ATOMIC FORCE MICROSCOPY ON MICROBIAL SURFACES



Audrey BEAUSSART & Sofiane EL-KIRAT-CHATEL

LIEC, UMR 7360, NANCY LCPME, UMR 7564, NANCY

Atomic Force Microscopy (AFM)



 This microscope works by touching objects with a sharp tip instead of looking at them »

The Nobel Prize in physics 1986



Gerd Binnig



Heinrich Rohrer

Atomic Force Microscopy (AFM)



Atomic Force Microscopy (AFM)







Geometry:	Poteted (Symmetric)
Tip Height (h):	2.5 - 8.0µm
Front Angle (FA):	15 ± 2.5°
Back Angle (BA):	25 ± 2.5 °
Side Angle (SA):	17.5 + 2.5 °
Tip Radius (Nom):	10 nm
Tip Radius (Max):	40 mm
Tip SetBack (TSB)(Nom):	4 µm
Tip Set Back (TSB)(RNG):	0 - 7 µm

Classical AFM

<image>



AFM mounted on an optical system







Imaging mode : principle



Imaging mode : contact vs peak force tapping

Contact



Peak force tapping





Color code



Immobilization techniques

Mammalian cells

(tend to naturally spread on the surface)



Micro-organisms

(round, difficult to deform)



AFM imaging of microbes

- Mechanically trapped in membrane
- Imaged under small force (100 pN)
- In physiological conditions (liquid)



Imaging mode : examples

Yeast

Candida albicans



Fungus

Aspergillus fumigatus Hydrophobic rodlets



Bacteria Lactococcus lactis peptidoglycan



Imaging artefacts

• Tip radius of curvature

B2*



A2

0.5 1.0

1.5

2.0

0.5 1.0 1.5 2.0 2.5 3 0 um

0.0

-750 nm





Imaging artefacts

• Tip contamination



Force Spectroscopy



Force-distance curves

Deflection sensitivity (nm/V)

Cantilever spring constant (N/m)

Hooke's law : $F = -k \times d$



Force-distance curves



Force-distance curves





Tip functionalisation



Mapping of individual molecules





Mapping of individual molecules

- Distribution
- Mechanical properties
- Dynamics

Single Cell Force Spectroscopy



Analytical Methods







A. Beaussart, P. Herman et al., 2013

Chemical force microscopy

Macroscopic measurements

Water contact angle







• Nanoscale measurements Chemical force microscopy







Macroscopic measurements

0.5

Chemical force microscopy of A. fumigatus





D. Alsteens et al., 2013

Nitzschia palea

- Photosynthetic microalgae
- Fresh water
- Motile
- Silica frustule with pores
- Adhere to surfaces and form mixed biofilms with other species
- Production of extracellular polymeric substances: EPS
- Bioindicator of water quality









What are the adhesive mechanisms of *N. palea* towards surfaces of different chemistry?

Chemical force microscopy of Nitzschia palea



M. Laviale, A. Beaussart, S. El-Kirat-Chatel, 2019

Adhesion of *Nitzschia palea*



Adhesion of *Nitzschia palea*

> Single cell force spectroscopy



> Single molecule probing by chemical force spectroscopy



CFM of Nitzschia palea



CFM of Nitzschia palea



Raman of Nitzschia palea

(Glyco)-lipids are the major EPS of *N. palea*



Proteins

Applications of AFM to green microalgae: Chlorella vulgaris



Applications of AFM to green microalgae: Chlorella vulgaris

