



HAL
open science

A time-lapse embryo dataset for morphokinetic parameter prediction

Tristan Gomez, Magalie Feyeux, Justine Boulant, Nicolas Normand, Laurent David, Perrine Paul-Gilloteaux, Thomas Fréour, Harold Mouchère

► **To cite this version:**

Tristan Gomez, Magalie Feyeux, Justine Boulant, Nicolas Normand, Laurent David, et al.. A time-lapse embryo dataset for morphokinetic parameter prediction. *Data in Brief*, 2022, pp.108258. 10.1016/j.dib.2022.108258 . hal-03667158

HAL Id: hal-03667158

<https://hal.science/hal-03667158>

Submitted on 24 May 2024

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Distributed under a Creative Commons Attribution 4.0 International License



Data Article

A time-lapse embryo dataset for morphokinetic parameter prediction



Tristan Gomez^{a,*}, Magalie Feyeux^b, Justine Boulant^c,
Nicolas Normand^a, Laurent David^b, Perrine Paul-Gilloteaux^b,
Thomas Fréour^d, Harold Mouchère^a

^a Nantes Université, Ecole Centrale Nantes, CNRS, LS2N, UMR 6004, F-44000 Nantes, France

^b University of Nantes, Nantes University Hospital, Inserm, CNRS, SFR Santé, Inserm UMS 016, CNRS UMS 3556, F-44000 Nantes, France. Address: 8 Quai Moncoussu, 44007 Nantes

^c University of Nantes, Nantes University Hospital, Inserm, DSN, Inserm UMR 1064, F-44000 Nantes, France. Address: 8 Quai Moncoussu, 44007 Nantes

^d University of Nantes, Nantes University Hospital, Inserm, CRTI, Inserm UMR 1064, F-44000 Nantes, France. Address: 8 Quai Moncoussu, 44007 Nantes

ARTICLE INFO

Article history:

Received 13 April 2022

Revised 4 May 2022

Accepted 5 May 2022

Available online 11 May 2022

Keywords:

Human reproduction

Deep learning

Time-lapse

Videos

IVF

Computer vision

ABSTRACT

One of the most common treatments for infertile couples is In Vitro Fertilization (IVF). It consists of controlled ovarian hyperstimulation, followed by ovum pickup, fertilization, and embryo culture for 2–6 days under controlled environmental conditions, leading to intrauterine transfer or freezing of embryos identified as having a good implantation potential by embryologists. To allow continuous monitoring of embryo development, Time-lapse imaging incubators (TLI) were first released in the IVF market around 2010. This time-lapse technology provides a dynamic overview of embryonic in vitro development by taking photographs of each embryo at regular intervals throughout its development. TLI appears to be the most promising solution to improve embryo quality assessment methods, and subsequently the clinical efficiency of IVF. In particular, the unprecedented high volume of high-quality images produced by TLI systems has already been leveraged using modern Artificial Intelligence (AI) methods, like deep learning (DL).

* Corresponding author.

E-mail address: tristan.gomez@univ-nantes.fr (T. Gomez).

Social media: [@perrinepgil](https://twitter.com/perrinepgil) (P. Paul-Gilloteaux), [@HaroldMouchere](https://twitter.com/HaroldMouchere) (H. Mouchère)

An important limitation to the development of AI-based solutions for IVF is the absence of a public reference dataset to train and evaluate deep learning (DL) models. In this work, we describe a fully annotated dataset of 704 TLI videos of developing embryos with all 7 focal planes available, for a total of 2,4M images. Of note, we propose highly detailed annotations with 16 different development phases, including early cell division phases, but also late cell divisions, phases after morulation, and very early phases, which have never been used before. This is the first public dataset that will allow the community to evaluate morphokinetic models and the first step towards deep learning-powered IVF. We postulate that this dataset will help improve the overall performance of DL approaches on time-lapse videos of embryo development, ultimately benefiting infertile patients with improved clinical success rates.

© 2022 The Authors. Published by Elsevier Inc.
 This is an open access article under the CC BY license (<http://creativecommons.org/licenses/by/4.0/>)

Specifications Table

Subject	Computer Vision and Pattern Recognition
Specific subject area	Time-lapse videos of human embryos with morphokinetic parameter annotations.
Type of data	Image Video
How the data were acquired	Images were acquired using the Embryoscope®, a time-lapse incubator system (Vitrolife®, Sweden) with a camera under a 635 nm LED light source passing through Hoffman's contrast modulation optics.
Data format	Raw Annotated
Description of data collection	The data comes from infertile couples that underwent Intracytoplasmic Sperm Injection (ICSI) cycles. Patient treatment and embryo culture protocol were described in a previous study [1]. Videos corresponding to embryos with less than 6 annotated phases were rejected to keep only videos with detailed annotations. Images were extracted using the manufacturer's (Vitrolife®) API.
Data source location	<ul style="list-style-type: none"> • Institution: Nantes University Hospital • City/Town/Region: Nantes • Country: France
Data accessibility	Repository name: The dataset is hosted at https://zenodo.org [2] Data identification number: 10.5281/zenodo.6390798 Direct URL to data: https://doi.org/10.5281/zenodo.6390798
Related research article	No related article

Value of the Data

- This dataset is composed of 704 TLI videos of developing embryos with all 7 focal planes available, for a total of 2,4M images.
- We propose highly detailed annotations with 16 different development phases, including early cell division phases, but also late cell divisions, phases after morulation, and very early phases, which have never been used before.
- This dataset can be used to train machine learning models to identify the various phases of embryo development from polar body appearance to blastocyst hatching.

- Researchers and developers of time-lapse automated analysis software can use this dataset to evaluate and compare new models against previously proposed models.
- To the best of our knowledge, this is the first embryo time-lapse dataset publicly available.

1. Data Description

This dataset is composed of 704 videos, each recorded at 7 focal planes, accompanied by the annotations of 16 cellular events.

First, we describe how the events were annotated, then how each frame was assigned a label, followed by the file descriptions and some dataset statistics and samples.

Dataset annotation. Each video was annotated by a qualified and experienced embryologist undergoing regular internal quality control. For each video, the annotation consists of the timing of 16 cellular events noted tPB2, tPNa, tPNf, t2, t3, t4, t5, t6, t7, t8, t9+, tM, tSB, tB, tEB, and finally tHB. We use the definition of the events proposed by Ciray et al. [3]: polar body appearance (tPB2), pronuclei appearance and disappearance (tPNa and tPNf), blastomere division from 2-cell stage to 9 (and more) cells-stage (t2,t3,t4,t5,t6,t8 and t9+), compaction (tM), blastocyst formation (tSB, tB), expansion and hatching (tEB and tHB). We chose to use more events than previous work [4–7] to develop models that can more precisely describe embryo development in a controlled environment.

We started prospective annotation of the database according to this reference work in 2014, while annotations made before 2014 were retrospectively checked.

From event timing to frame labels. We formulate the task as an image classification problem. This means that we need to assign a label to each frame that the model will be trained to predict. However, the annotations given by the biologists are timings in hours post-fertilization that indicates the temporal position of events in the video.

Knowing the timing at which each frame was taken, we identify the frames corresponding to each event and assigns them a label corresponding to the event they show (noted pPB2, pPNa, pPNf, p2, p3, p4, p5, p6, p7, p8, p9+, pM, pSB, pB, pEB or pHB), as illustrated in Fig. 1.

The other frames are assigned the label corresponding to the most recent event that has occurred in the previous frames. This labeling constructs the succession of embryo development phases, delimited by the cellular events.

Dataset files. The dataset is composed of several compressed folders:

- *embryo_dataset.tar.gz* contains 704 folders, each containing all the frames of one video. An embryo is a 3D object and the focal plane of the microscope can be changed to better visualize the embryo. This compressed folder contains images recorded at the central focal plane called F0. The images are JPEG files in shades of gray with a 500×500 resolution.
- *embryo_dataset_annotations.tar.gz* contains the 704 CSV annotations files. Each CSV is the annotation of one video and has three columns: phase, start frame index and end frame index that indicates the frame interval of each phase. Table 1 shows an example with the annotation file of the video AG274-2, which construction is illustrated in Fig. 1.
- Six other compressed folders containing the same videos as *embryo_dataset.tar.gz*, except recorded with different focal plane settings. There is 6 alternative focal planes setting available: F-45, F-30, F-15, F15, F30 and F45. The folders are named as follows *embryo_dataset_X.tar.gz* where X is one of the mentioned focal planes. Each folder contains 704 folders, each containing all the frames of one video, recorded at the focal plane X.

Dataset statistics. Deep learning models are heavily dependent on data and might provide poor performance on a specific class if the amount of input corresponding to it is too small. This is why for each label, we provide at least several thousand images (Fig. 2 (a)). Most videos have at least 8 annotated phases and approximately 360 videos have more than 13 phases annotated, illustrating the richness of annotation of our dataset (Fig. 2 (b)).

Dataset samples. Sample images allow one to have a clear view of the content of the dataset and the annotations associated with the images (Fig. 3). Note that, depending on their position

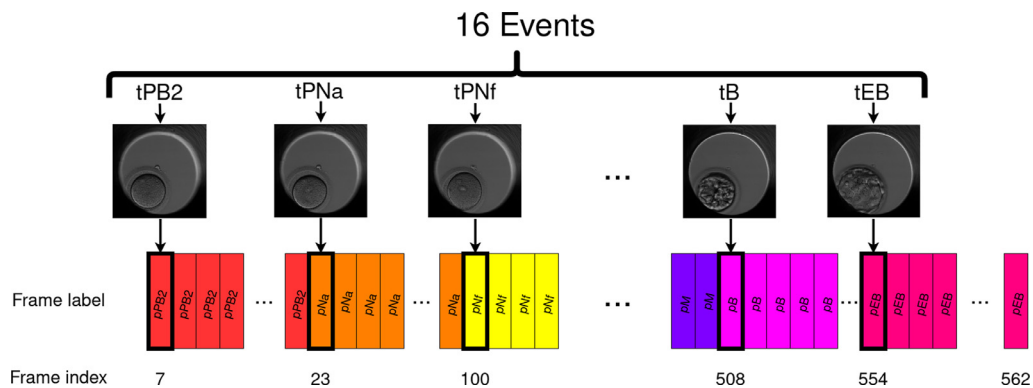


Fig. 1. The method used to assign a label to every frame of the video. First, we identify at which frame each event occurs and assign to these frames a label corresponding to the event they show. The other frames are assigned the label corresponding to the most recent event that has occurred in the previous frames. Note that all frames are labeled except the frames before tPB2 as they precede all the events. The video used as an example here is AG274-2.

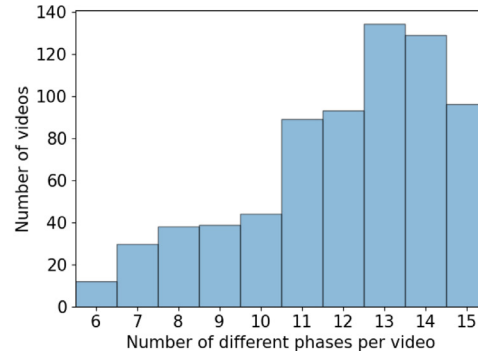
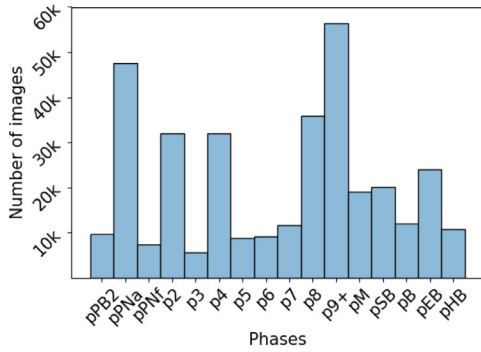


Fig. 2. Statistics of the dataset. (a) The number of images per phase in the dataset. (b) Distribution of the number of phases per video in the dataset.

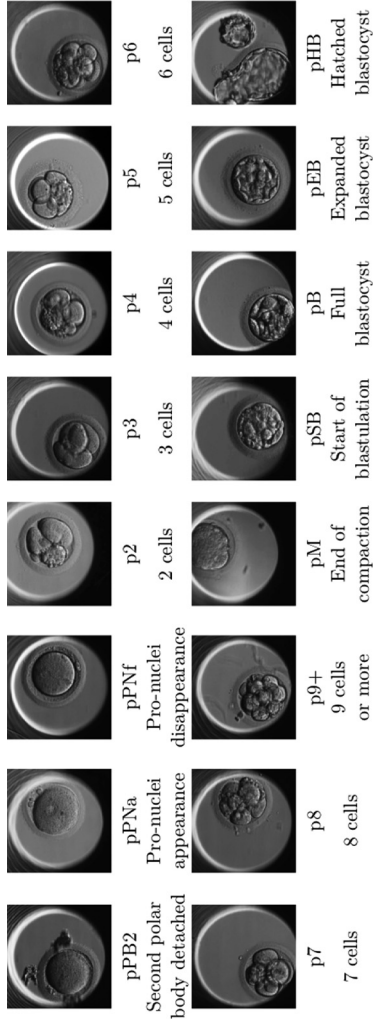


Fig. 3. Illustrations of the 16 development phases used.

Table 1

Example of an annotation CSV file. The first column indicates the phase (the label to predict) and the second and third columns indicate the frame index at which this phase starts and ends. Note that all frames are labeled except the frames before tPB2 as they precede all the events. The file shown here is the annotation of the video AG274-2 which is called AG274-2_phases.csv.

tPB2	7	22
tPNa	2	99
tPNf	100	111
t2	112	169
t3	170	171
t4	172	177
t5	178	228
t6	229	234
t7	235	318
t8	319	328
t9+	329	464
tM	465	491
tSB	492	507
tB	508	553
tEB	554	562

in the well, embryos can sometimes be partially occluded which is quite common in time-lapse videos. However, even when a part of the embryo is hidden, the images are sufficient to identify the development phase.

2. Experimental Design, Materials and Methods

Between 2011 and 2019, 716 infertile couples underwent Intracytoplasmic Sperm Injection (ICSI) cycles in our University-based IVF center and had all their embryos cultured and monitored up to blastocyst stage with a TLI system. We acknowledge that only ICSI cycles were included in our time-lapse devices over that period, as we considered that conventional IVF would lead to different developmental timings as compared to ICSI. We do not routinely use assisted hatching. There were no major lab changes over the study period. Patient treatment and embryo culture protocol were described in a previous study [1]. In brief, embryo culture was performed from fertilization (day 1) up to blastocyst stage (day 5 or day 6) at 37 °C with 5% O₂ and 6% CO₂ in a sequential culture medium, i.e. G1 plus (Vitrolife®, Sweden) from day 0 to day 3, followed by G2 plus (Vitrolife®, Sweden). We acknowledge that culture media might impact embryo development and have an evolving composition throughout embryo development. However, the available literature does not support the concept of medium-dependent morphokinetic patterns [8]. Although we agree that there is a need to clarify IVF culture media composition to enhance our understanding of embryo development [9], there is no evidence to our knowledge that the content of commercial culture media changes over time in ways that are important enough to consider. The images were acquired with a TLI system (Embryoscope®, Vitrolife®, Sweden) every 10 to 20 min by a camera under a 635 nm LED light source passing through Hoffman's contrast modulation optics.

To select the videos, we first excluded videos with strictly less than 6 phases annotated to keep only videos with highly detailed annotations and then randomly selected 10% of the remaining videos, which constitutes a dataset of 704 videos. Among these videos, 499 correspond to embryos considered to be morphologically viable and subsequently chosen for transfer, while the other videos correspond to discarded embryos because of poor development. These discarded embryos allowed us to study a variety of abnormal embryonic features (abnormal morphology, abnormal fertilization/number of pro-nuclei, necrosis, fragmentation, developmental delay, etc.) or problems during image acquisition (sharpness, change of focus, brightness, etc.). We subsequently extracted all images using the Application Programming Interface (API)

provided by the TLI manufacturer (Vitrolife®). The information about embryo viability is not included in this dataset as the purpose is to focus solely on morphokinetic parameter prediction.

Ethics Statements

The Local Institutional Review Board (GNEDS) (local ethics committee) approved this project before it started. The anonymised database is registered under CNIL approval number

1760497. All patients gave informed consents for the use of their anonymized clinical data in observational study. The research has been carried out in accordance with The Code of Ethics of the World Medical Association (Declaration of Helsinki).

CRediT Author Statement

Tristan Gomez: Methodology, Software, Data curation, Writing, Original Draft Supervision; **Magalie Feyeux:** Data curation, Writing; **Justine Boulant:** Software; **Nicolas Normand:** Reviewing; **Laurent David:** Supervision, Reviewing; **Perrine Paul-Gilloteaux:** Supervision, Reviewing; **Thomas Fréour:** Supervision, Reviewing; **Harold Mouchère:** Supervision, Reviewing.

Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

Data Availability

[Human embryo time-lapse video dataset \(Original data\)](#) (Zenodo).

Acknowledgments

The authors would like to thank the IVF staff at the University Hospital of Nantes, and more specifically Dr. Arnaud Reignier and Mrs. Jenna Lammers for the annotation of the database.

Funding: This work was funded by ANR - Next grant [DL4IVF \(2017\)](#).

References

- [1] T. Fréour, N. Le Fleuter, J. Lammers, C. Splingart, A. Reignier, P. Barrière, External validation of a time-lapse prediction model, *Fertil. Steril.* 103 (4) (2015) 917–922, doi:[10.1016/j.fertnstert.2014.12.111](#).
- [2] T. Gomez, M. Feyeux, J. Boulant, N. Normand, P. Paul-Gilloteaux, L. David, T. Fréour, H. Mouchère, Human embryo time-lapse video dataset, Zenodo v1 (2022), doi:[10.5281/zenodo.6390798](#).
- [3] HN. Ciray, A. Campbell, IE. Agerholm, J. Aguilar, S. Chamayou, M. Esbert, S. Sayed, Time-lapse user group, proposed guidelines on the nomenclature and annotation of dynamic human embryo monitoring by a time-lapse user group, *Hum. Reprod.* 29 (12) (2014) 2650–2660, doi:[10.1093/humrep/deu278](#).
- [4] T. Lau, N. Ng, J. Gingold, N. Desai, J. McAuley, Z.C. Lipton, Embryo staging with weakly-supervised region selection and dynamically-decoded predictions, *CoRR* abs/1904.04419 (2019) <http://arxiv.org/abs/1904.04419>.
- [5] J. Silva-Rodríguez, A. Colomer, M. Meseguer, V. Naranjo, Predicting the success of blastocyst implantation from Morphokinetic parameters estimated through cnns and sum of absolute differences, in: 2019 27th European Signal Processing Conference, 2019, pp. 1–5, doi:[10.23919/EUSIPCO.2019.8902520](#).
- [6] A. Khan, S. Gould, M. Salzman, Deep convolutional neural networks for human embryonic cell counting, in: G. Hua, H. Jégou (Eds.), *Computer Vision – ECCV 2016 Workshops*. ECCV. Lecture Notes in Computer Science, 9913, Springer, Cham, 2016, pp. 339–348, doi:[10.1007/978-3-319-46604-0_25](#).

- [7] R.M. Rad, P. Saeedi, J. Au, J. Havelock, Blastomere cell counting and centroid localization in microscopic images of human embryo, in: 2018 IEEE 20th International Workshop on Multimedia Signal Processing (MMSP), 2018, pp. 1–6, doi:[10.1109/MMSP2018.8547107](https://doi.org/10.1109/MMSP2018.8547107).
- [8] N. Basile, D. Morbeck, J. García-Velasco, F. Bronet, M. Meseguer, Type of culture media does not affect embryo kinetics: a time-lapse analysis of sibling oocytes, *Hum. Reprod.* 28 (3) (2013) 634–641, doi:[10.1093/humrep/des462](https://doi.org/10.1093/humrep/des462).
- [9] A. Sunde, D. Brison, J. Dumoulin, J. Harper, K. Lundin, M.C. Magli, E. Van den Abbeel, A. Veiga, Time to take human embryo culture seriously, *Hum. Reprod* 31 (10) (2016) 2174–2182, doi:[10.1093/humrep/dew157](https://doi.org/10.1093/humrep/dew157).