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Bianca C. Maniglia, Dâmaris C. Lima, Manoel da Matta Júnior, Anthony Oge, Patricia Le-Bail, Pedro E.D. Augusto, Alain Le-Bail

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1 ***Dry heating treatment: a potential tool to improve the wheat starch properties for 3D food***  
2 ***printing application***

3 Bianca C. MANIGLIA <sup>a,b,c,d,\*</sup>, Dâmaris C. LIMA<sup>d</sup>, Manoel da MATTA JÚNIOR<sup>d</sup>, Anthony  
4 OGE<sup>a,c</sup>, Patricia LE-BAIL<sup>b,c</sup>; Pedro E. D. AUGUSTO<sup>d,e</sup>, Alain LE-BAIL<sup>a,c,\*</sup>

5  
6 <sup>a</sup>ONIRIS-GEPEA UMR CNRS 6144 Nantes - France

7 <sup>b</sup>BIA-INRAE UR 1268 Nantes – France

8 <sup>c</sup>SFR IBSM INRA CNRS 4202

9 <sup>d</sup>Department of Agri-food Industry, Food and Nutrition (LAN), Luiz de Queiroz, College of Agriculture  
10 (ESALQ), University of São Paulo (USP), Piracicaba, SP – Brazil

11 <sup>e</sup> Food and Nutrition Research Center (NAPAN), University of São Paulo (USP), São Paulo, SP, Brazil

12 \*biancamaniglia@usp.br; alain.lebail@oniris-nantes.fr  
13

14 ***Abstract***

15 The futuristic technology of three-dimensional (3D) printing is an additive manufacturing that  
16 allows **obtaining** creative and personalized food products. In this context, the study of food  
17 formulations (named as “inks”) to be processed through 3D printing is necessary. This work  
18 investigated the use of dry heating treatment (DHT), a simple and safe method, to improve the  
19 wheat starch properties **aiming** to produce hydrogels to be used as “inks” for 3D printing.  
20 Wheat starch was processed by dry heating for 2 (DHT\_2h) and 4 h (DHT\_4h) at 130 °C.  
21 Modified wheat starches showed an increase in granule size, but **processing** did not alter the  
22 granule’s shape nor surface, neither alter the molecular functional groups. On the other hand,  
23 DHT promoted slight molecular depolymerization, and **reduction of starch crystallinity**.  
24 Hydrogels “inks” based on the modified starches showed **lower peak apparent viscosity**  
25 **during pasting**, higher structural strength at rest, higher resistance to external stresses, higher  
26 gel firmness, and lower syneresis than hydrogels based on native starch. The hydrogels based  
27 on starch DHT\_4h showed the best printability (**greater ability to make a 3D-object by layer-**  
28 **by-layer deposition and to support its structure once printed**) and this “ink” showed better  
29 reproducibility. Another point observed is that DHT extended the texture possibilities of  
30 printed samples based on wheat starch hydrogels. These results suggested that DHT is a  
31 relevant process to improve the properties of hydrogels based on wheat starch, making this  
32 ink suitable for 3D printing application.

33 **Keywords:** modified starch, 3D food printing, additive manufacturing, personalized food,  
34 food texture.

## 35 1. Introduction

36 The three-dimensional (3D) printing is an emerging technology to create personalized  
37 objects from digital models through the successive layer-by-layer deposition of materials such  
38 as plastic, metal, ceramics, or even living cells (Bhatia & Ramadurai, 2017). **Currently, 3D**  
39 **printing technology has been used for food printing, defining** new borders for food processing  
40 and being able to deliver a product that suits special consumer's criteria of taste, texture, cost,  
41 convenience and nutrition. **For instance,** 3D food printing can create unique novel textured  
42 foods, healthy foods, and smooth and easy-to-swallow foods **for elderly people** (Dankar,  
43 Haddarah, Omar, Sepulcre, & Pujola, 2018).

44 Extrusion **is the** most common 3D printing method for food applications, **being**  
45 **relatively** easy to develop and **presenting different alternatives of “inks”** (Guvendiren, Molde,  
46 Soares, & Kohn, 2016; Tan, Toh, Wong, & Lin, 2018). This process depends heavily on the  
47 physical properties of the material to be processed, which is one of the most important  
48 research topics **about** 3D printing (Liu, Liang, Saeed, Lan, & Qin, 2019). **Therefore, it is**  
49 **important to study** physicochemical and rheological aspects of **possible “inks”** used to print -  
50 in other words, their printability, **i.e. their ability to make a 3D-object by layer-by-layer**  
51 **deposition, to obtain a defined shape and to support its structure once printed** (Godoi,  
52 Prakash, & Bhandari, 2016).

53 In this context, it is important to study the ability of starch sources to be used as  
54 **“inks”**. As one of the most important carbohydrates in human diets and **one of the most**  
55 **available in nature,** starch is widely used as thickening and gelling agents in many food  
56 formulations (BeMiller, 2011). The gelatinized starch paste shows a shear-thinning behavior  
57 and instant responses to the applied shear strains, being easily extruded from the fine nozzle  
58 (Evans & Haisman, 1980; Le Tohic et al., 2018). **Finally, the starch hydrogel can present**  
59 **good printability, depending on the sources and modification processes.**

60 In fact, some studies have already explored starchy products as “inks” for 3D printing,  
61 like potato starch (Yang, Zhanga, & Bhandari, 2018), mashed potatoes (Liu et al., 2018),  
62 potato puree (Dankar et al., 2018), wheat flour (Liu, Liang, Saeed, Lan, & Qin, 2019a), rice  
63 flour (Anukiruthika, Moses, & Anandharamakrishnan, 2020). Moreover, our group has  
64 evaluated some alternatives to modify starches and improve the 3D printability, such as the  
65 ozone and dry heating treatment of cassava starch (Maniglia et al. (2019) and Maniglia et al.  
66 (2020)). However, the approach of starch modification for 3D printing application is still  
67 under-explored in the literature, considering different sources and modification technologies.

68 Wheat starch is the third most-produced starch in the world, being a coproduct in the  
69 vital gluten industry (Shevkani, Singh, Bajaj, & Kaur, 2017). It has an important role in many  
70 food products (Agama-Acevedo, Flores-Silva, & Bello-Perez, 2019; Ubeyitogullari & Ciftci,  
71 2016), being frequently used in gluten-free baked products (Houben, Höchstötter, & Becker,  
72 2012). However, native wheat starch produces weak-bodied, cohesive, rubbery pastes when  
73 heated and undesirable gels when the pastes are cooled (Punia, Sandhu, Dhull, & Kaur, 2019).

74 In this way, modification processes can be used to improve wheat starch properties  
75 and applicability. Among different possibilities, dry heat treatment (DHT) is a simple physical  
76 method, considered a “green” technology (chemical-free, simple, safe and pollution-free) to  
77 alter starch structure and functionality (Chandanasree, Gul, & Riar, 2016; Gou et al., 2019; K.  
78 Liu, Hao, Chen, & Gao, 2019; Maniglia, Castanha, Le-Bail, Le-Bail, & Augusto, 2020; Oh,  
79 Bae, & Lee, 2018). Moreover, Maniglia et al. (2020) described that cassava starch DHT  
80 resulted in stronger gels with better 3D printability than the native starch. It is worth mention  
81 each starch source reacts differently to each modification technology, justifying the present  
82 work.

83 Wheat starch modification by DHT has scarcely been reported in the literature,  
84 although some studies are available with wheat flour (Nakamura, Koshikawa, & Seguchi,  
85 2008; Ozawa, Kato, & Seguchi, 2009; Seguchi, 2001) – where the observed changes are  
86 mainly associated to the proteins.

87 Based on this context, this study aimed to use the DHT as a tool to improve the 3D  
88 printability of “inks” based on wheat starch hydrogels. To achieve it, firstly the effect of DHT  
89 was studied in relation to the wheat starch structure and properties. Then, the potential for 3D  
90 printing application was evaluated. The innovation of this study involves developing  
91 functional ingredients to be used as gelling and thickening agents to bring better printability  
92 for food products, also expanding the application and potential of wheat starch.

93

## 94 **2. Material and methods**

### 95 **2.1 Material and dry heating treatment (DHT)**

96 Native wheat starch (moisture content: 10.1 g/100 g) was purchased from Merck  
97 KGaA (Germany). All the chemicals were of analytical grade.

98 The dry heating treatment (DHT) was based on the work of Chandanasree et al.  
99 (2016). Wheat starch (50 g) was distributed evenly in a thin layer (~ 1 mm) and packaged on  
100 aluminum foil, as an envelope (30 x 20 cm), which was closed with a high-temperature tape  
101 to avoid moisture loss. The envelope was then placed in a hot-air convective oven (XU032,

102 Frances Etuves, Chelles, France) with air at 130 °C and  $1 \pm 0.1 \text{ m. s}^{-1}$  for 2 and 4 h (named  
103 DHT\_2h and DHT\_4h, respectively).

104 The temperature was recorded using a datalogger with six K-type thermocouples  
105 (AOIP Datalog, Ris-Orangis, France) of 0.5 mm diameter for measuring the temperature in  
106 different locations within the sample (Supplementary Fig. S1A). The treatment time was  
107 counted from the moment where all parts of the envelope reached the processing temperature  
108 (130 °C) - the heat-up time was 20 min (Supplementary Fig. S1B). After processing, the  
109 starch was cooled, sieved (250  $\mu\text{m}$ ), and stored in glass containers for further analysis.

110

## 111 2.2 Granule characterization: morphology and size distribution

112 The starch granules morphology (control and modified) was observed using a light  
113 microscope (model L1000, Bioval, Curitiba, Brazil; with a 20 W halogen lamp, the  
114 magnification of 40 x) and a digital camera of 5.1 megapixels (MT9P001, Aptina, Colorado,  
115 USA). The starch granules were dispersed in distilled water (1:1, v/v). Then, they were placed  
116 on a glass slide, covered by a glass coverslip, and directly evaluated. A polarized light filter  
117 coupled to the system was used to observe the Maltese crosses.

118 The granule size distribution was determined using the dynamic light scattering (DLS)  
119 of Malvern Master Size (3000, Malvern Instruments Ltd, Worcestershire, UK). The samples  
120 were dispersed in the air. The analyse was carried out 4 times for each sample. From the  
121 obtained data, the volume-based mean (D[4,3]) and the area-based mean (D[3,2]) diameters  
122 were calculated. The diameters d(0.1), d(0.5), and d(0.9) were also obtained, describing the  
123 dimension that 10 %, 50 %, and 90 % of the granules were smaller, respectively.

124

## 125 2.3 Molecular characterization: pH, functional groups, molecular size distribution

126 pH values were measured in the starch suspension of 10.7% (w/w) in distilled water,  
127 under constant stirring at room temperature (25 °C), using a potentiometer (model TEC-5  
128 mode, Tecnal, Piracicaba, Brazil).

129 The carboxyl and carbonyl contents were determined by the method of Smith (1967)  
130 and Chattopadhyay, Singhal, & Kulkarni (1997), respectively, with some modifications, as  
131 described by Maniglia et al. (2020). Their content was expressed as the number of carboxyl  
132 (COOH / 100 GU) or carbonyl (CO/100 GU) groups in relation to 100 units of glucose. The  
133 analyses were done in triplicate.

134 The changes in the functional groups were also evaluated using the infrared  
135 spectroscopy technique. Fourier transform infrared (FTIR) spectroscopy was conducted using  
136 a Spectrum 100™ FTIR instrument (Perkin-Elmer, Shelton, USA) equipped with an  
137 attenuated total reflection (ATR) accessory. All the spectra were the average of 16 scans from  
138 4000 cm<sup>-1</sup> to 650 cm<sup>-1</sup> and were acquired at a resolution of 4 cm<sup>-1</sup>.

139 The molecular size distribution profile (control, DHT\_2h, and DHT\_4h) was  
140 determined using a gel permeation chromatography (GPC) system, according to Song & Jane  
141 (2000), with some modifications described by Maniglia, Lima, Junior, et al. (2019).

142

#### 143 **2.4 Thermal and crystalline properties of starch**

144 The thermal properties during starch gelatinization were determined using a Multi-Cell  
145 Differential Scanning Calorimeter (MC-DSC) – (TA Instruments, Lindon, USA). The starch  
146 samples were weighed and hydrated directly into the ampoules (10 g dry starch / 100 g  
147 suspension). An ampoule with deionized water was used as a reference and three runs for  
148 each sample were analysed. The MC-DSC heating program consisted of going from 20 to 120  
149 °C at a rate of 2 °C/min. The onset temperature (To), the peak temperature (Tp), the  
150 conclusion temperature (Tc), and the enthalpy (ΔH) associated with the starch gelatinization  
151 interval were calculated with the aid of the Universal Analyzer software (TA Instruments,  
152 New Castle, USA).

153 The crystallinity of starch samples was determined by X-ray diffraction (XRD).  
154 Before the XRD analysis, the starch samples were maintained in a desiccator containing a  
155 saturated BaCl<sub>2</sub> solution (25 °C, a<sub>w</sub> = 0.900) for 7 days to ensure constant water activity. The  
156 sample was examined by X-ray diffraction equipment Inel X-ray equipment (Inel, Centre-Val  
157 de Loire, France) operated at 40 kV and 30 mA. The CuKα radiation (0.15405 nm) was  
158 selected using a quartz monochromator. Diffracted intensities were monitored (2θ) by a  
159 sensitive detector (CPS 120, Inel, Centre-Val de Loire, France). The resulting diffraction  
160 diagrams were normalized between 3 and 30 ° (2θ). The curves obtained were smoothed using  
161 the Origin software, version 2018 (Microcal Inc., Northampton, USA). The relative  
162 crystallinity was calculated as the ratio of upper diffraction peak area to the total diffraction  
163 area, following the method described by Nara & Komiya (1983) and considering 2θ ranging  
164 from 3 to 30 °.

#### 165 **2.5 Starch pasting behaviour and hydrogel properties (rheology and firmness)**

166 Starch pasting properties were determined using a Rapid Visco Analyzer (RVA4,  
167 Newport Scientific Pvt. Ltd., Warriewood, Australia) with ThermoLine for Windows  
168 software, version 3.0. The RVA analysis consisted of heating, retention, and cooling cycle  
169 under constant shear (160 rpm). Following the standard conditions for starch characterization,  
170 starch suspensions of 10.7 g starch / 100 g (correct to 14 % moisture basis) were prepared for  
171 the RVA analyses. The procedure used was: keep at 50 °C for 1 min, then heat up to 95 °C (6  
172 °C.min<sup>-1</sup> and maintained 5 min), followed by cooling to 50 °C (6 °C.min<sup>-1</sup>), and finally,  
173 maintained at 50 °C for 2 min.

174 Different parameters are obtained to describe the pasting behaviour: peak apparent  
175 viscosity – PAV (the maximum paste apparent viscosity achieved in the heating stage),  
176 through apparent viscosity – TAV (minimum paste apparent viscosity achieved after holding  
177 at the maximum temperature), final apparent viscosity – FAV (the apparent viscosity at end of  
178 run), breakdown – BD (difference between PAV and TAV), setback – SB (the difference  
179 between FAV and PAV), and pasting temperature – PT (the temperature at which starch  
180 granules begin to swell and gelatinize due to water uptake). For a better interpretation of these  
181 parameters, the relative breakdown (RBD) and relative setback (RSB) were also calculated,  
182 following the study of Castanha, Matta Junior, & Augusto (2017). The RBD (calculated as the  
183 ratio between the BD and PAV values) can be associated with the facility of starch granules to  
184 disrupt, while the RSB (calculated as the ratio between the SB and TAV values) can be  
185 associated to the retrogradation tendency.

186 Hydrogels were prepared by dispersing starch in distilled water (10.7 g dry starch/ 100  
187 g, being the starch mass corrected to 14 % moisture basis), and then heating it at 85 ± 2 °C in  
188 glass containers in a water bath for 20 min. Then, the pastes were placed in plastic cups  
189 (40 mm diameter × 20 mm height) and stored for 24 h in the refrigerator (5 ± 2 °C) to gelling.  
190 The cups were kept in a desiccator with water at the bottom to ensure uniform moisture. The  
191 obtained hydrogels were evaluated in relation to their viscoelasticity and firmness behaviours.

192 The viscoelastic properties were analysed by oscillatory stress sweep using a HAAKE  
193 MARS rheometer (Vane FL40, ThermoFisher, Karlsruhe, Germany). The hydrogels were  
194 loaded between the stainless steel parallel plates geometry (with a diameter of 25 mm and a  
195 gap of 1 mm), trimmed, and their edge was covered with silicon oil to prevent water  
196 vaporization. The stress sweep measurements were carried out at 22 °C with a logarithmically  
197 increasing shear stress at a frequency of 0.1 Hz (initial stress 0.001 Pa). This analysis was  
198 conducted in triplicate.

199 The hydrogel firmness was determined by a puncture assay using a texture analyzer  
200 TA TX Plus (Stable Micro Systems Ltd., Surrey, UK) with a load cell of 50 kgf (490.3 N).  
201 The samples were removed from the refrigerator and analysed immediately to ensure a  
202 constant temperature. The samples were penetrated until a distance of 4 mm using a  
203 cylindrical probe (P/0.5R, 0.5 in of diameter) at  $1 \text{ mm} \cdot \text{s}^{-1}$ . The equipment measured the force  
204 as a function of penetration depth. Gel firmness was evaluated by the energy required to  
205 penetrate the material (calculated by the area below the curve force versus distance of  
206 penetration). This analysis was conducted in triplicate.

207

## 208 2.6 Syneresis measurement

209 Syneresis was determined according to the method of Yeh & Yeh (1992). Briefly,  
210 starch suspensions (5 % w/w) were gelatinized at 85 °C for 20 min in a temperature-  
211 controlled water bath followed by cooling to room temperature. The pastes were then stored  
212 for 24 h at 4 °C. Syneresis was measured gravimetrically as the amount of water (g water/ 100  
213 g gel) released after centrifugation at 1500 g for 15 min. Triplicate of the results were read  
214 and the mean reported.

215

## 216 2.7 3D printing process

217 The “inks” (starch hydrogels) were prepared identically as described on the last  
218 section. After heating, the hydrogels were transferred for the printer syringes (60 mL), cooled  
219 to room temperature to form a weak gel-like structure, and stored in the refrigerator ( $5 \pm 2 \text{ }^\circ\text{C}$ )  
220 for 24 h before the printing.

221 The printing process was carried out in a 3D printer (Stampante 3DRAG V1.2, Futura  
222 Elettronica, Gallarate, Italy) presented in Fig. 1. The syringes with the hydrogels were  
223 removed from the refrigerator and immediately positioned in the 3D printer. A nozzle with  
224 diameter of 0.8 mm and height of 18 mm was coupled in the syringes. The 3D printing  
225 conditions were: nozzle movement of 5 mm/s and an extrusion rate of 4.5 mg/s at 20 °C.

226 Three physical models (heart, star, and cylinder shapes) were created using the  
227 Repetier Host V2.0.1 and Slic3r software (Hot-World GmbH & Co. KG, Willich, Germany).  
228 The dimensions of the cylinder shape were 2 cm x 4 cm (diameter x height), for the star shape  
229 were 2.5 cm x 2.5 cm x 2 mm (Length x Width x Height), and for the heart shape were  
230 5 cm x 6 cm x 2 mm (Length x Width x Height). 5 samples were printed for each hydrogel and



231 shape. All the printing experiments were carried out at room temperature. After printing, the  
232 samples were photographed.

233 **The reproducibility** of the 3D printed samples was evaluated considering the weigh  
234 and the dimensions of the sample printed. Each printed cylinder was weighed in analytical  
235 balance (**AZ214, Sartorius, Göttingen, Germany**) and their dimensions (height and diameter)  
236 were measured in five different positions using a **digital paquimeter (CD-6 CSX-B model,**  
237 **Mitutoyo, Roissy-en-France, France**).

238 Printed samples in cylinder shape were conditioned in a refrigerator (at  $5 \pm 2$  °C) for  
239 24 h, inside of a desiccator with water to avoid dehydration. Then, these printed samples were  
240 used for texture analysis.

241

## 242 **2.8 *Texture profile of the printed hydrogels***

243 A texture **analyser** TA-XT+ (Stable Microsystems, Surrey, UK) was used to assess the  
244 printed cylinders. These analyses were conducted immediately in the samples removed from  
245 the refrigerator in triplicates for each sample.

246 The texture profile analysis (TPA) measurement consisted of two compression-  
247 decompression cycles separated by a time interval of 10 s, at a rate of 1 mm/s, using a  
248 **cylindric** probe (25 mm **diameter**, Code P/25, Stable Micro System Ltd., **Surrey, UK**). The  
249 probe compressed the sample to 25 % (6.25 **mm**) of the initial height (25 mm) before  
250 decompression using a 50 kgf (490.3 N) load cell.

251 All the textural parameters were measured and calculated by the instrument software  
252 from the resulting force-deformation curves, including hardness, adhesiveness, cohesiveness,  
253 springiness, gumminess, and chewiness.

254

## 255 **2.9 *Experimental design and Statistical analysis***

256 A completely randomized design was conducted. All processes and **analyses** were  
257 carried out at least in triplicate. To evaluate differences, analysis of variance (ANOVA) and  
258 Tukey's test at a 5 % significance level were accomplished by using the software *Statistic 13*  
259 (StatSoft, USA).

260

## 261 **3. Results and Discussion**

### 262 3.1 Starch granule: size distribution and morphology

263 Fig. 2 shows the granule size distribution (A) and specific diameters (B) for the  
264 control and modified wheat starch. The samples displayed a unimodal size distribution and  
265 DHT resulted in larger granules sizes (Fig. 2A), being this effect consistent with the duration  
266 of the dry heating process: the D[4,3] varied from 21.7  $\mu\text{m}$  for the control starch to 22.6 and  
267 24.1  $\mu\text{m}$  for DHT\_2h and DHT\_4h, respectively (Fig. 2B). The same was observed for the  
268 D[3,2], varied from 13.3  $\mu\text{m}$  for the control starch to 13.7 and 14.1  $\mu\text{m}$  for DHT\_2h and  
269 DHT\_4h, respectively (Fig. 2B). Since the parameter D[4,3] is more sensitive to larger  
270 granules, while D[3,2] is more sensitive to the fraction of smaller granules (Lopez-Sanchez et  
271 al., 2011), it is noted that the greatest effect of the DHT occurred for the larger granules. This  
272 behavior is similar to that observed for cassava starch treated with DHT (Maniglia et al.,  
273 2020). It can be associated with a possible expansion in the starch granule due to water  
274 vaporization, such as during flash vaporization. In fact, part of the water content in the  
275 samples was vaporized during DHT, as reflected by the moisture of modified wheat starches  
276 (control: 10.1 g/100 g; DHT\_2h: 4.7 g/100 g and DHT\_4h: 4.5 g/100 g). The dry heating  
277 process also increased the number of granules with bigger sizes considering the d0.1, d0.5,  
278 and d0.9 values.

279 Fig. 3 shows the optical microscopy of the control and treated wheat starches using  
280 both nonpolarized and polarized light. The microscopies show wheat starch granules with  
281 lenticular morphology with a large variety of granules size, which is consistent with previous  
282 observation (Hruska, Arevalo, Muñoz, & Avila, 2019). While the birefringence effect (Maltese  
283 cross is visible) can be seen in all the images, the processed samples showed a shine  
284 reduction. Chung, Liu, & Hoover (2010) attributed this behavior to the temperature elevation  
285 at the center of the starch granule, which supposedly damage the amylopectin double helices  
286 structure. The same behavior was observed by Liu et al. (2019) in the case of dry heating  
287 treatment for waxy potato starch. Once the Maltese crosses still can be seen, it indicates the  
288 thermal energy provided was not sufficient to destroy amylopectin double helices structure.

289 These results indicated that DHT did not alter the shape nor altered the granules  
290 surface; however, it slightly affected the internal microstructure of wheat starch granules and  
291 promoted an expansion of the granule size – which can affect starch properties.

292

### 293 3.2 Starch molecular properties

294 DHT processing did not change the pH of the wheat starches suspensions (control:  
295  $5.90 \pm 0.10$ , DHT\_2h:  $6.06 \pm 0.15$ , and DHT\_4h:  $5.98 \pm 0.18$ ), and no carbonyl and carboxyl  
296 formation was found. It indicates that DHT did not promote starch oxidation. It is interesting  
297 to observe that cassava starch, at the same processing conditions, presented carbonyl groups  
298 formation (Maniglia et al., 2020), highlighting each starch source are differently affect by  
299 processing.

300 Fig. 4(A) shows the vibrational spectra of the control and modified starches. All  
301 samples showed the same characteristic bands at  $3323\text{ cm}^{-1}$ ,  $2934\text{ cm}^{-1}$ ,  $1648\text{ cm}^{-1}$ ,  $1339$   
302  $\text{cm}^{-1}$ ,  $1150\text{ cm}^{-1}$ ,  $1020\text{ cm}^{-1}$ , and  $996\text{ cm}^{-1}$ . The bands at  $3500$  and  $2900\text{ cm}^{-1}$  are associated  
303 with OH bonds and  $\text{CH}_2$  deformation, respectively (Kačuráková & Mathlouthi, 1996). Bands  
304 at  $1650\text{ cm}^{-1}$  are associated with water molecules absorbed in the amorphous region (Kizil,  
305 Irudayaraj, & Seetharaman, 2002) and the stretching vibration of the CO band (amide I). The  
306 peak at  $1339\text{ cm}^{-1}$  can be associated to OH bending due to the primary or secondary alcohols  
307 (Muscat, Adhikari, Adhikari, & Chaudhary, 2012). The fingerprint region of starch is the  
308 bands at  $1200\text{-}900\text{ cm}^{-1}$  (showed in Fig. 4(B)) and this region provided information about  
309 changes in the polymeric structure and conformation of starch (Dankar et al., 2018). The  
310 intense IR band at  $996\text{ cm}^{-1}$  can be mainly attributed to skeletal mode vibrations of  $\alpha\text{ 1} \rightarrow 4$   
311 skeletal glycoside bonds (Jao & Ko, 2002). In addition, the middle strong peaks at  $1150\text{ cm}^{-1}$   
312 and  $1060\text{ cm}^{-1}$  are mainly attributed to the stretching vibration of CO and bending vibration  
313 of C-O-H (Hou et al., 2019). The band at  $1060\text{ cm}^{-1}$  has been linked to amorphous structures  
314 (Xiong, Li, Shi, & Ye, 2017). On the other hand, the band at  $996\text{ cm}^{-1}$  can be associated with  
315 the single helix crystalline related to hydrogen bonds of anhydroglucose unit (Xiong et al.,  
316 2017).

317 It was not observed a significant difference between the presence and intensity of the  
318 bands in control and modified starches. These results indicate and confirm that the dry heating  
319 treatment did not modify the functional groups.

320 Fig. 5 shows the molecular size distribution of the control and treated wheat starches.  
321 The first peak consists of molecules of larger size and more ramifications, which can be  
322 associated with amylopectins, while the second peak represents molecules of smaller size and  
323 a linear structure, which can be associated with amyloses. DHT reduced the molecular size  
324 (Fig. 5): the first peak was slightly reduced, the fraction of molecules with intermediary size  
325 was also reduced, and the second peak was increased; moreover, the first and second peaks

326 scrolled to the right. It indicates the DHT promoted the formation of molecules with lower  
327 size (which are retained longer in the chromatographic column) due to depolymerization  
328 promoted by cleavage of glycosidic linkages. The depolymerization affected mainly the  
329 intermediate size molecules. Maniglia et al. (2020) also observed this behaviour for cassava  
330 starch, although with a much more accentuated effect. These results are also consistent with  
331 the study of Lei et al. (2020) involving maize starch. The authors observed the temperature  
332 increasing of DHT reduced drastically the long-amylose chains, while more short-amylose  
333 chains were formed.

334 The molecular results reinforce each starch source are differently affect by processing.  
335 Consequently, further structure and properties evaluations are needed.

336

### 337 3.3 *Thermal and crystalline properties of starch*

338 The differential scanning calorimetry (DSC) thermograms (A) and the X-ray  
339 diffractograms (B) of the control and modified wheat starches are shown in Fig. 6. The  
340 gelatinization properties were obtained (Table 1) from Fig. 6(A). There was no difference in  
341 the onset ( $T_o$ ) and the final ( $T_f$ ) temperatures among treatments. The peak temperature ( $T_p$ )  
342 increased significantly ( $p < 0.05$ ) as the treatment period increased, and the gelatinization  
343 enthalpy ( $\Delta H$ ) was reduced ( $p < 0.05$ ) only for the DHT\_4h starch. Lei et al. (2020) also  
344 observed a reduction of the gelatinization enthalpy ( $\Delta H$ ) for maize starch treated by DHT.

345 The enthalpy of gelatinization is an indirect measure of the amount of starch  
346 crystalline region (or double helices structure) which are broken during heating (Liu, Yu, Xie,  
347 & Chen, 2006). Therefore,  $\Delta H$  reduction can indicates the formation of weaker structures,  
348 which were less crystalline. In fact, Fig. 6(B) show the typical A-type diffraction pattern of  
349 wheat starch, with two singlet peaks ( $2\theta$ ) at  $15^\circ$  and  $22.8^\circ$ , a doublet peak ( $2\theta$ ) at  $16.8^\circ$  and  
350  $17.8^\circ$ , and two weak peaks ( $2\theta$ ) at  $5.6^\circ$  and  $20^\circ$ . In addition, Fig. 6(B) shows the relative  
351 crystallinity of DHT\_4h was lower than control and DHT\_2h starches. Therefore, the XRD  
352 results are consistent with the DSC results, indicating that DHT reduced the double-helical  
353 crystals and decreased the order of starch crystalline structure – as also observed by (Lei et  
354 al., 2020).

355 The thermal and crystalline results are consistent with the behavior observed by  
356 optical microscopy about the reduction of Maltese crosses brightness in modified starches. All  
357 these results pointed out that the DHT reduced the crystalline zones of wheat starch. Even so,  
358 the starch properties must be evaluated.

359

### 360 **3.4 Starch pasting properties**

361 Fig. 7 (A) shows the RVA curves for control and wheat starch treated by DHT. Table  
362 2 shows the pasting parameters obtained from RVA and the parameters.

363 The dry heating treatment significantly ( $p < 0.05$ ) decreased the peak apparent  
364 viscosity (PAV), and there was a greater effect on this parameter as treatment time increased.  
365 This effect can be associated with cleavage of glycosidic linkages which resulted in  
366 weakening of starch granules resulting in minor capacity to maintain the granule integrity  
367 (Chung, Min, Kim, & Lim, 2007).

368 The pasting temperature (PT) is the temperature at which starch granules begin to  
369 swell due to water uptake (Gałkowska & Juszczak, 2019). DHT applied for 4 h slightly  
370 increased PT, which can be related with the presence of a greater fraction of small-sized  
371 molecules, that require more energy to gelatinize than large-size molecules similar to that  
372 observed by Lima et al., (2020).

373 Moreover, as the dry heating treatment period increased, a significant reduction ( $p <$   
374  $0.05$ ) of the through apparent viscosity (TAV) and final apparent viscosity (FAV) occurred. It  
375 can be associated with the thermal degradation of crystalline structures (observed in DSC and  
376 XRD analysis) and depolymerization (cleavage of glycosidic linkages) promoted by DHT  
377 (Zou, Xu, Tian, & Li, 2019).

378 The increase of the DHT period also increased the RBD and reduced the RSB  
379 parameters, when compared to control starch (data shown in Table 2), indicating that the  
380 granules are more fragile, more susceptible to rupture, and less prone to re-associate.  
381 According to Castanha et al. (2017), this behavior can be attributed to the change in the  
382 molecular size distribution.

383 Therefore, pasting properties confirm the structural modifications due to DHT  
384 processing, as well as they indicate hydrogels can present unique properties.

385

### 386 **3.5 Hydrogel properties: rheology, firmness, and syneresis**

387 Fig. 7 shows the results of (B) oscillatory stress sweep analysis, (C) gel firmness, and  
388 (D) the energy required in a puncture assay, considering the control and the wheat starch  
389 modified by DHT (2 and 4 h). Table 2 shows the parameters obtained from oscillatory stress  
390 sweep analysis.

391 Fig. 7(B) shows the representative storage ( $G'$ ) and loss ( $G''$ ) modulus curves: the  $G'$   
392 in this plateau region is a measure of the elasticity of material at very small deformation and  
393 it can be taken as a measure of the structural strength of mechanical rigidity of material at rest  
394 (Lille, Nurmela, Nordlund, Metsä-Kortelainen, & Sozer, 2018; Mezger, 2006). Moreover, for  
395 both control and modified starches,  $G' > G''$  which indicates that all samples show a  
396 solid/elasticity dominating gel-like structure.  $G'$  increased with the DHT period, as well as the  
397 ratio  $G'/G''$ , which indicates this treatment promoted the formation of a gel with greater  
398 rigidity and capacity to recover the shape after suffering stress. Therefore, this hydrogel  
399 probably shows good properties to be printed, because it can recover better the shape when it  
400 is deposited on the surface – being this evaluated in the next section.

401 The yield stress ( $\sigma_{0,G'}$ ) is the stress where  $G'$  starts to decrease after the plateau region  
402 (Table 2), which indicates the stress at which the network/structure of the material starts to  
403 break down. The  $\sigma_{0,G'}$  can be related to the ability of the material to keep its shape under  
404 gravity and the stresses generated by material layers deposited on top of it (Lille et al. 2018).  
405 The  $\sigma_{0,G'}$  value was greater as the DHT period increases. According to Lille et al. (2018), a  
406 material with high structural strength at rest ( $G'$ ) and a certain degree of resistance to external  
407 stresses ( $\sigma_{0,G'}$ ) is required to ensure good printability of a material. In this way, we can  
408 observe that the DHT improved these rheological parameters of the wheat starch hydrogel,  
409 indicating that it can be a better material to be used in 3D printing.

410 Fig. 7(C) and 7(D) show the gels produced with modified starches showed higher  
411 firmness than the control, and this behavior was more accentuated as the treatment period  
412 increased. The molecular depolymerization promoted by DHT (Fig. 5) probably resulted in  
413 size distribution that better re-associate and packing, forming a stronger three-dimensional  
414 network structure. In this way, DHT promoted the formation of stronger gels, expanding the  
415 industrial applications of wheat starch.

416 During starch gels cooling, molecular retrogradation and amylose tendency to  
417 crystallize releases water from the gels – which is called syneresis. The syneresis prejudices  
418 the printability of the hydrogels, as well as the consumer perception. DHT reduced syneresis,  
419 which is an interesting result: control:  $7.38 \pm 0.71$  g water/ 100 g gel, DHT\_2h:  $5.19 \pm 0.19$  g  
420 water/ 100 g gel, DHT\_4h:  $3.62 \pm 0.13$  g water/ 100 g gel). As a result, these “inks” appeared  
421 well adapted to 3D printing use - in fact, this behavior can be seen in Fig. 8. Pictures of the  
422 syneresis in the hydrogels are shown in the Supplementary Fig. S2.

423

### 424 **3.6 3D printing performance and characterization**

425 Fig. 8 shows the star, heart, and cylinder shaped structures, printed using the hydrogels  
426 based on control and modified starches as “inks”. Overall, the printed samples in our work  
427 were very similar to the pre-designed virtual model, indicating that the 3D printing could  
428 achieve precise printing with a personalized design using wheat hydrogels. All hydrogels  
429 could be smoothly extruded from the nozzle. However, the hydrogel based on control starch  
430 showed a scattering on the surface and a syneresis effect (visible in the star shape, which is  
431 consistent with the results presented in Section 3.4). The modified starches showed higher gel  
432 consistency, without spreading on the surface. The hydrogel based on DHT\_4h starch showed  
433 the best resolution with the lines having higher cohesiveness and being more continuous. The  
434 behavior of the hydrogels after 3D printing can be associated with the rheological parameters:  
435 the hydrogels with higher  $G'$  and  $G''$  showed the best printability.

436 Texture is an important sensory attribute of food acceptability (Yu, Ren, Zhao, Cui, &  
437 Liu, 2020). In this way, the printed cylinders were evaluated in relation to its textural  
438 parameters (Table 3). Results showed there was no significant difference in the cohesiveness,  
439 springiness, and chewiness values of printed samples. On the other hand, the increase of the  
440 heating period caused a significant ( $p < 0.05$ ) increase on hardness. According to Yang, Zhang,  
441 Prakash, & Liu (2018b), higher  $G'$  indicates stronger mechanical strength often accompanied  
442 by greater hardness, behavior also observed in our results. The modified starches resulted in  
443 printed samples with less adhesiveness. According to Anukiruthika, Moses, &  
444 Anandharamakrishnan (2020), less adhesive “inks” flow more easily through the nozzle,  
445 contributing to the 3D printing process. In this way, DHT extended the possibility of textures  
446 of printed samples based on wheat starch hydrogels.

447 About reproducibility, the hydrogels based on modified starches showed lower weight  
448 and higher height than hydrogel based on control starch. The diameter did not show a  
449 significant difference among the treatments. Moreover, by comparing the standard deviation,  
450 we observed lower values for DHT\_4h, indicating this treatment formed hydrogel were more  
451 reproducible 3D printing.

452 Finally, comparing this study with the previous of our group (Maniglia et al., 2020),  
453 we can observe that the same physical modification, DHT, showed different effects for each  
454 starch source, affecting the final printability characteristic. The modified cassava starch was  
455 more depolymerized, suffered oxidation, and showed gel firmness much higher than the

456 native one, when compared with the changes here reported for wheat starch. Modified wheat  
457 starch showed a reduction in syneresis, which is important for 3D printing application. In this  
458 way, our results provide information on the benefits DHT modification can bring to each  
459 source, mainly focusing on the application of 3D printing.

460

#### 461 **4. Conclusion**

462 This work investigated the use of dry heating treatment (DHT) as a tool to improve the  
463 printability of hydrogels (“inks”) based on wheat starch. DHT caused a slightly expansion of  
464 granule size; however, it did not alter the shape of the starch granules nor their surface. DHT  
465 did not change the functional groups of the starch, indicating a negligible oxidation. However,  
466 slight depolymerization of starch was observed, in special in the middle size molecules, as  
467 well as a reduction of the crystalline zones in starch granule. The hydrogels based on the  
468 modified starches showed lower apparent viscosity during pasting and greater structural  
469 strength at rest. The longer duration of DHT processing, higher was the firmness of the  
470 hydrogels, and more reduced was the syneresis. The changes promoted by DHT in wheat  
471 starch, in special in 4 h treatment, resulted in hydrogels with better printability and  
472 reproducibility in 3D printing.

473 In this study, we demonstrated that the DHT is a simple alternative to improve the  
474 properties of wheat starch, as well as extend the texture possibilities of printed samples based  
475 on wheat starch hydrogels. Finally, this study shows the possibilities to expand the potential  
476 application of this starchy source.

477

#### 478 **Declaration of Conflict of Interest**

479 The authors declare no conflict of interest.

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490

## 491 **7. References**

492 Agama-Acevedo, E., Flores-Silva, P. C., & Bello-Perez, L. A. (2019). Cereal Starch

493 Production for Food Applications. In *Starches for Food Application* (pp. 71–102).

494 Elsevier.

495 Anukiruthika, T., Moses, J. A., & Anandharamkrishnan, C. (2020). 3D printing of egg yolk

496 and white with rice flour blends. *Journal of Food Engineering*, *265*, 109691.

497 BeMiller, J. N. (2011). Pasting, paste, and gel properties of starch–hydrocolloid combinations.

498 *Carbohydrate Polymers*, *86*(2), 386–423.

499 Bhatia, S. K., & Ramadurai, K. W. (2017). 3D printing and bio-based materials in global

500 health. *SpringerBriefs in Materials*. *Google Scholar*.

501 Castanha, N., Matta Junior, M. D. da, & Augusto, P. E. D. (2017). Potato starch modification

502 using the ozone technology. *Food Hydrocolloids*, *66*, 343–356.

503 <https://doi.org/10.1016/j.foodhyd.2016.12.001>

504 Chandanasree, D., Gul, K., & Riar, C. S. (2016). Effect of hydrocolloids and dry heat

505 modification on physicochemical, thermal, pasting and morphological characteristics of

506 cassava (*Manihot esculenta*) starch. *Food Hydrocolloids*, *52*, 175–182.

507 <https://doi.org/10.1016/j.foodhyd.2015.06.024>

508 Chattopadhyay, S., Singhal, R. S., & Kulkarni, P. R. (1997). Optimisation of conditions of

509 synthesis of oxidised starch from corn and amaranth for use in film-forming applications.

510 *Carbohydrate Polymers*, *34*(4), 203–212. [https://doi.org/10.1016/S0144-8617\(97\)87306-](https://doi.org/10.1016/S0144-8617(97)87306-7)

511 [7](https://doi.org/10.1016/S0144-8617(97)87306-7)

512 Chung, H.-J., Liu, Q., & Hoover, R. (2010). Effect of single and dual hydrothermal treatments

513 on the crystalline structure, thermal properties, and nutritional fractions of pea, lentil, and

514 navy bean starches. *Food Research International*, *43*(2), 501–508.

515 <https://doi.org/10.1016/j.foodres.2009.07.030>

516 Chung, H.-J., Min, D., Kim, J.-Y., & Lim, S.-T. (2007). Effect of minor addition of xanthan

517 on cross-linking of rice starches by dry heating with phosphate salts. *Journal of Applied*

518 *Polymer Science*, *105*(4), 2280–2286. <https://doi.org/10.1002/app.26237>

519 Dankar, I., Haddarah, A., Omar, F. E. L., Sepulcre, F., & Pujola, M. (2018). 3D printing

520 technology: The new era for food customization and elaboration. *Trends in Food Science*

521           & *Technology*, 75, 231–242.

522 Evans, I. D., & Haisman, D. R. (1980). Rheology of gelatinised starch suspensions. *Journal of*  
523           *Texture Studies*, 10(4), 347–370.

524 Fanli Yang, Min Zhanga, Bhesh Bhandari, Y. L. (2018). Investigation on lemon juice gel as  
525           food material for 3D printing and optimization of printing parameters. *LWT - Food*  
526           *Science and Technology*, 87, 67–76. <https://doi.org/10.1016/j.lwt.2017.08.054>

527 Gałkowska, D., & Juszczak, L. (2019). Effects of amino acids on gelatinization, pasting and  
528           rheological properties of modified potato starches. *Food Hydrocolloids*, 92, 143–154.

529 Godoi, F. C., Prakash, S., & Bhandari, B. R. (2016). 3d printing technologies applied for food  
530           design: Status and prospects. *Journal of Food Engineering*, 179, 44–54.  
531           <https://doi.org/10.1016/j.jfoodeng.2016.01.025>

532 Gou, M., Wu, H., Saleh, A. S. M., Jing, L., Liu, Y., Zhao, K., ... Li, W. (2019). Effects of  
533           repeated and continuous dry heat treatments on properties of sweet potato starch.  
534           *International Journal of Biological Macromolecules*, 129, 869–877.  
535           <https://doi.org/10.1016/j.ijbiomac.2019.01.225>

536 Guvendiren, M., Molde, J., Soares, R. M. D., & Kohn, J. (2016). Designing biomaterials for  
537           3D printing. *ACS Biomaterials Science & Engineering*, 2(10), 1679–1693.

538 Hou, S.-W., Wei, W., Wang, Y., Gan, J.-H., Lu, Y., Tao, N.-P., ... Xu, C.-H. (2019).  
539           Integrated recognition and quantitative detection of starch in surimi by infrared  
540           spectroscopy and spectroscopic imaging. *Spectrochimica Acta Part A: Molecular and*  
541           *Biomolecular Spectroscopy*, 215, 1–8.

542 Houben, A., Höchstötter, A., & Becker, T. (2012). Possibilities to increase the quality in  
543           gluten-free bread production: an overview. *European Food Research and Technology*,  
544           235(2), 195–208.

545 Hruska, J. S., Arevalo, S., Muñoz, F., & Avila, A. (2019). Impedance characterization of  
546           wheat starch at various water contents. *Powder Technology*, 346, 425–432.

547 Jao, C.-L., & Ko, W.-C. (2002). 1, 1-Diphenyl-2-picrylhydrazyl (DPPH) radical scavenging  
548           by protein hydrolyzates from tuna cooking juice. *Fisheries Science*, 68(2), 430–435.

549 Kačuráková, M., & Mathlouthi, M. (1996). FTIR and laser-Raman spectra of oligosaccharides  
550           in water: characterization of the glycosidic bond. *Carbohydrate Research*, 284(2), 145–  
551           157.

552 Kizil, R., Irudayaraj, J., & Seetharaman, K. (2002). Characterization of irradiated starches by  
553           using FT-Raman and FTIR spectroscopy. *Journal of Agricultural and Food Chemistry*,  
554           50(14), 3912–3918.

555 Le Tohic, C., O'Sullivan, J. J., Drapala, K. P., Chartrin, V., Chan, T., Morrison, A. P., ...  
556 Kelly, A. L. (2018). Effect of 3D printing on the structure and textural properties of  
557 processed cheese. *Journal of Food Engineering*, *220*, 56–64.  
558 <https://doi.org/10.1016/j.jfoodeng.2017.02.003>

559 Lei, N., Chai, S., Xu, M., Ji, J., Mao, H., Yan, S., ... Sun, B. (2020). Effect of dry heating  
560 treatment on multi-levels of structure and physicochemical properties of maize starch: A  
561 thermodynamic study. *International Journal of Biological Macromolecules*, *147*, 109–  
562 116. <https://doi.org/10.1016/j.ijbiomac.2020.01.060>

563 Lille, M., Nurmela, A., Nordlund, E., Metsä-Kortelainen, S., & Sozer, N. (2018).  
564 Applicability of protein and fiber-rich food materials in extrusion-based 3D printing.  
565 *Journal of Food Engineering*, *220*, 20–27.  
566 <https://doi.org/10.1016/j.jfoodeng.2017.04.034>

567 Lima, D. C., Castanha, N., Maniglia, B. C., Matta Junior, M. D., La Fuente, C. I. A., &  
568 Augusto, P. E. D. (2020). Ozone Processing of Cassava Starch. *Ozone: Science &*  
569 *Engineering*, 1–18. <https://doi.org/10.1080/01919512.2020.1756218>

570 Liu, H., Yu, L., Xie, F., & Chen, L. (2006). Gelatinization of cornstarch with different  
571 amylose/amylopectin content. *Carbohydrate Polymers*, *65*(3), 357–363.  
572 <https://doi.org/10.1016/j.carbpol.2006.01.026>

573 Liu, K., Hao, Y., Chen, Y., & Gao, Q. (2019). Effects of dry heat treatment on the structure  
574 and physicochemical properties of waxy potato starch. *International Journal of*  
575 *Biological Macromolecules*, *132*, 1044–1050.  
576 <https://doi.org/10.1016/j.ijbiomac.2019.03.146>

577 Liu, Y., Liang, X., Saeed, A., Lan, W., & Qin, W. (2019). Properties of 3D printed dough and  
578 optimization of printing parameters. *Innovative Food Science & Emerging Technologies*,  
579 *54*, 9–18. <https://doi.org/10.1016/j.ifset.2019.03.008>

580 Liu, Z., Bhandari, B., Prakash, S., & Zhang, M. (2018). Creation of internal structure of  
581 mashed potato construct by 3D printing and its textural properties. *Food Research*  
582 *International*, *111*(March), 534–543. <https://doi.org/10.1016/j.foodres.2018.05.075>

583 Lopez-Sanchez, P., Nijse, J., Blonk, H. C. G., Bialek, L., Schumm, S., & Langton, M.  
584 (2011). Effect of mechanical and thermal treatments on the microstructure and  
585 rheological properties of carrot, broccoli and tomato dispersions. *Journal of the Science*  
586 *of Food and Agriculture*, *91*(2), 207–217. <https://doi.org/10.1002/jsfa.4168>

587 Maniglia, B. C., Castanha, N., Le-Bail, P., Le-Bail, A., & Augusto, P. E. D. (2020). Starch  
588 modification through environmentally friendly alternatives: a review. *Critical Reviews in*

589 *Food Science and Nutrition*, 0(0), 1–24. <https://doi.org/10.1080/10408398.2020.1778633>

590 Maniglia, B. C., Lima, D. C., Matta Junior, M. D., Le-Bail, P., Le-Bail, A., & Augusto, P. E.  
591 D. (2019). Hydrogels based on ozonated cassava starch: Effect of ozone processing and  
592 gelatinization conditions on enhancing 3D-printing applications. *International Journal of*  
593 *Biological Macromolecules*, 138, 1087–1097.  
594 <https://doi.org/10.1016/j.ijbiomac.2019.07.124>

595 Maniglia, B. C., Lima, D. C., Matta Junior, M. D., Le-Bail, P., Le-Bail, A., & Augusto, P. E.  
596 D. (2020). Preparation of cassava starch hydrogels for application in 3D printing using  
597 dry heating treatment (DHT): A prospective study on the effects of DHT and  
598 gelatinization conditions. *Food Research International*, 128((accepted for publication)),  
599 108803. <https://doi.org/10.1016/j.foodres.2019.108803>

600 Mezger, T. G. (2006). *The rheology handbook: for users of rotational and oscillatory*  
601 *rheometers*. Vincentz Network GmbH & Co KG.

602 Muscat, D., Adhikari, B., Adhikari, R., & Chaudhary, D. S. (2012). Comparative study of  
603 film forming behaviour of low and high amylose starches using glycerol and xylitol as  
604 plasticizers. *Journal of Food Engineering*, 109(2), 189–201.  
605 <https://doi.org/10.1016/j.jfoodeng.2011.10.019>

606 Nakamura, C., Koshikawa, Y., & Seguchi, M. (2008). Increased volume of Kasutera cake  
607 (Japanese sponge cake) by dry heating of wheat flour. *Food Science and Technology*  
608 *Research*, 14(5), 431.

609 Nara, S., & Komiya, T. (1983). Studies on the Relationship Between Water-saturated State and  
610 Crystallinity by the Diffraction Method for Moistened Potato Starch. *Starch - Stärke*,  
611 35(12), 407–410. <https://doi.org/10.1002/star.19830351202>

612 Oh, I. K., Bae, I. Y., & Lee, H. G. (2018). Effect of dry heat treatment on physical property  
613 and in vitro starch digestibility of high amylose rice starch. *International Journal of*  
614 *Biological Macromolecules*, 108, 568–575.  
615 <https://doi.org/10.1016/j.ijbiomac.2017.11.180>

616 Ozawa, M., Kato, Y., & Seguchi, M. (2009). Investigation of Dry-Heated Hard and Soft  
617 Wheat Flour. *Starch-Stärke*, 61(7), 398–406.

618 Punia, S., Sandhu, K. S., Dhull, S. B., & Kaur, M. (2019). Dynamic, shear and pasting  
619 behaviour of native and octenyl succinic anhydride (OSA) modified wheat starch and  
620 their utilization in preparation of edible films. *International Journal of Biological*  
621 *Macromolecules*, 133, 110–116. <https://doi.org/10.1016/j.ijbiomac.2019.04.089>

622 Seguchi, M. (2001). Oil binding ability of chlorinated and heated wheat starch granules and

623 their use in breadmaking and pancake baking. *Starch-Stärke*, 53(9), 408–413.

624 Shevkani, K., Singh, N., Bajaj, R., & Kaur, A. (2017). Wheat starch production, structure,  
625 functionality and applications-a review. *International Journal of Food Science &*  
626 *Technology*, 52(1), 38–58. <https://doi.org/10.1111/ijfs.13266>

627 Smith, R. J. (1967). Production and use of hypochlorite oxidized starches. *Starch Chemistry*  
628 *and Technology*, Vol. 2, pp. 620–625. New York: Academic Press.

629 Song, Y., & Jane, J.-L. (2000). Characterization of barley starches of waxy, normal, and high  
630 amylose varieties. *Carbohydrate Polymers*, 41(4), 365–377.  
631 [https://doi.org/10.1016/S0144-8617\(99\)00098-3](https://doi.org/10.1016/S0144-8617(99)00098-3)

632 Tan, C., Toh, W. Y., Wong, G., & Lin, L. (2018). Extrusion-based 3D food printing –  
633 Materials and machines. *International Journal of Bioprinting*, 4(2).  
634 <https://doi.org/10.18063/ijb.v4i2.143>

635 Ubeyitogullari, A., & Ciftci, O. N. (2016). Formation of nanoporous aerogels from wheat  
636 starch. *Carbohydrate Polymers*, 147, 125–132.

637 Xiong, J., Li, Q., Shi, Z., & Ye, J. (2017). Interactions between wheat starch and cellulose  
638 derivatives in short-term retrogradation: Rheology and FTIR study. *Food Research*  
639 *International*, 100, 858–863.

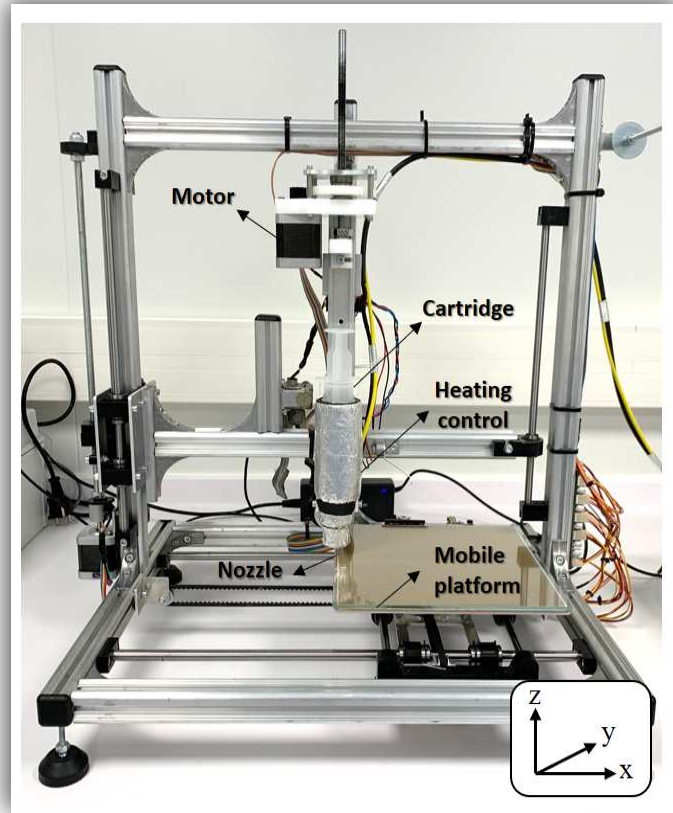
640 Yang, F., Zhang, M., Prakash, S., & Liu, Y. (2018). Physical properties of 3D printed baking  
641 dough as affected by different compositions. *Innovative Food Science & Emerging*  
642 *Technologies*, 49, 202–210. <https://doi.org/10.1016/j.ifset.2018.01.001>

643 Yeh, A.-I., & Yeh, S.-L. (1992). Some Characteristics of Hydroxypropylated and Crosslinked  
644 Rice Starches. *Cereal Chemistry*, 70(5), 596–601.

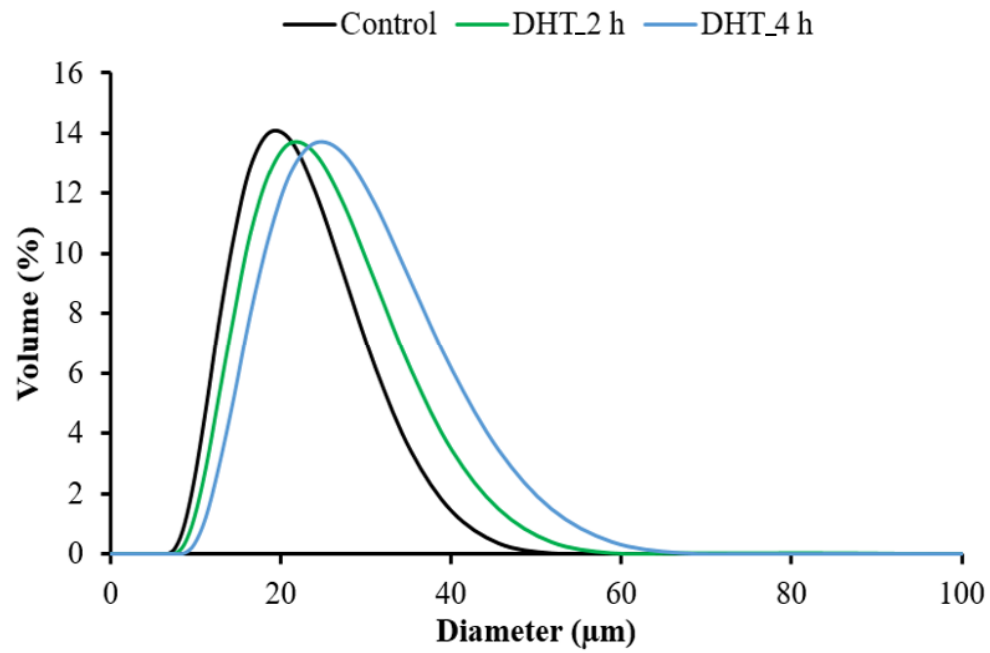
645 Yu, B., Ren, F., Zhao, H., Cui, B., & Liu, P. (2020). Effects of native starch and modified  
646 starches on the textural, rheological and microstructural characteristics of soybean  
647 protein gel. *International Journal of Biological Macromolecules*, 142, 237–243.

648 Zou, J., Xu, M., Tian, J., & Li, B. (2019). Impact of continuous and repeated dry heating  
649 treatments on the physicochemical and structural properties of waxy corn starch.  
650 *International Journal of Biological Macromolecules*, 135, 379–385.  
651 <https://doi.org/10.1016/j.ijbiomac.2019.05.147>

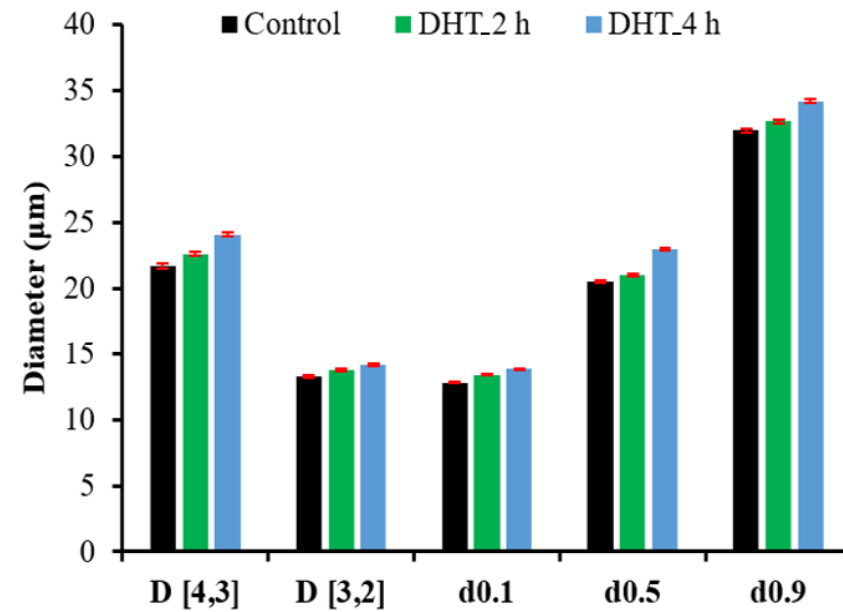
652



**Fig. 1.** The picture of 3D printer with the main parts: the extruder motor, syringe (cartridge) with the nozzle (movement in z direction), heating control, and mobile platform (movement in x and y directions).

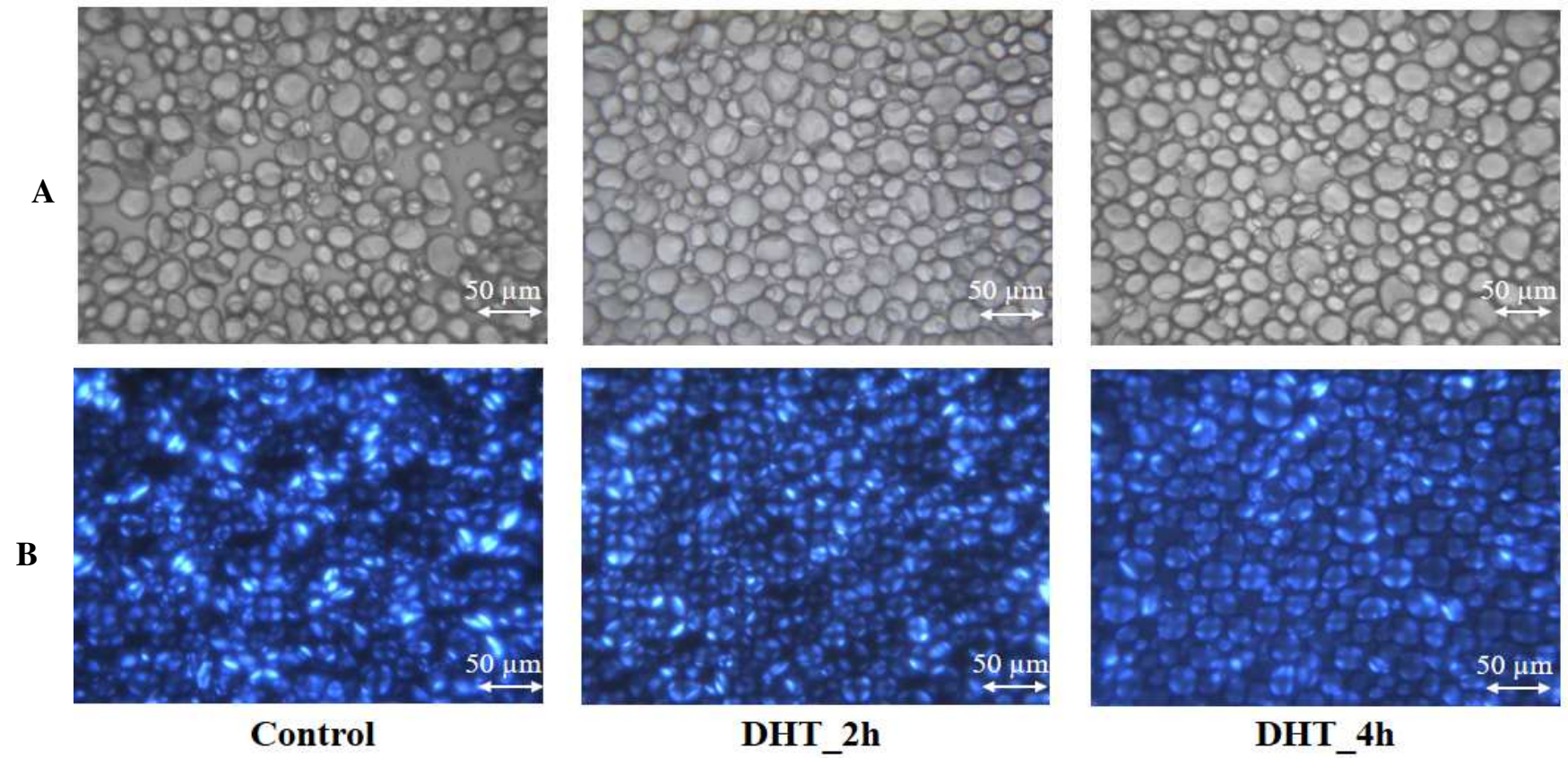


(A)



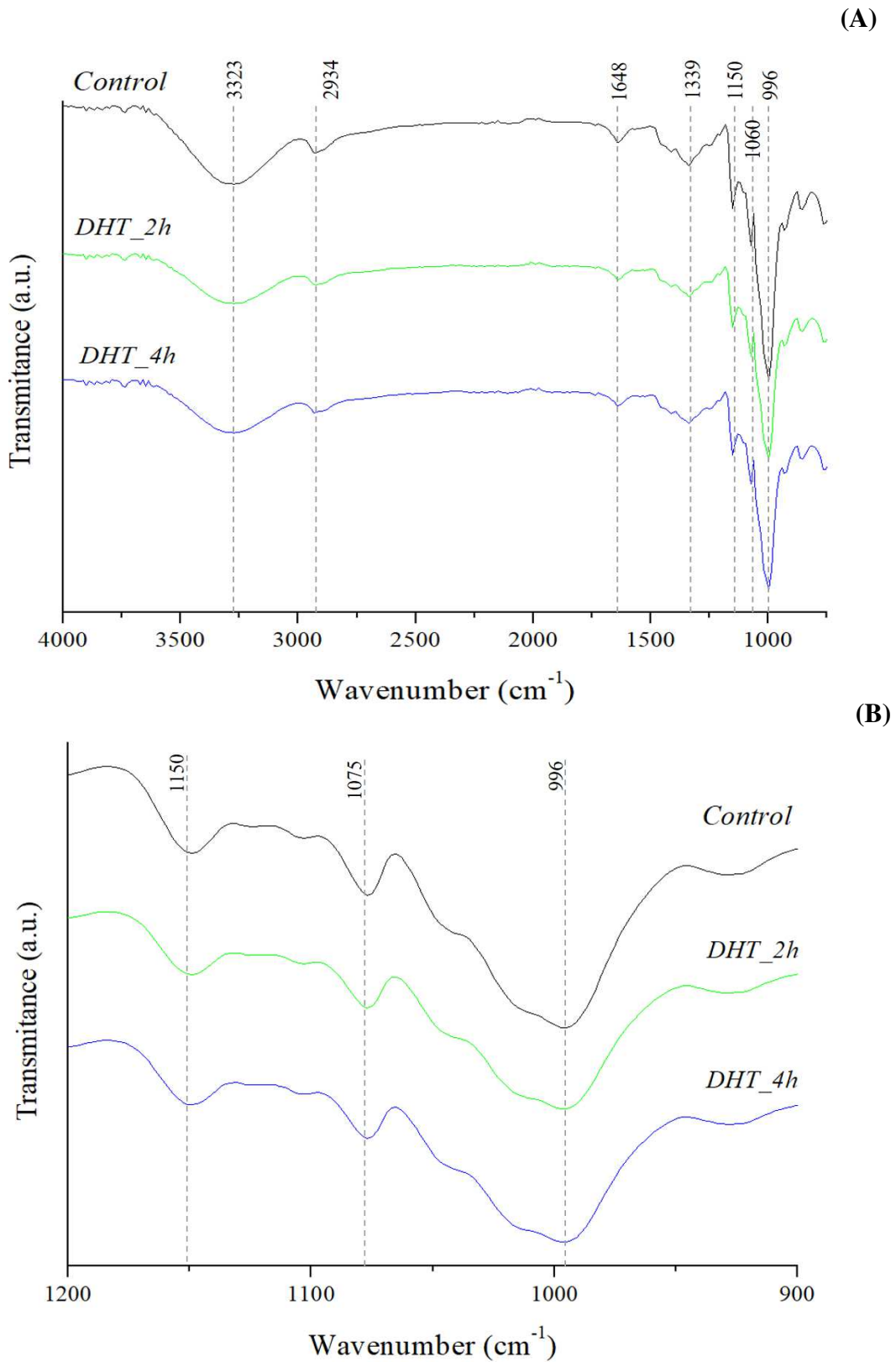
(B)

**Fig. 2.** (A) Granule size distribution (B) Specific diameters (D[4,3], D[3,2], d0.1, d0.5, d0.9 - vertical red bars are the standard deviation) of the control and modified starches by dry heating (DHT\_2h and DHT\_4h):

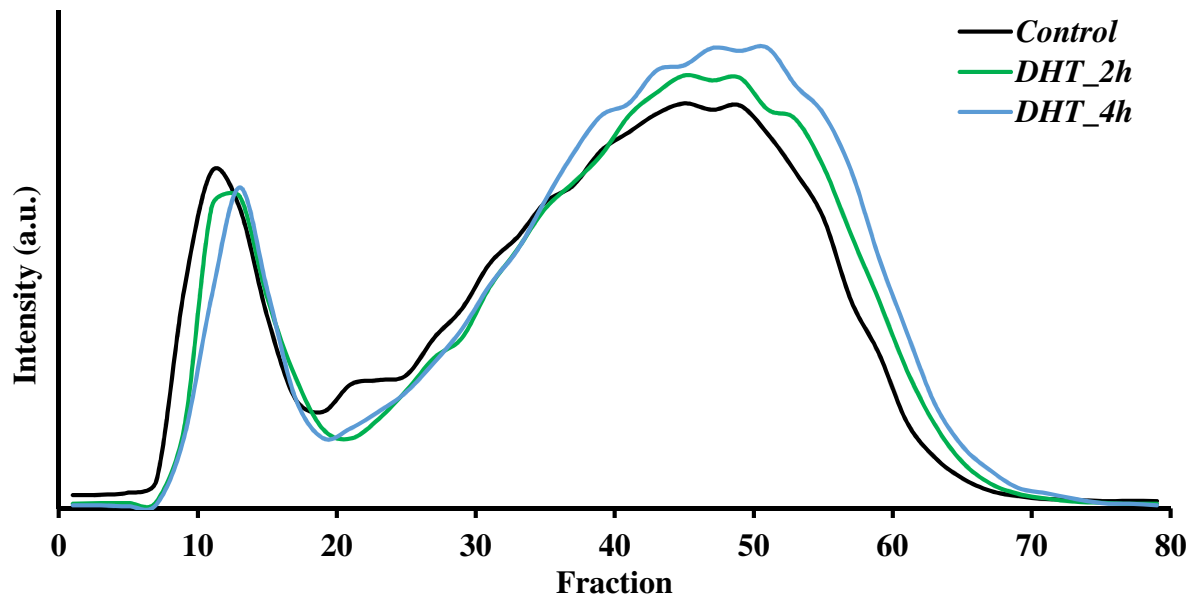


**Fig. 3.** Photographs from light microscopy (40×) using (A) nonpolarized and (B) polarized light of the **control and modified starches** by dry heating (DHT\_2h and DHT\_4h).

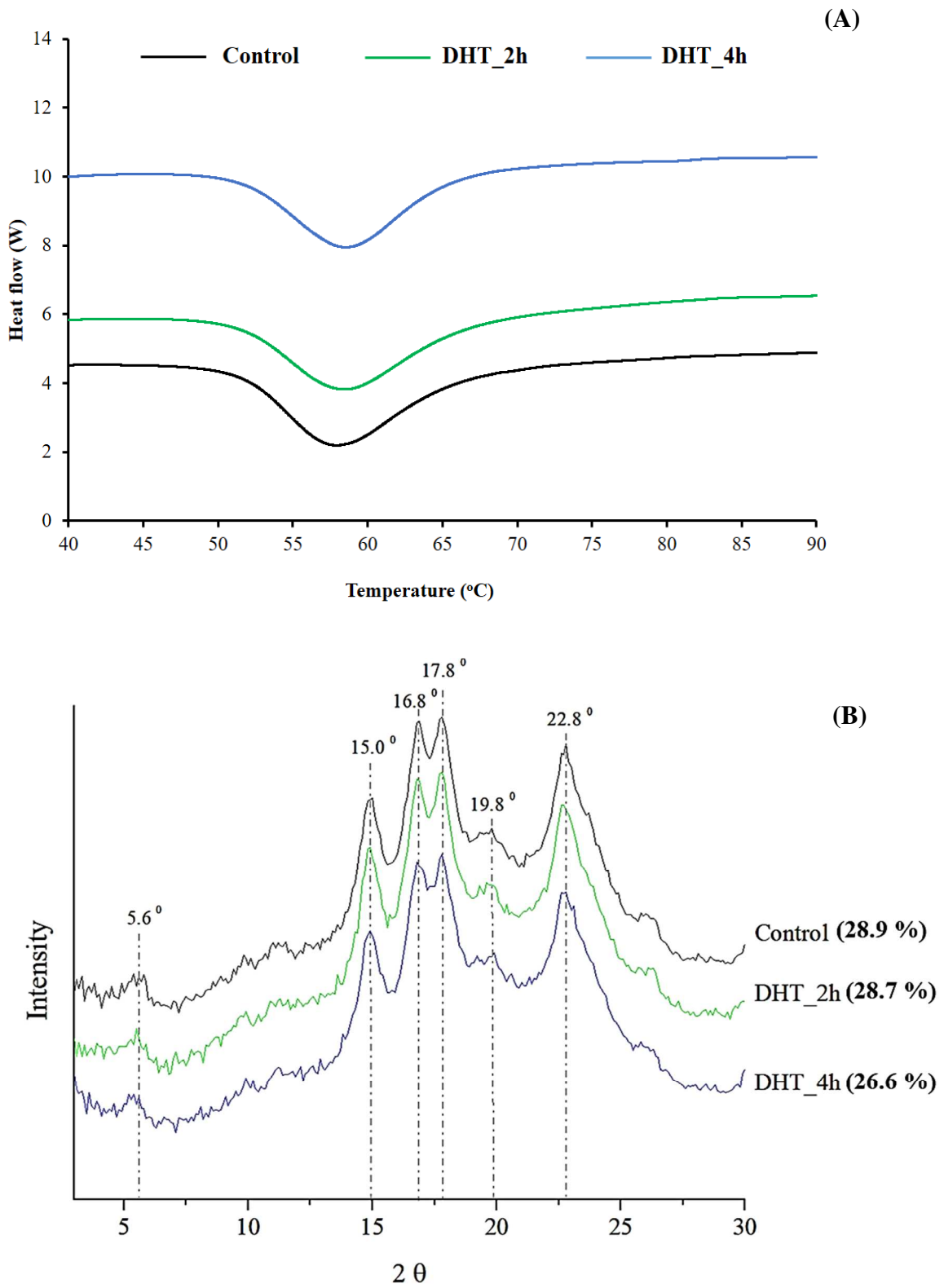




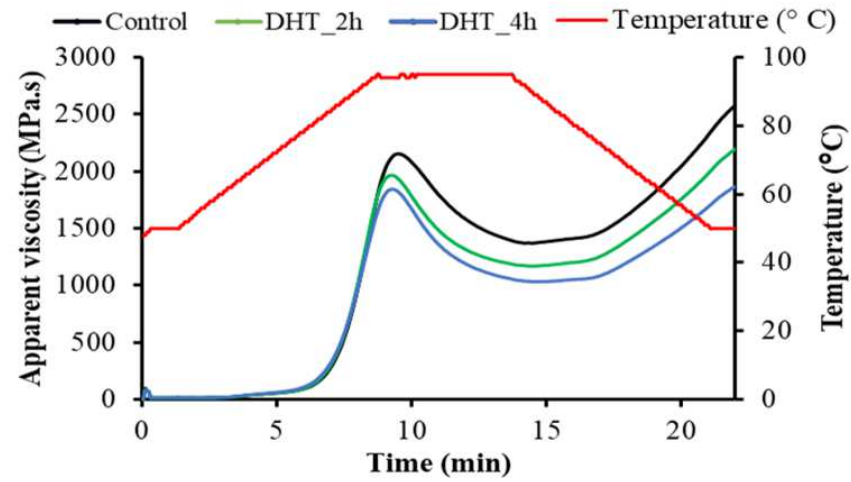
**Fig. 4.** (A) Vibrational spectra in wavenumber interval between (A) 4000 until 900  $\text{cm}^{-1}$ ) and between (B) 1200 until 900  $\text{cm}^{-1}$  of the control and modified starches by dry heating treatment (DHT\_2h and DHT\_4h).



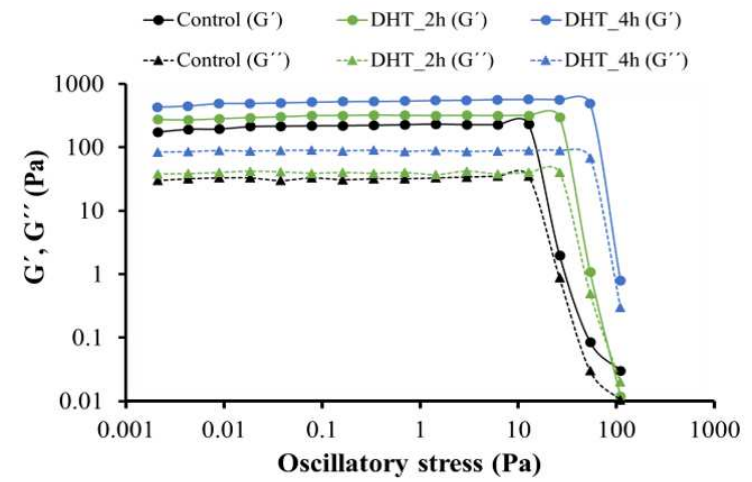
**Fig. 5.** Molecular size distribution of the control and modified starches by dry heating (DHT\_2h and DHT\_4h).



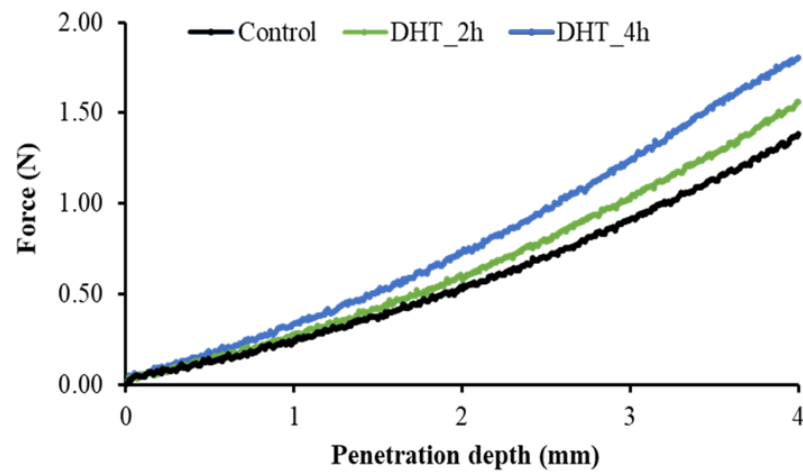
**Fig. 6. (A) DSC thermograms and (B) XRD of control and wheat starch treated by dry heating (DHT\_2 and DHT\_4 h).**



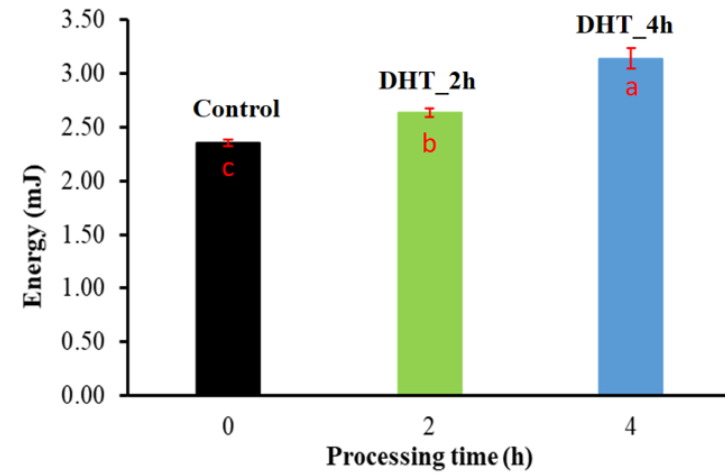
(A)



(B)

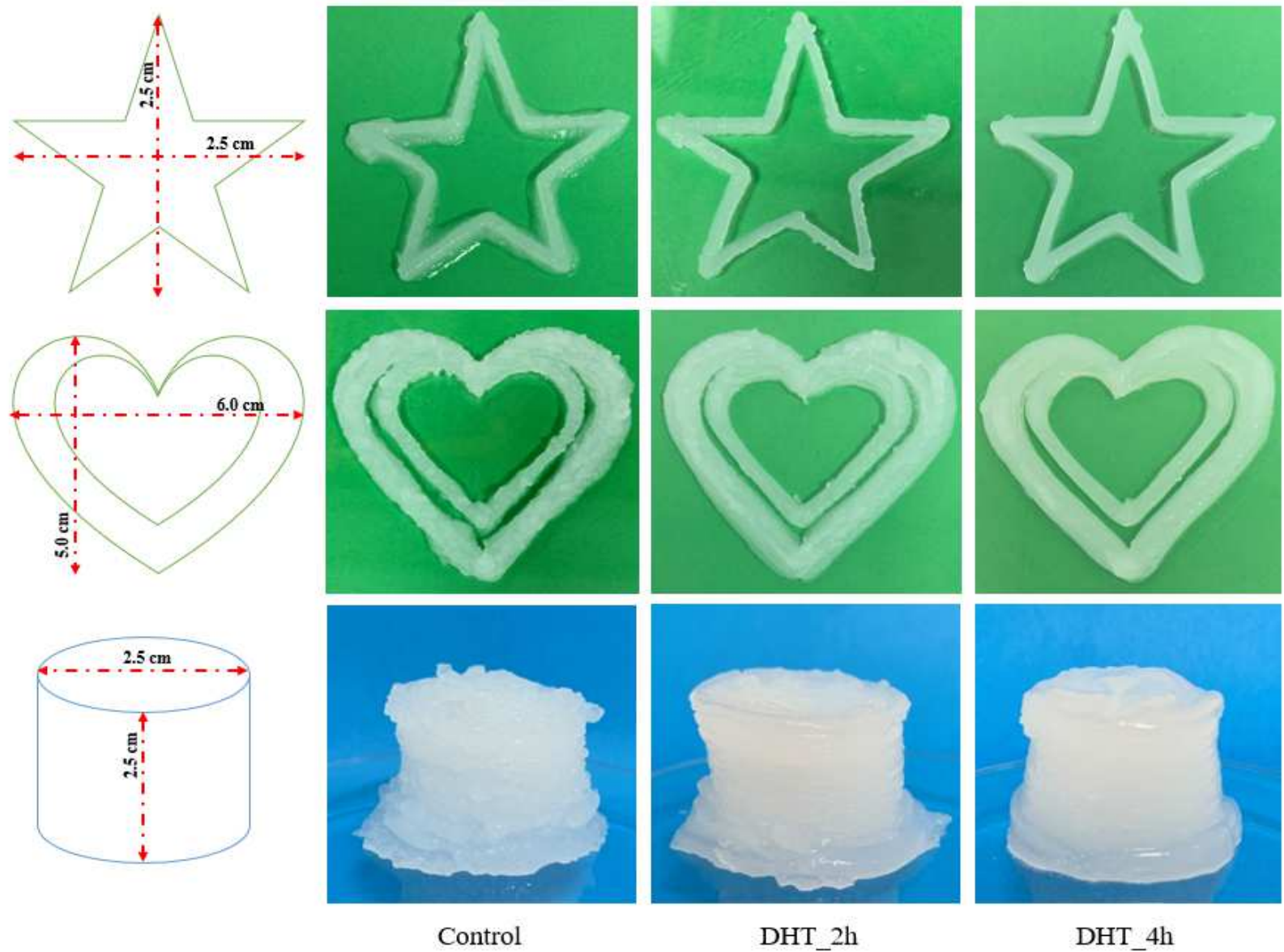


(C)



(D)

**Fig. 7.** (A) RVA curves, (B) oscillatory stress sweep analysis ( $G'$ : storage modulus and  $G''$ : loss modulus), (C) Gel firmness, and (D) the energy required in a puncture assay of the control and the wheat starch treated by dry heating (DHT\_2 and DHT\_4 h).



**Fig. 8.** The 3D printed hydrogels (star, heart, and cylindrical shapes) based on control wheat starch and modified wheat starch by dry heating treatment (DHT\_2h and DHT\_4h).

**Table 1.** Gelatinization properties of **the control and modified starches by dry heating** (DHT\_2h and DHT\_4h) (average  $\pm$  standard deviation).

<i>Samples</i>	<b>To (°C)</b>	<b>Tp (°C)</b>	<b>Tf (°C)</b>	<b><math>\Delta H</math> (J/g)</b>
<b>Control</b>	47.99 $\pm$ 0.28 <sup>a</sup>	58.05 $\pm$ 0.10 <sup>c</sup>	74.32 $\pm$ 0.56 <sup>a</sup>	9.85 $\pm$ 0.12 <sup>a</sup>
<b>DHT_2 h</b>	47.43 $\pm$ 0.37 <sup>a</sup>	58.30 $\pm$ 0.06 <sup>b</sup>	74.65 $\pm$ 0.62 <sup>a</sup>	9.89 $\pm$ 0.28 <sup>a</sup>
<b>DHT_4 h</b>	47.23 $\pm$ 0.43 <sup>a</sup>	59.85 $\pm$ 0.10 <sup>a</sup>	74.42 $\pm$ 0.35 <sup>a</sup>	7.57 $\pm$ 0.10 <sup>b</sup>

a, b, c: different letters in the same column indicates significant difference among samples, as revealed by Tukey's test,  $p < 0.05$

To = onset temperature, Tp = peak temperature, Tf = final temperature and  $\Delta H$  = enthalpy.

DHT\_2h: wheat starch dry heating treated for 2 h. DHT\_4h: wheat starch dry heating treated for 4 h.

**Table 2.** Pasting parameters from RVA and parameters from oscillatory stress sweep analysis of **the control and modified starches by dry heating (DHT\_2h and DHT\_4h)** (average  $\pm$  standard deviation)

<i>Samples</i>	<b>PAV (mPa.s)</b>	<b>TAV (mPa.s)</b>	<b>RBD (%)</b>	<b>FAV (mPa.s)</b>	<b>RSB (%)</b>	<b>PT (°C)</b>	<b>G' (Pa)</b>	<b>G'' (Pa)</b>	<b><math>\sigma_{0,G'}</math> (Pa)</b>
<b>Control</b>	2250.40 $\pm$ 40.92 <sup>a</sup>	1378.43 $\pm$ 18.30 <sup>a</sup>	37.36 $\pm$ 1.09 <sup>c</sup>	2757.50 $\pm$ 14.95 <sup>a</sup>	40.71 $\pm$ 0.72 <sup>a</sup>	73.53 $\pm$ 0.32 <sup>b</sup>	175 $\pm$ 25 <sup>c</sup>	33.4 $\pm$ 2.2 <sup>c</sup>	12.74 $\pm$ 0.05 <sup>c</sup>
<b>DHT_2 h</b>	2017.00 $\pm$ 30.15 <sup>b</sup>	1172.80 $\pm$ 22.30 <sup>b</sup>	40.45 $\pm$ 0.82 <sup>b</sup>	2257.00 $\pm$ 70.21 <sup>b</sup>	31.91 $\pm$ 0.49 <sup>b</sup>	73.68 $\pm$ 0.38 <sup>b</sup>	257 $\pm$ 32 <sup>b</sup>	40.1 $\pm$ 1.6 <sup>b</sup>	26.37 $\pm$ 0.20 <sup>b</sup>
<b>DHT_4 h</b>	1820.72 $\pm$ 22.17 <sup>c</sup>	1034.02 $\pm$ 10.12 <sup>c</sup>	43.93 $\pm$ 0.65 <sup>a</sup>	1927.50 $\pm$ 82.73 <sup>c</sup>	18.73 $\pm$ 0.42 <sup>c</sup>	74.53 $\pm$ 0.25 <sup>a</sup>	430 $\pm$ 14 <sup>a</sup>	88.9 $\pm$ 4.5 <sup>a</sup>	54.56 $\pm$ 0.08 <sup>a</sup>

Peak Apparent Viscosity (PAV), Through Apparent Viscosity (TAV), Relative Breakdown (RBD), Final Apparent Viscosity (FAV), **Relative Setback (RSB)** and Pasting Temperature (PT), Storage modulus (G'), Loss modulus (G''), **Yield stress ( $\sigma_{0,G'}$ )**

a - c: different letters in the same column indicates significant difference among samples, as revealed by Tukey's test,  $p < 0.05$ .

**DHT\_2h:** wheat starch dry heating treated for 2 h. **DHT\_4h:** wheat starch dry heating treated for 4 h.

**Table 3.** Texture parameters and reproducibility of the starch hydrogels printed in cylinder shape. The hydrogels are based on **control and modified starches by dry heating (DHT\_2h and DHT\_4h)** (average  $\pm$  standard deviation)

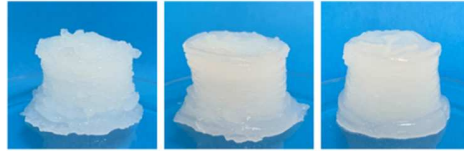
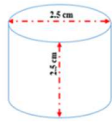
<i>Starch hydrogels</i>	<i>Texture</i>					<i>Reproducibility</i>		
	<i>Hardness (N)</i>	<i>Adhesiveness (N.s)</i>	<i>Cohesiveness (-)</i>	<i>Springiness (-)</i>	<i>Chewiness (-)</i>	<i>Weight (g)</i>	<i>Diameter (mm)</i>	<i>Height (mm)</i>
<b>Control</b>	0.68 $\pm$ 0.08 <sup>c</sup>	-1.67 $\pm$ 0.01 <sup>a</sup>	0.41 $\pm$ 0.01 <sup>a</sup>	0.81 $\pm$ 0.03 <sup>a</sup>	0.27 $\pm$ 0.01 <sup>a</sup>	15.38 $\pm$ 0.36 <sup>a</sup>	2.50 $\pm$ 0.16 <sup>a</sup>	2.08 $\pm$ 0.08 <sup>b</sup>
<b>DHT_2h</b>	0.82 $\pm$ 0.03 <sup>b</sup>	-2.16 $\pm$ 0.15 <sup>b</sup>	0.41 $\pm$ 0.01 <sup>a</sup>	0.84 $\pm$ 0.04 <sup>a</sup>	0.27 $\pm$ 0.01 <sup>a</sup>	14.74 $\pm$ 0.34 <sup>b</sup>	2.58 $\pm$ 0.16 <sup>a</sup>	2.34 $\pm$ 0.11 <sup>a</sup>
<b>DHT_4h</b>	1.03 $\pm$ 0.05 <sup>a</sup>	-2.23 $\pm$ 0.23 <sup>b</sup>	0.41 $\pm$ 0.02 <sup>a</sup>	0.82 $\pm$ 0.04 <sup>a</sup>	0.32 $\pm$ 0.03 <sup>a</sup>	14.81 $\pm$ 0.18 <sup>b</sup>	2.48 $\pm$ 0.08 <sup>a</sup>	2.36 $\pm$ 0.11 <sup>a</sup>

a - c: different letters in the same column indicates significant difference among samples, as revealed by Tukey's test,  $p < 0.05$

**DHT\_2h: wheat starch dry heating treated for 2 h. DHT\_4h: wheat starch dry heating treated for 4 h.**



*DHT improved the 3D printability and extended the texture possibilities of wheat starch hydrogels*



*Control*

*DHT\_2h*

*DHT\_4h*