

## Cover crops combined with soil tillage impact the spontaneous species density, richness and diversity in banana cover cropping systems

Diane Rakotomanga, Médina Lacoma, Marc Dorel, Gaëlle Damour

### ▶ To cite this version:

Diane Rakotomanga, Médina Lacoma, Marc Dorel, Gaëlle Damour. Cover crops combined with soil tillage impact the spontaneous species density, richness and diversity in banana cover cropping systems. Agronomy for Sustainable Development, 2021, 41 (3), pp.37. 10.1007/s13593-021-00692-1. hal-03662040

## HAL Id: hal-03662040 https://hal.science/hal-03662040v1

Submitted on 9 May 2022

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés. **RESEARCH ARTICLE** 



### Cover crops combined with soil tillage impact the spontaneous species density, richness and diversity in banana cover cropping systems

Diane Rakotomanga<sup>1,2,3</sup> • Médina Lacoma<sup>2,3</sup> • Marc Dorel<sup>2,3</sup> • Gaëlle Damour<sup>1,3</sup>

Accepted: 6 April 2021 / Published online: 5 May 2021 © INRAE and Springer-Verlag France SAS, part of Springer Nature 2021

### Abstract

Soil tillage and cover crops impact spontaneous vegetation in agrosystems. Soil tillage destroys seedlings and can affect the soil seedbank. Cover crops compete with naturally occurring spontaneous species. Our aim was to gain insight into how soil tillage (here, the false seedbed technique) and cover crops impact the spontaneous vegetation in fallows preceding banana crops. In a fallowed field in Guadeloupe (French West Indies), two factors were combined in a split-plot design: the soil tillage modality before cover crop sowing (with or without the false seedbed technique) and the cover crop species (Crotalaria spectabilis, Pueraria phaseoloïdes, a 50/50 mix of Brachiaria decumbens + Brachiaria ruziziensis, a spontaneous vegetation control). On a monthly basis, in three permanent quadrats per sub-plot, all spontaneous species were identified and their numbers counted, while calculating the species richness and Shannon's diversity index. The spontaneous vegetation dynamics contrasted depending on the treatment. From 7 months post-sowing the species density, richness and Shannon index were significantly lower in treatments with B. decumbens + B. ruziziensis and P. phaseoloïdes, while in treatments with C. spectabilis, the species density and richness were significantly higher with than without the false seedbed technique. These effects were probably related to differences in the competitive ability of the cover crop used and, to a lesser extent, to the false seedbed effects on the soil seedbank. This is the first study to show that under a tropical climate, sown cover crop species combined with different soil tillage modalities have substantial impacts on the species density, richness and diversity of the spontaneous plant community over a 1-year period. These results could help farmers in their weed control strategies by providing them with a deeper understanding of the effects of practices on the spontaneous plant community.

**Keywords** Brachiaria decumbens · Brachiaria ruziziensis · Crotalaria spectabilis · Species diversity · False seedbed technique · Pueraria phaseoloïdes · Species richness · Weed

### 1 Introduction

In agrosystems, spontaneous species may compete for resources with crops and decrease yields. Chemical management has long been the main control strategy, but the detrimental effects on the environment have prompted exploration of alternative strategies. Apart from herbicide applications,

Diane Rakotomanga diane.rakotomanga@hotmail.fr

- <sup>1</sup> UPR GECO, CIRAD, Cedex 5 34398 Montpellier, France
- <sup>2</sup> UPR GECO, CIRAD, 97130 Capesterre-Belle-Eau, France
- <sup>3</sup> UPR GECO, CIRAD, University of Montpellier, Montpellier, France

several non-chemical control methods have been used for a relatively long time (Petit et al. 2011).

Among them, soil tillage is one of the main methods used to control spontaneous vegetation. The effects of this approach on the spontaneous species depend on the tools used, the soil tillage depth and the spontaneous species composition (Cordeau et al. 2017; Senarathne and Sangakkara 2009). Tillage is known to affect the seed depth distribution and abundance in soils (e.g. Colbach et al. 2000), while also destroying the growing vegetation and upsetting seed production, consequently affecting the viable soil seedbank. However, tillage can also churn old seeds up to the surface and promote the germination of seed cohorts near the surface (e.g. Colbach et al. 2014). The well-known false seedbank (seeds and rhizomes) in the topsoil (Ringselle et al. 2019).





Successive false seedbed operations promote seed germination and destroy seedlings, hence often decreasing the seedbank in the topsoil (Lamour and Lotz 2007; Ringselle et al. 2019). This technique is efficient for controlling of annual species (Lamour and Lotz 2007). It can also be used to control perennial species that reproduce via underground storage and proliferation organs by fragmenting and thus weakening them (Ringselle et al. 2019).

The use of cover crops is another method for controlling the development and composition of the spontaneous species community. There is considerable evidence on the short- and long-term effects of the sown crops and soil tillage on the spontaneous vegetation (Gunton et al. 2011). Cover crops also provide other beneficial services such as erosion mitigation and soil nitrogen enhancement via atmospheric fixation (Lu et al. 2000; Bàrberi and Mazzoncini 2001; Snapp et al. 2005; Damour et al. 2015). Cover crops can affect the development and growth of spontaneous species through competition for above- and below-ground resources or by providing a physical barrier to seed germination and seedling emergence (Teasdale et al. 2007; Médiène et al. 2011). The race for height and soil cover between weeds and cover crops that happens during the first weeks following plant emergence determines which plants will prevail over the others throughout the rest of their cycles (e.g. den Hollander et al. 2007). However, cover crop control of the spontaneous vegetation is often incomplete, leading to the development of mixed communities of the sown cover crop and some spontaneous species (Damour et al. 2018).

The combined use of cover crops with other methods with different mechanisms of action could enhance the control of spontaneous species. For example, tillage followed by sowing cover crops appears to be a promising solution. Some studies have shown that soil tillage is more effective than growing cover crops with regard to weed control under temperate climatic conditions (Bàrberi and Mazzoncini 2001). Indeed, under these conditions, when tillage is used to control spontaneous species, seasonal effects characterised by substantial temperature and humidity changes markedly decrease seed germination and plant growth due to the low temperatures and humidity (Cordeau et al. 2017; Fried et al. 2019). In the tropics, however, seasonal effects are negligible on temperature, light and humidity variations, which are less pronounced. To our knowledge, no studies have been conducted to test the impact of soil tillage followed by cover crop sowing in tropical conditions.

In banana cropping systems, cover crops can be used during fallows or banana crop cycles to provide services such as enhanced soil fertility or pest control (Damour et al. 2015). In these systems, managing the spontaneous vegetation during the fallow period will reduce the presence of these plants and

INRA

thus their impact on subsequent banana crop cycles, while decreasing herbicide treatments, which are commonly used and problematic in banana cropping systems (Risède et al. 2010). Some studies have focused on describing the potential effects of cover crops on the growth of spontaneous species in banana cropping systems based on their morphological and physiological features (e.g. Tardy et al. 2015). Yet to our knowledge, no studies have assessed the impact of combined management practices on the development of the spontaneous plant community, despite the fact that the findings could help farmers tailor their management practices according to their objectives (Snapp et al. 2005).

The main objective of this study was to gain insight into how soil tillage (here, the false seedbed technique) and cover crops impact spontaneous plant communities in fallows preceding banana cropping. We thus monitored spontaneous vegetation growth over 12 months in a fallowed field where a cover crop species (three species tested) was combined with one soil tillage modality (two modalities tested) and calculated the density of the spontaneous plant community, its species richness, diversity and composition.

### 2 Material and methods

### 2.1 Experimental site and design

The experiment was carried out in a banana plantation in Capesterre-Belle-Eau in Guadeloupe, French West Indies (16° 04' 39.1" N; 61° 36' 17.3" W; 285 m a.s.l.), from June 2018 to June 2019. This is the standard duration of a fallow in banana cropping systems in the French West Indies. The chosen field was representative of the agroclimatic conditions and of the crop management techniques of the banana production area in Guadeloupe. Over this 1-year study period, the cumulated precipitation was 4489 mm (annual 10-year mean of 4500 mm), with a mean daily temperature of 24.2 °C (range 21.5-27.5 °C) and a mean solar radiation of 422.30 MJ/m<sup>2</sup>/month. The soil was an andosol (FAO 2014). A 0.9-ha field that had been under fallow since October 2017 was selected for the experiment. The plant community was visually observed to be homogeneously distributed within the field. A split-plot design with five blocks was adopted. The field was divided into ten strips of four sub-plots each. Two factors were applied: the soil tillage modality (applied in strips, two modalities, one strip per modality and per block) (see Fig. 1 and SM1) and the sown cover crop species (applied on a sub-plot in strips, four modalities, sown at the same date in all sub-plots). The blocks were separated by 1-m-wide alleys, and each sub-plot was 196 m<sup>2</sup>. In the whole field, the vegetation was cut with a rotary slasher 7 weeks before the cover crops were sown to facilitate soil tillage. In five strips (one per block), the soil was ploughed to 40 cm depth with a



**Fig. 1** The two soil tillage modalities tested in a banana field under fallow in Guadeloupe (French West Indies). The false seedbed technique (modality M2: "with false seedbed") was applied with a seedbed cultivator (equipment on the left side), and coarse tillage not followed

spading machine the day before sowing to destroy and bury the current vegetation and improve the soil seedbed for the future crop (modality M1: "without false seedbed") (SM1). In the five other strips (one per block), the soil was tilled four times at different depths and with different tools (modality M2: "with false seedbed") (SM1). Seven weeks before the cover crop was sown, the soil was ploughed to 40 cm depth with a spading machine. Then, the superficial soil layer was fragmented three times to 5 cm depth to eliminate seedlings that had emerged and deplete the soil seedbank (false seedbed technique): (i) 5.5 weeks before sowing with a seedbed cultivator, (ii) 4 weeks before sowing with a power harrow and (iii) the day before sowing with a seedbed cultivator. In late June 2018, three tropical cover crop species were sown, each on a sub-plot per strip: a 50-50 mixture of Brachiaria decumbens (Stapf, 1919) and Brachiaria ruziziensis (Germ. & C.M.Evrard, 1953) (modality B), Crotalaria spectabilis (Roth, 1821) (modality C) and Pueraria phaseoloïdes (Roxb.) Benth. (modality P). The main species characteristics, sowing densities and sowing depths are presented in Table 1. In each strip, a fourth sub-plot was not sown and was used as a spontaneous vegetation control (modality T). M1T was the control without false seedbed, and M2T was the control with false seedbed. The eight treatments were named according to the abbreviations of the two factors. Note that caterpillar attacks on C. spectabilis leaves were observed between the 4th and the 5th months post-sowing without being quantified, and this may have affected plant survival.

by the false seedbed technique (modality M1: "without false seedbed") was applied with a rotary spading machine (equipment on the right side). Photograph by Diane Rakotomanga

### 2.2 Measurements

In each sub-plot, three 25 cm  $\times$  25 cm permanent quadrats were installed far from the borders and marked with construction tape on stakes. Each month from July 2018 to June 2019 (dates t1 to t12, with t3 and t6 missing values due to bad weather, thus preventing us from obtaining measurements), the spontaneous species growing in the quadrats were identified and the number of individuals of each species was counted. The number of cover crop individuals was also counted. For each quadrat, at each date, the density of each species  $(d_i)$ for species *i*) was calculated as the number of individuals of that species per m<sup>2</sup>. The total densities of all species and of spontaneous species ( $d_{tot}$  and  $d_{spont}$ , respectively) were calculated as the sum of the densities of all species and spontaneous species, respectively. The relative density of each species  $(p_i)$ for species *i*) was calculated as  $p_i = d_i / d_{tot}$ . The species richness of the spontaneous plant community (S) was calculated as the number of spontaneous species counted in the quadrat. Shannon's diversity index for the spontaneous plant community (H') was calculated as follows:

 $H' = -\sum_{i=1}^{S} p_i \log_2 p_i,$ 

with i being the spontaneous species i

The frequency of occurrence of each species ( $F_i$  for species *i*) per treatment was also calculated as the number of quadrats of the treatment where a species occurred divided by the total number of sampled quadrats of the treatment (Fried et al. 2020).



Code of the modality	Abbreviation	Full name	Family	Cycle	Growth habit	Sowing density (kg/ha)	Sowing depth (cm)
В	BD	Brachiaria decumbens (Stapf.1919)	Poaceae	Perennial	Semi-erect	10	3
	BR	Brachiaria ruziziensis (Germ. & C.M.Evrard. 1953)	Poaceae	Perennial	Semi-erect	10	3
С	CS	Crotalaria spectabilis (Roth. 1821)	Fabaceae	Annual (6 months)	Erect	30	3
Р	PPh	Pueraria phaseoloïdes (Roxb.) Benth.	Fabaceae	Perennial	Twining	30	3

Table 1 Main cover crop characteristics, sowing densities and depth used in the experiment

Modality B is a 50–50 mixture of two species

### 2.3 Data analysis

All statistical analyses were performed with R software (version 3.5.3) (R Core Team 2018). The effects of cover crop species and soil tillage modalities on  $d_{\text{spont}}$ , species richness and Shannon's diversity index were tested at each date with a nested ANOVA with the cover crop species factor nested in the soil tillage modality factor. Tukey post hoc tests were used to assess differences between the treatment means. The effects of the monitoring date on  $d_{\text{spont}}$ , species richness and Shannon's diversity index were tested, for each treatment, with a one-way ANOVA, followed by a Tukey post hoc test. Beforehand, the effects of blocks and sub-plots were tested with ANOVA (results not presented). They were not significant and thus not included in subsequent models. The relative difference between the densities of spontaneous species observed in a treatment and the one observed in the control treatment M1T (RelDiff  $_{d_{snont}}$ ) was calculated as

$$\text{RelDiff}_{d_{\text{spont}}} = 100 \cdot \left[ d_{\text{spont}}(\text{trt}_i) - d_{\text{spont}}(\text{M1T}) \right] / d_{\text{spont}}(\text{M1T})$$

with trt<sub>*i*</sub> in {M1B, M2B, M1P, M2P, M1C, M2C}.

The relative difference between the species richness observed in a treatment and the one observed in the control treatment M1T (RelDiff<sub>S</sub>) was calculated similarly. Lastly, a principal component analysis (PCA) was performed on the densities of the 13 spontaneous species whose maximal frequency of occurrence was over 80% (combined dates and treatments). For each species, the contribution of variables to the first two axes was also assessed. Differences in the PCA axis 1 and 2 scores among treatments were tested by one-way ANOVA, followed by a Tukey HSD post hoc test.

### 3 Results

# 3.1 Dynamics of the density, species richness and diversity of the spontaneous plant community over 12 months

The spontaneous species community dynamics were contrasted over the 1-year monitoring period, depending on the treatment

Deringer

INRAO

(Table 2). The first 2 months post-sowing the spontaneous plant densities in all treatments decreased significantly. Thereafter, the density of spontaneous species ( $d_{spont}$ ) of communities without false seedbed associated with *C. spectabilis* (M1C) and the control (M1T) varied slightly but not significantly. The  $d_{spont}$  of communities with false seedbed associated with *C. spectabilis* (M2C) and the control (M2T) increased until 7 months post-sowing and then stabilised. Conversely, the  $d_{spont}$  of spontaneous species associated with *P. phaseoloides* (P) and the *Brachiaria* mixture (B) decreased significantly until 4 months post-sowing and then stabilised. The species richness (S) and Shannon's diversity index (H') presented the same variation pattern as  $d_{spont}$  (Table 2).

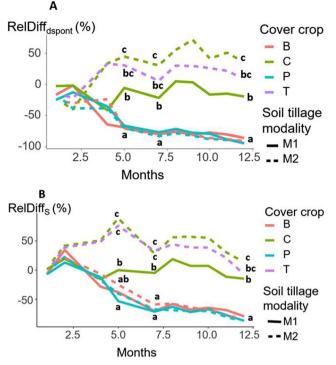
# **3.2 Effects of treatments on the density, species richness and diversity of the spontaneous species community**

The sown cover crop species, soil tillage modality and their interaction affected the species density, richness and diversity (Shannon's diversity index) (Fig. 2, SM2). For these three variables, differences between treatments appeared 2 months post-sowing. At this date, the interaction between the sown cover crop species and the soil tillage modality was significant. From 4 months post-sowing the interaction remained significant and the sown cover crop species was always highly significant thereafter.

From 2 months post-sowing the densities of spontaneous species ( $d_{spont}$ ) associated with *P. phaseoloïdes* (P) and the *Brachiaria* mixture (B) were lower than those in the control M1T (about 80% and 75%, respectively, 12 months post-sowing). On the contrary, the densities of spontaneous species ( $d_{spont}$ ) with false seedbed associated with *C. spectabilis* (M2C) and the control (M2T) were higher than those in the control M1T (about 50 and 40%, respectively, 12 months post-sowing) (Fig. 2A). From 5 months post-sowing the relative difference in  $d_{spont}$  with the control M1T (RelD*iff*  $d_{spont}$ ) for treatments associated with *P. phaseoloïdes* (P) and the *Brachiaria* mixture (B) was significantly lower than that observed in the other treatments. From 7 months post sowing, RelD*iff*  $d_{spont}$  was significantly lower for spontaneous

Table 2 correspo	<u> </u>	(d <sub>spont</sub> ), species ricl	hness (S) and Shann	Mean densities ( $d_{\text{spont}}$ ), species richness (S) and Shannon's diversity index (H) of the spontaneous species communities in each treatment and at each measurement date (ti), followed by the dding standard deviation	(H') of the spontan	eous species comm	unities in each treat	ment and at each n	neasurement date (ti	), followed by the
	t1	t2	t4	t5	t7	t8	t9	t10	t11	t12
$d_{\rm spont}$ M1R	250 7 + 123 1 c	173 6 + 100 2 b	27 7 + 34 4 a	45 6 + 27 5 a	45 1 + 34 8 a	411+754a	33 0 + 77 6 a	41 3 + 30 7 a	371+357a	e 996 + 86
MIC	$304.5 \pm 84.0$ b	$173.6 \pm 91.2$ a	96.8 ± 45.2 a		$181.3 \pm 99.0$ a	$185.9 \pm 57.2$ a	$185.6 \pm 80.2$ a			
MIP	235.4 ± 70.0 d		$107.1 \pm 67.0 \ bc$	$50.2 \pm 32.1$ ab	52.3 ± 37.9 ab		40.0 ± 24.8 a	38.8 ± 26.8 a	22.2 ± 20.6 a	
MIT	$313.3 \pm 75.1 \text{ c}$	$177.4 \pm 78.5 \text{ ab}$	$165.6 \pm 55.3 \text{ ab}$	$150.7 \pm 46.8$ a	$231.7 \pm 66.8 \text{ b}$	$177.5 \pm 43.2$ ab	$180.5 \pm 69.5 \text{ ab}$	$192.0 \pm 58.7 \text{ ab}$	$194.9 \pm 45.8 \text{ ab}$	$206.4 \pm 78.7 \text{ ab}$
M2B	$228.5 \pm 76.7 c$	$123.7 \pm 68.5 \text{ b}$	$125.6 \pm 104.5 \text{ b}$	43.2 ± 32.3 a	$45.6 \pm 36.3 \text{ a}$	$41.1 \pm 29.7 a$	$33.6 \pm 22.1 \text{ a}$	$26.4 \pm 19.1$ a	22.9 ± 24.3 a	$16.3 \pm 16.1 \text{ a}$
M2C	$241.3 \pm 73.5 \text{ b}$	$106.1 \pm 37.5 a$	$221.1 \pm 94.9 \text{ ab}$	$217.9 \pm 133.9$ ab	$302.1 \pm 132.6 \text{ b}$	$276.5 \pm 67.6 \text{ b}$	$309.9 \pm 102.7 \text{ b}$	$272.5 \pm 81.1 \text{ b}$	$294.7 \pm 105.6 \ b$	$279.2 \pm 120.3 \text{ b}$
M2P	$266.4\pm68.5~\mathrm{c}$	$109.3 \pm 63.8 \text{ b}$	$101.1 \pm 71.8 \ b$	43.7 ± 32.6 a	$38.4\pm30.8~a$	$36.8\pm22.8$ a	$30.7 \pm 13.2$ a	$23.5\pm13.5~\mathrm{a}$	$20.8\pm18.0~a$	$16.0 \pm 13.4 \text{ a}$
M2T	$269.3 \pm 90.1 \text{ b}$	$137.1 \pm 59.1$ a	$219.5\pm55.0~ab$	$196.0 \pm 102.8$ ab	$244.3 \pm 112.3 b$	$230.4 \pm 69.5 \text{ ab}$	$232.5 \pm 64.8 \text{ ab}$	$242.1 \pm 79.5 \text{ b}$	$236.5 \pm 78.5 \text{ b}$	$222.7 \pm 106.2$ ab
S										
MIB	$5.6\pm1.8~b$	$5.9 \pm 1.6 \text{ b}$	$3.4 \pm 2.3 \ a$	$2.1 \pm 1.3$ a	$1.9 \pm 0.9 a$	$2.3 \pm 1.2 a$	$2.1\pm0.9~a$	$2.1 \pm 1.1 a$	$2.3 \pm 1.7$ a	2. ± 1.2 a
MIC	$6.1 \pm 1.3$ ad	$5.3 \pm 1.9$ ac	$4.0 \pm 1.6 \text{ ab}$	$3.5 \pm 1.1$ a	$6.1 \pm 2.7$ ad	$6.6 \pm 2.6$ bcd	$7.1 \pm 3.1 \text{ cd}$	$6.5 \pm 2.6$ bcd	$6.7 \pm 3.4$ bcd	8.6 ± 3.5 d
MIP	$5.5 \pm 1.2 \text{ b}$	$5.0 \pm 1.9 \text{ b}$	$4.1\pm2.7~b$	$1.6\pm0.8~a$	$1.9 \pm 1.4 a$	$2.1 \pm 1.3$ a	$1.9 \pm 1.1 \text{ a}$	$2.0 \pm 1.4 a$	$1.8 \pm 1.5 a$	$1.4 \pm 0.8 a$
MIT	$6.0 \pm 1.3$ bc	$4.4 \pm 1.7 ab$	$4.7 \pm 1.6 \text{ ab}$	$3.5 \pm 1.1$ a	$6.5 \pm 1.9 \text{ bc}$	$5.6 \pm 1.6 \text{ ac}$	$6.6 \pm 2.8 \text{ bc}$	$6.1 \pm 2.0 \text{ bcd}$	$7.5 \pm 2.4 \text{ c}$	$10.1 \pm 2.7 \text{ d}$
M2B	$6.4 \pm 1.1 \text{ c}$	$5.4 \pm 1.1$ bc	$4.4 \pm 2.4 \text{ b}$	$2.6 \pm 1.3$ a	$2.7 \pm 1.5 a$	$2.4 \pm 1.1$ a	$2.4 \pm 1.0 a$	$2.0 \pm 1.0 a$	$1.6 \pm 0.9 a$	$1.9 \pm 1.3 a$
M2C	$6.1\pm0.8~a$	$6.3\pm1.7~a$	$7.1 \pm 1.8 \text{ ab}$	$6.5 \pm 2.5$ a	$8.3 \pm 2.3$ abc	$8.7 \pm 2.9 \text{ ad}$	$10.3 \pm 2.0 \text{ cd}$	$9.4 \pm 2.0$ bd	$9.8 \pm 2.6$ bd	11.3 ± 3.3 d
M2P	$5.7\pm0.8$ c	$5.4 \pm 1.4 \text{ c}$	$3.9 \pm 1.7 b$	$2.1 \pm 1.7 a$	$2.1 \pm 1.6$ a	$1.7\pm0.9~a$	$1.9 \pm 0.7 a$	$1.8\pm0.8~a$	$1.5\pm0.7$ a	$1.2\pm0.7~\mathrm{a}$
M2T	$5.8\pm0.8~a$	$6.1 \pm 1.3 ab$	$7.0 \pm 1.6 \text{ ac}$	$6.1 \pm 1.9 \text{ ab}$	$8.6 \pm 1.7 \text{ cd}$	$8.1 \pm 1.5 bcd$	$9.1 \pm 1.9 \text{ cd}$	$8.4 \pm 1.7 \text{ cd}$	$9.1 \pm 3.0 \text{ cd}$	$9.3 \pm 2.8 \text{ d}$
$H^{\prime}$										
MIB	$1.8\pm1.8~\mathrm{b}$	$1.9 \pm 1.6 \text{ b}$	$1.3 \pm 2.3 \text{ ab}$	$0.7 \pm 1.3$ a	$0.6 \pm 0.9 a$	$0.8 \pm 1.2 \ a$	$0.8 \pm 0.9 \ a$	$0.7 \pm 1.1$ a	$0.8 \pm 1.7 \ a$	$0.7 \pm 1.2 \text{ a}$
M1C	$1.8 \pm 1.3$ ac	$1.6 \pm 1.9 \text{ ab}$	$1.5 \pm 1.6 \text{ ab}$	1.3 ± 1.1 a	$1.9 \pm 2.7$ ac	$2.2\pm2.6\ c$	$2.2 \pm 3.1 \text{ c}$	$2.1 \pm 2.6 \text{ bc}$	$2.1 \pm 3.4 \text{ bc}$	$2.4\pm3.5~c$
M1P	$1.9 \pm 1.2 \text{ c}$	$1.6 \pm 1.9 c$	$1.3 \pm 2.7 \text{ bc}$	$0.4\pm0.8~a$	$0.5\pm1.4$ a	$0.7 \pm 1.3$ ab	$0.6 \pm 1.1$ ab	$0.6 \pm 1.4 \text{ ab}$	$0.6 \pm 1.5 \text{ ab}$	$0.3\pm0.8~a$
M1T	$1.9 \pm 1.3  bd$	$1.5 \pm 1.7 ab$	$1.6 \pm 1.6 \text{ bc}$	$1.1 \pm 1.1$ a	$2.0 \pm 1.9 \text{ cd}$	$1.9 \pm 1.6$ bd	$2.1 \pm 2.8 \text{ cd}$	$2.0 \pm 2.0$ bd	$2.2 \pm 2.4 \text{ de}$	$2.7 \pm 2.7 \text{ e}$
M2B	$1.9 \pm 1.1 \text{ c}$	$1.8 \pm 1.1 \text{ c}$	$1.7 \pm 2.4 \text{ bc}$	$1.0 \pm 1.3$ ab	$1.0 \pm 1.5$ ab	$0.9 \pm 1.1$ ab	$1.0 \pm 1.0$ ab	$0.7\pm1.0~\mathrm{a}$	$0.5\pm0.9~a$	$0.7 \pm 1.3$ a
M2C	$1.9\pm0.8$ a	$2.2 \pm 1.7 ab$	$2.4 \pm 1.8 \text{ ac}$	$2.1 \pm 2.5 \text{ ab}$	$2.4 \pm 2.3 \ bc$	$2.4 \pm 2.9 \text{ bc}$	$2.7 \pm 2.0 \text{ c}$	$2.7 \pm 2.0 \text{ c}$	$2.7 \pm 2.6 \text{ c}$	$2.8\pm3.3~c$
M2P	$1.7\pm0.8~\mathrm{b}$	$2.0\pm1.4~b$	$1.5 \pm 1.7 \text{ b}$	$0.6\pm1.7~\mathrm{a}$	$0.7\pm1.6~a$	$0.5 \pm 0.9 \text{ a}$	$0.7\pm0.7$ a	$0.6 \pm 0.8$ a	$0.5 \pm 0.7$ a	$0.3 \pm 0.7$ a
M2T	$1.7\pm0.8~a$	$2.1 \pm 1.3$ ac	$2.4 \pm 1.6$ bcd	$2.0 \pm 1.9 \text{ ab}$	$2.5 \pm 1.7 \text{ cd}$	$2.4 \pm 1.5 \text{ bcd}$	2.7 ± 1.9 d	$2.5 \pm 1.7 \text{ cd}$	$2.5 \pm 3.0 \text{ cd}$	$2.5 \pm 2.8 \text{ cd}$
Differe	int letters indicate a	significant differentes est	Different letters indicate a significant difference between dates for micture. MIC. without false seadbad 4. Contraine manufallie. MI		ey HSD test with $\alpha$	(= 0.05); all tests w	one treatment (Tukey HSD test with $\alpha = 0.05$ ); all tests were highly significant ( $p < 0.001$ ). M1B: without false seedbed + <i>Brachiaria</i> D. without false seedbed + <i>Dimension thresolvicher</i> M1T: without false seedbed + constructions wavefordow. M2B: with false seedbed +	It $(p < 0.001)$ . M11	B: without false see	lbed + Brachiaria th false seadhed ±
Brachic	e; MIUC: WILDOUL 1a <i>aria</i> mixture; M2C	ise seedded + <i>Crott</i> : with false seedbed	mixture; MIC: Without tailse seedbed + Crotataria spectabilis; MI Brachtaria mixture; M2C: with false seedbed + Crotalaria spectal		abea + <i>Fueraria p</i> . se seedbed + <i>Puera</i>	naseototaes; 14111: tria phaseoloïdes; N	r: without tailse seedbed + <i>Fueraria phaseoloides</i> ; M111. Without tailse seedbed + spontaneous vegetation; I alis; M2P: with false seedbed + <i>Pueraria phaseoloides</i> ; M2T: with false seedbed + spontaneous vegetation	ed + spontaneous v lbed + spontaneous	egetation; MZB: WI	un talse seedbed +

🖄 Springer



**Fig. 2** Relative difference between the densities of spontaneous species observed in a treatment and the one observed in the control treatment M1T (RelD*iff*  $d_{upout}$ ) (**A**) and relative difference between the species richness observed in a treatment and that observed in the control treatment M1T (RelDiff<sub>S</sub>) (**B**) in the eight treatments over 12 months for the 120 quadrats (8 × 15). Different letters indicate a significant difference between treatments (Tukey HSD test with  $\alpha = 0.05$ ). M1B: without false seedbed + *Brachiaria* mixture; M1C: without false seedbed + *Crotalaria spectabilis*; M1P: without false seedbed + *Pueraria phaseoloïdes*; M1T: without false seedbed + spontaneous vegetation; M2B: with false seedbed + *Brachiaria* mixture; M2C: with false seedbed + *Pueraria phaseoloïdes*; M2T: with false seedbed + spontaneous vegetation

species associated with *C. spectabilis* (C) without false seedbed (M1C) than with false seedbed (M2C).  $d_{\text{spont}}$  in M2C stayed far higher than that in the control M1T (30 to 50%), while  $d_{\text{spont}}$  in M1C stayed slightly lower than that in the control M1T.

The species richness (S) of spontaneous species generally showed the same trends as  $d_{\text{spont}}$ , except that differences in S between M2C and M2T compared to the other treatments where more pronounced.

Shannon's diversity index (H') was significantly lower for treatments with *Brachiaria* sp. (B) and *P. phaseoloides* (P) than that for the treatments with *C. spectabilis* (C) and the control (T) 7 months and 12 months post-sowing (SM2).

#### 3.3 Community composition 12 months post-sowing

The cover crop species and soil tillage modality affected the composition of the monitored communities after 12 months (Figs. 3 and 4). All of the identified spontaneous species,

🖉 Springer

INRA

along with their name abbreviations, are presented in the Supplementary Material (SM3).

The first two PCA axes constructed on the densities of the 13 most frequent spontaneous species explained 41.6% of the total variance. *Bidens alba* (BA), *Ludwigia octovalvis* (LO) and *Fimbristylis dichotoma* (FD) contributed the most to axis 1 (5.64%, 12.75% and 11.65%, respectively). *Mikania micrantha* (MM), *Lindernia crustacea* (LC), *Kyllinga erecta* (KE) and *Filicophyta* sp. (F) contributed the most to axis 2 (20.25%, 16.15%, 15.52% and 15.30%, respectively). *Fimbristylis dichotoma* (FD) and *Peperomia pellucida* (PPe) densities appeared to be closely correlated according to their positions on the ACP (Fig. 3A, close arrows). The same results were obtained for *Digitaria ciliaris* (DCi), *Mecardonia procumbens* (MP) and *Ludwigia octovalvis* (LO) on the one hand, and *Filicophyta* sp. (F) and *Kyllinga erecta* (KE) on the other.

Two groups of treatments were clearly distinguished on the PCA: treatments with *Brachiaria* sp. (B) and *P. phaseoloïdes* (P) on the left side and treatments with *C. spectabilis* (C) and the control (T) on the right. These two groups were significantly different on both axes (Tukey test performed on the axis 1 and 2 coordinates, Fig. 3C and D, p < 0.05). Moreover, M2C did not overlap with M1C and M1T on both axes (Fig. 3D).

No species were specifically present in one treatment, but two spontaneous species succeeded in becoming established in every treatment, i.e. Bidens alba (BA) and Mikania micrantha (MM) (Fig. 4). Communities associated with P. phaseoloïdes (P) included only a few species (see species richness results). Only the two ubiquitous species Bidens alba (BA) and Mikania micrantha (MM) were found in M1P, while Digitaria ciliaris (DCi) was also found in M2P. Spontaneous plant communities in treatments associated with Brachiaria sp. (B) were slightly richer, with Bidens alba (BA), Mikania micrantha (MM), Digitaria ciliaris (DCi), Fimbristylis dichotoma (FD) and Ludwigia octovalvis (LO) in M1B and the same species as well as Hedyotis lancifolia (HL) and Mecardonia procumbens (MP) in M2B. Several spontaneous species were only present in treatments with C. spectabilis (C) and the control (T).

### **4 Discussion**

The results of our study were obtained over a 1-year period on a single site. However, considering that during this year there were no extreme climatic events, that rainfall was similar to the 10-year annual mean and that the chosen field and its management were representative of banana crop fields in Guadeloupe, we consider that our results would be transposable to other years and fields with similar characteristics.

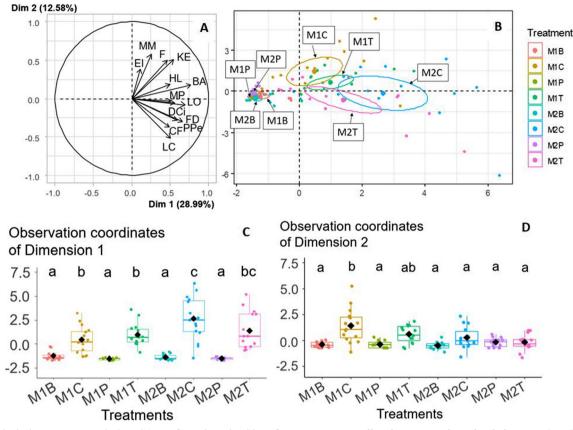


Fig. 3 Principal component analysis (PCA) performed on densities of the 13 most frequent spontaneous species of the communities monitored at 12 months post-sowing: A correlation circle between variables on the first two axes; B observations in the first two axes with representation of the treatments (ellipses and colours); Observation coordinates in dimension 1 (C) and dimension 2 (D). EI: *Eleusine indica*; MM: *Mikania micrantha*; F: *Filicophyta* sp.; KE: *Kyllinga erecta*; HL: Hedyotis lancifolia; BA: *Bidens alba*; MP: *Mecardonia procumbens*; LO: *Ludwigia octovalvis*; DCi: *Digitaria ciliaris*; PPe:

# 4.1 The cover crop affects the density, richness and diversity of the spontaneous vegetation

In this study, we showed that the spontaneous plant communities had contrasted dynamics over 1 year, depending on the sown cover crop species. However, the first 2 months post-sowing, the spontaneous plant densities in all treatments decreased markedly. This could possibly be explained by a high level of insect predation on the seedlings just after emergence (Petit et al. 2011), or even mollusc predation, which we observed during the study. After these 2 months, a marked difference was observed between communities associated with P. phaseoloïdes (P) and Brachiaria sp. (B) on the one hand, and communities associated with C. spectabilis (C) and the control (T) on the other, i.e. the species richness and diversity decreased with time in communities associated with P. phaseoloides (P) and Brachiaria sp. (B), whereas these metrics increased with time in communities associated with C. spectabilis (C) and the control (T). These results suggest differences in the abilities of these cover crop species to compete

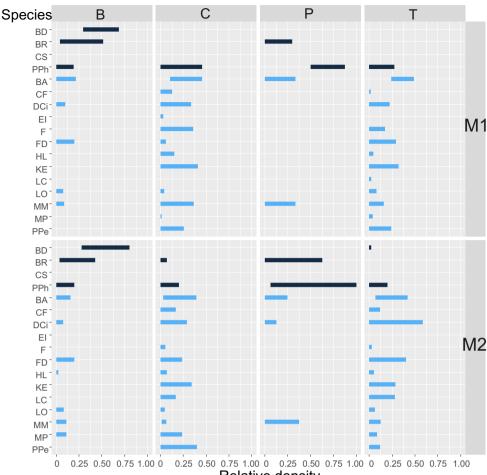
Peperomia pellucida; FD: Fimbristylis dichotoma; CF: Cardamine flexuosa; LC: Lindernia crustacea; M1B: without false seedbed + Brachiaria mixture; M1C: without false seedbed + Crotalaria spectabilis; M1P: without false seedbed + Pueraria phaseoloïdes; M1T: without false seedbed + spontaneous vegetation; M2B: with false seedbed + Brachiaria mixture; M2C: with false seedbed + Crotalaria spectabilis; M2P: with false seedbed + Pueraria phaseoloïdes; M2T: with false seedbed + spontaneous vegetation

for resources (mainly light) or to develop a physical barrier to seed germination and seedling emergence, i.e. the two main processes by which cover crops can limit the growth of neighbouring species (Teasdale et al. 2007; Médiène et al. 2011).

Indeed, *Pueraria phaseoloïdes* (PPh) is a perennial twining liana with large leaves that climb on other species and stifle them by occupying the space and reducing their access to light. This species appears to be a particularly strong competitor, according to the very low species richness and density of the spontaneous plant community we observed. Tardy et al. (2015) showed that *Pueraria phaseoloïdes* invests carbohydrates in support and light acquisition structures (stems and leaves, respectively), while rapidly producing substantial biomass. These growth characteristics could be associated with efficient light interception and acquisition to the detriment of the neighbouring plants. In our experiment, the only species that succeeded to grow with *Pueraria phaseoloïdes* were (i) the liana *Mikania micrantha* (MM), which has the same growth habit as *Pueraria phaseoloïdes* 



Fig. 4 Relative densities of the 13 most frequent spontaneous species and of the cover crop species at 12 months post-sowing. Data correspond to the densities measured on the quadrats of each treatment. BD: Brachiaria decumbens: BR: Brachiaria ruziziensis; CS: Crotalaria spectabilis; PPh: Pueraria phaseoloïdes; EI: Eleusine indica; MM: Mikania micrantha; F: Filicophyta; KE: Kyllinga erecta; HL: Hedyotis lancifolia; BA: Bidens alba; MP: Mecardonia procumbens; LO: Ludwigia octovalvis; DCi: Digitaria ciliaris; PPe: Peperomia pellucida; FD: Fimbristylis dichotoma; CF: Cardamine flexuosa; LC: Lindernia crustacea: B: modality with Brachiaria mixture; C: modality with Crotalaria spectabilis; P: modality with Pueraria phaseoloïdes; T: modality with control; M1: without false seedbed; M2: with false seedbed; in dark blue: the four cover crops; in pale blue: the 13 most common spontaneous species of the study



Relative density

and seems to grow more quickly, and (ii) and the erect species *Bidens alba* (BA), whose rapid growth may allow it to rise above the *Pueraria phaseoloïdes* cover, thereby avoiding being stifled by this species.

Brachiaria decumbens (BD) and Brachiaria ruziziensis (BR) are perennial semi-erect species with a high growth rate and biomass that is invested in expanding the leaf area (Damour et al. 2016; Tardy et al. 2015). They tend to maximise light interception with light acquisition organs (Tardy et al. 2015). They also densely cover the soil by tillering and emitting stolons, which consequently prevent seed germination and emergence. Indeed, under Brachiaria decumbens and Brachiaria ruziziensis cover, we observed high humidity and low incident light, which could have prevented seed germination and growth. It likely also prevented small spontaneous species from colonising the area, which was the case for Cardamine flexuosa (CF) and Mecardonia procumbens (MP) (Fig. 4) and could explain the low species richness and diversity observed in these communities. However, other species succeeded in growing within Brachiaria decumbens and Brachiaria ruziziensis cover. The liana Mikania micrantha (MM) had likely grown by twinning on the supporting Brachiaria decumbens and Brachiaria *ruziziensis* plants. The erect species *Bidens alba* (BA) likely succeeded in growing because of its tall habit, which enabled it to pierce through the *Brachiaria* cover.

*Crotalaria spectabilis* (CS) is an annual erect plant with a low crown width (Tardy et al. 2015). This low crown width was probably responsible for poor soil shading and poor space and light competition. This poor competitive ability, in addition to the caterpillar attack that occurred between the 4th and the 5th months post-sowing which probably contributed to the decrease in the *C. spectabilis* population, may explain the marked development of spontaneous species in these treatments. Moreover, *Crotalaria spectabilis* is an annual species with a 6-month cycle. This may explain why communities in treatments associated with *C. spectabilis* (C) thereafter tended to have the same dynamics as communities in the control treatment (T) without cover crops.

## 4.2 Soil tillage affects spontaneous plant communities depending on the cover crop sown

Our results showed that the soil tillage operations affected the spontaneous species density and the species richness of the



spontaneous plant community only when the *Crotalaria spectabilis* cover crop had been sown. These results could probably be explained by an effect of soil tillage on the seedbank, which may germinate and grow if the cover crop competition intensity is moderate.

Without false seedbed (modality M1), the soil was ploughed with a spading machine at 40 cm depth 1 day before sowing. Therefore, mixed spontaneous species seeds were likely present within the 0-40-cm soil layer, i.e. a depth at which they would be unable to germinate because of their small size. When the false seedbed technique was used (M2), the 5-cm upper soil layer was fragmented three times before sowing at 5 cm depth with a seedbed cultivator and a power harrow. These supplementary soil operations took place after seedling emergence so as to destroy them and prevent further growth, as well as to deplete the spontaneous species soil seedbank (seeds and rhizomes) (Ringselle et al. 2019). Our results showed that spontaneous species densities either did not differ between the two soil tillage modalities (treatments associated with P. phaseoloïdes (P) and Brachiaria sp. (B) and the control (T)), or they were higher with the false seedbed technique than without (treatments associated with C. spectabilis (C)). This suggests that the spontaneous species soil seedbank was not depleted sufficiently and that seed germination was favoured. Indeed, seed germination could have been favoured in treatments with false seedbed (M2) via improvement in the ground-seed contact (Senarathne and Sangakkara 2009) and consequently also in seed moistening. Otherwise, seed germination and seedling emergence could have been promoted with false seedbed because seeds were placed in a more fragmented superficial soil layer with better access to light and oxygen. Moreover, the false seedbed technique may have failed to destroy rhizomes present in the superficial layers (Lamour and Lotz 2007; Ringselle et al. 2019). Conversely, the successive soil operations may have resulted in these rhizomes being chopped into small pieces and disseminated across the sub-plots. We observed more rhizome species like Fimbristylis dichotoma (FD) and Digitaria ciliaris (DCi) with false seedbed (M2) than without false seedbed (M1) (Fig. 4). Consequently, we concluded that successive passages of the power harrow and seedbed cultivator were probably responsible for the higher density and richness of spontaneous species we observed with the false seedbed modality (M2) because the soil seedbank depletion by repeated destruction of emerging seedlings did not offset the favourable effects on seed germination and weed dissemination by rhizome fragmentation.

Finally, this is the first study to show that, under a tropical climate, sown cover crop species combined with different soil tillage modalities have strong impacts on the species density, richness and diversity of the spontaneous plant community over a 1-year period.

### 4.3 Taxonomic diversity vs functional diversity

In this experimental study, we tested three cover crop species and revealed their contrasted effects. However, a broad range of cover crop species may be used in agrosystems and testing all of them is not possible. Above we discussed that the plant growth habit (erect, semi-erect, liana) and some cover crop morphological characteristics reported in the literature could be a useful basis for discussing the competitive ability of species and the effects of cover crops on the spontaneous plant community composition and density. A trait-based approach, as described in Duarte et al. (1995) and Garnier et al. (2018), could be used to describe species (cover crops and spontaneous species) according to traits related to competition relationships, to gain in genericity and save research time. Among these traits, the flowering dates of the cover crops and spontaneous species (Storkey 2006), their heights (Storkey 2006) (proxy of light availability for neighbours) and biomass (Bàrberi and Mazzoncini 2001; Senarathne and Sangakkara 2009) (proxy of competition intensity) would warrant measurement, in addition to the growth habit.

The analysis of such traits could also help measure the functional diversity of the communities and not only their species diversity. Functional diversity would have the advantage of shedding light on the mechanisms by which cover crops affect the spontaneous plant community (Díaz et al. 2007; Petchey and Gaston 2006). The species diversity we observed in some treatments (e.g. treatments associated with *C. spectabilis* (C) with high diversity or slightly higher richness, although not significant, in treatments associated with *Brachiaria* sp. (B) compared to treatments associated with *P. phaseoloïdes* (P)) may have hidden functional redundancy, e.g. a similar response of some spontaneous species to a treatment.

### 4.4 Management strategies to control spontaneous species

We showed that two management strategies (the use of cover crops and soil tillage) could partially control spontaneous species and that combining them could, in some conditions, improve this control. Other strategies like regular vegetation cutting with a rotary slasher could be combined to further enhance control.

Some of the spontaneous species observed in this study seemed be very hard to control. In tropical areas, lianas represent a major functional type, which could be responsible for serious economic problems in crop production (Wyka et al. 2013). This is the case with regard to *Mikania micrantha* (MM), a liana that can twine on banana plants and slow down its growth, particularly young banana plants. This liana did not respond well to the management practices we tested in this



study. Other alternatives would be required to control them, such as manual detouring of each banana plant, which is a long and fastidious task.

Beyond the efficiency of these strategies, the targeted level of control of spontaneous species should be defined, depending on the farmer's objectives and on the nuisance these species generate. Maintaining some diversity in the field could improve the delivery of ecosystem services and the system resilience (Jackson et al. 2010; Gaba et al. 2014). For example, maintaining a certain level of spontaneous species in a diversified community may provide shelter for auxiliary fauna to help control pests or food for pollinator insects (Gaba et al. 2014). In our study, *Bidens alba* (BA) was present in all treatments—this species produces flowers year-round and may be a substantial pollination service.

### **5** Conclusion

Here, we showed for the first time that, under a tropical climate, sown cover crop species combined with different soil tillage modalities had substantial impacts on the species density, richness and diversity of the spontaneous plant community over 1 year, during the fallow period in banana cropping systems. This impact differed according to the cover crop and the soil tillage modality, probably related to differences in the competitive ability of the cover crop used and, to a lesser extent, to soil tillage effects on the soil seedbank. A functional approach could be used in further studies to gain further insight into the mechanisms by which plants compete and become established in the field. Ultimately, it could help farmers in their weed control strategies by boosting their understanding of the associated mechanisms.

**Supplementary Information** The online version contains supplementary material available at https://doi.org/10.1007/s13593-021-00692-1.

Acknowledgements We thank Steewy Lakhia and Jérôme Saint-Marc, technicians at the CIRAD research station in Guadeloupe, for their help in setting up the field experiments. We also thank Nathalie Colbach and the two anonymous reviewers for their constructive comments that helped us improve this manuscript.

Author contribution Conceptualization, D.R, M.D. and G.D.; Methodology, D.R. and G.D.; Resources, M.D.; Investigation, D.R and M.L.; Data curation, D.R.; Formal analysis, D.R.; Writing of the original draft, D.R., M.D and G.D.; Funding acquisition, M.D. and G.D.; Supervision, M.D. and G.D.

Funding This work was funded by the European Regional Development Fund for the RIVAGE project.

**Data availability** The datasets generated during and/or analysed during the current study are only available from the corresponding author on reasonable request.

Code availability Not applicable

#### Declarations

Ethics approval and consent to participate Not applicable

Consent for publication Not applicable

Conflict of interest The authors declare no competing interests.

### References

- Bårberi P, Mazzoncini M (2001) Changes in weed community composition as influenced by cover crop and management system in continuous corn. Weed Sci 49:491–499. https://doi.org/10.1614/0043-1745(2001)049[0491:CIWCCA]2.0.CO;2
- Colbach N, Roger-Estrade J, Chauvel B, Caneill J (2000) Modelling vertical and lateral seed bank movements during mouldboard ploughing. Eur J Agron 13:111–124. https://doi.org/10.1016/ S1161-0301(00)00069-1
- Colbach N, Busset H, Roger-Estrade J, Caneill J (2014) Predictive modelling of weed seed movement in response to superficial tillage tools. Soil Tillage Res 138:1–8. https://doi.org/10.1016/j.still.2013.12.002
- Cordeau S, Smith RG, Gallandt ER, Brown B, Salon P, DiTommaso A, Ryan MR (2017) How do weeds differ in their response to the timing of tillage? A study of 61 species across the northeastern United States. Ann Appl Biol 171:340–352. https://doi.org/10. 1111/aab.12377
- Damour G, Garnier E, Navas ML, Dorel M, Risède J-M (2015) Using functional traits to assess the services provided by cover plants. In: Advances in agronomy. Elsevier, Amsterdam, pp 81–133. https:// doi.org/10.1016/bs.agron.2015.06.004
- Damour, G., Quoc, H.T., Tardy, F., Meynard, C., Dorel, M., 2018. A trait-based approach for the choice of cover crops in banana cropping systems: theoretical developments and practical applications. Acta Horticulturae 33–40. https://doi.org/10.17660/ ActaHortic.2018.1196.4
- den Hollander NG, Bastiaans L, Kropff MJ (2007) Clover as a cover crop for weed suppression in an intercropping design: I. Characteristics of several clover species. Eur J Agron 26:92–103. https://doi.org/10. 1016/j.eja.2006.08.011
- Díaz S, Lavorel S, Chapin FS, Tecco PA, Gurvich DE, Grigulis K (2007) Functional diversity — at the crossroads between ecosystem functioning and environmental filters. In: Canadell JG, Pataki DE, Pitelka LF (eds) Terrestrial ecosystems in a changing world, global change — the IGBP series. Springer, Berlin, pp 81–91. https://doi. org/10.1007/978-3-540-32730-1\_7
- Duarte CM, Sand-Jensen K, Nielsen SL, Enríquez S, Agustí S (1995) Comparative functional plant ecology: rationale and potentials. Trends Ecol Evol 10:418–421. https://doi.org/10.1016/S0169-5347(00)89163-6
- FAO (2014) World reference base for soil resources 2014: international soil classification system for naming soils and creating legends for soil maps. FAO, Rome
- Fried G, Cordeau S, Metay A, Kazakou E (2019) Relative importance of environmental factors and farming practices in shaping weed communities structure and composition in French vineyards. Agric

Deringer

INRAe

Ecosyst Environ 275:1–13. https://doi.org/10.1016/j.agee.2019.01. 006

- Fried, Chauvel M, Reboud X (2020) Which traits make weeds more successful in maize crops? Insights from a three-decade monitoring in France. Plants 9:40. https://doi.org/10.3390/plants9010040
- Gaba S, Fried G, Kazakou E, Chauvel B, Navas M-L (2014) Agroecological weed control using a functional approach: a review of cropping systems diversity. Agron Sustain Dev 34:103–119. https://doi.org/10.1007/s13593-013-0166-5
- Garnier E, Navas M-L, Grigulis K (2018) Plant functional diversity: organism traits, community structure, and ecosystem properties. In: The quarterly review of biology. Oxford University Press, Oxford, p 51
- Gunton RM, Petit S, Gaba S (2011) Functional traits relating arable weed communities to crop characteristics: traits relating weed communities to crops. J Veg Sci 22:541–550. https://doi.org/10.1111/j.1654-1103.2011.01273.x
- Jackson L, van Noordwijk M, Bengtsson J, Foster W, Lipper L, Pulleman M, Said M, Snaddon J, Vodouhe R (2010) Biodiversity and agricultural sustainagility: from assessment to adaptive management. Curr Opin Environ Sustain 2:80–87. https://doi.org/10.1016/j.cosust. 2010.02.007
- Lamour A, Lotz LAP (2007) The importance of tillage depth in relation to seedling emergence in stale seedbeds. Ecol Model 201:536–546. https://doi.org/10.1016/j.ecolmodel.2006.10.015
- Lu Y-C, Watkins KB, Teasdale JR, Abdul-Baki AA (2000) Cover crops in sustainable food production. Food Rev Int 16:121–157. https:// doi.org/10.1081/FRI-100100285
- Médiène S, Valantin-Morison M, Sarthou J-P, de Tourdonnet S, Gosme M, Bertrand M, Roger-Estrade J, Aubertot J-N, Rusch A, Motisi N, Pelosi C, Doré T (2011) Agroecosystem management and biotic interactions: a review. Agron Sustain Dev 31:491–514. https://doi. org/10.1007/s13593-011-0009-1
- Petchey OL, Gaston KJ (2006) Functional diversity: back to basics and looking forward. Ecol Lett 9:741–758. https://doi.org/10.1111/j. 1461-0248.2006.00924.x
- Petit S, Boursault A, Le Guilloux M, Munier-Jolain N, Reboud X (2011) Weeds in agricultural landscapes. A review. Agron Sustain Dev 31: 309–317. https://doi.org/10.1051/agro/2010020

- R Core Team (2018) R: a language and environment for statistical computing. R Foundation for Statistical Computing, Vienna
- Ringselle B, Berge TW, Stout D, Breland TA, Hatcher PE, Haugland E, Koesling M, Mangerud K, Lunnan T, Brandsæter LO (2019) Effects of renewal time, taproot cutting, ploughing practice, false seedbed and companion crop on docks (Rumex spp.) when renewing grassland. Eur J Agron 103:54–62. https://doi.org/10.1016/j.eja.2018.11. 005
- Risède, J.-M., Lescot, T., Cabrera Cabrera, J., Guillon, M., Tomekpé, K., Kema, G.H.J., Côte, F.-X., 2010. Challenging short and mid-term strategies to reduce pesticides in bananas. Sci Field Banana Case Study – Guide Number 1, 8.
- Senarathne SHS, Sangakkara RU (2009) Effect of different weed management systems on the weed populations, and seedbank composition and distribution in tropical coconut plantations. Weed Biol Manag 9:209–216. https://doi.org/10.1111/j.1445-6664.2009. 00341.x
- Snapp SS, Swinton SM, Labarta R, Mutch D, Black JR, Leep R, Nyiraneza J, O'Neil K (2005) Evaluating cover crops for benefits, costs and performance within cropping system niches. Agron J 97: 322–332. https://doi.org/10.2134/agronj2005.0322a
- Storkey J (2006) A functional group approach to the management of UK arable weeds to support biological diversity. Weed Res 46:513–522. https://doi.org/10.1111/j.1365-3180.2006.00528.x
- Tardy F, Moreau D, Dorel M, Damour G (2015) Trait-based characterisation of cover plants' light competition strategies for weed control in banana cropping systems in the French West Indies. Eur J Agron 71:10–18. https://doi.org/10.1016/j.eja.2015.08.002
- Teasdale, J.R., Brandsaeter, L.O., Calegari, A., Skora Neto, F., 2007. Cover crops and weed management, non chemical weed management: principles, concepts and technology. Cabi Publishing-C a B Int, Wallingford. https://doi.org/10.1079/9781845932909.0049
- Wyka TP, Oleksyn J, Karolewski P, Schnitzer SA (2013) Phenotypic correlates of the lianescent growth form: a review. Ann Bot 112: 1667–1681. https://doi.org/10.1093/aob/mct236

**Publisher's note** Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

