



**HAL**  
open science

## Landscape effects on the thermotolerance of carabid beetles and the role of behavioral thermoregulation

Lucy Alford, Sacha Roudine, Jean-Sébastien Pierre, Françoise Burel, Joan van Baaren

► **To cite this version:**

Lucy Alford, Sacha Roudine, Jean-Sébastien Pierre, Françoise Burel, Joan van Baaren. Landscape effects on the thermotolerance of carabid beetles and the role of behavioral thermoregulation. *Insect Science*, 2023, 30 (1), pp.251-263. 10.1111/1744-7917.13044 . hal-03659680

**HAL Id: hal-03659680**

**<https://hal.science/hal-03659680>**

Submitted on 12 May 2022

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Distributed under a Creative Commons Attribution - NonCommercial 4.0 International License

Author running head: *L. Alford et al.*

Title running head: *Landscape effects on carabid thermotolerance*

Correspondence: Lucy Alford, School of Biological Sciences, University of Bristol, 24 Tyndall Avenue, Bristol BS8 1TQ, UK. Email: [lucy.alford@bristol.ac.uk](mailto:lucy.alford@bristol.ac.uk)

ORIGINAL ARTICLE

**Landscape effects on the thermotolerance of carabid beetles and the role of behavioral thermoregulation**

Lucy Alford<sup>1,2</sup>, Sacha Roudine<sup>1</sup>, Jean-Sébastien Pierre<sup>1</sup>, Françoise Burel<sup>1</sup> and Joan van Baaren<sup>1</sup>

<sup>1</sup>*UMR 6553 ECOBIO, Université de Rennes I, Avenue du Général Leclerc, 35042 Rennes Cedex, France;*

<sup>2</sup>*School of Biological Sciences, University of Bristol, 24 Tyndall Avenue, Bristol BS8 1TQ, UK*

**Abstract** Physiological thermotolerance and behavioral thermoregulation are central to seasonal cold adaptation in ectothermic organisms. For species with enhanced mobility, behavioral responses may be of greater importance in the cold stress response. Employing the carabid beetles as a study organism, the current study compared physiological thermotolerance and behavioral thermoregulation in carabid species inhabiting cereal fields in different landscape contexts, from fine grain heterogeneous ‘complex’ landscapes to homogenous ‘simple’ landscapes. Physiological thermotolerance was determined via measurement of the  $CT_{min}$  and chill coma temperature. Behavioral responses to cold temperature exposure were determined employing a purpose built arena, and thoracic temperature measured to estimate the efficacy of the behavior as a form of behavioral thermoregulation. Results revealed an influence of landscape composition on the cold tolerance of carabid beetles, although species differed in their sensitivity to landscape intensification. A reduced effect of landscape on the thermotolerance of larger carabid beetles was observed, thought to be the consequence of greater mobility preventing local acclimation to microclimatic variation along the landscape intensification gradient. Investigation into behavioral thermoregulation of the three largest species revealed burrowing behavior to be the main behavioral response to cold stress, acting to significantly raise carabid body temperature. This finding highlights the importance of behavioral thermoregulation as a strategy to evade cold stress. The use of behavioral thermoregulation may negate the need to invest in physiological thermotolerance, further offering explanation for the lack of landscape effect on the physiological thermotolerance of larger carabids.

**Key words** agroecosystems; local adaptation; physiological thermal tolerance; plasticity; winter resistance

## Introduction

When faced with predictable unfavourable cold temperatures associated with temperate winters, a range of coping strategies are available to ectothermic organisms to enable them to persist in a thermally stressful environment. Firstly, the individual may avoid exposure to the unfavourable conditions altogether and migrate to a more suitable climate (Larsen & Lee, 1994). Alternatively, the individual may remain in the thermally stressful environment and, in doing so, must either enter into diapause, a genetically determined state of decreased metabolic activity and developmental arrest (Denlinger, 2002; Teets & Denlinger, 2013), or instead maintain activity. Central to the ability to remain and endure the challenges associated with temperate winters is the capacity to respond to environmental cues such as shortening day lengths and declining temperatures to trigger seasonal cold adaptation (Teets & Denlinger, 2013). Such seasonal adaptation is incurred via adaptive phenotypic plasticity and may encompass short-term changes to the physiology or behavior of the individual that act to enhance thermotolerance (May, 1979; Overgaard & MacMillan, 2017), to longer-term transgenerational plasticity (Massamba-N'Siala *et al.*, 2014; Walsh *et al.*, 2014). In ectothermic organisms, physiological changes to enhance thermotolerance may include changes to homeostatic balance, lipid membrane composition, cryoprotective osmolytes and heat shock protein expression (Overgaard & MacMillan, 2017). In addition to physiological plasticity, phenotypic plasticity may also encompass behavioral alterations which further act to enhance thermotolerance and assist in seasonal cold adaptation. Indeed, behavioral changes such as alterations to foraging behavior (Andrew *et al.*, 2013), overwintering strategy (Tougeron *et al.*, 2017), choice of microhabitat (Alford *et al.*, 2017) and level of sociality (Schurch *et al.*, 2016) may offset the effects of changing temperatures.

Studies into the plasticity of thermotolerance are commonly performed at the local scale (e.g., Powell & Bale, 2005, 2006; Sinclair & Chown, 2006; Andrew *et al.*, 2011; Renault *et*

*al.*, 2012). However, with landscape components such as topography, vegetation type and cover known to directly impact the microclimate (Chen *et al.*, 1999), studies have more recently examined physiological thermotolerance at the landscape scale to elucidate how landscape composition can impact the acclimatory response of ectotherms (Tougeron *et al.*, 2016; Alford *et al.*, 2018). Indeed, simplified landscapes of intensive farming regimes display significantly different temperature means and extremes to wooded landscapes, with species reacting in accordance to their respective sensitivity to mean and extreme temperatures (Alford *et al.*, 2018). Furthermore, plasticity in physiological thermotolerance, although adaptive in winter when individuals are exposed to cold stress, becomes markedly reduced in spring, with any effect of landscape being reduced or lost completely (Tougeron *et al.*, 2016; Alford *et al.*, 2018). Ectothermic species may thus respond to microclimatic variation brought about by variation in landscape composition, and alter seasonal acclimation accordingly. In two related studies into the effect of landscape complexity on insect physiological thermotolerance, it was found that the cereal aphids *Metopolophium dirhodum*, *Rhopalosiphum padi* and *Sitobion avenae* (Hemiptera: Aphididae) and their parasitoid wasps *Aphidius avenae*, *A. ervi*, *A. matricariae*, *A. rhopalosiphi* (Hymenoptera: Braconidae) differed in their response. Here, aphids from more ‘complex’ farming landscapes (landscapes which retained a high proportion of semi-natural features such as hedgerows and grasslands), displayed an enhanced cold tolerance compared to their counterparts from intensively farmed ‘simple’ landscapes (Alford *et al.*, 2018). The reverse was true of aphid parasitoids, with parasitoids from intensively farmed landscapes displaying the greatest cold tolerance (Tougeron *et al.*, 2016), indicating that insects may not respond in a similar manner to landscape change. Cereal aphids and parasitoid wasps are comparatively small insects (Alford *et al.*, 2018) dependent upon a host plant and host aphid respectively, and thus with limited mobility. This is particularly true of aphids in their apterous form which predominate

in the study area. How larger insect species with enhanced mobility respond to landscape composition is currently unknown. However, any ability to utilize behavioral thermoregulation to buffer thermal variation may see a reduced plasticity in physiological thermotolerance and, as such, any landscape effect on physiological thermotolerance may be reduced or lacking completely.

Employing the Ground beetles (Coleoptera: Carabidae) *Agonum muelleri*, *Amara aenea*, *Anchomenus dorsalis*, *Bembidion lampros*, *Bembidion quadrimaculatum*, *Bembidion tetracolum*, *Harpalus rufipes*, *Poecilus cupreus*, *Pterostichus melanarius*, and *Trechus quadristriatus* as a study group, the current study aimed to investigate landscape effects on physiological thermotolerance and the role of behavioral thermoregulation in the cold stress response of a group of insects considered highly mobile at the local scale (Wallin & Ekblom, 1988; Jopp & Reuter, 2005). With over 40 000 species (Lövei & Sunderland, 1996), the carabids represent a diverse and cosmopolitan family of insects, exhibiting great diversity in traits including size (Jelaska & Durbešić, 2009), breeding season (Den Boer & Den Boer-Daanje, 1990), and diet (Sunderland, 1975; Jelaska *et al.*, 2014) to name a few. Many species of carabids are generalist predators and opportunistic feeders, capable of altering the relative consumption of prey species to match their seasonal abundance (Sunderland, 1975; Chiverton, 1987; Wheeler, 1989; King *et al.*, 2010; Jelaska *et al.*, 2014). For this reason, carabids play an important role in the biological control of herbivorous pests in temperate agroecosystems (Collins *et al.*, 2002; Sunderland, 2002; Al Hassan *et al.*, 2013). During winter months, carabids commonly overwinter within the soil in a state of diapause (Holland *et al.*, 2007). However, recent evidence suggests that carabids are becoming increasingly winter active in temperate agricultural landscapes as a consequence of warming temperatures (Damien, 2018). This increase in winter activity renders carabids increasingly susceptible to the cold stresses imposed by a temperate winter and places increased importance on

physiological and behavioral thermotolerance to withstand unfavourable cold temperatures. Using cereal fields (maize, wheat) of North-West France as a study system, we report on a two year study, comprising four biannual sampling seasons, investigating landscape effects on carabid beetle physiological thermotolerance along a landscape complexity gradient and the role of behavioral thermoregulation in the cold stress response. By employing the same ecosystem, region and landscape gradient as previously utilized by Tougeron *et al.* (2016) and Alford *et al.* (2018), results obtained for carabid beetles in the current study will be directly comparable to responses previously observed for the insects of other studied families. The following hypotheses were tested: (1) Due to complex landscapes having lower mean temperatures than simple landscapes, it is hypothesized that carabid physiological thermotolerance will significantly differ with landscape complexity with carabids originating from complex landscapes being the most cold tolerant. (2) Exposure to unfavourable cold temperatures will be less frequent in spring months compared to autumn months, eliciting a lower phenotypic response in carabid thermotolerance. Landscape effects on carabid thermotolerance will therefore be more pronounced in autumn and less pronounced or absent in spring. (3) There will be a reduced effect of landscape on the thermotolerance of larger carabids as a result of enhanced mobility to move through the landscape and evade unfavourable thermal conditions. (4) Carabid beetles will display behavioral thermoregulation to escape unfavourable cold temperatures. Active engagement in behavioral thermoregulation will enable the carabid to alter its core body temperature compared to carabids unable to engage in behavioral thermoregulation. (v) More cold tolerant carabid species will be less reliant on behavioral thermoregulation than less cold tolerant carabids.

## **Materials and methods**

### *Carabid collection and storage*

The current study utilized a landscape gradient previously established as part of the long-running project “FARMLAND” (farmland-biodiversity.org) in the Long-Term Ecological Research (LTER) site ZA Armorique (48°36'N, 1°32'W) (<https://osur.univ-rennes1.fr/za-armorique/>) in the Brittany region of France. Here, complex farming landscapes are classified as fine grain heterogeneous landscapes characterized by high diversity, high hedgerow density (> 3 200 m), small field sizes (< 0.93 ha) and the presence of grassland areas (> 45%). In contrast, intensified farming landscapes are classified as more homogenous landscapes characterized by low diversity, large field sizes (> 2.70 ha) and few semi-natural elements (grassland density < 20% and low hedgerow density < 550 m) (Tougeron *et al.*, 2016; Alford *et al.*, 2018). Utilizing this pre-existing landscape gradient, carabid beetles were sampled over four consecutive sampling seasons: autumn 2013 (commencing 9th September), spring 2014 (commencing 24th March), autumn 2014 (commencing 3rd September) and spring 2015 (commencing 30th March) for a duration of 4–5 weeks. In each of the four sampling seasons, a total of nine cereal fields from conventional, non-organic farms were selected along the landscape intensification gradient, with 3 fields selected to represent a ‘complex’ landscape, 3 fields an intensified ‘simple’ landscape, and 3 fields an ‘intermediate’ landscape. Due to sampling in agricultural landscapes, the fields selected for sampling were dictated by the crop rotation regime of the farmer. As a consequence, sampling fields varied season on season. During each sampling season, a total of 8 pitfall traps were dug in each field for the collection of carabids, spaced at a distance of 1 m apart and 5 m from the field margin. Traps comprised a plastic cup measuring 85 mm in diameter and 110 mm in depth. Traps were dug to a depth so that the entrance of the trap was flush with ground level. A ‘roof’, comprising a piece of Perspex measuring approximately 150 mm × 100 mm attached to a plastic stick, was positioned over each trap to prevent the trap filling with rainwater. A slice of apple and three to four pieces of dry dog food (Brekies Excel, Multicroc by Affinity Petcare) were placed



within each trap to provide a food source for captured carabids until the traps were emptied. Trap contents were collected every other day, approximately twice per week on a Wednesday and Friday, for the 4–5 week duration of the sampling season. At the end of a sampling week on the Friday, traps were closed with a plastic lid and re-opened the following week on the Monday to prevent carabids being trapped in the pitfall traps for extended periods of time over the weekends.

Collected adult carabids were returned to the laboratory at the Université de Rennes 1. Individual Perspex boxes were created to house the carabids and measured 175 mm × 115 mm × 65 mm with a ventilated lid. Boxes were lined with a layer of compost, approximately 20 mm deep, and roughly one quarter of the surface area was covered with moss. Carabids were crudely sorted into species and placed within boxes at densities of approximately 20 per box to reduce the possibility of cannibalism. All carabids were fed twice weekly on sliced apple and dry dog food (Brekies Excel, Multicroc by Affinity Petcare) and the boxes misted with water. All boxes were housed in a temperature-controlled climate chamber at  $20 \pm 1$  °C and LD 16 : 8 h for the minimum duration until use in experiments.

#### *Cold tolerance measurements*

Two indices of cold tolerance were determined for carabid beetles collected in the field. These thresholds included (1) the temperature at which spontaneous movement ceases and the insect is rendered immobile (referred to here as the Critical Thermal Minima or  $CT_{\min}$ ); and (2) the temperature at which the insect enters a cold induced torpor (chill coma), commonly defined by the last twitch of an appendage. These two thresholds refer to stages 2 and 4 in Hazell and Bale's four stages of the chill coma process (Hazell & Bale, 2011).

Critical thermal minima and chill coma thresholds were measured using a glass column, as described in Powell and Bale (2006). The glass column (35 × 5 cm), similar in design to a

condenser column, was connected to a programmable alcohol bath (Haake F3, Thermo Electron Corp., Karlsruhe, Baden-Württemberg, Germany), enabling the circulation of ethylene glycol around the outer chamber and thus fine control over the air temperature experienced within the inner column. In all experiments, the glass column was positioned horizontally and pre-set to the culture temperature of 20°C. Carabid beetles were inserted into the column at densities of 2–3 individuals (depending on size) and the column was subsequently closed with a sponge stopper to reduce air flow and maintain a stable thermal environment within the inner column. Densities of 2–3 carabids per test run enabled a more rapid test rate, whilst ensuring minimal interaction during threshold measurement. On the rare occasion that test carabids grouped together during threshold measurement, these data were excluded and the experiment was restarted with fresh carabids. Following a 10 min acclimatization period, the programmable alcohol bath was set to decrease the temperature of the column from 20°C to –10 °C at a rate of 0.5°C/min. The cooling rate was chosen to prevent the rapid cold hardening of test insects (Powell & Bale, 2006). During the cooling phase, the temperature of  $CT_{min}$  was determined for each carabid. Chill coma was determined for the larger species of carabid only (*H. rufipes*, *P. cupreus* and *P. melanarius*) since the larger size facilitated detection of the last twitch of an appendage. Critical thermal minima and coma threshold temperatures were recorded manually using a type T thermocouple probe (VWR, Fontenay-sous-Bois, France), accurate to 0.01°C, connected to a digital display.

Following completion of the experiment, each carabid (now in a state of chill coma) was removed from the glass tube and placed individually in an Eppendorf tube containing 70% ethanol solution for preservation. The experimental procedure was repeated to obtain  $CT_{min}$  and coma threshold temperatures for all carabids collected during a sampling season, which equated to approximately 450–500 carabids per season. This resulted in data for 1962

carabids over the two years. Species identification was performed for all carabids under a stereomicroscope using the identification key of Roger *et al.* (2012).

### *Carabid size*

Following determination of cold tolerance traits, carabid size was subsequently determined by measuring the distance between the labrum and the posterior end of the elytra (Roger *et al.*, 2012). Measurements were made using a camera (Zeiss AxioCam ERc5s<sup>®</sup> HD) mounted on an  $\times 9$  binocular microscope. Photographs were taken using Intelcam software and morphological measurements made using the image processing software Image J<sup>®</sup> software (v. 1.48). Size was determined for a total of 948 carabids over the two-year sampling period and the effect of size on carabid thermotolerance was investigated using this subset. The size of *B. quadrimaculatum* was not determined due to a lack of individuals when performing the size calculations.

### *Behavioral thermoregulation*

Behavioral thermoregulation was monitored in a subset of carabids comprising the three largest species collected during the spring 2015 sampling season from the intermediate landscape: *H. rufipes*, *P. cupreus* and *P. melanarius*. The largest species were chosen for ease of behavioral observations. All carabids were returned to the laboratory and stored as detailed above. A purpose-built observation chamber was designed that comprised two cylinders of Plexiglas which could be connected to a programmable alcohol bath. The doubled-walled nature of the arena, in conjunction with a secondary circuit built into the base, enabled the circulation of a cooling fluid (ethylene glycol) around the entirety of the cylinder with the exception of the open top of the cylinder which was eventually closed with a Plexiglas lid and rubber seal (Fig. 1). The base of the arena was divided into 4 sections, enabling the recreation

of 4 artificial habitats: (1) wheat grown in potting compost at a density of 15 blades of wheat per section and approximately 15 cm in height; (2) bare potting compost; (3) bare potting compost in which 3 holes of 1 cm in width and depth had been drilled; and (4) potting compost with an artificial shelter made of thin cardboard at a height of 3 cm. An artificial shelter was used to crudely represent leaf litter since use of leaf litter obstructed the carabid from view and interfered with behavioral observations.

For all experiments, the observation arena was housed in a temperature-controlled room held at  $15 \pm 1^\circ\text{C}$  under homogeneous white light, 250 lux. Prior to the experiment, an individual carabid was placed in the arena and allowed to acclimate to their new environment for a duration of 30 minutes. After this initial acclimation period, the programmable alcohol bath was programmed to decrease the temperature from the starting temperature of  $15^\circ\text{C}$  to  $0^\circ\text{C}$  at a rate of  $0.25^\circ\text{C}/\text{min}$  to produce an experimental duration of 1 h. A thermocouple was placed in the center of the arena to record the air temperature within the arena. During the cooling phase, carabid behavior was recorded every  $0.5^\circ\text{C}$  and the following behaviors were scored: walking, running, burying, grooming or inactive. Following completion of the temperature ramp, the carabid was removed and the body temperature of the beetle was immediately measured by inserting a thermocouple probe into the thorax of the carabid. A temperature control group was set up in which the carabids were not subjected to the declining temperature regime, but instead remained at the starting temperature of  $15^\circ\text{C}$ .

To test whether the ability to actively interact with the environment enabled the carabid to engage in behavioral thermoregulation and alter its body temperature, a second behavioral control was performed in which the bottom of the arena was filled with a layer of potting compost covered by a nylon mesh. The arena contained no features which enabled the carabid to climb or shelter and the nylon mesh acted to prevent the carabid from burrowing. For the behavioral control, a carabid was placed in the arena and the temperature was

decreased as detailed above. On reaching 0°C, the thoracic temperature of the carabid was measured as above.

### *Statistical analysis*

Since  $CT_{min}$  is fundamentally a delay,  $CT_{min}$  data were analyzed using a Cox Proportional Hazard model which belongs to the class of generalized linear models typically applied to survival data which are not normally distributed (exponential, Weibull, Gamma, etc.) (Alford *et al.*, 2018). Of the 31 and 38 carabid species collected in the autumn and spring sampling period respectively, 10 species were present across the three landscape types with at least 10 individuals for each type. A Cox Proportional Hazard model was subsequently built with data from the 10 species to determine any effect of landscape type and sampling season on the cold tolerance (measured by  $CT_{min}$ ) of carabid beetles using the *coxph* function of the “*survival*” package of R software (R Core team, 2019). Here, we take advantage of the confusion between time and temperature to treat the  $CT_{min}$  as a failure time. Cox models give access to all classic models of variance (deviance) analysis, covariance and regression. The cox model can be interpreted as the failure rate at time  $t$  and can be estimated as follows:

$$\lambda(t) = \lambda_0(t) \exp \sum_{i=1}^p \beta_i X_i \quad ,$$

with  $\lambda_0$  the baseline hazard rate that corresponds to the value of the hazard rate if all the covariates are equal to zero and  $\beta_i$  the regression coefficients that measure the impact of the covariates. A stepwise model selection was used based on the Akaike information criterion (AIC) to estimate predictor importance. Each field was considered as a frailty factor (random factor) and the significance of the interaction between the species and landscape type was tested. The overall model tested was the following:  $Surv(CT_{min}) = \text{intercept} + \text{species} \times$

landscape + sampling season+ frailty(field) + error. Analysis of deviance tests are issued from the  $\chi^2$  approximation of the deviance likelihood ratio. As the interaction was significant in the selected model, the crossed levels species  $\times$  landscape type was studied. The “survfit” function was used to calculate estimates of the different ‘survival’ curves (hereafter referred to as failure curves since the current study applies the method to cold tolerance data) using the Kaplan-Meier estimators. Pairwise differences between failure curves were studied using the log-rank test with the function *pairwise\_survdif* of the package *survminer* (Kassambara *et al.*, 2021), adjusting *P*-value with the Benjamini & Hochberg’s method (Benjamini & Hochberg, 1995).

Looking at carabid cold tolerance, a high interaction between landscape type and carabid species was found. As a consequence, landscape types were considered as strata to test interspecies differences in cold tolerance. As the number of carabid beetles was unbalanced between landscape type and in order to compare stratified samples of several species, the method of moments estimators was used. An unbiased estimator of the mean  $CT_{\min}$  for each species was independently built on the basis of the combination of a clustered sampling (fields) nested within a stratified sampling (landscape type). The details of the calculus are given in Appendix 1.  $CT_{\min}$  of all species were then compared two by two with a Bonferroni posthoc procedure.

To assess the effect of cold exposure on the behavior of carabids, as well as behavioral difference between the three species, Generalized Linear Models (GLM) with a Poisson distribution error were built. Separate GLM were performed for each behavior (i.e. walking, running, burrowing, inactive and grooming). Preference for substrate type was also analyzed using another GLM. To compare body temperature of carabids that were prevented from burrowing into the soil with the addition of a nylon mesh to carabids allowed to engage in burrowing behavior, as well as interspecific differences, a LM was performed. The

significance of each model term was assessed by a likelihood-ratio test (LRT), the distribution of which, as usual, was considered as Chi-square. A multiple comparison using the post hoc Tukey test was then performed using the emmeans package (Lenth, 2016) to evaluate differences among the three treatments for body temperature.

## Results

### *Carabid cold tolerance*

A total of 31 species were collected during the autumn sampling seasons and 38 during the spring sampling seasons (see Table S1). Of all the species collected, 10 species were present across the three landscape types [with at least 10 individuals for each type: simple (S.), intermediate (I.) and complex (C.)]: *Agonum muelleri* ( $n = 311$ // S. = 119, I. = 99, C. = 93), *Amara aenea* ( $n = 45$ // S.= 10, I.= 16, C.= 19), *Anchomenus dorsalis* ( $n = 126$ // S.= 58, I. = 46, C. = 22), *Bembidion lampros* ( $n = 294$ // S. = 143, I. = 88, C. = 63), *Bembidion quadrimaculatum* ( $n = 81$ // S. = 16, I. = 26, C. = 39), *Bembidion tetracolum* ( $n = 84$ // S.= 14, I. = 45, C. = 25), *Harpalus rufipes* ( $n = 172$ // S. = 58, I. = 75, C. = 39), *Poecilus cupreus* ( $n = 249$ // S.= 95, I. = 53, C. = 101), *Pterostichus melanarius* ( $n = 233$ // S. = 99, I. = 105, C. = 29) and *Trechus quadristriatus* ( $n = 100$ // S.= 30, I. = 39, C. = 31). This represented a total of 1 695 carabids out of the 1 927 sampled (642 in the simple landscape, 592 in the intermediate landscape and 461 in the complex landscape).

Sampling period had no effect on carabid cold tolerance as measured by  $CT_{\min}$  ( $\chi^2 = 2.20$ ,  $df = 1$ ,  $P = 0.14$ ). There was a significant effect of landscape type (simple, intermediate, complex) on the onset of carabid  $CT_{\min}$  and significant inter-species differences in the overall model. However, these effects were not interpretable as a high interaction effect was observed between landscape type and species ( $\chi^2 = 127.09$ ,  $df = 31$ ,  $P < 0.001$ ), suggesting that species differ in their sensitivity to landscape intensification. Carabid cold tolerance as

measured by  $CT_{min}$  differed between landscape type for six of the 10 species considered in the model, based on pairwise comparisons (Benjamini-Hochberg adjustment). No effect of landscape was observed for *A. aenea*, *B. tetracolum*, *H. rufipes* and *P. cupreus* (Fig. 2). For three species, the temperature of  $CT_{min}$  was significantly lower in complex landscapes compared to the two other landscape types (*B. lampros* and *T. quadristriatus*) or to the intermediate landscape type only (*A. muelleri*) (Fig. 2). This effect was most pronounced for *T. quadristriatus* for which individuals from complex landscapes ( $CT_{min}$  of  $0.73 \pm 0.22$  °C) maintained spontaneous movement to temperatures approximately 1.8 °C lower than individuals from intermediate and simple landscapes ( $CT_{min}$  of  $2.48 \pm 0.34$ °C and  $CT_{min}$  of  $2.56 \pm 0.49$ °C respectively) (Fig. 2). For three species, *A. dorsalis*, *B. quadrimaculatum* and *P. melanarius*, values of  $CT_{min}$  obtained for individuals from the intermediate landscape were almost double what were obtained for individuals in the simple landscape (Fig. 2).

*Poecilus cupreus* ( $CT_{min}$  of  $2.79 \pm 0.07$ °C) and *H. rufipes* ( $CT_{min}$  of  $3.53 \pm 0.16$ °C) showed the highest mean values of  $CT_{min}$  of the tested species. *Poecilus cupreus* and *H. rufipes* were thus rendered immobile at significantly warmer temperatures than the other species. No significant difference was observed between *B. lampros* ( $CT_{min}$  of  $1.94 \pm 0.08$ °C), *T. quadristriatus* ( $CT_{min}$  of  $1.97 \pm 0.22$  °C), *B. quadrimaculatum* ( $CT_{min}$  of  $1.43 \pm 0.15$ °C), *P. melanarius* ( $CT_{min}$  of  $1.25 \pm 0.08$ °C) and *A. aenea* ( $CT_{min}$  of  $1.48 \pm 0.30$ °C), although these species showed an increased cold tolerance compared to *P. cupreus* and *H. rufipes*. The three remaining species displayed an increased cold tolerance: *A. dorsalis* ( $CT_{min}$  of  $0.66 \pm 0.09$ °C), *A. muelleri* ( $CT_{min}$  of  $0.69 \pm 0.05$  °C) and *B. tetracolum* ( $CT_{min}$  of  $0.03 \pm 0.09$ °C). The  $CT_{min}$  of *B. tetracolum* was significantly lower than the two other species.

Sampling period had a significant effect on carabid cold tolerance as measured by chill coma (the temperature at which the insect enters a cold-induced torpor) ( $\chi^2 = 4.54$ ,  $df=1$ ,  $P = 0.03$ ) with the mean temperature of chill coma for carabids sampled in autumn/winter ( $-5.81$



$\pm 0.04^{\circ}\text{C}$ ) being significantly lower than the chill coma for carabids sampled in spring and summer ( $-5.70 \pm 0.08^{\circ}\text{C}$ ). Landscape type had no significant effect on carabid cold tolerance as measured by chill coma ( $\chi^2 = 2.28$ ,  $\text{df} = 2$ ,  $P = 0.32$ ). An interaction effect was also observed between landscape type and species ( $\chi^2 = 32.30$ ,  $\text{df} = 12.37$ ,  $P = 0.001$ ) although with not enough variance between modalities to be detected in pairwise tests. A negative beta coefficient was obtained for *P. cupreus* only ( $-5.96 \pm 0.07^{\circ}\text{C}$ ) when compared to *H. rufipes* as a baseline ( $-5.75 \pm 0.08^{\circ}\text{C}$ ). For *P. melanarius*, the temperature of chill coma ( $-5.61 \pm 0.06^{\circ}\text{C}$ ) was not significantly different from *H. rufipes*.

Carabid size was determined as follows: *A. aenea*  $6.25 \pm 0.23$  mm ( $n = 35$ ); *A. muelleri*  $7.52 \pm 0.08$  mm ( $n = 273$ ); *A. dorsalis*  $6.28 \pm 0.12$  mm ( $n = 113$ ); *B. lampros*  $2.72 \pm 0.03$  mm ( $n = 194$ ); *B. tetracolum*  $5.14 \pm 0.11$  mm ( $n = 73$ ); *H. rufipes*  $12.50 \pm 0.32$  mm ( $n = 25$ ); *P. cupreus*  $9.74 \pm 0.19$  mm ( $n = 102$ ); *P. melanarius*  $14.98 \pm 1.11$  mm ( $n = 5$ ); *T. quadristriatus*  $2.93 \pm 0.10$  mm ( $n = 2$ ). There was a significant effect of carabid size on cold tolerance ( $\chi^2 = 48.39$ ,  $\text{df} = 1$ ,  $P < 0.001$ ) with carabid size being positively correlated with the temperature of  $\text{CT}_{\text{min}}$  (positive coefficients  $\beta_i$ ). A reduced effect of landscape on the thermotolerance of larger carabids was observed. The larger species were either not affected by landscape type (*P. cupreus* and *H. rufipes*) or only in intermediate landscapes compared to simple landscapes (*P. melanarius*) (Fig. 2). Two of the three species for which cold tolerance was affected by complex landscape were among the smallest species sampled (*T. quadristriatus* and *B. lampros*) (Fig. 2).

#### *Carabid behavioral thermoregulation*

The activity level, as determined by exhibited walking behavior, significantly differed between the three carabid species ( $\chi^2 = 22.36$ ,  $\text{df} = 2$ ,  $P < 0.001$ ), and between the treatment group (cold stress vs. temperature control) ( $\chi^2 = 6.01$ ,  $\text{df} = 1$ ,  $P = 0.01$ ). Here, *P. cupreus* was

shown to be three times more active than *H. rufipes* (Tukey,  $P < 0.001$ ), engaging in more frequent walking behavior while other inter-species differences were not significant (Tukey,  $P > 0.05$ ). Cold stressed carabids significantly engaged in less frequent walking behavior than carabids in the control group (Tukey,  $P = 0.01$ ).

Very little grooming or running behavior or periods of inactivity were observed during the duration of the experiment regardless of the temperature treatment applied. When exposed to a cold stress, carabid beetles did not display different patterns of grooming ( $\chi^2 = 0.07$ ,  $df = 2$ ,  $P = 0.79$ ), running ( $\chi^2 = 0.41$ ,  $df = 2$ ,  $P = 0.52$ ) or inactivity ( $\chi^2 = 1.28$ ,  $df = 2$ ,  $P = 0.25$ ) compared to the control group. No inter-specific differences were measured for grooming ( $\chi^2 = 1.88$ ,  $df = 2$ ,  $P = 0.39$ ) and running ( $\chi^2 = 3.29$ ,  $df = 2$ ,  $P = 0.19$ ) but *P. melanarius* was significantly more inactive than the other 2 species ( $\chi^2 = 24.39$ ,  $df = 2$ ,  $P < 0.001$ ).

An interspecific preference for substrate type was also suggested for three of the substrate types: wheat ( $\chi^2 = 41.86$ ,  $df = 2$ ,  $P < 0.001$ ), bare soil without shelter ( $\chi^2 = 15.43$ ,  $df = 2$ ,  $P < 0.001$ ) and bare soil with shelter ( $\chi^2 = 80.83$ ,  $df = 2$ ,  $P < 0.001$ ), but not for the substrate with drilled holes ( $\chi^2 = 3.74$ ,  $df = 2$ ,  $P = 0.15$ ). *Pterostichus melanarius* was observed to spend more time in the substrate containing bare soil and an artificial shelter than either *P. cupreus* or *H. rufipes* ( $P < 0.001$ , Tukey) and at least four times less in the substrate containing wheat than the two other species ( $P < 0.001$ , Tukey). *P. cupreus* spent significantly more time in substrate containing bare soil without shelter than the two others species (Tukey,  $P < 0.01$ ).

Of the different behaviors investigated, burrowing behavior appeared to be the behavior most commonly employed by carabid beetles when exposed to a cold stress, with carabids from the cold stress treatment group spending significantly more time buried in the substrate ( $19.35 \pm 1.30$  min) than carabids in the control group ( $14.60 \pm 2.76$  min) ( $\chi^2 = 8.37$ ,  $df = 1$ ,  $P = 0.003$ ). Burrowing time also significantly differed between the three species ( $\chi^2 = 16.64$ ,  $df = 2$ ,  $P < 0.001$ ), with *H. rufipes* remaining buried in the substrate for a longer period of time

(21.06 ± 1.44 min) than *P. cupreus* (17.06 ± 1.76 min) (Tukey,  $P = 0.02$ ) and *P. melanarius* (13.00 ± 4.46 min) (Tukey,  $P < 0.001$ ). Cold stressed *P. cupreus*, the carabid displaying the lowest temperature of chill coma, burrowed into the soil later and thus at colder temperatures (14.25 ± 3.06 min) than cold stressed *H. rufipes* (7.37 ± 2.01 min) (Tukey,  $P < 0.001$ ) and cold stressed *P. melanarius* (4.40 ± 1.60 min) (Tukey,  $P < 0.001$ ). Burial latency between *P. melanarius* and *H. rufipes* were not significantly different (Tukey,  $P = 0.11$ ).

Body temperature was found to significantly differ between the treatment groups ( $\chi^2 = 133.76$ ,  $df = 2$ ,  $P < 0.001$ ), although did not differ between the three species ( $\chi^2 = 0.11$ ,  $df = 2$ ,  $P = 0.89$ ). When carabids were prevented from burrowing into the soil with the addition of a nylon mesh (the behavioral control), recorded body temperature (5.78 ± 0.29 °C) was significantly lower than the body temperature of carabids allowed to engage in burrowing behavior (8.13 ± 0.29°C) (Tukey,  $P < 0.001$ ). In addition, the body temperature of the temperature control group (14.91 ± 0.30°C) was significantly higher than the body temperature of carabids in the cold stress group and the behavioral control group (Tukey,  $P < 0.001$ ).

## Discussion

The current study revealed a landscape composition effect on the cold tolerance of carabid beetles, although species differed in their sensitivity to landscape composition, thus partly confirming hypothesis 1. Season did not influence the landscape effect on carabid cold tolerance, refuting hypothesis 2. Hypothesis 3, which predicted a reduced effect of landscape complexity on the thermotolerance of larger-bodied species, was partially supported since there was no effect of landscape on the  $CT_{min}$  of *H. rufipes* and *P. cupreus* or the chill coma temperature of *H. rufipes* and *P. cupreus*, *P. melanarius*; the three largest species of carabid in the current study. In support of Hypothesis 4, which predicted that carabid beetles would

Accepted Article

display behavioral thermoregulation to escape unfavorable cold temperatures, burrowing behavior was found to be the primary behavioral response to cold stress, acting to significantly alter carabid body temperature. Furthermore, *P. cupreus*, the carabid possessing the lowest chill coma temperature, burrowed into the soil at lower temperatures (i.e., remained exposed for longer durations) than *H. rufipes* and *P. melanarius*, offering support to Hypothesis 5 that more cold tolerant carabids will be less reliant on behavioral thermoregulation.

#### *Landscape effects on cold tolerance*

Landscape components such as topography, vegetation type and cover impact the microclimate of the landscape (Chen *et al.*, 1999). Indeed, complex agricultural landscapes have been shown to be significantly colder, but less variable, than simple landscapes (Tougeron *et al.*, 2016; Alford *et al.*, 2018). This is owing, in part, to the windbreak function of trees and hedgerows, resulting in a reduction in wind speed and subsequent retention of denser, cooler air (Quénol & Beltrando, 2006). Furthermore, as a landscape becomes increasingly closed, it is exposed to less short radiation from the sun which would otherwise raise the local temperature (Chen *et al.*, 1999; Suggitt *et al.*, 2011). Due to the colder mean temperatures experienced in complex landscapes, it was hypothesized that carabids originating from complex landscapes would be more cold tolerant than carabids originating from intermediate and simple landscapes (hypothesis 1). This hypothesis was supported by results on  $CT_{min}$  obtained for *A. muelleri*, *B. lampros* and *T. quadristriatus*. However, for *A. dorsalis*, *B. quadrimaculatum* and *P. melanarius*, individuals were more cold tolerant when originating from both complex and simple landscapes, although this difference was only significant for *A. dorsalis* and *P. melanarius* between simple and intermediate landscapes. When exposed to cold temperatures, a number of factors influence insect thermotolerance

and survival including temperature extremes, cooling rate, exposure duration and the extent to which temperature fluctuates around freezing (Sinclair *et al.*, 2003; Terblanche *et al.*, 2007). The importance of temperature extremes in dictating ectotherm thermal resistance is commonly accepted (Bahrndorff *et al.*, 2006; Paaijmans *et al.*, 2013; Estay *et al.*, 2014), although increasing research is suggesting that insects may differ in their sensitivity to aspects of the thermal environment and alter their physiological thermotolerance accordingly. Indeed, recent research has indicated that increased temperature variation increases the cold tolerance of the fruit fly *Drosophila melanogaster* (Diptera: Drosophilidae) and the moth *Helicoverpa armigera* (Lepidoptera: Noctuidae), whilst reducing the cold tolerance of the aphid *Acyrtosiphon pisum* (Hemiptera: Aphididae) (Estay *et al.*, 2014), indicating that whilst temperature extremes are important cues to the fruit fly and the moth, it is constant mean temperatures that are the more salient cue in aphid thermotolerance. Likewise, in a field study in temperate agricultural landscapes, the thermotolerance of *Aphidius* wasps responded to temperature fluctuations (Tougeron *et al.*, 2016), whilst the thermotolerance of cereal aphids responded to temperature means (Alford *et al.*, 2018). It is possible that carabid beetles may also differ in their sensitivity to temperature means and extremes, offering explanation as to why enhanced thermotolerance was observed in carabids originating from complex and simple landscapes.

Interestingly, the observed landscape effect on carabid thermotolerance (as determined by  $CT_{min}$ ) was not influenced by season, refuting hypothesis 2 which stated that landscape effects on carabid thermotolerance would be more pronounced in autumn and less pronounced or absent in spring. In temperate environments, environmental cues such as declining temperatures and shortening day lengths trigger seasonal cold adaptation in insects, enabling tolerance to the cold temperatures experienced during winter months (Teets & Denlinger, 2013). On the onset of spring, mean temperatures become warmer and less

extreme. Any acquired thermotolerance, which is costly to maintain (Block, 1991; Voituron *et al.*, 2002; Schou *et al.*, 2015), should be lost on cessation of winter temperatures, and any effect of landscape on thermotolerance should be reduced or lacking. This has previously been reported for parasitoid wasps (Tougeron *et al.*, 2017) and cereal aphids (Alford *et al.*, 2018), whereby landscape effects on thermotolerance were significantly reduced in spring months compared to winter months. This was not observed for carabids in the current study, with season having no effect on carabid  $CT_{min}$  along a landscape intensification gradient. Since carabids were sampled commencing September and March for an autumn and spring population respectively, it is possible that sampling did not capture peak seasonal cold acclimation in carabids, thereby preventing a seasonal effect from being elucidated.

Whilst an effect of landscape on the cold tolerance of *A. muelleri*, *A. dorsalis*, *B. lampros*, *B. quadrimaculatum*, *P. melanarius* and *T. quadristriatus* was reported, there was no effect of landscape on the  $CT_{min}$  of *A. aenea*, *B. tetracolum*, *H. rufipes* and *P. cupreus*, nor the chill coma temperature of *H. rufipes*, *P. cupreus* and *P. melanarius*. This finding partially supports hypothesis 3 which stated that there would be a reduced effect of landscape on the thermotolerance of larger carabids, since *H. rufipes*, *P. cupreus* and *P. melanarius* were the three largest species collected in the study area. *A. aenea* and *B. tetracolum* were intermediate sized carabids. Any increased ability to move throughout the landscape may hinder local acclimation to microclimatic variation along a landscape intensification gradient. In a tracking study conducted in cereal fields of Uppsala, Sweden, it was found that *P. melanarius* traveled average distances of 5.3 m, and *H. rufipes* average distances of 7.3 m, during their main activity period, although distances of up to 44.0 m were reported for *P. melanarius* (Wallin & Ekbohm, 1988). At least for the largest two carabids in the current study, comparatively large-scale movements throughout the landscape are thus known and, as such, may prevent patterns in physiological thermotolerance at the landscape scale. Indeed,

enhanced mobility of larger insects preventing local patterns in thermotolerance has recently been reported in bees. In a study into the critical thermal maximum ( $CT_{max}$ ) of the bees *Agapostemon sericeus* (Hymenoptera: Halictidae), *Apis mellifera* (Hymenoptera: Apidae) and *Bombus impatiens* (Hymenoptera: Apidae) along an urbanization gradient, Burdine and McCluney (2019) found no difference in thermotolerance between urban and rural sites. This led authors to conclude that greater mobility may prevent local acclimation and adaptation in the study bees. Furthermore, any local adaptation that did occur could be erased by genetic mixing.

#### *Carabid behavioral thermoregulation*

Burrowing behavior was revealed to be the primary behavioral response exhibited by *H. rufipes*, *P. cupreus* and *P. melanarius* when faced with a cold stress. Indeed, soil represents a more stable thermal environment than air, with thermal stability increasing with soil depth (Scharringa, 1976; Illston & Fiebrich, 2017) and vegetation cover further acting to reduce temperature variation (Scharringa, 1976; Oliver *et al.*, 1987). In the current study, engaging in burrowing behavior enabled carabids to maintain body temperature approximately 2 °C higher than carabids prevented from engaging in burrowing behavior, thus supporting hypothesis 4 which stated that carabid beetles will display behavioral thermoregulation to escape unfavourable cold temperatures. Burrowing behavior therefore provides thermoregulatory benefits to carabids during a cold stress. Indeed, behavioral thermoregulation more generally enables species to deal with variable environments. Nonetheless, for many species, behavioral thermoregulation is considered an unavailable or insufficient mechanism, with small body size considered a primary factor in limiting the ability of organisms to regulate body temperature (Gilchrist, 1995). Indeed, Merrick and Smith (2004) suggest that beetles below 2 g in weight are incapable of maintaining thoracic

temperature independently of ambient temperature, most likely the consequence of a larger surface area to volume ratio (Merrick & Smith, 2004). Authors further state that, for smaller beetles less than 1 g in weight, wing loading and wing beat frequency may play an important role in the maintenance of body temperature (Merrick & Smith, 2004). In the current study, *P. melanarius* represents one of the larger of the carabid species studied, weighing approximately 160 mg (Jonsson *et al.*, 2018); far below the 2 g threshold required for maintenance of body temperature as suggested by Merrick and Smith (2004). However, the current study revealed that burrowing behavior can significantly raise the thoracic temperature of carabids by approximately 2 °C, providing evidence that behavioral thermoregulation is a valuable method of body temperature regulation in beetles weighing 160 mg and below.

With burrowing behavior established as an important method of behavioral thermoregulation in carabid beetles, it was further hypothesized (hypothesis 5) that more cold tolerant species will be less reliant on behavioral thermoregulation, and instead utilize their inherent physiological thermotolerance to withstand a cold stress. Tentatively supporting this hypothesis was the finding that the species with the highest temperature of chill coma, *H. rufipes*, remained buried in the substrate for longer periods of time than the species with the lowest temperature of chill coma, *P. cupreus*. In a previous study investigating beetle overwintering in Japan, it was found that more accomplished diggers could avoid cold temperatures by burrowing deeper into the ground, thereby negating the need to develop frost resistance (Hoshikawa *et al.*, 1988). Since incurring seasonal cold acclimation is costly (Block, 1991; Voituron *et al.*, 2002; Schou *et al.*, 2015), it is possible that burrowing behavior results in a reduced investment in physiological mechanisms of cold tolerance, suggesting that a trade-off between physiological thermotolerance and behavioral



thermoregulation may exist. A reduced cold tolerance would thus be the consequence of burrowing behavior, and not instead a prerequisite to engage in burrowing behavior.

## **Conclusion**

A clear effect of landscape on the thermotolerance of carabid beetles was observed in the current study. However, carabid beetles did not respond in a similar manner to thermal conditions and changes in microhabitat and microclimate. The lack of landscape effect on the thermotolerance of larger species suggests that increased mobility may hinder local acclimation to thermal conditions. Furthermore, burrowing behavior, although revealed to be an important method of carabid behavioral thermoregulation, may further act to hinder local acclimation. If behavioral thermoregulation impedes an acclimatory response in carabid thermotolerance along a microclimatic gradient, it is likely that physiological mechanisms involved in thermotolerance are not under climate-induced selection pressures, with implications for the long-term evolutionary potential of carabid thermotolerance.

## **Acknowledgments**

The authors gratefully acknowledge Mathilde Méheut, Adrien Bonvin, Thomas Franco, Olivier Jambon, Romain Georges, Stephanie Llopis, Kévin Tougeron, Annabelle Androdias, Cécile Carré, Diab Al Hassan and Charlotte Alford for assistance in the field. Thanks also to Stephanie Llopis for technical assistance, Valérie Briand for bibliographic assistance, and Sandra Rigaud for handling the administrative aspects of the grant. This study was funded by a Marie Skłodowska-Curie Actions Intra-European Fellowship for the project ‘Climland’ (FP7-PEOPLE-2012-IEF-326943) awarded to L Alford, F Burel and J van Baaren and an Individual Fellowship for the project ‘FAB’ (H2020-MSCA-IF-2018-841952) awarded to L Alford and J van Baaren.

**Disclosure**

The authors declare that they have no conflict of interest.

## References

- Alford, L., Farahani, H.K., Pierre, J.S., Burel, F. and van Baaren, J. (2017) Why is there no impact of the host species on the cold tolerance of a generalist parasitoid? *Journal of Insect Physiology*, 103, 71–77.
- Alford, L., Tougeron, K., Pierre, J.S., Burel, F. and van Baaren, J. (2018) The effect of landscape complexity and microclimate on the thermal tolerance of a pest insect. *Insect Science*, 25, 905–915.
- Al Hassan, D., Georgelin, E., Delattre, T., Burel, F., Plantegenest, M., Kindlmann, P., *et al.* (2013) Does the presence of grassy strips and landscape grain affect the spatial distribution of aphids and their carabid predators? *Agricultural and Forest Entomology*, 15, 24–33.
- Andrew, N.R., Hart, R.A., Jung, M.P., Hemmings, Z. and Terblanche, J.S. (2013) Can temperate insects take the heat? A case study of the physiological and behavioral responses in a common ant, *Iridomyrmex purpureus* (Formicidae), with potential climate change. *Journal of Insect Physiology*, 59, 870–880.
- Andrew, N.R., Hart, R.A. and Terblanche, J.S. (2011) Limited plasticity of low temperature tolerance in an Australian cantharid beetle *Chauliognathus lugubris*. *Physiological Entomology*, 36, 385–391.
- Bahrndorff, S., Holmstrup, M., Petersen, H. and Loeschcke, V. (2006) Geographic variation for climatic stress resistance traits in the springtail *Orchesella cincta*. *Journal of Insect Physiology*, 52, 951–959.
- Benjamini, Y. and Hochberg, Y. (1995) Controlling the false discovery rate: A practical and powerful approach to multiple testing. *Journal of the Royal Statistical Society, Series B*, 57, 289–300.

- Block, W. (1991) To freeze or not to freeze? Invertebrate survival of sub-zero temperatures. *Functional Ecology*, 5, 284–290.
- Burdine, J.D. and McCluney, K.E. (2019) Differential sensitivity of bees to urbanization-driven changes in body temperature and water content. *Scientific Reports*, 9, 1643.
- Chen, J.Q., Saunders, S.C., Crow, T.R., Naiman, R.J., Brosofske, K.D., Mroz, G.D., *et al.* (1999) Microclimate in forest ecosystem and landscape ecology: variations in local climate can be used to monitor and compare the effects of different management regimes. *Bioscience*, 49, 288–297.
- Chiverton, P.A. (1987) Predation on *Rhopalosiphum padi* (Hom: Aphididae) by polyphagous predatory arthropods during the aphids' pre-peak period in spring barley. *Annals of Applied Biology*, 111, 257–269.
- Collins, K.L., Boatman, N.D., Wilcox, A., Holland, J.M. and Chaney, K. (2002) Influence of beetle banks on cereal aphid predation in winter wheat. *Agriculture, Ecosystems & Environment*, 93, 337–350.
- Damien, M. (2018) Favoriser les ennemis naturels de ravageurs par la diversité végétale dans un contexte hivernal. PhD Thesis L'universite De Rennes 1, Rennes.
- Den Boer, P.J. and Den Boer-Daanje, W. (1990) On life-history tactics in carabid beetles: are there only spring and autumn breeders? *The Role of Ground Beetles in Ecological and Environmental Studies* (ed. N.E. Stork), pp. 247–258. Intercept, Andover.
- Denlinger, D.L. (2002) Regulation of diapause. *Annual Review of Entomology*, 47, 93–122.
- Estay, S.A., Lima, M. and Bozinovic, F. (2014) The role of temperature variability on insect performance and population dynamics in a warming world. *Oikos*, 123, 131–140.
- Gilchrist, G.W. (1995) Specialists and generalists in changing environments. I. Fitness landscapes of thermal sensitivity. *The American Naturalist*, 146, 252–270.

- Hazell, S.P. and Bale, J.S. (2011) Low temperature thresholds: are chill coma and  $CT_{min}$  synonymous? *Journal of Insect Physiology*, 57, 1085–1089.
- Holland, J.M., Thomas, C.F.G., Birkett, T. and Southway, S. (2007) Spatio-temporal distribution and emergence of beetles in arable fields in relation to soil moisture. *Bulletin of Entomological Research*, 97, 89–100.
- Hoshikawa, K., Tsutsui, H., Honma, K. and Sakagami, S.F. (1988) Cold resistance in four species of beetles overwintering in the soil, with notes on the overwintering strategies of some soil insects. *Applied Entomology and Zoology*, 23, 273–281.
- Illston, B.G. and Fiebrich, C.A. (2017) Horizontal and vertical variability of observed soil temperatures. *Geoscience Data Journal*, 4, 40–46.
- Jelaska, L.S. and Durbešić, P. (2009) Comparison of the body size and wing form of carabid species (Coleoptera: Carabidae) between isolated and continuous forest habitats. *Annales Societe Entomologique de France*, 45, 327–338.
- Jelaska, L.S., Franjević, D., Jelaska, S.D. and Symondson, W.O.C. (2014) Prey detection in carabid beetles (Coleoptera: Carabidae) in woodland ecosystems by PCR analysis of gut contents. *European Journal of Entomology*, 111, 631–638.
- Jonsson, T., Kaartinen, R., Jonsson, M. and Bommarco, R. (2018) Predictive power of food web models based on body size decreases with trophic complexity. *Ecology Letters*, 21, 702–712.
- Jopp, F. and Reuter, H. (2005) Dispersal of carabid beetles: Emergence of distribution patterns. *Ecological Modelling*, 186, 389–405.
- Kassambara, A., Kosinski, M., Biecek, P. and Fabian, S. (2021) survminer: Drawing survival curves using ‘ggplot2’. R package version 0.4.9.

- King, R.A., Vaughan, I.P., Bell, J.R., Bohan, D.A. and Symondson, W.O.C. (2010) Prey choice by carabid beetles feeding on an earthworm community analysed using species- and lineage-specific PCR primers. *Molecular Ecology*, 19, 1721–1732.
- Larsen, K.J. and Lee, R.E. (1994) Cold tolerance including rapid cold-hardening and inoculative freezing of fall migrant monarch butterflies in Ohio. *Journal of Insect Physiology*, 40, 859–864.
- Lövei, G.L. and Sunderland, K.D. (1996) Ecology and behavior of ground beetles (Coleoptera: Carabidae). *Annual Review of Entomology*, 41, 231–236.
- Massamba-N’Siala, G., Prevedelli, D. and Simonini, R. (2014) Trans-generational plasticity in physiological thermal tolerance is modulated by maternal pre-reproductive environment in the polychaete *Ophryotrocha labronica*. *The Journal of Experimental Biology*, 217, 2004–2012.
- May, M.L. (1979) Insect thermoregulation. *Annual Review of Entomology*, 24, 313–349.
- Merrick, M.J. and Smith, R.J. (2004) Temperature regulation in burying beetles (*Nicrophorus* spp.: Coleoptera: Silphidae): Effects of body size, morphology and environmental temperature. *Journal of Experimental Biology*, 207, 723–733.
- Oliver, S.A., Oliver, H.R., Wallace, J.S. and Roberts, A.M. (1987) Soil heat flux and temperature variation with vegetation, soil type and climate. *Agricultural and Forest Meteorology*, 39, 257–269.
- Overgaard, J. and MacMillan, H.A. (2017) The integrative physiology of insect chill tolerance. *Annual Review of Physiology*, 79, 187–208.
- Paaijmans, K.P., Heinig, R.L., Seliga, R.A., Blanford, J.I., Blanford, S., Murdock, C.C., *et al.* (2013) Temperature variation makes ectotherms more sensitive to climate change. *Global Change Biology*, 19, 2373–2380.

- Powell, S.J. and Bale, J.S. (2005) Low temperature acclimated populations of the grain aphid *Sitobion avenae* retain ability to rapidly cold harden with enhanced fitness. *The Journal of Experimental Biology*, 208, 2615–2620.
- Powell, S.J. and Bale, J.S. (2006) Effect of long-term and rapid cold hardening on the cold torpor temperature of an aphid. *Physiological Entomology*, 31, 348–352.
- Quénol, H. and Beltrando, G. (2006) Microclimate in forest ecosystem and landscape ecology. *Climatologie*, 3, 9–23.
- Renault, D., Bijou, A. and Hervant, F. (2012) Impact of different acclimation temperatures and duration on the chill coma temperature and oxygen consumption in the tenebrionid beetle *Alphitobius diaperinus*. *Physiological Entomology*, 37, 354–359.
- Roger, J.L., Jambon, O. and Bouger, G. (2012) Clé de détermination des carabidés: Paysages agricoles du Nord Ouest de la France. *Rennes: SAD-Paysage, Institut National de Recherche Agronomique*.
- Scharringa, M. (1976) On the representativeness of soil temperature measurements. *Agricultural Meteorology*, 16, 263–276.
- Schou, M.F., Loeschke, V. and Kristensen, T.N. (2015) Strong costs and benefits of winter acclimatization in *Drosophila melanogaster*. *PLoS ONE*, 10, e0130307.
- Schurch, R., Accleton, C. and Field, J. (2016) Consequences of a warming climate for social organisation in sweat bees. *Behavioral Ecology and Sociobiology*, 70, 1131–1139.
- Sinclair, B.J. and Chown, S.L. (2006) Rapid cold-hardening in a Karoo beetle, *Afrinus* sp. *Physiological Entomology*, 31, 98–101.
- Sinclair, B.J., Vernon, P., Klok, C.J. and Chown, S.L. (2003) Insects at low temperatures: an ecological perspective. *Trends in Ecology and Evolution*, 18, 257–262.

- Suggitt, A.J., Gillingham, P.K., Hill, J.K., Thomas, C.D., Huntley, B., Kunin, W.E., *et al.* (2011) Habitat microclimates drive fine-scale variation in extreme temperatures. *Oikos*, 120, 1–8.
- Sunderland, K.D. (1975) The diet of some predatory arthropods in cereal crops. *Journal of Applied Ecology*, 12, 507–515.
- Sunderland, K.D. (2002) Invertebrate pest control by Carabids. *The Agroecology of Carabid Beetles* (ed. J.M. Holland), pp. 165–214. Intercept Publishers, U.K.
- Teets, N.M. and Denlinger, D.L. (2013) Physiological mechanisms of seasonal and rapid cold-hardening in insects. *Physiological Entomology*, 38, 105–116.
- Terblanche, J.S., Deere, J.A., Clusella-Trullas, S., Janion, C. and Chown, S.L. (2007) Critical thermal limits depend on methodological context. *Proceedings of the Royal Society B: Biological Sciences*, 274, 2935–2942
- Tougeron, K., van Baaren, J., Burel, F. and Alford, L. (2016) Comparing thermal tolerance across contrasting landscapes: first steps towards understanding how landscape management could modify ectotherm thermal tolerance. *Insect Conservation and Diversity*, 9, 171–180.
- Voituron, Y., Mouquet, N., de Mazancourt, C. and Clobert, J. (2002) To freeze or not to freeze? An evolutionary perspective on the cold hardiness strategies of overwintering ectotherms. *American Naturalist*, 160, 255–270.
- Wallin, H. and Ekbom, B.S. (1988) Movements of carabid beetles (Coleoptera: Carabidae) inhabiting cereal fields: A field tracing study. *Oecologia*, 77, 39–43.
- Walsh, M.R., Whittington, D. and Funkhouser, C. (2014) Thermal transgenerational plasticity in natural populations of *Daphnia*. *Integrative and Comparative Biology*, 54, 822–829.



Wheater, C.P. (1989) Prey detection by some predatory Coleoptera (Carabidae and Staphylinidae). *Journal of Zoology*, 218, 171–185.

Manuscript received December 16, 2021

Final version received February 25, 2022

Accepted March 22, 2022

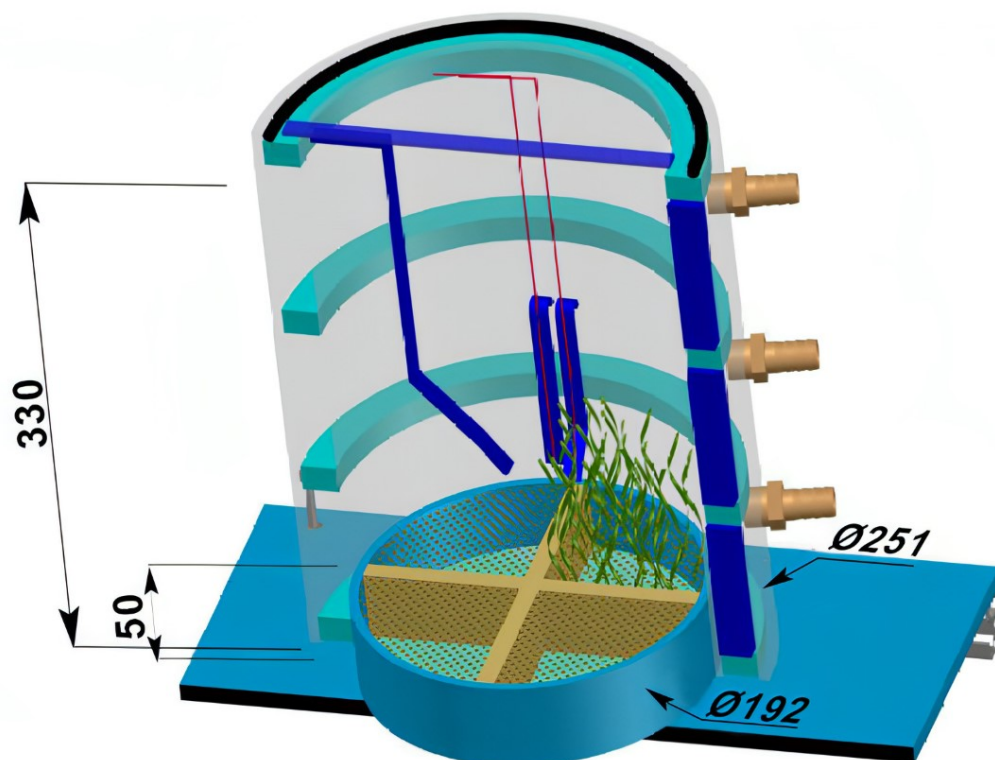
### **Supporting Information**

**Appendix 1** evaluation and comparison of the mean  $CT_{\min}$  of each species of carabids.

**Table S1** Carabid beetle species and associated  $n$  numbers collected during (a) autumn 2013 and 2014 and (b) spring 2014 and 2015.

### Figure legends

**Fig. 1** Observation arena developed to study carabid behavioral thermoregulation. The doubled-walled nature of the arena, in conjunction with a secondary circuit built into the base, enabled the circulation of ethylene glycol from a programmable alcohol bath around the cylinder to control the temperature experienced within the arena. The floor of the arena was divided into four different substrates: (1) wheat grown in potting compost at a density of 15 blades of wheat and approximately 15 cm in height; (2) bare potting compost; (3) bare potting compost in which 3 holes of 1 cm in width and depth were drilled; and (4) potting compost with an artificial shelter. Dimensions are provided in mm.



**Fig. 2** Carabid cold tolerance as measured by  $CT_{min}$  ( $\pm$  SE) sampled in the three landscape types: complex (comp), intermediate (inter) and simple (simp) considering all sampling seasons. Species are ordered by size, from smallest to largest. Each pairwise comparison between group level was calculated using a log-rank test using the `pairwise_survdif` function in R. Asterisks indicate significant  $P$  values: \* $P \leq 0.05$ , \*\* $P \leq 0.01$ , \*\*\* $P \leq 0.001$ .

