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Identification of dabigatran etexilate major degradation pathways by liquid chromatography coupled to multi stage high-resolution mass spectrometry

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Dabigatran etexilate (DABET) is an oral direct thrombin inhibitor that has been approved for the prevention of blood clot formation. As the active pharmaceutical ingredient (API) may undergo degradation, leading to drug activity loss or to the occurrence of adverse effects associated with degradation products, thorough knowledge of the API's stability profile is required. Since very few studies have been reported on the drug stability profile, a study related to DABET's behaviour under stress conditions was carried out in order to identify its major degradation pathways. DABET was subjected to hydrolytic (acidic and alkaline), oxidative, photolytic and thermal stress, as per ICH-specified conditions. Up to ten degradation products along with dabigatran, the active metabolite of DABET, were formed and detected by reverse phase liquid chromatography in gradient mode (LC) coupled to UV and mass spectrometry (UV-MS). Structures were determined by elemental composition determination and study of the fragmentation patterns, using high-resolution mass spectrometry in multistage mode (HR-MSⁿ). Under hydrolytic stress conditions, *O*-dealkylation may occur and formation of benzimidic acid derivatives was also observed. DABET was shown to be much less susceptible to photolysis and oxidative stress, even if *N*-dealkylation was highlighted. In view of the structures identified, various degradation pathways of DABET have been proposed.

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Introduction

Dabigatran etexilate (DABET) (ethyl 3-((2-((4-((hexyloxy)carbonyl)carbamimidoyl)phenyl)amino)methyl)-1-methyl-1*H*-benzimidazol-5-yl)carbonyl[(pyridin-2-yl)amino]propanoate) is a pro-drug, rapidly converted to dabigatran after oral administration. As a direct thrombin inhibitor, DABET is used in the prevention of thromboembolic events for patients with atrial fibrillation as well as in the prevention of venous thromboembolism.¹⁻⁴ It has been developed as an alternative to warfarin, an

anticoagulant with a narrow therapeutic index and hence trickier to use. As the API may experience degradation, resulting in activity loss or occurrence of adverse effects associated with the appearance of degradation products, a thorough knowledge of the API's stability profile is one of the key factors to prevent these risks during manufacture, transportation and storage.⁵ This is why study of the API's major degradation pathways should be carried out under stress conditions, such as those recommended by the International Conference of Harmonization guidelines.⁶

Liquid chromatography combined with multi-stage mass spectrometry (LC-MSⁿ) has been successfully used for the identification and characterization of degradation products generated by the API.⁷⁻¹² The comprehensive fragmentation pattern of API can be compared to the fragment ions of degradation products thus enabling their characterization.

Liquid chromatography methods have been used for the determination of dabigatran in plasma and other biological matrices.¹³⁻¹⁶ Impurities formed during the synthesis of DABET¹⁷ and drug metabolites¹⁸ have also been determined by LC and reported in literature. However, no study has been published on the formation and characterization of the DABET degradation products. That's why various stress conditions have

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been applied in order to simulate the degradation of DABET, such as hydrolysis, thermal, photolysis and oxidative conditions. The degradation products were to be detected and characterized. In view of the structures identified, major degradation pathways of DABET were to be proposed.

Experimental

Chemicals and reagent

Dabigatran etexilate (MW: 627.7332 g mol⁻¹) tablets (Pradaxa®) are marketed by Boehringer Ingelheim (France). Analytical grade methanol and formic acid came from Sigma-Aldrich (St Quentin-Fallavier, France). Ultrapure water was produced by the Q-Pod Milli-Q system (Millipore, Molsheim, France) and used for dissolution or as a mobile phase component. Hydrogen peroxide (H₂O₂) 30% was provided by Carlo Erba SDS (Val de Reuil, France).

LC-UV-HR-MS conditions

LC analyses were performed using a Dionex Ultimate 3000 system (DIONEX, Ulis, France) coupled to UV and MS detections. LC is equipped with a quaternary pump, a degasser, an autosampler and a thermostated column compartment, a 50 µL injection loop and a photo-diode array (PDA) detector. Kinetex™, C18 (100 Å, 50 mm × 2.1 mm, 2.6 µm) column accounted for the stationary phase. Mobile phase was composed of 2.5 mM formic acid (A) and methanol (B). The optimized gradient chromatographic program was the following: B 5% from 0 to 5 min; B 100% from 5 to 25 min with a fixed level from 25 to 28 min; B 5% from 28 to 35 min. The flow rate was set at 0.4 mL min⁻¹. LC-HR-MSⁿ was performed coupling this same LC system to an electrospray (ESI)-LTQ-Orbitrap Velos Pro system, composed of a double linear trap and an orbital trap (Thermo Fisher Scientific, CA, USA). Analyses were carried out in positive ion mode with the following conditions: the source voltage was set at 3.4 kV, the source and the capillary temperatures were fixed at 500 °C and 600 °C, respectively. S-lens was set at 60%. 30–40% CEL were set for high-resolution fragmentation studies. The MS data were processed using Xcalibur® software (version 2.2 SP 1.48).

Forced degradation protocol

Stock standard solutions were prepared by disintegrating 10 tablets, accounting for 1500 mg DABET, in 1500 mL of ultrapure water/methanol 50/50 (v/v). The working solutions were prepared by diluting stock solution in sort to obtain a final concentration of 0.5 mg mL⁻¹. For each condition, samples were made up in triplicate and allocated in 15 mL hermetically sealed glass vials.

Four stress conditions were tested: thermal, hydrolytic, photolytic and oxidative conditions. Thermal stress was achieved at 80 °C up to 28 days. Hydrolysis was studied at 40 °C over a period of 72 hours using HCl 0.1 mol L⁻¹ or NaOH 0.1 mol L⁻¹. Oxidation was tested in the presence of an equivalent of 3% (v/v) H₂O₂, at room temperature for 72 hours. Photo-degraded

samples resulted from exposure of working solutions for 45 hours to light using a xenon test chamber Q-SUN Xe-1 (Q-Lab Westlake, California, USA) operating in window mode. Wavelengths ranged from 300 to 800 nm. The light intensity delivered was at 1.50 W m⁻².

Results and discussion

Degradation of DABET along with formation of degradation products

Fig. 1 shows LC-MS extracted ion chromatograms obtained by analysis of samples submitted to various stress conditions. A total of ten degradation products, along with DABET and dabigatran were detected when taken at a degradation rate near to 20%. Even if degradation continues beyond, the study has been deliberately limited to that of the degradation products formed precociously in stress conditions, insofar as the others, sometimes secondarily formed, can be considered as less likely with respect to real-storage.¹⁹

Thereafter, the degradation products are named “DP-*n*” and numbered according to their elution order. Their relative retention times (rrT) and the HR-MSⁿ data (origin, exact mass, accurate mass and relative errors of degradation products and relevant product ions) are gathered in Table 1. Eight degradation products and dabigatran were eluted ahead DABET, whereas two degradation products eluted after. During the implementation stages of the separation method, it was noticed that DABET's retention time varied slightly from one day to another and showed certain sensitivity to temperature. Thermostatisation of the column at 40 °C had allowed to effectively remedy this fluctuation, but this approach was not used to minimize risk of *in situ* degradation during analysis. Nevertheless, to control any co-elution risk and to be sure of the method capacity to highlight the main degradation products formed with each of the stress conditions, purity of DABET's peak and mass balance were systematically monitored. A component detection algorithm (CODA) was run and outcome showed that the main peak always contains the only signals pertaining to DABET. In addition, mass balance (%assay + % total degradation products) of all the stressed samples of DABET was obtained in the range of 98.7–99.8%, suggesting that for each condition tested, most of the degradation products formed have been detected (Tables 1 and 2).

The hydrolytic conditions showed a strong impact on DABET's stability (Tables 1 and 2). In basic condition, after 1 hour of exposition to NaOH 0.1 mol L⁻¹, 24% degradation has occurred, resulting in the formation of DP-1 (rrT = 0.52), dabigatran (rrT = 0.58) and DP-6 (rrT = 0.89). The acidic conditions yielded DP-7 (rrT = 0.92), DP-8 (rrT = 0.97), DP-9 (rrT = 1.08) and DP-10 (rrT = 1.10) after 24 hours of exposure to HCl 0.1 mol L⁻¹, accounting for 25% DABET loss (Table 1).

To the other degradation conditions, degradation seems to be slower. After three days of exposure to H₂O₂ 3%, DP-3 (rrT = 0.64) was formed along with 22% DABET loss. After 45 hours of exposure inside the light chamber, DP-2 (rrT = 0.63) and DP-5 (rrT = 0.78) appeared along with 11% DABET loss. After 28 days of exposure at 80 °C, DP-4 (rrT = 0.67) and

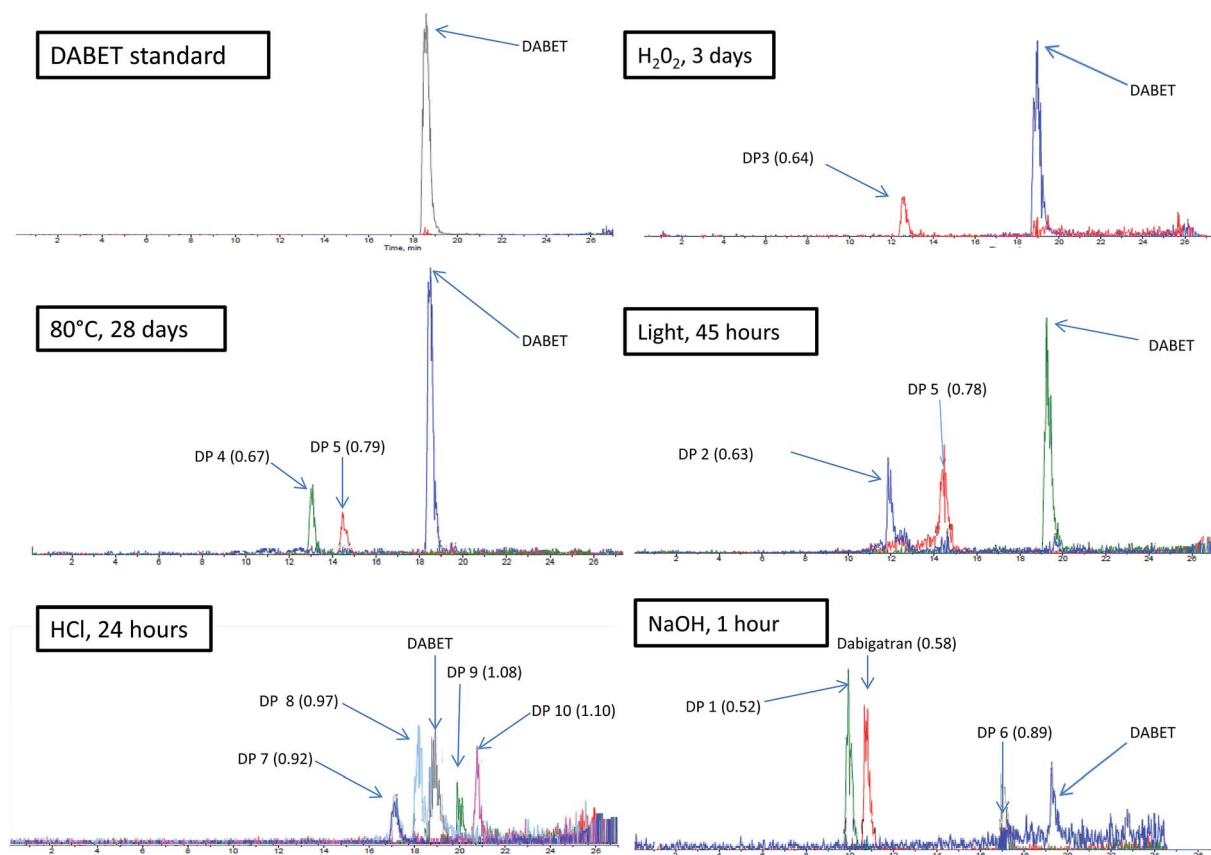


Fig. 1 Extracted ion chromatograms of DABET and degradation products according to stress conditions.

DP-5 (rrT = 0.78) were produced and only 8% of DABET loss was observed (Tables 1 and 2).

Characterization of DABET'S fragmentation pattern

The fragmentation scheme of DABET, which has not been studied in detail so far, was determined using ESI⁺ high-resolution multi-stage mass spectrometry (ESI⁺/HR-MSⁿ). The comprehension of various mechanisms that result, can, to a large extent, assist in the structural elucidation of the aforementioned degradation products. Hence, a study in depth of

DABET'S fragmentation pattern had been achieved in order to help assign, by comparison, the structures of the major product ions coming from the degradation products ions. The structures proposed for the product ions were systematically confirmed by the elemental compositions determined from accurate mass measurement and from the corresponding errors levels (systematically inferior to 4 ppm), as well as by logical or plausible losses. These data have been reported in Tables 2 and 3 and the proposed fragmentation pattern for the drug has been built based upon multi-stage HR-MS studies (Fig. 2 and 3). However, for the sake of visibility in terms of graphical

Table 1 Degradation outcome

Stress condition	Time	Average assay of API (%)	Average total impurities (% , area normalisation)	Average mass balance (assay + total impurities%)	Commentaries
Base hydrolysis (0.1 mol L ⁻¹ NaOH)	1 hour	76.0	22.7	98.7	Degradation accompanied by appearance of DP-1, dabigatran and DP-6
Acid hydrolysis (0.1 mol L ⁻¹ HCl)	24 hours	75.1	24.4	99.5	Degradation accompanied by appearance of DP-7, DP-8, DP-9 and DP-10
Oxidation (3% H ₂ O ₂)	72 hours	78.1	21.2	99.3	Degradation accompanied by appearance of DP-3
Thermal (80 °C)	28 days	91.8	8.0	99.8	Degradation accompanied by appearance of DP-4 and DP-5
Photolysis (UV light)	45 hours	88.6	10.3	98.9	Degradation accompanied by appearance of DP-2 and DP-5

Table 2 Relative retention times (rrT), accurate masses with errors, elemental compositions and MSⁿ relevant product ions of the degradation products

DP Name and rrT	Origin of ions	Best possible elemental formula	Theoretical mass <i>m/z</i>	Accurate mass <i>m/z</i>	Error (ppm)	
DP-1 (0.5)	Precursor ion	C ₂₅ H ₂₃ N ₆ Na ₂ O ₄ ⁺	517.15707	517.15679	-0.54	
	MS ¹	C ₂₅ H ₂₄ N ₆ O ₄ ⁺	473.19318	473.19285	-0.70	
	MS ² (517 →)	C ₂₂ H ₁₅ N ₄ Na ₂ O ₂ ⁺	445.13594	445.13535	-1.33	
	MS ² (517 →)	C ₁₈ H ₁₆ N ₄ Na ₂ O ₃ ⁺	382.10123	382.10101	-0.58	
	MS ² (517 →)	C ₁₇ H ₁₅ N ₄ Na ₂ O ₃ ⁺	369.09341	369.09322	-0.51	
	MS ³ (517 → 382 →)	C ₁₇ H ₁₃ N ₄ Na ₂ O ₃ ⁺	367.07776	367.07721	-1.50	
	MS ³ (517 → 445 →)	C ₁₆ H ₁₅ N ₄ Na ₂ O ⁺	325.10358	325.10331	-0.83	
	MS ³ (517 → 445 →)	C ₁₅ H ₁₂ N ₄ Na ₂ O ⁺	310.08100	310.07978	-3.93	
Dabigatran (0.58)	MS ³ (517 → 369 →)	C ₁₄ H ₁₁ N ₄ Na ₂ O ⁺	297.07228	297.07178	-1.68	
	Precursor ion	C ₂₅ H ₂₆ N ₇ O ₃ ⁺	472.20916	472.20771	-3.07	
	MS ² (472 →)	C ₂₂ H ₂₂ N ₇ O ⁺	400.18803	400.18768	-0.87	
		C ₁₇ H ₁₆ N ₅ O ⁺	306.13494	306.13483	-0.36	
DP-2 (0.61)		C ₁₇ H ₁₃ N ₄ O ⁺	289.10839	289.10829	-0.35	
	Precursor ion	C ₂₀ H ₂₁ N ₄ O ₄ ⁺	381.15573	381.15568	-0.13	
	MS ² (381 →)	C ₁₈ H ₁₅ N ₄ O ₃ ⁺	335.11387	335.11342	-1.34	
	MS ² (381 →)	C ₁₅ H ₁₃ N ₄ O ₂ ⁺	281.10330	281.10295	-1.25	
DP-3 (0.65)	MS ² (381 →)	C ₁₀ H ₇ N ₂ O ₂ ⁺	187.05020	187.04989	-1.66	
	Precursor ion	C ₂₇ H ₃₀ N ₇ O ₃ ⁺	500.24046	500.23944	-2.04	
	MS ² (500 →)	C ₂₇ H ₂₇ N ₆ O ₃ ⁺	483.21392	483.21381	-0.23	
		C ₂₂ H ₂₂ N ₇ O ⁺	400.18803	400.18788	-0.37	
DP-4 (0.67)		C ₂₀ H ₂₁ N ₄ O ₃ ⁺	365.16082	365.16070	-0.33	
		C ₁₇ H ₁₆ N ₅ O ⁺	306.13494	306.13488	-0.20	
	Precursor ion	C ₁₇ H ₁₃ N ₄ O ⁺	289.10839	289.10828	-0.38	
	MS ² (501 →)	C ₂₇ H ₂₉ N ₆ O ₄ ⁺	501.22448	501.22358	-1.80	
		C ₂₇ H ₂₆ N ₅ O ₄ ⁺	484.19793	484.19765	-0.58	
		C ₂₀ H ₂₁ N ₄ O ₃ ⁺	365.16082	365.16081	-0.03	
DP-5 (0.78)		C ₁₇ H ₁₇ N ₄ O ₃ ⁺	325.12952	325.12957	0.15	
		C ₁₇ H ₁₅ N ₄ O ₂ ⁺	307.11895	307.11914	0.62	
		C ₁₇ H ₁₃ N ₄ O ⁺	289.10839	289.10825	-0.48	
	Precursor ion	C ₂₁ H ₂₅ N ₄ O ₅ ⁺	413.18195	413.18088	-2.59	
	MS ² (413 →)	C ₂₁ H ₂₃ N ₄ O ₄ ⁺	395.17138	395.17124	-0.35	
	MS ² (413 →)	C ₂₀ H ₂₁ N ₄ O ₄ ⁺	381.15573	381.15560	-0.34	
	MS ³ (413 → 395 →)	C ₂₀ H ₂₀ N ₄ O ₄ ^{*+}	380.14791	380.14791	0.00	
	MS ³ (413 → 395 →)	C ₂₀ H ₁₉ N ₄ O ₄ ⁺	379.14008	379.14010	0.05	
	MS ³ (413 → 395 →)	C ₂₀ H ₂₃ N ₄ O ₃ ⁺	367.17647	367.17642	-0.14	
	MS ³ (413 → 395 →)	C ₂₀ H ₁₉ N ₄ O ₃ ⁺	363.14152	363.14157	0.15	
	MS ³ (413 → 395 →)	C ₁₉ H ₁₇ N ₄ O ₃ ⁺	349.12952	349.12945	-0.20	
	MS ³ (413 → 381 →)	C ₁₈ H ₁₅ N ₄ O ₃ ⁺	335.11387	335.11342	-1.34	
	MS ⁴ (413 → 395 → 349 →)	C ₁₈ H ₁₃ N ₄ O ₃ ⁺	333.09822	333.09724	-2.94	
	MS ⁴ (413 → 395 → 349 →)	C ₁₈ H ₁₃ N ₄ O ₂ ⁺	317.10330	317.10296	-1.07	
	MS ³ (413 → 395 →)	C ₁₆ H ₁₅ N ₄ O ₂ ⁺	295.11895	295.11801	-3.19	
	MS ³ (413 → 381 →)	C ₁₅ H ₁₃ N ₄ O ₂ ⁺	281.10330	281.10295	-1.25	
	MS ³ (413 → 395 →)	C ₁₅ H ₁₁ N ₄ O ₂ ⁺	279.08765	279.08727	-1.36	
	MS ⁴ (413 → 395 → 295 →)	C ₁₅ H ₁₅ N ₄ O ⁺	267.12404	267.12350	-2.02	
DP-6 (0.89)	MS ³ (413 → 395 →)	C ₁₅ H ₁₁ N ₄ O ⁺	263.09274	263.09241	-1.25	
	MS ² (413 →)	C ₁₁ H ₁₁ N ₂ O ₃ ⁺	219.07642	219.07612	-1.37	
	MS ³ (413 → 219 →)	C ₁₁ H ₉ N ₂ O ₂ ⁺	201.06585	201.06583	-0.10	
	MS ³ (413 → 381 →)	C ₁₀ H ₇ N ₂ O ₂ ⁺	187.05020	187.05021	0.05	
	MS ⁴ (413 → 395 → 295 →)	C ₁₀ H ₉ N ₂ O ⁺	173.07094	173.07082	-0.69	
	MS ³ (413 → 219 →)	C ₉ H ₇ N ₂ O ⁺	159.05529	159.05511	-1.13	
	Precursor ion	C ₂₉ H ₃₁ N ₆ O ₆ ⁺	559.22996	559.22840	-2.79	
	MS ² (559 →)	C ₂₈ H ₂₇ N ₆ O ₅ ⁺	527.20374	527.20362	-0.23	
	MS ³ (559 → 527 →)	C ₂₇ H ₂₉ N ₅ O ₄ ⁺	484.19793	484.19778	-0.31	
	MS ³ (559 → 527 →)	C ₂₀ H ₂₁ N ₄ O ₃ ⁺	365.16082	365.16059	-0.63	
	MS ² (559 →)	C ₁₈ H ₁₃ N ₄ O ₃ ⁺	333.09822	333.09829	0.22	
	MS ² (559 →)	C ₁₈ H ₁₆ N ₃ O ₃ ⁺	322.11862	322.11811	-1.58	
	DP-7 (0.92)	Precursor ion	C ₃₂ H ₃₈ N ₇ O ₅ ⁺	600.29289	600.29180	-1.82
		MS ² (600 →)	C ₂₆ H ₂₄ N ₇ O ₄ ⁺	498.18843	498.18830	-0.26
		C ₂₅ H ₂₃ N ₆ O ₃ ⁺	455.18262	455.18230	-0.70	
		C ₂₅ H ₂₆ N ₇ O ₃ ⁺	472.20916	472.20920	0.08	
	C ₁₈ H ₁₇ N ₄ O ₃ ⁺	337.12952	337.12898	-1.60		

Table 2 (Contd.)

DP Name and rrT	Origin of ions	Best possible elemental formula	Theoretical mass m/z	Accurate mass m/z	Error (ppm)
DP-8 (0.96)	Precursor ion MS ² (614 →)	C ₁₇ H ₁₃ N ₄ O ⁺	289.10839	289.10813	-0.90
		C ₃₃ H ₄₀ N ₇ O ₅ ⁺	614.30854	614.30705	-2.43
		C ₂₇ H ₂₆ N ₇ O ₄ ⁺	512.20408	512.20351	-1.11
		C ₂₆ H ₂₅ N ₆ O ₃ ⁺	469.19827	469.19782	-0.96
		C ₂₄ H ₂₈ N ₅ O ₃ ⁺	434.21867	434.21844	-0.53
		C ₂₃ H ₂₇ N ₄ O ₂ ⁺	391.21285	391.2126	-0.64
		C ₁₈ H ₁₄ N ₅ O ₂ ⁺	383.16149	383.16132	-0.44
		C ₁₉ H ₁₉ N ₄ O ₃ ⁺	351.14152	351.14131	-0.59
		C ₁₈ H ₁₄ N ₅ O ₂ ⁺	332.11420	332.11409	-0.33
		C ₁₆ H ₂₁ N ₂ O ₂ ⁺	273.15975	273.15954	-0.77
Dabigatran etexilate DP-9 (1.08)	API Precursor ion MS ² (615 →)	C ₃₄ H ₄₂ N ₇ O ₅ ⁺	628.32419	628.32236	-2.91
		C ₃₃ H ₃₉ N ₆ O ₆ ⁺	615.29256	615.29137	-1.93
		C ₂₇ H ₂₅ N ₆ O ₅ ⁺	513.18809	513.1873	-1.54
		C ₂₆ H ₂₄ N ₅ O ₄ ⁺	470.18228	470.18152	-1.62
		C ₂₄ H ₂₇ N ₄ O ₄ ⁺	435.20268	435.20226	-0.97
		C ₂₃ H ₂₆ N ₃ O ₃ ⁺	392.19687	392.19673	-0.36
		C ₂₂ H ₁₈ N ₅ O ₂ ⁺	384.145501	384.145102	-1.04
		C ₁₉ H ₁₉ N ₄ O ₃ ⁺	351.14517	351.14493	-0.68
		C ₁₈ H ₁₃ N ₄ O ₃ ⁺	333.09822	333.09811	-0.33
		C ₁₆ H ₂₁ N ₂ O ₂ ⁺	273.15975	273.15945	-1.10
DP-10 (1.10)	Precursor ion MS ² (629 →)	C ₃₄ H ₄₁ N ₆ O ₆ ⁺	629.30821	629.30712	-1.73
		C ₂₈ H ₂₇ N ₆ O ₅ ⁺	527.20374	527.20298	-1.44
		C ₂₇ H ₂₆ N ₅ O ₄ ⁺	484.19793	484.19669	-2.56
		C ₂₄ H ₂₇ N ₄ O ₄ ⁺	435.20268	435.20175	-2.14
		C ₂₃ H ₂₆ N ₃ O ₃ ⁺	392.19687	392.19598	-2.27
		C ₂₀ H ₂₁ N ₄ O ₃ ⁺	365.16082	365.16060	-0.60
		C ₁₈ H ₁₇ N ₄ O ₃ ⁺	337.12952	337.12891	-1.81
		C ₁₈ H ₁₃ N ₄ O ₃ ⁺	333.09822	333.0981	-0.36
		C ₁₇ H ₁₄ N ₃ O ₃ ⁺	308.10297	308.10292	-0.16
		C ₁₆ H ₂₁ N ₂ O ₂ ⁺	273.15975	273.15920	-2.01
		C ₁₅ H ₁₃ N ₄ O ⁺	265.10839	265.10825	-0.53

representations, the mass-to-charge values linked to each of the structures presented in Fig. 3–8 have been written in the form of nominal values.

Analysed in positive ion mode, DABET was detected as protonated [M + H]⁺ ion (m/z 628) and sodium adduct [M + Na]⁺

ion (m/z 650). Its HR-MS² spectrum yields 6 product ions with m/z of 526, 483, 434, 332, 289 and 273 (Table 3). However, as it will be demonstrated thereafter, the ions at m/z 483, 332, and 289 came from the ion at m/z 526, whereas m/z 273 ion turns out to be the fragmentation product of m/z 434 ion.

Table 3 Origin, best possible elemental formulae, exact and accurate masses of relevant DABET fragment ions along with relative errors in ppm

Product ions	Origin of fragments	Best possible elemental formulae	Theoretical mass m/z	Accurate mass m/z	Error (ppm)
628	(M + H) ⁺	C ₃₄ H ₄₂ N ₇ O ₅ ⁺	628.32419	628.32236	-2.91
526	MS ² (628 →)	C ₂₈ H ₂₈ N ₇ O ₄ ⁺	526.21973	526.21920	-1.01
483	MS ³ (628 → 526 →)	C ₂₇ H ₂₇ N ₆ O ₃ ⁺	483.21392	483.21327	-1.35
434	MS ² (628 →)	C ₂₄ H ₂₈ N ₅ O ₃ ⁺	434.21867	434.21824	-0.99
391	MS ³ (628 → 434 →)	C ₂₃ H ₂₇ N ₄ O ₂ ⁺	391.21285	391.21247	-0.97
365	MS ³ (628 → 526 →)	C ₂₀ H ₂₁ N ₄ O ₃ ⁺	365.16082	365.16060	-0.60
337	MS ³ (628 → 526 →)	C ₁₈ H ₁₇ N ₄ O ₃ ⁺	337.12952	337.12891	-1.81
332	MS ³ (628 → 526 →)	C ₁₈ H ₁₄ N ₅ O ₂ ⁺	332.11420	332.11401	-0.57
307	MS ⁴ (628 → 434 → 391)	C ₁₇ H ₁₅ N ₄ O ₂ ⁺	307.11895	307.11832	-2.05
289	MS ² (628 →)	C ₁₇ H ₁₃ N ₄ O ⁺	289.10839	289.10829	-0.35
273	MS ⁴ (628 → 434 → 391 →)	C ₁₆ H ₂₁ N ₂ O ₂ ⁺	273.15975	273.15920	-2.01
265	MS ³ (628 → 526)	C ₁₅ H ₁₃ N ₄ O ⁺	265.10839	265.10825	-0.53
261	MS ⁴ (628 → 434 → 289 →)	C ₁₆ H ₁₃ N ₄ ⁺	261.11347	261.11289	-2.22
172	MS ⁴ (628 → 434 → 289 →)	C ₁₀ H ₈ N ₂ O ⁺	172.06311	172.06273	-2.23

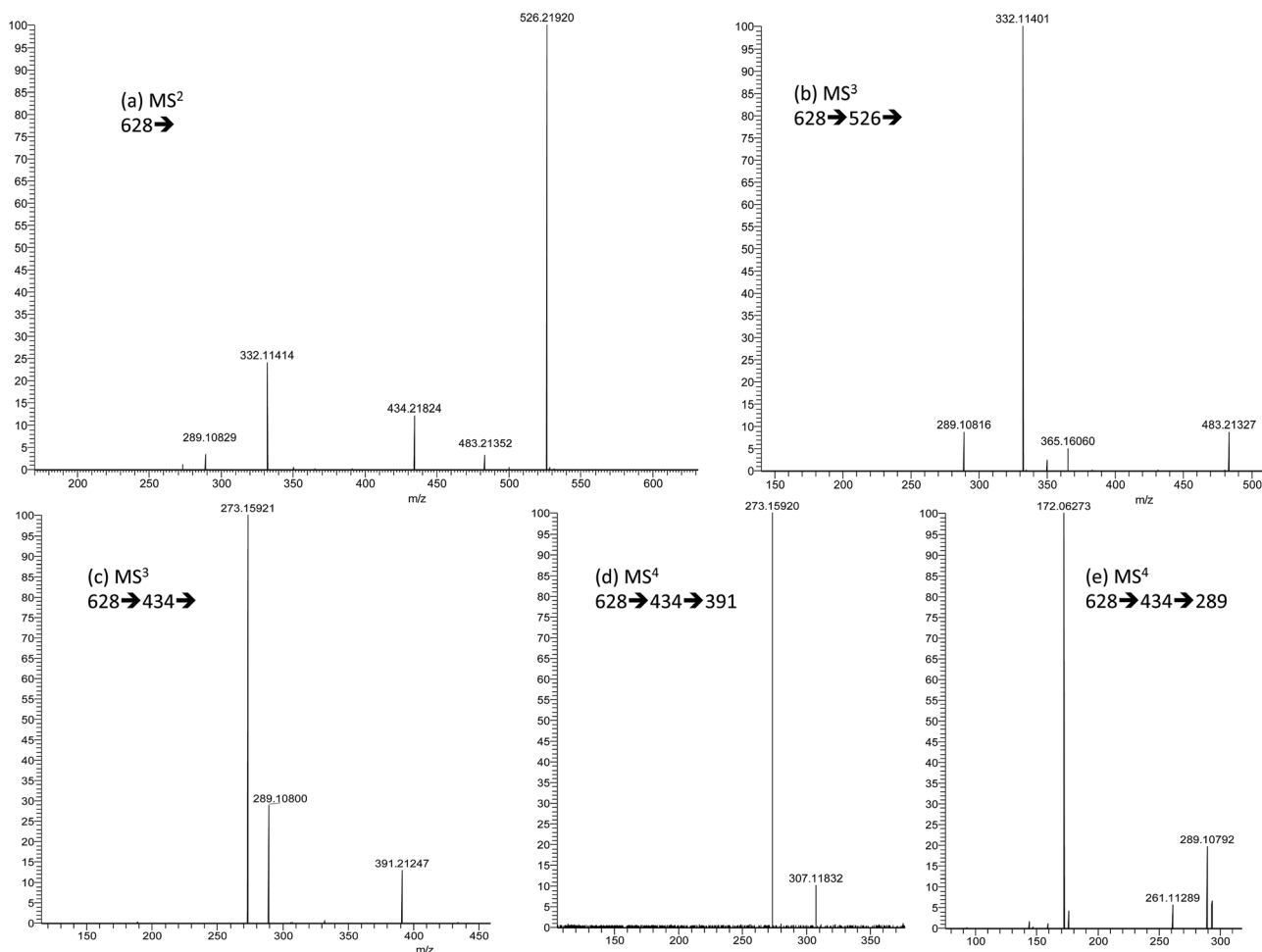


Fig. 2 HR-MS spectra of protonated DABET.

As shown in Fig. 3, the formation of m/z 526 ion ($C_{28}H_{28}N_7O_4^+$) may be due to an elimination of hexan-1-ol from protonated DABET ion. However, considering the product ions formed from m/z 526 ion, it appears that the way that protonated DABET was fragmented would be closely dependent upon the protonation site of the molecule. Although there are several possibilities, the system formed from the protonation of the amine function would facilitate the elimination of hexan-1-ol (-102 Da) through a rearrangement process involving a six-member centre, leading to the formation of an intermediate amino-acylium ion with m/z of 526, as shown in Fig. 3. From there, an internal rearrangement involving migration of adjacent double bonds would take place so to obtain a stable carbocation. Taken in turn as precursor for HR-MS³ studies, m/z 526 ion could lose isocyanic acid to give m/z 483 ($C_{27}H_{27}N_6O_3^+$) carbocation, whose configuration, such as is proposed in Fig. 3, would allow the departure of a 118 Da moiety by heterolytic cleavage of the C–N bond along with formation of a π -bond between N and C. This neutral loss would correspond to the departure of 4-(iminomethylene)cyclohexa-2,5-dienimine or equivalent, generating m/z 365 ion ($C_{20}H_{21}N_4O_3^+$). Thereafter, the fragmentations that would

involve the ester and amide functions were highlighted by the detection of m/z 337 ion ($C_{18}H_{17}N_4O_3^+$) and of m/z 265 ion ($C_{15}H_{13}N_4O^+$), formed by loss of ethylene and of ethyl acrylate, respectively. Parallel to this series of fragmentation, protonation of *O*-carbamimide would have facilitated the loss of ethyl 3-(pyridin-2-ylamino)propanoate (-194 Da), leading to the formation of m/z 332 ion ($C_{18}H_{14}N_5O_2^+$). At MS⁴, m/z 332 ion would lose isocyanic acid to afford m/z 289 ion ($C_{17}H_{13}N_4O^+$). Taken in turn as precursor to go further in the fragmentation study, thereof could be transformed into m/z 261 ion ($C_{16}H_{13}N_4^+$) by loss of CO. The ion at m/z 289 could also give way to m/z 172 radical dynconic ion ($C_{10}H_8N_2O^{*+}$) by homolytic cleavage leading to the departure of radical 4-(iminomethylene)cyclohexa-2,5-dienimine (-117 Da).

Parallel to loss of hexan-1-ol, protonated DABET ion could undergo direct loss of ethyl 3-(pyridin-2-ylamino)propanoate to yield m/z 434 ion ($C_{24}H_{28}N_5O_3^+$), according to a rearrangement process involving the proton carried by *O*-carbamimide (Table 3 and Fig. 3). Taken as a precursor for HR-MS³ studies, m/z 434 ion resulted in the formation of three intense product ions with m/z of 391, 289 and 273, while the hexane-1-ol loss seemed to be more difficult, given the very low intensity of the

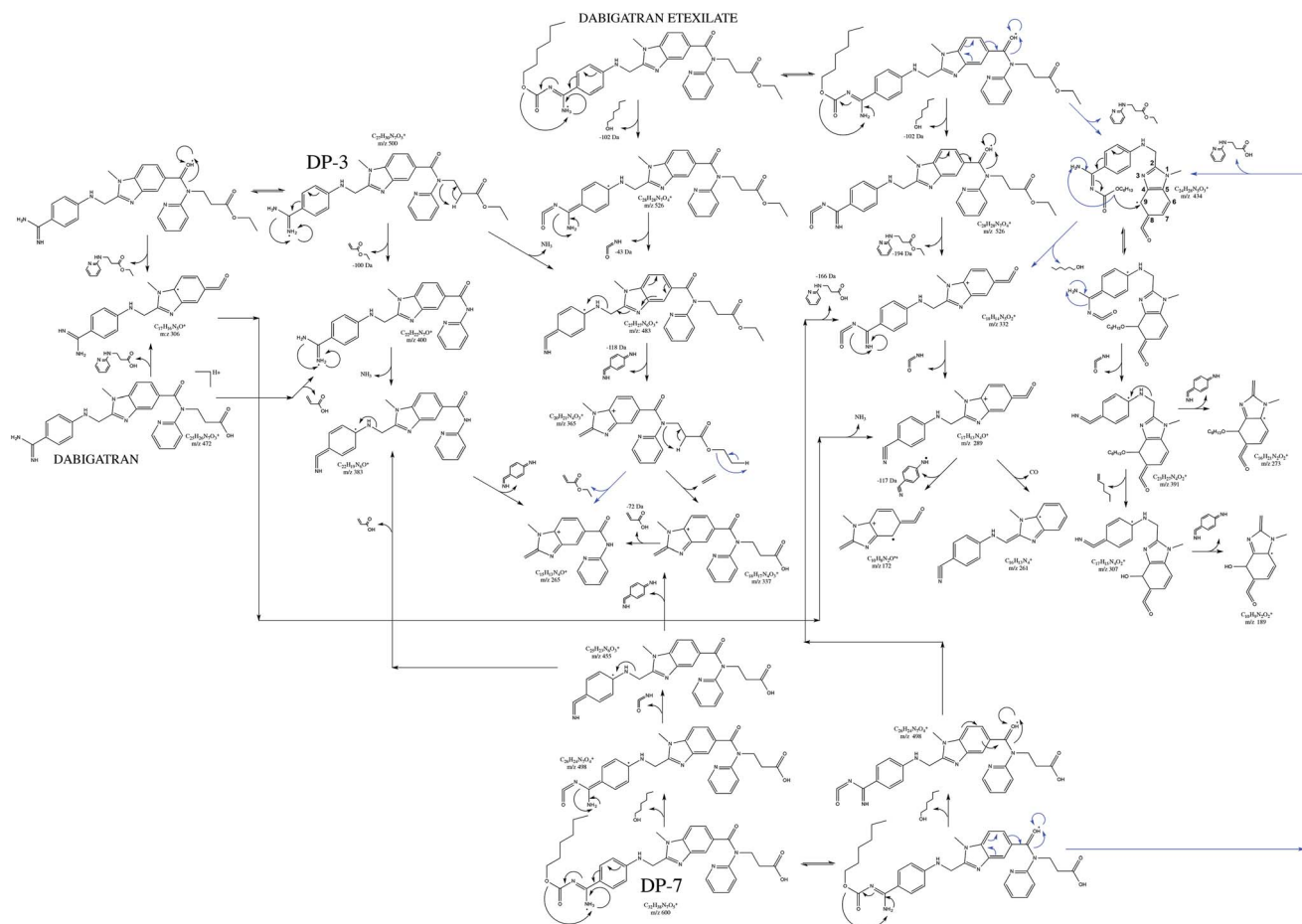


Fig. 3 Proposed fragmentation patterns of protonated DABET, dabigatran, DP-3 and DP-7.

resulting ion, supposed to have a mass-to-charge of 332 (Fig. 2). However, the surprising element is tied to the direct elimination of isocyanic acid to generate m/z 391 ion ($C_{23}H_{27}N_4O_2^+$), knowing that such a loss could logically occur only after elimination of hexan-1-ol. Therefore, the only plausible explanation has been to consider the prior migration of hexanolate towards another site of the structure, favoured by the configuration of the product ion. So the premise proposed here would be that the elimination of the 194 Da moiety would have resulted in rearrangements involving the successive migration of the adjacent double bonds, hence leading to a configuration where C_9 had become electron-deficient. C_9 would have then undergone nucleophile attack materialized by the transfer of hexan-1-ol to form an etheroxyde function, *via* a rearrangement mechanism implying a twelve-member centre, such was proposed in Fig. 3. This assumption was not meaningless insofar as the product ion at m/z 273 ($C_{16}H_{21}N_2O_2^+$), resulting from the loss of 4-(iminomethylene)cyclohexa-2,5-dienimine group from m/z 391 ion, would still have conserved the hexan-1-ol moiety. Parallel to the path leading to m/z 273 ion, m/z 391 ion could also successively lose hex-1-ene and 4-(iminomethylene)

cyclohexa-2,5-dienimine to generate m/z 307 ion ($C_{17}H_{15}N_4O_2^+$) and m/z 189 ion ($C_{10}H_9N_2O_2^+$), respectively.

Identification of the degradation products by LC-HR-MSⁿ

DP-1. DP-1 degradation product was the first eluted. The accurate mass measured (m/z 517.15679) was consistent with elemental formula $C_{25}H_{23}N_6Na_2O_4^+$ (relative error: -0.54 ppm). The corresponding ion would then account for $[DP_1-H + 2Na]^+$ ion. The MS spectrum acquired at the relative retention time of 0.5 also included a weakly intense peak at m/z 473.19285, which could be attributed to the protonated $[DP_1 + H]^+$ ion, as the accurate mass value is consistent with elemental formula $C_{25}H_{24}N_6O_4^+$ (Table 2). Because of the low information level obtained for the protonated ion, the fragmentation studies have been performed on the sodium-adduct ion, in order to highlight information useful to the identification of the degradation product. The HR-MS² spectrum of $[DP_1-H + 2Na]^+$ ion is mainly characterized by the presence of three intense peaks with m/z of 445, 382 and 369, which, in all likelihood, should correspond to the product ions having as elemental formulae $C_{22}H_{15}N_4Na_2O_2^+$, $C_{18}H_{16}N_4Na_2O_3^+$ and $C_{17}H_{15}N_4Na_2O_3^+$, respectively (Fig. 4). Similarly to what has been already described for DABET protonated ion, m/z 445 ion

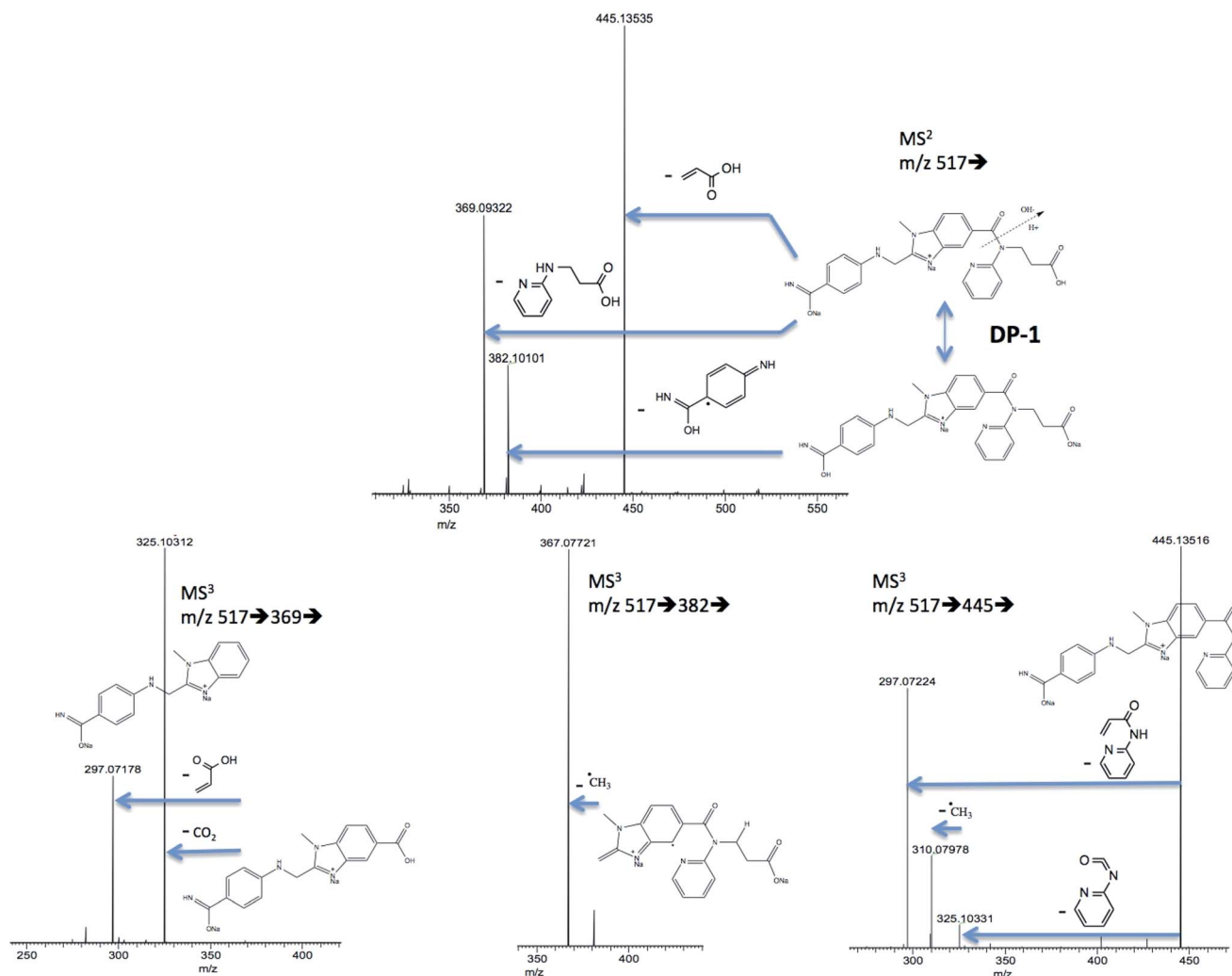


Fig. 4 Proposed fragmentation pattern of $[\text{DP}_1\text{-H} + 2\text{Na}]^+$.

would be produced by loss of acrylic acid, according to *N*-dealkylation process. When taken as precursor for MS^3 studies, m/z 445 could lose 2-isocyanatopyridine, to result m/z 325 ion ($\text{C}_{16}\text{H}_{15}\text{N}_4\text{Na}_2\text{O}^+$). From there, an elimination of a radical methyl was formulated to try to explain the presence of m/z 310 dystonic ion ($\text{C}_{15}\text{H}_{12}\text{N}_4\text{Na}_2\text{O}^+$). In parallel, the presence of m/z 297 could be attributed to the departure of *N*-(pyridin-2-yl) acrylamide ($\text{C}_{14}\text{H}_{11}\text{N}_4\text{Na}_2\text{O}^+$). Due to the nature of the losses observed, it was possible to locate sites that had interacted with the sodium ions.

Apart from the fragmentation in m/z 445 ion, $[\text{DP}_1\text{-H} + 2\text{Na}]^+$ ion could also generated m/z 369 ion by loss 3-(pyridin-2-ylamino)propanoic acid (-166 Da). This seems to be due to hydrolysis of the amide bond, by involvement of a water molecule. Subjected to further stage of fragmentation, m/z 369 ion lost CO_2 to yield m/z 325 ion and this helped confirm the previous assumption. Transition $517 \rightarrow 382$ means the possible loss of 4-iminocyclohexa-2,5-dienecarboxamide radical group by homolytic cleavage of the N-C bond. The

next fragmentation stage leads to the formation of m/z 367 ion ($\text{C}_{17}\text{H}_{13}\text{N}_4\text{Na}_2\text{O}_3^+$), by possible loss of radical $\cdot\text{CH}_3$.

These data seem to be consistent with the structure of 3-(2-((4-(hydroxy(imino)methyl)phenylamino)methyl)-1-methyl-*N*-(pyridin-2-yl)-1*H*-benzo[*d*]imidazole-5-carboxamido)propanoic acid.

DP-2. DP-2 yielded a protonated ion with an accurate mass of 381.15568, consistent with elemental formula $\text{C}_{20}\text{H}_{21}\text{N}_4\text{O}_4^+$ (relative error: -0.13 ppm). Due to strong similarities that exist between the protonated DP-2 MS^2 spectrum and that of one of the MS^3 product ion related to protonated DP-5, the data that pertain to DP-2 are therefore discussed below in the part devoted to DP-5 (Table 2). They show that DP-2 may be ethyl 3-(*N*-(pyridin-2-yl)-1,3-dihydrobenzo[*d*]oxazolo[3,4-*a*]imidazole-6-carboxamido)propanoate (Table 2 and Fig. 5 and 6).

DP-3. DP-3 gave a protonated ion with an accurate mass of 500.23944, consistent with elemental formula $\text{C}_{27}\text{H}_{30}\text{N}_7\text{O}_3^+$ (relative error: -2.04 ppm). Compared to that of DABET, the gap should correspond to a loss equivalent to 7C, 12H and 2O

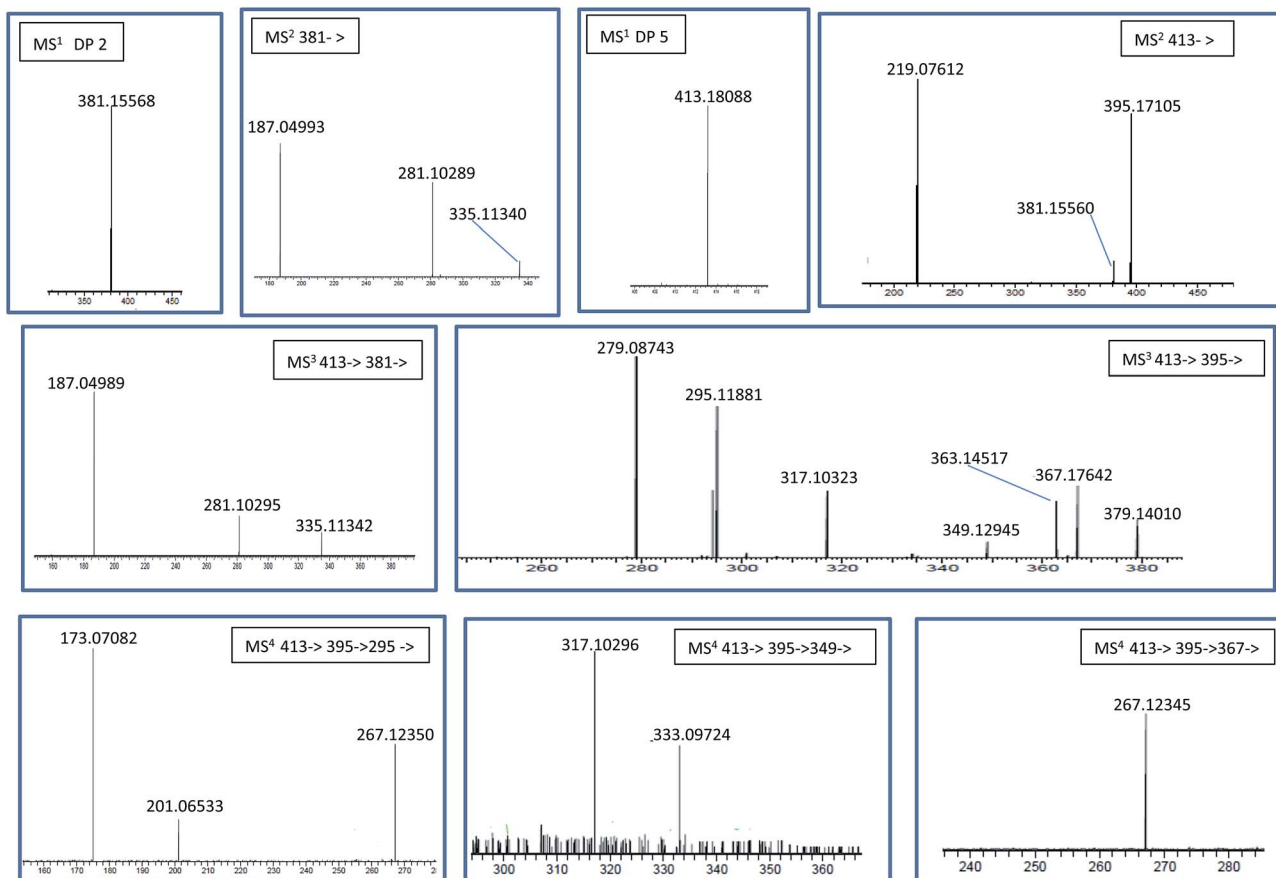


Fig. 5 HR-MS spectra of protonated DP-2 and DP-5.

(Tables 2 and 3). Moreover, the transitions that have been attributed to the departure of hexanol (-102 Da) or of isocyanic acid (-43 Da) are totally absent (Tables 2 and 3, Fig. 3). As a result, DP-3 could be considered as may be ethyl 3-(2-((4-carbamimidoylphenylamino)methyl)-1-methyl-*N*-(pyridin-2-yl)-1*H*-benzo[*d*]imidazole-5-carboxamido)propanoate.

DP-4. DP-4 gave a protonated ion with m/z of 501.22358, consistent with elemental formula $C_{27}H_{29}N_6O_4^+$ (relative error: -1.80 ppm). By drawing a parallel with that of DP-3, it appears that the difference is due to the substitution of NH_2 by OH (Table 2). During the fragmentation process, DP-4 can give rise, on the one hand, to the formation of the ions common to that of DP-3 and on the other hand, to that of the ions having one mass unit greater than their NH_2 counterparts (Table 2). Protonated DP-4 can undergo deamination giving birth to m/z 484 ion ($C_{27}H_{26}N_5O_4^+$) instead of m/z 483 ion ($C_{27}H_{27}N_6O_3^+$) that has been previously reported for protonated DP-3. Loss of ethyl acrylate was highlighted by transition $501 \rightarrow 401$, all as transition $500 \rightarrow 400$ for DP-3 (Fig. 3 and 7). After loss of ethyl 3-(pyridin-2-ylamino)propanoate (-194 Da), loss of water was observed, which has allowed, at this point, to say that DP-4 rather carries a benzimidic acid function. In the same way, loss of (4-iminocyclohexa-2,5-dienylidene)methanone (-119 Da) from m/z 484 ion to give m/z 365 ion, has confirmed the assumption inherent in this type of change (Table 2 and

Fig. 7). Considering these results, DP-4 could be 4-((5-((3-ethoxy-3-oxopropyl)(pyridin-2-yl)carbamoyl)-1-methyl-1*H*-benzo[*d*]imidazol-2-yl)methylamino)benzimidic acid.

DP-5. Protonated DP-5 has an accurate mass-to-charge value of 413.18088, particularly well correlated with elemental formula $C_{21}H_{25}N_4O_5^+$ (relative error: -2.59 ppm). Subjected to MS^2 studies (Table 2 and Fig. 5 and 6), m/z 413 ion yielded three main product ions at m/z 395 ($C_{21}H_{23}N_4O_4^+$), m/z 381 ($C_{20}H_{21}N_4O_4^+$) and m/z 219 ($C_{11}H_{11}N_2O_3^+$). The other fragments turn out to have been secondarily formed, as discussed later. The peak at m/z 219 accounts for the base peak. Transition $413 \rightarrow 219$ would be the reflect of the neutral loss previously attributed to ethyl 3-(pyridin-2-ylamino)propanoate. This result has allowed deducing that this part of the structure had not been impacted during the degradation having led to the formation of DP-5. When taken as precursor for MS^3 studies, m/z 219 ion produced two other ions with m/z of 201 and 187 by loss of water and of methanol, respectively, which is in line with the hypothesis for its structure. The ion at m/z 159 would be, as for it, secondarily formed from m/z 187 ion, by loss of CO (Table 2).

The ions that were produced either by dehydration (m/z 395) or by loss of methanol (m/z 381) have been fragmented (Table 2 and Fig. 5). But the usual losses related to the departure of isocyanic acid (-43 Da) or of 4-(iminomethylene)

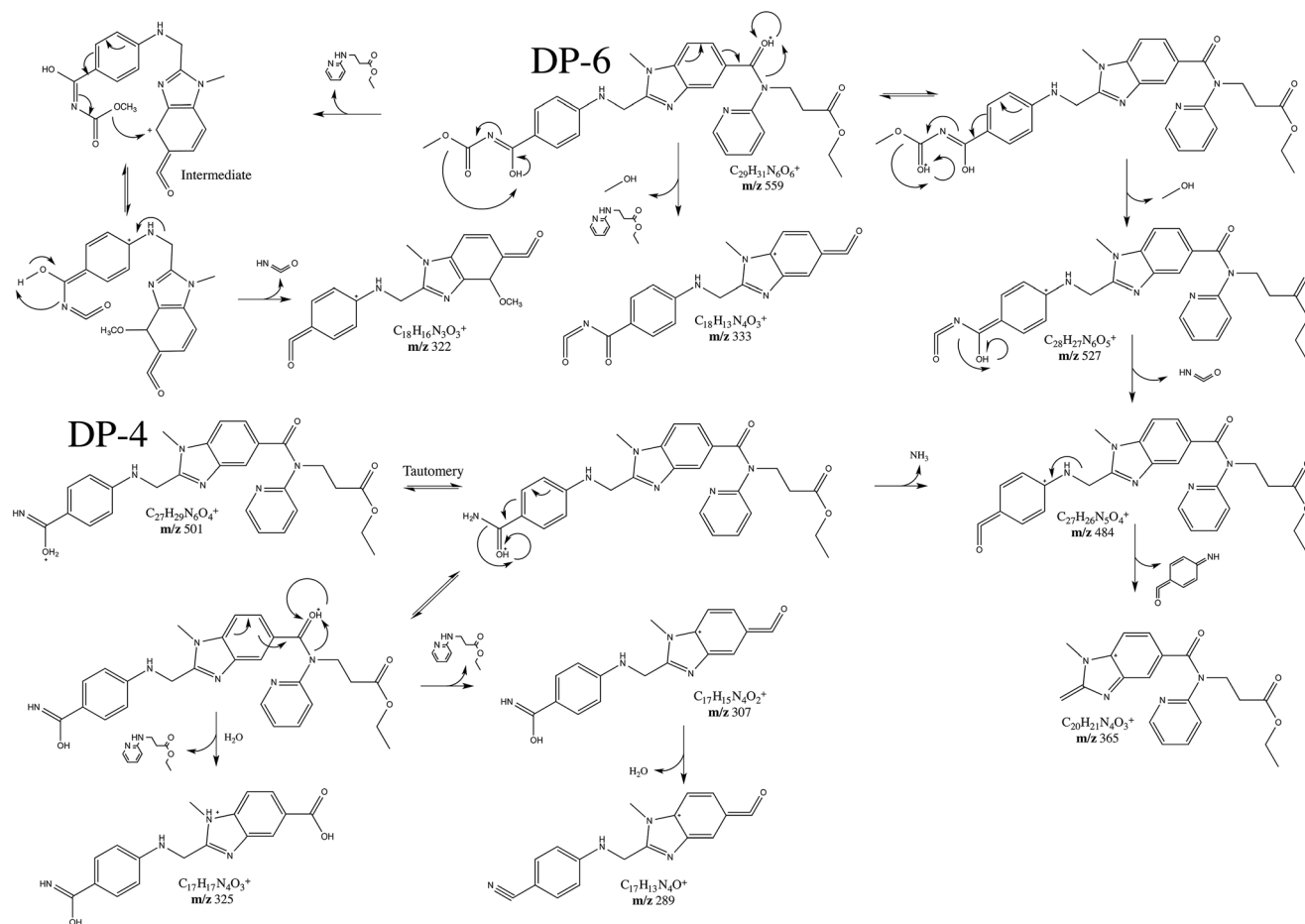


Fig. 7 Proposed fragmentation patterns of protonated DP-4 and DP-6.

part of the molecule that was the cause, has remained identical to that of DABET. Similarly as what has been described above regarding the intra-molecular transfer mechanism of a chemical group from one site to another during the fragmentation process, formation of m/z 322 ion is an additional example that justifies the assumption previously posed (Fig. 3 and 7).

Taken together, these data have allowed proposing the derivative 4-((5-((3-ethoxy-3-oxopropyl)(pyridin-2-yl)carbamoyl)-1-methyl-1*H*-benzo[*d*]imidazol-2-yl)methylamino)-*N*-(methoxycarbonyl)benzimidic acid as may correspond to DP-6.

DP-7. DP-7 gave a protonated ion with an accurate mass of 600.29180, consistent with elemental formula $C_{32}H_{38}N_7O_5^+$ (relative error: -1.82 ppm). A gap of 28 Da with respect to that of protonated DABET, should in all likelihood, be due to hydrolysis of the ester function. As shown in Table 2, the HR-MS² spectrum of protonated DP-7 comprises a lot of common ions to that of protonated DABET. Absence of the transition that reflects the departure of ethylene group has been noted though, which is in line with the above statement (Fig. 3). As a result, DP-7 can be identified as 3-(2-((4-(*N*'-(hexyloxycarbonyl)carbamimidoyl)phenylamino)methyl)-1-methyl-*N*-(pyridin-2-yl)-1*H*-benzimidazole-5-carboxamido)propanoic acid.

DP-8 and DP-9. All like the parallel drawn between DP-3 and DP-4, the same thing has been observed between DP-8

and DP-9 regarding their elemental compositions and respective fragmentation patterns (Table 2 and Fig. 8). Protonated DP-8 is characterized by an accurate mass of m/z 614.30705, while protonated DP-9, by m/z 615.29137, consistent with elemental formulae $C_{33}H_{40}N_7O_5^+$ (relative error: -2.43 ppm) and $C_{33}H_{39}N_6O_6^+$ (relative error: -1.93 ppm), respectively, thus showing that DP-9 carries an imidic acid function instead of the aminimido group carried by DP-8 (Table 2). But as shown in Fig. 8, they both lost methyl 3-(pyridin-2-ylamino)propanoate (-180 Da) rather than ethyl 3-(pyridin-2-ylamino)propanoate (-194 Da). This information has allowed identifying them as methyl propanoate derivatives and not the ethyl propanoate ones as is the case of DABET. This is in line with loss of methylacrylate as show transitions $469 \rightarrow 383$ and $470 \rightarrow 384$, pertaining to DP-8 and DP-9, respectively. For the rest, the fragmentation scheme of protonated DP-8 is comparable to that of protonated DABET, showing that DP-8 could be identified as 4-((5-((3-methoxy-3-oxopropyl)(pyridin-2-yl)carbamoyl)-1-methyl-1*H*-benzo[*d*]imidazol-2-yl)methylamino)-*N*-(hexyloxycarbonyl)benzimidic acid and consequently, DP-9 as *N*-(hexyloxycarbonyl)-4-((5-((3-methoxy-3-oxopropyl)(pyridin-2-yl)carbamoyl)-1-methyl-1*H*-benzo[*d*]imidazol-2-yl)methylamino)benzimidic acid.

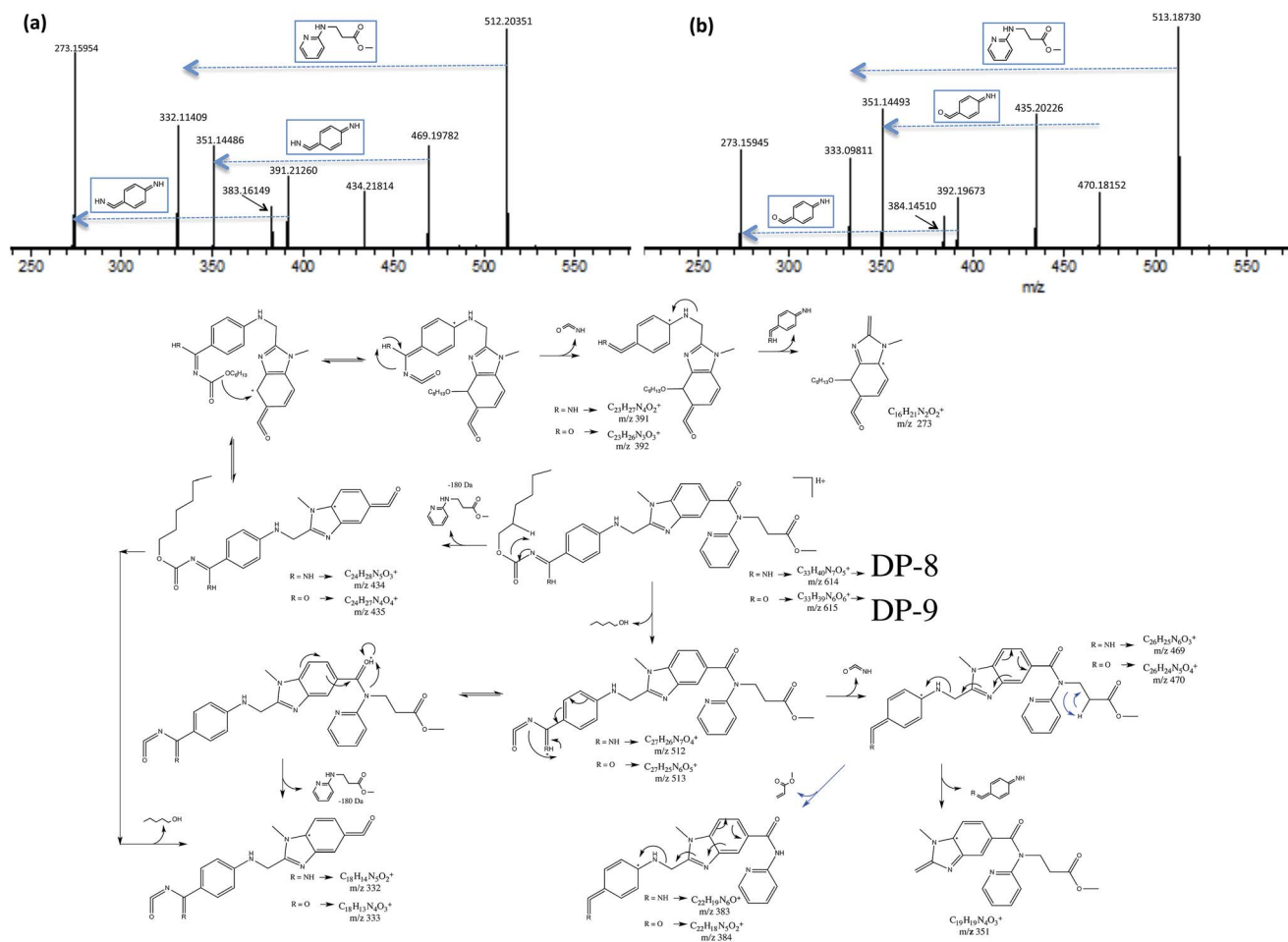


Fig. 8 Proposed fragmentation patterns of protonated DP-8 and DP-9.

DP-10. Similarly to what was drawn for DP-3/DP-4 and DP-8/DP-9, compared to DABET, DP-10 should be its OH counterpart (Table 2). As a result, DP-10 may correspond to 4-((5-((3-ethoxy-3-oxopropyl)(pyridin-2-yl)carbamoyl)-1-methyl-1H-benzo[d]imidazol-2-yl)methylamino)-N-(hexyloxy-carbonyl)benzimidic acid.

Proposed degradation pathways of dapixaban etexilate

Under influence of various stress conditions, DABET was degraded as per several degradation routes producing ten major degradation products in addition to dabigatran (Tables 1 and 2). Thereof turns out to be the compound, whose DABET is the pro-drug. The ten unknowns have been successfully characterized by use of multistage high-resolution mass spectrometry. On the whole, DABET was shown susceptible to hydrolysis, involving the ester function as well as the carbamimido group. *O*-Dealkylation may occur and formation of benzimidic acid derivatives was also observed. The schematic representations of mechanism of formation of the degradation products under hydrolytic stress (DP-1, DP-4, DP-6, DP-7, DP-8, DP-9 and DP-10) are shown in Fig. 9.

Over the other tested conditions, degradations by photo-catalysis and that in the presence of H_2O_2 were less significant. Nevertheless, DP-2, DP-3 and DP-5 were still produced, along the degradation pathways proposed in Fig. 10. In solution, photo-catalytic conditions may put into play a certain number of reactions that would be radical initiated. A number of reactions that followed this initiation stage may be rather of molecular nature, involving nucleophile attacks. Reaching a certain excited state, DABET could undergo auto-oxidation by radical initiation, which resulted in the abstraction of a proton from the α -carbon linked to the arylamine-nitrogen.^{20,21} The radical reaction was prolonged by reaction with O_2 to generate peroxide radical and then peroxide, by abstraction of a proton from solvent. From there, intermediate etheroxide DP-2 would have been formed by recombination mechanisms in the presence of water, such as stipulated in Fig. 10. A photo-catalytic methanolysis reaction has been proposed to explain the formation of DP-5. DP-2 was also found in oxidative condition in the presence of hydrogen peroxide, likely due to attack on the α -carbon linked to the arylamine-nitrogen, as shown in Fig. 10.

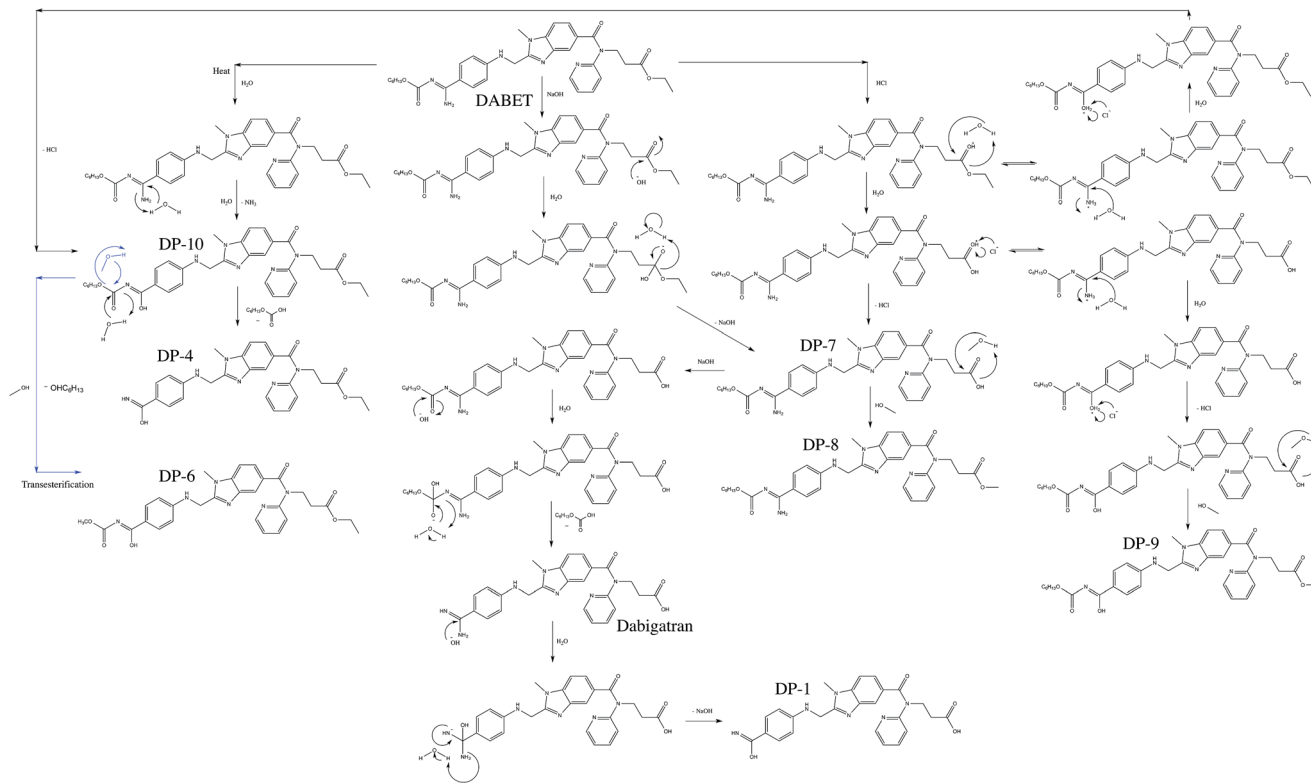


Fig. 9 Proposed degradation pathways of DABET under thermal and hydrolytic stress conditions.

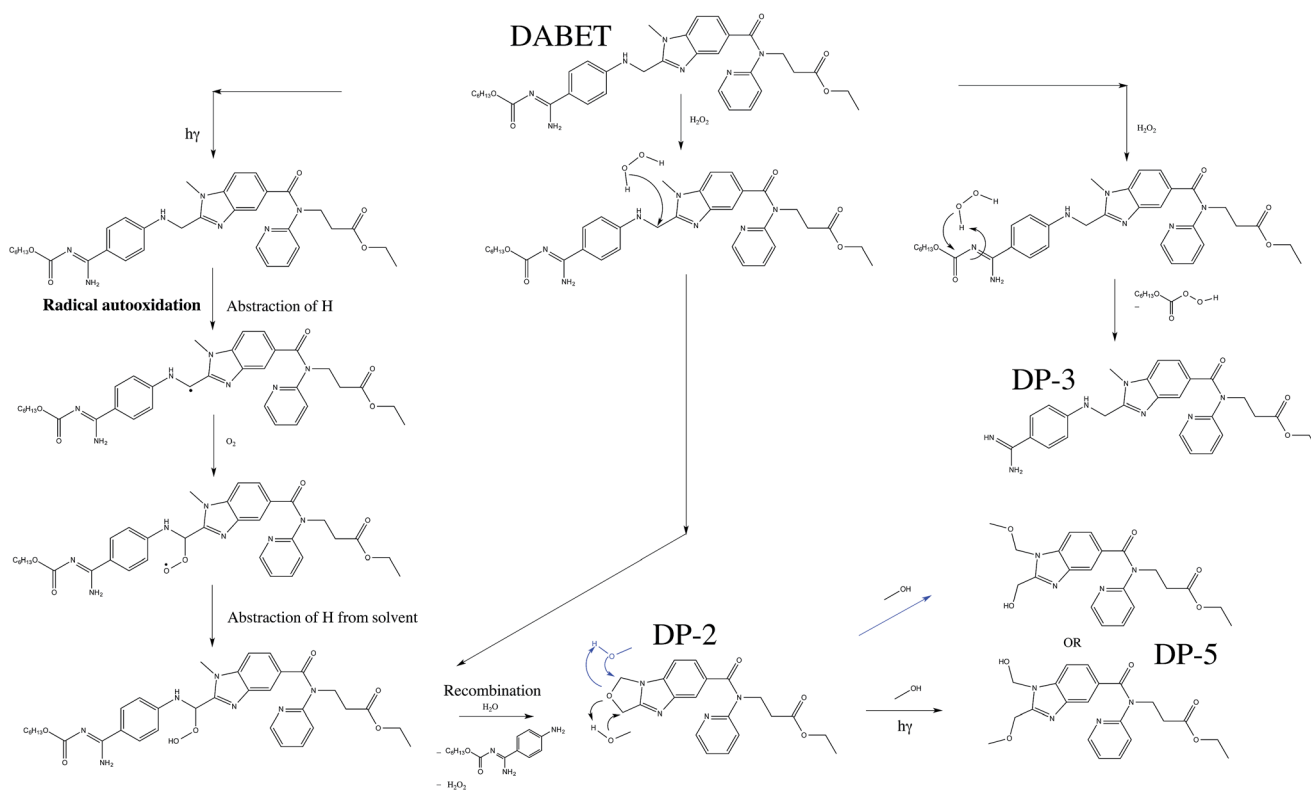


Fig. 10 Proposed degradation pathways of DABET under photolysis and oxidative stress conditions.

Conclusion

The degradation behaviour of DABET under hydrolytic (acid, base), oxidative, photolytic and thermal stress conditions was studied. Its MSⁿ fragmentation scheme was studied in depth in order to help assign, by comparison, the structures of the product ions formed from the degradation products. A total of ten degradation products were formed under stress conditions and characterized. Under hydrolytic stress conditions, O-dealkylation may occur and formation of benzimidic acid derivatives was also observed. DABET was shown much less susceptible to photolysis and oxidative stress, even if N-dealkylation was highlighted through the identification of the degradation products. Based on identification, it was possible to deduct major degradation mechanisms in the context of stress testing.

References

- 1 C. T. January, L. S. Wann, J. S. Alpert, H. Calkins, J. E. Cigarroa, J. C. Cleveland, J. B. Conti, P. T. Ellinor, M. D. Ezekowitz, M. E. Field, K. T. Murray, R. L. Sacco, W. G. Stevenson, P. J. Tchou, C. M. Tracy and C. W. Yancy, *J. Am. Coll. Cardiol.*, 2014, **64**, 2246.
- 2 J. W. M. Cheng and H. Vu, *Clin. Ther.*, 2012, **34**, 766.
- 3 C. S. Miller, S. M. Grandi, A. Shimony, K. B. Filion and M. J. Eisenberg, *Am. J. Cardiol.*, 2012, **110**, 453.
- 4 M. Ganetsky, K. M. Babu, S. D. Salhanick, R. S. Brown and E. W. Boyer, *J. Med. Toxicol.*, 2011, **7**, 281.
- 5 S. W. Baertschi, K. M. Alsante and R. A. Reed, in *Pharmaceutical stress testing: predicting drug degradation*, Informa Healthcare, New York, 2nd edn, 2011, ch. 2, pp. 10–49.
- 6 ICH Q1A(R2), *International Conference on Harmonization*, ICPMA, Geneva, 2003.
- 7 S. Singh, T. Handa, M. Narayanam, A. Sahu, M. Junwal and R. P. Shah, *J. Pharm. Biomed. Anal.*, 2012, **69**, 148.
- 8 D. Jain and P. K. Basniwal, *J. Pharm. Biomed. Anal.*, 2013, **86**, 11.
- 9 E. Nägele and R. Moritz, *J. Am. Soc. Mass Spectrom.*, 2005, **16**, 1670.
- 10 H. Sadou Yaye, P. H. Secrétan, T. Henriët, M. Bernard, F. Amrani, W. Akrouf, P. Tilleul, N. Yagoubi and B. Do, *J. Pharm. Biomed. Anal.*, 2015, **105**, 74.
- 11 I. Gana, A. Dugay, T. Henriët, I. B. Rietveld, M. Bernard, C. Guechot, J.-M. Teulon, F. Safta, N. Yagoubi, R. Céolin and B. Do, *J. Pharm. Biomed. Anal.*, 2014, **96**, 58.
- 12 R. P. Shah, V. Kumar and S. Singh, *Rapid Commun. Mass Spectrom.*, 2008, **22**, 613.
- 13 J. Li, J. Fang, F. Zhong, W. Li, Y. Tang, Y. Xu, S. Mao and G. Fan, *J. Chromatogr. B: Anal. Technol. Biomed. Life Sci.*, 2014, **973**, 110.
- 14 J. P. Antovic, M. Skeppholm, J. Eintrei, E. E. Boija, L. Söderblom, E.-M. Norberg, L. Onelöv, Y. Rönquist-Nii, A. Pohanka, O. Beck, P. Hjemdahl and R. E. Malmström, *Eur. J. Clin. Pharmacol.*, 2013, **69**, 1875.
- 15 M. Korostelev, K. Bihan, L. Ferreol, N. Tissot, J.-S. Hulot, C. Funck-Brentano and N. Zahr, *J. Pharm. Biomed. Anal.*, 2014, **100**, 230.
- 16 X. Delavenne, J. Moracchini, S. Laporte, P. Mismetti and T. Basset, *J. Pharm. Biomed. Anal.*, 2012, **58**, 152.
- 17 Y.-Y. Zheng, C.-W. Shen, M.-Y. Zhu, Y.-M. Zhou and J.-Q. Li, *Org. Process Res. Dev.*, 2014, **18**, 744.
- 18 Z.-Y. Hu, R. B. Parker, V. L. Herring and S. C. Laizure, *Anal. Bioanal. Chem.*, 2013, **405**, 1695.
- 19 M. Blessy, R. D. Patel, P. N. Prajapati and Y. K. Agrawal, *J. Pharm. Anal.*, 2014, **4**, 165.
- 20 G. A. Russe, *J. Am. Chem. Soc.*, 1957, **79**, 3871.
- 21 F. Minisci and F. Fontana, *La Chimica e l'Industria*, 1998, **80**, pp. 1309.