

Age-specific fecundity under parasitic threat in an insect: terminal investment versus reproductive restraint

Charly Jehan, Camille Sabarly, Thierry Rigaud, Yannick Moret

▶ To cite this version:

Charly Jehan, Camille Sabarly, Thierry Rigaud, Yannick Moret. Age-specific fecundity under parasitic threat in an insect: terminal investment versus reproductive restraint. Journal of Animal Ecology, 2022, 91 (1), pp.101-111. 10.1111/1365-2656.13604 . hal-03520427

HAL Id: hal-03520427 https://hal.science/hal-03520427

Submitted on 11 Jan2022

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

Age-specific fecundity under parasitic threat in an insect: terminal investment *versus* reproductive restraint

Charly Jehan, Camille Sabarly, Thierry Rigaud, Yannick Moret*

UMR CNRS 6282 BioGéoSciences, Équipe Écologie Évolutive, Université Bourgogne-Franche Comté, Dijon, France

* Corresponding author: yannick.moret@u-bourgogne.fr

1 Abstract

The terminal investment hypothesis predicts that animals increase their reproductive effort
 as they age, and their chances for survival and reproduction decline. An infection predicts
 further diminished residual reproductive value, and could therefore trigger increased
 breeding effort at the expense of somatic defences and survival. Attempts to test this
 hypothesis have produced mixed results, which also often relied on potential changes in the
 breeding effort only. Alternatively, animals may restrain their current reproduction to
 sustain immunity and so later reproduction.

9 2. We tested these possibilities in females of the yellow mealworm beetle, *Tenebrio molitor*,
10 an iteroparous insect with reproductive tactics similar to that of long-lived organisms.

3. For this purpose we mimicked pathogenic bacterial infections early or late in life in
breeding females by injecting them with a suspension of inactivated *Bacillus cereus*, a
known natural pathogen of *T. molitor*, and measured female age-specific fecundity,
survival and immunity.

4. Inconsistent with a terminal investment, early or late-life immune challenges of females did
not reduce their survival or enhance their reproductive output. Female fecundity declined
with age and was reduced by the early but not the late immune challenge. Both early and
late-life fecundity correlated positively with life expectancy. Finally, young and old
females exhibited similar antibacterial immune responses, suggesting that they both
restrained reproduction to sustain immunity.

5. Our results clearly demonstrate that age-specific reproduction of *T. molitor* females under parasitic threat is discrepant with a terminal investment. By contrast, our results rather suggest that females used a reproductive restraint strategy to sustain immunity and therefore subsequent reproductive opportunities. However, as infections were mimicked only, the fitness benefit of this reproductive restraint could not be shown.

- 26 Keywords: Cost of innate immunity, life history shift, reproductive restraint, senescence,
- 27 terminal investment.

28 Introduction

29 Trade-offs are of paramount importance in life history evolution (Roff, 1993; Stearns, 1992). 30 Because of resource and time scarcity, organisms cannot simultaneously maximize all life 31 history traits and natural selection should favour optimal combinations of those to maximize 32 fitness. In iteroparous species, optimal investment between current and future reproduction 33 likely depends on the relative expected lifespan (Williams, 1966). According to the terminal 34 investment hypothesis (Clutton-Brock, 1984), individuals increase their current reproductive 35 effort as their residual reproductive value (i.e., opportunity for future reproduction) decreases. 36 Such a terminal investment is expressed at the expense of longevity, which would no longer 37 provide fitness benefit (Clutton-Brock, 1984).

38 Terminal investment may occur as a response to a survival threat such as a parasite or pathogen infection (Agnew et al., 2000; Duffield et al., 2017; Forbes, 1993; Hochberg et al., 39 40 1992; Minchella, 1985; Perrin et al., 1996). Parasite and pathogen typically reduce host 41 survival or fecundity (Hamilton & Zuk, 1982) and infection might be a relevant cue of a 42 rapidly diminishing life span (Grenfell & Dobson, 1995). There is ample empirical evidence 43 that parasitized host can make adaptive adjustments of their reproductive effort towards a 44 terminal reproductive investment to reduce the fitness cost of parasitism (Adamo, 1999; Blair 45 & Webster, 2007; Chadwick & Little, 2005; Giehr et al., 2017; Minchella & Loverde, 1981; 46 Schwanz, 2008). Similar outcomes were found from immune stimulations with non-living immune elicitors (Adamo, 1999; Bonneaud et al., 2004; Duffield et al., 2017; Moret & 47 Schmid-Hempel, 2000). Indeed, the immune system can act as a signalling mechanism 48 49 alerting the host about a virulent infection threat (Adamo, 1999; Bonneaud et al., 2004; 50 Moret, 2003), or about a higher risk of subsequent infection by the same parasite or pathogen 51 (Lui, 1998). While most of the studies that provided evidence of a terminal investment did so 52 by revealing increased allocation to current reproduction in response to a parasitic threat 53 (Duffield et al., 2017), they often overlooked two other key predictions. First, the resulting 54 increased allocation to current reproduction should be negatively correlated to post-55 reproduction survival (Jehan et al., 2021; Tarwater & Arcese, 2017). Second, the increased 56 reproductive effort should be accompanied by a reduced investment into somatic defences 57 such as immune defence (Duffield et al., 2017; Jehan et al., 2021), because of trade-offs 58 between reproduction and immunity (Norris & Evans, 2000; Sheldon & Verhulst, 1996).

59 As an alternative to terminal reproductive investment, immune-challenged hosts may reduce 60 investment into current reproduction and sustain immune defence to maintain subsequent 61 reproductive opportunities, a strategy in accordance with the reproductive restraint hypothesis (Curio, 1983; McNamara et al., 2009). Such a strategy involves that organisms may develop a 62 63 long-term immunity to the parasite after recovery, thus deferring death and maintaining later 64 life reproduction. Long-term immunity shall be achieved through the acquired immune 65 system of vertebrates or the immune priming response of invertebrates (Coustau et al., 2016; 66 McDade et al., 2016; Milutinović & Kurtz, 2016).

67 Here, we tested for these two alternative hypotheses by mimicking bacterial infections 68 in females of the mealworm beetle, Tenebrio molitor. T. molitor is a cosmopolitan pest of 69 stored grain product that is currently raising considerable interests for its use as an alternative 70 protein source for food and feed (Vigneron et al., 2019). Females begin to reproduce from 71 their fifth day post-eclosion, and can mate many times with several males within weeks of 72 their adult life. T. molitor is iteroparous and is an income breeder insect (Zanchi et al., 2019): 73 it reproduces using resources gained as an adult, as opposed to a capital breeder insect, for 74 which reproduction depends on resources cumulated during the juvenile stages (Jönsson, 1997; Stearns, 1992). T. molitor feeds at both the larval and the adult stages and females lay 75 76 eggs in several reproductive bouts along its lifespan (Dick, 1937; Hughes, 2017). These 77 features possibly allow females of this insect to adjust their lifetime reproductive effort

78 according to variation in their physiological state and environmental conditions. T. molitor is 79 a relatively long-lived insect species that exhibits reproductive tactics similar to that of long-80 lived mammalians (Jehan et al., 2021), for which the cost of current reproduction may 81 compromise future reproduction without jeopardizing survival (Hamel et al., 2010). Previous 82 work found that ageing T. molitor females tended to restrain reproduction late in life in favour 83 of longevity (Jehan et al., 2021). However, in this study, females were not exposed to cues 84 mimicking a survival threat like would do a pathogenic challenge. It is therefore not known 85 whether T. molitor females, exposed to a parasitic threat, are maintaining a reproductive restraint strategy or are switching toward a terminal investment. 86

87 In the present study, individual T. molitor females were immune challenged with the 88 inactivated bacterium, Bacillus cereus, an important natural and virulent pathogen of T. 89 molitor (Du & Laing, 2011; Jurat-Fuentes & Jackson, 2012). This challenge was made either 90 early or late in life to examine the effect of a pathogenic threat at different ages on fecundity, 91 post-reproduction survival and immunity. Indeed, since young females (i.e., below c.a. 15 92 days post-eclosion – Jehan et al., 2020) still have a high residual reproductive value, we 93 expect that an immune challenge occurring early in life should not trigger a reproductive shift 94 toward a terminal investment (Duffield et al., 2017). As the immune response is costly, we 95 even predict that early challenged females should exhibit reduced fecundity (Moreau et al., 96 2012). By contrast, as the residual reproductive value of old females becomes low, exposure 97 to an immune challenge is expected to trigger a life history shift resulting in an increased 98 reproductive investment in line with the terminal investment hypothesis (Duffield et al., 99 2017). Such an increased reproductive investment late in life should be accompanied by both 100 a reduced immunity and a reduced survival. If these assumptions are not fulfilled, then T. 101 *molitor* females may alternatively use the reproductive restraint strategy. The reproductive 102 restraint strategy shall be supported if the following results are found. First, late-life reproductive output should not increase in response to the late-life immune challenge. Second,
late-life reproductive output should not correlate negatively with late-life survival (Jehan et
al., 2021; Tarwater & Arcese, 2017). Finally, immune investment should be maintained late in
life (Duffield et al., 2017).

107

108 Material and Methods

109 INSECT CULTURE AND EXPERIMENTAL DESIGN

Virgin adult beetles of controlled age (10 ± 2 days post-eclosion) were obtained from pupae, randomly sampled from a stock culture and maintained in standard laboratory conditions (see below). Prior to the experiments, all the experimental insects were maintained separately, at $24 \pm 1^{\circ}$ C, 70% RH in permanent darkness, supplied with *ad libitum* bran flour and water, and supplemented once a week with apple.

This study involved two separate experiments because it was not possible to assess the influence of age-specific immune challenges on reproduction, longevity and immunity on the same individuals. For this reason, a first experiment investigated the influence of age-specific immune challenges on fecundity and longevity, and a second one examined the influence of age-specific immune challenges on induced antibacterial activity.

120

121 EXPERIMENT 1: AGE-SPECIFIC IMMUNE CHALLENGE, FECUNDITY AND122 LONGEVITY

Here, we assessed the age-specific reproductive performance of *T. molitor* females exposed to early or late-life immune challenges, mimicking reduced expected life span, while tacking into account for variation in individual quality. To this purpose, sixty 10-day-old virgin adult females were individually immune challenged with a single injection of a 5µL suspension of

the inactivated bacterium, Bacillus cereus (108 bacteria.mL-1 in Phosphate Saline Buffer: PBS 127 128 10 mM, pH 7.4). A second group of 60 females received a 5-uL injection of sterile PBS only 129 as a procedural control (see below for inactivation procedure). Immediately after their 130 respective immune treatment, females were allowed to reproduce for a 20-days period (from 131 day 10 to day 30 post-eclosion) to assess their early-life reproductive performance. To this 132 end, each female was paired with a 10 ± 2 -day old virgin male for 10 days. Then, the male 133 was replaced by a new 10 ± 2 -day old virgin male, with which the female spent 10 additional 134 days. Couples were maintained in a Petri dish containing bleached flour and provided ad libitum food and water under standard laboratory conditions (see above). Female reproductive 135 136 output was estimated by counting the number of eggs produced by the female every 3 or 4 137 days along the 20-day episode of reproduction. For this purpose, couples were transferred into 138 a new Petri dish and eggs were searched by sieving the flour (600 µm) and counted (Moreau 139 et al., 2012). At the end of this early reproductive episode, each female was kept alone in a 140 new Petri dish with ad libitum food and water for 10 days (until day 40 post-eclosion) 141 awaiting their assessment for their late-life reproductive performance.

142 Prior to the assessment of their late-life reproductive performance, half of the early 143 challenged and control females were challenged with inactivated B. cereus whereas the other 144 half was injected with sterile PBS, as described above. Thus, the late-life reproductive 145 performance was assessed on females that have been either immune challenged twice ('early 146 and late' in life), once (either 'early' or 'late' in life) or never ('control'). Late-life 147 reproductive performance of females was assessed from day 40 to day 60 post-eclosion as 148 described above for the early reproductive performance of females. At the end of their late 149 reproductive period, females were isolated into a new Petri dish with ad libitum food and 150 water, checked twice a week until death. Prior to each reproductive period and immune

151 treatment, female body mass was measured to the nearest 1 mg using an OHAUS balance152 (discovery series, DU114C).

153

154 EXPERIMENT 2: AGE-SPECIFIC IMMUNE CHALLENGE ON ANTIBACTERIAL155 ACTIVITY

156 T. molitor immunity relies on both constitutive and inducible innate mechanisms with cellular 157 and humoral components (Vigneron et al., 2019). An important aspect of T. molitor immunity 158 is immune priming, through which individuals improve their immune response as a function 159 of their immunological experience. This reduces their susceptibility to re-infection (Moret & 160 Siva-Jothy, 2003). In T. molitor, the acquired immune protection in response to a bacterial 161 immune challenge is pathogen-dependent and is mainly caused by a persistent antibacterial 162 response, which can last at least 20 days after the immune challenge (Dhinaut et al., 2018). 163 We therefore assessed the effect of age-specific challenge on immunity of T. molitor females 164 through the measurement of the antibacterial activity of their haemolymph.

165 Six groups of females were created, according to their respective age-specific immune 166 treatment and the age at which their haemolymph was sampled to assess antibacterial activity. 167 Four groups of 20 females were treated as described above (immune-challenged early, late, 168 both early and late, and not immune-challenged as control) to estimate antibacterial activity in 169 their haemolymph late in life. Females were allowed to reproduce as previously described, 170 and provided a sample of haemolymph immediately after their late reproductive episode (at 171 60 days post-eclosion). Two additional groups of 25 females were created to estimate 172 antibacterial activity early in life. Here, the sample of haemolymph was taken at 30 days post-173 eclosion (i.e., immediately after their early reproductive episode). In one group, females were 174 immune challenged early at 10 days post-eclosion, while in the second group they were not 175 immune challenged. These two additional groups were created to test the effect of the immune 176

challenge on antibacterial activity according to age when compared to the groups of old 177 females that were immune challenged late only (at 40 days post-eclosion).

178

179 **BACTERIAL CULTURES FOR IMMUNE CHALLENGES**

180 The bacterium Bacillus cereus used to perform the immune challenges was obtained from the 181 Pasteur institute (CIP69.12). The bacteria were grown overnight at 28°C in liquid Broth 182 medium (10 g/L bacto-tryptone, 5 g/L yeast extract, 10 g/L NaCl, pH 7). Bacteria were then 183 inactivated in 0.5% formaldehyde prepared in phosphate-buffered saline (PBS 10 mM, pH 184 7.4) for 30 minutes, rinsed three times in PBS, and their concentration adjusted to 10^8 bacteria 185 per mL using a Neubauer improved cell counting chamber (Dubuffet et al., 2015). The 186 success of the inactivation was tested by plating a sample of the bacterial solution on sterile 187 Broth medium with 1% of bacterial agar and incubated at 28°C for 24 hours. Aliquots were 188 kept at -20 °C until use. Immune challenges were performed by injection through the pleural 189 membrane between the second and the third abdominal tergites using sterile glass capillaries 190 that had been pulled out to a fine point with an electrode puller (Narashige PC-10). Immune 191 challenge with inactivated B. cereus using this method has proven its efficacy to efficiently 192 stimulate the immune response of *T. molitor* (Jehan et al., 2021).

193

194 HAEMOLYMPH COLLECTION AND ANTIBACTERIAL ACTIVITY

195 Haemolymph collection was done as described by (Dhinaut et al., 2018). Briefly, beetles were 196 chilled on ice for 10 minutes before a 3-µL sample of haemolymph was collected from a 197 wound made in the beetle's neck. This sample was flushed into an N-phenylthiourea-coated 198 microcentrifuge tube (P7629; Sigma-Aldrich, St Louis, MO, USA), flash-frozen in liquid 199 nitrogen and then stored at -80 °C for later examination of its antibacterial activity. 200 Antimicrobial activity of the haemolymph was measured using the inhibition zone 201 assay described by (Moret, 2006). Briefly, haemolymph samples were thawed on ice, and 2µL 202 of the sample solution was used to measure antibacterial activity on zone inhibition plates 203 seeded with Arthrobacter globiformis from the Pasteur Institute (CIP105365). An overnight 204 culture of the bacterium was added to Broth medium containing 1 % agar to achieve a final concentration of 10⁵ cells.mL⁻¹. Six millilitres of this seeded medium were subsequently 205 206 poured per Petri dish. After solidification, sample wells were made using a Pasteur pipette 207 fitted with a ball pump. Two microliters of sample were deposited in each well. Plates were 208 then incubated overnight at 28°C and the diameter of each zone of inhibition was measured.

209

210 STATISTICS

211 Female longevity was analysed as a function of their immune treatment (occurring early or 212 late in life) using a mixed effects Cox model (Therneau, 2020) including early-life immune 213 treatment (yes/no), late-life immune treatment (yes/no) and their interaction as independent 214 categorical covariates. This allowed us to test, in a full factorial design, whether the impact of 215 the late immune challenge could be dependent on the early immune challenge, which would 216 be revealed as a significant interaction term since the effect of early and late immune 217 challenge on survival might not be a priori additive. We also included female body mass 218 measured prior to each reproductive period (at 10 and 40 days old) as an independent 219 continuous covariate, which was tested as a main effect only. To account for potential 220 variation in frailty among female individuals, their identity was included as a random effect. 221 This Cox regression analysis was performed using R version 3.6.3 (R Core Team, 2020) and 222 COXME package version 2.2-16. Model reduction was limited to non-significant interaction 223 terms, whereas non-significant main effects were kept in the final model. Proportional

Page 12 of 38

assumption was checked using cox.zph() function in the R Survival package version 3.1-12(Therneau, 2020).

We investigated potential change in female body mass according to age at which they initiate each reproductive period (10 day old *versus* 40 day old, respectively) using a mixed linear model that included female age, the early immune challenge (yes/no) and their interaction term as independent categorical variables. Female identity was included as a random effect. Note that the late immune challenge was not included as an explanatory variable in the model as this treatment was applied after body mass measurements.

232 Age-specific female fecundity was analysed using a mixed linear model as a function 233 of age at which females initiated each reproductive period (10 day old versus 40 day old), 234 whether the females were exposed to an early immune challenge (yes/no), a late immune 235 challenge (yes/no) and their interaction as fixed categorical factors. Female body mass 236 measured prior to each reproductive period was included as an independent continuous 237 covariate tested as a main effect only. Based on the survival data, we calculated life 238 expectancy (in days) of each female at the beginning of each reproductive period (here 239 corresponding to the total adult lifespan minus 10 for early-life expectancy and minus 40 for 240 late-life expectancy) as an indication of the general state of each individual. We included it in 241 the model as an independent continuous covariate tested as a main effect and as two-way and 242 three-way interaction terms with the early and late immune challenges. A negative 243 relationship between fecundity and life expectancy would be indicative of a terminal 244 investment (Jehan et al., 2021; Tarwater & Arcese, 2017).

Antibacterial activity of the haemolymph was analysed as a function of immune challenge (occurring early or late in life), age and initial body mass using Generalized Linear Models (GLM) with a Tweedie distribution and a Log link function. Fecundity, body mass and antibacterial activity data were analysed using IBM® SPSS® statistics 19 for Macintosh.

Model simplification was achieved by removing higher order non-significant interactionsbased on Akaike's information criterion.

251

252 Results

253 LONGEVITY REMAINED UNALTERED BY AGE-SPECIFIC IMMUNE CHALLENGE

Whether the immune challenge occurred early or late in life, it had no effect on female longevity either as main effects (Fig. S1, Table S1, mixed effects Cox regression analysis: early challenge z = 0.61, df = 1, P = 0.540, late challenge z = 0.83, df = 1, P = 0.410) or as an interaction (effect before being removed from the last model z = 0.52, df = 1, P = 0.610), while accounting for inter-individual variation by including female identity as random effects. Female body mass measured prior to each reproductive period did not explain any variation of longevity (mass z = -1.01, df = 1, P = 0.310).

261

262 BODY MASS DECREASED WITH AGE

Female body mass significantly decreased with age (estimate of 10 day old *versus* 40 day old ± s.e. 5.66 ± 3.61 , $F_{1,187} = 6.50$ P = 0.012, Fig. 1). However, the early immune challenge did not affect female body mass either as a main effect ($F_{1,187} = 0.12$ P = 0.734) or in interaction with age ($F_{1,187} = 0.07$ P = 0.793).

267

268 REPRODUCTION DECLINED WITH AGE AND AS A RESULT OF AN IMMUNE269 CHALLENGE EARLY IN LIFE

Overall, female fecundity significantly depended on female age and female life expectancy as main effects (Table 1). On the one hand, female fecundity declined with age (Fig. 2) and, on the other hand, females that exhibited the longest life expectancy were also those that produced the largest number of eggs (Fig. 3). The early immune challenge significantly 274 affected female fecundity, but as two-way interaction terms with age and with life expectancy 275 (Table 1). The interaction term between the early immune challenge and age shows that the 276 early immune challenge induced a decline in female fecundity at early age only (Fig. 2, Fig. 277 3A, B). The significant interaction term between early challenge and life expectancy (Table 1) 278 shows that females with the longest life expectancy were those that exhibited the largest 279 decline in fecundity after an immune challenge early in life (Fig. 3A). The late immune 280 challenge had no significant effect on late-life female fecundity, whether they were previously 281 exposed to an early challenge or not (Table 1, Fig. 2). Female body mass significantly 282 explained variation in female fecundity, as the heaviest females laid more eggs than the 283 lightest ones (Table 1).

284

285 ANTIBACTERIAL IMMUNE RESPONSE

286 The challenge with inactivated B. cereus significantly induced an antibacterial immune 287 response in young (assessed at 30 days post-eclosion) and old (assessed at 60 days post-288 eclosion) females (Table 2a, Fig. 4). The analysis revealed a significant interaction between 289 female age and the immune challenge (Table 2a), which mainly results from the fact that old 290 control females exhibited a slightly higher antibacterial activity than young control ones (Fig. 291 4). This is also the reason why antibacterial activity was marginally increased with age (Table 292 2a, Fig. 4), since the level of antibacterial activity in young and old immune-challenged 293 females was similar (Fig. 4).

Among old females (assessed at 60 days post-eclosion), antibacterial activity was marginally increased by both the early and the late immune challenge (Table 2b, Fig. 4). The analysis revealed a significant interaction between the early and the late immune challenges (Table 2b). Indeed, old females that were exposed to an early immune challenge (occurring 50 days earlier) still exhibited high level of antibacterial activity in their haemolymph (Fig. 4),

299 possibly as a result of an immune priming response (Dhinaut et al., 2018).

300

301 Discussion

302 Challenging the immune system of breeding T. molitor females early or late in life did not 303 induce measurable effects on their longevity in our experimental conditions. Reduced 304 longevity was expected if the immune response imposed a substantial survival cost or if the 305 challenged females shifted toward a terminal reproductive investment. The immune function 306 is known to be costly for organisms (Moret & Schmid-Hempel, 2000) and could cause self-307 damage (named immunopathology) (Sadd & Siva-Jothy, 2006), which is expected to reduce 308 longevity (Pursall & Rolff, 2011; Urabe et al., 1994). Previous studies found that the immune 309 response causes self-damages in T. molitor (Khan et al., 2017; Sadd & Siva-Jothy, 2006) and 310 that females exposed to an early life immune challenge suffered from a reduced longevity 311 (Khan et al., 2017). Our results, contrasting with those of these previous studies, might have 312 at least two main causes in addition to many other small differences. First, Khan et al. (2017) 313 did not allow females to reproduce, whereas our experiment allowed all the females to 314 reproduce early and late in life. It was previously found that reproduction significantly 315 stimulates immune activity in T. molitor females (Jehan et al., 2021), which may have 316 buffered the effect of the immune challenge in our experiment. Second, Khan et al. (2017) 317 kept their insects at a suboptimal temperature condition (30°C) compared to the optimum 318 range of temperature of 25-28°C usually used to grow T. molitor (Punzo, 1975; Ribeiro et al., 319 2018) and that we have used in the present study. Actually, in the study by Khan et al. (2017), 320 control (non-immune challenged) females of T. molitor showed a 90th percentile survival time 321 of 83.1 days, whereas it was 112 days in our study, a difference suggesting that in their study, 322 females were maintained under stressful conditions, allowing the expression of the cost of an

arly immune activation on longevity. In general, detrimental effects of the immune response
on survival are revealed in stressful conditions (Moret & Schmid-Hempel, 2000). Rearing
conditions in our study were probably too favourable to highlight such a survival cost of the
immune response.

327 Nevertheless, despite the relatively favourable conditions of our study, the immune 328 challenge had an age-specific effect on female reproduction. Only the early-life immune 329 challenge had a measurable effect on female reproduction by reducing their early-life 330 fecundity, a result consistent with the predicted trade-off between reproduction and immunity 331 (Sheldon & Verhulst, 1996). Accordingly, early challenged females exhibited stronger level 332 of antibacterial activity in their haemolymph than control females. However, despite having 333 induced an antibacterial immune response of similar magnitude to that of the early challenge, 334 the late-life challenge had no further impact on the following female's clutch size. This 335 suggests that old females paid a lower cost to the immune response than young ones.

336 It should be noted that fecundity of old females was about half that of young ones 337 among controls. This is consistent with previous observations, showing that despite being 338 continuously sexually receptive during adulthood, T. molitor female fecundity declines after 339 about two weeks but does not necessary stop (Dick, 1937; Drnevich et al., 2001; Jehan et al., 340 2020). This might be due to senescence. The age-specific decline of female body mass, which 341 positively predicted female fecundity, may support this view. However, ageing females are 342 also expected to exhibit reduced somatic protection, such as a reduced immune activity. This 343 was clearly not the case here, although our measure of immune activity was restricted to the 344 ability to produce an antibacterial immune response. Antimicrobial activity found in early 345 challenged females is still high late in life. This suggests that old females are still protected by 346 the antibacterial activity induced 50 days earlier, which is, at least, twice as long as previously 347 observed in this insect species (Dhinaut et al., 2018; Haine et al., 2008) and is in line with other observations in dragonflies (Bulet et al., 1992). Such a relatively high level of immune activity maintained late in life is not consistent with the terminal investment hypothesis, but is rather in support of the reproductive restraint hypothesis. Indeed, it seems that reduced fecundity late in life allows females to afford the cost of a late-life immune response, which presumably helps to prolong lifespan and, consequently, future reproductive opportunities.

353 Another important prediction of the terminal investment hypothesis is that individuals 354 should increase their reproductive investment at the expense of their subsequent lifespan. A 355 negative relationship between reproduction and life expectancy from the moment at which the 356 terminal investment is initiated is therefore expected (Tarwater & Arcese, 2017). We found no 357 evidence of such a relationship either early or late in life in T. molitor females. At the 358 opposite, the relationship between female fecundity and life expectancy was always positive, 359 illustrating that the females with the lowest life expectancy were also those that had the lowest 360 fecundity. This result is similar to the one previously found in another experimental context 361 (Jehan et al., 2021). The slope of this positive relationship was decreased in response to the 362 early immune challenge, suggesting that females with the longest life expectancy were also 363 those that exhibited the largest reduction in fecundity caused by the early challenge. These 364 females, presumably also those of the best relative quality, were likely those that invested the 365 most into immunity in response to the early challenge. This suggests that they might have 366 diverted resources from current reproduction to sustain somatic defence. We may hypothesise 367 that these females may gain fitness by improving their survival to the pathogenic threat to 368 reproduce later, as predicted by the reproductive restraint hypothesis. However, since our 369 immune treatment only mimicked a survival threat without being real, our experimental 370 design cannot reveal the true associated fitness benefit of diverting resources from current 371 reproduction to immunity. In any case, this result, and the fact that female close to death did 372

not produce the highest clutch-size, strongly suggests that T. molitor females did not perform 373 a terminal investment.

374 As noted earlier, our study comforts the results of a previous study which found no 375 evidence of a terminal investment with age in T. molitor females (Jehan et al., 2021). 376 However, this previous study did not involve cues, such as an immune challenge, that would 377 represent a terminal investment trigger (Duffield et al., 2017). Here, we have used *B. cereus*, 378 recognized as an important natural pathogen of T. molitor (Du & Laing, 2011; Jurat-Fuentes 379 & Jackson, 2012), to challenge the females' immune system. Such an immune challenge was 380 expected to mimic a systemic infection by the bacterium, representing a significant cue 381 predicting imminent death. Accordingly, experimental infections with this bacterium can kill 382 90% of T. molitor females in less than three days (Dhinaut et al., 2018; Jehan et al., 2021). 383 Here, the use of B. cereus immune challenge did not trigger terminal investment either in 384 young or old females. However, we cannot dismiss that the way the immune challenge was 385 performed might have been too artificial to accurately mimic a deadly infection by *B. cereus*. 386 B. cereus infects T. molitor mostly orally and more accidentally by wounding through the 387 cuticle. The host immune response can greatly vary according to the route of infection 388 (Behrens et al., 2014), as host adaptation can be contingent on the infection route taken by 389 pathogens (Martins et al., 2013). Future studies may need to perform oral immune challenges 390 to mimic the trigger that would induce terminal investment. Nevertheless, *Bacilli* bacteria that 391 include numerous entomopathogens such as *B. cereus*, appear to have been of paramount 392 importance in the evolution of T. molitor immunity, especially through the evolution of 393 immune priming (Dubuffet et al., 2015). This is shown by the fact that they induce highly 394 persistent antibacterial immune responses compared to other bacterial pathogens (Dhinaut et 395 al., 2018). The very long-lasting antibacterial response resulting from the *B. cereus* challenge observed in this study suggests that the immune system of *T. molitor* females integrated theinfectious signal appropriately.

398 In conclusion, our work does not support the triggering of a terminal investment in T. 399 molitor females in response to a challenge with a natural bacterial pathogen of this insect. 400 Indeed, we showed that, while investing in their immune function, immune-challenged 401 females early in life restrained their reproduction and maintained their longevity. Moreover, 402 when immune challenged late in life, females maintained their immunity and longevity, which 403 may provide future reproductive opportunities instead of performing terminal investment. On 404 the basis of this work and a previous one (Jehan et al., 2021), the reproductive strategy of T. 405 molitor females appears to rely on reproductive restraint rather than on terminal investment. 406 This strategy also seems to be the opposite of the one used by males, which were found to 407 produce a terminal reproductive effort in comparable circumstances (Kivleniece et al., 2010; 408 Krams et al., 2011; Sadd et al., 2006). However, it has been proposed that males may have 409 higher fitness payoffs from terminal investment than females (Bateman, 1948; Kivleniece et 410 al., 2010; Sadd et al., 2006), a statement that looks true in the light of the present study.

411

412 Acknowledgements

413 We thank B. O'Driscoll for proofreading our manuscript. This study was supported by the 414 CNRS and a grant from the ANR (ANR-14-CE02-0009) to Y.M. The authors declare no 415 conflict of interest.

416

417 Authors' contributions

CJ, TR and YM conceived the ideas and designed methodology; CJ and CS collected the data;
CJ and YM analysed the data; CJ, TR and YM led the writing of the manuscript. All authors
contributed critically to the drafts and gave final approval for publication.

421

422 Data accessibility

423 Data will be available from the Dryad Digital Repository.

424

425 **References**

- Adamo, S. A. (1999). Evidence for adaptive changes in egg laying in crickets exposed to
 bacteria and parasites. *Animal Behaviour*, 57, 117–124.
 https://doi.org/10.1006/anbe.1998.0999
- Agnew, P., C Koella, J., & Michalakis, Y. (2000). Host life history responses to parasitism. *Microbes and Infection*, *2*, 891–896. https://doi.org/10.1016/s1286-4579(00)00389-0
- 431 Bateman, A. J. (1948). Intra-sexual selection in Drosophila. *Heredity*, 2, 349–368.
 432 https://doi.org/10.1038/hdy.1948.21
- 433 Behrens, S., Peuß, R., Milutinović, B., Eggert, H., Esser, D., Rosenstiel, P., Schulenburg, H.,
- Bornberg-Bauer, E., & Kurtz, J. (2014). Infection routes matter in population-specific
 responses of the red flour beetle to the entomopathogen *Bacillus thuringiensis*. *BMC Genomics*, *15*, 445. https://doi.org/10.1186/1471-2164-15-445
- Blair, L., & Webster, J. P. (2007). Dose-dependent schistosome-induced mortality and
 morbidity risk elevates host reproductive effort. *Journal of Evolutionary Biology*, *20*,
 54–61. https://doi.org/10.1111/j.1420-9101.2006.01230.x
- Bonneaud, C., Mazuc, J., Chastel, O., Westerdahl, H., & Sorci, G. (2004). Terminal
 investment induced by immune challenge and fitness traits associated with major
 histocompatibility complex in the house sparrow. *Evolution*, *58*, 2823–2830.
 https://doi.org/10.1111/j.0014-3820.2004.tb01633.x
- Bulet, P., Cociancich, S., Reuland, M., Sauber, F., Bischoff, R., Hegy, G., Van Dorsselaer, A.,
 Hetru, C., & Hoffmann, J. A. (1992). A novel insect defensin mediates the inducible

Page 21 of 38

446 antibacterial activity in larvae of the dragonfly Aeschna cyanea (Paleoptera, Od	donata).
--	----------

447 European Journal of Biochemistry, 209, 977–984. https://doi.org/10.1111/j.1432-

448 1033.1992.tb17371.x

- Chadwick, W., & Little, T. J. (2005). A parasite-mediated life-history shift in *Daphnia magna. Proceedings of the Royal Society B: Biological Sciences*, 272, 505–509.
 https://doi.org/10.1098/rspb.2004.2959
- 452 Clutton-Brock, T. H. (1984). Reproductive Effort and terminal investment in iteroparous
 453 animals. *The American Naturalist*, *123*, 212–229. https://doi.org/10.1086/284198

454 Coustau, C., Kurtz, J., & Moret, Y. (2016). A novel mechanism of immune memory unveiled

- 455 at the invertebrate-parasite interface. *Trends in Parasitology*, *32*, 353-355.
 456 https://doi.org/10.1016/j.pt.2016.02.005
- 457 Curio, E. (1983). Why de young birds reproduce less well? *Ibis*, *125*, 400–404.
 458 https://doi.org/10.1111/j.1474-919X.1983.tb03130.x
- Dhinaut, J., Chogne, M., & Moret, Y. (2018). Immune priming specificity within and across
 generations reveals the range of pathogens affecting evolution of immunity in an
 insect. *The Journal of Animal Ecology*, 87, 448–463. https://doi.org/10.1111/1365-
- 462 2656.12661
- 463 Dick, J. (1937). Oviposition in certain Coleoptera. *Annals of Applied Biology*, 24, 762–796.
 464 https://doi.org/10.1111/j.1744-7348.1937.tb05055.x
- Drnevich, J. M., Papke, R. S., Rauser, C. L., & Rutowski, R. L. (2001). Material benefits
 from multiple mating in female mealworm beetles (*Tenebrio molitor* L.). *Journal of Insect Behavior*, 14, 215–230. https://doi.org/10.1023/A:1007889712054
- 468 Du, N., & Laing, M. (2011). Determination of insecticidal toxicity of three species of
 469 entomopathogenic spore-forming bacterial isolates against *Tenebrio molitor* L.

- 470 (Coleoptera: Tenebrionidae). African Journal of Microbiology Research 15, 2222-
- 471 2228.
- Dubuffet, A., Zanchi, C., Boutet, G., Moreau, J., Teixeira, M., & Moret, Y. (2015). Transgenerational immune priming protects the eggs only against gram-positive bacteria in
 the mealworm beetle. *PLOS Pathogens*, *11*, e1005178.
- 475 https://doi.org/10.1371/journal.ppat.1005178
- 476 Duffield, K. R., Bowers, E. K., Sakaluk, S. K., & Sadd, B. M. (2017). A dynamic threshold
 477 model for terminal investment. *Behavioral Ecology and Sociobiology*, *71*, 185.
 478 https://doi.org/10.1007/s00265-017-2416-z
- 479 Forbes, M. R. L. (1993). Parasitism and host reproductive effort. *Oikos*, 67, 444–450.
 480 https://doi.org/10.2307/3545356
- 481 Giehr, J., Grasse, A. V., Cremer, S., Heinze, J., & Schrempf, A. (2017). Ant queens increase
 482 their reproductive efforts after pathogen infection. *Royal Society Open Science*, *4*,
 483 170547. https://doi.org/10.1098/rsos.170547
- 484 Grenfell, B. T., & Dobson, A. P. (Eds.). (1995). Ecology of infectious diseases in natural
 485 populations. Cambridge University Press.
 486 https://doi.org/10.1017/CBO9780511629396
- Haine, E. R., Pollitt, L. C., Moret, Y., Siva-Jothy, M. T., & Rolff, J. (2008). Temporal
 patterns in immune responses to a range of microbial insults (*Tenebrio molitor*). *Journal of Insect Physiology*, 54, 1090–1097.
- 490 https://doi.org/10.1016/j.jinsphys.2008.04.013
- 491 Hamel, S., Gaillard, J.-M., Yoccoz, N. G., Loison, A., Bonenfant, C., & Descamps, S. (2010).
- 492 Fitness costs of reproduction depend on life speed: empirical evidence from
 493 mammalian populations. *Ecology Letters*, *13*, 915-935. doi:10.1111/j.1461494 0248.2010.01478.x

495	Hamilton,	W.	D.,	& Z	Zuk,	M.	(1982).	Heritable	true	fitness	and	bright	birds:	a	role	for
496	par	asite	es? So	cienc	ce, 2	18, 3	384–387	. https://do	i.org/	10.1126	/scie	nce.712	23238			

- Hochberg, M. E., Michalakis, Y., & Meeus, T. D. (1992). Parasitism as a constraint on the
 rate of life-history evolution. *Journal of Evolutionary Biology*, *5*, 491–504.
 https://doi.org/10.1046/j.1420-9101.1992.5030491.x
- Hughes, P. W. (2017). Between semelparity and iteroparity: Empirical evidence for a
 continuum of modes of parity. *Ecology and Evolution*, 7, 8232–8261.
 https://doi.org/10.1002/ece3.3341
- Jehan, C., Chogne, M., Rigaud, T., & Moret, Y. (2020). Sex-specific patterns of senescence in
 artificial insect populations varying in sex-ratio to manipulate reproductive effort.
 BMC Evolutionary Biology, 20, 18. https://doi.org/10.1186/s12862-020-1586-x
- Jehan, C., Sabarly, C., Rigaud, T., & Moret, Y. (2021). Late-life reproduction in an insect:
 Terminal investment, reproductive restraint or senescence. *Journal of Animal Ecology*,

508 90, 282–297. https://doi.org/10.1111/1365-2656.13367

- Jönsson, K. I. (1997). Capital and income breeding as alternative tactics of resource use in
 reproduction. *Oikos*, 78, 57–66. https://doi.org/10.2307/3545800
- 511 Jurat-Fuentes, J. L., & Jackson, T. A. (2012). Chapter 8 Bacterial entomopathogens. In F. E.
- 512 Vega & H. K. Kaya (Eds.), *Insect pathology (Second Edition)* (pp. 265–349).
 513 Academic Press. https://doi.org/10.1016/B978-0-12-384984-7.00008-7
- Khan, I., Agashe, D., & Rolff, J. (2017). Early-life inflammation, immune response and
 ageing. *Proceedings of the Royal Society B: Biological Sciences*, 284, 20170125.
 https://doi.org/10.1098/rspb.2017.0125
- 517 Kivleniece, I., Krams, I., Daukšte, J., Krama, T., & Rantala, M. J. (2010). Sexual 518 attractiveness of immune-challenged male mealworm beetles suggests terminal

- 519 investment in reproduction. Animal Behaviour, 80, 1015–1021.
 520 https://doi.org/10.1016/j.anbehav.2010.09.004
- 521 Krams, I., Daukšte, J., Kivleniece, I., Krama, T., Rantala, M., Ramey, G., & Šauša, L. (2011).
- 522 Female choice reveals terminal investment in male mealworm beetles, *Tenebrio* 523 *molitor*, after a repeated activation of the immune system. *Journal of Insect Science*,
- 524 *11*. https://doi.org/10.1673/031.011.5601
- 525 Lui, K. J. (1998). Interval estimation of the risk ratio between a secondary infection, given a 526 primary infection, and the primary infection. *Biometrics*, *54*, 706–711.
- Martins, N. E., Faria, V. G., Teixeira, L., Magalhães, S., & Sucena, É. (2013). Host
 Adaptation is contingent upon the infection route taken by pathogens. *PLOS Pathogens*, 9, e1003601. https://doi.org/10.1371/journal.ppat.1003601
- McDade, T. W., Georgiev, A. V., & Kuzawa, C. W. (2016). Trade-offs between acquired and
 innate immune defenses in humans. *Evolution, Medicine, and Public Health, 2016*, 1–
 16. https://doi.org/10.1093/emph/eov033
- McNamara, J. M., Houston, A. I., Barta, Z., Scheuerlein, A., & Fromhage, L. (2009).
 Deterioration, death and the evolution of reproductive restraint in late life. *Proceedings of the Royal Society B: Biological Sciences*, 276, 4061–4066.
 https://doi.org/10.1098/rspb.2009.0959
- 537 Milutinović, B., & Kurtz, J. (2016). Immune memory in invertebrates. *Seminars in*538 *Immunology*, 28, 328–342. https://doi.org/10.1016/j.smim.2016.05.004
- Minchella, D. J. (1985). Host life-history variation in response to parasitism. *Parasitology*, *90*,
 205–216. https://doi.org/10.1017/S0031182000049143
- 541 Minchella, D. J., & Loverde, P. T. (1981). A cost of increased early reproductive effort in the
 542 snail *Biomphalaria glabrata*. *The American Naturalist*, *118*, 876–881.
 543 https://doi.org/10.1086/283879

- Moreau, J., Martinaud, G., Troussard, J.-P., Zanchi, C., & Moret, Y. (2012). Transgenerational immune priming is constrained by the maternal immune response in an insect. *Oikos*, *121*, 1828–1832. https://doi.org/10.1111/j.1600-0706.2011.19933.x
- Moret, Y. (2003). Explaining variable costs of the immune response: selection for specific
 versus non-specific immunity and facultative life history change. *Oikos*, *102*, 213–
 216. https://doi.org/10.1034/j.1600-0706.2003.12496.x
- Moret, Y. (2006). 'Trans-generational immune priming': specific enhancement of the
 antimicrobial immune response in the mealworm beetle, *Tenebrio molitor*. *Proceedings of the Royal Society B: Biological Sciences*, 273, 1399–1405.
 https://doi.org/10.1098/rspb.2006.3465
- Moret, Y., & Schmid-Hempel, P. (2000). Survival for immunity: the price of immune system
 activation for bumblebee workers. *Science*, 290, 1166–1168.
 https://doi.org/10.1126/science.290.5494.1166
- Moret, Y., & Siva-Jothy, M. T. (2003). Adaptive innate immunity? Responsive-mode
 prophylaxis in the mealworm beetle, *Tenebrio molitor*. *Proceedings of the Royal Society* B: Biological Sciences, 270, 2475–2480.
 https://doi.org/10.1098/rspb.2003.2511
- Norris, K., & Evans, M. R. (2000). Ecological immunology: life history trade-offs and
 immune defense in birds. *Behavioral Ecology*, *11*, 19–26.
 https://doi.org/10.1093/beheco/11.1.19
- 564 Perrin, N., Christe, P., & Richner, H. (1996). On host life-history response to parasitism.
 565 *Oikos*, 75, 317–320. https://doi.org/10.2307/3546256
- Punzo, F. (1975). Effects of temperature, moisture and thermal acclimation on the biology of
 Tenebrio molitor (Coleoptera: Tenebrionidae) [PhD Thesis, Iowa State University].
 https://lib.dr.iastate.edu/rtd/5438

- Pursall, E. R., & Rolff, J. (2011). Immune responses accelerate ageing: proof-of-principle in
 an insect model. *PLOS ONE*, *6*, e19972. https://doi.org/10.1371/journal.pone.0019972
- R Core Team (2020). A language and environment for statistical computing. R Foundation for
 Statistical Computing, Vienna, Austria. http://www.R-project.org/
- Ribeiro, N., Abelho, M., & Costa, R. (2018). A review of the scientific literature for optimal
 conditions for mass rearing *Tenebrio molitor* (Coleoptera: Tenebrionidae). *Journal of Entomological Science*, 53, 434–454. https://doi.org/10.18474/JES17-67.1
- 576 Roff, D. (1993). Evolution of life histories: Theory and analysis. Springer US.
 577 https://www.springer.com/gp/book/9780412023910
- Sadd, B. M., Holman, L., Armitage, H., Lock, F., Marland, R., & Siva-Jothy, M. T. (2006).
 Modulation of sexual signalling by immune challenged male mealworm beetles
 (*Tenebrio molitor*, L.): evidence for terminal investment and dishonesty. *Journal of Evolutionary Biology*, *19*, 321–325. https://doi.org/10.1111/j.1420-9101.2005.01062.x
- Sadd, Ben M, & Siva-Jothy, M. T. (2006). Self-harm caused by an insect's innate immunity. *Proceedings of the Royal Society B: Biological Sciences*, 273, 2571–2574.
- 584 https://doi.org/10.1098/rspb.2006.3574
- Schwanz, L. E. (2008). Chronic parasitic infection alters reproductive output in deer mice. *Behavioral Ecology and Sociobiology*, *62*, 1351–1358.
- Sheldon, B. C., & Verhulst, S. (1996). Ecological immunology: costly parasite defences and
 trade-offs in evolutionary ecology. *Trends in Ecology & Evolution*, 11, 317–321.
- 589 https://doi.org/10.1016/0169-5347(96)10039-2
- 590 Stearns, S. C. (1992). *The evolution of life histories*. Oxford University Press.
- Tarwater, C. E., & Arcese, P. (2017). Age and years to death disparately influence
 reproductive allocation in a short-lived bird. *Ecology*, *98*, 2248–2254.
 https://doi.org/10.1002/ecy.1851

- Therneau, T. M. (2020). A package for survival analysis in R. R package version 3.1-12.
 Retrieved from https://CRAN.R-proje ct.org/package=survival
- 596 Urabe, K., Aroca, P., Tsukamoto, K., Mascagna, D., Palumbo, A., Prota, G., & Hearing, V. J.
- 597 (1994). The inherent cytotoxicity of melanin precursors: a revision. *Biochimica Et*598 *Biophysica Acta*, 1221, 272–278. https://doi.org/10.1016/0167-4889(94)90250-x
- 599 Vigneron, A., Jehan, C., Rigaud, T., & Moret, Y. (2019). Immune defenses of a beneficial
- pest: the mealworm beetle, *Tenebrio molitor. Frontiers in Physiology*, 10, 138.
 https://doi.org/10.3389/fphys.2019.00138
- Williams, G. C. (1966). Natural selection, the costs of reproduction, and a refinement of
 Lack's principle. *The American Naturalist*, *100*, 687–690.
 https://doi.org/10.1086/282461
- Zanchi, C., Moret, Y., & Gillingham, M. A. F. (2019). Relationship between early body
 condition, energetic reserves and fitness in an iteroparous insect. *BioRxiv*, 774893.
 https://doi.org/10.1101/774893

Table 1. Mixed linear model investigating variance in female egg production as a function of their age, their experience of an immune challenge occurring early and/or late in life, body mass measured before each reproductive episode and residual life expectancy calculated at each reproductive episode. Female identity was included as a random factor. Values $P \le 0.05$ are given in bold.

614

Source	Estimate \pm s.e.	d.f. _{num.}	d.f. den.	F	Р
Agea	-49.93 ± 6.06	1	183	79.28	< 0.001
Early challenge ^b	-8.45 ± 10.02	1	183	0.07	0.796
Late challenge ^b	-2.69 ± 4.12	1	183	0.43	0.515
Life expectancy	0.77 ± 0.12	1	183	57.35	< 0.001
Body mass	0.26 ± 0.13	1	183	4.26	0.040
Age *Early challenge	21.60 ± 8.64	1	183	6.25	0.013
Early challenge*Life expectancy	0.38 ± 0.15	1	183	6.16	0.014

 a Estimate of old female relative to young females as the reference (estimate = 0).

^bEstimate of challenged female relative to control females as the reference (estimate = 0).
617

618

010

Table 2. Generalized Linear Model (GLM, Tweedie distribution, Log link function) analysing antibacterial activity of (a) *Tenebrio molitor* females assessed early (30 day old) and late (60 day old) in life after a single immune challenge occurring at 10 day old and 40 day old, respectively, as a function of their age and their age-specific immune treatment (N = 113), and of (b) females assessed late in life (60 days old) as a function of their immune treatment early and late in life (N = 77). Body mass did not explain any variation in antibacterial activity and was therefore removed from the models. Values $P \le 0.05$ are given in bold.

627

Source of variation	β	s.e.	Wald χ^2	d.f.	Р
(a) Young and old females					
Age ^a	1.87	0.81	3.64	1	0.05
Challenge ^b	3.54	0.71	32.67	1	< 0.0
Age* Challenge	-2.07	0.88	5.58	1	0.01
(b) Old females					
Early Challenge ^b	1.43	0.43	7.14	1	0. 00
Late Challenge ^b	1.51	0.43	8.65	1	0.00
Early Challenge* Late Challenge	-1.37	0.56	6.02	1	0.01

⁶⁴⁴ ^aEstimate of old female relative to young females as the reference (estimate = 0).

^bEstimate of challenged female relative to control females as the reference (estimate = 0).

646 647

648

649

Figure 1. Mean body mass (\pm s.e.) of females at the age at which they started their early (at 10 days post-eclosion) and their late (at 40 days post-eclosion) reproductive episodes. Numbers at the bottom of the bars are sample size.

654

Figure 2. Mean number of eggs (\pm s.e.) produced by females early in life (between day 10 and day 30 post-eclosion) and late in life (between day 40 and day 60 post-eclosion) as function of their life time immune treatment: control (white bars), immune challenged early (grey bars), immune challenged late (hatched white bar) and immune challenged both early and late in life (hatched grey bar). Numbers at the bottom of the bars are sample size.

660

661 Figure 3. Relationship between fecundity (number of eggs) and age-specific life expectancy 662 in days of (A) young and (B) old females that have been exposed to an early immune 663 challenge. In (A), the relationships between fecundity and life expectancy were y = 0.85 xdays of life expectancy + 38.32, $r^2 = 0.34$, P < 0.0001 and y = 0.40 x days of life expectancy + 664 665 34.04, $r^2 = 0.15$, P = 0.0017 among controlled and early-challenged females, respectively. In 666 (B), the relationships between fecundity and life expectancy were y = 0.47 x days of life expectancy + 5.21, $r^2 = 0.21$, P = 0.0033 and y = 0.50 x days of life expectancy - 0.62, $r^2 =$ 667 668 0.20, P < 0.0084 among controlled and early-challenged females, respectively.

669

Figure 4. Mean antibacterial activity $(\log_{10} \text{ (mm)} \pm \text{s.e.})$ in the haemolymph of young (30 days post-eclosion) and old (60 days post-eclosion) females as function of their life time immune treatment: control (white bars), immune challenged early (at 10 post-eclosion, grey bars), immune challenged late (at 40 days post-eclosion, hatched white bar) and immune challenged both early and late in life (at 10 and 40 days post-eclosion, hatched grey bar). Numbers at the bottom of the bars are sample size.

676 Supplementary information

677

678 Table S1. Mixed effects Cox regression analysis for T. molitor female longevity according to 679 early challenge ('early reproduction': yes/no), late challenge ('Late reproduction': yes/no) and 680 their interaction as independent categorical covariates in a full factorial design. Female body 681 mass ('Mass') was also included as an independent continuous covariate, which was tested as 682 a main effect only. To account for female individual heterogeneity, female identity was 683 included as a random effect in the model. We limited model reduction to the interaction terms 684 when found not significant, whereas main effects were always kept in the final model (N =685 120).

686

Figure S1. Survival of *Tenebrio molitor* females as a function of their life-time immune
treatment: C0 control, C1 early-life, C2 late-life and C12 early and late immune challenges

689 with the inactivated bacterium *Bacillus cereus*.

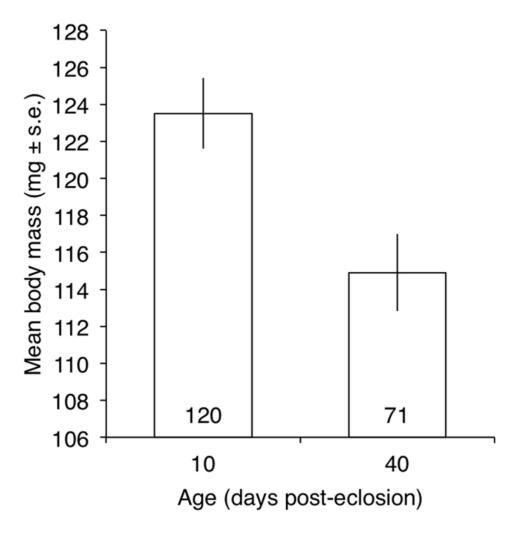


Figure 1. Mean body mass (\pm s.e.) of females at the age at which they started their early (at 10 days posteclosion) and their late (at 40 days post-eclosion) reproductive episodes. Numbers at the bottom of the bars are sample size.

70x70mm (300 x 300 DPI)

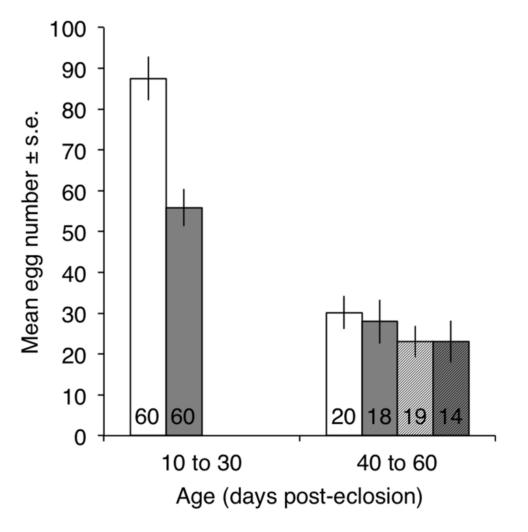


Figure 2. Mean number of eggs (± s.e.) produced by females early in life (between day 10 and day 30 posteclosion) and late in life (between day 40 and day 60 post-eclosion) as function of their life time immune treatment: control (white bars), immune challenged early (grey bars), immune challenged late (hatched white bar) and immune challenged both early and late in life (hatched grey bar). Numbers at the bottom of the bars are sample size.

64x65mm (300 x 300 DPI)

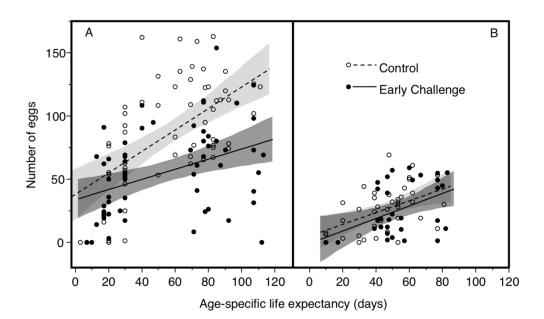


Figure 3. Relationship between fecundity (number of eggs) and age-specific life expectancy in days of (A) young and (B) old females that have been exposed to an early immune challenge. In (A), the relationships between fecundity and life expectancy were $y = 0.85 \times days$ of life expectancy + 38.32, r2 = 0.34, P < 0.0001 and $y = 0.40 \times days$ of life expectancy + 34.04, r2 = 0.15, P = 0.0017 among controlled and early-challenged females, respectively. In (B), the relationships between fecundity and life expectancy + 5.21, r2 = 0.21, P = 0.0033 and $y = 0.50 \times days$ of life expectancy - 0.62, r2 = 0.20, P < 0.0084 among controlled and early-challenged females, respectively.

109x68mm (600 x 600 DPI)

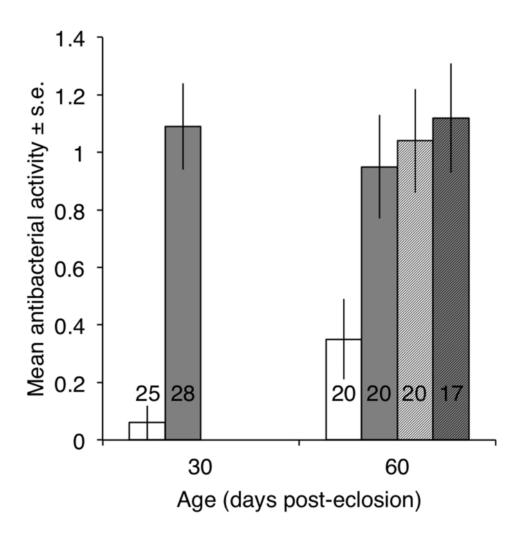


Figure 4. Mean antibacterial activity (log10 (mm) ± s.e.) in the haemolymph of young (30 days posteclosion) and old (60 days post-eclosion) females as function of their life time immune treatment: control (white bars), immune challenged early (at 10 post-eclosion, grey bars), immune challenged late (at 40 days post-eclosion, hatched white bar) and immune challenged both early and late in life (at 10 and 40 days post-eclosion, hatched grey bar). Numbers at the bottom of the bars are sample size.

65x66mm (300 x 300 DPI)

Supplementary Information for:

Age-specific fecundity under parasitic threat in an insect: terminal

investment versus reproductive restraint

Table S1. Mixed effects Cox regression analysis for *T. molitor* female longevity according to early challenge ('early reproduction': yes/no), late challenge ('Late reproduction': yes/no) and their interaction as independent categorical covariates in a full factorial design. Female body mass ('Mass') was also included as an independent continuous covariate, which was tested as a main effect only. To account for female individual heterogeneity, female identity was included as a random effect in the model. We limited model reduction to the interaction terms when found not significant, whereas main effects were always kept in the final model (N = 120).

Variable in the best model	ß	\pm s.e.	Z	d.f.	Р	Odd ratio
Mass	- 0.011	0.010	-1.01	1	0.310	0.99
Early challenge	0.218	0.356	0.61	1	0.540	1.24
Late challenge	0.295	0.357	0.83	1	0.410	1.34

Random effects (Female ID): SD 1.36, Variance 1.85

Proportional hazard assumption was checked as a non-significant relationship between residuals and time using the cox.zph() function in the R survival package (global test $\chi^2_{65.9} = 3.70 \text{ P} = 1$, the test is also non significant for each of the covariates).

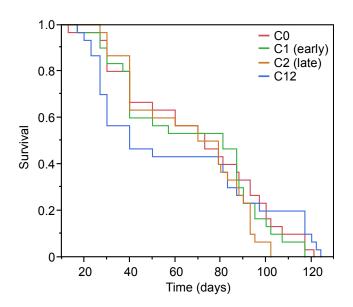


Fig. S1. Survival of *Tenebrio molitor* females as a function of their life-time immune treatment: C0 control, C1 early-life, C2 late-life and C12 early and late immune challenges with the inactivated bacterium *Bacillus cereus*.