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De novo assembling 19 maize inbred lines of the European germplasm

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Background and objectives

Characterizing the genomic diversity of maize is critical to understand the molecular origin of structural variation, and is a prerequisite to underpin the functional variation underlying phenotypic variation. Whole genome sequence assemblies at the chromosome scale with low amount of missing data are critical resources for answering such questions.

Whole genome sequence assemblies are now available for several maize inbred lines, but this is still missing for a number of key founders used in European maize breeding programs. This concerns the dent genetic pool and the European specific flint genetic group. Therefore, we have established a list of 13 priority lines for *de novo* sequencing as part of the SeqOccIn project.

One of the aim of the SeqOccIn project (Sequencing Occitanie Innovation, https://get.genotoul.fr/seqoccin/) is to acquire expertise on the optimal combination of long fragment sequencing technologies and associated applications to better characterize complex genomes for species of agronomical interest (maize, sheep, cow, pork). In the framework of this project, we are de novo assembling whole genome sequences of these 13 maize inbred lines. Here, we present results obtained for one inbred line, highlighting the strategy used to a "high quality assembled genome" with a combination of chosen technologies.

Sequencing data

		Links due s de	III: C woode			
	Long reads	Linked reads	Hi-C reads			
Preparation kit	SMRTbell Express Template Prep	Genome Reagent Kits v2 (10X	Dovetail Hi-C Kit			
rieparation kit	Kit 2.0	Genomics-chromium)	Dovetail Hi-C Kit			
Production plateforme	PacBio Sequel II	Illumina NovaSeq3000	Illumina NovaSeq3000			
Librairie size	15 kb	>50kb	NA			
Sequencing Layout	CSS mode	Paired End 2 × 150 bp	Paired End 2 × 150 bp			
Total number of reads	4,6 M	640 M paired-end	205 M paired-end			
Total Gb	67 Gb	197 Gb	62 Gb			

Our first goal is to validate the strategy choose to obtained a "high quality assembled genome" with our combination of technologies.

The table above summarizes the sequencing data produce for one of the maize line (MBS847) use to test our strategy.

Genome assembly protocole

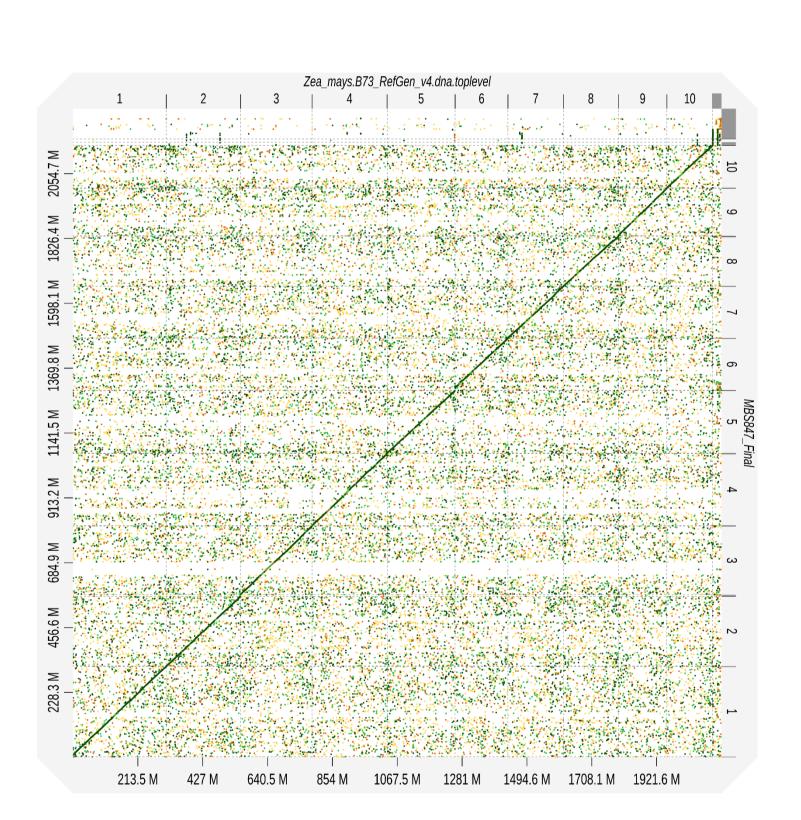
- 1. HiFiasm contig assembly with 28X HiFi reads
- 2. 3d-dna scaffolding with 26X Hi-C reads
- 3. Manual modification of Hi-C map
- 4. Manual verification with 10X chromium map (65X)

PacBio HiFi reads were assembled with Hifiasm v0.9 with default parameters. The assembly was then scaffolded using Hi-C reads which were first aligned to the assembly using juicer with default parameters. A pre-scaffolded assembly was then generated with 3D-DNA (version 180114) with -r 0 flag (no iterative rounds for mis-join correction). In addition, 10X chromium reads were aligned on the scaffolds and using tag continuity as 3d-dna input file was generated to help scaffolding validation. Both 10X chromium and Hi-C links were uploaded in Juicebox to manually review the assembly. Genome completeness was qualified using Benchmarking Universal Single-Copy Orthologs (BUSCO) v4.0.2 based on 4,896 BUSCO orthologs derived from the poales lineage.

Final Assembly statistics

	MBS847	B73
Number of scaffolds	2 564	685
Total size of scaffolds	2 282 968 551	2 182 075 994
Total scaffold length as percentage of assumed genome size	95,1 %	90,9%
% of estimated genome that is useful	94,7%	90,9%
N50 scaffold length	219 798 000	226 353 449
Scaffold %N	0,01	0,17
Number of contigs	2 981	2787
N50 contig length	53 262 804	1 279 966

Contiguity statistics of MBS847 assembly versus B73 reference sequence: The use of HiFi reads allowed us to obtain significantly longer contigs compared to the maize reference sequence. With a longer total assembly size (the expected size is ~2.4Gb) and lower gap percentage, our results offer a more complete image of the maize genome. As presented in Figure 2, the first 10 scaffolds represent an almost complete assembly of the maize chromosomes and the difference in scaffold number between our assembly and the reference sequence is due to a high number of short scaffolds originated from highly repetitive regions that have not been assembled in the reference.



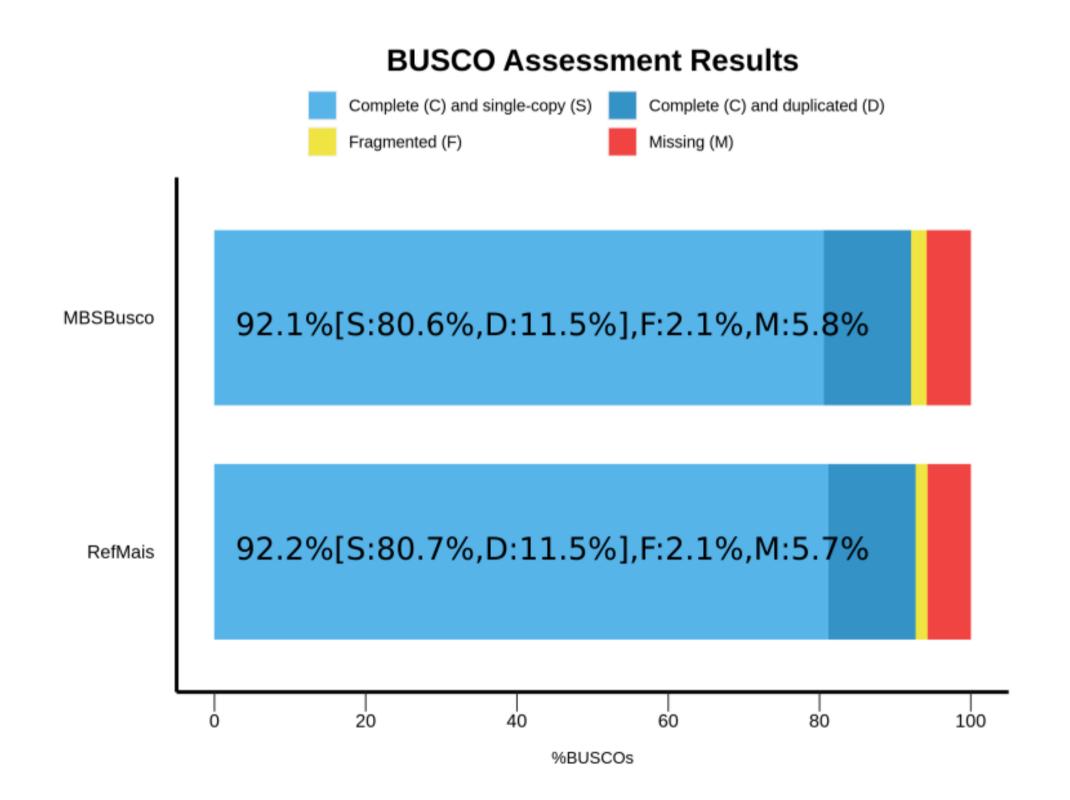


Figure 2: Alignment results of MBS847 assembly to the B73 reference sequence: The correctness of our assembly is indicated by the high quality alignment to the maize reference genome. The first 10 scaffolds represent an almost complete assembly of the maize chromosomes and we obtained also a significant number of short scaffolds originated from highly repetitive regions that have not been assembled in the reference.

Figure 1: Busco statistics of MBS847 assembly versus B73 reference sequence: The number of genes founded in our assembly is similar to the results of the maize reference sequence. A potential cause for the missing genes in our assembly is the variability between different maize lines.

Inbred line	CO255	3IIH6	LH82	B14	B37	F7130	F283	F120	PHN82	PHP02	LH123	PHR03	DKMM50 1D	PHG35	A632	DKFBLL	DKPB80	PHG39	PHW52
Group	Flint	lodent	Lancaster	SSS	SSS	Flint	Flint	Flint	lodent	lodent	Lancaster	Lancaster	Lancaster	Lancaster	SSS	SSS	SSS	SSS	SSS
Family	Flint	lodent	Lanc-Oh43	B14	B37	Aranga	F7	Flint	Iodent	lodent	Lancaster	Lanc-lod	Oh43	Oh43	B14	B73	B73	B73	B73

Inbred lines and type of sequencing to be applied: We identified by an enquiry process important founder lines of ongoing breeding programs for which genomic data are currently missing to our knowledge. Genetic groups encompass Flint lines specific to North European breeding, and several Dent heterotic groups adapted to different European regions.

More about the SeqOccin program

The overall SeqOccIn project should enable us to acquire expertise on the optimal combination of long fragment sequencing technologies and associated applications to better characterize complex genomes in agronomical field. Three majors topics are treated in this project: genome variability analysis, epigenetic mark analysis and metagenome analysis. The SeqOccIn project is carried out by Get-PlaGe and Genotoul Bioinfo platforms. It is financed by Feder funds (Programme Opérationnel FEDER-FSE_Midi-Pyrénées et Garonne 2014-2020).

