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Dendrimer nanoplatfoms for veterinary medicine applications: a concise overview

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Teaser. We discuss opportunities to develop dendrimer nanoplatfoms for veterinary medicine applications.

Within the nanoparticle (NP) space, dendrimers are becoming increasingly important in the field of nanomedicine, not only to treat human diseases, but also in veterinary medicine, which represents a new therapeutic approach. Major applications include using dendrimers to tackle highly contagious foot-and-mouth disease virus (FMDV) and swine fever virus (SFV) in pigs, FMDV in cattle, hypothermic circulatory arrest (HCA) in dogs, rabies, and H9N2 avian influenza virus in chickens. As we review here, intramuscular (im) subcutaneous (sc), intravenous (iv), and intraperitoneal (ip) routes of administration can be used for the successful application of dendrimers in animals.

Introduction

Until recently, veterinary drugs were developed simultaneously with human drugs, with no clear distinction between the two, regardless of the physical differences between the species. However, the trend now is for a clear distinction between drugs intended for humans and those intended for animals. Nevertheless, some biologically active molecules are used in both branches of medicine. As for human health, the efficacy and safety of biologically active molecules are also paramount for veterinary drugs. To be approved, these molecules must go through a validation process involving high-quality scientific research and extensive testing.

To ensure animal health and well-being, the development of new and specific veterinary pharmaceuticals is becoming increasingly important. A significant current focus of the pharmaceutical industry is the development of nanomedicines, involving nanotechnology. Nanotechnology involves a range of technologies and materials in the design, production, and application of nanodevices and system, and we are now surrounded by many applications of nanotechnology.¹ Indeed, during the past decade, nanotechnology has significantly impacted the precise delivery of therapeutic agents, diagnostics, and molecular imaging agents.² Approved nanoformulations have been developed, such as liposomal formulations (e.g., Abelect, AmBisome, and Amphoytec), pegylated liposomes (e.g., Doxil, and DaunoXome), nanocrystal formulations (e.g., Megace EM, Triplide, and Tricor), NP formulations (e.g., Abraxane and Estrasorb), and dendrimer formulations (e.g., VivaGel® and Viraleze). Recently, the successful of nanomedicine was demonstrated by the development of vaccines against severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2), such as liposomes encapsulating strands of mRNA encoding the 'spike' docking S protein of the virus. The BioNTech/Pfizer (BNT162b2) and Moderna mRNA-based vaccines^{3,4} are proving to be highly effective against the current coronavirus 2019 (COVID-19) pandemic.

A major aim of drug delivery is to improve efficiency by increasing the drug concentration at the site of action, improving the selectivity of the encapsulated or conjugated biologically active ingredients, and reducing their limiting toxic effects via dose reduction, consequently maximizing their therapeutic efficiency index and their preferential accumulation at target and tissue sites versus healthy tissues. This efficient drug reformulation strategy has been used for low water-soluble hydrophobic drugs and unstable drugs, as well as soluble toxic drugs, in aqueous solutions, increasing their dosage for parental administration and decreasing their systemic toxicity. Another advantage of NPs is the reduced clearance over parent drugs. In veterinary applications, numerous nanocarriers have been developed for the delivery of antibacterials, antifungals, antivirals, anesthetics, antiparasitics, and antineoplastic drugs, as well as vaccines. The simultaneous incorporation of hydrophobic and hydrophilic molecules has also been reported.

Based on the rapid emergence of nanotechnology, many promising domains are being introduced into medicine (so-called 'nanomedicine'). Nanotechnology represents an effective strategy for the cell-specific delivery and sustained release of drugs,⁵ as well as the co-delivery of antigens in vaccine development.^{6,7}

Within the numerous polymers, such as linear polymers and dendritic polymers,^{8,9} with polydispersity obtained by polymerization reactions or monodispersity frameworks obtained by step reactions (Figure 1), dendrimers are carefully assembled NPs of synthetic monomers with low polydispersity. There are over 100 families of dendrimers, the most well known being polyamidoamine (PAMAM), polylysine (PL), phosphorus dendrimers (PD), and polysilane dendrimers forming branch-like structures and repetitive geometrical units under radial layers, in so-called 'generations' (e.g., G0-Gn). The nanostructure of these materials has a significant impact on their physicochemical landscape.

Here, we review the most relevant milestones in the research and development of dendrimers as nanoplatforms for veterinary medicine. We discuss the general aspects of dendrimers in nanomedicine and the use of dendrimers in veterinary medicine. A search of the literature shows that the major veterinary applications of dendrimers are to tackle FMDV and SFV in pigs, FMDV in cattle, HCA in dogs, as well as rabies, and H9N2 avian influenza virus (AIV) in chickens. Intramuscular, subcutaneous, intravenous, and intraperitoneal routes of administrations are used.

Dendrimers in nanomedicine: general aspects

Cascade molecules were first synthesized by Vögtle *et al.* and described in 1978.¹⁰ Subsequently, these cascade structures have provided crucial low-molecular-weight, branched precursors to higher-molecular-weight, monodispersed, symmetrical nanoscale structures, now recognized as dendrimers. The first examples of these nanostructures were synthesized in 1979 [11] and given the name 'dendrimer' by Tomalia *et al.*^{11,12} The globular architecture of dendrimers enables hollow cavities within the branching structures to harbor synthetic drugs, biomacromolecules, natural bioactive compounds, and imaging agents, among others, as exemplified by PAMAM dendrimers (Figure 2). Numerous examples have been described, many in the oncology domain, including the delivery of small interfering (si)RNAs, complexed on the surface of dendrimers, forming dendriplexes, and of conjugated drugs on the surface of dendrimers. Importantly, dendrimers can be used as drugs themselves as anticancer, antibacterial, antiviral, anti-inflammatory, microbicide, antitubercular, and anti-Alzheimer agents.¹³ Modified PAMAM dendrimers have been successfully demonstrated to deliver self-amplifying mRNA to produce vaccines against H1N1 influenza and Ebola virus, among others.^{14,15} Tomalia *et al.* reported how emerging concepts in dendrimer-based nanomedicine provide fascinating opportunities in clinical translation, nanotoxicology, and theranostic nanodevices.¹⁶ Recently, the same authors emphasized the key influence of the cell symmetry of the dendrimer branches on the interior packing properties and, consequently, on their solubility and efficiency for drug delivery and nanomedical applications.¹² Altering five parameters (flexibility/rigidity, size, shape, surface chemistry, and elemental composition) offers the possibility to optimize the physicochemical profiles of dendrimers for their development in a variety of therapeutic applications.¹⁷

The nature of the terminal chemical groups (e.g., polycationic, polyanionic, and neutral functional groups) of dendrimers has a central role in both their *in vitro* and *in vivo* therapeutic applications, and delineates new 'druggable' spaces within this huge chemical space. The size of the dendrimers can be tightly regulated, and their surface can be highly functionalized with many attached group types for therapeutic agent delivery and active targeting (vectorization) purposes, and bioavailability improvements. Examples of the use of this approach include phosphorus dendrimers as nanoplatforms in therapeutic areas such as Alzheimer's and prion diseases,^{18,19} inflammatory diseases,^{20,21} cancers,^{22–24} and TB²⁵. Some dendrimers have also entered clinic trials. For example, Starpharma Holdings developed and marketed the first-in-class VivaGel (SPL7013), which is a G3 3 L-lysine dendrimer for antiviral and antibacterial applications in the treatment and prevention of bacterial vaginosis.^{26–28} VivaGel has 32 sodium 1-(carboxymethoxy)naphthalene-3,6-disulfonate on its surface. Recently, Starpharma Holdings announced the development of VivaGel as a potent antiviral agent against respiratory syncytial virus (RSV) using nasal spray technology named VIRALEZE™.

Dendrimers in veterinary medicine

The number of papers reporting the development of NPs in veterinary medicine have increased significantly over the past ten years or so, from ~100 in 2010 to ~800 in 2020 (based on a search of Scopus). The number of papers related to liposomes (57 versus 278), dendrimers (5 versus 38), polymeric NPs (3 versus 129), and metallic NPs (1 versus 37) showed a similar pattern of increase evolution from 2004 to 2020.

Most NP systems used in the veterinary medicine domain involve mainly liposomes (in a variety of animals, including dogs, cats, pigs, birds, sheep, horses and cattle), emulsions, magnetic NPs, dendrimers, water-based NP adjuvants, polystyrene nano-beads, and chitosan NPs.^{29–32} Liposomes can carry drugs, such as doxorubicin and cisplatin (various tumors), muramyl tripeptide (adenocarcinoma melanoma, osteosarcoma, and hemangiosarcoma), photosensitizers (carcinoma), 99m-technetium (imaging of sarcomas), IL-2 DNA (chronic rhinitis and pulmonary neoplasia), ribavirin (infections), hydromorphone (analgesia), trifluralin, amphotericin B, and meglumine antimoniate (leishmaniasis), clodronate (hemolytic anemia), amphotericin B (blastomycosis), α -tocopherol (vitamin E supplementation), streptomycin (brucellosis), gentamicin and staphylococcal antigens (*Staphylococcus aureus* mastitis treatment and vaccine), adriamycin (leukemia), diclofenac (analgesia and anti-inflammatory), diamidine (babesiosis), *Toxoplasma gondii* antigen (*T. gondii* vaccine), , avian colibacillosis vaccine, DNA vaccine (infectious

disease), *Mycoplasma gallisepticum* vaccine (mycoplasmosis), butorphanol (arthritis), and bupivacaine (antineoplastic and anesthesia). Most dendrimers are administered via iv, but other routes include im, ip, intratumoral, oral, intramammary, intranasal, transdermal, intraocular, and nebulization. These examples highlight the development of NPs in veterinary medicine, also detailed in Table 1.

Dendrimers to tackle HCA in dogs

Classically, during complex open-heart surgery, blood must continue to flow throughout the body, except in the surgical area, where the major blood vessels are clamped to prevent blood flow.⁴⁴ For instance, in aortic surgery, it is necessary to perform technical procedures to avoid clamping the aorta and to keep the surgical field free of blood. The technique known as deep HCA was developed to provide neuroprotection, but entails an ischemic period inducing excitotoxicity,⁴⁵ neuroinflammation,⁴⁶ and subsequent neurological injury as neurological sequelae. Consequently, neurocognitive dysfunction and injury remain problematic during HCA, although strokes, flawed neurocognitive function, neurological problems, and congenital malformations were observed in only 1–3% of patients after cardiac surgery.^{44,47} Unfortunately, several potential treatments to mitigate the neurological impact of HCA have been performed in the clinic but have been largely unsuccessful.⁴⁸ Consequently, no viable therapeutic options are available. This is mainly because of the limited blood–brain barrier (BBB) penetration, as well the systemic toxicity of therapeutic drugs. In addition, circulatory arrest increases toxicity of drugs at doses that are normally well tolerated, because of changes in their clearance and therapeutic effects.^{49,50}

Clinically relevant large animal models of HCA, such as the canine model, have been developed to induce brain injury, neuroinflammation, and excitotoxicity.⁵¹ Significant differences exist between large and small animal models, such as cardiac characteristics, including heart rate, oxygen consumption, adrenergic receptor ratios, and contractile protein expression. Canine models are closer to the human model in terms of physiology, anatomy, and function, and consequently represent adequate translations from bench to bedside.

Notably, in early studies, Mishra *et al.* developed non-cytotoxic conjugated bifunctional G4 hydroxy-PAMAM dendrimers encapsulating two clinically approved drugs, valproic acid (VPA) and *N*-acetyl cysteine (NAC), which attenuate excitotoxicity (epilepsy/bipolar disorder) and neuroinflammation, respectively.³⁸ Importantly, NAC exhibited poor BBB penetration, as well as defective localization, demanding large doses of drugs to gain neuroprotection.⁵² In addition to the neuroprotective effects, a large dose of VPA and NAC induced adverse effects, including perioperative hypotension involving fluid resuscitation and inotropic effect, and pulmonary edema/hemorrhage. To increase the therapeutic effects and decrease adverse effects, a strategy to tackle both excitotoxicity and neuroinflammation was developed using combination therapy strategies. This study is based on previous work by the same team showing that the G4 hydroxyl-PAMAM dendrimers without any encapsulated drugs were selectively localized in the injured brain and retina at the microglia level.⁵³ Two different bifunctional dendrimer types (D-VPA and D-NAC) were designed (Figure 3). These dendrimers have a disulfide-based glutathione sensitive linker enabling fast intracellular release of VPA and NAC. A polyethylene glycol (PEG)-valproic acid derivative (PEG-VPA) was prepared in a one-step synthesis by the esterification of VPA with PEG-SH (DCC/DMAP). In a brain injury canine model following circulatory arrest, the systemic administration of D-VPA and D-NAC was selectively assumed in injured neurons and microglia in the brain. In addition, combination therapy based on the association of D-VPA and D-NAC using 10 mg/kg of VPA in 70 mg/kg of dendrimer, and 10 mg/kg of NAC in 50 mg/kg of dendrimer, respectively, decreased systemic hemodynamic adverse effects and increased neurological outcomes based on a 24-h neurological deficit score compared with free drugs at the higher doses of tenfold (VPA) and 30-fold (NAC) or in combination therapy with 100 mg/kg of VPA and 300 mg/kg of NAC. D-VPA and D-NAC showed comparable improvements in neurological outcomes at one tenth of the dose used with reduced drug adverse effects over free VPA at the dose of 100 mg/kg.

Then, in 2016, Grimm and colleagues highlighted G6 hydroxyl-PAMAM dendrimers conjugated with the near-IR imaging agent Cy5 in a canine HCA-induced brain injury model.³⁹ Interestingly, by itself, this dendrimer interrupted the cascade process inducing neurocognitive dysfunction. The G6 hydroxyl-PAMAM dendrimer (Figure 3) was systematically administered, initially by an intravenous bolus dose (10% of the total 200 mg dose) and then, after peripheral aortic cannulation for 90 min, by the infusion administration of 70% of the total dose during a period of 6 h. The final bolus dose of 20% of the overall dose was given 24 h post-HCA. Analysis of G6 hydroxyl-Cy5 PAMAM dendrimer biodistribution showed that this dendrimer was specifically localized to the local macrophages of the nervous system microglia within the dorsal hippocampus, which is strongly connected with HCA. Microglia are the major effectors of inflammation and propagators of injury. The G6 hydroxyl-Cy5 PAMAM dendrimer crossed the BBB and targeted injured neurons and microglia after systemic administration.

The same team then investigated the hydroxyl-PAMAM dendrimer size effect of G4 versus G6 PAMAM dendrimers on brain uptake and kinetics in a canine model of HCA.⁴⁰ It is well accepted that both the size and the plasma-binding profile of compounds are major factors influencing their *in vivo* clearance and secretion pathways.

During systemic circulation, the strongest plasma protein interactions with NPs in general, and dendrimers in particular, occurred with hydrophobic or charged surfaces inducing high reticuloendothelial system (RES) clearance and strong liver uptake through the opsonization process.⁵⁴ G6 and G4 dendrimers are ~6.7 nm and ~4.3 nm in size, respectively. Using the same experimental procedure as the G6 hydroxyl-PAMAM dendrimer in the canine model of post-HCA dog brain injury, the G6 hydroxyl-PAMAM dendrimer displayed both high blood circulation times and strong accumulation in the cerebrospinal fluid (CSF) in injured animal brain versus the G4

hydroxyl-PAMAM dendrimers, which were undetectable in the brain after 48 h. Interestingly, the area under the curve (AUC) analysis showed that the CSF/serum ratio was ~20% at its peak and strongly correlated with the severity of neuroinflammation event. This ratio is higher compared with well-known clinical neurological drugs, such as penicillin, cloxacillin, amoxicillin, piperacillin, clavulanate, and cephalosporins. Based on confocal microscopy, G6 hydroxyl-PAMAM dendrimers displayed a decrease in renal clearance rate and a slightly increased uptake in the liver and spleen compared with G4 hydroxyl-PAMAM dendrimers.

Interestingly, as presented by Wang *et al.*, the topical cervical application of noncytotoxic G4 hydroxyl-PAMAM dendrimers (G4-PAMAM-OH) showed efficiency against *Escherichia coli* uterine infection in a guinea pig model of chorioamnionitis.⁴² The authors showed that cytokine release (TNF α and IL-6) was comparable between the treatment group over the negative control, clearly showing the efficiency of this approach for the treatment of *E. coli* infections. Based on an outer and inner membrane permeability assay, G4-PAMAM-OH dendrimers induced important structural changes to the outer membrane.

Dendrimers to tackle viral infections in pigs, dogs, and chickens

Another direction for the use of dendrimers in veterinary applications is to treat viral infections. Viral infections and immunization processes with vaccines usually induce high levels of circulating immunoglobulin G (IgG) as neutralizing antibodies, which correlates with protection against related viruses.⁵⁵ Thus, peptide vaccines represent an attractive strategy to treat viral infections. Peptide vaccines have short peptide fragments that induce highly targeted immune responses.⁵⁶ Generally, biodegradable NPs protect antigens from degradation, promote cellular absorption, and enhance immune responses, for instance as oral nanocarriers.⁵⁷

An interesting example is the development of vaccines to prevent endemic SFV (CSFV), one of the most important diseases for the pig industry worldwide.⁵⁸ Despite oral vaccination of the wild boar population to control this destructive disease, the development of vaccines is still required to allow the differentiation between vaccinated and infected animals (named DIVA).⁵⁹ Neutralizing antibodies specifically recognize the structural E₁ and E₂ envelope glycoproteins, and their presence has been associated with a protective effect.^{60,61} The E₂ glycoprotein is the primary target for neutralizing antibodies during natural CSFV infection.⁶²

Tarradas and colleagues reported the immunogenic effect of three synthetic PEGylated dendrimeric peptide vaccines against CSFV in pigs (Figure 4). The peptide vaccine had four copies of a B cell epitope from the CSF E₂ glycoprotein linked to a T cell epitope from the NS3 protein of the virus (residues 1446–1460). The pigs were challenged with a lethal dose of CSFV. The four copies of the B cell epitope from the E₂ envelope glycoprotein were E₂ (694–712) (Construct 1); 2: E₂ (712–727) (Construct 2); and E₂ (829–842) (Construct 3) (Figure 4).³⁵

Dendrimers have desirable features for the multimeric presentation of epitopes in candidate vaccines.^{63,64} In addition, the combination of B and T cell epitopes in dendrimers has been shown to confer significant protection against FMDV in domestic pigs.⁶⁵ This strategy aims to facilitate interactions between B and T epitopes, and to increase the immune response. Two doses [1.4 mg, im route of administration] of the corresponding construct were mixed with Montanide v206 adjuvant and were administered at days 1 and 21 post infection (p.i.). Challenge with virus was performed 15 days after the second dose. Im injection in the neck region was used for all inoculations. To evaluate the immunogenicity of the different constructions, the detection of specific antibodies was performed using ELISA. Partial protection and antibody responses were observed in the three groups of peptide-immunized animals, although construct 1 showed the highest levels of antibody response against the dendrimeric peptide. Thus, compared with control pigs developing important clinical signs of the disease, including anorexia, conjunctivitis, diarrhea, constipation, abdominal petechiae, and prostration, all pigs vaccinated with the construct 1 developed a strong antibody response recognizing the original dendrimeric immunogen as well as the E₂ epitope. This protection (only for one-third of the pigs) was related to the induction of IFN- γ -producing cells as well as neutralizing antibodies.

Based on the previous work, the same team developed four dendritic constructions to immunize pigs.³⁶ These constructions were prepared bearing either the NS3 T cell epitope alone or two different B cell epitopes without this T cell epitope (Figure 4). Construct 1', containing the NS3 T cell epitope and four copies of the B cell epitope, significantly reduced the clinical scores as well the RNA viral loads. Thus, partial protection was observed with half of the pigs, and the protection was associated with the action of IFN- γ -producing cells and neutralizing antibodies. Construct 2', bearing four copies of the B epitope of construct 1 but without the T cell motif, induced more severe clinical signs. Constructs 3' and 4', which included four copies of a B epitope and differed from Constructs 1' and 2' with or without the NS3 T cell epitope (Figure 4), showed low levels of cellular responses, an absence of neutralizing antibodies, and a lack of protection. Even so, the clinical score during the first week after challenge was less severe for animals vaccinated with Construct 3' than for those given Construct 4'. Collectively, these results strongly confirm the protection of Constructs 1' and 2' against CSFV, based on the important role of the B cell epitope in residues 694–712 of the glycoprotein E₂.

Interestingly, Asgary and colleagues advocated PEGylated G2 citric-acid based dendrimers as adjuvants of veterinary vaccines against rabies virus in NMRI mice. Rabies is a viral disease that attacks the central nervous system and is common in both humans and animals, with wild animals considered to be the source of infection.⁴¹ The main strategy for control of human rabies is vaccination.⁶⁶ Adjuvants are usually added to the vaccine formulation to increase the immunogenicity of the vaccines as well the strength of the immune response.⁶⁷ Currently, aluminum salts, such as aluminum hydroxide ('alum'), are used as an adjuvant, for instance, in

veterinary rabies vaccines. Although alum enhances the immune response, the aluminum adjuvant-containing vaccines can cause effects on local tissues, prolong inflammation at the injection site, and cause post-immunization headache, arthralgia, and myalgia.⁶⁸ These globular dendrimers are citric acid-polyethylene glycol-citric acid (CPEGC) triblock dendrimers (generation 2) (Figure 5).⁶⁹

In this study, to determine the best formulation for the survival of mice after viral challenge, different concentrations of G2 citric-acid based dendrimers were used. The increase in neutralizing antibodies was also checked by a fluorescent technique. Importantly, the *in vitro* toxicity of the G2 dendrimer was investigated using the MTT assay on J774A.1 macrophage-like cells and showed no significant toxic effects. An *in vivo* assay in mice, intraperitoneally injected on days 0 and 7, showed relevant adjuvanticity of the G2 dendrimer. The mortality assay of mice at different dendrimer concentrations showed that the highest viability percentage was observed at 12.5 mg/kg compared with the PBS group, which was confirmed by the fluorescent technique. However, there was no significant difference in the numbers of live and dead mice at higher concentrations compared with the PBS group. Finally, specific rabies-neutralizing antibodies against the rabies virus were confirmed by a neutralizing antibody titration assay.

Interestingly, Cubillos and colleagues designed and developed a synthetic dendrimeric peptide, named B₄T, to protect pigs against FMDV.³³ FMDV is a picornavirus producing a highly transmissible and devastating disease in farm animals, such as cattle and swine, and is one of the most devastating animal diseases worldwide.^{70,71} B₄T contains one copy of a FMDC– T cell epitope and branches bearing four copies of a B cell epitope (Figure 6). Interestingly, the immunized domestic pigs did not develop significant clinical signs, such as fever and viremia, based on RT-PCR viral RNA amplification in respiratory tract samples. B₄T specifically induced high titers of FMDV-neutralizing antibodies of T cells and a potent anti-FMDV IgA systemic and mucosal response. B₄T also inhibited airborne excretion of the virus. Interestingly, no contact transmission was observed from naive animals housed with peptide-vaccinated pigs, inducing no development of clinical signs of disease. Consequently, synthetic peptides mimicking protective B and T cell epitopes are good candidates for safer, more effective FMDV vaccines.

Recently, the same team developed dendrimer peptides for the protection of cattle against FMDV.³⁴ Several studies showed effective protection using peptide-based vaccines,⁷² synthetic peptides, or recombinant protein vaccines against FMDV in pigs but limited efficiency in cattle.⁷³ The dendrimeric peptides used had two or four copies of a peptide corresponding to the B cell epitope linked through thioether bonds to a single copy of the T cell epitope (Figure 6). The dendrimeric peptides B₂T⁺ and B₄T⁺ induced antipeptide and FMDV-specific antibodies (IgG1/2 responses) in cattle after subcutaneous injection. Overall, B₄T⁺ and B₂T⁺ elicited humoral and cellular immune responses in cattle, inducing partial protection against a heterogenous virus challenge, which was associated with both T cell and antibody responses.

Recently, Zhao *et al.* designed and developed a DNA vaccine based on dendrigraft poly-L-lysines against H9N2 AIV infection in chickens.³⁷ DNA vaccines are considered to be third-generation vaccines and are relatively safe over classical vaccines, and lack serious adverse effects.⁷⁴ DNA vaccines have been developed to treat many diseases caused by pathogens, including Zydus' ZyCoV-D, which recently received emergency use authorization (EUA) in India against COVID-19.⁷⁵ DNA vaccines increase DNA insertion and translation in the cell, activate CD4⁺ T cells, and enhance cytotoxic T lymphocyte induction and specific antibody production.⁷⁶ This study involved the construction of a specific plasmid by insertion of an HA gene fragment of H9N2 AIV into a vector that was packaged with G3 dendrigraft poly-L-lysines, which contain 123 primary amino groups. This construct allowed high stability of the system and high *in vitro* and *in vivo* transfection efficiency. Strong cellular immune responses were observed in immunized chickens. T lymphocytes were activated and proliferated, and CD3⁺/CD4⁺ and CD4⁺/CD8⁺ were increased, allowing complete protection of chickens after intramuscular administration against H9N2 AIV virus infection.

Concluding remarks and perspectives

Veterinary medicine ensures the health and well-being of all animals and, therefore, contributes to the improvement of public health, particularly with regard to diseases transmissible from animals to humans and the health and safety of animal products. As for humans, the effectiveness and safety of care depend on the quality of the veterinary diagnosis, which forms the basis of treatment. The two main specific applications of nanotechnologies in veterinary medicine are for veterinary pharmacology and therapeutics and veterinary vaccinology using viral NPs, including dendrimers. The major applications of dendrimers include against FMDV and SFV in pigs, FMDV in cattle, HCA in dogs, rabies, and H9N2 AIV in chickens. Im, sc, iv, and ip routes of administration are used. The oral route has never been a focus, although it is the main route of drug administration used in veterinary medicine. Within the dendrimer space defined elsewhere,⁷⁷ dendrimers can be delivered via several routes of administration using enteral, parenteral, and topical routes, such as oral, transdermal, ocular, and transmucosal routes.⁷⁸ The main potential challenges to the development of NPs for veterinary medicine application in general, and of dendrimers in particular, are as important as in human medicine, and include: (i) targeted transport of nano-encapsulated drugs; (ii) therapeutic nanomaterials active by themselves; (iii) vectorization of vaccine antigens (viral NPs); (iv) *in vivo* imaging; and (v) *in vitro* and *in vivo* diagnosis. The increasing use of these nanomaterials also requires an assessment of their environmental risks as well as their large-scale production, including the high costs of the manufacturing process compared with conventional drugs. In addition, the development of analytical procedures for in-process quality control and

adequate final drug quality of the nanomedicine must be controlled. Consequently, the future is promising for veterinary medicine applications using dendrimer nanoplatfroms.

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Declaration of interests

The authors declare no conflict of interest.

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Figure 1. Comparison of (a) linear polymers versus (b) dendritic polymers.

Figure 2. Chemical structure of 2D G4 polyamidoamine (PAMAM) dendrimers, their functional core, surface functions, internal cavities, and monomer elements.

Figure 3. Schematic of bifunctional (a) D-valproic acid (VPA), (b) D-N-acetyl cysteine (NAC), and (c) G6 hydroxyl-Cy5 polyamidoamine (PAMAM) dendrimers.

Figure 4. Schematic of constructs 1, 2, 3, 1', 2', and 3' based on dendrimer architecture developed against foot and mouth disease virus in pigs.

Figure 5. Chemical structure of citric acid-polyethylene glycol-citric acid (CPEGC) triblock dendrimers (generation 2) developed to treat rabies.

Figure 6. 2D chemical structure of (a) B₄T developed by Cubillos *et al.*,³³ and (b) B₄T⁺ and B₂T⁺, developed by Soria *et al.*³⁴ against foot and mouth disease virus in cattle.

Table 1. Studies of dendrimers in veterinary applications

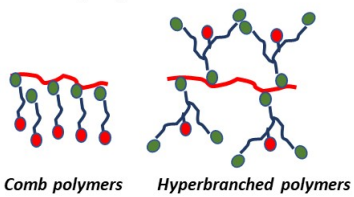
Species	Agent	Disease	Administration route	Refs
Cattle	Dendrimer peptide vaccine	FMDV	sc	34
Chickens	Dendrigrraft poly-L-lysines	H9N2 avian influenza virus	im	37
Dogs	VPA and NAC encapsulated in G4 hydroxy-PAMAM dendrimers	HAC	iv	38
	G6 hydroxyl-PAMAM dendrimers	HAC	iv	39,40
	PEGylated G2 citric-acid based dendrimers	Rabies	ip	41
	Construction (recombinant C1 antigen/G4 PAMAM dendrimer/CYS-SAM/gold surface) of SPR immunosensor to evaluate reaction kinetics between <i>Leishmania infantum</i> and specific antibodies (IgG anti-C1) for detection of visceral leishmaniasis	Visceral leishmaniasis		43
Guinea pigs	G4 hydroxyl-PAMAM dendrimer	<i>Escherichia coli</i>	Topical cervical application	42
Pigs	Dendrimer peptide vaccine	FMDV	im	33
	PEGylated dendrimeric peptide	SFV	im	35,36

Linear polymers

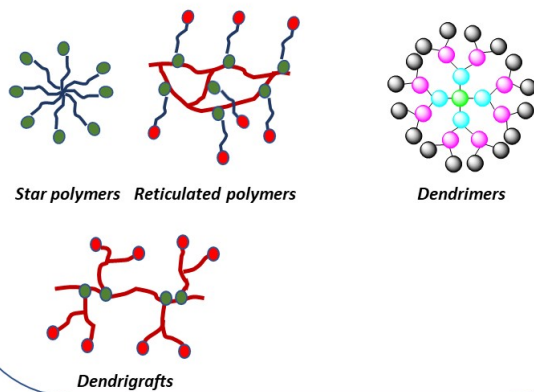


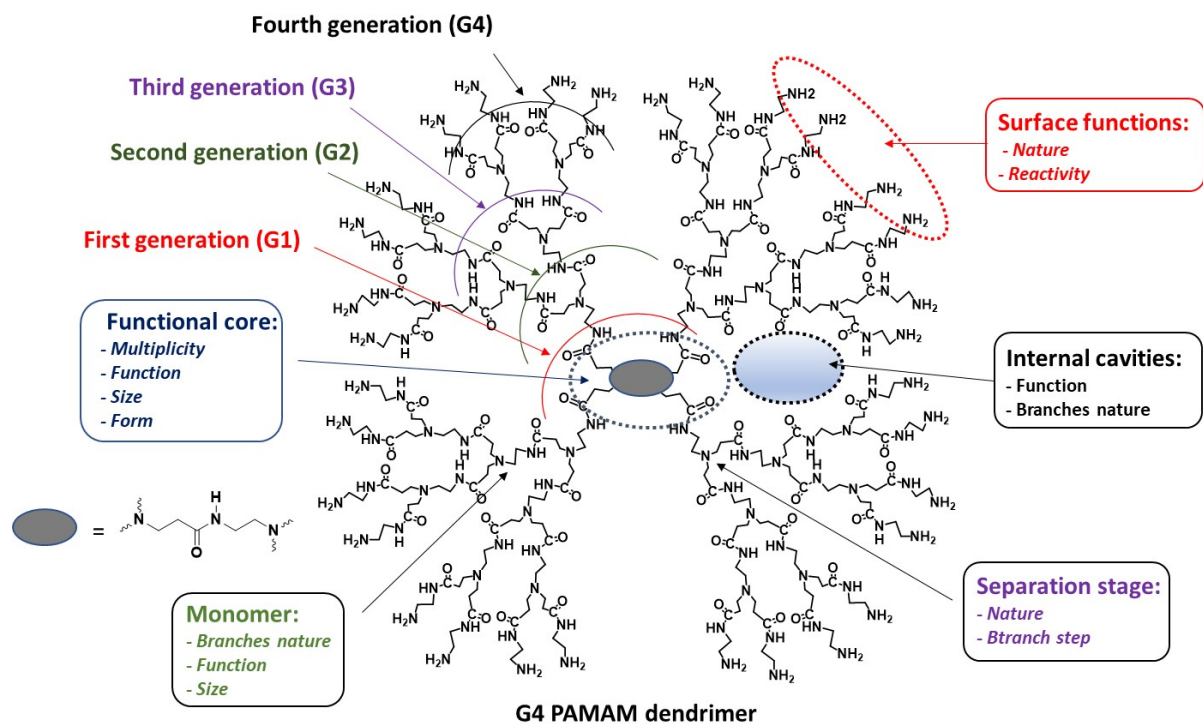
Dendritic polymers

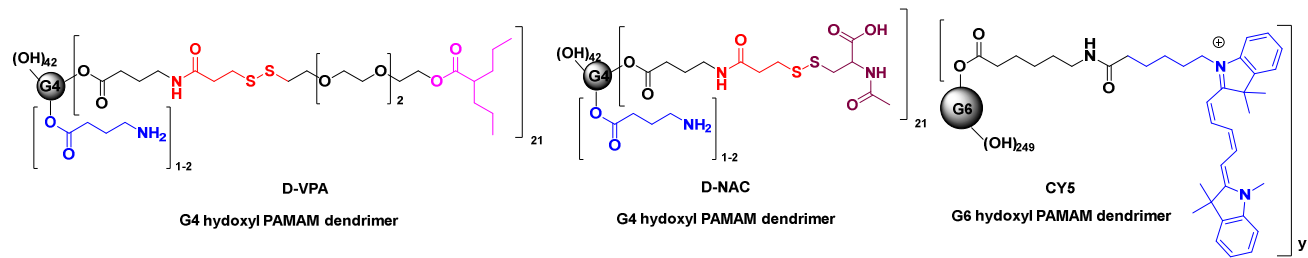
Polydisperse frameworks

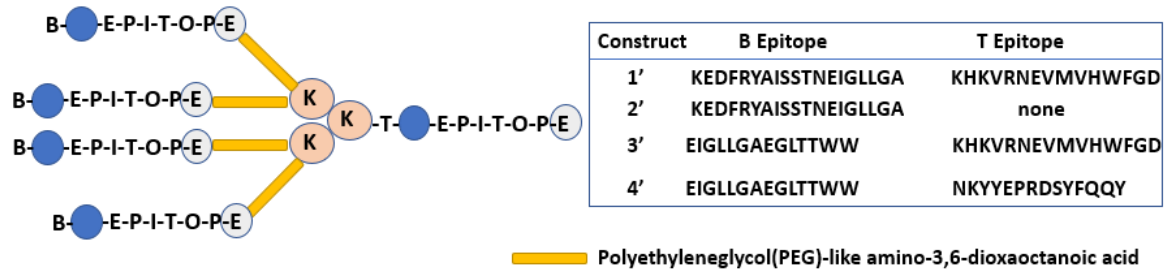
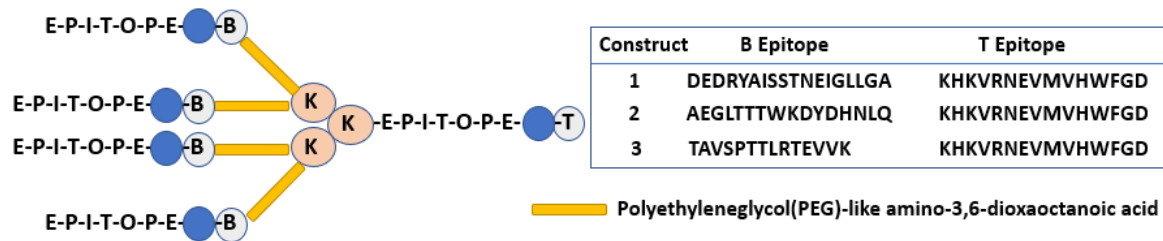


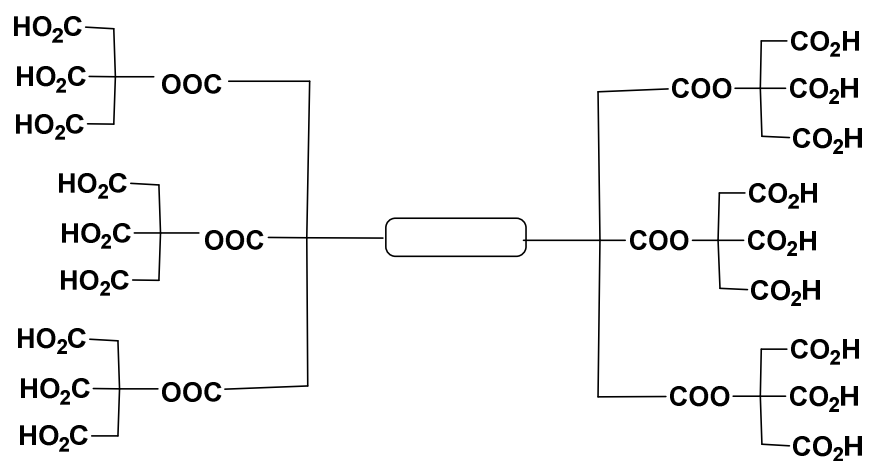
Monodisperse frameworks

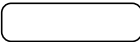






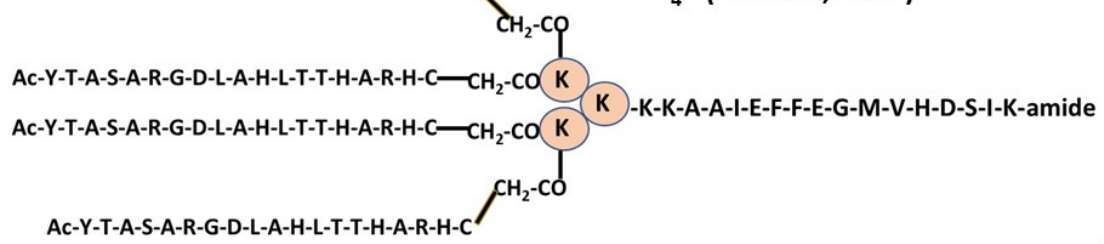




 : $-\text{OOCCH}_2(\text{OCH}_2-\text{OCH}_2)_n\text{OCH}_2-\text{COO}-$
 PEG_{600}

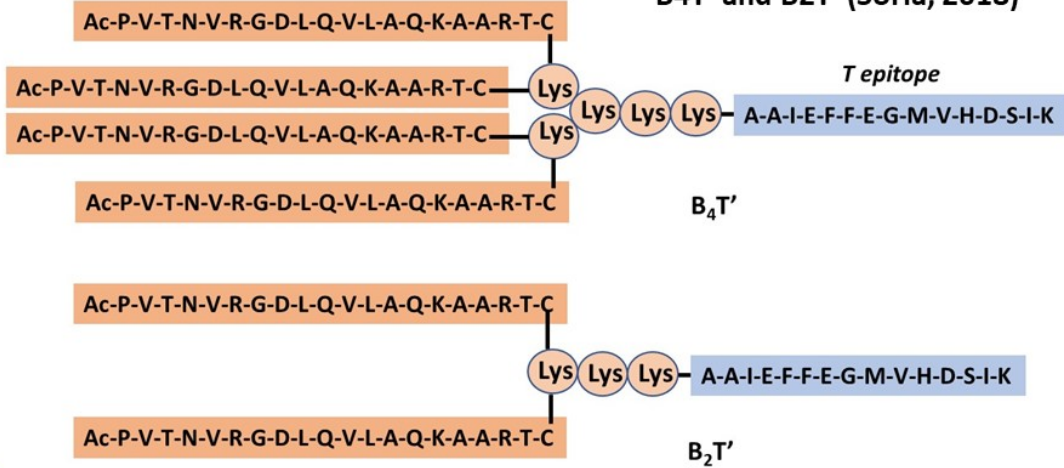
Ac-Y-T-A-S-A-R-G-D-L-A-H-L-T-T-H-A-R-H-C

B₄T (Cubillos, 2008)

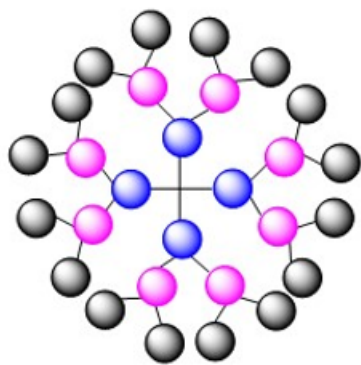
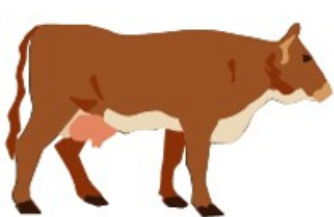


B epitope

B₄T' and B₂T' (Soria, 2018)



Veterinary medicine applications



Dendrimer platforms