

# Blood mercury concentrations in four sympatric gull species from South Western France: Insights from stable isotopes and biologging

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1	Blood mercury concentrations in four sympatric gull species from
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# 25 ABSTRACT

Mercury (Hg) is a toxic trace element widely distributed in the environment, which 26 particularly accumulates in top predators, including seabirds. Among seabirds, large gulls 27 (*Larus sp*) are generalist feeders, foraging in both terrestrial and marine habitats, making 28 them relevant bioindicators of local coastal Hg contamination. In the present study, we 29 reported blood Hg concentrations in adults and chicks of four different gull species 30 breeding on the French Atlantic coast: the European herring gull (Larus argentatus), the 31 Lesser black-backed gull (L. fuscus), the Great black-backed gull (L. marinus) and the 32 33 Yellow-legged gull (L. michahellis). We also investigated the potential role of foraging ecology in shaping Hg contamination across species, using the unique combination of three 34 dietary tracers (carbon, nitrogen and sulfur stable isotopes) and biologging (GPS tracking). 35 A high concentration of Hg was associated with high trophic position and a marine diet in 36 gulls, which was corroborated by birds' space use strategy during foraging trips. Adults of 37 all four species reached Hg concentrations above reported toxicity thresholds. Specifically, 38 adults of Great black-backed gulls had a high trophic marine specialized diet and 39 significantly higher Hg concentrations than the three other species. Blood Hg was 4 to 7 40 41 times higher in adults than in chicks, although chicks of all species received mainly marine and high trophic position prey, which is expected to be the cause of blood Hg 42 concentrations of toxic concern. By using both stable isotopes and GPS tracking, the 43 44 present study provides compelling insights on the main feeding habits driving Hg contamination in a seabird assemblage feeding in complex coastal environments. 45

- 47 Key words: Contaminant, Trace element, Seabird, Habitat, Movement ecology, Stable
- 48 isotopes

#### 49 INTRODUCTION

Mercury (Hg) is a non-essential trace element released by both natural and 50 anthropogenic processes (Pirrone et al., 2010). Organic Hg (mostly methylmercury; MeHg) 51 is the main bioavailable and toxic form for living organisms (Díez, 2008; Thompson and 52 Furness, 1989). MeHg is incorporated in organisms mainly via ingestion of contaminated 53 food (Eagles-Smith et al., 2018). In birds, accumulation of MeHg has been associated with 54 a wide range of behavioral and physiological detrimental consequences, impacting 55 individual fitness, and ultimately, population dynamics (Tan et al., 2009; Whitney and 56 57 Cristol, 2017). Due to a relatively long biological half-life in tissues (Stickel et al., 1977), Hg tends to bioaccumulate in organisms over time and biomagnify up trophic food webs, 58 leading long-lived top predators to exhibit elevated Hg concentrations (Atwell et al., 1998; 59 Cherel et al., 2018; Evers et al., 2005). In coastal environments, seabirds are therefore 60 considered as excellent sentinels of the local contamination (Burger and Gochfeld, 2004; 61 62 Furness and Camphuysen, 1997; Monteiro and Furness, 1995).

Seabirds show a large interspecific variation in foraging strategies (Ceia and 63 Ramos, 2015), leading to varied degrees of Hg exposure among species (Carravieri et al., 64 65 2014b; Monteiro et al., 1998; Stewart et al., 1997). Large gulls of the Larus genus are omnivorous and opportunistic feeders. Larus spp. are known to exploit both marine and 66 terrestrial habitats, and can scavenge from multiple anthropogenic sources including waste 67 68 dumps, landfill sites and fishing discards, as well as predating upon eggs or chicks (Buckley, 1990; Mudge and Ferns, 1982; Ramos et al., 2009). Assessing Hg concentrations 69 70 in sympatric *Larus* species with differences in foraging ecology thus enables to have an 71 overview of the local environmental contamination, particularly in complex coastal

72 habitats (Binkowski et al., 2020). Additionally, in seabirds, individual variation in foraging behavior appears to be highly prevalent, with individuals differing in their resource use, 73 habitat selection, and their fidelity to foraging sites (Ceia and Ramos, 2015; Phillips et al., 74 2017). Such variation in foraging behavior may be consistent over time, and linked to 75 individuals' characteristics, such as their age or sex (Bolnick et al., 2003). As Hg exposure 76 largely varies with feeding strategies and habitat use, this variability shapes intraspecific 77 differences in Hg contamination (Bustamante et al., 2016; Ceia and Ramos, 2015; Stewart 78 et al., 1997), with consequences for individual fitness. Characterizing foraging ecology at 79 80 the individual level is therefore essential to understand patterns of Hg contamination within and among species. 81

Tracking devices, including Global Positioning System (GPS) loggers, enable fine 82 scale monitoring of an animal's movements, which facilitates a direct characterization of 83 its habitat use (Burger and Shaffer, 2008). Furthermore, recently developed analytical tools 84 - including hidden Markov models (HMMs) - facilitate behavioral classification of GPS 85 tracking data (Langrock et al., 2012), enabling the identification of the precise locations 86 and habitats in which animals are foraging. The analysis of stable isotopes - used as a 87 proxy for the trophic niche (Newsome et al., 2007) – can also represent an effective tool to 88 define the foraging habitats (carbon:  $\delta^{13}$ C and sulfur:  $\delta^{34}$ S) and trophic positions (nitrogen: 89  $\delta^{15}$ N) of seabirds (Hobson et al., 1994; Kelly, 2000; Lott et al., 2003). A marine diet is 90 characterized by higher  $\delta^{13}$ C,  $\delta^{15}$ N and  $\delta^{34}$ S values, as opposed to lower levels indicating 91 terrestrial food intakes (Chisholm et al., 1982; Hobson, 1987; MacAvoy et al., 2000; 92 Schoeninger and DeNiro, 1984). More specifically,  $\delta^{34}S$  is preferentially used to 93 94 differentiate marine vs terrestrial foraging habitats, since it does not present a stepwise 95 enrichment within the food chain, and therefore shows a greater discriminating power than 96  $\delta^{13}$ C and  $\delta^{15}$ N (Hobson, 1999; McCutchan Jr et al., 2003). This characteristic makes  $\delta^{34}$ S 97 particularly interesting to depict seabirds' use of coastal habitats. Therefore, the use of GPS 98 trackers combined with isotopic measurements enables a holistic overview of birds' 99 feeding strategies with respect to multiple available foraging habitats, as well as a high 100 discriminant power (Bracey et al., 2021; Caron-Beaudoin et al., 2013; Ceia et al., 2018; 101 Mendes et al., 2018).

In France, Hg contamination of seabirds has been largely documented in overseas 102 103 departments and territories (e.g. French Guiana, Sebastiano et al., 2016; 2017; Scattered Islands, Kojadinovic et al. 2007a; Réunion Island, Kojadinovic et al. 2007b; 2007c) and 104 French Southern and Antarctic Territories (e.g., Blévin et al., 2013; Carravieri et al., 2014; 105 2016; Goutte et al., 2014a; 2014b; Tartu et al., 2014; 2015a). Although the coasts of 106 metropolitan France host large numbers of breeding seabirds (e.g., 174 000 pairs, Cadiou, 107 2011), data on Hg concentrations in seabird tissues are scarce and limited to chicks of four 108 gull species (Larus argentatus, Larus fuscus, Larus marinus and Larus michahellis) from 109 the Southern Bay of Biscay and the English Channel (Binkowski et al., 2020; Zorrozua et 110 111 al., 2020). In these studies, high blood Hg concentrations were associated with a diet of marine origin and of high trophic position. Yet, our knowledge on Hg concentrations in 112 other potentially contaminated areas remains very limited. 113

In the present study, we investigated Hg concentrations in adults and chicks of four gull species from a South Western France colony: Herring gulls *Larus argentatus argenteus*, Lesser black-backed gulls *L. fuscus graellsii*, Great black-backed gulls *L. marinus* and Yellow-legged gulls *L. michahellis*. We were particularly interested to i)

118 describe Hg concentrations of the species during the breeding period; ii) identify the factors driving inter- and intraspecific Hg variations among adults and chicks - in relation to their 119 different feeding habitats inferred from blood  $\delta^{13}$ C,  $\delta^{15}$ N and  $\delta^{34}$ S stable isotopes, and in 120 relation to their individuals characteristics as age and sex; and iii) explore whether fine-121 scale habitat partitioning using GPS tracking further elucidates the potential variation in 122 adult Hg contamination. Due to the known differences in foraging ecology among the four 123 species, as well as the individual differences within species, we expected a high inter- and 124 intraspecific variation in Hg concentrations. We further predicted that i) individuals feeding 125 at higher trophic positions (indicated by higher  $\delta^{15}$ N values) or foraging in marine habitats 126 (indicated by both high  $\delta^{13}$ C and  $\delta^{34}$ S values, as well as GPS tracking) would exhibit higher 127 Hg concentrations; and that ii) these patterns would persist both between and within 128 species. 129

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# 131 MATERIALS AND METHODS

#### 132 Field methodology

In the breeding seasons of 2016-2019, we sampled gulls breeding in "Lilleau des 133 Niges" National Nature reserve, Ile de Ré, South Western France (46°13'53" N, 1°30'22" 134 W; Figure 1): European herring gulls (EHG, approx. 400 pairs), Lesser black-backed gulls 135 (LBBG, approx. 400 pairs), Great black-backed gulls (GBBG, approx. 200 pairs), and 136 137 Yellow-legged gull (YLG, approx. 100 pairs). Adult birds were sampled in May (n = 140), and chicks in July (n = 107). All individuals were captured and sampled only once over the 138 139 four years. Annual sample sizes for adults and chicks of each species can be found in the 140 Supporting Information (SI) Table S1. Adults incubating a full-size clutch (i.e. 2 to 3 eggs) 141 were caught on their nests, using a trap placed over the eggs. Chicks of approximately 1 month of age were caught by hand, within the colony. From an early age, gull chicks are 142 developed enough to explore their environment and do not stay in their nest, therefore we 143 were unable to assign relatedness among chicks and adults. Skull and tarsus were measured 144 using a caliper ( $\pm 0.1$  mm). Wing length was also measured using a ruler ( $\pm 1$  mm), and 145 birds were weighted using a Pesola spring balance ( $\pm$  5g). Blood samples (2 mL, <1 % of 146 the body mass of LBBG, the smallest species in this study) were taken from the brachial 147 vein of adults and chicks using heparinized syringes. Blood was transferred into a 2 mL 148 Eppendorf<sup>®</sup> tube and immediately stored in a cool box. At the end of each sampling day, 149 whole blood was centrifuged and red blood cells (hereafter "blood") were stored at -20°C 150 until laboratory analyses. Blood samples were then used to measure Hg and stable isotopes 151 and for molecular sexing. 152

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#### 154 Mercury analysis

The analysis of total Hg in lyophilized blood was carried out using an Altec AMA 155 254 spectrophotometer following Bustamante et al. (2006). Each sample (mass: ~0.5 mg 156 157 dry weight, dw) was analysed twice and Hg concentrations were then averaged. For each individual, the relative standard deviation was <10 %. Accuracy was checked at the 158 beginning and the end of each measurement session using a certified reference material 159 160 (2016-2018: DOLT-5 dogfish liver - Hg certified concentration (mean  $\pm$  SD): 0.44  $\pm$  0.18  $\mu$ g g<sup>-1</sup> dw; 2019: TORT-3 lobster hepatopancreas - Hg certified concentration:  $0.29 \pm 0.02$ 161  $\mu$ g g<sup>-1</sup> dw; obtained from NRC, Canada). Recoveries were respectively 0.422  $\pm$  0.005 and 162  $0.318 \pm 0.014 \ \mu g \ g^{-1}$  dw. Blanks were also run before and after each set of samples. The 163

limit of detection of the AMA was 0.1 ng. Hg concentrations are expressed as  $\mu g g^{-1}$  dw. 164 In several bird species, MeHg represents most of the total Hg in blood (Renedo et al., 2018; 165 Rimmer et al., 2005). Chicks accumulate Hg from hatching, being fed by their parents from 166 local food items. For this reason, they are relevant bioindicators of the local environment 167 contamination (Binkowski et al., 2020; Blévin et al., 2013). By contrast to chicks, using 168 adult gulls as bioindicators of their local environment during breeding is still debated as 169 the half-life of Hg in avian tissues is 1~3 months, and contaminants acquired during 170 migration might still be present in blood (Bearhop et al., 2000; Monteiro and Furness, 2001; 171 172 Stickel et al., 1977). However, all species used in the present study were on the breeding grounds more than 3 months before sampling, therefore blood Hg should mainly represent 173 local contamination. 174

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#### 176 Stable isotope analysis

Sub-samples of blood were weighed (mean  $\pm$  SD: 0.29  $\pm$  0.06 mg dw for  $\delta^{13}$ C and 177  $\delta^{15}$ N; 0.71 ± 0.05 mg dw for  $\delta^{34}$ S) with a microbalance and then packed into tin containers 178 for combustion. Relative abundances of carbon, nitrogen and sulfur isotopes were 179 measured using a continuous flow mass spectrometer (Thermo Scientific Delta V 180 Advantage) coupled with an elemental analyzer (carbon and nitrogen: Thermo Scientific 181 Flash EA 1112; sulfur: Thermo Scientific Flash IRMS EA IsoLink). The delta ( $\delta$ ) notation 182 relative to Vienna PeeDee Belemnite, atmospheric N2 and Vienna Cañon Diablo troilite 183 for  $\delta^{13}$ C,  $\delta^{15}$ N and  $\delta^{34}$ S respectively, was used to present isotopic results. Accuracy was 184 checked by replicate measurements of the internal laboratory standard, and analytical 185 precision was <0.10 % for  $\delta^{13}$ C, <0.15 % for  $\delta^{15}$ N and <0.20 % for  $\delta^{34}$ S values. The stable 186

isotopes values were computed as  $\delta X = \left[\left(\frac{R_{Sample}}{R_{standard}}\right) - 1\right] x 1000$ , where X stands for <sup>13</sup>C, <sup>15</sup>N or <sup>34</sup>S and *R* being the ratio <sup>13</sup>C/<sup>12</sup>C, <sup>15</sup>N/<sup>14</sup>N or <sup>34</sup>S/<sup>32</sup>S. Turnover time for  $\delta^{13}$ C and  $\delta$ <sup>15</sup>N in blood is 11~30 days (Barquete et al., 2013; Hobson and Clark, 1992; 1993), therefore stable isotopes values are representative of the breeding period in adults.

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# 192 GPS tracking analysis

A subset (n = 43) of the captured adults (18 EHG, 18 LBBG and 7 GBBG, see SI Table S1) were equipped with GPS-UHF loggers (HARRIER, Ecotone<sup>®</sup>) in 2017 and 2018. Among the 43 GPS equipped birds, 17 EHG, 18 LBBG and 6 GBBG were blood sampled and included in Hg and stable isotopes analyses. The total mass of the logger and the harness (used for attachment) weighted on average 13g (i.e. <2 % of the body mass of LBBG, the smallest species in this study).

199 GPS loggers were set to record locations every 5 min. For each tracked individual, locations were filtered to the period between the day after blood sampling to the last day 200 before either nest failure or egg hatching had been recorded (Figure 1 & SI Figure S1 & 201 202 Table S2). Foraging sites were identified from GPS tracks by assigning a behavioral state to each location using Hidden Markov Models (hereafter "HMMs", Franke et al. 2004) 203 fitted using the moveHMM R package (Michelot et al., 2016). Additional information on 204 205 the spatial data treatment and the behavioral classification can be found in the SI. The habitat for each of these locations was assigned as either "pelagic"; "coastal" (i.e. lagoons, 206 207 estuaries, coastal salt marshes, intertidal flats), or "terrestrial" (all others) based on the CORINE Land Cover dataset (CORINE Land Cover; Feranec et al. 2016). Ratios of pelagic 208 vs coastal and marine (i.e., pelagic and coastal) vs terrestrial habitats used by each 209

210 individual were then calculated.

For an overview of the feeding behaviors leading to the observed Hg concentrations 211 in the tracked species, we estimated birds' foraging site fidelity. As well as giving important 212 insight into the degree of specialization in birds' uses of foraging habitat, this measure of 213 site fidelity served to test whether individuals' foraging behavior in the period after blood 214 215 sampling was likely to be representative of their behavior prior to blood sampling (see SI). Individual foraging site fidelity was assessed following the randomization procedure of 216 Harris et al. (2020). For each foraging site, the distance to a randomly selected site from 217 218 the same individual during another trip (within-individual comparison), as well as to a randomly selected site from each other individual of the same species (between-individual 219 comparisons) was measured. The site fidelity index corresponds to the proportion of 220 between-individual sites that were farther from the focal site than the within-individual 221 site. This index is bounded between 0 and 1, with high values representing high site fidelity 2.2.2 (an individual forages more closely to its own foraging sites than to the sites of other 223 individuals), and low values representing low site fidelity (an individual forages more 224 closely to the foraging sites of other individuals than to its own foraging sites). Additional 225 information can be found in the SI. 226

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#### 228 Molecular sexing

All adults and chicks (n = 247) were sexed using blood samples by polymerase chain reaction (PCR) amplification of parts of two highly conserved genes (CHD) of sex chromosomes, following Fridolfsson and Ellegren (1999).

# 233 Statistical analyses

Statistical analyses were performed using R 4.0.0 (R Core Team, 2020). One YLG chick was removed from the dataset due to outliers in  $\delta^{13}$ C and  $\delta^{34}$ S values. In all species, adults had much higher Hg concentrations than chicks, and therefore chicks and adults were treated separately.

Firstly, relationships between Hg concentrations and diet (inferred by stable isotope 238 values) were investigated. As  $\delta^{13}$ C,  $\delta^{15}$ N and  $\delta^{34}$ S were positively correlated in adults 239 (Pearson correlation >0.70, p < 0.001 for all relationships) and partially correlated in chicks 240 (correlation: C-N = 0.72, p < 0.001; S-C = -0.15; S-N = -0.18, both p > 0.05), principal 241 components analyses ("FactoMineR" R package, version 2.3; Lê et al., 2008) were 242 performed on the three isotopes to reduce the number of explanatory variables. Principal 243 components (PCs) which explained more than 80 % of the total variance were retained. In 244 adults, principal components analysis reduced  $\delta^{13}$ C,  $\delta^{15}$ N and  $\delta^{34}$ S to one PC (PC<sub>1</sub>) 245 explaining 83.6 % of the variance and equally influenced by  $\delta^{13}$ C,  $\delta^{15}$ N and  $\delta^{34}$ S (SI Figure 246 S2 & Table S3). High PC<sub>1</sub> scores reflected high  $\delta^{13}$ C,  $\delta^{15}$ N and  $\delta^{34}$ S. In chicks, principal 247 components analyses reduced  $\delta^{13}$ C,  $\delta^{15}$ N and  $\delta^{34}$ S to two PCs explaining 92.0 % of the 248 variance (SI Figure S2 & Table S3). High PC<sub>1</sub> scores indicated high  $\delta^{13}$ C and  $\delta^{15}$ N values, 249 and high PC<sub>2</sub> scores indicated high  $\delta^{34}$ S values. The isotopic niches, which are essential 250 for an overview of each species' local feeding habits, were investigated, and the foraging 251 252 ecology of each species were illustrated and described in the SI. Generalized least square (GLS) models were then used to investigate the effects of the selected PCs on Hg ("nlme" 253 R package, version 3.1-151; Pinheiro et al., 2020). For that, using Hg as the response 254 255 variable, we included as explanatory covariates the selected PCs and the species. Year was

not included as a predictor in the models to improve statistical power, and also because 256 four years of sampling was insufficient to evaluate temporal trends in Hg concentration. 257 We used GLS models instead of linear models (LM) due to heteroscedasticity in Hg 258 concentrations among species (see SI for further information). For both adults and chicks, 259 after running GLS models, Tukey post-hoc pairwise comparison tests were performed to 260 quantify the differences in Hg concentrations among species ("emmeans" R package, 261 version 1.7.1-1; Lenth et al., 2021). Although performed on only three variables, PCA 262 enabled the inclusion of ecological information provided by the three stable isotopes rather 263 264 than exploring the effect of each stable isotopes in independent models (similar conclusions were obtained using both methods, see SI) 265

Secondly, as Hg levels were found to be highly variable among species, we ran one 266 LM per species to investigate differences between the sexes. With Hg as the response 267 variable, we included the sex as an explanatory variable. This model was not fitted for 268 269 adults of YLG as only a single female was sampled. In previous studies, chick age has been found to be a key predictors of Hg contamination, as blood Hg strongly fluctuates during 270 chick growth, reaching highest concentrations when feathers stop growing (Spalding et al., 271 272 2000). Morphology is commonly used as a proxy of chick age, and we therefore also included the skull length (head and bill) in the LMs for chicks as an additional explanatory 273 variable. Where Hg differed significantly between sexes, an additional LM was built to 274 275 investigate whether this was linked to sex differences in diet, with the selected PCs fitted as the response and sex as an explanatory variable. 276

Finally, in GPS-tracked adults, the impact of the use of marine or terrestrial habitats on Hg concentrations was investigated using a GLS model: Hg was fitted as the response variable, and the proportional use of marine *vs* terrestrial habitat and species (to control for the variation among species) were included as explanatory covariates. A similar model was built to investigate the impact of the use of coastal or pelagic habitats on Hg concentrations, including coastal *vs* pelagic habitats and species (to control for the variation among species) as explanatory covariates. Before modelling, two individuals that were tracked but not blood sampled were removed from the dataset.

For all models, a set of candidate models including all possible combinations of the 285 predictors, ranging from the full model (that includes each of the stated variables) to the 286 287 null model was built. In each case, the best candidate model -i.e., the one with the lowest second-order Akaike's Information Criterion value for small sample size (AICc; 288 "AICcmodavg" R package, version 2.2-2; Mazerolle, 2017), or the most parsimonious 289 model among those with a  $\Delta AICc \leq 2$  – was retained for inference. Homogeneity and 290 normality of the residuals were visually checked using plots of residuals vs fitted values 291 and histograms of the residuals (Zuur et al., 2007). In post-hoc tests, an  $\alpha < 0.05$  threshold 292 was used to assess the significance of the tests. 293

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# 295 **RESULTS**

# 296 Mercury in relation to feeding habits as inferred by stable isotopes

In chicks, Hg concentrations ranged from 0.27  $\mu$ g g<sup>-1</sup> dw in EHG to 3.86  $\mu$ g g<sup>-1</sup> dw in GBBG (Table 1). Model selection indicated that PC<sub>1</sub> and species were significant predictors of Hg concentrations, but not PC<sub>2</sub> (SI Table S4). Post-hoc comparison test showed that GBBG had the highest Hg concentrations, followed by YLG that was not significantly different from EHG and LBBG, but LBBG had lower Hg concentrations than EHG (Tables 1 & 2, Figure 2). Chicks also exhibited a positive relationship between Hg concentration and PC<sub>1</sub> (estimate  $\pm$  SE = 0.09  $\pm$  0.03, *t* = 3.49, *p* <0.001; Table 2 & Figure 3). Sex was not retained as a significant predictor of Hg concentration for any species, but skull size (as a proxy for chick age) was included in the best model for GBBG only (SI Table S5), with Hg significantly increasing with skull size in this species (*F*<sub>1, 26</sub> = 6.51, *p* = 0.017; estimate  $\pm$  SE = 0.04  $\pm$  0.02).

In adults, individual blood Hg concentrations ranged from 0.89  $\mu$ g g<sup>-1</sup> dw in EHG 308 to 21.5  $\mu$ g g<sup>-1</sup> dw in GBBG (Table 1). Model selection showed that PC<sub>1</sub> and species were 309 significant predictors of Hg concentrations in adult gulls (SI Table S4). Post-hoc 310 comparison test indicated that GBBG had the highest Hg concentrations, while lower 311 concentrations were found in YLG, LBBG and EHG (not significantly different among 312 them; Tables 1 & 2, Figure 2). Adults also showed a significant and positive association 313 between Hg concentration and PC<sub>1</sub> (estimate  $\pm$  SE = 0.55  $\pm$  0.06, t = 9.25, p < 0.001; Table 314 2 & Figure 3). Sex was retained for GBBG only (SI Table S5), with males having higher 315 Hg concentrations than females in adults of this species ( $F_{1,35} = 19.2, p < 0.001$ ; estimate  $\pm$ 316 SE =  $4.62 \pm 1.05$ ). However, sex was not retained as a significant predictor of stable 317 isotopes in GBBG (PC<sub>1</sub>; SI Table S6). 318

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#### 320 Mercury in relation to habitat use as inferred by GPS tracking

From GPS tracked adults, a total of 1069 foraging trips from 43 individuals were recorded (612 in EHG, 304 in LBBG and 154 in GBBG). The three tracked species showed different patterns of habitat use: GBBG forage predominantly at sea and showed low interindividual variation (average use of marine habitats  $\pm$  SD: 0.93  $\pm$  0.05 %), while EHG (0.63 325  $\pm$  0.23 %) and LBBG (0.78  $\pm$  0.17 %) individuals were more spread on a gradient from strictly terrestrial to strictly marine (Figure 1 & SI Figure S3). The use of marine habitats 326 in EHG and GBBG was mainly coastal (proportion of coastal habitats use  $\pm$  SD: 0.58  $\pm$ 327 0.25 and  $0.90 \pm 0.07$  respectively), while LBBG used pelagic and coastal habitats 328 (proportion of coastal habitats use  $\pm$  SD: 0.23  $\pm$  0.16; Figure 1 & SI Figure S3). Model 329 selection indicated that the proportional use of marine vs terrestrial habitats and the species 330 were significant predictors of Hg variations (SI Table S7), Hg concentrations increased 331 with the use of marine habitats in all three species (Estimate  $\pm$  SE = 2.59  $\pm$  0.99, t = 2.62, 332 p = 0.01; Figure 4). Conversely, the coastal vs pelagic gradient was not a significant 333 predictor of Hg variations in the three gull species. (Table S7). GBBG, EHG and LBBG 334 exhibited a relatively high fidelity to foraging sites (fidelity index  $\pm$  SD = 0.71  $\pm$  0.18; 0.59 335  $\pm 0.13$  and  $0.72 \pm 0.13$  respectively). 336

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#### 338 **DISCUSSION**

Blood Hg concentrations of four sympatric gull species were investigated in 339 relation to their foraging ecology inferred from stable isotopes  $\delta^{13}$ C,  $\delta^{15}$ N and  $\delta^{34}$ S blood 340 341 values in adults and chicks, and from GPS tracking of adult birds. High Hg concentrations were related to stable isotopes values representative of a high trophic position food, mainly 342 of marine origin. Foraging movements tracked during breeding corroborated the 343 344 relationship between Hg and the birds' foraging strategies with relation to terrestrial vs marine habitat use. All species showed foraging site fidelity, indicating a degree of 345 specialization in their use of foraging sites, and thus high potential for consistent individual 346 347 differences in contaminant level to emerge. This led some adult individuals to exceed reported toxicity thresholds in all four species, particularly in GBBG, showing extremely high levels of Hg. These findings are also concerning for chicks, which predominantly receive marine food from their parents, and are therefore likely to accumulate high concentrations of Hg. Together, these results support the use of sympatric species of gulls as bioindicators of contamination in complex coastal habitats.

353

# 354 Mercury concentrations

LBBG and GBBG chicks had blood Hg concentrations similar to those from the 355 356 French coast of the English Channel, although EHG chicks had higher Hg concentrations, even compared to the colonies from the Seine estuary, one of the most polluted rivers in 357 Europe (Binkowski et al., 2020; Cossa et al., 2002). Toxicological benchmarks are usually 358 assessed for adult birds, meaning that chicks Hg concentrations can only be compared to 359 published thresholds from adults with caution. However, much lower blood Hg 360 361 concentrations have been associated with an impaired physiological condition in LBBG chicks (lower metabolism markers, increased oxidative stress and increased anaerobic 362 metabolism; Santos et al., 2020). In the present study, all four species showed similar Hg 363 364 levels between sexes, likely because sex difference in contaminant exposure is usually visible later in life. However, in GBBG we detected increased levels of Hg with skull size 365 (a proxy of chick age), but this result was not found in other species. Blood Hg is known 366 367 to increase with the chicks age when most of the feathers have grown (Ackerman et al., 2011). This relationship may be easier to detect in GBBG as they grow larger, thus their 368 369 among individual variation in size is higher than the other investigated species.

370 Hg concentrations in adults ranged from similar (EHG and LBBG) to much higher (YLG and GBBG) than in gulls from the Baltic Sea including EHG and GBBG, or Brown 371 skuas (Stercorarius antarcticus lonnbergi) from three colonies of the Southern Ocean 372 (Mills et al., 2022; Szumiło-Pilarska et al., 2017). Notably, we found GBBG to have 373 average blood Hg concentrations similar to those of the Wandering albatross (Diomedea 374 exulans; Anderson et al., 2009), the highest trophic position seabird in the Southern Ocean 375 (Cherel et al., 2017) and one of the species with the highest known Hg levels worldwide 376 (Blévin et al., 2013; Carravieri et al., 2014a; 2014b; Cherel et al., 2018). Such Hg 377 378 concentrations are known to significantly impact reproduction, including breeding success in various bird species (Goutte et al., 2015; Tartu et al., 2013; 2014; 2015b; Ackerman et 379 al., 2016). As no physiological endpoints were measured in the present study, it is not 380 possible to directly draw conclusions on the impact of Hg concentrations of this population. 381 Other factors may also contribute to a reduced toxicity of Hg. For instance, selenium -382 383 which was not measured in the present study - is an essential trace element which may protect birds against Hg toxicity (Dietz et al., 2000; Manceau et al., 2021). However, Hg is 384 known to affect almost all aspects of avian physiology and life history traits (Sebastiano et 385 386 al., 2022; Tartu et al., 2015a; Whitney and Cristol, 2017). We therefore emphasize further work to investigate whether the reported high blood Hg concentrations are detrimental to 387 the different species of birds in the region. Similarly to what we observed in GBBG, higher 388 389 concentrations of Hg in males than in females were previously found in other bird species (Ackerman et al., 2007; Burgess et al., 2005; La Sala et al., 2011). Interestingly, incubating 390 391 GBBGs males and females were not found to differ in terms of stable isotopes 392 concentration, suggesting diet is not the main driver of this difference. A possible explanation may be represented by the fact that female gulls deposit Hg into their eggs
therefore lowering their circulating contaminants (Lewis et al., 1993).

395

# 396 Mercury concentrations in relation to foraging ecology

The positive relationship between blood Hg concentrations and  $\delta^{13}$ C or  $\delta^{15}$ N values 397 observed in gull chicks is in agreement with previous studies on the same four species in 398 the English Channel (Binkowski et al., 2020) and the southern Bay of Biscay (Zorrozua et 399 al., 2020). However, blood  $\delta^{34}$ S was not found to be an important driver of Hg 400 contamination at Ile de Ré, contrary to chicks in the English Channel (Binkowski et al., 401 2020) and Spain (Ramos et al., 2013). In the present study, chick isotopic values were in 402 the highest range values of those of their parents for  $\delta^{13}$ C,  $\delta^{15}$ N and  $\delta^{34}$ S, meaning that they 403 predominantly received food at a higher trophic position and of marine origin. A diet shift 404 after hatching – reflecting different food requirement before and after hatching – has 405 previously been described in EHG and other species of gulls (Annett and Pierotti, 1989; 406 Bukacińska et al., 1996; Pierotti and Annett, 1987). This specialized chick diet of low 407 marine prey diversity, provided by the parents, may explain why the terrestrial vs marine 408 409 gradient is not a good predictor of Hg variations in chicks.

Isotopic data showed that, similarly to chicks, blood Hg contamination was higher in adult gulls feeding on marine than terrestrial items. This is consistent with previous studies investigating the relationship between Hg and  $\delta^{13}$ C,  $\delta^{15}$ N and  $\delta^{34}$ S in coastal seabirds and their prey (Gongora et al., 2018; Peterson et al., 2017). These patterns likely reflect the biomagnifying characteristics of Hg along food webs, with longer and more complex food webs in the marine environment (Bełdowska and Falkowska, 2016; Morel et al., 1998), as 416 well as that MeHg is mainly produced in the ocean (Chen et al., 2008). Conversely, some previous studies did not report such relationship in other aquatic birds (Einoder et al., 2018; 417 Soldatini et al., 2020), however, the association between Hg and  $\delta^{13}$ C or  $\delta^{15}$ N is complex 418 and largely depends on the species or their environments. Different environments likely 419 differ in their  $\delta^{15}$ N baseline (Elliott et al., 2021), thus precluding comparing directly 420 consumers'  $\delta^{15}$ N values as reflecting their trophic position. This is likely the case of gulls 421 from Ile de Ré, meaning that any blood  $\delta^{15}$ N difference at the species or individual level 422 must be interpreted with caution, especially if their  $\delta^{13}$ C and  $\delta^{34}$ S values indicate foraging 423 in different habitats. 424

In the present study, EHG, LBBG and YLG showed important inter-individual 425 variations in blood  $\delta^{13}$ C,  $\delta^{15}$ N and  $\delta^{34}$ S values, that characterize populations of generalist 426 species made of specialist individuals (Jaeger et al., 2009). In contrast, GBBG exhibited 427 narrower ranges for all isotopic values, representative of a specialist species. GBBG's 428 isotopic values were in the highest range of those of the other gull species, that characterize 429 a diet based on marine items ( $\delta^{34}$ S: +15 to +19 ‰), whereas EHG, LBBG and YLG 430 individuals showed a diet of mixed terrestrial, intertidal and marine sources (Nehlich, 2015; 431 432 Rees et al., 1978). The very high Hg concentrations found in GBBG are therefore most likely related to its marine diet. These isotopic results are supported by the analyses of 433 foraging trips in EHG, LBBG and GBBG, as the use of a marine habitats was related to 434 435 higher blood Hg concentrations. In other species of coastal gulls equipped with tracking devices, including the American Herring gull (Larus smithsonianus) and the Western gull 436 437 (Larus occidentalis), a similar pattern was recently observed, with individuals mainly 438 foraging in marine environments exhibiting the highest Hg concentrations (Clatterbuck et 439 al., 2021; Thorne et al., 2021). EHG showed higher use of coastal foraging sites but similar 440 Hg concentrations compared to the more pelagic LBBG. The coastal specialists GBBG 441 also presented the highest Hg concentrations, further suggesting that "marine *vs* terrestrial" 442 may represent a better gradient than "coastal *vs* pelagic" to explain Hg variations in gull 443 species. As  $\delta^{34}$ S highly discriminates between terrestrial and marine habitats (McCutchan 444 Jr et al., 2003), we further stress the importance of the use of  $\delta^{34}$ S as relevant index of 445 foraging habitats.

Nevertheless, Figures 3 & 4 reveal a weaker relationship between blood Hg and PC<sub>1</sub> in GBBGs. It was however not possible to include the interaction between PC<sub>1</sub> and species in the models to study if this relationship was different among species, due to the relatively different range in PC<sub>1</sub> and Hg values between GBBG and the other species. In chicks, the additive effect of maternal Hg initially transferred in the egg may contribute to blur the relationship in all species.

Surprisingly, the range of Hg concentrations seems to be inversely correlated to the 452 isotopic niche width in adults, i.e., with the foraging habitat diversity. In the specialist 453 GBBG, high fidelity to foraging sites may explain the high inter-individual variation in Hg 454 455 concentration despite a narrow isotopic niche, i.e., birds feeding on a similar trophic position prey but specialized in areas of different contamination intensities (river estuarine, 456 salt-marshes, intertidal area, open sea,...; Mitchell and Gilmour, 2008), generating 457 458 individual variation in Hg exposure. EHG and LBBG also exhibited a relatively high fidelity to foraging sites, but the lower range of Hg concentrations compared to GBBG 459 suggests that the foraging areas provided by movement analyses are important to 460 461 understand the sources of Hg in specialist species.

462

# 463 CONCLUSION

In the present study, we have highlighted the need to refine ecotoxicological studies 464 with stable isotope analyses, as the trophic position (using  $\delta^{15}$ N as a proxy) and the feeding 465 habitat ( $\delta^{34}$ S) were important drivers of blood Hg contamination. We also point out that not 466 only the use of GPS data highly reflects the results on Hg concentrations and habitat 467 selection through isotopic analyses, but it may further provides fundamental insights on 468 inter-individual contamination through habitat fidelity. We argue for the concomitant use 469 470 of both GPS and the stable isotopes method to characterize the sources of contaminants in seabirds. 471

Our results also call for a potential toxicological risk in both chicks and adults. As 472 chicks receive high trophic position prey of marine origin from their parents, such diet 473 could lead to Hg concentrations of concern during their development. In adults, GBBG 474 showed particularly high concentrations of Hg due to its almost exclusively marine food. 475 Nonetheless, several adults of all four species might be at risk as exceeded previously 476 reported Hg toxicity thresholds. It is therefore fundamental to further investigate the 477 potential detrimental effects of Hg exposure on physiological and life-history traits of the 478 studied species. 479

480

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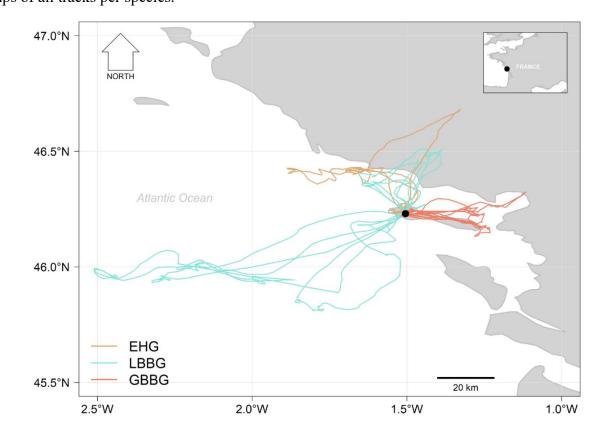
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Figure 1. Map of the Ile de Ré and surroundings, France, visited by the adults of four species of gulls from the "Lilleau des Niges" colony during their breeding period; European herring gulls (EHG), Lesser black-backed gulls (LBBG), Great black-backed gulls (GBBG) and Yellow-legged gulls (YLG). The studied colony is represented by the black point. YLG were not tracked and so are not presented, but were included in analyses of Hg and stable isotopes. Tracks presented here are a random selection of 5 trips per species, see SI Figure S1 for maps of all tracks per species.



507	<b>Table 1.</b> Hg concentrations ( $\mu$ g g <sup>-1</sup> dw) and carbon ( $\delta^{13}$ C), nitrogen ( $\delta^{15}$ N) and sulfur ( $\delta^{34}$ S) stable isotope values (‰) measured in blood
508	of adults and chicks of four gull species from the Ile de Ré, France: sample size ( $n$ ), mean $\pm$ standard deviation ( $SD$ ), median and range

509 (min-max). For adults, Hg values are presented separately for females ( $\bigcirc$ ) and males ( $\bigcirc$ ).

			Adu	lts				
		Hg	$\delta^{13}$ C		$\delta^{15}$ N		$\delta^{34}$ S	
	n	Mean ± SD Min/max	Mean ± SD	Min/max	Mean ± SD	Min/max	Mean ± SD	Min/max
EHG (Larus argentatus)	44	$2.29 \pm 1.06  0.89/5.01$	$\textbf{-19.4} \pm 1.93$	-22.9/-15.5	$12.7\pm1.82$	9.76/16.0	$13.1\pm2.23$	8.72/17.0
Ŷ	23	$2.05 \pm 0.96  0.90/4.31$						
8	21	$2.55 \pm 1.12  0.89/5.01$			-			
LBBG (Larus fuscus)	54	$3.08 \pm 1.31  0.95/6.42$	$\textbf{-19.1} \pm 1.34$	-22.6/-17.6	$13.8\pm1.30$	10.9/15.8	$15.9\pm2.37$	9.90/18.6
Ŷ	27	$2.82 \pm 1.03  0.98/4.78$						
8	27	$3.35 \pm 1.52  0.95/6.42$			-			
GBBG (Larus marinus)	37	$12.5\pm 3.93  6.97/21.5$	$\textbf{-16.8} \pm \textbf{0.83}$	-19.7/-14.7	$16.4\pm0.41$	15.8/17.8	$16.9 \pm 1.07$	14.6/18.7
Ŷ	19	$10.3\pm 2.20  6.97/13.8$						
8	18	$14.9\pm 4.00  7.26/21.5$			-			
YLG (Larus michahellis)	5	$6.00 \pm 2.72  2.25/9.63$	$\textbf{-19.4} \pm \textbf{3.51}$	-25.2/-16.7	$15.1\pm1.32$	12.76/15.91	$12.6\pm4.66$	5.87/16.7
<b>P</b>	1	$2.25 \pm NA$ 2.25/2.25						
8	4	$6.94 \pm 2.00  5.26/9.63$			-			

Chicks

		Hg	$\delta^{13}$ C		$\delta^{15}{ m N}$		$\delta^{34}$ S	
	n	Mean ± SD Min/max	Mean ± SD	Min/max	Mean ± SD	Min/max	Mean ± SD	Min/max
EHG (Larus argentatus)	30	$0.68 \pm 0.25  0.27/1.19$	$-18.3 \pm 0.66$ -	19.9/-17.1	$13.8\pm0.75$	12.1/15.0	$16.8\pm1.21$	14.0/18.7
LBBG (Larus fuscus)	37	$0.58 \pm 0.15  0.35/0.94$	$-18.3 \pm 0.27$ -	18.8/-17.7	$14.5\pm0.41$	13.7/15.2	$18.4\pm0.93$	16.7/19.8
GBBG (Larus marinus)	28	$1.91 \pm 0.76  0.81/3.86$	$-17.1 \pm 0.77$ -1	19.33/-16.7	$15.6\pm0.48$	14.0/16.6	$16.5\pm1.32$	12.8/18.6
YLG (Larus michahellis)	11	$0.90 \pm 0.34  0.48/1.75$	$-18.0 \pm 0.61$ -1	9.37/-17.42	$14.9 \pm 1.07$	12.4/16.1	$16.5\pm1.46$	14.07/18.4

Table 2. Outputs of linear models examining the variables affecting Hg concentration in
four gull species from the Ile de Ré, France: European herring gulls (EHG), Lesser blackbacked gulls (LBBG), Great black-backed gulls (GBBG) and Yellow-legged gulls (YLG).
Significant *p*-values are in bold.

Adults						
Parameter	Estimate	SE	<i>t</i> -value	<i>p</i> -value		
PC <sub>1</sub>	0.55	0.06	9.25	<0.001		
EHG - LBBG	0.10	0.19	0.56	0.943		
EHG - GBBG	8.56	0.67	12.70	<0.001		
EHG - YLG	2.65	0.79	3.38	0.089		
LBBG - GBBG	8.45	0.66	12.79	<0.001		
LBBG - YLG	2.55	0.79	3.22	0.099		
GBBG - YLG	-5.90	1.02	-5.78	<0.001		

Chicks

CHICKS						
Parameter	Estimate	SE	<i>t</i> -value	<i>p</i> -value		
PC <sub>1</sub>	0.09	0.03	3.49	<0.001		
EHG - LBBG	-0.13	0.05	-2.72	0.043		
EHG - GBBG	1.01	0.16	6.33	<0.001		
EHG - YLG	0.11	0.12	0.93	0.792		
LBBG - GBBG	1.14	0.15	7.45	<0.001		
LBBG - YLG	0.24	0.11	2.16	0.189		
GBBG - YLG	-0.90	0.18	-5.05	<0.001		

**Figure 2**. Blood concentrations of Hg in adults and chicks of four gull species from the Ile de Ré, France; European herring gulls (EHG), Lesser black-backed gulls (LBBG), Great black-backed gulls (GBBG) and Yellow-legged gulls (YLG). For adults (left), Hg values are shown separately for females ( $\bigcirc$ ) and males ( $\bigcirc$ ). Interspecific (dark-grey) and between sex (light grey) statistically significant differences are indicated by the asterisk: \*, \*\*, \*\*\*; indicating a *p*-value < 0.05, <0.01, and <0.001, respectively. Values are median, 25<sup>th</sup> and 75<sup>th</sup> percentiles and range.



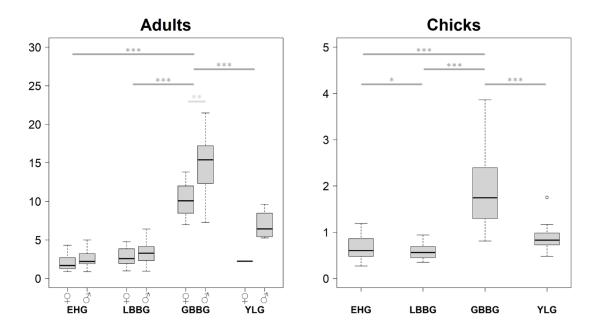
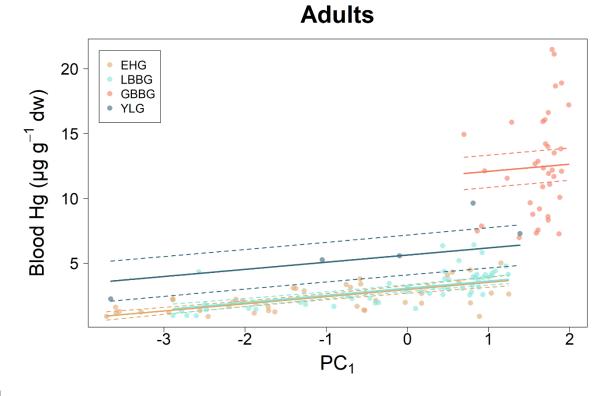
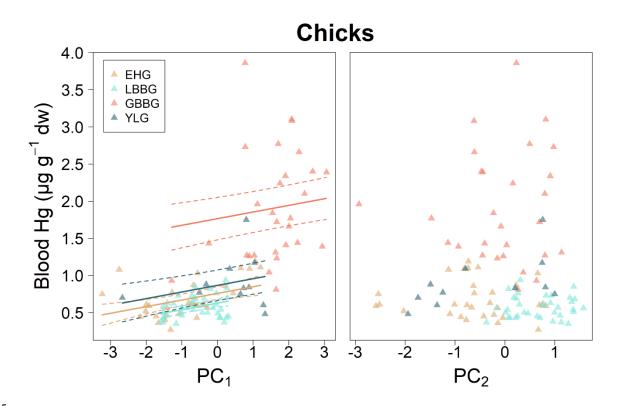


Figure 3. Relationship between blood Hg concentrations and PC<sub>1</sub> ( $\delta^{13}$ C,  $\delta^{15}$ N and  $\delta^{34}$ S values in adults and  $\delta^{13}$ C,  $\delta^{15}$ N in chicks) or PC<sub>2</sub> ( $\delta^{34}$ S values in chicks) of four gull species from Ile de Ré, France: European herring gulls (EHG), Lesser black-backed gulls (LBBG), Great black-backed gulls (GBBG) and Yellow-legged gulls (YLG). For each species, the solid line refers to the fitted models obtained and dotted lines represent 95 % confident intervals.

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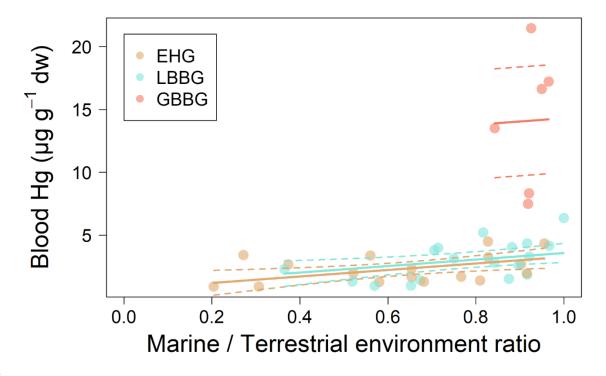






**Figure 4.** Relationship between blood Hg concentrations and the proportion of foraging trips in marine (1: exclusively marine) *vs* terrestrial (0: exclusively terrestrial) habitats obtain by GPS tracking in adults of three gull species from Ile de Ré, France; European herring gulls (EHG), Lesser black-backed gulls (LBBG) and Great black-backed gulls (GBBG). For each species, the solid line refers to the fitted models obtained and dotted lines represent 95 % confident intervals.





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