



HAL
open science

Advances in the treatment of inflammatory bowel disease: Focus on polysaccharide nanoparticulate drug delivery systems

Hicheme Hadji, Kawthar Bouchemal

► **To cite this version:**

Hicheme Hadji, Kawthar Bouchemal. Advances in the treatment of inflammatory bowel disease: Focus on polysaccharide nanoparticulate drug delivery systems. *Advanced Drug Delivery Reviews*, 2022, 181, pp.114101. 10.1016/j.addr.2021.114101 . hal-03524042

HAL Id: hal-03524042

<https://hal.science/hal-03524042v1>

Submitted on 13 Jan 2022

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

1 Advances in the Treatment of Inflammatory Bowel Disease:
2 Focus on Polysaccharide Nanoparticulate Drug Delivery
3 Systems

4 *Hicheme Hadji, Kawthar Bouchemal **

5 Institut Galien Paris Saclay, CNRS UMR 8612, Université Paris-Saclay, Faculté de Pharmacie, 5 rue
6 J-B Clément, 92296, Châtenay-Malabry France

7 Corresponding Author

8 * kawthar.bouchemal@universite-paris-saclay.fr

1 **Abstract**

2 The complex pathogenesis of inflammatory bowel disease (IBD) explains the several hurdles
3 for finding an efficient approach to cure it. Nowadays, therapeutic protocols aim to reduce
4 inflammation during the hot phase or maintain remission during the cold phase. Nonetheless,
5 these drugs suffer from severe side effects or poor efficacy due to low bioavailability in the
6 inflamed region of the intestinal tract. New protocols based on antibodies that target
7 proinflammatory cytokines are clinically relevant. However, besides being expensive, their
8 use is associated with a primary nonresponse or a loss of response following a long
9 administration period. Accordingly, many researchers exploited the physiological changes of
10 the mucosal barrier for designing nanoparticulate drug delivery systems to target inflamed
11 tissues. Others exploited biocompatibility and relative affordability of polysaccharides to test
12 their intrinsic anti-inflammatory and healing properties in IBD models. This critical review
13 updates state of the art on advances in IBD treatment. Data on using polysaccharide
14 nanoparticulate drug delivery systems for IBD treatment are reviewed and discussed.

15 **Keywords:** Inflammatory bowel disease; Nanoparticles; Drug delivery systems;
16 Polysaccharides.

1 **Contents**

2 Introduction..... 4

3 1. Overview of the anatomy and physiology of the colon 5

4 2. Pathophysiology and complications of IBD 7

5 3. General overview of conventional therapeutic agents for IBD and their shortcomings 13

6 4. Implication of pathophysiological changes of the GI tract during active IBD for oral drug

7 delivery strategies 15

8 4.1. *pH changes*..... 15

9 4.2. *Mucus*..... 16

10 4.3. *Enzymatic secretions*..... 19

11 4.4. *Epithelial barrier* 19

12 4.5. *Modifications of immune cell surface receptors* 20

13 4.6. *Production of reactive oxygen species* 21

14 5. Polysaccharide-based drug delivery systems for IBD 26

15 5.1. *Chitosan* 28

16 5.2. *Hyaluronan* 29

17 5.3. *Pectins*..... 31

18 5.4. *Inulin*..... 32

19 7. Challenges and future directions..... 36

20 8. Conclusion 38

21 9. Funding 39

22 10. References..... 39

23 11. Abbreviations 58

24

1 Introduction

2 Inflammatory bowel disease (IBD) is a broad term used to describe a chronic
3 inflammatory state that occurs mainly in the colon, although the other portions of the
4 intestinal tract were also concerned. IBD mainly encompasses two inflammatory conditions,
5 either ulcerative colitis (UC) or Crohn's disease (CD). From an anatomical point of view, UC
6 patients mainly suffer from an inflammation of the rectal and sigmoid colon.^{1,2} In contrast,
7 the ileo-caecal area remains the most affected region in CD,^{3,4} although many cases have also
8 been reported in the small intestine.⁵ Another aspect that characterizes CD is the skip lesions,
9 where the affected region alternates between inflamed and non-inflamed segments.⁶ From a
10 histological point of view, UC is a superficial inflammation, whereas CD is transmural,
11 affecting all the layers of the colon, which makes the complications more severe.^{7,8}

12 Despite the tremendous efforts made to comprehend the pathophysiological state
13 triggering the occurrence of IBD, the exact etiology remains unclear. Up to date, several
14 parameters are incriminated as potential causative factors. They can be classified as being
15 either endogenous or exogenous. Endogenous factors are mainly represented by genetic
16 predisposition or immunoregulatory disorders. However, several studies suggested that
17 microbiome dysregulation could also have a considerable impact on the development of
18 IBD.^{9,10} Exogenous factors, on the other hand, are linked to exposure to external stress.¹¹ The
19 stress can be environmental (e.g., alcohol, chemicals, drugs, industrial diet style),
20 psychological (e.g., depression, anxiety), or even microbial through the intestine invasion by
21 pathogenic bacteria.

22 With more than 6.8 million people suffering from the complications of IBD at a global
23 scale,¹² a curative treatment of this pathology is still missing, and current treatment protocol
24 aims at relieving acute attacks (e.g., the combination of 5-aminosalicylic acid (5-ASA) and
25 corticosteroids, TNF- α antibodies) or preventing relapse episodes (e.g., 5-ASA therapy for
26 UC and corticosteroids for CD). However, the active pharmaceutical ingredients (API)
27 delivery still rests on conventional drug delivery systems (DDS) such as tablets, capsules,
28 enemas, or solutions. These formulations have shown several limitations as the yield of the
29 API in the colonic region is low.^{13,14} Consequently, working on alternative therapeutic
30 approaches remains necessary to offer a better treatment protocol and reduce patient
31 discomfort while diminishing the multiple administrations of medications or the recurrence to
32 less hospitalization.

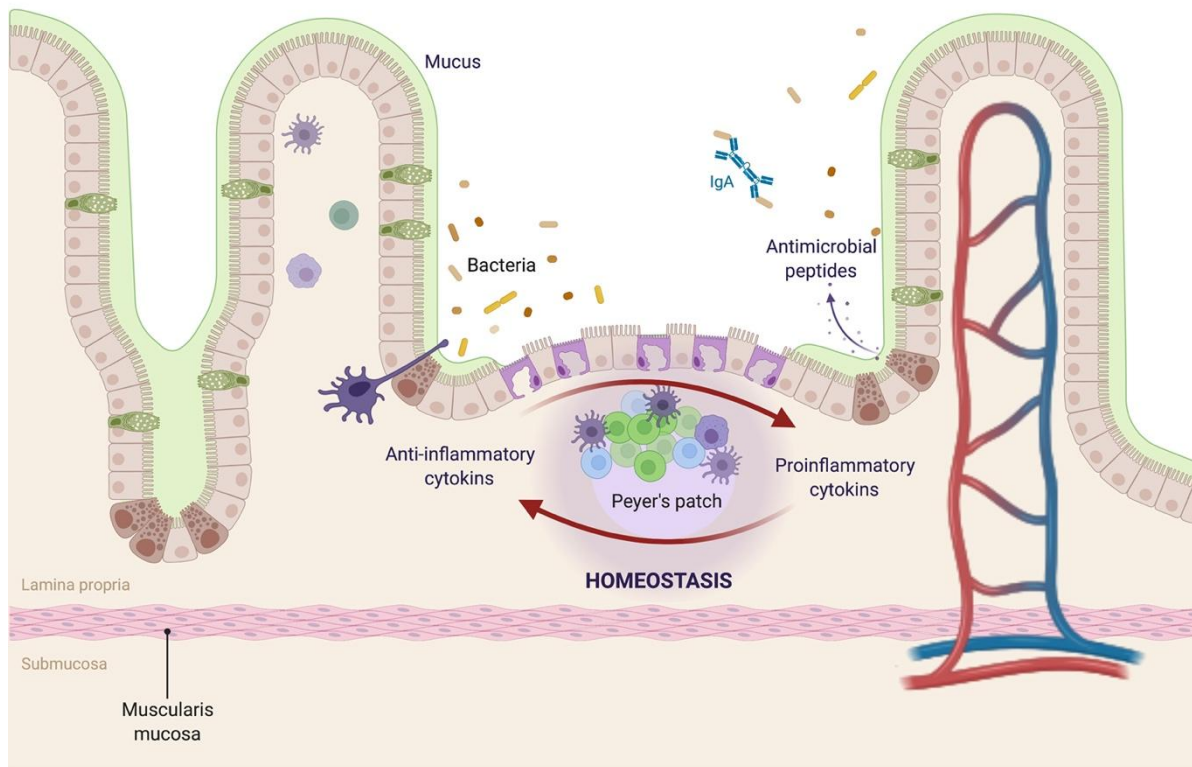
1 Over the last decade, many researchers have taken advantage of the rapid technological
2 advances in nanomedicine by designing novel DDS. The DDS aim at enhancing the drug
3 concentration in the targeted region through the control of API release or cell-specific
4 targeting strategies.¹⁵ However, many of these new approaches have failed to circumvent the
5 several hurdles of the gastro-intestinal (GI) tract microenvironment in IBD-suffering patients.
6 In this context, nanoparticles composed of polysaccharides showed promising results to
7 alleviate IBD. They combine the advantages of nanoparticulate systems with intrinsic
8 biological activities of polysaccharides. On one side, the nanoparticles offer different
9 functionalities such as mucoadhesion, mucopenetration, passive targeting of inflamed tissues
10 in the GI tract, and capture by the immune cells (For review articles on nanoparticles used for
11 IBD, see references¹⁶⁻²²). On the other hand, polysaccharides exhibit intrinsic biological
12 activity such as anti-inflammatory, healing, immunomodulation, and regulation of the
13 intestinal microbiota (For a review article on polysaccharide-based DDS for IBD, see
14 reference²³).

15 In this review, we will first discuss the up-to-date knowledge on pathophysiology of IBD
16 and its implication in the design of oral DDS. Physiological changes of the gastro-intestinal
17 tract during active IBD and their implications for designing oral DDS strategies will be
18 underscored. After that, we will analyze the undergoing research strategies using
19 nanoparticles and polysaccharides for IBD. Finally, challenges and perspectives in the oral
20 delivery of nanoparticles and polysaccharides for IBD treatment will be discussed.

21 **1. Overview of the anatomy and physiology of the colon**

22 The colon, or large intestine, constitutes the final part of the GI tract. The colon is
23 approximately 150 cm in length and is divided into six main segments.²⁴ The first segment is
24 the caecum, where the colon connects to the small intestine. Then comes the ascending colon,
25 the transverse colon, the descending colon, the sigmoid colon, and finally the rectum. The
26 appendix is another part of the colon connected to the caecum. Studies identified it as a
27 component of the immune system.²⁵ From a histological point of view, the colon comprises
28 four layers. The outermost layer that overlooks the intestinal lumen is the mucosa, then
29 comes the sub-mucosa, the muscularis, and finally the serosa.²⁶ Above the colon tissue lays a
30 thick mucus layer involved in food digestion, immunity, and protects the GI tract from
31 virulent microbes for example. On the top of the mucus layer lay millions of bacteria which
32 constitute the gut microbiota.²⁷

1 Colon histology revealed that its epithelium is arranged in crypts, where different cells are
 2 unequally distributed.²⁸ In the basement of the crypts lays the stem cells that differentiate
 3 while migrating up in the crypts to mainly four different types of functional cells (Figure 1).²⁹
 4 Enterocytes represent the most abundant cells in the colon epithelium (Figure 1). They are
 5 responsible for nutrient absorption. Goblet cells are responsible for mucus secretion.
 6 Enteroendocrine cells are involved in the secretion of GI hormones. Paneth cells oversee the
 7 secretion of antimicrobial peptides in the colon's lumen. Antimicrobial peptides protect the
 8 colon microenvironment from bacteria, viruses, fungi, and even cancerous cells.
 9 Antimicrobial peptides play a role in immunomodulation and actively participate in the
 10 immune system.³⁰ Microfold cells (M cells) are other types of cells. They are mainly
 11 distributed in the small intestine but can be found in the colon in the gut-associated lymphoid
 12 tissue (GALT) of the Peyer's patches.



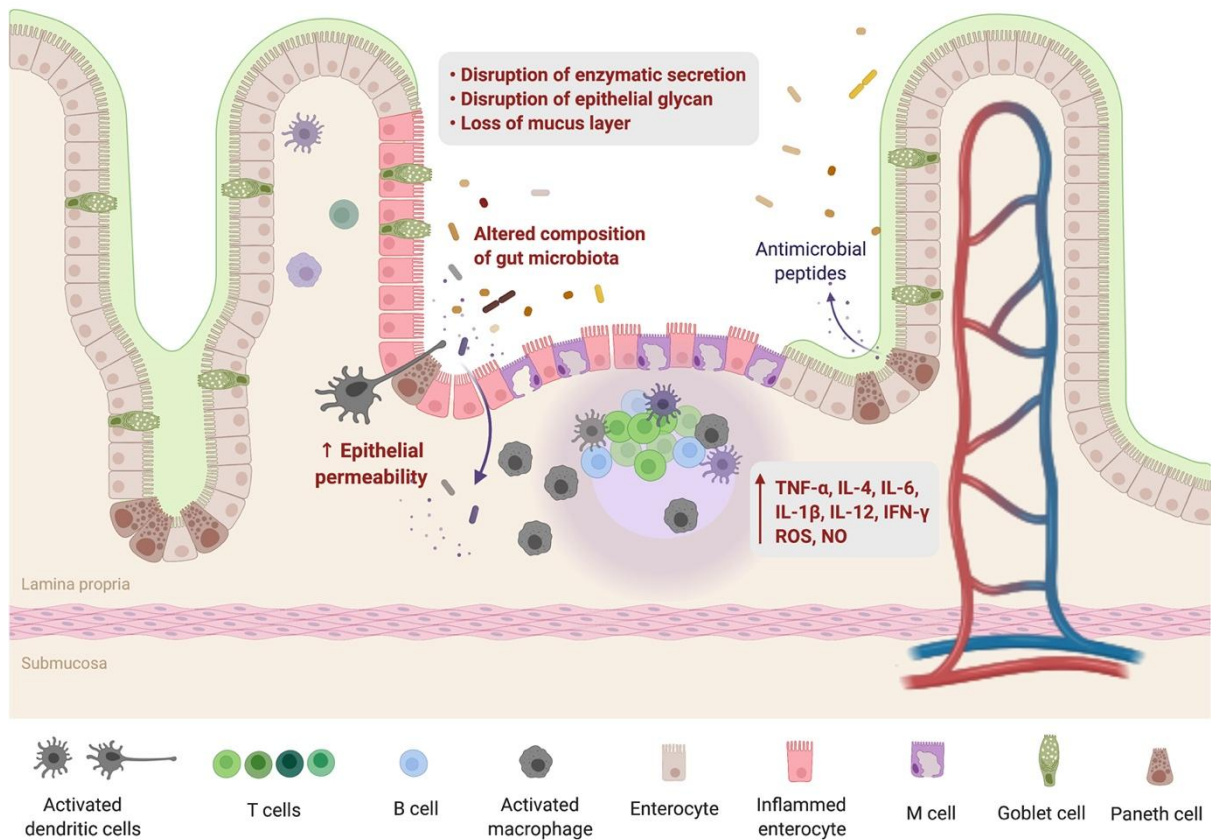
13

14 *Figure 1: Scheme summarizing the anatomy and physiology in the intestine during a homeostatic state.*

2. Pathophysiology and complications of IBD

The pathophysiology of both UC and CD is not clearly elucidated. At the same time, many gaps toward the comprehension of these diseases have been filled in the last years. These fundamental findings opened new avenues for accelerating the race toward developing new therapeutic protocols. One of the most studied areas in IBD relies on the factors predisposing its development. These triggers could be either genetic or environmental, as it is described in Table 1. Following an exposition to one of the previously described genetic or environmental triggers, the microbial peptides that are typically secreted in the intestinal lumen translocate in the lamina propria, either through dysfunction of M cells or because of an impaired barrier function (Figure 2). The barriers' function impairment also allows the translocation of luminal bacteria into the lamina propria.^{31, 32} The translocated bacteria activate the professional immune cells (e.g., dendritic cells, macrophages) that engulf the pathogens and present their epitopes to CD4 T-cells. Once activated, both CD4 T-cells and professional immune cells start releasing proinflammatory cytokines that trigger the local and systemic complications of IBD (Figure 2). Each cytokine is involved in the inflammatory response by modulating one or more specific functions. Table 2 briefly summarizes the mechanism of action of the main cytokines implicated in IBD.

IBD can trigger both local and systemic complications. Local complications are a consequence of the epithelial cells swelling and may differ according to the type of pathology. UC and CD share some similar local complications such as blood or mucus in the stool,³³ perforation of the bowel,³⁴ predisposition to colon cancer,³⁵ and loss of body weight³⁶ due to the disruption of reabsorption of water and nutrients by the intestine. However, there are some other complications that are typical for each pathology. During a CD episode, the patient may also encounter an abscess formation,³⁷ fistula,³⁸ a lymph adenopathy,³⁹ and bowel wall obstruction.⁴⁰ During a UC episode, one can observe a toxic megacolon⁴¹ as well as a loss of haustra form.⁴² The high proinflammatory cytokines level in the blood during the active phase of IBD is responsible for the various systemic complications such as conjunctivitis,⁴³ mouth ulcer,⁴⁴ the presence of an abscess in the liver,⁴⁵ portal vein thrombosis,⁴⁶ large joint arthritis,⁴⁷ and pyoderma gangrenosum.⁴⁸



1
2

Figure 2: Pathophysiological process and cytokines productions during an IBD.

Table 1: Review of the genetic and environmental factors implicated in IBD.

Factors	Expression	Pathophysiology	Ref
Genetic factors	Autophagic dysfunction	Many autophagy gene variants were linked to IBD, among which <i>ATG16L1</i> (promotes autophagosome formation and participates in suppressing Paneth cells) and <i>IRGM</i> (responsible for phagosome maturation).	⁴⁹
	Cytokine imbalance	Cytokines are essential in the modulation of the intestinal immune system. During an IBD episode, there is an imbalance between pro and anti-inflammatory cytokines, which favor the progress of the proinflammatory process.	^{50, 51}
	Bacterial imbalance	During an IBD episode, bacterial dysbiosis occurs mainly in mucosal samples. In that regard, an increase in the abundance of certain types of bacteria (e.g., <i>Enterobacteriaceae</i> , <i>Pasteurellaceae</i> , and <i>Veillonellaceae</i>) was noticed, accompanied by a decrease of other types (e.g., <i>Erysipelotrichales</i> , <i>Bacteroidales</i> , and <i>Clostridiales</i>).	⁵²
Environmental factors	Diet	The dietary habits influence the gut fatty acids equilibrium, modifying many parameters such as the pro and anti-inflammatory cytokines balance, the gut microbiota, and the intestinal permeability. For example, a study highlighted that a Mediterranean diet is less likely to induce colitis than a western diet.	^{53, 54}
	Stress	The role of the brain-gut axis in the induction of colitis was more recently studied than other factors. During a stress episode, hypothalamic secretion of corticotrophin-releasing factor (CRF) influences the secretion of cortisol by the adrenal cortex. High cortisol levels are involved in IBD by disrupting the gut microbiota, damaging intestinal barrier function.	^{11, 55}
	Non-steroids anti-inflammatory	Prostaglandins play a crucial role in the mucosal and immune defenses in the colon. During the therapy, there is selective inhibition of COX1 and COX2 receptors. These two receptors are implicated in prostaglandin production. Low levels of prostaglandins are correlated to the development or the exacerbation of colitis.	^{56, 57}
	Smoking	Cigarette smoke plays an important role in colorectal neoplasia development in IBD patients. Furthermore, smoking significantly increases the risk of developing and worsens CD.	⁵⁸⁻⁶⁰
	Antibiotic	Antibiotics favor the development of IBD through several mechanisms. They can alter the gut microbiome balance, favoring the intestinal proinflammatory phenotype. Studies reported that developing IBD is antibiotic dose-dependent, with a higher prevalence while using broad-spectrum antibiotics. From another standpoint, certain antibiotics such as ciprofloxacin or rifaximin also favor remission from an IBD by targeting specific bacteria linked with the progression of the inflammation.	^{61, 62}

Table 2: Main cytokines involved during an IBD episode.

Cytokine	Mechanism of action
TNF- α	TNF- α is a significant cytokine produced mainly by CD14 ⁺ macrophages. TNF- α plays a significant role in the pathogenesis of both UC and CD. Binding to NTF-a receptors (TNFR1 and TNFR2) activated the nuclear factor- κ B (NF- κ B) family, which plays a significant role in the regulation of genes involved in inflammation. ^{63, 64}
IL-6	IL-6 is secreted mainly by macrophages but can also be produced by CD4 ⁺ T cells. ^{65, 66} Intestinal inflammation is induced following the interaction of IL-6 with membrane glycoprotein (gp130) on the surface of CD4 T-cells. ^{67, 68} IL-6 also plays a homeostatic role as it promotes the proliferation of intestinal epithelial cells. ⁶⁹
IL-1	During the active phase of IBD, lamina propria dendritic cells and macrophages, in addition to endothelial cells, increase the production of IL-1 cytokines. There are three subtypes of IL-1 cytokines, two agonistic proteins (IL-1 α , IL-1 β), which favor inflammation, and one antagonistic protein (IL1-Ra) that reduces the inflammatory response. ⁷⁰ Although their mechanism of action is not clearly elucidated; it was shown in several studies that this higher production of IL-1 cytokines during IBD induces an imbalance between IL-1 agonists and their natural antagonist, ⁷¹⁻⁷³ which favors the progress of the inflammation in the colon.
IL-12	IL-12 cytokine family is mainly produced by dendritic cells and macrophages. IL-12 participates in the progress of inflammation after binding to its receptors (IL-12R- β 2). IL-12R- β 2 are overexpressed during inflammation by the T cells in the mucosa and the lamina propria. Accordingly, their stimulation favors a Th1 immune response, thus promoting mucosal degradation and lamina propria expansion. In addition to Th1 activation, IL-12 stimulates innate lymphoid cells (ILCs) to produce IFN- γ , as recent studies demonstrated that inhibiting IL-12 results in a lower production of IFN- γ during a CD episode. ⁷⁴ IL-23 binds its receptor (a combination of IL-12R β 1 and IL-23R ⁷⁵) and induces the formation of Th17 cells out of their naïve CD4+ T counterparts. ⁷⁶ Th17 cells have both biochemical role by the further induction of proinflammatory cytokines (IL-17, TNF- α , and IL-6), ⁷⁷ and an immunological role by promoting the activation of CD8+ T, ⁷⁸ natural killer (NK) ⁷⁹ and ILCs ⁸⁰
IFN- γ	IFN- γ is mainly produced by T _H 1-cells of the lamina propria, ^{81, 82} but also by NK cells. Its level increased during IBD. ^{83, 84} Also, it stimulates macrophages to produce proinflammatory cytokines. ⁸⁵ It was recently reported that IFN- γ worsen the pathogenesis of IBD by breaking down the vascular barrier while targeting adherents junction protein VE-cadherin. ⁸⁶
IL-4	IL-4 is a cytokine primarily produced by T _H 2 cells. Although lack of understanding of the activity of IL-4, it is known that IL-4 plays a role in promoting B-cells and T-cells. IL-4 also has a well-documented immunosuppressive effect in the intestine. ⁵⁰ Other studies depicted the role of IL-4 in the inhibition of vascular endothelial growth factor during IBD. ⁸⁷
IL-13	IL-13 is another T _H 2-related cytokine. During UC, IL-13 is implicated in the aggravation of the pathology as it impairs the epithelial barrier function while affecting the tight junctions and inducing epithelial apoptosis. ⁸⁸ It is also reported in another study that tissue fibrosis occurrence is correlated to the binding of IL-13 with a novel

	cell surface receptor IL-13 α . ⁸⁹
IL-17	IL-17 is a proinflammatory cytokine exclusively produced by T-lymphocytes (CD45RO+ cells). ⁹⁰ During an inflammatory process, IL-17 further stimulates the secretion of characteristic proinflammatory cytokines such as IL-1 β and TNF- α . ⁹¹
TGF- β	TGF- β is secreted by lymphoid and non-lymphoid cells ^{92, 93} (e.g., macrophages, fibroblast, epithelial cells). It is synthesized in an inactive form which is activated after binding latency-associated peptides. TGF- β plays a crucial regulatory role in the immune system while controlling the production of several immune systems. On the one hand, they promote the proliferation of regulatory T cells (Treg), ⁹⁴ favoring the production of anti-inflammatory cytokines (e.g., IL-10). On the other hand, they participate in the neutralization of pathogens by promoting the generation and maturation of IgA producing B-cells ⁹⁵ and boosting the activity of dendritic cells and macrophages. ^{96, 97}
IL-10	IL-10 is an anti-inflammatory cytokine. It is produced primarily by monocytes. It monitors IBD by the inhibition of proinflammatory cytokines production by T _H 1 cells. Other studies reported that, following a bacterial invasion, IL-10 expresses an inhibitory effect on the secretion of proinflammatory cytokines by Toll-like receptor-triggered myeloid lineage cells. ^{98, 99}

1

1

Table 3: Current therapeutic protocols for the monitoring of UC and CD.

	UC	CD	Ref
Active phase	<p>During an extensive UC:</p> <ul style="list-style-type: none"> - Oral 5-ASA - Oral corticosteroids <p>During a severe UC:</p> <ul style="list-style-type: none"> - Rehydration - intravenous corticosteroids - intravenous antibiotics - TNF-α antagonist (anti- TNF-α) - Surgery 	<p>Adjusting diet:</p> <ul style="list-style-type: none"> - A fiber rich meal - Avoiding excess proteins <p>During a severe CD:</p> <ul style="list-style-type: none"> - Corticosteroids - Immunosuppression - Surgery (less favorable due to the non-continuous character of CD) - TNF-α antagonist for chronic patients 	100
Cold phase	<p>Maintenance protocol consists of:</p> <ul style="list-style-type: none"> - Oral 5-ASA therapy 	<p>Maintenance protocol aims at reducing the immune response:</p> <ul style="list-style-type: none"> - Thiopurine - Methotrexate 	

2

3. General overview of conventional therapeutic agents for IBD and their shortcomings

The management of IBD aims to treat the acute attacks during the inflammatory process or prevent it from recurring during the cold phase of the disease. Depending on the type of IBD, the therapeutic protocol is different, as illustrated in table 3. For several years, the standard treatment consisted of administering steroids anti-inflammatory, immunomodulators such as thiopurines and methotrexate, cyclosporine, aminosalicylates (e.g., 5-ASA), and surgery by removing the damaged section of the intestine.

5-ASA exerts its effect through antioxidant activity and modulation of inflammatory mediators. Unfortunately, the use of 5-ASA is linked to diverse side effects (e.g., fever, nausea, diarrhea, cramping, headaches, rashes, and in some cases, hair loss, nephritis, pancreatitis, and pancytopenia). 5-ASA is rapidly absorbed from the small intestine leading to low local availability in the colon. It is thus necessary to formulate 5-ASA in adequate DDS to increase local bioavailability, lower applicable 5-ASA doses, and decrease side effects.

Corticosteroids are another standard drug class used in IBD patients. Although corticosteroids provide a robust anti-inflammatory response, their use is commonly related to strong side effects that arise only a few weeks following the start of a topical or systemic treatment.¹⁰¹⁻¹⁰⁴ Those side effects (e.g., Cushing's syndrome, infection, adrenal suppression, sleep disorders, osteoporosis, and renal function impairment) limit their application in long-term therapy. Recent endeavors suggest that using nanoparticles to deliver corticosteroids during an IBD allows keeping their steroid anti-inflammatory response while minimizing the systemic side effects by targeted delivery of the API into the inflamed colon. From another perspective, certain corticosteroids (e.g., budesonide) have a low mucus penetrating capability, justifying the use of nanoparticles to enhance their penetrability.¹⁰⁵

Immunosuppressants can target specific proteins in the body that induce inflammation. Monoclonal antibodies targeting specific cytokines were used to monitor IBD. Indeed, anti-TNF- α (e.g., infliximab, adalimumab, golimumab) constitute the mainstream of biological treatment during the active phase of IBD and as maintenance therapy.¹⁰⁶ Also, anti-IL-12 (e.g., ustekinumab) is used only in the case of CD.¹⁰⁷ One major drawback of biologics protocol is the primary nonresponse or a loss of response during treatments. The best example would be anti-TNF- α based therapy, where 30% of the patients present a primary nonresponse,¹⁰⁸ while 30 to 50% lost the response over time.¹⁰⁹ Up to date, many biological-

1 based therapeutics are available in the market (e.g., certolizumab, infliximab, adalimumab,
2 natalizumab, vedolizumab). Nonetheless, the administration of biologics is commonly applied
3 by the parenteral routes (e.g., subcutaneous, intramuscular, or intravenous), and their long-
4 lasting use is associated with numerous side effects such as immunosuppression,¹¹⁰⁻¹¹³
5 infection,¹¹⁴ cancer,¹¹⁵ and the formation of antibodies against biologics, reducing their
6 efficacy. Consequently, a novel drug delivery strategy relying on nanoparticles has been more
7 and more investigated in the last year to administrate the API orally, besides reducing their
8 dose through a more targeted delivery to the inflamed regions.¹¹⁶⁻¹¹⁸

9 Using nanoparticles for the oral delivery of immunosuppressants to IBD patients is a
10 promising approach to reduce their side effects. Indeed, monitoring the aberrant balance of the
11 immune system during UC or CD relies on controlling various immune cells, enzymes, and
12 cytokines through the administration of immunosuppressants (e.g., methotrexate, cyclosporine
13 A, tacrolimus). However, their poor solubility often limits their performance,¹¹⁹⁻¹²¹ urging to
14 increase the administered dose. Also, misusing these therapeutic agents and systemic
15 administration led to off-target severe side effects (e.g., immunodeficiency, allergies, a loss of
16 activity following a long systemic circulation). Nanoparticle development offers the
17 possibility of selectively targeting the colon's inflamed region and a controlled release of the
18 immunosuppressant.¹²² They also play a role in protecting the drugs from degradation during
19 their journey in the GI tract.^{123, 124} Table 4 summarizes the significant works in designing
20 nanoparticle-based IBD therapy.

4. Implication of pathophysiological changes of the GI tract during active IBD for oral drug delivery strategies

Active IBD could induce several changes in the GI tract barriers that should be fully understood and considered for developing efficient DDS and identifying therapeutic targets.¹²⁵⁻¹²⁷ Those changes concern the pH, degradative enzymes,¹²⁷ mucus barrier function, microbiota composition^{20, 128} and the permeability of the intestinal epithelium.¹²⁹

4.1. pH changes

In healthy subjects, the stomach pH ranges from 1.0 to 2.5 (fasted). It rises from 6.6 to 7.5 in the small and the large intestine. More precisely, luminal pH in the proximal small bowel ranges from 5.5 – 7.0 and gradually rises to 6.5 – 7.5 in the distal ileum. In the caecum, the pH decreases to 5.5 – 7.5 and rises from 6.87 in the proximal colon to 7.2 in the distal colon. In UC and CD patients, the pH of the stomach was found higher than in healthy subjects.¹³⁰ This change was correlated to a decrease in the acidic secretions of the stomach. In UC and CD patients, no significant changes in the pH of the small intestine were reported compared to healthy subjects.¹³⁰⁻¹³² Although the colonic pH of patients with IBD broadly fluctuates depending on the individuals and the disease state, patients are generally subject to acidic pH in the colon.^{133, 134} The pH in UC and CD patients ranged from 5.5 to as low as 2.3.^{133, 135, 136} The decrease in the colonic pH is due to disruption of factors including intestinal volume, transit time, microbial fermentation, bile acid metabolism of fatty acids, bicarbonate, and lactate secretions.¹³⁴ Changes in colonic pH in UC and CD patients could affect the composition of the microbiome and consequently the colonic transit. The residence time of the drugs in the colon, drug release, and absorption are thus modified. Additionally, pH changes could affect drug release from pH-sensitive formulations.

The pH change along the GI tract was exploited to design DDS able to disintegrate at a specific pH range to release the API. A general strategy consists in coating the DDS with polymers that are pH-sensitive. Polymers used for colon delivery are methacrylic acid, polymethacrylic acid, polyacrylic acid, and their derivatives which respond to the high pH of the colon (≥ 6.8).¹⁹ So far, the most common pH-sensitive systems for oral drug delivery in IBD treatment are composed of methacrylic acid co-polymers (Eudragit[®]). Eudragit S100 (ES100), which should dissolve in the distal intestine at a pH of 7, is the most used polymer for colon drug delivery. ES100-containing DDS exhibited pH-sensitive properties by limiting the burst release at the gastric pH. Most of the drug was released at pH higher than 7.^{137, 138}

1 The pH at which the drug is released could be adjusted to lower values by mixing ES100 to
2 other methacrylic acid co-polymers that dissolve at low pH.¹³⁹ Eudragit L100 is soluble at pH
3 6, while EL30-D55 was used to manipulate the drug release within the desired pH range of
4 5.5 – 7.0 in the distal small intestine. The appropriate colon dissolution pH of delivery
5 systems can be controlled by optimizing the ratio of EL30-D55 and ES100 for targeted drug
6 delivery to the inflamed colon.¹³⁹ More recently, coating nanoparticulate DDS with Eudragit
7 FS 30D conferred a sustained release of the encapsulated drugs.¹⁴⁰ Indeed, Eudragit FS 30D
8 dissolves similarly to Eudragit S100 at pH 7 but in a more sustained and controlled mode. The
9 Eudragit-coated strategy of nanoparticulate DDS was applied for many types of
10 nanostructures,¹⁹ including polymer,¹⁴¹ lipidic,¹⁴² polysaccharide-based,¹⁴³ or inorganic
11 nanoparticles.¹³⁸

12 Other types of pH-sensitive materials were used for designing nanoparticles that can target
13 and remain in the inflamed colonic tissue through multiple mechanisms. Hybrid nanoparticles
14 composed of pH and enzyme-sensitive polymers were designed by combining ES100 and
15 azopolyurethane polymers.^{144, 145} However, as mentioned above, the colonic pH in patients
16 with IBD could be more acidic than in healthy subjects. Consequently, the DDS coated with
17 Eudragit that dissolves at pH higher than 6 might not release the drug in vivo. Indeed, in most
18 investigations on Eudragit-based nanoparticles, the drug release was evaluated in vitro in
19 simulated colonic media without mimicking IBD disorders. In the next section, we will
20 discuss the interest of polysaccharides for designing pH-sensitive DDS that could be an
21 alternative to synthetic polymers.

22 **4.2. Mucus**

23 In healthy subjects, mucus is a viscoelastic hydrogel composed of water (90 – 98 wt%),
24 mucins (2 – 5 wt%), lipids (1 – 2 wt% primarily associated with hydrophobic domains of
25 mucin glycoproteins), mineral salts (2 wt%), and proteins (immunoglobulins A and M,
26 lysozyme) that contribute to the mucus elasticity.^{146, 147} The degree of mucus hydration is a
27 significant determinant of mucus viscoelasticity. Mucin fibers (typically 10 – 40 MDa ,
28 3 – 10 nm in diameter)¹⁴⁸ are crosslinked and entangled macromolecules secreted by both
29 goblet cells and the seromucinous glands of the lamina propria at the apical epithelium.¹⁴⁹
30 Mucins are proteins glycosylated by *O*-linked *N*-acetyl galactosamine and *N*-linked sulfate-
31 bearing glycans.¹⁵⁰ Dense glycan coverage of mucins represents up to 80% of the dry weight
32 of mucus. For 100 amino acid residues, carbohydrate chains represent 25 – 30,¹⁵¹ and
33 contribute up to 80% of the dry weight of mucus (For review articles on mucus, see

1 references¹⁵²⁻¹⁵⁴). The glycocalyx that protects cell surfaces comprises cell-associated mucins
2 and mucus gels formed by secreted gel-forming mucins.¹⁵³

3 Significant changes in mucus composition, thickness, physical properties, and function
4 were reported in IBD. CD is characterized by goblet cell hypertrophy, while patients with UC
5 are characterized by a reduction and depletion of goblet cells. In this latter case, mucus
6 production is significantly decreased, and the thickness of mucus layers was reduced in the
7 colon and the rectum.^{155, 156} In UC, the substantial increase in mucus production leads to a
8 thicker mucus layer in the inflamed areas. Consequently, a higher amount of nanoparticle
9 adherence was reported for inflamed areas compared to healthy mucosa. Due to their small
10 size, nanoparticles were better retained in the intestinal tract than large particles. Smaller
11 nanoparticles are known to exhibit better mucoadhesion and mucopenetration compared to
12 larger particles.^{157, 158} Lamprecht et al.,¹⁵⁹ demonstrated an inverse relationship between
13 particle size and binding to the inflamed intestinal mucosa. They revealed that particles
14 smaller than 100 nm exhibited the highest binding affinity to inflamed colonic tissue. The
15 reduction of the DDS size to the nanometer scale improved colonic residence time in inflamed
16 regions of the intestine and provided benefits for IBD treatment. Particles with a size larger
17 than 200 μm are subjected to diarrhea symptoms, resulting in a decreased GI transit time and,
18 therefore, decreased efficiency.^{160, 161}

19 PEGylation strategy improves mucopenetration properties of nanoparticles. Being
20 hydrophilic and uncharged, the presence of poly(ethylene glycol) (PEG) confers to
21 nanoparticulate DDS mucus penetrating properties.^{157, 162-164} The behaviors of PEG nano-
22 delivery systems toward mucus are dependent on the molecular weight. Low PEG molecular
23 weight (2-5 kDa) provided a non-mucoadhesive coating on nanoparticles. Whereas coatings
24 using 10 kDa PEG resulted in strong particle mucoadhesion.^{163, 164} The surface charge of
25 PEGylated nanoparticles showed a significant impact on their accumulation in the inflamed
26 colon of a dextran sulfate sodium (DSS)-induced mice model of acute UC.¹⁶⁵ Nanoparticles
27 had a 100-nm diameter and were composed of poly(ethylene glycol)_{5k}-*b*-poly(lactic-co-
28 glycolic acid)_{10k} (PEG_{5k}-*b*-PLGA_{10k}). They were loaded with TNF- α siRNA.¹⁶⁵ In this study,
29 cationic PEGylated PLGA nanoparticles showed a higher accumulation in the colon and a
30 better silencing capacity than nanoparticles with a neutral charge.

31 In patients with active UC, the carbohydrate content of the epithelial glycan, including
32 mucus glycoproteins, is altered compared with healthy controls (For reviews on epithelial
33 glycans in IBD, see references¹⁶⁶⁻¹⁶⁸). Epithelial glycans are a major component of the

1 intestinal mucosa. Glycans typically form glycoconjugates (e.g., glycoproteins and
2 glycolipids) by attaching to other molecules (e.g., cytosolic lipids or membrane-associated
3 lipids and proteins). Thanks to their position at the interface between the epithelial cells and
4 the outer mucus layer, glycans serve as attachment sites for nutrients and ligand-binding
5 proteins (e.g., antibodies, lectins). They also play an essential role in the interaction between
6 intestinal epithelia and the commensal flora in the mucus. The major class of glycan in the gut
7 (80 wt% of MUC2) is composed of mucin-type O-glycans.

8 In patients with active disease, intestinal epithelial glycosylation is disrupted.¹⁶⁷⁻¹⁶⁹
9 Alterations include truncated mucin-type O-glycans and a reduction of oligosaccharide chain
10 length and sulfatation, conferring less negative charge to mucins.^{169, 170} Reduced sulfatation
11 was correlated to increased activity of mucin sulfatase. Structural and immunohistochemical
12 studies revealed that patients with active IBD were characterized by a simplified O-
13 glycome.¹⁷¹ The amount of smaller glycans was increased, while more complex structures
14 were lowered. Glycan disruption altered intestinal immunity, disrupted glycan-lectin and host-
15 microorganism interactions, and altered MUC2 synthesis¹⁶⁸ and stability. Glycome
16 simplification makes mucins more accessible to host and bacterial proteases.¹⁷² The loss of
17 mucus viscoelasticity and reduction of barrier properties lead to enhanced interactions of
18 bacteria and intestinal epithelia. Furthermore, a significant correlation was found between
19 changes in glycosylation and the level of inflammation.¹⁶⁹

20 Understanding the disorders of epithelial glycosylation in IBD patients with active disease
21 is fundamental for developing strategies for ‘remodeling’ the disease-associated glycan by
22 targeting intestinal epithelial glycosylation. Those approaches include direct methods (e.g.,
23 inhibitors of glycosidases,¹⁷³ or sulfatases^{174, 175}) and indirect methods (e.g., normalizing the
24 gut microbiota by faecal transplantation or prebiotics, or by bacterial products). Inhibiting
25 protein-glycan interactions that contribute to recruiting pathogenic bacteria or inflammatory
26 cells seems to be a promising strategy to alleviate inflammation in IBD patients.¹⁶⁸ For
27 example, galactose interferes with the interaction of *Fusobacterium nucleatum* (a Gram-
28 negative commensal bacteria) with the carbohydrate epitope galactose or *N*-acetyl-
29 galactosamine (Gal, GalNAc).^{176, 177} It was found that the interaction of *F. nucleatum* Fap-2
30 (fibroblast activation protein 2), and Gal or GalNAc was implicated in numerous diseases,
31 including IBD and cancer.¹⁷⁸ These findings should trigger further investigations to design
32 DDS to block Fap-2/Gal or Fap-2/GalNAc interaction. Similarly, it was demonstrated that
33 blocking the interaction of *Escherichia coli* FimH lectin with epithelial glycan prevents

1 mucosal inflammation associated with CD.¹⁷⁹ Knowing that mannose derivatives are able to
2 interact with FimH, several research works were dedicated to developing mannose derivatives
3 to alleviate inflammation. Interestingly, the FimH blocker Sibofimloc (EB8018/TAK-018)
4 derived from the patent WO2014100158A1 has entered the clinical trials for CD.¹⁸⁰ Thanks to
5 safety and pharmacokinetic results obtained in 2020,¹⁸⁰ the clinical development program is
6 continuing with a phase 2 trial in patients with CD.

7 On the other hand, ulcerated tissues contain high concentrations of positively charged
8 proteins that increase the affinity to negatively charged substances.¹⁸¹ In particular, inflamed
9 colon contains high concentrations of transferrin and eosinophil cationic protein.¹⁸²⁻¹⁸⁵ This
10 property was exploited in designing negatively charged DDS that preferentially adhered
11 inflamed tissue *in vivo* through electrostatic interactions with positively charged proteins.^{159,}
12 ^{186, 187} However, conflicting results were obtained in the literature on the impact of
13 nanoparticle charge on their accumulation in the inflamed colon since the net charge of mucus
14 remains negative due to the presence of mucins. Iqbal et al.,¹⁶⁵ revealed that cationic
15 nanoparticles exhibited remarkably higher accumulation in the inflamed colon compared to
16 anionic nanoparticles.¹⁶⁵

17 **4.3. Enzymatic secretions**

18 In CD patients, the enzymatic secretion was significantly modified in comparison with
19 healthy subjects. Indeed, for digestive purposes, the lumen of the upper GI tract contains large
20 amounts of pancreatic proteases, but studies have also demonstrated increased proteolytic
21 activity into mucosal tissues (both in the upper and lower GI tract), associated with
22 pathological conditions such as IBD. This upregulation was correlated with the degradation of
23 tight junctions.¹⁸⁸ In that context, protease inhibition was used as a therapeutic approach in
24 IBD treatment.¹⁸⁸ Additionally, increased fluid secretion can dilute the digestive enzymes
25 implicated in intestinal transit.

26 **4.4. Epithelial barrier**

27 As detailed above, in IBD patients, the intestinal epithelium is characterized by epithelial
28 defects, including a loss of continuous epithelium,¹⁸⁹ TNF- α mediated epithelial apoptosis in
29 the colon,¹⁹⁰ and damage of epithelial tight junctions. In particular, abnormal expression of
30 tight junction proteins (e.g., occluding, ZO-1 and claudins)¹⁹¹ is an essential characteristic in
31 IBD. They lead to increased permeability of colonic epithelial mucosa and a loss of the barrier
32 function of the mucosa toward microbes, toxic substances such as lipoproteins, causing

1 inflammatory response. A consequence of epithelial shedding and impaired paracellular
2 epithelial barrier in IBD was exploited by different research groups to passively target API
3 and nanoparticles to the inflamed tissue through the so-called epithelial enhanced
4 permeability and retention (eEPR) effect. A study implied that the recurrence of budesonide-
5 loaded PLGA/Eudragit nanoparticles improved the anti-inflammatory effect of budesonide.¹⁹²
6 The authors suggest that these observations could be linked to the high adhesion and uptake of
7 the nanoparticles mediated by the eEPR effect, which increased the levels of budesonides in
8 the inflamed site.

9 In a more recent investigation, Ahmad et al.¹⁹³ used polycaprolactone (PCL) covalently
10 grafted with aminocellulose and used to prepare nanoparticles loaded with anti-inflammatory
11 drugs (e.g., glycyrrhizic acid, budesonide). Nanoparticles designed by solvent evaporation had
12 a mean hydrodynamic diameter of ~230 nm and positive zeta potential (24 to 29 mV). This
13 study revealed that nanoparticles had a preferential accumulation in the inflamed colon in
14 mice with DSS-induced colitis. Higher accumulation in the inflamed colon could be due to
15 passive targeting by eEPR since no accumulation of nanoparticles in a healthy colon was
16 reported.¹⁹³ These nanoparticles improved disease activities like occult blood in the stool,
17 length of the colon, and fecal properties. Also, the nanoparticles decreased colonic mast
18 cellular infiltration, significantly maintained mucin protection, improved histological features
19 of the colon, and ameliorated the markers of inflammation (e.g., induced nitric oxide (NO)
20 synthetase (iNOS), cyclooxygenase (COX)-2, IL1- β , TNF- α , and myeloperoxidase
21 (MPO)).¹⁹³

22 **4.5. Modifications of immune cell surface receptors**

23 In inflamed tissues, the immune cells are subject to different modifications of the
24 expression of cell receptors. In the inflamed state of UC, the glycoprotein CD98 is
25 overexpressed by colonic epithelial cells and macrophages. CD44 receptor is another
26 glycoprotein overexpressed on the activated inflammatory cells in colitis tissues. Active
27 targeting by nanoparticulate DDS consists of the intentional orientation of the localization of
28 nanoparticles to inflamed tissues.^{194, 195} This results from a high concentration of
29 nanoparticles in the disease site and consequently high therapeutic efficacy of a drug, while
30 the adverse effects on normal tissue are reduced.^{195, 196}

31 Another strategy targets receptors naturally expressed by macrophages and dendritic cells.
32 Because macrophages and dendritic cells are abundant in inflamed tissues, targeting receptors

Table 4: Table summarizing the recent work on the use of nanoparticles for alleviating IBD.

1 (e.g., dectin-1, galactose agglutinin, and mannose receptor) promoted the distribution of
2 targeted nanoparticles in the inflamed tissues.¹⁹⁷⁻²⁰¹ The ligands that can target receptors on
3 macrophages include saccharides (e.g., mannose,¹⁹⁸⁻²⁰⁰ hyaluronan, galactose²⁰¹), dectin-1
4 able to recognize β -glucan, lectins, or monoclonal antibodies.

5 **4.6. Production of reactive oxygen species**

6 In response to inflammatory stimuli through specialized enzyme complexes called
7 NADPH oxidase, reactive oxygen species (ROS) can be produced by epithelial cells and
8 innate immune cells, including resident monocytes/macrophages, dendritic cells, or infiltrated
9 neutrophils. Those cells are able to produce ROS into the mucus and intestinal lumen.²⁰² The
10 intestinal mucosa of patients with IBD is characterized by severe disorders of antioxidants and
11 ROS overexpression by immune cells (e.g., neutrophils and macrophages), leading to
12 oxidative damage. ROS concentration was up to 100-fold higher than in control patients.²⁰³ In
13 this context, several researchers exploited this property to design ROS-scavenger DDS able to
14 reduce oxidative stress and inflammatory response. Those strategies include colon-targeted
15 delivery of nanoparticles containing donors of NO radicals (e.g., NO-nitrosothiols) or
16 antioxidant molecules (e.g., bilirubin, curcumin, ginger, resveratrol).

API	Materials composing the nanoparticles	Inflammation model	Delivery route	Observations	Ref
Corticosteroids (budesonide)	PLGA/ Eudragit S100	Trinitrobenzenesulfonic acid (TNBS) (mice) DSS (mice) Oxazolone (mice)	Oral	Comparing budesonide-loaded nanoparticles to the free drug revealed: <ul style="list-style-type: none"> - A better histological, biological, and endoscopic data regarding inflammation reduction while using nanoparticles. - Nanoparticles could be tuned into pH-responsive nanosystems allowing targeted delivery, considering the acidic pH environment of the inflamed area. 	141
Corticosteroids (Budesonide)	PLGA and Eudragit S100	TNBS (rats)	Oral	<ul style="list-style-type: none"> - Budesonide-loaded nanospheres had a higher impact on the reduction of TNBS induced colitis. - The inflammation reduction could be linked to the inflamed mucosa's higher systemic availability and cellular uptake of nanospheres. 	192
Corticosteroids (Budesonide)	Mannosylated nanostructured lipid carrier system (Mn-NLCs)	Oxazolone (mice)	Oral	The in vivo data following the administration of a budesonide-loaded Mn-NLCs and a budesonide suspension revealed: <ul style="list-style-type: none"> - A lower clinical, macroscopic, and microscopic scoring with the budesonide Mn-NLCs treated group. - A lower level of proinflammatory cytokines and myeloperoxidase. 	204
Corticosteroids (Prednisolone)	ϵ - polylysine-coated mesoporous silica	RAW 264 cells Caco-2 cells	NA	<ul style="list-style-type: none"> - A high inhibition of inflammation was observed at the cellular level. 	205
Corticosteroids (Betamethasone)	Lectin-conjugated PLGA	TNBS (mice)	Oral	<ul style="list-style-type: none"> - Lectin-decorated PLGA-nanoparticles selectivity increased adhesion to inflamed tissue compared to plain nanoparticles, thus improving their therapeutic effect. 	206
5-ASA (Mesalazine)	Polycaprolactone	TNBS (mice)	Oral	<ul style="list-style-type: none"> - A higher therapeutic effect of encapsulated 5-ASA was observed compared to the free drug. 	207
5-ASA (Mesalazine)	Silica	TNBS (mice)	Oral	The use of silica nanoparticles to deliver mesalazine allowed to: <ul style="list-style-type: none"> - Selectively target the inflamed tissue to deliver the API and spare the healthy tissue. - A higher anti-inflammatory effect was obtained with a lower dose of the API. 	208

5-ASA	Silicon dioxide (SiO ₂)	DSS (mice)	Oral	<ul style="list-style-type: none"> - A lower inflammation score was observed with the groups treated by the nanoparticles or with high doses of 5-ASA. - Myeloperoxidase and cytokines (TNF-α and IL-6) levels were also much lower with the 5-ASA-nanoparticles treated group suggesting a higher colon-targeting efficacy of 5-ASA-nanoparticles. 	209
5-ASA (Mesalazine)	Redox particles	DSS (mice)	Oral	<ul style="list-style-type: none"> - Tempol or mesalamine-loaded nanoparticles reduced the inflammation strongly compared with free tempol or mesalazine. - The nanosystems specifically accumulated in the colon during a colitis episode. 	210
Tempol					
Tempol	Methoxy-poly(ethylene glycol)-b-poly [4-(2,2,6,6-tetramethylpiperidine-1-oxyl)oxymethylstyrene] (MeO-PEG-b-PMOT) block copolymers	DSS (mice)	Oral	<ul style="list-style-type: none"> - Following a DSS-induced inflammation, an increase of commensal bacteria (<i>Escherichia coli</i>, <i>Staphylococcus spp.</i>) was noticed. - The nanoparticle administration reduced the prevalence of these commensal bacteria. 	211
Biologics (Anti-TNF- α)	Natural polyphenol tannic acid and poly(ethylene glycol)	DSS (mice)	Oral	<ul style="list-style-type: none"> - The oral administration of anti-TNF-α nanoparticles had a significantly higher reduction of inflammation than free anti-TNF-α. 	212
Biologics (TNF- α siRNA)	Thioketal	DSS (mice)	Oral	<ul style="list-style-type: none"> - Using thioketal nanoparticles as a carrier to orally deliver siRNA against TNF-α into the colon reduces the TNF-α RNA level in the inflamed region, preventing the worsening of UC. 	213
Biologics (TNF- α siRNA)	Poly(ethylene glycol)-block-PLGA <ul style="list-style-type: none"> - Cationic charge: aminated particles - Anionic charge: carboxylate particles - Neutral charge: plain particles 	DSS (mice)	Oral	<ul style="list-style-type: none"> - Aminated nanoparticles expressed the highest accumulation in the colon, translated by increased inhibition of TNF-α secretion and mRNA levels. - Animals treated with aminated nanoparticles had similar histological scores to healthy mice. - The results suggest the significant influence of surface charge on colonic accumulation. 	165
Biologics (CD98 siRNA)	polyethyleneimine (PEI)	DSS (mice)	Oral	<ul style="list-style-type: none"> - Significant reduction of colitis occurs through CD98 downregulation in the intestinal 	214

				epithelial cells.	
Biologics (CD98 siRNA) + Curcumin	PLGA	DSS (mice)	Oral	<ul style="list-style-type: none"> - PLGA nanoparticles offered a tuneable platform to deliver CD98 siRNA and curcumin in the inflamed colon selectively. - This nanoparticle-based therapy showed promising results in the monitoring of IBD. 	215
Curcumin	PLGA/Eudragit S100	DSS (mice)	Oral	<p>While compared to groups treated with non-encapsulated curcumin was used, it was noticed:</p> <ul style="list-style-type: none"> - Decrease of immune cells infiltration. - Reduction of TNF-α levels. 	216
Curcumin + celecoxib	Eudragit S100	TNBS (mice)	Oral	<p>Eudragit nanoparticles showed the following events:</p> <ul style="list-style-type: none"> - Targeted delivery to the colon. - Reduced body weight loss and diarrheic stool. - When delivered in a nanoparticulate form, much lower doses of API were needed to obtain the desired effect than a separate administration of drugs. 	217
Peptides (Lysine-proline-valine) KPV	Alginate and chitosan	DSS (mice)	Oral	<p>KPV-loaded nanoparticles reduced the anti-inflammatory response in DSS pre-treated mice. KPV loaded into nanoparticles can be delivered at a concentration 12,000-fold-lower than a free solution of KPV.</p>	218
Immuno-modulator (Rolipram)	PLGA	TNBS (mice)	Oral	<ul style="list-style-type: none"> - The nanoparticle formulation expressed a similar anti-inflammatory effect to the drug in solution. - Five days post-treatment, the animals that received the nanoparticle formulation showed less relapse than those that received the drug solution. - The animals that received nanoparticle formulation suffered less from the adverse effect caused by rolipram. 	219
Immuno-modulator (Tacrolimus)	PLGA, Eudragit P-4135F (pH-sensitive)	DSS (mice)	Oral	<ul style="list-style-type: none"> - Oral tacrolimus-loaded nanoparticles attenuated colitis more efficiently than oral tacrolimus solution. - Lower efficiency of oral tacrolimus-loaded nanoparticles was observed than the drug solution administered subcutaneously. - Encapsulating tacrolimus inside nanoparticles reduced its adverse 	220

				<p>effect.</p> <ul style="list-style-type: none"> - Both nanoparticles (PLGA and pH-sensitive Eudragit) showed similar data in reducing colon inflammation. Nonetheless, pH-sensitive nanoparticles were slightly less nephrotoxic. 	
<p>Immuno-modulator (Cyclosporine A) CYA</p>	Lipids	DSS (mice)	Oral	<ul style="list-style-type: none"> - While compared to an oral administration of a commercial solution of Sandimmun (Neoral[®]), CYA-loaded lipid nanoparticles did not improve the therapeutic effect. 	221
	PLGA	DSS (mice)	Oral	<p>Compared to drug-free particles and commercial formulation Sandimmun (Neoral[®]), CYA-loaded PLGA nanoparticle exhibited:</p> <ul style="list-style-type: none"> - An unchanged body weight. - The concentration of proinflammatory cytokine in the plasma was undetectable. <p>The authors concluded that CYA-loaded PLGA nanoparticles yielded a similar reduction of inflammation at a half dose of the commercial product.</p>	222
<p>Immuno-modulator (Methotrexate)</p>	Grapefruit-derived nanovesicles (GDNs)	DSS (mice)	Oral	<p>After their selective internalization by macrophages, GDNs decrease the inflammation by:</p> <ul style="list-style-type: none"> - Upregulation of heme oxygenase-1 (HO-1) expression. - Inhibition of IL-1β and TNF-α production - Loading methotrexate in GDNs further enhanced their anti-inflammatory property while maintaining the homeostasis of intestinal macrophages. 	223
<p>Immuno-modulator (Azathioprine)</p>	Chitosan beads	Acetic acid (rabbit)	Oral	<p>Azathioprine has numerous side effects, such as hepatotoxicity, bone marrow suppression, and allergic reaction. The oral administration of Azathioprine-chitosan beads allowed to:</p> <ul style="list-style-type: none"> - Reduce the systemic side effects of azathioprine. - Decrease the levels of myeloperoxidase and proinflammatory cytokines to the levels of healthy animals. - Restore the microscopic structure of the colon. 	224
<p>Antibiotics (Rifaximin)</p>	Chitosan	<i>In vitro</i>	NA	<p>Encapsulating the drug into chitosan nanoparticles expressed the following</p>	225

				<p>data:</p> <ul style="list-style-type: none"> - Improved the solubility of rifaximin. - Expressed high stability of the formulation. - The formulation offers the possibility of a controlled release of the drug in the targeted region. <p>Nevertheless, <i>in vivo</i> investigation still lacks to confirm these observations.</p>	
Probiotic extract	Chitosan-coated PLGA	TNBS (rats)	Oral	<ul style="list-style-type: none"> - The administration of probiotic extract and nanoparticles significantly mitigated colitis. - The medium dose of probiotic extract-loaded nanoparticles reduced the inflammation more efficiently than the high dose of its free administered counterpart. 	226
Prohibitin	Ply(lactic acid) (PLA)	DSS (mice)	Oral	<ul style="list-style-type: none"> - Prohibitin has an anti-inflammatory property, but its levels decrease during an IBD episode. - Oral prohibitin-loaded PLA particles or rectal prohibitin-loaded adenovirus increased the prohibitin levels in the inflamed colonic region. - Increased prohibitin levels correlated with decreased inflammation intensity translated by lower clinical and histological scores, reduced myeloperoxidase activity, proinflammatory cytokines, and protein carbonyl content. 	227
	Adenovirus		enema		
Ginger active compound (6-shogaol)	<p>Poly(lactic-co-glycolic acid)/poly(lactic acid)-polyethylene glycol-folic acid</p> <p>PLGA/PLA-PEG-FA</p>	DSS (mice)	Oral	<p>Oral delivery of 6-shogaol loaded into nanoparticles reduced the virulence of the colitis by:</p> <ul style="list-style-type: none"> - Diminishing the levels of proinflammatory cytokines (TNF-α, IL-6, IL-1β) - Increasing the level of anti-inflammatory factors such as the nuclear factor (erythroid-derived 2)-like 2 (Nrf-2) and HO-1. - Accelerating wound repair 	228

1

2

5. Polysaccharide-based drug delivery systems for IBD

3

Polysaccharides are naturally produced by animals, plants, microorganisms, fungus, and

4

algae. Among the polysaccharides' diverse intrinsic biological activities (e.g.,

1 immunomodulatory, anti-inflammatory, regulation of intestinal flora imbalance, and healing
2 properties), their use to alleviate IBD has gained significant interest over the last years.^{23, 229}
3 Table 5 summarizes the main polysaccharides studied for their role in IBD modulation.
4 Polysaccharides manage IBD by different mechanisms. From one side, they can regulate the
5 different proinflammatory signalling pathways such as Toll-like receptors (TLR), mitogen-
6 activated protein kinase (MAPK), nuclear factor- κ B (NF- κ B), and G-protein coupled
7 receptors. On the other side, polysaccharides play a crucial role in the intestinal flora balance.
8 They increase the symbiotic bacteria (e.g., *Lactobacillus*) and decrease the pathogenic
9 bacteria (e.g., *Facklamia*, *Clostridium*, and *Enterococcus* species).^{230, 231} Polysaccharide
10 metabolites, known as short-chain fatty acids (SCFAs); can also modulate the homeostasis of
11 the GI tract as several described their role in the inhibition of pathogenic microorganisms.²³²⁻
12 ²³⁴ Acetic acid, propionic acid, and butyric acid are the main SCFAs. Lactic acid and valeric
13 acid are also metabolites produced by microbial fermentation of undigested carbohydrates and
14 dietary fibers.²³⁵⁻²³⁷ SCFAs decrease the pH of the intestine. They improve the homeostasis of
15 the intestinal flora by inhibiting the growth of pathogenic bacteria and enabling the
16 proliferation of beneficial bacteria.²³⁸ SCFAs also have direct antimicrobial activity against
17 bacterial pathogens by diffusing across the bacterial membrane and reducing the intracellular
18 pH. In particular, butyrate has a positive effect on epithelial integrity and tight junction
19 permeability.^{229, 239} Butyrate enhances the intestinal barrier by facilitating tight junction
20 assembly via activation of AMP-activated protein kinase²⁴⁰ and up-regulating the tight
21 junction protein Claudin-1 transcription.²⁴¹ Additionally, butyrate increased mRNA
22 expression and protein abundance of claudins-3 and 4 and influenced intracellular ATP
23 concentration in a dose-dependent manner.

24 From another perspective, it is well documented that several polysaccharides (e.g.,
25 dextran, inulin, chondroitin sulfate, cyclodextrin, and hyaluronan) are stable within the
26 stomach and intestinal environment but are degraded in the colon by the colonic bacterial
27 flora.^{242, 243} These characteristics have driven scientists to investigate the possibility of using
28 them as drug carriers to deliver API in the inflamed region of the colon selectively.
29 Accordingly, many studies have flourished while formulating different polysaccharide-
30 micro/nanocarriers.²⁴⁴⁻²⁴⁸

31 The main polysaccharides used for designing nanoparticles for IBD treatment are
32 chitosan, hyaluronic acid, pectins, alginates, and inulin (Table 5).²⁴⁹ Other polysaccharides for
33 IBD have also been reported, such as xanthan gum and guar gum.^{250, 251} Hereafter,

1 polysaccharide nanoparticles for IBD are summarized, and the structure-activity relationship
2 is discussed.

3 **5.1. Chitosan**

4 Chitosan is obtained by the partial deacetylation of chitin which is mainly derived from
5 the shells of crustaceans such as lobster, shrimp, or crab. It can also be found in fungi and
6 insects. Chitin is the second largest and most abundant polysaccharide in nature after
7 cellulose. It is responsible for maintaining the fungal shape, strength, and cell's structural
8 integrity. Chitosan is a linear heteropolymer composed of β -(1-4) linked *N*-acetyl-D-
9 glucosamine and D-glucosamine. The molecular weight and the degree of acetylation (DA)
10 are essential characteristics of chitosan. The DA of chitosan (> 50%) represents the fraction
11 of *N*-acetyl-D- glucosamine relative to the total number of units. Contrary to chitin (DA <
12 50%), which is insoluble in aqueous and many organic solvents, chitosan is hydrophilic and
13 soluble in acidic solutions, while the amine groups present on the macromolecule are
14 protonated. Chitosan with low DA and low molecular weight has increased solubility in
15 water.

16 However, the physicochemical properties of chitosan derived from the deacetylation of
17 chitin from crustaceans are heterogeneous, depending on the raw material source.²⁵² More
18 recently, chitosan was produced from chitin contained in selected fungi' cell walls and septa.
19 *Ascomycetes*, *Zygomycetes*, *Basidiomycetes*, and *Deuteromycetes*. Chitosan from fungi is
20 characterized by low molecular weight (e.g., 45 kDa)²⁵³ can be directly extracted from the raw
21 material without needing thermal or chemical depolymerization.

22 Due to its intrinsic anti-inflammatory, healing, and immunomodulatory activity, chitosan
23 is the most cited polysaccharide in developing nanoparticulate DDS for IBD.²⁴⁹ It is the only
24 cationic polysaccharide used to develop positively charged nanoparticulate DDS. The net
25 positive charge of chitosan nanoparticles confers mucoadhesive properties through the
26 electrostatic interactions with negatively charged mucins.²⁵⁴ Chitosan interacts with
27 negatively charged proteoglycan on the cell surface. It increases cell permeability by
28 reversibly acting on paracellular and intracellular pathways of epithelial cells.^{255, 256}
29 Nanoparticle intrinsic activity was further improved by functionalizing chitosan to natural
30 molecules such as gingerol, used for its anti-inflammatory and antioxidant properties.²⁵⁷ The
31 nanoparticles composed of chitosan covalently linked to gingerol (~50 – 78 nm, -21 mV)
32 were successfully used for the encapsulation and the controlled release of 5-ASA. Although in

1 vivo evaluations were not yet performed using those nanoparticles, size and charge properties,
2 as well as drug release control, could be favorable in vivo for the treatment of IBD.²⁵⁷

3 Due to its solubility in acidic media, chitosan-based nanoparticles could exhibit a
4 premature release of the drug before reaching the colon. Different strategies were thus
5 envisioned to improve colon-specific drug delivery in IBD. Chemical crosslinking (e.g.,
6 glutaraldehyde) or ionic crosslinking (e.g., tripolyphosphate (TPP)) are the most commonly
7 used methods to make chitosan nanoparticles insoluble in acidic media.^{258, 259}

8 Hybrid multifunctional nanoparticles composed of a blend of chitosan and anionic
9 polysaccharides such as alginate^{218, 260-264}, hyaluronan,^{265, 266} pectins,²⁵⁸ fucoidans,^{267, 268} or
10 carboxymethyl starch²⁶⁹ showed promise to deliver the drug to the colon and to control its
11 release. Ahmed et al.,²⁵⁸ demonstrated that the administration of taurine encapsulated in
12 chitosan/pectin nanoparticles exerted beneficial effects in induced colitis in rats thanks to their
13 anti-inflammatory and antioxidant activities. The nanoparticles (~74 nm, 48 mV) were
14 prepared by ionic gelation and crosslinked with sodium TPP. However, at pH 6.4, a burst
15 release was observed after the first 30 min followed by a slower release over 4 h.²⁵⁸ Coating
16 of nanoparticles composed of chitosan/TPP with Eudragit (FS30D),¹⁴⁰ confers a sustained
17 release and a pH stimulated delivery of the encapsulated active drug in addition to a good
18 accumulation in the colonic region of UC-induced rats.¹⁴⁰ Nanoparticles composed of
19 multilayers of chitosan oligosaccharide, alginate, and Eudragit S100 demonstrated pH-
20 dependent release of dexamethasone with a limited initial burst release in acidic pH. The
21 nanoparticles exhibited significant therapeutic activity in a colitis-induced mouse model.²⁷⁰

22 The chemical grafting of recognition ligands on chitosan nanoparticles further improved
23 the targeting of the immune cells in the inflamed colon. Galactosylated chitosan nanoparticles
24 were mainly used for the encapsulation of TNF- α siRNA or Map4k4 siRNA to increase the
25 cellular uptake by activated macrophages through the galactose-mediated receptor.^{201, 271, 272}
26 Major results revealed that galactosylated nanoparticles that have a high binding affinity for
27 siRNA showed enhanced in vivo anti-inflammatory efficacy in a mouse model of UC.^{201, 271}
28 To learn more about chitosan nanoparticle-mediated gene therapy strategies for mitigating
29 IBD, see the review of Verma et al.¹⁶

30 **5.2. Hyaluronan**

31 Hyaluronan (HA), a nonsulfated glycosaminoglycan, is an essential component of the
32 vertebrate extracellular matrix, where it is naturally present at relatively high concentrations

1 in the extracellular matrix, especially of soft connective tissues (e.g., skin, vitreous humor,
2 synovial fluid, umbilical cord). Viruses and bacteria also produced HA.²⁷³ HA is a linear
3 heteropolysaccharide composed of repeating β -1,4 bond disaccharide units of D-glucuronic
4 acid and *N*-acetyl-D-glucosamine linked together through β -1,3 glycosidic bonds. HA is a
5 polyanionic polysaccharide. The HA molecule is negatively charged when the pH is higher
6 than the pKa of HA carboxyl groups (3 – 4).

7 HA is implicated in various physiological functions in the intercellular matrix (e.g.,
8 homeostasis of water and plasma protein). HA is either anchored firmly in the plasma
9 membrane or bond via HA-specific binding proteins. Among those receptors, called
10 hyalderins, CD44 is a transmembrane receptor known to play a critical role in
11 inflammation.²⁷⁴

12 HA has a variable molecular weight depending on the source of production. It is highly
13 polydisperse and varies from 2 kDa to > 6000 kDa. The rheological properties of HA are
14 dependent on the HA molecular weight. In water, HA has a semi-flexible structure adopting a
15 worm-like random coil. The radius of gyration, R_g depends on the molecular weight of HA as
16 follows $R_g \approx 1.3 \text{ nm} \times M_w^{0.6}$.²⁷⁵ HA has intrinsic healing and anti-inflammatory properties
17 that rely on the HA molecular weight. While high molecular weight HA (HMw-HA) has anti-
18 inflammatory activity, low molecular weight HA (LMw-HA) activates an innate immune
19 response. Also, the affinity toward hyalderins such as CD44 and RHAMM (receptor for HA-
20 mediated motility, designated as CD168) was controlled by the Mw of HA in solution or
21 grafted on the particle surface.^{276, 277} Surface plasmon resonance experiments revealed that
22 low molecular weight HA nanoparticles (6.4 kDa) exhibited low binding to CD44 receptor,
23 while high molecular weight HA nanoparticles (1500 kDa) had a high binding affinity.²⁷⁶

24 Intensive research investigations were focused on designing HA functionalized
25 nanoparticles as a vehicle for active drugs to alleviate IBD (e.g., bilirubin,²⁷⁸ Lysine-proline-
26 valine (KPV tripeptide),²⁷⁹ budesonide,^{280, 281} methotrexate²⁶⁵). The drug is either
27 encapsulated or covalently linked to HA nanoparticles. It was demonstrated that CD44
28 overexpression on the surface of inflamed intestinal epithelial cells²⁸¹ and proinflammatory
29 macrophages²⁷⁸ contributed to the accumulation of HA nanoparticles in the inflamed colonic
30 tissue in DSS-induced colitis mice.²⁷⁸ In addition to the eEPR effect, the glycoside cluster
31 provided by dense HA on nanoparticle surface contributes to higher receptor-mediated
32 interaction with cells.²⁸² This property was exploited for the intracellular delivery of nucleic

1 acids. CD44 targeting HA nanoparticles can selectively deliver siRNA (silencing TNF- α or
2 CD98) to peritoneal macrophages leading to downregulation of proinflammatory cytokines.²¹⁵

3 HA nanoparticles act on the tight junction proteins ZO-1 and occludin-1, which are tight
4 junction-associated proteins that play pivotal roles in gut homeostasis.²⁷⁹ DSS-colitis mice
5 orally administered with HA nanoparticles covalently linked to bilirubin normalized the
6 expression patterns and messenger RNA levels of ZO-1 and occludin-1, while other control
7 groups, including free HA, and the association of HA and bilirubin had minimal impact.²⁷⁸
8 Furthermore, the same study reported the role of HA nanoparticles conjugated to bilirubin in
9 restoring intestinal barrier functions and reducing the level of apoptosis in the colonic
10 epithelium. This formulation significantly reduced the local levels of proinflammatory
11 cytokines, such as IL-1 β , TNF- α , and IL-6, while increasing the levels of anti-inflammatory
12 IL-10 and TGF- β cytokines.²⁷⁸

13 HA nanoparticles also modulate the gut microbiota, increasing the overall richness and
14 diversity. HA nanoparticles covalently linked to bilirubin markedly increased the abundance
15 of *Akkermansia muciniphila* (known to be associated with protective intestinal barrier
16 functions²⁸³) and *Clostridium XIVa*, which are microorganisms with crucial roles in gut
17 homeostasis.

18 **5.3. Pectins**

19 Pectins are a class of complex polysaccharides extracted from most terrestrial plants' cell
20 walls, where they exert a function of controlling the movement of water and cementing for the
21 cellulosic network.²⁸⁴ They are known for their ability to form gels in the presence of calcium
22 ions, solutes, or at low pH. Pectins are mainly composed of a linear chain of a homopolymer
23 of α -(1,4)-D-galacturonic acid. The carboxylic acid groups on the backbone are methyl
24 esterified at various degrees, thus controlling the gelling properties of pectins.²⁸⁵ Pectins are a
25 class of block copolymers because the linear chain termed 'smooth region' is occasionally
26 interrupted by side chains composed of rhamnogalactomannan. This heteropolymer alternates
27 (1,2)- α -L-rhamnosyl-(1-4)- α -D-galacturonic acid disaccharide units. These neutral chains tend
28 to be gathered into particular areas of the pectin molecule are called 'hairy regions'. There
29 may be different branched blocks in pectins from one cell wall or even within a single pectin
30 molecule. Notably, pectins with high hairy regions are less stable at low pH than pectins with
31 smooth regions.

1 Pectins possess intrinsic anti-inflammatory activity depending on the degree of
2 methylesterification.²⁸⁶ Low methylesterification pectins would be a more efficient anti-
3 inflammatory agent than high methylesterification pectins upon oral administration.²⁸⁶ Low
4 methylesterification pectin inhibited local and systemic inflammation, whereas high
5 methylesterification pectins prevented intestinal inflammation.²⁸⁶ When administered orally,
6 pectins are subject to biodegradation by colonic bacteria. This property makes it ideal for
7 targeted drug delivery to the colon.²⁸⁷ In particular, pectins with low hairy regions and low
8 methylesterification are more subject to degradation in the colon have been widely used to
9 design colon-specific nanoparticulate DDS for the treatment of IBD. In addition to these
10 advantages, pectins are known for their mucoadhesive properties,²⁸⁸ making them ideal
11 candidates to design colon-specific nanoparticulate DDS to alleviate IBD.

12 In a recent work conducted by Yener et al.,²⁸⁹ pectin-based nanoparticles containing
13 melatonin ameliorated the TNBS-induced IBD in rats by decreasing colonic fibrosis,
14 oxidative stress, and inflammatory parameters of the colon. Melatonin was used as an
15 antioxidant, anti-inflammatory, and radical scavenger drug. The nanoparticles formed by the
16 polyelectrolyte complexation method using CaCl₂ had a mean hydrodynamic diameter of 75
17 nm and zeta potential of 24 mV. The same study revealed that pectin nanoparticles were 40
18 times more adherent to the inflamed rat's colon than the healthy colon mucosa, where the
19 drug is released.²⁸⁹

20 The efficacy of pectin nanoparticles was further improved by adding HA in the
21 formulation. The resulting hybrid particles (284 nm in size) were used to encapsulate Rhein as
22 a natural anti-inflammatory ingredient.²⁹⁰ According to physicochemical characterizations, the
23 HA is located on the nanoparticles' outer shell that actively targets macrophages through
24 CD44-mediated endocytosis. Biological evaluations revealed that the nanoparticles
25 accumulated in the inflamed area of the colon of a DSS-induced UC mouse model. The
26 nanoparticles protected the intestinal barrier of UC mice by acting on tight junction proteins.

27 **5.4. Inulin**

28 Inulin is a natural fructose polymer mainly derived from plants. Examples of plants that
29 contain large quantities of inulin are Jerusalem artichoke, chicory root, asparagus root. It can
30 also be found in consumed vegetables such as onion, banana, leek, garlic, rye, barley, and
31 wheat. Inulin consists of linear chains of D-fructose units ($n = 2 - 60$) linked by β -(2,1)-
32 glycosidic bonds and a terminal glucose residue (For review article on inulin structure and

1 physicochemical properties see reference²⁹¹). Inulin containing a maximum 10 fructose units
 2 is also referred to as oligofructose. Inulin had several intrinsic biological properties, such as
 3 anti-inflammatory activity. It can promote the proliferation of beneficial intestinal bacteria
 4 (e.g., *Bifidobacterium and Lactobacillus*) and maintain intestinal micro-ecological balance
 5 and host health.^{292, 293} In a pilot study, inulin-type fructans orally administered to patients with
 6 active UC induced functional but not compositional shifts of the gut microbiota. High dose
 7 administration of inulin-type fructans increased *Bifidobacteriaceae* and *Lachnospiraceae*
 8 abundance and increased SCFA levels.²⁹⁴

9 Inulin is a potential candidate for drug delivery to the colon because the presence of β -
 10 (2,1)-glycosidic bonds prevents its degradation in the upper part of the GI tract, while it gets
 11 degraded in the colon by colonic enzymes (e.g., ptyalin and amylase, which are adapted to
 12 digesting starch)²⁹⁵ and bacteria (*Bifidobacteria*).²⁹⁶

13 Inulin has moderate solubility in water which decreases with temperature.²⁹⁷ Inulin was
 14 hydrophobically modified by grafting carboxymethyl groups²⁹⁸ hydrophobic
 15 dehydropeptide,²⁹⁹ on hydroxyls present on inulin backbone to deliver hydrophobic drugs like
 16 glucocorticoids (e.g., budesonide) or nitroimidazole antibiotics (e.g., ornidazole)³⁰⁰ for IBD
 17 treatment. In vivo results revealed that budesonide-loaded carboxymethyl inulin nanoparticles
 18 accumulate in inflamed colon tissue. Those systems showed enzyme- redox- and/or pH-
 19 sensitive properties allowing a specific drug release into the inflamed colon.^{298, 299}

20 It has been reported that apremilast-loaded inulin nanoparticles can be coupled with
 21 mannose as a macrophage-targeting ligand.¹⁹⁷ Apremilast was used as a model anti-
 22 inflammatory drug for the treatment of IBD.³⁰¹ Mannosylated nanoparticles showed greater
 23 uptake in inflamed macrophages than the untreated macrophage and mannose receptor-
 24 negative cell lines. In vivo biodistribution exhibited 60% of mannosylated nanoparticles
 25 accumulated in the inflamed colon of UC mice. In addition to the colon-targeting property and
 26 anti-inflammatory activity of inulin, macrophage-targeted drug delivery could have promise
 27 for the treatment of IBD.¹⁹⁷

28 *Table 5: A list of significant polysaccharides used to monitor IBD.*

Polysaccharides	Origin	Role in IBD
Chitosan	Exoskeleton of crustaceans ^{302, 303} The cell wall of	Different aspects of chitosan were explored for the monitoring of IBD: 1- As a carrier of conventional drugs due to its upper GI tract resistance properties ³⁰⁶

	fungi ^{304, 305}	<p>2- As a drug, where it can regulate different mechanisms:</p> <ul style="list-style-type: none"> - Increasing the number of probiotics that regenerate the microbiota balance^{307, 308} - Protecting the intestinal integrity by modulating the inflammation and oxidative stress³⁰⁹ - Down-regulating the levels of macrophage-inflammatory protein (MIP)-2 in the serum. - Up-regulating the frequency of FoxP3+ T cells, which prevent the auto-immune attacks.
Hyaluronan (HA)	<p>Various origins</p> <p>Traditionally: extraction from rooster combs</p> <p>Nowadays: streptococcal fermentation</p>	<p>HA is one of the glycosaminoglycans that is a significant component of the extracellular matrix of the intestinal mucosa,³¹⁰ where it has a crucial function of maintaining the translocation of luminal content into the general circulation. Although its mechanism of action is not yet fully understood, HA plays a significant role in immuno-inflammatory process due to its high affinity to the CD44 receptors that are overexpressed by the endothelial cells of the inflamed colonic segments. These receptors are crucial for infiltrating immune cells (e.g., monocytes, macrophages, neutrophils) in the inflamed tissues to aggravate the inflammation. Accordingly, the selective binding of HA to endothelial CD44 receptors during an inflammatory process will hamper the recruitment of immune cells.³¹¹⁻³¹³ In another study, Chen et al., explored the anti-inflammatory potential of high molecular HA (HM_w-HA) in reducing inflammation through a Toll-like receptor 4 (TLR4) modulation.³¹⁴ Indeed, TLR4 are widely distributed in the colon,³¹⁵ and they play a pivotal role in the colonic defense system. Their results confirmed their theory as HM_w-HA reduced the TNBS induced colitis in wild-type mice but not in TLR4^{-/-} mice.</p>
Pectins	<p>Various origins (e.g., orange, citrus, apple)</p>	<p>According to the origin of the extracted pectins, their role in IBD was reported to be different. Indeed, in one study, authors reported that pre-feeding mice with a side chain content of orange-extracted pectins had a protective effect of a TNBS-induced inflammation, whereas the pre-feeding with citrus-extracted pectins did not improve the clinical symptoms.³¹⁶ Pectins have a powerful impact on the downregulation of nuclear factor (NF)-κB p65 that is directly implicated in the immune response.³¹⁷</p>
Fructan	<p>Angiosperms³¹⁸ (e.g., wheat, garlic, leeks, artichokes, agave)</p>	<p>Fructan, is reported to protect the intestinal mucosa due to its ability to enrich the mucosal intestine with probiotics. Indeed, Hino et al., suggested that a TNBS-induced colitis healing in rats could be a consequence of an increase in the number of lactic acid-producing bacteria following the ingestion of short-chain inulin-like fructan.³¹⁹ More recently, similar data were obtained at the clinical level, where inulin-type fructan showed</p>

		clinical benefits in UC, as 77% of the patients showed a clinical response. ²⁹⁴
Guar gum	Guar beans ²⁵¹	Guar gum participates in preventing and reducing UC by promoting the production of SCFAs. In a TNBS colitis model, it was demonstrated that partially hydrolyzed guar gum participated in the reduction of proinflammatory-cytokines, immune cells infiltration in the intestinal mucosa, and myeloperoxidase activity. Furthermore, another study reflected the wound healing of colonic epithelial cells and the repair of intestinal mucosa following the administration of partially hydrolyzed guar gum. The authors suggested that these observations would be a consequence of an up-regulation of extracellular signal-regulated kinases 1 and 2.

1

7. Challenges and future directions

Nowadays, monitoring IBD relies on conventional drugs to prevent inflammation, alleviate the active phase, or modulate the immune response while using immunosuppressants. Nonetheless, using these drugs is often limited by either poor clinical efficiency or numerous side effects. Although the new biological therapies relying on the administration of monoclonal antibodies offer better management of the disease, their cost remains an obstacle for their ubiquitous use. Technological advances in designing nanoparticle-based therapies permitted the oral delivery of various drugs that would have found hurdles to reach the inflamed segment of the intestine after an oral administration. Nanoparticles exhibited many properties that make them suitable for the oral delivery of active drugs to alleviate IBD (e.g., ability to diffuse through mucus layer, mucoadhesive properties, passive targeting of the inflamed colon, and cell penetrability).

Polysaccharides, initially used for colon-specific drug delivery due to their biodegradability by bacterial enzymes, appear to be promising materials for designing nanoparticulate DDS for IBD treatment. The biocompatibility and the intrinsic activity of the polysaccharides, together with the advantages of the nanoparticulate systems, offer exceptional results in alleviating IBD. The most cited polysaccharides for IBD are chitosan and alginates. More recently, other polysaccharides were also investigated to alleviate IBD, such as hyaluronan, pectins, and inulin. Using those polysaccharides in combination (e.g., pectin and HA) is particularly interesting because it leads to multifunctional nanosystems with controlled properties such as drug release and receptor-mediated targeting. Furthermore, the synergism between the drug and the polysaccharide that have an intrinsic biological activity should be further explored in the future. Cui et al.,¹²⁶ recently reviewed this synergism between the drugs and polysaccharide-based carriers.

Overall, the results depicted from the numerous studies which investigated the use of polysaccharides as delivery systems are encouraging. Nonetheless, several hurdles need to be tackled. Most of the polysaccharides are both polymolecular and polydisperse. Their composition varies with the source and conditions of extraction, location, and other environmental factors. Thanks to fungi and bacteria fermentation processes, the sources of polysaccharides production have been diversified over recent years. Additional efforts should be made in this area to produce polysaccharides in reproducible and controlled manners.

1 A significant area of concern is to develop standardized methods for the in vitro
2 evaluation of drug behaviors in simulated media of disease patients. Indeed, the simulated
3 media usually used for in vitro investigations rather mimic healthy patients than patients
4 suffering from IBD. IBD patients suffer from several physiological changes compared to
5 healthy subjects. Those changes include pH in the GI tract, gut microbiota alteration, and
6 variations in the colonic enzymes. This standardization should allow a better comparison
7 between different research groups and a better correlation between in vitro and in vivo
8 situations in humans. Furthermore, the models used to evaluate the mucoadhesion and the
9 mucopenetration of nano-drug delivery were usually performed on mucus from healthy
10 animals. In contrast, several changes in mucus, structure, composition, and viscoelastic
11 properties were reported with the mucus of patients suffering from IBD. Alternative methods
12 to evaluate the behaviors of nanoparticles toward mucus from animal models with induced
13 inflammation should be developed.

14 Due to the complexity of IBD pathophysiology, the DSS and TNBS colitis models are far
15 from mimicking the disease in humans. Passive targeting using eEPR was found to be weaker
16 in humans compared to animal models. It has been increasingly clear that the intestinal
17 microbiome is different in mice and humans. The pH in the mouse stomach varies from pH
18 2.7 to 4.1, while in humans, it can go down to pH 1.³²⁰ However, the intestinal pH in mice
19 was lower than in humans. Transit time, the mucus growth rate, and thickness are also
20 different in mice and humans.³²¹ Mice mucus shows different levels of glycan profiles than
21 humans. For a review article see reference ³²². Furthermore, only a few percent of the
22 bacterial gene are shared between mice and humans.³²² Some receptors expressed by cells in
23 mice are different from those in humans. Accurate animal models that mimic human IBD
24 disease need to be developed in the future. Efforts were recently stressed on developing
25 humanized mouse models,³²³⁻³²⁵ or using large mammalian models such as pigs.³²⁶

26 Over the recent years, receptor-mediated targeting of immune cells showed promising
27 preclinical results compared to non-targeted nanoparticulate DDS. However, the analysis of
28 research articles in this field revealed that targeting strategies mainly focus on mannose
29 receptor, CD44, and galactose agglutinin, while there are various receptors or cell adhesion
30 molecules that are upregulated in the colon tissues of IBD.³²⁷⁻³³⁰ Those receptors and cell
31 adhesion molecules are still not applied to design targeted nanoparticles. Also, efforts should
32 be devoted to understanding additional molecules implicated in the pathological process of
33 IBD.

1 Another hurdle is related to the low encapsulation efficiency and the premature drug
2 release of the encapsulated drug. Low encapsulation efficiency and premature drug release
3 were tackled through the covalent linkage of the drug to the polysaccharide.³³¹⁻³³⁴ Compared
4 with physical encapsulation, the prodrug strategy was explored for developing smart-
5 responsive drug-polysaccharides conjugates.³³¹ In these systems, the drug was released under
6 stimulus (e.g., pH, ROS, enzymes) for specific drug delivery at the site of inflammation.^{298,}
7 ^{331, 335} However, the prodrug strategy could be limited by the availability of the appropriate
8 functional groups of the drugs to be conjugated to the polysaccharide.

9 Finally, from a technological point of view, there are several well-known hurdles related
10 to complex manufacturing processes of nanoparticles, the difficulty of scalability, and poor
11 reproducibility.³³⁶ These issues should be addressed by taking advantage of the considerable
12 progress in the nanomedicine field these two last years. The psychological barrier to the
13 clinical development of nanomedicines has been broken with the planetary commercialization
14 of COVID vaccines composed of lipidic nanoparticles. The latest advances in lipid
15 nanoparticles will undoubtedly orient future research works in the nanotechnology field.
16 Today, we can rapidly overcome the technological and regulatory issues related to
17 nanoparticle preparation and treatment processes after their preparations.

18 **8. Conclusion**

19 IBD remains one of the most challenging and less understandable diseases of the 21st
20 century. Over the last couple of years, numerous medical and technological advances have
21 allowed a better comprehension of the pathophysiology of IBD. Understanding
22 immunological and proinflammatory signalling cascades has opened new avenues in
23 monitoring the disease. Developing biological treatments relying on administrating
24 monoclonal antibodies or siRNA targeting proinflammatory cytokines has gained much
25 interest. Nonetheless, biologics are associated with some shortcomings, such as their
26 expensive treatment or their reported loss of efficacy, partly because of the formation of anti-
27 drug antibodies (ADAbs). Within these circumstances, new strategies are under investigation
28 to develop new management protocols for IBD symptoms. Such strategies include
29 nanotechnology or polysaccharide-based therapies. The principal target behind using
30 engineered nanoparticles is to deliver a lower drug dosage to reach a higher therapeutic
31 response. This strategy is possible because nanoparticles can target the API directly to the
32 inflamed region of the colon, allowing to avoid the numerous systemic side effects of the

1 current medicines. Another characteristic of nanoparticles is their ability to protect fragile
2 drugs from degradation in the harsh GI tract environments, allowing them to reach their site
3 of action. More recently, polysaccharide-based therapies are gaining more interest as potential
4 molecules for the monitoring of IBD. In fact, in addition to their high safety and low systemic
5 side effects, polysaccharides were reported in numerous studies to play a prime role in
6 regulating the inflammatory cytokines, the intestinal microbiome, and the immune system.
7 These characteristics put polysaccharides on the spot of being an exciting alternative for
8 currently available drugs.

9 **9. Funding**

10 Author K.B. received funding from the Institut Universitaire de France and ANR-17-
11 CE09-0038-1.

12 **10. References**

- 13 1. Farmer, R. G., Long-term prognosis for patients with ulcerative proctosigmoiditis (ulcerative
14 colitis confirmed to the rectum and sigmoid colon). *Journal of clinical gastroenterology* **1979**, *1* (1),
15 47-50.
- 16 2. Farmer, R. G.; Brown, C. H., Ulcerative colitis confined to the rectum and sigmoid flexure.
17 *Diseases of the Colon & Rectum* **1967**, *10* (3), 177-182.
- 18 3. Dsouza, R.; Varghese, G.; Korula, D. R.; Dutta, A. K., Crohn's disease associated
19 adenocarcinoma of ileocaecal region: a miscalculated approach. *BMJ Case Reports CP* **2020**, *13* (4),
20 e234512.
- 21 4. Sturm, E.; Cobben, L.; Meijssen, M.; van der Werf, S.; Puylaert, J., Detection of ileocecal
22 Crohn's disease using ultrasound as the primary imaging modality. *European radiology* **2004**, *14* (5),
23 778-782.
- 24 5. Voderholzer, W.; Beinhoelzl, J.; Rogalla, P.; Murrer, S.; Schachschal, G.; Lochs, H.; Ortner,
25 M., Small bowel involvement in Crohn's disease: a prospective comparison of wireless capsule
26 endoscopy and computed tomography enteroclysis. *Gut* **2005**, *54* (3), 369-373.
- 27 6. Allan, A.; Andrews, H.; Hilton, C.; Keighley, M.; Allan, R.; Alexander-Williams, J., Segmental
28 colonic resection is an appropriate operation for short skip lesions due to Crohn's disease in the
29 colon. *World journal of surgery* **1989**, *13* (5), 611-614.
- 30 7. Qin, X., Why is damage limited to the mucosa in ulcerative colitis but transmural in Crohn's
31 disease? *World journal of gastrointestinal pathophysiology* **2013**, *4* (3), 63-64.
- 32 8. Sanders, D. S. A., 2 The differential diagnosis of Crohn's disease and ulcerative colitis.
33 *Baillière's clinical gastroenterology* **1998**, *12* (1), 19-33.
- 34 9. Walker, A. W.; Sanderson, J. D.; Churcher, C.; Parkes, G. C.; Hudspith, B. N.; Rayment, N.;
35 Brostoff, J.; Parkhill, J.; Dougan, G.; Petrovska, L., High-throughput clone library analysis of the
36 mucosa-associated microbiota reveals dysbiosis and differences between inflamed and non-inflamed
37 regions of the intestine in inflammatory bowel disease. *BMC microbiology* **2011**, *11* (1), 1-12.
- 38 10. Ryan, F. J.; Ahern, A.; Fitzgerald, R.; Laserna-Mendieta, E.; Power, E.; Clooney, A.;
39 O'Donoghue, K.; McMurdie, P.; Iwai, S.; Crits-Christoph, A., Colonic microbiota is associated with
40 inflammation and host epigenomic alterations in inflammatory bowel disease. *Nature*
41 *communications* **2020**, *11* (1), 1-12.
- 42 11. Sgambato, D.; Miranda, A.; Ranaldo, R.; Federico, A.; Romano, M., The role of stress in
43 inflammatory bowel diseases. *Current pharmaceutical design* **2017**, *23* (27), 3997-4002.

- 1 12. Jairath, V.; Feagan, B. G., Global burden of inflammatory bowel disease. *The Lancet*
2 *Gastroenterology & Hepatology* **2020**, *5* (1), 2-3.
- 3 13. Tremaine, W. J., Failure to yield: drug resistance in inflammatory bowel disease.
4 *Gastroenterology* **2002**, *122* (4), 1165-1167.
- 5 14. Carty, E.; Rampton, D., Evaluation of new therapies for inflammatory bowel disease. *British*
6 *journal of clinical pharmacology* **2003**, *56* (4), 351-361.
- 7 15. Li, X.; Lu, C.; Yang, Y.; Yu, C.; Rao, Y., Site-specific targeted drug delivery systems for the
8 treatment of inflammatory bowel disease. *Biomedicine & Pharmacotherapy* **2020**, *129*, 110486.
- 9 16. Verma, P.; Srivastava, A.; Srikanth, C. V.; Bajaj, A., Nanoparticle-mediated gene therapy
10 strategies for mitigating inflammatory bowel disease. *Biomaterials science* **2021**, *9* (5), 1481-1502.
- 11 17. Dar, M. J.; Ali, H.; Khan, A.; Khan, G. M., Polymer-based drug delivery: the quest for local
12 targeting of inflamed intestinal mucosa. *Journal of drug targeting* **2017**, *25* (7), 582-596.
- 13 18. Talaei, F.; Atyabi, F.; Azhdarzadeh, M.; Dinarvand, R.; Saadatzadeh, A., Overcoming
14 therapeutic obstacles in inflammatory bowel diseases: a comprehensive review on novel drug
15 delivery strategies. *European Journal of Pharmaceutical Sciences* **2013**, *49* (4), 712-722.
- 16 19. Zeeshan, M.; Ali, H.; Khan, S.; Khan, S. A.; Weigmann, B., Advances in orally-delivered pH-
17 sensitive nanocarrier systems; an optimistic approach for the treatment of inflammatory bowel
18 disease. *International journal of pharmaceutics* **2019**, *558*, 201-214.
- 19 20. Hua, S.; Marks, E.; Schneider, J. J.; Keely, S., Advances in oral nano-delivery systems for
20 colon targeted drug delivery in inflammatory bowel disease: selective targeting to diseased versus
21 healthy tissue. *Nanomedicine: nanotechnology, biology and medicine* **2015**, *11* (5), 1117-1132.
- 22 21. Lautenschläger, C.; Schmidt, C.; Fischer, D.; Stallmach, A., Drug delivery strategies in the
23 therapy of inflammatory bowel disease. *Advanced drug delivery reviews* **2014**, *71*, 58-76.
- 24 22. Yang, M.; Zhang, Y.; Ma, Y.; Yan, X.; Gong, L.; Zhang, M.; Zhang, B., Nanoparticle-based
25 therapeutics of inflammatory bowel diseases: a narrative review of the current state and prospects.
26 *Journal of Bio-X Research* **2020**, *3* (4), 157-173.
- 27 23. Yang, W.; Zhao, P.; Li, X.; Guo, L.; Gao, W., The potential roles of natural plant
28 polysaccharides in inflammatory bowel disease: A review. *Carbohydrate Polymers* **2021**, 118821.
- 29 24. Jung, A., Colon and rectum normal anatomy and congenital variants. *Abdominal Imaging*
30 **2013**, 785-796.
- 31 25. Kooij, I.; Sahami, S.; Meijer, S.; Buskens, C.; Te Velde, A., The immunology of the vermiform
32 appendix: a review of the literature. *Clinical & Experimental Immunology* **2016**, *186* (1), 1-9.
- 33 26. Rao, J.; Wang, J., Intestinal architecture and development. *Regulation of Gastrointestinal*
34 *Mucosal Growth* **2010**, 3-8.
- 35 27. Rodríguez-Piñeiro, A. M.; Johansson, M. E., The colonic mucus protection depends on the
36 microbiota. *Gut microbes* **2015**, *6* (5), 326-330.
- 37 28. Hosoyamada, Y.; Sakai, T., Structural and mechanical architecture of the intestinal villi and
38 crypts in the rat intestine: integrative reevaluation from ultrastructural analysis. *Anatomy and*
39 *embryology* **2005**, *210* (1), 1-12.
- 40 29. Potten, C.; Gandara, R.; Mahida, Y.; Loeffler, M.; Wright, N., The stem cells of small
41 intestinal crypts: where are they? *Cell proliferation* **2009**, *42* (6), 731-750.
- 42 30. Hancock, R.; Falla, T., Antimicrobial peptides: broad-spectrum antibiotics from nature. *Clinical*
43 *Microbiology and Infection* **1996**, *1* (4), 226-229.
- 44 31. Liang, W.; Diana, J., The dual role of antimicrobial peptides in autoimmunity. *Frontiers in*
45 *immunology* **2020**, *11*, 2077.
- 46 32. Muniz, L. R.; Knosp, C.; Yeretsian, G., Intestinal antimicrobial peptides during homeostasis,
47 infection, and disease. *Frontiers in immunology* **2012**, *3*, 310.
- 48 33. Foell, D.; Wittkowski, H.; Roth, J., Monitoring disease activity by stool analyses: from occult
49 blood to molecular markers of intestinal inflammation and damage. *Gut* **2009**, *58* (6), 859-68.
- 50 34. Makkar, R.; Bo, S., Colonoscopic perforation in inflammatory bowel disease. *Gastroenterol*
51 *Hepatol (N Y)* **2013**, *9* (9), 573-83.

- 1 35. Kim, E. R.; Chang, D. K., Colorectal cancer in inflammatory bowel disease: the risk,
2 pathogenesis, prevention and diagnosis. *World J Gastroenterol* **2014**, *20* (29), 9872-81.
- 3 36. Elsherif, Y.; Alexakis, C.; Mendall, M., Determinants of Weight Loss prior to Diagnosis in
4 Inflammatory Bowel Disease: A Retrospective Observational Study. *Gastroenterol Res Pract* **2014**,
5 *2014*, 762191.
- 6 37. Garcia, J. C.; Persky, S. E.; Bonis, P. A.; Topazian, M., Abscesses in Crohn's disease: outcome
7 of medical versus surgical treatment. *J Clin Gastroenterol* **2001**, *32* (5), 409-12.
- 8 38. Gecse, K.; Khanna, R.; Stoker, J.; Jenkins, J. T.; Gabe, S.; Hahnloser, D.; D'Haens, G.,
9 Fistulizing Crohn's disease: Diagnosis and management. *United European Gastroenterol J* **2013**, *1* (3),
10 206-13.
- 11 39. Gourtsoyianni, S.; Papanikolaou, N.; Amanakis, E.; Bourikas, L.; Roussomoustakaki, M.;
12 Grammatikakis, J.; Gourtsoyiannis, N., Crohn's disease lymphadenopathy: MR imaging findings. *Eur J*
13 *Radiol* **2009**, *69* (3), 425-8.
- 14 40. Petagna, L.; Antonelli, A.; Ganini, C.; Bellato, V.; Campanelli, M.; Divizia, A.; Efrati, C.;
15 Franceschilli, M.; Guida, A.; Ingallinella, S., Pathophysiology of Crohn's disease inflammation and
16 recurrence. *Biology direct* **2020**, *15* (1), 1-10.
- 17 41. Desai, J.; Elnaggar, M.; Hanfy, A. A.; Doshi, R., Toxic Megacolon: Background,
18 Pathophysiology, Management Challenges and Solutions. *Clin Exp Gastroenterol* **2020**, *13*, 203-210.
- 19 42. Parray, F. Q.; Wani, M. L.; Malik, A. A.; Wani, S. N.; Bijli, A. H.; Irshad, I.; Nayeem Ul, H.,
20 Ulcerative colitis: a challenge to surgeons. *Int J Prev Med* **2012**, *3* (11), 749-63.
- 21 43. Mady, R.; Grover, W.; Butrus, S., Ocular complications of inflammatory bowel disease.
22 *ScientificWorldJournal* **2015**, *2015*, 438402.
- 23 44. Chandan, J. S.; Thomas, T., The impact of inflammatory bowel disease on oral health. *Br Dent*
24 *J* **2017**, *222* (7), 549-553.
- 25 45. Lin, J. N.; Lin, C. L.; Lin, M. C.; Lai, C. H.; Lin, H. H.; Kao, C. H., Pyogenic liver abscess in
26 patients with inflammatory bowel disease: a nationwide cohort study. *Liver Int* **2016**, *36* (1), 136-44.
- 27 46. Sinagra, E.; Aragona, E.; Romano, C.; Maisano, S.; Orlando, A.; Virdone, R.; Tesè, L.;
28 Modesto, I.; Criscuoli, V.; Cottone, M., The role of portal vein thrombosis in the clinical course of
29 inflammatory bowel diseases: report on three cases and review of the literature. *Gastroenterol Res*
30 *Pract* **2012**, *2012*, 916428.
- 31 47. Kontny, E.; Dmowska-Chalaba, J.; Kwiatkowska, B.; Maśliński, W., Cytokines and integrins
32 related to inflammation of joint and gut in patients with spondyloarthritis and inflammatory bowel
33 disease. *Reumatologia* **2017**, *55* (6), 276-283.
- 34 48. States, V.; O'Brien, S.; Rai, J. P.; Roberts, H. L.; Paas, M.; Feagins, K.; Pierce, E. J.;
35 Baumgartner, R. N.; Galandiuk, S., Pyoderma Gangrenosum in Inflammatory Bowel Disease: A
36 Systematic Review and Meta-Analysis. *Dig Dis Sci* **2020**, *65* (9), 2675-2685.
- 37 49. Iida, T.; Onodera, K.; Nakase, H., Role of autophagy in the pathogenesis of inflammatory
38 bowel disease. *World journal of gastroenterology* **2017**, *23* (11), 1944.
- 39 50. Rogler, G.; Andus, T., Cytokines in inflammatory bowel disease. *World journal of surgery*
40 **1998**, *22* (4), 382-389.
- 41 51. Guan, Q.; Zhang, J., Recent advances: the imbalance of cytokines in the pathogenesis of
42 inflammatory bowel disease. *Mediators of inflammation* **2017**, *2017*.
- 43 52. Hofer, U., Bacterial imbalance in Crohn's disease. *Nature Reviews Microbiology* **2014**, *12* (5),
44 313-313.
- 45 53. Lo, C.-H.; Lochhead, P.; Khalili, H.; Song, M.; Tabung, F. K.; Burke, K. E.; Richter, J. M.;
46 Giovannucci, E. L.; Chan, A. T.; Ananthakrishnan, A. N., Dietary inflammatory potential and risk of
47 Crohn's disease and ulcerative colitis. *Gastroenterology* **2020**, *159* (3), 873-883. e1.
- 48 54. Chicco, F.; Magrì, S.; Cingolani, A.; Paduano, D.; Pesenti, M.; Zara, F.; Tumbarello, F.; Urru,
49 E.; Melis, A.; Casula, L., Multidimensional impact of Mediterranean diet on IBD patients.
50 *Inflammatory bowel diseases* **2021**, *27* (1), 1-9.
- 51 55. Sun, Y.; Li, L.; Xie, R.; Wang, B.; Jiang, K.; Cao, H., Stress triggers flare of inflammatory
52 bowel disease in children and adults. *Frontiers in pediatrics* **2019**, *7*, 432.

- 1 56. Klein, A.; Eliakim, R., Non steroidal anti-inflammatory drugs and inflammatory bowel disease.
2 *Pharmaceuticals* **2010**, *3* (4), 1084-1092.
- 3 57. Long, M.; Kappelman, M.; Martin, C.; Chen, W.; Anton, K.; Sandler, R., Role of Non-
4 Steroidal Anti-inflammatory Drugs in Exacerbations of Inflammatory Bowel Disease: 1742. *Official*
5 *journal of the American College of Gastroenterology| ACG* **2013**, *108*, S526.
- 6 58. van der Sloot, K. W.; Tiems, J. L.; Visschedijk, M. C.; Festen, E. A.; van Dullemen, H. M.;
7 Weersma, R. K.; Kats-Ugurlu, G.; Dijkstra, G., Cigarette smoke increases risk for colorectal neoplasia
8 in inflammatory bowel disease. *Clinical Gastroenterology and Hepatology* **2021**.
- 9 59. Mahid, S. S.; Minor, K. S.; Soto, R. E.; Hornung, C. A.; Galandiuk, S. In *Smoking and*
10 *inflammatory bowel disease: a meta-analysis*, Mayo Clinic Proceedings, Elsevier: 2006; pp 1462-1471.
- 11 60. Boyko, E.; Perera, D.; Koepsell, T.; Keane, E.; Inui, T., Effects of cigarette smoking on the
12 clinical course of ulcerative colitis. *Scandinavian journal of gastroenterology* **1988**, *23* (9), 1147-1152.
- 13 61. Sokol, H., Antibiotics: a trigger for inflammatory bowel disease? *The Lancet Gastroenterology*
14 *& Hepatology* **2020**, *5* (11), 956-957.
- 15 62. Nitzan, O.; Elias, M.; Peretz, A.; Saliba, W., Role of antibiotics for treatment of inflammatory
16 bowel disease. *World journal of gastroenterology* **2016**, *22* (3), 1078.
- 17 63. Schreiber, S.; Nikolaus, S.; Hampe, J., Activation of nuclear factor κ B in inflammatory bowel
18 disease. *Gut* **1998**, *42* (4), 477-484.
- 19 64. Rogler, G.; Brand, K.; Vogl, D.; Page, S.; Hofmeister, R.; Andus, T.; Knuechel, R.; Baeuerle,
20 P. A.; Schölmerich, J.; Gross, V., Nuclear factor κ B is activated in macrophages and epithelial cells of
21 inflamed intestinal mucosa. *Gastroenterology* **1998**, *115* (2), 357-369.
- 22 65. Kai, Y.; Takahashi, I.; Ishikawa, H.; Hiroi, T.; Mizushima, T.; Matsuda, C.; Kishi, D.; Hamada,
23 H.; Tamagawa, H.; Ito, T., Colitis in mice lacking the common cytokine receptor γ chain is mediated
24 by IL-6-producing CD4⁺ T cells. *Gastroenterology* **2005**, *128* (4), 922-934.
- 25 66. Ogino, T.; Nishimura, J.; Barman, S.; Kayama, H.; Uematsu, S.; Okuzaki, D.; Osawa, H.;
26 Haraguchi, N.; Uemura, M.; Hata, T., Increased Th17-inducing activity of CD14⁺ CD163^{low} myeloid
27 cells in intestinal lamina propria of patients with Crohn's disease. *Gastroenterology* **2013**, *145* (6),
28 1380-1391. e1.
- 29 67. Atreya, R.; Neurath, M. F., Involvement of IL-6 in the pathogenesis of inflammatory bowel
30 disease and colon cancer. *Clinical reviews in allergy & immunology* **2005**, *28* (3), 187-195.
- 31 68. Mudter, J.; Neurath, M. F., IL-6 signaling in inflammatory bowel disease: pathophysiological
32 role and clinical relevance. *Inflammatory bowel diseases* **2007**, *13* (8), 1016-1023.
- 33 69. Jeffery, V.; Goldson, A. J.; Dainty, J. R.; Chieppa, M.; Sobolewski, A., IL-6 Signaling Regulates
34 Small Intestinal Crypt Homeostasis. *J Immunol* **2017**, *199* (1), 304-311.
- 35 70. Apte, R. N.; Voronov, E., Interleukin-1--a major pleiotropic cytokine in tumor-host
36 interactions. *Semin Cancer Biol* **2002**, *12* (4), 277-90.
- 37 71. Casini-Raggi, V.; Kam, L.; Chong, Y. J.; Fiocchi, C.; Pizarro, T. T.; Cominelli, F., Mucosal
38 imbalance of IL-1 and IL-1 receptor antagonist in inflammatory bowel disease. A novel mechanism of
39 chronic intestinal inflammation. *J Immunol* **1995**, *154* (5), 2434-40.
- 40 72. Ludwiczek, O.; Vannier, E.; Borggraefe, I.; Kaser, A.; Siegmund, B.; Dinarello, C.; Tilg, H.,
41 Imbalance between interleukin-1 agonists and antagonists: relationship to severity of inflammatory
42 bowel disease. *Clinical & Experimental Immunology* **2004**, *138* (2), 323-329.
- 43 73. Andus, T.; Daig, R.; Vogl, D.; Aschenbrenner, E.; Lock, G.; Hollerbach, S.; Köllinger, M.;
44 Schölmerich, J.; Gross, V., Imbalance of the interleukin 1 system in colonic mucosa--association with
45 intestinal inflammation and interleukin 1 receptor antagonist [corrected] genotype 2. *Gut* **1997**, *41*
46 (5), 651-7.
- 47 74. Aggeletopoulou, I.; Assimakopoulos, S. F.; Konstantakis, C.; Triantos, C., Interleukin
48 12/interleukin 23 pathway: biological basis and therapeutic effect in patients with Crohn's disease.
49 *World journal of gastroenterology* **2018**, *24* (36), 4093.
- 50 75. Hoeve, M. A.; de Boer, T.; Langenberg, D. M.; Sanal, O.; Verreck, F. A.; Ottenhoff, T. H., IL-
51 12 receptor deficiency revisited: IL-23-mediated signaling is also impaired in human genetic IL-12
52 receptor beta1 deficiency. *Eur J Immunol* **2003**, *33* (12), 3393-7.

- 1 76. Imamura, E.; Taguchi, K.; Sasaki-Iwaoka, H.; Kubo, S.; Furukawa, S.; Morokata, T., Anti-IL-23
2 receptor monoclonal antibody prevents CD4(+) T cell-mediated colitis in association with decreased
3 systemic Th1 and Th17 responses. *Eur J Pharmacol* **2018**, *824*, 163-169.
- 4 77. Iwakura, Y.; Ishigame, H., The IL-23/IL-17 axis in inflammation. *J Clin Invest* **2006**, *116* (5),
5 1218-22.
- 6 78. Vanden Eijnden, S.; Goriely, S.; De Wit, D.; Willems, F.; Goldman, M., IL-23 up-regulates IL-
7 10 and induces IL-17 synthesis by polyclonally activated naive T cells in human. *Eur J Immunol* **2005**,
8 *35* (2), 469-75.
- 9 79. van de Wetering, D.; de Paus, R. A.; van Dissel, J. T.; van de Vosse, E., IL-23 modulates
10 CD56+/CD3- NK cell and CD56+/CD3+ NK-like T cell function differentially from IL-12. *Int Immunol*
11 **2009**, *21* (2), 145-53.
- 12 80. Geremia, A.; Arancibia-Cárcamo, C. V.; Fleming, M. P.; Rust, N.; Singh, B.; Mortensen, N. J.;
13 Travis, S. P.; Powrie, F., IL-23-responsive innate lymphoid cells are increased in inflammatory bowel
14 disease. *J Exp Med* **2011**, *208* (6), 1127-33.
- 15 81. Parrello, T.; Monteleone, G.; Cucchiara, S.; Monteleone, I.; Sebkova, L.; Doldo, P.; Lizza,
16 F.; Pallone, F., Up-regulation of the IL-12 receptor beta-2 in Crohn's disease. **2000**.
- 17 82. Neurath, M. F.; Weigmann, B.; Finotto, S.; Glickman, J.; Nieuwenhuis, E.; Iijima, H.;
18 Mizoguchi, A.; Mizoguchi, E.; Mudter, J.; Galle, P. R.; Bhan, A.; Autschbach, F.; Sullivan, B. M.;
19 Szabo, S. J.; Glimcher, L. H.; Blumberg, R. S., The transcription factor T-bet regulates mucosal T cell
20 activation in experimental colitis and Crohn's disease. *J Exp Med* **2002**, *195* (9), 1129-43.
- 21 83. Fuss, I. J.; Neurath, M.; Boirivant, M.; Klein, J. S.; de la Motte, C.; Strong, S. A.; Fiocchi, C.;
22 Strober, W., Disparate CD4+ lamina propria (LP) lymphokine secretion profiles in inflammatory bowel
23 disease. Crohn's disease LP cells manifest increased secretion of IFN-gamma, whereas ulcerative
24 colitis LP cells manifest increased secretion of IL-5. *J Immunol* **1996**, *157* (3), 1261-70.
- 25 84. Breese, E.; Braegger, C. P.; Corrigan, C. J.; Walker-Smith, J. A.; MacDonald, T. T., Interleukin-
26 2- and interferon-gamma-secreting T cells in normal and diseased human intestinal mucosa.
27 *Immunology* **1993**, *78* (1), 127-31.
- 28 85. Cavalcanti, Y. V.; Brelaz, M. C.; Neves, J. K.; Ferraz, J. C.; Pereira, V. R., Role of TNF-Alpha,
29 IFN-Gamma, and IL-10 in the Development of Pulmonary Tuberculosis. *Pulm Med* **2012**, *2012*,
30 745483.
- 31 86. Langer, V.; Vivi, E.; Regensburger, D.; Winkler, T. H.; Waldner, M. J.; Rath, T.; Schmid, B.;
32 Skottke, L.; Lee, S.; Jeon, N. L.; Wohlfahrt, T.; Kramer, V.; Tripal, P.; Schumann, M.; Kersting, S.;
33 Handtrack, C.; Geppert, C. I.; Suchowski, K.; Adams, R. H.; Becker, C.; Ramming, A.; Naschberger,
34 E.; Britzen-Laurent, N.; Stürzl, M., IFN-γ drives inflammatory bowel disease pathogenesis through VE-
35 cadherin-directed vascular barrier disruption. *J Clin Invest* **2019**, *129* (11), 4691-4707.
- 36 87. Griga, T.; Hebler, U.; Voigt, E.; Tromm, A.; May, B., Interleukin-4 inhibits the increased
37 production of vascular endothelial growth factor by peripheral blood mononuclear cells in patients
38 with inflammatory bowel disease. *Hepato-gastroenterology* **2000**, *47* (36), 1604-1607.
- 39 88. Heller, F.; Florian, P.; Bojarski, C.; Richter, J.; Christ, M.; Hillenbrand, B.; Mankertz, J.;
40 Gitter, A. H.; Bürgel, N.; Fromm, M.; Zeitz, M.; Fuss, I.; Strober, W.; Schulzke, J. D., Interleukin-13 is
41 the key effector Th2 cytokine in ulcerative colitis that affects epithelial tight junctions, apoptosis, and
42 cell restitution. *Gastroenterology* **2005**, *129* (2), 550-64.
- 43 89. Fichtner-Feigl, S.; Fuss, I. J.; Young, C. A.; Watanabe, T.; Geissler, E. K.; Schlitt, H. J.; Kitani,
44 A.; Strober, W., Induction of IL-13 triggers TGF-beta1-dependent tissue fibrosis in chronic 2,4,6-
45 trinitrobenzene sulfonic acid colitis. *J Immunol* **2007**, *178* (9), 5859-70.
- 46 90. Kennedy, J.; Rossi, D. L.; Zurawski, S. M.; Vega, F., Jr.; Kastelein, R. A.; Wagner, J. L.;
47 Hannum, C. H.; Zlotnik, A., Mouse IL-17: a cytokine preferentially expressed by alpha beta TCR + CD4-
48 CD8-T cells. *J Interferon Cytokine Res* **1996**, *16* (8), 611-7.
- 49 91. Chabaud, M.; Lubberts, E.; Joosten, L.; van Den Berg, W.; Miossec, P., IL-17 derived from
50 juxta-articular bone and synovium contributes to joint degradation in rheumatoid arthritis. *Arthritis*
51 *Res* **2001**, *3* (3), 168-77.

- 1 92. Di Sabatino, A.; Pickard, K. M.; Rampton, D.; Kruidenier, L.; Rovedatti, L.; Leakey, N. A.;
2 Corazza, G. R.; Monteleone, G.; MacDonald, T. T., Blockade of transforming growth factor beta
3 upregulates T-box transcription factor T-bet, and increases T helper cell type 1 cytokine and matrix
4 metalloproteinase-3 production in the human gut mucosa. *Gut* **2008**, *57* (5), 605-12.
- 5 93. Babyatsky, M. W.; Rossiter, G.; Podolsky, D. K., Expression of transforming growth factors
6 alpha and beta in colonic mucosa in inflammatory bowel disease. *Gastroenterology* **1996**, *110* (4),
7 975-84.
- 8 94. Marie, J. C.; Liggitt, D.; Rudensky, A. Y., Cellular mechanisms of fatal early-onset
9 autoimmunity in mice with the T cell-specific targeting of transforming growth factor-beta receptor.
10 *Immunity* **2006**, *25* (3), 441-54.
- 11 95. Cazac, B. B.; Roes, J., TGF-beta receptor controls B cell responsiveness and induction of IgA in
12 vivo. *Immunity* **2000**, *13* (4), 443-51.
- 13 96. Ramalingam, R.; Larmonier, C. B.; Thurston, R. D.; Midura-Kiela, M. T.; Zheng, S. G.;
14 Ghishan, F. K.; Kiela, P. R., Dendritic cell-specific disruption of TGF- β receptor II leads to altered
15 regulatory T cell phenotype and spontaneous multiorgan autoimmunity. *J Immunol* **2012**, *189* (8),
16 3878-93.
- 17 97. Rani, R.; Smulian, A. G.; Greaves, D. R.; Hogan, S. P.; Herbert, D. R., TGF- β limits IL-33
18 production and promotes the resolution of colitis through regulation of macrophage function. *Eur J*
19 *Immunol* **2011**, *41* (7), 2000-9.
- 20 98. Opp, M. R.; Smith, E. M.; Hughes, T. K., Jr., Interleukin-10 (cytokine synthesis inhibitory
21 factor) acts in the central nervous system of rats to reduce sleep. *J Neuroimmunol* **1995**, *60* (1-2),
22 165-8.
- 23 99. Aste-Amezaga, M.; Ma, X.; Sartori, A.; Trinchieri, G., Molecular mechanisms of the induction
24 of IL-12 and its inhibition by IL-10. *J Immunol* **1998**, *160* (12), 5936-44.
- 25 100. Baumgart, D. C., The diagnosis and treatment of Crohn's disease and ulcerative colitis. *Dtsch*
26 *Arztebl Int* **2009**, *106* (8), 123-33.
- 27 101. Dubois-Camacho, K.; Ottum, P. A.; Franco-Muñoz, D.; De la Fuente, M.; Torres-Riquelme,
28 A.; Díaz-Jiménez, D.; Olivares-Morales, M.; Astudillo, G.; Quera, R.; Hermoso, M. A.,
29 Glucocorticosteroid therapy in inflammatory bowel diseases: From clinical practice to molecular
30 biology. *World J Gastroenterol* **2017**, *23* (36), 6628-6638.
- 31 102. Rutgeerts, P. J., Review article: the limitations of corticosteroid therapy in Crohn's disease.
32 *Aliment Pharmacol Ther* **2001**, *15* (10), 1515-25.
- 33 103. Buchman, A. L., Side effects of corticosteroid therapy. *J Clin Gastroenterol* **2001**, *33* (4), 289-
34 94.
- 35 104. Moghadam-Kia, S.; Werth, V. P., Prevention and treatment of systemic glucocorticoid side
36 effects. *Int J Dermatol* **2010**, *49* (3), 239-48.
- 37 105. Zhou, H.; Qian, H., Preparation and characterization of pH-sensitive nanoparticles of
38 budesonide for the treatment of ulcerative colitis. *Drug Des Devel Ther* **2018**, *12*, 2601-2609.
- 39 106. Peyrin-Biroulet, L., Anti-TNF therapy in inflammatory bowel diseases: a huge review. *Minerva*
40 *Gastroenterol Dietol* **2010**, *56* (2), 233-43.
- 41 107. Kashani, A.; Schwartz, D. A., The Expanding Role of Anti-IL-12 and/or Anti-IL-23 Antibodies in
42 the Treatment of Inflammatory Bowel Disease. *Gastroenterol Hepatol (N Y)* **2019**, *15* (5), 255-265.
- 43 108. Gaujoux, R.; Starosvetsky, E.; Maimon, N.; Vallania, F.; Bar-Yoseph, H.; Pressman, S.;
44 Weisshof, R.; Goren, I.; Rabinowitz, K.; Waterman, M.; Yanai, H.; Dotan, I.; Sabo, E.; Chowers, Y.;
45 Khatri, P.; Shen-Orr, S. S., Cell-centred meta-analysis reveals baseline predictors of anti-TNF α non-
46 response in biopsy and blood of patients with IBD. *Gut* **2019**, *68* (4), 604-614.
- 47 109. Vulliamoz, M.; Brand, S.; Juillerat, P.; Mottet, C.; Ben-Horin, S.; Michetti, P., TNF-Alpha
48 Blockers in Inflammatory Bowel Diseases: Practical Recommendations and a User's Guide: An
49 Update. *Digestion* **2020**, *101 Suppl 1*, 16-26.
- 50 110. Targan, S. R.; Hanauer, S. B.; van Deventer, S. J.; Mayer, L.; Present, D. H.; Braakman, T.;
51 DeWoody, K. L.; Schaible, T. F.; Rutgeerts, P. J., A short-term study of chimeric monoclonal antibody

1 cA2 to tumor necrosis factor alpha for Crohn's disease. Crohn's Disease cA2 Study Group. *N Engl J*
2 *Med* **1997**, 337 (15), 1029-35.

3 111. Present, D. H.; Rutgeerts, P.; Targan, S.; Hanauer, S. B.; Mayer, L.; van Hogezaand, R. A.;
4 Podolsky, D. K.; Sands, B. E.; Braakman, T.; DeWoody, K. L.; Schaible, T. F.; van Deventer, S. J.,
5 Infliximab for the treatment of fistulas in patients with Crohn's disease. *N Engl J Med* **1999**, 340 (18),
6 1398-405.

7 112. Van Assche, G.; Vermeire, S.; Rutgeerts, P., Safety issues with biological therapies for
8 inflammatory bowel disease. *Curr Opin Gastroenterol* **2006**, 22 (4), 370-6.

9 113. Roblin, X.; Marotte, H.; Rinaudo, M.; Del Tedesco, E.; Moreau, A.; Phelip, J. M.; Genin, C.;
10 Peyrin-Biroulet, L.; Paul, S., Association between pharmacokinetics of adalimumab and mucosal
11 healing in patients with inflammatory bowel diseases. *Clinical Gastroenterology and Hepatology*
12 **2014**, 12 (1), 80-84. e2.

13 114. Bongartz, T.; Sutton, A. J.; Sweeting, M. J.; Buchan, I.; Matteson, E. L.; Montori, V., Anti-TNF
14 antibody therapy in rheumatoid arthritis and the risk of serious infections and malignancies:
15 systematic review and meta-analysis of rare harmful effects in randomized controlled trials. *Jama*
16 **2006**, 295 (19), 2275-85.

17 115. Micic, D.; Komaki, Y.; Alavanja, A.; Rubin, D. T.; Sakuraba, A., Risk of Cancer Recurrence
18 Among Individuals Exposed to Antitumor Necrosis Factor Therapy: A Systematic Review and Meta-
19 Analysis of Observational Studies. *J Clin Gastroenterol* **2019**, 53 (1), e1-e11.

20 116. Schoellhammer, C. M.; Schroeder, A.; Maa, R.; Lauwers, G. Y.; Swiston, A.; Zervas, M.;
21 Barman, R.; DiCiccio, A. M.; Brugge, W. R.; Anderson, D. G.; Blankschein, D.; Langer, R.; Traverso,
22 G., Ultrasound-mediated gastrointestinal drug delivery. *Sci Transl Med* **2015**, 7 (310), 310ra168.

23 117. Zhang, S.; Ermann, J.; Succi, M. D.; Zhou, A.; Hamilton, M. J.; Cao, B.; Korzenik, J. R.;
24 Glickman, J. N.; Vemula, P. K.; Glimcher, L. H.; Traverso, G.; Langer, R.; Karp, J. M., An
25 inflammation-targeting hydrogel for local drug delivery in inflammatory bowel disease. *Sci Transl*
26 *Med* **2015**, 7 (300), 300ra128.

27 118. Ali, H.; Weigmann, B.; Collnot, E. M.; Khan, S. A.; Windbergs, M.; Lehr, C. M., Budesonide
28 Loaded PLGA Nanoparticles for Targeting the Inflamed Intestinal Mucosa--Pharmaceutical
29 Characterization and Fluorescence Imaging. *Pharm Res* **2016**, 33 (5), 1085-92.

30 119. Giri, B. R.; Kim, J. S.; Park, J. H.; Jin, S. G.; Kim, K. S.; Din, F. U.; Choi, H. G.; Kim, D. W.,
31 Improved Bioavailability and High Photostability of Methotrexate by Spray-Dried Surface-Attached
32 Solid Dispersion with an Aqueous Medium. *Pharmaceutics* **2021**, 13 (1).

33 120. Czogalla, A., Oral cyclosporine A--the current picture of its liposomal and other delivery
34 systems. *Cell Mol Biol Lett* **2009**, 14 (1), 139-52.

35 121. Tamura, S.; Ohike, A.; Ibuki, R.; Amidon, G. L.; Yamashita, S., Tacrolimus is a class II low-
36 solubility high-permeability drug: the effect of P-glycoprotein efflux on regional permeability of
37 tacrolimus in rats. *J Pharm Sci* **2002**, 91 (3), 719-29.

38 122. Pichai, M. V.; Ferguson, L. R., Potential prospects of nanomedicine for targeted therapeutics
39 in inflammatory bowel diseases. *World J Gastroenterol* **2012**, 18 (23), 2895-901.

40 123. Wang, Y.; Pi, C.; Feng, X.; Hou, Y.; Zhao, L.; Wei, Y., The Influence of Nanoparticle Properties
41 on Oral Bioavailability of Drugs. *Int J Nanomedicine* **2020**, 15, 6295-6310.

42 124. Laroui, H.; Wilson, D. S.; Dalmasso, G.; Salaita, K.; Murthy, N.; Sitaraman, S. V.; Merlin, D.,
43 Nanomedicine in GI. *Am J Physiol Gastrointest Liver Physiol* **2011**, 300 (3), G371-83.

44 125. Nunes, R.; Neves, J. d.; Sarmiento, B., Nanoparticles for the regulation of intestinal
45 inflammation: Opportunities and challenges. *Nanomedicine* **2019**, 14 (19), 2631-2644.

46 126. Cui, M.; Zhang, M.; Liu, K., Colon-targeted drug delivery of polysaccharide-based
47 nanocarriers for synergistic treatment of inflammatory bowel disease: A review. *Carbohydrate*
48 *Polymers* **2021**, 118530.

49 127. Zhang, Y.; Thanou, M.; Vllasaliu, D., Exploiting disease-induced changes for targeted oral
50 delivery of biologics and nanomedicines in inflammatory bowel disease. *European Journal of*
51 *Pharmaceutics and Biopharmaceutics* **2020**, 155, 128-138.

- 1 128. Franzosa, E. A.; Sirota-Madi, A.; Avila-Pacheco, J.; Fornelos, N.; Haiser, H. J.; Reinker, S.;
2 Vatanen, T.; Hall, A. B.; Mallick, H.; McIver, L. J., Gut microbiome structure and metabolic activity in
3 inflammatory bowel disease. *Nature microbiology* **2019**, *4* (2), 293-305.
- 4 129. Turpin, W.; Lee, S.-H.; Garay, J. A. R.; Madsen, K. L.; Meddings, J. B.; Bedrani, L.; Power, N.;
5 Espin-Garcia, O.; Xu, W.; Smith, M. I., Increased intestinal permeability is associated with later
6 development of Crohn's disease. *Gastroenterology* **2020**, *159* (6), 2092-2100. e5.
- 7 130. Press, A.; Hauptmann, I.; Hauptmann, L.; Fuchs, B.; Fuchs, M.; Ewe, K.; Ramadori, G.,
8 Gastrointestinal pH profiles in patients with inflammatory bowel disease. *Alimentary pharmacology*
9 *& therapeutics* **1998**, *12* (7), 673-678.
- 10 131. Ewe, K.; Schwartz, S.; Petersen, S.; Press, A. G., Inflammation does not decrease intraluminal
11 pH in chronic inflammatory bowel disease. *Digestive diseases and sciences* **1999**, *44* (7), 1434-1439.
- 12 132. Fallingborg, J.; Pedersen, P.; Jacobsen, B. A., Small intestinal transit time and intraluminal pH
13 in ileocecal resected patients with Crohn's disease. *Digestive diseases and sciences* **1998**, *43* (4), 702-
14 705.
- 15 133. McConnell, E. L.; Fadda, H. M.; Basit, A. W., Gut instincts: explorations in intestinal
16 physiology and drug delivery. *International journal of pharmaceutics* **2008**, *364* (2), 213-226.
- 17 134. Nugent, S.; Kumar, D.; Rampton, D.; Evans, D., Intestinal luminal pH in inflammatory bowel
18 disease: possible determinants and implications for therapy with aminosalicylates and other drugs.
19 *Gut* **2001**, *48* (4), 571-577.
- 20 135. Sasaki, Y.; Hada, R.; Nakajima, H.; Fukuda, S.; Munakata, A., Improved localizing method of
21 radiopill in measurement of entire gastrointestinal pH profiles: colonic luminal pH in normal subjects
22 and patients with Crohn's disease. *American Journal of Gastroenterology (Springer Nature)* **1997**, *92*
23 (1).
- 24 136. Fallingborg, J.; Christensen, L. A.; Jacobsen, B. A.; Rasmussen, S. N., Very low intraluminal
25 colonic pH in patients with active ulcerative colitis. *Digestive diseases and sciences* **1993**, *38* (11),
26 1989-1993.
- 27 137. Qelliny, M. R.; Aly, U. F.; Elgarhy, O. H.; Khaled, K. A., Budesonide-loaded Eudragit S 100
28 nanocapsules for the treatment of acetic acid-induced colitis in animal model. *AAPS PharmSciTech*
29 **2019**, *20* (6), 1-17.
- 30 138. Qu, Z.; Wong, K. Y.; Moniruzzaman, M.; Begun, J.; Santos, H. A.; Hasnain, S. Z.; Kumeria, T.;
31 McGuckin, M. A.; Popat, A., One-Pot Synthesis of pH-Responsive Eudragit-Mesoporous Silica
32 Nanocomposites Enable Colonic Delivery of Glucocorticoids for the Treatment of Inflammatory Bowel
33 Disease. *Advanced Therapeutics* **2021**, *4* (2), 2000165.
- 34 139. Gao, C.; Yu, S.; Zhang, X.; Dang, Y.; Han, D.-d.; Liu, X.; Han, J.; Hui, M., Dual Functional
35 Eudragit® S100/L30D-55 and PLGA Colon-Targeted Nanoparticles of Iridoid Glycoside for Improved
36 Treatment of Induced Ulcerative Colitis. *International Journal of Nanomedicine* **2021**, *16*, 1405.
- 37 140. Raj, P. M.; Raj, R.; Kaul, A.; Mishra, A. K.; Ram, A., Biodistribution and targeting potential
38 assessment of mucoadhesive chitosan nanoparticles designed for ulcerative colitis via scintigraphy.
39 *RSC advances* **2018**, *8* (37), 20809-20821.
- 40 141. Ali, H.; Weigmann, B.; Neurath, M. F.; Collnot, E. M.; Windbergs, M.; Lehr, C. M.,
41 Budesonide loaded nanoparticles with pH-sensitive coating for improved mucosal targeting in mouse
42 models of inflammatory bowel diseases. *J Control Release* **2014**, *183*, 167-77.
- 43 142. Mishra, R. K.; Ahmad, A.; Kumar, A.; Vyawahare, A.; Raza, S. S.; Khan, R., Lipid-based
44 nanocarrier-mediated targeted delivery of celecoxib attenuate severity of ulcerative colitis. *Materials*
45 *Science and Engineering: C* **2020**, *116*, 111103.
- 46 143. Zhang, L.; Sang, Y.; Feng, J.; Li, Z.; Zhao, A., Polysaccharide-based micro/nanocarriers for
47 oral colon-targeted drug delivery. *Journal of drug targeting* **2016**, *24* (7), 579-589.
- 48 144. Naeem, M.; Kim, W.; Cao, J.; Jung, Y.; Yoo, J.-W., Enzyme/pH dual sensitive polymeric
49 nanoparticles for targeted drug delivery to the inflamed colon. *Colloids and Surfaces B: Biointerfaces*
50 **2014**, *123*, 271-278.

- 1 145. Naeem, M.; Cao, J.; Choi, M.; Kim, W. S.; Moon, H. R.; Lee, B. L.; Kim, M.-S.; Jung, Y.; Yoo,
2 J.-W., Enhanced therapeutic efficacy of budesonide in experimental colitis with enzyme/pH dual-
3 sensitive polymeric nanoparticles. *International journal of nanomedicine* **2015**, *10*, 4565.
- 4 146. Girod, S.; Zahm, J.; Plotkowski, C.; Beck, G.; Puchelle, E., Role of the physicochemical
5 properties of mucus in the protection of the respiratory epithelium. *European Respiratory Journal*
6 **1992**, *5* (4), 477-487.
- 7 147. Harbitz, O.; Jenssen, A.; Smidsrød, O., Lysozyme and lactoferrin in sputum from patients with
8 chronic obstructive lung disease. *European journal of respiratory diseases* **1984**, *65* (7), 512-520.
- 9 148. Shogren, R.; Gerken, T. A.; Jentoft, N., Role of glycosylation on the conformation and chain
10 dimensions of O-linked glycoproteins: light-scattering studies of ovine submaxillary mucin.
11 *Biochemistry* **1989**, *28* (13), 5525-5536.
- 12 149. Thornton, D. J.; Sheehan, J. K., From mucins to mucus: toward a more coherent
13 understanding of this essential barrier. *Proceedings of the American Thoracic Society* **2004**, *1* (1), 54-
14 61.
- 15 150. Carlstedt, I.; Sheehan, J. K., Macromolecular properties and polymeric structure of mucus
16 glycoproteins. *Mucus and mucosa* **1984**, *109*, 157-172.
- 17 151. Lamblin, G.; Lhermitte, M.; Klein, A.; Houdret, N.; Scharfman, A.; Ramphal, R.; Roussel, P.,
18 The carbohydrate diversity of human respiratory mucins: a protection of the underlying mucosa. *Am*
19 *Rev Respir Dis* **1991**, *144* (3 Pt 2), S19-S24.
- 20 152. Lai, S. K.; Wang, Y.-Y.; Wirtz, D.; Hanes, J., Micro-and macrorheology of mucus. *Advanced*
21 *drug delivery reviews* **2009**, *61* (2), 86-100.
- 22 153. Cone, R. A., Barrier properties of mucus. *Advanced drug delivery reviews* **2009**, *61* (2), 75-85.
- 23 154. Leal, J.; Smyth, H. D.; Ghosh, D., Physicochemical properties of mucus and their impact on
24 transmucosal drug delivery. *International journal of pharmaceuticals* **2017**, *532* (1), 555-572.
- 25 155. Pullan, R.; Thomas, G.; Rhodes, M.; Newcombe, R.; Williams, G.; Allen, A.; Rhodes, J.,
26 Thickness of adherent mucus gel on colonic mucosa in humans and its relevance to colitis. *Gut* **1994**,
27 *35* (3), 353-359.
- 28 156. Johansson, M. E.; Gustafsson, J. K.; Holmén-Larsson, J.; Jabbar, K. S.; Xia, L.; Xu, H.;
29 Ghishan, F. K.; Carvalho, F. A.; Gewirtz, A. T.; Sjövall, H., Bacteria penetrate the normally
30 impenetrable inner colon mucus layer in both murine colitis models and patients with ulcerative
31 colitis. *Gut* **2014**, *63* (2), 281-291.
- 32 157. Suk, J. S.; Lai, S. K.; Boylan, N. J.; Dawson, M. R.; Boyle, M. P.; Hanes, J., Rapid transport of
33 muco-inert nanoparticles in cystic fibrosis sputum treated with N-acetyl cysteine. *Nanomedicine*
34 **2011**, *6* (2), 365-375.
- 35 158. Bandi, S. P.; Kumbhar, Y. S.; Venuganti, V. V. K., Effect of particle size and surface charge of
36 nanoparticles in penetration through intestinal mucus barrier. *Journal of Nanoparticle Research*
37 **2020**, *22* (3), 1-11.
- 38 159. Lamprecht, A.; Schäfer, U.; Lehr, C.-M., Size-dependent bioadhesion of micro-and
39 nanoparticulate carriers to the inflamed colonic mucosa. *Pharmaceutical research* **2001**, *18* (6), 788-
40 793.
- 41 160. Hardy, J.; Davis, S.; Khosla, R.; Robertson, C., Gastrointestinal transit of small tablets in
42 patients with ulcerative colitis. *International journal of pharmaceuticals* **1988**, *48* (1-3), 79-82.
- 43 161. Watts, P.; Barrow, L.; Steed, K.; Wilson, C.; Spiller, R.; Melia, C.; Davies, M., The transit rate
44 of different-sized model dosage forms through the human colon and the effects of a lactulose-
45 induced catharsis. *International journal of pharmaceuticals* **1992**, *87* (1-3), 215-221.
- 46 162. Cu, Y.; Saltzman, W. M., Controlled surface modification with poly (ethylene) glycol enhances
47 diffusion of PLGA nanoparticles in human cervical mucus. *Molecular pharmaceuticals* **2009**, *6* (1), 173-
48 181.
- 49 163. Tang, B. C.; Dawson, M.; Lai, S. K.; Wang, Y.-Y.; Suk, J. S.; Yang, M.; Zeitlin, P.; Boyle, M. P.;
50 Fu, J.; Hanes, J., Biodegradable polymer nanoparticles that rapidly penetrate the human mucus
51 barrier. *Proceedings of the National Academy of Sciences* **2009**, *106* (46), 19268-19273.

- 1 164. Lai, S. K.; Wang, Y.-Y.; Hanes, J., Mucus-penetrating nanoparticles for drug and gene delivery
2 to mucosal tissues. *Advanced drug delivery reviews* **2009**, *61* (2), 158-171.
- 3 165. Iqbal, S.; Du, X.; Wang, J.; Li, H.; Yuan, Y.; Wang, J., Surface charge tunable nanoparticles for
4 TNF- α siRNA oral delivery for treating ulcerative colitis. *Nano Research* **2018**, *11* (5), 2872-2884.
- 5 166. Theodoratou, E.; Campbell, H.; Ventham, N. T.; Kolarich, D.; Pučić-Baković, M.; Zoldoš, V.;
6 Fernandes, D.; Pemberton, I. K.; Rudan, I.; Kennedy, N. A., The role of glycosylation in IBD. *Nature*
7 *reviews Gastroenterology & hepatology* **2014**, *11* (10), 588-600.
- 8 167. Campbell, B. J.; Yu, L.-G.; Rhodes, J. M., Altered glycosylation in inflammatory bowel disease:
9 a possible role in cancer development. *Glycoconjugate journal* **2001**, *18* (11), 851-858.
- 10 168. Kudelka, M. R.; Stowell, S. R.; Cummings, R. D.; Neish, A. S., Intestinal epithelial glycosylation
11 in homeostasis and gut microbiota interactions in IBD. *Nature Reviews Gastroenterology &*
12 *Hepatology* **2020**, *17* (10), 597-617.
- 13 169. Larsson, J. M. H.; Karlsson, H.; Crespo, J. G.; Johansson, M. E.; Eklund, L.; Sjövall, H.;
14 Hansson, G. C., Altered O-glycosylation profile of MUC2 mucin occurs in active ulcerative colitis and is
15 associated with increased inflammation. *Inflammatory bowel diseases* **2011**, *17* (11), 2299-2307.
- 16 170. Raouf, A.; Tsai, H.; Parker, N.; Hoffman, J.; Walker, R.; Rhodes, J. M., Sulphation of colonic
17 and rectal mucin in inflammatory bowel disease: reduced sulphation of rectal mucus in ulcerative
18 colitis. *Clinical Science* **1992**, *83* (5), 623-626.
- 19 171. Clamp, J.; Fraser, G.; Read, A., Study of the carbohydrate content of mucus glycoproteins
20 from normal and diseased colons. *Clinical Science* **1981**, *61* (2), 229-234.
- 21 172. Bergstrom, K.; Fu, J.; Johansson, M. E.; Liu, X.; Gao, N.; Wu, Q.; Song, J.; McDaniel, J. M.;
22 McGee, S.; Chen, W., Core 1- and 3-derived O-glycans collectively maintain the colonic mucus barrier
23 and protect against spontaneous colitis in mice. *Mucosal immunology* **2017**, *10* (1), 91-103.
- 24 173. Zhao, M.; Xiong, X.; Ren, K.; Xu, B.; Cheng, M.; Sahu, C.; Wu, K.; Nie, Y.; Huang, Z.;
25 Blumberg, R. S., Deficiency in intestinal epithelial O-GlcNAcylation predisposes to gut inflammation.
26 *EMBO molecular medicine* **2018**, *10* (8), e8736.
- 27 174. Schelwies, M.; Brinson, D.; Otsuki, S.; Hong, Y.-H.; Lotz, M. K.; Wong, C.-H.; Hanson, S. R.,
28 Glucosamine-6-sulfamate Analogs of Heparan Sulfate as Inhibitors of Endosulfatases. *Chembiochem:*
29 *a European journal of chemical biology* **2010**, *11* (17), 2393.
- 30 175. Parker, R. B.; Kohler, J. J., Regulation of intracellular signaling by extracellular glycan
31 remodeling. *ACS chemical biology* **2010**, *5* (1), 35-46.
- 32 176. Weiss, E.; Shanitzki, B.; Dotan, M.; Ganeshkumar, N.; Kolenbrander, P.; Metzger, Z.,
33 Attachment of *Fusobacterium nucleatum* PK1594 to mammalian cells and its coaggregation with
34 periodontopathogenic bacteria are mediated by the same galactose-binding adhesin. *Oral*
35 *microbiology and immunology* **2000**, *15* (6), 371-377.
- 36 177. Mongiello, J.; Falkler Jr, W., Sugar inhibition of oral *Fusobacterium nucleatum*
37 haemagglutination and cell binding. *Archives of oral biology* **1979**, *24* (7), 539-545.
- 38 178. Stokowa-Sottys, K.; Wojtkowiak, K.; Jagiełło, K., *Fusobacterium nucleatum*—Friend or foe?
39 *Journal of Inorganic Biochemistry* **2021**, *224*, 111586.
- 40 179. Chevalier, G.; Laveissière, A.; Desachy, G.; Barnich, N.; Sivignon, A.; Maresca, M.; Nicoletti,
41 C.; Di Pasquale, E.; Martinez-Medina, M.; Simpson, K. W., Blockage of bacterial FimH prevents
42 mucosal inflammation associated with Crohn's disease. *Microbiome* **2021**, *9* (1), 1-16.
- 43 180. Reinisch, W.; Hébuterne, X.; Buisson, A.; Schreiber, S.; Desreumaux, P.; Paillarse, J.; Bonny,
44 C., P568 An open-label, multicenter, phase Ib, pharmacokinetic (pk) and safety study of a fimh
45 blocker, Sibofimloc (TAK-018/EB8018), in patients with Crohn's disease (CD). *Journal of Crohn's and*
46 *Colitis* **2020**, *14* (Supplement_1), S479-S480.
- 47 181. Nagashima, R., Mechanisms of action of sucralfate. *Journal of clinical gastroenterology* **1981**,
48 *3* (Suppl 2), 117-127.
- 49 182. Tirosh, B.; Khatib, N.; Barenholz, Y.; Nissan, A.; Rubinstein, A., Transferrin as a luminal target
50 for negatively charged liposomes in the inflamed colonic mucosa. *Molecular pharmaceuticals* **2009**, *6*
51 (4), 1083-1091.

- 1 183. Carlson, M.; Raab, Y.; Peterson, C.; Hällgren, R.; Venge, P., Increased intraluminal release of
2 eosinophil granule proteins EPO, ECP, EPX, and cytokines in ulcerative colitis and proctitis in
3 segmental perfusion. *The American journal of gastroenterology* **1999**, *94* (7), 1876-1883.
- 4 184. Peterson, C. G.; Eklund, E.; Taha, Y.; Raab, Y.; Carlson, M., A new method for the
5 quantification of neutrophil and eosinophil cationic proteins in feces: establishment of normal levels
6 and clinical application in patients with inflammatory bowel disease. *The American journal of*
7 *gastroenterology* **2002**, *97* (7), 1755-1762.
- 8 185. Casado-Bedmar, M.; de-Faria, F. M.; Biskou, O.; Lindqvist, C. M.; Ranasinghe, P. D.;
9 Bednarska, O.; Peterson, C.; Walter, S. A.; Carlson, M.; Keita, Å. V., Elevated F-EDN correlates with
10 mucosal eosinophil degranulation in patients with IBS—A possible association with microbiota?
11 *Journal of Leukocyte Biology* **2021**.
- 12 186. Lamprecht, A.; Ubrich, N.; Yamamoto, H.; Schäfer, U.; Takeuchi, H.; Maincent, P.;
13 Kawashima, Y.; Lehr, C.-M., Biodegradable nanoparticles for targeted drug delivery in treatment of
14 inflammatory bowel disease. *Journal of Pharmacology and Experimental Therapeutics* **2001**, *299* (2),
15 775-781.
- 16 187. Beloqui, A.; Coco, R.; Alhouayek, M.; Solinís, M. Á.; Rodríguez-Gascón, A.; Muccioli, G. G.;
17 Préat, V., Budesonide-loaded nanostructured lipid carriers reduce inflammation in murine DSS-
18 induced colitis. *International journal of pharmaceutics* **2013**, *454* (2), 775-783.
- 19 188. Vergnolle, N., Protease inhibition as new therapeutic strategy for GI diseases. *Gut* **2016**, *65*
20 (7), 1215-1224.
- 21 189. Lee, J. Y.; Wasinger, V. C.; Yau, Y. Y.; Chuang, E.; Yajnik, V.; Leong, R. W., Molecular
22 pathophysiology of epithelial barrier dysfunction in inflammatory bowel diseases. *Proteomes* **2018**, *6*
23 (2), 17.
- 24 190. Yao, X.; Cadwell, K., Tumor Necrosis Factor- α -Induced Apoptosis in the Intestinal Epithelium
25 due to Chronic Nuclear Factor Kappa B Signaling Is Mediated by Receptor Interacting
26 Serine/Threonine Kinase 1. *Cellular and molecular gastroenterology and hepatology* **2020**, *9* (2), 337-
27 338.
- 28 191. Zeissig, S.; Bürgel, N.; Günzel, D.; Richter, J.; Mankertz, J.; Wahnschaffe, U.; Kroesen, A. J.;
29 Zeitz, M.; Fromm, M.; Schulzke, J. D., Changes in expression and distribution of claudin 2, 5 and 8
30 lead to discontinuous tight junctions and barrier dysfunction in active Crohn's disease. *Gut* **2007**, *56*
31 (1), 61-72.
- 32 192. Makhlof, A.; Tozuka, Y.; Takeuchi, H., pH-Sensitive nanospheres for colon-specific drug
33 delivery in experimentally induced colitis rat model. *Eur J Pharm Biopharm* **2009**, *72* (1), 1-8.
- 34 193. Ahmad, A.; Ansari, M. M.; Kumar, A.; Bishnoi, M.; Raza, S. S.; Khan, R., Aminocellulose-
35 grafted polycaprolactone-coated core-shell nanoparticles alleviate the severity of ulcerative colitis: a
36 novel adjuvant therapeutic approach. *Biomaterials Science* **2021**, *9* (17), 5868-5883.
- 37 194. Zhang, M.; Merlin, D., Nanoparticle-based oral drug delivery systems targeting the colon for
38 treatment of ulcerative colitis. *Inflammatory bowel diseases* **2018**, *24* (7), 1401-1415.
- 39 195. Liu, P.; Gao, C.; Chen, H.; Vong, C. T.; Wu, X.; Tang, X.; Wang, S.; Wang, Y., Receptor-
40 mediated targeted drug delivery systems for treatment of inflammatory bowel disease: opportunities
41 and emerging strategies. *Acta Pharmaceutica Sinica B* **2020**.
- 42 196. Zhang, X.; Wu, W., Ligand-mediated active targeting for enhanced oral absorption. *Drug*
43 *discovery today* **2014**, *19* (7), 898-904.
- 44 197. Sun, Q.; Arif, M.; Chi, Z.; Li, G.; Liu, C.-G., Macrophages-targeting mannosylated
45 nanoparticles based on inulin for the treatment of inflammatory bowel disease (IBD). *International*
46 *Journal of Biological Macromolecules* **2021**, *169*, 206-215.
- 47 198. Yang, H.; Zhu, C.; Yuan, W.; Wei, X.; Liu, C.; Huang, J.; Yuan, M.; Wu, Y.; Ling, Q.;
48 Hoffmann, P. R., Mannose-rich Oligosaccharides-functionalized Selenium Nanoparticles Mediates
49 Macrophage Reprogramming and Inflammation Resolution in Ulcerative Colitis. *Chemical Engineering*
50 *Journal* **2021**, 131715.

- 1 199. Mukhtar, M.; Zeshan, M.; Khan, S.; Shahnaz, G.; Khan, S. A.; Sarwar, H. S.; Pasha, R. A.;
2 Ali, H., Fabrication and optimization of pH-sensitive mannose-anchored nano-vehicle as a promising
3 approach for macrophage uptake. *Applied Nanoscience* **2020**, *10* (11), 4013-4027.
- 4 200. Gan, J.; Dou, Y.; Li, Y.; Wang, Z.; Wang, L.; Liu, S.; Li, Q.; Yu, H.; Liu, C.; Han, C., Producing
5 anti-inflammatory macrophages by nanoparticle-triggered clustering of mannose receptors.
6 *Biomaterials* **2018**, *178*, 95-108.
- 7 201. Zhang, J.; Tang, C.; Yin, C., Galactosylated trimethyl chitosan–cysteine nanoparticles loaded
8 with Map4k4 siRNA for targeting activated macrophages. *Biomaterials* **2013**, *34* (14), 3667-3677.
- 9 202. Dang, P. M.-C.; Rolas, L.; El Benna, J., The dual role of ROS-generating NADPH oxidases in
10 gastrointestinal inflammation and therapeutic perspectives. *Antioxidants and Redox Signaling* **2020**,
11 *33* (5), 354-373.
- 12 203. Simmonds, N. J.; Allen, R. E.; Stevens, T. R.; Niall, R.; Van Someren, M.; Blake, D. R.;
13 Rampton, D. S., Chemiluminescence assay of mucosal reactive oxygen metabolites in inflammatory
14 bowel disease. *Gastroenterology* **1992**, *103* (1), 186-196.
- 15 204. Sinhmar, G. K.; Shah, N. N.; Rawal, S. U.; Chokshi, N. V.; Khatri, H. N.; Patel, B. M.; Patel, M.
16 M., Surface engineered lipid nanoparticle-mediated site-specific drug delivery system for the
17 treatment of inflammatory bowel disease. *Artif Cells Nanomed Biotechnol* **2018**, *46* (sup2), 565-578.
- 18 205. Nguyen, C. T.; Webb, R. I.; Lambert, L. K.; Strounina, E.; Lee, E. C.; Parat, M. O.; McGuckin,
19 M. A.; Popat, A.; Cabot, P. J.; Ross, B. P., Bifunctional Succinylated ϵ -Polylysine-Coated Mesoporous
20 Silica Nanoparticles for pH-Responsive and Intracellular Drug Delivery Targeting the Colon. *ACS Appl*
21 *Mater Interfaces* **2017**, *9* (11), 9470-9483.
- 22 206. Moulari, B.; Béduneau, A.; Pellequer, Y.; Lamprecht, A., Lectin-decorated nanoparticles
23 enhance binding to the inflamed tissue in experimental colitis. *J Control Release* **2014**, *188*, 9-17.
- 24 207. Pertuit, D.; Moulari, B.; Betz, T.; Nadaradjane, A.; Neumann, D.; Ismaïli, L.; Refouvelet, B.;
25 Pellequer, Y.; Lamprecht, A., 5-amino salicylic acid bound nanoparticles for the therapy of
26 inflammatory bowel disease. *J Control Release* **2007**, *123* (3), 211-8.
- 27 208. Moulari, B.; Pertuit, D.; Pellequer, Y.; Lamprecht, A., The targeting of surface modified silica
28 nanoparticles to inflamed tissue in experimental colitis. *Biomaterials* **2008**, *29* (34), 4554-60.
- 29 209. Tang, H.; Xiang, D.; Wang, F.; Mao, J.; Tan, X.; Wang, Y., 5-ASA-loaded SiO₂ nanoparticles-a
30 novel drug delivery system targeting therapy on ulcerative colitis in mice. *Mol Med Rep* **2017**, *15* (3),
31 1117-1122.
- 32 210. Vong, L. B.; Tomita, T.; Yoshitomi, T.; Matsui, H.; Nagasaki, Y., An orally administered redox
33 nanoparticle that accumulates in the colonic mucosa and reduces colitis in mice. *Gastroenterology*
34 **2012**, *143* (4), 1027-36.e3.
- 35 211. Vong, L. B.; Yoshitomi, T.; Morikawa, K.; Saito, S.; Matsui, H.; Nagasaki, Y., Oral
36 nanotherapeutics: effect of redox nanoparticle on microflora in mice with dextran sodium sulfate-
37 induced colitis. *Journal of gastroenterology* **2014**, *49* (5), 806-813.
- 38 212. Wang, X.; Yan, J.; Wang, L.; Pan, D.; Xu, Y.; Wang, F.; Sheng, J.; Li, X.; Yang, M., Oral
39 delivery of anti-TNF antibody shielded by natural polyphenol-mediated supramolecular assembly for
40 inflammatory bowel disease therapy. *Theranostics* **2020**, *10* (23), 10808-10822.
- 41 213. Wilson, D. S.; Dalmaso, G.; Wang, L.; Sitaraman, S. V.; Merlin, D.; Murthy, N., Orally
42 delivered thioketal nanoparticles loaded with TNF- α -siRNA target inflammation and inhibit gene
43 expression in the intestines. *Nat Mater* **2010**, *9* (11), 923-8.
- 44 214. Laroui, H.; Geem, D.; Xiao, B.; Viennois, E.; Rakhya, P.; Denning, T.; Merlin, D., Targeting
45 intestinal inflammation with CD98 siRNA/PEI-loaded nanoparticles. *Molecular Therapy* **2014**, *22* (1),
46 69-80.
- 47 215. Xiao, B.; Zhang, Z.; Viennois, E.; Kang, Y.; Zhang, M.; Han, M. K.; Chen, J.; Merlin, D.,
48 Combination Therapy for Ulcerative Colitis: Orally Targeted Nanoparticles Prevent Mucosal Damage
49 and Relieve Inflammation. *Theranostics* **2016**, *6* (12), 2250-2266.
- 50 216. Beloqui, A.; Coco, R.; Memvanga, P. B.; Ucakar, B.; des Rieux, A.; Pr at, V., pH-sensitive
51 nanoparticles for colonic delivery of curcumin in inflammatory bowel disease. *Int J Pharm* **2014**, *473*
52 (1-2), 203-12.

- 1 217. Gugulothu, D.; Kulkarni, A.; Patravale, V.; Dandekar, P., pH-sensitive nanoparticles of
2 curcumin-celecoxib combination: evaluating drug synergy in ulcerative colitis model. *J Pharm Sci*
3 **2014**, *103* (2), 687-96.
- 4 218. Laroui, H.; Dalmaso, G.; Nguyen, H. T. T.; Yan, Y.; Sitaraman, S. V.; Merlin, D., Drug-loaded
5 nanoparticles targeted to the colon with polysaccharide hydrogel reduce colitis in a mouse model.
6 *Gastroenterology* **2010**, *138* (3), 843-853. e2.
- 7 219. Lamprecht, A.; Ubrich, N.; Yamamoto, H.; Schäfer, U.; Takeuchi, H.; Maincent, P.;
8 Kawashima, Y.; Lehr, C. M., Biodegradable nanoparticles for targeted drug delivery in treatment of
9 inflammatory bowel disease. *J Pharmacol Exp Ther* **2001**, *299* (2), 775-81.
- 10 220. Meissner, Y.; Pellequer, Y.; Lamprecht, A., Nanoparticles in inflammatory bowel disease:
11 particle targeting versus pH-sensitive delivery. *Int J Pharm* **2006**, *316* (1-2), 138-43.
- 12 221. Guada, M.; Beloqui, A.; Alhouayek, M.; Muccioli, G. G.; Dios-Viéitez Mdel, C.; Préat, V.;
13 Blanco-Prieto, M. J., Cyclosporine A-loaded lipid nanoparticles in inflammatory bowel disease. *Int J*
14 *Pharm* **2016**, *503* (1-2), 196-8.
- 15 222. Melero, A.; Draheim, C.; Hansen, S.; Giner, E.; Carreras, J. J.; Talens-Visconti, R.; Garrigues,
16 T. M.; Peris, J. E.; Recio, M. C.; Giner, R.; Lehr, C. M., Targeted delivery of Cyclosporine A by
17 polymeric nanocarriers improves the therapy of inflammatory bowel disease in a relevant mouse
18 model. *Eur J Pharm Biopharm* **2017**, *119*, 361-371.
- 19 223. Wang, B.; Zhuang, X.; Deng, Z. B.; Jiang, H.; Mu, J.; Wang, Q.; Xiang, X.; Guo, H.; Zhang, L.;
20 Dryden, G.; Yan, J.; Miller, D.; Zhang, H. G., Targeted drug delivery to intestinal macrophages by
21 bioactive nanovesicles released from grapefruit. *Mol Ther* **2014**, *22* (3), 522-534.
- 22 224. Helmy, A. M.; Elsabahy, M.; Soliman, G. M.; Mahmoud, M. A.; Ibrahim, E. A., Development
23 and in vivo evaluation of chitosan beads for the colonic delivery of azathioprine for treatment of
24 inflammatory bowel disease. *Eur J Pharm Sci* **2017**, *109*, 269-279.
- 25 225. Kumar, J.; Newton, A. M. J., Rifaximin - Chitosan Nanoparticles for Inflammatory Bowel
26 Disease (IBD). *Recent Pat Inflamm Allergy Drug Discov* **2017**, *11* (1), 41-52.
- 27 226. Saadatzadeh, A.; Atyabi, F.; Fazeli, M. R.; Dinarvand, R.; Jamalifar, H.; Abdolghaffari, A. H.;
28 Mahdavian, P.; Mahbod, M.; Baeeri, M.; Baghaei, A., Biochemical and pathological evidences on the
29 benefit of a new biodegradable nanoparticles of probiotic extract in murine colitis. *Fundamental &*
30 *clinical pharmacology* **2012**, *26* (5), 589-598.
- 31 227. Theiss, A. L.; Laroui, H.; Obertone, T. S.; Chowdhury, I.; Thompson, W. E.; Merlin, D.;
32 Sitaraman, S. V., Nanoparticle-based therapeutic delivery of prohibitin to the colonic epithelial cells
33 ameliorates acute murine colitis. *Inflamm Bowel Dis* **2011**, *17* (5), 1163-76.
- 34 228. Zhang, M.; Viennois, E.; Prasad, M.; Zhang, Y.; Wang, L.; Zhang, Z.; Han, M. K.; Xiao, B.;
35 Xu, C.; Srinivasan, S.; Merlin, D., Edible ginger-derived nanoparticles: A novel therapeutic approach
36 for the prevention and treatment of inflammatory bowel disease and colitis-associated cancer.
37 *Biomaterials* **2016**, *101*, 321-40.
- 38 229. Yan, H.; Ajuwon, K. M., Butyrate modifies intestinal barrier function in IPEC-J2 cells through a
39 selective upregulation of tight junction proteins and activation of the Akt signaling pathway. *PLoS one*
40 **2017**, *12* (6), e0179586.
- 41 230. Wang, Y.; Guan, M.; Zhao, X.; Li, X., Effects of garlic polysaccharide on alcoholic liver fibrosis
42 and intestinal microflora in mice. *Pharm Biol* **2018**, *56* (1), 325-332.
- 43 231. Wang, Y.; Fei, Y.; Liu, L.; Xiao, Y.; Pang, Y.; Kang, J.; Wang, Z., Polygonatum odoratum
44 Polysaccharides Modulate Gut Microbiota and Mitigate Experimentally Induced Obesity in Rats. *Int J*
45 *Mol Sci* **2018**, *19* (11).
- 46 232. Lamas, A.; Regal, P.; Vázquez, B.; Cepeda, A.; Franco, C. M., Short chain fatty acids
47 commonly produced by gut microbiota influence Salmonella enterica motility, biofilm formation, and
48 gene expression. *Antibiotics* **2019**, *8* (4), 265.
- 49 233. Sun, Y.; O'Riordan, M. X., Regulation of bacterial pathogenesis by intestinal short-chain fatty
50 acids. *Advances in applied microbiology* **2013**, *85*, 93-118.
- 51 234. Tsugawa, H.; Kabe, Y.; Kanai, A.; Sugiura, Y.; Hida, S.; Taniguchi, S.; Takahashi, T.; Matsui,
52 H.; Yasukawa, Z.; Itou, H.; Takubo, K.; Suzuki, H.; Honda, K.; Handa, H.; Suematsu, M., Short-chain

1 fatty acids bind to apoptosis-associated speck-like protein to activate inflammasome complex to
2 prevent Salmonella infection. *PLoS Biol* **2020**, *18* (9), e3000813.

3 235. LeBlanc, J. G.; Chain, F.; Martín, R.; Bermúdez-Humarán, L. G.; Courau, S.; Langella, P.,
4 Beneficial effects on host energy metabolism of short-chain fatty acids and vitamins produced by
5 commensal and probiotic bacteria. *Microbial cell factories* **2017**, *16* (1), 1-10.

6 236. Macfarlane, S.; Macfarlane, G. T., Regulation of short-chain fatty acid production.
7 *Proceedings of the Nutrition Society* **2003**, *62* (1), 67-72.

8 237. McDonald, J. A.; Mullish, B. H.; Pechlivanis, A.; Liu, Z.; Brignardello, J.; Kao, D.; Holmes, E.;
9 Li, J. V.; Clarke, T. B.; Thursz, M. R., Inhibiting growth of *Clostridioides difficile* by restoring valerate,
10 produced by the intestinal microbiota. *Gastroenterology* **2018**, *155* (5), 1495-1507. e15.

11 238. Guo, C.; Wang, Y.; Zhang, S.; Zhang, X.; Du, Z.; Li, M.; Ding, K., *Crataegus pinnatifida*
12 polysaccharide alleviates colitis via modulation of gut microbiota and SCFAs metabolism.
13 *International Journal of Biological Macromolecules* **2021**, *181*, 357-368.

14 239. Ma, X.; X. Fan, P.; Li, L.; Qiao, S.; Zhang, G.; Li, D., Butyrate promotes the recovering of
15 intestinal wound healing through its positive effect on the tight junctions. *Journal of animal science*
16 **2012**, *90* (suppl_4), 266-268.

17 240. Peng, L.; Li, Z.-R.; Green, R. S.; Holzman, I. R.; Lin, J., Butyrate enhances the intestinal barrier
18 by facilitating tight junction assembly via activation of AMP-activated protein kinase in Caco-2 cell
19 monolayers. *The Journal of nutrition* **2009**, *139* (9), 1619-1625.

20 241. Wang, H.-B.; Wang, P.-Y.; Wang, X.; Wan, Y.-L.; Liu, Y.-C., Butyrate enhances intestinal
21 epithelial barrier function via up-regulation of tight junction protein Claudin-1 transcription. *Digestive*
22 *diseases and sciences* **2012**, *57* (12), 3126-3135.

23 242. Lai, H.; Zhu, F.; Yuan, W.; He, N.; Zhang, Z.; Zhang, X.; He, Q., Development of multiple-unit
24 colon-targeted drug delivery system by using alginate: in vitro and in vivo evaluation. *Drug Dev Ind*
25 *Pharm* **2011**, *37* (11), 1347-56.

26 243. Kimura, M.; Maeshima, T.; Kubota, T.; Kurihara, H.; Masuda, Y.; Nomura, Y., Absorption of
27 Orally Administered Hyaluronan. *J Med Food* **2016**, *19* (12), 1172-1179.

28 244. García-González, C. A.; Jin, M.; Gerth, J.; Alvarez-Lorenzo, C.; Smirnova, I., Polysaccharide-
29 based aerogel microspheres for oral drug delivery. *Carbohydr Polym* **2015**, *117*, 797-806.

30 245. Kotla, N. G.; Burke, O.; Pandit, A.; Rochev, Y., An Orally Administered Hyaluronan
31 Functionalized Polymeric Hybrid Nanoparticle System for Colon-Specific Drug Delivery.
32 *Nanomaterials (Basel)* **2019**, *9* (9).

33 246. Bowman, K.; Leong, K. W., Chitosan nanoparticles for oral drug and gene delivery. *Int J*
34 *Nanomedicine* **2006**, *1* (2), 117-28.

35 247. Chalasani, K. B.; Russell-Jones, G. J.; Jain, A. K.; Diwan, P. V.; Jain, S. K., Effective oral
36 delivery of insulin in animal models using vitamin B12-coated dextran nanoparticles. *J Control*
37 *Release* **2007**, *122* (2), 141-50.

38 248. Debele, T. A.; Mekuria, S. L.; Tsai, H. C., Polysaccharide based nanogels in the drug delivery
39 system: Application as the carrier of pharmaceutical agents. *Mater Sci Eng C Mater Biol Appl* **2016**,
40 *68*, 964-981.

41 249. Lima, I. B.; Moreno, L. C. G.; Silva-Filho, E. C.; Irache, J. M.; Veiga, F. J.; Rolim, H. M.; Nunes,
42 L. C., Development of nanostructured systems using natural polymers to optimize the treatment of
43 inflammatory bowel diseases: A prospective study. *Journal of Drug Delivery Science and Technology*
44 **2021**, 102590.

45 250. Mutalik, S.; Suthar, N. A.; Managuli, R. S.; Shetty, P. K.; Avadhani, K.; Kalthur, G.; Kulkarni,
46 R. V.; Thomas, R., Development and performance evaluation of novel nanoparticles of a grafted
47 copolymer loaded with curcumin. *International journal of biological macromolecules* **2016**, *86*, 709-
48 720.

49 251. Mudgil, D.; Barak, S.; Khatkar, B. S., Guar gum: processing, properties and food
50 applications—a review. *Journal of food science and technology* **2014**, *51* (3), 409-418.

51 252. Kou, S.; Peters, L.; Mucalo, M., Chitosan: A review of sources and preparation methods.
52 *International Journal of Biological Macromolecules* **2020**.

- 1 253. Niederhofer, A.; Müller, B. W., A method for direct preparation of chitosan with low
2 molecular weight from fungi. *European Journal of Pharmaceutics and Biopharmaceutics* **2004**, *57* (1),
3 101-105.
- 4 254. Hasnain, M. S.; Nayak, A. K., Chitosan as mucoadhesive polymer in drug delivery. In *Chitosan*
5 *in Drug Delivery*, Elsevier: 2022; pp 225-246.
- 6 255. Dodane, V.; Khan, M. A.; Merwin, J. R., Effect of chitosan on epithelial permeability and
7 structure. *International journal of pharmaceutics* **1999**, *182* (1), 21-32.
- 8 256. Lang, X.; Wang, T.; Sun, M.; Chen, X.; Liu, Y., Advances and applications of chitosan-based
9 nanomaterials as oral delivery carriers: A review. *International journal of biological macromolecules*
10 **2020**, *154*, 433-445.
- 11 257. Markam, R.; Bajpai, A., Functionalization of ginger derived nanoparticles with chitosan to
12 design drug delivery system for controlled release of 5-amino salicylic acid (5-ASA) in treatment of
13 inflammatory bowel diseases: An in vitro study. *Reactive and Functional Polymers* **2020**, *149*, 104520.
- 14 258. Ahmed, O.; Abdel-Halim, M.; Farid, A.; Elamir, A., Taurine loaded chitosan-pectin
15 nanoparticle shows curative effect against acetic acid-induced colitis in rats. *Chemico-Biological*
16 *Interactions* **2021**, 109715.
- 17 259. Soni, J. M.; Sardoiwala, M. N.; Choudhury, S. R.; Sharma, S. S.; Karmakar, S., Melatonin-
18 loaded chitosan nanoparticles endows nitric oxide synthase 2 mediated anti-inflammatory activity in
19 inflammatory bowel disease model. *Materials Science and Engineering: C* **2021**, *124*, 112038.
- 20 260. Gamboa, A.; Araujo, V.; Caro, N.; Gotteland, M.; Abugoch, L.; Tapia, C., Spray freeze-drying
21 as an alternative to the ionic gelation method to produce chitosan and alginate nano-particles
22 targeted to the colon. *Journal of pharmaceutical sciences* **2015**, *104* (12), 4373-4385.
- 23 261. Laroui, H.; Viennois, E.; Xiao, B.; Canup, B. S.; Geem, D.; Denning, T. L.; Merlin, D., Fab'-
24 bearing siRNA TNF α -loaded nanoparticles targeted to colonic macrophages offer an effective therapy
25 for experimental colitis. *Journal of Controlled Release* **2014**, *186*, 41-53.
- 26 262. Rivera, M. C.; Pinheiro, A. C.; Bourbon, A. I.; Cerqueira, M. A.; Vicente, A. A., Hollow
27 chitosan/alginate nanocapsules for bioactive compound delivery. *International journal of biological*
28 *macromolecules* **2015**, *79*, 95-102.
- 29 263. Xiao, B.; Laroui, H.; Viennois, E.; Ayyadurai, S.; Charania, M. A.; Zhang, Y.; Zhang, Z.;
30 Baker, M. T.; Zhang, B.; Gewirtz, A. T., Nanoparticles with surface antibody against CD98 and carrying
31 CD98 small interfering RNA reduce colitis in mice. *Gastroenterology* **2014**, *146* (5), 1289-1300. e19.
- 32 264. Zhang, M.; Xu, C.; Liu, D.; Han, M. K.; Wang, L.; Merlin, D., Oral delivery of nanoparticles
33 loaded with ginger active compound, 6-shogaol, attenuates ulcerative colitis and promotes wound
34 healing in a murine model of ulcerative colitis. *Journal of Crohn's and Colitis* **2018**, *12* (2), 217-229.
- 35 265. Boni, F. I.; Almeida, A.; Lechanteur, A.; Sarmiento, B.; Cury, B. S. F.; Gremião, M. P. D.,
36 Mucoadhesive nanostructured polyelectrolytes complexes modulate the intestinal permeability of
37 methotrexate. *European Journal of Pharmaceutical Sciences* **2018**, *111*, 73-82.
- 38 266. Xiao, B.; Zhang, Z.; Viennois, E.; Kang, Y.; Zhang, M.; Han, M. K.; Chen, J.; Merlin, D.,
39 Combination therapy for ulcerative colitis: orally targeted nanoparticles prevent mucosal damage
40 and relieve inflammation. *Theranostics* **2016**, *6* (12), 2250.
- 41 267. Lee, M.-C.; Huang, Y.-C., Soluble eggshell membrane protein-loaded chitosan/fucoidan
42 nanoparticles for treatment of defective intestinal epithelial cells. *International journal of biological*
43 *macromolecules* **2019**, *131*, 949-958.
- 44 268. Wu, S.-J.; Don, T.-M.; Lin, C.-W.; Mi, F.-L., Delivery of berberine using chitosan/fucoidan-
45 taurine conjugate nanoparticles for treatment of defective intestinal epithelial tight junction barrier.
46 *Marine drugs* **2014**, *12* (11), 5677-5697.
- 47 269. Saboktakin, M. R.; Tabatabaie, R. M.; Maharramov, A.; Ramazanov, M. A., Synthesis and in
48 vitro evaluation of carboxymethyl starch-chitosan nanoparticles as drug delivery system to the
49 colon. *International journal of biological macromolecules* **2011**, *48* (3), 381-385.
- 50 270. Oshi, M. A.; Naeem, M.; Bae, J.; Kim, J.; Lee, J.; Hasan, N.; Kim, W.; Im, E.; Jung, Y.; Yoo,
51 J.-W., Colon-targeted dexamethasone microcrystals with pH-sensitive chitosan/alginate/Eudragit S

1 multilayers for the treatment of inflammatory bowel disease. *Carbohydrate polymers* **2018**, *198*, 434-
2 442.

3 271. Cheng, W.; Tang, C.; Yin, C., Effects of particle size and binding affinity for small interfering
4 RNA on the cellular processing, intestinal permeation and anti-inflammatory efficacy of polymeric
5 nanoparticles. *The journal of gene medicine* **2015**, *17* (10-12), 244-256.

6 272. Huo, X.; Liu, K., Renal organic anion transporters in drug–drug interactions and diseases.
7 *European Journal of Pharmaceutical Sciences* **2018**, *112*, 8-19.

8 273. Takahama, R.; Kato, H.; Tajima, K.; Tagawa, S.; Kondo, T., Biofabrication of a
9 Hyaluronan/Bacterial Cellulose Composite Nanofibril by Secretion from Engineered
10 *Gluconacetobacter*. *Biomacromolecules* **2021**.

11 274. Krolkoski, M.; Monslow, J.; Puré, E., The CD44-HA axis and inflammation in atherosclerosis:
12 a temporal perspective. *Matrix Biology* **2019**, *78*, 201-218.

13 275. Takahashi, R.; Kubota, K.; Kawada, M.; Okamoto, A., Effect of molecular weight distribution
14 on the solution properties of sodium hyaluronate in 0.2 M NaCl solution. *Biopolymers: Original*
15 *Research on Biomolecules* **1999**, *50* (1), 87-98.

16 276. Mizrahy, S.; Raz, S. R.; Hasgaard, M.; Liu, H.; Soffer-Tsur, N.; Cohen, K.; Dvash, R.;
17 Landsman-Milo, D.; Bremer, M. G.; Moghimi, S. M., Hyaluronan-coated nanoparticles: the influence
18 of the molecular weight on CD44-hyaluronan interactions and on the immune response. *Journal of*
19 *controlled release* **2011**, *156* (2), 231-238.

20 277. Tavianatou, A. G.; Caon, I.; Franchi, M.; Piperigkou, Z.; Galesso, D.; Karamanos, N. K.,
21 Hyaluronan: molecular size-dependent signaling and biological functions in inflammation and cancer.
22 *The FEBS journal* **2019**, *286* (15), 2883-2908.

23 278. Lee, Y.; Sugihara, K.; Gilliland, M. G.; Jon, S.; Kamada, N.; Moon, J. J., Hyaluronic acid–
24 bilirubin nanomedicine for targeted modulation of dysregulated intestinal barrier, microbiome and
25 immune responses in colitis. *Nature materials* **2020**, *19* (1), 118-126.

26 279. Xiao, B.; Xu, Z.; Viennois, E.; Zhang, Y.; Zhang, Z.; Zhang, M.; Han, M. K.; Kang, Y.; Merlin,
27 D., Orally targeted delivery of tripeptide KPV via hyaluronic acid-functionalized nanoparticles
28 efficiently alleviates ulcerative colitis. *Molecular Therapy* **2017**, *25* (7), 1628-1640.

29 280. Li, W.; Li, Y.; Liu, Z.; Kerdsakundee, N.; Zhang, M.; Zhang, F.; Liu, X.; Bauleth-Ramos, T.;
30 Lian, W.; Mäkilä, E., Hierarchical structured and programmed vehicles deliver drugs locally to
31 inflamed sites of intestine. *Biomaterials* **2018**, *185*, 322-332.

32 281. Vafaei, S. Y.; Esmaeili, M.; Amini, M.; Atyabi, F.; Ostad, S. N.; Dinarvand, R., Self assembled
33 hyaluronic acid nanoparticles as a potential carrier for targeting the inflamed intestinal mucosa.
34 *Carbohydrate polymers* **2016**, *144*, 371-381.

35 282. Diaz-Salmeron, R.; Michel, J.-P.; Hadji, H.; Gout, E.; Vivès, R. R.; Ponchel, G.; Bouchemal, K.,
36 Role of the interactions with CD44 and supported bilayer membranes in the cellular uptake of soft
37 multivalent hyaluronan nanoparticles. *Colloids and Surfaces B: Biointerfaces* **2021**, 111916.

38 283. Cani, P. D.; de Vos, W. M., Next-generation beneficial microbes: the case of *Akkermansia*
39 *muciniphila*. *Frontiers in microbiology* **2017**, *8*, 1765.

40 284. Mualikrishna, G.; Tharanathan, R. N., Characterization of pectic polysaccharides from pulse
41 husks. *Food chemistry* **1994**, *50* (1), 87-89.

42 285. Thakur, B. R.; Singh, R. K.; Handa, A. K.; Rao, M., Chemistry and uses of pectin—a review.
43 *Critical Reviews in Food Science & Nutrition* **1997**, *37* (1), 47-73.

44 286. Popov, S. V.; Markov, P. A.; Popova, G. Y.; Nikitina, I. R.; Efimova, L.; Ovodov, Y. S., Anti-
45 inflammatory activity of low and high methoxylated citrus pectins. *Biomedicine & Preventive*
46 *Nutrition* **2013**, *3* (1), 59-63.

47 287. Liu, L.; Fishman, M. L.; Kost, J.; Hicks, K. B., Pectin-based systems for colon-specific drug
48 delivery via oral route. *Biomaterials* **2003**, *24* (19), 3333-3343.

49 288. Thirawong, N.; Nunthanid, J.; Puttipatkhachorn, S.; Sriamornsak, P., Mucoadhesive
50 properties of various pectins on gastrointestinal mucosa: an in vitro evaluation using texture
51 analyzer. *European journal of Pharmaceutics and Biopharmaceutics* **2007**, *67* (1), 132-140.

- 1 289. Yener, S.; Akbulut, K. G.; Karakuş, R.; Erdoğan, D.; Acartürk, F., Development of melatonin
2 loaded pectin nanoparticles for the treatment of inflammatory bowel disease: In vitro and in vivo
3 studies. *Journal of Drug Delivery Science and Technology* **2021**, 102861.
- 4 290. Luo, R.; Lin, M.; Fu, C.; Zhang, J.; Chen, Q.; Zhang, C.; Shi, J.; Pu, X.; Dong, L.; Xu, H.,
5 Calcium pectinate and hyaluronic acid modified lactoferrin nanoparticles loaded rhein with dual-
6 targeting for ulcerative colitis treatment. *Carbohydrate Polymers* **2021**, 263, 117998.
- 7 291. Mensink, M. A.; Frijlink, H. W.; van der Voort Maarschalk, K.; Hinrichs, W. L., Inulin, a flexible
8 oligosaccharide I: Review of its physicochemical characteristics. *Carbohydrate polymers* **2015**, 130,
9 405-419.
- 10 292. Tremaroli, V.; Bäckhed, F., Functional interactions between the gut microbiota and host
11 metabolism. *Nature* **2012**, 489 (7415), 242-249.
- 12 293. Liu, Z.; Liu, F.; Wang, W.; Sun, C.; Gao, D.; Ma, J.; Hussain, M. A.; Xu, C.; Jiang, Z.; Hou, J.,
13 Study of the alleviation effects of a combination of Lactobacillus rhamnosus and inulin on mice with
14 colitis. *Food & function* **2020**, 11 (5), 3823-3837.
- 15 294. Valcheva, R.; Koleva, P.; Martínez, I.; Walter, J.; Gänzle, M. G.; Dieleman, L. A., Inulin-type
16 fructans improve active ulcerative colitis associated with microbiota changes and increased short-
17 chain fatty acids levels. *Gut Microbes* **2019**, 10 (3), 334-357.
- 18 295. McKellar, R.; Modler, H., Metabolism of fructo-oligosaccharides by Bifidobacterium spp.
19 *Applied microbiology and biotechnology* **1989**, 31 (5), 537-541.
- 20 296. Roberfroid, M. B., Introducing inulin-type fructans. *British journal of nutrition* **2005**, 93 (S1),
21 S13-S25.
- 22 297. Naskar, B.; Dan, A.; Ghosh, S.; Moulik, S. P., Viscosity and solubility behavior of the
23 polysaccharide inulin in water, water+ dimethyl sulfoxide, and water+ isopropanol media. *Journal of*
24 *Chemical & Engineering Data* **2010**, 55 (7), 2424-2427.
- 25 298. Sun, Q.; Luan, L.; Arif, M.; Li, J.; Dong, Q.-J.; Gao, Y.; Chi, Z.; Liu, C.-G., Redox-sensitive
26 nanoparticles based on 4-aminothiophenol-carboxymethyl inulin conjugate for budesonide delivery
27 in inflammatory bowel diseases. *Carbohydrate polymers* **2018**, 189, 352-359.
- 28 299. Shivhare, K.; Garg, C.; Priyam, A.; Gupta, A.; Sharma, A. K.; Kumar, P., Enzyme sensitive
29 smart inulin-dehydropeptide conjugate self-assembles into nanostructures useful for targeted
30 delivery of ornidazole. *International journal of biological macromolecules* **2018**, 106, 775-783.
- 31 300. Rutgeerts, P.; Van Assche, G.; Vermeire, S.; D'Haens, G.; Baert, F.; Noman, M.; Aerden, I.;
32 De Hertogh, G.; Geboes, K.; Hiele, M., Ornidazole for prophylaxis of postoperative Crohn's disease
33 recurrence: a randomized, double-blind, placebo-controlled trial. *Gastroenterology* **2005**, 128 (4),
34 856-861.
- 35 301. Laird, M. E.; Tong, L. X.; Sicco, K. I. L.; Kim, R. H.; Meehan, S. A.; Franks, A. G., Novel use of
36 apremilast for adjunctive treatment of recalcitrant pyoderma gangrenosum. *JAAD case reports* **2017**,
37 3 (3), 228-229.
- 38 302. Teli, M. D.; Sheikh, J., Extraction of chitosan from shrimp shells waste and application in
39 antibacterial finishing of bamboo rayon. *Int J Biol Macromol* **2012**, 50 (5), 1195-200.
- 40 303. Puvvada, Y. S.; Vankayalapati, S.; Sukhvasi, S., Extraction of chitin from chitosan from
41 exoskeleton of shrimp for application in the pharmaceutical industry. *International Current*
42 *Pharmaceutical Journal* **2012**, 1 (9), 258-263.
- 43 304. Pochanavanich, P.; Suntornsuk, W., Fungal chitosan production and its characterization. *Lett*
44 *Appl Microbiol* **2002**, 35 (1), 17-21.
- 45 305. Nwe, N.; Stevens, W. F., Production of fungal chitosan by solid substrate fermentation
46 followed by enzymatic extraction. *Biotechnology letters* **2002**, 24 (2), 131-134.
- 47 306. Chen, S.-q.; Song, Y.-q.; Wang, C.; Tao, S.; Yu, F.-y.; Lou, H.-y.; Hu, F.-q.; Yuan, H., Chitosan-
48 modified lipid nanodrug delivery system for the targeted and responsive treatment of ulcerative
49 colitis. *Carbohydrate polymers* **2020**, 230, 115613.
- 50 307. Tang, D.; Wang, Y.; Kang, W.; Zhou, J.; Dong, R.; Feng, Q., Chitosan attenuates obesity by
51 modifying the intestinal microbiota and increasing serum leptin levels in mice. *Journal of Functional*
52 *Foods* **2020**, 64, 103659.

- 1 308. Wang, J.; Zhang, C.; Guo, C.; Li, X., Chitosan ameliorates DSS-induced ulcerative colitis mice
2 by enhancing intestinal barrier function and improving microflora. *International journal of molecular*
3 *sciences* **2019**, *20* (22), 5751.
- 4 309. Lan, R.; Chang, Q.; Wei, L.; Zhao, Z., The Protect Effects of Chitosan Oligosaccharides on
5 Intestinal Integrity by Regulating Oxidative Status and Inflammation under Oxidative Stress. *Marine*
6 *Drugs* **2021**, *19* (2), 57.
- 7 310. Riehl, T. E.; Ee, X.; Stenson, W. F., Hyaluronic acid regulates normal intestinal and colonic
8 growth in mice. *Am J Physiol Gastrointest Liver Physiol* **2012**, *303* (3), G377-88.
- 9 311. Wittig, B. M.; Stallmach, A.; Zeitz, M.; Günthert, U., Functional involvement of CD44 variant
10 7 in gut immune response. *Pathobiology* **2002**, *70* (3), 184-9.
- 11 312. Johnson, P.; Ruffell, B., CD44 and its role in inflammation and inflammatory diseases.
12 *Inflamm Allergy Drug Targets* **2009**, *8* (3), 208-20.
- 13 313. Wittig, B. M.; Johansson, B.; Zöller, M.; Schwärzler, C.; Günthert, U., Abrogation of
14 experimental colitis correlates with increased apoptosis in mice deficient for CD44 variant exon 7
15 (CD44v7). *J Exp Med* **2000**, *191* (12), 2053-64.
- 16 314. Chen, H.; Mahaseth, M.; Zhang, Y., Hyaluronic acid as a rescue therapy for trinitrobenzene
17 sulfonic acid-induced colitis through Cox-2 and PGE2 in a Toll-like receptor 4-dependent way. *J*
18 *Zhejiang Univ Sci B* **2011**, *12* (9), 712-9.
- 19 315. Janeway, C. A., Jr.; Medzhitov, R., Lipoproteins take their toll on the host. *Curr Biol* **1999**, *9*
20 (23), R879-82.
- 21 316. Ishisono, K.; Mano, T.; Yabe, T.; Kitaguchi, K., Dietary Fiber Pectin Ameliorates Experimental
22 Colitis in a Neutral Sugar Side Chain-Dependent Manner. *Front Immunol* **2019**, *10*, 2979.
- 23 317. Miao, X. P.; Sun, X. N.; Cui, L. J.; Cao, Q. F.; Zhuang, G. F.; Deng, T. Z.; Zhang, D. Y.,
24 Suppressive effect of pectic polysaccharides extracted from *Rauwolfia verticillata* (Lour.)
25 *Baill.var.hainanensis* Tsiang on inflammation by regulation of NF- κ B pathway and interleukin-17 in
26 mice with dextran sulphatesodium-induced ulcerative colitis. *Asian Pac J Trop Med* **2015**, *8* (2), 147-
27 52.
- 28 318. Hendry, G., The ecological significance of fructan in a contemporary flora. *New Phytologist*
29 **1987**, *106*, 201-216.
- 30 319. Hino, S.; Ito, H.; Bito, H.; Kawagishi, H.; Morita, T., Ameliorating effects of short-chain inulin-
31 like fructans on the healing stage of trinitrobenzene sulfonic acid-induced colitis in rats. *Bioscience,*
32 *biotechnology, and biochemistry* **2011**, *75* (11), 2169-2174.
- 33 320. McConnell, E. L.; Basit, A. W.; Murdan, S., Measurements of rat and mouse gastrointestinal
34 pH, fluid and lymphoid tissue, and implications for in-vivo experiments. *Journal of Pharmacy and*
35 *Pharmacology* **2008**, *60* (1), 63-70.
- 36 321. Gustafsson, J. K.; Ermund, A.; Johansson, M. E.; Schütte, A.; Hansson, G. C.; Sjövall, H., An
37 ex vivo method for studying mucus formation, properties, and thickness in human colonic biopsies
38 and mouse small and large intestinal explants. *American Journal of Physiology-Gastrointestinal and*
39 *Liver Physiology* **2012**, *302* (4), G430-G438.
- 40 322. Hugenholtz, F.; de Vos, W. M., Mouse models for human intestinal microbiota research: a
41 critical evaluation. *Cellular and Molecular Life Sciences* **2018**, *75* (1), 149-160.
- 42 323. Cuthbertson, P.; Adhikary, S. R.; Geraghty, N. J.; Guy, T. V.; Hadjiashrafi, A.; Fuller, S. J.; Ly,
43 D.; Watson, D.; Sluyter, R., Increased P2X7 expression in the gastrointestinal tract and skin in a
44 humanised mouse model of graft-versus-host disease. *Clinical Science* **2020**, *134* (2), 207-223.
- 45 324. Park, J. C.; Im, S.-H., Of men in mice: the development and application of a humanized
46 gnotobiotic mouse model for microbiome therapeutics. *Experimental & molecular medicine* **2020**, *52*
47 (9), 1383-1396.
- 48 325. Jain, K.; Sinha, P.; Mohan, K. V.; Upadhyay, P. K., Mouse Models of the Humanized Immune
49 System. In *Essentials of Laboratory Animal Science: Principles and Practices*, Springer: 2021; pp 725-
50 742.

- 1 326. Yong, Y.; Li, J.; Gong, D.; Yu, T.; Wu, L.; Hu, C.; Liu, X.; Yu, Z.; Ma, X.; Gooneratne, R.,
2 ERK1/2 mitogen-activated protein kinase mediates downregulation of intestinal tight junction
3 proteins in heat stress-induced IBD model in pig. *Journal of Thermal Biology* **2021**, *101*, 103103.
- 4 327. Langelueddecke, C.; Roussa, E.; Fenton, R. A.; Thévenod, F., Expression and function of the
5 lipocalin-2 (24p3/NGAL) receptor in rodent and human intestinal epithelia. *PLoS One* **2013**, *8* (8),
6 e71586.
- 7 328. Dalmaso, G.; Charrier-Hisamuddin, L.; Nguyen, H.; Yan, Y.; Sitaraman, S.; Merlin, D., PepT1
8 mediated tripeptide KPV uptake reduces intestinal inflammation: P-078. *Inflammatory Bowel*
9 *Diseases* **2008**, *14*, S32.
- 10 329. Danese, S.; Semeraro, S.; Marini, M.; Roberto, I.; Armuzzi, A.; Papa, A.; Gasbarrini, A.,
11 Adhesion molecules in inflammatory bowel disease: therapeutic implications for gut inflammation.
12 *Digestive and Liver Disease* **2005**, *37* (11), 811-818.
- 13 330. Biagioli, M.; Carino, A.; Cipriani, S.; Francisci, D.; Marchianò, S.; Scarpelli, P.; Sorcini, D.;
14 Zampella, A.; Fiorucci, S., The bile acid receptor GPBAR1 regulates the M1/M2 phenotype of
15 intestinal macrophages and activation of GPBAR1 rescues mice from murine colitis. *The Journal of*
16 *Immunology* **2017**, *199* (2), 718-733.
- 17 331. Shen, C.; Zhao, L.; Du, X.; Tian, J.; Yuan, Y.; Jia, M.; He, Y.; Zeng, R.; Qiao, R.; Li, C., Smart
18 responsive quercetin-conjugated glycol chitosan prodrug micelles for treatment of inflammatory
19 bowel diseases. *Molecular Pharmaceutics* **2021**, *18* (3), 1419-1430.
- 20 332. Varshosaz, J.; Emami, J.; Fassihi, A.; Tavakoli, N.; Minaiyan, M.; Ahmadi, F.; Mahzouni, P.;
21 Dorkoosh, F., Effectiveness of budesonide-succinate-dextran conjugate as a novel prodrug of
22 budesonide against acetic acid-induced colitis in rats. *International journal of colorectal disease* **2010**,
23 *25* (10), 1159-1165.
- 24 333. Nalinbenjapun, S.; Ovatlarnporn, C., Chitosan-5-aminosalicylic acid conjugates for colon-
25 specific drug delivery: Methods of preparation and in vitro evaluations. *Journal of Drug Delivery*
26 *Science and Technology* **2020**, *57*, 101397.
- 27 334. Hartzell, A. L.; Maldonado-Gómez, M. X.; Yang, J.; Hutkins, R. W.; Rose, D. J., In vitro
28 digestion and fermentation of 5-formyl-aminosalicylate-inulin: A potential prodrug of 5-
29 aminosalicylic acid. *Bioactive Carbohydrates and Dietary Fibre* **2013**, *2* (1), 8-14.
- 30 335. Kotla, N. G.; Rana, S.; Sivaraman, G.; Sunnapu, O.; Vemula, P. K.; Pandit, A.; Rochev, Y.,
31 Bioresponsive drug delivery systems in intestinal inflammation: State-of-the-art and future
32 perspectives. *Advanced drug delivery reviews* **2019**, *146*, 248-266.
- 33 336. Leroux, J. C., Drug delivery: too much complexity, not enough reproducibility? Wiley Online
34 Library: 2017.

35

1 **11. Abbreviations**

- 2 5-ASA: 5-aminosalicylic acid
- 3 API: Active pharmaceutical ingredients
- 4 CD: Crohn disease
- 5 COX: Cyclooxygenase
- 6 CRF: Corticotrophin-releasing factor
- 7 CYA: Cyclosporine A
- 8 DA: Degree of acetylation
- 9 DDS: Drug delivery systems
- 10 eEPR: Epithelial enhanced permeability and retention
- 11 ES100: Eudragit S100
- 12 GALT: Gut-associated lymphoid tissue
- 13 GDNs: Grapefruit-derived nanovesicles
- 14 HA: Hyaluronan
- 15 HMw-HA: high molecular weight HA
- 16 IBD: Inflammatory bowel disease
- 17 Ig: Immunoglobulin
- 18 ILC: Innate lymphoid cells
- 19 KPV: Lysine-proline-valine
- 20 LMw-HA: low molecular weight HA
- 21 MAPK: Mitogen-activated protein kinase
- 22 MeO-PEG-b-PMOT: Methoxy-poly(ethylene glycol)-b-poly [4-(2,2,6,6-
- 23 tetramethylpiperidine-1-oxyl)oxymethylstyrene]
- 24 NF- κ B: nuclear factor- κ B
- 25 NK: Natural killer
- 26 NLC: Nanostructured lipid carriers

- 1 NO: Nitroxide
- 2 PCL: Polycaprolactone
- 3 PEG: Poly(ethylene glycol)
- 4 PEI: Polyethyleneimine
- 5 PLGA/PLA-PEG-FA: Poly(lactic-*co*-glycolic acid)/poly(lactic acid)-polyethylene glycol-folic
6 acid
- 7 ROS: Reactive oxygen species
- 8 SCFAs: Short-chain fatty acids
- 9 Silicon dioxide: SiO₂
- 10 TLR: Toll-like receptors
- 11 TNBS: Trinitrobenzosulfonic acid
- 12 TPP: Tripolyphosphate
- 13 UC: Ulcerative colitis