms that are found in marine and freshwater environments which agrees with Barnes propositions (Barnes 1989).

Thus, to explain the ability of some diatom taxa to live in high salinity concentration and fluctuation environments, Potapova (2011) made the hypothesis of the innate ability of diatoms to adapt to changing salinity or high salinity and the better conservation of this ability by some lineages. Indeed, ocean salinity was not modelled as stable (Hay et al. 2006), a great decrease of salinity was estimated during the Mesozoic because of the younger basin after the Pangée breaking. But it seems to have several other decreasing salinity events in the past that suggests that diatoms and other micro-organisms were confronted to several fluctuations of salinity in their evolution.

## Conclusion

The studied of the Poix spring is known to be bitumen and associated with salt water, methane and traces of hydrogen sulphide. This environment with a conductivity higher than the ocean could be considered as extreme for diatom species. Moreover, this spring, as others, could be also considered as isolated aquatic ecosystem in the landscape whose conditions have certainly changed very little over the past millennia. The Poix spring showed a relatively high species richness with 31 taxa observed during the three sampling campaigns. The diatom community was mainly dominated by different *Navicula* and *Nitzschia* species. The species are mainly halophilous to brackish such as *Crenotia thermalis*, *Navicula meulemansii*, *N. salinarum*, *N. salinicola* and *N. veneta* observed for some of them in coastal brackish environments or estuaries. Because of their living environment, these species maintain high ability of adaptation of high salinity concentrations and fluctuations which could be the result of well conserved ability of ionic and osmotic stress resistance. This inland brackish environment is a geological and also ecological particularity of the French Massif central. For these arguments, this type of environment needs to be preserved.

## Acknowledgements

This study was funded by a grant from the Centre National de la Recherche Scientifique (CNRS, NEEDS, project Acronym: DISCOVER). Funding for this research was also partly provided by Clermont Auvergne Métropole and in the framework of the DIATOMS project (LIST - Luxembourg Institute of Science and Technology). We thank the Conservatoire des espaces naturels d'Auvergne, which manages the spring.

## References

- Alverson A.J., Jansen R.K. & Theriot E.C., 2007. Bridging the Rubicon: Phylogenetic analysis reveals repeated colonizations of marine and fresh waters by thalassiosiroid diatoms. *Molecular Phylogenetics and Evolution*, **45**(1): 193-210. <https://doi.org/10.1016/j.ympev.2007.03.024>
- Angel A., Vila I., Díaz C., Molina X. & Sepúlveda P., 2018. Geothermal diatoms: seasonal variability in the El Tatio geothermal field (Altiplano, Chile). *Advances in Microbiology*, **8**(3): 211-234. <https://doi.org/10.4236/aim.2018.83015>
- Angeli N., Cantonati M., Spitale D. & Lange-Bertalot H., 2010. A comparison between diatom assemblages in two groups of carbonate, low-altitude springs with different levels of anthropogenic disturbances. *Fottea*, **10**(1): 115-128. <https://doi.org/10.5507/fot.2010.006>
- Barnes R.S.K., 1989. What, if anything, is a brackish-water fauna? *Earth and Environmental Science Transactions of the Royal Society of Edinburgh* **80**(3-4): Environments and Physiology of Fossil Organisms: 235-240. <https://doi.org/10.1017/S0263593300028674>
- Beauger A., Voldoire O., Mertens A., Le Cohu R. & Van de Vijver B., 2015. Two new *Navicula* species (Bacillariophyceae) from Western Europe. *Phytotaxa*, **230**(2): 172-182. <https://doi.org/10.11646/phytotaxa.230.2.4>
- Beauger A., Wetzel C.E., Voldoire O., Garreau A. & Ector L., 2016. *Sellaphora labernardierei* (Sellaphoraceae, Bacillariophyta), a new epilithic species from French spring and four new combinations within the genus *Sellaphora*. *Phytotaxa*, **260**(3): 235-246. <https://doi.org/10.11646/phytotaxa.260.3.3>
- Beauger A., Wetzel C.E., Voldoire O., Garreau A. & Ector L., 2017. Morphology and ecology of *Craticula lecohui* sp. nov. (Bacillariophyceae) from hydrothermal springs (Puy-de-Dôme, Massif Central, France) and comparison with similar *Craticula* species. *Nova Hedwigia, Beiheft*, **146**: 7-22. <https://doi.org/10.1127/1438-9134/2017/007>
- Beauger A., Wetzel C.E., Voldoire O. & Ector L., 2019. *Pseudostaurosira bardii* (Fragilariaceae, Bacillariophyta), a new species from a saline hydrothermal spring of the Massif Central (France). *Botany Letters*, **166**(1): 3-13. <https://doi.org/10.1080/23818107.2018.1460278>
- Beauger A., Voldoire O., Carlos W., Allain E., Millan F., Breton V., Kolovi S. & Ector L., 2020. Biodiversity and ecology of diatoms in mineral springs of the area of Sainte Marguerite (Saint-Maurice-ès-Allier, Massif central, France). *BIOM*, **1**(1): 21-34. <https://dx.doi.org/10.18145/biom.v1i1.250>
- Bourrelly P. & Manguin E., 1952. *Algues d'eau douce de la Guadeloupe et dépendances*. Centre National de la Recherche Scientifique, Société d'Édition d'Enseignement Supérieur, Paris, 281: 1952 p.
- Cantonati M., Angeli N., Bertuzzi E., Spitale D. & Lange-Bertalot H., 2012. Diatoms in springs of the Alps: spring types, environmental determinants, and substratum. *Freshwater science*, **31**(2): 499-524. <https://doi.org/10.1899/11-065.1>
- Cognetti G. & Maltagliati F., 2000. Biodiversity and adaptive mechanisms in brackish water fauna. *Marine Pollution Bulletin*, **40**(1): 7-14. [https://doi.org/10.1016/S0025-326X\(99\)00173-3](https://doi.org/10.1016/S0025-326X(99)00173-3)
- Denys L. & Oosterlynck P., 2015. Diatom assemblages of non-living substrates in petrifying *Cratoneurion* springs from lower



- Belgium. *Fottea*, **15**(2): 123-138. <http://dx.doi.org/10.5507/fot.2015.014>
- Foets J., Stanek-Tarkowska J., Teuling A.J., Van de Vijver B., Wetzel C.E. & Pfister L., 2021. Autecology of terrestrial diatoms under anthropic disturbance and across climate zones. *Ecological Indicators*, **122**: 107-248. <https://doi.org/10.1016/j.ecolind.2020.107248>
- Gonnard F., 1906. Sur le quartz du calcaire bitumineux de la Limagne. *Bulletin de la Société française de Minéralogie*, **29**(7): 362-365. <https://doi.org/10.3406/bulmi.1906.2792>
- Guiry M.D. & Guiry G.M., 2022. AlgaeBase. World-wide electronic publication, National University of Ireland, Galway. <http://www.algaebase.org>; searched on 2 February 2022.
- Hay W.W., Migdisov A., Balukhovskiy A.N., Wold C.N., Flögel S. & Söding E., 2006. Evaporites and the salinity of the ocean during the Phanerozoic: Implications for climate, ocean circulation and life. *Palaeogeography, Palaeoclimatology, Palaeoecology*, **240**(1-2): 3-46. <https://doi.org/10.1016/j.palaeo.2006.03.044>
- Kelly M.G., Trobajo R., Rovira L. & Mann D.G., 2015. Characterizing the niches of two very similar *Nitzschia* species and implications for ecological assessment. *Diatom Research*, **30**(1): 27-33. <http://doi.org/10.1080/0269249X.2014.951398>
- Krammer K., 2000. *The genus Pinnularia. Diatoms of Europe* 1, A.R.G. Gantner Verlag Kommanditgesellschaft, Königstein: 1-703.
- Krammer K., 2002. *Cymbella. Diatoms of Europe* 3, A.R.G. Gantner Verlag Kommanditgesellschaft, Königstein: 1-584.
- Krammer K., 2003. *Cymbopyleura, Delicata, Navicymbula, Gomphocymbellopsis, Afrocybella. Diatoms of Europe* 4, A.R.G. Gantner Verlag Kommanditgesellschaft, Königstein: 1-530.
- Krammer K. & Lange-Bertalot H., 1997a. Bacillariophyceae. 1. Teil: Naviculaceae. In: Ettl H., Gerloff J., Heynig H. & Mollenhauer D. (eds.), *Süßwasserflora von Mitteleuropa*, **2**(1), Gustav Fischer Verlag, Stuttgart, 2th ed., 876 p.
- Krammer K. & Lange-Bertalot H., 1997b. Bacillariophyceae. 2. Teil: Bacillariaceae, Epithemiaceae, Surirellaceae. In: Ettl H., Gerloff J., Heynig H. & Mollenhauer D. (eds.), *Süßwasserflora von Mitteleuropa*, **2**(2), Gustav Fischer Verlag, Stuttgart, 2th ed., 611 p.
- Krammer K. & Lange-Bertalot H., 2000a. Bacillariophyceae. 3. Teil: Centrales, Fragilariaceae, Eunotiae. In: Ettl H., Gerloff J., Heynig H. & Mollenhauer D. (eds.), *Süßwasserflora von Mitteleuropa*, **2**(3), Gustav Fischer Verlag, Stuttgart, 2th ed., 576 p.
- Krammer K. & Lange-Bertalot H., 2000b. Bacillariophyceae. 5. Teil: English and French translation of the keys. In: Büdel B., Gärtner G., Krienitz L. & Lokhorst G.M. (eds.), *Süßwasserflora von Mitteleuropa*, **2**(5), Spektrum Akademischer Verlag, Heidelberg, 599 p.
- Krammer K. & Lange-Bertalot H., 2004. Bacillariophyceae. 4. Teil: Achnanthaceae, kritische Ergänzungen zu *Navicula* (Lineolatae) und *Gomphonema*. In: Ettl H., Gärtner G., Heynig H. & Mollenhauer D. (eds.), *Süßwasserflora von Mitteleuropa*, **2**(4), Spektrum Akademischer Verlag, Heidelberg, 2th ed., 468 p.
- Krivograd Klemenčič A., Vrhovšek D. & Smolar-Žvanut N., 2006. Contribution on the algal flora of Slovenia, with emphasis on newly recorded algal taxa. *Acta Musei Nationalis Pragae, Series B, Historia Naturalis* **62**, (1-2): 71-80.
- Lai G.G., Padedda B.M., Wetzel C.E., Lugliè A., Sechi N. & Ector L., 2016. Epilithic diatom assemblages and environmental quality of the Su Gologone karst spring (central-eastern Sardinia, Italy). *Acta Botanica Croatica*, **75**(1): 129-143. <https://doi.org/10.1515/botcro-2016-0008>
- Lai G.G., Beauger A., Wetzel C.E., Padedda B.M., Voldoire O., Lugliè A., Allain E. & Ector L., 2019a. Diversity, ecology and distribution of benthic diatoms in thermo-mineral springs in Auvergne (France) and Sardinia (Italy). *PeerJ*, **7**: e7238. <https://doi.org/10.7717/peerj.7238>
- Lai G.G., Padedda B.M., Wetzel C.E., Cantonati M., Sechi N., Lugliè A. & Ector L., 2019b. Diatom assemblages from different substrates of the Casteldoria thermo-mineral spring (Northern Sardinia, Italy). *Botany Letters*, **166**(1): 14-31. <https://doi.org/10.1080/23818107.2018.1466726>
- Lai G.G., Padedda B.M., Ector L., Wetzel C.E., Lugliè A., Cantonati M., 2019c. Mediterranean karst springs: Diatom biodiversity hotspots under the pressure of hydrological fluctuations and nutrient enrichment, *Plant Biosystems*, **154**(5): 673-684. <https://doi.org/10.1080/11263504.2019.1674402>
- Lange-Bertalot H., 2001. *Navicula sensu stricto*, 10 genera separated from *Navicula sensu lato* *Frustulia. Diatoms of Europe* 2, A.R.G. Gantner Verlag Kommanditgesellschaft, Königstein: 1-526.
- Lange-Bertalot H., Hofmann G., Werum M. & Cantonati M., 2017. *Freshwater benthic diatoms of Central Europe: over 800 common species used in ecological assessments. English edition with updated taxonomy and added species.* Koeltz Botanical Books, Schmitt-Oberreifenberg, 942 p.
- Lavoie I., Vincent W.F., Pienitz R. & Painchaud J., 2003. Dynamique temporelle des algues périphytiques dans une rivière influencée par les activités agricoles. *Revue des sciences de l'eau*, **16**: 55-77. <https://doi.org/10.7202/705498ar>
- Lecointe C., Coste M. & Prygiel J., 1993. "Omnidia": software for taxonomy, calculation of diatom indices and inventories management. *Hydrobiologia*, **269**: 509-513. <https://doi.org/10.1007/BF00028048>
- Lecoq H., 1864. *Les eaux minérales du massif central de la France, considérées dans leurs rapports avec la chimie et la géologie.* J. Rothschild, Paris, 256 p.
- Legrand R., 2019. *Note synthétique de gestion. Puy de la Poix (Clermont-Ferrand). Bilan des enjeux, objectifs et actions.* Conservatoire d'espaces naturels. CEN Auvergne, Riom, 16 p.
- Leira M., Meijide-Failde R. & Torres E., 2017. Diatom communities in thermo-mineral springs of Galicia (NW Spain). *Diatom Research*, **32**(1): 29-42. <https://doi.org/10.1080/0269249X.2017.1286266>
- Levkov Z., Mitić-Kopanja D. & Reichardt E., 2016. The diatom genus *Gomphonema* from the Republic of Macedonia. *Diatom of Europe* **8**: 1-552.
- Levkov Z., Vidaković D., Cvetkoska A., Mitić-Kopanja D., Krstić S., Van de Vijver B. & Hamilton P.B., 2019. Observations of the genus *Muelleria* (Bacillariophyceae) from the Republic of North Macedonia. *Plant Ecology and Evolution*, **152**(2): 293-312. <https://doi.org/10.5091/plecevo.2019.1611>
- Mertens A., Witkowski A., Lange-Bertalot H., Ribeiro L. & Rhiel E., 2014. *Navicula meulemansii* sp. nov., (Bacillariophyceae) from brackish waters in Europe and the U.S.A. *Nova Hedwigia*, **98**(1-2): 201-212. <https://doi.org/10.1127/0029-5035/2013/0152>
- Michard G., 1990. Behaviour of major elements and some trace elements (Li, Rb, Cs, Sr, Fe, Mn, W, F) in deep hot waters from granitic areas. *Chemical Geology*, **89**(1-2): 117-134. [https://doi.org/10.1016/0009-2541\(90\)90062-C](https://doi.org/10.1016/0009-2541(90)90062-C)

- Nivet V., 1846. Suite du dictionnaire des eaux minérales du département du Puy-de-Dôme. *Annales scientifiques, littéraires et industrielles de l'Auvergne*, **19**: 17-208.
- Oksanen J., Simpson G.L., Blanchet F.G., Kindt R., Legendre P., Minchin P.R., O'Hara R.B., Solymos P., Stevens M. H. H., Szoecs E., Wagner H., Barbour M., Bedward M., Bolker B., Borcard D., Carvalho G., Chirico M., De Caceres M., Durand S., Antoniazzi Evangelista H. B., FitzJohn R., Friendly M., Furneaux B., Hannigan G., O. Hill M., Lahti L., McGlenn D., Ouellette M.-H., Ribeiro Cunha E., Smith T., Stier A., Ter Braak C. J.F. & Weedon J., 2022. *vegan: Community Ecology Package*. R package version 2.6-2. <https://CRAN.R-project.org/package=vegan>
- O'Leary J.W. & Glenn E.P., 1994. Global distribution and potential for halophytes. In: Squires V.R. & Ayoub A.T. (eds.), *Halophytes as resource for livestock and rehabilitation of degraded lands. Tasks for Vegetation Science*, **32**: 7-17. [https://doi.org/10.1007/978-94-011-0818-8\\_2](https://doi.org/10.1007/978-94-011-0818-8_2)
- Owen R.B., Renaut R.W. & Jones B., 2008. Geothermal diatoms: a comparative study of floras in hot spring systems of Iceland, New Zealand, and Kenya. *Hydrobiologia*, **610**: 175-192. doi: <https://doi.org/10.1007/s10750-008-9432-y>
- Petersen J.B., 1915. Studier over danske ærofile alger. *Det Kongelige Danske videnskabernes selskabs skrifter. Naturvidenskabelig og matematisk afdeling*, ser. 7, **12**: 272-379, 4 pls.
- Piernik A., 2012. *Ecological pattern of inland salt marsh vegetation in central Europe*. Wydawnictwo Naukowe Uniwersytetu Mikołaja Kopernika, Toruń, 229 p.
- Poljakoff-Mayber A. & Gale J. (eds.), 1975. Plants in saline environments. *Ecological Studies*, **15**. Springer, Berlin, Heidelberg, 219 p. <https://doi.org/10.1007/978-3-642-80929-3>
- Potapova M., 2011. Patterns of diatom distribution in relation to salinity. In: Seckbach J. & Kociolek P. (eds), *The Diatom World. Cellular Origin, Life in Extreme Habitats and Astrobiology*, **19**: 313-332. Springer, Dordrecht. [https://doi.org/10.1007/978-94-007-1327-7\\_14](https://doi.org/10.1007/978-94-007-1327-7_14)
- Prygiel J. & Coste M., 2000. *Guide méthodologique pour la mise en oeuvre de l'indice Biologique Diatomées NF T 90-354*. Etude Agences de l'Eau-Cemagref Bordeaux, mars 2000. Douai: Agences de l'Eau; 134 p. + clés de détermination (90 planches couleurs) + cdrom bilingue français-anglais (Tax'IBD).
- Quintela A., Almeida S., Terroso D., Ferreira da Silva E., Forjaz V. & Rocha F., 2013. Diatom assemblages of thermal and mineral waters from volcanic environments in São Miguel Island, Azores. *Diatom Research*, **28**(4): 407-417. <https://doi.org/10.1080/0269249X.2013.822833>
- Rakowska B., 1997. Diatom communities in a salt spring at Pełczyska (Central Poland). *Biologia*, **52**(4): 489-493.
- Reimold R.J. & Queen W.H. (eds), 1974. *Ecology of halophytes*. Academic Press, INC, New York-London, 607 p. <https://doi.org/10.1016/B978-0-12-586450-3.X5001-X>
- Remane A., 1940. Einführung in die zoologische Ökologie der Nord- und Ostsee. In: Grimpe G. & Wagler E. (eds.), *Die Tierwelt der Nord- und Ostsee*. vol. 1 (1a). Akademische Verlagsgesellschaft Becker & Erler, Leipzig, 238 p.
- Ribeiro L., 2010. *Intertidal benthic diatoms of the Tagus estuary: Taxonomic composition and spatial-temporal variation*. PhD Thesis, Universidade de Lisboa, 311 p.
- Round F.E. & Sims P.A., 1981. The distribution of diatom genera in marine and freshwater environments and some evolutionary considerations. In: Ross, R. (ed.), *Proceedings of the Sixth Symposium on Recent and Fossil Diatoms. Budapest, September 1-5, 1980. Taxonomy Morphology Ecology Biology*. Otto Koeltz, Koenigstein, 301-320.
- Sabaka T.J., Olsen N., Tyler R.H. & Kuvshinov A., 2015. CM5, a pre-Swarm comprehensive geomagnetic field model derived from over 12 yr of CHAMP, Ørsted, SAC-C and observatory data. *Geophysical Journal International*, **200**(3): 1596-1626. <https://doi.org/10.1093/gji/ggu493>
- Sabater S. & Roca J.R., 1992. Ecological and biogeographical aspects of diatom distribution in Pyrenean springs. *British Phycological Journal*, **27**(2): 203-213. <https://doi.org/10.1080/00071619200650201>
- Schnepf N.R., Manoj C., Kuvshinov A., Toh H. & Maus S., 2014. Tidal signals in ocean-bottom magnetic measurements of the Northwestern Pacific: observation versus prediction. *Geophysical Journal International*, **198**(2): 1096-1110. <https://doi.org/10.1093/gji/ggu190>
- Shevah Y., 2014. 1.4 - Adaptation to Water Scarcity and Regional Cooperation in the Middle East. In: *Comprehensive Water Quality and Purification 1*, Satinder Ahuja, Elsevier, 40-70. <https://doi.org/10.1016/B978-0-12-382182-9.00004-9>.
- Sims P.A., Mann D.G. & Medlin L.K., 2006. Evolution of the diatoms: insights from fossil, biological and molecular data. *Phycologia*, **45**(4): 361-402. <https://doi.org/10.2216/05-22.1>
- Spaulding S.A. & Stoermer E.F., 1997. Taxonomy and distribution of the genus *Muelleria* Frenguelli. *Diatom Research*, **12**(1): 95-113 <https://doi.org/10.1080/0269249X.1997.9705405>
- Spaulding S.A., Kociolek J.P. & Wong D., 1999. A taxonomic and systematic revision of the genus *Muelleria* (Bacillariophyta). *Phycologia*, **38**(4): 314-341. <https://doi.org/10.2216/i0031-8884-38-4-314.1>
- Sylvestre F., Beck-Eichler B., Duleba W. & Debenay J.-P., 2001. Modern benthic diatom distribution in a hypersaline coastal lagoon: the Lagoa de Araruama (R.J.), Brazil. *Hydrobiologia*, **443**: 213-231. <https://doi.org/10.1023/A:1017558914971>
- Targett N.M., Bishop S.S., McConnell O.J. & Yoder J.A., 1983. Antifouling agents against the benthic marine diatom, *Navicula salinicola* Homarine from the gorgonians *Leptogorgia virgulata* and *L. setacea* and analogs. *Journal of Chemical Ecology*, **9**(7): 817-829. <https://doi.org/10.1007/BF00987807>
- Taukulis F.E. & John J., 2006. Diatoms as ecological indicators in lakes and streams of varying salinity from the wheatbelt region of Western Australia. *Journal of the Royal Society of Western Australia*, **89**(1): 17-25.
- Thomas P., 2008. Source et ruisseau d'hydrocarbures, Puy de la Poix, Limagne de Clermont-Ferrand (Puy de Dôme). <https://planet-terre.ens-lyon.fr/image-de-la-semaine/lmg220-2008-01-07.xml>
- Trobajo R., Rovira L., Mann D.G. & Cox E.J., 2011. Effects of salinity on growth and on valve morphology of five estuarine diatoms. *Phycological Research* **59**(2): 83-90. <https://doi.org/10.1111/j.1440-1835.2010.00603.x>
- Truchot P., 1878. *Dictionnaire des eaux minérales du département du Puy-de-Dôme*. Delahaye A., Paris, 320 p.
- Van Dam H., Mertens A. & Sinkeldam J., 1994. A coded checklist and ecological indicator values of freshwater diatoms from The Netherlands. *Netherlands Journal of Aquatic Ecology* **28**: 117-133. <https://doi.org/10.1007/BF02334251>
- Waisel Y., 1972. *The Biology of Halophytes*. Academic Press, New York and London, 395 p. <https://doi.org/10.1016/B978-0-12->

730850-0.X5001-6

- Werum M. & Lange-Bertalot H., 2004. Diatoms in springs from Central Europe and elsewhere under the influence of hydrogeology and anthropogenic impacts. *Iconographia Diatomologica*, **13**: 1-417.
- Wetzel C.E., Ector L., Van de Vijver B., Compère P. & Mann D.G., 2015. Morphology, typification and critical analysis of some ecologically important small naviculoid species (Bacillariophyta). *Fottea*, **15**(2): 203-234. <https://doi.org/10.5507/fot.2015.020>
- Witkowski A., Pilzen M., Kort R., Rhiel E., Wawrzyniak-Wydrowska B., Lange-Bertalot H., Bak M. & Daniszewska G., 2004. Investigations on the seasonal succession of Wadden Sea inhabiting diatoms at Dangast (North Sea, German Bight) over a one year period. *Vie et Milieu*, **54**(2-3): 181-189.
- Wojtal A.Z., 2013. Species composition and distribution of diatom assemblages in spring waters from various geological formations in southern Poland. *Bibliotheca Diatomologica*, **59**: 1-436.
- Żelazna-Wieczorek J., 2011. Diatom flora in springs of Łódź Hills (Central Poland). Biodiversity, taxonomy, and temporal changes of epipsammic diatom assemblages in springs affected by human impact. *Diatom Monographs*, **13**: 1-419.
- Żelazna-Wieczorek J., Olszyński R.M. & Nowicka-Krawczyk P., 2015. Half a century of research on diatoms in athalassic habitats in central Poland. *Oceanological and Hydrobiological Studies*, **44**(1): 51-67. <https://doi.org/10.1515/ohs-2015-0006>