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1	Recent developments in preparative-scale supercritical fluid- and liquid chromatography for
2	chiral separations
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22	chromatography
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28 Abstract

This review covers preparative-scale chromatography for enantioseparations in both supercritical fluid and liquid chromatography published between 2016 and 2020. When pertinent, some fundamental publications from outside this time window are cited. The recent developments in instrumentation, novelties in method development and chiral stationary- and mobile phases are placed in context, while throughput, scale-up and sample issues are presented for supercritical fluid chromatography, and large-batch techniques together with computer-assisted method development, are described for liquid chromatography. Preparative-scale applications are reported for natural products, pharmaceutical compounds and pesticides, as well some other small molecules. Finally, a section is devoted to reports comparing supercritical fluid- and liquid chromatography and the strategies of various commercial users are briefly described. The goal of this review is to present current practices in the field of medium to large-scale chiral separations and offer indications as to how the best approach may be chosen for a given application.

53	
54	Abbreviations
55	AcN acetonitrile
56	API active pharmaceutical ingredient
57	CAMP chiral additive at the mobile phase
58	CPME cyclopentylmethylether
59	CSP chiral stationary phase
60	DCM dichloromethane
61	DEA diethylamine
62	EA ethyl acetate
63	EtOH ethanol
64	iPrOH isopropanol
65	MeOH methanol
66	MTBE methyl tert-butylether
67	<i>n</i> -Hept <i>n</i> -Heptane
68	<i>n</i> -Hex <i>n</i> -Hexane
69	TFA trifluoroacetic acid
70	TFE trifluorethanol
71	THF tetrahydrofuran
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109 **1. Introduction**

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Why did evolution favor L-amino acids and D-sugars as the homochiral building blocks of proteins 111 and nucleic acids in all living species? While its origins remain obscure, chirality exerts a profound 112 influence on biology and defines many of the key mechanisms of life. Chiral molecules consist of at 113 least two forms which cannot be superimposed on their mirror image, known as enantiomers or 114 115 optical isomers, and different enantiomers can have very different pharmacological effects. The global market for chiral chemicals was valued at USD 39.79 billion in 2015 and is projected to 116 expand at a compound annual growth rate of 13.67% to be USD 96.89 billion by 2023. Within this 117 118 market are flavors and fragrances, agrochemicals such as herbicides, pesticides, plant growth regulators and fungicides (30% of registered pesticides are chiral compounds [1]), and lastly 119 pharmaceuticals within which chiral compounds accounted for 72.5% of the global market in 2015. 120 121 Among the top-selling drugs on the market, most of the pharmaceutical blockbusters are pure stereoisomers, clearly showing that methods which separate enantiomers are needed for biological 122 testing [2]. 123

Among the different methods developed for enantioseparation, chromatographic resolution of the 124 racemic mixture is the most widely used technology (for more on the available methods, the reader 125 126 is referred to a comprehensive review from 2018 detailing both analytical- and preparative-scale enantioseparation by chromatography and related methods [3]). Two approaches can be pursued: 127 the first, "indirect" one forms true diasteroisomers, through derivatization of the compound using a 128 chiral agent (CDA), which can then be subsequently separated in an achiral environment; the 129 130 second, "direct" one, forms transient diatereoisomers between the solute and either a chiral stationary phase (CSP) or a chiral additive to the mobile phase (CAMP). While the CAMP method 131 is classically employed in capillary electrophoresis (CE), this direct approach is not very popular in 132 either supercritical fluid chromatography (SFC) or high-performance liquid chromatography 133 (HPLC), mainly because it is only suitable for analytical-scale and not preparative-scale 134

separations. However CE presents many advantages over chromatography, primarily derived from 135 136 the small dimensions of the silica capillary. These include high flexibility and separation power, short migration times, low consumption of analyte and chemicals, and a wealth of available chiral 137 selector types, while the use of capillaries results in very high plate numbers. The first CE chiral 138 separation was reported in 1985 by Gassman et al. [4] on the resolution of amino acids using a 139 ligand-exchange separation mechanism. Three years later, Snopek et al. achieved the separation of 140 141 pseudoephedrine enantiomers using β -cyclodextrin and heptakis(2,6-di-O-methyl)- β -cyclodextrin as additives to the leading electrolyte in isotachophoresis [5]. Recently, the major contributions to 142 chiral resolution by CE have been reported by the groups such of Fanali, Chankvetadze, Fillet and 143 144 Scriba, among others [6-9].

While HPLC with single and multi-column processes has been used for many years [10-12], SFC is starting to carve out a niche [13] thanks, in particular, to the work of Berger and colleagues in the mid-1990s [14], and has found its main field of application in chiral separation at the preparative scale [15; 16].

This review will report the recent developments in preparative-scale chromatographic separations, covering preparative SFC and HPLC publications over the last five years and highlighting the emerging trends at the dawn of the 2020s. Readers will find a summary table at the end of the manuscript summarizing the publications cited hereinand one at the end of each of the three applications sections (Section 4) summarizing the methods used.

154 **2. Recent developments in instrumentation**

2.1 Supercritical fluid chromatography

The first application of a supercritical fluid as a mobile phase for chromatography was reported by Klesper *et al.* [17] in 1962. However, because of poor availability of accurate and reliable commercial devices, use of supercritical fluid chromatography was confined for many years to academic research [18] with home-made devices. It was not until the early 1990s that a first generation of SFC devices was commercialized by several suppliers such as Gilson or Hewlett-

Packard (HP) then by Berger, Jasco, Pic Solution or Novasep, which permitted rapid development 161 162 of this technique in industry and especially in pharmaceutical discovery. The first devices dedicated to purification by SFC were essentially of the same design and enabled purification of a few 163 milligrams for the semi-preparative systems, up to several kilograms of mixtures consisting of 2, 3 164 or 4 compounds for the pilot, preparative-scale devices. They were all equipped with a binary 165 166 solvent delivery pump, an injector suitable for multiple injection of one feed solution, a UV or 167 diode array detector, a Back-Pressure Regulator (BPR), a Gas Liquid Separator (GLS) and a limited set of fraction collectors (from 3 to 8). The choice of preparative SFC systems is growing but is still 168 limited to 6 main suppliers providing equipment which accept columns with dimension from 1cm to 169 170 80cm internal diameter (ID).

Most of the research investments and innovations in SFC have been made by these suppliers to 171 reduce the extra column volumes and/or to enhance detector sensitivity. Nowadays, the 172 173 performance of analytical SFC devices is close to UHPLC systems and SFC is now suitable for like metabolite identification, metabolomic, pharmacokinetic, 174 different applications or 175 environmental research. However, the suppliers of preparative SFC devices have innovated too. Recently, thanks to the development of a new generation of GLS preventing the formation of 176 aerosols and allowing fraction collection at atmospheric pressure, open bed collectors are now 177 178 available. The commercialization of such devices (by Jasco, PIC Solution, Sepiatec, Waters or recently by Shimadzu) enables higher throughput and represents a step forward for the development 179 of the preparative SFC. Meanwhile, the coupling of preparative SFC apparatuses to mass 180 spectrometers by suppliers like Sepiatec, Waters or PIC Solution has extended the range of 181 applications for this technology. Indeed, the combination of the open bed collector and the mass 182 spectrometry-directed collection is a configuration suitable for both the purification of libraries 183 184 (achiral chromatography) and the separation of a set of racemic mixtures (chiral chromatography). Until now, racemic mixtures have usually been purified one-by-one due to the limited number of 185 fractions collectable by compatible apparatus, resulting in a relative low turn around despite the 186

numerous advantages of this technique. Thanks to the development of these new devices, preparative SFC-MS is becoming a real alternative for the separation of a series of compounds. This type of device, equipped with an autosampler and a mass detector, is dedicated to the separation of small amount of compounds (< 50 mg) requiring one or two injection per sample, however some of them have additional features to perform stacked injections and thereby increase yield to several grams.

It is worth mentioning that Shimadzu has recently released a new preparative-scale SFC system. This is the result of a collaboration with the Enabling Technologies ConsortiumTM (ETC), a "forum for pharmaceutical and biotechnology companies to discuss ideas, share information, and collaborate on the development of new enabling technologies". Like the previously mentioned preparative SFC-MS systems, this device is suitable for research and for yields of up to a few grams.

However, the price of SFC devices remains high compared to preparative HPLC devices and SFC is not suitable for all compounds, often making the acquisition of both HPLC and SFC preparativescale devices necessary. Hence, hybrid systems have been developed to do both analytical scale and preparative scale SFC on the same device (Waters, PIC Solution or Jasco), or else dual systems to perform both HPLC and SFC on one system (Agilent, Shimadzu). In addition, ABsys [19-21] has developed several features to economically and efficiently set up a preparative SFC device from an LC device.

206 2.2 Liquid chromatography

Much has changed since the first complete separation of the enantiomers of a racemate by ligandexchange liquid chromatography was reported by Rogozhin and Davankov in the late 1960s [22]. Preparative-scale chromatography technologies implemented the most frequently today are batch-HPLC, steady state recycling (SSR) and simulated moving bed (SMB). From a practical point of view, batch HPLC involves a single column process, while SSR is semi-continuous chromatography on a single column and SMB is a continuous, multicolumn chromatographicapproach.

From an equipment point of view, large-batch HPLC systems mirror their smaller-scale laboratory counterparts but use large diameter, pre-packed columns of 20, 30 or 50 mm I.D (or larger).

SSR liquid chromatography is used to support production of early-stage drug development. The 216 217 principle is that only the purified fractions are collected: the non-separated portion of the 218 chromatogram is recycled at the column inlet and a new injection of racemate is performed while the unresolved portion is recycled. The steady state is reached when the amounts of product injected 219 and collected are equal and the productivity will not decrease with the number of cycles. From an 220 221 instrumental point of view, an SSR device is merely a Batch HPLC with a recycling loop and a 4port manifold which allows the recycling and the injection of new racemate, along with two 222 223 collectors for the pure enantiomers.

224 Chiral separation using the SMB principle employs a series of columns connected in a loop to 225 continuously collect two fractions, each containing a pure enantiomer, from a continuously-eluting 226 racemic mixture. The remaining mixture of enantiomers that has not been separated is recycled 227 through the columns while the racemic mixture is newly injected. The advantage here is that there is no need to have a complete separation since only a fraction of the enantiomer is collected as a 228 "pure" fraction and the unresolved mixture remains in the loop. An SMB device is equipped with 6 229 to 8 columns in series, with a recycling loop and 2 inlet (for feed and eluent) and 2 outlet (for 230 extract and raffinate) streams connected between the columns. The countercurrent movement of the 231 solid is simulated by moving the inlet and outlet ports in the direction of the liquid flow. This 232 approach has been optimized by Novasep, and their Varicol® device allows the use of 5 or 6 233 columns with a claimed 30% increase in productivity over conventional systems. 234

Compared to batch HPLC, since no complete separation is needed, the racemate concentration in
SSR or SMB (and hence the productivity) can be substantially higher than with a standard batch
process, and the volume of solvent consumed substantially lower. While SMB technology allows

the separation of 100s of grams, even kilograms of racemate, it requires several days of development before production and a standard batch process remains the most suitable when only a few grams are to be separated.

Obviously liquid chromatography (LC) at the preparative scale is a mature technology and noteworthy advances in recent years are few. Further complicating the compilation of a review such as this is the commercial and proprietary nature of many technologies. Publicity from one of the biggest manufacturers in this sector cites preparative LC installations still in operation after 40 years, but few specific details of instrumentation are available and little mention made of enantioseparation. The Prochrom® package of products and services and the Hipersep® pilot are two such products.

While Mattrey et al. [23] do not deal exclusively with chiral nor preparative LC in their 248 comprehensive and wide-ranging review, their authoritative tour d'horizon of computer-assisted 249 250 method development deserves a mention. They report on the increasing potential, accessibility and perhaps the inevitability – of this technology as an important tool in screening for all aspects of 251 252 chromatographic method. They note that chiral analyses, and in particular the sometimesunpredictable performance of CSPs in their early days, contributed much to drive what is fast-253 becoming an industry in its own right: Bennett et al. [24] make the case for one such product (see 254 255 section 3.2).

256 **3. Recent method developments**

257 **3.1 Supercritical fluid chromatography**

258 3.1.1 Chiral stationary phases and mobiles phases

The choice of a preparative chiral stationary phase (CSP), in both SFC and HPLC, is based upon several criteria: the CSP must be universal to separate different types of compounds with a high resolution; the utilization of the CSP in preparative chromatography requires a high loading capacity; and finally, it is best to select robust CSPs to handle many purifications – indeed, the only thing all CSPs have in common is their high cost, making it expensive to change them often. A few evolutions in CSPs have been observed recently and will be discussed briefly here, as more detailsare available in a recent review by West focusing on the analytical scale [25].

In addition to the introduction of chlorinated CSPs providing different selectivity [26], and the 266 development of the generic polysaccharide based CSPs with the same chiral selector but from 267 several suppliers, the main improvement in recent years is the development of immobilized 268 polysaccharide-based stationary phases. Polysaccharide-based stationary phases are increasingly 269 270 recognized as a powerful media for the separation of enantiomers, however the first generation of these stationary phases consisted of polysaccharide coatings on the silica support, leading to 271 compatibility issues with some solvents like dichloromethane or THF. Indeed, as the polysaccharide 272 273 is soluble in these solvents, there is a major risk of bleeding of the phase which is not covalently 274 linked to the silica. A new generation of these phases, with the polysaccharide immobilized on the silica, is extending their application range. The first preparation of an immobilized type CSP was 275 276 described more than 30 years ago by Okamoto et al. [27], but initial results showed a progressive degradation of enantioselectivity. Different immobilization processes have been investigated to 277 278 obtain new CSPs with an enantiomeric recognition capacity close to the original coated phases. The first immobilized polysaccharide-based CSPs, commercialized in 2005, were Chiralpak IA, based 279 on the 3,5 dimethylphenylcarbamate of amylose [28], followed by Chiralpak IB (3,5 280 281 dimethylphenylcarbamate of cellulose). Both are immobilized versions of well-known coated phases (Chiralpak AD and Chiralcel OD, respectively). Immobilized versions of amylose tris (S)- α -282 methylbenzylcarbamate, amylose tris (3-chloro-4-methylphenycarbamate) and 283 tris (4methylbenzoate) have been commercialized very recently. Immobilized CSPs with chiral selectors, 284 285 unavailable in coated versions, were also developed such as Chiralpak IC or Lux-I cellulose-5 or 286 Chiral ART cellulose SC or reflect i-cellulose C (tris-3,5 dichlorophenylcarbamate of cellulose), ID chlorophenylcarbamate amylose), 287 Chiralpak (tris-3 of Chiralpak IE (tris-3,5 dichlorophenylcarbamate of amylose) and, the newest, Chiralpak IG or Lux-I amylose-3 (tris-3 288 chloro 5 methylphenyl carbamate) commercialized in 2016 [29]. These immobilized-289

polysaccharide-based CSPs provide different selectivities and are now commonly used in the 290 291 screening approach for preparative-scale enantiomeric separations [30]. This new type of chiral stationary phase also represents an opportunity to explore new organic solvents as modifiers such as 292 293 DCM, THF or MTBE (all miscible with scCO₂) in a quest for novel enantioselectivities [28; 31; 32]. In addition, these "exotic" solvents may also enhance productivity in preparative 294 chromatography thanks to the higher solubility of the racemates [33], and recently published papers 295 296 on this aspect will be presented in section 4.1 [34-36]. These solvents are now frequently used in preparative SFC, even if alcohols are still more commonly used. 297

In addition to the polysaccharide-based CSPs, specific CSPs have been recently developed and used in SFC: (i) the anion exchange phases Chiralpak QD-AX and QN-AX [37] provide a good separation of chiral acids; (ii) the zwitterionic CSPs Chiralpak ZWIX(+) and Chiralpak ZWIX(-) dedicated to the separations of amino-acids [38]; (iii) Chirobiotic T2 for the separation of aromatic amino acids [39] and (iv) cyclofructan CSPs for enantiomeric separations of α -aryl-ketone. However, no paper showing the utilization of these CSPs in preparative SFC has yet been published.

Regarding the mobile phase, it is widely acknowledged that SFC is perhaps better described as 305 306 Subcritical rather than Supercritical fluid chromatography: indeed, the supercritical state only occurs above the critical pressure and temperature and the addition of a modifier to the scCO₂ 307 influences these critical points. However, as there is no discontinuous transition between liquid and 308 309 supercritical states, the consequence on the physical properties of the mobile phase are negligible. For more polar compounds the proportion of modifier can reach 50% while for highly hydrophilic 310 analytes (peptides, sugars or nucleobases), the addition of water to the mobile phase is needed to 311 achieve reasonable retention. A recent example used a mobile phase containing a high percentage of 312 modifier (above 50%), including 10% water, to separate chiral aromatic amino acids on a 313 Chirobiotic T column [39]. However, the proportion of water is limited due to miscibility limits and 314 315 increases in detection noise. Some applications using a percentage of modifier above 50% mixed with liquid CO₂ and a high percentage of water were reported by Olesik [40]. This approach, known as Enhanced fluidity liquid chromatography (EFLC), was used by Bennett *et al.* in the first application of EFLC for preparative-scale separation of hydrophilic analytes. They report that HILIC-type columns can be used on SFC instrumentation with EFLC methods for both analysis and purification of very polar and hydrophilic compounds [41]. No chiral separation has so far been reported using EFLC, however the works of Bennett demonstrates that a specific class of compounds may be explored with this technique.

323 3.1.2 Increasing throughput and scale-up

There is a need for fast chiral separation in early-stage drug discovery and rapid separation of the 324 325 pure enantiomer. To reduce the process time, several options or strategies can be developed: the first is to reduce the screening time. To do this, both column dimension and particle size have 326 327 reduced, year-on-year, from 250×4.6 mm with 10µm particle size to 50×3 mm packed with 1.6µm 328 particle size [23], to achieve run times of less than 3 minutes. The run time in some cases has been more drastically reduced with chiral separation reported by T. Berger of less than 30 seconds (7.5s 329 330 for trans-stilbene oxide enantiomer separation) [42; 43]. In this race to ultrafast separation, the interesting concept of a segmented chiral-achiral liquid chromatography column, introduced in 2018 331 by Chankvetadze et al., is of particular note with baseline separation of enantiomers achieved in a 332 333 time of between 1-2 s [44].

This last, extreme strategy requires the modification of the devices to reduce dead volumes to a 334 minimum and cannot be used routinely. However, Zawatzki et al. [45] showed that a very short run 335 time is a way to set up a Multiple Injection in a Single Experimental Run (MISER). Some analytical 336 337 SFC devices are designed for this and permit the injection of many samples of the same mixture in one run. MISER can be used to monitor enantiomerisation or to analyze several fractions from a 338 preparative separation. Zawatzki and colleagues report the analysis of 96 samples of Trögger's base 339 on a Chiralpak AD $(4 \times 10 \text{ mm})$ in 33 minutes. This high throughput is also essential for rapid 340 screening of the numerous and diverse CSPs available. 341

A second option to enhance throughput in SFC is the introduction of open-access both for analytical 342 343 screening and preparative scale enantioseparation [46]. A platform with both an analytical system, consisting of only four chiral columns for screening plus one modifier, and a preparative-scale, 344 345 mass-triggered SFC system was developed. About 70% of the separation can be carried out thanks to this process. The use of a preparative SFC-MS platform with autosampler and open-bed 346 collection is also an opportunity to increase the throughput of specialized chiral separation 347 348 laboratories (full service platform). Indeed, with the standard batch preparative SFC system, compounds are handled one by one, the number of fractions it is possible to collect is limited, and 349 the whole system requires flushing between purifications. With an SFC-MS prep' platform there is 350 351 no limit to the number of fractions collectable, and the target is tracked by its molecular weight and purification can thus be performed in a continuous flow. 352

Another way to increase productivity was reported by Da Silva *et al.* [47]. They showed that the use of high linear velocities in preparative SFC (above 10mm/s) is efficient for the separation of the enantiomers of the Trögger's base on the Chiralpak AD. Productivity was similar when using a 2cm ID column with 20µm particles and high linear velocity when compared with a 3cm ID column with 5µm particles. In addition, solvent consumption was lower with the smaller column.

In these examples we have been talking about batch purification, meaning one column was used to 358 separate the isomers, but continuous mode or Simulated moving bed (SMB) is also used in SFC. 359 The SMB process has been already described for HPLC in section 2.2, and SMB-SFC also requires 360 the use of several columns (usually 6 or 8). A comparison of these approaches in SFC was made in 361 2007 by Peper et al. [48]; more recently, Johannsen et al. [49] compared productivity them using 362 363 stacked injections, concluding that the choice of technique depended on the separation required. While the solvent consumption with SMB is usually lower, an economic comparison of these 364 365 approaches for the separation of two mixtures showed that, in spite of a better productivity with SMB-SFC, the batch process remained more economic. 366

The scale-up in SFC is more complex than that of HPLC due to the compressibility of the mobile 367 368 phase which alters its density and hence the pressure or temperature along the column. According to Tarafder *et al.* [50], the strategy for reliable and accurate SFC scale-up is to maintain equivalent 369 370 mobile phase density in both the analytical and the preparative systems by keeping column length and particle size, eluent composition, flow-rate, temperature and pressure the same. However, as 371 372 noted by Enmark *et al.* [51], scale-up in SFC is complicated by differences in instrumentation: the 373 analytical SFC units use a volume flowmeter while the preparative systems use a mass flowmeter. An accurate measurement of the volumetric fraction of modifier inside the column is therefore 374 required to reliably scale-up in SFC. 375

Comparing SMB-SFC and SMB-LC, it is clear that the first approach presents more technical challenges, notably controlling the flow-rate and proportioning the flow between the outlets and the following column. However both the solvent usage and time spent evaporating the collected fractions are generally lower in SMB-SFC than in SMB-LC, while higher productivities are also reported for SFC-SMB relative to SMB-LC.

381 3.1.3 Sample integrity before, during and after purification

The wider range of solvents usable with the newer CSPs has gone some way to aid the preparativescale separation of poorly-soluble compounds: it is nevertheless still necessary to avoid sample precipitation and the resultant blockage of the inlet or any impact on the concentration of the feed solution.

Leek *et al.* [52] modified their injection system with the replacement of the sample loop by an extraction vessel, containing the sample adsorbed onto silica or celite[®], and the quantity of compound injected depends on the volume of mobile phase passing through this vessel. This example shows that the injection is a key step in preparative-scale purification. Indeed, the sample concentration is usually high to maximize productivity, and several studies have been carried out on the impact on enantioresolution of the injection mode and the modifier- and mixed streams [53]. Two recent papers described the impact of the nature of the solvent used to dissolve the sample: Desfontaines *et al.* [54] showed that MTBE, DCM, AcN or CPME were for the most part welladapted to large volume injections, while MeOH was the poorest in this regard. They also reported that the choice of the solvent depended on the stationary phase used. Meanwhile, Shalliker *et al.* [54] described the impact of the viscosity of the solvent on the peak shape.

The solubility of the sample is a key point, however another key issue in preparative SFC is sample stability: preventing its degradation, epimerization, racemization or interconversion is essential in preparative chiral separation. These phenomena usually occur in the column during the separation, in the collection flask, or during the subsequent evaporation step, but the modifier may also play a role.

402 As explained previously, the nature of the modifier impacts the retention, the selectivity and the solubility of the racemates but this modifier can also induce the degradation of some categories of 403 404 compounds such as esters, anhydrides, phenol esters and thiol esters. An approach developed by 405 Byrne *et al.* [56] is the replacement of the standard alcohol by 2,2,2 trifluoroethanol (TFE). They report that the presence of the fluoro- substituent strongly reduces the nucleophilic reactivity of the 406 407 oxygen of the hydroxyl group. This strategy was also used by Asokan et al. [57] for the purification of boronate ester. Boronic acid picanol esters were successfully separated without degradation or 408 epimerization using TFE as modifier. It is worth noting that thanks to its stronger eluting strength 409 410 compared with MeOH, the percentage of TFE in the mobile phase was lower and the stack injection cycle time reduced, with sharper peaks leading to higher loading per injection and hence higher 411 412 throughput.

Another potential source of degradation is linked to the acidity of the mobile phase. The combined presence of CO_2 and MeOH results in the formation of methyl carbonic acid which may favor degradation either inside the column or in the fraction collector. As described in a second paper, Asokan *et al.* [58] report that the concentration of CO_2 in the fraction collector depends on the system and the pressure used in the phase separator. For the compounds potentially degraded by methyl carbonic acid, and purified using a phase separator requiring a pression around 50bar, degradation can be avoided by replacing MeOH with another solvent, by bubbling nitrogen into the
fraction collector to chase CO₂ from the MeOH, or finally by adding a basic additive to neutralize
the solvent in the mobile phase or collected fraction.

422 The choice of the modifier is thus key to ensure the integrity of the analyte during purification. In addition, it is essential to control the kinetics of the degradation after separation. For instance, 423 Bajpai et al. [59] developed a process to prevent the degradation of one of their internal 424 425 compounds: purification required diethylamine (DEA) in the mobile phase, however a degradation of the target was observed during the post-purification process due to this basic additive. To 426 neutralize the base, a post column make-up introduced TFA (and, at the same time, DEA was 427 428 substituted by ammonium hydroxide to avoid the formation of TFA-DEA adducts in the final compound). Another separation of atropisomers posed particular challenges due to the potential 429 interconversion of the isomers depending on their rotational energy barriers. The R&D team of 430 431 Bristol-Mayer-Squibb succeeded in separating and isolating atropisomers by SFC, with good purity, thanks to simple ice baths added at the fraction collection reservoir to slow this interconversion. 432

433 **3.2 Liquid chromatography**

434 3.2.1 HPLC, SMB and CCC

The maturity of HPLC technologies and the commercial nature of preparative-scale methods for
chiral separations means few publications describe in any detail the recent advances in this sector. *Mode of injection*

In the field of natural products, the advantages of at-columnn dilution in preparative-scale separations of highly polar, water-soluble compounds were presented by Jaffuel *et al.* [60], while Ferrera-Queiroz *et al.* [60] investigated the use of a dry load injection method for poorly watersoluble compounds. While neither study was strictly concerned with chiral compounds, and the dry load injection method is perhaps too involved (as described) at anything beyond the semipreparative scale, they merit inclusion to illustrate the advantages of creative thinking with respect to sample introduction. Innovative methods for injecting samples have been a staple of preparativescale chromatography for some time, and Taheri *et al.* [62] described the optimization of a stackedinjection method for the semi-preparative enantioseparation of tramadol by chiral LC. The isomers of this poorly soluble molecule were separated with acceptable throughput by means of saturated injections of the racemate, and their study offers a model for enantioseparation of other poorly soluble molecules.

450 *Simulation*

In 2018, Forssén and Fornsted [63] published a paper including both chiral and non-chiral 451 possibilities from a comprehensive modelling study of the impact of LC column and chiral 452 stationary phase type on maximal productivity (P_{R-MAX}) of enantioseparations. Using Monte Carlo 453 454 simulations, they concluded that column efficiencies should be at least 500 theoretical plates but that nothing is to be gained by increasing separation efficiency above this limit. While increases in 455 $P_{\text{R-MAX}}$ can be achieved by increases in selectivity if $P_{\text{R-MAX}} < 2$, increasing selectivity if it is already 456 457 >3 will have the opposite effect. Similarly, they report that substantial gains in $P_{\text{R-MAX}}$ can be achieved by increasing relative saturation of the enantioselective site if this is $\langle ca.40\%$, but will 458 459 yield only moderate productivity gains if it is above this limit. Perhaps most pertinent to this review, they reported that predicting the productivity of up-scaling of analytical-scale injections on 460 CSPs is not straightforward – good analytical performance, they warn, does not necessarily predict 461 462 good productivity at the preparative scale.

463 *Chiral stationary phases*

In one of the most comprehensive recent reviews of monolithic CSPs, Guo *et al.* [64] extensively reviewed the preparation of this class of stationary phases for enantioseparation covering cyclodextrin-, polysaccharide-, protein- and antibiotic-functionalized CSPs on various supports, as well as ligand-, ion-exchange- and brush-type CSPs. Superficially porous silica as a carrier for polysaccharide CSP development can be found in a recent work by Chankvetadze *et al.* [65]. These two papers are necessarily focussed upon analytical-scale LC, given the relative youth of the field, but they deserve a mention here since these CSPs display low flow resistance and good selectivity, while the speed of progress among non-chiral, monolithic stationary phases of this type potentiallypromises a rapid evolution of this sector for chiral applications.

473 In an innovative study aimed at achieving genuinely preparative-scale chiral LC at reasonable cost, 474 Malik et al. [66] describe what a cookery book might describe as a 'store cupboard recipe' for a robust, reusable CSP consisting of bovine serum albumin (BSA) bonded to silica gel. Bemoaning 475 476 the fact that over 200 CSPs had been produced but that few (if any) had been successfully used for 477 "large-scale preparations in real terms", the authors noted that many CSPs developed for LC contain proteins. They rightly point out that many are also expensive (particularly true at the 478 preparative scale) and that they are often constrained in terms of percentage of organic modifier, 479 480 loading capacity, pressure limits and robustness. While not trivial, Malik et al. [66] described the preparation of a CSP where BSA is covalently bonded to silica gel using ingredients found in many 481 laboratories' cupboards. They used this CSP in an open column set-up for the direct 482 483 enantioresolution of 30mg of propranolol and atenolol and 3g of phenylalanine with >97% isomeric purity. 484

485 Xie et al. [67] also cite the high cost of CSPs as one of the motivations for their review of less wellknown CSPs for LC. Focussing on stationary phases based on chitosan- and cyclofructan 486 derivatives and chiral porous materials, their review covers mostly analytical-scale methods; they 487 488 nonetheless cite the cyclofructan-based stationary phases, in particular the functionalised, sixmembered (CF6) CSPs, as having potential for preparative-scale LC owing to their stability and 489 high loading capacity. While the native CF6 possesses only limited chiral recognition capability, 490 aliphatic- and aromatic-functionalized CF6 are able to separate enantiomers of primary amines and 491 492 other racemic compounds such as acids, secondary- and tertiary amines and alcohols.

493 *Minor hardware adaptations*

Building on work outside the period of this review (*ie.* published before 2016), Wrzosek *et al.* [68] sought to overcome the inherent "50% maximum yield" of chiral chromatography where the preferred enantiomer is selected from the racemate by LC and the undesired isomer discarded. The 497 authors described a proof-of-concept study combining preparative, chiral LC in series with an 498 enzymatic reactor for racemization of the unwanted isomer. Using mandelic acid as their model 499 compound, the authors also modelled their method to increase the potential transfer of this 500 technique to other applications. In simple terms, the model's key elements consisted of just two 501 parameters – the absorption isotherms of the enantiomers on the LC column and the rate of 502 racemization in the enzymatic reactor.

503 Simulated Moving Bed chromatography

The conventional SMB system is considered as the gold standard for separations in the range of hundreds of kilograms to hundreds of tons and is currently used commercially to produce Union Chimique Belge (UCB)'s antiepileptic active pharmaceutical ingredient (API) Keppra, Lundbeck A/S's Lexapro/Cipralex, Pfizer's Zoloft and Cephalon's Armodafinil, among others.

508 Chen et al. [69] reported considerable solvent savings when they compared preparative SMB and 509 LC to separate metalaxyl enantiomers from the racemate; they achieved excellent purity of the two isomers (>99%) from a concentrated racemate solution (15 mg/mL) and consumption of mobile 510 511 phase was 9-fold lower using SMB. Cunha et al. [70] investigated SMB to separate the L and D forms of praziquantel at high purity (100%) and semi-preparative scale (> 0.250 kkd). They built 512 several versions of their SMB unit around a cellulose tris CSP (Chiralcel OZ) and found that, in all 513 the scenarios investigated, at least one of the outlet streams produced one enantiomer at high purity 514 (often 100% optical purity). They compared their results with previous studies and reported at least 515 a doubling of productivity with much shorter run times, opening the way for studies with the 516 individual enantiomers of this drug. 517

518 Counter-Current Chromatography

This technique is considered as having a low environmental impact as it uses liquid-liquid partitioning to separate different compounds with minimal flow, and thus consumes less solvent. In their short review on the topic, Wu *et al.* [71] only briefly mention preparative-scale counter-current

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522 chromatography (CCC) for enantioseparation, concluding that its future in this field of application523 may rest primarily as a complement to HPLC.

More recently, Huang et al. [72] provided a comprehensive review of chiral separation by CCC, 524 525 citing its loading capacity, low cost, scalability and the complete recoverability of both enantiomers, as well as the chiral selector, as the principal advantages of the technique. While they acknowledge 526 527 that separation efficiency remains below that of HPLC, its potential usefulness at the preparative 528 scale makes it a worthy complement to both SFC and LC. They cite several studies from within the period covered by the present review which employed β -cyclodextrin derivatives as a chiral selector 529 530 in high-speed CCC (HSCCC) enantioseparations of ibuprofen, 2-(substituted phenyl)propanoic acids, naringenin and pheniramine (respectively: Rong et al. [73], Tong et al. [74], Wang et al. [75] 531 and Xu *et al.* [76]). 532

Recycling elution mode was used by Rong *et al.* [73], with hydroxypropyl-β-cyclodextrin (HP-β-CD) dissolved in the lower phase, in an HSCCC separation of ibuprofen enantiomers. The relatively low number of theoretical plates and very similar *K*-values of the (*R*)- and (*S*)-ibuprofen enantiomers made recycling necessary, and the authors reported a high enantiopurity (>97.5%) and recovery (>82%) at a multi-milligram production scale for both isomers.

Tong *et al.* [74] also used HP- β -CD as a chiral selector, in a continuation of their previous work on 2-phenyl propionic acid derivatives, for the enantioseparation of ten 2-(substituted phenyl)propanoic acid racemates. While not strictly preparative in scale (only 1 mmol/L solutions of racemate mixtures were investigated), they nevertheless offer a proof-of-concept for this class of chiral molecules, achieving enantiopurity >97% and recovery of each enantiomer of *ca*.80%, whilst providing a detailed investigation of optimum separation conditions.

Wang *et al.* [75] also employed HP-β-CD as a chiral ligand, but exploited the synergy between it and a Cu(II)-complexed amino acid ionic liquid (Cu(II)-[1-butyl-3-methylimidazolium][L-Pro]) for the enantioseparation of intractable naringenin racemic mixtures. Achieving enantiomeric purity of 98% at a multi-milligram production scale, they used UV-vis and NMR spectra to elucidate the role

of thermodynamic stability of the quaternary complexes amongst the metal ion, the ionic liquid, 548 HP- β -CD and the racemate in this enantioseparation. 549

550 Again at the multi-milligram scale of production, Xu et al. [76] described a method for the almost complete separation of enantiomers of pheniramine using carboxymethyl-β-cyclodextrin (CM-β-551 CD): they obtained 99% enantiopurity and a recovery of 85-90% from 20mg of racemate.

552

3.2.2 Computer-assisted Method Development 553

Despite the analytical scale of many publications dealing with this topic, the supported scalability of 554 555 some of these studies is of particular interest to this review – as Fornstedt et al. [77] observe in their 556 excellent primer on the topic, the complexity of preparative-scale enantioseparations can be too 557 expensive to optimize without computer simulations. Both Bennett et al. [24] and Tsay et al. [78] describe the use of a simulation package from ACD/Labs to manage scale-up from analytical- to 558 preparative scales, covering several key techniques and obstacles to this process. While not directly 559 560 concerned with enantioseparation, their studies reflect the growing use of this aspect of LC method 561 development and are the more pertinent because trial-and-error method development for preparative-scale chiral LC is often unfeasible – as remarked by Mattrey et al. [23], in many 562 laboratories it is more a case of "when" and not "if" this technology will be used. 563

4. Recent applications at the preparative-scale from 2016 to 2020 564

565 The present article covers the published works from 2016 to 2020, but the reader may refer to a review by Speybrouck and Lipka for studies published before 2016 [16]. 566

567 4.1 Supercritical fluid chromatography

568 All the experimental conditions in the following articles are summarized in Table 1. As SFC is 569 considered an eco-friendly method, some sort of ecological metric is increasingly reported in published articles in order to evaluate the environmental impact of the chromatographic method. At 570 571 the preparative scale, productivity is also a parameter of key importance.

The environmental factor (E-Factor) is defined as a waste-to-product ratio for any chromatographic 572 procedure. Solvent usage corresponds to the volume of solvent consumed to purify a known amount 573

of racemate (expressed in L/g). Productivity is defined as the amount of product separated *per* unit time *per* amount of stationary phase, and is expressed in kg racemate *per* kg CSP per day (kkd), while production rate is defined as the amount of product separated *per* unit time (kg product/day). In early-stage development, productivity is often very low (0.1 kkd or even poorer), while with a good separation productivity is generally in the range of 1 kkd and an exceptional separation might have a productivity greater than 10 kkd [79]: these values can be considered as useful guidelines when assessing productivity.

581 Natural products

In 2017, Xin et al. reported the separation and the purification of four lignan diastereoisomers from 582 583 Piper kadsura by chiral SFC: (-)-Galbelgin (compound 1), (-)-Ganschisandrin (compound 2), Galgravin (compound 3) and (-)-Veraguensin (compound 4) [36]. These four lignans are known for 584 their anti-neuroinflammatory activity and the four diastereoisomers are present together in the *Piper* 585 586 kadsura extract. In this work, chiral SFC was tried because successive achiral-HPLC and achiral-SFC methods had failed to resolve the diastereoisomers. Thus a three-step method was carried-out, 587 separating two pairs of diastereoisomers (compounds 1 and 2 from compounds 3 and 4) on a first 588 CSP (CSP-1) then, in two subsequent steps, separating compound 1 from compound 2 and then 589 compound **3** from compound **4** on a second CSP (CSP-2). 590

591 Step 1 was performed on CSP-1 column, injecting 90mg.

Thanks to stacked injections, the extract mixture could be injected four times in one 30min cycle, and the productivity increased by 277%; indeed, while a single injection was completed in 22.5 min four were completed in 32.5 min using stacked injections. The four diastereoisomers were collected into two parts (Fraction 1 and Fraction 2) weighing 150 mg and 470 mg, respectively, with four injections implemented twice.

597 Step 2 was performed on CSP-2 injecting 12mg using stacked injections. The cycle time was 598 reduced, with the productivity increased by 291% as one injection was completed in 7.7 min while 599 fourteen injections were completed in 37 min. Following those conditions, a 150 mg mixture was purified and 103.1 mg of compound 1 and 10.0 mg of compound 2 were respectively obtained in 37
min.

Step 3 was also performed on the CSP-2 column, injecting 45mg, and once again stacked injections were performed. The cycle time was reduced, and productivity increased by 235% as one injection was completed in 8 min while ten injections were completed in 34 min. Following those conditions, a 470 mg mixture was purified and 152.3 mg of compound **3** and 178.6 mg of compound **4** were respectively obtained in 34 min. The four lignans were obtained with a purity >98%.

Though not particularly original, the novelty of this work resides in its use of the "exotic" solventDCM to increase sample solubility.

609 In 2019, Lin et al. published the semi-preparative separation of enantiomers of dihydromyricetin [80]. Dihydromyricetin is a natural dihydroflavonoid compound mainly isolated from traditional 610 Chinese medicines (TMC), namely, Ampelopsis grossedentata and Hovenia dulcis. This 611 612 dihydroflavonoid compound exhibits many beneficial pharmacological activities, such as antioxidation, anti-inflammation, hepatoprotection and anti-hypertensive activities. A productivity of 613 614 0.07 kkd was achieved through stacked injections of 40mg per 12.8 min cycle. The solvent use was 5.86 L/g. The productivity was improved to 0.17 kkd through stacked injections of 80mg per 11.4 615 min cycle and solvent use was reduced to 2.75 L/g. Lastly a productivity of 0.27 kkd was achieved 616 through stacked injections of 120mg per 10.5 min cycle and solvent use further reduced to 1.76 L/g. 617 While this study is a good example of how productivity can be improved by injecting more of the 618 racemate, this increase in productivity (and decrease in MeOH consumption) nevertheless came at a 619 cost: the enantiomeric purities of two enantiomers were 98.5% and 98.4% at 40mg injected, falling 620 621 slightly to 98.5% and 97.3% at 80mg injected, but were both less than 95% when 120mg was injected. 622

623 Small pharmaceutical molecules or intermediates

E. Francotte reported the preparative separation of 50 g of racemic 1-benzyl-3-methyl-2-piperidone,
a drug intermediate [81]. Stacked injections were again implemented with a 1.8 min cycle time and

626 240 injections of 208 mg each were performed. The method demonstrated a high efficiency, leading 627 to a productivity up to 2.3 kkd. The enantiomeric excess of the separated enantiomer was greater 628 than 99%. This work also reported the preparative separation of two iodo-aryl regioisomers and 629 highlights the ability of chiral stationary phase to separate regioisomers.

In 2016, Wu et al. [26] reported the separation of three synthetic intermediates. The first was a non-630 polar, linear hydrocarbon, carboxylic acid (A) containing two chiral centers. The enantiomeric ratio 631 632 of the active trans-target to the undesired trans-isomer was 5:1. A productivity of 0.15 kkd was achieved thanks to stacked injections (6.5 min per cycle) with a recovery >80%. Interestingly this is 633 the first report of the use of a mixture of carbone dioxide and *n*-heptane (and iPrOH) as a mobile 634 635 phase. The second example was a two-step preparative separation to obtain a chirally pure drug intermediate. Indeed, the studied compound (B) contain two chiral centers and among the four 636 diastereoisomers (B1 to B4) only one was desired (B2). The optimized conditions led to 637 638 unsatisfactory loading and productivity when the four isomers were tested on the Chiralpak IC, so another four-step approach was tried. The first step was synthesis of a pair of B3 and B4 isomers; a 639 640 second step was chiral SFC of the B3 and B4 mixture to produce pure B3; a third step was epimerization of B3 to the mixture containing target B2 and B3 isomers, and the final step was 641 chiral SFC of the B2 and B3 mixture to obtain pure B2. A productivity rate of 204 g/day was 642 643 achieved with a high recovery through stacked injections of 440mg per 3.1 min cycle. In total, 1kg was separated in one week, and the targeted B2 isomer was recovered with an enantiomer excess 644 >99%. At the end of the process, the undesired B3 isomer was epimerized in a B2 and B3 mixture 645 for recycling. The third compound described by Wu et al. was a polar, chiral API (C). A 646 647 productivity rate of 22.8 g/day was achieved with a high recovery. In this particular case, although the loading capacity per injection cycle could have been increased, it was not investigated as the 648 649 target enantiomer eluted second.

Interestingly, this is the first time that 5% DCM has been successfully used on a coated CSP. Theauthors stated that no performance deterioration was observed on the column after extended runs

with this ternary, co-solvent system. In addition, this work underlines the complementarity between chlorinated and non-chlorinated CSPs, as chlorinated phases permitted the authors to successfully resolve several structurally diverse compounds by chiral SFC which were not well resolved on the non-chlorinated CSPs.

The same year, the same research group reported on the preparative separations of 20 propanoic 656 acids [82], developed as novel modulators of the glucocorticoid receptor. Among the 20 propanoic 657 658 acids, the preparative-scale resolution of compounds 4 and 14 were reported. Preparative separation of compound 4 (without TFA) had a productivity rate of 67.2 g/day achieved through stacked 659 injections of 330mg per 7 min cycle. The solvent use was 0.76 L/g. Preparative separation of the 660 661 weakly basic compound 14 had a productivity of 0.312 kkd and a production rate of 111 g/day achieved through stacked injections of 247mg per 8 min cycle. The flow-rate was equal to 100 662 mL/min and the solvent use was 0.65 L/g. Further scale-up on a 5 cm internal diameter column was 663 664 calculated and would have resulted in a production rate higher than 120g/day, considering a scaleup factor of 2.78. 665

In order to improve selectivity (previously 2.0), a mobile phase consisting of 30% iPrOH and 0.1% 666 DEA was tried and a selectivity of 3.3 was achieved. These conditions were scaled-up to the 667 Chiralcel OJ-H column (250 \times 50 mm; 5 μ m). The sample was dissolved at 62 mg/mL in 668 iPrOH/DEA leading to a productivity of 0.209 kkd and a production rate of 81.1 g/day achieved 669 through stacked injections of 620mg per 11 min cycle. The flow-rate was equal to 220 mL/min and 670 the solvent use was 1.17 L/g. Despite an improved resolution, the productivity was nevertheless 671 lower and solvent consumption higher with this alternative mobile phase. This is mainly due to the 672 peak tailing of the 2nd eluting enantiomer and the large injected volume (10 mL) required by the low 673 674 solubility of the sample in the iPrOH/DEA mixture. This example illustrates once again the importance of the sample solubility (*i.e* the feed concentration) in productivity. 675

In 2017, Dai *et al.* reported on the resolution at the preparative scale of atropisomers [83]. These
atropisomeric compounds contain two chiral axes, resulting in four atropisomers to be separated,

- and are pharmaceutical inhibitors of Bruton's tyrosine kinase (BTK) which plays a critical role in multiple cell types and is implicated in numerous autoimmune diseases. They found that isolated fractions were unstable and racemized to give two peaks. Thus, a second preparative SFC was then carried out on a Chiralpak AD column to which ice baths were added at the fraction collection reservoirs to slow the interconversion.
- In 2017, Zehani *et al.* published a small-scale chiral separation of four isoxazole derivatives [84].
- These four 3-carboxamido-5-aryl isoxazole analogues are novel potential ligands of sub-type 2
- 685 cannabinoid receptors (CB2), designed as treatments for Inflammatory Bowel Diseases (IBD).
- 686 Preparative separation of compound 1:
- 687 A productivity of 0.40 kkd and a production rate of 0.70 mg/min (1 g/day) were achieved. The
- solvent use was 0.87 L/g and the E-Factor was calculated as being 4.82.
- 689 Preparative separation of compound 2:
- A productivity of 0.23 kkd and a production rate of 0.39 mg/min (0.56 g/day) were achieved. The
- solvent use was 1.01 L/g and the E-Factor was calculated to be 7.86.
- 692 Preparative separation of compound **3**:
- A productivity of 0.34 kkd and a production rate of 0.58 mg/min (0.84 g/day) were achieved. The
- solvent use was 0.48 L/g and the E-Factor was 5.28.
- 695 Preparative separation of compound 4:
- A productivity of 0.24 kkd and a production rate of 0.40 mg/min (0.58 g/day) were achieved. The
- solvent used was 0.98 L/g and the E-Factor was 7.55.
- 698 While in this article the E-Factors are calculated, these calculations remain scarce in the literature.
- 699 In 2019, Yip *et al.* published the evaluation and comparison of two possible approaches to large
- scale SFC separation of a key intermediate in the synthesis of two $S1P_1$ -active pharmaceutical
- ingredients, namely BMS-986166 and compound 2 [35]. Recently, the S1P₁ (sphingosine-1-
- 702 phosphate-subtype 1) receptor has been highlighted as an attractive drug target in treatment of
- autoimmune diseases. The authors describe the development of an efficient chiral SFC for purifying

kilogram-quantities of a shared, key intermediate in preparation of the two S1P₁ drug candidates.
Figure 1 in Supplementary Information presents their two possible routes to prepare the
enantiomerically pure alcohol (Int 2A):

Route 1) SFC of methyl esters (Int 1A and 1B) followed by conversion of Int 1A to the alcohol (Int
2A) and route 2) conversion of the methyl esters to alcohols (Int 2A and 2B) followed by SFC to
obtain the desired Int 2A and the two desired APIs.

710 Comparing the two routes, separation of Int 1A and 1B led to a throughput of 94.08 g/day (0.72 kkd) as compared with 1.7 g/h (0.31 kkd) for Int 2A and 2B. The solvent consumption in SFC of Int 711 1A and 1B was 1.43 L/g, which was 63% of 2.27 L/g in Int 2A and 2B. Thus, route 1 was chosen 712 713 and around 1 kg of the ester intermediate was purified by SFC on the 3 cm Chiralpak IC column in less than twelve days, with a high recovery and a diastereoisomeric excess superior to 99% of the 714 715 desired Int 1A isomer. Hundreds of grams of the enantiomerically pure Int 1A were converted to the 716 Int 2A alcohol and then carried through subsequent steps to synthesize both BMS-986166 and compound 2. 717

718 Pesticides

In 2017, Yan *et al.* reported the preparative separation of four β -cypermethrin stereoisomers [85].

Synthetic pyrethroids are a family of important chiral pesticides containing multiple stereoisomers, and β -cypermethrin in particular is one of the most commonly used pesticides due to its persistence, photostability, high toxicity for insects, and strong endocrine disruptive ability for aquatic organisms.

A three-steps approach was developed: the four isomers of β -cypermethrin were separated in a first step leading to two peaks, each containing two stereoisomeric pairs. β -Cypermethrin was injected in 10 stacked injection volumes of 2 mL with a 5 min cycle time. Two stereoisomeric pairs (labeled as P1 and P2) were collected separately and P1 was subsequently separated. The mobile phase was 20% EtOH in CO₂ and the P1 sample was injected in 10 stacked 2 mL injections with 5 min cycle time leading to two enantiopure products. The P2 was also separated and injected in 10 stacked 2 mL injections with a 2 min cycle time leading to two enantiopure products. The four isomer configurations were determined according to the literature and were denoted 1R-cis- α S, 1R-trans- α S, 1S-cis- α R, and 1S-trans- α R based on their elution order from the chiral column. Their recoveries were about 94.5, 97.7, 96.5, and 95.6% and their enantiomeric purities were about 97.39, 97.53, 92.19, and 95.61%, respectively.

735 **4.2 Liquid chromatography**

All the experimental conditions depicted in the following articles are summarized in Table 2.

737 Natural products

In 2017, Gao *et al.* published their work on the separation of the isoborneol enantiomers [86].
Isoborneol is a monoterpenoid compound with good biological activity as an anti-inflammatory,
antibacterial, and analgesic.

741 In 2020, Tanno *et al.* reported the semi-preparative separation of α -tocopherol [87]. Tocopherol is a 742 lipophilic antioxidant and oxidation products of α -Toc, such as 8a-hydroperoxy- α -tocopherone (Toc-OOH) exert an antioxidative effect, but accumulation of such products can alter food quality. 743 Toc-OOH possesses chiral centers at the 2-,4- and 8-carbon along with the hydroperoxide binding 744 8a-carbon, thus constituting two stereoisomeric forms. In order to optimize and validate the LC-745 MS/MS quantification method of the stereoisomers in extra-virgin olive oil (marketed or oxidized), 746 747 standards of these Toc-OOH isomers were needed in significant quantities but were not commercially available. The separation of the 8a(S)-hydroperoxy- α -tocopherone (8a(S)-Toc-OOH) 748 749 and 8a(R)-hydroperoxy-a-tocopherone (8a(R)-Toc-OOH) stereoisomers was performed by 750 connecting two columns in series, which is rarely reported in LC owing to the increase in back-751 pressure it creates. This purification was repeated several times to provide sufficient quantities of each standard. 752

753 Small pharmaceutical molecules or intermediates

In 2016, Procopiou *et al.* reported the preparative separation of enantiomers of N-CBZ-3fluoropyrrolidine-3-MeOH [88]. This compound is a building block for synthesis of $\alpha_v\beta_6$ integrin inhibitors used in the treatment of idiopathic pulmonary fibrosis. The injection mass was 38
g/injection, and the run-time was 24 min. Their enantiomeric purities were 99.5 and 99.8%
respectively. Thus, to purify 5 kg, 132 injections were carried-out in 52.8 hours, consuming 1626 L
of EtOH and 6505 L of *n*-heptane for elution, underlying high consumption of alkane, requiring
high amounts of energy to evaporate.

761 In 2016, Kurka *et al.* published the semipreparative separation of cis-itraconazole [89]. Itraconazole 762 displays anticancer and antiangiogenic properties but is also an antifungal drug, administered as a racemate, and the molecule presents three chiral centers resulting in eight stereoisomers. However, 763 for clinical purposes only the cis-racemate mixture is used, containing four stereoisomers: (+)-764 765 2R,4S,2'R; (+)-2R,4S,2'S; (-)-2S,4S,2'R; (-)-2S,4R,2'S. The chiral separation of these four stereoisomers is required in order to comply with current legislation and describe their individual 766 767 biological properties. A four-step HPLC approach was developed for the semi-preparative 768 purification of four stereoisomers (Fig. 2 in Supplementary Information). The elution order observed on Lux Cellulose-1 column was: the two (+)-2R,4S stereoisomers eluted first in one peak 769 770 (Fraction 1), followed by the two (-)-2S,4R stereoisomers separated into two peaks. The first step consisted in the collection of Fraction 1 and the two levorotary stereoisomers. Subsequently, 771 Fraction 1 was separated and the (+)-2R,4S,2'R and (+)-2R,4S,2'S isomers were collected in step 772 two. In the case of the Cellulose-3 column, the two 2'R stereoisomers co-eluted first (Fraction 1'), 773 774 followed by the two 2'S stereoisomers, again separated into two peaks, such that three peaks were 775 collected in step 3. Finally, Fraction 1' containing the (+)-2R,4S,2'R and (-)-2S,4R,2'R isomers was separated and the two isomers collected (step 4). 776

During these steps, triple mobile phase recycling was used leading to a 50% reduction of mobilephase consumption. The chiral purity of each of the four isomers was >97%.

It is worth noting that "*artisanal*", small-scale chromatography is always implemented on analytical
columns, illustrating their high loading capacity. In 2016, Lal and Bhushan published the semipreparative separation of ketorolac (Ket) [90], a nonsteroidal anti-inflammatory drug (NSAID). The

therapeutic activity of ketorolac resides almost exclusively in the (S)-(-) enantiomer while the drug is marketed and administered as a racemic mixture. Fractions pertaining to the first-eluting enantiomer (En-I) and the second-eluting enantiomer (En-II) were collected, leading to 150 mg of En-I and 158 mg of En-I. Another example was published in 2017 by Chi *et al.* [91] reporting the semi-preparative resolution of (S)- and (R)-tetrahydro- α -(1-methylethyl)-2-oxo-1(2H)-pyrimidine acetic acids ((S)-TPA and (R)-TPA, respectively), known as useful intermediates in the synthesis of human immunodeficiency virus (HIV) protease inhibitors like lopinavir.

In 2016, Sadutto *et al.* [92] reported the semi-preparative separation of bicalutamide, which is a selective nonsteroidal, anti-androgen drug already used for treatment of prostate cancer, but for which only the R-enantiomer presents biological activity. A volume of 2 mL containing 21.6 mg of bicalutamide was purified. The yield was 90% and the throughput was 960 mg per day.

793 In 2016 Silva et al. [93] reported the chiral separation of some cathinone derivatives, which are 794 novel psychotropic drugs. In particular, 3,4-methylenedioxypyrovalerone (MDPV) was purified. 795 Thanks to multiple injections, 100 mg of MDPV were separated, resulting in 45.5 mg of the first-796 eluting enantiomer and 41.3 mg of the second. In 2018 Silva et al. [94] published more of their work on cathinone derivatives in "legal highs", this time studying the compounds pentedrone and 797 methylone. Over 14 hours, multiple injections of sample solution, totaling 9.80 mg of pentedrone, 798 799 resulted in 3.53 mg of the first-eluting pentedrone enantiomer (72% recovery) and 3.48 mg of the second (71% recovery). Regarding methylone, 12 mg were injected on the device, resulting in 4.80 800 801 mg of the first-eluting enantiomer (80% recovery) and 4.74 mg of the second (79% recovery). It is noteworthy that the column was unable to separate the two racemates with the same performance 802 after 14 hours and a washing process was needed to regenerate the column. 803

In 2019, Onyameh *et al.* [95] reported the preparative separation of a ligand with a high affinity for the 5HT₇ receptor: 5-chloro-2-{2-[3,4-dihydroisoquinolin-2(1H)-yl]ethyl}-2-methyl-2,3-dihydro-1H-inden-1-one (SYA40247). This racemic compound needs to be separated as the 5HT₇ receptor is a known target of drug development for the treatment of schizophrenia. Two repeated injections
were made and the enantiomeric purity of each evaporated fraction was 100%.

4.3 Comparing supercritical fluid- and liquid chromatography

All the experimental conditions depicted in the following articles are summarized in Table 3.

811 Small pharmaceutical molecules or intermediates

In 2016, Leek et al. [52] reported the large-scale isolation of enantiomers of compound A, an 812 undisclosed, key intermediate used in AstraZeneca R&D. The preparative SFC separation was 813 implemented through stacked injections of 500mg with a 2 min cycle time. The complete resolution 814 815 of the 5kg batch was estimated to require 53 hours with a MeOH use of 0.08 L/g and a productivity equal to 6.4 kkd. However, after ten injections a slow increase in the pressure drop was observed 816 and eventually led to the complete obstruction of the tubing from sample loop to the column. 817 Different strategies were tested to circumvent this precipitation of the poorly soluble compound in 818 819 the carbon-based mobile phase. An apolar solvent (DCM) was added to the modifier to improve 820 solubility of the sample, but no improvement was observed and in addition a decrease in selectivity was observed. A lower concentration of feed solution was tested (i.e 100mg/mL instead of 250 821 mg/mL), but this approach was also unsuccessful. 822

823 Building on the work of Cox and Shaimi, who described injection by extraction to solve this problem of low solubility in the carbon-based mobile phase [96], the authors used an extraction 824 vessel (100×10 mm) filled with 4.4 g compound A:silica 1:5 mixture ratio. The principle of this 825 approach is that only the soluble amount in the mobile phase is extracted and injected, thus avoiding 826 precipitation. The same CSP and mobile phase were used, but the flow-rate was increased to 150 827 828 g/min. With a dynamic mode of injection (40 seconds, 52 mg racemate): a productivity of 0.2 kkd 829 was achievable and 190 days were calculated as being needed to purify the 5 kg batch. Even though precipitation was avoided by the extraction injection approach, it was not possible to further 830 improve the scale-up of the extraction column and an HPLC method was thus evaluated. At a flow-831 rate of 600mL/min, the authors calculated that with an injected amount of 4.8 g and a cycle time of 832

833 10 min, the time needed to resolve the 5kg batch would be 174 hours, but productivity of only 0.4 834 kkd was achievable. In this particular case the main issue was not the time required but the solvent 835 consumption: more than 6000 L of mobile phase would have been required, mainly composed of 836 heptane. In this article neither of the two methods were adopted by AstraZeneca and the resolution 837 was eventually performed by crystallization of a diastereomeric salt.

838 In 2016, Rossi *et al.* reported the semi-preparative purification of 20 mg of a novel potent σ 1 839 receptor agonist by SFC and HPLC [97]. The aim was to elucidate its role as a therapeutic target in 840 several neurodegenerative diseases or mood disorders. In small-scale, preparative SFC, 20mg of the 841 racemate were purified in 40 cycles of 10 min each, leading to 9.1 mg of the (S) enantiomer and 8.2 mg of the (R) enantiomer at an overall yield of 86.5%, and an enantiomeric excess >99.5%. In 842 small-scale, preparative HPLC 21mg of racemate were separated in 7 cycles of 16 min each, 843 leading to 8.7 mg of the first-eluting enantiomer and 9.1 mg of the second, with an overall yield of 844 88.8 % and an enantiomeric excess >99.9%. 845

In this study, semi-preparative HPLC was found to outperform semi-preparative SFC, as productivities were equal to 0.0270 kkd and 0.0072 kkd respectively, with an average recovery of around 45%. Another method was developed in SFC with other conditions of mobile and stationary phases but the productivity obtained was even lower at 0.0065 kkd.

But it should be noted that the comparison in this case is not optimal for two reasons: the CSPs were not chemically identical and the SFC method had not been fully optimized. But in any case, neither of the two SFC methods exhibited productivity greater than the 0.0270 kkd obtained using semi-preparative HPLC.

In 2017, Leek and Andersson published three case studies of new candidate drugs developed at AstraZeneca [34]. Case study No.1 dealt with the requirement for 100 g of pure enantiomer from the optimized active pharmaceutical ingredient (API) destined to treat acute myeloid leukemia. The racemic intermediate, compound A, was a weak base with low solubility in alcohol. The preliminary experiments carried-out by SFC showed insufficient loadability due to the low solubility of compound A in the carbon dioxide-based mobile phase and normal-phase LC was thus
investigated. The quantity injected was 9.45g with a cycle time of 13 min and a fraction volume of
1.8L/injection. The productivity was quite low (0.9 kkd) and the solvent consumption was 830 L/kg
racemate. In this example, normal phase LC outperformed SFC which failed to separate the desired
compound.

Case study No.2 concerned the dextrorotary enantiomer of the racemic API (compound B) for the development of a first-line anti-inflammatory therapy for asthma patients. Preparative SFC was chosen because the desired enantiomer eluted first. The quantity injected was 3.4g using an injection cycle time of 155 s, with a fraction volume of 70mL/injection. The productivity was quite high (5.7 kkd) and the solvent consumption was 70 L/kg racemate. A total of 1.1 kg of the API with an overall yield of 96% and an enantiomeric excess >99.9% was achieved.

870 Case study No.3 aimed to address the shortcomings of case No.2, since it can be seen that, even if 871 successful, the SFC method developed above is uneconomical and wasteful with only half the API being used after the separation step. Thus, a combination of chiral SFC and racemization was 872 873 investigated in order to evaluate the toxicology of compound C. The quantity injected was 0.5g for each 190 s injection cycle and the productivity remained quite high (4.4 kkd) and solvent 874 consumption lower than with normal-phase LC. The second-eluting and unwanted enantiomer was 875 evaporated to dryness after chiral separation and then dissolved at 30mg/mL in MeOH/TEA 3/1 876 877 molar equivalent before being left overnight under rotation. More than 95% was thereby racemized, and after 3 cycles of chromatography and racemization, a total yield of 87% was obtained. 878

In 2018, Yan *et al.* reported the preparative separation of enantiomers of lenalidomide [98]. Over the past 10 years, Lenalidomide has been used for the treatment of multiple myeloma, other hematological malignancies and myelodysplastic syndromes, with significantly improved overall survival in myeloma. In this work, two different SFC methods were developed on two different CSPs and compared.

34

Method 1: many conditions were not entirely favorable: the maximum injection mass was about 8 mg and the elution time was about 20 min, while the cycle time for the stacked injections was estimated to be at least 14 min. Thus, a second approach was tested on another chiral stationary phase type.

Method 2: the injection volume was set at 3 mL leading to a productivity of 0.16 kkd achieved 888 through 10 stacked injections of 30mg per 5 min cycle. The solvent use was 2.6 L/g. The recoveries 889 890 for each enantiomer were about 81.7 and 79.5%, and their enantiomeric purities were 97.4, and 97.5%, respectively. This work was compared to previous publications which used HPLC [99; 100] 891 and many features were better in SFC. In the first HPLC study, Tojo and co-workers who reported a 892 893 productivity of 0.015 kkd [99]. In the SFC study, the productivity was about ten times higher (0.16 894 g kkd), while consumption of MeOH was only 2.6 L/g, much less than the second HPLC study 895 which reported a solvent use of 17.4 L/g of EA [100], indicating that SFC separation was both more 896 productive and economical than HPLC.

In 2019, Cheng et al. described the preparative separations of 6-(4-aminophenyl)-5-methyl-4, 5-897 dihydro-3(2H)-pyridazinone (SKF 93505) [101]. SKF 93505 is a key synthetic intermediate for 898 cardiotonic agent levosimendan which is the levorotatory enantiomer of simendan, a positive 899 inotropic drug and calcium sensitizer for the treatment of short-term, decompensated heart failure. 900 The analysis time of a single injection in SFC and HPLC was about 7.5 and 9 min, respectively. 901 Stacked injection mode was used to increase the productivity with an injection cycle of 5 min, 902 which increased productivity to 33 and 44% in SFC and HPLC, respectively. To separate 1 g of 903 racemate SKF 93505, the SFC technique took about 2.8 h with 4.6 L/g MeOH consumption, and the 904 905 HPLC technique took about 2.8 h with 3.4 L/g solvent use. The productivity of both modes was equal to 0.171 kkd using a concentration of feed solution equal to 15 mg/mL. The enantiomeric 906 907 excess values of each enantiomer were 100% for both isomers in SFC, and were 100% (R isomer) and 98.82% (S isomer) in HPLC. In this particular case, then, both techniques enabled fast, low-cost 908

separations of (R) (-) SKF 93505 with high yields (98 % for SFC and 95% for HPLC) and very
high enantiopurity.

Also in 2019, Wang et al. reported a comparison of the preparative separation of Corey lactone diol 911 [102]. Purification of Corey lactone diol (CLD) is of utmost importance in the pharmaceutical 912 industry as the levorotary enantiomer has become the key intermediate in the syntheses of 913 prostaglandins (PGs). PGs derivatives are an important class of drugs, involved in the treatment of 914 915 idiopathic pulmonary hypertension and gastrointestinal tract illnesses. The dextrorotary enantiomer, on the other hand, is a key intermediate in the synthesis of entecavir, a medicine for anti-viral 916 917 therapy in chronic hepatitis B patients. In both SFC and HPLC, the separation was run on an ADMPC30 column (250×20 mm, 5 µm). The preparation of the stationary phase was conducted in 918 house and ADMPC (amylose tris-(3,5-dimethylphenylcarbamate) was coated on 5 µm silica gel. 919 The notation used by the authors of ADMPC-10, ADMPC-20, ADMPC-30, and ADMPC-40 920 921 corresponds to the ratio of the derivative to the silica gel support (w/w) equal to 1:10; 2:10; 3:10, and 4:10, respectively. In SFC the injection was of 130mg and the HPLC the injection quantity was 922 160mg. The analysis time of a single injection in both SFC and HPLC was 3 min. Stacked injection 923 mode was used, with the injection cycle set to 3 min in both SFC and HPLC. To separate 1 g of 924 racemate CLD, the SFC technique took about 23 min with 0.46 L/g MeOH consumption, and the 925 926 HPLC technique took about 20 min with 0.39 L/g solvent use of AcN. The productivity in SFC was 927 1.248 kkd and 1.536 kkd in HPLC. Concerning the enantiomeric purities, values of each enantiomer 928 in SFC were 99.02% and 94.26% and in HPLC were 98.33% and 99.41%: in this particular case, 929 then, HPLC enabled a higher productivity and a slightly lower solvent consumption.

930 Conclusion

The new developments and applications in preparative chiral SFC and chiral HPLC have been reviewed in the previous sections and the *pros* and *cons* of each method have been summarized in Figure 1. We have described how the advances in preparative SFC and HPLC are being led by the appearance on the market of new CSPs, able to be used with more nonpolar solvents like DCM

while at the same time exhibiting improved robustness and loadability. Innovative injection 935 936 techniques and intelligent recycling and fraction-collecting approaches have also been important over the last five years, with several offering the potential to avoid the inherently large "carbon 937 938 footprint" of preparative-scale enantioseparations, both in terms of solvent usage but also in the way that only half the racemate is frequently required (at best) and collected. We have shown that the 939 940 proportion of preparative enantiomer separations carried out by SFC is growing. The benefits of 941 SFC over HPLC are widely described, Francotte [81] summarizing well the advantages of this technique when writing that in SFC, "the consumption of solvent is reduced by 60-70% and the 942 running cost by 70-80%". However, in spite of the advantages of SFC, chiral LC remains useful for 943 many applications. 944

One of the motivations for this review concerned the rationale for selecting either SFC or HPLC for 945 a preparative-scale, enantioselective separation. Leek and colleagues from AstraZeneca R&D 946 947 describe a typical industrial setting, where large-scale chiral separations are performed by SFC for 95% of their racemates [52]; a similar observation was made by Miller and co-workers from Amgen 948 949 [103], who reported a success rate greater than 98% in SFC for their discovery program, in which only SFC is screened and HPLC evaluated only when SFC is unsuccessful. This is also the case at 950 Reachseparations: since January 2020, their split between SFC and HPLC for purification projects 951 is approximatively 80% SFC and 20% HPLC. For Janssen, the ratio of SFC/HPLC depends on the 952 953 department: in discovery more than 99% of the chiral separations are performed by SFC, while in development about 70% of the chiral separations are handled by HPLC. For other pharmaceutical 954 laboratories, like Eli Lilly, SFC is clearly the future, with the utilization of SFC for almost all 955 956 batches less than 50g, and huge investment in facilities to handle 1kg batches by SFC. For other users, sample quantity is also a criterion of selection. 957

The selection of the technique is necessarily multifactorial and SFC screening and HPLC screening are sometimes performed in parallel, with the physical properties of the target compound sometimes also leading to an *a priori* choice of either SFC or HPLC. Thus, for hydrophilic compounds, chiral

RP-HPLC would be the first option. Some chiral SFC separations reviewed above required up to 961 962 50% modifier and the advantage of SFC over HPLC regarding solvent consumption is thus largely negated. Solubility is also a criterion – a racemate is often reported as being less soluble in a CO₂-963 964 enriched mobile phase, especially at high concentrations, and several users report that it is easier to handle a compound with poor solubility in HPLC than in SFC. Furthermore, it is obvious that the 965 solubility cannot be easily assessed in the mobile phase of SFC with a large proportion of 966 967 Supercritical fluid CO₂. However, thanks to the development of immobilized polysaccharide CSPs, the use of solvents like DCM, THF or even DMSO is now possible both in SFC and in HPLC, 968 resulting in a wider range of options for increasing the solubility of the racemates. The stability of 969 970 the compound is also a criterion for selecting either SFC or HPLC, notably the reactivity of CO₂ or acidity of the mobile phase in SFC. However, unlike several industry experts who favor SFC for 971 972 preparative separation of small quantities, others favor HPLC for small quantities because of the 973 relatively low recovery rate in SFC, while the equipment available in the laboratory also guides the 974 selection of technique, as does the relative expense of each and the familiarity of laboratory 975 personnel with one or the other.

To sum up, if the sample stability and solubility is suitable for SFC, most small quantities are 976 handled by SFC. For larger quantities both SFC and HPLC are often screened, with a majority of 977 compounds still separated by SFC if the laboratory has the equipment available to handle large 978 979 batches with this technique. New CSPs, compatible with a wider range of solvents, are broadening the range of compounds suitable for both techniques while innovative injection, recycling and 980 collection techniques have made significant advances in enantiomeric separations using both SFC 981 982 and HPLC. With SFC bound to become more accessible, and perhaps more scalable over the coming years, it will be fascinating to see how these two keystone techniques of preparative-scale, 983 984 chiral separation compare in another 5 years' time.

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Pros & Cons of preparative-scale SFC

High **efficiency** due to larger diffusion coefficients of solutes and/or the possibility of longer columns

Faster **flow-rate** and **column equilibration** due to lower viscosity leading to shorter analysis and cycle times, without loss of resolution

Higher column **loadability**, hence productivity, than HPLC because the equilibrium isotherms of compounds between supercritical fluids and the preparative CSPs are flatter

Solubility in supercritical fluid decreases rapidly as the pressure decreases near the critical point, facilitating recovery of the isomer fractions and **increasing yield**

Mobile phase is mainly constituted of carbon dioxide which is **non-flammable** & **less expensive** and **toxic** than common organic solvents, the use of which is also reduced

Instrumentation allows stacked or overlapped injections improving **productivity** and reducing solvent consumption

Pros & Cons of preparative-scale HPLC

When required, **reversed-phase chromatography** is easier to implement in HPLC because of the lack of polar supercritical fluid

From a theoretical point of view, **knowledge** of the physical chemistry of liquid/solid adsorption is widespread and established, whereas it remains lacking for supercritical fluid/solid adsorption

In HPLC, while time-consuming, the **scale-up is straightforward**, based on rules of thumb such as scaling volumetric flow and injection volume to the square of the ratio of the column radius

Although non-toxic, **massive leaks of carbon dioxide** remain a constant risk for the operator together with the storage of CO₂ under pressure

Instrumentation is **simpler** and **cheaper** in HPLC than in SFC, making the technique more accessible

HPLC benefits from a developmental and experiential legacy generating a **huge technical expertise** and **mass of scientific data** available in the literature

Analyte	Preparative device	Stationnary phase	Mobile phase	Pout	Flow-rate	Concentration of feed solution	Injected Volume	Reference
Natural products								
Calbalain ()	SFC prep 80 system	CSP-1 (250×20mm, 5µm)	CO ₂ :MeOH/DCM (1/1) 85:15			50mg/mL 1/1 MeOH/DCM	1.8mL	
Ganbergin, (-)- Ganschisandrin, Galgravin and (-)- Veraguensin	pressure fraction- collection cyclones	CSP-2 (250×20mm, 5µm)	CO ₂ :MeOH/DCM (1/1) 80:20	150bar	60g/min	15mg/mL 1/1 MeOH/DCM	0.8mL	[35]
veraguensm	(Waters Corporation, Milford, MA, USA)	CSP-2 (250×20mm, 5µm)	CO ₂ :MeOH/DCM (1/1) 80:20			30mg/mL 1/1 MeOH/DCM	1.5mL	
Dihydromyricetin	SFC prep 80 system and six, high- pressure fraction- collection cyclones (Waters Corporation, Milford, MA, USA)	EnantioPak AD (250×20mm, 10µm)	CO ₂ :MeOH 60:40	100bar	40g/min	40mg/mL MeOH	-	[79]
Small p	harmaceutical molecul	es or intermediates						
1-benzyl-3-methyl- 2-piperidone	N.R.	Chiralpak AD-H (250×30 mm, 5µm)	CO ₂ :EtOH 95:5	NR 1	N.R. 120mL/min	N.R.	N.R.	[80]
Iodoaryl regioisomers		Chiralpak IC (250×30 mm, 5µm)	CO ₂ :1PrOH/DEA 75:25/0.25%					
3 Synthetic intermediates Compound A	Berger MGII and	Lux TM Cellulose-4 (250×30mm; 5µm).	CO ₂ : <i>n</i> -Hept/iPrOH/TFA 88:12 (80/20)/0.10%			29.2mg/mL 8/2 <i>n</i> -Hept/iPrOH		
Compound B Compound C	Thar SFC Prep 350 device (Waters,	Chiralpak IC (250×30 mm, 5µm)	CO ₂ :MeOH 90:10	100bar	N.R. 220mg/mL AcN/DCM	220mg/mL 1/1 AcN/DCM	/1 N.R. /2 M	[25]
	Boston, MA, USA)	Lux TM Amylose-2 (250×30 mm; 5µm).	CO ₂ : MeOH/AcN/DCM 50:50 (20/20/10)			50mg/mL 1/2 MeOH/DCM		
2 Propanoic acids Compound 4	Berger MGII and	Chiralcel OJ-H (250×30mm; 5µm)	CO ₂ :MeOH 70:30	100h a 1	120mL/min	100mg/mL MeOH	ND	[01]
Compound 14	device	Chiralcel OJ-H (250×30mm; 5µm)	CO ₂ :MeOH 80:20	TUUDar	100mL/min	75mg/mL MeOH	N.R.	[81]

Atropisomeric compounds	Waters MGII preparative device (Waters, Boston, MA, USA)	Chiralpak AD-H (250×30mm; 5µm)	CO ₂ :MeOH 70:30	100bar	85mL/min	N.R.	N.R.	[82]
	SFC-PICLAB hybrid		CO ₂ :EtOH 85:15			18mg/mL EtOH	250µL	
4 Isoxazole	10-20 device (PIC	Chiralpak AD-H	CO ₂ :EtOH 90:10	150bar	4 mL/min	15.8mg/mL EtOH	250µL	[83]
derivatives	Solution, Avignon,	(250×4.6mm; 5µm)	CO ₂ :EtOH 93:7	10000	, , , , , , , , , , , , , , , , , , , ,	13.2mg/mL EtOH	485µL	[05]
	France		CO ₂ :EtOH 90:10			13.5mg/mL EtOH	485µL	
S1D final ADI	Thar SFC Prep 350 device (Waters, Boston, MA, USA)	Chiralpak IC (250×30mm; 5µm)	CO ₂ :MeOH 48:52	100bar	180mL/min	145mg/mL 1/1 MeOH/DCM	N.R.	[24]
$S1P_1$ Illiai AP1		Chiralpak AD-H (250×4.6mm; 5µm)	CO ₂ :MeOH 50:50		130mL/min	80mg/mL MeOH	1mL	[34]
Pesticides								
	SFC prep 80 system and a six high- pressure fraction collection cyclones (Waters Corporation, Milford, MA, USA)	EnantioPak OD (250×20mm; 5µm)	CO ₂ :iPrOH 95:5					
4 β-cypermethrin stereoisomers		EnantioPak AD (250×20mm; 5µm)	CO ₂ :EtOH 80:20	100bar	40g/min	10mg/mL iPrOH	2mL	[84]
		EnantioPak AD (250×20mm; 5µm)	CO ₂ :EtOH 85:15					

N.R: Not Reported

Analyte	Preparative device	Stationnary phase	Mobile phase	Flow-rate	Concentration of feed solution	Injected Volume	Reference		
Natural products									
Isoborneol	Agilent system 1200 series (Palo Alto, CA, USA)	EnantioPak OD (250×10mm; 5µm)	<i>n</i> -Hex:EtOH 99:1	60g/min	50mg/mL <i>n</i> -Hex	N.R.	[85]		
α-Tocopherol	2010 A HT series HPLC instrument from Shimadzu Technologies (Kyoto, Japan)	Connected Silica SG120 and Inertsil SIL-100A (250×10 mm; 5µm)	<i>n</i> -Hex:iPrOH 98:2	10mL/min	N.R.	N.R.	[86]		
Small pharma	aceutical molecules or	intermediates							
N-CBZ-3- fluoropyrrolidine-3- MeOH	N.R.	Chiralpak AD (250×20mm; 10µm)	<i>n</i> -Hept:EtOH 80:20	2567 mL/min	151.5g/mL 6/4 <i>n</i> -Hept/EtOH	250mL	[87]		
		Lux TM Cellulose-1 (250×10mm; 5µm)	AcN:MeOH 1:9	5mL/min	830mg/mL 1:9 AcN:MeOH	1mL			
	Smartline HPI C	Lux TM Cellulose-3 (250×10mm; 5µm)	AcN:MeOH 1:14	5mL/min	1000mg/mL 1:14 AcN:MeOH	1mL	[88]		
cis-itraconazole	system (Knauer, Berlin, Germany)	Lux [™] Cellulose-1 (250×10mm; 5µm)	AcN:MeOH 1:10	5mL/min	1500mg/mL 1:10 AcN:MeOH	1mL			
		Lux TM Cellulose-3 (250×10mm; 5µm)	AcN:MeOH 1:5	5mL/min	1500mg/mL 1:5 AcN:MeOH	1mL			
Ketorolac	2010 A HT series HPLC instrument from Shimadzu Technologies (Kyoto, Japan)	Lux TM Amylose-2 (250×4.6 mm; 5µm)	<i>n</i> -Hex:iPrOH/FA 85:15/0.1%	1 mL/min	N.R.	20µL	[89]		

Tetrahydro-α-(1- methylethyl)-2-oxo- 1(2H)-pyrimidine acetic acids	Hitachi L-2000 device (Tokyo, Japan)	Chiralcel OD-H (250×4.6mm; 5µm)	n-Hex:EtOH/AA 920:80/0.2%	N.R.	N.R.	N.R.	[90]	
Bicalutamide	200LC Perkin-Elmer (Norwalk, CT, USA)	Chiralpak IA (250×10 mm, 5µm)	<i>n</i> -Hex:EA:EtOH 100:30:5	5.5 mL/min	32 g/mL EA	N.R.	[91]	
Cathinone derivatives	JASCO 880-PU pump (JASCO, Tokyo, Japan), with a Rheodyne 7125 injector (Rheodyne, Rohnert Park, CA, USA) and a JASCO 880-30 solvent mixer	amylose tris-3,5- dimethylphenylcarba mate (200×70 mm, 7µm)	<i>n</i> -Hex:EtOH/TEA 97:3/0.1%	1.5 mL/min	10 mg/mL EtOH	100µL	[92]	
Cathinone derivatives: pentedrone	Dionex UltiMate instrument (Dionex GmbH, Germany)	Dionex UltiMate instrument (Dionex GmbH, Germany) with a 3000	Chiralpak AS	<i>n</i> -Hex:iPrOH 97:3		1 mg/mL 97:30/0.4% <i>n</i> -Hex:iPrOH/DEA		
methylone	quaternary pump and a 3000 Automated Fraction Collector LC system	(250×10mm; 10µm)	n-Hex:iPrOH 85:15	2 mL/min	1 mg/mL 85:15/0.4% <i>n</i> -Hex:iPrOH/DEA	N.R.	[93]	
5-chloro-2-{2-[3,4- dihydroisoquinolin-2 (1H)-yl]ethyl}-2-met hyl-2,3-dihydro-1H- inden-1-one	Gilson 306 HPLC system (Gilson, Middleton, WI, USA)	Chiralpak AD-H (250×10mm; 5µm)	EtOH: <i>n</i> -Hex/DEA 5:95/0.1%	N.R.	3 mg/mL 5:95/0.1% EtOH: <i>n</i> -Hex/DEA	1 mL	[94]	

N.R: Not Reported

Analyte	Preparative device	Stationnary phase	Mobile phase	Pout	Flow-rate	Concentration of feed solution	Injected Volume	Reference
• Small	pharmaceutical molecul	es or intermediates						
Undisclosed	Thar 70 (Waters, Milford, MA, USA) SuperSep 600 unit (Novasep, Pompey, France)	Chiralpak IC (250×30mm, 5µm)	CO ₂ :MeOH/DEA 85:25/0.1%	80bar	70g/min	250mg/mL MeOH	2mL	. [51]
compound A	Hipersep LAB LC110 (Novasep, Pompey, France)	Chiralpak AD (250×110mm, 20µm)	<i>n</i> -Hept:EtOH:MeOH/TEA 80:10:10/0.10%		600mL/min	N.R.	N.R.	
	Analytical/semi-prep device (Waters, Spa, Milan, Italy)	Chiralpak IA (250×10 mm, 5µm)	CO ₂ :(<i>n</i> -Hept:EtOH/DEA) 70:30 (90:10/0.10%)	N.R.	10mL/min	10mg/mL iPrOH	50µL	[96]
σl receptor agonist	Jasco PU-2089 Plus pump, AS-2055 Plus autosampler and MD- 2010 Plus detector (Jasco Europe, Cremella, Italy)	Chiralcel OJ-H (250×10mm; 5µm)	MeOH/DEA 99.9/0.10%		3mL/min	3mg/mL MeOH	1mL	
Case study n°1 Insufficient solubility of compound A in CO ₂ -based mobile phase precluded use of SFC								
API compound A	Hipersep LAB LC110 (Novasep, Pompey, France)	Chiralpak AD (250×110mm, 20µm)	<i>n</i> -Hept:EA/DEA 50:50/0.50%		600mL/min	270mg/mL DCM	N.R.	[33]

Case study n°2 API compound B	SuperSep 600 unit (Novasep, Pompey, France)	Chiralpak IC (250×50mm, 5µm)	CO ₂ :EtOH/FA 80:20/0.50%	N.R.	450g/min	200mg/mL EtOH	N.R.	[33]
Case study n°3 API compound C	SuperSep 600 unit (Novasep, Pompey, France)	Chiralart SA (250×20mm; 5µm)	CO ₂ :iPrOH 60:40	0:40 N.R. 80g/min 100mg/mL AcN N.R.		[55]		
	SFC prep 80 system	Chiralpak AD-H (250×20mm, 5µm)	CO ₂ :MeOH 50:50	100bar	30g/min	2mg/mL MeOH	1, 2, 3 or 4mL	
Lenalidomide	(Waters, Milford, MA, USA)	(4-chloro- phenylcarbamoylated β-cyclodextrin) (250×20mm, 5μm)	CO ₂ :MeOH 60:40	100bar	50g/min	10mg/mL (25/75) DMSO/MeOH	3mL	[97]
Lenandonnue	Bischoff Corp HPD Pump & Lambda 1010	Chiralpak IC (250×20 mm; 5µm)	EA 100		10 mL/min	2.3mg/mL AcN	5mL	[98]
	Agilent 1260 Infinity HPLC system (Agilent Technologies, Waldbronn, Germany).	Chiralpak AD-H (250×4.6mm, 5µm)	MeOH 100		0.8 mL/min	5mg/mL MeOH	40µL	[99]
6-(4- aminophenyl)-5-	SFC prep 80 system (Waters, Milford, MA, USA)	Chiralpak AS (250×20 mm, 10µm)	CO ₂ :MeOH 55:45	170bar	60g/min	15mg/mL MeOH	2mL	
methyl-4, 5- dihydro-3(2H)- pyridazinone	Waters Autopurification LC System (Waters, Milford, MA, USA)	Chiralcel OJ (250×20 mm, 10µm)	MeOH 100		20 mL/min	15mg/mL MeOH	2mL	[100]
Corey lactone	ne SFC prep 80 system (Waters, Milford, MA, USA) Waters Autopurification LC System (Waters, Milford, MA, USA) Amylose tris-(3,5- dimethylphenyl carbamate (250×20 mm, 5µm)	CO ₂ :MeOH 80:20	200bar	60g/min	10mg/mL MeOH	N.R.		
diol		carbamate (250×20 mm, 5µm)	AcN:H ₂ O 93:7		20 mL/min	10mg/mL (93/7) AcN/H ₂ O	N.R.	[101]

Table 4 : Synopsis of the references by theme.

Field	Preparative SFC references	Preparative HPLC references	Other references
Mobile phase	[32-35] ; [40] ; [52] ; [56 ; 57]		[31] ; [39] ; [53-55]
CSP	[25]		[26-30] ; [36 ; 37] ; [42-44] ; [63-66]
Chiral capillary electrophoresis			[4-8]
Instrumentation	[18-20] ; [58]	[67]	
Reviews	[14 ; 15] ; [17]	[9-11]	[1;2;3];[12];[22];[24]
Processes	[45]	[23] ; [59-61] ; [76 ; 77]	[44]
Theory	[46] ; [49 ; 50]	[62]	[13]
Environmental parameters	[32] ; [78]		
Large-batch	[47 ; 48]	[68-75]	
Applications	[46] ; [78-84] ; [102]	[85-94]	[16]; [18]; [21]; [38]; [51]; [95-101]