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Nelly Béchir, Émilie Pecchi, Christophe Vilmen, Monique Bernard, David Bendahan, et al.. Activin type IIB receptor blockade does not limit adenosine triphosphate supply in mouse skeletal muscle in Vivo. Muscle & nerve. Supplement., 2018, 58 (6), pp.834-842. <10.1002/mus.26306>. <hal-03548285>

HAL Id: hal-03548285

<https://hal.science/hal-03548285v1>

Submitted on 30 Jan 2022

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ACTIVIN TYPE IIB RECEPTOR BLOCKADE DOES NOT LIMIT ADENOSINE TRIPHOSPHATE SUPPLY IN MOUSE SKELETAL MUSCLE *IN VIVO*

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Accepted 15 July 2018

ABSTRACT: *Introduction:* Postnatal activin/myostatin type IIB receptor (ActRIIB) blockade increases skeletal muscle mass and strength but also increases muscle fatigability and impairs oxidative metabolism. The objective of this study was to determine *in vivo* whether this increased fatigability is due to energy supply limitation. *Methods:* The impact of 8-week ActRIIB blockade with soluble receptor (sActRIIB-Fc) on muscle function and adenosine triphosphate (ATP) fluxes was investigated noninvasively by using multimodal magnetic resonance and indirect calorimetry measurements in wild-type mice. *Results:* Activin/myostatin type IIB receptor blockade reduced (−41%) the muscle apparent mitochondrial capacity and increased (+11%) the basal body energy expenditure. During a fatiguing exercise, ActRIIB blockade decreased both oxidative ATP production rate (−32%) and fatigue resistance (−36%), but these changes affected neither the total ATP production rate nor the contractile ATP cost. *Discussion:* These findings demonstrate that the increased fatigability after ActRIIB blockade is not due to limitation in energy supply and/or disturbance in contractile ATP cost.

Muscle Nerve 58:834–842, 2018

Skeletal muscle overgrowth is limited by various mechanisms, especially through the activin type IIB receptor (ActRIIB), a transmembrane serine-threonine kinase receptor highly expressed in skeletal muscle and adipose tissue. Activin type IIB receptor mediates the signaling for secreted molecules of the transforming growth factor- β family involved in the negative regulation of muscle development, the most important being myostatin (GDF-8).¹ Postnatal blockade of ActRIIB signaling by using systemic injection of soluble ActRIIB decoy receptor (sActRIIB-Fc) or neutralizing antibodies against ActRIIB has been shown to induce a rapid and massive muscle hypertrophy leading to absolute strength improvement in healthy human and in normal and dystrophic mice.^{2–9} These findings have raised the possibility of pharmaceutical

blockade of ActRIIB as a promising means for improving the quality of life and reducing mortality of patients with muscle wasting associated with conditions such as neuromuscular disorders, cancer, human immunodeficiency virus, and sarcopenia.^{4,10–12}

The beneficial effect of ActRIIB blockade on muscle strength is limited by concomitant impairments in mechanical performance and bioenergetics. Activin type IIB receptor inhibition in healthy mice has been shown to reduce fatigue resistance in hind limb muscle electrically stimulated *in situ*^{2,9} and during running tests.³ Nevertheless, the origin of this increased fatigability remains unknown. Long-term sActRIIB-Fc treatment in mice has been reported to reduce the expression of genes involved in oxidative phosphorylation^{13,14} (via reduction of proliferator-activated receptors and peroxisome-proliferator-activated receptor (PPAR)- γ -coactivator 1 α as a consequence of PPAR- β downregulation), the level of porin³ (a mitochondrial protein involved in adenosine triphosphate [ATP] transport between the mitochondrial matrix and the cytosol), and mitochondrial oxidative capacity *in vivo*,⁹ hence indicating mitochondrial dysfunction. Because limitation in energy supply during muscular activity is considered a major contributor to the development of fatigue,^{15,16} this raises the question regarding whether a link exists between increased fatigability and mitochondrial dysfunction in muscle undergoing ActRIIB blockade. This warrants consideration not only to facilitate a better understanding of the physiological impact of ActRIIB blockade but also to clarify whether the upregulation of oxidative metabolism would be justified in that context. Such an approach has been envisaged as complementary to ActRIIB blockade to reverse mitochondrial dysfunction;¹⁷ however, no study has yet been devoted to the investigation of muscle performance with respect to energy fluxes; the effect of the impact of mitochondrial dysfunction induced by ActRIIB blockade over energy expenditure *in vivo* has not been well studied and remains controversial. Indirect calorimetry experiments have shown that 4-week ActRIIB blockade does not change basal oxygen consumption per animal despite substantial muscle hypertrophy and body weight increase, thereby suggesting that muscle mitochondrial dysfunction does not affect global metabolic rate.¹⁸ On the contrary, it has been proposed on the basis of similar experiments that ActRIIB blockade reduces energy

Additional supporting information may be found in the online version of this article.

Key words: activin myostatin receptor, energy expenditure, exercise, fatigue, muscle wasting therapy, oxidative capacity

Abbreviations: ³¹P, 31-phosphorus; ActRIIB, activin type IIB receptor; ANCOVA, analysis of covariance; ATP, adenosine triphosphate; EE, energy expenditure; FID, free induction decays; MR, magnetic resonance; PBS, phosphate-buffered saline; PCr, phosphocreatine; Pi, inorganic phosphate; sActRIIB-Fc, soluble ActRIIB decoy receptor; SR, sarcoplasmic reticulum; VCO₂, CO₂ production; VO₂, oxygen consumption

Funding: This work was financially supported by the French Muscular Dystrophy Association (AFM-Téléthon; to BG) and the France Life Imaging network (Grant ANR-11-INBS-0006; to MB).

Conflicts of Interest: None of the authors have any conflicts of interest to disclose.

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Published online 19 July 2018 in Wiley Online Library (wileyonlinelibrary.com). DOI 10.1002/mus.26306

efficiency,¹⁹ which echoes previous studies in myostatin-deficient mice showing that muscle hypertrophy is associated with a dramatic increase in ATP cost of contraction.^{20,21} Moreover, because energy cost of contraction is an important factor in muscle performance and physical capacity,^{22–24} any increase in this variable might contribute to the increased fatigability induced by ActRIIB blockade.

In this study, we noninvasively investigated the impact of ActRIIB blockade on muscle function and bioenergetics in normal mice. In particular, we sought to determine whether a limitation in energy supply and/or an increase in contractile ATP cost could explain the increase in muscle fatigability already reported in this model. For this purpose, we have used a global *in vivo* approach, assessing whole-body energy expenditure and ATP production rates in working muscle.

MATERIALS AND METHODS

Animal Care and Feeding. Fifteen C57BL/6 3-month-old female mice (Janvier Labs, Le Genest Saint Isle, France) were used for these experiments, which were conducted in strict accordance with the guidelines of the European Communities Council Directive 86/609/EEC for care and use of laboratory animals. All experiments were performed according to protocols reviewed and approved by the institutional animal care committee of Aix-Marseille University (93-21122012) and under the supervision of B. Giannesini (license number 13.164 2008/11/25). Mice were housed 3 or 4 per cage in an environmentally controlled facility (12-h light/dark cycle, 22°C) with free access to commercial standard food and water. After experiments, animals were humanely killed by cervical dislocation after isoflurane anesthesia, and gastrocnemius muscles were immediately removed, freeze clamped with nitrogen-chilled metal tongs, and stored at –80°C for *in vitro* measurement.

ActRIIB Signaling Blockade. The blockade was performed by using a soluble fusion protein generated by linking the ActRIIB extracellular domain to the mouse immunoglobulin Fc region (sActRIIB-Fc; provided by Acceleron Pharma, Cambridge, Massachusetts). Mice were randomly assigned to 2 groups and were injected subcutaneously twice weekly for a total of 8 weeks with sActRIIB-Fc (10 mg/kg body weight; $n = 8$) or an equal volume of vehicle (10 mM phosphate-buffered saline [PBS]; control group; $n = 7$).

NONINVASIVE INVESTIGATION OF GASTROCNEMIUS MUSCLE FUNCTION AND BIOENERGETICS

Muscle anatomy and changes in bioenergetics associated with force output were investigated with multimodal magnetic resonance (MR) data acquisition throughout a standardized fatiguing bout of exercise consisting of 6 min of repeated maximal isometric contractions induced by transcutaneous electrostimulation at a frequency of 1.7 Hz.

Each mouse was initially anesthetized in an induction chamber by using an air flow (3 L/min) containing 4% isoflurane. The left hind limb was shaved, and electrode cream was applied at the knee

and heel regions to optimize electrostimulation. The anesthetized animal was then placed supine in a custom-built experimental cradle that had been designed for the noninvasive MR investigation of gastrocnemius muscle function.²⁵ Corneas were protected from drying by applying ophthalmic cream, and the animal's head was placed in a facemask that was continuously supplied with 1.75% isoflurane in 33% O₂ (0.2 L/min) and 66% N₂O (0.4 L/min). The left foot was positioned on the pedal of an ergometer, the lower hind limb was centered inside a 20-mm-diameter ¹H Helmholtz imaging coil, and the belly of the gastrocnemius muscle was located above an elliptic (8 × 12 mm²) ³¹P-MR spectroscopy surface coil. Body temperature was monitored and maintained at physiological level throughout the entire experiment by using a feedback loop including electrical heating, a temperature control unit, and a rectal thermometer.

Muscle contractions were achieved *in vivo* by using 2 rod-shaped transcutaneous electrodes integrated in the experimental cradle and connected to a constant-current stimulator (DS7A; Digitimer, Hertfordshire, United Kingdom): 1 electrode was located above the knee and the other electrode was placed under the heel. Mechanical response to electrostimulation was measured by amplifying (operational amplifier AD620; Analog Devices, Norwood, Massachusetts) the electrical signal coming out from the force transducer. The signal was then digitized (PCI-6220; National Instrument, Austin, Texas) and continuously monitored and recorded on a personal computer in WinATS version 6.5 (Sysma, Aix-en-Provence, France). The digitalized signal was converted to force according to a linear calibration curve. At the beginning of each experiment, resting muscle was passively stretched by adjusting the angle between the foot and the lower hind limb to reach maximal isometric twitch amplitude after supramaximal square wave pulses (1-ms duration). The individual maximal electrostimulation intensity was determined by progressively increasing the stimulus intensity until there was no further peak twitch force increase.

Magnetic resonance explorations were performed in the 4.7-Tesla horizontal magnet of a 47/30 Biospec Avance MR system (Bruker, Karlsruhe, Germany) equipped with a Bruker 120-mm BGA12SL (200 mT/m) gradient insert. Ten consecutive noncontiguous axial slices (1-mm thickness, 0.5-mm interslice gap) covering the region from the knee to the ankle were selected across the hind limb. Magnetic resonance images of these slices (rapid acquisition with relaxation enhancement sequence; 8 echoes; 67.9-ms effective echo time; 16.7-ms actual echo time; 2,000-ms repetition time; 1 accumulation; 20 × 15 mm² field of view; 256 × 256 matrix size) were recorded at rest. ³¹P-MR spectra

(8 kHz sweep width; 2,048 data points) from the gastrocnemius region were dynamically acquired at rest (6 min), during the 6-min fatiguing bout of exercise, and during the following 15-min recovery period. Magnetic resonance data acquisition was gated to muscle electrostimulation for reducing potential motion artifacts due to contraction. A fully relaxed spectrum (12 scans, 20-s repetition time) was acquired at rest, followed by a total of 768 saturated free induction decays (FID; 1.875-s repetition time). The first 64 FIDs were acquired at rest and added together. The next 192 FIDs were acquired during the stimulation period and were added by packets of 32, allowing a temporal resolution of 60 s. The remaining 512 FIDs were obtained during the post-stimulation recovery period and were added as 7 packets of 32 FIDs followed by 3 packets of 64 FIDs (120-s temporal resolution) and 1 packet of 96 FIDs (180-s temporal resolution).

Magnetic resonance data were processed by using custom-written analysis algorithms developed under the IDL environment (Interactive Data Language; Harris Geospatial Solutions, Broomfield, Colorado). For each MR image, the region of interest was manually outlined so that the corresponding cross-sectional area of the gastrocnemius muscle was measured. Muscle volume was then calculated by adding the volumes included between consecutive slices by using the truncated cone formula. Intracellular pH was calculated from the chemical shift of the inorganic phosphate (P_i) signal relative to phosphocreatine (PCr).²⁶ Relative concentrations of PCr, P_i , and ATP were obtained from ^{31}P -MR spectra by a time-domain fitting routine by using the AMARES-MRUI Fortran code and appropriate prior knowledge for the ATP multiplets.²⁷ Absolute amounts of phosphorylated compounds were expressed relative to a resting β -ATP concentration determined in muscle extract by using a bioluminescence-based method (Supp. Info.). The rates of ATP production from oxidative and anaerobic pathways were calculated *in vivo* during the fatiguing bout of exercise according to the quantitative interpretation of bioenergetics data^{28,29} (Supp. Info.). Adenosine triphosphate cost of contraction was calculated as total ATP production (oxidative plus anaerobic) scaled to force generation during the same period of time.

In Vivo Quantitation of Body Fat Content. Magnetic resonance image acquisition was performed with a whole-body imaging coil (PRK 200 RES 200; Bruker) inserted in the 4.7-Tesla horizontal magnet of the 47/30 Biospec Avance MR system. Mice were kept anesthetized by using isoflurane inhalation as previously described. Axial MR images were acquired across the whole body (excluding the tail) by using a high-resolution 3-dimensional (turbo spin echo)

sequence (5.53-ms echo time; 77.85-ms effective echo time; 300-ms repetition time; 2 averages; $40 \times 40 \times 80 \text{ mm}^3$ field of view; $128 \times 128 \times 64$ matrix size). Fat volume was quantitated by using an automatic segmentation method that was based on a pixel-intensity analysis of MR images with the FMRIB Software Library v5.0.2.2, (Oxford University, United Kingdom; <http://www.fmrib.ox.ac.uk/fsl>). Fat mass was calculated by using a density of 0.92 g/cm^3 for adipose tissue.³⁰ Adiposity index was calculated as fat mass relative to total body weight.

Body Energy Expenditure. Measurements were performed at room temperature (23°C) by using a 2-chamber Oxylet system (Panlab, Barcelona, Spain) with a constant inlet flow of room air (5 ml/min). Animals were individually acclimated and kept for 24 h in metabolic cages with free access to food and water. Oxygen consumption (VO_2) and CO_2 production (VCO_2) were continuously analyzed by using the following sequence: 5 min from cage No. 1, 5 min from cage No. 2, and 5 min from room air. Energy expenditure (EE) was calculated as $\text{EE} = (3.815 + 1.232 \text{ RQ}) \times \text{VO}_2$, in which RQ is the respiratory quotient of VCO_2 to VO_2 . Ambulatory activity (locomotion and rearing) was determined simultaneously with the collection of indirect calorimetry data.

Statistical Analysis. All data are mean \pm SEM (except in Fig. 1A, in which data are reported as mean \pm SD for clarity). The overall time courses of variables changing with respect to time were compared with two-factor (group \times time) analysis of variance with repeated measures on time followed when appropriated (P -value < 0.05) by Tukey-Kramer *post hoc* multiple comparison tests for determining pairwise differences between groups. The influence of body mass or lean mass on energy expenditure was performed by using analysis of covariance (ANCOVA).^{31,32} Other variables were compared with paired or unpaired two-tailed Student's t tests. The significance level was set at $P < 0.05$.

RESULTS

Anatomical Changes And Body Energy Expenditure.

Mice treated with sActRIIB-Fc had a significant increase in body weight that began after a week of treatment (Fig. 1A). At the termination of the 8-week treatment period, body weight (Fig. 1A) and body lean mass (Fig. 1B) were larger (+32% and +33%, respectively) in animals treated with sActRIIB-Fc, whereas the body fat content did not differ between groups (Fig. 1C). As a result, the adiposity index was 14% lower in mice treated with sActRIIB-Fc (Fig. 1D). Treatment with sActRIIB-Fc also dramatically increased (+60%) the gastrocnemius

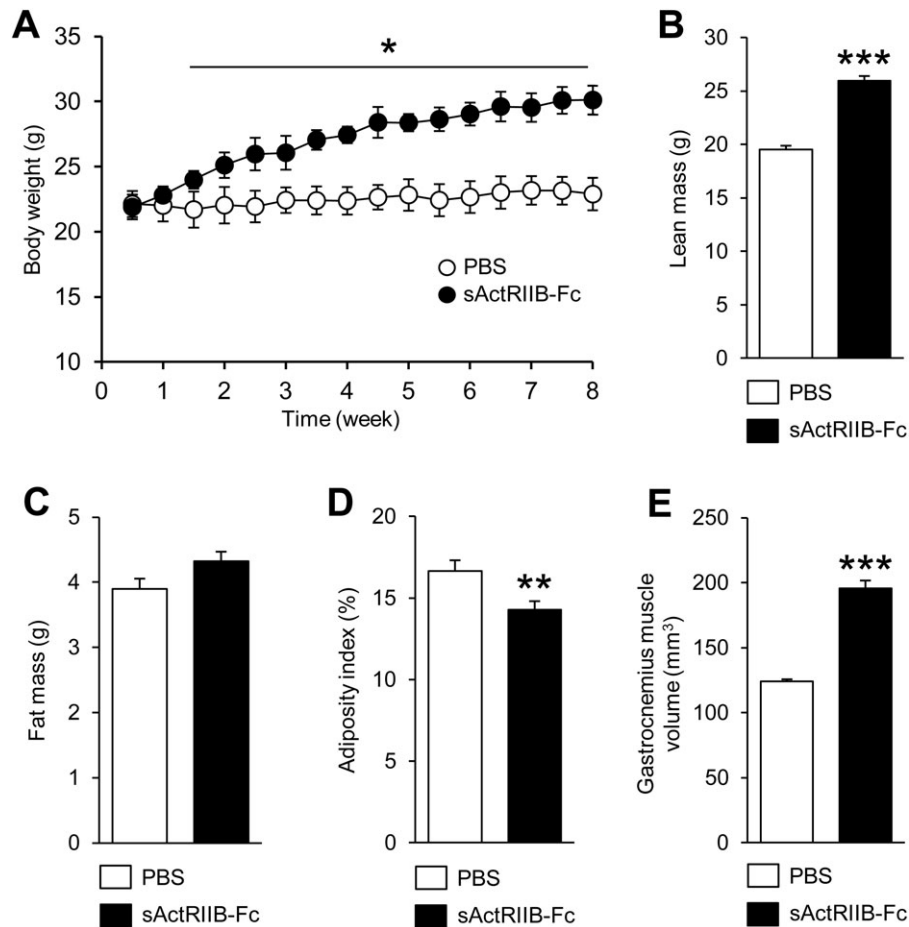


FIGURE 1. Physiological changes. Changes in body weight throughout the 8-week treatment with PBS or sActRIIB-Fc (**A**). Lean mass (**B**), fat mass (**C**), adiposity index (**D**), and gastrocnemius muscle volume (**E**) at the end of the 8-week treatment. All data are mean \pm SEM (except for **A** in which data are reported as mean \pm SD for clarity). * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$ vs. PBS-treated animals. PBS, phosphate-buffered saline; sActRIIB-Fc, soluble activin type IIB decoy receptor.

muscle volume (Fig. 1E). O_2 consumption (100 ± 4 ml/h, PBS; 111 ± 3 ml/h, sActRIIB-Fc), CO_2 release (78 ± 3 mL/h, PBS; 88 ± 3 ml/h sActRIIB-Fc), and energy expenditure (0.48 ± 0.02 kcal/h, PBS;

0.53 ± 0.02 kcal/h, sActRIIB-Fc) per animal were larger (+11%, +13% and +11%, respectively) in the group receiving sActRIIB-Fc. After adjustment for total body weight by ANCOVA (Fig. 2A), mice treated with

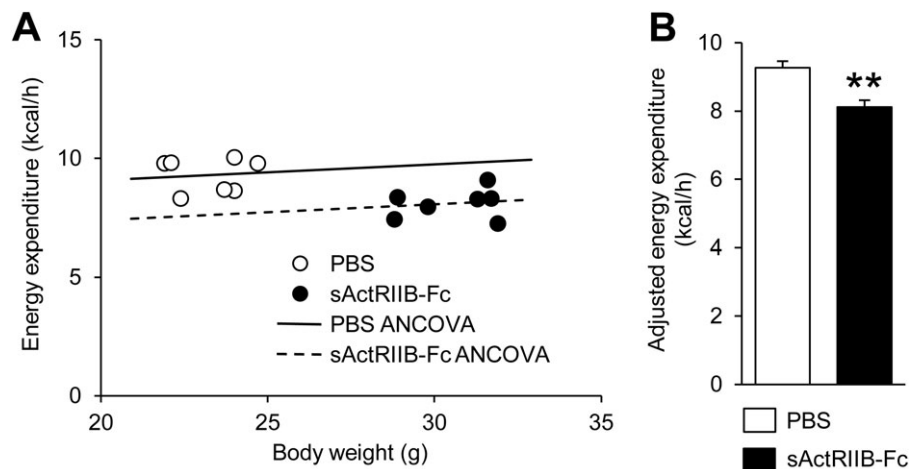


FIGURE 2. Adjustment of energy expenditure for body weight. Energy expenditure per animal during a 24-h period was plotted against body weight (**A**), and group comparison was adjusted for body weight by using ANCOVA (**B**). Data are mean \pm SEM. ** $P < 0.01$ vs. PBS-treated animals. ANCOVA, analysis of covariance; PBS, phosphate-buffered saline; sActRIIB-Fc, soluble activin type IIB decoy receptor.

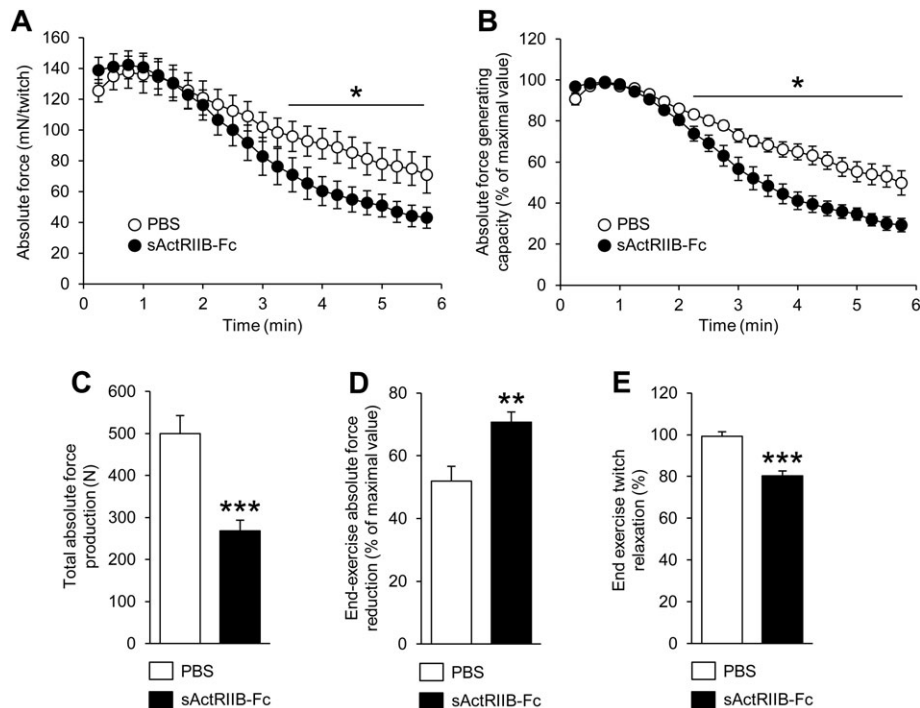


FIGURE 3. In vivo gastrocnemius muscle force-generating capacity. Absolute force (A) and absolute force-generating capacity (B) throughout the 6-min fatiguing bout of exercise performed *in vivo* simultaneously with ^{31}P -MR spectra acquisition. Total amount of absolute force produced during the whole exercise (C) and extent of force reduction (D) and muscle relaxation (E) measured at the end of exercise. Data are mean \pm SEM. * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$ vs. PBS-treated animals. ^{31}P -MR, 31-phosphorus; MR, magnetic resonance; PBS, phosphate-buffered saline; sActRIIB-Fc, soluble activin type IIB decoy receptor.

sActRIIB-Fc exhibited lower energy expenditure (Fig. 2B). Adjustment for lean mass produced similar differences (9.3 ± 0.2 kcal/h, PBS; 8.1 ± 0.2 kcal/h, sActRIIB-Fc). There were no differences between groups for respiratory quotient (0.78 ± 0.01 , PBS; 0.80 ± 0.02 , sActRIIB-Fc), horizontal activity (581 ± 92 arbitrary unit, PBS; 806 ± 190 arbitrary unit, sActRIIB-Fc), and rearing (110 ± 19 counts/h, PBS; 92 ± 15 counts/h, sActRIIB-Fc).

Gastrocnemius Muscle Function And Bioenergetics.

Treatment with sActRIIB-Fc increased by 22% the gastrocnemius muscle tension (122 ± 7 mN, PBS; 150 ± 7 mN, sActRIIB-Fc) produced during a single twitch performed before MR multimodal acquisition but did not affect the corresponding contraction (47 ± 2 ms, PBS; 50 ± 2 ms, sActRIIB-Fc) and half-relaxation (30 ± 2 ms, PBS; 35 ± 2 ms, sActRIIB-Fc) times. Changes in absolute force and force-generating capacity throughout the 6-min fatiguing bout of exercise are illustrated in Figure 3A,B. In mice treated with sActRIIB-Fc, total force produced during the whole exercise was 46% lower compared with the control group (Fig. 3C). Furthermore, the extent of force reduction at the end of exercise was 36% larger in sActRIIB-Fc-treated mice (Fig. 3D), indicating a lower resistance to fatigue. Twitch relaxation (i.e., the return to rest tension between consecutive twitches) was achieved in the early stage of exercise for both groups. At the end of

exercise, relaxation reached $99\% \pm 2\%$ and $80\% \pm 2\%$ in PBS- and sActRIIB-Fc-treated mice, respectively (Fig. 3E).

Mitochondrial oxidative capacity and metabolic fluxes were inferred from the *in vivo* changes in PCr

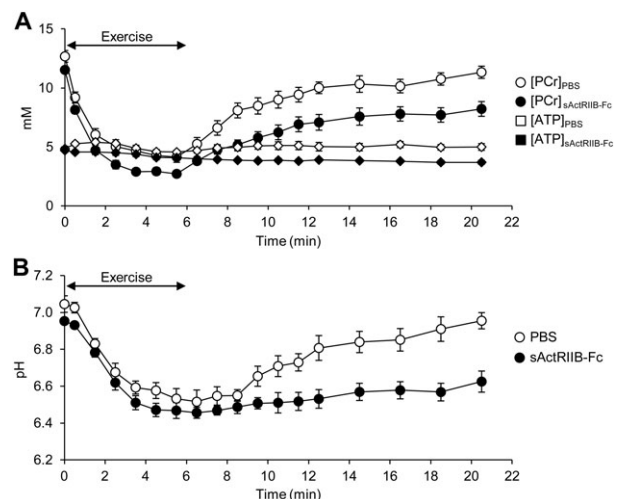


FIGURE 4. Noninvasive investigation of gastrocnemius muscle bioenergetics with dynamic ^{31}P -MR spectroscopy. Changes in PCr and ATP levels (A) and pH (B) were measured throughout the 6-min fatiguing bout of exercise and the 15-min postexercise recovery period. For each panel, the first time-point ($t = 0$) indicates the basal value. Data are mean \pm SEM. ATP, adenosine triphosphate; ^{31}P -MR, 31-phosphorus magnetic resonance; PBS, phosphate-buffered saline; PCr, phosphocreatine; sActRIIB-Fc, soluble activin type IIB decoy receptor

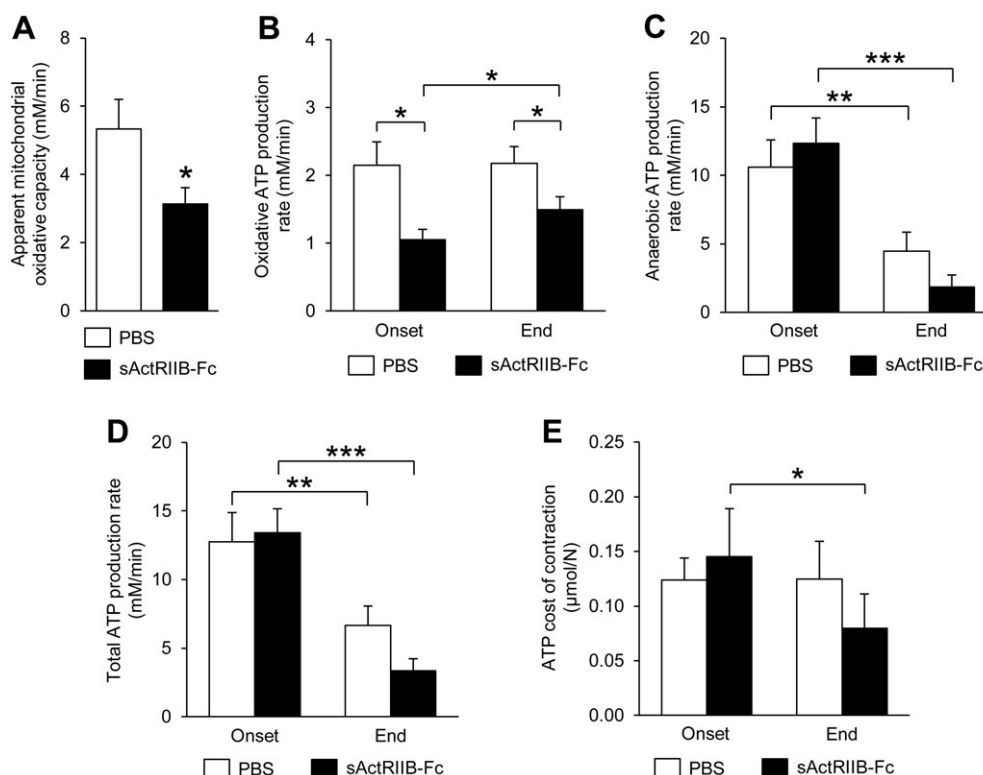


FIGURE 5. Metabolic fluxes and ATP cost of contraction. Apparent mitochondrial oxidative capacity (**A**), oxidative (**B**) and anaerobic (**C**) ATP production rates, total ATP production rate (**D**), and ATP cost of contraction (**E**) calculated at the start and at the end of the 6-min fatiguing bout of exercise. Data are mean \pm SEM. * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$ vs. PBS-treated animals. ATP, adenosine triphosphate; PBS, phosphate-buffered saline.

and ATP levels (Fig. 4A) and pH (Fig. 4B) throughout the rest, exercise, and postexercise recovery periods. The apparent mitochondrial oxidative capacity was 41% lower in mice treated with sActRIIB-Fc (Fig. 5A) and was associated with a reduction of the oxidative ATP production rate at the start (–51%) and at the end (–32%) of the exercise (Fig. 5B). In contrast, sActRIIB-Fc affected neither the anaerobic ATP production rate (Fig. 5C) nor the total ATP production rate (Fig. 5D) at the start and end of exercise. Overall, there were no differences between groups for the ATP cost of contraction at the start and at the end of the exercise (Fig. 5E). However, the ATP cost of contraction was decreased by 44% between the start and the end of the exercise in sActRIIB-Fc-treated mice, whereas it remained unchanged during this interval in PBS-treated mice (Fig. 5E).

DISCUSSION

The major findings from this study are that long-term ActRIIB blockade reduces the muscle apparent mitochondrial capacity *in vivo* and disturbs the basal body energy expenditure, whereas the voluntary locomotor activity remains unchanged. In exercising muscle, ActRIIB blockade enhances the development of fatigue and reduces the oxidative ATP production rate, but this reduction affects neither energy supply nor the ATP cost of contraction. An

additional important finding is that ActRIIB blockade impairs twitch relaxation in fatigued muscle.

In agreement with previously published reports, we found that ActRIIB blockade increases muscle strength and produces dramatic morphological changes, including muscle hypertrophy and increase in both lean mass and body weight, whereas body fat content remains unchanged.^{18,19,33,34} Just as we had expected, we found that the blockade strongly reduces the muscle apparent mitochondrial capacity *in vivo*, thereby corroborating previous *in vitro* studies in mice showing that biomarkers of oxidative function are impaired after postnatal ActRIIB inhibition^{3,13,14} and myostatin gene deficiency.^{21,35,36} Because skeletal muscle plays an important role in body energy homeostasis, our results suggest that long-term sActRIIB-Fc treatment affects basal energy expenditure, similarly to what has been reported in myostatin-null mice.^{37,38} We performed indirect calorimetry experiments to test this assumption and found that ActRIIB blockade increases basal oxygen consumption and energy expenditure per animal. Our findings differ from previous experiments showing that ActRIIB blockade has no effect upon basal VO_2 per mouse.¹⁸ However, this discrepancy could be linked to the duration of treatment (i.e., 8 weeks here vs. 4 weeks in the previous experiments);¹⁸ our results are closer to those obtained in myostatin-

deficient mice in which basal metabolic rate is increased relative to wild-type controls.³⁸ We found that this increase (11%) in energy expenditure was not proportional to the rise (32%–33%) in both body weight and lean mass. It is tempting to attribute this imbalance to mitochondrial dysfunction, but this phenomenon might also result from the differences in oxygen consumption among tissues. Basal oxygen consumption in skeletal muscle is much lower than the corresponding consumption in high-energy tissues such as brown adipose tissue and heart so that muscle hypertrophy is not expected to induce a substantial rise in resting energy consumption. Thus, after adjustment for body weight or lean mass by covariance analysis, we found that basal energy expenditure was reduced in animals treated with sActRIIB-Fc. This reduction occurred, however, without a corresponding change in voluntary ambulatory activity, which raises the possibility that ActRIIB blockade increases the animal's physical capacity through a reduction in the energy cost of ambulation. These considerations are in opposition to the reduction in muscle fatigue resistance evidenced here and in previous studies in mice treated with ActRIIB blocker.^{2,3} Nevertheless, the situation could be different in exercising muscle because of the large increase in energy demand.

To answer this question, we assessed ATP fluxes at the start (i.e., before fatigue development) and at the end of a fatiguing bout of exercise generating large changes in force production and energy demand. As an illustration, force reduction at the end of exercise reached 52% and 71% in PBS and sActRIIB-Fc groups, respectively. In mice treated with sActRIIB-Fc, we found that the oxidative ATP production rate was much lower at these 2 times, thereby providing evidence that the deleterious effect of ActRIIB blockade on mitochondrial function directly impacts bioenergetics in working muscle. However, this lower oxidative rate was compensated for by ATP production from anaerobic processes so that total energy production throughout the exercise was not disturbed in animals treated with sActRIIB-Fc. In addition, we found that ActRIIB blockade did not affect the ATP cost of contraction before and after the development of fatigue. Overall, our findings demonstrate that, despite impairing muscle oxidative function, ActRIIB blockade does not limit energy supply during a sustained exercise. However, the mouse gastrocnemius muscle is predominantly glycolytic, so the results of the present study might not be applicable to pure oxidative muscle. In any case, our results support the concept of upregulating oxidative metabolism as a complementary approach to ActRIIB blockade.¹⁷

We found that the contractile ATP cost decreased by 45% between the start and the end of the

fatiguing exercise in mice treated with sActRIIB-Fc, whereas it did not differ significantly during this interval in control mice. Shift in the pattern of fiber type recruitment and alteration of activation–relaxation processes have been proposed to explain the reduction of contractile ATP cost throughout a fatiguing exercise.^{39–41} Mouse gastrocnemius muscle is a mixed muscle composed mainly of fast glycolytic type-2b fibers (about 85%) with low oxidative capacity; the remaining muscle is composed of fast-oxidative type-2a fibers with higher oxidative capacity.^{9,42} Because a fiber is more fatigue resistant and contracts more economically when its metabolism is oxidative,^{43,44} change in muscle typology and/or preferential recruitment of type-2a fibers could explain the decrease in contractile ATP cost in fatigued muscle. This was unlikely in the present study for at least 2 reasons. First, we recently demonstrated that an 8-week treatment with sActRIIB-Fc does not affect fiber type distribution in the gastrocnemius muscle of wild-type mice.⁹ Second, our methodology allows stimulation of the whole gastrocnemius muscle,²⁵ and there is no reason that 1 type of fiber would be recruited more than the other in sActRIIB-Fc-treated animals compared with controls. On the other hand, we have observed in animals that have been treated with sActRIIB-Fc that twitch relaxation (i.e., the return to rest tension between consecutive twitches) was not fully achieved at the end of exercise, thereby indicating an alteration of calcium fluxes across the sarcoplasmic reticulum (SR) membrane. Such an alteration has been linked to fatigue development and/or structural anomalies at the SR level.^{45–47} It should be noted that the accumulation of tubular aggregates has been observed in the SR calcium release channel of myostatin-deficient mice, and this accumulation could alter calcium handling during excitation–contraction cycles.^{48,49} It is then conceivable that ActRIIB blockade produces similar accumulation of tubular aggregates, hence reducing calcium availability during excitation–contraction cycles. Because 20%–50% of the ATP consumed during muscle contraction is used for calcium pumping across the SR membrane,^{39,50} alteration in the activation–relaxation process might contribute to the reduction in contractile ATP cost in sActRIIB-Fc-treated animals.

In conclusion, these data demonstrate *in vivo* that, despite the decrease in oxidative capacity, muscle fatigue enhancement after ActRIIB treatment cannot be attributed to any limitation in energy supply and/or disturbance in contractile ATP cost. Additional investigations are required for determining the origin of this increased fatigability. This issue warrants consideration even if it can be argued that absolute increase in strength caused by ActRIIB blockade constitutes a functional advantage over the

increased fatigability. The possibility of attenuating or preventing muscle fatigability is an important clinical consideration for improving the quality of life of patients who are treated for muscle wasting and weakness.

We thank Acceleron Pharma Inc for the gift of sActRIIB-Fc; and Delphine Bastelica and Marie-Christine Alessi are both PhD (NORT, Aix-Marseille Université, INSERM U1062/INRA 1260, Marseille, France) for sharing the indirect calorimetry system.

Ethical Publication Statement: All authors have read the Journal's position on issues involved in ethical publication and affirm that this report is consistent with those guidelines.

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