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Cellular and molecular complementary immune stress markers for the model species *Dreissena polymorpha*

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1 Full title: **Cellular and molecular complementary immune stress markers for the model**
2 **species *Dreissena polymorpha*.**

3

4 Short title: **New insight on hemocyte markers in zebra mussel.**

5

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27 **Abstract**

28 This study aimed to combine cellular and molecular analyses for better detail the effects of
29 various stresses on a sentinel species of freshwater invertebrate. For this purpose, the
30 hemocytes of the zebra mussel, *Dreissena polymorpha*, were exposed to different stresses
31 at two different intensities, high or low: chemical (cadmium and ionomycin), physical
32 (ultraviolet B), or biological ones (*Cryptosporidium parvum* and *Toxoplasma gondii*). After
33 exposure, flow cytometry and droplet digital PCR analyses were performed on the same
34 pools of hemocytes. Several responses related to necrosis, apoptosis, phagocytosis,
35 production of nitric oxide and expression level of several genes related to the antioxidant,
36 detoxification and immune systems were evaluated. Results showed that hemocyte integrity
37 was compromised by both chemical and physical stress, and cellular markers of
38 phagocytosis reacted to ionomycin and protozoa. While cadmium induced oxidative stress
39 and necrosis, ionomycin tends to modulate the immune response of hemocytes. Although
40 both biological stresses led to a similar immune response, *C. parvum* oocysts induced more
41 effects than *T. gondii*, notably through the expression of effector caspases gene and an
42 increase in hemocyte necrosis. This suggests different management of the two protozoa by
43 the cell. This work provides new knowledge of biomarkers in the zebra mussel, at both
44 cellular and molecular levels, and contributes to elucidate the mechanisms of action of
45 different kinds of stress in this species.

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47 **Keywords:** Bivalve; ddPCR; flow cytometry; hemocyte; protozoa.

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55 **1. Introduction**

56 Over the last years, ecotoxicology has gained increasing interest to assess the impact of
57 anthropogenic contaminants on organisms and ecosystems. Among the tools developed for
58 environmental risk assessment, biomarkers are of great interest, especially for their potential
59 as early warning indicators of contamination [1]. These tools are therefore increasingly
60 developed on sentinel organisms to better characterize the potential effects of contaminants.
61 In conjunction with chemical and biocenotic analyses, biomarkers contribute to being aware
62 of the global impact of this contamination on ecosystems.

63 However, this risk prediction requires a good knowledge of the biomarkers used as well as
64 an explanation of the mechanical bases of xenobiotic effects on well-known sentinel species
65 [2]. It is therefore essential to understand the modes of action of the various xenobiotics at
66 infra-individual levels (cellular, molecular) before being able to extrapolate to the scale of the
67 population or the community.

68 Among biomarkers, those linked to the immune system can reflect the state of health of an
69 organism under stress. Indeed, the immune system is responsible for the integrity of
70 organism, protects it against infections and parasitism and is linked to various other
71 biological functions, such as nervous or endocrine systems [3-6]. These immune markers are
72 often i) functional and assessed at the cellular level, such as the capacity or the efficiency of
73 phagocytosis, ii) structural in terms of the integrity of the immune effector cells, or iii) linked to
74 the expression of genes coding for immune proteins. They are therefore mainly assessed on
75 two levels of biological organization: the cell and the molecule. The analysis of multi-level
76 markers has several advantages. First, it allows a more complete diagnosis of contamination
77 effects on an organism and can allow their extrapolation on a population [7, 8]. Then, this
78 multi-level analysis can provide information on the impact kinetics of a contamination within
79 an organism [9]. Finally, it can be used to determine whether the responses are due to the
80 exposure or only due to a homeostatic modification [10].

81 In this study, cellular and molecular analyses were combined to understand the effects of
82 various stress on a sentinel species of freshwater invertebrate, the zebra mussel *Dreissena*
83 *polymorpha*. This invasive species is known to have colonized several regions in the
84 northern hemisphere (America, Europe) and to have negatively impacted the invaded
85 ecosystems [11]. Our objective was to understand the immune response capabilities of
86 hemocytes of the zebra mussel to various stresses (chemical, physical and biological), *via*
87 cellular and molecular markers. For this purpose, several parameters were set upstream of
88 the study. First, bivalves were chosen because they have several advantages to be used in
89 biomonitoring programs as sentinel species [12-15]: i) these animals are directly in contact
90 with the contamination (water and sediment) and can accumulate pollutants [16]; ii) they are
91 sessile, making possible a spatial and temporal assessment of surrounding contamination
92 [13, 14, 17]. For freshwater biomonitoring, *Dreissena polymorpha* is a good sentinel species
93 since it occupies a large geographical area, it resists to variations in external conditions and
94 is considered to be the counterpart of *Mytilus edulis* for ecotoxicological studies [18-20].
95 Second, cellular and molecular markers were evaluated on the same measurement scale,
96 namely on hemocytes. In bivalves, hemocytes are the effector cells of immunity. Exposures
97 to various stresses were conducted in *ex vivo* conditions to overcome the complexity of the
98 whole organism and to minimize the exposure bias. Third, all analyses (cellular and
99 molecular ones) were performed on the same samples to provide a better comprehension of
100 stressors' modes of action. This criterion required to work with pools of hemolymph to obtain
101 enough biological material for analysis. Finally, hemocytes from *D. polymorpha* were
102 exposed to various stresses (chemical, physical and biological), whose expected effects are
103 known. Hemocytes were exposed to i) cadmium, a cytotoxic metal [21]; ii) ionomycin, an
104 antibiotic which induces apoptosis and alters immune functions [22, 23]; iii) ultra-violet B
105 (UVb) which are known to be cytotoxic and immune suppressor [24, 25]; and iv) two
106 protozoa, *Cryptosporidium parvum* and *Toxoplasma gondii*, whose effects on the hemocytes
107 of mussels have already been evaluated [26].

108 Regarding mussel immunity, two processes have been studied *via* cellular and molecular
109 markers, namely:

110 - Phagocytosis, through the internalization of microbeads (capacity, efficiency and
111 avidity of phagocytosis) and the indirect stimulating action of nitric oxide (NO, [27]) for
112 cellular markers, as well as by the expression of several genes involved in this response: the
113 complex *arp 2/3*, involved in the regulation of cytoskeleton actin; *cdc 42*, involved in cell
114 migration and endocytosis; and *gal 2*, a galactose-binding lectin, playing many roles
115 including cell adhesion and signalling [28-30].

116 - Apoptosis, *via* the study of the cellular activation of an effector with cysteine effector,
117 caspases 3/7, and the expression of its gene. Galectin-2 also plays a role in apoptosis by
118 being part of anti-apoptotic molecules [31].

119 The molecular markers were selected in accordance with the work of Leprêtre *et al.* [32] who
120 observed the involvement of the proteins encoded by these genes in the cell compartment of
121 the hemolymph of the zebra mussel.

122 Cell necrosis marker, detoxification and antioxidant systems were also evaluated, by flow
123 cytometry and droplet digital polymerase chain reaction (ddPCR). This latter technology is
124 based on the massive fragmentation of the genetic material, which facilitates the analysis of
125 low-expressed genes [33, 34]. ddPCR was preferred to other PCR techniques because of
126 the low expression levels of certain genes such as *cdc 42* or *casp 3/7*, as well as being able
127 to express the absolute result in a number of copies per hemocyte. Thanks to the partitioning
128 of the sample and Poisson's law, ddPCR makes possible to express the quantification results
129 directly as the number of copies of the target DNA per μL , or in our case as the number of
130 copies per cell. This technique improves the measurement accuracy without the need for a
131 prior calibration curve [34, 35]. This simplifies quantification and reduces the risk of error.
132 Compared to other PCR techniques, it offers other advantages such as high accuracy and an
133 increase in the signal-to-noise ratio [34, 36]. This strategy, developed and used in medicine,
134 is pioneer in ecotoxicology. This opportunity to assess cellular and molecular markers on the

135 same scale, namely the hemocyte, allows a better understanding of the response capacities
136 of these cells to stress.

137

138 **2. Materials and methods**

139 *2.1. Mussel and hemolymph collection*

140 Zebra mussels (3.18 ± 0.41 g; 2.72 ± 0.16 cm) were collected on 25th August 2017 from Der
141 lake (51290 Giffaumont-Champaubert, France, N 48°33'35"; E 4°45'11") and brought back to
142 the laboratory, where they were maintained in 20 L aerated glass tanks in Cristaline Aurele
143 drinking water (14.41 ± 0.67 °C; pH 7.47 ± 0.07 ; 570 ± 40 mS/cm; spring Jandun, France).
144 Four weeks of acclimation were respected before *ex vivo* experiments. Throughout the study,
145 mussels were fed, twice per week, with two microalga species: *Scenedesmus obliquus* and
146 *Chlorella pyrenoidosa* (10^6 microalgae of each species per mussel and per day). Water (6 L)
147 was changed every week, after assessment of nitrites (0.12 ± 0.02 mg/L), nitrates ($34.32 \pm$
148 13.16 mg/L), and ammonium concentrations (0.03 ± 0.01 mg/L).

149 Hemolymph recovery was realised before feeding to not compromise cytometric analyses.
150 Hemolymph from ten mussels was withdrawn from adductor muscle with 0.3 mL syringes (30
151 G x 8 mm, Becton Dickinson, NJ, USA), kept on ice and pooled to obtain enough hemolymph
152 volume for both flow cytometric and ddPCR analysis. Cell concentration was determined with
153 KOVA slide (KOVA International, CA, USA), then adjusted to 375,000 cells per mL and
154 samples were separated into microplate (U-bottom, VWR, PA, USA) for flow cytometry and
155 into tubes for ddPCR. To discard cell debris and dead cells caused by sampling and starting
156 with healthy samples, plasma was removed after 20 minutes of hemocyte sedimentation (16
157 °C, in the dark [37]). Then, 190 μ L (or 200 μ L for UVb exposure) of modified Leibovitz 15
158 medium (L15, Sigma-Aldrich, MO, USA) were added for flow cytometry [38, 39]. For ddPCR
159 analyses, samples were placed into three 1.5 mL tubes with phosphate buffer saline (Sigma-
160 Aldrich) q.s. 500 μ L per tube.

161

162 2.2. Nature of stress and exposure

163 Stresses of different natures were selected: two chemical ones, represented by cadmium
 164 chloride (Agilent technologies, CA, USA) and an antibiotic from *Streptomyces globatus*
 165 named ionomycin (VWR); a physical stress with exposure to ultraviolet B (UVb, 254 nm, at a
 166 distance of 10 cm, tubes or plates closed to prevent evaporation and kept on ice, Camag,
 167 NC, USA). Finally, two human protozoan parasites were selected as biological stressors,
 168 *Toxoplasma gondii* (generously gifted by J.P. Dubey, USDA, Beltsville, USA) and
 169 *Cryptosporidium parvum* (INRAE, Val de Loire research center, France). Hemocytes were
 170 submitted to the different stress conditions for four hours (Table 1): i) to each solvent for
 171 control conditions, ii) to cadmium chloride (in phosphate buffer saline, Sigma-Aldrich), to
 172 ionomycin (in phosphate buffer saline with 0.2 % dimethylsulfoxide, ThermoFisher Scientific,
 173 MA, USA) or to protozoan solutions (in phosphate buffer saline), and iii) hemocytes were
 174 exposed to a UVb light for 0 (control), 3 or 10 minutes.

175

176 **Table 1:** Nature and intensities of each stress imposed on zebra mussel hemocytes.

177

Stress nature	Stress intensity			Reference
	null	medium	high	
Cadmium chloride µg/L	0	25	250	Sokolova <i>et al.</i> (2004) Fang <i>et al.</i> (2010)
Ionomycin µM	0	1	10	Opper <i>et al.</i> (2010)
Ultraviolet B min	0	3	10	Estevez-Calvar <i>et al.</i> (2013)
<i>Toxoplasma gondii</i> protozoa:cell ratio	0:1	1:25	1:5	Le Guernic <i>et al.</i> (2020)
<i>Cryptosporidium parvum</i> protozoa:cell ratio	0:1	1:1	5:1	Le Guernic <i>et al.</i> (2020)

178

179 2.3. Flow cytometric analyses

180 Analyses with flow cytometer were based on Barjhoux *et al.* [37] protocol with minor
 181 modifications. Briefly, hemocyte samples were placed in nine wells (three stress conditions x

182 three analyses), at 75.000 cells per well. After UVb exposure or immediately after adding
 183 other stressors, hemocytes were incubated four hours according to the following conditions:
 184 i) 2 µm diameter yellow-green fluorescent latex microbeads with a cell:bead ratio of 1:50 (16
 185 °C, in the dark, Polysciences Inc., PA, USA, [37]), ii) 0.5 µM caspases 3/7 green flow
 186 cytometric reagent (30 min, 16 °C, in the dark, Thermofisher Scientific), and iii) with 5 µM of
 187 4-amino-5-methylamino-2',7'-difluorofluorescein diacetate (DAF-fm, 30 min, 16 °C, in the
 188 dark, Thermofisher Scientific). Ten minutes before analyses, cells were removed into new
 189 wells with an anti-aggregate solution (Trypsin-EDTA solution, 0.0625 % final, Sigma-Aldrich
 190 [37]). Just before flow cytometric analyses, propidium iodide (PI, 1 % final, Sigma-Aldrich)
 191 was added to mark dead cells in double labeling with microbeads. The oocysts were
 192 removed from flow cytometry analyses by their size, complexity and autofluorescence,
 193 different from those of hemocytes, as previously describes [26]. Flow cytometric data were
 194 evaluated with 10,000 cells counted for each suspension sample thanks to Accuri™ C6
 195 SORP flow cytometer (Becton Dickinson, N.J., USA, with BD Accuri™ C6 software
 196 (v1.0.264.21)).

197 All immune markers and their corresponding method of analysis are described in Table 2.

198

199 **Table 2:** List of immune markers used in this study with corresponding method of analysis by flow

Immune markers	Abbreviations	Analysis methods	Units	Ex/Em (nm)
Hemocyte necrosis	PI+	percentage of cells labelled with propidium iodide among total cells	%	533/617
Phagocytic capacity	P.cap	percentage of cells that ingested at least one microbead among total cells	%	441/486
Phagocytic efficiency	P.eff	percentage of cells that ingested at least three microbead among total cells	%	441/486
Phagocytic avidity	P.nb	mean number of ingested beads by hemocytes that ingested at least one bead	number	441/486
Hemocyte apoptosis	Casp+	percentage of hemocytes with activated caspases 3/7, fluorescence is induced by cleavage of the associated DEVD peptide by activated caspases.	%	503/530
NO basal production	NO+	DAF-FM is modified to benzotriazole derivative (fluorescent) by the presence of nitric oxide in hemocytes	mean of fluorescence	495/515

200 cytometry, units, excitation wavelength (Ex), and emission wavelength (Em) used.

201

202 *2.4. Gene expression analysis*

203 2.4.1. RNA extraction and quantification

204 After exposure, the supernatant was removed from the sample after centrifugation (3,000 g,
205 5 min, 4 °C), and hemocytes were lysed with trizol reagent (Molecular Research Center Inc.,
206 OH, USA). At this stage, samples were frozen (-80 °C) for several weeks until resumption of
207 RNA extraction.

208 After thawing, chloroform (VWR) was added and vortexed with samples and then incubated
209 15 minutes at room temperature. The following steps have been carried out on ice to better
210 preserve the integrity of the biological material. The aqueous phase containing the nucleic
211 material was recovered after centrifugation (12,000 g, 8 min, 4 °C). Total RNA was
212 precipitated using isopropanol (10 minutes on ice, VWR) and the supernatant was kept after
213 centrifugation (12,000 g, 8 min, 4 °C). RNA was washed with 70 % ethanol (Charbonneaux-
214 Brabant Chimie, Reims, France), removed from the sample by centrifugation (7,500 g, 5 min,
215 4 °C). To remove ethanol, the pellet was air-dried on ice for about thirty minutes and was
216 then mixed with RNase free water (40 µL, United States Biological, MA, USA). A water bath
217 step was then respected (15 min, 57 °C) to dissolve RNA in water. RNA quantification was
218 assessed using Genova Nano spectrophotometer (Jenway, Staffordshire, United Kingdom),
219 and samples were homogenized at 100 ng RNA / µL. Quality index (absorbance ratio
220 OD_{260}/OD_{280}) was checked and samples that did not meet quality standards were removed
221 from the analyses. RNA purity was observed after electrophoresis and UV revelation (1.5 %
222 agarose gel and TBE buffer (0.5 X, Euromedex, Souffelweyersheim, France)). RNA samples
223 were frozen (-20 °C) until reverse transcription polymerase chain reaction.

224

225 2.4.2. Reverse transcription polymerase chain reaction

226 The reverse transcription was performed with 400 ng of total RNA using Verso cDNA
 227 Synthesis Kit (ThermoFisher Scientific). The reaction was conducted at 42 °C for 30 min
 228 followed by 2 minutes at 95 °C (Eppendorf Mastercycler EP Gradient S, Eppendorf France,
 229 Montesson, France). Reverse transcription polymerase chain reaction was not coupled with
 230 PCR to reduce final variability [35].

231

232 2.4.3. Droplet digital polymerase chain reaction analyses

233 All primers were obtained from Eurogentec (Liège, Belgium). All characteristics concerning
 234 the primers are present in table 3.

235

236 **Table 3:** List and characteristics of primers used for ddPCR analyses. SOD: superoxide dismutase;
 237 CAT: catalase; GPx: glutathione peroxidase; GST: glutathione-S-transferase; Casp3/7: caspases 3/7;
 238 Arp 2/3: actin-related protein 2/3; Cdc 42: cell division control 42; Gal 2: galectin 2. Primers were
 239 defined via the zebra mussel transcriptome compared to the sequences of the genes of interest in
 240 other bivalves of the databases.

Primers	Sequences (5'-3')	Amplicon size	Accession reference	Reference
SOD - F100 SOD - R100	GACAGCATGGCTTCCATGTG AGATTCTGGGCCAGTCAGAG	212 pb	AY377970.2	Geret <i>et al.</i> (2004)
CAT - F100 CAT - R100	ACGGCTATGGAAGCCACACG AGGTCGCGCATCGCATAGTC	157 pb	EF681763	Peuthert (2007)
Se - GPx - F100 Se - GPx - R100	TAGACCGCCGCTCGGTAATC CACTCCCAGCACCTGGAAAC	205 pb	EF194204.2	Doyen <i>et al.</i> (2008)
GST - pi -F100 GST - pi -R100	TCCGCTATATCTGCCTGGAC GCTCCTTCAGACCTGCTTTC	186 pb	DQ459993	Doyen <i>et al.</i> (2008)
MT - F200 MT - R200	GCGTTGAAACCGGTGATTGC GCCACAACAGTTGGGTTTGC	104 pb	U67347.1	Engelken and Hildebrandt (1999)
Casp3/7 - F Casp3/7 - R	CTCCGTCTAACATACAGGCC GCGTTGACAGAATTCAGTTGC	249 pb		Present study
Arp 2/3 - F Apr 2/3 - R	GCCAAAAGAATTAAGTACA GGGTATTCGGATGTGTGACA	241 pb		Present study
Cdc 42 - F Cdc 42 - R	GCCTAGTGTGGGCACTACAG ACCCACCACTGTCAACGGAC	247 pb		Present study
Gal 2 - F Gal 2 - R	GGAGTCATCATCCACCGTGA CAACGCGGGGCGGAAGCAGA	158 pb		Present study

241

242

243 cDNA samples, diluted to the tenth, were combined according to the manufacturer's
244 instructions on ice with a mix of primers (final concentration of 150 nM), Eva Green solution
245 (QX200 ddPCR EvaGreen Supermix RXN, BioRad, CA, USA) and osmosed water. Droplet
246 formation was then automatized with BioRad apparatus. PCR was then performed within 40
247 cycles (BioRad thermocycler). All reads with less than 13,000 accepted droplets were
248 removed from results [35]. The results are then expressed in gene copies per hemocyte, with
249 reference to the count carried out on each pool.

250 According to the Lievens *et al.* [35] study, many criteria for validating ddPCR data have been
251 taken into account and are presented in annex 1 (the number of accepted partition; the
252 number of outliers; the results of NTC, the mean number of copies per partition; the
253 percentage of "rain" and the maximum threshold).

254

255 *2.5. Statistical analysis*

256 Statistical analyses were performed using R 3.4.1 software. An analysis of variance
257 (ANOVA) was performed and normality and variance homogeneity were tested on its
258 residuals thanks to Shapiro-Wilk and Levene tests respectively ($\alpha \leq 5\%$). If normality and
259 homoscedasticity were respected, results from ANOVA were kept and followed by a post-hoc
260 test to discriminate significant differences inside each group (TukeyHSD with Holm
261 correction, $\alpha \leq 5\%$). If one of these criteria was not respected, ANOVA was replaced by a
262 Kruskal-Wallis test followed by Nemenyi post-hoc test ($\alpha \leq 5\%$). Statistics were performed
263 on ten biological replicas (pool).

264

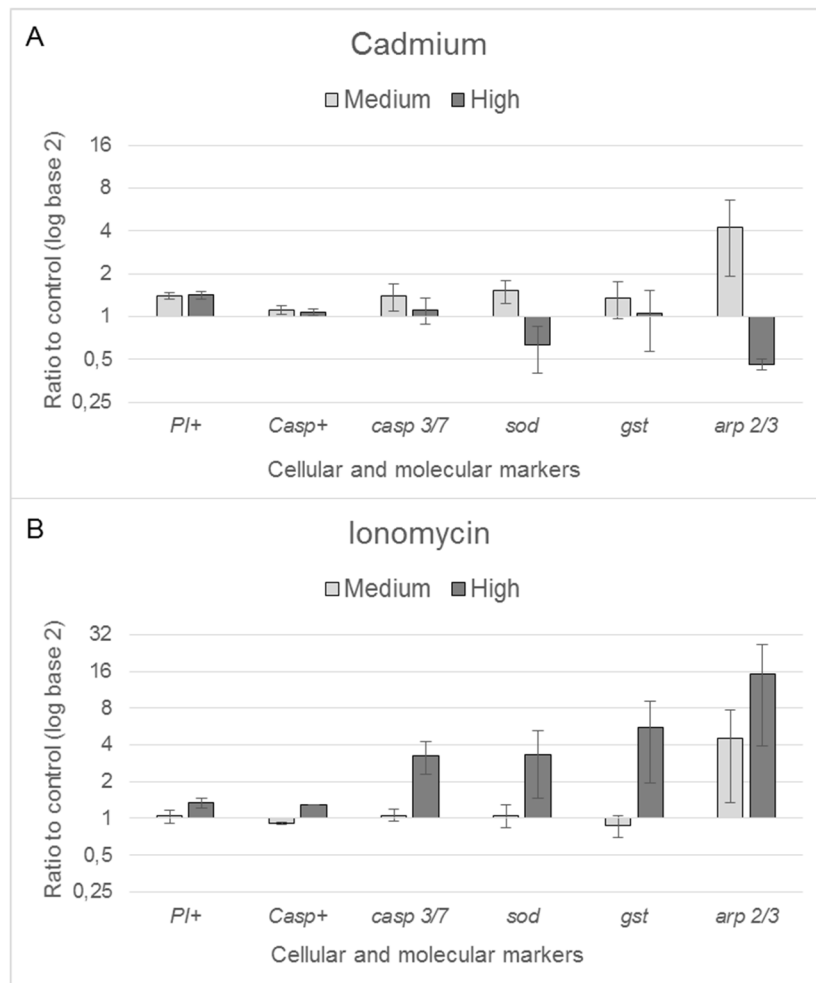
265 **3. Results**

266 *3.1. Chemical stress (cadmium and ionomycin)*

267 Both chemical stresses were cytotoxic. While cadmium induced hemocyte necrosis from
268 medium intensity, only the highest concentration of ionomycin was cytotoxic (Table 4).
269 Contrary to cadmium exposure, ionomycin induced other effects. First, a significant increase

270 of 3/7 caspases activation was observed when the hemocytes were exposed to 10 μ M
271 ionomycin compared to the control condition (evolution rate on means: +118.0 %, *p-value* =
272 8.40×10^{-3} , Table 4). Then, at this concentration, ionomycin also induced a decrease in all
273 cell markers linked to phagocytosis (the capacity (-23.7 %), efficiency (-56.6 %) and avidity (-
274 51.7 %) of this immune process, *p-value* = 4.30×10^{-2} ; *p-value* = 1.00×10^{-2} ; *p-value* = $1.00 \times$
275 10^{-2} , respectively, Table 4). The intermediate concentration of cadmium chloride induced a
276 higher *sod* gene expression compared to the control condition (evolution rate: -62.8 %, *p-*
277 *value* = 1.67×10^{-2} , Table 5) while no significant change was observed for the highest
278 concentration. No significant effect on gene expression was observed after ionomycin
279 exposure. Nonetheless, both chemical stressors tended to modify some gene expressions
280 (Figure 1). At the highest concentrations, opposite effects on gene expression were observed
281 between cadmium and ionomycin exposures compared to their control. While cadmium
282 would tend to decrease the gene expression of *sod* (-49.8 %), *gst* (-34.0 %), and *arp 2/3* (-
283 56.5 %), ionomycin would increase it (*sod* (+163.7 %), *gst* (+152.3 %), *casp 3/7* (+141.2 %)
284 and *arp 2/3* (+1075.7 %)). This is also observable when the ratios of the two stress intensities
285 is carried out with respect to the control condition for each pool (Figure 1).

286



287

288 **Figure 1:** Ratio to control (log base 2) for medium and high stresses intensities for each pool (mean ±
 289 SEM) submitted: A to cadmium and B to ionomycin. The light gray bars correspond to the ratio of
 290 medium intensity, and the dark gray ones correspond to the ratios of high intensity compared to the
 291 control. *PI+*: percentage of cells labelled with propidium iodide; *Casp+*: percentage of cells labelled
 292 with caspases 3/7 reagent; *casp 3/7*: capases 3/7; *sod*: superoxide dismutase; *gst*: glutathione-S-
 293 transferase; *arp 2/3*: actin-related protein 2/3.

294

295 **3.2. Physical stress (UVb)**

296 From the intermediate intensity (3 min), ultraviolet B induced significant necrosis and
 297 apoptosis (via the activation of 3/7 caspases, Table 4). These effects persisted at the highest
 298 intensity (10 min). The phagocytic avidity can be impaired by UVb exposure, after 3 (-26.9 %)

299 and 10 minutes (-34.3 %) exposure (p -value = 3.04×10^{-2} and p -value = 8.36×10^{-3}
300 compared to control, respectively, Table 4).

301

302 3.3. Biological stress (*C. parvum* and *T. gondii*)

303 Contrary to *C. parvum*, oocysts of *T. gondii* did not induce hemocyte necrosis (Table 4).

304 These two protozoa, at the highest stress intensity, disrupted phagocytic parameters: i)

305 phagocytic capacity (evolution rate: -20.3 %, p -value = 2.26×10^{-3} for *T. gondii* and -27.5 %, p -value = 1.20×10^{-2} for *C. parvum* compared to control), ii) efficiency (-41.0 %, p -value =

306 2.10×10^{-2} for *C. parvum* compared to control), and iii) avidity (-24.2 %, p -value = 7.78×10^{-3}

307 for *T. gondii* and -29.0 %, p -value = 3.47×10^{-2} for *C. parvum* compared to control, Table 4).

308 Finally, exposure to both biological stress at the highest stress intensity induced higher NO

309 basal production than without stress (evolution rate: +126.8 %, p -value = 2.50×10^{-3} and

310 +41.5 %, p -value = 1.24×10^{-2} , for *T. gondii* and *C. parvum* respectively, Table 4).

311 In addition to cellular effects, exposure to *C. parvum* oocysts, at the highest stress intensity,

312 induced increase of caspases 3/7 gene expression (+282.3 %, p -value = 3.50×10^{-2}

313 compared to control, Table 5). No effect on gene expression was observed after *T. gondii*

314 oocyst exposure.

315

316

317 **Table 4:** Immune marker responses after exposure to different stress at three conditions. Values are means \pm standard deviations ($N = 10$). For each immune
 318 parameter, and inside each stress, the difference between conditions is indicated by different letters with $a > b > c$ ($\alpha = 5\%$). Absence of letter means that no
 319 significant difference has been observed between these conditions. PI+: percentage of cells labelled with propidium iodide; Casp+: percentage of cells
 320 labelled with caspases 3/7 reagent; P.cap: percentage of cells that have ingested at least one microbead (phagocytic capacity); P.eff: percentage of cells that
 321 have ingested at least three microbeads (phagocytic efficiency); P.nb: mean number of ingested microbeads (phagocytic avidity); NO+: NO basal production.
 322

Immune marker		PI+ (%)		Casp+ (%)		P.cap (%)		P.eff (%)		P.nb		NO+ (FU)	
Stress	Condition	Mean \pm SD	Diff	Mean \pm SD	Diff	Mean \pm SD	Diff	Mean \pm SD	Diff	Mean \pm SD	Diff	Mean \pm SD	Diff
Cadmium chloride	0 μ g/L	16,0 \pm 3,8	b	10,5 \pm 1,9		53,0 \pm 3,0		30,4 \pm 3,3		5,9 \pm 0,6		12125 \pm 5453	
	25 μ g/L	21,9 \pm 2,9	a	11,8 \pm 2,8		54,7 \pm 2,9		31,3 \pm 3,1		5,8 \pm 0,4		13821 \pm 3481	
	250 μ g/L	22,2 \pm 2,7	a	11,3 \pm 2,8		54,5 \pm 3,0		31,6 \pm 3,5		6,0 \pm 0,7		16230 \pm 12251	
Ionomycin	0 μ M	19,4 \pm 3,5	b	13,0 \pm 6,2	b	42,3 \pm 5,4	a	22,1 \pm 4,1	a	5,8 \pm 0,7	a	9414 \pm 8966	
	1 μ M	19,4 \pm 4,9	ab	16,7 \pm 3,4	ab	38,8 \pm 11,2	ab	19,3 \pm 5,4	ab	5,3 \pm 0,8	a	8835 \pm 5408	
	10 μ M	25,4 \pm 2,7	a	28,4 \pm 6,4	a	32,5 \pm 4,9	b	9,6 \pm 3,4	b	2,8 \pm 0,4	b	6955 \pm 2753	
Ultraviolet B	0 min	17,2 \pm 2,8	b	18,5 \pm 3,1	b	50,8 \pm 7,7		28,7 \pm 8,6		6,7 \pm 1,2	a	8028 \pm 3151	
	3 min	22,9 \pm 2,9	a	29,7 \pm 3,1	a	46,8 \pm 5,3		20,5 \pm 4,4		4,9 \pm 0,8	b	8596 \pm 3011	
	10 min	25,9 \pm 1,8	a	34,9 \pm 3,3	a	47,3 \pm 2,8		18,7 \pm 3,1		4,4 \pm 0,9	b	8874 \pm 1062	
<i>Toxoplasma gondii</i>	0 : 1	12,8 \pm 6,1		8,5 \pm 1,4		65,4 \pm 7,3	a	52,1 \pm 8,1		6,6 \pm 1,4	a	8162 \pm 2240	b
	1 : 25	16,2 \pm 6,6		8,6 \pm 1,2		57,7 \pm 6,7	ab	42,1 \pm 14,3		5,5 \pm 0,5	ab	12273 \pm 5172	ab
	1 : 5	14,1 \pm 5,8		8,6 \pm 2,2		52,1 \pm 8,1	b	33,6 \pm 12,6		5,0 \pm 1,0	b	18468 \pm 8782	a
<i>Cryptosporidium parvum</i>	0 : 1	10,1 \pm 3,0	b	15,1 \pm 7,9		55,3 \pm 8,0	a	30,7 \pm 8,1	a	6,9 \pm 1,8	a	9269 \pm 2538	b
	1 : 1	13,4 \pm 3,1	ab	15,3 \pm 6,7		48,7 \pm 10,1	ab	23,3 \pm 10,0	ab	6,2 \pm 1,5	ab	11387 \pm 2522	ab
	5 : 1	14,1 \pm 4,0	a	15,2 \pm 6,2		40,1 \pm 14,0	b	18,1 \pm 10,1	b	4,9 \pm 1,1	b	13111 \pm 3039	a

323
 324
 325

326 **Table 5:** ddPCR results after exposure of hemocytes to different stress at three conditions. Values are means \pm standard error of the mean (N = between 8
 327 and 10). For each measured parameter, and inside each stress, the difference between conditions is indicated by different letters with a > b > c ($\alpha = 5\%$).
 328 Absence of letter means that no significant difference has been observed between these conditions. *sod*: superoxide dismutase; *cat*: catalase; *gpx*:
 329 glutathione peroxidase; *gst*: glutathione-S-transferase; *casp 3/7*: caspases 3/7; *arp 2/3*: actin-related protein 2/3; *cdc 42*: cell division control 42; *gal 2*: galectin

Molecular marker		<i>sod</i> (10-2 copies/hemocyte)		<i>cat</i> (10-2 copies/hemocyte)		<i>gpx</i> (10-2 copies/hemocyte)		<i>gst</i> (10-2 copies/hemocyte)		<i>mt</i> (10-2 copies/hemocyte)	
Stress	Condition	Mean \pm SEM	Diff	Mean \pm SEM	Diff	Mean \pm SEM	Diff	Mean \pm SEM	Diff	Mean \pm SEM	Diff
Cadmium chloride	0 μ g/L	17.61 \pm 2.63	ab	3.65 \pm 0.69		4.32 \pm 2.35		74.22 \pm 9.09		797.03 \pm 218.43	331
	25 μ g/L	23.78 \pm 3.70	a	5.35 \pm 0.65		3.16 \pm 0.68		88.09 \pm 18.86		1023.27 \pm 482.92	
	250 μ g/L	8.84 \pm 2.87	b	3.08 \pm 0.61		2.28 \pm 0.69		48.98 \pm 19.99		679.67 \pm 289.63	
Ionomycin	0 μ M	4.60 \pm 1.44		2.46 \pm 0.45		1.03 \pm 0.67		13.44 \pm 4.70		118.39 \pm 58.39	
	1 μ M	4.13 \pm 1.02		2.80 \pm 0.57		1.65 \pm 1.11		11.23 \pm 4.73		125.91 \pm 61.52	
	10 μ M	12.13 \pm 6.10		2.98 \pm 0.66		2.05 \pm 0.63		33.91 \pm 13.41		188.48 \pm 74.39	
Ultraviolet B	0 min	13.95 \pm 5.69		3.84 \pm 0.82		2.01 \pm 1.05		22.54 \pm 2.16		876.73 \pm 419.65	
	3 min	18.48 \pm 4.64		3.66 \pm 1.05		3.74 \pm 1.00		53.85 \pm 22.64		780.98 \pm 254.24	
	10 min	17.28 \pm 7.03		3.86 \pm 0.82		2.86 \pm 1.13		62.42 \pm 33.06		472.30 \pm 155.10	
<i>Toxoplasma gondii</i>	0 : 1	18.10 \pm 5.78		4.70 \pm 1.16		2.89 \pm 0.91		14.02 \pm 4.65		746.23 \pm 311.75	
	1 : 25	12.46 \pm 2.82		3.45 \pm 0.38		2.03 \pm 0.21		23.86 \pm 6.13		542.38 \pm 133.37	
	1 : 5	9.15 \pm 2.03		2.83 \pm 0.37		3.01 \pm 0.26		21.51 \pm 5.53		407.53 \pm 111.06	
<i>Cryptosporidium parvum</i>	0 : 1	29.86 \pm 14.99		5.75 \pm 2.06		4.25 \pm 1.40		80.42 \pm 38.29		138.85 \pm 51.56	
	1 : 1	44.85 \pm 30.81		5.25 \pm 0.91		9.07 \pm 5.41		110.91 \pm 67.98		350.18 \pm 112.44	
	5 : 1	77.24 \pm 32.27		11.64 \pm 4.73		31.24 \pm 26.51		176.15 \pm 74.80		370.46 \pm 88.04	

2.

Molecular marker		<i>casp 3/7</i> (10-2 copies/hemocyte)		<i>arp 2/3</i> (10-2 copies/hemocyte)		<i>cdc 42</i> (10-2 copies/hemocyte)		<i>gal 2</i> (10-2 copies/hemocyte)	
Stress	Condition	Mean \pm SEM	Diff	Mean \pm SEM	Diff	Mean \pm SEM	Diff	Mean \pm SEM	Diff
Cadmium chloride	0 μ g/L	4.61 \pm 1.08		43.72 \pm 24.10		5.54 \pm 1.62		543.39 \pm 243.64	
	25 μ g/L	4.95 \pm 0.68		85.49 \pm 22.86		4.75 \pm 1.47		622.05 \pm 309.40	
	250 μ g/L	4.17 \pm 0.86		19.02 \pm 3.02		4.82 \pm 3.95		388.71 \pm 204.43	
Ionomycin	0 μ M	1.87 \pm 0.69		2.35 \pm 2.17		0.67 \pm 0.32		84.50 \pm 49.09	
	1 μ M	2.17 \pm 0.90		6.06 \pm 5.91		0.52 \pm 0.20		92.13 \pm 50.47	
	10 μ M	4.51 \pm 1.10		27.63 \pm 19.16		0.91 \pm 0.28		152.18 \pm 58.40	
Ultraviolet B	0 min	8.88 \pm 5.09		19.09 \pm 18.60		1.38 \pm 0.25		322.69 \pm 214.44	
	3 min	7.15 \pm 3.64		59.96 \pm 39.44		3.60 \pm 0.66		212.86 \pm 147.02	
	10 min	3.01 \pm 0.75		5.00 \pm 3.73		0.62 \pm 0.27		124.87 \pm 92.37	
<i>Toxoplasma gondii</i>	0 : 1	2.55 \pm 0.87		59.62 \pm 26.00		7.28 \pm 1.81		278.70 \pm 64.58	
	1 : 25	2.59 \pm 0.50		43.27 \pm 13.72		5.55 \pm 0.86		465.59 \pm 82.17	
	1 : 5	2.16 \pm 0.52		36.11 \pm 10.35		4.82 \pm 1.27		300.97 \pm 70.90	
<i>Cryptosporidium parvum</i>	0 : 1	2.48 \pm 0.86	b	52.78 \pm 17.88		5.88 \pm 0.41		371.40 \pm 128.08	
	1 : 1	6.94 \pm 1.84	ab	52.07 \pm 11.35		4.42 \pm 0.66		384.06 \pm 135.49	
	5 : 1	9.48 \pm 3.43	a	58.51 \pm 9.05		3.35 \pm 0.57		334.38 \pm 58.13	

332 4. Discussion

333 4.1. Chemical stress induced different effects

334 Many authors have investigated the effects of cadmium on animals, including bivalves.
335 These organisms are known to accumulate high doses of this metal, and thus are used as
336 sentinel species for metallic biomonitoring [40, 41]. The effects of Cd on mussel physiology
337 are numerous and complex, depending on the dose and bioavailability of metal as well as the
338 size and reproductive status of mussels [41, 42]. Nonetheless, several effects on bivalve
339 hemocytes are often highlighted by authors. Cadmium has deleterious effects on hemocyte
340 viability, on the one hand being pro-apoptotic, and on the other hand by inducing necrosis
341 [43-45]. This impact was observed in this study with an increase in necrosis following
342 hemocyte exposure to 25 µg/L Cd and more, but no effect on hemocyte apoptosis (assessed
343 by activation of caspases 3/7) was reported contrary to the literature [43-45]. Nevertheless,
344 the conditions for exposing cells to cadmium were different. Also, Sokolova *et al.* [43] have
345 reported the induction of apoptosis after exposure to Cd without caspase activation. Cd²⁺
346 would induce apoptosis in a caspase-independent manner [46]. Disturbance of SOD
347 production after exposure to Cd is a common feature, highlighted in our study and by several
348 authors [44, 45, 47]. Cadmium can also alter immune functions in bivalves. Indeed, this metal
349 can disturb cytoskeletal actin and thus modify adhesion and phagocytic process, leading
350 sometimes to immune suppression [48-50]. No significant effect of Cd on *D. polymorpha*
351 immune function was reported in this study, probably explained by several factors. First, Cd
352 concentrations were higher than those of other studies with effects on immune functions [48,
353 50]. Then, exposure conditions and duration were different (*ex vivo* versus *in vivo*), and
354 finally the necrosis and phagocytosis double-labelling allowed to remove dead cells for
355 phagocytic parameters. It should be noted that despite the importance of inter-pool
356 variations, the highest concentration of cadmium seemed to decrease the expression of part
357 of the genes studied. The expression levels of *sod*, *gpx*, *gst*, *arp 2/3* and *gal2* decreased at
358 the highest concentration of Cd.

359 Results obtained after exposure of hemocytes to ionomycin are consistent with those of the
360 literature. Ionomycin is a Ca^{2+} ionophore that increases intracellular Ca^{2+} , leading to pH
361 modifications inside cells, such as hemocytes [51-54]. These effects induce the phenomenon
362 of programmed death, named apoptosis. This was revealed in our study, with a dose-
363 dependent activation of caspases 3/7 inside hemocytes and with an increase of *casp 3/7*
364 gene expression, as revealed in several studies [51-53]. Ionomycin induces the intrinsic
365 pathway of apoptosis by stimulating the activation of the initiator caspase 9 and executor
366 caspases (caspases 3 and 7) as well as the release of cytochrome c from mitochondria, thus
367 limiting the anti-apoptotic factors [52]. These authors highlighted that three hours after
368 ionomycin exposure is enough to induce apoptosis and necrosis, as compared to our results
369 [52, 55]. Few studies, however, have observed a decrease in phagocytosis after cell
370 exposure to this antibiotic, contrary to our results. These decreases of phagocytic
371 parameters assessed in mussel hemocyte can be explained by several hypotheses. First, it
372 has been noticed that a part of phagocytosis can be Ca^{2+} -dependent, at least in human
373 neutrophils [56-58]. Ca^{2+} would act as a second messenger for phagocytosis and superoxide
374 anion formation [59]. The disruption in intracellular transport caused by ionomycin may thus
375 alter this immune function. Secondly, apoptotic hemocytes, rapidly ingested after ionomycin
376 exposure [55], can be preferred to microbeads by viable hemocytes. Finally, although
377 propidium iodide-labelled hemocytes (in necrosis) are removed from the phagocytosis assay,
378 those labelled with caspase 3/7 reagent are not. The increase of apoptotic cells after
379 exposure to ionomycin may directly lead to a decrease in phagocytosis of the microbeads.
380 This hypothesis was supported by a tendency to increase the expression level of caspases
381 3/7 when the ionomycin concentration rises up. This was also observed with other genes
382 under study, such as *sod*, *gst*, *arp 2/3* and *gal2*, with induction factor compared to control
383 condition from 2 to 11 for *arp 2/3*.

384 Cellular activities are coordinated by the information contained in genetic material. This
385 creates a close link between gene expression and protein activity. However, many stages of
386 gene regulation exist. In eukaryotes, they are found at the level of chromatin structure,

387 transcription, maturation of mRNAs and their temporal stability, at the level of translation and
388 post-translational modifications that can also impact the activity of encoded protein [60]. In
389 this study, no significant correlation between gene expression (*casp 3/7* on one side and *arp*
390 *2/3*, *cdc 42* and *gal 2* on the other) and protein or cellular activity (activation of caspases 3/7
391 and phagocytosis, respectively) had been observed. However, the comparison of the results
392 obtained by flow cytometry and by ddPCR can provide information on the synergy of action
393 of different studied stress. Indeed, the higher mortality rate (necrosis and apoptosis analysed
394 by flow cytometry), caused by high concentrations of cadmium and ionomycin, could explain
395 the modifications, although not significant, of the gene expressions (*sod* [$p\text{-value} = 1.67 \times 10^{-2}$],
396 *gst* and *arp 2/3* for cadmium at 250 µg/L; *sod*, *gst*, *casp 3/7*, and *arp 2/3* for ionomycin at
397 10 µM). Both stress, at the highest intensities, had opposite effects. Cadmium tends to
398 decrease gene expression while ionomycin would increase it (Figure 1). Despite a similar
399 cytotoxic effect, this suggests that these two chemical stressors do not have the same
400 modes of action. Cadmium is known to induce oxidative stress in cells and this oxidative
401 stress would certainly impair the integrity of hemocytes. This could be the cause of the
402 insignificance of the results in gene expression for the highest concentration of cadmium.
403 Otherwise, activation of calcium channels ultimately leads to cell apoptosis by ionomycin
404 further impacts immune cell functions and could induce the expression of genes linked to
405 phagocytosis. Nevertheless, *arp 2/3*, *cdc 42* and *gal 2* have other functions than those
406 related to phagocytosis. *cdc 42* and *arp 2/3* play roles in the cellular cycle and maintaining
407 cellular integrity, while Gal 2 regulates the inflammation, cell homeostasis and cell growth
408 processes [28, 61, 62].

409 Regarding caspases 3/7, no similarity of effect was observed between the results obtained in
410 cytometry and in ddPCR. It is likely that the stress exerted on the hemocytes only influenced
411 the inactive procaspases already present in the hemocytes without inducing modulation of
412 their gene expression. Ionomycin, which induced significant activation of 3/7 caspases at the
413 highest concentration, also seemed to induce the gene expression of these caspases.

414 However, this induction was just a trend. A larger number of biological replicas could improve
415 the discernment of these effects.

416

417 4.2. A cytotoxic physical stress

418 The physical stress, ultraviolet B, induced hemocyte necrosis and apoptosis as well as
419 decrease of phagocytic avidity. These effects are in agreement with those reported by
420 several studies. Clydesdale *et al.* [63] have synthesized effects of ultraviolet on human cells,
421 such as apoptosis and necrosis induction and immunomodulatory effects. These effects have
422 been also reported on aquatic invertebrates, such as crustaceans [64, 65], and bivalves [66,
423 67]. Apoptosis caused by UVb light seems to be activated by the intrinsic way, *via* DNA
424 fragmentation, caspases activation, modulation of cytoplasmic calcium, increase in
425 mitochondrial membrane potential, and modifications of pro- and anti-apoptotic gene
426 expression [64-67]. UV are thus immune toxic as well as genotoxic, as observed by Rank
427 and Jensen [68] on hemocytes of *Mytilus edulis* with the same emission wave length (254
428 nm).

429

430 4.3. Different protozoa, different interactions?

431 Few studies are looking at protozoan:hemocyte interaction when the host is paratenic.
432 Previously, we have described the effects of *C. parvum* and *T. gondii* oocysts on hemocytes
433 of two bivalves, *D. polymorpha* and *M. edulis* [26]. The vast majority of effects previously
434 observed have been also reported in this study. We observed a decrease of phagocytic
435 parameters after exposure to both protozoa and an increase of basal NO production after
436 exposure to *C. parvum* oocysts [26].

437 Decreases of phagocytosis can be explained by the establishment of immune process by
438 hemocytes. Palos Ladeiro *et al.* [15] have reported that hemocytes of zebra mussels are able
439 to engulf *C. parvum* oocysts. This protozoan has been used as a marker of phagocytosis for

440 *Crassostrea virginica* [69]. The *T. gondii* oocyst being about twice the size of that of *C.*
441 *parvum*, it seems unlikely that it can be ingested. Nevertheless, immune cells of zebra
442 mussels can aggregate around the oocysts to realize the phenomenon of encapsulation [70].
443 These immune functions are accompanied by an overproduction of NO, which can be
444 produced against microorganisms [71]. Not observed in previous studies [26, 70], oocysts of
445 *C. parvum* induced hemocyte necrosis. Although the number of activated caspases 3/7 did
446 not differ among the condition of *C. parvum* exposure, this protozoan induced gene
447 expression of these enzymes. Terahara and Takahashi [72] have reported an over-
448 expression of caspase genes as an immune response of mollusc host against parasite
449 infection. Induction of apoptosis with parasitic exposure was often highlighted in edible
450 bivalves [73-75]. If this exposure is prolonged, apoptosis can even be suppressed [75, 76].
451 Nonetheless, these results were reported with specific hosts (i.e. *Crassostrea virginica* for
452 *Perkinsus marinus*) and not for paratenic hosts. We previously evidenced that *T. gondii* was
453 able to activate caspases 3/7 in hemocytes of the blue mussel [26]. Functional changes in
454 the immunity of *Dreissena polymorpha* after exposure to both protozoa were more numerous
455 than changes in the expression of genes linked to the immune system. *T. gondii* and *C.*
456 *parvum* oocysts did not significantly impact immune gene expression in hemocytes of zebra
457 mussels. Many authors have reported over-expression of genes following the exposure of
458 molluscs to parasites, particularly lectins MTs or GSTs [77-81]. However, a big difference
459 between these results and ours is that all of these studies were focused on a specific host,
460 which could be subject to infection by the parasite. Recently, Leprêtre *et al.* [82] compared
461 the protein expression profiles on the hemolymphatic compartments of zebra mussels
462 between unexposed samples and others exposed to *C. parvum*. The changes in protein
463 expression after exposure to this protozoan were mainly located at the cell level, and
464 included proteins linked to cell motility, to the internalization and destruction of pathogens,
465 apoptosis, and immune signalization, including an up-regulation of arp 2/3 complex, cdc 42
466 and a galectin proteins, confirming our results obtained in flow cytometry. Nonetheless,
467 Leprêtre *et al.* [82] emphasized the importance of the plasma compartment in the immune

468 responses of the zebra mussel. Some immune proteins have been up or down modulated
469 after a 4-hour exposure to *C. parvum*, including arp 2/3, galectin, serine, clathrin and
470 complement-like proteins. In our study, no correlation between the disturbances of
471 phagocytosis by hemocytes exposed to both biological stressors and the expression of
472 genes potentially linked to this process (*arp 2/3*, *cdc 42*, *gal 2*) has been reported. This can
473 be explained by the fact that phagocytosis included several immune processes, such as
474 vesicle transport, cell motility, cell adhesion, cytoskeleton organisation, etc., and by a large
475 number of genes and proteins involved in this defence system [32, 83]. The complementarity
476 between the two hemolymphatic compartments (plasma and hemocytes) is significant. Thus,
477 the exposure to the various stresses carried out without plasma may block this
478 complementarity, in particular for the recognition of pathogens [32]. It would be interesting to
479 expose the hemolymph of the zebra mussels as a whole before separating plasma and cells.
480 The cellular and molecular markers studied on hemocytes after this protocol may be different
481 from those analysed without the plasma-hemocyte complementarity.

482

483 4.4. Improvements and future tracks

484 It is important to note the high variability of gene expression among pools. This variability
485 was reported between pools inside a condition (SEM) and between stress conditions (Table
486 5). This variability was not identical according to the studied genes. Comparing gene
487 expression for control conditions of each exposure, ratio of 2 (*cat*) to 30 (*arp 2/3*) were
488 reported. Expression levels of *casp 3/7*, *cat*, *gal 2* and *mt* seemed to be less variable than
489 those of *sod*, *gpx*, *gst*, *arp 2/3* and *cdc 42* genes. This strong variability can be explained by
490 three hypotheses. The main reason for explaining this variation would be linked to
491 interactions between hemocytes from different individuals within the same pool. We decided
492 to work with pool to have enough hemocytes to expose the same sample and analyse all the
493 markers on the same group. Barjhoux *et al.* [37] have observed a reduction in the variability
494 of the immune markers analysed by flow cytometry in this species. However, the interaction

495 between hemocytes from different individuals may have caused this strong variability in
496 molecular markers. Volume improvements on the hemolymph sample for this species would
497 make it possible to analyse the expression of several genes individually.

498 Although the production of proteins in response to external stimuli comes from a rapid
499 expression of genes, the regulatory steps induce a strong variation in the kinetics between
500 gene expression and expression of the protein formed [84]. This is especially important when
501 studying a biological process (i.e. phagocytosis), involving a large number of molecules [85].
502 Thus, after exposure to a xenobiotic, the expression profiles vary depending on the post-
503 exposure period at which analyses are carried out [86]. The kinetics of analysis can also
504 modify our gene expression results. In our study, hemocytes were exposed for four hours
505 before genetic material extraction. However, this duration may be too long to observe a
506 modulation of gene expression. Morga *et al.* [80] highlighted over-expression of several
507 genes (*gst*, *p450 cytochrome*, *galectin* and *lysozyme*) in hemocytes of *Ostrea edulis* after
508 one hour *in vitro* exposure to *Bonamia ostreae*. These expressions were returned to basic
509 levels 2 or 4 hours after exposure [80]. The kinetics of exposure (four hours exposure) could
510 explain that the stress had more effects on the hemocyte functions than on the gene
511 expression. On the other hand, this modulation of gene expression can appear later than four
512 hours post-stress. Indeed, Farcy *et al.* [87] reported, in *C. gigas*, an over-expression of genes
513 coding for stress proteins 4 to 8 hours after heat shock, this expression returning to basic
514 levels after 48 hours. Correlatively, the expression of immune genes from the same bivalve
515 (*C. gigas*) was stimulated after 24 to 48 hours after exposure to a virus (Ostreid herpesvirus
516 1 mVar), and the return to baseline levels took place after 120 hours [88]. Nonetheless, these
517 two studies were conducted *in vivo*. Although the 4-hours post-stress exposure period was
518 established in previous work [26], this sampling time is ideal for cytometric analyses and to
519 keep healthy cells. However, for the analysis of gene expression, it would be interesting to
520 follow the kinetics of post-exposure expression over several hours to confirm our results.

521 These biases are mainly due to a narrow vision of the complex processes involved in the
522 response to stress. In the last decade, the omics approaches (from genomics to

523 interactomics) permit to obtain the entire toxicological signature of a xenobiotic. These
524 approaches would allow a larger view of the interaction between chemical, physical and
525 biological stress and hemocytes. Even if the mechanisms which govern remains unclear,
526 they are mainly controlled at the protein level (i.e. cytokines, immune recognition factors,
527 etc.). The proteomic approach would answer several questions: i) do the reductions in
528 phagocytosis parameters following exposure to biological stress and ionomycin have the
529 same mechanisms? and ii) despite fairly similar cellular effects, are the oocysts of *T. gondii*
530 and *C. parvum* managed in the same way by cell (recognition, destruction, etc.)? Finally,
531 proteomics, flow cytometry and ddPCR could be performed jointly on proteins, activities and
532 genes of interest previously chosen, during post-exposure kinetics to characterize the links
533 between the different levels of organization.

534

535 **5. Conclusion**

536 One of the challenges of ecotoxicology today is to be able to link molecular biomarkers,
537 cellular biomarkers and pathophysiological 'endpoints' with higher level ecological
538 consequences [89]. This requires a good understanding of the effects of various xenobiotics
539 as well as the responses of the organisms subjected to them. This study aimed to
540 understand the immune response capabilities of zebra mussel hemocytes to various stresses
541 (chemical, physical and biological), *via* cellular and molecular markers. Ultraviolet b are
542 cytotoxic since they induced necrosis, apoptosis and reduced the phagocytosis of
543 hemocytes. The oocysts from both protozoa induced nitric oxide production by exposed
544 hemocytes and phagocytic parameters decreases, as already reported. Nonetheless,
545 hemocytes appear to be more affected by oocysts of *C. parvum*, suggesting a difference in
546 the management of these two protozoa. New gene sequences related to the immune system
547 of zebra mussels were obtained in connection with apoptosis and cell motility and
548 phagocytosis. Omics approaches could provide more details on the links between gene
549 expression and cellular activity, particularly for complex processes such as phagocytosis. It

550 would be interesting to encompass the entire immune system by taking into account the
551 plasma compartment. Indeed, plasma contains immune proteins which can play an important
552 role in defence against xenobiotics [90, 91]. These two compartments could work together to
553 eliminate some microorganisms [92]. The responses obtained on the entire hemolymphatic
554 compartment may be different than those assessed exclusively at the cellular level.

555

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561

562 **References**

- 563 1. Van der Oost R, Beyer J, Vermeulen NPE. Fish bioaccumulation and biomarkers in
564 environmental risk assessment: a review. *Environ Toxicol Pharmacol*. 2003 13:57-149.
- 565 2. Forbes VE, Palmqvist A, Bach L. The use and misuse of biomarkers in ecotoxicology.
566 *Environmental toxicology and chemistry*. 2006 25:272-80.
- 567 3. Kouassi É, Revillard J, Fournier M, Ayotte P, Roy R, Brousseau P, et al. Système immunitaire.
568 *Environnement et santé publique: Fondements et pratiques*. 2003.
- 569 4. Song L, Wang L, Qiu L, Zhang H. Bivalve immunity. In: Söderhäll K, editor. *Invertebrate*
570 *immunity*: Springer; 2010, p. 44-65.
- 571 5. Delves PJ, Roitt IM. The immune system. *New England journal of medicine*. 2000 343:37-49.
- 572 6. Su T-P, London ED, Jaffe JH. Steroid binding at sigma receptors suggests a link between
573 endocrine, nervous, and immune systems. *Science*. 1988 240:219-21.
- 574 7. Samanta P, Im H, Na J, Jung J. Ecological risk assessment of a contaminated stream using
575 multi-level integrated biomarker response in *Carassius auratus*. *Environmental Pollution*. 2018
576 233:429-38.
- 577 8. Kim W-K, Lee S-K, Park J-W, Choi K, Cargo J, Schlenk D, et al. Integration of multi-level
578 biomarker responses to cadmium and benzo[k]fluoranthene in the pale chub (*Zacco platypus*).
579 *Ecotoxicology and Environmental Safety*. 2014 110:121-8.
- 580 9. Lebrun JD, De Jesus K, Rouillac L, Ravelli M, Guenne A, Tournebize J. Single and combined
581 effects of insecticides on multi-level biomarkers in the non-target amphipod *Gammarus fossarum*
582 exposed to environmentally realistic levels. *Aquatic Toxicology*. 2020 218:105357.
- 583 10. Damásio J, Fernández-Sanjuan M, Sánchez-Avila J, Lacorte S, Prat N, Rieradevall M, et al.
584 Multi-biochemical responses of benthic macroinvertebrate species as a complementary tool to
585 diagnose the cause of community impairment in polluted rivers. *Water Research*. 2011 45:3599-613.
- 586 11. Quinn A, Gallardo B, Aldridge DC. Quantifying the ecological niche overlap between two
587 interacting invasive species: the zebra mussel (*Dreissena polymorpha*) and the quagga mussel

588 (*Dreissena rostriformis bugensis*). Aquatic Conservation: Marine and Freshwater Ecosystems. 2014
589 24:324-37.

590 12. Besse J-P, Geffard O, Coquery M. Relevance and applicability of active biomonitoring in
591 continental waters under the Water Framework Directive. TrAC Trends in Analytical Chemistry. 2012
592 36:113-27.

593 13. Salazar MH, Salazar SM. Using caged bivalves to characterize exposure and effects associated
594 with pulp and paper mill effluents. Water Science and Technology. 1997 35:213-20.

595 14. Bervoets L, Voets J, Covaci A, Chu S, Qadah D, Smolders R, et al. Use of transplanted zebra
596 mussels (*Dreissena polymorpha*) to assess the bioavailability of microcontaminants in Flemish surface
597 waters. Environmental science & technology. 2005 39:1492-505.

598 15. Palos Ladeiro M, Bigot-Clivot A, Geba E, Le Foll F, Le Guernic A, Leprêtre M, et al. Mollusc
599 bivalves as indicators of contamination of water bodies by protozoan parasites. In: Elias S, editor.
600 Reference Module in Earth Systems and Environmental Sciences: Elsevier; 2018.

601 16. Palos-Ladeiro M, Aubert D, Villena I, Geffard A, Bigot A. Bioaccumulation of human
602 waterborne protozoa by zebra mussel (*Dreissena polymorpha*): interest for water biomonitoring.
603 water research. 2014 48:148-55.

604 17. Kraak MH, Martin CT, Peeters WH, De Kock WC. Biomonitoring of heavy metals in the
605 Western European rivers Rhine and Meuse using the freshwater mussel *Dreissena polymorpha*.
606 Environmental pollution. 1991 74:101-14.

607 18. Binelli A, Della Torre C, Magni S, Parolini M. Does zebra mussel (*Dreissena polymorpha*)
608 represent the freshwater counterpart of *Mytilus* in ecotoxicological studies? A critical review.
609 Environmental Pollution. 2015 196:386-403.

610 19. Kerambrun E, Palos Ladeiro M, Bigot-Clivot A, Dedourge-Geffard O, Dupuis E, Villena I, et al.
611 Zebra mussel as a new tool to show evidence of freshwater contamination by waterborne
612 *Toxoplasma gondii*. Journal of applied microbiology. 2016 120:498-508.

613 20. Binelli A, Bacchetta R, Vailati G, Galassi S, Provini A. DDT contamination in Lake Maggiore (N.
614 Italy) and effects on zebra mussel spawning. Chemosphere. 2001 45:409-15.

615 21. Olabarrieta I, L'azou B, Yuric S, Cambar J, Cajaraville M. *In vitro* effects of cadmium on two
616 different animal cell models. Toxicology in vitro. 2001 15:511-7.

617 22. Tennant I, Pound J, Marr L, Willems J, Petrova S, Ford C, et al. Innate recognition of apoptotic
618 cells: novel apoptotic cell-associated molecular patterns revealed by crossreactivity of anti-LPS
619 antibodies. Cell Death & Differentiation. 2013 20:698-708.

620 23. Stankov K, Bogdanovic G, Stankov S, Draskovic D, Grubor-Lajsic G, Spasic M, et al. Expression
621 analysis of genes involved in apoptosis, proliferation and endoplasmic reticulum stress in
622 ionomycin/PMA treated Jurkat cells. Journal of BU ON: official journal of the Balkan Union of
623 Oncology. 2012 17:369-76.

624 24. Steinert SA. Contribution of apoptosis to observed dna damage in mussel cells. Marine
625 Environmental Research. 1996 42:253-9.

626 25. Rana S, Byrne SN, MacDonald LJ, Chan CY-Y, Halliday GM. Ultraviolet B suppresses immunity
627 by inhibiting effector and memory T cells. The American journal of pathology. 2008 172:993-1004.

628 26. Le Guernic A, Geffard A, Le Foll F, Ladeiro MP. Comparison of viability and phagocytic
629 responses of hemocytes withdrawn from the bivalves *Mytilus edulis* and *Dreissena polymorpha*, and
630 exposed to human parasitic protozoa. International Journal for Parasitology. 2020 50:75-83.

631 27. Müllerbner A, Dorighello GG, Kozlov AV, Duvigneau JC. Interaction between mitochondrial
632 reactive oxygen species, heme oxygenase, and nitric oxide synthase stimulates phagocytosis in
633 macrophages. Frontiers in medicine. 2018 4:252.

634 28. Machesky LM, Gould KL. The Arp2/3 complex: a multifunctional actin organizer. Current
635 opinion in cell biology. 1999 11:117-21.

636 29. Almkvist J, Karlsson A. Galectins as inflammatory mediators. Glycoconjugate journal. 2002
637 19:575-81.

638 30. Hemalatha A, Mayor S. Recent advances in clathrin-independent endocytosis.
639 F1000Research. 2019 8.

- 640 31. Villeneuve C, Baricault L, Canelle L, Barboule N, Racca C, Monsarrat B, et al. Mitochondrial
641 proteomic approach reveals galectin-7 as a novel BCL-2 binding protein in human cells. *Molecular*
642 *biology of the cell*. 2011 22:999-1013.
- 643 32. Leprêtre M, Almunia C, Armengaud J, Salvador A, Geffard A, Palos-Ladeiro M. The immune
644 system of the freshwater zebra mussel, *Dreissena polymorpha*, decrypted by proteogenomics of
645 hemocytes and plasma compartments. *Journal of proteomics*. 2019 202:103366.
- 646 33. Sanders R, Huggett JF, Bushell CA, Cowen S, Scott DJ, Foy CA. Evaluation of digital PCR for
647 absolute DNA quantification. *Analytical chemistry*. 2011 83:6474-84.
- 648 34. Huggett JF, Foy CA, Benes V, Emslie K, Garson JA, Haynes R, et al. The digital MIQE guidelines:
649 minimum information for publication of quantitative digital PCR experiments. *Clinical chemistry*.
650 2013 59:892-902.
- 651 35. Lievens A, Jacchia S, Kagkli D, Savini C, Querci M. Measuring digital PCR quality: performance
652 parameters and their optimization. *PLoS One*. 2016 11:e0153317.
- 653 36. Miotke L, Lau BT, Rumma RT, Ji HP. High sensitivity detection and quantitation of DNA copy
654 number and single nucleotide variants with single color droplet digital PCR. *Analytical chemistry*.
655 2014 86:2618-24.
- 656 37. Barjhoux I, Rioult D, Geffard A, Ladeiro MP. A new protocol for the simultaneous flow
657 cytometric analysis of cytotoxicity and immunotoxicity on zebra mussel (*Dreissena polymorpha*)
658 hemocytes. *Fish & Shellfish Immunology*. 2020 98:224-35.
- 659 38. Quinn B, Costello MJ, Dorange G, Wilson JG, Mothersill C. Development of an in vitro culture
660 method for cells and tissues from the zebra mussel (*Dreissena polymorpha*). *Cytotechnology*. 2009
661 59:121-34.
- 662 39. Evariste L, Auffret M, Audonnet S, Geffard A, David E, Brousseau P, et al. Functional features
663 of hemocyte subpopulations of the invasive mollusk species *Dreissena polymorpha*. *Fish & Shellfish*
664 *Immunology*. 2016 56:144-54.
- 665 40. Marie V, Baudrimont M, Boudou A. Cadmium and zinc bioaccumulation and metallothionein
666 response in two freshwater bivalves (*Corbicula fluminea* and *Dreissena polymorpha*) transplanted
667 along a polymetallic gradient. *Chemosphere*. 2006 65:609-17.
- 668 41. Roseman EF, Mills EL, Rutzke M, Gutenmann WH, Lisk DJ. Absorption of cadmium from water
669 by North American zebra and quagga mussels (*Bivalvia: Dreissenidae*). *Chemosphere*. 1994 28:737-
670 43.
- 671 42. Amiard-Triquet C, Berthet B, Metayer C, Amiard JC. Contribution to the ecotoxicological
672 study of cadmium, copper and zinc in the mussel *Mytilus edulis*. *Marine Biology*. 1986 92:7-13.
- 673 43. Sokolova I, Evans S, Hughes F. Cadmium-induced apoptosis in oyster hemocytes involves
674 disturbance of cellular energy balance but no mitochondrial permeability transition. *Journal of*
675 *Experimental Biology*. 2004 207:3369-80.
- 676 44. Gómez-Mendikute A, Cajaraville MP. Comparative effects of cadmium, copper, paraquat and
677 benzo[a]pyrene on the actin cytoskeleton and production of reactive oxygen species (ROS) in mussel
678 haemocytes. *Toxicology in Vitro*. 2003 17:539-46.
- 679 45. Qin Q, Qin S, Wang L, Lei W. Immune responses and ultrastructural changes of hemocytes in
680 freshwater crab *Sinopotamon henanense* exposed to elevated cadmium. *Aquatic Toxicology*. 2012
681 106-107:140-6.
- 682 46. Kiss T. Apoptosis and its functional significance in molluscs. *Apoptosis*. 2010 15:313-21.
- 683 47. Fang Y, Yang H, Wang T, Liu B, Zhao H, Chen M. Metallothionein and superoxide dismutase
684 responses to sublethal cadmium exposure in the clam *Macraa veneriformis*. *Comparative*
685 *Biochemistry and Physiology Part C: Toxicology & Pharmacology*. 2010 151:325-33.
- 686 48. Bouilly K, Gagnaire B, Bonnard M, Thomas-Guyon H, Renault T, Miramand P, et al. Effects of
687 cadmium on aneuploidy and hemocyte parameters in the Pacific oyster, *Crassostrea gigas*. *Aquatic*
688 *toxicology*. 2006 78:149-56.
- 689 49. Ivanina AV, Hawkins C, Sokolova IM. Immunomodulation by the interactive effects of
690 cadmium and hypercapnia in marine bivalves *Crassostrea virginica* and *Mercenaria mercenaria*. *Fish*
691 *& Shellfish Immunology*. 2014 37:299-312.

692 50. Rocha TL, Gomes T, Cardoso C, Letendre J, Pinheiro JP, Sousa VS, et al. Immunocytotoxicity,
693 cytogenotoxicity and genotoxicity of cadmium-based quantum dots in the marine mussel *Mytilus*
694 *galloprovincialis*. Marine Environmental Research. 2014 101:29-37.

695 51. Yamada H, Seki G, Taniguchi S, Uwatoko S, Suzuki K, Kurokawa K. Effect of ionomycin on cell
696 pH in isolated renal proximal tubules. Biochemical and Biophysical Research Communications. 1996
697 225:215-8.

698 52. Gil-Parrado S, Fernández-Montalván A, Assfalg-Machleidt I, Popp O, Bestvater F, Holloschi A,
699 et al. Ionomycin-activated calpain triggers apoptosis: a probable role for Bcl-2 family members.
700 Journal of Biological Chemistry. 2002.

701 53. Park H, Makepeace C, Lyons J, Song C. Effect of intracellular acidity and ionomycin on
702 apoptosis in HL-60 cells. European Journal of Cancer. 1996 32:540-6.

703 54. Opper B, Németh P, Engelmann P. Calcium is required for coelomocyte activation in
704 earthworms. Molecular Immunology. 2010 47:2047-56.

705 55. Hirt UA, Gantner F, Leist M. Phagocytosis of nonapoptotic cells dying by caspase-independent
706 mechanisms. The Journal of Immunology. 2000 164:6520-9.

707 56. Lew DP, Andersson T, Hed J, Di Virgilio F, Pozzan T, Stendahl O. Ca²⁺-dependent and Ca²⁺-
708 independent phagocytosis in human neutrophils. Nature. 1985 315:509.

709 57. Jaconi M, Lew DP, Carpentier J-L, Magnusson K, Sjögren M, Stendahl O. Cytosolic free calcium
710 elevation mediates the phagosome-lysosome fusion during phagocytosis in human neutrophils. The
711 Journal of cell biology. 1990 110:1555-64.

712 58. Sawyer DW, Sullivan JA, Mandell GL. Intracellular free calcium localization in neutrophils
713 during phagocytosis. Science. 1985 230:663-6.

714 59. Finkel TH, Pabst MJ, Suzuki H, Guthrie LA, Forehand JR, Phillips W, et al. Priming of
715 neutrophils and macrophages for enhanced release of superoxide anion by the calcium ionophore
716 ionomycin. Implications for regulation of the respiratory burst. Journal of Biological Chemistry. 1987
717 262:12589-96.

718 60. De Nadal E, Ammerer G, Posas F. Controlling gene expression in response to stress. Nature
719 Reviews Genetics. 2011 12:833-45.

720 61. Liu F-T. Regulatory roles of galectins in the immune response. International archives of
721 allergy and immunology. 2005 136:385-400.

722 62. Erickson JW, Cerione RA. Multiple roles for Cdc42 in cell regulation. Current opinion in cell
723 biology. 2001 13:153-7.

724 63. Clydesdale GJ, Dandie GW, Muller HK. Ultraviolet light induced injury: immunological and
725 inflammatory effects. Immunology and cell biology. 2001 79:547-68.

726 64. Schramm H, Jaramillo ML, de Quadros T, Zeni EC, Müller YM, Ammar D, et al. Effect of UVB
727 radiation exposure in the expression of genes and proteins related to apoptosis in freshwater prawn
728 embryos. Aquatic Toxicology. 2017 191:25-33.

729 65. Gouveia GR, Marques DS, Cruz BP, Geracitano LA, Nery LE, Trindade GS. Antioxidant defenses
730 and DNA damage Induced by UV-A and UV-B radiation in the crab *Chasmagnathus granulata*
731 (*Decapoda, Brachyura*). Photochemistry and photobiology. 2005 81:398-403.

732 66. Gervais O, Renault T, Arzul I. Induction of apoptosis by UV in the flat oyster, *Ostrea edulis*.
733 Fish & shellfish immunology. 2015 46:232-42.

734 67. Estevez-Calvar N, Romero A, Figueras A, Novoa B. Genes of the mitochondrial apoptotic
735 pathway in *Mytilus galloprovincialis*. PloS one. 2013 8:e61502.

736 68. Rank J, Jensen K. Comet assay on gill cells and hemocytes from the blue mussel *Mytilus*
737 *edulis*. Ecotoxicology and Environmental Safety. 2003 54:323-9.

738 69. Abbott Chalew TE, Galloway JF, Graczyk TK. Pilot study on effects of nanoparticle exposure
739 on *Crassostrea virginica* hemocyte phagocytosis. Marine Pollution Bulletin. 2012 64:2251-3.

740 70. Le Guernic A, Geffard A, Rioult D, Bonnard I, Le Foll F, Palos Ladeiro M. First evidence of
741 cytotoxic effects of human protozoan parasites on zebra mussel (*Dreissena polymorpha*) hemocytes.
742 Environmental Microbiology Reports. 2019 11:414-8.

- 743 71. Xian JA, Zhang XX, Sun JF, Wang L, Wang DM, Li JT, et al. Flow cytometric analysis of *Penaeus*
744 *monodon* haemocyte responses to poly I:C. *Fish Shellfish Immunol.* 2018 74:62-8.
- 745 72. Terahara K, Takahashi KG. Mechanisms and immunological roles of apoptosis in molluscs.
746 *Current pharmaceutical design.* 2008 14:131-7.
- 747 73. Carella F, Feist SW, Bignell JP, De Vico G. Comparative pathology in bivalves: Aetiological
748 agents and disease processes. *Journal of Invertebrate Pathology.* 2015 131:107-20.
- 749 74. Yee A, Dungan C, Hamilton R, Goedken M, de Guise S, Sunila I. Apoptosis of the protozoan
750 oyster pathogen *Perkinsus marinus* *in vivo* and *in vitro* in the Chesapeake Bay and the Long Island
751 Sound. *Journal of Shellfish Research.* 2005 24:1035-42.
- 752 75. Roberts SB, Sunila I, Wikfors GH. Immune response and mechanical stress susceptibility in
753 diseased oysters, *Crassostrea virginica*. *Journal of Comparative Physiology B.* 2012 182:41-8.
- 754 76. Soudant P, Chu F-LE, Volety A. Host-parasite interactions: Marine bivalve molluscs and
755 protozoan parasites, *Perkinsus* species. *Journal of invertebrate pathology.* 2013 114:196-216.
- 756 77. Perrigault M, Tanguy A, Allam B. Identification and expression of differentially expressed
757 genes in the hard clam, *Mercenaria mercenaria*, in response to quahog parasite unknown (QPX). *BMC*
758 *Genomics.* 2009 10:377.
- 759 78. Guillou F, Mitta G, Galinier R, Coustau C. Identification and expression of gene transcripts
760 generated during an anti-parasitic response in *Biomphalaria glabrata*. *Developmental & Comparative*
761 *Immunology.* 2007 31:657-71.
- 762 79. Desclaux-Marchand C, Paul-Pont I, Gonzalez P, Baudrimont M, De Montaudouin X.
763 Metallothionein gene identification and expression in the cockle (*Cerastoderma edule*) under
764 parasitism (trematodes) and cadmium contaminations. *Aquatic Living Resources.* 2007 20:43-9.
- 765 80. Morga B, Renault T, Faury N, Chollet B, Arzul I. Cellular and molecular responses of
766 haemocytes from *Ostrea edulis* during *in vitro* infection by the parasite *Bonamia ostreae*.
767 *International Journal for Parasitology.* 2011 41:755-64.
- 768 81. Kang Y-S, Kim Y-M, Park K-I, Cho SK, Choi K-S, Cho M. Analysis of EST and lectin expressions in
769 hemocytes of Manila clams (*Ruditapes philippinarum*)(Bivalvia: Mollusca) infected with *Perkinsus*
770 *olseni*. *Developmental & Comparative Immunology.* 2006 30:1119-31.
- 771 82. Leprêtre M, Almunia C, Armengaud J, Le Guernic A, Salvador A, Geffard A, et al. Identification
772 of immune-related proteins of *Dreissena polymorpha* hemocytes and plasma involved in host-
773 microbe interactions by differential proteomics. *Scientific Reports.* 2020 10:1-18.
- 774 83. Groves E, Dart A, Covarelli V, Caron E. Molecular mechanisms of phagocytic uptake in
775 mammalian cells. *Cellular and molecular life sciences.* 2008 65:1957-76.
- 776 84. Weake VM, Workman JL. Inducible gene expression: diverse regulatory mechanisms. *Nature*
777 *Reviews Genetics.* 2010 11:426-37.
- 778 85. Moore MN, Depledge MH, Readman JW, Leonard DP. An integrated biomarker-based
779 strategy for ecotoxicological evaluation of risk in environmental management. *Mutation*
780 *Research/Fundamental and Molecular Mechanisms of Mutagenesis.* 2004 552:247-68.
- 781 86. Huang X, Xu H, Sun X, Ohkusu K, Kawamura Y, Ezaki T. Genome-wide scan of the gene
782 expression kinetics of *Salmonella enterica* serovar Typhi during hyperosmotic stress. *International*
783 *Journal of Molecular Sciences.* 2007 8:116-35.
- 784 87. Farcy É, Voiseux C, Lebel J-M, Fiévet B. Transcriptional expression levels of cell stress marker
785 genes in the Pacific oyster *Crassostrea gigas* exposed to acute thermal stress. *Cell Stress and*
786 *Chaperones.* 2009 14:371-80.
- 787 88. He Y, Jouaux A, Ford SE, Lelong C, Sourdain P, Mathieu M, et al. Transcriptome analysis
788 reveals strong and complex antiviral response in a mollusc. *Fish & Shellfish Immunology.* 2015
789 46:131-44.
- 790 89. Moore MN. Biocomplexity: the post-genome challenge in ecotoxicology. *Aquatic toxicology.*
791 2002 59:1-15.
- 792 90. Fernández-Boo S, Villalba A, Cao A. Protein expression profiling in haemocytes and plasma of
793 the Manila clam *Ruditapes philippinarum* in response to infection with *Perkinsus olseni*. *Journal of*
794 *fish diseases.* 2016 39:1369-85.

- 795 91. Hartman R, Espinosa EP, Allam B. Identification of clam plasma proteins that bind its
796 pathogen Quahog Parasite Unknown. *Fish & shellfish immunology*. 2018 77:214-21.
797 92. Allam B, Espinosa EP. Bivalve immunity and response to infections: Are we looking at the
798 right place? *Fish & shellfish immunology*. 2016 53:4-12.

799