

## Neurophysiology of gaze orientation: Core neuronal networks

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## Neurophysiology of gaze orientation:

## Core neuronal networks

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## Summary

The appearance of an object in the visual field triggers a shift of gaze toward its location. This orienting response consists of a rapid rotation of the eyes, the saccade, which can be accompanied by a rotation of the head. If the target moves, the saccade is followed by a slow movement of the eyes and a catch-up saccade. In this chapter, instead of describing the path leading from the target-evoked retinal activity to the changes in muscle tension, we shall take the reverse path. Starting from the muscle contractions, we shall proceed upstream and describe the tremendous organization that, in the brainstem and cerebellum, enables us to rapidly and accurately orient the foveae towards visual targets located at different eccentricities and depths. This chapter is an attempt to synthesize the considerable knowledge that neurophysiologists and neuroanatomists gathered in the monkey during the last six decades.

Wrapped within the orbits by fibrous fascia and fat cushions, the eyeballs owe their orientation to the tensions that six extraocular muscles (EOM) exert upon them. Their steady tension acts as a kind of basal tone on top of which the orientation of the eyes can change when the balance of activity carried by their innervation also changes. Whenever modifications in the firing rate of motor neurons cause asymmetrical changes in the contraction of extraocular muscle fibers, the eye starts to move.

The figure 1A shows the approximate location of extra-ocular muscles. Four rectus muscles attach to the sclera anteriorly with respect to the eyeball equator whereas the oblique muscles insert posteriorly. The insertions of the lateral (LR) and medial (MR) rectus muscles are located on opposite sides of each eyeball, making their primary actions antagonist with each other. Although there is no rigid axis about which the eyes rotate, their movements are, by convention, described as rotations about three virtual axes: a vertical axis for horizontal (leftward and rightward) movements, a horizontal axis for vertical (upward and downward) movements and an anteroposterior axis for torsional movements (intorsion and extorsion). Kinematic studies in healthy subjects tested with the head upright have measured the orientation of eyeballs and found that two rotation axes suffice to describe their changes. These axes lie within a plane called Listing's plane, which is perpendicular to the anteroposterior axis of the eyeball. Some rare subjects are able to make voluntary torsional eye movements. However, in most cases, such movements are observed in case of particular brain lesions or when healthy subjects tilt their head.

#### Figure 1 about here

With the head upright and gaze directed straight ahead, the contraction of LR fibers and the relaxation of MR fibers causes the abduction of the eyeball while the contraction of MR fibers and the relaxation of LR fibers of the other eye causes its adduction. If the tensions exerted by the other muscles do not change, gaze direction is expected to move only along a plane orthogonal to the head's sagittal plane because, unlike the other four muscles, the LR and MR muscles exert no secondary action. In fact, precise measurements made in human subjects reveal a transient torsion during horizontal saccades, indicating changes in the activation state of the other muscles.

The situation is more complicated when gaze moves vertically. Vertical movements involve not only steady tension of LR and MR muscles, but also a complex cooperation between the four other muscles. The contraction of superior and inferior rectus muscles elevate and depress each eye, respectively. These actions are stronger when the eye is in abduction. The contraction of oblique muscles causes torsional movements but also elevate or depress the eye, more strongly when the eye is in adduction. If we consider the movements made when the eyes are centered in the orbit, in addition to elevating the optic axis (turquoise arrows in Fig. 1B), the contraction of the superior rectus (SR) muscle fibers causes adduction (black arrows) and intorsion (blue arrows) of the eyeball. Likewise, in addition to lowering the optic axis (yellow arrows in Fig. 1B), the contraction of the inferior rectus (IR) muscle fibers causes adduction and extorsion (green arrows). These secondary actions can be counteracted by the contraction of oblique muscles, for the contraction of inferior oblique (IO) muscle fibers causes elevation, abduction and extorsion of the eyeball, whereas the contraction of superior oblique (SO) muscle fibers causes depression, abduction and intorsion.

Binocular conjugate movements are made when the head rotates while a visual target is being fixated (vestibulo-ocular reflex), when the visual surround is moving (optokinetic reflex) and whenever a target appears or is imagined in the peripheral visual field (saccades). We shall first examine the neurophysiology of saccade generation when the head does not move. Then, we shall study the case when a head rotation accompanies the saccade. During these combined eye-head movements, two couplings can happen: the eyes and the head move either in the same direction or in opposite directions. Next, we shall see the neurophysiology of disconjugate movements of the eyes, which occur when gaze is shifted between targets located at different depths (vergence eye movements). Finally, we shall have an overview of the complex network that enables one to fixate and track a moving target, i.e., an object whose spatial coordinates keep changing. (smooth pursuit eye movements).

### Binocular conjugate movements

#### Horizontal saccades

Figure 2 schematizes the current view of the premotor neurophysiology of horizontal saccades in the pontine and mesencephalic reticular formation. During leftward saccades (Fig. 2A), the fast-twitch muscle fibers of the right eye's MR and of the left eye's LR contract while

those of antagonist muscles (right eye's LR and left eye's MR) relax. Motor neurons in the abducens nucleus (ABD, located in the pontine reticular formation) and in the oculomotor nucleus (OMN in the midbrain tegmentum) emit bursts of action potentials that respectively cause the phasic contraction of muscle fibers. The burst discharge of cells in the OMN is triggered and supported by spikes emitted by internuclear neurons (AIN) also located in the abducens nucleus. In return, the bursts of abducens motor (MN) and internuclear (AIN) neurons are driven by afferent input from excitatory burst neurons (EBN) located in the ipsilateral paramedian pontine reticular formation (PPRF, synapses a). The abducens neurons also receive excitatory input from internuclear burst-tonic neurons in the contralateral oculomotor nucleus (not shown). Discharging during saccades and increasing their firing rate during convergence, these cells are possibly involved in the motor drive for conjugate saccades because in addition to cause exophoria, lidocaine injection in the medial rectus subdivision of the oculomotor nucleus causes hypometria and slowing of abducting saccades of the contralateral eye.

Otherwise silent, the EBN fire trains of action potentials whose interspike interval is smaller for larger saccades. Contrary to statements found in numerous review articles, the instantaneous firing rate of EBN is not a replica of instantaneous eye velocity. For each saccade, the interspike interval is much more constant than expected if this claim were true (i.e, that EBN rate equals instantaneous eye velocity). The suggestion that the firing rate of motor neurons would alone determine the instantaneous eye velocity is also brought into question by a study in the cat, which showed that a model factoring in muscle tension and its first derivative accounted for the firing rate of motor neurons better than a model factoring in eye position and its first and second derivatives (velocity and acceleration).

Regarding the relaxation of antagonist muscles, it results from a pause in the firing of motor neurons innervating them. This pause is itself caused by inhibitory input from burst neurons (IBN) located in the ipsilateral dorsal paragigantocellular reticular formation (dPGRF, synapses b). In addition to bursting during ipsiversive horizontal saccades (so called "on-direction" response), approximately half of these inhibitory burst neurons also emit spikes during contralateral horizontal saccades (off-direction response). Thus, the agonist motor drive is a combination of excitatory and inhibitory presynaptic input from the ipsilateral EBN and contralateral IBN, respectively. The relative strength of these antagonist premotor inputs

is under the descending influence of saccade-related burst neurons located in regions involved in other functions, such as the left and right caudal fastigial nuclei (located in the medial cerebellum) and the contralateral deep superior colliculus (in the midbrain tectum). The bilateral fastigial input exerts an influence that is more modulatory than the collicular influence because, contrary to lesions in the superior colliculus, fastigial lesions do not suppress or delay the generation of saccades.

#### Figure 2 about here

During rightward saccades (Fig. 2B), the active circuit is mirror reversed. The agonist muscles are the right eye's LR and the left eye's MR, whereas the antagonist muscles are the right eye's MR and the left eye's LR. IBN in the right dPGRF cause the relaxation of antagonist muscles. Under the excitatory influence of EBN in the right PPRF and the inhibitory influence of IBN in the left dPGRF, motor and internuclear neurons in the right abducens nucleus provide the motor commands to the ipsilateral LR and contralateral MR, respectively. The circuits producing rightward and leftward saccades are presumably not completely symmetric insofar as the number of active neurons and the number of synapses and neuromuscular junctions likely differ between them. By contrast, we shall see next that the networks involved in the generation of downward and upward eye movements are not at all symmetrical with respect to each other.

#### Vertical cardinal saccades

The neuronal network underlying the generation of vertical saccades is located in the midbrain tegmentum. For upward saccades (Fig. 3A), the agonist muscles are the IO and the SR and the antagonist muscles the IR and the SO. As reported above, in addition to elevating the eye, the contraction of SR muscle fibers causes intorsion, whereas contraction of IO muscle fibers causes extorsion. Thus, the intorsion caused by the motor commands to the SR combines with the extorsion caused by the commands to the IO. The observation of negligible torsional component during upward saccades suggests that the two torsions cancel each other out. Such a balance of force likely involves bilateral activity in the midbrain because for each eye, the motor neurons innervating the SR muscle are located in the contralateral OMN whereas the motor neurons innervating the IO muscle are located in the ipsilateral OMN. The

two sets of commands during upward saccades must be adjusted in a timely manner, and any asymmetry in the bilateral motor drive would engender a torsional component.

#### Figure 3 about here

The excitatory commands responsible for generating upward saccades are issued by burst neurons located in the left and right rostral interstitial nucleus of the medial longitudinal fasciculus (RIMLF). These commands are doubly bilateral. Indeed, on the one hand, the burst emitted by premotor neurons (uEBN) in the left RIMLF excite the motoneurons innervating the SR muscles of *both* eyes (black synapses a) and motor neurons innervating the IO muscles of *both* eyes as well (black synapses b in Fig. 3). On the other hand, the burst emitted by uEBNs in the opposite RIMLF excite the same motoneuronal group (blue synapses a and b). Moreover, in addition to recruiting the motor neurons, the uEBN also excite neurons that inhibit the motor neurons that innervate the antagonist muscles (IR and SO, synapses c). Located in the interstitial nucleus of Cajal, these cells (uIBN) inhibit the motor neurons in the contralateral oculomotor nucleus, i.e., cells responsible for the contraction of the IR of the contralateral eye (synapses d) and motor neurons responsible for the contraction of the SO of the ipsilateral eye (synapses e). However, contrary to uEBN, uIBN do not send bilateral projections to the motor neurons innervating the SO muscle fibers. Their projection make synaptic contacts only with motor neurons on the contralateral side.

As for the upward saccades, the discharge of neurons located in each side of the brain is capable of causing a downward movement of *both* eyes (Fig. 3B). The premotor excitatory burst neurons leading to downward saccades (dEBN) are also located in the RIMLF. However, unlike the uEBN, the dEBN do not send bilateral projections to the motoneurons; their projection is ipsilateral only. During downward saccades, the SO and IR muscle fibers of *both* eyes contract because the dEBN excite 1) the motor neurons in the ipsilateral OMN, which cause the contraction of IR muscle fibers of the *ipsilateral* eye (synapses a), and 2) the motor neurons which, in the ipsilateral trochlear nucleus, cause the contraction of SO muscle fibers of the *contralateral* eye (synapses b). Moreover, in addition to lowering the eyes, the contraction of SO muscle fibers causes an intorsion of the eye while the contraction of IR muscle fibers causes their extorsion. The commands from the ipsilateral OMN for extorsion combine with commands from the contralateral trochlear nucleus for intorsion. Precisely how

these opposite torsional components cancel each other out during downward saccades remains unexplained. A bilateral adjustment at the level of premotor burst neurons seems inevitable. Moreover, the neural processes in charge of the relaxation of antagonist muscles during downward saccades remain to be characterized.

Even though the number of cells and the number of synaptic boutons likely differ between the left and right sides of the reticular formation, the connectivity patterns involved in generating leftward and rightward eye movements are symmetrical with respect to the brainstem midsagittal plane (see Fig. 2 and 5). By contrast, the networks involved in the generation of downward and upward eye movements are not symmetrical with respect to the midsagittal plane (Fig 3).

During vertical saccades, the EBN involved in the generation of horizontal saccades do not fire. Some IBN in the dPGRF emit action potentials whose number increases with the saccade size. For vertical saccades to remain straight while their size increases, one possibility is that the motor and internuclear cells in the abducens nucleus emit additional spikes, thus reducing the contrapulsive effect of IBN spikes. This does not seem to be the case because MN and AIN in the abducens nucleus do not emit action potentials during vertical saccades. A more likely possibility is that the contrapulsion exerted by IBN in the left and right dPGRF cancel each other out. If no perfect symmetry exists between the set of active neural elements (number of neurons and of synaptic boutons) located on each part of the reticular formation, then the observation of strictly vertical saccades leads us to infer that a process adjusts the bilateral activity of IBN during vertical saccades.

This bilateral control is not restricted to the group of IBNs, which are primarily involved in the generation of horizontal saccades. In addition to elevating both eyes, the discharge of motor neurons in the right OMN causes, as a secondary action and via the contraction of the right eye's IO and the left eye's SR, a rightward deflection of both eyes during vertical saccades while the discharge of motor neurons in the opposite side causes a leftward deflection (Fig. 3A, top row). Likewise, in addition to lowering both eyes, the dEBN in the right RIMLF lead to the contraction of the left eye's SO and the right eye's IR, causing a leftward deflection of their trajectory while dEBN in the left RIMLF cause a rightward deflection during vertical saccades (Fig. 3B, top row). The observation of straight vertical saccades (with no horizontal deviation)

implies that the horizontal deflections caused by the secondary action of the muscles elevating or depressing the eyes are also canceled in the motor drives.

Therefore, the generation of straight vertical saccades rests upon a balance of bilateral motor activation, which itself depends upon an adjustment of premotor commands that takes into account the structural (neurons and their connectivity) and functional (firing properties, secondary and tertiary consequence of muscle contractions) asymmetries between the oculomotor territories that are distributed on either side of the brain stem. The saccade-related neurons in the left and right fastigial nuclei and in the oculomotor vermis regulate this bilateral balance. Indeed, most of them burst during vertical saccades and their asymmetrical perturbation horizontally deflects the trajectory of vertical saccades. Bilateral activity in the two deep superior colliculi is also involved with their medial halves for driving upward saccades and their lateral halves driving downward saccades. The contribution of the deep superior colliculi and of the caudal fastigial nuclei differ insofar as horizontal saccades involve activity of bursting cells in only the superior colliculus on the contralateral side, whereas saccade-related neurons burst in the caudal fastigial nuclei on both sides.

#### Combined horizontal and vertical saccades

When a target appears off the cardinal visual axes, the orienting response does not consist of horizontal and vertical saccades made in sequential order. Both horizontal and vertical components are generated in such a manner that their onset and offset are relatively simultaneous. These combined horizontal-vertical saccades are conventionally called "oblique" saccades. Considering the muscle diversity (lengths, contractile properties and orbital attachments) involved in the horizontal and vertical components and the fact that their motor innervations are not located in nearby regions of the brainstem, their simultaneous onsets and the limited curvature of their spatial trajectory are empirical observations suggesting coupling mechanisms.

In the medial pontine reticular formation, the nucleus raphe interpositus (RIP) hosts a population of cells that exhibit a sustained firing rate during intersaccadic intervals but pauses during all saccades, regardless of their direction and amplitude. These cells are called omnidirectional pause (or omnipause) neurons (OPN) and are thought to prevent the eyes from moving during visual fixation through their inhibitory post-synaptic influence upon the

premotor burst neurons (Fig. 2). Moreover, with their pause, OPN could also be responsible for the synchronous onsets of horizontal and vertical saccades, as they project to burst neurons in both the horizontal and vertical gaze centers. So far, however, experimental studies have failed to document onset asynchronies during inactivation or lesion of RIP. No fixation instability was reported either. The absence of deficits in visual fixation may have resulted from the particular testing conditions. Indeed, the consequence of suppressing OPN sustained activity during delayed or memory-guided saccade tasks was not tested. Indeed, the observation of irrepressible saccades to the peripheral target during the delay interval would unambiguously confirm the role of OPN in visual fixation; a result that electrical microstimulation studies have suggested. Unfortunately, these stimulation studies are not sufficiently conclusive because it is possible that the interruption and the delaying of saccades by electrical stimulation result from undesired retrograde recruitment of inhibitory burst neurons.

Turning back to the generation of combined horizontal and vertical saccades, one study reported crucial observations signaling a process that couples their time course during oblique saccades. When the horizontal component is experimentally slowed by the local injection of a pharmacological agent in the pontine reticular formation, the vertical component was also slowed. In some of these experiments, the duration of both components was prolonged and the accuracy of saccades was preserved, supporting further the persistence and preservation of the target-related commands. The neural connectivity underlying this "component stretching mechanism" have not yet been elucidated.

## Combined eye and head movements

Saccades are frequently associated with a head movement when the target eccentricity approaches and exceeds the limits of the oculomotor range. Moving the head enable to look at targets located more eccentrically in the visual field, and to re-centers the eyes in their orbit. The directions of the eyes in the orbit and of the head relative to the trunk add together to determine the direction of gaze.

Two functional modes characterize orienting gaze movements toward a static target: a mode during which both eyes and the head rotate in the same direction and a mode during which they move in opposite directions. The second mode happens sometimes when the head

movement precedes the eye saccade, mostly when the target location is known or foreseeable. More frequently, it happens later because the eyes move much faster than the head. The equilibrium of bilateral activity that characterizes visual fixation has been reached before the head has terminated its movement. Before describing how gaze is stabilized at the end of orienting movements, we shall examine the first mode, i.e., how despite the vestibulo-ocular reflex, the eyes and the head can move in the same direction during horizontal gaze shifts (Figure 4).

#### Combining a head movement with a saccade

In this section, we limit our description to the horizontal eye movements that are made while the head also moves horizontally. Whether the head is moving or not, behavioral studies show that the gaze movements are equally accurate. In the monkey, around twenty muscles are involved to change the orientation of the head. Among them are the muscles whose contraction changes the orientation of the head relative to the trunk, the muscles that stabilize the head during its rotation and the muscles that do not exhibit any change. The cocontraction of the left and right rectus capitis posterior minor (RCPm) stabilize the atlas while the contraction of the rectus capitis posterior major (RCPM) and obliquus capitis inferior (OCI) produce an ipsilateral horizontal rotation of the head. The observation that the contralateral muscles exhibit no electromyographic change indicates that stopping the head rotation is not caused by muscle co-contraction. For movement amplitudes greater than 20 degrees, the contraction of the splenius capitis muscle adds to that of the RCPM and OCI muscles. At the end of the orienting movement, the co-contraction of the RCPM, OCI and obliquus capitis superior (OCS) muscles altogether stabilizes the head.

Thus, orienting gaze shifts mobilize several muscles in parallel when a movement of the head accompanies the saccade. The synergy results from activities distributed within a complex set of motor and premotor neurons located in the reticular and vestibular nuclei. Contrary those involved in the cat, the premotor processes remain incompletely characterized in the monkey. Contemporary knowledge is not sufficient to reach a synthetic understanding that is comparable to the neurophysiology of saccades.

The recruitment of EBN and reticulo-spinal neurons (RSN) by descending commands from bursting cells in the deep superior colliculus (grey-colored synapses a in Fig. 4) leads to

the excitation of the pools of extraocular motoneurons (ABD) in the abducens nucleus and neck motoneurons (NMN) in the cervical spinal cord (green and blue synapses b), and thus, to the contraction of the agonist extraocular and neck muscle fibers, respectively. Contrary to the cat, collicular connections with EBN do not seem to be direct in the monkey. The EBN and RSN also excite the IBN (synapse a'), thereby suppressing the activity of cells that otherwise act to facilitate the contraction of antagonist extraocular (synapses c) and neck muscles (synapse g), respectively. Finally, they also excite secondary vestibular neurons which are inhibitory (IVN-2; blue and green colored synapses d) and located in the ipsilateral medial vestibular nucleus. Within the vestibular nuclei, the discharge of IVN-2 inhibits the secondary vestibular neurons, IVN-1 and EVN-1 (purple-colored synapse e). By inhibiting IVN-1, the motor neurons innervating the agonist extraocular and neck muscles fibers (red synapses f and f') are disinhibited, and the contraction of these muscles is facilitated. By inhibiting the EVN-1, the IVN-2 prevent the primary vestibular neurons (PVN), whose firing is enhanced by the head movement, from causing a counter-rotation of the eyes in the orbits (synapse h). The contraction of antagonist neck muscles through vestibulospinal pathways is also prevented (synapse h').

#### Figure 4 about here

#### Stopping gaze while the head terminates its motion

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The second mode of combined eye head movements is reminiscent of the vestibuloocular reflex recorded when the head is passively rotated while subjects maintain their gaze
directed toward a visual target. During active eye and head movements, the premotor input
to motor neurons makes a transition from commands fed by target-related (visual or auditory)
signals to commands fed by head movement-related signals. The latter consist of trains of
action potentials that originate in the three semicircular canals buried within the temporal
bone. Each semicircular canal consists of a ring within which a fluid (endolymph) flows and
mechanically deflects the stereocilia of multiple cells whenever the head rotation accelerates
or decelerates. This deflection either increases or decreases the discharge of primary
vestibular neurons (PVN) with a rate proportional to head velocity.

The two lateral canals are located in the left and right inner ears and lie roughly in a "plane" which is tilted not only relative to the Reid plane, but also to the "plane" passing

through the medial and lateral rectus muscles. These canals are often called "horizontal" because they are primarily activated during head rotations about the Earth gravity axis. During such rotations, PVN on the side toward which the nose is rotating (ipsilateral side) increase their firing rate while those on the contralateral side decrease their firing rate. These bilateral changes modify the firing rate of post-synaptic neurons (secondary vestibular neurons) located in the medial and superior vestibular nuclei. On either side of the brain's midsagittal plane, these nuclei are bilaterally organized and house two categories of neurons: type 1 and type 2 neurons. Type 1 neurons increase their firing rate during head rotations toward the side (left or right) containing the nucleus in which their discharge is recorded. During contraversive head rotations, they decrease their discharge. In addition, they pause during saccades. In the monkey, these neurons have been called Position Vestibular Pause (PVP) neurons. Type 2 neurons exhibit the opposite pattern, increasing their firing rate during contraversive head rotations and decreasing it during ipsiversive head rotations.

#### Figure 5 about here

Figure 5 schematizes the mechanism by which vestibular signals contribute to ending a leftward movement of gaze toward a visual target. The end of the orienting movement is characterized by a time interval during which the gaze direction remains stationary relative to the trunk while the head slowly terminates its motion. The saccades by both eyes give way to their rotation in the opposite direction. As the head rotates, the primary vestibular neurons (PVN) excite the type I vestibular neurons (EVN-1) in the ipsilateral medial and ventrolateral vestibular nuclei. In turn, the EVN-1 initiate and drive the counter-rotation of both eyes in the orbit by exciting the motor (LRMN) and internuclear (AIN) neurons in the contralateral abducens nucleus (red-colored synapses a). Thus, the abducens neurons can resume their activity from the pause imposed by the inhibitory input from IBN in the left Med RF (see bluecolored synapses b in Fig. 2). The LR muscle fibers of the right eye can contract, as well as the MR muscle fibers of the left eye, which are activated via motoneurons located in the OMN. The latter cells (MRMN) are also excited by neurons in the ipsilateral nucleus of the ascending tract of Deiters (ATDN; green-colored synapse b). In parallel, the spikes emitted by other type 1 vestibular neurons (IVN-1) choke the activity the LRMN and AIN in the abducens nucleus (red-colored synapses c), preventing them from responding to any residual descending (target-related) commands. The recruitment of type 2 neurons (IVN-2) in the contralateral vestibular nuclei (red-colored synapse d) complements the cessation of agonist commands by inhibiting the activity of EVN-1, thereby reducing their excitatory influence upon the motor and internuclear neurons (synapses e). IVN-2 also inhibit the activity of IVN-1, which in turn favors the disinhibition of cells (synapses e) in charge of the counter-rotation of the eyes under the influence of EVN-1 (synapses a).

The functional anatomy schematized in Fig. 5 enables us to see how the disinhibition of type 1 vestibular neurons can quickly arrest the eye saccade during ipsilateral orienting movements. The spikes of IVN-1 (inhibitory synapses c) choke the burst commands while the spikes of EVN-1 order the eyes to counter-rotate as the head terminates its motion (excitatory synapses a). This mechanism also helps us to understand the hypometria of contralateral eye and head movements after unilateral lesion of the caudal fastigial nucleus (for instance the right nucleus). Contralateral (leftward) saccades are hypometric because the agonist drive from EBN lacks of the excitatory input from inactivated (right) fastigial cells, and because the saccade-related burst of neurons in the opposite (unaffected) fastigial nucleus excite IBN in the contralateral (right) reticular formation. These IBN inhibit IVN-2 in the opposite (left) vestibular nuclei, disinhibiting the excitatory influence of EVN-1 (initiating the counterrotation of the eyes) and the inhibitory influence of IVN-1 upon ipsilateral motor neurons (inhibiting the saccade-related drive). The latter mechanism also explains the hypometria of head movements. Their amplitude is truncated because the IVN-1 inhibits the ipsilateral neck motor neurons (NMN; synapse e) while the EVN-1 facilitates the contraction of contralateral neck muscles (synapse f).

#### Figure 6 about here

As for the generation of upward and downward saccades (Fig. 3), the neural connectivity involved in the generation of downward and upward slow eye movements are not symmetrical. Figure 6A schematizes the network that stabilizes gaze during downward gaze movements. While the head pitches down and slowly terminates its motion, both eyes rotate upward in the orbits. The upward eye movement is the outcome of contracting SR and IO muscles fibers. Driven by the increased firing of PVN connected with the left anterior canal, excitatory vestibular neurons (EVN-U) in the left medial vestibular nucleus activate the motor neurons innervating the left eye's SR and the right eye's IO muscle fibers (synapses a).

In addition to elevating both eyes, the contraction of these fibers causes an intorsion of the left eye and an extorsion of the right eye, as if both eyes were counteracting a tilt of the head toward the left shoulder. This clockwise torsion of both eyes can be attenuated by the torsion caused by the excitatory input from EVN-U in the right MVN. Because of their excitation by the right anterior canal, PVN drive the firing rate of EVN-U, which in turn excite the motor neurons that innervate the right eye's SR and the left eye's IO muscle fibers (synapses b), causing the right eye's intorsion and left eye's extorsion. The absence of torsion during strictly upward slow eye movements suggests that the output of the left and right vestibulo-oculomotor channels have been adjusted so that the torsional signals balance out.

Regarding the antagonist muscles, their inhibition is promoted by IVN-U located in the superior vestibular nuclei. These cells inhibit the motor neurons innervating the IR of the ipsilateral eye (synapses c) and those innervating the SO muscle of the contralateral eye (synapse d). Contrary to the network involved in horizontal eye and head movements (Fig. 5), the neural mechanisms triggering the counter-rotation of the eyes when the head pitches down are not yet elucidated. However, the inputs are necessarily bilateral and balanced since the eyes move by the same amount and their torsion canceled.

Finally, figure 6B schematizes the neuronal network involved in stopping an upward gaze shift while the head slowly terminates its upward motion. Both eyes rotate downward in the orbits because of the contraction of IR and SO muscle fibers. This is driven by the increased firing of primary vestibular neurons (PVN) connected with the left posterior canal. Excitatory vestibular neurons (EVN-D) in the left medial vestibular and ventral lateral vestibular nuclei activate the motor neurons innervating the fibers of the right eye's IR and the left eye's SO muscles (synapses a).

As above, in addition to depressing both eyes, the contraction of these fibers cause an extorsion of the right eye and an intorsion of the left eye, as if the eyes are counteracting a tilt of the head toward the right shoulder. Once again, the binocular torsion is attenuated by a torsion made in the opposite direction and caused by excitatory input from the right posterior canal to the motor neurons innervating the left eye's IR and the right eye's SO muscle fibers (synapses b). Inhibitory vestibular neurons (IVN-D) located in the superior vestibular nuclei prevent the antagonist muscles from contracting by inhibiting the motor neurons innervating

the SR of the contralateral eye (synapses c) and possibly also the motor neurons innervating the IO muscle of the ipsilateral eye (synapse d). As for the neural mechanisms ending downward gaze shifts, the neural mechanisms triggering the counter-rotation of the eyes when the head pitches up are not completely identified. Nevertheless, if the eyes do not exhibit any torsional component, the motor commands are necessarily bilateral and balanced.

In summary, by studying the networks underlying the generation of slow eye movements, we discover a complexity that radically differs from the straightforward solutions that human engineers would have designed. One puzzling question concerns the constraints that led the torsional components caused by the contraction of elevator or depressor muscles to cancel each other out. Moreover, the connectivity that we have described is not complete, because new eye movements can be produced in response to "unusual" combinations of head rotations and visual stimulation. Experiments in the cat have shown that the visual stimulation exerts a significant influence on the functional connectivity yielding vestibulo-oculomotor responses. Horizontal compensatory eye movements can be generated during vertical head rotations in the dark after training sessions during which the visual field moved horizontally whenever the head rotated vertically.

### Fixating a static visual target

Considering the line of sight as the direction of gaze measured when a subject fixates a visual target, the binocular fixation point designates the intersection between the two lines of sight. Of course, the quantification of movements must be distinguished from their neurophysiological correspondence. On the retinal side, looking at a visual target obviously does not reduce to the excitation of one single photoreceptor on the fovea of each eye. On the oculomotor side, the orientation of the eyes is maintained steady thanks to the discharge of a tonic neurons distributed in the nucleus prepositus hypoglossi (NPH), the vestibular nuclei (in the medullary reticular formation) and in the interstitial nucleus of Cajal. The more deviated the eyes in the orbit, the higher their firing rate. In addition to connective tissues that stabilize each eyeball relative to the orbit, two neuronal networks are involved in maintaining the rotation axes of the eyes within Listing's plane and deviating them horizontally and vertically. These networks are relatively independent from each other insofar as a lesion in the NPH primarily affects the ability to maintain gaze direction within the

horizontal plane, with marginal effects on vertical gaze direction. After the saccades, whose amplitude is unaffected, the eyes exhibit an exponential drift toward their primary orientation (roughly straight ahead). By contrast, lesion in the NIC causes a failure to hold vertical deviations of the eyes (i.e. both upward and downward) but not horizontal deviations. A torsional drift of the eyes is also observed when the lesion is unilateral.

Considering now the case when the foveation changes between targets or locations situated at different depths in the three-dimensional physical space, three visuomotor processes occur: 1) vergence, 2) accommodation and 3) variation of pupil diameter. Convergence designates the symmetrical eye movements made when the MR muscle fibers of both eyes contract, diminishing the distance of the binocular fixation zone relative to the head. Divergence designates those movements made when the distance increases because of a binocular relaxation of MR muscle fibers. The contraction level of intraocular muscles also changes. During convergence, the contraction of ciliary muscles (CM) reduces the tension in zonular fibers, changing the refractive power of the lens as its anterior surface bulges forward (increase in convexity). Thus, the target images are brought into focus on the foveae. This accommodation is associated with a myosis, i.e., the constriction of the iris and the reduction of the pupil diameter, thereby increasing the depth of field. The myosis itself results from the contraction of the sphincter pupillae (SP) combined with the relaxation of the dilator pupillae (DP). Altogether, these three visuomotor processes constitute the so-called the near response (or near triad).

Although these processes are generated jointly, clinical studies report pathological cases in which they are dissociated and the neuroanatomy helps us to understand why. Figure 7 shows that the contraction of MR muscle fibers is controlled by the firing rate of motor neurons (MRMN) located in the oculomotor nucleus whereas the intrinsic musculature of each eye is controlled by motor neurons located in the ciliary ganglions (CG) and premotor neurons in the Edinger-Westphal preganglionic nucleus (EWpg). In both cases, the innervation is ipsilateral. The binocular control and the near triad starts with their afferent input from two major regions of the midbrain tegmentum: the supraoculomotor area (SOA) and the central mesencephalic reticular formation (cMRF).

The SOA is primarily involved in the generation of the slow near response and its sustained maintenance. Therein, we find burst neurons and tonic neurons, with separate subgroups characterized by different discharge patterns during the generation of diverging and converging eye movements. More common just lateral to the MLF, four categories of neurons are found: convergence burst neurons, divergence burst neurons, convergence tonic neurons and divergence tonic neurons. Whether divergence neurons exert an inhibitory influence upon the firing rate of convergence neurons is not known yet. Neurons in SOA send projections to the medial rectus MN that are ipsilateral, whereas the projections to the EWpg are bilateral.

In the cMRF, we find neurons that emit a burst discharge whenever the near response accompanies a saccadic eye movement: saccade convergence burst neurons and saccade divergence burst neurons. The former fire during asymmetrical saccades associated with an increase of the vergence angle and the latter fire during asymmetrical saccades associated with a divergence. Each cMRF innervates the SOA and the EWpg bilaterally. It also sends ipsilateral projections to the sites in the OMN in which the motor neurons that innervate the singly-innervated muscle fibers are located.

The bilateral projections from SOA to the EWpg, and from the cMRF to both EWpg and SOA, are completed by bilateral afferents from caudal fastigial nuclei (CFN) whose contribution in converging eye movements has been reported by unit recording and lesion studies. In addition to send bilateral projections to the midbrain tegmentum and contralateral projections to the pontomedullary reticular formation (where EBN and IBN are located, Fig. 2), the CFN also project bilaterally to the rostral part of the deep superior colliculi, with a contralateral predominance. Therein, some cells either increase (convergence neurons) or decrease (divergence neurons) their firing rate during vergence eye movements.

In addition to this role in vergence, recording studies in the rostral superior colliculi (SC) also revealed the presence of cells that burst during miniature saccades (also called microsaccades), just like more caudal cells discharge during larger saccades. Local pharmacological inactivation confirmed a causal role in the generation of microsaccades. However, it also unraveled an implication of rostral SC in specifying the direction of gaze when a target is presented (or even imagined) in the central visual field, static or moving.

Unbalanced activity between the two rostral SC causes a fixation offset: gaze direction is not directed to the target center but toward an offset location. The same deficit happens after unilateral pharmacological perturbation of CFN, suggesting that the sustained firing rate of its neurons adjusts the balance of activity between the two rostral SC during visual fixation. Thus, the direction of gaze would correspond to an equilibrium of commands that counterbalance with each other, involving fastigial projections to the left and right rostral SC but also to the left and right nuclei *reticularis tegmenti pontis* (NRTP). Through their projections to NRTP, the two rostral SC and CFN have access to neurons which, in the flocculus and paraflocculus, influence the firing rate of tonic neurons that innervate the motor and internuclear neurons in the abducens nuclei.

In summary, the knowledge of the detailed neuronal circuitry underlying the generation of disjunctive eye movements is not advanced as much as the circuitry for conjugate saccades. However, significant progress has been made during the last decade, helping to better understand the neurophysiology of vergence eye movements and strabismus.

## Tracking a moving visual target

A target drifting in the visual field often elicits a primary saccade (called interceptive saccade) at the end of which gaze does *not* land ahead of the moving target. Interceptive saccades either accurately capture (foveate) or fall behind the target. Then, a slow eye movement follows with a velocity that rarely matches the target's. A textured background (yielding visual motion in the direction opposite to the eye movement) does not seem to be responsible for the pursuit slower than target velocity because the consequences of its suppression are very weak. A subsequent correction saccade (called catch-up saccade) restores the foveation. Thus, unless the target trajectory is already known and foreseeable, gaze lags behind the target most of the time, especially when the target is small: the visual tracking is mostly composed of catch-up saccades interspersed by intervals during which the eyes drift roughly in the same direction as the target and with a lesser speed. In well-trained subjects, the catch-up saccades can be eliminated. Moreover, increasing the target size enhances the velocity of slow eye movements and reduces the size and number of catch-up

saccades. This point is important because most experimental studies use a small spot of light as a target.

Contrary to a static target which yields a bounded activity, a moving spot evokes on the retina a streak of activity the tail of which corresponds to past locations of the target and the leading edge the most recent location. In the deep superior colliculus, the population of saccade-related burst neurons is expanded as for a static target. However, it does not recruit cells that fire during saccades to future target locations. The population consists of a continuum of cells ranging from neurons issuing commands related to past locations of the target to neurons issuing commands related to its current location. At the level of motor neurons, other input must therefore complement the collicular signals. This input stems, at least in part, from motion signals processed in area MT because its lesion leads to saccades landing at locations that the target crossed earlier. Thus, the ability to capture the visual target here and now (at the time of saccade landing) may consist of adding a "predictive" component based on the integration of putative target-related "velocity" signals. However, an alternative explanation is possible. Accurate saccades toward a moving target may result from attenuating the synaptic weight of spikes emitted by cells that were the earliest to be excited by the passage of the target across their response field combined with promoting the synaptic weight of action potentials emitted by the cells that are more recently excited.

The dorsolateral pontine nucleus (DLPN) is one of the nuclei that relays signals from motion-related areas of the cerebral cortex to the cerebellum. When a chemical lesion is placed in the DLPN, saccades toward a moving target are hypometric whereas saccades to a static target remain unchanged. The hypometria suggests that the saccades were directed toward the past location of the target. Neurons in DLPN project to the floccular and medio-posterior regions of the cerebellum. Ablation of the floccular region (flocculus and paraflocculus) impairs the velocity of slow eye movements, seemingly without affecting catch-up saccades. By contrast, unilateral inactivation of the output nucleus of the medio-posterior cerebellum, the caudal fastigial nucleus, impairs both contralesional saccadic and pursuit eye movements. Gaze direction always lags behind the moving target because the saccades are hypometric and the pursuit velocity slowed. Unfortunately, the interpretation of the hypometria is complicated by the fact that saccades toward a static target are also

hypometric. More experiments are required to refine the neuronal mechanisms underlying the generation of these two saccade categories.

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#### Figure 8 about here

Concerning the generation of slow pursuit eye movements, the figure 8 schematizes the complex neuronal network involved in their generation when the movement direction is horizontal. The schema is not complete because our current knowledge prevents from explaining how neurons in the oculomotor vermis and the rostral superior colliculi are recruited during the generation of tracking (saccadic and slow) eye movements.

Some cells firing tonically in the nucleus prepositus hypoglossi (NPH) and the nearby medial vestibular nucleus (MVN) increase their firing rate during ipsilateral pursuit. Although called "eye-head velocity" neurons in the monkey, we shall name them tonic neurons for simplicity and for distinguishing them from the burst neurons and EVN-1 described in the previous paragraphs. Some of the tonic neurons are believed to provide an excitatory input to the abducens neurons for contracting the agonist muscle fibers during ipsilateral pursuit. While documented in the cat, this ipsilateral excitatory tonic input (ETN) has been questioned in the monkey. Most NPH/MVN neurons seem to be inhibitory and to project to the contralateral side (synapses a and b). When the sustained firing rate of tonic neurons is considered, we understand that the reciprocal commissural inhibition between the NPH/MVN in the left and right sides yields a push-pull organization, such that a reduced firing rate on one side enhances the firing rate on the opposite side. Thus, by suppressing the activity of the neurons that innervate the antagonist muscle fibers (red synapses a), the inhibitory tonic neurons (ITN) in the NPH/MVN contribute to the relaxation of these muscles, while disinhibiting the ipsilateral abducens and internuclear neurons (blue synapses a) and promoting an ipsiversive movement of both eyes.

Neurons in NPH/MVN are monosynaptically inhibited by Purkinje cells in the flocculus and the ventral paraflocculus (synapse b), which receive excitatory input from the three major pontine nuclei (synapses c) involved in relaying visual motion signals to the cerebellum: the rostral part of nucleus reticularis tegmenti pontis (rNRTP), the dorsolateral pontine nucleus (DLPN) and the dorsomedian pontine nucleus (DMPN). These pontine nuclei contribute to the inhibition that Purkinje cells exert upon NPH/MVN neurons. After a chemical lesion in the

dorsolateral pontine nucleus (DLPN) or in the rostral nucleus reticularis tegmenti pontis (rNRTP), ipsilesional pursuit eye movements are severely impaired. The suppression of pontine input likely reduces the floccullar inhibition of ITN neurons, which in turn impede the recruitment of agonist motor and internuclear neurons. The consequences of suppressing the input from DMPN have not been tested yet. However, an impairment of ipsilesional pursuit is also expected because the DMPN and the contralateral caudal fastigial nucleus exhibit strong reciprocal connections, and fastigial inactivation severely impairs the generation of contralesional pursuit eye movements.

The NPH/MVN are also the target of signals from neurons in the pretectal nucleus of the optic tract (NOT; synapse d) some of which are sensitive to the motion of a small visual target. These neurons fire during ipsilateral target motions and their loss severely impairs the ability to generate ipsilesional slow pursuit eye movements. NOT receives visual signals from the retina and from the middle temporal (MT) and medial superior temporal (MST) areas of cerebral cortex (synapses e and f) wherein numerous cells are sensitive to target motion as well. Projections of the frontal eye field (FEF) to NOT and to NPH have been documented (synapses e and e'). This access of frontal cortex to premotor neurons does not indicate a voluntary control of pursuit eye movements since the presence of a target is required for initiating a smooth pursuit. The NOT contributes to the maintenance of ipsilateral pursuit eye movements by its direct projections to tonic neurons in the ipsilateral NPH/MVN (synapse d), but also to the ipsilateral pontine nuclei (synapses g).

Through their projections to the ipsilateral rostral SC (synapse j), Gaba-ergic cells in NOT may contribute to suppression of the generation of saccades made in the direction opposite to the target motion. Simultaneously, they may depress the excitatory drive that NRTP exerts upon the ipsilateral paraflocculus (synapse k).

Finally, the secondary excitatory vestibular neurons (EVN-1 or PVP cells) that are involved in moving the eyes in the direction opposite to the head movement (Fig. 5) increase their firing rate during contraversive pursuit eye movements. However, as they do not seem to be inhibited by Purkinje cells located in the floccular region, the origin of the discharge modulation during contraversive pursuit remains to be identified. Direct projections from the frontal eye fields to the NPH/MVN might support this influence.

#### Conclusion

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Studying orienting movements of gaze is a convenient means for investigating how intertwined networks of neurons in the central nervous system transforms sensory signals into motor commands, but also, how it enables an entire animal to visually locate and capture an object in the physical environment and to adapt when intrinsic damage or contextual changes alter their execution.

During the last decades, technical developments offered the possibility to measure precisely the time course of these movements and to study correlations between the firing rate of neurons and kinematic parameters such as movement amplitude, velocity, acceleration and various differences (e.g., gaze error, velocity error, etc.). However, these correlation studies should not lead us to believe that a one-to-one correspondence exists between the multiple neuronal networks within which activities propagate, and the extrinsic and homogeneous medium (mathematical space) with which we quantify the motion of rigid objects such as the eyeballs, the head and targets. Such a mapping is not at all obvious; it is even quite questionable. Contrary to the physical space, the medium of neuronal activity is neither homogeneous nor passive. Unlike most objects at which we look and that we manipulate, the corresponding brain activity is not rigid. Even the mere spot of activity that a small static object evokes on the retina yields multiple parallel flows of activity that make its correspondence in the brain spatially distributed, temporally extended and contextdependent. Moreover, reducing neurons or neuronal chains to units encoding geometric or kinematic relations between gaze and target conceals the muscle forces and the multiple antagonisms that we have described between muscles. Furthermore, the velocity profile of the movement and the time course of force development do not match. During horizontal saccades for example, the duration of the interval during which the muscle force increases is on average 60% shorter than the duration of the total change in eye orientation. A force decay happens well before the change in eye position ends and the larger the saccade, the longer its lead-time relative to saccade arrest. Finally, the temporal series of numerical values quantifying the changes during eye reorientation hides the antagonisms between those multiple channels that cause each movement. To our knowledge, no study has reported identical time courses between the kinematic and neurophysiological recordings that would support the assumption of an isomorphism.

In the majority of models proposed during the last decades, the movements of the eyes and the head were considered as driven by (intrinsic) error signals encoding displacement vectors in physical space. These models have definitely been useful to communicate concepts and to bring a comprehensive picture of the complexity underlying the generation of movements. However, embedding geometric and kinematic notions within the inner functioning of the brain (i.e., mapping intrinsic neuronal signals with extrinsic behavioral measurements) may be neurophysiologically misleading because different constraints characterize the neurophysiological and kinematic descriptions. In fact, rather than outcomes of processes reducing geometric or kinematic errors, the saccade and pursuit acceleration may resume merely to transitions between equilibria opposing populations of neurons whose activity leads to mutually antagonist movement tendencies.

Further empirical investigation is still required to determine and explain several other issues: i.e., whether and how the networks underlying orienting movements of the eyes and head interact with those generating other types of goal-directed action such as reaching movements of the hand or locomotion; whether and how they interact with the networks involved in the navigation and the memory of locations; and whether and how they support the learning of new skills and possibly the acquisition of more abstract knowledge such as geometry or counting. With the recent multiplication of cognitive studies that use eye-tracking techniques to explore the so-called "inner space" with quantitative methods, more effort is required to characterize what exactly are those covert processes that eye movements would express. At the same time, neurophysiological and neuroanatomical works must continue to reveal the fine structure of the brain connectivity and to unravel the multiplicity of channels and networks that enable a living organism to connect with its natural and cultural environments as well as to reactivate the memory of past events.

By studying the simple and seemingly straightforward problem of orienting the eyes toward the location of a physical object, we see how the neurophysiological and neuroanatomical studies performed with animals enables us to understand the symptoms exhibited by patients suffering from eye movement disorders. They also show that evolutionary and embryological developments led to patterns that are different from the strategies of human engineers. Contrary to the case where a design precedes the realization of a machine for solving a specific problem, the physiology teaches us that any behavioral

observation is the outcome of a complex network of multiple and parallel chains of neurons. It also teaches us how the multiplicity and the diversity of elements enable an organism to solve several problems in parallel and even take up new challenges.

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### Figure captions

**Figure 1**: Schematic representation of the extraocular muscles (A) and pulling directions engaged by the contraction of their fibers when the eyes are centered in the orbit (B). LR: lateral rectus, MR: medial rectus, SR: superior rectus, IR: inferior rectus, SO: superior oblique, IO: inferior oblique, tr: trochlea. The red arrows indicate the muscles whose contraction abducts the eye, the black arrows the muscles whose contraction adducts the eye. The turquoise arrows indicate the muscles whose contraction elevates the eye, the yellow arrows those whose contraction lowers the eye. The blue arrows indicate the muscles whose contraction causes an intorsion of the eye, the green arrows those whose contraction causes an extorsion.

Figure 2: Neuronal network involved during horizontal saccades toward the left (A) and the right (B). Connecting lines ended by an arrow indicate excitatory connections; those ended by a circle indicate inhibitory synaptic connections. Blue color indicates the agonist neuronal elements, red color the antagonist ones. The thickness of connecting lines schematizes the strength with which the neurons fire. OPN: omnipause neurons, EBN: excitatory burst neurons, IBN: inhibitory burst neurons, MN: abducens motoneurons, AIN: abducens internuclear neurons. In the bottom, the thickness of arrows attached to the eyeballs schematizes the strength of muscle contraction. LR: lateral rectus, MR: medial rectus, SR: superior rectus, IR: inferior rectus, SO: superior oblique, IO: inferior oblique.

 **Figure 3**: Neuronal network involved during vertical saccades toward the top (A) and the bottom (B). Connecting lines ended by an arrow indicate excitatory connections; those ended by a circle indicate inhibitory synaptic connections. Blue color indicates the agonist neuronal elements, red color the antagonist ones. The thickness of connecting lines schematizes the strength with which the neurons fire. uEBN: upward excitatory burst neurons, dEBN: downward excitatory burst neurons, uIBN: upward inhibitory burst neurons, MN: motoneurons. In the top, the thickness of arrows attached to the eyeballs schematizes the strength of muscle contraction. LR: lateral rectus, MR: medial rectus, SR: superior rectus, IR: inferior rectus, SO: superior oblique, IO: inferior oblique.

Figure 4: Neuronal network involved during combined horizontal eye and head movements toward the left. Connecting lines ended by an arrow indicate excitatory connections; those ended by a circle indicate inhibitory synaptic connections. Blue color indicates the agonist neuronal oculomotor elements, red color the antagonist ones. Green color indicates the cephalomotor elements. The thickness of connecting lines schematizes the strength with which the neurons fire. dSC: deep superior colliculus, RSN: reticulospinal neurons, EBN: excitatory burst neurons, IBN: inhibitory burst neurons, ABD: abducens motor and internuclear neurons, NMN: neck motoneurons, PVN: primary vestibular neurons, IVN: inhibitory vestibular neurons, EVN: excitatory vestibular neurons, VIII: eight cranial nerve.

**Figure 5**: Neuronal network involved in ending a leftward gaze shift while the head terminates its rotation (stabilized gaze direction). Connecting lines ended by an arrow indicate excitatory connections; those ended by a circle indicate inhibitory synaptic connections. Blue color indicates the agonist neuronal oculomotor elements, red color the antagonist ones. Green color indicates the neurons of the ascending tract of Deiters (ATDN). The thickness of connecting lines schematizes the strength with which the neurons fire. LR: lateral rectus,MR: medial rectus, VI, abducens nerve, MRMN: motor neurons innervating the medial rectus muscle, LRMN: motor neurons innervating the lateral rectus muscle, AIN: abducens internuclear neurons, NMN: neck motoneurons, PVN: primary vestibular neurons, IVN: inhibitory vestibular neurons, EVN: excitatory vestibular neurons, VIII: eight cranial nerve.

Figure 6: Neuronal network involved in ending a downward (A) and upward (B) gaze shift while the head terminates its movement (stabilized gaze direction). Connecting lines ended by an arrow indicate excitatory connections; those ended by a circle indicate inhibitory synaptic connections. Blue color indicates the agonist neuronal oculomotor elements, red color the antagonist ones. IOMN: motor neurons innervating the inferior oblique muscle, IRMN: motor neurons innervating the inferior rectus muscle, SRMN: motor neurons innervating the superior oblique muscle, PVN: primary vestibular neurons, IVN: inhibitory vestibular neurons, EVN: excitatory vestibular neurons, VIII: eight cranial nerve.

**Figure 7**: Neuronal network involved in the near response. LR: lateral rectus, MR: medial rectus, SP: sphincter pupillae, DP: dilator pupillae, CM: ciliary muscle, CG: ciliary ganglion, MRMN: motor neurons innervating the medial rectus muscle, EWpg: Edinger-Westphal paraganglionic nucleus, cMRF: central mesencephalic reticular formation, SOA: supraoculomotor area, cFN: caudal fastigial nucleus, SC: superior colliculus.

**Figure 8**: Neuronal network involved in the generation of horizontal slow pursuit eye movements. LR: lateral rectus, MR: medial rectus, MN: motor neurons, AIN: abducens internuclear neurons, NHP/MV: complex nucleus prepositus hypoglossi-medial vestibular nucleus, ETN: excitatory tonic neurons, ITN: inhibitory tonic neurons, cFN: caudal fastigial nucleus, NOT: nucleus of the optic tract, rSC: rostral superior colliculus, rNRTP: rostral part of nucleus reticularis tegmenti pontis, DMPN: dorsomedian pontine nucleus, DLPN: dorsolateral pontine nucleus, VPL-Th: ventroposterolaral part of thalamus, FEF: frontal eye field, MST: medial superior temporal area of cerebral cortex.

















