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Article

Antimicrobial Activity and In Silico Molecular Docking Studies of Pentacyclic Spiro[oxindole-2,3'-pyrrolidines] Tethered with Succinimide Scaffolds

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Abstract: Three-component cascade reactions of (*E*)-3-arylidene-1-methyl-pyrrolidine-2,5-diones, *L*-valine and various isatin derivatives are described. A series of 17 spiropyrrolidine derivatives with wide structural complexity and diversity have been thus obtained in moderate to excellent yields under mild reaction conditions. The structure and stereochemistry of these *N*-heterocyclic cycloadducts has been established by spectroscopic techniques and unambiguously confirmed by a single-crystal X-ray diffraction analysis performed on one derivative. UV-visible spectra have been recorded for all new compounds. Furthermore, the synthesized *N*-heterocyclic compounds have been screened for their in vitro antibacterial and antifungal activities. Several derivatives exhibited moderate to good activities, comparable to those of the known standard drugs Amphotericin B and Tetracycline. Structural activity relationships (SARs) and molecular docking of the most promising derivatives into the binding sites of glucosamine 6-phosphate synthase (GlcN6P) and methionyl-trna-synthetase (1PFV) were also established. Furthermore, pharmacokinetic studies indicate that the heterocycles exhibit acceptable predictive ADMET (absorption, distribution, metabolism, excretion, and toxicity) properties and good drug ability.

Keywords: spirooxindolepyrrolidines; multicomponent reaction; 1,3-dipolar cycloaddition; antimicrobial activity; molecular docking analysis



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1. Introduction

Spirocyclic oxindoles, which are widely distributed in natural alkaloids and bioactive compounds, also represent attractive synthetic targets [1–7]. As a subset of this family, spiro[oxindole-2,3'-pyrrolidine] constitute a privileged structural unit which is present in numerous natural isolates and synthetic bioactive compounds [8,9]. They exhibit a wide spectrum of biological properties, such as antitumoral [10,11], anti-inflammatory [12], antimycobacterial [13], antimicrobial and anticancer activities (Figure 1) [14–17]. Furthermore, they are also known for acetylcholinesterase inhibition [18,19].

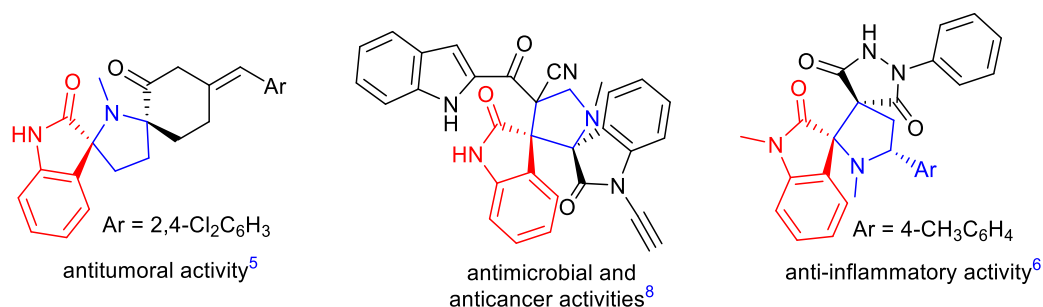
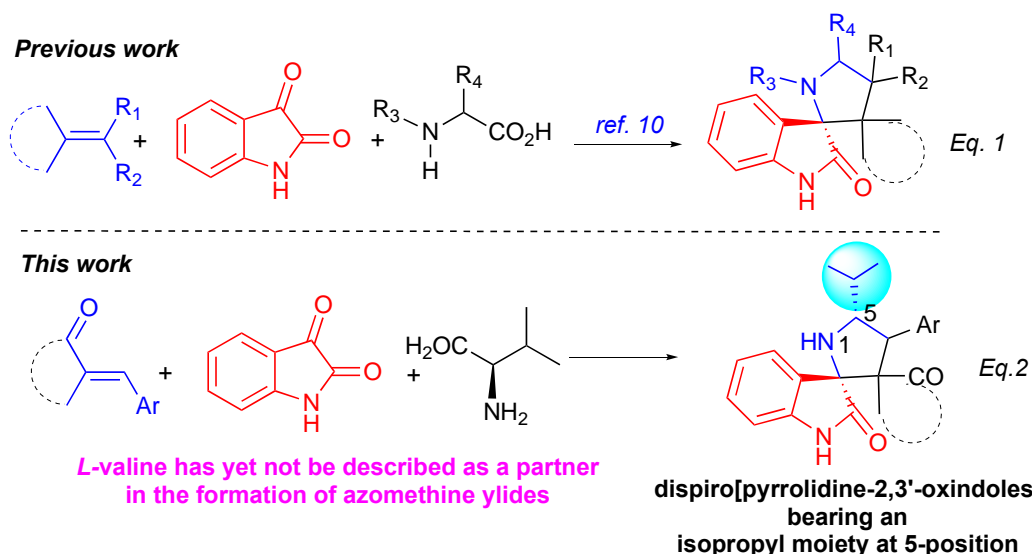


Figure 1. Examples of biologically active compounds containing spiro[oxindole-2,3'-pyrrolidine] units.

Among the current methodologies for the synthesis of spiro[oxindole-2,3'-pyrrolidine] derivatives, multicomponent (3+2) cycloaddition reactions (MCRs) of electron-deficient exocyclic alkenes with azomethine ylides, generated in situ by decarboxylative condensation of isatin derivatives with α -amino acids, have emerged as one of the most powerful tools (Scheme 1, eq. 1) [20–27]. However, a thorough literature survey of the decarboxylative process reveals that *L*-valine received only little attention as a component in the formation of azomethine ylides (Scheme 1, eq. 2). Considering this challenge, we were intrigued as to whether α -arylidene succinimides could be employed as dipolarophiles to undergo MCRs with azomethine ylides derived from *L*-valine and isatin derivatives, thus yielding dispiro[pyrrolidine-2,3'-oxindoles] bearing an isopropyl moiety at 5-position.



Scheme 1. MCR as tool for the synthesis of pentacyclic spiro[oxindole-2,3'-pyrrolidines].

Furthermore, our targeted formal (3+2) cycloaddition products **5** bear succinimide scaffolds. These latter five-membered (CH₂)₂(CO)₂NH or (CH₂)₂(CO)₂NR cycles constitute important skeletons in numerous alkaloids, including Hirsutellone A and B **7**, drug molecules and various other bioactive synthetic compounds. Some succinimide derivatives, for instance, are used as antimycobacterial agents for the treatment of mycobacteria infections [28,29] and for the suppression of cancer cell proliferation [30]. Several succinimides are furthermore used as anticonvulsant drugs, including ethosuximide (Zarontin), phensuximide, and methsuximide (Petinitin) [31,32]. Recently, we showed that some spirooxindolepyrrolidine and spirooxindolepyrrolizidine derivatives incorporating the pyrrolidine-2,5-dione motif are promising for acetylcholinesterase (AChE) inhibition [33] and possess both antimicrobial and anticoagulant activities [34].

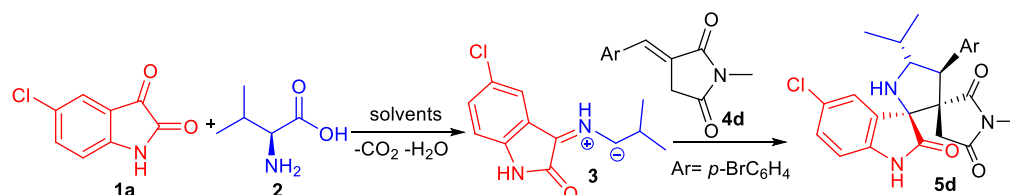
Herein, and in continuation of our research in the domain of *one-pot* MCRs of azomethine ylides [33–41], we report on the synthesis of various spiro[oxindole-2,3'-pyrrolidines]

tethered with succinimide derivatives. The latter were assembled by a *one-pot* three-component [3+2]-cycloaddition reaction of isatin itself or halogenated isatin derivatives **1**, *L*-valine **2** and (*E*)-3-arylidene-1-methyl-succinimides **4**. Note that isatin and its analogues also constitute an important class of organic compounds featuring biological and pharmacological properties and serve as building blocks for further transformation. To valorize the synthetic results, some selected heterocyclic compounds were screened *in vitro* to evaluate their antimicrobial activity and the structure-activity relationship (SAR) was supported by a molecular docking analysis.

2. Results and Discussion

2.1. Synthetic Chemistry

At the onset of our work, 5-chloroisatin **1a**, *L*-valine **2** and dipolarophile **4d** were probed as model substrates to optimize the reaction conditions leading to pentacyclic compound **5d** (Scheme 2). The effects of solvents and temperature on the reaction time and yield were examined and the results are summarized in Table 1.



Scheme 2. 1,3-Dipolar cycloaddition affording spirooxindole **5d**.

Table 1. Optimization of the reaction parameters for the synthesis of **5d**^a.

Entry	Solvent	T (°C)	Time (h)	Yield (%) ^b
1	PhCH ₃	110	24	69
2	CH ₃ CN	80	13	83
3	EtOH	78	12	65
4	MeOH	64	3	96
5	THF	70	36	20

^a The reaction of **1a** (0.5 mmol), **2** (0.6 mmol) and **4d** (0.5 mmol) carried out in various solvents (5.0 mL). ^b Yield of the isolated product after purification.

Compound **5d** was isolated only with a low yield when the reaction was carried out in tetrahydrofuran (Table 1, entry 5). However, in refluxing methanol, the 1,3-dipolar cycloaddition reaction proceeds smoothly affording the targeted spiro[oxindole-2,3'-pyrrolidine] **5d** with an excellent yield of 96% (Table 1, entry 4) as a racemic mixture.

Having once established the optimal conditions (Table 1, entry 4), we next examined the scope of the reaction in regard to diverse α -arylidene-succinimides **4**, as well as with isatin **1c** and various halogenated isatin derivatives **1a,b** (Scheme 3, Table 2).

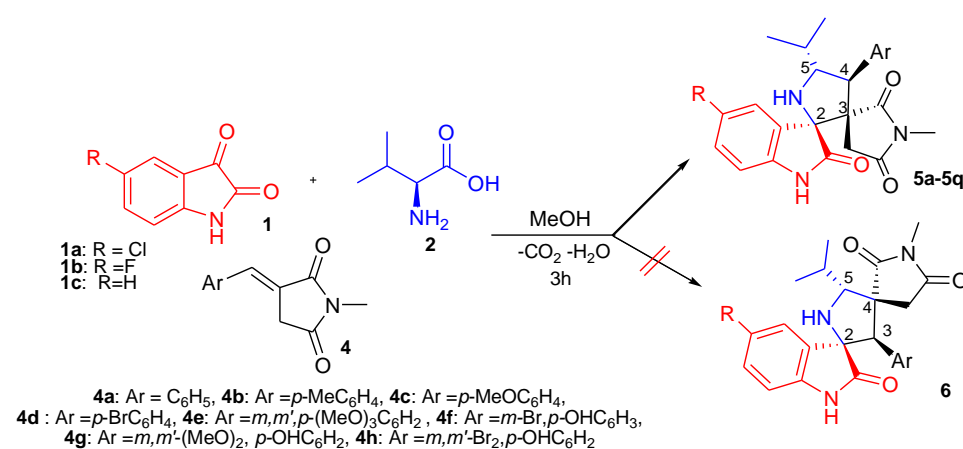
Table 2. Synthesis of the spirooxindole pyrrolidines **5**^a.

Entry	Compounds	R	Ar	Yield (%) ^b
1	5a	Cl	C ₆ H ₅	72
2	5b	Cl	<i>p</i> -MeC ₆ H ₄	66
3	5c	Cl	<i>p</i> -MeOC ₆ H ₄	86
4	5d	Cl	<i>p</i> -BrC ₆ H ₄	96
5	5e	Cl	<i>m,m',p</i> - (MeO) ₃ C ₆ H ₂	75
6	5f	Cl	<i>m-Br,p</i> -OHC ₆ H ₃	73
7	5g	Cl	<i>m,m'</i> -(MeO) ₂ , <i>p</i> - OHC ₆ H ₂	66
8	5h	Cl	<i>m,m'</i> -Br ₂ , <i>p</i> - OHC ₆ H ₂	70
9	5i	F	C ₆ H ₅	52
10	5j	F	<i>p</i> -Me C ₆ H ₄	53
11	5k	F	<i>p</i> -MeOC ₆ H ₄	55
12	5l	F	<i>p</i> -BrC ₆ H ₄	52
13	5m	F	<i>m,m',p</i> - (MeO) ₃ C ₆ H ₂	58
14	5n	F	<i>m-Br,p</i> -OHC ₆ H ₃	80
15	5o	F	<i>m,m'</i> -(MeO) ₂ , <i>p</i> - OHC ₆ H ₂	56
16	5p	F	<i>m,m'</i> -Br ₂ , <i>p</i> - OHC ₆ H ₂	51
17	5q	H	<i>p</i> -MeOC ₆ H ₄	83

^a Reactions were carried out with **1** (1 mmol), **2** (1.2 mmol) and **4** (1 mmol) in methanol (10 mL) at 64 °C for 3 h.

^b Isolated yield after purification by column chromatography.

The electronic properties and position of the substituent on the aromatic cycle of dipolarophiles **4** had no substantial influence on the efficiency of this reaction. As summarized in Table 2, arylidene-succinimides **4** bearing an electron-neutral (H), electron-donating (e.g., 4-Me, 3-OMe, 4-OMe or 4-OH) groups or electron-withdrawing substituent (4-Br, 3-Br) reacted smoothly to afford the desired spiro[oxindole-2,3'-pyrrolidine] derivatives **5a–5q** in good to excellent yields. In addition, 5-fluoroisatin **1c** was a suitable substrate under standard conditions, although compounds **5i–5p** were obtained with somewhat lower yields (Table 2, entries 9–16). No spectroscopic evidence for a competing formation of isomeric compounds **6** bearing the succinimide ring at 4-position (Scheme 3).

**Scheme 3.** 1,3-Dipolar cycloaddition route for the synthesis of spirooxindolepyrrolidines **5**.

2.2. Spectroscopic and Crystallographic Characterization of the Cycloadducts **5**

The structure and the relative configuration of the spiro-adducts **5** was deduced by NMR spectroscopy. As a representative example, **5b** has been chosen and selected ^1H and ^{13}C chemical shifts along with some characteristic coupling constants are shown in Figure 2.

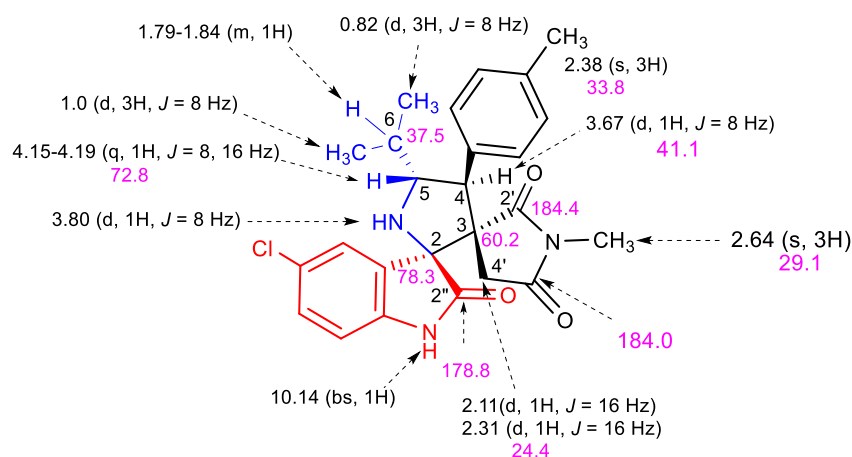


Figure 2. ^1H (black) chemical shifts with coupling constants and ^{13}C NMR (pink) data (ppm) of **5b**.

The ^1H NMR spectrum of **5b** shows a singlet at δ 2.64 ppm due to the *N*-methyl protons. Two mutually coupled doublets are observed at δ 2.10 and 2.31 ppm ($J = 16$ Hz) corresponding to the diastereotopic $4'$ - CH_2 group of the pyrrolidine-2,5-dione cycle. A doublet at δ 3.67 ppm ($J = 8$ Hz) and quadruplet-like multiplet at δ 4.15–4.19 ppm ($J = 6.8$ Hz, $J = 14$ Hz) are assigned to the pyrrolidine H-4 and H-5 protons, respectively. The multiplicity of these signals clearly demonstrates the regiochemistry of the 1,3-dipolar cycloaddition reaction. If the alternative other regioisomer **6b** (Scheme 3) would have been formed, the pyrrolidinyll H-3 and H-5 protons should give rise to a singlet and doublet of doublet pattern in the ^1H NMR spectrum.

The aromatic protons give rise to a multiplet in the region δ 6.77–7.31 ppm and a broad singlet at δ 10.62 ppm is attributed to the NH proton of the oxindole motif. In addition, a doublet at δ 3.80 ppm can be assigned to the NH proton of the pyrrolidine ring.

The proton decoupled ^{13}C NMR spectrum of **5b** exhibits signals at δ 41.1 and 72.8 ppm corresponding to the C-4 and C-5 carbons. The two spiranic carbons C-3 and C-2 resonate at δ 60.2 and 78.8 ppm, respectively. In addition, the oxindole carbonyl carbon is recognized at δ 178.8 and two further signals resonating at δ 184.0 and 184.4 ppm are attributed to the succinimide carbonyl carbons.

The regio- and stereochemical outcome of the reaction was finally ascertained by an X-ray study, allowing elucidation of the molecular structure of **5b**, and confirming that the isopropyl and aryl group stemming from the dipolarophile are in a transoid configuration.

The crystallographic data and the structural refinement details are reported in Table S1 (Supplementary Materials). The asymmetric unit contains two independent molecules. A thermal ellipsoid plot with the atom numbering is shown in Figure 3, and selected bond distances and angles are listed in Table S2.

The solid-state structure of compound **5b** is stabilized through significant intermolecular interactions such as $\text{N}-\text{H}\cdots\text{N}$, $\text{N}-\text{H}\cdots\text{O}$, $\text{C}-\text{H}\cdots\text{Cl}$ and $\text{C}-\text{H}\cdots\text{O}$ hydrogen bonding (Table S2). Within the supramolecular crystal structure of **5b** (Figure 4), the two independent molecules are linked together via weak non-conventional intermolecular H-bonding interactions. Thus, the N2 atom of the pyrrolidine ring of the first molecule is involved in a $\text{N}4-\text{H}4\cdots\text{N}2$ H-bond with the H4 atom of the pyrrolidine ring of the second molecule with a separation of 2.871(2) Å. The H5A atom of the pyrrolidine ring in the second molecule is also involved in another $\text{N}5-\text{H}5\text{A}\cdots\text{O}6$ H-bond with the O6 oxygen atom of pyrrolidine-2,5-dione of another nearby molecule with a length of 2.957(2) Å (Figure 4a). In addition, this crystal packing is reinforced by several weak $\text{C}-\text{H}\cdots\text{O}$ interactions between the H16A

atom of the pyrrolidine-2,5-dione group of one molecule and the O5 oxygen atom of the same group of another molecule, with a C16-H16A \cdots O5 distance of 2.943 Å.

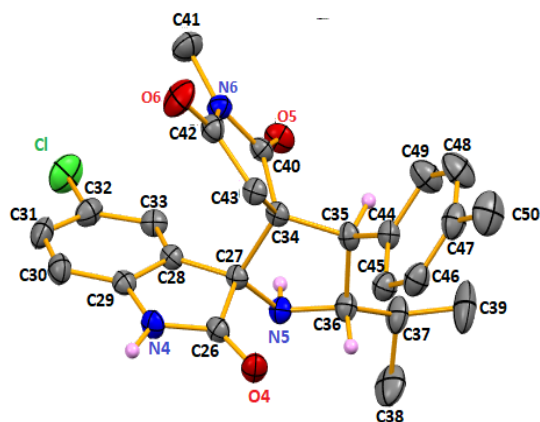


Figure 3. The molecular structure of one of the two independent molecules of **5b**, showing the atom-numbering scheme. The displacement ellipsoids drawn at the 50% probability level. Only relevant H atoms are shown for clarity.

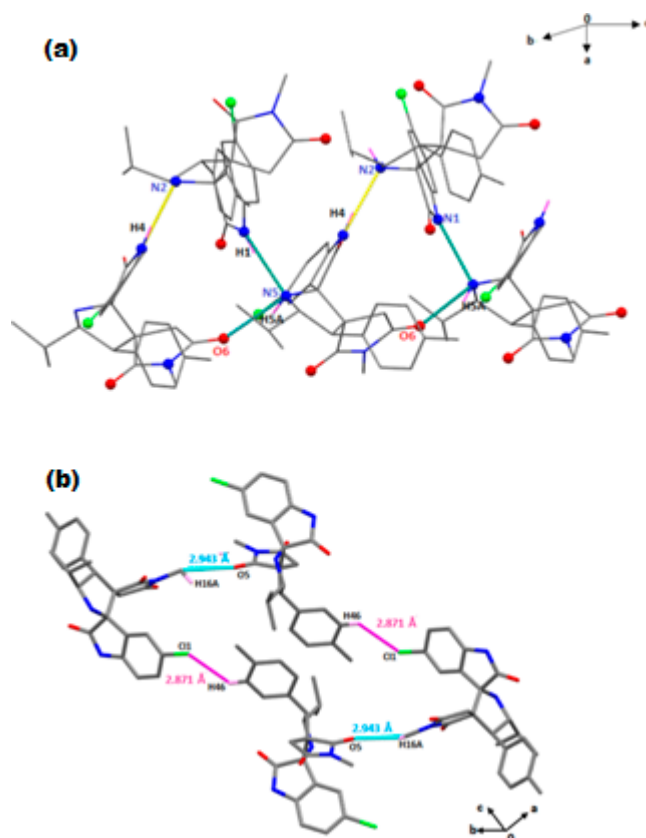
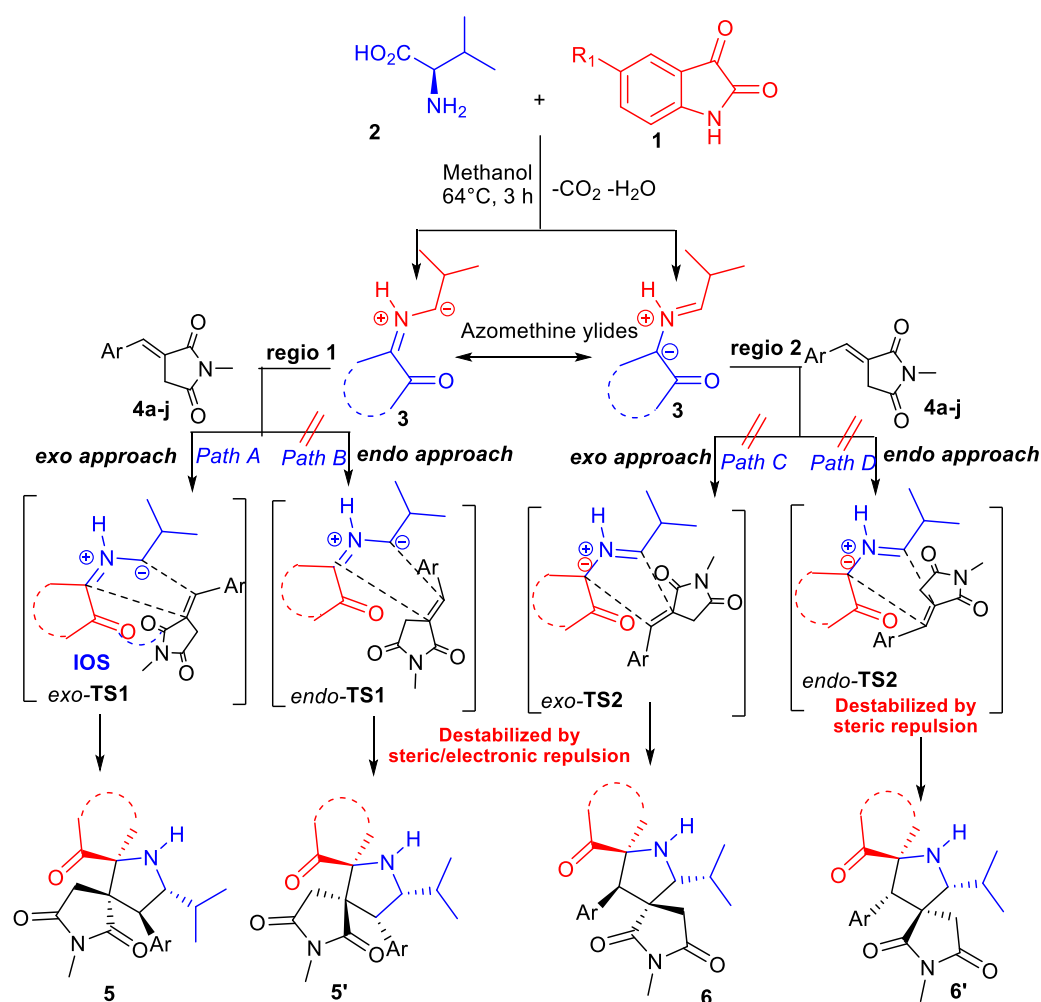


Figure 4. (a) Representation of the N-H \cdots N and N-H \cdots O bonding between two adjacent molecules of **5b**. Intermolecular interactions contacts are shown as dashed lines. (b) Presentation of the intermolecular C-H \cdots Cl and C-H \cdots O contacts occurring in **5b**.

A similar contact also occurs between the H46 atom of the phenyl ring of one molecule and the *p*-chloroaryl substituent of a neighbored molecule (C46-H46 \cdots Cl1, 2.871 Å (Figure 4b). Further details of the intermolecular interactions present in **5b** are listed in Table S2.

2.3. Mechanistic Discussion

To explain the stereochemical outcome in line with antecedent studies and our previous experimental findings [33–41], we propose in Scheme 4 a plausible mechanism for the regioselective and stereoselective formation of the spiro[oxindole-2,3'-pyridine].



Scheme 4. Suggested mechanism for the 1,3-dipolar cycloaddition of azomethine ylides across (*E*)-3-arylidene-1-methyl-pyrrolidine-2,5-diones **4**.

First, the in situ decarboxylative condensation of isatin derivatives **1** with *L*-valine **2** generates the (*Z,E*)-dipole **3**. Then the azomethine ylide subsequently undergoes a 1,3-dipolar cycloaddition reaction across the dipolarophile **4**. *A priori*, two regioisomeric and two stereoisomeric pathways can be taken in consideration. The *exo*- and *endo*-approaches refer to the approach of the dipolarophile with its electron-withdrawing group (carbonyl group) oriented toward the EWG group of the dipole (carboxamide group of oxindole) or away from it, respectively. The presence of a secondary orbital interaction (SOI) [42], via path *A*, favors more the formation of the *exo*-regioisomer **5**, which occurs between the oxygen atom of the carbonyl of the diketone and the carbon atom of the carbonyl succinimide acting as dipolarophile according to Scheme 4. The formation of the alternative regioisomers or diastereoisomers **5'** or **6/6'** via pathways *B*, *C* and *D* is probably less favorable because of steric or electronic repulsion occurring in their corresponding transition states. We are aware that, apart from SOI interactions, other parameters, such as reaction temperature [35,40] or the choice of the solvent [43], may also impact the stereochemical outcome. To confirm our mechanistic proposal, DFT computing is intended in prospective work.

2.4. Electronic Properties: UV-Visible Absorption Spectroscopy

The absorption spectral data recorded in DMSO as solvent for the spiro[pyrrolidin-2,3-oxindoles]**5** are summarized in Table 3. As representative example, the superposition of the electronic absorption spectra of **5c**, **5g** and **5h** using a dilute concentration around 10^{-5} M are depicted in Figure 5.

Table 3. UV-vis maxima and optical gap (eV) for series 5.

Compounds	λ_{\max} (nm)	λ_{abs} (nm) ($\log(\epsilon)$ $\text{M}^{-1}\text{cm}^{-1}$)	E _g (eV)
5a	300		
5b	300		
5c	307		
5d	320		
5e	310		
5f	320	420	2.60
5g	340	454	2.44
5h	340	410	2.53
5i	290		
5j	300		
5k	300		
5l	295		
5m	297		
5n	320	418	2.67
5o	320	420	2.60
5p	340	409	2.69
5q	298		

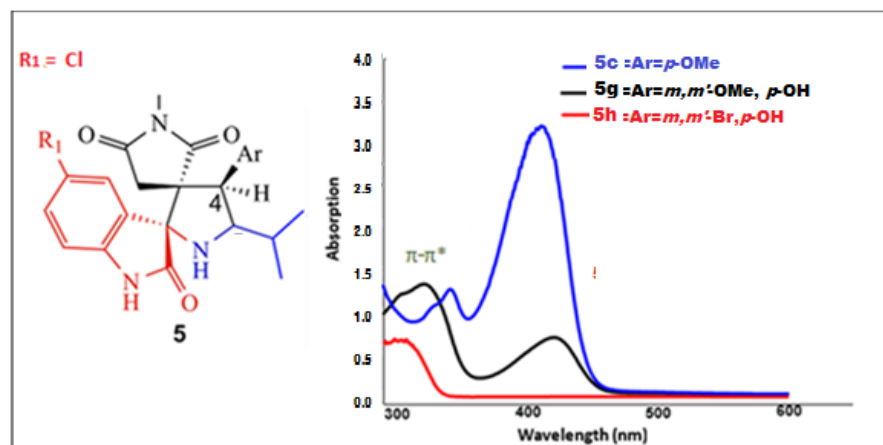


Figure 5. UV-visible absorption spectra in DMSO ($c = 10^{-5}$ M) at 298 K of compounds **5c**, **5g** and **5h**.

The broad absorption band with λ_{abs} in the range 290–340 nm can be assigned due to π - π^* transitions. Additionally, in the case of *p*-hydroxyspiro[pyrrolidin-2,3-oxindoles] **5f–h** and **5n–p**, the appearance of a second high intensity band within the 409–454 nm region is noticeable. The intense absorptions can be attributed to an intramolecular charge transfer (ICT) transition involving to hydroxy group [44] (Figure 5) and (Table 3).

This CT-band is red-shifted for compounds **5h** and **5o**, where the methoxy group is attached at 3,5-positions. The bathochromic effect on the charge transfer band is mainly due to the electron-withdrawing propensity of OCH_3 in an inductive sense [45]. In contrast, a hypsochromic shift is noticed for *p*-hydroxyspiro[pyrrolidin-2,3-oxindoles] **5h** and **5p** bearing bromo substituents.

The optical gap ($E_{\text{gap-op}}$) refers to the energetic separation between the HOMO and the LUMO frontier orbitales of the spiropyrrolidine oxindoles. To determine the optical

bandgap, we used the UV-visible spectrum, employing the Tauc plot method [46]. The E_{gap} values in DMSO solution are 2.44 eV and 2.69 eV for **5i** and **5p**, respectively (Table 3).

2.5. Antimicrobial Screening

The antibacterial activity of the pentacyclic compounds was screened against four bacteria strains, namely *Staphylococcus aureus* ATCC 25,923 (S.A.), *Micrococcus luteus* NCIMB 8166 (M.L), *Escherichia coli* ATCC 25,922 (E.C.) and *Pseudomonas aeruginosa* ATCC 27,853 (P.A.). Tetracycline (Tet. <https://en.wikipedia.org/wiki/Tetracycline>, accessed on 13 December 2021) was used as standard antibacterial agent. Furthermore, the antifungal activity of the products was tested against two pathogenic reference yeasts, namely *Candida albicans* ATCC 90,028 (C.A.) and *Candida krusei* ATCC 6258 (C.K.). Amphotericin B (Amp B https://en.wikipedia.org/wiki/Amphotericin_B, accessed on 13 December 2021.) has been used as a standard antifungal agent.

The determinations of the minimal inhibition concentrations (MIC) and the minimal bactericidal/fungicidal concentrations (MBC/MFC) were conducted following the technique of dilution in liquid medium, as described by Snoussi et al. [47].

When tested on different microorganism strains, the compounds were found to display significant activity. The potencies were qualitatively assessed by the presence or the absence of an inhibition zone diameter (Figures 6 and 7). The MIC, MBC and MFC values are presented in Tables 4–6, respectively.

Table 4. Antibacterial and antifungal activity (MIC in $\mu\text{g}/\text{mL}$).

Comp.	Bacterial Strains					
	Gram-Positive Bacteria		Gram-Negative Bacteria		Yeast Strains	
	<i>S. aureus</i> ATCC 25923	<i>M. luteus</i> NCIMB 8166	<i>E. coli</i> ATCC 25922	<i>P. aeruginosa</i> ATCC 27853	<i>C. albicans</i> ATCC 90028	<i>C. krusei</i> ATCC 6258
5a	3.9	NA	NA	250	7.8	31.5
5b	500	250	250	NA	250	125
5c	NA	250	NA	NA	62.5	NA
5d	31.5	250	NA	NA	125	125
5e	31.5	250	500	NA	NA	NA
5f	NA	125	NA	NA	7.8	250
5g	31.5	125	NA	250	125	125
5h	250	31.5	NA	NA	250	250
5i	3.9	125	250	250	7.8	62.5
5j	500	250	250	NA	250	1000
5k	1000	500	NA	NA	125	250
5l	62.5	500	NA	NA	NA	NA
5m	125	1000	NA	NA	250	250
5n	NA	125	NA	NA	NA	NA
5o	250	31.5	NA	NA	500	250
5p	250	62.5	NA	NA	31.5	125
5q	500	500	NA	NA	125	250
Tet.	24 \pm 0	12 \pm 0.1	9 \pm 0	11 \pm 1.1	NT	NT
Amp. B	NT	NT	NT	NT	15 \pm 0.58	14 \pm 0.1

Tet.: Tetracycline; Amp. B: Amphotericin B; MIC: Minimum Inhibition Concentration; NA: no activity.

Table 5. Antibacterial and antifungal activity of series 5 (Inhibition zone in mm).

Comp.	Bacterial Strains				Yeast Strains	
	<i>S. aureus</i> ATCC 25923	<i>M. luteus</i> NCIMB 8166	<i>E. coli</i> ATCC 25922	<i>P. aeruginosa</i> ATCC 27853	<i>C. albicans</i> ATCC 90028	<i>C. krusei</i> ATCC 6258
5a	31 ± 0.03	NA	NA	9 ± 0	12 ± 0	11 ± 0
5b	17 ± 0	10 ± 0	8 ± 0	NA	14 ± 0	11 ± 0
5c	NA	10 ± 0	NA	NA	7 ± 0.2	NA
5d	12 ± 0	17 ± 0.01	NA	NA	11 ± 0	10 ± 0
5e	11 ± 0	9 ± 0	15 ± 0	NA	NA	NA
5f	NA	11 ± 0	NA	NA	12 ± 0	8 ± 0
5g	18 ± 0	12 ± 0	NA	8 ± 0	12 ± 0	14 ± 0
5h	20 ± 0	10 ± 0	NA	NA	7 ± 0	8 ± 0
5i	19 ± 0	11 ± 0	8 ± 0	8 ± 0.02	12 ± 0.1	12 ± 0
5j	12 ± 0	12 ± 0	9 ± 0	NA	12 ± 0	11 ± 0
5k	11 ± 0	10 ± 0	NA	NA	9 ± 0	9 ± 0
5l	12 ± 0	13 ± 0	NA	NA	NA	NA
5m	9 ± 0.1	9 ± 0	NA	NA	11 ± 0	11 ± 0
5n	NA	13 ± 0	NA	NA	6 ± 0	NA
5o	19 ± 0	14 ± 0	NA	NA	12 ± 0.1	12 ± 0
5p	20 ± 0	15 ± 0	NA	NA	9 ± 0	9 ± 0
5q	9 ± 0	9 ± 0	NA	NA	10 ± 0	10 ± 0
Tet.	24 ± 0	12 ± 0.1	9 ± 0	11 ± 1.1	NT	NT
Amp. B	NT	NT	NT	NT	15 ± 0.58	14 ± 0.1

Tet.: Tetracycline; Amp. B: Amphotericin B; NA: no activity; NT: not probed.

Table 6. Antibacterial and antifungal activity (MBC and MFC in µg/mL).

Comp.	Bacterial Strains				Yeast Strains	
	Gram-Positive Bacteria		Gram-Negative Bacteria		<i>C. albicans</i> ATCC 90028	<i>C. krusei</i> ATCC 6258
	<i>S. aureus</i> ATCC 25923	<i>M. luteus</i> NCIMB 8166	<i>E. coli</i> ATCC 25922	<i>P. aeruginosa</i> ATCC 27853		
5a	31.5	1000	500	250	7.8	31.5
5b	1000	>1000	>1000	1000	125	500
5c	NA	>1000	NA	NA	62.5	NA
5d	62.5	>1000	NA	NA	125	125
5e	500	>1000	1000	NA	NA	NA
5f	NA	>1000	NA	NA	250	250
5g	500	>1000	NA	500	125	125
5h	1000	>1000	NA	NA	250	250
5i	>1000	NA	NA	>1000	31.5	62.5
5j	1000	>1000	>1000	NA	250	1000
5k	>1000	>1000	NA	NA	125	250
5l	1000	>1000	NA	NA	NA	NA
5m	1000	>1000	>1000	NA	1000	500
5n	NA	>1000	NA	NA	NA	NA
5o	1000	>1000	NA	NA	500	500
5p	500	>1000	NA	NA	250	250
5q	1000	>1000	NA	NA	125	250
Tet.	24 ± 0	12 ± 0.1	9 ± 0	11 ± 1.1	NT	NT
Amp. B	NT	NT	NT	NT	15 ± 0.58	14 ± 0.1

Tet.: Tetracycline; Amp. B: Amphotericin B; MBC: Minimum Bactericidal activity; MFC: minimum fungicidal activity; NA: no activity.

Yeast strains: test 6c-*Candida krusei* ATCC 6258test 9c-*Candida albicans* ATCC 90028

Figure 6. Antibacterial activity of some derivatives via determination of the inhibition zone (mm) around discs impregnated with **5a** (M1V), **5b** (M2V), **5c** (M3V), **5d** (M4V), **5e** (M7V), **5f** (H1V), **5g** (H2V), **5h** (H3V), **5i** (M1VF), **5j** (M2VF), **5k** (M3VF), **5l** (M4VF), **5m** (M7VF), **5n** (H1VF), **5o** (H2VF), **5p** (H3VF), and **5q** (M3VI) concentration (10 mg/disk). Tetracycline (10 mg/mL) or Amphotericin B (10 mg/mL) expressed as mean \pm SD (n = 3).

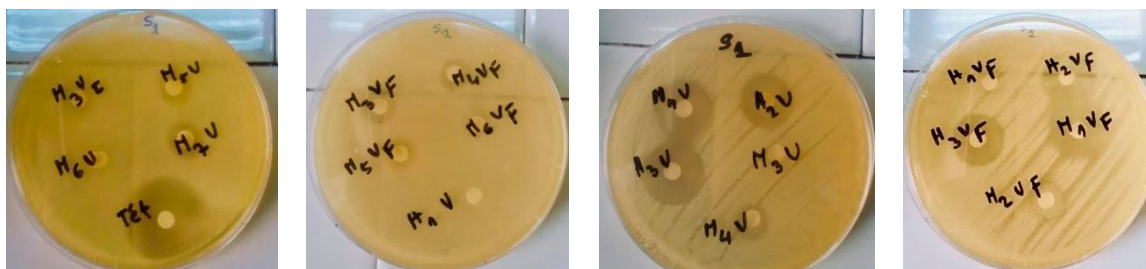
Gram-positive bacteria: test s1-*Staphylococcus aureus* ATCC 25923

Figure 7. Antimicrobial activity of some samples determined via the inhibition zone (mm) around discs impregnated with **5a** (M1V), **5b** (M2V), **5c** (M3V), **5d** (M4V), **5e** (M7V), **5f** (H1V), **5g** (H2V), **5h** (H3V), **5i** (M1VF), **5j** (M2VF), **5k** (M3VF), **5l** (M4VF), **5m** (M7VF), **5n** (H1VF), **5o** (H2VF), **5p** (H3VF), and **5q** (M3VI) compounds (10 mg/disk); Tetracycline (10 mg/mL) or Amphotericin B (10 mg/mL) expressed as mean \pm SD (n = 3).

The highest antibacterial activity of the cycloadducts is displayed against gram-positive bacteria (*Staphylococcus aureus* ATCC 25,923 and *Micrococcus luteus* NCIMB 8166), whereas towards gram-negative bacteria (*Escherichia coli* ATCC 25,922 and *Pseudomonas aeruginosa* ATCC 27853), the probed compounds are in general less active compared to gram-positive ones.

In the first screening phase against *Staphylococcus aureus* ATCC 25923, **5a** was noted to be the most sensitive derivative, with an inhibition zone of 31 mm, the MIC being 3.9 μ g/mL and the MBC being 31.5 μ g/mL, respectively (Tables 4–6). Then, against the same pathogenic bacteria, four compounds **5d**, **5e**, **5h** and **5l** showed an excellent activity with MIC values in the range of 31.5–62.5 μ g/mL as MBC with an inhibition zone between 11 and 20 mm, compared to standard antibiotic Tetracycline.

When testing against *Micrococcus luteus* NCIMB 8166, compounds **5f**, **5g**, **5n** and **5o** were equipotent as Tetracycline. Thus, **5h** and **5p** were found to be the most active, with

MIC values of 31.5 and 62.5 $\mu\text{g}/\text{mL}$, respectively (Table 4). Concerning the evaluation of the inhibition zone, the compounds **5d**, **5o** and **5p** were found to be the most sensitive, with 17mm to 14 mm. Additionally, the different compounds were evaluated against two gram-negative bacteria (*Escherichia coli* ATCC 25,922 and *Pseudomonas aeruginosa* ATCC 27853). In this case, **5e** was found to be the most sensitive, with an inhibition zone of 15 mm. All compounds were found to be equal or less active (125– >1000 $\mu\text{g}/\text{mL}$) concerning the MIC or MBC compared to standard antibiotic Tetracycline.

On the other hand, the in vitro antifungal activity data reveal that **5a**, **5f** and **5i** display the highest activity with MIC 7.812 $\mu\text{g}/\text{mL}$ against Amphotericin B.

Compounds **5d**, **5k** and **5q** showed a somewhat less efficient activity, when compared to compounds **5i** and **5p** with MIC 31.5 $\mu\text{g}/\text{mL}$ relating to *Candida albicans* ATCC 90028. Additionally, regarding the results of antifungal screening, the MFC is noted to have good antifungal activity on **5a**, **5i** and **5c** with 7.81, 31.5 and 62.5 $\mu\text{g}/\text{mL}$, respectively. Furthermore, **5b** appeared to be the most sensitive heterocycle (14 mm). In contrast, as indicated in Table 5, **5n** was revealed to be the least sensitive heterocycle, with an inhibition zone of 6 mm.

Finally, we discuss the last yeast strains is *Candida krusei* ATCC 6258. In fact, **5g**, **5i** and **5p** showed a superior sensitivity with an inhibition zone of 14–12 mm. It is also worth mentioning the excellent antifungal activity of compounds **5a**, displaying both a MIC and MFC of 31.5 $\mu\text{g}/\text{mL}$ (Table 6).

2.6. Molecular Docking Studies

In order to provide a comprehensive rationale for the obtained biological activities, we docked the most active compounds, **5a** and **5i**, into glucosamine 6-phosphate synthase (GlcN6P) and methionyl-trna-synthetase (1PFV). GlcN6P catalyzes a multifarious reaction comprising both sugar isomerization (fructosamine-6-phosphate to glucosamine-6-phosphate) and ammonia transfer (*L*-glutamine to Fru-6-P) [48–50]. This reaction initiates a pathway that finally drives the production of uridine 5'-diphospho-*N*-acetyl-*D*-glucosamine. This nucleotide sugar is used to assemble the macromolecules essential for constructing cell walls, such as the macromolecules with several amino sugar units, like chitin or proteins in fungi, and lipopolysaccharides and peptide glycan in bacteria. GlcN6P synthase inhibition is fatal, even for a short time, in prokaryotic cells but not in human cells. Therefore, it is considered as a promising target for developing antibacterial and antifungal agents.

The compounds, except **5a** enantiomer (which has the most potent antibacterial activity among all the synthesized compounds), exhibit interactions with several amino acid residues at the active site of GlcN6P and reveal low binding energy. They also share several amino acids with the positive controls, Ampicillin and Tetracycline (Table 7) and (Figure 8).

Infections caused by antibiotic-resistant pathogens have raised markedly in recent years. Over 60% of staphylococcal infections in intensive care units in the United States, for example, are caused by methicillin-resistant bacterial strains. The incidence of the emerging methicillin-resistant staphylococcus aureus strains in community has raised many concerns.

These strains were found to express a wide range of virulence factors (https://en.wikipedia.org/wiki/Virulence_factor, accessed on 15 January 2021) and have an enhanced tendency toward progression into invasive ailments. The action takes place over two-step reactions. In the first step, methionine and ATP bind together to the enzyme's active site, resulting in the formation of methionyl adenylate, releasing pyrophosphate (PPi). In the following reaction, the activated methionyl adenylate is moved to tRNAMet and its 3' end, releasing an AMP molecule. MetRS has the unique ability to identify and load two tRNA substrates; namely, tRNAMet and tRNAfMet [51]. For these reasons, the design of alternative antibacterial agents exhibiting an activity against drug-resistant *staphylococci* represents an important area for the development of novel drugs.

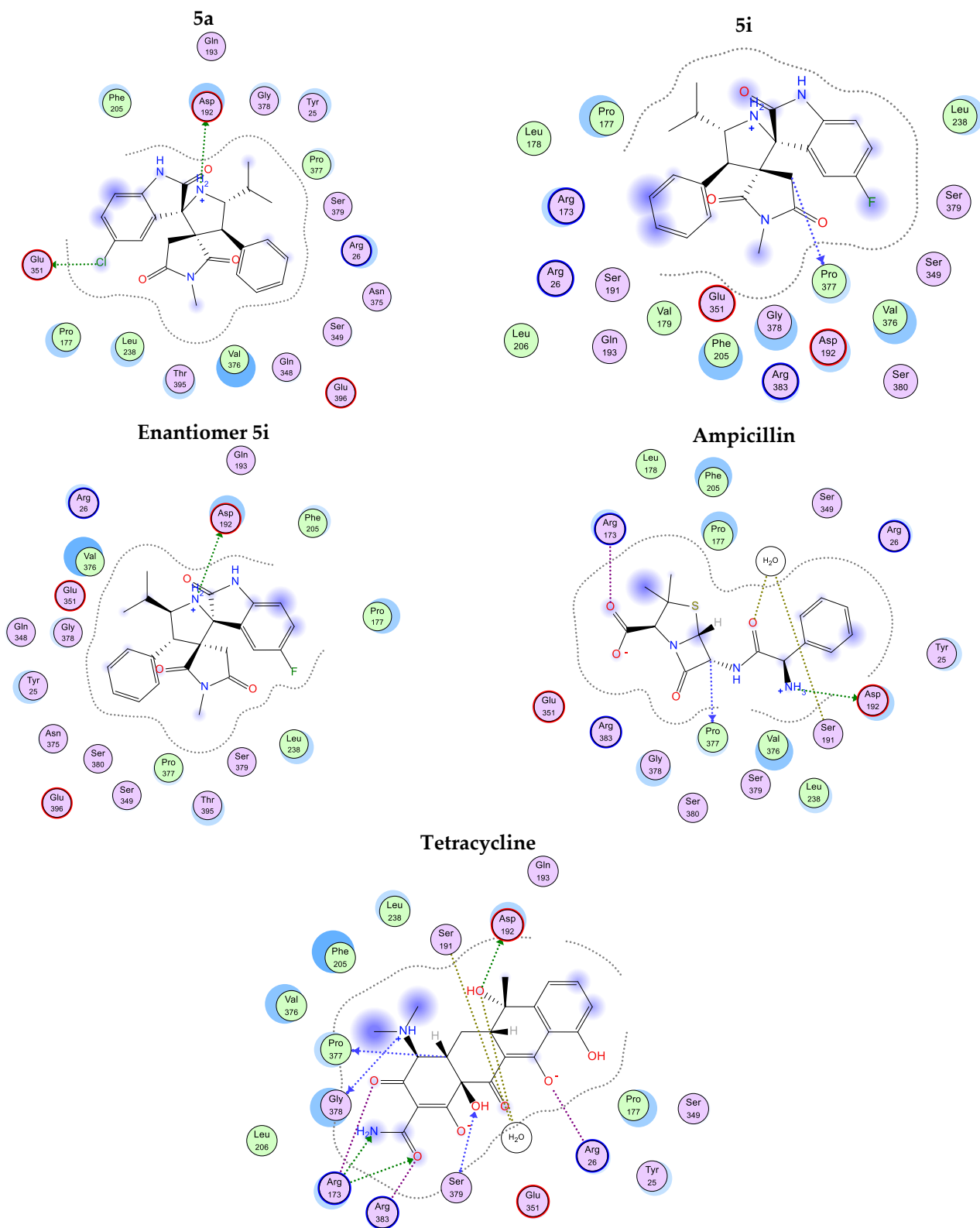


Figure 8. 2D-interactions of compounds **5a**, **5i**, **5i enantiomer**, Ampicillin and Tetracycline with amino acid residues on glucosamine 6-phosphate synthase (1jxa).

Table 7. Scoring function of docking poses and the interactions of the docked compounds **5a**, **5i**, **5i enantiomer**, Ampicillin and Tetracycline with amino acid residues on glucosamine 6-phosphate synthase (1jxa) and methionyl-trna-synthetase (1PFV).

Comp.	1JXA		1PFV	
	Ligand Interactions	Score Function	Ligand Interactions	Score Function
5a	ASP 192 (H-bonding) GLU 351 (H-bonding)	−15.33	TYR 15 (Hydrophobic bonding and H-bonding through solvent) HIS 24 (H-bonding through solvent)	−12.09
5a enantiomer		Failed		Failed
5i	PRO 377 (H-bonding)	−13.40	HIS 24 (Hydrophobic bonding) HIS 21 (Hydrophobic bonding)	−7.45
5i enantiomer	ASP 192 (H-bonding)	−15.11	TYR 15 (H-bonding through solvent) HIS 24 (H-bonding through solvent)	−12.51
Ampicillin	ASP 192 (H-bonding) SER 191 (H-bonding through solvent) ARG 173 (H-bonding)	−14.83	TYR 15 (H-bonding through solvent) HIS 24 (H-bonding through solvent) HIS 21 (H-bonding) LYS 335 (H-bonding and salt bridge)	−9.99
Tetracycline	PRO 377 (H-bonding) ASP 192 (H-bonding) SER 191 (H-bonding through solvent) ARG 173 (H-bonding and salt bridge) SER 379 (H-bonding) GLY 378 (H-bonding) ARG 383 (Salt bridge) ARG 26 (Salt bridge)	−19.71	LYS 335 (Salt bridge) HIS 21 (H-bonding)	−15.11

Again, our *in silico* study results revealed that the screened compounds, except the **5a enantiomer**, exhibited low binding energy when docked into methionyl-trna-synthetase (1PFV) and shared similar interactions with several amino acids similar to the positive control, Ampicillin and Tetracycline (Table 7) and Figure 9.

Interestingly, all compounds obeyed to the parameters of the rule of 5 (Lipinski rule) [52]. These include molecular weight, number of hydrogen bonds, accepting (lip_acc) and donating (lip_don) groups, and the drug-likeness potential (Table 8).

Table 8. Results of the Lipinski parameters of compounds **5a**, **5i**, **5i enantiomer**, Ampicillin and Tetracycline.

Compound	lip_acc	lip_don	lip_druglike	Weight	lip_violation
	≤10	≤5	=1	≤500	
5a	6	3	1	438.94	0
5a enantiomer	6	3	1	438.94	0
5i	6	3	1	422.48	0
5i enantiomer	6	3	1	422.48	0
Ampicillin	7	4	1	349.41	0
Tetracycline	10	6	1	443.43	1

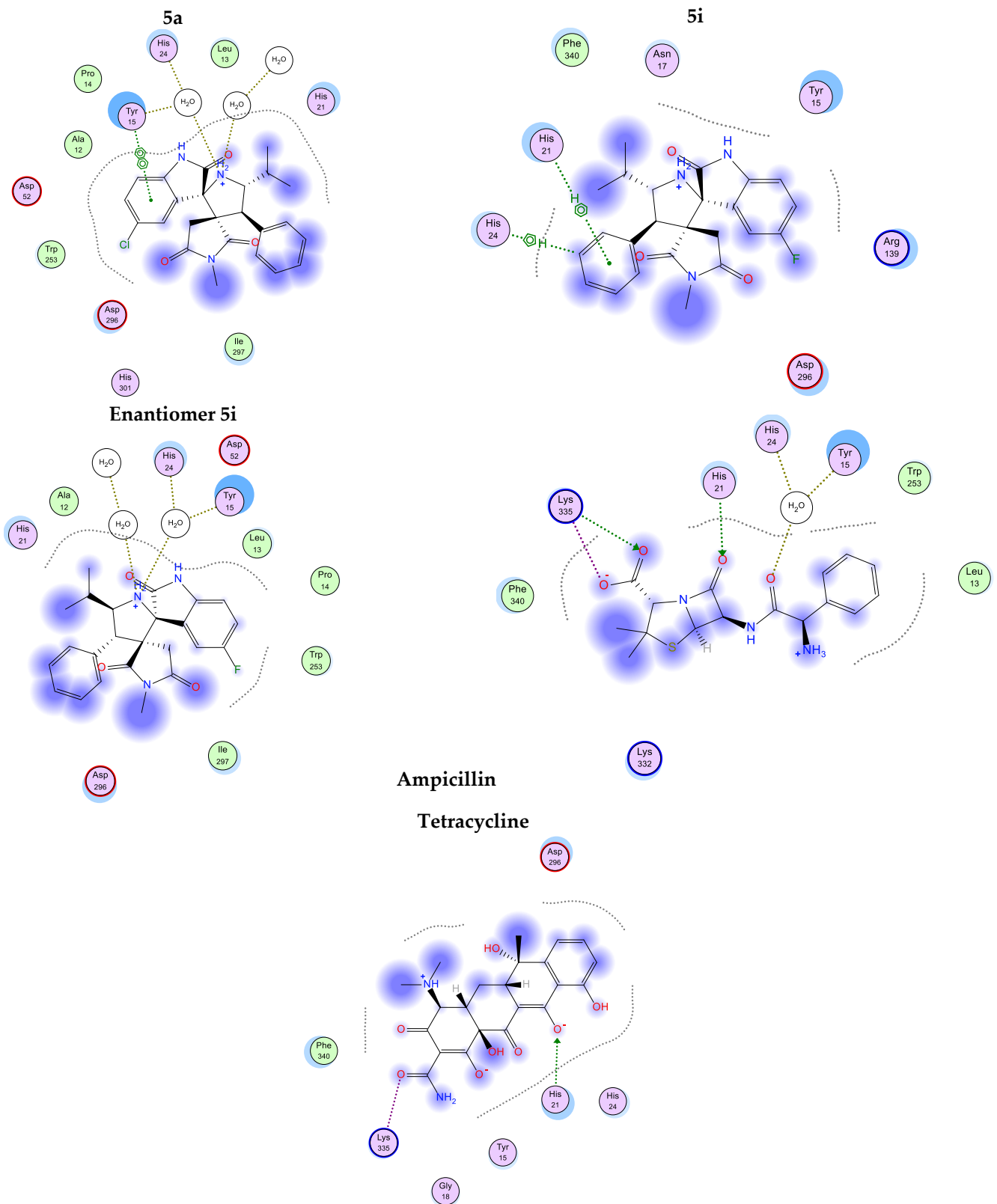


Figure 9. 2D-interactions of compounds **5a**, **5i**, **5i enantiomer**, Ampicillin and Tetracycline with amino acid residues on methionyl-trnasynthetase (1PFV).

3. Experimental

3.1. General Experimental Methods

The ^1H NMR spectra were recorded at 400 MHz using a Bruker Avance III-400 machine (Rheinstetten, Germany). The chemical shifts were recorded in ppm relative to TMS and with the solvent resonance as the internal standard. Data were reported as follows: chemical shift, multiplicity (bs = broad singlet, s = singlet, d = doublet, dd = doublet of doublet, m = multiplet), coupling constants (Hz), and integration. $^{13}\text{C}\{^1\text{H}\}$ NMR data were collected at 100 MHz with complete proton decoupling with the solvent resonance as the internal standard. Optical absorption spectra were recorded at an ambient temperature, using a Shimadzu UV-3600 double-beam UV-visible spectrophotometer and the concentrations of spiro[pyrrolidin-2,3'-oxindole] derivatives were controlled to be of 10^{-5} M in DMSO. The materials used were: thin-layer chromatography (TLC): TLC plates (Merck, silica gel 60 F254 0.2 mm 200×200 nm), and substances were detected using UV light at 254 nm. High Resolution Mass Spectra (ES-HRMS) were obtained with Micromass LCT (ESI technique, positive mode) spectrometers.

3.2. X-ray Crystallography

X-ray suitable crystal of **5b** for the diffraction study were obtained by slow evaporation of a diethyl ether solution containing **5b**. The crystallographic data were collected at 193(2)K on a Bruker-AXS D8-Venture diffractometer equipped with a Photon III-C14 detector using $\text{MoK}\alpha$ radiation ($\lambda = 0.71073\text{\AA}$). Phi- and omega-scans were used. Space group was determined on the basis of systematic absences and intensity statistics. Semi-empirical absorption correction was employed [53]. The structure was solved using an intrinsic phasing method (SHELXT) [54] and refined using the least-squares method on F [55]. All non-H atoms were refined with anisotropic displacement parameters. Hydrogen atoms were refined isotropically at calculated positions using a riding model with their isotropic displacement parameters constrained to be equal to 1.5 times the equivalent isotropic displacement parameters of their pivot atoms for terminal sp^3 carbon and 1.2 times for all other carbon atoms. H on nitrogen atoms were located by difference Fourier maps.

CCDC-2063918 contains the supplementary crystallographic data for this paper. These data can be obtained free of charge from The Cambridge Crystallographic Data Centre via <https://www.ccdc.cam.ac.uk/structures> (accessed on 19 Mars 2021).

Crystal data for **5b**: $\text{C}_{25}\text{H}_{26}\text{ClN}_3\text{O}_3$ ($M = 451.94$ g/mol): Monoclinic, space group $P2_1/c$, $a = 12.8755(8)$ \AA , $b = 25.4246(16)$ \AA , $c = 14.9546(9)$ \AA , $\beta = 100.370(2)^\circ$, $V = 4815.5(5)\text{\AA}^3$, $Z = 8$, crystal size $0.40 \times 0.08 \times 0.06$ mm^3 , $\mu(\text{MoK}\alpha) = 0.189$ mm^{-1} , $D_{\text{calc}} = 1.247$ g/cm^3 , 163,328 reflections collected, 13,498 unique ($R_{\text{int}} = 0.1293$), 601 parameters, $R1 [I > 2s(I)] = 0.0584$, $wR2 [\text{all data}] = 0.1428$, and the largest differential peak and hole: 0.516 and -0.343 $\text{e}\text{\AA}^{-3}$.

3.3. Antimicrobial Screening

First, the optical density of each microorganism suspensions was adjusted to 0.1 at OD600 for bacteria and 0.4 at OD540 for yeasts. Then, 500 μL of inoculums were dropped onto adequate agar plates. Sterile filter discs (diameter 6 mm, Biolife, Italy) were placed at the surface of the appropriate agar mediums and 10 mg/disc of the product dissolved in 10% of dimethyl sulfoxide was dropped onto each disc. Tetracycline (10 mg/mL; 10 μL /disc) and Amphotericin B (10 mg/mL; 10 μL /disc), were used as reference drugs. After incubation for 18–24 h at 37 $^\circ\text{C}$, the antibacterial activities were evaluated by measuring an inhibition zone formed around the disc. Each assay was performed in triplicate.

For this, the microbial inoculums were prepared from 12 h broth cultures and spectrophotometrically adjusted to 10^7 UFC/mL.

Serial two-fold dilutions of the different amounts of the compounds (1000 to 0.09 $\mu\text{g/mL}$) were prepared in adequate broth. Then, 10 μL of the inocula of each reference strain were added to the plates containing the serial dilution and were incubated aerobically at 37 $^\circ\text{C}$ for 24 h. MIC was defined as the lowest concentration that completely inhibited visible cell growth during 24 h incubation period at 37 $^\circ\text{C}$.

MBC and MFC values were determined by inoculating 10 μ L of each well medium with no visible growth on Müller Hinton or Sabouraud chromphenicol agar plates (https://en.wikipedia.org/wiki/Sabouraud_agar, accessed on 17 January 2021) and were defined as the lowest concentration at which 99% of the tested strains were killed after 24 h of incubation at 37 °C.

3.4. General Procedure for the Preparation of Cycloadducts 5

The (*E*)-3-arylidene-1-methylpyrrolidine-2,5-diones **4a-h** have been synthesized by Wittig reaction between *N*-methylmaleimide and the corresponding aromatic aldehyde [56].

A mixture of isatin derivatives **1** (1 mmol), *L*-valine **2** (1.2 mmol) and dipolaphile **4** (1 mmol) was refluxed in methanol (10 mL) for 3h. After completion of the reaction as monitored from TLC, the solvent was removed under reduced pressure and the residue was chromatographed on silica gel employing ethylacetate-heptane (3:7 *v/v*) as eluent to obtain the pure products **5**. Spectroscopic data for all compounds are presented in the ESI.

4. Conclusions

To resume, we reported the synthesis of a series spiro[oxindole-2,3'-pyrrolidine] tethered with succinimide scaffolds via a three component 1,3-dipolar cycloaddition reaction of (*E*)-3-arylidene-1-methyl-pyrrolidine-2,5-diones, *L*-valine and cyclic ketones of the isatin type. The antimicrobial activity of these heterocyclic compounds was investigated against six microbial strains. The screening results showed that most of the tested compounds showed moderate to excellent antibacterial and antifungal activities. Molecular docking studies indicates that two compounds, **5a** and **5i**, had the most potent antibacterial and antifungal activities among all the synthesized compounds, revealing interactions with several amino acid residues at active site of GlcN6P and revealing low binding energy. The predicted ADME profiles of the compounds **5a** and **5i** are in line with the Lipinski rules.

Supplementary Materials: The following supporting information can be downloaded at: <https://www.mdpi.com/article/10.3390/app12010360/s1>, compound characterization data and Figures S1–S34: ¹H and ¹³C-NMR spectra, additional crystallographic data.

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