

Metabolic heat loss in southern elephant seals (Mirounga leonina) differs with stage of moult and between habitats

William D Paterson, Laureline Chaise, Chris Mcknight, John I Currie, Dave Thompson, André Ancel, Caroline Gilbert, Dominic J Mccafferty

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- 3
- 4 WILLIAM D. PATERSON^{a, e}, LAURELINE L. CHAISE^b, CHRIS McKNIGHT^a, JOHN I. CURRIE^c,
- 5 DAVE THOMPSON^a, ANDRÉ ANCEL^d, CAROLINE GILBERT^{b, f} and DOMINIC J. McCAFFERTY^e
- 6 ^aSea Mammal Research Unit, Scottish Oceans Institute, University of St. Andrews, St. Andrews, Fife,
- 7 KY16 8LB, UK
- 8 ^bUMR 7179 Laboratoire MECADEV CNRS/MNHN, 1 avenue du petit château, 91800 Brunoy, France
- 9 ^cSchool of Engineering and the Built Environment, Edinburgh Napier University, Merchiston Campus,
- 10 Colinton Road, Edinburgh, EH10 5DT, UK
- ^dUniversité de Strasbourg, CNRS, IPHC UMR 7178, F-67000 Strasbourg, France
- 12 ^eScottish Centre for Ecology and the Natural Environment, Institute of Biodiversity, Animal Health and
- 13 Comparative Medicine, University of Glasgow, Rowardennan, Glasgow G63 0AW, UK
- 14 ^fEcole Nationale Vétérinaire d'Alfort, Ethologie, 7 avenue du Général de Gaulle, 94700, Maisons-Alfort,
- 15 France
- 16
- 17 Corresponding author:
- 18 WILLIAM D. PATERSON
- 19 Email address:
- 20 <u>WilliamDavidPaterson@gmail.com</u>
- 21 Telephone number:
- **22** +44 (0) 7709602314
- 23 ORCID ID number:

24 https://orcid.org/0000-0001-5646-3782

25 Highlights

26	•	Heat loss in moulting southern elephant seals (SES) differed between habitat types.
27	•	Greatest heat loss was in beach habitat during the late moult.
28	•	Heat loss was lower in wallow and lower still in vegetation habitats.
29	•	Body condition and skin temperature both reduced at the end of the moult.

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31 Abstract

32 The moult in southern elephant seals (SES, Mirounga leonina) represents an especially energetically demanding period during which seals must maintain high skin temperature to facilitate complete 33 34 replacement of body fur and upper dermis. In this study, heat loss was measured on the body surface of 35 18 moulting SES in three different habitats (beach, wallow or vegetation) to estimate heat loss. 36 Temperature data loggers were also deployed on 10 SES to monitor skin surface temperature. On average, 37 heat loss of animals on the beach was greater than in wallows or vegetation, and greater in wallows than 38 in vegetation. Average heat loss across all habitats during the moult equated to 1.8 x resting metabolic 39 rate. The greatest heat loss of animals was recorded in the beach habitat during the late moult, that was 40 estimated to represent 2.3 x RMR. Mean mass loss was 3.6 ± 0.3 kg per day, resulting in changes in body 41 condition as the moult proceeded. As body condition decreased, skin surface temperature also decreased, 42 suggesting that as animals approached the end of the moult blood flow to the skin surface was no longer 43 required for growth of hair follicles.

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47 Keywords

48 *Mirounga leonina*, marine mammals, pinnipeds, heat loss, skin temperature, moult, thermoregulation

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71

72 1. Introduction

73 The annual moult in phocid seals represents a distinct, energetically demanding period during which 74 animals spend an increased amount of time on land as they shed and renew their entire coat (Hindell and 75 Burton 1988; Thompson et al. 1989; Boyd et al. 1993). Conditions for the proliferation of hair follicles 76 and skin cells during this time are optimised by increasing skin surface temperature closer to core body 77 temperature (Feltz and Fay 1966; Paterson et al. 2012). This is achieved by increasing perfusion of blood 78 at the skin surface (Khamas et al. 2012). Thermal conduction in water is 25 times that in air (Nadel 1984) 79 meaning that increasing skin surface temperature while at sea would result in a physiologically 80 unsustainable heat loss (Boily 1995; Watts 1996). High skin temperature with minimal heat loss is 81 therefore most efficiently achieved while animals are on land. For some seal species, moulting involves a 82 diffuse process lasting several weeks with intermittent trips to sea, such as for the grey seal (Halichoerus 83 grypus) (Boily, 1996) and the harbour seal (Phoca vitulina) (Ashwell-Erickson et al., 1986). In these 84 species, moulting is visually apparent during a four-week period (Thompson and Rothery 1987). 85 However, the underlying process of hair follicle regrowth, which is much less conspicuous, may be 86 evident for a period of up to 12 weeks (Ashwell-Erickson et al. 1986). For species such as the northern 87 elephant seal (Mirounga angustirostris) and the southern elephant seal (Mirounga leonina) (hereafter 88 referred to as SES) that undergo a catastrophic moult, hair and the upper dermis are shed together over a 89 period of 25-32 days with the majority if not the entire moult process being completed on land (Worthy et 90 al. 1992; Boyd et al. 1993).

91 The behavioural adaptation of spending more time on land during the moult allows phocid seals to 92 overcome the thermoregulatory constraints of moulting in an aquatic environment (Boily 1996; Watts 93 1996). However, for species found in more extreme climatic conditions at higher latitudes, further 94 behavioural adaptations may be necessary to minimise heat loss. In Antarctic and sub-Antarctic regions 95 SES that come ashore to moult initially search for muddy pools or wallows (Boyd et al. 1993; Chaise et 96 al. 2017) where they often aggregate in tight groups (Riedman 1990; Cruwys and Davis 1995). This 97 behaviour may serve to minimise the amount of skin surface area exposed to the air by either being in 98 contact with other animals or being partially covered in mud warmed by body heat from large groups of

99 seals. Huddling is an adaptive response seen in many species of birds and mammals (Canals et al. 1997; 100 Ostner 2002; Gilbert et al., 2010) that minimises the metabolic input required for thermoregulation 101 (Gilbert et al. 2010). If moulting SES elevate skin temperature as is seen in other phocid species (Paterson 102 et al. 2012) then huddling in wallows may help to conserve heat when actively perfusing the skin with 103 blood. The final stage of the moult process in SES is characterised by animals remaining on land but 104 moving out of wallows and closer to shore (Boyd et al. 1993; Chaise et al. 2017). Choice of habitat type 105 and stage of moult are therefore important factors when considering the environmental conditions 106 affecting the moult in SES.

107 Several studies have relied on theoretical heat flux models to estimate heat loss from phocid seals while 108 hauled out (Boily 1995; Harding et al. 2005). However, these models can significantly overestimate or 109 underestimate heat transfer rates (Kvadsheim et al. 1997; Boily et al. 2000, respectively). More accurate 110 estimates are achieved when theoretical models are validated with concurrent recordings of metabolic rate 111 (Kvadsheim and Folkow 1997; Kvadsheim et al. 1997) or by measuring heat flux directly (Kvadsheim 112 and Folkow 1997; Hindle et al. 2015). Direct measurements of heat flux in phocid seals have quantified 113 how heat is differentially lost from parts of the body in contact with the ground (Mellish et al. 2015) and 114 with the air (Hindle et al. 2015). Meteorological conditions and substrate temperature can then be used to 115 explain changes in heat transfer of animals.

116 Direct measurement of heat loss in moulting seals in the wild has rarely been successful (though see 117 Walcott et al. 2020) due to the difficulties of gluing devices to hair that is being shed or that is newly 118 grown and too weak to support device attachment. New research is required to determine heat loss of 119 moulting phocid seals to the surrounding environment to be able to estimate the energetic cost of the 120 moult associated with thermoregulation and the choice of habitat type when hauling out. An increased 121 metabolic rate during the moult has been demonstrated in previous studies (Slip et al. 1992; Boyd et al. 122 1993; Paterson et al. 2012; Paterson et al. 2021). This increased energetic demand is compounded by the 123 fact that animals fast on land (Slip et al. 1992; Hindell et al. 1994). As a result, body condition decreases 124 as blubber reserves become depleted during the moult (Hindell et al. 1994; Carlini et al. 2005). 125 Meteorological conditions and choice of habitat that increase heat loss from animals to the ground and air will increase energy costs as a greater metabolic rate will be required to maintain high skin temperaturesnecessary for hair and skin growth.

The aim of this study was to determine the energetic cost of a catastrophic moult in SES and how this may be influenced by terrestrial habitat and climate while moulting. Measurements of heat flux and body surface temperature were used to determine the extent to which environmental drivers such as habitat type and meteorological conditions influenced the energetic cost of moult in this species. Furthermore, because the moult fast decreases body condition which may compromise their ability to thermoregulate, we investigated how body surface temperature varied with body condition.

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135 2. Materials and methods

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137 2.1. Study animals

138 Study SES were captured during the early part of the moult, shortly after they arrived onshore and as they 139 moved away from the beach passing through vegetation en route to the wallows. Where recaptures were 140 possible in the latter part of the moult, animals were captured either while still in wallows or when they 141 had moved back to the beach and were likely to leave. A total of 18 adult females, 4 sub-adult females 142 and 1 sub-adult male SES were sampled between 14/01/2014 and 26/02/2016 at Point Suzanne, Kerguelen Islands (49°26'S, 70°26'E) during this study. Animals were anaesthetised with a 1:1 143 combination of Tiletamine and Zolazepam (Zoletil[®] 100), either administered intramuscularly with a 144 145 blow dart at a dose rate of 1ml/100kg or intravenously at a dose rate of 0.5ml/100kg (Baker et al. 1990). 146 Moult stage of each individual was estimated at capture and recapture based on the percentage area of the 147 body over which old skin and hair had been shed and new hair was growing through (Chaise et al. 2019). 148 Upon capture when animals were heavily moulting and recapture when the moult process was complete, 149 individuals were weighed (± 0.5kg) using a tripod and scales (HST Mini-Weigher, HST Scales, Milton 150 Keynes, UK). Measurements of body length (\pm 1.0cm) from nose to tail were also taken (Table 1).

SES work (Program IPEV 1037 HENERGES) was approved by The Antarctic Committee for Environmental Protection and the Ethics Committee (Cometh ANSES/ENVA/UPEC n°16: n°14-055 and n°15-061). The program was also authorised by the French Southern and Antarctic Lands (Decree 2014-131, 15th October 2014; Decree 2015-110, 4th September 2015). All applicable international, national and institutional guidelines for the care and use of animals were followed. All procedures performed in studies involving animals were in accordance with the ethical standards of the institution or practice at which the studies were conducted.

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2.2. Heat flux measurements

Heat flux (Wm⁻²) from the body of animals to the surrounding environment was measured while animals 159 160 were under anaesthesia, using heat flux discs (HFP01, Huxseflux Thermal Sensors B. V., Delft, 161 Netherlands) with data recorded to a logger (SQ2010, Omni Instruments Ltd., Dundee, UK). Heat flux 162 measurements were taken while animals were in a prone position. To measure heat flux from the body 163 surface to air, a heat flux disc was placed on the dorsal midline posterior to the axial line between the fore 164 flippers. A second heat flux disc was also placed posterior to the axial line but on the ventral surface 165 measuring heat flux to the ground (Fig. 1). Data were recorded at a sampling interval of once every 166 minute.

167 Animals were captured in three different types of habitat to determine differences in heat flux dependent 168 on substrate. The three main substrate types found at the Point Suzanne study site are; "vegetation" 169 composed of the plant species Azorella selago, Acaena magellanica and Taraxacum officinale (Chapuis 170 et al. 2004), "beach" composed of basalt pebbles (Nicolaysen et al. 2000) and "wallows" that are 171 normally characterised as depressions in the land in which aggregations of seals create wet, muddy 172 conditions. It should be noted that animals captured in the present study had to be isolated from groups in 173 order to be able to take measurements safely. This included those in wallows which resulted in heat flux 174 measurements being taken of isolated animals, usually at the edge of wallows, rather than in the centre. 175 Effects of heat flux to other animals while huddling and heat flux affected by large areas of the body 176 covered in mud were therefore not possible. However, the substrate type and ground temperature 177 differences at the edge of wallows was sufficiently different from vegetation and beach habitat types to 178 make valid comparisons (Chaise et al. 2019).

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2.3. Conversion of heat flux measurements to whole body heat flux

181 To extrapolate measurements of heat flux taken on the dorsal and ventral surfaces to the whole body 182 surface required a measure of the whole body surface area of animals. Previously, this has been done by 183 predicting maximal surface area based on mass according to the following allometric equation developed 184 for phocid seals by Innes et al. (1990): $\log A = \log 0.14 + 0.51 * \log m$ where A is body surface area (m²) 185 and m is the mass (kg) of the animal. This allometric relationship was based on post-mortem measures of 186 surface area of 56 skins removed from individuals of five different seal species including harbour seals, 187 harp seals (Pagophilus groenlandicus), hooded seals (Cystophora cristata), ringed seals (Pusa hispida) 188 and grey seals.

A photogrammetry method was developed to test whether the previous equation held true for SES by creating fully scaled 3D models using the software package AutoDesk® Photo RecapTM. To create 3D models, approximately 40 photographs were taken at different angles around the circumference of animals while they were anaesthetised. Accurate 3D models of animals were then generated and scaled using nose to tail length measurements taken during captures. Fig. 2 shows examples of 3D models generated for animals in each of the three habitat types; vegetation, beach and wallow.

195 Previous theoretical heat flux studies have assumed that 20% of the body surface of pinnipeds was in 196 contact with the substrate (Luecke et al. 1975; Paterson et al. 2012) and the remaining 80% exposed to the 197 surrounding environment. The 3D models were used to calculate the amount of model mesh, and therefore area of the body, that was in contact with the air and with the ground. Heat flux measurements 198 taken on the ventral and dorsal side of animals (Wm⁻²) were extrapolated over the area of skin in contact 199 200 with the air and the ground and weighted accordingly to give an overall estimate of whole-body heat flux 201 (W). For animals that had no photogrammetry taken, 3D models could not be created. In these cases, the 202 proportion of surface area in contact with the ground was assumed to be the mean of the proportions 203 calculated for all animals with photogrammetry measurements.

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205 2.4. Body surface temperature measurements

206 Measurements of body surface temperature for the duration between capture and recapture were recorded 207 (± 0.5°C) using miniaturised temperature loggers (iButton® DS1922L-F5, Maxim Integrated, San Jose, 208 USA). In 2014 and 2015, average temperature was sampled at an interval of 10 minutes. However, in 209 2016 this was increased to an interval of every one minute. This type of logger was chosen due to its 210 small size (diameter = 17.4mm, thickness = 5.9mm, weight = 3g) and its reliable use in a range of other 211 studies (McCafferty et al. 2015). Each logger was located over newly grown hair on either flank, dorsal to 212 the fore flipper and held in place by covering with a fine nylon mesh and a layer of two-part epoxy (Araldite® AW 2101, Huntsman International LLC, Texas, USA) (Fig. 3). 213

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2.5. Environmental measurements

Environmental conditions were recorded within one metre of animals during captures. Air temperature (\pm 0.1°C), ground temperature (\pm 0.1°C), relative humidity (\pm 1%), wind speed (\pm 0.1ms-⁻¹) and solar radiation (W.m⁻² error \pm 5%) were all measured using handheld instruments (Kestrel 3000 Pocket Weather Meter, Kestrel Instruments, Pennsylvania, USA; Pyranometer SKS111, Skye Instruments Ltd., Llandrindon Wells, UK). Measurements were taken at the beginning and end of captures with the average of those two values being used.

Environmental conditions in the vicinity of the study site (air temperature ($\pm 0.1^{\circ}$ C), relative humidity (\pm 1%), wind speed (± 0.1 ms-⁻¹) and solar radiation (Wm⁻² error \pm 5%)) were recorded as spot measurements every 30 minutes. Rainfall (mm error <6%) was recorded as the total every 30 minutes. These environmental conditions were logged using an automatic weather station (Minimet, Skye Instruments Ltd., Powys, UK) in a fixed location.

227 All means reported in the results below are given as
$$\pm$$
 SE

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- 230 **2.6.** Statistical analyses
- 231 2.6.1. 3D models and predictions of surface area

232 3D models generated for animals caught in 2016 were used to estimate whole body surface area. A multiple linear regression model was used to predict whole body surface area (m²) as a function of mass 233 234 (kg), length (cm) and girth (cm). Results were compared to predictions of surface area derived from 235 Equation 1 developed by Innes et al. (1990) which relied solely on mass as the predictor. All dependent 236 and explanatory variables were log-transformed for multiple regression analyses. The full model used for 237 selection was therefore log (surface area) ~ $\log (mass) + \log (length) + \log (girth)$ with all variables being 238 treated as continuous. Model selection was performed in a step-wise backwards selection process 239 dropping non-significant explanatory variables as appropriate. The final model could then be used to 240 predict surface area while accounting for variation in morphometrics. Table 1 summarises captures during 241 which photogrammetry images were taken that were then developed into 3D models.

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243 **2.6.2.** Heat flux

244 Whole body heat flux (W) was converted to kcal/day. This allowed for comparisons with estimations of 245 resting metabolic rate based on mass according to Kleiber (1932): $RMR = 70 * M^{0.75}$

where RMR is resting metabolic rate in kcal.day⁻¹ and M is mass (kg). Estimations of metabolic rate were
expressed as a multiple of resting metabolic rate (RMR multiplier), defined as dividing whole body heat
flux (kcal.day⁻¹) based on measurements by predictions of RMR (kcal.day⁻¹) from Kleiber's equation. In
this definition, Kleiber's prediction is referred to as 1 x Kleiber.

250 Analysis of co-variance (ANCOVA) was used to determine differences in the continuous response 251 variable, RMR multiplier, dependent on habitat type consisting of three levels; vegetation, wallow and 252 beach. A post-hoc Tukey's test was used to make pairwise comparisons of RMR multiplier across the 253 three habitat types. To reduce the number of explanatory variables used, air temperature and wind speed were combined to create the variable windchill, defined by the equation: windchill = 13.12+(0.6215*Ta)-254 $11.37*(v^{0.16})+(0.3965*Ta*v^{0.16})$ where Ta is air temperature (°C) and v is wind speed (kmh⁻¹) 255 (Environment and Climate Change Canada, Government of Canada; NOAA's National Weather Service, 256 257 USA). The environmental variables included as covariates were therefore windchill, ground temperature, 258 relative humidity and solar radiation. Time since initiation of general anaesthesia (minutes) was also included as a continuous covariate to determine how changes in body temperature associated with
anaesthesia may have influenced recorded heat flux. Moult stage at capture (20-60% of old skin and hair
shed) and recapture (90-100% of old skin and hair shed) was treated as a two-level factor. The full model
before selection was RMR multiplier ~ habitat type + windchill + ground temperature + relative humidity
+ solar radiation + time since anaesthesia + moult stage. ANCOVA model selection was performed in a
step-wise backwards selection process dropping non-significant explanatory variables as appropriate.

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2.6.3. Body surface temperature

267 Changes in body surface temperature (response variable) dependent on body condition were modelled 268 using a Generalised Additive Model (GAM) approach from the mgcv library (Wood 2004). In this case, a 269 proxy for body condition was used which was body mass index (BMI = mass $(kg)/length^2$ (m)). This 270 index was chosen for consistency with a previous study of habitat use and movements of the same study 271 animals (Chaise et al. 2018). Calculations of BMI between captures assumed that mass changed linearly 272 between measurements at capture and recapture. In the full GAM model, body surface temperature was 273 fitted as a smooth (thin plate regression spline) over values of BMI. Pre-screening of the measured 274 environmental variables showed that air temperature and solar radiation were correlated (Pearson, r=0.55, 275 p < 0.001). Air temperature was therefore used and solar radiation was excluded from the analysis. To 276 further reduce the number of explanatory variables used, air temperature and wind speed were combined 277 to create the smoothed variable windchill (as above). Rainfall was included as a continuous explanatory 278 variable and relative humidity as a smoothed explanatory variable. Moult stage was included as a 279 continuous explanatory variable which was estimated at capture and recapture as the percentage of body 280 surface where old skin and hair had been shed and new hair growth was evident. Animals were assumed 281 to shed and renew skin and hair linearly between those observations allowing moult stage to be treated as 282 a continuous variable rather than a categorical variable as in the heat flux analysis. Similarly, rate of 283 change of moult was also included as a continuous explanatory variable calculated as the change in 284 percentage moult per day between capture and recapture. The full GAM model before selection was body 285 surface temperature ~ smooth(BMI) + moult stage + rate of moult + s (windchill) + rainfall +

- smooth(relative humidity). Model selection during GAM analyses were performed by step-wisebackwards selection of candidate models with the lowest AIC scores.
- All statistical analyses were carried out using the statistical package R (R Development Core Team 2017).
- 289
- **3.** Results
- 291
- **3.1. Environmental measurements**

Environmental measurements taken with handheld instruments during captures averaged: air temperature 9.7 \pm 0.4 °C, ground temperature 8.9 \pm 0.2 °C, wind speed 6.9 \pm 0.7 ms⁻¹, relative humidity 78.5 \pm 0.9 % and solar radiation 320.8 \pm 33.5 Wm⁻² respectively (Table 2). Longer term measurements from the fixed weather station were air temperature 7.3 \pm 0.1 °C, wind speed 6.0 \pm 0.1 ms⁻¹ and relative humidity 76.2 \pm 0.2 %. Solar radiation during daylight hours averaged 313.6 \pm 6.9 W.m⁻² and there were 34 days of rain (>0.2mm) with a total rainfall of 122.0 mm.

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300 3.2. 3D models and predictions of surface area

A total of 18 scaled 3D models involving 12 individuals were created for animals captured in 2016 (Table 1). The mean surface area measured using photogrammetry and 3D modelling software $(3.20 \pm 0.08m^2)$ was higher than that predicted using equation by Innes et al. (1990) (2.54 ± 0.04m²). Similarly, mean surface area in contact with the ground measured using 3D modelling software was $1.27 \pm 0.04m^2$, equivalent to 39.7% of total surface area and greater than an assumed 20% of total surface area used previously (0.64 ± 0.02m²) (Luecke et al. 1975, Paterson et al. 2012).

307 Model selection during multiple linear regression analyses showed that girth measurements did not 308 explain a significant amount of the observed variation in surface area. After omission of girth 309 measurements during the selection process, the final model was log (surface area) ~ log (mass) + log 310 (length). The explanatory variable log (mass) was also not found to be significant during model selection 311 (t(15) = 0.85, p=0.41) but was retained due to it being the primary explanatory variable of interest. The 312 final model showed that surface area had a positive relationship with mass, increasing by 0.16 (SE = 0.19) 313 units of log (surface area) for every unit of log (mass) when controlling for length (Fig. 4). Surface area 314 was related to length, increasing by 1.34 (SE = 0.40) units of log (surface area) for every unit of log 315 (length) when controlling for mass (Fig. 5). The final model explained 59% of the variation in log 316 (surface area) overall (F = 10.93, df = 2,15, p=0.001). This final model was then used to predict surface 317 area using mass and length measurements for the animals captured throughout the study period that were 318 subsequently used for heat flux analyses.

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320 **3.3. Heat flux**

321 Of the 23 SES used in this study overall, a total of 18 individuals (13 adult females, 4 juvenile females 322 and 1 juvenile male) were used for measuring heat flux to the environment (Table 1). During model 323 selection, time since start of anaesthesia was not found to be a significant explanatory variable and was 324 subsequently excluded. The final model after selection was therefore RMR multiplier ~ habitat type + 325 windchill + ground temperature + relative humidity + solar radiation + moult stage. ANCOVA results 326 showed that RMR multiplier was different between the three habitat types sampled (F = 21.86, df = 2.77, 327 p < 0.001). Post-hoc Tukey comparisons indicated that the RMR multiplier was greater for animals on the 328 beach (2.3 ± 0.1) than in both vegetation (1.3 ± 0.1) (p<0.001) and wallow (1.8 ± 0.1) (p<0.001) habitats 329 and greater in wallows compared with vegetation (p=0.001). Across all habitat types, the mean predicted 330 RMR multiplier was 1.8. There was a decrease in RMR multiplier with increasing windchill (F = 20.53, 331 df = 1,77, p<0.001), ground temperature (F = 19.19, df = 1,77, p<0.001), relative humidity (F = 6.40, df = 1,77, p<0.001), relative humidity (F = 6.40, df = 1,77, p<0.001), relative humidity (F = 6.40, df = 1,77, p<0.001), relative humidity (F = 6.40, df = 1,77, p<0.001), relative humidity (F = 6.40, df = 1,77, p<0.001), relative humidity (F = 6.40, df = 1,77, p<0.001), relative humidity (F = 6.40, df = 1,77, p<0.001), relative humidity (F = 6.40, df = 1,77, p<0.001), relative humidity (F = 6.40, df = 1,77, p<0.001), relative humidity (F = 6.40, df = 1,77, p<0.001), relative humidity (F = 6.40, df = 1,77, p<0.001), relative humidity (F = 6.40, df = 1,77, p<0.001), relative humidity (F = 6.40, df = 1,77, p<0.001), relative humidity (F = 6.40, df = 1,77, p<0.001), relative humidity (F = 6.40, df = 1,77, p<0.001), relative humidity (F = 6.40, df = 1,77, p<0.001), relative humidity (F = 6.40, df = 1,77, p<0.001), relative humidity (F = 6.40, df = 1,77, p<0.001), relative humidity (F = 6.40, df = 1,77, p<0.001), relative humidity (F = 6.40, df = 1,77, p<0.001), relative humidity (F = 6.40, df = 1,77, p<0.001), relative humidity (F = 6.40, df = 1,77, p<0.001), relative humidity (F = 6.40, df = 1,77, p<0.001), relative humidity (F = 6.40, df = 1,77, p<0.001), relative humidity (F = 6.40, df = 1,77, p<0.001), relative humidity (F = 6.40, df = 1,77, p<0.001), relative humidity (F = 6.40, df = 1,77, p<0.001), relative humidity (F = 6.40, df = 1,77, p<0.001), relative humidity (F = 6.40, df = 1,77, p<0.001), relative humidity (F = 6.40, df = 1,77, p<0.001), relative humidity (F = 6.40, df = 1,77, p<0.001), relative humidity (F = 6.40, df = 1,77, p<0.001), relative humidity (F = 6.40, df = 1,77, humidity (F = 6.40, Humidity (F 332 1,77, p=0.013) and solar radiation (F = 4.40, df = 1,77, p=0.039). The RMR multiplier increased (F = 333 4.89, df = 1,77, p=0.030) as animals progressed in their moult stage. Mean RMR multiplier with 95%334 confidence intervals adjusted for the effects of covariates are presented for each habitat type in Fig. 6. The 335 final model explained 51.4% of the variation in RMR multiplier across the three habitat types. The final 336 ANCOVA model was checked and found to meet assumptions of homogeneity and normal distribution of 337 residual errors.

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339 3.4. Body surface temperature

340 A total of 10 adult females were instrumented with iButton® temperature loggers with a mean 341 deployment duration of 5.1 ± 0.7 days, resulting in 59900 individual measurements (Table 1). A further 342 14 deployments were attempted but instruments were either quickly lost due to hair not being strong 343 enough to support device attachment or water damage causing instruments to fail. During model 344 selection, all explanatory variables were retained, meaning the final GAM model selected was body 345 surface temperature \sim smooth(BMI) + moult stage + rate of moult + s (windchill) + rainfall + s (relative humidity). GAM model predictions showed there to be an overall non-linear decrease in body surface 346 temperature with decreasing BMI (p<0.001). Animals with a BMI of between 55 and 66 kg m⁻² 347 maintained a surface temperature of between 26.7°C and 28.8°C which then rapidly declined to 9.0°C 348 when animals had a BMI of between 47 and 54 kg m⁻² (Fig. 7). Body surface temperature decreased 349 350 linearly with increasing moult stage (p<0.001) and rainfall (p<0.001) and increased linearly with 351 increasing rate of moult (p<0.001). BMI, windchill and relative humidity were all significant smooth 352 terms (all p<0.001) included in the final model. The amount of variability in body surface temperature 353 explained by the final GAM model was 19.5% indicating that the extent to which external conditions 354 affect surface temperature is relatively small and that other behavioural/physiological factors may be 355 more important.

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360 **4.** Discussion

361 SES that come ashore to moult have been shown previously to initially reside in vegetation before 362 seeking out muddy wallows (Boyd et al. 1993; Chaise et al. 2017) where they aggregate in tight groups 363 (Riedman 1990; Cruwys and Davis 1995). When the moult is nearing completion, they move towards the 364 beach before eventually going to sea (Boyd et al. 1993; Chaise et al. 2017). In this study, animals showed 365 a similar temporal and spatial pattern of behaviour while moulting in that the initial part of the moult was 366 characterised by movements across vegetation towards wallows when initial captures were carried out. 367 Having completed the moult, animals then left the wallows and moved back through vegetation towards 368 the beach where they would reside for several days before leaving the study site (Chaise et al. 2017). All 369 second captures of animals took place either as animals were in wallows or on the beach as they were 370 getting ready to leave. This study sought to determine possible drivers of these behaviours by examining 371 heat loss in each of the three different habitat types and by assessing temperature profiles of animals as 372 body condition decreased during the moult fast.

373 Heat loss from animals to their surrounding environment resulted in predictions of RMR multiplier that 374 were 38% higher in individuals that had already moved to wallow habitat (1.8 x Kleiber) compared with 375 those that were moving towards wallows through vegetation (1.3 x Kleiber). This may be partly explained 376 by the fact that the largest visual changes in terms of renewing skin and hair occurred while animals were 377 in wallows and so could be considered to be at the peak of moulting. At this time, phocid seals are 378 expected to be maximally perfusing blood to the skin surface which facilitates the renewal of skin and 379 hair (Paterson et al. 2012). Animals captured around wallows may therefore simply have had a higher 380 skin temperature gradient with the surrounding environment because they were at the peak of the moult, 381 resulting in higher heat flux measurements. Conversely, animals moving towards wallow habitat through 382 vegetation had lower heat fluxes. This could have been due to the fact that animals were at an earlier 383 stage in the moult process and so elevation of skin temperature was less evident. Additionally, the thermal 384 properties of vegetation may mean that conduction of heat from animals' bodies was reduced.

Heat loss from animals to the surrounding environment resulted in predictions of RMR multiplier that were 77% higher when animals had appeared visually to have completed the moult and moved to beach habitat (2.3 x Kleiber) compared with vegetation habitat (1.3 x Kleiber), and 28% higher compared with wallow habitat (1.8 x Kleiber). Visually, study animals appeared to have completed the moult process when they left wallow and vegetation habitats and arrived at the beach. However, previous studies on other phocid species have shown that the period during which hair follicles are active extends beyond the point when visual signs of moulting are apparent (Ashwell-Erickson et al. 1986; Ling 2012). It is therefore likely that animals captured in the beach habitat were still actively moulting despite appearing to
have completed the moult process. RMR on the beach, measured at 2.3 x Kleiber, approached the 2.8 x
Kleiber reported by Boyd et al. in 1993 for animals captured at a lower latitude in South Georgia.
However, the average RMR across the three habitats in the present study was lower overall at 1.8 x
Kleiber, which may be partly due to study animals moulting in a less extreme environment at a higher
latitude.

398 Predictions of heat flux expressed as RMR multiplier in different habitat types were performed using a 399 modelling approach that accounted for the effects of environmental variables. Choice of habitat type 400 would therefore appear to influence how heat is lost to the environment in moulting SES. Behavioural 401 adaptations to reduce heat loss are rarely assessed in hauled out phocids, though lair use in ringed seals 402 (Phoca hispida) (Kelly and Quakenbush 1990) represents a rare example. A lack of research in this area 403 may be due to the thick insulating blubber layer in phocid seals being considered to provide sufficient 404 insulation in cold environments (Hart and Irving 1959; Hansen et al. 1995). However, the results of this 405 study suggest that differences in heat conduction between habitat types was enough to influence where 406 animals chose to reside at certain stages of the moulting process. Mellish et al. (2015) predicted that in 407 Weddell seals (Leptonychotes weddellii), conduction of heat to the substrate accounted for 28% of total 408 heat loss while hauled out on ice. In the present study, average conduction of heat to the substrate was 409 found to average 37% after accounting for the increased surface area in contact with the ground shown by 410 3D models. It may therefore be the case that differential conduction of heat away from animals dependent 411 on substrate type and localised environmental conditions was a significant driver of habitat choice while 412 animals were moulting. The data also suggest that the relatively high energetic cost of moulting in SES 413 described by Boyd et al. (1993) may be more evident when animals have established themselves in 414 wallows and when they are on the beach before heading out to sea.

415 Surface temperature decreased with decreasing BMI between capture and recapture. The body condition 416 and thickness of blubber of animals that come ashore to begin moulting is expected to be relatively high 417 and decrease thereafter as the moult proceeds (Slip et al. 1992; Hindell et al. 1994). This is 418 physiologically important as the blubber layer is the primary means of insulation due to its low thermal 419 conductivity and high insulative properties when not perfused (Ling 1968; Kvadsheim and Aarseth, 420 2002). However, the insulative properties of blubber may be bypassed to some extent through venous 421 return from the skin surface when animals must lose excess heat (Kvadsheim and Folkow 1997; Mauck et 422 al. 2003). Heat may also be lost by perfusion of blood through anastomoses at the skin surface to facilitate 423 hair renewal during the moult (Paterson et al. 2012). Mean mass loss in the present study was 3.6 ± 0.3 kg day⁻² which was similar to that found for moulting SES in other studies (Carlini et al. 1999; Chaise et al. 424 2019). The fact that body surface temperature decreased at BMI values of less than 54 kg m⁻² suggests 425 426 that perfusion of blood to the skin surface was reduced beyond this point. However, it is unlikely that a 427 lowered BMI and a resultant reduction in blubber layer thickness would occur to the extent that animals 428 were forced to reduce perfusion of blood to the skin surface as a thermoregulatory response. It is therefore 429 more likely that that lower body surface temperature at the end of the moult was the result of a reduced 430 need to elevate skin temperature for hair and skin growth.

431 Higher body surface temperatures at the beginning and middle of deployments were likely a reflection of 432 increased perfusion of blood to the skin surface as part of the moult process. Additionally, behavioural 433 changes associated with moulting in SES could have contributed to elevated body surface temperatures. 434 SES huddle during the moult (Riedman 1990; Cruwys and Davis 1995; Chaise et al. 2019) which results 435 in a large proportion of the body being in contact with other individuals and not the ground or air. The 436 behavioural adaptation of aggregating (huddling) in wallows by SES is considered to minimise heat loss 437 through sharing body heat from other individuals (Riedman 1990). However, Chaise et al. (2019) found 438 that stomach and skin temperature were in fact lower in aggregated seals compared with solitary ones. 439 Huddling animals may be able to reduce core and skin temperature as a means of saving energy while 440 also benefitting from heat shared between individuals in contact with each other. Animals in the present 441 study were likely to have been huddling some of the time in wallow habitats and vegetation but no 442 behavioural observations between capture and recapture were recorded.

443

444 5. Conclusions

445 This study showed that heat transfer between skin surface and the surrounding environment in SES was446 dependent on habitat selection. Moving from the haulout beach to aggregate in wallows or in vegetation

447 may have allowed animals to increase skin surface temperature while minimising heat loss. The relatively 448 high body condition of animals during the initial stages of moult would have afforded animals higher 449 energy stores to cope better with the thermoregulatory costs at this time. As the moult proceeded, body 450 condition declined which was concomitant with a lowering of skin surface temperature. In late moult, this 451 may have been due to vasoconstriction of anastomoses in the skin to minimise heat loss when moving 452 back to the beach before heading out to sea. However, the main reason for reduced skin temperatures was 453 likely a reduced need to perfuse blood to the skin surface as the moult was nearing completion. This study 454 provides valuable insight into the ecophysiology of the catastrophic moult in SES, showing that the 455 behaviour of animals coming onto land and moving from the beach across vegetation and into wallows 456 influences heat transfer as they make that transition. Longitudinal behavioural observations concurrent 457 with heat flux and surface temperature measurements are recommended as the next step to examine more 458 fully the relationship between behavioural and physiological adaptations that facilitate the moult process. 459 This research may be particularly relevant for SES adapted to changes in environmental conditions in 460 polar regions.

461

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474	care and use of animals were followed. All procedures performed in studies involving animals were in
475	accordance with the ethical standards of the institutions or practice at which the studies were conducted.
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633	Tables	
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	Da	ate	Ma	ss (kg)		Gir	th (cm)		Heat flu	$(W.m^{-2})$
Animal	Capture	Recapture	Capture	Recapture	Length	Capture	Recapture	iButton® (°C)	Capture	Recapture
sex										
9 A ♀	14/01/2014	23/01/2014	313.0	279.6	228	171	164	21.1 ± 0.4	NA	NA
10 A ♀	15/01/2014	22/01/2014	289.4	259.4	230	170	158	19.7 ± 0.3	NA	NA
30 J ∂	24/12/2014	NA	279.0	NA	217	165	NA	NA	78.05 ± 3.86	NA
31 J ♀	24/12/2014	NA	222.0	NA	212	150	NA	NA	115.67 ± 8.63	NA
32 J ♀	24/12/2014	NA	268.0	NA	207	170	NA	NA	185.62 ± 4.46	NA
33 J ♀	24/12/2014	NA	183.0	NA	192	144	NA	NA	167.10 ± 6.67	NA
34 J ♀	25/12/2014	NA	185.0	NA	166	153	NA	NA	175.15 ± 4.04	NA
38 A ♀	09/01/2015	12/01/2015	323.0	313.0	222	172	175	26.4 ± 0.1	NA	NA
39 A ♀	09/01/2015	13/01/2015	297.0	284.0	221	165	163	26.8 ± 0.1	NA	NA
40 A ♀	09/01/2014	13/01/2014	259.0	249.0	211	161	160	21.4 ± 0.1	NA	NA
43 A ♀	25/01/2016 (P)	01/02/2016 (P)	298.0	275.0	236	166	152	NA	97.54 ± 1.96	NA
44 A ♀	25/01/2016 (P)	NA	326.5	NA	247	177	NA	NA	96.00 ± 2.15	NA
45 A ♀	27/01/2016 (P)	02/02/2016 (P)	319.0	300.5	218	177	169	23.9 ± 0.1	126.66 ± 3.48	NA
47 A ♀	27/01/2016 (P)	02/02/2016 (P)	298.5	288.5	231	158	168	26.2 ± 0.1	NA	161.94 ± 1.66
50 A ♀	27/01/2016 (P)	01/02/2016 (P)	286.0	273.0	237	136	155	25.8 ± 0.1	93.51 ± 2.38	NA
51 A ♀	27/01/2016	31/01/2016 (P)	313.5	299.0	226	163	174	23.3 ± 0.1	200.38 ± 2.06	NA
52 A ♀	28/01/2016 (P)	02/02/2016 (P)	265.5	250.5	230	154	167	NA	103.54 ± 4.72	NA
53 A ♀	28/01/2016 (P)	NA	284.0	NA	227	170	NA	NA	134.07 ± 6.50	NA
54 A ♀	28/01/2016 (P)	01/02/2016	322.5	311.0	236	176	172	27.4 ± 0.1	165.23 ± 1.36	NA
56 A ♀	16/02/2016	22/02/2016 (P)	308.5	290	228	170	171	NA	NA	277.22 ± 4.18
57 A ♀	17/02/2016 (P)	NA	420.0	NA	250	193	NA	NA	NA	145.18 ± 2.93
58 A ♀	17/02/2016 (P)	25/02/2016 (P)	265.5	248.0	203	210	165	NA	132.82 ± 1.81	128.86 ± 3.11
59 A ♀	21/02/2016	26/02/2016	NA	383.5	253	NA	187	NA	146.39 ± 1.75	NA

Table 1. Sex and age class are indicated in individual codes (A = adult, J= juvenile). Mass, length and girth measurements as well as mean \pm SE heat flux (W.m⁻²) are

shown for each individual on the date of capture and recapture. Also given are mean ± SE iButton® (°C) measurements over the course of deployment. P next to dates

indicates	3D	photogrammetry.	NA	values	indicate	no	data	were	collected.
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			Habitat type	
	Environmental measure	Vegetation	Beach	Wallow
	Air temperature (°C)	8.8 (0.3)	9.2 (0.3)	11.7 (1.1)
	Ground temperature (°C)	7.4 (0.1)	10.5 (0.3)	9.5 (0.2)
	Relative humidity (%)	76.5 (1.8)	81.5 (1.5)	77.2 (1.0)
	Wind speed (ms ⁻¹)	5.5 (0.7)	5.7 (1.0)	10.6 (1.9)
	Solar radiation (Wm ⁻²)	211.2 (42.5)	260.6 (43.1)	568.1 (76.9)
637	Table 2. Mean ± SE of environmenta	al conditions recorded	using handheld ins	struments during cap
638	of the three habitat types: vegetation,			<i>o</i> - T
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658 Figure Captions

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Fig. 1. Placement of heat flux disc on the dorsal midline posterior to the axial line between the fore flippers to
measure heat flux from the animal to the air. Heat flux to the ground was measured with a heat flux disc placed
on the ventral midline (not shown) directly opposite to the dorsal surface.

- **Fig. 2.** 3D models using the software package AutoDesk[®] Photo Recap[™] to measure surface area of animals in
- 664 contact with the air and with the ground. From top to bottom models shown are for each of the three habitats:
- vegetation (Individual 45 at recapture), beach (Individual 56 at recapture) and wallow (Individual 58 at capture).

666 Fig. 3. Location of iButtons® placed on the flank of animals, posterior to the fore flipper. iButtons® were

667 covered with a combination of fine nylon mesh and a layer of two-part epoxy. Also attached on the dorsal

- surface were a GPS transmitter and a VHF transmitter for locating animals.
- **Fig. 4.** Partial residual plot of log (area) against log (mass). The black solid line is the predicted linear

relationship between log (area) and log (mass) with 95% confidence intervals shaded grey. Points represent the

prediction of log (area) dependent on log (mass) for each capture. The predicted linear relationship is given after

accounting for the other explanatory variable used, which in this plot was log (length).

Fig. 5. Partial residual plot of log (area) over log(length). The black solid line is the predicted linear relationship
between log (area) and log(length) with 95% confidence intervals shaded grey. Points represent the prediction of
log (area) dependent on log (length) for each capture. The predicted linear relationship is given after accounting
for the other explanatory variable used, which in this plot was log (mass).

677 Fig. 6. ANCOVA predictions of corrected mean multiple of resting metabolic rate (RMR multiplier) according

to allometric equations by Kleiber (1932) (left y-axis) in three different habitat types; vegetation, wallow and

beach (x-axis) based on heat flux calculations. Also shown are predicted values for RMR multiplier converted to

- 680 kJ.day⁻¹ (right y-axis). 95% confidence intervals of corrected means are also shown.
- 681 Fig. 7. Predictions (solid black line) of body surface temperature (°C) (y-axis) dependent on body mass index

(BMI (kg.m⁻²)) (x-axis). 95% confidence intervals of predictions (grey) are also shown. Black circles indicate

683 mean body surface temperature for each value of BMI.

684



Fig. 1







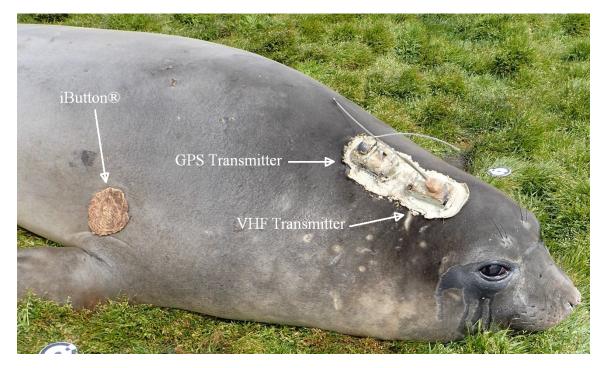


Fig. 3

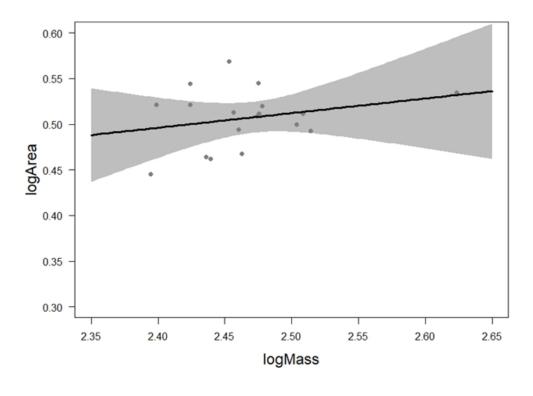
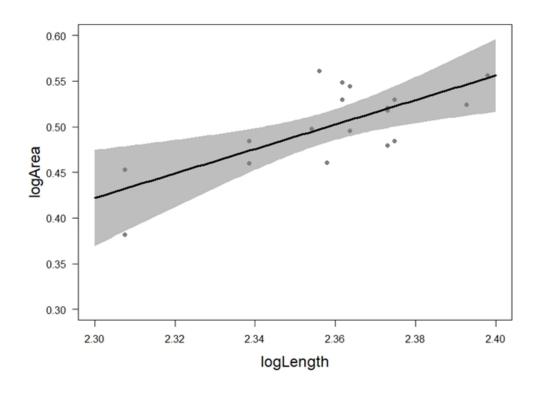
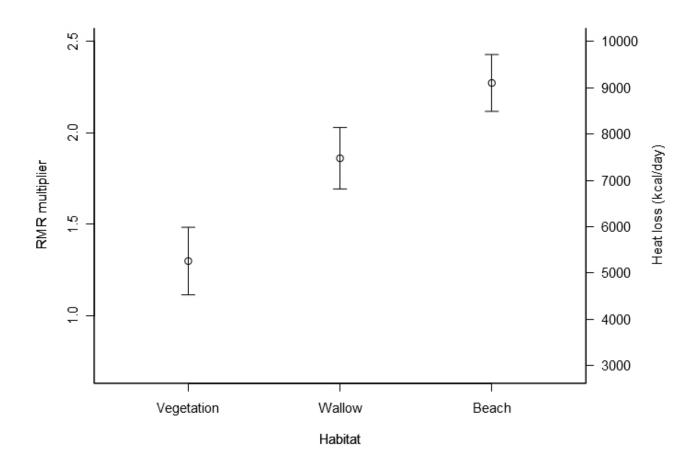


Fig. 4







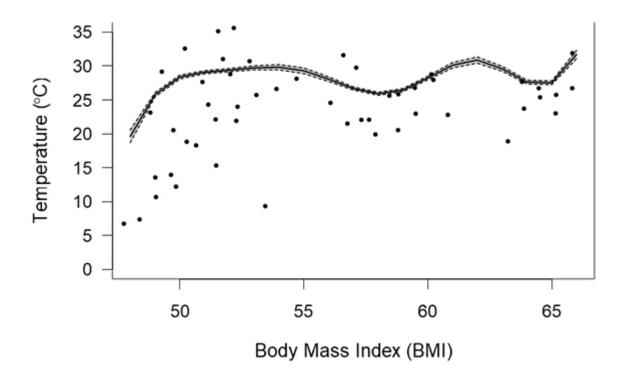


Fig. 7