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Quentin Sallembien, Laurent Bouteiller, Jeanne Crassous, Matthieu Raynal. Possible chemical and physical scenarios towards biological homochirality. *Chemical Society Reviews*, 2022, 51 (9), pp.3436-3476. 10.1039/D1CS01179K . hal-03632805

HAL Id: hal-03632805

<https://hal.science/hal-03632805>

Submitted on 6 Apr 2022

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Possible Chemical and Physical Scenarios Towards Biological Homochirality

Received 00th January 20xx,
Accepted 00th January 20xx

Quentin Sallembien,*^a Laurent Bouteiller,^a Jeanne Crassous*^b and Matthieu Raynal*^a

DOI: 10.1039/x0xx00000x

The single chirality of biological molecules in terrestrial biology raises more questions than certitudes about its origin. The emergence of biological homochirality (BH) and its connection with the appearance of Life has elicited a large number of theories related to the generation, amplification and preservation of a chiral bias in molecules of Life under prebiotically relevant conditions. However, a global scenario is still lacking. Here, the possibility of inducing a significant chiral bias “from scratch”, *i.e.* in absence of pre-existing enantiomerically-enriched chemical species, will be considered first. It includes phenomena that are inherent to the nature of matter itself, such as the infinitesimal energy difference between enantiomers as a result of violation of parity in certain interactions within nucleus of atoms, and physicochemical processes related to interactions between chiral organic molecules and physical fields, polarized particles, polarized spins and chiral surfaces. The spontaneous emergence of chirality in absence of detectable chiral physical and chemical sources has recently undergone significant advances thanks to the deracemization of conglomerates through Viedma ripening and asymmetric auto-catalysis with the Soai reaction. All these phenomena are commonly discussed as plausible sources of asymmetry under prebiotic conditions and are potentially accountable for the primeval chiral bias in molecules of Life. Then, several scenarios will be discussed that are aimed to reflect the different debates about the emergence of BH: extra-terrestrial or terrestrial origin (where?), nature of the mechanisms leading to the propagation and enhancement of the primeval chiral bias (how?) and temporal sequence between chemical homochirality, BH and Life emergence (when?). The last point encompasses various theories setting the emergence of optically pure molecules at the stage of building blocks of Life, of instructed/functional polymers or later. The underlying principles and the experimental evidences will be commented for each scenario with a particular attention on those leading to the induction and enhancement of enantiomeric excesses in proteinogenic amino acids, natural sugars, their intermediates or derivatives. The aim of this review is to propose an updated and timely synopsis in order to stimulate new efforts in this interdisciplinary field.

Dedicated to the memory of Sandra Pizzarello (1933-2021)

1. Introduction

In 1884, Lord Kelvin used the word chirality, —derived from the Proto-Indo-European *ǵ^hesr- through the Ancient Greek χείρ (kheír), that both mean ‘hand’— and gave the following definition: “an object is chiral if and only if it is not superimposable on its mirror image”.¹ Additionally, chirality can be described based on symmetry aspects: an object is chiral if it possesses no symmetry elements of the second kind (*i.e.* if it is devoid of any improper axis of rotation).² Whilst the manifestation of chirality at the macroscopic scale sparked human’s curiosity from antiquity, its observation at the molecular and sub-atomic levels is relatively recent. In the 19th-century, advances made in optics,³ crystallography⁴ and

chemistry⁵ paved the way to the scientific study of molecular chirality (named ‘molecular dissymmetry’ by Louis Pasteur)⁶ which soon after manifested itself in a variety of studied phenomena. The term “chirality” took additionally almost 80 years to be introduced in chemistry by Kurt Mislow (1962).⁷ Chirality is found at all scales in matter, from elementary particles to cucumber tendrils,⁸ from screws to spiral galaxies, in living and inert systems.⁹ It is also an everyday concern in industry (*e.g.* pharma, agribusiness, cosmetics)^{10–14} as well as in fundamental research (visible in countless conferences encompassing not only chemistry, physics, and biology but also economy and arts).¹⁵

Homochirality of Life refers to the fact that Nature has chosen a specific handedness. Homochirality is a fascinating aspect of terrestrial biology: All living systems are composed of L-amino acids and D-sugars⁸ to such an elevated extent that the occurrence of the molecules of Life with different configurations (*e.g.* D-amino acids) is seen as a curiosity.¹⁶ Clearly, the perfect level of selectivity reached by evolution and preserved along billion years, is out of reach for currently

^a Sorbonne Université, CNRS, Institut Parisien de Chimie Moléculaire, Equipe Chimie des Polymères, 4 Place Jussieu, 75005 Paris (France)

E-mail: quentin.sallembien.2017@enscbp.fr, matthieu.raynal@sorbonne-universite.fr

^b Univ Rennes, CNRS, Institut des Sciences Chimiques de Rennes, ISCR-UMR 6226, F-35000 Rennes, France. E-mail: jeanne.crassous@univ-rennes1.fr

developed artificial systems. Homochirality and Life are so closely related that homochirality in Nature is considered as a

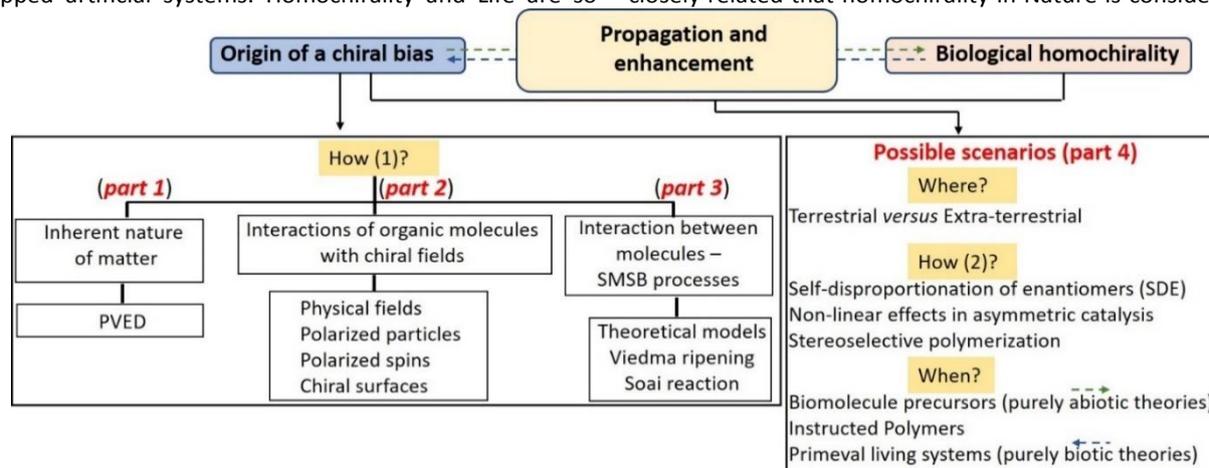


Figure 1 Schematic representation of the questions and potential answers which are fundamentally related to the conundrum of the origin of the homochirality of Life. The review is divided into 4 parts as indicated in the scheme. PVED: Parity-Violating Energy Difference. SMSB: Spontaneous Mirror Symmetry Breaking.

stereochemical imperative.¹⁷ As a matter of example, D-sugars are building blocks of helically-shaped DNA and RNA macromolecules, which store genetic information and encode the synthesis of proteins through the ligation of their constituting amino acids. Glucose monomers in glycogen, starch and cellulose also have a D configuration. This suggests that chirality, structure, and functions of these biomacromolecules are intimately related.¹⁸

In 1857, Louis Pasteur revealed the dramatic difference in the fermentation rate of the two tartaric acid enantiomers with a yeast microorganism, thus uncovering biological enantioselectivity.^{19–21} Pasteur was convinced that chirality was a manifestation of Life, and unsuccessfully looked for the link between physical forces ruling out the Cosmos and the molecular dissymmetry observed in natural products. In 1886, the Italian chemist Arnaldo Piutti²² succeeded to isolate (*R*)-asparagine, mirror-image of the tasteless amino-acid (*S*)-asparagine, and found that it was intensely sweet.²³ These discoveries refer to the link between the handedness of chiral substances and their biological properties but does not explain the origin of biological homochirality (BH).

Despite an extensive literature, the emergence of BH remains a conundrum.^{24–44} The key points of this intricate topic can be summarized as: how, when and where did single chirality appear and eventually lead to the emergence of Life (Figure 1).^{45–48} Along this line, the question of the creation of the original chiral bias appears critical (box “how?” in Figure 1). Huge efforts have been dedicated to decipher which processes may lead to the generation of a chiral bias without the action of pre-existing enantiomerically-enriched chemical species, that is without using the commonly employed routes in stereoselective synthesis. The creation of a chiral bias “from scratch”, often referred to as absolute asymmetric synthesis^{30,31,49,50} and spontaneous deracemization,^{41,51,52} actually encompasses a large variety of phenomena. Here, a distinction can be made between chiral biases that: (i) are inherent to the nature of matter itself, (ii) originate from the

interaction of molecules with physical fields, particles, spins or surfaces, or (iii) emerge from the mutual interaction between molecules (Figure 1). The first category (i) corresponds to the fact that a racemate deviates infinitesimally from its ideal equimolar composition deterministically, *i.e.* in direction of the same enantiomer for a given racemate, as a result of parity violation in certain interactions within nuclei.^{53,54} The second category (ii) refers to natural physical fields (gravitational, magnetic, electric), light and their combinations, which under certain conditions constitute truly chiral fields,³⁰ but also to a range of inherently chiral sources such as chiral light and polarized particles (mostly electrons), polarized electron spins, vortices, or surfaces.⁴⁴ The third category (iii) encompasses processes that lead to the spontaneous emergence of chirality in absence of detectable chiral physical and chemical sources, upon destabilization of the racemic state and stabilization of a scalemic or homochiral state. Such spontaneous mirror symmetry breaking (SMSB) phenomena⁴⁰ involve interactions between molecules through auto-catalytic processes which under far-from-equilibrium conditions may lead to the emergence of enantiopure molecules. The topic has recently undergone significant progress thanks to numerous theoretical models and experimental validations, namely the deracemization of conglomerates through Viedma ripening⁵⁵ and the asymmetric auto-catalysis with the Soai reaction.⁵⁶ Importantly, the plausibility of the aforementioned chirality induction processes in the context of BH will depend on several parameters such as: the extent of asymmetric induction they may provide, their mode of action, *i.e.* if they are unidirectional (deterministic towards a single enantiomer) or bidirectional (leading to either type of enantiomers), their relevance according to prebiotic conditions present on Earth 4 billion years ago, the scope of molecules it could be applied to, and their validation by experimental evidences. The first three parts of this review will provide an updated version of phenomena i-iii that are commonly discussed as plausible sources of asymmetry under prebiotic conditions and can thus

be potentially accountable for the primeval chiral bias in molecules of Life.

However, uncovering plausible mechanisms towards the emergence of a chiral bias is not enough *per se* for elucidating the origin of BH. Additional fundamental challenges such as the extra-terrestrial or terrestrial origin of molecule of Life precursors (box “where?” in Figure 1), the mechanism(s) for the propagation and enhancement of the original chiral bias (box “how?” in Figure 1) and the chemical/biological pathways leading to functional bio-relevant molecules are key aspects to propose a credible scenario. The detection of amino acids and sugars with preferred L and D configuration, respectively, on carbonaceous meteorites⁵⁷ instigated further research for determining plausible mechanisms for the production of chiral molecules in interstellar environment and their subsequent enantiomeric enrichment.^{58,59} Alternatively, hydrothermal vents in primeval Oceans constitute an example of reaction domains often evoked for prebiotic chemistry which may also include potential sources of asymmetry such as high-speed microvortices.⁶⁰ Some mechanisms are known for increasing an existing *e.e.*, such as the self-disproportionation of enantiomers (SDE),⁶¹ non-linear effects in asymmetric catalysis,^{62,63} and stereoselective polymerization.⁶⁴ Noteworthy in the present context, these processes may be applied to increase the optical purity of prebiotically relevant molecules. However, a general amplification scheme which is valid for all molecules of Life is lacking.

The temporal sequence between chemical homochirality, BH and Life emergence is another intricate point (box “when?” in Figure 1). Tentative explanations try to build-up either abiotic theories considering that single chirality is created before the living systems or biotic theories suggesting that Life preceded homochirality.⁴⁴ Purely abiotic theories refer to reactions or physicochemical processes involving low-molecular weight organic molecules presumably present in the prebiotic soup.^{38,65} From a different angle, polymerization of activated building blocks is also discussed as a possible stage for the induction/enhancement of chirality,⁶⁴ even though prebiotic mechanisms towards these essential-to-Life macromolecules remain highly elusive.^{45–48} In the fourth part of this review, we will propose an update of the most plausible chemical and physical scenarios towards BH, with an emphasis on the underlying principles and the experimental evidences, showing merits and limitations of each mechanism. Notably, relevant experimental investigations conducted with building blocks of Life: proteinogenic amino acids, natural sugars, their intermediates or derivatives, will be commented in regards of the different scenarios.

Ultimately, the aim of this literature review is to familiarize the novice with research dealing with BH, and to propose to the expert an updated and timely synopsis of this interdisciplinary field.

1. Parity Violation (PV) and Parity-Violating Energy Difference (PVED)

“Videmus nunc per speculum in aenigmate,” (Holy Bible, I Cor. XIII, 12) which can be translated into “At present, we see indistinctly, as in a mirror” refers to the intuition that a mirror reflection is a distorted representation of the reality. The perception of the different nature of mirror-image objects is also found in the modern literature. In his famous novel “Through the Looking-Glass” by Lewis Carroll Alice raises important questions: ‘How would you like to live in Looking-glass House, Kitty? I wonder if they’d give you milk in there? Perhaps Looking-glass milk isn’t good to drink...’ These sentences refer to the intuition that a mirror reflection is a distorted representation of the reality. The perception of the different nature of mirror-image objects is also found in the modern literature.^{9,24}

The Universe is constituted of elementary particles which interact through fundamental forces, namely the electromagnetic, strong, weak, and gravitational forces. Until the mid-20th century, fundamental interactions were thought to equally operate in a physical system and its image built through space inversion. Indeed, these laws were assumed by physicists to be conserved under the parity operator P (which transforms the spatial coordinates x, y, z into $-x, -y, -z$), *i.e.* parity-even. However, in 1956, Lee and Yang highlighted that parity was only conserved for strong and electromagnetic forces, and proposed experiments to test it for weak interactions.⁶⁶ A few months later, Wu experimentally demonstrated that the parity symmetry is indeed broken in weak forces (which are hereby parity-odd),⁶⁷ by showing that the transformation of unstable ⁶⁰Co nuclei into ⁶⁰Ni, through the β -decay of a neutron into a proton, emit electrons of only *left-handedness*. In fact, solely *left-handed* electrons were emitted since W^+ and W^- bosons (abbreviated as W^\pm bosons), which mediate the weak charged-current interactions, only couple with *left-handed* particles. *Right-handed* particles are not affected by weak interactions carried by W^\pm bosons and consequently, neutrinos, that are only generated by processes mediated by W^\pm bosons, are all *left-handed* in the universe.⁶⁸

The weak neutral current interactions, mediated by the Z^0 boson (sometimes called Z forces), are without charge exchange and, just like the charged ones, violate the parity symmetry.^{69–73} Thus, all weak interactions, carried by W^\pm or Z^0 bosons, break the fundamental parity symmetry.

Parity violation has been observed in nuclear⁶⁷ and atomic Physics.^{74–77} In consequence, the contribution of the Z force between the nuclei and electrons produces an energy shift between the two enantiomers of a chiral molecule. The lower-energy enantiomer would thus be present in slight excess in an equilibrium mixture; this imbalance may provide a clue to the origin of biomolecular homochirality, *i.e.* why chiral molecules usually occur in a single enantiomeric form in nature. Such a tiny parity violation energy difference (PVED of about 10^{-17} kT at 300 K) should be measurable by any absorption spectroscopy provided that ultra-high resolution is reached.^{78–81}

Over the past decades, various experiments have been proposed to observe parity violation in chiral molecules, including measurements of PV frequency shifts in NMR spectroscopy,⁸² measurements of the time dependence of

optical activity,⁸³ and direct measurement of the absolute PV energy shift of the electronic ground state.^{79–81,84}

However, it has never been unequivocally observed at the molecular level to date. Note that symmetry violation of time reversal (T) and of charge parity (CP) is actually recovered in the CPT symmetry, *i.e.*, in the “space-inverted anti-world made of antimatter”.⁸⁵ Quantitative calculations of this parity-violating energy difference between enantiomers have been improved during the last four decades,^{86–90} to give for example about 10^{-12} J/mol for CHFCIBr .^{91,92} Although groups of Crassous/Darquié in France^{78,93–98} and Quack in Switzerland^{99–103} have been pursuing an experimental effort to measure PVED, thanks to approaches based on spectroscopic techniques and/or tunneling processes, no observation has unambiguously confirmed it yet. However, thanks to the combination of the contribution from the weak interaction Hamiltonian (Z^3) and from the spin orbit coupling (Z^2), the parity violating energy difference strongly increases with increasing nuclear charge with a commonly accepted Z^5 scaling law, thus chiral heavy metal complexes might be favourable candidates for future observation of PV effects in chiral molecules.^{94,96} Other types of experiments have been proposed to measure PV effects, such as nuclear magnetic resonance (NMR), electron paramagnetic resonance (EPR), microwave (MW) or Mössbauer spectroscopy.⁷⁹ Note that other phenomena have been taken into consideration to measure PVED such as in Bose-Einstein condensation, but those were not conclusive.^{104,105}

The tempting idea that PVED could be the source of the tiny enantiomeric excess amplified to the asymmetry of Life was put forward by Ulbricht in 1959^{106,107} and by Yamagata in 1966.¹⁰⁸ With this in mind, Mason, Tranter and MacDermott^{109–122} defended in the eighties and early nineties that (*S*)-amino acids, *D*-sugars, α -helix or β -sheet secondary structures, as well as other natural products and secondary structures of biological importance, are more stable than their enantiomorph due to PVED.⁵⁴ However, Quack^{89,123} and Schwerdtfeger^{124,125} independently refuted these results on the strength of finer calculations, and Lente^{126,127} asserted that a PVED of around 10^{-13} J/mol causes an excess of only 6×10^6 molecules in one mole (against 1.9×10^{11} for the standard deviation). In reply, MacDermott claimed, by means of a new generation of PVED computations, that the enantiomeric excess of four gaseous amino acids found in the Murchison meteorite (in the solid state) could originate from their PVED.^{128,129} Whether PVED could have provided a sufficient bias for the emergence of BH likely depends on the related amplification mechanism, a point that will be discussed into more details in part 3.

2. Chiral fields

Physical fields, polarized particles, polarized spins and surfaces are commonly discussed as potential chiral inducers of enantiomeric excesses in organic molecules. The aim of this part is to present selected chiral fields along with experimental

observations which are relevant in the context of elucidating BH.

2.1 Physical fields

a True and False Chirality

Chirality's definitions based on symmetry arguments are adequate for stationary objects, but not when motion comes into play. To address the potential chiral discriminating nature of physical fields, Barron defined true and false chirality as follows: the “true chirality is shown by systems existing in two distinct enantiomeric states that are interconverted by space inversion (P), but not by time reversal (T) combined with any proper spatial rotation (R_π)”.¹³⁰ Along this line, a stationary and a translating rotating cone are prototypical representations of false and true chirality, respectively (Figure 2a). Cones help to better visualize the true chiral nature of vortices, but the concept is actually valid for any translating spinning objects, *e.g.* photons and electrons.^{85,131} All experimental attempts to produce any chiral bias using a static uniform magnetic or electric field, or unpolarized light failed and this can be explained by the non-chiral nature of these fields.^{30,31,49} In addition, the parallel or antiparallel combination of static uniform magnetic and electric fields constitute another example of false chirality (Figure 2b).³⁰

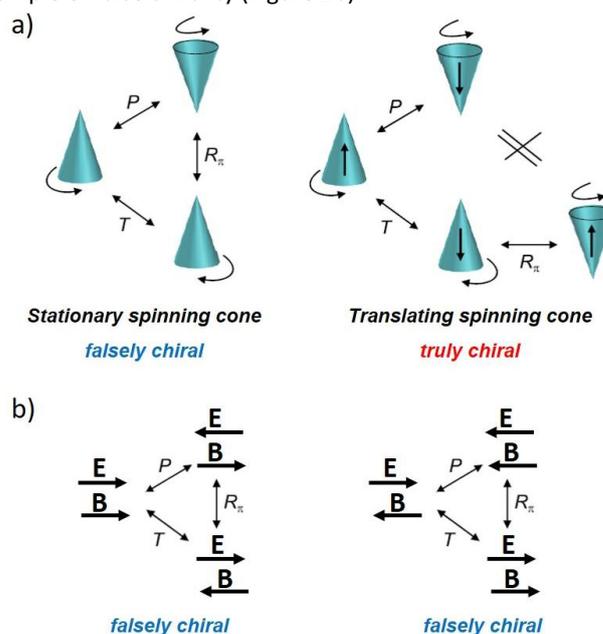


Figure 2 Distinction between “true” and “false” chirality by considering the effect of parity (P) and time (T) reversal on spinning cones (a) and aligned magnetic and electric fields (b). Figure 2a is reprinted from reference¹³¹ with permission from American Chemical Society. Figure 2b is adapted from reference³⁰ with permission from American Chemical Society.

Importantly, only when interacting with a truly chiral system the energy of enantiomeric probes can be different (corresponding to diastereomeric situations), while no loss of degeneration in energy levels can happen in a falsely chiral system; however, asymmetry could be obtained for processes out of thermodynamic equilibrium.^{30,31} Based on these definitions, truly chiral forces may lift the degeneracy of enantiomers and induce enantioselection in a reaction system reaching its stationary state, while an influence of false

chirality is only possible for kinetically controlled reaction outputs, since in this case the enantiomers remain strictly degenerate and only the breakdown of the reaction path microreversibility occurs.⁴¹ Furthermore, the extent of chiral induction that can be achieved by a chiral physical field is intimately related to the nature of its interaction with matter, *i.e.* with prebiotically relevant organic molecules in the context of BH. A few examples of physical fields for absolute asymmetric synthesis are mentioned in the next paragraphs.

b Magneto-chiral effects

A light beam of arbitrary polarization (with k as wavevector) propagating parallel to a static magnetic field (B) also possesses true chirality ($k \cdot B$), exploited by the magneto-chiral dichroism (MChD, Figure 3a).¹³² MChD was first observed by Rikken and Raupach in 1997 for a chiral europium(III) complex, and was further extended to other metal compounds and a few aggregates of organic molecules.^{132–136} Photoresolution of Δ - and Λ -chromium(III) tris(oxalato) complexes thanks to magneto-chiral anisotropy was accomplished in 2000 by the same authors,¹³⁷ with an enantioenrichment proportional to the magnetic field, *e.e.*/ B being equal to $1 \times 10^{-5} \text{ T}^{-1}$ (Figure 4b).

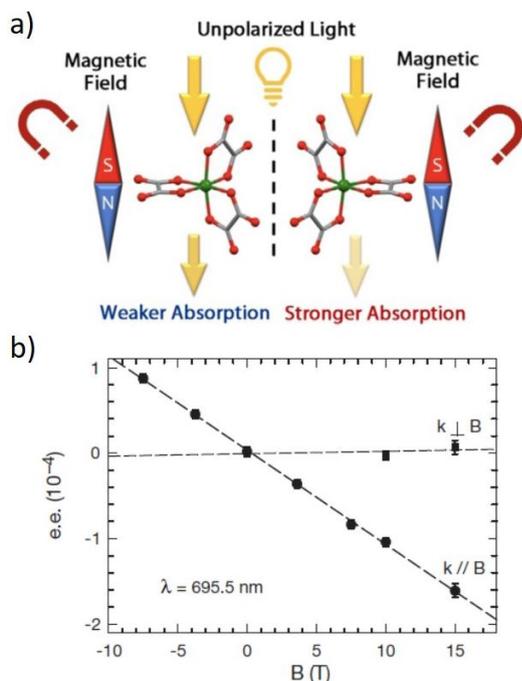


Figure 3 a) Schematic representation of MChD for a racemate of a metal complex: the unpolarised light is preferentially absorbed by the Δ versus Λ enantiomers. Reprinted from ref.¹³⁶ with permission from Wiley-VCH. b) Photoresolution of the chromium(III) tris(oxalato) complex. Plot of the *e.e.* after irradiation with unpolarised light for 25 min at $\lambda = 695.5 \text{ nm}$, as a function of magnetic field, with an irradiation direction k , either parallel or perpendicular to the magnetic field. Reprinted from reference¹³⁷ with permission from Nature publishing group.

c Mechanical chiral interactions

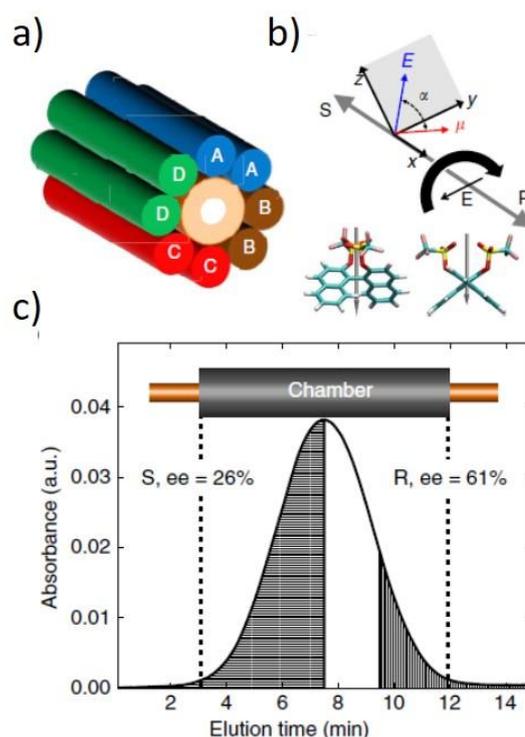


Figure 4 a) Schematic representation of the experimental set-up for the separation of chiral molecules placed in a microfluidic capillary surrounded by rotating electric fields (A-D electrodes). b) Expected directions of motion for the enantiomers of 1,1'-bi-2-naphthol bis(trifluoromethanesulfonate) for the indicated direction of rotation of the REF (curved black arrow). α is the relative angle between the electric dipole moment and electric field. The grey arrows show the opposite directions of motion for the enantiomers. c) Absorbance chromatogram from the in-line detector of a slug of (*rac*)-1,1'-bi-2-naphthol bis(trifluoromethanesulfonate) after exposure to clockwise REF for 45 h. The sample collected from the shaded left side of the chromatogram had *e.e.* of 26% in favour of the (*S*) enantiomer, while the right shaded section of the chromatogram had *e.e.* of 61% for the (*R*) enantiomer. Reprinted from reference¹³⁵ with permission from Nature publishing group.

Whilst mechanical interactions of chiral objects with their environment is well established at the macroscale, the ability of

these interactions to mediate the separation of molecular enantiomers remains largely under-explored.¹³⁹ A few experimental reports indicate that fluid flows can discriminate not only large chiral objects,^{140–142} but also helical bacteria,¹⁴³ colloidal particles,¹⁴⁴ and supramolecular aggregates.^{145,146} It has been indeed found that vortices, being induced by stirring, microfluidics or temperature gradients, are capable of controlling the handedness of supramolecular helical assemblies.^{60,145–159}

Laminar vortices have been recently employed as the single chiral discriminating source for the emergence of homochiral supramolecular gels in milliseconds.⁶⁰ High speed vortices have been evoked as potential sources of asymmetry present in hydrothermal vents, presumed key reaction sites for the generation of prebiotic molecules. However, the propensity of shear flow to prevent the Brownian motion and allow for the discrimination of small molecules remain to be demonstrated. Grzybowski and co-workers showed that *s*-shaped μm -size particles located at the oil/air interface parallel to the shear plane migrate to different positions in a Couette cell.¹⁶⁰ The proposed chiral drift mechanism may in principle allow the separation of smaller chiral objects, with size on the order of

the ten of nanometres. In 2015, a new molecular parameter called hydrodynamic chirality was introduced to characterize the coupling of rotational motion of a chiral molecule to its translational motion and quantify the direction and velocity of such motion.¹³⁸ The concept concerns the possibility to control the motion of chiral molecules by orienting and aligning their dipole moment with the electric field position leading to their rotation. The so-called molecular propeller effect allows enantiomers of two binaphthyl derivatives, upon exposition to rotating electric fields (REF), to propel in opposite directions, leading to a local enrichment of up to 60% *e.e.* (Figure 4). It would be essential to probe interactions of vortices, shear flows and rotating physical fields with biologically relevant molecules in order to uncover whether it could have played a role in the emergence of a chiral bias on early Earth.

d Combined action of gravity, magnetic field and rotation

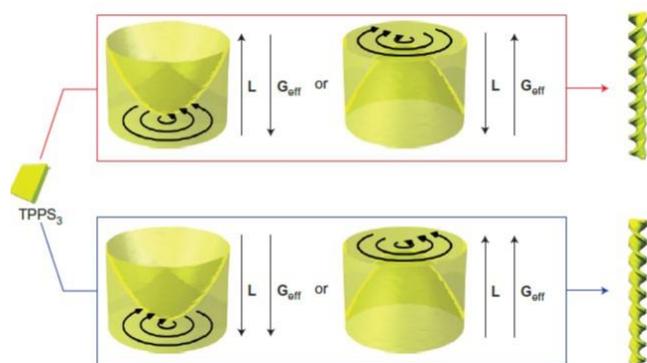


Figure 5 Control of the handedness of TPPS₃ helical assemblies by the relative orientation of the angular momentum of the rotation (L) and the effective gravity (G_{eff}). TPPS₃: tris-(4-sulfonatophenyl)phenyl porphyrin. Reprinted from reference¹⁶¹ with permission from Nature publishing group.

Micali et al. demonstrated in 2012 that the combination of gravity, magnetic field and rotation can be used to direct the handedness of supramolecular helices generated upon assembly of an achiral porphyrin monomer (TPPS₃, Figure 5).¹⁶¹ It was presumed that the enantiomeric excess generated at the onset of the aggregation was amplified by autocatalytic growth of the particles during the elongation step. The observed chirality is correlated to the relative orientation of the angular momentum and the effective gravity, the direction of the former being set by clockwise or anticlockwise rotation. The role of the magnetic field is fundamentally different to that in MChD effect (2.1 a) since its direction does not influence the sign of the chiral bias. Its role is to provide tunable magnetic levitation force and alignment of the supramolecular assemblies. These results therefore seem to validate experimentally the prediction by Barron that falsely chiral influence may lead to absolute asymmetric synthesis after enhancement of an initial chiral bias created under far-from-equilibrium conditions.¹³⁰ According to the authors, control experiments performed in absence of magnetic field discard macroscopic hydrodynamic chiral flow, *i.e.* a true chiral force (see 2.1 c), as the driving force for chirality induction; a point that has been recently disputed by other authors.⁴¹ Whatever the true or false nature of the combined action of

gravity, magnetic field and rotation, its potential connection to BH is hard to conceive at this stage.

e Through plasma-triggered chemical reactions

Plasma, produced by the impact of extra-terrestrial objects on Earth, has been investigated as a potential source of asymmetry. Price and Furukawa teams reported, in 2013 and 2015, respectively, that nucleobases and/or proteinogenic amino acids were formed under conditions which presumably reproduced the conditions of impact of celestial bodies on primitive Earth.^{162,163} When shocked with a steel projectile fired at high velocities in a light gas gun, ice mixtures made of NH₄OH, CO₂ and CH₃OH, were found to produce equal amounts of (*R*)- and (*S*)-alanine, α -aminoisobutyric acid and isovaline as well as their precursors.¹⁶² Importantly, only the impact shock is responsible for the formation of amino-acids because post-shot heating is not sufficient. A richer variety of organic molecules, including nucleobases, was obtained by shocking ammonium bicarbonate solution under nitrogen (representative of the Hadean ocean and its atmosphere) with various metallic projectiles (as simplified meteorite materials).¹⁶³ The production of amino-acids is correlated to the concentration of ammonium bicarbonate concentration, acting as the C1-source. The attained pressure and temperature (up to 60 GPa and thousands Kelvin) allowed chemical reactions to proceed, as well as racemization as evidenced later,¹⁶⁴ but were not enough to trigger plasma processes. A meteorite impact was reproduced in the laboratory by Wurz and co-workers in 2016,¹⁶⁵ by firing projectiles of pure ¹³C synthetic diamond to a multilayer target consisting of ammonium nitrate, graphite and steel. The impact generated a pressure of 170 GPa and a temperature of 3 to 4 × 10⁴ K, enough to form a plasma torch, through the interaction between the projectile and target materials and their subsequent atomization and ionization. The most striking result is certainly the formation of ¹³C-enriched alanine which is claimed to be obtained with *e.e.* values ranging from 7 to 25%. The exact source of asymmetry is uncertain: the far-from-equilibrium nature of the plasma-triggered reactions and the presence of spontaneously generated electromagnetic fields in the reactive plasma torch may have led to the observed chiral biases.¹⁶⁶ This first report of an impact-produced enantioenrichment needs to be confirmed experimentally and supported theoretically.

2.2 Polarized radiations and spins

a Circularly Polarized Light (CPL)

A long time before the discussions on the true or false chiral nature of physical fields, Le Bel and van't Hoff already proposed, at the end of the nineteenth century, to use circularly polarized light, a truly chiral electromagnetic wave existing in two enantiomorphic forms (*i.e.* the *left*- and *right*-handed CPL), as chiral bias to induce enantiomeric excess.^{31,167–169} Cotton strengthened this idea in 1895,^{170–172} when he reported the circular dichroism (CD) of an aqueous solution of potassium chromium(III) tartrate.

Circular dichroism is a phenomenon, corresponding to the differential absorption of *l*-CPL and *r*-CPL at a given wavelength

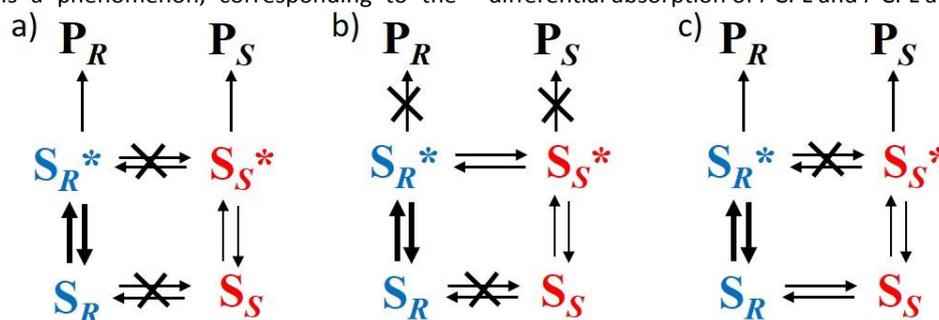


Figure 6 Simplified kinetic schemes for asymmetric (a) photolysis, (b) photoresolution and (c) photosynthesis with CPL. S_R and S_S are substrate enantiomers and S_R^* and S_S^* are their photoexcited states. P_R and P_S are the products generated from the respective photoexcited states. The thick line represents the preferential absorption of CPL by one of the enantiomers. $[S_S] > [S_R]$ for asymmetric photolysis and photoresolution processes whilst $[P_R] > [P_S]$ for asymmetric photosynthesis.

in the absorption region of an optically active material, as well as the spectroscopic method that measures it.^{173,174} Enantiomers absorbing CPL of one handedness constitute non-degenerated diastereoisomeric systems, based on the interaction between two distinct chiral influences, one chemical and the other physical. Thus, one state of this system is energetically favoured, and one enantiomer preferentially absorbs CPL of one polarization state (*l*- or *r*-CPL).

The dimensionless Kuhn anisotropy (or dissymmetry) factor g allows to quantitatively describe the chiroptical response of enantiomers (**Equation 1**). The Kuhn anisotropy factor is expressed by the ratio between the difference in molar extinction coefficients of *l*-CPL and *r*-CPL ($\Delta\epsilon$), and the global molar extinction coefficient (ϵ), where ϵ_L and ϵ_R are the molar extinction coefficients for *left*- and *right*-handed CPL, respectively.¹⁷⁵ It ranges from -2 to +2, for a total absorption of *right*- and *left*-handed CPL, respectively, and is wavelength-dependent. Enantiomers have equal but opposite g values, corresponding to their preferential absorption of one CPL handedness.

$$g = \frac{\Delta\epsilon}{\epsilon} = \frac{\epsilon_L - \epsilon_R}{\left(\frac{\epsilon_L + \epsilon_R}{2}\right)} \quad (1)$$

The preferential excitation of one over the other enantiomer in presence of CPL allows the emergence of a chiral imbalance from a racemate (by asymmetric photoresolution or photolysis) or from rapidly interconverting chiral conformations (by asymmetric photosynthesis).^{176–178}

Asymmetric *photolysis* is based on the irreversible photochemical consumption of one enantiomer at a higher rate, within a racemic mixture which does not racemize during the process (Figure 6a). In most cases, the (enantio-enriched) photo products are not identified. Thereby, the enantioenrichment comes from the accumulation of the slowly reacting enantiomer. It depends both on the unequal molar extinction coefficients (ϵ_R and ϵ_S) for CPL of the (*R*)- and (*S*)-enantiomers, governing the different rate constants, as well as the extent of reaction ξ . Asymmetric *photoresolution* occurs within a mixture of enantiomers that interconvert in their excited states (Figure 6b). Since the reverse reactions, from the excited to the ground states, should not be enantiodifferentiating, the deviation from the racemic mixture

is only due to the difference of extinction coefficients (ϵ_R and ϵ_S). While the total concentration in enantiomers ($C_R + C_S$) is constant during the photoresolution, the photostationary state (p_{ss}) is reached after a prolonged irradiation irrespective of the initial enantiomeric composition.¹⁷⁷ In absence of side reactions, the p_{ss} is reached for $\epsilon_R C_R = \epsilon_S C_S$, which allows to determine *e.e.*_{*pss*}, *e.e.* at the photostationary state, as being equal to $(C_R - C_S)/(C_R + C_S) = g/2$. Asymmetric *photosynthesis* or *asymmetric fixation* produces an enantio-enriched product by preferentially reacting one enantiomer of a substrate undergoing fast racemization (Figure 6c). Under these conditions, the (*R*)/(*S*) ratio of the product is equal to the excitation ratio ϵ_R/ϵ_S , and the *e.e.* of the photoproduct is thus equal to $g/2$. The chiral bias which can be reached in asymmetric photosynthesis and photoresolution processes is thus related to the g value of enantiomers, whereas the *e.e.* in asymmetric photolysis is influenced by both g and ξ values.

The first CPL-induced asymmetric partial resolution dates back to 1968 thanks to Stevenson and Verdick, who worked with octahedral oxalato complexes of chromium(III).¹⁷⁹ Asymmetric photoresolution was further investigated for small organic molecules,^{180,181} macromolecules¹⁸² and supramolecular assemblies.¹⁸³ A number of functional groups such as overcrowded alkene, azobenzene, diarylethene, α,β -unsaturated ketone or fulgide were specifically-designed to enhance the efficiency of the photoresolution process.⁵⁸

Kagan *et al.* pioneered the field of asymmetric photosynthesis with CPL in 1971, through examining hexahelicene photocyclization in the presence of iodine.¹⁸⁴ The following year, Calvin *et al.* reported *e.e.* up to 2% for an octahelicene produced under similar conditions.¹⁸⁵ Enantioenrichment by photoresolution and photosynthesis with CPL is limited in scope, since it requires molecules with high g values to be detected, and in intensity, since it is limited to $g/2$.

Since its discovery by Kuhn *et al.* ninety years ago,^{186,187} through the enantioselective decomposition of ethyl- α -bromopropionate and *N,N*-dimethyl- α -azidopropionamide, the asymmetric photolysis of racemates has attracted a lot of interest. In the common case of two competitive pseudo-first order photolytic reactions with unequal rate constants, k_S and k_R , for the (*S*) and (*R*) enantiomers, respectively, and if the anisotropies are close to zero, the enantiomeric excess

induced by asymmetric photolysis can be approximated as Equation 2:¹⁸⁸

$$e.e. \geq \left(1 - (1 - \xi)^{\frac{g}{2}}\right) \times 100 (\%) \quad (2)$$

where ξ is the extent of reaction.

In 1974, the asymmetric photodecomposition of racemic camphor reported by Kagan *et al.* reached 20% *e.e.* at 99% completion, a long-lasting record in this domain.¹⁸⁹ Three years later, Norden¹⁹⁰ and Bonner *et al.*¹⁹¹ independently showed that enantioselective photolysis by UV-CPL was a viable source of symmetry-breaking for amino acids, by inducing *e.e.* up to 2% in aqueous solutions of alanine and glutamic acid,¹⁹¹ or 0.2% with leucine.¹⁹⁰ Leucine was then intensively studied thanks to a high anisotropy factor in the UV region.¹⁹² The *e.e.* was increased up to 1.3% in 2001 ($\xi = 0.55$) by Inoue *et al.* by exploiting the pH-dependence of the g value.^{193,194} Since the early 2000s, Meierhenrich *et al.* got closer to astrophysically relevant conditions by irradiating samples in the solid state with synchrotron vacuum ultraviolet (VUV)-CPL (below 200 nm). It made it possible to avoid the water absorption in the VUV, and allowed to reach electronic transitions having higher anisotropy factors (Figure 7).¹⁹⁵ In 2005, a solid racemate of leucine was reported to reach 2.6% of *e.e.* after illumination with *r*-CPL at 182 nm (ξ not reported).¹⁹⁶ More recently, the same team improved the selectivity of the photolysis process thanks to amorphous samples of finely-tuned thickness, providing *e.es.* of $5.2 \pm 0.5\%$ and $4.2 \pm 0.2\%$ for leucine,¹⁹⁷ and alanine,^{198,199} respectively. A similar enantioenrichment was reached in 2014 with gaseous photoionized alanine,²⁰⁰ which constitutes an appealing result taking into account the detection of interstellar gases such as propylene oxide²⁰¹ and glycine²⁰² in star-forming regions.

Important studies in the context of BH reported the direct formation of enantio-enriched amino acids generated from simple chemical precursors, when illuminated with CPL. Takano *et al.* showed in 2007 that eleven amino acids could be generated upon CPL irradiation of macromolecular compounds, originating from proton-irradiated gaseous mixtures of CO, NH₃ and H₂O.²⁰³ Small *e.es.*, $+0.44 \pm 0.31\%$ and $-0.65 \pm 0.23\%$, were detected for alanine upon irradiation with *r*- and *l*-CPL, respectively. Nuevo *et al.* irradiated interstellar ice analogues composed of H₂O, ¹³CH₃OH and NH₃ at 80 K with CPL centred at 187 nm, which led to the formation of alanine with an *e.e.* of $1.34 \pm 0.40\%$.²⁰⁴ The same team also studied the effect of CPL on regular ice analogues or organic residues coming from their irradiation in order to mimic the different stages of asymmetric

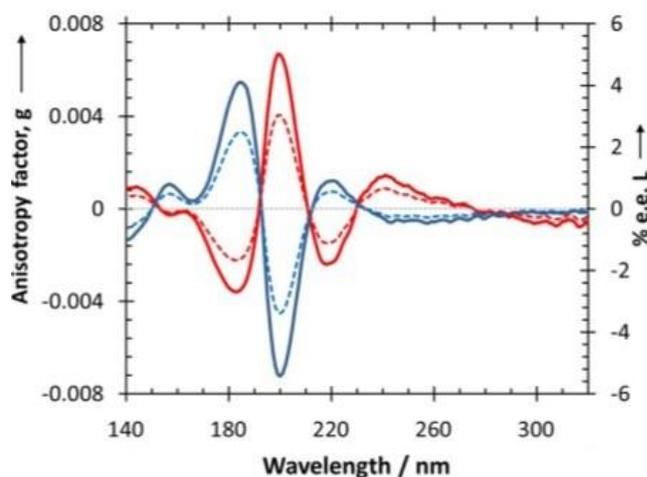


Figure 7 Anisotropy spectra (thick lines, left ordinate) of isotropic amorphous (*R*)-alanine (red) and (*S*)-alanine (blue), in the VUV and UV spectral region. Dashed lines represent the enantiomeric excess (right ordinate) that can be induced by photolysis of *rac*-alanine with either *left*- (in red) or *right*- (in blue) circularly polarized light at $\xi = 0.9999$. Positive *e.e.* value corresponds to scalemic mixture biased in favour of (*S*)-alanine. Note that enantiomeric excesses are calculated from Equation (2). Reprinted from reference¹⁹⁵ with permission from Wiley-VCH.

induction in interstellar ices.²⁰⁵ Sixteen amino acids were identified, and five of them (including alanine and valine) were analysed by enantioselective two-dimensional gas chromatography, GC×GC,²⁰⁶ coupled to TOF mass spectrometry, to show enantioenrichments up to $2.54 \pm 0.28\%$ *e.e.* Optical activities likely originated from the asymmetric photolysis of the amino acids initially formed as racemates. Advantageously, all five amino acids exhibited *e.e.* of identical sign for a given polarization and wavelength, suggesting that irradiation by CPL could constitute a general route towards amino acids with a single chirality. Even though the chiral biases generated upon CPL irradiation are modest, these values can be significantly amplified through different physicochemical processes, notably those including autocatalytic pathways (see parts 3 and 4).

b Spin-polarized particles

In the cosmic scenario, it is believed that the action of polarized quantum radiation in space, such as circularly polarized photons or spin-polarized particles, may have induced asymmetric conditions in the primitive interstellar media resulting in terrestrial bioorganic homochirality. In particular, nuclear-decay- or cosmic-ray-derived leptons (*i.e.* electrons, muons, and neutrinos) in nature have a specified helicity, that is, they have a spin angular momentum polarized parallel or antiparallel to their kinetic momentum due to parity violations (PV) in the weak interaction (part 1).

Of the leptons, electrons are one of the most universally present particles in ordinary materials. Spin-polarized electrons in nature are emitted with β^- decay from radioactive nuclear particles derived from PV involving the weak nuclear interaction and spin-polarized positrons (the anti-particle of electrons) from β^+ decay. In β^-/β^+ decay, with the weak interaction, the spin angular momentum vectors of electron/positron are perfectly polarized as antiparallel/parallel to the vector direction of the kinetic momentum. In this meaning, spin-polarized

electrons/positrons are “chiral radiation”, as well as are muons and neutrinos, which will be mentioned below. It is expected that the spin-polarized leptons will induce reactions different from those triggered by CPL. For example, β^- decay from ^{60}Co is accompanied by circularly polarized gamma-rays.²⁰⁷ Similarly, spin-polarized muon irradiation has the potential to induce novel types of optical activities different from those of polarized photon and spin-polarized electron irradiation.

Single-handed polarized particles produced by supernovae explosions may thus interact with molecules in the proto-solar clouds.^{35,208–210,207} Left-handed electrons generated by β^- -decay impinge on matter to form a polarized electromagnetic radiation through *bremstrahlung*. At the end of fifties, Vester and Ulbricht suggested that these circularly-polarized “Bremsstrahlen” photons can induce and direct asymmetric processes towards a single direction upon interaction with organic molecules.^{107,211} From the sixties to the eighties,^{212–220}

many experimental attempts to show the validity of the “V–U hypothesis”, generally by photolysis of amino acids in presence of a number of β -emitting radionuclides or through self-irradiation of ^{14}C -labeled amino acids, only led to poorly conclusive results.^{44,221,222} During the same period, the direct effect of high-energy spin-polarized particles (electrons, protons, positrons and muons) have been probed for the selective destruction of one amino acid enantiomer in a racemate, but without further success as reviewed by Bonner.^{44,54} More recent investigations by the international collaboration RAMBAS (Radiation Mechanism of Biomolecular ASymmetry) claimed minute *e.es.* (up to 0.005%) upon irradiation of various amino acid racemates with (natural) left-handed electrons.^{223,224}

Other fundamental particles have been proposed to play a key role in the emergence of BH.^{207,209,225} Amongst them, electron antineutrinos have received particular attention, through the Supernova Neutrino Amino Acid Processing (SNAAP) model.^{226–228} Electron antineutrinos are emitted after a supernova explosion, to cool the nascent neutron star, and, by a similar reasoning to that applied with neutrinos, they are all right-handed.

According to the SNAAP scenario, right-handed electron antineutrinos generated in the vicinity of neutron stars with strong magnetic and electric fields were presumed to selectively transform ^{14}N into ^{14}C , and this process depended on whether the spin of the ^{14}N was aligned or anti-aligned with that of the antineutrinos. Calculations predicted enantiomeric excesses for amino acids from 0.02% to a few percent, and a preferential enrichment in (*S*)-amino acids. No experimental evidences have been reported to date in favour of a deterministic scenario for the generation of a chiral bias in prebiotic molecules.

c Chirality Induced Spin Selectivity (CISS)

An electron in helical roto-translational motion with spin–orbit coupling (*i.e.* translating in a “ballistic” motion with its spin projection parallel or antiparallel to the direction of propagation) can be regarded as chiral, existing as two possible enantiomers, corresponding to the α and β spin

configurations, which do not coincide upon space *and* time inversion. Such

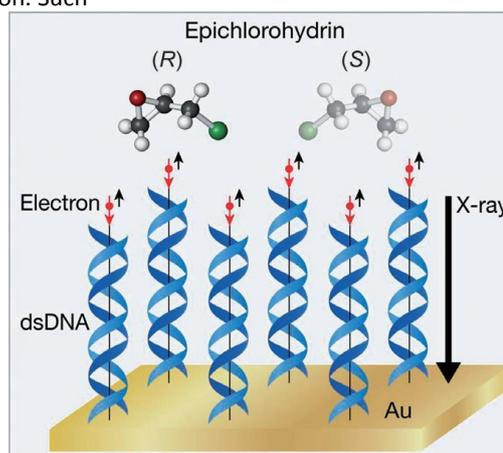


Figure 8 Enantioselective dissociation of epichlorohydrin by spin polarized SEs. Red (black) arrows indicate the electron's spin (motion direction, respectively). Reprinted from reference²²⁹ with permission from Wiley-VCH.

peculiar “chiral actor” is the object of spintronics, the fascinating field of modern physics which deals with the active manipulation of spin degrees of freedom of charge carriers.²³⁰ The interaction between polarized spins of secondary electrons (SEs) and chiral molecules leads to chirality induced spin selectivity (CISS), a recently reported phenomenon.

In 2008, Rosenberg *et al.*²³¹ irradiated adsorbed molecules of (*R*)-2-butanol or (*S*)-2-butanol on a magnetized iron substrate with low-energy SEs (10–15% of spin polarization) and measured a difference of about ten percent in the rate of CO bond cleavage of the enantiomers. Extrapolations of the experimental results suggested that an *e.e.* of 25% would be obtained after photolysis of the racemate at 98.6% of conversion. Importantly, the different rates in the photolysis of the 2-butanol enantiomers depend on the spin polarization of the SE, showing the first example of CISS.^{232–234} Later, SEs with a higher degree of spin polarization (60%) were found to dissociate Cl from epichlorohydrin (Epi) with a quantum yield 16% greater for the *S* form.²²⁹ To achieve this, electrons are produced by X-ray irradiation of a gold substrate and spin-filtered by a self-assembled overlayer of DNA before they reach the adlayer of Epi (Figure 8).

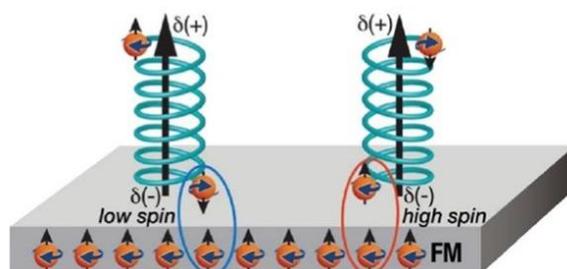


Figure 9 Scheme of the enantiospecific interaction triggered by chiral-induced spin selectivity. Enantiomers are sketched as opposite green helices, electrons as orange spheres with straight arrows indicating their spin orientation, which can be reversed for surface electrons by changing the magnetization direction. In contact with the perpendicularly magnetized FM surface, molecular electrons are redistributed to form a dipole, the spin orientation at each pole depending on the chiral potentials of enantiomers. The interaction between the FM

substrate and the adsorbed molecule (circled in blue and red) is favoured when the two spins are antiparallel leading to the preferential adsorption of one enantiomer over the other. Reprinted from reference²³⁵ with permission from the American Association for the Advancement of Science

In 2018, Banerjee-Ghosh *et al.* showed that a magnetic field perpendicular to a ferromagnetic (FM) substrate can generate enantioselective adsorption of polyalanine, *ds*-DNA and cysteine.²³⁵ One enantiomer was found to be more rapidly adsorbed on the surface depending on the magnetization direction (Figure 9). The effect is not attributed to the magnetic field *per se* but to the exchange interaction between the adsorbed molecules and surface electrons spins, *i.e.* CISS.

Enantioselective crystallization of initially racemic mixtures of asparagine, glutamic acid, and threonine, known to crystallize as conglomerates, was also observed on a ferromagnetic substrate surface (Ni(120 nm)/Au(10 nm)).²³⁰ The racemic mixtures were crystallized from aqueous solution on the ferromagnetic surfaces in the presence of two magnets, one pointing north and the other south, located at different sites of the surface. A clear enantioselective effect was observed in the formation of an excess of *d*- or *l*-crystals, depending on the direction of the magnetization orientation.

In 2020, the CISS effect was successfully applied to several asymmetric chemical processes, SEs acting as chiral reagents.²³⁶ Spin-polarized electrons, produced by a magnetized Ni/Au substrate coated with an achiral self-assembled monolayer (SAM) of carboxyl-terminated alkanethiols [HS-(CH₂)_{x-1}-COO⁻], caused an enantiospecific association of 1-amino-2-propanol enantiomers, leading to an *e.e.* of 20% in the reactive medium. The enantioselective electro-reduction of (1*R*/1*S*)-10-camphorsulfonic acid (CSA) into isborneol was also governed by the spin orientation of SEs, injected through an electrode, with an *e.e.* of about 11.5% after the electrolysis of 80% of the initial amount of CSA.

Electrochirogenesis links CISS process to biological homochirality through several theories, all based on an initial bias stemming from spin polarized electrons.^{232,237} Strong fields and radiations of neutron stars could align ferrous magnetic domains in interstellar dust particles, and produce spin-polarized electrons, able to create an enantiomeric excess into adsorbed chiral molecules. One enantiomer from a racemate in a cosmic cloud would merely accrete on a magnetized domain in an enantioselective manner as well. Alternatively, magnetic minerals of the prebiotic world, like pyrite (FeS₂) or greigite (Fe₃S₄), might serve as an electrode in the asymmetric electrosynthesis of amino acids or purines, or as spin-filter in the presence of an external magnetic field, *e.g.* in hydrothermal vents.

2.3 Chiral surfaces

a Inorganic minerals and clays

Chiral surfaces constitute unequivocal true asymmetry sources that are widespread over the Earth surface under the form of various minerals, α -quartz, calcite, gypsum, and some clays notably. The implication of chiral surfaces in the context of BH has been debated^{44,238-242} along two main axes: (i) the preferential adsorption of prebiotically relevant molecules, and (ii) the potential unequal distribution of *left*-handed and

right-handed surfaces for a given mineral or clay on the Earth surface.

Selective adsorption is generally the consequence of reversible and preferential diastereomeric interactions between the chiral surface and one of the enantiomers,²³⁹ commonly described by the simple three-point model. But this model, assuming that only one enantiomer can present three groups that match three active positions of the chiral surface,²⁴³ fails to fully explain chiral recognition which are the fruit of more subtle interactions.²⁴⁴ In the second part of the XXth century, a large number of studies have focused on demonstrating chiral interactions between biological molecules and inorganic mineral surfaces.

Quartz is the only common mineral which is composed of enantiomorphic crystals. *Right*-handed (*d*-quartz) and *left*-handed (*l*-quartz) can be separated (similarly to the tartaric acid salts of the famous Pasteur experiment) and investigated independently in adsorption studies of organic molecules. The process of separation is made somewhat difficult by the presence of "Brazilian twins" (also called chiral or optical twins),²⁴² which might bias the interpretation of the experiments. Bonner *et al.* in 1974^{245,246} measured the differential adsorption of alanine derivatives defined as % adsorbed on *d*-quartz – % adsorbed on *l*-quartz. These authors reported on the small but significant $1.4 \pm 0.4\%$ preferential adsorption of (*R*)-alanine over *d*-quartz and (*S*)-alanine over *l*-quartz, respectively. A more precise evaluation of the selectivity with radiolabelled (*RS*)-alanine hydrochloride led to higher levels of differential adsorption between *l*-quartz and *d*-quartz (up to 20%).²⁴⁷ The hydrochloride salt of alanine isopropyl ester was also found to be adsorbed enantiospecifically from its chloroform solution leading to chiral enrichment varying between 1.5 and 12.4%.²⁴⁸ Furuyama and co-workers also found preferential adsorption of (*S*)-alanine and (*S*)-alanine hydrochloride over *l*-quartz from their ethanol solutions.^{249,250} Anhydrous conditions are required to get sufficient adsorption of the organic molecules onto α -quartz crystals which, according to Bonner, discards α -quartz as a suitable mineral for the deracemization of building blocks of Life.²⁵¹ According to Hazen and Scholl,²³⁹ the fact that these studies have been conducted on powdered quartz crystals (*i.e.* polycrystalline quartz) have hampered a precise determination of the mechanism and magnitude of adsorption on specific surfaces of α -quartz. Some of the faces of quartz crystals likely display opposite chiral preferences which may have reduced the experimentally-reported chiral selectivity. Moreover, chiral indices of the commonest crystal growth surfaces of quartz, as established by Downs and Hazen, are relatively low (or zero), suggesting that potential of enantiodiscrimination of organic molecules by quartz is weak in overall.²⁵² Quantum-mechanical studies using density functional theories (DFT) have also been performed to probe the enantiospecific adsorption of various amino acids on hydroxylated quartz surfaces.²⁵³⁻²⁵⁶ In short, the computed differences in the adsorption energies of the enantiomers are modest (on the order of 2 kcal.mol⁻¹ at best) but strongly depend on the nature of amino acids and quartz surfaces. A

final argument against the implication of quartz as a deterministic source of chiral discrimination of the molecules of Life comes from the fact that *d*-quartz and *l*-quartz are equally distributed on Earth.^{257,258}

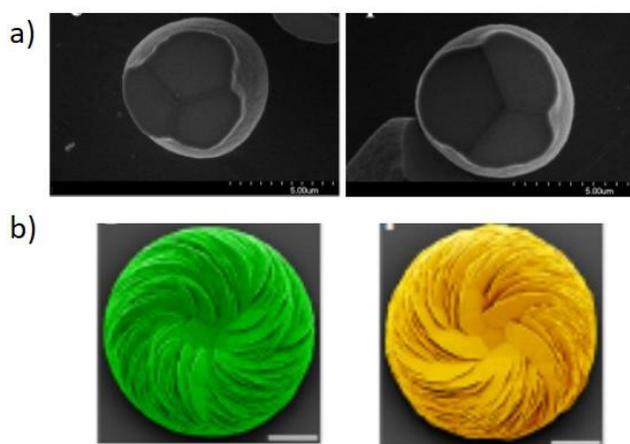


Figure 10 Asymmetric morphologies of CaCO_3 -based crystals induced by enantiopure amino acids. a) Scanning electron micrographs (SEM) of calcite crystals obtained by electrodeposition from calcium bicarbonate in the presence of magnesium and (*S*)-aspartic acid (left), and (*R*)-aspartic acid (right). Reproduced from reference²⁵⁷ with permission from American Chemical Society. b) SEM images of vaterite helicoids obtained by crystallization in presence of non-racemic solutions (40% *e.e.*) biased in favour of (*S*)-aspartic acid (left) and (*R*)-aspartic acid. Reproduced from reference²⁶⁰ with permission from Nature publishing group.

Calcite (CaCO_3), as the most abundant marine mineral in the Archaean era, has potentially played an important role in the formation of prebiotic molecules relevant to Life. The trigonal scalenohedral crystal form of calcite displays chiral faces which can yield chiral selectivity. In 2001, Hazen *et al.*²⁶¹ reported that (*S*)-aspartic acid adsorbs preferentially on the $(3\bar{1}\bar{2}1)$ face of calcite, whereas (*R*)-aspartic acid adsorbs preferentially on the $(21\bar{3}1)$ face. An *e.e.* value on the order of 0.5% on average was measured for the adsorbed aspartic acid molecules. No selectivity was observed on a centric surface that served as control. The experiments were conducted with aqueous solutions of (*rac*)-aspartic acid and selectivity was greater on crystals with terraced surface textures, presumably because enantiomers concentrated along step-like linear growth features. The calculated chiral indices of the (214) scalenohedral face of calcite was found to be the highest amongst 14 surfaces selected from various minerals (calcite, diopside, quartz, orthoclase) and face-centred cubic (FCC) metals.²⁵² In contrast, DFT studies revealed negligible difference in adsorption energies of enantiomers ($< 1 \text{ kcal.mol}^{-1}$) of alanine on the $(21\bar{3}1)$ face of calcite because alanine cannot establish three points of contact on the surface.²⁶² Conversely, it is well established that amino acids modify the crystal growth of calcite crystals in a selective manner leading to asymmetric morphologies, *e.g.* upon crystallization^{263,264} or electrodeposition (Figure 10a).²⁵⁹ Vaterite helicoids, produced by crystallization of CaCO_3 in presence of non-racemic mixtures of aspartic acid, were found to be single-handed (Figure 10b).²⁶⁰ Enantiomeric ratio are identical in the helicoids and in solution, *i.e.* incorporation of aspartic acid in vaterite displays no chiral amplification effect. Asymmetric growth was

also observed for various organic substances with gypsum, another mineral with a centrosymmetric crystal structure.²⁶⁵

As expected, asymmetric morphologies produced from amino acid enantiomers are mirror image (Figure 10).

Clay minerals, which for some of them display high specific surface area, adsorption and catalytic properties, are often invoked as potential promoters of the transformation of prebiotic molecules. Amongst the large variety of clays, serpentine and montmorillonite were likely the dominant ones on Earth prior to Life's origin.²⁴¹ Clay minerals can exhibit non-centrosymmetric structures, such as the A and B forms of kaolinite, which correspond to the enantiomeric arrangement of the interlayer space. These chiral organizations are, however, not individually separable. All experimental studies claiming asymmetric inductions by clay minerals reported in the literature have raised suspicion about their validity, with no exception.²⁴² This is because these studies employed either racemic clay or clays which have no established chiral arrangement, *i.e.* presumably achiral clay minerals. Asymmetric adsorption and polymerization of amino acids reported with kaolinite^{266–270} and bentonite^{271–273} in the 1970s–1980s actually originated from experimental errors or contaminations. Supposedly enantiospecific adsorptions of amino acids with allophane,²⁷⁴ hydrocalcite-like compound,²⁷⁵ montmorillonite,²⁷⁶ and vermiculite^{277,278} also likely belong to this category.

Experiments aimed at demonstrating deracemization of amino acids in absence of any chiral inducers or during phase transition under equilibrium conditions have to be interpreted cautiously (see the Chapter 4.2 of the book written by Meierhenrich for a more comprehensive discussion on this topic).²⁴ Deracemization is possible under far-from-equilibrium conditions but a set of repeated experiments must then reveal a distribution of the chiral biases (see Part 3). The claimed specific adsorptions for racemic mixtures of amino acids likely originated from the different purities between (*S*)- and (*R*)-amino acids; or contaminants of biological origin such as microbial spores.²⁷⁹ Such issues are not old-fashioned and despite great improvement in analytical and purification techniques, the difference in enantiomer purities is most likely at the origin of the different behaviour of amino acid enantiomers observed in the crystallization of wulfingite ($\epsilon\text{-Zn(OH)}_2$)²⁸⁰ and CaCO_3 ^{281,282} in two recent reports.

Very impressive levels of selectivities (on the range of 10% *e.e.*) were recently reported for the adsorption of aspartic acid on brushite, a mineral composed of achiral crystals of $\text{CaHPO}_4 \cdot 2\text{H}_2\text{O}$.²⁸³ In that case, selective adsorption was observed under supersaturation and undersaturation conditions (*i.e.* non-equilibrium states) but not at saturation (equilibrium state). Likewise, opposite selectivities were observed for the two non-equilibrium states. It was postulated that mirror symmetry breaking of the crystal facets occurred during the dynamic events of crystal growth and dissolution. Spontaneous mirror symmetry breaking is not impossible under far-from-equilibrium conditions but again a distribution of the selectivity outcome is expected upon repeating the experiments under strictly achiral conditions (Part 3).

Ribó and co-workers proposed that chiral surfaces could have been involved in the chiral enrichment of prebiotic molecules on carbonaceous chondrites present on meteorites.²⁸⁴ In their scenario, mirror symmetry breaking during the formation of planetesimal bodies and comets may have led to a bias in the distribution of chiral fractures, screw distortions or step-kink chiral centres on the surfaces of these inorganic matrices. This in turn would have led to a bias in the adsorption of organic compounds. Their study was motivated by the fact that the enantiomeric excesses measured for organic molecules vary according to their location on the meteorite surface.²⁸⁵ Their measurement of the optical activity of three meteorite samples by circular birefringence (CB) indeed revealed a slight bias towards negative CB values for the Murchinson meteorite. The optically active areas are attributed to serpentines and other poorly identified phyllosilicate phases, whose formation may have occurred concomitantly to organic matter. The implication of inorganic minerals in biasing the chirality of prebiotic molecules remains uncertain given that no strong asymmetric adsorption values have been reported to date and that certain minerals were even found to promote the racemization of amino acids²⁸⁶ and secondary alcohols.²⁸⁷ However, evidences exist that minerals could have served as hosts and catalysts for prebiotic reactions, including the polymerization of nucleotides.²⁸⁸ In addition, minute chiral biases provided by inorganic minerals could have driven SMSB processes into a deterministic outcome (Part 3).

b Organic crystals

Organic crystals may have also played a role in biasing the chirality of prebiotic chemical mixtures. Along this line, glycine appears as the most plausible candidate given its probable dominance over more complex molecules in the prebiotic soup.

α -Glycine crystallizes from water into a centrosymmetric form. In the 1980s, Lahav, Leiserowitch and co-workers demonstrated that amino acids were occluded to the basal faces (010 and $0\bar{1}0$) of glycine crystals with exquisite selectivity.^{289–291} For example, when a racemic mixture of leucine (1–2% wt/wt of glycine) was crystallized with glycine at an air/water interface, (*R*)-Leu was incorporated only into those floating glycine crystals whose (010) faces were exposed to the water solution, while (*S*)-Leu was incorporated only into the crystals with exposed ($0\bar{1}0$) faces. This results into the nearly perfect resolution (97–98% *e.e.*) of Leu enantiomers. In presence of a small amount of an enantiopure amino-acid (*e.g.* (*S*)-Leu), all crystals of Gly exposed the same face to the water solution leading to one enantiomer of a racemate being occluded in glycine crystals while the other remains in solution. These striking observations led the same authors to propose a scenario in which the crystallization of supersaturated solutions of glycine in presence of amino-acid racemates would have led to the spontaneous resolution of all amino acids (Figure 11).

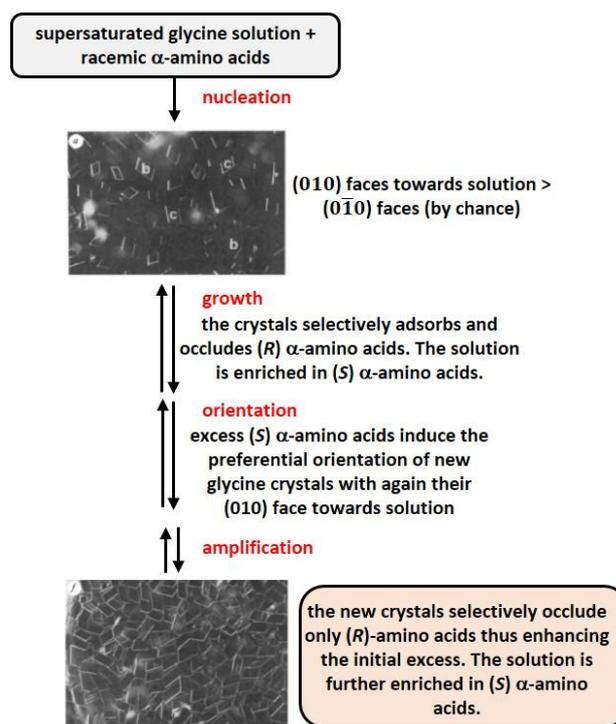


Figure 11 Resolution of amino acid enantiomers following a “by chance” mechanism including enantioselective occlusion into achiral crystals of glycine. Adapted from references²⁸⁰ and ²⁸⁹ with permission from American Chemical Society and Nature publishing group, respectively

This can be considered as a “by chance” mechanism in which one of the enantiotopic face (010) would have been exposed preferentially to the solution in absence of any chiral bias. From then, the solution, enriched into (*S*)-amino acids, enforces all glycine crystals to expose their (010) faces to water, eventually leading to all (*R*)-amino acids being occluded in glycine crystals.

A somewhat related strategy was disclosed in 2010 by Soai and co-workers.²⁹² Dehydration of centrosymmetric crystals of cytosine monohydrate yielded enantio-enriched anhydrous cytosine chiral crystals when only one of two enantiotopic faces of the crystal was put into direct contact with the hot plate. Enrichment was also observed if the dehydration was performed under reduced pressure but with a surprising inversion of the configuration of the chiral crystal relatively to thermal dehydration.²⁹³ Rearrangement of the hydrogen bond network during the transition between the centrosymmetric and chiral forms of the cytosine crystals is likely at the origin of this selective process. Reactions occurring at the enantiotopic surface of an achiral compound also yields enantio-enriched compounds whose optical purity can be enhanced by SMSB processes.^{294,295}

c Metal surfaces

The last two decades have seen the emergence of chiral metal surfaces displaying high adsorption enantioselectivities.^{239,244,296} The choice between metal surfaces can be oriented based on computational predictions of their enantiospecificity as reported recently for a variety of α -amino acids on Ag/Cu/Pd/Pt(531) surfaces.²⁹⁷ Mechanisms leading to specific adsorptions on chiral metal surfaces can be

well established. As a matter of example, the decomposition of tartaric acid, enantiospecifically adsorbed on Cu(651)⁵ surfaces, follows non-linear kinetics, suggesting auto-catalysis.²⁹⁸ Likewise, adsorption of scalemic mixtures of aspartic acid onto an achiral Cu(111) surface occurs with amplification of the chirality, *i.e.* the *e.e.* of the adsorbed enantiomers of the surface is largely higher than the *e.e.* of enantiomers in the gas phase.²⁹⁹ This effect, which presents some similarity to non-linear effects observed in asymmetric catalysis,⁶² is related to the formation of homochiral adsorbate clusters. Auto-catalysis and chiral amplification are two key requirements for spontaneous mirror symmetry breaking (Part 3) and consequently such mechanistic studies on chiral metal surfaces may reveal important pathways towards homochirality.³⁰⁰

3. Spontaneous Mirror-Symmetry Breaking (SMSB)

3.1 Definition, models and the Soai reaction

Spontaneous mirror-symmetry breaking (SMSB) phenomenon is the process that leads to the preferential formation of one chiral state over its enantiomeric form in absence of a detectable chiral bias or enantiomeric imbalance. As defined by Ribó and co-workers, SMSB concerns the transformation of “metastable racemic non-equilibrium stationary states (NESS) into one of two degenerate but stable enantiomeric NESSs”.³⁰¹ Despite this definition being in somewhat contradiction with the textbook statement that enantiomers need the presence of a chiral bias to be distinguished, it was recognized a long time ago that SMSB can emerge from reactions involving asymmetric self-replication or auto-catalysis. The connection between SMSB and BH is appealing,^{25,40,51,301–308} since SMSB is the unique physicochemical process that allows for the emergence and retention of enantiopurity from scratch. It is also intriguing to note that the competitive chiral reaction networks that might give rise to SMSB could exhibit replication, dissipation and compartmentalization,^{301,309} *i.e.* fundamental functions of living systems.

Systems able to lead to SMSB consist of enantioselective autocatalytic reaction networks, described through models dealing with either the transformation of achiral to chiral compounds, or the deracemization of racemic mixtures.³⁰¹ As early as 1953, Frank described a theoretical model dealing with the former case. According to Frank model, SMSB emerges from a system involving homochiral self-replication (one enantiomer of the chiral product accelerates its own formation) and heterochiral inhibition (the replication of the other product enantiomer is prevented).³⁰³ It is now well-recognized that the Soai reaction,⁵⁶ an auto-catalytic asymmetric process (Figure 12a), disclosed 42 years later,³¹⁰ is an experimental validation of the Frank model. The reaction between pyrimidine-5-carbaldehyde and diisopropyl zinc (two achiral reagents) is strongly accelerated by their zinc alkoxy product, which is found to be enantiopure (>99% *e.e.*) after a few cycles of reaction/addition of reagents (Figure 12b and

c).^{310–312} Kinetic models based on the stochastic formation of homochiral and heterochiral dimers^{313–315} of the zinc alkoxy product provide

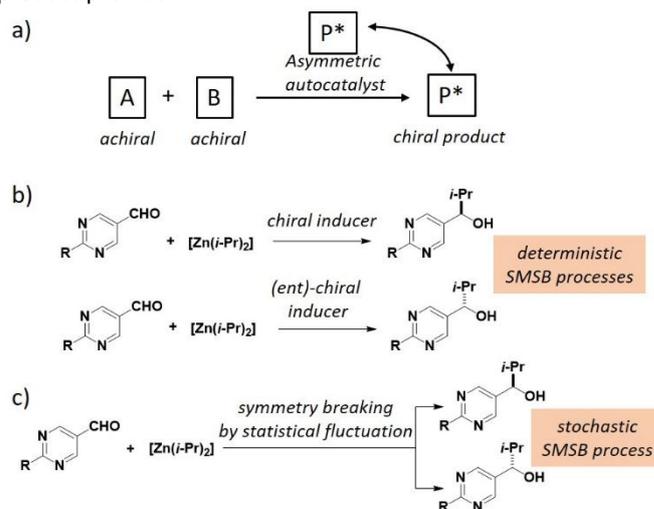


Figure 12 a) General scheme for an auto-catalysed asymmetric reaction. b) Soai reaction performed in the presence of detected chirality, leading to highly enantio-enriched alcohol with the same configuration in successive experiments (deterministic SMSB). c) Soai reaction performed in absence of detected chirality, leading to highly enantio-enriched alcohol with a bimodal distribution of the configurations in successive experiments (stochastic SMSB). c) Adapted from reference 311 with permission from D. Reidel Publishing Co.

good fits of the kinetic profile even though the involvement of higher species has gained more evidence recently.^{316–324} In this model, homochiral dimers serve as auto-catalyst for the formation of the same enantiomer of the product whilst heterochiral dimers are inactive and sequester the minor enantiomer, a Frank model-like inhibition mechanism. A hallmark of the Soai reaction is that direction of the auto-catalysis is dictated by extremely weak chiral perturbations: quartz, cryptochiral molecules, circularly polarized light, chiral isotopomers amongst others (Figure 12b).³¹² In addition, the apparent outcome of the Soai reaction performed in absence of detectable chiral species is stochastic as expected for a truly SMSB process (Figure 12c).^{325–333} On the one hand, the Soai reaction offers a credible mechanistic scenario from which homochiral biomolecules at the origin of Life would have been created on a deterministic manner through a SMSB process coupled to an infinitesimal chiral bias (*vide infra*). This bias would have survived from a larger one despite significant erosion through racemization processes. On the other hand, the Soai reaction is more the exception than the rule in the chemical space explored up to date.^{334–342} The exergonic and irreversible nature of the organozinc addition reaction are key for pushing the system far-from-equilibrium and for the generation and preservation of the homochiral state. On the contrary, it is assumed that prebiotic chemical reactions would have been only weakly exergonic, *i.e.* their products would have been more prone to racemization or to side reactions occurring in solution.^{37,46,301}

Many other models of spontaneous emergence of homochirality in far-from-equilibrium systems have been proposed in the literature.^{343–345} Most of them are derived from the Frank model but do not include any mutual inhibition

reaction. The limited enantioselective (LES) model^{306,346} assumes that both the asymmetric auto-catalysis (similar to the homochiral self-replication in the Frank model) and the non-enantioselective auto-catalysis (the accelerated formation of both enantiomers of the product) can co-exist. SMSB emerges if these two autocatalytic processes are: i) individually compartmentalized within regions experiencing different temperature,^{347,348} or ii) are driven by a constant concentration of external reagents.³⁴⁹ Required conditions for SMSB through LES model could have been present in deep ocean hydrothermal plumes. Likewise, a chemical scenario has been proposed for LES based on coupled Strecker-type reactions for amino acid synthesis and degradation which have been postulated to be accelerated by a heterogeneous catalytic support such as phyllosilicates.³⁴⁹ However, the LES model has found no experimental evidence up to date. Models for enantioselective hypercyclic replicators were recently disclosed in which the inhibition reaction in the Frank model has been replaced by mutual cross-catalytic processes occurring between families of coupled replicators.^{350,351} These models support a scenario in which the combination of SMSB, formation of the first (coupled) self-replicators and the emergence of their functions would have led to BH.³⁰¹ This intriguing concept may foster experimental investigations of SMSB processes in polymerization/depolymerization reactions. Imposed boundary conditions for SMSB involve “either systems open to matter exchange, or closed systems unable to equilibrate energy with their surroundings”.³⁰¹ In the absence of any chiral influence, the obtained metastable NESS are exposed to statistical fluctuations, and evolve towards scalemic or homochiral NESSs, as long as the systems are far-from-equilibrium. It is important to note that in absence of these boundary conditions, systems will be able to equilibrate with their surrounding and the deviation from the racemic state will be lost, *e.g.* racemization would occur under classically employed reaction workups operated in solution.^{41,352} This is probably the main reason why a single SMSB process has been identified to date for a reaction performed in solution (the Soai reaction). On the contrary, SMSB processes have been observed more frequently in crystals (*vide infra*) or in supramolecular assemblies,³⁵³ *i.e.* processes involving phase transition. Asymmetric reactions performed with catalytic single-handed supramolecular assemblies obtained through a SMSB process were found to yield enantio-enriched products whose configuration is left to chance.^{157,354} SMSB processes leading to homochiral crystals as the final state appear particularly relevant in the context of BH and will thus be discussed separately in the following section.

3.2 Homochirality by crystallization

Havinga postulated that just one enantiomorph can be obtained upon a gentle cooling of a racemate solution: i) when the crystal nucleation is rare and the growth is rapid and, ii) when fast inversion of configuration occurs in solution (*i.e.* racemization). Under these circumstances, only monomers with matching chirality to the primary nuclei crystallize leading

to SMSB.^{55,355} Havinga reported in 1954 a set of experiments aimed at demonstrating his hypothesis with *N,N,N*-

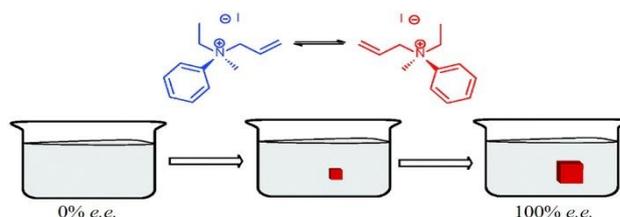


Figure 13 Enantiomeric preferential crystallization of *N,N,N*-allylethylmethylanilinium iodide as described by Havinga. Fast racemization in solution supplies the growing crystal with the appropriate enantiomer. Reprinted from reference⁵⁵ with permission from the Royal Society of Chemistry.

allylethylmethylanilinium iodide – an organic molecule which crystallizes as a conglomerate from chloroform (Figure 13).³⁵⁵ Fourteen supersaturated solutions were gently heated in sealed tubes, then stored at 0°C to give crystals which were in 12 cases inexplicably more dextrorotatory (measurement of optical activity by dissolution in water, where racemization is not observed). Seven other supersaturated solutions were carefully filtered before cooling to 0°C, but no crystallization occurred after one year. Crystallization occurred upon further cooling: three crystalline products with no optical activity were obtained, while the four other ones showed a small optical activity ($[\alpha]_D = +0.2^\circ; +0.7^\circ; -0.5^\circ; -3.0^\circ$). More successful examples of preferential crystallization of one enantiomer appeared in the literature notably with tri-*o*-thymotide,³⁵⁶ and 1,1'-binaphthyl.^{357,358} In the latter case, the distribution of specific rotations recorded for several independent experiments is centred to zero.

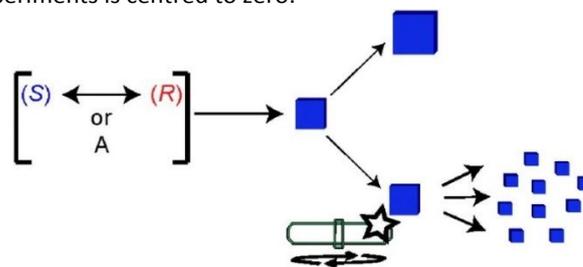


Figure 14 Primary nucleation of an enantiopure 'Eve crystal' of random chirality, slightly amplified by growing under static conditions (top, Havinga-like), or strongly amplified by secondary nucleation thanks to magnetic stirring (bottom, Kondepudi-like). Reprinted from reference⁵⁵ with permission from the Royal Society of Chemistry.

Sodium chlorate (NaClO_3) crystallizes by evaporation of water into a conglomerate ($P2_13$ space group).^{359–361} Preferential crystallization of one of the crystal enantiomorph over the other was already reported by Kipping and Pope in 1898.^{362,363} From static (*i.e.* non-stirred) solution, NaClO_3 crystallization seems to undergo an uncertain resolution, similar to Havinga findings with the aforementioned quaternary ammonium salt. However, a statistically significant bias in favour of *d*-crystals was invariably observed, likely due to the presence of bio-contaminants.³⁶⁴ Interestingly, Kondepudi *et al.* showed in 1990 that magnetic stirring, during the crystallization of sodium chlorate, randomly oriented the crystallization to only

one enantiomorph, with a virtually perfect bimodal distribution over several samples (± 1).³⁶⁵ Further studies^{366–369} revealed that the maximum degree of supersaturation is solely reached once, when the first primary nucleation occurs. At this stage, the magnetic stirring bar breaks up the first nucleated crystal into small fragments that have the same chirality than the ‘Eve crystal’, and act as secondary nucleation centres whence crystals grow (Figure 14). This constitutes a SMSB process coupling homochiral self-replication plus inhibition through the supersaturation drop during secondary nucleation, precluding new primary nucleation and the formation of crystals of the mirror-image form.³⁰⁷ This deracemization strategy was also successfully applied to 4,4'-dimethyl-chalcone,³⁷⁰ and 1,1'-binaphthyl (from its melt).³⁷¹

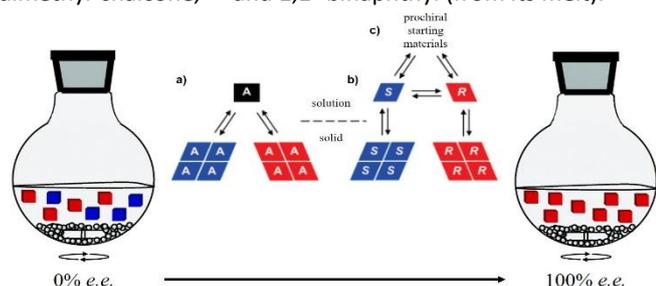


Figure 15 Schematic representation of Viedma ripening and solution-solid equilibria of an intrinsically achiral molecule (a) and a chiral molecule undergoing solution-phase racemization (b). The racemic mixture can result from chemical reaction involving prochiral starting materials (c). Adapted from references³⁵⁶ and³⁸² with permission from the Royal Society of Chemistry and the American Chemical Society, respectively.

In 2005, Viedma reported that solid-to-solid deracemization of NaClO_3 proceeded from its saturated solution by abrasive grinding with glass beads.³⁷³ Complete homochirality with bimodal distribution is reached after several hours or days.³⁷⁴ The process can also be triggered by replacing grinding with ultrasound,³⁷⁵ turbulent flow,³⁷⁶ or temperature variations.^{376,377} Although this deracemization process is easy to implement, the mechanism by which SMSB emerges is an ongoing highly topical question that falls outside the scope of this review.^{40,41,378–381}

Viedma ripening was exploited for deracemization of conglomerate-forming achiral or chiral compounds (Figure 15).^{55,382} The latter can be formed *in situ* by a reaction involving a prochiral substrate. For example, Vlieg *et al.* coupled an attrition-enhanced deracemization process with a reversible organic reaction (an aza-Michael reaction) between prochiral substrates under achiral conditions to produce an enantiopure amine.³⁸³ In a recent review, Buhse and co-workers identified a range of conglomerate-forming molecules that can be potentially deracemized by Viedma ripening.⁴¹ Viedma ripening also proves to be successful with molecules crystallizing as racemic compounds at the condition that conglomerate form is energetically accessible.³⁸⁴ Furthermore, a promising mechanochemical method to transform racemic compounds of amino acids into their corresponding conglomerates has been recently found.³⁸⁵ When valine, leucine and isoleucine were milled one hour in the solid state, in a Teflon jar with a zirconium ball and in the decisive

presence of zinc oxide, their corresponding conglomerates eventually formed.

Shortly after the discovery of Viedma, aspartic acid³⁸⁶ and glutamic acid^{387,388} were deracemized up to homochiral state starting from biased racemic mixtures. The chiral γ -polymorph of glycine³⁸⁹ was obtained with a preferred handedness by Ostwald ripening, albeit with a stochastic distribution of the optical activities.³⁹⁰ Salts or imine derivatives of alanine,^{391,392} phenylglycine^{372,384} and phenylalanine^{391,393} were desymmetrized by Viedma ripening with DBU (1,8-diazabicyclo[5.4.0]undec-7-ene) as the racemization catalyst. Successful deracemization was also achieved with amino acid precursors such as α -aminonitriles,^{394–396} α -iminonitriles,³⁹⁷ *N*-succinopyridine³⁹⁸ and thiohydantoin.³⁹⁹ The first three classes of compounds could be obtained directly from prochiral precursors by coupling synthetic reactions and Viedma ripening. In the preceding examples, the direction of SMSB process is selected by biasing the initial racemic mixtures in favour of one enantiomer or by seeding the crystallization with chiral chemical additives. In the next sections, we will consider the possibility to drive the SMSB process towards a deterministic outcome by means of PVED, physical fields, polarized particles, and chiral surfaces, *i.e.* the sources of asymmetry depicted in Part 2 of this review.

3.3 Deterministic SMSB processes

a Parity violation coupled to SMSB

In the 1980s, Kondepudi and Nelson, constructed stochastic models of a Frank-type autocatalytic network which allowed them to probe the sensitivity of the SMSB process to very weak chiral influences.^{304,400–402} Their estimated energy values for biasing the SMSB process into a single direction was in the range of PVED values calculated for biomolecules. Despite the competition with the bias originated from random fluctuations (as underlined later by Lente),¹²⁶ it appears possible that such a very weak ‘‘asymmetric factor can drive the system to a preferred asymmetric state with high probability’’.³⁰⁷ Recently, Blackmond and co-workers performed a series of experiments with the objective of determining the energy required for overcoming the stochastic behaviour of well-designed Soai⁴⁰³ and Viedma ripening experiments.⁴⁰⁴ This was done by performing these SMSB processes with very weak chiral inductors, isotopically chiral molecules and isotopologue enantiomers for the Soai reaction and the Viedma ripening, respectively. The calculated energies, 0.15 kJ/mol (for Viedma) and 2×10^{-8} kJ/mol (for Soai), are considerably higher than PVED estimates (*ca.* 10^{-12} – 10^{-15} kJ/mol). This means that the two experimentally SMSB processes reported to date are not sensitive enough to detect any influence of PVED and questions the existence of an ultra-sensitive auto-catalytic process, as the one described by Kondepudi and Nelson.

The possibility to bias crystallization processes with chiral particles emitted by radionuclides was probed by several groups as summarized in the reviews of Bonner.^{44,54} Kondepudi-like crystallization of NaClO_3 in presence of β particles from a ^{90}Sr source notably yielded a distribution of

(+) and (-)-NaClO₃ crystals, largely biased in favour of (+) crystals.⁴⁰⁵ It was presumed that spin polarized electrons produced chiral nucleating sites, albeit chiral contaminants cannot be excluded.

b Chiral surfaces coupled to SMSB

The extreme sensitivity of the Soai reaction to chiral perturbations is not restricted to soluble chiral species.³¹² Enantio-enriched or enantiopure pyrimidine alcohol was generated with determined configuration when the auto-catalytic reaction was initiated with chiral crystals, such as (α)-quartz,⁴⁰⁶ γ-glycine,⁴⁰⁷ N-(2-thienylcarbonyl)glycine,⁴⁰⁸ cinnabar,⁴⁰⁹ anhydrous cytosine,²⁹² or triglycine sulfate,⁴¹⁰ or with enantiotopic faces of achiral crystals such as CaSO₄·2H₂O (gypsum).⁴¹¹ Even though the selective adsorption of product to crystal faces has been observed experimentally⁴⁰⁹ and computed,⁴⁰⁸ the nature of the heterogeneous reaction steps that provide the initial enantiomer bias remains to be determined.³⁰⁰

The effect of chiral additives on crystallization processes, in which the additive inhibits one of the enantiomer growth thereby enriching the solid phase with the opposite enantiomer is well established as “the rule of reversal”.^{412,413} In the realm of the Viedma ripening, Noorduin *et al.* discovered in 2020 a way of propagating homochirality between α-iminonitriles, possible intermediates in the Strecker synthesis of α-amino acids.⁴¹⁴ These authors demonstrated that an enantiopure additive (1-20 mol%) induces an initial enantio-imbalance, which is then amplified by Viedma ripening up to a complete mirror-symmetry breaking. In contrast to the “rule of reversal”, the additive favours the formation of the product with identical configuration. The additive is actually incorporated in a thermodynamically controlled way into the bulk crystal lattice of the crystallized product of the same configuration, *i.e.* a solid solution is formed enantiospecifically.

c CPL coupled to SMSB

Coupling CPL-induced enantioenrichment and amplification of chirality has been recognized as a valuable method to induce a preferred chirality to a range of assemblies and polymers.^{182,183,354,415,416} On the contrary, the implementation of CPL as a trigger to direct auto-catalytic processes towards enantiopure small organic molecules has been scarcely investigated.

CPL was successfully used in the realm of the Soai reaction to direct its outcome, either by using a chiroptical switchable additive, or by asymmetric photolysis of a racemic substrate. In 2004, Soai *et al.* illuminated during 48h a photoresolvable chiral olefin with *l*- or *r*-CPL, and mixed it with the reactants of the Soai reaction to afford (S)- or (R)-5-pyrimidyl alkanol, respectively, in *e.e.* higher than 90%.⁴¹⁷ In 2005, the photolyzate of a pyrimidyl alkanol racemate acted as an asymmetric catalyst for its own formation reaching *e.e.* greater than 99.5%.⁴¹⁸ The enantiomeric excess of the photolyzate was below the detection level of chiral HPLC instrument but was amplified thanks to the SMSB process.

In 2009, Vlieg *et al.* coupled CPL with Viedma ripening to achieve complete and deterministic mirror-symmetry breaking.⁴¹⁹ Previous investigation revealed that the deracemization by attrition of the Schiff base of phenylglycine amide (**rac-1**, Figure 16a) always occurred in the same direction, the (R)-enantiomer, as a probable result of minute levels of chiral impurities.³⁷² CPL was envisaged as potent chiral physical field to overcome this chiral interference. Irradiation of solid-liquid mixtures of **rac-1**

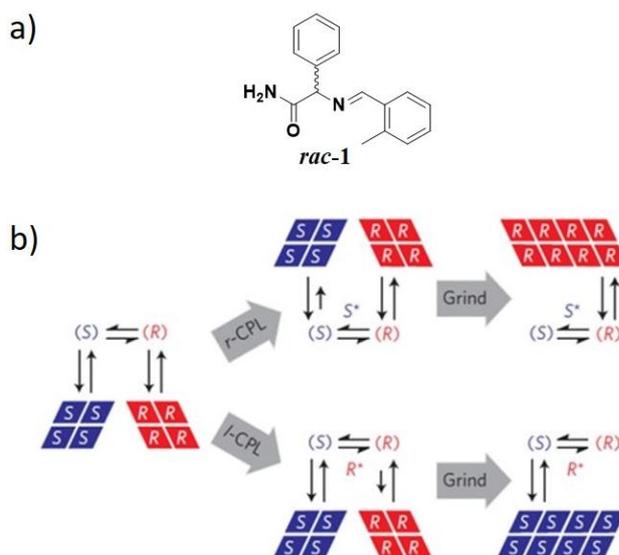


Figure 16 a) Molecular structure of **rac-1**. b) CPL-controlled complete attrition-enhanced deracemization of **rac-1**. (S) and (R) are the enantiomers of **rac-1** and S* and R* are chiral photoproducts formed upon CPL irradiation of **rac-1**. Reprinted from reference⁴¹⁹ with permission from Nature publishing group.

indeed led to complete deracemization, the direction of which was directly correlated to the circular polarization of light. Control experiments indicated that the direction of the SMSB process is controlled by a non-identified chiral photoproduct generated upon irradiation of (**rac-1**) by CPL. This photoproduct (S* or R* in Figure 16b) then serves as an enantioselective crystal-growth inhibitor which mediates the deracemization process towards the other enantiomer (Figure 16b). In the context of BH, this work highlights that asymmetric photosynthesis by CPL is a potent mechanism that can be exploited to direct deracemization processes when coupled to an amplification phenomenon.

4. Theories for the emergence of BH

Physical fields, CPL, polarized particles, polarized spins, chiral surfaces and SMSB processes have been presented as potential candidates for the emergence of chiral biases in prebiotic molecules. Their main properties are summarized in Table 1. The plausibility of the occurrence of these biases under the conditions of the primordial universe have also been evoked for certain physical fields (such as CPL or CISS). However, it is important to provide a more global overview of the current theories that tentatively explain the following

puzzling questions: where, when and how did the molecules of Life reach a homochiral state? At which point of this undoubtedly intricate process did Life emerge?

4.1 Terrestrial or Extra-Terrestrial Origin of BH?

The enigma of the emergence of BH might potentially be solved by finding the location of the initial chiral bias, might it be on Earth or elsewhere in the universe. The 'panspermia'

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Table 1 Potential sources of asymmetry and “by chance” mechanisms for the emergence of a chiral bias in prebiotic and biologically-relevant molecules.

Type	Truly/ Falsely chiral	Direction	Extent of induction	Scope	Importance in the context of BH	Selected references
PV	Truly	Unidirectional, deterministic, (+) or (-) for a given molecule	Minute ^a	Any chiral molecules	PVED : theo. calculations (natural) polarized particles : asymmetric destruction of racemates ^b	52, 53, 124
MChD	Truly	Bidirectional, (+) or (-) depending on the relative orientation of light and magnetic field	Minute ^c	Chiral molecules with high g_{NCD} and g_{MCD} values	Proceed with unpolarised light	137
Aligned magnetic field, gravity and rotation	Falsely	Bidirectional, (+) or (-) depending on the relative orientation of angular momentum and effective gravity	Minute ^d	Large supramolecular aggregates	Ubiquitous natural physical fields	161
Vortices	Truly	Bidirectional, (+) or (-) depending on the direction of the vortices	Minute ^d	Large objects or aggregates	Ubiquitous natural physical field (pot. present in hydrothermal vents)	149, 158
CPL	Truly	Bidirectional, (+) or (-) depending on the direction of CPL	Low to Moderate ^e	Chiral molecules with high g_{NCD} values	Asymmetric destruction of racemates	58
Spin-polarized electrons (CISS effect)	Truly	Bidirectional, (+) or (-) depending on the polarization	Low to high ^f	Any chiral molecules	Enantioselective adsorption/crystallization of racemate, asymmetric synthesis	233
Chiral surfaces	Truly	Bidirectional, (+) or (-) depending on surface chirality	Low to excellent	Any adsorbed chiral molecules	Enantioselective adsorption of racemates	237, 243
SMSB (crystallization)	<i>na</i>	Bidirectional, stochastic distribution of (+) or (-) for repeated processes	Low to excellent	Conglomerate-forming molecules	Resolution of racemates	54, 367
SMSB (asymmetric auto- catalysis)	<i>na</i>	Bidirectional, stochastic distribution of (+) or (-) for repeated processes	Low to excellent	Soai reaction	<i>to be demonstrated</i>	312
Chance mechanisms	<i>na</i>	Bidirectional, stochastic distribution of (+) or (-) for repeated processes	Minute ^g	Any chiral molecules	<i>to be demonstrated</i>	17, 412–414

(a) $PVED \approx 10^{-12} - 10^{-15}$ kJ/mol.⁵³ (b) However, experimental results are not conclusive (see part 2.2b). (c) $e.e._{\text{MChD}} = g_{\text{MChD}}/2$ with $g_{\text{MChD}} \approx (g_{\text{NCD}} \times g_{\text{MCD}})/2$. NCD: Natural Circular Dichroism. MCD: Magnetic Circular Dichroism. For the resolution of Cr complexes,¹³⁷ $e.e. = k \times B$ with $k = 10^{-5} \text{ T}^{-1}$ at $\lambda = 695.5 \text{ nm}$. (d) The minute chiral induction is amplified upon aggregation leading to homochiral helical assemblies.⁴²³ (e) For photolysis, $e.e.$ depends both on g and the extent of reaction (see equation 2 and the text in part 2.2a). Up to a few $e.e.$ percent have been observed experimentally.^{197,198,199} (f) Recently, spin-polarized SE through the CISS effect have been implemented as chiral reagents with relatively high $e.e.$ values (up to a ten percent) reached for a set of reactions.²³⁶ (g) The standard deviation for 1 mole of chiral molecules is of 1.9×10^{11} .¹²⁶ *na*: not applicable.

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hypothesis,⁴²⁴ for which living organisms were transplanted to Earth from another solar system, sparked interest into an extra-terrestrial origin of BH, but the fact that such a high level of chemical and biological evolution was present on celestial objects have not been supported by any scientific evidence.⁴⁴ Accordingly, terrestrial and extra-terrestrial scenarios for the original chiral bias in prebiotic molecules will be considered in the following.

a Terrestrial origin of BH

A range of chiral influences have been evoked for the induction of a deterministic bias to primordial molecules generated on Earth. Enantiospecific adsorptions or asymmetric syntheses on the surface of abundant minerals have long been debated in the context of BH^{44,238–242} since no significant bias of one enantiomorphic crystal or surface over the other has been measured when counting is averaged over several locations on Earth.^{257,258} Prior calculations supporting PVED at the origin of excess of *l*-quartz over *d*-quartz^{114,425} or favouring the A-form of kaolinite⁴²⁶ are thus contradicted by these observations. Abyssal hydrothermal vents during the Hadean/Eo-Archaeon eon are argued as the most plausible regions for the formation of primordial organic molecules on the early Earth.⁴²⁷ Temperature gradients may have offered the different conditions for the coupled autocatalytic reactions and clays may have acted as catalytic sites.³⁴⁷ However, chiral inductors in these geochemically reactive habitats are hypothetical, even though vortices⁶⁰ or CISS occurring at the surfaces of greigite have been mentioned recently.⁴²⁸ CPL and MChD are not potent asymmetric forces on Earth as a result of low levels of circular polarization detected for the former and small anisotropic factors of the latter.^{429–431} PVED is an appealing “intrinsic” chiral polarization of matter but its implication in the emergence of BH is questionable (Part 1).¹²⁶ Alternatively, theories suggesting that BH emerged from scratch, *i.e.* without any involvement of the chiral discriminating sources mentioned in Part 1-2 and SMSB processes (Part 3), have been evoked in the literature since a long time,⁴²⁰ and variant versions appeared sporadically. Herein, these mechanisms are named as “random” or “by chance” and are based on probabilistic grounds only (Table 1). The prevalent form comes from the fact that a racemate is very unlikely made of exactly equal amounts of enantiomers, due to natural fluctuations described statistically like coin tossing.^{126,432} One mole of chiral molecules actually exhibits a standard deviation of 1.9×10^{11} . Putting into relation this statistical variation and putative strong chiral amplification mechanisms and evolutionary pressures, Siegel suggested that homochirality is an imperative of molecular evolution.¹⁷ However, the probability to get both homochirality and Life

emerging from statistical fluctuations at the molecular scale appears very unlikely.^{35,59,433} SMSB phenomena may amplify statistical fluctuations up to homochiral state, yet the direction of process for multiple occurrences will be left to chance in absence of a chiral inducer (Part 3). Other theories suggested that homochirality emerges during the formation of biopolymers “by chance”, as a consequence of the limited number of sequences that can be possibly contained in a reasonable amount of macromolecules (see 4.3).^{17,421,422} Finally, kinetic processes have also been mentioned in which a given chemical event would have occurred to a larger extent for one enantiomer over the other under achiral conditions (see one possible physicochemical scenario in Figure 11). Hazen notably argued that nucleation processes governing auto-catalytic events occurring at the surface of crystals are rare and thus a kinetic bias can emerge from an initially unbiased set of prebiotic racemic molecules.²³⁹ Random and by chance scenarios towards BH might be attractive on a conceptual view but lack experimental evidences.

b Extra-terrestrial origin of BH

Scenarios suggesting a terrestrial origin behind the original enantiomeric imbalance leave a question unanswered: how an Earth-based mechanism can explain enantioenrichment in extra-terrestrial samples?^{433,59} However, to stray from “geocentrism” is still worthwhile; another plausible scenario is the exogenous delivery on Earth of enantio-enriched molecules relevant for the appearance of Life. The body of evidence grew from the characterization of organic molecules, especially amino acids and sugars, and their respective optical purity, in meteorites,⁵⁹ comets, and laboratory-simulated interstellar ices.⁴³⁴

The 100-kg Murchison’s meteorite that fell to Australia in 1969 is generally considered as the standard reference for extra-terrestrial organic matter (Figure 17a).⁴³⁵ In fifty years, analyses of its composition revealed more than ninety α , β , γ and δ -isomers of C₂ to C₉ amino acids, diamino acids, and dicarboxylic acids as well as numerous polyols including sugars (ribose,⁴³⁶ a building block of RNA), sugar acids and alcohols, but also α -hydroxycarboxylic acids,⁴³⁷ and deoxy acids.⁴³⁴ Unequal amounts of enantiomers were also found with a quasi-exclusively predominance for (*S*)-amino acids^{57,285,438–440} ranging from 0 to 26.3±0.8% *e.es.* (highest *e.e.* being measured for non-proteinogenic α -methyl amino acids);⁴⁴¹ and, when they are not racemates, only *D*-sugar acids with *e.e.* up to 82% for xylonic acid have been detected.⁴⁴² These measurements are relatively scarce for sugars and in general need to be repeated, notably to definitely exclude their potential contamination by terrestrial environment. Future space

missions to asteroids, comets and Mars, coupled with more advanced analytical techniques,⁴⁴³ will

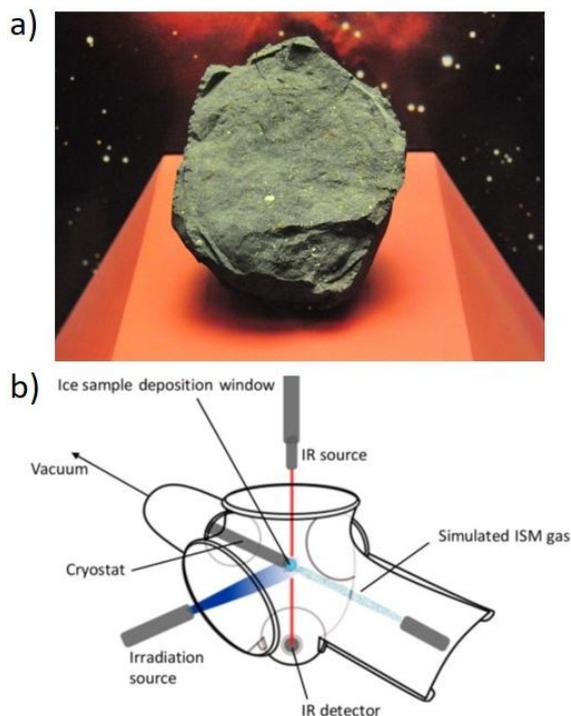


Figure 17 a) A fragment of the meteorite landed in Murchison, Australia, in 1969, and exhibited at the National Museum of Natural History (Washington). b) Scheme of the preparation of interstellar ice analogues. A mixture of primitive gas molecules is deposited and irradiated under vacuum on a cooled window. Composition and thickness are monitored by infrared spectroscopy. Reprinted from reference⁴⁵⁴ with permission from MDPI.

indubitably lead to a better determination of the composition of extra-terrestrial organic matter. The fact that major enantiomers of extra-terrestrial amino acids and sugar derivatives have the same configuration as the building blocks of Life constitutes a promising set of results.

To complete these analyses of the difficult-to-access outer space, laboratory experiments have been conducted by reproducing the plausible physicochemical conditions present on astrophysical ices (Figure 17b).⁴⁴⁴ Natural ones are formed in interstellar clouds,^{445,446} on the surface of dust grains from which condensates a gaseous mixture of carbon, nitrogen and oxygen-based molecules (e.g. H₂O, CH₃OH, CH₄, NH₃, CO₂),⁴⁴⁷ under the influence of very low temperature (5–15 K)⁴⁴⁸ and pressure. Subsequent photochemical processes in this mantle of frost are assumed to lead to complex molecules.⁴⁴⁹ Since collapsing clouds gave birth to our solar system through the aggregation of dust grains,⁴⁵⁰ studies of their composition receive a keen interest to broaden our knowledge about the prebiotic environment. Experiments on simulated interstellar ices support the formation of many proteinaceous amino acids,^{451,452} and building blocks of RNA and DNA such as sugars,⁴⁵³ like ribose⁴⁵⁴ and deoxyribose,⁴⁵⁵ as well as nucleobases (adenine, cytosine, uracil and thymine for example).⁴⁵⁶ These molecules were obtained with no

significant bias from the ideal racemic composition which supports their abiotic origin.⁴⁴⁴ However, when similar experiments are conducted under CPL irradiation, amino acids are generated with significant biases towards one enantiomer as described in 2.2a.^{204,205}

The occurrence of CPL-driven photochirogenesis on interstellar dust grains was supported by the detection of near-infrared light with significant circular polarization degrees (up to 22%),⁴⁵⁷ in parsec-sized star-forming regions, such as massive molecular clouds.^{458–464} Cosmic circularly polarized photons arise from synchrotron radiations emitted by neutron stars, remnants of supernovae explosions,³⁵ through *bremstrahlung*, dichroic scattering and/or light extinction (along lined up grains).⁴⁶⁵ In addition, although it was not directly observed due to dust shielding, models predicted the generation of vacuum ultraviolet (VUV) and UV-CPL under these conditions,⁴⁵⁹ i.e. spectral regions of light absorbed by amino acids and sugars. Photolysis by broad band and optically impure CPL is expected to yield lower enantioenrichments than those obtained experimentally by monochromatic and quasiperfect circularly polarized synchrotron radiation (see 2.2a).¹⁹⁸ However, a broad band CPL is still capable of inducing chiral bias by photolysis of an initially abiotic racemic mixture of aliphatic α -amino acids as previously debated.^{466,467} Likewise, CPL in the UV range will produce a wide range of amino acids with a bias towards the (S) enantiomer,¹⁹⁵ including α -dialkyl amino acids.⁴⁶⁸

l- and *r*-CPL produced by a neutron star are equally emitted in vast conical domains in the space above and below its equator.³⁵ However, appealing hypotheses were formulated against the apparent contradiction that amino acids have always been found as predominantly (S) on several celestial bodies,⁵⁹ and the fact that CPL is expected to be portioned into *left*- and *right*-handed contributions in equal abundance within the outer space. In the 1980s, Bonner and Rubenstein proposed a detailed scenario in which the solar system, revolving around the centre of our galaxy, had repeatedly traversed a molecular cloud and accumulated enantioenriched incoming grains.^{430,469} The same authors assumed that this enantioenrichment would come from asymmetric photolysis induced by synchrotron CPL emitted by a neutron star at the stage of planets formation. Later, Meierhenrich remarked in addition that, in molecular clouds, regions of homogeneous CPL polarization can exceed the expected size of a protostellar disk – or of our solar system,^{458,470} allowing a unidirectional enantioenrichment within our solar system, including comets.²⁴ A solid scenario towards BH thus involves CPL as a source of chiral induction for biorelevant candidates, through photochemical processes on the surface of dust grains, and delivery of the enantioenriched compounds on primitive Earth by direct grain accretion or by impact⁴⁷¹ of larger objects (Figure 18).^{472–474}

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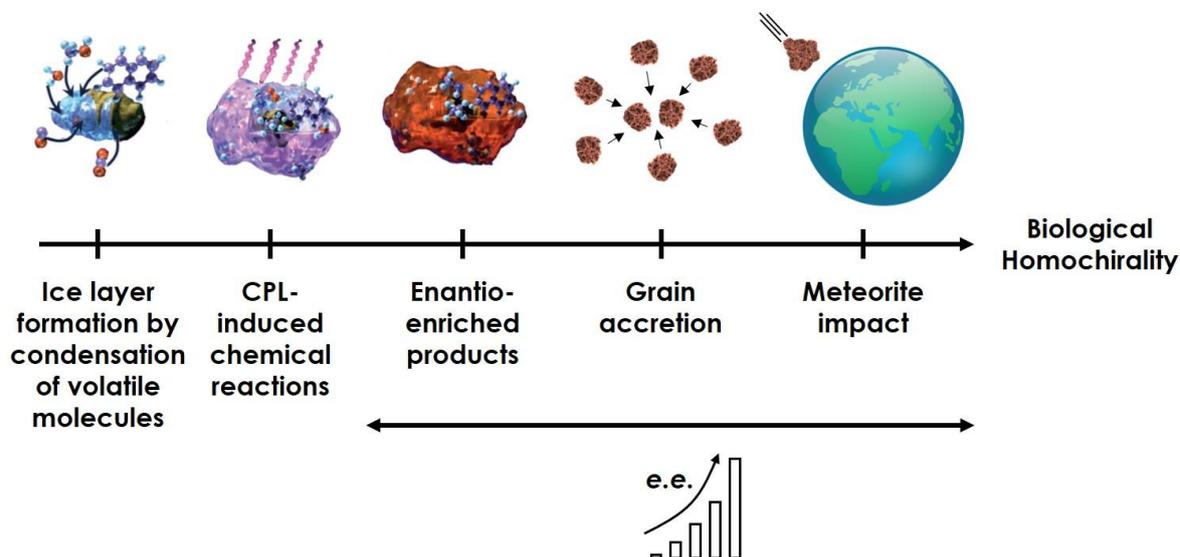


Figure 18 CPL-based scenario for the emergence of BH following the seeding of the early Earth with extra-terrestrial enantio-enriched organic molecules. Adapted from reference⁴⁷⁴ with permission from Wiley-VCH.

The high enantiomeric excesses detected for (*S*)-isovaline in certain stones of the Murchison's meteorite (up to $15.2 \pm 0.2\%$) suggested that CPL alone cannot be at the origin of this enantioenrichment.²⁸⁵ The broad distribution of *e.e.s.* (0–15.2%) and the abundance ratios of isovaline relatively to other amino acids also point to (*S*)-isovaline (and probably other amino acids) being formed through multiple synthetic processes that occurred during the chemical evolution of the meteorite.⁴⁴⁰ Finally, based on the anisotropic spectra,¹⁸⁸ it is highly plausible that other physicochemical processes, *e.g.* racemization coupled to phase transitions or coupled non-equilibrium/equilibrium processes,^{378,475} have led to a change in the ratio of enantiomers initially generated by UV-CPL.⁵⁹ In addition, a serious limitation of the CPL-based scenario shown in Figure 18 is the fact that significant enantiomeric excesses can only be reached at high conversion, *i.e.* by decomposition of most of the organic matter (see equation 2 in 2.2a). Even though there is a solid foundation for CPL being involved as an initial inducer of chiral bias in extra-terrestrial organic molecules, chiral influences other than CPL cannot be excluded. Induction and enhancement of optical purities by physicochemical processes occurring at the surface of meteorites and potentially involving water and the lithic environment have been evoked but have not been assessed experimentally.²⁸⁵

Asymmetric photoreactions⁴³¹ induced by MChD can also be envisaged notably in neutron stars environment, of tremendous magnetic fields (10^8 – 10^{12} T) and synchrotron radiations.^{35,476} Spin-polarized electrons (SPEs), another

potential source of asymmetry, can potentially be produced upon ionizing irradiation of ferrous magnetic domains present in interstellar dust particles, aligned by the enormous magnetic fields produced by a neutron star. One enantiomer from a racemate in a cosmic cloud could adsorb enantiospecifically on the magnetized dust particle. In addition, meteorites contain magnetic metallic centres that can act as asymmetric reaction sites upon generation of SPEs. Finally, polarized particles such as antineutrinos (SNAAP model^{226–228}) have been proposed as a deterministic source of asymmetry, at work in the outer space. Radoracemization must potentially be considered as a jeopardizing factor in that specific context.^{44,477,478} Further experiments are needed to probe whether these chiral influences may have played a role in the enantiomeric imbalances detected in celestial bodies.

4.2 Purely abiotic scenarios

Emergences of Life and biomolecular homochirality must be tightly linked,^{46,479,480} but in such a way that needs to be cleared up. As recalled recently by Glavin, homochirality by itself cannot be considered as a biosignature.⁵⁹ Non proteinogenic amino acids are predominantly (*S*) and abiotic physicochemical processes can lead to enantio-enriched molecules. However, it has been widely substantiated that polymers of Life (proteins, DNA, RNA) as well as lipids need to be enantiopure to be functional. Considering the NASA definition of Life, “a self-sustaining chemical system capable of Darwinian evolution”,⁴⁸¹ and the “widespread presence of ribonucleic acid (RNA) cofactors and catalysts in today's terran

biosphere”,⁴⁸² a strong hypothesis for the origin of Darwinian evolution and Life is “the

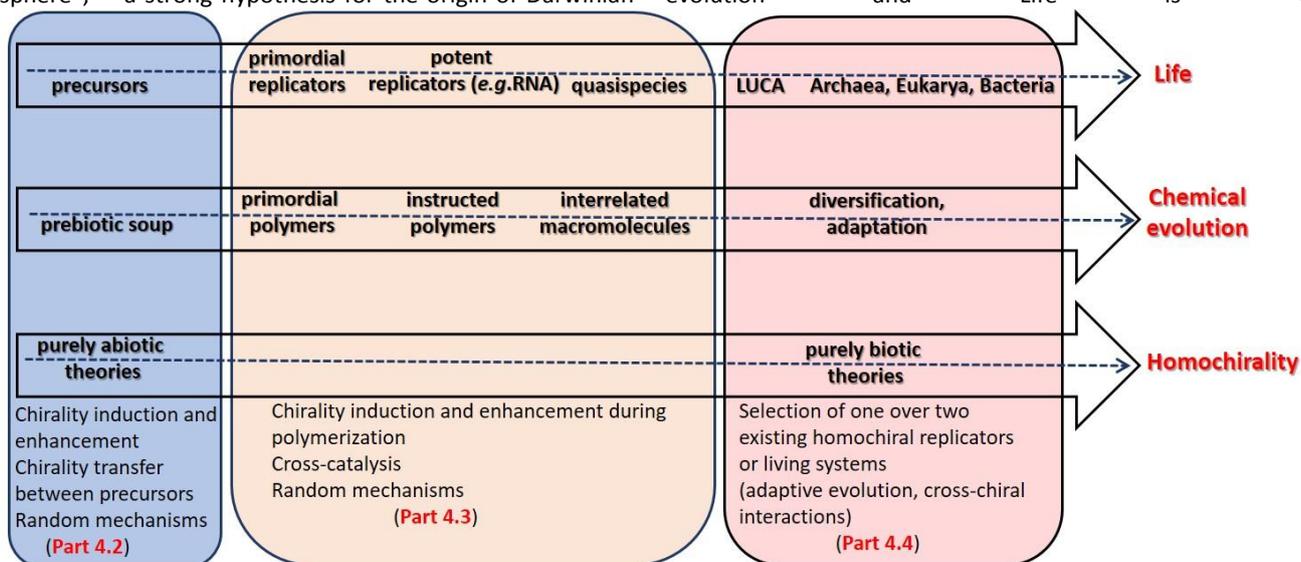


Figure 19 Possible connections between the emergences of Life and homochirality at the different stages of the chemical and biological evolutions. Possible mechanisms leading to homochirality are indicated below each of the three main scenarios. Some of these mechanisms imply an initial chiral bias which can be of terrestrial or extra-terrestrial origins as discussed in 4.1. LUCA = Last Universal Cellular Ancestor.

abiotic formation of long-chained RNA polymers” with self-replication ability.³⁰⁹ Current theories differ by placing the emergence of homochirality at different times of the chemical and biological evolutions leading to Life. Regarding on whether homochirality happens before or after the appearance of Life discriminates between purely abiotic and biotic theories, respectively (Figure 19). In between these two extreme cases, homochirality could have emerged during the formation of primordial polymers and/or their evolution towards more elaborated macromolecules.

a Enantiomeric cross-inhibition

The puzzling question regarding primeval functional polymers is whether they form from enantiopure, enantio-enriched, racemic or achiral building blocks. A theory that has found great support in the chemical community was that homochirality was already present at the stage of the primordial soup, *i.e.* that the building blocks of Life were enantiopure. Proponents of the purely abiotic origin of homochirality mostly refer to the inefficiency of polymerization reactions when conducted from mixtures of enantiomers. More precisely, the term enantiomeric cross-inhibition was coined to describe experiments for which the rate of the polymerization reaction and/or the length of the polymers were significantly reduced when non-enantiopure mixtures were used instead of enantiopure ones.^{24,44} Seminal studies were conducted by oligo- or polymerizing α -amino acid *N*-carboxy-anhydrides (NCAs) in presence of various initiators. Idelson and Blout observed in 1958 that (*R*)-glutamate-NCA added to reaction mixture of (*S*)-glutamate-NCA led to a significant shortening of the resulting polypeptides, inferring that (*R*)-glutamate provoked the chain termination of (*S*)-glutamate oligomers.⁴⁸³ Lundberg and Doty also observed that the rate of polymerization of (*R*)/(*S*) mixtures

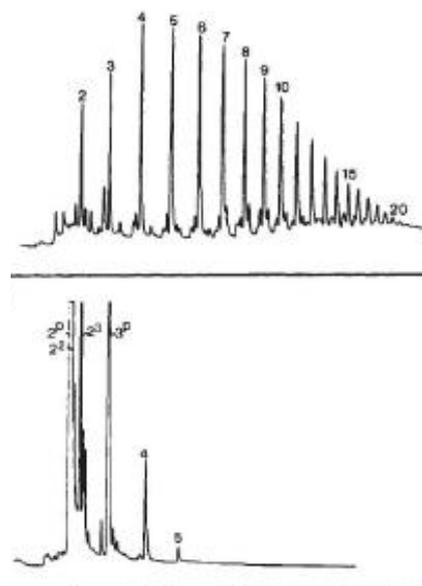


Figure 20 HPLC traces of the condensation reactions of activated guanosine mononucleotides performed in presence of a complementary poly-D-cytosine template. Reaction performed with D (top) and L (bottom) activated guanosine mononucleotides. The numbers labelling the peaks correspond to the lengths of the corresponding oligo(G)s. Reprinted from reference⁴⁸⁴ with permission from Nature publishing group.

of a glutamate-NCA and the mean chain length reached at the end of the polymerization were decreased relatively to that of pure (*R*)- or (*S*)-glutamate-NCA.^{485,486} Similar studies for oligonucleotides were performed with an enantiopure template to replicate activated complementary nucleotides. Joyce *et al.* showed in 1984 that the guanosine oligomerization, directed by a poly-D-cytosine template, was inhibited when conducted with a racemic mixture of activated mononucleotides.⁴⁸⁴ The L residues are predominantly located at the chain-end of the oligomers, acting as chain terminators, thus decreasing the yield in oligo-D-guanosine (Figure 20). A

similar conclusion was reached by Goldanskii and Kuz'min upon studying the dependence of the length of enantiopure oligonucleotides on the chiral composition of the reactive monomers.⁴²⁹ Interpolation of their experimental results with a mathematical model led to the conclusion that the length of potent replicators will dramatically be reduced in presence of enantiomeric mixtures reaching a value of 10 monomer units at best for a racemic medium.

Finally, the oligomerization of activated racemic guanosine was also inhibited on DNA and PNA templates.⁴⁸⁷ The latter being achiral, it suggests that enantiomeric cross-inhibition is intrinsic to the oligomerization process involving complementary nucleobases.

b Propagation and enhancement of the primeval chiral bias

Studies demonstrating enantiomeric cross-inhibition during polymerization reactions have led the proponents of purely abiotic origin of BH to propose several scenarios for the formation building blocks of Life in an enantiopure form. In this regards, racemization appears as a redoubtable opponent considering that harsh conditions – intense volcanism, asteroid bombardment and scorching heat^{488,489} – prevailed between the Earth formation, 4.5 billion years ago, and the appearance of Life, 3.5 billion years ago at the latest.^{490,491} At that time, deracemization inevitably suffered from its nemesis, racemization, which may take place in days or less in hot alkaline aqueous medium.^{35,301,492–494}

Several scenarios have considered that initial enantiomeric imbalances have probably been decreased by racemization but not eliminated. Abiotic theories thus rely on processes that would be able to amplify tiny enantiomeric excesses (likely $\ll 1\%$ *e.e.*) up to homochiral state. Intermolecular interactions cause enantiomer and racemate to have different physicochemical properties and this can be exploited to enrich a scalemic material into one enantiomer under strictly achiral conditions. This phenomenon of self-disproportionation of the enantiomers (SDE) is not rare for organic molecules and may occur through a wide range of physicochemical processes.⁶¹

SDE with molecules of Life such as amino acids and sugars is often discussed in the framework of the emergence of BH. SDE often occurs during crystallization as a consequence of the different solubility between racemic and enantiopure crystals, and its implementation to amino acids was exemplified by Morowitz as early as 1969.⁴⁹⁵ It was confirmed later that a range of amino acids displays high eutectic *e.e.* values which allows very high *e.e.* values to be present in solution, even from moderately biased enantiomeric mixtures.⁴⁹⁶ Serine is the most striking example since a virtually enantiopure solution ($> 99\%$ *e.e.*) is obtained at 25°C under solid-liquid equilibrium conditions starting from a 1% *e.e.* mixture only.⁴⁹⁷ Enantioenrichment was also reported for various amino acids after consecutive evaporations of their aqueous solutions⁴⁹⁸ or preferential kinetic dissolution of their enantiopure crystals.⁴⁹⁹ Interestingly, the eutectic *e.e.* values can be increased for certain amino acids by addition of simple achiral molecules such as carboxylic acids.⁵⁰⁰ DL-Cytidine, DL-adenosine and DL-uridine also form racemic crystals and their scalemic mixture can thus be enriched towards the D enantiomer in the same

way at the condition that the amount of water is small enough that the solution is saturated in both D and DL.⁵⁰¹ SDE of amino acids does not occur solely during crystallization;⁵⁰² *e.g.* sublimation of near-racemic samples of serine yields a sublimate which is highly enriched in the major enantiomer.⁵⁰³ Amplification of *e.e.* by sublimation has also been reported for other scalemic mixtures of amino acids,^{504–506} or for a racemate mixed with a non-volatile optically pure amino acid.⁵⁰⁷ Alternatively, amino acids were enantio-enriched by simple dissolution/precipitation of their phosphorylated derivatives in water.⁵⁰⁸

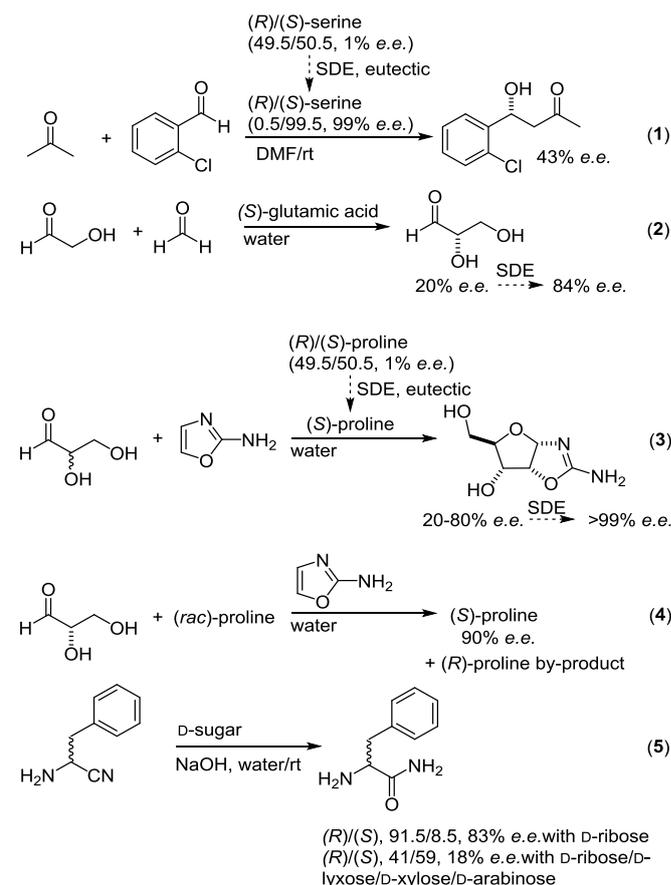


Figure 21 Selected catalytic reactions involving amino acids and sugars, and leading to the enantioenrichment of prebiotically relevant molecules.

It is likely that prebiotic chemistry have linked amino acids, sugars and lipids in a way that remains to be determined. Merging the organocatalytic properties of amino acids with the aforementioned SDE phenomenon offers a pathway towards enantiopure sugars.⁵⁰⁹ The aldol reaction between 2-chlorobenzaldehyde and acetone was found to exhibit a strongly positive non-linear effect, *i.e.* the *e.e.* in the aldol product is drastically higher than that expected from the optical purity of the engaged amino acid catalyst.⁴⁹⁷ Again, the effect was particularly strong with serine since nearly racemic serine (1% *e.e.*) and enantiopure serine provided the aldol product with the same enantioselectivity (*ca.* 43% *e.e.*, Figure 21, (1)). Enamine catalysis in water was employed to prepare glyceraldehyde, the probable synthon towards ribose and

other sugars, by reacting glycolaldehyde and formaldehyde in presence of various enantiopure amino acids. It was found that all (*S*)-amino acids, except (*S*)-proline, provided glyceraldehyde with a predominant *R* configuration (up to 20% *e.e.* with (*S*)-glutamic acid, Figure 21, (2)).^{65,510} This result coupled to SDE furnished a small fraction of glyceraldehyde with 84% *e.e.* Enantio-enriched tetrose and pentose sugars are also produced by means of aldol reactions catalysed by amino acids and peptides in aqueous buffer solutions, albeit in modest yields.⁵⁰³⁻⁵⁰⁵

The influence of α -amino acids on the synthesis of RNA precursors was also probed. Along this line, Blackmond and co-workers reported that ribo- and arabino-amino oxazolines were

enantio-enriched towards the expected *D* configuration when 2-aminooxazole and (*RS*)-glyceraldehyde were reacted in presence of (*S*)-proline (Figure 21, (3)).⁵¹⁴ When coupled with the SDE of the reacting proline (1% *e.e.*) and of the enantio-enriched product (20-80% *e.e.*), the reaction yielded enantiopure crystals of ribo-amino-oxazoline. (*S*)-proline does not act as a mere catalyst in this reaction but rather traps the (*S*)-enantiomer of glyceraldehyde, thus accomplishing a formal resolution of the racemic starting material. The latter reaction can also be exploited in the opposite way to resolve a racemic mixture of proline in presence of enantiopure glyceraldehyde (Figure 21, (4)). This dual substrate/reactant behaviour motivated the same group to test the possibility of synthesizing enantio-enriched amino acids with *D*-sugars. The hydrolysis of 2-benzyl α -amino nitrile yielded the corresponding α -amino amide (precursor of phenylalanine) with various *e.e.* values and configurations depending on the nature of the sugars.⁵¹⁵ Notably, *D*-ribose provided the product with 70% *e.e.* biased in favour of unnatural (*R*)-configuration (Figure 21, (5)). This result, which is apparently contradictory with such process being involved in the primordial synthesis of amino acids, was solved by finding that the mixture of four *D*-pentoses actually favoured the natural (*S*) amino acid precursor. This result suggests an unanticipated role of prebiotically relevant pentoses such as *D*-lyxose in mediating the emergence of amino acid mixtures with a biased (*S*) configuration.

How the building blocks of proteins, nucleic acids and lipids would have interacted between each other before the emergence of Life is a subject of intense debate. The aforementioned examples by which prebiotic amino acids, sugars, and nucleotides would have mutually triggered their formation is actually not the privileged scenario of 'origin of Life' practitioners. Most theories infer relationships at a more advanced stage of the chemical evolution. In the "RNA World",⁵¹⁶ a primordial RNA replicator catalysed the formation of the first peptides and proteins. Alternative hypotheses are that proteins ("metabolism first" theory) or lipids⁵¹⁷ originated first⁵¹⁸ or that RNA, DNA and proteins emerged simultaneously by continuous and reciprocal interactions, *i.e.* mutualism.^{519,520} It is commonly considered that homochirality would have arisen through stereoselective interactions between the different types of biomolecules, *i.e.* chirally matched

combinations would have conducted to potent living systems whilst the chirally mismatched combinations would have declined. Such theory has notably been proposed recently to explain the splitting of lipids into opposite configurations in archaea and bacteria (known as the 'lipide divide')⁵²¹ and their persistence.⁵²² However, these theories do not address the fundamental question of the initial chiral bias and its enhancement.

SDE appears as a potent way to increase the optical purity of some building blocks of Life but its limited scope, efficiency (initial bias $\geq 1\%$ *e.e.* is required) and productivity (high optical purity is reached at the cost of the mass of material) appear detrimental for explaining the emergence of chemical homochirality. An additional drawback of SDE is that the enantioenrichment is only local, *i.e.* the overall material remains unenriched. SMSB processes as those mentioned in Part 3 are consequently considered as more probable alternatives towards homochiral prebiotic molecules. They disclose two major advantages: i) a tiny fluctuation around the racemic state might be amplified up to the homochiral state in a deterministic manner, ii) the amount of prebiotic molecules generated throughout these processes is potentially very high (*e.g.* in Viedma-type ripening experiments).³⁸³ Even though experimental reports of SMSB processes have appeared in the literature in the last 25 years, none of them display conditions that appear relevant to prebiotic chemistry. The quest for small-molecule reactions, exhibiting asymmetric replication and persisting high selectivity, compatible with primeval conditions has recently been suggested as a key challenge for organic chemists.⁵²³ Studying complex networks of organic chemical reactions⁵²⁴ instead of single auto-catalytic events might shed light on cooperative systems from which homochirality might emerge.³⁰² In this context, open systems with a continuous supply of reactants are better suited to reach homochiral NESS and it is expected that current progresses made in studying self-assembly process under dissipative conditions⁵²⁵ will be extended to chiral reaction networks.⁴¹

4.3 Homochirality through polymerization

Purely abiotic theory is based on the argument that enantiomeric cross-inhibition will ineluctably impede the formation of potent replicators. However, the fact that chemical processes may follow dramatically different mechanisms depending on the conditions has been overlooked. Likewise, stereoselective and non-selective polymerization reactions which allow regular and random arrangements of the monomer enantiomers along the polymer backbone, respectively, are ubiquitous in polymer science, and cross-inhibition is likely to be the exception rather than the norm.^{526,527}

a Stereoselective polymerization or ligation

Along this line, experimental efforts have been devoted to demonstrate the preferential formation of isotactic over heterochiral macromolecules for reactions initiated with racemic mixture of amino acids. An additional objective of

these studies was to demonstrate the emergence of homochiral oligomers, of a sufficient size to sustain a secondary structure. It is indeed well established that the helical configuration present in peptides tends to improve further the stereoselectivity of the polymerization process, through preferential helical growth.^{485,528} Goldberg studied the ligation of enantiopure amino esters, dipeptides and tripeptides (derived from alanine, aspartic acid and glycine) to racemic mixtures of activated alanine or aspartic acid amino esters in DMF and found a modest but significant bias towards the formation of homochiral peptides in the majority of cases.⁵²⁹ More recent investigations by the group of Luisi on the polymerization of racemic α -amino acid NCAs of leucine (Leu), isoleucine (Ile), tryptophan (Trp) and glutamic acid (Glu) in buffered aqueous solution also indicated a slight bias towards homochiral sequences.⁵³⁰ Excess factors, calculated relatively to a stereorandom polymerization process, were higher for the longer oligomers.⁵³¹ In the case of Leu, the presence of (α)-quartz as 1:1 mixture of the *d* and *l* enantiomorphs was found to improve the stereoselectivity of the polymerization process thanks to the selective adsorption of the more regular homochiral peptides on the quartz surface.⁵³² The combination of (α)-quartz and a reaction mixture biased in favour of one of the amino-acid enantiomer (20% *e.e.*) was necessary to get homochiral sequences as the major component of the peptide stereoisomers.⁵³³ The length of peptides reached under these conditions remains limited ($n < 10$) which lets the question of how long and well-structured homochiral peptides sequences emerged from the prebiotic soup unanswered. One possibility is that their formation was triggered by a ribozyme, *i.e.* that the construction of functional and catalytic RNAs preceded the generation of peptides and proteins.⁵¹⁶

Synthetic chemistry aimed at mimicking prebiotic conditions for the synthesis of RNA oligomers has provided some support along this direction. Oligomers of up to 55 nucleotides can be synthesized by successive elongation of a decanucleotide with enantiopure nucleotides on Na^+ -montmorillonite.²⁸⁸ Subsequent experiments have then been conducted directly from racemic mixtures of activated mononucleotides in order to probe the possibility of generating homochiral RNA oligomers, again with Na^+ -montmorillonite. Activated racemic adenosine oligomerized with comparable efficiency to enantiopure D-monomers discarding significant enantiomeric cross-inhibition.⁵³⁴ The distribution of oligomer stereoisomers (up to 8 units) appeared to be biased in favour of homochiral sequences. Deeper investigation of these reactions confirmed important and modest chiral selection in the oligomerization of activated adenosine^{535–537} and uridine, respectively.⁵³⁷ Co-oligomerization reaction of activated adenosine and uridine exhibited greater efficiency (up to 74% homochiral selectivity for the trimers) compared with the separate reactions of enantiomeric activated monomers.⁵³⁸ Again, the length of

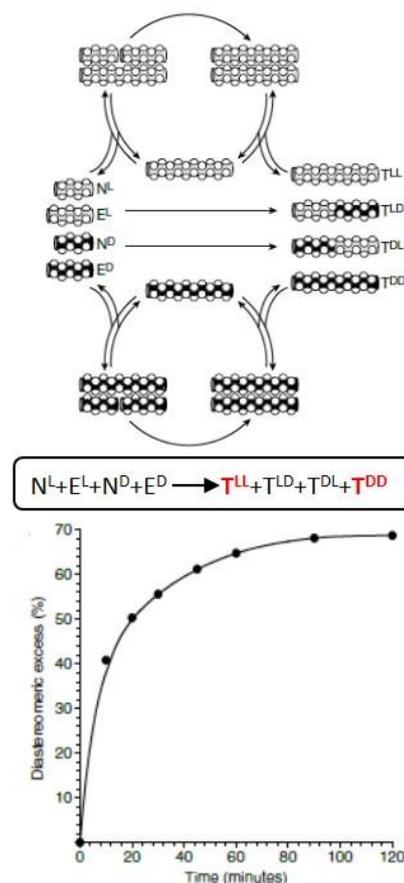


Figure 22 Top: schematic representation of the stereoselective replication of peptide residues with the same handedness. Below: diastereomeric excess (*de*) as a function of time. $de (\%) = [(T^L + T^{DD}) - (T^D + T^{LL})] / T^{\text{total}}$. Adapted from reference⁵³⁹ with permission from Nature publishing group.

oligomers detected in these experiments is far below the estimated number of nucleotides necessary to instigate chemical evolution.⁵⁴⁰ This questions the plausibility of RNA as the primeval informational polymer. Joyce and co-workers evoked the possibility of a more flexible chiral polymer based on acyclic nucleoside analogues as an ancestor of the more rigid furanose-based replicators but this hypothesis has not been probed experimentally.⁵⁴¹

Replication provided an advantage for achieving stereoselectivity at the condition that reactivity of chirally mismatched combinations are disfavoured relative to homochiral ones. A 32-residue peptide replicator was designed to probe the relationship between homochirality and self-replication.⁵³⁹ Electrophilic and nucleophilic 16-residue peptide fragments of the same handedness were preferentially ligated, even in the presence of their enantiomers (*ca.* 70% of diastereomeric excess was reached when peptide fragments E^L , E^D , N^E , and N^D were engaged, Figure 22). The replicator entails a stereoselective autocatalytic cycle, for which all bimolecular steps are faster for matched *versus* unmatched pairs of substrate enantiomers, thanks to self-recognition driven by hydrophobic interactions.⁵⁴² The process is very sensitive to the optical purity of the substrates, fragments embedding a single (*S*)/(*R*) amino acid substitution lacked significant auto-catalytic properties. On the contrary,

stereochemical mismatches were tolerated in the replicator: single mutated templates were able to couple homochiral fragments, a process referred to as “dynamic stereochemical editing”.

Templating also appeared to be crucial for promoting the oligomerization of nucleotides in a stereoselective way. The complementarity between nucleobase pairs was exploited to achieve homochiral sequences of pyranosyl-RNA.⁴²¹ Activated homochiral tetramers containing hemi self-complementary base sequences (pr(GCCG)-2'3'cyclophosphate, pr= pyranosyl-ribo) yielded relatively long oligomers (a ten of units) under mild conditions. Heterochiral tetramers (*e.g.* DDDL, DDLD, DLDD stereoisomers) were found to be poorly reactive under the same conditions. Importantly, the oligomerization of the homochiral tetramer was only slightly affected when conducted in the presence of heterochiral tetramers. These results raised the possibility that a similar experiment performed with the whole set of stereoisomers would have generated “predominantly homochiral” (*L*) and (*D*) sequences libraries of relatively long p-RNA oligomers. The studies with replicating peptides or auto-oligomerizing pyranosyl tetramers undoubtedly yield peptides and RNA oligomers that are both longer and optically purer than in the aforementioned reactions (part 4.2) involving activated monomers. Further work is needed to delineate whether these elaborated molecular frameworks could have emerged from the prebiotic soup.

Replication in the aforementioned systems stems from the stereoselective non-covalent interactions established between products and substrates. Stereoselectivity in the aggregation of non-enantiopure chemical species is a key mechanism for the emergence of homochirality in the various states of matter.⁵⁴³ The formation of homochiral *versus* heterochiral aggregates with different macroscopic properties led to enantioenrichment of scalemic mixtures through SDE as discussed in 4.2. Alternatively, homochiral aggregates might serve as templates at the nanoscale. In this context, the ability of serine (Ser) to form preferentially octamers when ionized from its enantiopure form is intriguing.⁵⁴⁴ Moreover, (*S*)-Ser in these octamers can be substituted enantiospecifically by prebiotic molecules (notably *D*-sugars)⁵⁴⁵ suggesting an important role of this amino acid in prebiotic chemistry. However, the preference for homochiral clusters is strong but not absolute and other clusters form when the ionization is conducted from racemic Ser,^{546,547} making the implication of serine clusters in the emergence of homochiral polymers or aggregates doubtful.

Lahav and co-workers investigated into details the correlation between aggregation and reactivity of amphiphilic activated racemic α -amino acids.⁵⁴⁸ These authors found that the stereoselectivity of the oligomerization reaction is strongly enhanced under conditions for which β -sheet aggregates are initially present⁵⁴⁹ or emerge during the reaction process.^{550–552} These supramolecular aggregates serve as templates in the propagation step of chain elongation leading to long peptides and co-peptides with a significant bias towards homochirality. Large enhancement of the homochiral content was detected,

notably for the oligomerization of *rac*-Val NCA in presence of 5% of an initiator (Figure 23).⁵⁵¹ Racemic mixtures of isotactic peptides are desymmetrized by adding chiral initiators⁵⁵¹ or by biasing the initial enantiomer composition.^{553,554} The interplay between aggregation and reactivity might have played a key role for the emergence of primeval replicators.

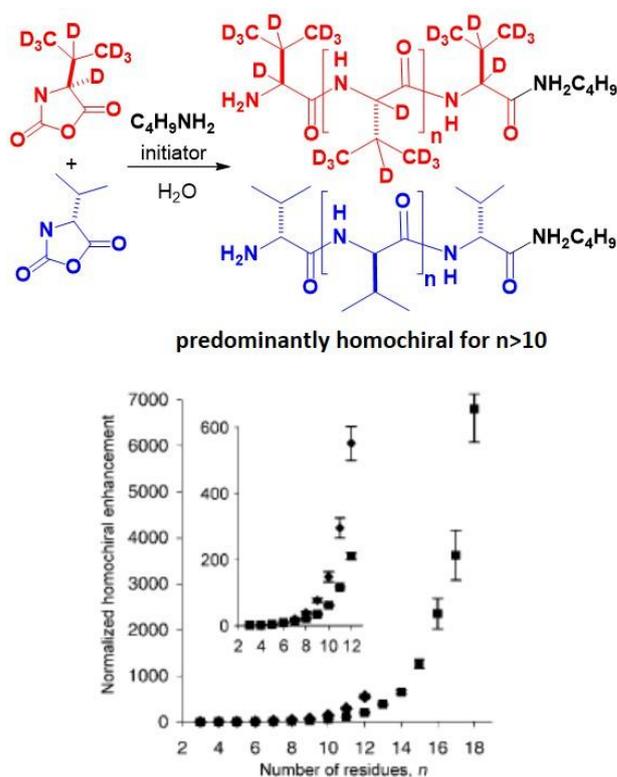


Figure 23 Stereoselective polymerization of *rac*-Val N-carboxyanhydride in presence of 5 mol% (square) or 25 mol% (diamond) of *n*-butylamine as initiator. Homochiral enhancement is calculated relatively to a binomial distribution of the stereoisomers. Reprinted from reference⁵⁵¹ with permission from Wiley-VCH.

b Heterochiral polymers

DNA and RNA duplexes as well as protein secondary and tertiary structures are usually destabilized by incorporating chiral mismatches, *i.e.* by substituting the biological enantiomer by its antipode. As a consequence, heterochiral polymers, which can hardly be avoided from reactions initiated by racemic or quasi racemic mixtures of enantiomers, are mainly considered in the literature as hurdles for the emergence of biological systems. Several authors have nevertheless considered that these polymers could have formed at some point of the chemical evolution process towards potent biological polymers. This is notably based on the observation that the extent of destabilization of heterochiral *versus* homochiral macromolecules depends on a variety of factors, including the nature, number, location, and environment of the substitutions,⁵⁵⁵ *e.g.* certain *D* to *L* mutations are tolerated in DNA duplexes.⁵⁵⁶ Moreover, simulations recently suggested that “demi-chiral” proteins, which contain 1:1 ratio of (*R*) and (*S*) α -amino acids, even though less stable than their homochiral analogues, exhibit

structural requirements (folding, substrate binding and active site) suitable for promoting early metabolism (e.g. t-RNA and DNA ligase activities).⁵⁵⁷ Likewise, several

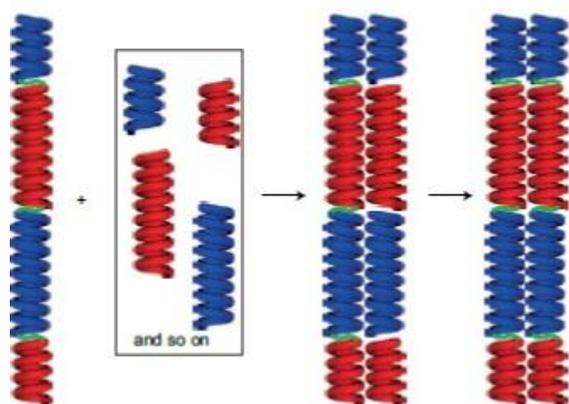


Figure 24 Principle of chiral encoding in the case of a template consisting of regions of alternating helicity. The initially formed heterochiral polymer replicates by recognition and ligation of its constituting helical fragments. Reprinted from reference⁵⁵⁷ with permission from Nature publishing group.

racemic membranes, *i.e.* composed of lipid antipodes, were found to be of comparable stability than homochiral ones.⁵²¹

Several scenarios towards BH involve non-homochiral polymers as possible intermediates towards potent replicators. Joyce proposed a three-phase process towards the formation of genetic material assuming the formation of flexible polymers, constructed from achiral or prochiral acyclic nucleoside analogues, as intermediates towards RNA and finally DNA.⁵⁴¹ It was presumed that ribose-free monomers would be more easily accessed from the prebiotic soup than ribose ones and that the conformational flexibility of these polymers would work against enantiomeric cross-inhibition. Other simplified structures, relatively to RNA, have been proposed by others.⁵⁵⁸ However, the molecular structures of the proposed building blocks is still complex relatively to what is expected to be readily generated from the prebiotic soup. Davis hypothesized a set of more realistic polymers that could have emerged from very simple building blocks such as formaldehyde, α -substituted ketones, aldehydes, alkenes, amino-acids or α -hydroxy acids.⁴²² Polymers with random arrangement of (*R*) and (*S*) stereogenic centres are expected to be replicated through recognition of their chiral sequence. Such chiral encoding⁵⁵⁹ might allow the emergence of replicators with specific catalytic properties. If one considers that the large number of possible sequences exceeds the number of molecules present in a reasonably sized sample of these chiral informational polymers, then their mixture will not constitute a perfect racemate since certain heterochiral polymers will lack their enantiomers. This argument of the emergence of homochirality or of a chiral bias “by chance” mechanism through the polymerization of a racemic mixture was also put forward previously by Eschenmoser⁴²¹ and Siegel.¹⁷ This concept has been sporadically probed notably through the template-controlled copolymerization of the racemic mixtures of two different activated amino acids.^{560–562}

However, in absence of any chiral bias, it is more likely that this mixture will yield informational polymers with pseudo enantiomeric like structures rather than the idealized chirally uniform polymers (see part 4.4). Finally, Davis also considered that pairing and replication between heterochiral polymers could operate through interaction between their helical structures, rather than on their individual stereogenic centres (Figure 24).⁴²² On this specific point, it should be emphasized that the helical conformation adopted by the main chain of certain types of polymers can be “amplified”, *i.e.* that single handed fragments may form even if composed of non-enantiopure building blocks.⁵⁶³ For example, synthetic polymers embedding a modestly biased racemic mixture of enantiomers adopt a single-handed helical conformation thanks to the so-called “majority-rules” effect.^{564–566} This phenomenon might have helped to enhance the helicity of the primeval heterochiral polymers relatively to the optical purity of their feeding monomers.

c. Theoretical models of polymerization

Several theoretical models accounting for the homochiral polymerization of a molecule in the racemic state, *i.e.* mimicking a prebiotic polymerization process, were developed by means of kinetic or probabilistic approaches. As early as 1966, Yamagata proposed that stereoselective polymerization coupled with different activation energies between reactive stereoisomers will “accumulate” the slight difference in energies between their composing enantiomers (assumed to originate from PVED) to eventually favour the formation of a single homochiral polymer.¹⁰⁸ Amongst other criticisms,¹²⁴ the unrealistic condition of perfect stereoselectivity has been pointed out.⁵⁶⁷ Yamagata later developed a probabilistic model which (i) favours ligations between monomers of the same chirality without discarding chirally mismatched combinations, (ii) gives an advantage of bonding between L monomers (again thanks to PVED) and (iii) allows racemization of the monomers and reversible polymerization. Homochirality in that case appears to develop much more slowly than the growth of polymers.⁵⁶⁸ This conceptual approach neglects enantiomeric cross-inhibition and relies on the difference in reactivity between enantiomers which has not been observed experimentally. The kinetic model developed by Sandars⁵⁶⁹ in 2003 received deeper attention as it revealed some intriguing features of homochiral polymerization processes. The model is based on the following specific elements: (i) chiral monomers are produced from an achiral substrate, (ii) cross-inhibition is assumed to stop polymerization, (iii) polymers of a certain length *N* catalyses the formation of the enantiomers in an enantiospecific fashion (similarly to a nucleotide synthetase ribozyme), and (iv) the system operates with a continuous supply of substrate and a withhold of polymers of length *N* (*i.e.* the system is open). By introducing a slight difference in the initial concentration values of the (*R*) and (*S*) enantiomers, bifurcation³⁰⁴ readily occurs, *i.e.* homochiral polymers of a single enantiomer are formed. The required conditions are sufficiently high values of the kinetic constants associated with enantioselective production of the enantiomers and cross-inhibition.

The Sandars model was modified in different ways by several groups^{570–574} to integrate more realistic parameters such as the possibility for polymers of all lengths to act catalytically in the breakdown of the achiral substrate into chiral monomers (instead of solely polymers of length N in the model of

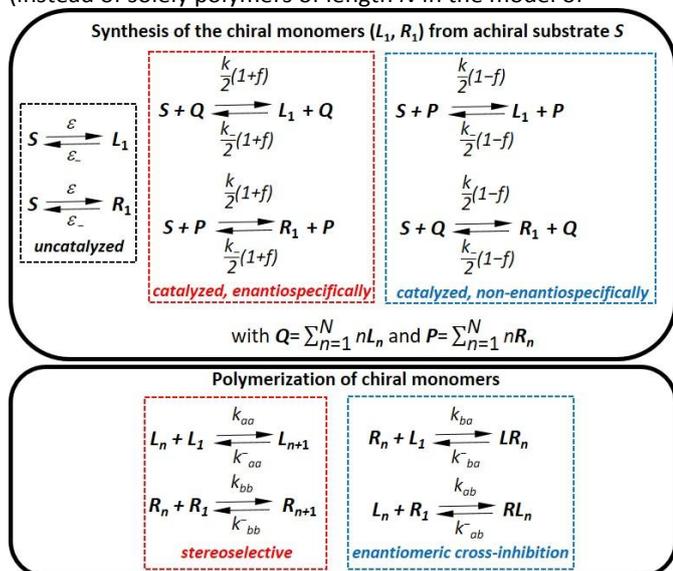


Figure 25 Hochberg model for chiral polymerization in closed systems. N = maximum chain length of the polymer, f = fidelity of the feedback mechanism, Q and P are the total concentrations of *left*-handed and *right*-handed polymers, respectively. ε (ε_{-}), k (k_{-}), k_{aa} (k_{aa}), k_{bb} (k_{bb}), k_{ba} (k_{ba}), k_{ab} (k_{ab}) denote the forward (reverse) reaction rate constants. Adapted from reference⁵⁷⁵ with permission from the Royal Society of Chemistry.

Sandars).^{64,575} Hochberg considered in addition a closed chemical system (*i.e.* the total mass of matter is kept constant) which allows polymers to grow towards a finite length (see reaction scheme in Figure 25).⁶⁴ Starting from an infinitesimal *e.e.* bias ($e.e.^0 = 5 \times 10^{-8}\%$), the model shows the emergence of homochiral polymers in an absolute but temporary manner. The reversibility of this SMSB process was expected for an open system. Ma and co-workers recently published a probabilistic approach which is presumed to better reproduce the emergence of the primeval RNA replicators and ribozymes in the RNA World.⁵⁷⁶ The D-nucleotide and L-nucleotide precursors are set to racemize to account for the behaviour of glyceraldehyde under prebiotic conditions; and the polynucleotide synthesis is surface- or template-mediated. The emergence of RNA polymers with RNA replicase or nucleotide synthase properties during the course of the simulation led to amplification of the initial chiral bias. Finally, several models show that cross-inhibition is not a necessary condition for the emergence of homochirality in polymerization processes. Higgs and co-workers considered all polymerization steps to be random (*i.e.* occurring with the same rate constant) whatever the nature of condensed monomers and that a fraction of homochiral polymers catalyzes the formation of the monomer enantiomers in an enantiospecific manner.⁵⁷⁷ The simulation yielded homochiral polymers (of both antipodes) even from a pure racemate under conditions which favour the catalyzed over non-catalyzed synthesis of the monomers. These polymers are referred as “chiral living polymers” as the result of their auto-catalytic properties. Hochberg modified its

previous kinetic reaction scheme drastically by suppressing cross-inhibition (polymerization operates through a stereoselective and cooperative mechanism only), and by allowing fragmentation and fusion of the homochiral polymer chains.⁵⁷⁸ The process of fragmentation is irreversible for the longest chains, mimicking a mechanical breakage. This breakage represents an external energy input to the system. This binary chain fusion mechanism is necessary to achieve SMSB in this simulation from infinitesimal chiral bias ($e.e.^0 = 5 \times 10^{-11}\%$). Finally, even though not specifically designed for a polymerization process, a recent model by Ribó and Hochberg show how homochiral replicators could emerge from two or more catalytically coupled asymmetric replicators, again with no need of the inclusion of a heterochiral inhibition reaction.³⁵⁰ Six homochiral replicators emerge from their simulation by means of an open flow reactor incorporating six achiral precursors and replicators in low initial concentrations and minute chiral biases ($e.e.^0 = 5 \times 10^{-18}\%$). These models should stimulate the quest of polymerization pathways which include stereoselective ligation, enantioselective synthesis of the monomers, replication and cross-replication, *i.e.* hallmarks of an ideal stereoselective polymerization process.

4.4 Purely biotic scenarios

In the previous two sections, the emergence of BH was dated at the level of prebiotic building blocks of Life (for purely abiotic theories) or at the stage of the primeval replicators, *i.e.* at the early or advanced stages of the chemical evolution, respectively. In most theories, an initial chiral bias was amplified yielding either prebiotic molecules or replicators as single enantiomers. Others hypothesized that homochiral replicators and then Life emerged from unbiased racemic mixtures by chance, basing their rationale on probabilistic grounds.^{17,421,559,577} In 1957, Fox,⁵⁷⁹ Rush⁵⁸⁰ and Wald⁵⁸¹ held a different view and independently emitted the hypothesis that BH is an inevitable consequence of the evolution of the living matter.⁴⁴ Wald notably reasoned that, since polymers made of homochiral monomers likely propagate faster, are longer and have stronger secondary structures (*e.g.* helices), it must have provided sufficient criteria to the chiral selection of amino acids thanks to the formation of their polymers in *ad hoc* conditions. This statement was indeed confirmed notably by experiments showing that stereoselective polymerization is enhanced when oligomers adopt a α -helix conformation.^{485,528} However, Wald went a step further by supposing that homochiral polymers of both handedness would have been generated under the supposedly symmetric external forces present on prebiotic Earth and that primordial Life would have originated under the form of two populations of organisms, enantiomers of each other. From then, the natural forces of evolution led certain organisms to be superior to their enantiomeric neighbours leading to Life in a single form, as we know it today. The purely biotic theory of emergence of BH thanks to biological evolution, instead of chemical evolution for abiotic theories, was accompanied with large scepticism in the literature even though the arguments of Wald were

developed later on by others⁴⁴ and notably by Ageno (sexual reproduction naturally resolves enantiomeric populations),⁵⁸² and Kuhn (the stronger enantiomeric form of Life survived in the “struggle”).⁵⁸³ More recently, Green and Jain summarized the Wald theory into the catchy formula “Two Runners, One Tripped”,⁵⁸⁴ and called for deeper investigation on routes towards racemic mixtures of biologically relevant polymers.

The Wald theory by its essence has been difficult to assess experimentally. On the one side, (*R*)-amino acids when found in mammals are often related to destructive and toxic effects suggesting a lack of complementarity with the current biological machinery in which (*S*)-amino acids are ultra-predominating. On the other side, (*R*)-amino acids have been detected in the cell wall peptidoglycan layer of bacteria⁵⁸⁵ and in various peptides of bacteria, archaea, and eukaryotes.¹⁶ (*R*)-amino acids in these various living systems have an unknown origin. Certain proponents of the purely biotic theories suggest that the small but general occurrence of (*R*)-amino acids in nowadays living organisms can be a relic of a time in which mirror-image living systems were “struggling”. Likewise, to rationalize the aforementioned “lipid divide”, it has been proposed that the LUCA of bacteria and archaea could have embedded a heterochiral lipid membrane, *i.e.* a membrane containing two sorts of lipid with opposite configurations.⁵²¹

Several studies also probed the possibility to prepare a biological system containing the enantiomers of the molecules of Life as we know it today. L-poly-nucleotides and (*R*)-polypeptides were synthesized; and expectedly they exhibited chiral substrate specificity and biochemical properties that mirrored those of their natural counterparts.^{586–588} In a recent example, Liu, Zhu and co-workers showed that a synthesized 174-residue (*R*)-polypeptide catalyzes the template-directed polymerization of L-DNA and its transcription into L-RNA.⁵⁸⁷ It was also demonstrated that the synthesized and natural DNA polymerase systems operate without any cross-inhibition when mixed together in presence of a racemic mixture of the constituents required for the reaction (D- and L primers, D- and L-templates and D- and L-dNTPs). From these impressive results, it is easy to imagine how mirror-image ribozymes would have worked independently in the early evolution times of primeval living systems.

One puzzling question concerns the feasibility for a biopolymer to synthesize its mirror-image. This has been addressed elegantly by the group of Joyce which demonstrated very recently the possibility for a RNA polymerase ribozyme to catalyze the templated synthesis of RNA oligomers of the opposite configuration.⁵⁸⁹ The D-RNA ribozyme was selected, through 16 rounds of selective amplification away from a random sequence, for its ability to catalyze the ligation of two L-RNA substrates on a L-RNA template. The D-RNA ribozyme exhibited sufficient activity to generate full-length copies of its enantiomer through the template-assisted ligation of 11 oligonucleotides. A variant of this cross-chiral enzyme was able to assemble a two-fragment form of a former version of the ribozyme from a mixture of trinucleotide building blocks.⁵⁹⁰ Again, no inhibition was detected when the ribozyme enantiomers were put together with the racemic substrates

and templates (Figure 26). In the hypothesis of a RNA world, it is intriguing to consider the possibility of a primordial ribozyme with cross-catalytic polymerization activities. In such a case, one can consider the possibility that enantiomeric ribozymes would

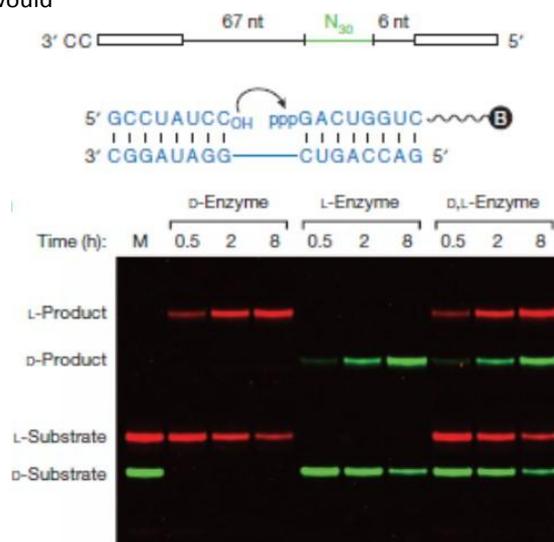


Figure 26 Cross-chiral ligation: the templated ligation of two oligonucleotides (shown in blue, B: biotin) is catalyzed by a RNA enzyme of the opposite handedness (open rectangle: primers, N₃₀: optimized nucleotide (nt) sequence). The D and L substrates are labelled with either fluoresceine (green) or boron-dipyrromethene (red), respectively. No enantiomeric cross-inhibition is observed when the racemic substrates, enzymes, and templates are mixed together. Adapted from reference⁵⁸⁹ with permission from Nature publishing group.

have existed concomitantly and that evolutionary innovation would have favoured the systems based on D-RNA and (*S*)-polypeptides leading to the exclusive form of BH present on Earth nowadays. Finally, a strongly convincing evidence for the standpoint of the purely biotic theories would be the discovery in sediments of primitive forms of Life based on a molecular machinery entirely composed of (*R*)-amino acids and L-nucleic acids.

5. Conclusions and Perspectives of Biological Homochirality Studies

Questions accumulated while considering all the possible origins of the initial enantiomeric imbalance that have ultimately led to biological homochirality. When some hypothesize a reason behind its emergence (such as for informational entropic reasons, resulting in evolutionary advantages towards more specific and complex functionalities),^{25,350} others wonder whether it is reasonable to reconstruct a chronology 3.5 billion years later.³⁷ Many are circumspect in front of the pile-up of scenarios and assert that the solution is likely not expected in a near future (due to the difficulty to do all required control experiments, and fully understand the theoretical background of the putative selection mechanism).⁵³ In parallel, the existence, and the extent, of a putative link between the different configurations of biologically-relevant amino acids and sugars also remain unsolved,⁵⁹¹ and only Goldanskii and Kuz'min studied the

effects of a hypothetical global loss of optical purity in the future.⁴²⁹

Nevertheless, great progress has been made recently for a better perception of this long-standing enigma. The scenario involving circularly polarized light as a chiral bias inducer is more and more convincing thanks to operational and analytical improvements. Increasingly accurate computational studies supply precious information, notably about SMSB processes, chiral surfaces, and other truly chiral influences. Asymmetric autocatalytic systems and deracemization processes have also undoubtedly grown in interest (notably thanks to the discoveries of the Soai reaction and the Viedma ripening). Space missions are also an opportunity: to study *in situ* organic matter, its conditions of transformations, and possible associated enantio-enrichment; to elucidate the solar system origin and its history; and maybe, to find traces of chemicals with “unnatural” configurations in celestial bodies, what could indicate that the chiral selection of terrestrial BH could be a mere coincidence.

The current state-of-the-art indicates that further experimental investigations of the possible effect of other sources of asymmetry are needed. Photochirogenesis is attractive in many respects: CPL has been detected in space, *e.es.* have been measured for several prebiotic molecules found on meteorites or generated in laboratory-reproduced interstellar ices. However, this detailed postulated scenario still faces pitfalls related to the variable sources of extra-terrestrial CPL, the requirement of finely-tuned illumination conditions (almost full extent of reaction at the *right* place and moment of the evolutionary stages), and the unknown mechanism leading to the amplification of the original chiral biases. Strong calls to organic chemists are thus necessary, to discover new asymmetric autocatalytic reactions, maybe through the investigation of complex and large chemical systems,⁵⁹² that can meet the criteria of primordial conditions.^{40,41,302,312}

Anyway, the quest of the biological homochirality origin is fruitful in many aspects. The first concerns one consequence of the asymmetry of Life: the contemporary challenge of synthesizing enantiopure bioactive molecules. Indeed, many synthetic efforts are directed towards the generation of optically-pure molecules, to avoid potential side effects of racemic mixtures due to the enantioselectivity of biological receptors. These endeavors can undoubtedly draw inspiration from the range of deracemization and chirality induction processes conducted in connection with biological homochirality. One example is the Viedma ripening, which allows the preparation of enantiopure molecules displaying potent therapeutic activities.^{55,593} Other efforts are devoted to the building-up of sophisticated experiments and pushing their measurement limits to be able to detect tiny enantiomeric excesses, thus strongly contributing to important improvements in scientific instrumentation and acquiring fundamental knowledge at the interface between chemistry, physics, and biology. Overall, this joint endeavor at the frontier of many fields is also beneficial to material sciences notably for

the elaboration of biomimetic materials and emerging chiral materials.^{594,595}

Abbreviations

BH: Biological Homochirality
 CISS: Chiral-Induced Spin Selectivity
 CD: circular dichroism
de: diastereomeric excess
 DFT: Density Functional Theories
 DNA: DeoxyriboNucleic Acid
 dNTPs: deoxyNucleotide TriPhosphates
e.e(s).: enantiomeric excess(es)
 EPR: Electron Paramagnetic Resonance
 Epi: epichlorohydrin
 FCC: Face-Centred Cubic
 GC: Gas Chromatography
 LES: Limited EnantioSelective
 LUCA: Last Universal Cellular Ancestor
 MCD: Magnetic Circular Dichroism
 MChD: Magneto-Chiral Dichroism
 MS: Mass Spectrometry
 MW: MicroWave
 NESS: Non-Equilibrium Stationary States
 NCA: *N*-carboxy-anhydride
 NMR: Nuclear Magnetic Resonance
 OEEF: Oriented-External Electric Fields
 Pr: Pyranosyl-ribo
 PV: Parity Violation
 PVED: Parity-Violating Energy Difference
 REF: Rotating Electric Fields
 RNA: RiboNucleic Acid
 SDE: Self-Disproportionation of the Enantiomers
 SEs: Secondary Electrons
 SMSB: Spontaneous Mirror Symmetry Breaking
 SNAAP: Supernova Neutrino Amino Acid Processing
 SPEs: Spin-Polarized Electrons
 VUV: Vacuum UltraViolet

Author Contributions

QS selected the scope of the review, made the first critical analysis of the literature and wrote the first draft of the review. JC modified parts 1 and 2 according to her expertise to the domains of chiral physical fields and parity violation. MR re-organized the review into its current form and extended parts 3 and 4. All authors were involved in the revision and proof-checking of the successive versions of the review.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

The French Agence Nationale de la Recherche is acknowledged for funding the project AbsoluCat (ANR-17-CE07-0002) to MR. The GDR 3712 Chirafun from Centre National de la recherche Scientifique (CNRS) is acknowledged for allowing a collaborative network between partners involved in this review. J.C. warmly thanks Dr. Benoît Darquié from the Laboratoire de Physique des Lasers (Université Sorbonne Paris Nord) for fruitful discussions and precious advice.

Notes and references

& Proteinogenic amino acids and natural sugars are usually mentioned as L-amino acids and D-sugars according to the descriptors introduced by Emil Fischer. It is worth noting that the natural L-cysteine is (R) using the Cahn–Ingold–Prelog system, due to the sulfur atom in the side chain which changes the priority sequence. In the present review, (R)/(S) and D/L descriptors will be used for amino acids and sugars, respectively, as commonly employed in the literature dealing with BH.

- W. Thomson, Baltimore Lectures on Molecular Dynamics and the Wave Theory of Light, Cambridge Univ. Press. Warehouse, 1894, Edition of 1904, pp 619.
- K. Mislow, in *Topics in Stereochemistry*, ed. S. E. Denmark, John Wiley & Sons, Inc., Hoboken, NJ, USA, 2007, pp. 1–82.
- B. Kahr, *Chirality*, 2018, **30**, 351–368.
- S. H. Mauskopf, *Trans. Am. Philos. Soc.*, 1976, **66**, 1–82.
- J. Gal, *Helv. Chim. Acta*, 2013, **96**, 1617–1657.
- L. Pasteur, *Comptes Rendus Hebd. Séances Académie Sci.*, 1848, **26**, 535–538.
- C. Djerassi, R. Records, E. Bunnenberg, K. Mislow and A. Moscowitz, *J. Am. Chem. Soc.*, 1962, 870–872.
- S. J. Gerbode, J. R. Puzey, A. G. McCormick and L. Mahadevan, *Science*, 2012, **337**, 1087–1091.
- G. H. Wagnière, *On Chirality and the Universal Asymmetry. Reflections on Image and Mirror Image*, VHCA, Verlag Helvetica Chimica Acta, Zürich (Switzerland), 2007.
- H.-U. Blaser, *Rendiconti Lincei*, 2007, **18**, 281–304.
- H.-U. Blaser, *Rendiconti Lincei*, 2013, **24**, 213–216.
- A. Rouf and S. C. Taneja, *Chirality*, 2014, **26**, 63–78.
- H. Leek and S. Andersson, *Molecules*, 2017, **22**, 158.
- D. Rossi, M. Tarantino, G. Rossino, M. Rui, M. Juza and S. Collina, *Expert Opin. Drug Discov.*, 2017, **12**, 1253–1269.
- Nature, Editorial: Asymmetry symposium unites economists, physicists and artists*, 2018, **555**, 414.
- Y. Nagata, T. Fujiwara, K. Kawaguchi-Nagata, Yoshihiro Fukumori and T. Yamanaka, *Biochim. Biophys. Acta BBA - Gen. Subj.*, 1998, **1379**, 76–82.
- J. S. Siegel, *Chirality*, 1998, **10**, 24–27.
- J. D. Watson and F. H. C. Crick, *Nature*, 1953, **171**, 737.
- L. Pasteur, *Comptes Rendus Hebd. Séances Académie Sci.*, 1857, **45**, 1032–1036.
- L. Pasteur, *Comptes Rendus Hebd. Séances Académie Sci.*, 1858, **46**, 615–618.
- J. Gal, *Chirality*, 2008, **20**, 5–19.
- J. Gal, *Chirality*, 2012, **24**, 959–976.
- A. Piutti, *Comptes Rendus Hebd. Séances Académie Sci.*, 1886, **103**, 134–138.
- U. Meierhenrich, *Amino acids and the asymmetry of life: caught in the act of formation*, Springer, Berlin, 2008.
- L. Morozov, *Orig. Life*, 1979, **9**, 187–217.
- S. F. Mason, *Nature*, 1984, **311**, 19–23.
- S. Mason, *Chem. Soc. Rev.*, 1988, **17**, 347–359.
- W. A. Bonner, in *Topics in Stereochemistry*, eds. E. L. Eliel and S. H. Wilen, John Wiley & Sons, Ltd, 1988, vol. 18, pp. 1–96.
- L. Keszthelyi, *Q. Rev. Biophys.*, 1995, **28**, 473–507.
- M. Avalos, R. Babiano, P. Cintas, J. L. Jiménez, J. C. Palacios and L. D. Barron, *Chem. Rev.*, 1998, **98**, 2391–2404.
- B. L. Feringa and R. A. van Delden, *Angew. Chem. Int. Ed.*, 1999, **38**, 3418–3438.
- J. Podlech, *Cell. Mol. Life Sci. CMLS*, 2001, **58**, 44–60.
- D. B. Cline, *Eur. Rev.*, 2005, **13**, 49–59.
- A. Guijarro and M. Yus, *The Origin of Chirality in the Molecules of Life: A Revision from Awareness to the Current Theories and Perspectives of this Unsolved Problem*, RSC Publishing, 2008.
- V. A. Tsarev, *Phys. Part. Nucl.*, 2009, **40**, 998–1029.
- D. G. Blackmond, *Cold Spring Harb. Perspect. Biol.*, 2010, **2**, a002147–a002147.
- M. Avalos, R. Babiano, P. Cintas, J. L. Jiménez and J. C. Palacios, *Tetrahedron Asymmetry*, 2010, **21**, 1030–1040.
- J. E. Hein and D. G. Blackmond, *Acc. Chem. Res.*, 2012, **45**, 2045–2054.
- P. Cintas and C. Viedma, *Chirality*, 2012, **24**, 894–908.
- J. M. Ribó, D. Hochberg, J. Crusats, Z. El-Hachemi and A. Moyano, *J. R. Soc. Interface*, 2017, **14**, 20170699.
- T. Buhse, J.-M. Cruz, M. E. Noble-Terán, D. Hochberg, J. M. Ribó, J. Crusats and J.-C. Micheau, *Chem. Rev.*, 2021, **121**, 2147–2229.
- G. Palyi, *Biological Chirality*, 1st Edition, Elsevier, 2019.
- M. Mauksch and S. B. Tsogoeva, in *Biomimetic Organic Synthesis*, John Wiley & Sons, Ltd, 2011, pp. 823–845.
- W. A. Bonner, *Orig. Life Evol. Biosph.*, 1991, **21**, 59–111.
- G. Zubay, *Origins of Life on the Earth and in the Cosmos*, Elsevier, 2000.
- K. Ruiz-Mirazo, C. Briones and A. de la Escosura, *Chem. Rev.*, 2014, **114**, 285–366.
- M. Yadav, R. Kumar and R. Krishnamurthy, *Chem. Rev.*, 2020, **120**, 4766–4805.
- M. Frenkel-Pinter, M. Samanta, G. Ashkenasy and L. J. Leman, *Chem. Rev.*, 2020, **120**, 4707–4765.
- L. D. Barron, *Science*, 1994, **266**, 1491–1492.
- J. Crusats and A. Moyano, *Synlett*, 2021, **32**, 2013–2035.
- V. I. Gol'danskiĭ and V. V. Kuz'min, *Sov. Phys. Uspekhi*, 1989, **32**, 1–29.
- D. B. Amabilino and R. M. Kellogg, *Isr. J. Chem.*, 2011, **51**, 1034–1040.
- M. Quack, *Angew. Chem. Int. Ed.*, 2002, **41**, 4618–4630.
- W. A. Bonner, *Chirality*, 2000, **12**, 114–126.
- L.-C. Sögütöglü, R. R. E. Steendam, H. Meekes, E. Vlieg and F. P. J. T. Rutjes, *Chem. Soc. Rev.*, 2015, **44**, 6723–6732.
- K. Soai, T. Kawasaki and A. Matsumoto, *Acc. Chem. Res.*, 2014, **47**, 3643–3654.
- J. R. Cronin and S. Pizzarello, *Science*, 1997, **275**, 951–955.
- A. C. Evans, C. Meinert, C. Giri, F. Goesmann and U. J. Meierhenrich, *Chem. Soc. Rev.*, 2012, **41**, 5447.
- D. P. Glavin, A. S. Burton, J. E. Elsilá, J. C. Aponte and J. P. Dworkin, *Chem. Rev.*, 2020, **120**, 4660–4689.
- J. Sun, Y. Li, F. Yan, C. Liu, Y. Sang, F. Tian, Q. Feng, P. Duan, L. Zhang, X. Shi, B. Ding and M. Liu, *Nat. Commun.*, 2018, **9**, 2599.
- J. Han, O. Kitagawa, A. Wzorek, K. D. Klika and V. A. Soloshonok, *Chem. Sci.*, 2018, **9**, 1718–1739.
- T. Satyanarayana, S. Abraham and H. B. Kagan, *Angew. Chem. Int. Ed.*, 2009, **48**, 456–494.
- K. P. Bryliakov, *ACS Catal.*, 2019, **9**, 5418–5438.
- C. Blanco and D. Hochberg, *Phys. Chem. Chem. Phys.*, 2010, **13**, 839–849.
- R. Breslow, *Tetrahedron Lett.*, 2011, **52**, 2028–2032.
- T. D. Lee and C. N. Yang, *Phys. Rev.*, 1956, **104**, 254–258.
- C. S. Wu, E. Ambler, R. W. Hayward, D. D. Hoppes and R. P. Hudson, *Phys. Rev.*, 1957, **105**, 1413–1415.

- 68 M. Drewes, *Int. J. Mod. Phys. E*, 2013, **22**, 1330019.
- 69 F. J. Hasert et al., *Phys. Lett. B*, 1973, **46**, 121–124.
- 70 F. J. Hasert et al., *Phys. Lett. B*, 1973, **46**, 138–140.
- 71 C. Y. Prescott et al., *Phys. Lett. B*, 1978, **77**, 347–352.
- 72 C. Y. Prescott et al., *Phys. Lett. B*, 1979, **84**, 524–528.
- 73 L. Di Lella and C. Rubbia, in *60 Years of CERN Experiments and Discoveries*, World Scientific, 2014, vol. 23, pp. 137–163.
- 74 M. A. Bouchiat and C. C. Bouchiat, *Phys. Lett. B*, 1974, **48**, 111–114.
- 75 M.-A. Bouchiat and C. Bouchiat, *Rep. Prog. Phys.*, 1997, **60**, 1351–1396.
- 76 L. M. Barkov and M. S. Zolotarev, *JETP*, 1980, **52**, 360–376.
- 77 C. S. Wood, S. C. Bennett, D. Cho, B. P. Masterson, J. L. Roberts, C. E. Tanner and C. E. Wieman, *Science*, 1997, **275**, 1759–1763.
- 78 J. Crassous, C. Chardonnet, T. Saue and P. Schwerdtfeger, *Org. Biomol. Chem.*, 2005, **3**, 2218.
- 79 R. Berger and J. Stohner, *Wiley Interdiscip. Rev. Comput. Mol. Sci.*, 2019, **9**, 25.
- 80 R. Berger, in *Theoretical and Computational Chemistry*, ed. P. Schwerdtfeger, Elsevier, 2004, vol. 14, pp. 188–288.
- 81 P. Schwerdtfeger, in *Computational Spectroscopy*, ed. J. Grunenberg, John Wiley & Sons, Ltd, pp. 201–221.
- 82 J. Eills, J. W. Blanchard, L. Bougas, M. G. Kozlov, A. Pines and D. Budker, *Phys. Rev. A*, 2017, **96**, 042119.
- 83 R. A. Harris and L. Stodolsky, *Phys. Lett. B*, 1978, **78**, 313–317.
- 84 M. Quack, *Chem. Phys. Lett.*, 1986, **132**, 147–153.
- 85 L. D. Barron, *Magnetochemistry*, 2020, **6**, 5.
- 86 D. W. Rein, R. A. Hegstrom and P. G. H. Sandars, *Phys. Lett. A*, 1979, **71**, 499–502.
- 87 R. A. Hegstrom, D. W. Rein and P. G. H. Sandars, *J. Chem. Phys.*, 1980, **73**, 2329–2341.
- 88 M. Quack, *Angew. Chem. Int. Ed. Engl.*, 1989, **28**, 571–586.
- 89 A. Bakasov, T.-K. Ha and M. Quack, *J. Chem. Phys.*, 1998, **109**, 7263–7285.
- 90 M. Quack, in *Quantum Systems in Chemistry and Physics*, eds. K. Nishikawa, J. Maruani, E. J. Brändas, G. Delgado-Barrio and P. Piecuch, Springer Netherlands, Dordrecht, 2012, vol. 26, pp. 47–76.
- 91 M. Quack and J. Stohner, *Phys. Rev. Lett.*, 2000, **84**, 3807–3810.
- 92 G. Rauhut and P. Schwerdtfeger, *Phys. Rev. A*, 2021, **103**, 042819.
- 93 C. Stoeffler, B. Darquié, A. Shelkovnikov, C. Daussy, A. Amy-Klein, C. Chardonnet, L. Guy, J. Crassous, T. R. Huet, P. Soulard and P. Asselin, *Phys. Chem. Chem. Phys.*, 2011, **13**, 854–863.
- 94 N. Saleh, S. Zrig, T. Roisnel, L. Guy, R. Bast, T. Saue, B. Darquié and J. Crassous, *Phys. Chem. Chem. Phys.*, 2013, **15**, 10952.
- 95 S. K. Tokunaga, C. Stoeffler, F. Auguste, A. Shelkovnikov, C. Daussy, A. Amy-Klein, C. Chardonnet and B. Darquié, *Mol. Phys.*, 2013, **111**, 2363–2373.
- 96 N. Saleh, R. Bast, N. Vanthuyne, C. Roussel, T. Saue, B. Darquié and J. Crassous, *Chirality*, 2018, **30**, 147–156.
- 97 B. Darquié, C. Stoeffler, A. Shelkovnikov, C. Daussy, A. Amy-Klein, C. Chardonnet, S. Zrig, L. Guy, J. Crassous, P. Soulard, P. Asselin, T. R. Huet, P. Schwerdtfeger, R. Bast and T. Saue, *Chirality*, 2010, **22**, 870–884.
- 98 A. Cournol, M. Manceau, M. Pierens, L. Lecordier, D. B. A. Tran, R. Santagata, B. Argence, A. Goncharov, O. Lopez, M. Abgrall, Y. L. Coq, R. L. Targat, H. Á. Martinez, W. K. Lee, D. Xu, P.-E. Pottie, R. J. Hendricks, T. E. Wall, J. M. Bieniewska, B. E. Sauer, M. R. Tarbutt, A. Amy-Klein, S. K. Tokunaga and B. Darquié, *Quantum Electron.*, 2019, **49**, 288.
- 99 C. Fábri, L. Horný and M. Quack, *ChemPhysChem*, 2015, **16**, 3584–3589.
- 100 P. Dietiker, E. Miloglyadov, M. Quack, A. Schneider and G. Seyfang, *J. Chem. Phys.*, 2015, **143**, 244305.
- 101 S. Albert, I. Bolotova, Z. Chen, C. Fábri, L. Horný, M. Quack, G. Seyfang and D. Zindel, *Phys. Chem. Chem. Phys.*, 2016, **18**, 21976–21993.
- 102 S. Albert, F. Arn, I. Bolotova, Z. Chen, C. Fábri, G. Grassi, P. Lerch, M. Quack, G. Seyfang, A. Wokaun and D. Zindel, *J. Phys. Chem. Lett.*, 2016, **7**, 3847–3853.
- 103 S. Albert, I. Bolotova, Z. Chen, C. Fábri, M. Quack, G. Seyfang and D. Zindel, *Phys. Chem. Chem. Phys.*, 2017, **19**, 11738–11743.
- 104 A. Salam, *J. Mol. Evol.*, 1991, **33**, 105–113.
- 105 A. Salam, *Phys. Lett. B*, 1992, **288**, 153–160.
- 106 T. L. V. Ulbricht, *Orig. Life*, 1975, **6**, 303–315.
- 107 F. Vester, T. L. V. Ulbricht and H. Krauch, *Naturwissenschaften*, 1959, **46**, 68–68.
- 108 Y. Yamagata, *J. Theor. Biol.*, 1966, **11**, 495–498.
- 109 S. F. Mason and G. E. Tranter, *Chem. Phys. Lett.*, 1983, **94**, 34–37.
- 110 S. F. Mason and G. E. Tranter, *J. Chem. Soc. Chem. Commun.*, 1983, 117–119.
- 111 S. F. Mason and G. E. Tranter, *Proc. R. Soc. Lond. Math. Phys. Sci.*, 1985, **397**, 45–65.
- 112 G. E. Tranter, *Chem. Phys. Lett.*, 1985, **115**, 286–290.
- 113 G. E. Tranter, *Mol. Phys.*, 1985, **56**, 825–838.
- 114 G. E. Tranter, *Nature*, 1985, **318**, 172–173.
- 115 G. E. Tranter, *J. Theor. Biol.*, 1986, **119**, 467–479.
- 116 G. E. Tranter, *J. Chem. Soc. Chem. Commun.*, 1986, 60–61.
- 117 G. E. Tranter, A. J. MacDermott, R. E. Overill and P. J. Speers, *Proc. Math. Phys. Sci.*, 1992, **436**, 603–615.
- 118 A. J. MacDermott, G. E. Tranter and S. J. Trainor, *Chem. Phys. Lett.*, 1992, **194**, 152–156.
- 119 G. E. Tranter, *Chem. Phys. Lett.*, 1985, **120**, 93–96.
- 120 G. E. Tranter, *Chem. Phys. Lett.*, 1987, **135**, 279–282.
- 121 A. J. MacDermott and G. E. Tranter, *Chem. Phys. Lett.*, 1989, **163**, 1–4.
- 122 S. F. Mason and G. E. Tranter, *Mol. Phys.*, 1984, **53**, 1091–1111.
- 123 R. Berger and M. Quack, *ChemPhysChem*, 2000, **1**, 57–60.
- 124 R. Wesendrup, J. K. Laerdahl, R. N. Compton and P. Schwerdtfeger, *J. Phys. Chem. A*, 2003, **107**, 6668–6673.
- 125 J. K. Laerdahl, R. Wesendrup and P. Schwerdtfeger, *ChemPhysChem*, 2000, **1**, 60–62.
- 126 G. Lente, *J. Phys. Chem. A*, 2006, **110**, 12711–12713.
- 127 G. Lente, *Symmetry*, 2010, **2**, 767–798.
- 128 A. J. MacDermott, T. Fu, R. Nakatsuka, A. P. Coleman and G. O. Hyde, *Orig. Life Evol. Biosph.*, 2009, **39**, 459–478.
- 129 A. J. MacDermott, *Chirality*, 2012, **24**, 764–769.
- 130 L. D. Barron, *J. Am. Chem. Soc.*, 1986, **108**, 5539–5542.
- 131 L. D. Barron, *Nat. Chem.*, 2012, **4**, 150–152.
- 132 K. Ishii, S. Hattori and Y. Kitagawa, *Photochem. Photobiol. Sci.*, 2020, **19**, 8–19.
- 133 G. L. J. A. Rikken and E. Raupach, *Nature*, 1997, **390**, 493–494.
- 134 G. L. J. A. Rikken, E. Raupach and T. Roth, *Phys. B Condens. Matter*, 2001, **294–295**, 1–4.
- 135 Y. Xu, G. Yang, H. Xia, G. Zou, Q. Zhang and J. Gao, *Nat. Commun.*, 2014, **5**, 5050.
- 136 M. Atzori, G. L. J. A. Rikken and C. Train, *Chem. – Eur. J.*, 2020, **26**, 9784–9791.
- 137 G. L. J. A. Rikken and E. Raupach, *Nature*, 2000, **405**, 932–935.
- 138 J. B. Clemens, O. Kibar and M. Chachisvilis, *Nat. Commun.*, 2015, **6**, 7868.
- 139 V. Marichez, A. Tassoni, R. P. Cameron, S. M. Barnett, R. Eichhorn, C. Genet and T. M. Hermans, *Soft Matter*, 2019, **15**, 4593–4608.

- 140 B. A. Grzybowski and G. M. Whitesides, *Science*, 2002, **296**, 718–721.
- 141 P. Chen and C.-H. Chao, *Phys. Fluids*, 2007, **19**, 017108.
- 142 M. Makino, L. Arai and M. Doi, *J. Phys. Soc. Jpn.*, 2008, **77**, 064404.
- 143 Marcos, H. C. Fu, T. R. Powers and R. Stocker, *Phys. Rev. Lett.*, 2009, **102**, 158103.
- 144 M. Aristov, R. Eichhorn and C. Bechinger, *Soft Matter*, 2013, **9**, 2525–2530.
- 145 J. M. Ribó, J. Crusats, F. Sagués, J. Claret and R. Rubires, *Science*, 2001, **292**, 2063–2066.
- 146 Z. El-Hachemi, O. Arteaga, A. Canillas, J. Crusats, C. Escudero, R. Kuroda, T. Harada, M. Rosa and J. M. Ribó, *Chem. - Eur. J.*, 2008, **14**, 6438–6443.
- 147 A. Tsuda, M. A. Alam, T. Harada, T. Yamaguchi, N. Ishii and T. Aida, *Angew. Chem. Int. Ed.*, 2007, **46**, 8198–8202.
- 148 M. Wolffs, S. J. George, Ž. Tomović, S. C. J. Meskers, A. P. H. J. Schenning and E. W. Meijer, *Angew. Chem. Int. Ed.*, 2007, **46**, 8203–8205.
- 149 O. Arteaga, A. Canillas, R. Purrello and J. M. Ribó, *Opt. Lett.*, 2009, **34**, 2177.
- 150 A. D'Urso, R. Randazzo, L. Lo Faro and R. Purrello, *Angew. Chem. Int. Ed.*, 2010, **49**, 108–112.
- 151 J. Crusats, Z. El-Hachemi and J. M. Ribó, *Chem. Soc. Rev.*, 2010, **39**, 569–577.
- 152 O. Arteaga, A. Canillas, J. Crusats, Z. El-Hachemi, J. Llorens, E. Sacristan and J. M. Ribo, *ChemPhysChem*, 2010, **11**, 3511–3516.
- 153 O. Arteaga, A. Canillas, J. Crusats, Z. El-Hachemi, J. Llorens, A. Sorrenti and J. M. Ribo, *Isr. J. Chem.*, 2011, **51**, 1007–1016.
- 154 P. Mineo, V. Villari, E. Scamporrino and N. Micali, *Soft Matter*, 2014, **10**, 44–47.
- 155 P. Mineo, V. Villari, E. Scamporrino and N. Micali, *J. Phys. Chem. B*, 2015, **119**, 12345–12353.
- 156 Y. Sang, D. Yang, P. Duan and M. Liu, *Chem. Sci.*, 2019, **10**, 2718–2724.
- 157 Z. Shen, Y. Sang, T. Wang, J. Jiang, Y. Meng, Y. Jiang, K. Okuro, T. Aida and M. Liu, *Nat. Commun.*, 2019, **10**, 1–8.
- 158 M. Kuroha, S. Nambu, S. Hattori, Y. Kitagawa, K. Niimura, Y. Mizuno, F. Hamba and K. Ishii, *Angew. Chem. Int. Ed.*, 2019, **58**, 18454–18459.
- 159 Y. Li, C. Liu, X. Bai, F. Tian, G. Hu and J. Sun, *Angew. Chem. Int. Ed.*, 2020, **59**, 3486–3490.
- 160 T. M. Hermans, K. J. M. Bishop, P. S. Stewart, S. H. Davis and B. A. Grzybowski, *Nat. Commun.*, 2015, **6**, 5640.
- 161 N. Micali, H. Engelkamp, P. G. van Rhee, P. C. M. Christianen, L. M. Scolaro and J. C. Maan, *Nat. Chem.*, 2012, **4**, 201–207.
- 162 Z. Martins, M. C. Price, N. Goldman, M. A. Sephton and M. J. Burchell, *Nat. Geosci.*, 2013, **6**, 1045–1049.
- 163 Y. Furukawa, H. Nakazawa, T. Sekine, T. Kobayashi and T. Kakegawa, *Earth Planet. Sci. Lett.*, 2015, **429**, 216–222.
- 164 Y. Furukawa, A. Takase, T. Sekine, T. Kakegawa and T. Kobayashi, *Orig. Life Evol. Biosph.*, 2018, **48**, 131–139.
- 165 G. G. Managadze, M. H. Engel, S. Getty, P. Wurz, W. B. Brinckerhoff, A. G. Shokolov, G. V. Sholin, S. A. Terent'ev, A. E. Chumikov, A. S. Skalkin, V. D. Blank, V. M. Prokhorov, N. G. Managadze and K. A. Luchnikov, *Planet. Space Sci.*, 2016, **131**, 70–78.
- 166 G. Managadze, *Planet. Space Sci.*, 2007, **55**, 134–140.
- 167 J. H. van 't Hoff, *Arch. Néerl. Sci. Exactes Nat.*, 1874, **9**, 445–454.
- 168 J. H. van 't Hoff, *Voorstel tot uitbreiding der tegenwoordig in de scheikunde gebruikte structuur-formules in de ruimte: benevens een daarmee samenhangende opmerking omtrent het verband tusschen optisch actief vermogen en chemische constitutie van organische verbindingen.*, Utrecht, J. Greven, 1874.
- 169 J. H. van 't Hoff, *Die Lagerung der Atome im Raume.*, Friedrich Vieweg und Sohn, Braunschweig, 2nd edn., 1894.
- 170 A. A. Cotton, *Comptes Rendus Hebd. Séances Académie Sci.*, 1895, **120**, 989–991.
- 171 A. A. Cotton, *Comptes Rendus Hebd. Séances Académie Sci.*, 1895, **120**, 1044–1046.
- 172 A. A. Cotton, *Ann. Chim. Phys.*, 1896, **7**, 347–432.
- 173 N. Berova, P. Polavarapu, K. Nakanishi and R. W. Woody, *Comprehensive Chiroptical Spectroscopy: Instrumentation, Methodologies, and Theoretical Simulations*, vol. 1, Wiley: Hoboken, NJ, USA, 2012.
- 174 N. Berova, P. Polavarapu, K. Nakanishi and R. W. Woody, Eds., *Comprehensive Chiroptical Spectroscopy: Applications in Stereochemical Analysis of Synthetic Compounds, Natural Products, and Biomolecules*, vol. 2, Wiley: Hoboken, NJ, USA, 2012.
- 175 W. Kuhn, *Trans. Faraday Soc.*, 1930, **26**, 293–308.
- 176 H. Rau, *Chem. Rev.*, 1983, **83**, 535–547.
- 177 Y. Inoue, *Chem. Rev.*, 1992, **92**, 741–770.
- 178 P. K. Hashim and N. Tamaoki, *ChemPhotoChem*, 2019, **3**, 347–355.
- 179 K. L. Stevenson and J. F. Verdick, *J. Am. Chem. Soc.*, 1968, **90**, 2974–2975.
- 180 B. L. Feringa, R. A. van Delden, N. Koumura and E. M. Geertsema, *Chem. Rev.*, 2000, **100**, 1789–1816.
- 181 W. R. Browne and B. L. Feringa, in *Molecular Switches*, 2nd Edition, W. R. Browne and B. L. Feringa, Eds., vol. 1, John Wiley & Sons, Ltd, 2011, pp. 121–179.
- 182 G. Yang, S. Zhang, J. Hu, M. Fujiki and G. Zou, *Symmetry*, 2019, **11**, 474–493.
- 183 J. Kim, J. Lee, W. Y. Kim, H. Kim, S. Lee, H. C. Lee, Y. S. Lee, M. Seo and S. Y. Kim, *Nat. Commun.*, 2015, **6**, 6959.
- 184 H. Kagan, A. Moradpour, J. F. Nicoud, G. Balavoine and G. Tsoucaris, *J. Am. Chem. Soc.*, 1971, **93**, 2353–2354.
- 185 W. J. Bernstein, M. Calvin and O. Buchardt, *J. Am. Chem. Soc.*, 1972, **94**, 494–498.
- 186 W. Kuhn and E. Braun, *Naturwissenschaften*, 1929, **17**, 227–228.
- 187 W. Kuhn and E. Knopf, *Naturwissenschaften*, 1930, **18**, 183–183.
- 188 C. Meinert, J. H. Bredehöft, J.-J. Filippi, Y. Baraud, L. Nahon, F. Wien, N. C. Jones, S. V. Hoffmann and U. J. Meierhenrich, *Angew. Chem. Int. Ed.*, 2012, **51**, 4484–4487.
- 189 G. Balavoine, A. Moradpour and H. B. Kagan, *J. Am. Chem. Soc.*, 1974, **96**, 5152–5158.
- 190 B. Norden, *Nature*, 1977, **266**, 567–568.
- 191 J. J. Flores, W. A. Bonner and G. A. Massey, *J. Am. Chem. Soc.*, 1977, **99**, 3622–3625.
- 192 I. Myrgorodska, C. Meinert, S. V. Hoffmann, N. C. Jones, L. Nahon and U. J. Meierhenrich, *ChemPlusChem*, 2017, **82**, 74–87.
- 193 H. Nishino, A. Kosaka, G. A. Hembury, H. Shitomi, H. Onuki and Y. Inoue, *Org. Lett.*, 2001, **3**, 921–924.
- 194 H. Nishino, A. Kosaka, G. A. Hembury, F. Aoki, K. Miyauchi, H. Shitomi, H. Onuki and Y. Inoue, *J. Am. Chem. Soc.*, 2002, **124**, 11618–11627.
- 195 U. J. Meierhenrich, J.-J. Filippi, C. Meinert, J. H. Bredehöft, J. Takahashi, L. Nahon, N. C. Jones and S. V. Hoffmann, *Angew. Chem. Int. Ed.*, 2010, **49**, 7799–7802.
- 196 U. J. Meierhenrich, L. Nahon, C. Alcaraz, J. H. Bredehöft, S. V. Hoffmann, B. Barbier and A. Brack, *Angew. Chem. Int. Ed.*, 2005, **44**, 5630–5634.
- 197 U. J. Meierhenrich, J.-J. Filippi, C. Meinert, S. V. Hoffmann, J. H. Bredehöft and L. Nahon, *Chem. Biodivers.*, 2010, **7**, 1651–1659.

- 198 C. Meinert, S. V. Hoffmann, P. Cassam-Chenaï, A. C. Evans, C. Giri, L. Nahon and U. J. Meierhenrich, *Angew. Chem. Int. Ed.*, 2014, **53**, 210–214.
- 199 C. Meinert, P. Cassam-Chenaï, N. C. Jones, L. Nahon, S. V. Hoffmann and U. J. Meierhenrich, *Orig. Life Evol. Biosph.*, 2015, **45**, 149–161.
- 200 M. Tia, B. Cunha de Miranda, S. Daly, F. Gaie-Levrel, G. A. Garcia, L. Nahon and I. Powis, *J. Phys. Chem. A*, 2014, **118**, 2765–2779.
- 201 B. A. McGuire, P. B. Carroll, R. A. Loomis, I. A. Finneran, P. R. Jewell, A. J. Remijan and G. A. Blake, *Science*, 2016, **352**, 1449–1452.
- 202 Y.-J. Kuan, S. B. Charnley, H.-C. Huang, W.-L. Tseng and Z. Kisiel, *Astrophys. J.*, 2003, **593**, 848.
- 203 Y. Takano, J. Takahashi, T. Kaneko, K. Marumo and K. Kobayashi, *Earth Planet. Sci. Lett.*, 2007, **254**, 106–114.
- 204 P. de Marcellus, C. Meinert, M. Nuevo, J.-J. Filippi, G. Danger, D. Deboffe, L. Nahon, L. Le Sergeant d'Hendecourt and U. J. Meierhenrich, *Astrophys. J.*, 2011, **727**, L27.
- 205 P. Modica, C. Meinert, P. de Marcellus, L. Nahon, U. J. Meierhenrich and L. L. S. d'Hendecourt, *Astrophys. J.*, 2014, **788**, 79.
- 206 C. Meinert and U. J. Meierhenrich, *Angew. Chem. Int. Ed.*, 2012, **51**, 10460–10470.
- 207 J. Takahashi and K. Kobayashi, *Symmetry*, 2019, **11**, 919.
- 208 A. G. W. Cameron and J. W. Truran, *Icarus*, 1977, **30**, 447–461.
- 209 P. Bargaño and R. Pérez de Tudela, *Orig. Life Evol. Biosph.*, 2007, **37**, 253–257.
- 210 P. Banerjee, Y.-Z. Qian, A. Heger and W. C. Haxton, *Nat. Commun.*, 2016, **7**, 13639.
- 211 T. L. V. Ulbricht, *Q. Rev. Chem. Soc.*, 1959, **13**, 48–60.
- 212 T. L. V. Ulbricht and F. Vester, *Tetrahedron*, 1962, **18**, 629–637.
- 213 A. S. Garay, *Nature*, 1968, **219**, 338–340.
- 214 A. S. Garay, L. Keszthelyi, I. Demeter and P. Hrasko, *Nature*, 1974, **250**, 332–333.
- 215 W. A. Bonner, M. a. V. Dort and M. R. Yearian, *Nature*, 1975, **258**, 419–421.
- 216 W. A. Bonner, M. A. van Dort, M. R. Yearian, H. D. Zeman and G. C. Li, *Isr. J. Chem.*, 1976, **15**, 89–95.
- 217 L. Keszthelyi, *Nature*, 1976, **264**, 197–197.
- 218 L. A. Hodge, F. B. Dunning, G. K. Walters, R. H. White and G. J. Schroepfer, *Nature*, 1979, **280**, 250–252.
- 219 M. Akaboshi, M. Noda, K. Kawai, H. Maki and K. Kawamoto, *Orig. Life Evol. Biosph.*, 1979, **9**, 181–186.
- 220 R. M. Lemmon, H. E. Conzett and W. A. Bonner, *Orig. Life Evol. Biosph.*, 1981, **11**, 337–341.
- 221 T. L. V. Ulbricht, *Nature*, 1975, **258**, 383–384.
- 222 W. A. Bonner, *Orig. Life Evol. Biosph.*, 1984, **14**, 383–390.
- 223 V. I. Burkov, L. A. Goncharova, G. A. Gusev, K. Kobayashi, E. V. Moiseenko, N. G. Poluhina, T. Saito, V. A. Tsarev, J. Xu and G. Zhang, *Orig. Life Evol. Biosph.*, 2008, **38**, 155–163.
- 224 G. A. Gusev, K. Kobayashi, E. V. Moiseenko, N. G. Poluhina, T. Saito, T. Ye, V. A. Tsarev, J. Xu, Y. Huang and G. Zhang, *Orig. Life Evol. Biosph.*, 2008, **38**, 509–515.
- 225 A. Dorta-Urra and P. Bargaño, *Symmetry*, 2019, **11**, 661.
- 226 M. A. Famiano, R. N. Boyd, T. Kajino and T. Onaka, *Astrobiology*, 2017, **18**, 190–206.
- 227 R. N. Boyd, M. A. Famiano, T. Onaka and T. Kajino, *Astrophys. J.*, 2018, **856**, 26.
- 228 M. A. Famiano, R. N. Boyd, T. Kajino, T. Onaka and Y. Mo, *Sci. Rep.*, 2018, **8**, 8833.
- 229 R. A. Rosenberg, D. Mishra and R. Naaman, *Angew. Chem. Int. Ed.*, 2015, **54**, 7295–7298.
- 230 F. Tassinari, J. Steidel, S. Paltiel, C. Fontanesi, M. Lahav, Y. Paltiel and R. Naaman, *Chem. Sci.*, 2019, **10**, 5246–5250.
- 231 R. A. Rosenberg, M. Abu Haija and P. J. Ryan, *Phys. Rev. Lett.*, 2008, **101**, 178301.
- 232 K. Michaeli, N. Kantor-Uriel, R. Naaman and D. H. Waldeck, *Chem Soc Rev*, 2016, **45**, 6478–6487.
- 233 R. Naaman, Y. Paltiel and D. H. Waldeck, *Acc. Chem. Res.*, 2020, **53**, 2659–2667.
- 234 R. Naaman, Y. Paltiel and D. H. Waldeck, *Nat. Rev. Chem.*, 2019, **3**, 250–260.
- 235 K. Banerjee-Ghosh, O. Ben Dor, F. Tassinari, E. Capua, S. Yochelis, A. Capua, S.-H. Yang, S. S. P. Parkin, S. Sarkar, L. Kronik, L. T. Baczewski, R. Naaman and Y. Paltiel, *Science*, 2018, **360**, 1331–1334.
- 236 T. S. Metzger, S. Mishra, B. P. Bloom, N. Goren, A. Neubauer, G. Shmul, J. Wei, S. Yochelis, F. Tassinari, C. Fontanesi, D. H. Waldeck, Y. Paltiel and R. Naaman, *Angew. Chem. Int. Ed.*, 2020, **59**, 1653–1658.
- 237 R. A. Rosenberg, *Symmetry*, 2019, **11**, 528.
- 238 A. Guijarro and M. Yus, in *The Origin of Chirality in the Molecules of Life*, 2008, RSC Publishing, pp. 125–137.
- 239 R. M. Hazen and D. S. Sholl, *Nat. Mater.*, 2003, **2**, 367–374.
- 240 I. Weissbuch and M. Lahav, *Chem. Rev.*, 2011, **111**, 3236–3267.
- 241 H. J. C. li, A. M. Scott, F. C. Hill, J. Leszczynski, N. Sahai and R. Hazen, *Chem. Soc. Rev.*, 2012, **41**, 5502–5525.
- 242 E. I. Klabunovskii, G. V. Smith and A. Zsigmond, *Heterogeneous Enantioselective Hydrogenation - Theory and Practice*, Springer, 2006.
- 243 V. Davankov, *Chirality*, 1998, **9**, 99–102.
- 244 F. Zaera, *Chem. Soc. Rev.*, 2017, **46**, 7374–7398.
- 245 W. A. Bonner, P. R. Kavasmaneck, F. S. Martin and J. J. Flores, *Science*, 1974, **186**, 143–144.
- 246 W. A. Bonner, P. R. Kavasmaneck, F. S. Martin and J. J. Flores, *Orig. Life Evol. Biosph.*, 1975, **6**, 367–376.
- 247 W. A. Bonner and P. R. Kavasmaneck, *J. Org. Chem.*, 1976, **41**, 2225–2226.
- 248 P. R. Kavasmaneck and W. A. Bonner, *J. Am. Chem. Soc.*, 1977, **99**, 44–50.
- 249 S. Furuyama, H. Kimura, M. Sawada and T. Morimoto, *Chem. Lett.*, 1978, **7**, 381–382.
- 250 S. Furuyama, M. Sawada, K. Hachiya and T. Morimoto, *Bull. Chem. Soc. Jpn.*, 1982, **55**, 3394–3397.
- 251 W. A. Bonner, *Orig. Life Evol. Biosph.*, 1995, **25**, 175–190.
- 252 R. T. Downs and R. M. Hazen, *J. Mol. Catal. Chem.*, 2004, **216**, 273–285.
- 253 J. W. Han and D. S. Sholl, *Langmuir*, 2009, **25**, 10737–10745.
- 254 J. W. Han and D. S. Sholl, *Phys. Chem. Chem. Phys.*, 2010, **12**, 8024–8032.
- 255 B. Kahr, B. Chittenden and A. Rohl, *Chirality*, 2006, **18**, 127–133.
- 256 A. J. Price and E. R. Johnson, *Phys. Chem. Chem. Phys.*, 2020, **22**, 16571–16578.
- 257 K. Evgenii and T. Wolfram, *Orig. Life Evol. Biosph.*, 2000, **30**, 431–434.
- 258 E. I. Klabunovskii, *Astrobiology*, 2001, **1**, 127–131.
- 259 E. A. Kulp and J. A. Switzer, *J. Am. Chem. Soc.*, 2007, **129**, 15120–15121.
- 260 W. Jiang, D. Athanasiadou, S. Zhang, R. Demichelis, K. B. Koziara, P. Raiteri, V. Nelea, W. Mi, J.-A. Ma, J. D. Gale and M. D. McKee, *Nat. Commun.*, 2019, **10**, 2318.
- 261 R. M. Hazen, T. R. Filley and G. A. Goodfriend, *Proc. Natl. Acad. Sci.*, 2001, **98**, 5487–5490.
- 262 A. Asthagiri and R. M. Hazen, *Mol. Simul.*, 2007, **33**, 343–351.
- 263 C. A. Orme, A. Noy, A. Wierzbicki, M. T. McBride, M. Grantham, H. H. Teng, P. M. Dove and J. J. DeYoreo, *Nature*, 2001, **411**, 775–779.

- 264 M. Maruyama, K. Tsukamoto, G. Sazaki, Y. Nishimura and P. G. Vekilov, *Cryst. Growth Des.*, 2009, **9**, 127–135.
- 265 A. M. Cody and R. D. Cody, *J. Cryst. Growth*, 1991, **113**, 508–519.
- 266 E. T. Degens, J. Matheja and T. A. Jackson, *Nature*, 1970, **227**, 492–493.
- 267 T. A. Jackson, *Experientia*, 1971, **27**, 242–243.
- 268 J. J. Flores and W. A. Bonner, *J. Mol. Evol.*, 1974, **3**, 49–56.
- 269 W. A. Bonner and J. Flores, *Biosystems*, 1973, **5**, 103–113.
- 270 J. J. McCullough and R. M. Lemmon, *J. Mol. Evol.*, 1974, **3**, 57–61.
- 271 S. C. Bondy and M. E. Harrington, *Science*, 1979, **203**, 1243–1244.
- 272 J. B. Youatt and R. D. Brown, *Science*, 1981, **212**, 1145–1146.
- 273 E. Friebele, A. Shimoyama, P. E. Hare and C. Ponnampertuma, *Orig. Life*, 1981, **11**, 173–184.
- 274 H. Hashizume, B. K. G. Theng and A. Yamagishi, *Clay Miner.*, 2002, **37**, 551–557.
- 275 T. Ikeda, H. Amoh and T. Yasunaga, *J. Am. Chem. Soc.*, 1984, **106**, 5772–5775.
- 276 B. Siffert and A. Naidja, *Clay Miner.*, 1992, **27**, 109–118.
- 277 D. G. Fraser, D. Fitz, T. Jakschitz and B. M. Rode, *Phys. Chem. Chem. Phys.*, 2010, **13**, 831–838.
- 278 D. G. Fraser, H. C. Greenwell, N. T. Skipper, M. V. Smalley, M. A. Wilkinson, B. Demé and R. K. Heenan, *Phys. Chem. Chem. Phys.*, 2010, **13**, 825–830.
- 279 S. I. Goldberg, *Orig. Life Evol. Biosph.*, 2008, **38**, 149–153.
- 280 G. Otis, M. Nassif, M. Zutta, A. Saady, S. Ruthstein and Y. Mastai, *Angew. Chem. Int. Ed.*, 2020, **59**, 20924–20929.
- 281 S. E. Wolf, N. Loges, B. Mathiasch, M. Panthöfer, I. Mey, A. Janshoff and W. Tremel, *Angew. Chem. Int. Ed.*, 2007, **46**, 5618–5623.
- 282 M. Lahav and L. Leiserowitz, *Angew. Chem. Int. Ed.*, 2008, **47**, 3680–3682.
- 283 W. Jiang, H. Pan, Z. Zhang, S. R. Qiu, J. D. Kim, X. Xu and R. Tang, *J. Am. Chem. Soc.*, 2017, **139**, 8562–8569.
- 284 O. Arteaga, A. Canillas, J. Crusats, Z. El-Hachemi, G. E. Jellison, J. Llorca and J. M. Ribó, *Orig. Life Evol. Biosph.*, 2010, **40**, 27–40.
- 285 S. Pizzarello, M. Zolensky and K. A. Turk, *Geochim. Cosmochim. Acta*, 2003, **67**, 1589–1595.
- 286 M. Frenkel and L. Heller-Kallai, *Chem. Geol.*, 1977, **19**, 161–166.
- 287 Y. Keheyhan and C. Montesano, *J. Anal. Appl. Pyrolysis*, 2010, **89**, 286–293.
- 288 J. P. Ferris, A. R. Hill, R. Liu and L. E. Orgel, *Nature*, 1996, **381**, 59–61.
- 289 I. Weissbuch, L. Addadi, Z. Berkovitch-Yellin, E. Gati, M. Lahav and L. Leiserowitz, *Nature*, 1984, **310**, 161–164.
- 290 I. Weissbuch, L. Addadi, L. Leiserowitz and M. Lahav, *J. Am. Chem. Soc.*, 1988, **110**, 561–567.
- 291 E. M. Landau, S. G. Wolf, M. Levanon, L. Leiserowitz, M. Lahav and J. Sagiv, *J. Am. Chem. Soc.*, 1989, **111**, 1436–1445.
- 292 T. Kawasaki, Y. Hakoda, H. Mineki, K. Suzuki and K. Soai, *J. Am. Chem. Soc.*, 2010, **132**, 2874–2875.
- 293 H. Mineki, Y. Kaimori, T. Kawasaki, A. Matsumoto and K. Soai, *Tetrahedron Asymmetry*, 2013, **24**, 1365–1367.
- 294 T. Kawasaki, S. Kamimura, A. Amihara, K. Suzuki and K. Soai, *Angew. Chem. Int. Ed.*, 2011, **50**, 6796–6798.
- 295 S. Miyagawa, K. Yoshimura, Y. Yamazaki, N. Takamatsu, T. Kuraishi, S. Aiba, Y. Tokunaga and T. Kawasaki, *Angew. Chem. Int. Ed.*, 2017, **56**, 1055–1058.
- 296 M. Forster and R. Raval, *Chem Commun*, 2016, **52**, 14075–14084.
- 297 C. Chen, S. Yang, G. Su, Q. Ji, M. Fuentes-Cabrera, S. Li and W. Liu, *J. Phys. Chem. C*, 2020, **124**, 742–748.
- 298 A. J. Gellman, Y. Huang, X. Feng, V. V. Pushkarev, B. Holsclaw and B. S. Mhatre, *J. Am. Chem. Soc.*, 2013, **135**, 19208–19214.
- 299 Y. Yun and A. J. Gellman, *Nat. Chem.*, 2015, **7**, 520–525.
- 300 A. J. Gellman and K.-H. Ernst, *Catal. Lett.*, 2018, **148**, 1610–1621.
- 301 J. M. Ribó and D. Hochberg, *Symmetry*, 2019, **11**, 814.
- 302 D. G. Blackmond, *Chem. Rev.*, 2020, **120**, 4831–4847.
- 303 F. C. Frank, *Biochim. Biophys. Acta*, 1953, **11**, 459–463.
- 304 D. K. Kondepudi and G. W. Nelson, *Phys. Rev. Lett.*, 1983, **50**, 1023–1026.
- 305 L. L. Morozov, V. V. Kuz Min and V. I. Goldanskii, *Orig. Life*, 1983, **13**, 119–138.
- 306 V. Avetisov and V. Goldanskii, *Proc. Natl. Acad. Sci.*, 1996, **93**, 11435–11442.
- 307 D. K. Kondepudi and K. Asakura, *Acc. Chem. Res.*, 2001, **34**, 946–954.
- 308 J. M. Ribó and D. Hochberg, *Phys. Chem. Chem. Phys.*, 2020, **22**, 14013–14025.
- 309 S. Bartlett and M. L. Wong, *Life*, 2020, **10**, 42.
- 310 K. Soai, T. Shibata, H. Morioka and K. Choji, *Nature*, 1995, **378**, 767–768.
- 311 T. Gehring, M. Busch, M. Schlageter and D. Weingand, *Chirality*, 2010, **22**, E173–E182.
- 312 K. Soai, T. Kawasaki and A. Matsumoto, *Symmetry*, 2019, **11**, 694.
- 313 T. Buhse, *Tetrahedron Asymmetry*, 2003, **14**, 1055–1061.
- 314 J. R. Islas, D. Lavabre, J.-M. Grevy, R. H. Lamonedá, H. R. Cabrera, J.-C. Micheau and T. Buhse, *Proc. Natl. Acad. Sci.*, 2005, **102**, 13743–13748.
- 315 O. Trapp, S. Lamour, F. Maier, A. F. Siegle, K. Zawatzky and B. F. Straub, *Chem. – Eur. J.*, 2020, **26**, 15871–15880.
- 316 I. D. Gridnev, J. M. Serafimov and J. M. Brown, *Angew. Chem. Int. Ed.*, 2004, **43**, 4884–4887.
- 317 I. D. Gridnev and A. Kh. Vorobiev, *ACS Catal.*, 2012, **2**, 2137–2149.
- 318 A. Matsumoto, T. Abe, A. Hara, T. Tobita, T. Sasagawa, T. Kawasaki and K. Soai, *Angew. Chem. Int. Ed.*, 2015, **54**, 15218–15221.
- 319 I. D. Gridnev and A. Kh. Vorobiev, *Bull. Chem. Soc. Jpn.*, 2015, **88**, 333–340.
- 320 A. Matsumoto, S. Fujiwara, T. Abe, A. Hara, T. Tobita, T. Sasagawa, T. Kawasaki and K. Soai, *Bull. Chem. Soc. Jpn.*, 2016, **89**, 1170–1177.
- 321 M. E. Noble-Terán, J.-M. Cruz, J.-C. Micheau and T. Buhse, *ChemCatChem*, 2018, **10**, 642–648.
- 322 S. V. Athavale, A. Simon, K. N. Houk and S. E. Denmark, *Nat. Chem.*, 2020, **12**, 412–423.
- 323 A. Matsumoto, A. Tanaka, Y. Kaimori, N. Hara, Y. Mikata and K. Soai, *Chem. Commun.*, 2021, **57**, 11209–11212.
- 324 Y. Geiger, *Chem. Soc. Rev.*, DOI:10.1039/D1CS01038G.
- 325 K. Soai, I. Sato, T. Shibata, S. Komiya, M. Hayashi, Y. Matsueda, H. Imamura, T. Hayase, H. Morioka, H. Tabira, J. Yamamoto and Y. Kowata, *Tetrahedron Asymmetry*, 2003, **14**, 185–188.
- 326 D. A. Singleton and L. K. Vo, *Org. Lett.*, 2003, **5**, 4337–4339.
- 327 I. D. Gridnev, J. M. Serafimov, H. Quiney and J. M. Brown, *Org. Biomol. Chem.*, 2003, **1**, 3811–3819.
- 328 T. Kawasaki, K. Suzuki, M. Shimizu, K. Ishikawa and K. Soai, *Chirality*, 2006, **18**, 479–482.
- 329 B. Barabas, L. Caglioti, C. Zucchi, M. Maioli, E. Gál, K. Micskei and G. Pályi, *J. Phys. Chem. B*, 2007, **111**, 11506–11510.
- 330 B. Barabás, C. Zucchi, M. Maioli, K. Micskei and G. Pályi, *J. Mol. Model.*, 2015, **21**, 33.
- 331 Y. Kaimori, Y. Hiyoshi, T. Kawasaki, A. Matsumoto and K. Soai, *Chem. Commun.*, 2019, **55**, 5223–5226.

- 332 A biased distribution of the product enantiomers has been observed for Soai (reference 332) and Soai related (reference 333) reactions as a probable consequence of the presence of cryptochiral species: D. A. Singleton and L. K. Vo, *J. Am. Chem. Soc.*, 2002, **124**, 10010–10011.
- 333 G. Rotunno, D. Petersen and M. Amedjkouh, *ChemSystemsChem*, 2020, **2**, e1900060.
- 334 M. Mauksch, S. B. Tsogoeva, I. M. Martynova and S. Wei, *Angew. Chem. Int. Ed.*, 2007, **46**, 393–396.
- 335 M. Amedjkouh and M. Brandberg, *Chem. Commun.*, 2008, 3043.
- 336 M. Mauksch, S. B. Tsogoeva, S. Wei and I. M. Martynova, *Chirality*, 2007, **19**, 816–825.
- 337 M. P. Romero-Fernández, R. Babiano and P. Cintas, *Chirality*, 2018, **30**, 445–456.
- 338 S. B. Tsogoeva, S. Wei, M. Freund and M. Mauksch, *Angew. Chem. Int. Ed.*, 2009, **48**, 590–594.
- 339 S. B. Tsogoeva, *Chem. Commun.*, 2010, **46**, 7662–7669.
- 340 X. Wang, Y. Zhang, H. Tan, Y. Wang, P. Han and D. Z. Wang, *J. Org. Chem.*, 2010, **75**, 2403–2406.
- 341 M. Mauksch, S. Wei, M. Freund, A. Zamfir and S. B. Tsogoeva, *Orig. Life Evol. Biosph.*, 2009, **40**, 79–91.
- 342 G. Valero, J. M. Ribó and A. Moyano, *Chem. – Eur. J.*, 2014, **20**, 17395–17408.
- 343 R. Plasson, H. Bersini and A. Commeyras, *Proc. Natl. Acad. Sci. U. S. A.*, 2004, **101**, 16733–16738.
- 344 Y. Saito and H. Hyuga, *J. Phys. Soc. Jpn.*, 2004, **73**, 33–35.
- 345 F. Jafarpour, T. Biancalani and N. Goldenfeld, *Phys. Rev. Lett.*, 2015, **115**, 158101.
- 346 K. Iwamoto, *Phys. Chem. Chem. Phys.*, 2002, **4**, 3975–3979.
- 347 J. M. Ribó, J. Crusats, Z. El-Hachemi, A. Moyano, C. Blanco and D. Hochberg, *Astrobiology*, 2013, **13**, 132–142.
- 348 C. Blanco, J. M. Ribó, J. Crusats, Z. El-Hachemi, A. Moyano and D. Hochberg, *Phys. Chem. Chem. Phys.*, 2013, **15**, 1546–1556.
- 349 C. Blanco, J. Crusats, Z. El-Hachemi, A. Moyano, D. Hochberg and J. M. Ribó, *ChemPhysChem*, 2013, **14**, 2432–2440.
- 350 J. M. Ribó, J. Crusats, Z. El-Hachemi, A. Moyano and D. Hochberg, *Chem. Sci.*, 2017, **8**, 763–769.
- 351 M. Eigen and P. Schuster, *The Hypercycle: A Principle of Natural Self-Organization*, Springer-Verlag, Berlin Heidelberg, 1979.
- 352 F. Ricci, F. H. Stillinger and P. G. Debenedetti, *J. Phys. Chem. B*, 2013, **117**, 602–614.
- 353 Y. Sang and M. Liu, *Symmetry*, 2019, **11**, 950–969.
- 354 A. Arlegui, B. Soler, A. Galindo, O. Arteaga, A. Canillas, J. M. Ribó, Z. El-Hachemi, J. Crusats and A. Moyano, *Chem. Commun.*, 2019, **55**, 12219–12222.
- 355 E. Havinga, *Biochim. Biophys. Acta*, 1954, **13**, 171–174.
- 356 A. C. D. Newman and H. M. Powell, *J. Chem. Soc. Resumed*, 1952, 3747–3751.
- 357 R. E. Pincock and K. R. Wilson, *J. Am. Chem. Soc.*, 1971, **93**, 1291–1292.
- 358 R. E. Pincock, R. R. Perkins, A. S. Ma and K. R. Wilson, *Science*, 1971, **174**, 1018–1020.
- 359 W. H. Zachariasen, *Z. Für Krist. - Cryst. Mater.*, 1929, **71**, 517–529.
- 360 G. N. Ramachandran and K. S. Chandrasekaran, *Acta Crystallogr.*, 1957, **10**, 671–675.
- 361 S. C. Abrahams and J. L. Bernstein, *Acta Crystallogr. B*, 1977, **33**, 3601–3604.
- 362 F. S. Kipping and W. J. Pope, *J. Chem. Soc. Trans.*, 1898, **73**, 606–617.
- 363 F. S. Kipping and W. J. Pope, *Nature*, 1898, **59**, 53–53.
- 364 J.-M. Cruz, K. Hernández-Lechuga, I. Domínguez-Valle, A. Fuentes-Beltrán, J. U. Sánchez-Morales, J. L. Ocampo-Espindola, C. Polanco, J.-C. Micheau and T. Buhse, *Chirality*, 2020, **32**, 120–134.
- 365 D. K. Kondepudi, R. J. Kaufman and N. Singh, *Science*, 1990, **250**, 975–976.
- 366 J. M. McBride and R. L. Carter, *Angew. Chem. Int. Ed. Engl.*, 1991, **30**, 293–295.
- 367 D. K. Kondepudi, K. L. Bullock, J. A. Digits, J. K. Hall and J. M. Miller, *J. Am. Chem. Soc.*, 1993, **115**, 10211–10216.
- 368 B. Martin, A. Tharrington and X. Wu, *Phys. Rev. Lett.*, 1996, **77**, 2826–2829.
- 369 Z. El-Hachemi, J. Crusats, J. M. Ribó and S. Veintemillas-Verdaguer, *Cryst. Growth Des.*, 2009, **9**, 4802–4806.
- 370 D. J. Durand, D. K. Kondepudi, P. F. Moreira Jr. and F. H. Quina, *Chirality*, 2002, **14**, 284–287.
- 371 D. K. Kondepudi, J. Laudadio and K. Asakura, *J. Am. Chem. Soc.*, 1999, **121**, 1448–1451.
- 372 W. L. Noorduin, T. Izumi, A. Millemaggi, M. Leeman, H. Meekes, W. J. P. Van Enckevort, R. M. Kellogg, B. Kaptein, E. Vlieg and D. G. Blackmond, *J. Am. Chem. Soc.*, 2008, **130**, 1158–1159.
- 373 C. Viedma, *Phys. Rev. Lett.*, 2005, **94**, 065504.
- 374 C. Viedma, *Cryst. Growth Des.*, 2007, **7**, 553–556.
- 375 C. Xiouras, J. Van Aeken, J. Panis, J. H. Ter Horst, T. Van Gerven and G. D. Stefanidis, *Cryst. Growth Des.*, 2015, **15**, 5476–5484.
- 376 J. Ahn, D. H. Kim, G. Coquerel and W.-S. Kim, *Cryst. Growth Des.*, 2018, **18**, 297–306.
- 377 C. Viedma and P. Cintas, *Chem. Commun.*, 2011, **47**, 12786–12788.
- 378 W. L. Noorduin, W. J. P. van Enckevort, H. Meekes, B. Kaptein, R. M. Kellogg, J. C. Tully, J. M. McBride and E. Vlieg, *Angew. Chem. Int. Ed.*, 2010, **49**, 8435–8438.
- 379 R. R. E. Steendam, T. J. B. van Benthem, E. M. E. Huijs, H. Meekes, W. J. P. van Enckevort, J. Raap, F. P. J. T. Rutjes and E. Vlieg, *Cryst. Growth Des.*, 2015, **15**, 3917–3921.
- 380 C. Blanco, J. Crusats, Z. El-Hachemi, A. Moyano, S. Veintemillas-Verdaguer, D. Hochberg and J. M. Ribó, *ChemPhysChem*, 2013, **14**, 3982–3993.
- 381 C. Blanco, J. M. Ribó and D. Hochberg, *Phys. Rev. E*, 2015, **91**, 022801.
- 382 G. An, P. Yan, J. Sun, Y. Li, X. Yao and G. Li, *CrystEngComm*, 2015, **17**, 4421–4433.
- 383 R. R. E. Steendam, J. M. M. Verkade, T. J. B. van Benthem, H. Meekes, W. J. P. van Enckevort, J. Raap, F. P. J. T. Rutjes and E. Vlieg, *Nat. Commun.*, 2014, **5**, 5543.
- 384 A. H. Engwerda, H. Meekes, B. Kaptein, F. Rutjes and E. Vlieg, *Chem. Commun.*, 2016, **52**, 12048–12051.
- 385 C. Viedma, C. Lennox, L. A. Cuccia, P. Cintas and J. E. Ortiz, *Chem. Commun.*, 2020, **56**, 4547–4550.
- 386 C. Viedma, J. E. Ortiz, T. de Torres, T. Izumi and D. G. Blackmond, *J. Am. Chem. Soc.*, 2008, **130**, 15274–15275.
- 387 L. Spix, H. Meekes, R. H. Blaauw, W. J. P. van Enckevort and E. Vlieg, *Cryst. Growth Des.*, 2012, **12**, 5796–5799.
- 388 F. Cameli, C. Xiouras and G. D. Stefanidis, *CrystEngComm*, 2018, **20**, 2897–2901.
- 389 K. Ishikawa, M. Tanaka, T. Suzuki, A. Sekine, T. Kawasaki, K. Soai, M. Shiro, M. Lahav and T. Asahi, *Chem. Commun.*, 2012, **48**, 6031–6033.
- 390 A. V. Tarasevych, A. E. Sorochinsky, V. P. Kukhar, L. Toupet, J. Crassous and J.-C. Guillemin, *CrystEngComm*, 2015, **17**, 1513–1517.
- 391 L. Spix, A. Alfring, H. Meekes, W. J. P. van Enckevort and E. Vlieg, *Cryst. Growth Des.*, 2014, **14**, 1744–1748.
- 392 L. Spix, A. H. J. Engwerda, H. Meekes, W. J. P. van Enckevort and E. Vlieg, *Cryst. Growth Des.*, 2016, **16**, 4752–4758.
- 393 B. Kaptein, W. L. Noorduin, H. Meekes, W. J. P. van Enckevort, R. M. Kellogg and E. Vlieg, *Angew. Chem. Int. Ed.*, 2008, **47**, 7226–7229.

- 394 T. Kawasaki, N. Takamatsu, S. Aiba and Y. Tokunaga, *Chem. Commun.*, 2015, **51**, 14377–14380.
- 395 S. Aiba, N. Takamatsu, T. Sasai, Y. Tokunaga and T. Kawasaki, *Chem. Commun.*, 2016, **52**, 10834–10837.
- 396 S. Miyagawa, S. Aiba, H. Kawamoto, Y. Tokunaga and T. Kawasaki, *Org. Biomol. Chem.*, 2019, **17**, 1238–1244.
- 397 I. Baglai, M. Leeman, K. Wurst, B. Kaptein, R. M. Kellogg and W. L. Noorduin, *Chem. Commun.*, 2018, **54**, 10832–10834.
- 398 N. Uemura, K. Sano, A. Matsumoto, Y. Yoshida, T. Mino and M. Sakamoto, *Chem. – Asian J.*, 2019, **14**, 4150–4153.
- 399 N. Uemura, M. Hosaka, A. Washio, Y. Yoshida, T. Mino and M. Sakamoto, *Cryst. Growth Des.*, 2020, **20**, 4898–4903.
- 400 D. K. Kondepudi and G. W. Nelson, *Phys. Lett. A*, 1984, **106**, 203–206.
- 401 D. K. Kondepudi and G. W. Nelson, *Phys. Stat. Mech. Its Appl.*, 1984, **125**, 465–496.
- 402 D. K. Kondepudi and G. W. Nelson, *Nature*, 1985, **314**, 438–441.
- 403 N. A. Hawbaker and D. G. Blackmond, *Nat. Chem.*, 2019, **11**, 957–962.
- 404 J. I. Murray, J. N. Sanders, P. F. Richardson, K. N. Houk and D. G. Blackmond, *J. Am. Chem. Soc.*, 2020, **142**, 3873–3879.
- 405 S. Mahurin, M. McGinnis, J. S. Bogard, L. D. Hulett, R. M. Pagni and R. N. Compton, *Chirality*, 2001, **13**, 636–640.
- 406 K. Soai, S. Osanai, K. Kadowaki, S. Yonekubo, T. Shibata and I. Sato, *J. Am. Chem. Soc.*, 1999, **121**, 11235–11236.
- 407 A. Matsumoto, H. Ozaki, S. Tsuchiya, T. Asahi, M. Lahav, T. Kawasaki and K. Soai, *Org. Biomol. Chem.*, 2019, **17**, 4200–4203.
- 408 D. J. Carter, A. L. Rohl, A. Shtukenberg, S. Bian, C. Hu, L. Baylon, B. Kahr, H. Mineki, K. Abe, T. Kawasaki and K. Soai, *Cryst. Growth Des.*, 2012, **12**, 2138–2145.
- 409 H. Shindo, Y. Shirota, K. Niki, T. Kawasaki, K. Suzuki, Y. Araki, A. Matsumoto and K. Soai, *Angew. Chem. Int. Ed.*, 2013, **52**, 9135–9138.
- 410 T. Kawasaki, Y. Kaimori, S. Shimada, N. Hara, S. Sato, K. Suzuki, T. Asahi, A. Matsumoto and K. Soai, *Chem. Commun.*, 2021, **57**, 5999–6002.
- 411 A. Matsumoto, Y. Kaimori, M. Uchida, H. Omori, T. Kawasaki and K. Soai, *Angew. Chem. Int. Ed.*, 2017, **56**, 545–548.
- 412 L. Addadi, Z. Berkovitch-Yellin, N. Domb, E. Gati, M. Lahav and L. Leiserowitz, *Nature*, 1982, **296**, 21–26.
- 413 L. Addadi, S. Weinstein, E. Gati, I. Weissbuch and M. Lahav, *J. Am. Chem. Soc.*, 1982, **104**, 4610–4617.
- 414 I. Baglai, M. Leeman, K. Wurst, R. M. Kellogg and W. L. Noorduin, *Angew. Chem. Int. Ed.*, 2020, **59**, 20885–20889.
- 415 J. Royes, V. Polo, S. Uriel, L. Oriol, M. Piñol and R. M. Tejedor, *Phys. Chem. Chem. Phys.*, 2017, **19**, 13622–13628.
- 416 E. E. Greciano, R. Rodríguez, K. Maeda and L. Sánchez, *Chem. Commun.*, 2020, **56**, 2244–2247.
- 417 I. Sato, R. Sugie, Y. Matsueda, Y. Furumura and K. Soai, *Angew. Chem. Int. Ed.*, 2004, **43**, 4490–4492.
- 418 T. Kawasaki, M. Sato, S. Ishiguro, T. Saito, Y. Morishita, I. Sato, H. Nishino, Y. Inoue and K. Soai, *J. Am. Chem. Soc.*, 2005, **127**, 3274–3275.
- 419 W. L. Noorduin, A. A. C. Bode, M. van der Meijden, H. Meeke, A. F. van Etteger, W. J. P. van Enckevort, P. C. M. Christianen, B. Kaptein, R. M. Kellogg, T. Rasing and E. Vlieg, *Nat. Chem.*, 2009, **1**, 729.
- 420 W. H. Mills, *J. Soc. Chem. Ind.*, 1932, **51**, 750–759.
- 421 M. Bolli, R. Micura and A. Eschenmoser, *Chem. Biol.*, 1997, **4**, 309–320.
- 422 A. Brewer and A. P. Davis, *Nat. Chem.*, 2014, **6**, 569–574.
- 423 A. R. A. Palmans, J. A. J. M. Vekemans, E. E. Havinga and E. W. Meijer, *Angew. Chem. Int. Ed. Engl.*, 1997, **36**, 2648–2651.
- 424 F. H. C. Crick and L. E. Orgel, *Icarus*, 1973, **19**, 341–346.
- 425 A. J. MacDermott and G. E. Tranter, *Croat. Chem. Acta*, 1989, **62**, 165–187.
- 426 A. Julg, *J. Mol. Struct. THEOCHEM*, 1989, **184**, 131–142.
- 427 W. Martin, J. Baross, D. Kelley and M. J. Russell, *Nat. Rev. Microbiol.*, 2008, **6**, 805–814.
- 428 W. Wang, *arXiv:2001.03532*.
- 429 V. I. Goldanskii and V. V. Kuz'min, *AIP Conf. Proc.*, 1988, **180**, 163–228.
- 430 W. A. Bonner and E. Rubenstein, *Biosystems*, 1987, **20**, 99–111.
- 431 A. Jorissen and C. Cerf, *Orig. Life Evol. Biosph.*, 2002, **32**, 129–142.
- 432 K. Mislow, *Collect. Czechoslov. Chem. Commun.*, 2003, **68**, 849–864.
- 433 A. Burton and E. Berger, *Life*, 2018, **8**, 14.
- 434 A. Garcia, C. Meinert, H. Sugahara, N. Jones, S. Hoffmann and U. Meierhenrich, *Life*, 2019, **9**, 29.
- 435 G. Cooper, N. Kimmich, W. Belisle, J. Sarinana, K. Brabham and L. Garrel, *Nature*, 2001, **414**, 879–883.
- 436 Y. Furukawa, Y. Chikaraishi, N. Ohkouchi, N. O. Ogawa, D. P. Glavin, J. P. Dworkin, C. Abe and T. Nakamura, *Proc. Natl. Acad. Sci.*, 2019, **116**, 24440–24445.
- 437 J. Bocková, N. C. Jones, U. J. Meierhenrich, S. V. Hoffmann and C. Meinert, *Commun. Chem.*, 2021, **4**, 86.
- 438 J. R. Cronin and S. Pizzarello, *Adv. Space Res.*, 1999, **23**, 293–299.
- 439 S. Pizzarello and J. R. Cronin, *Geochim. Cosmochim. Acta*, 2000, **64**, 329–338.
- 440 D. P. Glavin and J. P. Dworkin, *Proc. Natl. Acad. Sci.*, 2009, **106**, 5487–5492.
- 441 I. Myrgorodska, C. Meinert, Z. Martins, L. le Sergeant d'Hendecourt and U. J. Meierhenrich, *J. Chromatogr. A*, 2016, **1433**, 131–136.
- 442 G. Cooper and A. C. Rios, *Proc. Natl. Acad. Sci.*, 2016, **113**, E3322–E3331.
- 443 A. Furusho, T. Akita, M. Mita, H. Naraoka and K. Hamase, *J. Chromatogr. A*, 2020, **1625**, 461255.
- 444 M. P. Bernstein, J. P. Dworkin, S. A. Sandford, G. W. Cooper and L. J. Allamandola, *Nature*, 2002, **416**, 401–403.
- 445 K. M. Ferrière, *Rev. Mod. Phys.*, 2001, **73**, 1031–1066.
- 446 Y. Fukui and A. Kawamura, *Annu. Rev. Astron. Astrophys.*, 2010, **48**, 547–580.
- 447 E. L. Gibb, D. C. B. Whittet, A. C. A. Boogert and A. G. G. M. Tielens, *Astrophys. J. Suppl. Ser.*, 2004, **151**, 35–73.
- 448 J. Mayo Greenberg, *Surf. Sci.*, 2002, **500**, 793–822.
- 449 S. A. Sandford, M. Nuevo, P. P. Bera and T. J. Lee, *Chem. Rev.*, 2020, **120**, 4616–4659.
- 450 J. J. Hester, S. J. Desch, K. R. Healy and L. A. Leshin, *Science*, 2004, **304**, 1116–1117.
- 451 G. M. Muñoz Caro, U. J. Meierhenrich, W. A. Schutte, B. Barbier, A. Arcones Segovia, H. Rosenbauer, W. H.-P. Thiemann, A. Brack and J. M. Greenberg, *Nature*, 2002, **416**, 403–406.
- 452 M. Nuevo, G. Auger, D. Blanot and L. d'Hendecourt, *Orig. Life Evol. Biosph.*, 2008, **38**, 37–56.
- 453 C. Zhu, A. M. Turner, C. Meinert and R. I. Kaiser, *Astrophys. J.*, 2020, **889**, 134.
- 454 C. Meinert, I. Myrgorodska, P. de Marcellus, T. Buhse, L. Nahon, S. V. Hoffmann, L. L. S. d'Hendecourt and U. J. Meierhenrich, *Science*, 2016, **352**, 208–212.
- 455 M. Nuevo, G. Cooper and S. A. Sandford, *Nat. Commun.*, 2018, **9**, 5276.
- 456 Y. Oba, Y. Takano, H. Naraoka, N. Watanabe and A. Kouchi, *Nat. Commun.*, 2019, **10**, 4413.
- 457 J. Kwon, M. Tamura, P. W. Lucas, J. Hashimoto, N. Kusakabe, R. Kandori, Y. Nakajima, T. Nagayama, T. Nagata and J. H. Hough, *Astrophys. J.*, 2013, **765**, L6.
- 458 J. Bailey, *Orig. Life Evol. Biosph.*, 2001, **31**, 167–183.
- 459 J. Bailey, A. Chrysostomou, J. H. Hough, T. M. Gledhill, A. McCall, S. Clark, F. Ménard and M. Tamura, *Science*, 1998, **281**, 672–674.

- 460 T. Fukue, M. Tamura, R. Kandori, N. Kusakabe, J. H. Hough, J. Bailey, D. C. B. Whittet, P. W. Lucas, Y. Nakajima and J. Hashimoto, *Orig. Life Evol. Biosph.*, 2010, **40**, 335–346.
- 461 J. Kwon, M. Tamura, J. H. Hough, N. Kusakabe, T. Nagata, Y. Nakajima, P. W. Lucas, T. Nagayama and R. Kandori, *Astrophys. J.*, 2014, **795**, L16.
- 462 J. Kwon, M. Tamura, J. H. Hough, T. Nagata, N. Kusakabe and H. Saito, *Astrophys. J.*, 2016, **824**, 95.
- 463 J. Kwon, M. Tamura, J. H. Hough, T. Nagata and N. Kusakabe, *Astron. J.*, 2016, **152**, 67.
- 464 J. Kwon, T. Nakagawa, M. Tamura, J. H. Hough, R. Kandori, M. Choi, M. Kang, J. Cho, Y. Nakajima and T. Nagata, *Astron. J.*, 2018, **156**, 1.
- 465 K. Wood, *Astrophys. J.*, 1997, **477**, L25–L28.
- 466 S. F. Mason, *Nature*, 1997, **389**, 804.
- 467 W. A. Bonner, E. Rubenstein and G. S. Brown, *Orig. Life Evol. Biosph.*, 1999, **29**, 329–332.
- 468 J. H. Bredehöft, N. C. Jones, C. Meinert, A. C. Evans, S. V. Hoffmann and U. J. Meierhenrich, *Chirality*, 2014, **26**, 373–378.
- 469 E. Rubenstein, W. A. Bonner, H. P. Noyes and G. S. Brown, *Nature*, 1983, **306**, 118–118.
- 470 M. Buschermohle, D. C. B. Whittet, A. Chrysostomou, J. H. Hough, P. W. Lucas, A. J. Adamson, B. A. Whitney and M. J. Wolff, *Astrophys. J.*, 2005, **624**, 821–826.
- 471 J. Oró, T. Mills and A. Lazcano, *Orig. Life Evol. Biosph.*, 1991, **21**, 267–277.
- 472 A. G. Griesbeck and U. J. Meierhenrich, *Angew. Chem. Int. Ed.*, 2002, **41**, 3147–3154.
- 473 C. Meinert, J.-J. Filippi, L. Nahon, S. V. Hoffmann, L. D’Hendecourt, P. De Marcellus, J. H. Bredehöft, W. H.-P. Thiemann and U. J. Meierhenrich, *Symmetry*, 2010, **2**, 1055–1080.
- 474 I. Myrgorodska, C. Meinert, Z. Martins, L. L. S. d’Hendecourt and U. J. Meierhenrich, *Angew. Chem. Int. Ed.*, 2015, **54**, 1402–1412.
- 475 I. Baglai, M. Leeman, B. Kaptein, R. M. Kellogg and W. L. Noorduin, *Chem. Commun.*, 2019, **55**, 6910–6913.
- 476 A. G. Lyne, *Nature*, 1984, **308**, 605–606.
- 477 W. A. Bonner and R. M. Lemmon, *J. Mol. Evol.*, 1978, **11**, 95–99.
- 478 W. A. Bonner and R. M. Lemmon, *Bioorganic Chem.*, 1978, **7**, 175–187.
- 479 M. Preiner, S. Asche, S. Becker, H. C. Betts, A. Boniface, E. Camprubi, K. Chandru, V. Erastova, S. G. Garg, N. Khawaja, G. Kostyrka, R. Machné, G. Moggioli, K. B. Muchowska, S. Neukirchen, B. Peter, E. Pichlhöfer, Á. Radványi, D. Rossetto, A. Salditt, N. M. Schmelling, F. L. Sousa, F. D. K. Tria, D. Vörös and J. C. Xavier, *Life*, 2020, **10**, 20.
- 480 K. Michaelian, *Life*, 2018, **8**, 21.
- 481 NASA Astrobiology, <https://astrobiology.nasa.gov/research/life-detection/about/>, (accessed Decembre 22th, 2021).
- 482 S. A. Benner, E. A. Bell, E. Biondi, R. Brassler, T. Carell, H.-J. Kim, S. J. Mojzsis, A. Omran, M. A. Pasek and D. Trail, *ChemSystemsChem*, 2020, **2**, e1900035.
- 483 M. Idelson and E. R. Blout, *J. Am. Chem. Soc.*, 1958, **80**, 2387–2393.
- 484 G. F. Joyce, G. M. Visser, C. A. A. van Boeckel, J. H. van Boom, L. E. Orgel and J. van Westrenen, *Nature*, 1984, **310**, 602–604.
- 485 R. D. Lundberg and P. Doty, *J. Am. Chem. Soc.*, 1957, **79**, 3961–3972.
- 486 E. R. Blout, P. Doty and J. T. Yang, *J. Am. Chem. Soc.*, 1957, **79**, 749–750.
- 487 J. G. Schmidt, P. E. Nielsen and L. E. Orgel, *J. Am. Chem. Soc.*, 1997, **119**, 1494–1495.
- 488 M. M. Waldrop, *Science*, 1990, **250**, 1078–1080.
- 489 E. G. Nisbet and N. H. Sleep, *Nature*, 2001, **409**, 1083–1091.
- 490 V. R. Oberbeck and G. Fogleman, *Orig. Life Evol. Biosph.*, 1989, **19**, 549–560.
- 491 A. Lazcano and S. L. Miller, *J. Mol. Evol.*, 1994, **39**, 546–554.
- 492 J. L. Bada, in *Chemistry and Biochemistry of the Amino Acids*, ed. G. C. Barrett, Springer Netherlands, Dordrecht, 1985, pp. 399–414.
- 493 S. Kempe and J. Kazmierczak, *Astrobiology*, 2002, **2**, 123–130.
- 494 J. L. Bada, *Chem. Soc. Rev.*, 2013, **42**, 2186–2196.
- 495 H. J. Morowitz, *J. Theor. Biol.*, 1969, **25**, 491–494.
- 496 M. Klussmann, A. J. P. White, A. Armstrong and D. G. Blackmond, *Angew. Chem. Int. Ed.*, 2006, **45**, 7985–7989.
- 497 M. Klussmann, H. Iwamura, S. P. Mathew, D. H. Wells, U. Pandya, A. Armstrong and D. G. Blackmond, *Nature*, 2006, **441**, 621–623.
- 498 R. Breslow and M. S. Levine, *Proc. Natl. Acad. Sci.*, 2006, **103**, 12979–12980.
- 499 M. Levine, C. S. Kenesky, D. Mazori and R. Breslow, *Org. Lett.*, 2008, **10**, 2433–2436.
- 500 M. Klussmann, T. Izumi, A. J. P. White, A. Armstrong and D. G. Blackmond, *J. Am. Chem. Soc.*, 2007, **129**, 7657–7660.
- 501 R. Breslow and Z.-L. Cheng, *Proc. Natl. Acad. Sci.*, 2009, **106**, 9144–9146.
- 502 J. Han, A. Wzorek, M. Kwiatkowska, V. A. Soloshonok and K. D. Klika, *Amino Acids*, 2019, **51**, 865–889.
- 503 R. H. Perry, C. Wu, M. Neffliu and R. Graham Cooks, *Chem. Commun.*, 2007, 1071–1073.
- 504 S. P. Fletcher, R. B. C. Jagt and B. L. Feringa, *Chem. Commun.*, 2007, 2578–2580.
- 505 A. Bellec and J.-C. Guillemin, *Chem. Commun.*, 2010, **46**, 1482–1484.
- 506 A. V. Tarasevych, A. E. Sorochinsky, V. P. Kukhar, A. Chollet, R. Daniellou and J.-C. Guillemin, *J. Org. Chem.*, 2013, **78**, 10530–10533.
- 507 A. V. Tarasevych, A. E. Sorochinsky, V. P. Kukhar and J.-C. Guillemin, *Orig. Life Evol. Biosph.*, 2013, **43**, 129–135.
- 508 V. Dašková, J. Buter, A. K. Schoonen, M. Lutz, F. de Vries and B. L. Feringa, *Angew. Chem. Int. Ed.*, 2021, **60**, 11120–11126.
- 509 A. Córdova, M. Engqvist, I. Ibrahim, J. Casas and H. Sundén, *Chem. Commun.*, 2005, 2047–2049.
- 510 R. Breslow and Z.-L. Cheng, *Proc. Natl. Acad. Sci.*, 2010, **107**, 5723–5725.
- 511 S. Pizzarello and A. L. Weber, *Science*, 2004, **303**, 1151.
- 512 A. L. Weber and S. Pizzarello, *Proc. Natl. Acad. Sci.*, 2006, **103**, 12713–12717.
- 513 S. Pizzarello and A. L. Weber, *Orig. Life Evol. Biosph.*, 2009, **40**, 3–10.
- 514 J. E. Hein, E. Tse and D. G. Blackmond, *Nat. Chem.*, 2011, **3**, 704–706.
- 515 A. J. Wagner, D. Yu. Zubarev, A. Aspuru-Guzik and D. G. Blackmond, *ACS Cent. Sci.*, 2017, **3**, 322–328.
- 516 M. P. Robertson and G. F. Joyce, *Cold Spring Harb. Perspect. Biol.*, 2012, **4**, a003608.
- 517 A. Kahana, P. Schmitt-Kopplin and D. Lancet, *Astrobiology*, 2019, **19**, 1263–1278.
- 518 F. J. Dyson, *J. Mol. Evol.*, 1982, **18**, 344–350.
- 519 R. Root-Bernstein, *BioEssays*, 2007, **29**, 689–698.
- 520 K. A. Lanier, A. S. Petrov and L. D. Williams, *J. Mol. Evol.*, 2017, **85**, 8–13.
- 521 H. S. Martin, K. A. Podolsky and N. K. Devaraj, *ChemBioChem*, 2021, **22**, 3148–3157.
- 522 V. Sojo, *BioEssays*, 2019, **41**, 1800251.
- 523 A. Eschenmoser, *Tetrahedron*, 2007, **63**, 12821–12844.

- 524 S. N. Semenov, L. J. Kraft, A. Ainla, M. Zhao, M. Baghbanzadeh, V. E. Campbell, K. Kang, J. M. Fox and G. M. Whitesides, *Nature*, 2016, **537**, 656–660.
- 525 G. Ashkenasy, T. M. Hermans, S. Otto and A. F. Taylor, *Chem. Soc. Rev.*, 2017, **46**, 2543–2554.
- 526 K. Satoh and M. Kamigaito, *Chem. Rev.*, 2009, **109**, 5120–5156.
- 527 C. M. Thomas, *Chem. Soc. Rev.*, 2009, **39**, 165–173.
- 528 A. Brack and G. Spach, *Nat. Phys. Sci.*, 1971, **229**, 124–125.
- 529 S. I. Goldberg, J. M. Crosby, N. D. Iusem and U. E. Younes, *J. Am. Chem. Soc.*, 1987, **109**, 823–830.
- 530 T. H. Hitz and P. L. Luisi, *Orig. Life Evol. Biosph.*, 2004, **34**, 93–110.
- 531 T. Hitz, M. Blocher, P. Walde and P. L. Luisi, *Macromolecules*, 2001, **34**, 2443–2449.
- 532 T. Hitz and P. L. Luisi, *Helv. Chim. Acta*, 2002, **85**, 3975–3983.
- 533 T. Hitz and P. L. Luisi, *Helv. Chim. Acta*, 2003, **86**, 1423–1434.
- 534 H. Urata, C. Aono, N. Ohmoto, Y. Shimamoto, Y. Kobayashi and M. Akagi, *Chem. Lett.*, 2001, **30**, 324–325.
- 535 K. Osawa, H. Urata and H. Sawai, *Orig. Life Evol. Biosph.*, 2005, **35**, 213–223.
- 536 P. C. Joshi, S. Pitsch and J. P. Ferris, *Chem. Commun.*, 2000, 2497–2498.
- 537 P. C. Joshi, S. Pitsch and J. P. Ferris, *Orig. Life Evol. Biosph.*, 2007, **37**, 3–26.
- 538 P. C. Joshi, M. F. Aldersley and J. P. Ferris, *Orig. Life Evol. Biosph.*, 2011, **41**, 213–236.
- 539 A. Saghatelian, Y. Yokobayashi, K. Soltani and M. R. Ghadiri, *Nature*, 2001, **409**, 797–801.
- 540 J. E. Šponer, A. Mládek and J. Šponer, *Phys. Chem. Chem. Phys.*, 2013, **15**, 6235–6242.
- 541 G. F. Joyce, A. W. Schwartz, S. L. Miller and L. E. Orgel, *Proc. Natl. Acad. Sci.*, 1987, **84**, 4398–4402.
- 542 J. Rivera Islas, V. Pimienta, J.-C. Micheau and T. Buhse, *Biophys. Chem.*, 2003, **103**, 201–211.
- 543 M. Liu, L. Zhang and T. Wang, *Chem. Rev.*, 2015, **115**, 7304–7397.
- 544 S. C. Nanita and R. G. Cooks, *Angew. Chem. Int. Ed.*, 2006, **45**, 554–569.
- 545 Z. Takats, S. C. Nanita and R. G. Cooks, *Angew. Chem. Int. Ed.*, 2003, **42**, 3521–3523.
- 546 R. R. Julian, S. Myung and D. E. Clemmer, *J. Am. Chem. Soc.*, 2004, **126**, 4110–4111.
- 547 R. R. Julian, S. Myung and D. E. Clemmer, *J. Phys. Chem. B*, 2005, **109**, 440–444.
- 548 I. Weissbuch, R. A. Illos, G. Bolbach and M. Lahav, *Acc. Chem. Res.*, 2009, **42**, 1128–1140.
- 549 J. G. Nery, R. Eliash, G. Bolbach, I. Weissbuch and M. Lahav, *Chirality*, 2007, **19**, 612–624.
- 550 I. Rubinstein, R. Eliash, G. Bolbach, I. Weissbuch and M. Lahav, *Angew. Chem. Int. Ed.*, 2007, **46**, 3710–3713.
- 551 I. Rubinstein, G. Clodic, G. Bolbach, I. Weissbuch and M. Lahav, *Chem. – Eur. J.*, 2008, **14**, 10999–11009.
- 552 R. A. Illos, F. R. Bisogno, G. Clodic, G. Bolbach, I. Weissbuch and M. Lahav, *J. Am. Chem. Soc.*, 2008, **130**, 8651–8659.
- 553 I. Weissbuch, H. Zepik, G. Bolbach, E. Shavit, M. Tang, T. R. Jensen, K. Kjaer, L. Leiserowitz and M. Lahav, *Chem. – Eur. J.*, 2003, **9**, 1782–1794.
- 554 C. Blanco and D. Hochberg, *Phys. Chem. Chem. Phys.*, 2012, **14**, 2301–2311.
- 555 J. Shen, *Amino Acids*, 2021, **53**, 265–280.
- 556 A. T. Borchers, P. A. Davis and M. E. Gershwin, *Exp. Biol. Med.*, 2004, **229**, 21–32.
- 557 J. Skolnick, H. Zhou and M. Gao, *Proc. Natl. Acad. Sci.*, 2019, **116**, 26571–26579.
- 558 C. Böhler, P. E. Nielsen and L. E. Orgel, *Nature*, 1995, **376**, 578–581.
- 559 A. L. Weber, *Orig. Life Evol. Biosph.*, 1987, **17**, 107–119.
- 560 J. G. Nery, G. Bolbach, I. Weissbuch and M. Lahav, *Chem. – Eur. J.*, 2005, **11**, 3039–3048.
- 561 C. Blanco and D. Hochberg, *Chem. Commun.*, 2012, **48**, 3659–3661.
- 562 C. Blanco and D. Hochberg, *J. Phys. Chem. B*, 2012, **116**, 13953–13967.
- 563 E. Yashima, N. Ousaka, D. Taura, K. Shimomura, T. Ikai and K. Maeda, *Chem. Rev.*, 2016, **116**, 13752–13990.
- 564 H. Cao, X. Zhu and M. Liu, *Angew. Chem. Int. Ed.*, 2013, **52**, 4122–4126.
- 565 S. C. Karunakaran, B. J. Cafferty, A. Weigert-Muñoz, G. B. Schuster and N. V. Hud, *Angew. Chem. Int. Ed.*, 2019, **58**, 1453–1457.
- 566 Y. Li, A. Hammoud, L. Bouteiller and M. Raynal, *J. Am. Chem. Soc.*, 2020, **142**, 5676–5688.
- 567 W. A. Bonner, *Orig. Life Evol. Biosph.*, 1999, **29**, 615–624.
- 568 Y. Yamagata, H. Sakihama and K. Nakano, *Orig. Life*, 1980, **10**, 349–355.
- 569 P. G. H. Sandars, *Orig. Life Evol. Biosph.*, 2003, **33**, 575–587.
- 570 A. Brandenburg, A. C. Andersen, S. Höfner and M. Nilsson, *Orig. Life Evol. Biosph.*, 2005, **35**, 225–241.
- 571 M. Gleiser, *Orig. Life Evol. Biosph.*, 2007, **37**, 235–251.
- 572 M. Gleiser and J. Thorarinson, *Orig. Life Evol. Biosph.*, 2006, **36**, 501–505.
- 573 M. Gleiser and S. I. Walker, *Orig. Life Evol. Biosph.*, 2008, **38**, 293.
- 574 Y. Saito and H. Hyuga, *J. Phys. Soc. Jpn.*, 2005, **74**, 1629–1635.
- 575 J. A. D. Wattis and P. V. Coveney, *Orig. Life Evol. Biosph.*, 2005, **35**, 243–273.
- 576 Y. Chen and W. Ma, *PLOS Comput. Biol.*, 2020, **16**, e1007592.
- 577 M. Wu, S. I. Walker and P. G. Higgs, *Astrobiology*, 2012, **12**, 818–829.
- 578 C. Blanco, M. Stich and D. Hochberg, *J. Phys. Chem. B*, 2017, **121**, 942–955.
- 579 S. W. Fox, *J. Chem. Educ.*, 1957, **34**, 472.
- 580 J. H. Rush, *The dawn of life*, 1st Edition, Hanover House, Signet Science Edition, 1957
- 581 G. Wald, *Ann. N. Y. Acad. Sci.*, 1957, **69**, 352–368.
- 582 M. Ageno, *J. Theor. Biol.*, 1972, **37**, 187–192.
- 583 H. Kuhn, *Curr. Opin. Colloid Interface Sci.*, 2008, **13**, 3–11.
- 584 M. M. Green and V. Jain, *Orig. Life Evol. Biosph.*, 2010, **40**, 111–118.
- 585 F. Cava, H. Lam, M. A. de Pedro and M. K. Waldor, *Cell. Mol. Life Sci.*, 2011, **68**, 817–831.
- 586 B. L. Pentelute, Z. P. Gates, J. L. Dashnau, J. M. Vanderkooi and S. B. H. Kent, *J. Am. Chem. Soc.*, 2008, **130**, 9702–9707.
- 587 Z. Wang, W. Xu, L. Liu and T. F. Zhu, *Nat. Chem.*, 2016, **8**, 698–704.
- 588 A. A. Vinogradov, E. D. Evans and B. L. Pentelute, *Chem. Sci.*, 2015, **6**, 2997–3002.
- 589 J. T. Sczepanski and G. F. Joyce, *Nature*, 2014, **515**, 440–442.
- 590 K. F. Tjhung, J. T. Sczepanski, E. R. Murtfeldt and G. F. Joyce, *J. Am. Chem. Soc.*, 2020, **142**, 15331–15339.
- 591 F. Jafarpour, T. Biancalani and N. Goldenfeld, *Phys. Rev. E*, 2017, **95**, 032407.
- 592 G. Laurent, D. Lacoste and P. Gaspard, *Proc. Natl. Acad. Sci. USA*, 2021, **118**, e2012741118.

593 M. W. van der Meijden, M. Leeman, E. Gelens, W. L. Noorduyn, H. Meekes, W. J. P. van Enkevort, B. Kaptein, E. Vlieg and R. M. Kellogg, *Org. Process Res. Dev.*, 2009, **13**, 1195–1198.

594 J. R. Brandt, F. Salerno and M. J. Fuchter, *Nat. Rev. Chem.*, 2017, **1**, 1–12.

595 H. Kuang, C. Xu and Z. Tang, *Adv. Mater.*, 2020, **32**, 2005110.