

CF2H as hydrogen bond donor group for the fine tuning of peptide bond geometry with difluoromethylated pseudoprolines

N Malquin, K Rahgoshay, N Lensen, G Chaume, E Miclet, Thierry Brigaud

▶ To cite this version:

N Malquin, K Rahgoshay, N Lensen, G Chaume, E Miclet, et al.. CF2H as hydrogen bond donor group for the fine tuning of peptide bond geometry with diffuoromethylated pseudoprolines. Chemical Communications, 2019, 55 (83), pp.12487-12490. 10.1039/C9CC05771D . hal-03549249

HAL Id: hal-03549249 https://hal.science/hal-03549249v1

Submitted on 31 Jan2022

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

View Article Online

ChemComm

Chemical Communications

Accepted Manuscript

This article can be cited before page numbers have been issued, to do this please use: N. Malquin, K. Rahgoshay, N. Lensen, G. Chaume, E. Miclet and T. Brigaud, *Chem. Commun.*, 2019, DOI: 10.1039/C9CC05771D.



This is an Accepted Manuscript, which has been through the Royal Society of Chemistry peer review process and has been accepted for publication.

Accepted Manuscripts are published online shortly after acceptance, before technical editing, formatting and proof reading. Using this free service, authors can make their results available to the community, in citable form, before we publish the edited article. We will replace this Accepted Manuscript with the edited and formatted Advance Article as soon as it is available.

You can find more information about Accepted Manuscripts in the Information for Authors.

Please note that technical editing may introduce minor changes to the text and/or graphics, which may alter content. The journal's standard <u>Terms & Conditions</u> and the <u>Ethical guidelines</u> still apply. In no event shall the Royal Society of Chemistry be held responsible for any errors or omissions in this Accepted Manuscript or any consequences arising from the use of any information it contains.



rsc.li/chemcomm

Published on 23 September 2019. Downloaded by Bibliotehque de L'universite de Cergy Pontoise on 9/25/2019 10:00:44 AM

COMMUNICATION

CF₂H as hydrogen bond donor group for the fine tuning of peptide bond geometry with difluoromethylated pseudoprolines

Received 00th January 20xx, Accepted 00th January 20xx

N. Malquin,^a K. Rahgoshay,^a N. Lensen,^{*a} G. Chaume,^{*a} E. Miclet^b and T. Brigaud^{*a}

DOI: 10.1039/x0xx00000x

 CF_2H -pseudoprolines obtained from difluoroacetaldehyde hemiacetal and serine are stable proline surrogates. The consequence of the incorporation of the CF_2H group is an important decrease of the *trans* to *cis* amide bond isomerisation energy and a remarkable stabilisation of the *cis* conformer by an hydrogen bond.

Because of the unique properties of fluorine atom, fluorinated compounds are increasingly recognized as highly promising for the discovery of new bioactive compounds.¹ Considering the specific case of peptides, the incorporation of fluorinated amino acids in peptides has been shown to impart remarkable properties.² It can modulate the peptide conformation and assembly,³ the local hydrophobicity,^{2,4} and the resistance towards proteolysis.⁵ Moreover fluorine atoms are very useful as highly sensitive labels for ¹⁹F-NMR spectroscopy because of ¹⁹F 100% natural abundance and the absence of background signals in natural and biological media.⁶ Although several methods have been reported for the synthesis of trifluoromethylated amino acids, the preparation of difluoromethylated one's is much less common.^{2,7} It is a lack because the CF₂H group presents unique features and we believe that this group will play an increasing role in peptides chemistry. The main property of the CF₂H group is to act as a lipophilic hydrogen bond donor group bioisostere of a hydroxyl or a thiol.⁸ These original bioisosteric properties have recently been highlighted in medicinal chemistry.⁹ Because of its cyclic structure, the proline residue is recognized to play a unique and crucial role on peptide backbone conformation. When included into a peptide, the pyrrolidine ring cannot act as an hydrogen bond donor and restrain the ϕ diehedral angle to about -60°.10 The Xaa-Pro peptide bond is characterized by a small free energy difference (ΔG°_{tc}) between the *trans* and the





Fig. 1 Peptidyl bond isomerization and puckering of CF₃-YPro

^{a.} Laboratory of Chemical Biology (LCB, EA 4505), Université de Cergy-Pontoise, 5 Mail Gay-Lussac, 95000 Cergy-Pontoise, France.

^{b.} Sorbonne Université, École normale supérieure, PSL University, CNRS, Laboratoire desBiomolécules, 75005 Paris, France.

⁺ Footnotes relating to the title and/or authors should appear here.

Electronic Supplementary Information (ESI) available: [details of any supplementary information available should be included here]. See DOI: 10.1039/x0xx00000x

Published on 23 September 2019. Downloaded by Bibliotehque de L'universite de Cergy Pontoise on 9/25/2019 10:00:44 AM



Scheme 1 Synthesis of H-Ser($\psi^{CF2H,H}$ Pro)-OBn (*S*,*S*)-1 and (*R*,*S*)-1

Considering the growing importance of the difluoromethyl group, we wish to present herein our results dedicated to difluoromethylated pseudoprolines (CF₂H- Ψ Pro). A particular attention is paid to the possibility of interactions through hydrogen bonds.

The oxazolidine ring of the CF₂H-pseudoproline was built by condensation of difluoroacetaldehyde ethyl hemiacetal with serine benzyl ester under an acidic catalysis (Scheme 1). Both diastereomers (S,S)-1 and (R,S)-1 were conveniently separated by silica gel chromatography. According to our previous investigations in the trifluoromethylated series,^{17a} the cis relative configuration of (R,S)-1 has been assigned by nuclear Overhauser effect experiments showing a correlation between the C_{α} and the C_{δ} protons. In order to investigate the electronic and geometric consequences due to the CF₂H group incorporation of CF_2H - Ψ Pro in a peptide chain, the Ac- Ψ Pro-NHMe model tripeptides (S,S)-4 and (R,S)-4 have been synthesized in a multi-step procedure (Scheme 2). The Nacetylation of (S,S)-1 with acetic anhydride under iodine catalysis provided a mixture of Ac-ΨPro-OBn (S,S)-2 (21%) and (R,S)-2 (39%). In order to circumvent the epimerization of the C_{δ} atom through an acidic catalysis,^{17a,b} the reaction was performed with acetyl chloride in pyridine. These experimental conditions led to the expected N-acetylated oxazolidines (S,S)-2 and (R,S)-2 in 81% and 75% yield respectively without epimerisation.

The *cis/trans* ratio of the acetyl- Ψ Pro bond was determined by ¹H and ¹⁹F NMR integration in CDCl₃. For that purpose, we first assigned each conformers by comparing the $\alpha,~\beta,$ and γ ^{13}C chemical shifts of the cis and the trans isomers according to Lubell et al.^{18a} For the *trans* isomer, the α and β -carbons appear upfield while they appear downfield in the cis isomer. Conversely, the δ carbon for the *trans* isomer appears downfield relative to that of the cis isomer. Once acetylated on their N-terminal position, we checked the acidic stability of the CF_2H -oxazolidines (S,S)-2 and (R,S)-2. Experiments showed that, similarly to the CF_3 - Ψ Pro, they are stable in an acidic medium (5% TFA in CHCl₃), whereas pseudoprolines of the nonfluorinated series are generally prone to ring opening.^{16a} Even when exposed to more concentrated acidic medium for several hours (3h in TFA/H2O 95:5) no ring opening or epimerization occurred. The CF₂H-oxazolidines are therefore suitable for SPPS strategy.



 a cisitrans conformers ratio of the peptidyl- ${\tt PPro}$ bond measured by NMR in CDCI_3 Scheme 2 Synthesis of Ac- ${\tt PPro-NHMe}$ (S,S)-4 and (R,S)-4

After removal of the benzyl ester function by hydrogenolysis in quantitative yields, the *N*-methyl amides (S,S)-**4** and (R,S)-**4** were obtained in 40% and 33% yield using classical peptide coupling conditions (Scheme 2).

The *cis-trans* bond isomerisation of the CF₂H-oxazolidinecontaining peptide models (*S*,*S*)-**4** and (*R*,*S*)-**4** was studied by ¹H NMR analysis in water at various temperatures. The coalescence temperatures were determined to estimate the rotational barriers for the *cis-trans* isomerisation. The results are reported in the Table 1 and compared with those reported for regular proline¹⁸ and pseudoproline^{17c} as well as those previously obtained in the CF₃ series.^{17c} The difference of isomerisation energy between the two diastereomers of the CF₃ series (ca. 2.5 kcal.mol⁻¹) is no longer observed in the CF₂H series, the energy barriers are low for both CF₂Hpseudoprolines. It is interesting to note that the energy barriers for amide isomerisation of both CF₂H-pseudoproline is ca 5.0 kcal.mol⁻¹ lower than for proline.¹⁸

The *cis* amide bond contents were significantly higher when the CF₃ substituent was replaced by the CF₂H group. Indeed in the apolar chloroform solution, the *cis* population of (*S*,*S*)-**4** reached even 78%. This enrichment may be due to the presence of a new H-bond involving the CF₂H group.⁸ Conversely, it was previously demonstrated that the *trans* amide bond conformer of the 2-(*R*)-CF₃- Ψ Pro analogue was stabilized by a H-bond between the *C*-terminal amide NH and the carbonyl of the *N*-terminal acetyl group.^{17c} Two antagonist H-bonds may thus compete in the compound (*R*,*S*)-**4** (Fig. 2). This balance was slightly in favour of the CF₂H H-bond in water (53% *cis* population) and underlines the hydrophobicity of the fluorinated group which partially preserves the stability of the H-bond in polar solvents such as methanol and water (see supplementary information).

Finally, the puckerings of the CF₂H-five-membered rings were determined by NMR from the coupling constants between the α and the β protons as previously reported in the CF₃ series (see supplementary information).^{17c}

The results showed that the Ac-(2S,4S)-CF₂H- Ψ Pro-NHMe (S,S)-**4** adopted a γ -endo puckering conformation for the two

Table 1 Thermodynamic properties of (*S*,*S*)-4 and (*R*,*S*)-4 determined by NMR. ^(a) Populations (%) observed in CDCl₃. ^(b) Populations (%) and ^(c) energies observed in H₂O:D₂O 90:10. All energies are given in kcal.mol⁻¹

Pro	$\Psi \text{Pro}^{\text{(ref 17c)}}$	2-(S)-CF ₃ -ΨPro ^(ref 17c)	2-(S)-CF₂H-Ψpro (<i>S,S</i>)- 4	2-(<i>R</i>)-CF₂H-Ψpro (<i>R,S</i>)- 4	2-(<i>R</i>)-CF ₃ -ΨPro ^(ref 17c)
13:87 (ref 18b)	27:73	40:60	78:22	32:68	24:76
24:76 (ref 18b)	34:66	43:57	60:40	53:47	45:55
20.40 (ref 18a)	18.41	18.06	15.19	15.82	15.63
19.83 ^(ref 18a)	17.93	17.88	15.45	15.90	15.51
	Pro 13:87 (ref 18b) 24:76 (ref 18b) 20.40 (ref 18a) 19.83 (ref 18a)	Pro ΨPro ^(ref 17c) 13:87 ^(ref 18b) 27:73 24:76 ^(ref 18b) 34:66 20.40 ^(ref 18a) 18.41 19.83 ^(ref 18a) 17.93	Pro ΨPro (ref 17c) 2-{S}-CF ₃ -ΨPro (ref 17c) 13:87 (ref 18b) 27:73 40:60 24:76 (ref 18b) 34:66 43:57 20.40 (ref 18a) 18.41 18.06 19.83 (ref 18a) 17.93 17.88	$\begin{array}{c c c c c c c } & & & & & & & & & & & & & & & & & & &$	Pro $\Psi Pro^{(ref 17c)}$ $2 \cdot (S) - CF_3 - \Psi Pro^{(ref 17c)}$ $2 \cdot (S) - CF_2 H - \Psi pro$ $2 \cdot (R) - CF_2 H - \Psi pro$ 13:87 (ref 18b) 27:73 40:60 78:22 32:68 24:76 (ref 18b) 34:66 43:57 60:40 53:47 20.40 (ref 18a) 18.41 18.06 15.19 15.82 19.83 (ref 18a) 17.93 17.88 15.45 15.90

2 | J. Name., 2012, 00, 1-3

This journal is © The Royal Society of Chemistry 20xx

Published on 23 September 2019. Downloaded by Bibliotehque de L'universite de Cergy Pontoise on 9/25/2019 10:00:44 AM



Fig. 2 Possible intramolecular H-bonds for Ac- Ψ Pro-NHMe (S,S)-4 and (R,S)-4

geometries of the peptide bond. The same observation was made for the 2-(*S*)-CF₃- Ψ Pro analogue.^{17c} Conversely the compound (*R*,*S*)-**4** underwent a puckering switch when the amide bond geometry was reversed.

Instead of maintaining an *exo*-puckered oxazolidine ring for both the *cis* and the *trans* amide conformers, as observed with 2-(R)-CF₃- Ψ Pro,^{17c} the *cis* rotamer of (*R*,*S*)-**4** showed a slight preference for the *endo* puckering to allow the CF₂H H-bond.

These structural features can be understood by analyzing the unusual NMR parameters of the CF₂H groups. First, a high chemical shift difference ($\Delta\delta$) of the diastereotopic ¹⁹F informed about a restricted dynamic of the difluoromethyl group. For the *cis* conformers, $\Delta\delta$ was above 5 ppm, whereas $\Delta \delta \approx 3$ ppm for their corresponding *trans* conformations (see Fig. 3 and supplementary information). Second, the measured vicinal couplings reflected a very limited rotameric averaging of the CF2H group for the cis conformers. In particular the Ac-(2S,4S)-CF₂H- Ψ Pro-NHMe (S,S)-4 displayed the following values: ${}^{3}J_{H\delta-F1}$ = 3.2 Hz; ${}^{3}J_{H\delta-F2}$ = 18.2 Hz; ${}^{3}J_{H\epsilon-H\delta}$ = 1.0 Hz. These three couplings are compatible with a single $H^{\epsilon}-C^{\epsilon}-C^{\delta}-H^{\delta}$ dihedral angle of ~50° (see supplementary information). Such a restricted rotation of the CF₂H group likely arose from the Hbonding observed in the cis forms since the oxygen of the preceding acetyl group is facing the positively charged CF₂H proton in this conformation (Fig. 3). The endo puckering of the oxazolidine ring may be needed for the CF₂H H-bonding establishment in the cis conformers, whereas the exo puckering was required for the NH...O=C H-bond in the trans form.^{17c} This provides an explanation for the trans-exo to cisendo transition observed for the compound (R,S)-4. The (S,S)-4 diastereoisomer adopts preferentially the endo conformation which explains the high stability of the CF₂H...O=C H-bond in the cis conformer. In the trans conformers, the CF₂H group cannot establish any H-bond since the O=C acceptor of the acetyl is moved away and replaced by the methyl group. As a result, several rotamers of the CF₂H group are present as attested by the averaged NMR parameters (Fig. 3).

In order to expand the scope of the incorporation of CF₂Hpseudoprolines in peptides, a preliminary methodological study of the peptide coupling reactions at their N- and Cposition was undertaken. We already demonstrated that the presence of a CF₃ group at the δ position of an oxazolidine unit is detrimental for coupling reactions because of its powerful withdrawing effect. Although the CF₂H group is less deactivating, typical protocols using HOBt, EDCI, DIPEA or IBCF reagents failed to achieve the N-coupling reaction of (S,S)-1 and (R,S)-1 with Fmoc-Gly-OH. However, the peptide coupling could be achieved in good yields while using the most electrophilic Fmoc protected glycine and alanine amino acid chloride (Scheme 3). When the reaction was performed in the presence of pyridine, the epimerization of the $\boldsymbol{\delta}$ carbon of the oxazolidine ring through a dynamic kinetic resolution (DKR) process17b was avoided or limited. The expected dipeptides (S,S)-5, (R,S)-5, (S,S)-6 and (R,S)-6 were obtained in 86%, 76%, 57% and 88% yield respectively (Scheme 3). It should be





 $^{\rm a}$ cis/trans conformers ratio of the peptidyl- ΨPro bond measured by NMR in ${\rm CDCI}_3$

Scheme 3 Incorporation of CF₂H-pseudoprolines in a peptide chain

noticed that in any cases the cis conformer of the amide bond was the major one. The cis/trans conformers ratio of the peptidyl- Ψ Pro bond measured by NMR in CDCl₃ were ranging from 58/42 to 83/17. After having successfully solved the challenging coupling reaction at the N-terminal positions of the CF₂H-pseudoprolines, suitable experimental conditions were investigated for the C-terminal coupling. The removal of the benzyl ester was carried out by palladium mediated hydrogenolysis without impacting the N-Fmoc protecting group. The free carboxylic acid was then used in the next step without further purification. Typical protocols involving HOBt, EDCI, DIPEA at room temperature were not suitable for the coupling reaction between dipeptides 6 and HCl.H₂N-Ala-O^tBu. However, the coupling of the CF₂H-dipeptides using BOP-Cl in the presence of TEA afforded the expected tripeptides (S,S)-7 and (*R*,*S*)-7 in 56% and 63% yield respectively. ¹⁹F and ¹H NMR measurements in CDCl₃ revealed a 85/15 cis/trans ratio for (S,S)-7 and 59/41 for (R,S)-7.

In conclusion, original enantiopure (*S*,*S*) and (*R*,*S*) CF₂Hpseudoprolines have been synthesized as stable 5-substituted proline surrogates which can be incorporated in a peptide chain. The CF₂H group provides a strong decrease of the *trans* to *cis* amide bond isomerisation energy and enhances the *cis* conformation content compared to the CF₃ series, by acting as an efficient hydrogen bond donor. These CF₂H-proline surrogates will provide outstanding tools for the fine tuning of the dynamic and the secondary structure of peptides, in particular type VI β -turns.^{13d} Such conformations are known to play important roles in protein structures and molecular recognition. New non-natural amino acids able to stabilize the *cis*-isomer backbone geometry and/or type VI β -turns are still in demand for the design of potent and selective prototypes for drug discovery.¹⁹

The authors gratefully acknowledge Central Glass Co. for the gift of difluoroacetaldehyde hemiacetal and the Agence Nationale de la Recherche for funding (ANR CH2PROBE and ANR F-LAIR). K. R. thanks the MNREST for a fellowship.

Conflicts of interest

There are no conflicts to declare.

Notes and references

 (a) S. Purser, P. R. Moore, S. Swallow and V. Gouverneur, *Chem. Soc. Rev.*, 2008, **37**, 320; (b) N. A. Meanwell *J. Med. Chem.*, 2018, **61**, 5822; (c) E. P. Gillis, K. J. Eastman, M. D. Hill, D. J. Donnelly and N. A. Meanwell, *J. Med. Chem.*, 2015, **58**, 8315; (d) W. K. J. Hagmann, *J. Med. Chem.*, 2008, **51**,

Journal Name

4359; (e) J. Wang, M. Sánchez-Roselló, J. L. Aceña, C. del Pozo, A. E. Sorochinsky, S. Fustero, V. A. Soloshonok and H. Liu, Chem. Rev., 2014, 114, 2432; (f) Y. Zhou, J. Wang, Z. Gu, S. Wang, W. Zhu, J.L. Aceña, V. A. Soloshonok, K. Izawa and H. Liu, Chem. Rev., 2016, 116, 422.

- (a) A. A. Berger, J.-S. Völler, N. Budisa and B. Koksch, Acc. 2 Chem. Res., 2017, 50, 2093; (b) E. N. G. Marsh, Acc. Chem. Res., 2014, 47, 2878; (c) M. Salwiczek, E. K. Nyakatura, U. I. M. Gerling, S. Ye and B. Koksch, Chem. Soc. Rev., 2012, 41, 2135.
- U. I. M. Gerling, M. Salwiczek, C. D. Cadicamo, H. Erdbrink, C. 3 Czekelius, S. L. Grage, P. Wadhwani, A. S. Ulrich, M. Behrends, G. Haufe and B. Koksch, Chem. Sci., 2014, 5, 819.
- (a) C. Gadais, E. Devillers, V. Gasparik, E. Chelain, J. Pytkowicz and T. Brigaud, ChemBioChem, 2018, 19, 1026; (b) J. R. Robalo, S. Huhmann, B. Koksch and A. Vila Verde, Chem, 2017, 3, 881.
- (a) S. Huhmann and B. Koksch, Eur. J. Org. Chem., 2018, 5 3667, (b) V. Asante, J. Mortier, G. Wolber and B. Koksch, Amino Acids, 2014, 46, 2733; (c) G. Akcay and K. Kumar, J. Fluorine Chem., 2009, 130, 1178.
- For review on the use of ¹⁹F NMR, see: (a) E. N. G. Marsh and 6 Y. Suzuki, ACS Chem. Biol., 2014, 9, 1242; (b) A. Vulpetti and C. Dalvit, Drug Discov. Today, 2012, 17, 890; (c) K. Koch, S. Afonin, M. Ieronimo, M. Berditsch and A. S. Ulrich, Top. Curr. Chem., 2012, 306, 89; (d) K. E. Arntson and W. C. K. Pomerantz, J. Med. Chem., 2016, 59, 5158.
- Y.-G. Lou, A.-J. Wang, L. Zhao, L.-F. He, X.-F. Li, C.-Y. He and X. Zhang, Chem. Commun., 2019, 55, 3705.
- (a) Y. Zafrani, D. Yeffet, G. Sod-Moriah, A. Berliner, D. Amir, D. Marciano, E. Gershonov and S. Saphier, J. Med. Chem., 2017, 60, 797; (b) C. D. Sessler, M. Rahm, S. Becker, J. M. Goldberg, F. Wang and S. J. Lippard, J. Am. Chem. Soc., 2017, 139, 9325; (c) J. A. Erickson and J. I. McLoughlin, J. Org. Chem., 1995, 60, 1626; (d) Q. A. Huchet, B. Kuhn, B. Wagner, N. A. Kratochwil, H. Fischer, M. Kansy, D. Zimmerli, E. M. Carreira and K. Müller, J. Med. Chem., 2015, 58, 9041; (e) A. Meanwell, J. Med. Chem., 2011, 54, 2529.
- (a) B. Zheng, S. V. D'Andrea, L.-Q. Sun, A. X. Wang, Y. Chen, P. Hrnciar, J. Friborg, P. Falk, D. Hernandez, F. Yu, A. K. Sheaffer, J. O. Knipe, K. Mosure, R. Rajamani, A. C. Good, K. Kish, J. Tredup, H. E. Klei, M. Paruchuri, A. Ng, Q. Gao, R. A. Rampulla, A. Mathur, N. A. Meanwell, F. McPhee and P. M. Scola, ACS Med. Chem. Lett., 2018, 9, 143; (b) T. Naret, J. Bignon, G. Bernadat, M. Benchekroun, H. Levaique, C. Lenoir, J. Dubois, A. Pruvost, F. Saller, D. Borgel, B. Manoury, V. Leblais, R. Darrigrand, S. Apcher, J. D. Brion, E. Schmitt, F. R. Leroux, M. Alami and A. Hamze, Eur. J. Med. Chem., 2018, 143, 473; (c) F. Narjes, K. F. Koehler, U. Koch, B. Gerlach, S. Colarusso, C. Steinkühler, M. Brunetti, S. Altamura, R. De Francesco and V. G. Matassa, Bioorg. Med. Chem. Lett., 2002, 12, 701; (d) S. Thompson, S. A. McMahon, J. H. Naismith and D. O'Hagan, Bioorg. Chem., 2016, 64, 37
- 10 F. A. Momany, R. F. McGuire, A. W. Burgess and H. A. Scheraga, J. Phys. Chem., 1975, 79, 2361.
- 11 (a) S. S. Zimmerman and H. A. Scheraga, Macromolecules, 1976, 9, 408; (b) D. E. Stewart, A. Sarkar and J. E. Wampler, J. Mol. Biol., 1990, 214, 253; (c) S. Fischer, R. L. Dunbrack and M. Karplus, J. Am. Chem. Soc., 1994, 116, 11931; (d) D. Kern, M. Schutkowski and T. Drakenberg, J. Am. Chem. Soc., 1997, 119, 8403.
- 12 (a) M. Levitt, J. Mol. Biol., 1981, 145, 251; (b) A. J. Salahuddin, J. Biosci., 1984, 6, 349; (c) W. J. Wedemeyer, E. Welker and H. A. Scheraga, Biochemistry, 2002, 41, 14637; (d) Mechanisms of Protein Folding, Ed. R. H. Pain, Oxford University Press, New York, 1994; (e) J. F. Brandts, H. R. Halvorson and M. Brennan, Biochemistry, 1975, 14, 4953; (f) U. Reimer, G. Scherer, M. Drewello, S. Kruber, M.

Schutkowski and G. Fischer, J. Mol. Biol. 1998, 279, 449; (g) P. A. M. Schmidpeter, J. R. Koch and Fox Schmid, Biochim Biophys. Acta, 2015, 1850, 1973; (h) W. J. Wedemeyer, E. Welker and H. A. Scheraga, Biochemistry, 2002, 41, 14637.

- 13 (a) P. Karoyan, S. Sagan, O. Lequin, J. Quancard, S. Lavielle and G. Chassaing, in Targets in Heterocyclic Systems-Chemistry and Properties, O. A. Eds Attanasi, D. Spinelli, Royal Society of Chemistry, Cambridge, 2005, pp 216-273; (b) Cis-Trans Isomerization in Biochemistry, Ed. C. Dugave, Wiley-VCH, Weinheim, 2006; (c) C. Dugave and L. Demange, Chem. Rev., 2003, 103, 2475; (d) Y. Che and G. R. Marshall, Biopolymers, 2006, 81, 392.
- 14 (a) V. Kubyshkin, S. Pridma and N. Budisa, New J. Chem., 2018, 42, 13461 and refs cited; (b) N. A. Tolmachova, I. S. Kondratov, V. G. Dolovanyuk, S. O. Pridma, A. V. Chernykh, C. G. Daniliuc and G. Haufe, Chem. Commun., 2018, 54, 9683; (c) S. J. M. Verhoork, P. M. Killoran and C. R. Coxon, Biochemistry, 2018, 57, 6132; (d) M. Oliver, C. Gadais, J. García-Pindado, M. Teixidó, N. Lensen, G. Chaume and T. Brigaud, RSC Adv., 2018, 8, 14597; (e) C. Caupène, G. Chaume, L. Ricard and T. Brigaud, Org. Lett., 2009, 11, 209; (f) I. Jlalia, N. Lensen, G. Chaume, E. Dzhambazova, L. Astasidi, R. Hadjiolova, A. Bocheva and T. Brigaud, Eur. J. Med. Chem., 2013, 62, 122; (g) J. Simon, J. Pytkowicz, N. Lensen, G. Chaume and T. Brigaud, J. Org. Chem., 2016, 81, 5381; (h) H. Lubin, J. Pytkowicz, G. Chaume, G. Sizun-Thomé and T. Brigaud, J. Org. Chem., 2015, 80, 2700; (i) V. Kubyshkin, S. Afonin, S. Kara, N. Budisa, P. K. Mykhailiuk and A. S. Ulrich, Org. Biomol. Chem., 2015, 13, 3171; (j) I. S. Kondratov, V. G. Dolovanyuk, N. Tolmachova, I. I. Gerus, K. Bergander, R. Fröhlich and G. Haufe, Org. Biomol. Chem., 2012, 10, 8778; (k) N. G. Voznesenskaia, O. I. Shmatova, V. N. Khrustalev and V. G. Nenajdenko, Org. Biomol. Chem., 2018, 16. 7004: (/) J. Del Valle and M. Goodman. Angew. Chem. Int. Ed., 2002, 41, 1600; (m) X.-l. Qiu and F.-l. Qing, J. Org. Chem., 2002, 67, 7162.
- 15 (a) R. Nadano, Y. Iwai, T. Mori and J. Ichikawa, J. Org. Chem., 2006, 71, 8748; (b) X. L. Qiu and F. L. Qing, J. Org. Chem., 2003, 68, 3614; (c) R. C. McAtee, J. W. Beatty, C. C. McAtee and C. R. J. Stephenson, Org. Lett., 2018, 20, 3491.
- 16 (a) T. Wöhr and M. Mutter, Tetrahedron Lett., 1995, 36, 3847; (b) P. Dumy, M. Keller, D. E. Ryan, B. Rohwedder, T. Wöhr and M. Mutter, J. Am. Chem. Soc., 1997, 119, 91; (c) M. Keller, C. Sager, P. Dumy, M. Schutkowski, G. S. Fischer and M. Mutter, J. Am. Chem. Soc., 1998, 120, 2714.
- 17 (a) G. Chaume, O. Barbeau, P. Lesot and T. Brigaud, J. Org. Chem., 2010, 75, 4135; (b) G. Chaume, J. Simon, C. Caupene, N. Lensen, E. Miclet and T. Brigaud, J. Org. Chem., 2013, 78, 10144; (c) D. Feytens, G. Chaume, G. Chassaing, S. Lavielle, T. Brigaud, B. J. Byun, Y. K. Kang, E. Miclet, J. Phys. Chem. B, 2012, 116, 4069; (d) G. Chaume, D. Feytens, G. Chassaing, S. Lavielle, T. Brigaud and E. Miclet, New J. Chem., 2013, 37, 1336; (e) G. Chaume, J. Simon, N. Lensen, J. Pytkowicz, T. Brigaud and E. Miclet, J. Org. Chem., 2017, 82, 13602.
- 18 (a) E. Beausoleil and W. D. Lubell, J. Am. Chem. Soc., 1996, 118, 12902; (b) C. B. Braga, W. G. D. P. Silva and R. Rittner, New J. Chem. 2019, 43, 1757.
- 19 For a recent example see R. Chingle, M. Mulumba, N. N. Chung, T. M.-D. Nguyen, H. Ong, S. Ballet, P. W. Schiller and W. D. Lubell, J. Org. Chem., 2019, 84, 6006 and cited references.

Published on 23 September 2019. Downloaded by Bibliotehque de L'universite de Cergy Pontoise on 9/25/2019 10:00:44 AM