

Childhood asthma heterogeneity at the era of precision medicine: Modulating the immune response or the microbiota for the management of asthma attack

Stéphanie Lejeune, Antoine Deschildre, Olivier Le Rouzic, Ilka Engelmann,

Rodrigue Dessein, Muriel Pichavant, Philippe Gosset

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5	Lejeune Stéphanie ^{1,2} , Deschildre Antoine ^{1,2} , Le Rouzic Olivier ^{2,3} , Engelmann Ilka ⁴ ,				
6	Dessein Rodrigue ^{2,5} , Pichavant Muriel ² , Gosset Philippe ²				
7	¹ CHU Lille, Univ. Lille, Pediatric Pulmonology and Allergy Department. Hôpital Jeanne de				
8	Flandre, F-59000 Lille, France.				
9	² Univ. Lille, INSERM Unit 1019, CNRS UMR 9017 CHU Lille, Institut Pasteur de Lille,				
10	Center for infection and immunity of Lille, F-59019 Lille cedex, France.				
11	³ CHU Lille, Univ. Lille, Department of Respiratory Diseases, F- 59000 Lille cedex, France.				
12	⁴ Univ. Lille, Virology laboratory, EA3610, Institute of Microbiology, CHU Lille, F-59037 Lille				
13	cedex, France.				
14	⁵ Univ. Lille, Bacteriology departement, Institute of Microbiology, CHU Lille, F-59037 Lille				
15	cedex, France.				
16					
17	Corresponding author:				
18	Philippe Gosset, PhD				
19	OpInfIELD, CIIL, Institut Pasteur de Lille				
20	1, rue du Pr A. Calmette				
21	59019- Lille Cedex –FRANCE				
22	Tel : +33 320 87 79 65				
23	E-mail: philippe.gosset@pasteur-lille.fr				
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- 31 ORCID Numbers:
- 32 S. Lejeune: https://orcid.org/0000-0001-7009-2113
- 33 A Deschildre: https://orcid.org/0000-0003-2006-7221
- 34 O. Le Rouzic: https://orcid.org/0000-0003-4439-2950
- 35 M. Pichavant: https://orcid-org/0000-0002-1315-4397
- 36 P. Gosset: https://orcid.org/0000-0002-4043-6429

38 Abstract (249 words)

Exacerbations are a main characteristic of asthma. In childhood, the risk is increasing with severity. Exacerbations are a strong phenotypic marker, particularly of severe and therapy-resistant asthma. These early-life events may influence the evolution and be involved in lung function decline.

43 In children, asthma attacks are facilitated by exposure to allergens and pollutants, but are mainly triggered by microbial agents. Multiple studies have assessed immune responses 44 to viruses, and to a lesser extend bacteria, during asthma exacerbation. Research has 45 46 identified impairment of innate immune responses in children, related to altered pathogen recognition, interferon release, or anti-viral response. Influence of this host-microbiota dialog 47 on the adaptive immune response may be crucial, leading to the development of biased T 48 49 helper (Th)2 inflammation. These dynamic interactions may impact the presentations of 50 asthma attacks, and have long-term consequences.

51 The aim of this review is to synthesize studies exploring immune mechanisms impairment against viruses and bacteria promoting asthma attacks in children. The potential influence of 52 the nature of infectious agents and/or preexisting microbiota on the development of 53 54 exacerbation is also addressed. We then discuss our understanding of how these diverse host-microbiota interactions in children may account for the heterogeneity of endotypes and 55 clinical presentations. Finally, improving the knowledge of the pathophysiological processes 56 induced by infections has led to offer new opportunities for the development of preventive or 57 58 curative therapeutics for acute asthma. A better definition of asthma endotypes associated 59 with precision medicine might lead to substantial progress in the management of severe childhood asthma. 60

61 Introduction

62 Childhood asthma is a major public health issue in industrialized countries. 63 Development and progression of the disease is marked by exacerbations, also called 64 asthma attacks, which are one of the most prevalent causes of hospitalization in children 65 with consequently significant costs worldwide [1–3]. They constitute key events in the natural 66 history of childhood asthma and are major markers of its heterogeneity [4]. Mechanisms 67 leading to asthma attacks are not fully understood and may differ between children, 68 according to their age range or phenotype.

Pediatric literature on asthma usually distinguishes preschool recurrent wheeze / also 69 called preschool asthma in children aged 5 years and younger; from school-age children 70 71 asthma, in those aged 6 to 11 years; and adolescent asthma, in those aged 12-17 years 72 [3,5,6]. During preschool years, these events can be isolated, with few or no interval 73 symptoms, and their persistence despite conventional maintenance treatment, such as 74 inhaled corticosteroids (ICS), is a marker of severity [6,7]. Although viral infections are the 75 main triggers of acute asthma, early and multiple allergic sensitization is a strong risk factor 76 of presenting severe and frequent exacerbations [7,8]. In school-age children and 77 adolescents, acute asthma attacks are usually less frequent, and not only triggered by lung 78 infections, but also by exposure to allergens and pollutants [3,9,10]. Exacerbations rates 79 increase with the severity of the disease, which is defined according to treatment pressure to 80 obtain asthma control [4,5]. Repetition of frequent and severe episodes under high doses of ICS associated with another maintenance treatment (mainly long acting beta2 agonists) is a 81 criteria of severe asthma in these age-groups [11,12], as demonstrated in pediatric asthma 82 cohorts [13,14]. Additionally, it is now recognized that patients prone to exacerbations 83 constitute a distinct susceptibility phenotype, possibly associated with different endotypes, 84 including specific airway inflammation patterns, or inflammatory mechanisms in response to 85 infectious agents [15]. In some individuals, the severity of the disease and propensity to 86 develop acute asthma may also vary throughout the course of childhood [16]. These early-87 88 life events can influence the long-term evolution at adult-age, in terms of asthma

89 persistence, phenotype and alterations of lung function, that may lead to early chronic obstructive pulmonary disease (COPD) [17]. Early interventions could modify the natural 90 history of the disease. However, in children with severe asthma, conventional maintenance 91 therapies may be insufficient to prevent asthma attacks [7,18], and repeated or prolonged 92 93 oral corticosteroids are associated with many side effects, such as defective growth [19]. New treatment strategies are now needed. Interestingly, some studies have shown the 94 efficacy of biologics in pediatric asthma, mainly on severe exacerbations, and suggested 95 96 disease-modifying properties of some of them, making it a particularly attractive strategy in children [20]. 97

Viruses are closely linked to wheezing illnesses during early childhood [21]. 98 Respiratory syncytial virus (RSV), and rhinoviruses (RV), both RNA viruses, are among the 99 100 most common causes of lower respiratory tract infections, *i.e.* bronchiolitis and/or pneumonia, with a predominance of RSV in the first 12 months of life and then RV in the 101 following years [22]. Birth cohort studies and other epidemiological studies have shown a 102 strong association between early RV-induced wheeze, and to a lesser extent, RSV-induced 103 104 wheeze, and subsequent development of childhood asthma [23-26]. Moreover, pediatric 105 asthma attacks are mainly triggered by respiratory viruses, particularly RV, both in the 106 preschool and school-age [10,27-31]. The question of whether asthma-associated underlying inflammation promotes the pathogenic effect of viral infection or whether the virus 107 108 induces exaggerated inflammation is still under debate and has been the subject of many research studies over the past 25 years. 109

Alongside viruses, there is also a growing interest on the impact of bacteria in the onset of asthma attacks and the perpetuation of inflammation. Neonatal airway colonization with bacteria including *Haemophilus influenzae, Moraxella catarrhalis* and/or *Streptococcus pneumoniae,* has been associated with persistence of asthma [32]. Indeed, Kloepfer, *et al.* have shown frequent viral and bacterial co-infection in asthmatic children aged 2-18 years [33]. It is well known that viral infection can predispose to bacterial secondary infection. A few studies have begun to describe inflammatory phenotypes linked with altered microbiota,

known as dysbiosis, within the airways [34,35]. Interactions between viruses and bacteria, personal risk factors (*e.g.* genetic background and atopy), and environmental exposures may promote more severe acute asthma episodes that will influence the progression of asthma [21]. However, the precise role of each of these factors and their interplay with the host immune defenses remains to be elucidated.

The aim of this review was to synthesize studies exploring human innate immune mechanisms responses against viruses and bacteria during asthma attacks and to provide hypotheses to decipher how they may contribute to the phenotypes observed in childhood asthma. We will then discuss how therapeutic strategies targeting these pathways may improve the management of acute asthma in children.

128 1/ Impact of viruses on the onset of asthma attacks: chicken or egg?

Up to now, mechanisms explaining susceptibility to develop acute asthma upon respiratory viruses, in particular RV and to a lesser extend RSV, in asthmatic children prone to exacerbation are not fully understood. As innate immune cells play a central role in the onset of anti-viral defenses, they have been the focus of many studies in pediatric and adult asthma, conducted both *ex-vivo* and *in-vivo*, contributing to the identification of new asthma endotypes. The original studies presented in this review have been summarized in **Table 1**.

135

136 **1.1** Virus recognition during asthma attack is mainly driven by airway 137 epithelial cells

Before addressing the viral infections impact on asthma attacks, we will briefly 138 describe the virus-induced anti-viral responses. Airway epithelial cells (AEC) are the main 139 cellular targets of pathogens, and act as sentinel cells in response to infection (Fig 1) [36-140 38]. These structural cells are a central part of innate immune response [39]. They first act 141 as a complex physicochemical barrier, via the presence of tight intercellular junctions and 142 the muco-ciliary escalator, clearing foreign particles out. If not cleared, Pathogen Associated 143 144 Molecular Patterns (PAMP) on the surface of infectious agents, will then be recognized through Pattern Recognition Receptors (PRR), in particular Toll-Like Receptors (TLR) and 145 intracellular RNA helicases, which will trigger innate immune responses. Although AEC play 146 an early and central role in orchestrating innate immune defenses, innate immune cells 147 148 including alveolar macrophages (AM) and dendritic cells (DC), especially plasmacytoid DC 149 (pDC), are also involved in innate immune responses against pathogens [40-44].

150 Studies aiming to explore PRR recognition by epithelial cells and innate immune cells 151 from asthmatic children are multiple but heterogeneous. In cultures of primary AEC from 152 children with severe therapy resistant asthma (STRA), lower expression levels of 153 extracellular and intracellular PRR than healthy controls were observed at baseline [45]. 154 Expression of PRR was also decreased upon infection with RV on these cultures and in 155 those from adult asthmatics, suggesting a persistent defect in adulthood [45,46]. Hence, in

156 AM from adults with severe asthma, Rupani, et al. also showed a reduction of TLR7 expression after RV infection compared with healthy subjects, with an effect potentially 157 mediated by microRNAs [47]. Conversely, in ex vivo cultures of cells from broncho-alveolar 158 lavages (BAL) cells and peripheral blood mononuclear cells (PBMC) from adults with mild-to-159 160 moderate asthma, no impairment of PRR expression (TLR, RNA-helicases) was observed [48]. This discrepancy was also illustrated by our in vivo real-life study conducted in 72 161 school-age children with allergic asthma hospitalized for severe exacerbation, associated 162 with viral infection in 64% (RV: 51%) [49]. Whereas viral infection did not modulate PRR 163 expression and function on PBMC as compared with non-infected asthmatic patients, a 164 reduced expression of PRR was reported in a sub-group of children prone to reinfection 165 166 outside an exacerbation (steady state 8 weeks later), and displaying neutrophilic airway inflammation away from exacerbation [10,49]. Although present both at exacerbation and 167 168 steady state, this impairment of PRR expression as well as the inflammatory profile was not associated with the clinical evolution and risk of exacerbations in the year following 169 170 exacerbation [50].

Altogether, these data suggest that impairment of PRR recognition in asthmatic children could be limited to a subpopulation of patients, potentially severe, and may persist into adulthood. Rather than an altered baseline expression or synthesis of PRR, a downregulation of the expression and activation of the co-receptors, and/or enzymes involved in signaling may be determinant. Longitudinal *in vivo* studies could provide more insights into these mechanisms and allow studying their dynamics during the course of the disease.

177

178 **1.2 Modulation of anti-viral responses**

Activation of PRR on AEC and innate immune cells will onset various signaling pathways, such as interferon regulatory factors (IRF) 3 and 7, and nuclear factor-kappa B pathway (NF-kB). This will lead to the production of anti-viral molecules such as interferons (IFN) and anti-microbial peptides, including interferon-stimulated genes (ISG) as well as cathelicidins and defensins [51]. Anti-microbial peptides have immunomodulatory activities

and can alter host cellular responses. LL-37, also called human cationic antimicrobial 184 peptide is the only human peptide of the cathelicidin family and one of the most-studied 185 [52,53]. In *in vitro* models, LL-37 has the ability to reduce virus replication, notably for RSV 186 [53,54]. In a population of 77 asthmatic children, Arikoglu, et al. have shown that children 187 188 prone to develop acute asthma attacks displayed higher blood levels of cathelicidins and lower levels of vitamin D at baseline than children with controlled asthma, suggesting a role 189 for anti-microbial peptides in acute asthma [55]. However, the evolution of these baseline 190 191 levels during acute asthma attacks and their impact on the anti-viral and inflammatory 192 response remain unknown.

193 The majority of studies conducted in asthmatic subjects have had the aim to explore 194 anti-viral IFN response upon viral infection. They have observed a defect of production of type I IFN. In response to RV, most ex-vivo studies have demonstrated an impairment of 195 196 IFN- α and IFN- β production in AEC from pediatric asthmatic patients compared to levels observed in cells from healthy controls [45,56–58]. This impairment was also demonstrated 197 198 in adult cells [59-61]. Interestingly, similar results have been observed in ex-vivo cultures of 199 PBMC. As early as 2002, Bufe, et al. have shown in an ex-vivo model a defective production 200 of IFN- α in PBMC from pediatric asthmatic patients compared to healthy controls, upon 201 Newcastle disease virus (NDV) infection, an avian respiratory virus belonging to the Paramyxoviridae family and well-known potent inductor of IFN-α [62]. They later extended 202 these observations in whole blood cells cultures from adult asthmatics, upon infection with 203 either NDV of RSV [63]. In the later years, similar results have been shown in pediatric 204 studies [64,65] but also in adult studies [45,48,66]. So far, only a few studies have been 205 conducted upon in vivo inoculation and natural infection. In a cohort of preschool children, 206 IFN-α systemic levels were reduced at the time of RV-induced exacerbation in asthmatics, 207 as compared with healthy controls during an asymptomatic RV infection [30]. In our recently 208 published cohort of preschool asthmatic children with natural virus-induced exacerbation 209 (RV: 73%), the most severe children, who were prone-to-exacerbation, displayed lower 210 211 plasma concentrations of IFN-B at exacerbation than the others, suggesting a defect of 212 production in response to viruses limited to this sub-population [8]. In another study conducted in older children, we did not observe altered IFN responses in sputum, plasma 213 and cultures of stimulated-PBMC of patients presenting a virus-induced exacerbation, 214 compared with those with a non-virus-induced exacerbation [49]. In adult asthmatics, a 215 216 significant type I IFN defect was demonstrated in bronchial epithelium and sub-epithelium obtained from bronchial biopsies compared with healthy controls, after experimental infection 217 with RV-A16 [67]. In contrast, Schwantes, et al. described elevated rather than low RNA 218 levels of type I IFN in sputum from adult asthmatic patients, compared to those observed in 219 healthy subjects, early in the course of a virus-induced exacerbation.[68]. 220

221

222 Following their discovery in 2003, there has been increasing knowledge regarding the 223 implication of type III IFN implication in anti-viral responses [69]. Ex-vivo studies conducted 224 in AEC from asthmatic subjects have suggested a parallel impairment of IFN-λ1 and IFN- $\lambda 2/3$ production upon RV infection in pediatric patients [45,56,57], and this impairment 225 226 seemed to persist in adult patients [46,59]. In vivo, this defective response was not observed 227 at the time of asthma exacerbation in sputum and/or PBMC from atopic asthmatic children 228 [30,49,70], nor in adults [68,71], suggesting that there are compensatory mechanisms or 229 other sources of IFN, supplementing the defect observed in resident cells.

A few studies have explored the potential pathways leading to a defect of IFN production in asthmatic patients. Downregulation of IRF3 or IRF7, following virus infection in asthmatic cells, has been suggested [57,59]. Gielen, *et al.* have also demonstrated a role of the molecule suppressor of cytokine signaling 1 (SOCS1), exercising a negative-feedback on IFN production following viral infection in children with STRA [57].

235 Collectively, these studies suggest a potential impairment of anti-viral IFN response, 236 in particular type I IFN, during asthma attack. Rather than explained by immaturity in the first 237 years of life, this defective response could also be limited to a sub-phenotype of patients 238 who are prone-to-exacerbation, from early childhood to adulthood. The effect of age, the

kinetics of the IFN response following viral infection and the impact of viral load in theairways remain to be addressed.

241

1.3 Cytokines and epithelial-derived alarmins (IL-25, IL-33 and TSLP) response against

243 viral infection

Airway epithelial cells display immunoregulatory properties by producing pro-244 inflammatory cytokines, including alarmins interleukin (IL)-25, IL-33 and thymic stromal 245 lymphopeietin (TSLP), released upon viral infection, and implicated in the recruitment of 246 macrophages, DC, T cells and granulocytes, such as eosinophils and neutrophils. This will 247 promote the adaptive immune response through DC maturation and migration to draining 248 lymph nodes associated with the airways. Dendritic cells will thus induce naïve T cell 249 250 differentiation into antigen-specific T cells. This will drive the adaptive immune response within the airways to clear the virus out. The alarmins also contribute to a biased T helper 251 (Th)2 immune response by activating type 2 innate lymphoid cells (ILC2) and polarizing DC 252 in order to promote T cells differentiation (Fig 1). This specific cytokine environment in 253 254 asthmatic patients favor naïve T cells polarization into Th2 cells, producing IL-4, IL-5, IL-13, as opposed to Th1 cells, producing IFN-y and TNF-B [37,72]. These cytokines are 255 responsible for the type 2 inflammation and induce the pathophysiological features of 256 asthma, including eosinophil mobilization, mucus hypersecretion, smooth muscle 257 hyperplasia and airflow obstruction [3,73]. . Under the influence of IL-5, eosinophils will 258 enter the airways and perpetuate type 2 inflammation [49,74]. They will also contribute to 259 resolution of immune responses, including tissue repair [75]. Other cytokines, such as type 260 17 cytokines: IL-17, IL-21, IL-22, can also promote mucus hypersecretion and production of 261 cytokines and chemokines by AEC. They will favor the recruitment of neutrophils, through 262 the induction of C-X-C chemokines, conducting to a state of chronic inflammation [67]. In the 263 longer term, IL-17 may contribute to the development of airway remodeling during asthma by 264 enhancing the production of profibrotic cytokines, proangiogenic factors, proteases and 265 266 collagen [76,77].

267 In children, the influence of respiratory viruses and innate immune mechanisms on the adaptive immune response development may be crucial. First, some respiratory viruses 268 may induce the release of alarmins, and thus favor the development of Th2 inflammation. A 269 recently published study showed that children with RV-positive-bronchiolitis displayed 270 271 increased IL-4/IFN-y ratio in nasopharyngeal aspirates [78]. Upon RSV infection, Lee, et al. 272 observed a higher production of TSLP by AEC from asthmatic children compared to healthy 273 controls [79]. Hence, some respiratory viruses may induce Th2 inflammation in the airways of children and this may be increased in an asthmatic environment. In a cohort of preschool 274 asthmatic children, ILC markers, such as soluble ST2, were induced in vivo following RV 275 276 infection in preschool asthmatic children, and were correlated with nasal levels of IL-33 [80]. In our study of school-age children, at exacerbation, airway inflammation in infected patients 277 was characterized by significantly higher IL-5 concentration and eosinophil count than in 278 non-infected patients [49]. This may not be specific to childhood, as Jackson, et al. have also 279 280 demonstrated an induction of IL-4, IL-5 and IL-13 in BAL following RV inoculation in asthmatic subjects, associated with exacerbation severity [81]. Additional in vitro 281 experiments showed that IL-33 directly induced Th2 cytokine production by T cells and ILC2. 282 However, this RV-inducedTh2 inflammation could be limited to an asthmatic environment in 283 284 adults, as suggested by an *in vitro* study reproducing the induction of IL-4 and IL-13 by asthmatic PBMC after co-exposition with RV and IL-33, but showing no effect on PBMC from 285 healthy donors [82]. In addition, exposure to respiratory viruses has also been associated 286 with an increase in Th17 cytokines, as demonstrated in co-cultures of RSV-infected AEC 287 288 with T cells [83].

Interferon production is also closely linked to Th2 and inflammatory cytokines. In asthmatic preschool children prone to exacerbation, we observed simultaneously lower production of Th1 cytokines (IFN- γ), Th2 cytokines (IL-5), pro-inflammatory cytokines (TNF- α), regulatory cytokines (IL-10), and type 1 IFN (IFN- β) during virus-induced exacerbation in a subgroup of preschool children with characteristics of severe asthma and a history of

294 frequent exacerbations [8]. This association seems to persist in adults, as suggested by Parsons, et al. reporting a parallel decrease of type III IFNs and of IL-6, (C-X-C motif) ligand 295 (CXCL)8 and CXCL10 in adult asthmatic AEC inoculated with RV [46]. In a murine model of 296 allergic asthma, IRF3 controlled the parallel evolution of IFN and Th2 cytokines during acute 297 298 episodes induced by allergen exposure [84]. Some adult studies have suggested bidirectional influences during viral infection. Upon RV infection, pre-treatment with Th2 299 cytokines in cultures of AEC may impair TLR3 and IRF3 signaling pathway [59], or type 1 300 and 3 IFN expression [57]. Lower production of type I IFN, displayed by some children, could 301 also reduce the apoptosis of neighbored-infected cells and increase viral loads, 302 303 consequently amplifying airway inflammation, inflammatory cytokine production and thus, 304 asthma attack severity [58,60].

Altogether, these studies confirm close interactions between innate and adaptive immune mechanisms against viruses in an asthmatic environment. Modulating airway inflammation, particularly Th2 inflammation, could therefore directly improve anti-viral responses during asthma attacks in some children.

309

310 **1.4 Viral behavior during asthma attack**

Although the impairment of anti-viral responses plays a key role in asthma attack, 311 characteristics of the microorganisms are also important. First, pathogenicity of RV strongly 312 differs according to subtypes, probably because of the implication of different cellular targets. 313 Indeed, infection with RV-C strains, which use cadherin-related family member 3 (CDHR3) 314 as a receptor, may be associated with higher clinical severity [10,31,85], whereas B 315 serotypes seem to be responsible for milder symptoms. Interestingly, single nucleotide 316 polymorphism in the CDHR3 gene, which influences the efficiency of RV-C to infect its target 317 cells, has been associated with childhood asthma susceptibility [86]. Moreover, different viral 318 strains could have different replication rates and thus induce higher viral loads [87]. 319 Rhinovirus C strains could also modulate the airways inflammation, and induce higher levels 320 321 of Th2 and/or Th17 cytokines, as shown in preschool children with RV-C-induced wheeze 322 [88]. Second, systemic viremia has been observed in children infected with respiratory 323 viruses, such as RV-C, and young age was found to be a risk factor associated with viremia 324 [89]. This state could induce an enhanced inflammation in children by interaction of viruses 325 with systemic immune cells. Finally, viral-coinfection has been frequently observed at 326 exacerbation in asthmatic children, particularly in the preschool years [30,31]. The combined 327 effects of several viruses within the airways may modulate both the innate and adaptive 328 responses.

These data demonstrate that, in susceptible children, the nature of respiratory viruses, their spreading, as well as their interactions with the host cells may directly impact the inflammation and the severity of asthma exacerbations.

332

333 **1.5 Emerging role of the airways virome**

334 The role of viral carriage and/or asymptomatic infection at steady state, *i.e.* outside an exacerbation, in asthmatic patients has also been hypothesized. Hence, pediatric studies 335 have shown a frequent virus shedding in children, with RV positive samples observed in 19-336 29% of unselected asthmatic children and in 15-23% of healthy children [10,28,90,91]. In the 337 338 previously-cited cohort of preschool children prone to exacerbation, we observed even higher rates, with 67% of positive PCR (51% for RV) at steady state [8]. In 78 school-age 339 children, we also observed that 10% of children had positive RV sampling at exacerbation 340 and steady state, and that RV serotypes were systematically different at both time points, 341 pointing toward reinfection rather than persistence [10]. Consistent with these observations, 342 Bergauer, et al. also found 54% of RV at steady state in preschool children with asthma, vs 343 45% in healthy controls [30]. Interestingly, RV carriage in both asthmatics and healthy 344 controls was found to be associated with higher levels of type III IFN at baseline than in 345 subjects without. This increase in baseline inflammation seems to persist in adulthood. 346 Hence, in sputum cells from 57 asthmatic adults, da Silva, et al. have shown higher baseline 347 levels of IFN and anti-viral molecules compared with healthy subjects [92]. In all, one could 348 349 hypothesize that RV persistence in asthmatic patients could induce a pro-inflammatory state and/or a desensitization of PRR, which could reduce the response against pathogens withinthe airways upon new infection.

Alongside RV, other respiratory viruses, could also take part in these processes. 352 Metagenomic analyses have made it possible to identify hundreds of viral species within the 353 354 airways, constituting the respiratory tract virome [93]. In non-asthmatic children, Wang, et al. have shown multiple common epidemic respiratory viruses in children with severe acute 355 respiratory infection, whereas, the virome was less diverse and mainly dominated by the 356 Anelloviridae family in healthy children [94]. Anelloviruses are major components of the 357 virome and display immunomodulatory properties on both innate and adaptive immunity [95]. 358 Although their presence has been associated with an impaired lung function in asthmatic 359 children [96], to our knowledge, no study has evaluated their association with asthma 360 361 attacks. Thus, the contribution of these viruses and their interactions with both pathogenic 362 infectious agents, as well as commensal bacteria from the pulmonary microbiota on chronic inflammation and the onset of acute asthma remain to be determined. 363

365 **2/ The emerging role of pathogenic bacteria and microbiota in asthma**

Increasing evidence indicates that the respiratory microbiota, including bacterial and viral microorganisms has an important role in respiratory health and disease [97,98]. In addition, recent data underline the role of bacterial infections in the development and progression of asthma, as summarized in **table 2**.

During the first weeks of life, the respiratory tract begins to develop a niche-specific 370 community pattern, where Staphylococcus aureus and Corynebacterium replace the 371 originally colonizing bacteria and become the dominant microorganisms [98]. The most 372 373 dramatic changes occur during the first 2 months. In the following six months, the respiratory 374 microbiota continues to mature; relative abundance of S. aureus declines, and an increase in Moraxella, Streptococcus, Haemophilus, Dolosigranulum, Alloiococcus, and Prevotella sp. is 375 observed. This period is crucial and alterations of the microbiota have been implicated in the 376 377 development and the manifestations of chronic lung diseases. .

378

379 **2.1 Bacteria and asthma development**

380 Using bacterial culture techniques, it has been demonstrated that neonates born to 381 asthmatic mothers with upper airway colonization with S. pneumoniae, M. catarrhalis, and/or H. influenzae were at higher risk of presenting asthma at the age of 5 years [32]. This was 382 associated with an aberrant immune response, with increased IL-5 and IL-13 synthesis in 383 PBMC obtained at the age of 6 months [99]. Interestingly, evaluation of the gut microbiota in 384 infants included in the Canadian (CHILD) study showed that infants at risk of asthma 385 exhibited transient alterations of the microbiota, known as dysbiosis, in neonates [100]. More 386 specifically, the relative abundance of the commensal bacterial genera Lachnospira, 387 Veillonella, Faecalibacterium, and Rothia was significantly decreased in children at risk of 388 asthma. Furthermore, antibiotic-induced gut dysbiosis has been shown to facilitate the 389 development of allergic asthma in experimental murine models and clinical studies [101]. In 390 a population-based cohort, it has been reported that pre- and post-natal antibiotic exposure 391 392 was associated with a higher asthma risk [102]. No significant difference in bacterial diversity 393 was observed between samples from those with wheeze and healthy controls. Age and attendance at day care or kindergarten were important factors in driving bacterial diversity 394 whereas wheeze and viral infection were not found to be significantly associated to the 395 bacterial communities. In the Avon Longitudinal Study of Parents and Children (ALSPAC) 396 397 cohort, children for whom antibiotics course was reported before the age of 2 years were more likely to have asthma at age 7, although microbiota was not characterized [103]. 398 Similarly, in British Columbia (Canada), "controlled" antibiotic use during infancy was 399 400 associated with a reduction in the incidence of pediatric asthma, and was related with the 401 preservation of gut microbial communities [104]. Altogether, these data confirm the need to 402 preserve the microbiota in early infancy in order to limit the development of asthma.

403

404 **2.2 Relationship between bacteria and asthma attack**

405 Bacterial infections potentially contribute to the severity of symptomatic respiratory viral infections and asthma attacks in children. Indeed, there are bidirectional interactions 406 407 between viruses and airway bacteria that appear to modulate the severity of illness and the 408 likelihood of asthma attack [20]. Pediatric studies have shown that upper respiratory tract 409 colonization with Streptococcus, Haemophilus, and/or Staphylococcus genus during RSV and RV infection increases the risk of hospitalization independently of age and of the 410 presence of asthma [105,106]. In our previously-cited cohort of preschool children, asthma 411 attacks were frequently associated with viral infection (94%, mostly RV). Pathogenic bacteria 412 were isolated in induced sputum in 56%, mainly H. influenza, M. catharralis 413 or *S*. pneumoniae [21]. In adult asthmatics, asymptomatic RV infection was associated with 414 increased abundance of *Dolosigranulum* and *Corynebacterium*, whereas symptomatic RV 415 infection seemed to be related with a higher frequency of Moraxella [107]. In another adult 416 study, co-infection with virus and bacteria induced higher levels of cytokine/chemokine than 417 bacterial infection [108]. Interestingly, circulating CCL2 and CCL5 concentrations were 418 increased in infected asthmatic patients compared with non-asthmatic patients. 419 420 Streptococcus species may also influence the course of asthma exacerbation, as shown by

421 Kama, *et al.* in preschool children [109]. Indeed, patients with *S. pneumoniae* colonization in 422 the pharyngeal microbiota had significantly shorter wheezing episodes and reduced lung 423 inflammation (including lower levels of TNF- α).

In summary, these data suggest an ability for bacteria to promote asthma attacks and
modulate the inflammation during childhood. Targeting bacterial infections during asthma
attacks could thus be an alternative strategy in some children.

427

428 **2.3 Dysbiosis and impact on asthma natural history**

Early-life dysbiosis of the microbiota, in particular lung microbiota, may have long-429 430 term consequences on asthma natural history. During viral infection in children less than two 431 years old, detection of *Moraxella* was found to be associated with current wheeze at 5 years old and presence of Streptococcus, Moraxella, or Haemophilus was associated with a high 432 risk of asthma development [110]. Mansbach, et al. showed in a joint modeling analysis 433 adjusting for 16 covariates, including viral trigger, that a higher relative abundance of 434 Moraxella or Streptococcus species 3 weeks after day 1 of hospitalization for severe 435 436 bronchiolitis was associated with an increased risk of recurrent wheezing [111]. In contrast, 437 no significant difference in microbiota composition was observed between oro-pharyngeal microbiota from children with acute wheezing (0-14 years old) and healthy controls [112]. In 438 this study, the large range of patient ages and day care attendance were important 439 confounding factors, since the microbiota evolves throughout childhood. 440

In school-age asthmatic children, Zhou, et al. assessed by longitudinal 441 measurements the relationship between nasal airway microbiota and either loss of asthma 442 control, risk 443 or of severe exacerbations [113]. Whereas the commensal Corynebacterium / Dolosigranulum cluster characterized the patients with the lowest risk of 444 disease progression, a switch towards the Moraxella cluster was associated with the highest 445 risk of severe asthma exacerbations. Similarly, in another cohort of school-age asthmatic 446 children, nasal microbiota dominated by Moraxella species was associated with increased 447 448 exacerbation risk, and eosinophil activation at exacerbation [35]. Staphylococcus or

449 Corynebacterium species-dominated microbiota were linked with reduced respiratory illness and exacerbation event rates, whereas Streptococcus species-dominated microbiota 450 increased the risk of RV infection. Interestingly, using unbiased airway microbiome-driven 451 clustering, Abdel-Aziz, et al. recently described in adult asthmatics two distinct robust severe 452 453 asthma phenotypes that were associated with airway neutrophilia, and not related to the presence of *Moraxella* species [114]. Robinson, et al. assessed the lower airway microbiota 454 by performing BAL at steady state in severe preschool wheezers [34]. They reported two 455 profiles: a Moraxella species "dysbiotic" cluster, associated with airway neutrophilia and a 456 "mixed microbiota" cluster associated with macrophage- and lymphocyte-predominant 457 airway inflammation, thus suggesting that bacteria might influence the characteristics of lung 458 inflammation. 459

Altogether, these data show that airway dysbiosis in early life might contribute to recurrent wheeze and asthma, and to the severity of the disease. Among bacterial species, the early presence of *M. Catharralis* and *H. influenza*e during childhood seems to be a marker of evolution toward a severe disease, through the modulation of the inflammatory reaction. In contract, colonization with S. pneumoniae might have some anti-inflammatory effects. However, the role of bacteria is often exacerbated or modulated by intrinsic (atopy, inflammation characteristics) or extrinsic factors, such as viral infections and antibiotics.

467

469 **3**/ **Consequences on asthma phenotypes and endotypes**

It is now clear that phenotyping and even more so, endotyping asthmatic patients is an essential step to understand the diversity of immune responses at the time of an asthma attack. We will review the commonly described endotypes of childhood asthma, successively based on airway leukocyte infiltrate, profiles of allergic sensitization and nature of asthma trajectories.

Among airway inflammatory phenotypes, neutrophilic asthma appears to be a distinct 475 endotype, associated with asthma severity in adults, and possibly involved in corticosteroid 476 insensitivity [76]. In children, several studies have shown that association of airway 477 neutrophilia with severity is inconstant [115–118], and may not persist throughout the course 478 479 of the disease [116,119]. However, neutrophils present in the airways of severe asthmatic 480 children may have enhanced survival and proinflammatory functions that could increase baseline inflammation [120]. As previously stated, airway neutrophilia has also been 481 associated with a different IFN response, at baseline and at the time of asthma attack, both 482 483 in adults [66,92] and pediatric studies [49]. This could have an impact on lung infections. 484 Thus, we have shown that a subgroup of school-aged children with positive viral PCR both 485 at baseline and exacerbation displayed airway neutrophilia [49]. In severe adult asthmatics, 486 airway neutrophilia has also been associated with lesser microbiota diversity in BAL [114,121]. Finally, experimental RV infection in adult asthmatic subjects directly induces 487 488 bronchial mucosal neutrophilia [122]. Therefore, the neutrophilic endotype may be characterized by an alteration of airway microbiota and association with viral infections. 489

Airway eosinophilia is highly frequent in asthmatic children, observed in severe and non-severe asthma, often but not always associated with response to ICS, thus highlighting the diversity of eosinophilic profiles in pediatric asthma [116,118,123]. Mouse models and adult studies have suggested a role of IL-5 induced airway eosinophilia as a positive regulator of RV receptor intercellular adhesion molecule (ICAM)-1 on AEC, and a negative regulator of PRR and antiviral responses [124,125]. However, in children, it remains unclear whether airway eosinophils are involved in the pathophysiology of severe asthma, and in the 497 propensity to develop acute asthma attacks [126]. In addition, peripheral blood eosinophilia, currently used as the basis for selection of eosinophil-targeted biologics, may not be 498 representative of airway eosinophilia in children [127]. Most importantly, it has been reported 499 that eosinophilia is not necessarily associated with higher levels of Th2 cytokines [118], and 500 501 other cells such as alarmin-stimulated ILC2 may have a greater biological importance in the pathophysiology of asthma attacks [128]. Eosinophils can also have beneficial effects in 502 childhood asthma, as they display regulatory functions and contribute to lung repair and 503 mucosal homeostasis [75]. Peripheral blood eosinophilia has recently been identified has a 504 505 biomarker predicting the decrease of severity throughout the adolescent years [16]. Finally, it remains to be determined whether eosinophil-targeted biologics, such as anti-IL-5 506 507 mepolizumab, are efficient in reducing asthma attacks in children [129]. Overall, as opposed 508 to adults, airway eosinophilia does not appear as the preferential biomarker to classify 509 pediatric patients nor to assess their risk of acute asthma attacks.

Rather than through the recruitment of eosinophils, type 2 inflammation may play a 510 key role in some children's susceptibility to develop acute asthma attacks through IgE 511 512 sensitization. Hence, atopy, defined as allergic sensitization, is a key feature associated with 513 risk of asthma attacks in childhood [7,8,14]. As shown by latent-class analyses, atopy is also one of the main factors that may account for the heterogeneity of the disease, especially in 514 the preschool years [130,131] and could drive response to treatment [3]. Pre-existing atopy 515 516 may play a role in the modulation of IFN response upon viral infection in children, although studies have shown conflicting results [59,61,65]. Durrani, et al. have suggested a direct 517 implication of the high-affinity IgE receptor (FccRI) in this defect, as its cross-linking on pDC 518 from asthmatic allergic patients is associated with reduced IFN secretion upon RV infection 519 [65]. Following virus-induced exacerbation in atopic children, upregulation of FcERI receptor 520 on monocytes and DC was observed, associated with a concomitant upregulation of pro-521 inflammatory cytokines, including IL-4 and IL-13 [132], suggesting that respiratory viral 522 infection in atopic children may initiate an atopy-dependent cascade that amplifies virus-523 524 induced airway inflammation. This could initiate long-term remodeling and inflammation.

Hence, IgE sensitization, especially if occurring early in life and multiple, has been identified
as the main factor associated with long-term asthma risk (recurrence of exacerbations,
persistence of asthma, impaired lung function) in neonatal cohorts, including MAAS (Great
Britain), MAS (Germany), PARIS (France) and PASTURE (Europe) [133–136].

529 Describing asthma trajectories in neonatal longitudinal cohorts may precisely allow identifying novel endotypes. For instance, in a birth cohort of children at high risk of atopy, 530 cord blood cells were tested for their ability to produce IFN after exposure to polyinosinic-531 polycytidylic acid (Poly I:C), a double stranded RNA that mimics viral PAMP [137]. A group 532 of low producers of type I and III IFN response upon stimulation representing 24% of the 533 patients was identified. They displayed higher risk of lower respiratory tract infection and 534 persistent wheeze at the age of 5, suggesting that these clinical outcomes are partly 535 536 determined by constitutional mechanisms.

Altogether, beyond airway leukocyte and type 2 inflammation, childhood asthma endotypes seem to be associated with the characteristics of airway microbiota and of the associated immune response. Because these might involve dynamic processes, the study of asthma trajectories throughout childhood is crucial. A better characterization of asthma endotypes will be an essential step in order to achieve a personalized medicine.

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543 4/ In the future: machine learning and systemic biological approaches

Digital tools provide an innovative way to study the integrated immune response in asthmatic patients. For instance, Custovic, *et al.* used machine learning in an *in vitro* model of RV-A16 infection on PBMC extracted from 11 years old children and described 6 clusters of cytokine response to infection [138]. One cluster, with patients from whom PBMC displayed low IFN responses, high pro-inflammatory cytokines, low Th2 responses and moderate regulatory responses, was associated with early-onset asthma and the highest risk of asthma exacerbations and hospitalizations for lower respiratory tract infection.

551 Omic technologies, known as genomic, epigenomic, transcriptomic, proteomic, 552 single-cell omics, metabolomics, microbiomics, exposomics, are others fast-developing

553 tools, generating a massive amount of data reflecting the biological state of cell populations at a given time, that will certainly make possible in the future to identify new endotypes and 554 their biomarkers [139,140]. For instance, with the development of the Human Genome 555 Project, genome-wide association studies have allowed to link childhood asthma with 556 557 association signals in or near genes involved in innate immune response pathways, such as the TLR1 locus, and the IL-33 locus [141,142]. Transcriptomics analyses, which aim at 558 investigating all RNA transcripts in a biological sample, will certainly enable the identification 559 of a large quantity of genes modulated in certain conditions. A recent study has described 94 560 distinct gene-expression modules between virus-induced 561 and non-virus-induced exacerbation in nasal and sputum cells from a large cohort of school-age children [143]. In 562 563 asthma attacks, this tool may also provide information on the activated pathways linked with 564 inflammatory and anti-microbial response [144,145]. RNA sequencing in nasal lavage cells from asthmatic children at exacerbation led to identify some genes overexpressed as 565 relevant "nodes" linking genes of the inflammatory response, such as IRF7 [144]. 566 Transcriptomic analyses have also been used to identify genes associated with different 567 568 phenotypes of asthmatic patients [146]. Finally, metabolomics, the study of the metabolites 569 produced in living systems (both humans and microbiota) represents another promising 570 strategy in the upcoming years to address the role of immuno-metabolism as well as lung and gut microbiota in asthma pathophysiology [147,148]. 571

To sum up, systemic approaches integrating all biologic pathways are now increasingly used to study the mechanisms leading to susceptibility to acute asthma attacks in children. They may facilitate the identification of novel multimodal biomarkers linked with specific endotypes and therapeutic targets.

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577

579 5/ Therapeutic strategies modulating innate immune defenses during asthma attacks

580 Conventional maintenance treatment, such as ICS, may be insufficient to prevent 581 attacks in children, especially in severe asthma. Severe therapy-resistant asthma is 582 precisely characterized by repeated severe exacerbations despite high dose daily treatment. 583 As the knowledge about infection-underlying pathophysiological processes grows, new 584 therapeutic approaches that may contribute to either improve innate immune responses or 585 limit the occurrence and consequences of infections during asthma attacks have been 586 recently explored (**Figure 2**).

587

588 5.1 Immunostimulatory therapeutics

As we have shown earlier, asthmatic children may display an impairment of PRR 589 590 signaling that leads to susceptibility to infection-induced asthma attacks. One of the strategies to enhance their anti-viral responses could involve stimulation of PRR. This 591 strategy has only been tested in pre-clinical and adult studies. Following promising results 592 from pre-clinical studies [149], Silkoff, et al. reported results of a clinical trial assessing the 593 594 effect of CNTO3157, a TLR3 agonist, upon in vivo inoculation of RV-A16 in adult subjects 595 with mild-to-moderate asthma [150]. The molecule was ineffective in blocking the effect of RV-16 challenge on the onset of asthma exacerbation, without effect on asthma symptoms 596 nor lung function. Additionally, all post-inoculation exacerbations occurred in the CNTO3157 597 group, and two were severe, thus pointing toward a lack of efficacy and safety of the TLR3 598 targeting drugs. 599

Another way of enhancing PRR activation and activate different pathways could therefore be the administration of PAMP-like adjuvants or particles. Some teams have hypothesized that Poly I:C, a double stranded RNA that activates TLR3 and intracellular receptors, might modulate anti-viral responses. Preclinical studies have shown contrasting results, with promising effect in the context of Influenza A virus infection [52,151], but concerns regarding its safety. Indeed, Tian, *et al.* have described, in a mouse model, a prolonged IFN response upon *S. pneumonia* and *S. aureus* infection, inducing an impaired bacterial clearance [152]. We have also demonstrated in a murine model of allergic asthma that TLR3 ligands can enhance the Th2 response [153]. Thus, the exact consequences of administering exogenous TLR ligands (including TLR7-8) in humans are still unknown and their potential role in bacterial secondary infections in the lungs need to be addressed.

611 Another immunomodulatory strategy, particularly appealing in pediatric asthma, could rely on the administration of bacterial lysates [154]. Two main therapies have been 612 described: polyvalent mechanical bacterial lysates (PMBL®) and Broncho-Vaxom OM-85 613 BV. They contain lysates of 8 pathogenic bacteria including H. influenzae and S. 614 pneumoniae. Both PMBL® and OM-85 administration have proven to be efficient in 615 preventing respiratory tract infections in adult and pediatric patients with chronic bronchitis 616 617 and/or history of recurrent chronic infections [155–159]. Interestingly, in a trial conducted in elderly COPD patients, Ricci, et al. observed a reduced number of seroconversion against 618 619 numerous viral pathogens in patients treated with PMBL®, as well as a better control of the 620 number of all infectious episodes and COPD exacerbations, suggesting that the preventing effect of PMBL® on acute infections was not only on bacterial infections, but also on viral 621 622 infections [160]. In 152 school-aged children with allergic asthma, Emerick, et al. have 623 described a reduction of exacerbations after 12 weeks of treatment with PMBL®, and no serious adverse event, although no improvement of asthma control scores were achieved 624 [161]. In 75 preschool children with recurrent wheeze, Razi, et al. demonstrated that OM-85 625 626 BV reduced the rate and duration of wheezing episodes in the 12 months following the treatment initiation, and reduced the number of acute respiratory tract illnesses [162]. Finally, 627 in toddlers aged 6-18 months, a clinical trial is ongoing to assess the preventive effect of 628 OM-85V on the onset of first wheeze episode related to lower respiratory tract infection 629 during a 3 years observational period [163]. Regarding the mechanisms of action, prevention 630 of acute respiratory infections was not induced by purified LPS or bacterial DNA, suggesting 631 that other PAMP contained in bacterial lysates would activate innate immunity and lead to 632 promotion of type 1 inflammation by DC [164–166]. 633

Altogether, immunostimulatory therapeutics may represent a promising strategy as additional treatments to prevent asthma attacks but their safety and efficacy need to be properly addressed. Because multiple microorganisms may be involved in acute asthma, it remains also unclear whether they would have an impact on the onset and severity of acute asthma attacks in children.

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640 **5.2 Therapeutics enhancing anti-viral responses**

Anti-microbial peptides, among which LL-37, are key molecules of the innate immune 641 responses against both viruses and bacteria. Thus, it has been hypothesized that 642 supplementing with LL-37 or inducing its production among AEC could enhance anti-643 infectious defenses. Sousa, et al. have tested the effect of exogenous cathelicidin on RV-1B 644 replication after inoculation on cultures of AEC [167]. They reported that exogenous LL-37, 645 as well as homologous porcine cathelicidins, either administered prior or after infection, 646 displayed antiviral activity against RV by reducing the metabolic activity of infected cells. 647 648 However, one preclinical study in a mouse model of asthma showed in vivo a deleterious 649 effect of exogenous cathelicidin-related antimicrobial peptide in airway inflammation and 650 responsiveness [168], and to our knowledge, there is no published study *in vivo* in humans. Nevertheless, promising results regarding vitamin D supplementation on the release of LL-651 37 by AEC in *in vitro* experiments have been published [169–171]. In clinical studies, there is 652 evidence toward a protective effect of vitamin D supplementation on risk of acute respiratory 653 infections in children [172]. Although there are other effects of vitamin D that might explain 654 its clinical efficacy [80,173], there is a well-studied biological link between vitamin D and host 655 microbial peptides, via the translocation of vitamin D to the nucleus of AEC, where it 656 activates Vitamin v responsive elements in promoter regions of anti-microbial peptide genes 657 such as LL-37 [52,171,174]. Hence, targeting the vitamin D / cathelicidin axis could 658 represent a promising strategy during acute asthma in children. 659

660 As studies have shown deficient IFN responses in asthmatic patients, it has been 661 hypothesized that IFN supplementation upon viral infection could enhance anti-viral

662 responses and reduce asthma attacks severity. Interferon therapies (pegylated IFN-α in association with ribavirin) have first been developed in other diseases such as in hepatitis C 663 and human immunodeficiency virus [175,176]. Ribavirin is a synthetic guanosine nucleoside 664 that impairs the synthesis of viral mRNA [52]. Its clinical utility, upon respiratory virus 665 666 infection, has been shown in RSV, Adenovirus, RV, or in (SARS)-CoV immunocompromised patients [177–180]. Since exogenous IFNs reduced RV replication in *in vitro* cultures of RV-667 infected AEC from asthmatic patients [181], a trial was initiated in 147 adult asthmatic 668 patients developing symptoms of viral infection (68% RV) to evaluate the efficacy of inhaled 669 IFN-β treatment for 14 days [182]. Overall, it did not improve asthma symptoms, but it 670 enhanced morning peak expiratory flow recovery and reduced the need for additional 671 treatment, *i.e.* oral corticosteroids or antibiotics. In an exploratory analysis of the subset of 672 more difficult-to-treat patients, IFN supplementation improved asthma control questionnaires. 673 674 In addition, it boosted anti-viral innate immunity with persistent elevated serum levels of CXCL10, and decreased levels of pro-inflammatory molecules such as CCL4 and CXCL8. In 675 676 the INEXAS phase 2 trial, designed to assess efficacy of inhaled IFN-β1a for 14 days to 677 prevent asthma exacerbations in adults, McCrae, et al. also reported improvement in the 678 morning peak expiratory flow in the treated arm compared with placebo, but the study 679 stopped early [183,184]. No study has yet been conducted in pediatric patients.

680 Collectively, anti-viral therapies could represent potential therapies in the treatment of 681 acute asthma attacks in children. However, the exact timing of administration upon viral 682 infection and onset of acute asthma, and the population of children who might benefit from 683 these strategies remain to be determined.

684

685 **5.3 Role of anti-microbial therapeutics**

Apart from ribavirin, other anti-viral therapies, such as capside-targeting molecules pleconaril, BTA-798 and V-073 have been studied in enteroviruses infections. However, none of these compounds have proven sufficient efficacy and safety in clinical trials [185]. In particular, a phase 2 clinical trial with intranasal pleconaril in children aged 6 years and older, and adults, showed no impact of treatment on the onset of RV-induced cold symptoms
or acute asthma exacerbation [186]. Although their efficacy needs to be addressed in clinical
trials, DNAzyme therapies could also be promising strategies, as recently suggested by an *in vitro* study showing good catalytic activity of 2 candidates (Dua-01 and Dua-02) on various
RV strains [187].

695 As described above, bacteria and microbiota may be associated with asthma development and attacks, and antibacterial therapies have emerged as alternative strategies 696 697 to prevent and/or to treat asthma exacerbation [154,188,189]. Treatments aiming to 698 influence the composition of the microbiota are presently tested in different clinical situations 699 and recent data have suggested that these methods might have some interest in asthma. 700 Transplantation of healthy microbiota seems a promising alternative as demonstrated in gut 701 inflammatory diseases [190]. In preclinical study, Arrieta, et al. demonstrated that oral 702 inoculation with four bacterial species previously identified as associated with risk of childhood asthma to germ-free mice improved allergic airway inflammation in their adult 703 704 offspring [100]. Several studies have also evaluated the benefit of prebiotic and probiotic but 705 their interest during childhood asthma attack has yet to be evaluated [191,192]. Another 706 option may be to administer bacteria metabolites, which can mimic the effect of normal 707 microbiota. Thus, feeding pregnant mice with short chain fatty acid (SCFA), such as acetate, was associated with modulation of the maternal microbiota and protection from developing 708 allergic airway disease in the offspring [193]. Finally, an increasing number of studies in 709 experimental models of lung infections has demonstrated the potential interest of fecal 710 microbiota transplantation and/or SCFA supplementation, suggesting that they might be 711 efficient in bacteria- and virus-induced exacerbations [194-196]. 712

The potential modulatory effects of some antibiotics on asthma attacks in children are subject of intense research. First, in terms of prevention, Schwerk, *et al.* observed a positive impact of long-term antibiotics (mainly amoxicillin and amoxicillin-clavulanic acid) in preschoolers with severe wheeze who had positive bacterial infection on BAL samples, with reduced rates of exacerbations and hospitalizations at 6 months follow-up compared with

718 rates before treatment [197]. Second, immunomodulatory effects of macrolides are welldescribed [198], and have been evaluated in the treatment of acute asthma in children. 719 Hence, in the Early Administration of Azithromycin and Prevention of Severe Lower 720 Respiratory Tract Illnesses in Preschool Children (APRIL) study, Bacharier, et al. conducted 721 722 a pacebo-controlled trial in 607 preschool children with recurrent wheeze and no controller medication, testing the efficacy of azithromycin started prior to the development of lower 723 respiratory tract illness [199]. Macrolide administration reduced the risk of progressing to 724 severe lower respiratory tract illness (*i.e.* requiring a course of oral steroids), without 725 evidence for induction of azithromycin-resistant organisms, nor adverse events. In the 726 Copenhagen Prospective Studies on Asthma in Childhood 2010 (COPSAC2010) cohort, 727 728 children with recurrent asthma-like symptoms, some of whom under maintenance treatment, 729 were randomly assigned to receive either azithromycin or placebo in case of episode of 730 asthma-like symptoms, lasting at least 3 days [200]. Children treated with azithromycin displayed a significant reduction of episode length, and the effect size increased with early 731 administration of treatment. Moreover, Mandhane, et al. conducted a placebo-control trial in 732 preschool children aged 12 to 60 months, presenting to the emergency department with 733 734 wheeze, to assess the effect of azithromycin [201]. They did not show a reduction of duration of respiratory symptoms in the treated group, neither in the time to the next exacerbation 735 upon 6 months follow-up. Thus, there is still limited evidence on the use of azithromycin as a 736 preventive treatment of acute asthma attacks [202]. In addition, studies have been 737 conducted in adult patients to test the effect of azithromycin as a preventive strategy, with 738 conflicting results [203-205]. Notably, Gibson, et al. described in a placebo-controlled trial 739 that 48 weeks maintenance treatment with azithromycin decreased the frequency of asthma 740 exacerbations and improved asthma-related quality of life in symptomatic asthma despite 741 current maintenance treatment [204]. In a meta-analysis of the three main studies conducted 742 in adult patients [203-205], maintenance treatment with azithromycin was associated with a 743 reduced rate of exacerbations in asthma as well as in all subgroups (non-eosinophilic, 744 745 eosinophilic and severe asthma), and a good tolerance [206]. Examining each exacerbation

746 type separately, patients with eosinophilic asthma reported fewer oral corticosteroid courses, and patients with non-eosinophilic and severe asthma reported fewer antibiotic courses. 747 Additionally, a specific effect of macrolides in neutrophilic asthma had been reported 748 previously by Simpson, et al. in adult subjects with severe refractory asthma, treated with 749 750 clarithromycin for 8 weeks [207]. Clarithromycin therapy reduced sputum concentrations of IL-8 and neutrophil numbers leading to lower levels of neutrophil elastase and Matrix 751 metallopeptidase (MMP)-9 concentrations versus placebo. Finally, a decrease of 752 inflammatory cytokines, notably IL-1β, IL-6 and IL-33, was observed in vitro in cultures of 753 human AEC from asthmatic patients pretreated with clarithromycin, in a context of RV14 754 infection, suggesting a direct anti-viral effect [208]. Consistent with this hypothesis, another 755 756 in vitro study showed an effect of azithromycin to reduce RV replication after inoculation in AEC, associated with an increase in IFN and interferon-stimulated genes mRNA expression 757 758 and protein production [209].

To sum up, although macrolides and more particularly azithromycin constitute an appealing strategy in acute asthma, further studies are needed to identify the phenotypes of potential responders and the possible adverse events. Since antibiotics have the potential limitation of eliminating commensal bacteria that protect from asthma development, it must be emphasized that their use should only be considered in particular clinical situations [188].

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766 **5.4-** Potential role of biologics targeting type 2 inflammation in anti-microbial 767 response

We have highlighted the close interactions between underlying airway inflammation, host anti-microbial responses and microorganisms leading to asthma attacks. Therefore, novel therapeutics targeting inflammatory pathways in asthma, such as anti-Th2 inflammation biologics, may have a specific impact on lung infections.

772 Omalizumab is an anti-IgE monoclonal antibody, targeting the effector mechanisms 773 of Th2 inflammation. After proof of efficacy in pediatric clinical trials, it has been widely

774 prescribed in severe asthmatic atopic children aged older than 6 years, providing more insights on the longer-term evolution [210]. In real-life studies, we have observed higher 775 levels of asthma control after its initiation, and a significant decrease of asthma attacks in 776 children previously prone to exacerbation [211]. Hence, a direct effect of omalizumab on RV-777 778 induced exacerbations in children has been suggested [212,213]. In the Preventative Omalizumab or Step-up Therapy for Severe Fall Exacerbations (PROSE) study, a placebo-779 controlled trial conducted among 478 inner-city asthmatic children aged 6 to 17 years with 1 780 781 or more recent exacerbations, Esquivel, et al. observed a decrease of the duration of RV infections, viral shedding, and risk of RV-induced illnesses in omalizumab-treated patients 782 783 [212]. A recently published adult study including results from two clinical trials has compared 784 the effect of omalizumab on the response to an experimental inoculation with RV-16 in allergic asthmatic adults [214]. Omalizumab-treated patients displayed a reduction of 785 symptoms of lower respiratory tract infection, more pronounced in the four first days 786 following inoculation, suggesting that the protective effect of omalizumab on virus-induced 787 788 exacerbations occurs in the early stages of infection. Teach, et al. described other results 789 from the PROSE study that compared omalizumab with placebo and omalizumab with an 790 inhaled corticosteroid boost with regard to fall exacerbation rates when initiated 4 to 6 weeks before return to school [215]. They recorded 86 exacerbations, among which 89% were 791 associated with a respiratory tract virus detection, most commonly RV (81%). More 792 793 importantly, they observed a significantly lower fall exacerbation rate in the omalizumab group, but no difference between omalizumab and inhaled corticosteroid boost except in the 794 subgroup of patients with exacerbation during the run-in phase. In an ex vivo study of PBMC 795 taken during the intervention phase, the authors observed increased IFN-α responses 796 following RV-A16 inoculation in the omalizumab-treated group, suggesting that the 797 mechanism of Omalizumab protection against exacerbations could involve IFN-a dependent 798 antiviral effect [20]. Another study in children with exacerbation-prone asthma before and 799 during omalizumab treatment described the effect of ex vivo inoculation of RV and Influenza 800 801 A virus on PBMC and blood pDC, after IgE cross-linking on cell-surface FcERI [216]. The

authors observed that omalizumab increased both virus-induced IFN- α responses in PBMCs and pDC, and subsequently reduced FccRI expression on pDC. Altogether, the lower asthma exacerbation rates consecutive to omalizumab treatment were related to IFN- α responses and attenuated FccRI expression on pDC.

806 Omalizumab remains the most-studied biologic in pediatric asthma but others have been experimented in the past years [217]. Mepolizumab, an anti-IL-5 antibody therapy, has 807 proven to be efficient in reducing the exacerbation rate in adults and is now available in 808 children older than 6 years old with eosinophilic asthma in Europe and the US [218]. As 809 previously stated, its efficacy in reducing exacerbation rates in pediatric clinical trials and 810 real-life studies remain to be determined [129]. However, adult studies suggest it may have a 811 812 modifying effect on RV-induced inflammatory response. Hence, a placebo-controlled trial 813 was conducted in adult patients with mild asthma, randomized to receive mepolizumab versus placebo, followed by RV-16 inoculation, to study its impact on anti-viral responses 814 [219]. The authors showed that mepolizumab attenuated baseline blood eosinophils 815 816 numbers and activation status, but did not prevent the activation of remaining eosinophils 817 upon RV infection. However, after infection, it enhanced levels of airway B lymphocytes and 818 AM and modified BAL levels of CCL20 and IL-1RA. Effects of other drugs targeting Th2 cytokines and alarmins, including lebrikizumab (anti IL-13), dupilumab (anti IL-4R), 819 reslizumab (anti-IL-5), benralizumab (anti-IL-5R), tezepelumab (anti-TSLP), or anti-IL-33 820 821 monoclonal antibodies are currently studied in pre-clinical and clinical studies [217].

In the next years, new studies are needed regarding these novel treatments, both in allergic and non-allergic asthmatic children, to provide new evidence on their immunological effects and potential disease-modifying properties, and to fully determine the phenotypes/endotypes of good responders. Most importantly, specific studies in the youngest children need to be carried out, with appropriate end-points [129]. This effort will contribute to enter the era of precision medicine in pediatric asthma.

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830

6. Conclusion

In conclusion, asthma attacks are a main feature of pediatric asthma, especially of 831 severe asthma. We have highlighted studies suggesting impairment of the mucosal anti-832 microbial immune responses during acute asthma and/or at steady state. Interestingly, these 833 834 alterations seem to inconstantly persist into adulthood. These various altered pathways may reflect different endotypes underlying asthma heterogeneity and consequently, constitute 835 targets for new treatment approaches. Preventing, understanding and efficiently managing 836 acute asthma attacks remain one of the unmet needs in severe childhood asthma, especially 837 in the youngest children. At the era of precision medicine, early interventions targeting 838 mucosal immune responses, lung infections, and microbiota, in early childhood, could have 839 840 disease-modifying properties and positive impact on the trajectory of childhood asthma.

841

842

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849

851 **CrediT:**

- Lejeune S: Conceptualization, Data collection Data analysis, Writing, Review & Editing,
- 853 Deschildre A: Conceptualization, Data collection Data analysis, Writing, Review & Editing,
- Le Rouzic O: Data collection Data analysis, Writing, Review & Editing,
- 855 Engelmann I: Data collection Data analysis, Writing, Review & Editing,
- 856 Dessein R: Data collection Data analysis, Writing, Review & Editing,
- 857 Pichavant M: Data collection Data analysis, Writing, Review & Editing,
- 858 Gosset P[:] Conceptualization, Data collection Data analysis, Writing, Review & Editing.
- 859

860 **Figure Legends**:

Figure 1: Summary of the main mechanisms favoring asthma development and involved in asthma attack.

These mechanisms **(in red)** involve: (1) Impairment of innate immune responses; (2) Influence of the host-microbiota dialog on Th2 inflammation; (3) Pathogen characteristics; (4) Airway leukocyte inflammation. These dynamic interactions may impact the presentations of asthma attacks, and have long-term consequences.

867

AM: Alveolar macrophages; AEC: airway epithelial cells; DC: dendritic cells; IFN: Interferon;
IL: Interleukin; ILC2: type 2 innate lymphoid cells; IRF: interferon regulatory factor; PAMP:
Pathogen-associated molecular pattern; PRR: pattern recognition receptor; RV: Rhinovirus;
TSLP: Thymic stromal lymphopoietin.

872

Figure 2: Possible therapeutics strategies to limit asthma development/progression
and attacks.

These strategies (in green) target: (1) Enhancement of the innate immune responses; (2) Anti-infectious therapeutics and strategies to modulate the microbiota; (3) Alarmins and anti-Th2 biologics.

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882	
881	SCFA: Short Chain Fatty Acid, TSLP: Thymic stromal lymphopoietin.
880	Pathogen-associated molecular pattern; PRR: pattern recognition receptor; RV: Rhinovirus;
879	IL: Interleukin; ILC2: type 2 innate lymphoid cells; IRF: interferon regulatory factor; PAMP:
878	AM: Alveolar macrophages; AEC: airway epithelial cells; DC: dendritic cells; IFN: Interferon;

883 Capsule summary:

- 884 Characteristics of host-microbiota interactions in asthmatic children favor asthma attacks
- and may allow to define asthma endotypes. This knowledge offers promising perspectives to
- 886 treat asthmatic children using precision medicine.

888 Bibliographical references

- M.-C. Delmas, N. Guignon, B. Leynaert, M. Moisy, C. Marguet, C. Fuhrman, [Increase
 in asthma prevalence among young children in France], Rev Mal Respir. 34 (2017)
 525–534. https://doi.org/10.1016/j.rmr.2016.09.002.
- S. Ebmeier, D. Thayabaran, I. Braithwaite, C. Bénamara, M. Weatherall, R. Beasley,
 Trends in international asthma mortality: analysis of data from the WHO Mortality
 Database from 46 countries (1993-2012), Lancet. 390 (2017) 935–945.
 https://doi.org/10.1016/S0140-6736(17)31448-4.
- 896 [3] S. Saglani, L. Fleming, S. Sonnappa, A. Bush, Advances in the aetiology,
 897 management, and prevention of acute asthma attacks in children, Lancet Child Adolesc
 898 Health. 3 (2019) 354–364. https://doi.org/10.1016/S2352-4642(19)30025-2.
- [4] C.I. Bloom, F. Nissen, I.J. Douglas, L. Smeeth, P. Cullinan, J.K. Quint, Exacerbation
 risk and characterisation of the UK's asthma population from infants to old age, Thorax.
 73 (2018) 313–320. https://doi.org/10.1136/thoraxjnl-2017-210650.
- [5] Global Initiative for Asthma (GINA). 2020 Report: Global strategy for asthma
 management and prevention. https://ginasthma.org/ (last accessed: April 2020), (n.d.).
- 904 [6] P.L.P. Brand, D. Caudri, E. Eber, E.A. Gaillard, L. Garcia-Marcos, G. Hedlin, J.
 905 Henderson, C.E. Kuehni, P.J.F.M. Merkus, S. Pedersen, A. Valiulis, G. Wennergren, A.
 906 Bush, Classification and pharmacological treatment of preschool wheezing: changes
 907 since 2008, Eur. Respir. J. 43 (2014) 1172–1177.
 908 https://doi.org/10.1183/09031936.00199913
- 908 https://doi.org/10.1183/09031936.00199913.
- [7] L.B. Bacharier, B.R. Phillips, G.R. Bloomberg, R.S. Zeiger, I.M. Paul, M. Krawiec, T.
 Guilbert, V.M. Chinchilli, R.C. Strunk, Childhood Asthma Research and Education
 Network, National Heart, Lung, and Blood Institute, Severe intermittent wheezing in
 preschool children: a distinct phenotype, J. Allergy Clin. Immunol. 119 (2007) 604–610.
 https://doi.org/10.1016/j.jaci.2006.12.607.
- [8] S. Lejeune, M. Pichavant, I. Engelmann, L. Beghin, E. Drumez, O. Le Rouzic, R.
 Dessein, S. Rogeau, T. Beke, G. Kervoaze, C. Delvart, H. Ducoin, G. Pouessel, A. Le
 Mée, S. Boileau, J. Roussel, C. Bonnel, C. Mordacq, C. Thumerelle, P. Gosset, A.
 Deschildre, Severe preschool asthmatics have altered cytokine and anti-viral
 responses during exacerbation, Pediatr Allergy Immunol. (2020).
 https://doi.org/10.1111/pai.13268.
- [9] S.L. Johnston, P.K. Pattemore, G. Sanderson, S. Smith, F. Lampe, L. Josephs, P.
 Symington, S. O'Toole, S.H. Myint, D.A. Tyrrell, Community study of role of viral
 infections in exacerbations of asthma in 9-11 year old children, BMJ. 310 (1995) 1225–
 1229. https://doi.org/10.1136/bmj.310.6989.1225.
- [10] I. Engelmann, C. Mordacq, P. Gosset, I. Tillie-Leblond, A. Dewilde, C. Thumerelle, G.
 Pouessel, A. Deschildre, Rhinovirus and asthma: reinfection, not persistence, Am. J.
 Respir. Crit. Care Med. 188 (2013) 1165–1167. https://doi.org/10.1164/rccm.2013030585LE.
- [11] K.F. Chung, S.E. Wenzel, J.L. Brozek, A. Bush, M. Castro, P.J. Sterk, I.M. Adcock,
 E.D. Bateman, E.H. Bel, E.R. Bleecker, L.-P. Boulet, C. Brightling, P. Chanez, S.-E.
 Dahlen, R. Djukanovic, U. Frey, M. Gaga, P. Gibson, Q. Hamid, N.N. Jajour, T. Mauad,
 R.L. Sorkness, W.G. Teague, International ERS/ATS guidelines on definition,
 evaluation and treatment of severe asthma, Eur. Respir. J. 43 (2014) 343–373.
 https://doi.org/10.1183/09031936.00202013.
- [12] G. Hedlin, A. Bush, K. Lødrup Carlsen, G. Wennergren, F.M. De Benedictis, E. Melén,
 J. Paton, N. Wilson, K.-H. Carlsen, Problematic Severe Asthma in Childhood Initiative
 group, Problematic severe asthma in children, not one problem but many: a GA2LEN
 initiative, Eur. Respir. J. 36 (2010) 196–201.
- 938 https://doi.org/10.1183/09031936.00104809.
- [13] W.G. Teague, B.R. Phillips, J.V. Fahy, S.E. Wenzel, A.M. Fitzpatrick, W.C. Moore, A.T.
 Hastie, E.R. Bleecker, D.A. Meyers, S.P. Peters, M. Castro, A.M. Coverstone, L.B.
- Bacharier, N.P. Ly, M.C. Peters, L.C. Denlinger, S. Ramratnam, R.L. Sorkness, B.M.

- Gaston, S.C. Erzurum, S.A.A. Comhair, R.E. Myers, J. Zein, M.D. DeBoer, A.-M. Irani,
 E. Israel, B. Levy, J.C. Cardet, W. Phipatanakul, J.M. Gaffin, F. Holguin, M.L. Fajt, S.J.
 Aujla, D.T. Mauger, N.N. Jarjour, Baseline Features of the Severe Asthma Research
 Program (SARP III) Cohort: Differences with Age, J Allergy Clin Immunol Pract. 6
 (2018) 545-554.e4. https://doi.org/10.1016/j.jaip.2017.05.032.
- [14] L. Fleming, C. Murray, A.T. Bansal, S. Hashimoto, H. Bisgaard, A. Bush, U. Frey, G.
 Hedlin, F. Singer, W.M. van Aalderen, N.H. Vissing, Z. Zolkipli, A. Selby, S. Fowler, D.
 Shaw, K.F. Chung, A.R. Sousa, S. Wagers, J. Corfield, I. Pandis, A. Rowe, E.
 Formaggio, P.J. Sterk, G. Roberts, U-BIOPRED Study Group, The burden of severe
 asthma in childhood and adolescence: results from the paediatric U-BIOPRED cohorts,
 Eur. Respir. J. 46 (2015) 1322–1333. https://doi.org/10.1183/13993003.00780-2015.
- [15] L.C. Denlinger, P. Heymann, R. Lutter, J.E. Gern, Exacerbation-Prone Asthma, J
 Allergy Clin Immunol Pract. 8 (2020) 474–482.
- 955 https://doi.org/10.1016/j.jaip.2019.11.009.
- [16] K.R. Ross, R. Gupta, M.D. DeBoer, J. Zein, B.R. Phillips, D.T. Mauger, C. Li, R.E.
 Myers, W. Phipatanakul, A.M. Fitzpatrick, N.P. Ly, L.B. Bacharier, D.J. Jackson, J.C.
 Celedón, A. Larkin, E. Israel, B. Levy, J.V. Fahy, M. Castro, E.R. Bleecker, D. Meyers,
 W.C. Moore, S.E. Wenzel, N.N. Jarjour, S.C. Erzurum, W.G. Teague, B. Gaston,
 Severe asthma during childhood and adolescence: A longitudinal study, J. Allergy Clin.
 Immunol. 145 (2020) 140-146.e9. https://doi.org/10.1016/j.jaci.2019.09.030.
- 962 [17] D.C.M. Belgrave, I. Buchan, C. Bishop, L. Lowe, A. Simpson, A. Custovic, Trajectories
 963 of lung function during childhood, Am. J. Respir. Crit. Care Med. 189 (2014) 1101–
 964 1109. https://doi.org/10.1164/rccm.201309-1700OC.
- [18] A. Beigelman, L.B. Bacharier, Management of Preschool Children with Recurrent
 Wheezing: Lessons from the NHLBI's Asthma Research Networks, J Allergy Clin
 Immunol Pract. 4 (2016) 1–8; quiz 9–10. https://doi.org/10.1016/j.jaip.2015.10.003.
- [19] A. Bourdin, I. Adcock, P. Berger, P. Bonniaud, P. Chanson, C. Chenivesse, J. de Blic,
 A. Deschildre, P. Devillier, G. Devouassoux, A. Didier, G. Garcia, A. Magnan, Y.
 Martinat, T. Perez, N. Roche, C. Taillé, P. Val, P. Chanez, How can we minimise the
 use of regular oral corticosteroids in asthma?, Eur Respir Rev. 29 (2020).
 https://doi.org/10.1183/16000617.0085-2019.
- 973 [20] A. Licari, S. Manti, R. Castagnoli, A. Marseglia, T. Foiadelli, I. Brambilla, G.L.
 974 Marseglia, Immunomodulation in Pediatric Asthma, Front Pediatr. 7 (2019) 289.
 975 https://doi.org/10.3389/fped.2019.00289.
- 976 [21] T. Jartti, J.E. Gern, Role of viral infections in the development and exacerbation of
 977 asthma in children, J. Allergy Clin. Immunol. 140 (2017) 895–906.
 978 https://doi.org/10.1016/j.jaci.2017.08.003.
- [22] T. Jartti, M. Korppi, Rhinovirus-induced bronchiolitis and asthma development, Pediatr
 Allergy Immunol. 22 (2011) 350–355. https://doi.org/10.1111/j.13993038.2011.01170.x.
- [23] D.J. Jackson, R.E. Gangnon, M.D. Evans, K.A. Roberg, E.L. Anderson, T.E. Pappas,
 M.C. Printz, W.-M. Lee, P.A. Shult, E. Reisdorf, K.T. Carlson-Dakes, L.P. Salazar, D.F.
 DaSilva, C.J. Tisler, J.E. Gern, R.F. Lemanske, Wheezing rhinovirus illnesses in early
 life predict asthma development in high-risk children, Am. J. Respir. Crit. Care Med.
 178 (2008) 667–672. https://doi.org/10.1164/rccm.200802-309OC.
- M. Lukkarinen, A. Koistinen, R. Turunen, P. Lehtinen, T. Vuorinen, T. Jartti, Rhinovirus induced first wheezing episode predicts atopic but not nonatopic asthma at school age,
 J. Allergy Clin. Immunol. 140 (2017) 988–995.
- 990 https://doi.org/10.1016/j.jaci.2016.12.991.
- [25] E. Bergroth, M. Aakula, M. Korppi, S. Remes, J.E. Kivistö, P.A. Piedra, C.A. Camargo,
 T. Jartti, Post-bronchiolitis Use of Asthma Medication: A Prospective 1-year Follow-up
 Study, Pediatr. Infect. Dis. J. 35 (2016) 363–368.
 https://doi.org/10.1097/INE.0000000001017
- 994 https://doi.org/10.1097/INF.00000000001017.
- [26] F.J. Rubner, D.J. Jackson, M.D. Evans, R.E. Gangnon, C.J. Tisler, T.E. Pappas, J.E.
 Gern, R.F. Lemanske, Early life rhinovirus wheezing, allergic sensitization, and asthma

- risk at adolescence, J. Allergy Clin. Immunol. 139 (2017) 501–507.
- 998 https://doi.org/10.1016/j.jaci.2016.03.049.
- [27] C. Thumerelle, A. Deschildre, C. Bouquillon, C. Santos, A. Sardet, M. Scalbert, L.
 Delbecque, P. Debray, A. Dewilde, D. Turck, F. Leclerc, Role of viruses and atypical
 bacteria in exacerbations of asthma in hospitalized children: a prospective study in the
 Nord-Pas de Calais region (France), Pediatr. Pulmonol. 35 (2003) 75–82.
 https://doi.org/10.1002/ppul.10191.
- IO04 [28] N. Khetsuriani, N.N. Kazerouni, D.D. Erdman, X. Lu, S.C. Redd, L.J. Anderson, W.G.
 Teague, Prevalence of viral respiratory tract infections in children with asthma, J.
 Allergy Clin. Immunol. 119 (2007) 314–321. https://doi.org/10.1016/j.jaci.2006.08.041.
- [29] J.L. Kennedy, M. Shaker, V. McMeen, J. Gern, H. Carper, D. Murphy, W.-M. Lee, Y.A.
 Bochkov, R.F. Vrtis, T. Platts-Mills, J. Patrie, L. Borish, J.W. Steinke, W.A. Woods,
 P.W. Heymann, Comparison of viral load in individuals with and without asthma during
 infections with rhinovirus, Am. J. Respir. Crit. Care Med. 189 (2014) 532–539.
 https://doi.org/10.1164/rccm.201310-1767OC.
- [30] A. Bergauer, N. Sopel, B. Kroß, T. Vuorinen, P. Xepapadaki, S.T. Weiss, A. Blau, H.
 Sharma, C. Kraus, R. Springel, M. Rauh, S. Mittler, A. Graser, T. Zimmermann, V.O.
 Melichar, A. Kiefer, M.L. Kowalski, A. Sobanska, T. Jartti, H. Lukkarinen, N.G.
 Papadopoulos, S. Finotto, IFN-α/IFN-λ responses to respiratory viruses in paediatric
 asthma, Eur. Respir. J. 49 (2017). https://doi.org/10.1183/13993003.00969-2016.
- [31] S. Lejeune, I. Engelmann, R. Dessein, G. Pouessel, H. Ducoin, C. Delvart, C. Mordacq,
 C. Thumerelle, M. Pichavant, P. Gosset, A. Deschildre, Microbiological status in
 preschool asthma: the VIRASTHMA 2 study, European Respiratory Journal. 54 (2019).
 https://doi.org/10.1183/13993003.congress-2019.OA4938.
- 1021 [32] H. Bisgaard, M.N. Hermansen, F. Buchvald, L. Loland, L.B. Halkjaer, K. Bønnelykke,
 1022 M. Brasholt, A. Heltberg, N.H. Vissing, S.V. Thorsen, M. Stage, C.B. Pipper, Childhood
 1023 asthma after bacterial colonization of the airway in neonates, N. Engl. J. Med. 357
 1024 (2007) 1487–1495. https://doi.org/10.1056/NEJMoa052632.
- [33] K.M. Kloepfer, W.M. Lee, T.E. Pappas, T.J. Kang, R.F. Vrtis, M.D. Evans, R.E.
 Gangnon, Y.A. Bochkov, D.J. Jackson, R.F. Lemanske, J.E. Gern, Detection of
 pathogenic bacteria during rhinovirus infection is associated with increased respiratory
 symptoms and asthma exacerbations, J. Allergy Clin. Immunol. 133 (2014) 1301–1307,
 1307.e1–3. https://doi.org/10.1016/j.jaci.2014.02.030.
- [34] P.F.M. Robinson, C. Pattaroni, J. Cook, L. Gregory, A.M. Alonso, L.J. Fleming, C.M.
 Lloyd, A. Bush, B.J. Marsland, S. Saglani, Lower airway microbiota associates with
 inflammatory phenotype in severe preschool wheeze, J. Allergy Clin. Immunol. 143
 (2019) 1607-1610.e3. https://doi.org/10.1016/j.jaci.2018.12.985.
- [35] K. McCauley, J. Durack, R. Valladares, D.W. Fadrosh, D.L. Lin, A. Calatroni, P.K. 1034 1035 LeBeau, H.T. Tran, K.E. Fujimura, B. LaMere, G. Merana, K. Lynch, R.T. Cohen, J. Pongracic, G.K. Khurana Hershey, C.M. Kercsmar, M. Gill, A.H. Liu, H. Kim, M. Kattan, 1036 S.J. Teach, A. Togias, H.A. Boushey, J.E. Gern, D.J. Jackson, S.V. Lynch, National 1037 Institute of Allergy and Infectious Diseases-sponsored Inner-City Asthma Consortium. 1038 Distinct nasal airway bacterial microbiotas differentially relate to exacerbation in 1039 1040 pediatric patients with asthma, J. Allergy Clin. Immunol. 144 (2019) 1187–1197. https://doi.org/10.1016/j.jaci.2019.05.035. 1041
- 1042 [36] M. Vareille, E. Kieninger, M.R. Edwards, N. Regamey, The airway epithelium: soldier in
 1043 the fight against respiratory viruses, Clin. Microbiol. Rev. 24 (2011) 210–229.
 1044 https://doi.org/10.1128/CMR.00014-10.
- 1045 [37] A. Bush, Cytokines and Chemokines as Biomarkers of Future Asthma, Front Pediatr. 7 1046 (2019) 72. https://doi.org/10.3389/fped.2019.00072.
- 1047 [38] F.M.A. Hossain, J.Y. Choi, E. Uyangaa, S.O. Park, S.K. Eo, The Interplay between
 1048 Host Immunity and Respiratory Viral Infection in Asthma Exacerbation, Immune Netw.
 1049 19 (2019) e31. https://doi.org/10.4110/in.2019.19.e31.
- 1050 [39] J.A. Whitsett, T. Alenghat, Respiratory epithelial cells orchestrate pulmonary innate 1051 immunity, Nat. Immunol. 16 (2015) 27–35. https://doi.org/10.1038/ni.3045.

- [40] N. Kadowaki, S. Ho, S. Antonenko, R.W. Malefyt, R.A. Kastelein, F. Bazan, Y.J. Liu, Subsets of human dendritic cell precursors express different toll-like receptors and respond to different microbial antigens, J. Exp. Med. 194 (2001) 863–869. https://doi.org/10.1084/jem.194.6.863.
- [41] W. Barchet, A. Blasius, M. Cella, M. Colonna, Plasmacytoid dendritic cells: in search of their niche in immune responses, Immunol. Res. 32 (2005) 75–83. https://doi.org/10.1385/IR:32:1-3:075.
- 1059 [42] H. See, P. Wark, Innate immune response to viral infection of the lungs, Paediatr 1060 Respir Rev. 9 (2008) 243–250. https://doi.org/10.1016/j.prrv.2008.04.001.
- 1061 [43] C. Duez, P. Gosset, A.-B. Tonnel, Dendritic cells and toll-like receptors in allergy and 1062 asthma, Eur J Dermatol. 16 (2006) 12–16.
- [44] F.P. Siegal, N. Kadowaki, M. Shodell, P.A. Fitzgerald-Bocarsly, K. Shah, S. Ho, S.
 Antonenko, Y.J. Liu, The nature of the principal type 1 interferon-producing cells in human blood, Science. 284 (1999) 1835–1837.
 https://doi.org/10.1126/science.284.5421.1835.
- [45] M.R. Edwards, N. Regamey, M. Vareille, E. Kieninger, A. Gupta, A. Shoemark, S.
 Saglani, A. Sykes, J. Macintyre, J. Davies, C. Bossley, A. Bush, S.L. Johnston,
 Impaired innate interferon induction in severe therapy resistant atopic asthmatic
 children, Mucosal Immunol. 6 (2013) 797–806. https://doi.org/10.1038/mi.2012.118.
- [46] K.S. Parsons, A.C. Hsu, P. a. B. Wark, TLR3 and MDA5 signalling, although not
 expression, is impaired in asthmatic epithelial cells in response to rhinovirus infection,
 Clin. Exp. Allergy. 44 (2014) 91–101. https://doi.org/10.1111/cea.12218.
- [47] H. Rupani, R.T. Martinez-Nunez, P. Dennison, L.C.K. Lau, N. Jayasekera, T. Havelock,
 A.S. Francisco-Garcia, C. Grainge, P.H. Howarth, T. Sanchez-Elsner, Toll-like
 Receptor 7 Is Reduced in Severe Asthma and Linked to an Altered MicroRNA Profile,
 Am. J. Respir. Crit. Care Med. 194 (2016) 26–37. https://doi.org/10.1164/rccm.2015020280OC.
- [48] A. Sykes, M.R. Edwards, J. Macintyre, A. del Rosario, E. Bakhsoliani, M.-B. Trujillo-Torralbo, O.M. Kon, P. Mallia, M. McHale, S.L. Johnston, Rhinovirus 16-induced IFN-α and IFN-β are deficient in bronchoalveolar lavage cells in asthmatic patients, J. Allergy Clin. Immunol. 129 (2012) 1506-1514.e6. https://doi.org/10.1016/j.jaci.2012.03.044.
- [49] A. Deschildre, M. Pichavant, I. Engelmann, C. Langlois, E. Drumez, G. Pouessel, S.
 Boileau, D. Romero-Cubero, I. Decleyre-Badiu, A. Dewilde, D. Hober, V. Néve, C.
 Thumerelle, S. Lejeune, C. Mordacq, P. Gosset, Virus-triggered exacerbation in allergic
 asthmatic children: neutrophilic airway inflammation and alteration of virus sensors
 characterize a subgroup of patients, Respir. Res. 18 (2017) 191.
 https://doi.org/10.1186/s12931-017-0672-0.
- [50] S. Lejeune, C. Mordacq, E. Drumez, S. Brisset, G. Pouessel, M. Pichavant, I.
 Engelmann, L. Béghin, I. Decleyre-Badiu, V. Neve, C. Thumerelle, P. Gosset, A.
 Deschildre, Relationship between immune parameters during a severe exacerbation in
 allergic asthmatic children and asthma outcomes in the following year, Clin. Exp.
 Allergy. (2020). https://doi.org/10.1111/cea.13570.
- 1094 [51] D.J. Jackson, S.L. Johnston, The role of viruses in acute exacerbations of asthma, J.
 1095 Allergy Clin. Immunol. 125 (2010) 1178–1187; quiz 1188–1189.
 1096 https://doi.org/10.1016/j.jaci.2010.04.021.
- [52] V. Casanova, F.H. Sousa, C. Stevens, P.G. Barlow, Antiviral therapeutic approaches
 for human rhinovirus infections, Future Virol. 13 (2018) 505–518.
 https://doi.org/10.2217/fvl-2018-0016.
- 1100 [53] P.G. Barlow, E.G. Findlay, S.M. Currie, D.J. Davidson, Antiviral potential of 1101 cathelicidins, Future Microbiol. 9 (2014) 55–73. https://doi.org/10.2217/fmb.13.135.
- [54] S.M. Currie, E. Gwyer Findlay, A.J. McFarlane, P.M. Fitch, B. Böttcher, N. Colegrave,
 A. Paras, A. Jozwik, C. Chiu, J. Schwarze, D.J. Davidson, Cathelicidins Have Direct
 Antiviral Activity against Respiratory Syncytial Virus In Vitro and Protective Function In
 Vivo in Mice and Humans, J. Immunol. 196 (2016) 2699–2710.
- 1106 https://doi.org/10.4049/jimmunol.1502478.

- 1107 [55] T. Arikoglu, S. Kuyucu, E. Karaismailoglu, S.B. Batmaz, S. Balci, The association of
 vitamin D, cathelicidin, and vitamin D binding protein with acute asthma attacks in
 children, Allergy Asthma Proc. 36 (2015) 51–58.
 https://doi.org/10.2500/aap.2015.36.3848.
- [56] S. Baraldo, M. Contoli, E. Bazzan, G. Turato, A. Padovani, B. Marku, F. Calabrese, G. Caramori, A. Ballarin, D. Snijders, A. Barbato, M. Saetta, A. Papi, Deficient antiviral immune responses in childhood: distinct roles of atopy and asthma, J. Allergy Clin. Immunol. 130 (2012) 1307–1314. https://doi.org/10.1016/j.jaci.2012.08.005.
- [57] V. Gielen, A. Sykes, J. Zhu, B. Chan, J. Macintyre, N. Regamey, E. Kieninger, A.
 Gupta, A. Shoemark, C. Bossley, J. Davies, S. Saglani, P. Walker, S.E. Nicholson, A.H.
 Dalpke, O.-M. Kon, A. Bush, S.L. Johnston, M.R. Edwards, Increased nuclear
 suppressor of cytokine signaling 1 in asthmatic bronchial epithelium suppresses
 rhinovirus induction of innate interferons, J. Allergy Clin. Immunol. 136 (2015) 177188.e11. https://doi.org/10.1016/j.jaci.2014.11.039.
- [58] A. Kicic, P.T. Stevens, E.N. Sutanto, E. Kicic-Starcevich, K.-M. Ling, K. Looi, K.M.
 Martinovich, L.W. Garratt, T. Iosifidis, N.C. Shaw, A.G. Buckley, P.J. Rigby, F.J.
 Lannigan, D.A. Knight, S.M. Stick, Impaired airway epithelial cell responses from
 children with asthma to rhinoviral infection, Clin. Exp. Allergy. 46 (2016) 1441–1455.
 https://doi.org/10.1111/cea.12767.
- [59] M. Contoli, K. Ito, A. Padovani, D. Poletti, B. Marku, M.R. Edwards, L.A. Stanciu, G.
 Gnesini, A. Pastore, A. Spanevello, P. Morelli, S.L. Johnston, G. Caramori, A. Papi,
 Th2 cytokines impair innate immune responses to rhinovirus in respiratory epithelial
 cells, Allergy. 70 (2015) 910–920. https://doi.org/10.1111/all.12627.
- [60] P.A.B. Wark, S.L. Johnston, F. Bucchieri, R. Powell, S. Puddicombe, V. Laza-Stanca,
 S.T. Holgate, D.E. Davies, Asthmatic bronchial epithelial cells have a deficient innate
 immune response to infection with rhinovirus, J. Exp. Med. 201 (2005) 937–947.
 https://doi.org/10.1084/jem.20041901.
- [61] S. Moskwa, W. Piotrowski, J. Marczak, M. Pawełczyk, A. Lewandowska-Polak, M.
 Jarzębska, M. Brauncajs, A. Głobińska, P. Górski, N.G. Papadopoulos, M.R. Edwards,
 S.L. Johnston, M.L. Kowalski, Innate Immune Response to Viral Infections in Primary
 Bronchial Epithelial Cells is Modified by the Atopic Status of Asthmatic Patients, Allergy
 Asthma Immunol Res. 10 (2018) 144–154. https://doi.org/10.4168/aair.2018.10.2.144.
- [62] A. Bufe, K. Gehlhar, E. Grage-Griebenow, M. Ernst, Atopic phenotype in children is associated with decreased virus-induced interferon-alpha release, Int. Arch. Allergy Immunol. 127 (2002) 82–88. https://doi.org/10.1159/000048173.
- [63] K. Gehlhar, C. Bilitewski, K. Reinitz-Rademacher, G. Rohde, A. Bufe, Impaired virusinduced interferon-alpha2 release in adult asthmatic patients, Clin. Exp. Allergy. 36 (2006) 331–337. https://doi.org/10.1111/j.1365-2222.2006.02450.x.
- [64] K. likura, T. Katsunuma, S. Saika, S. Saito, S. Ichinohe, H. Ida, H. Saito, K. Matsumoto,
 Peripheral blood mononuclear cells from patients with bronchial asthma show impaired
 innate immune responses to rhinovirus in vitro, Int. Arch. Allergy Immunol. 155 Suppl 1
 (2011) 27–33. https://doi.org/10.1159/000327262.
- [65] S.R. Durrani, D.J. Montville, A.S. Pratt, S. Sahu, M.K. DeVries, V. Rajamanickam, R.E.
 Gangnon, M.A. Gill, J.E. Gern, R.F. Lemanske, D.J. Jackson, Innate immune
 responses to rhinovirus are reduced by the high-affinity IgE receptor in allergic
 asthmatic children, J. Allergy Clin. Immunol. 130 (2012) 489–495.
 https://doi.org/10.1016/j.jaci.2012.05.023.
- [66] J.L. Simpson, M. Carroll, I.A. Yang, P.N. Reynolds, S. Hodge, A.L. James, P.G.
 Gibson, J.W. Upham, Reduced Antiviral Interferon Production in Poorly Controlled
 Asthma Is Associated With Neutrophilic Inflammation and High-Dose Inhaled
 Corticosteroids, Chest. 149 (2016) 704–713.
- 1158 https://doi.org/10.1016/j.chest.2015.12.018.
- 1159 [67] J. Zhu, S.D. Message, P. Mallia, T. Kebadze, M. Contoli, C.K. Ward, E.S. Barnathan,
- M.A. Mascelli, O.M. Kon, A. Papi, L.A. Stanciu, M.R. Edwards, P.K. Jeffery, S.L.
 Johnston, Bronchial mucosal IFN-α/β and pattern recognition receptor expression in

- patients with experimental rhinovirus-induced asthma exacerbations, J. Allergy Clin.
 Immunol. 143 (2019) 114-125.e4. https://doi.org/10.1016/j.jaci.2018.04.003.
- 1164 [68] E.A. Schwantes, D.M. Manthei, L.C. Denlinger, M.D. Evans, J.E. Gern, N.N. Jarjour,
- S.K. Mathur, Interferon gene expression in sputum cells correlates with the Asthma
 Index Score during virus-induced exacerbations, Clin. Exp. Allergy. 44 (2014) 813–821.
 https://doi.org/10.1111/cea.12269.
- [69] S.V. Kotenko, G. Gallagher, V.V. Baurin, A. Lewis-Antes, M. Shen, N.K. Shah, J.A.
 Langer, F. Sheikh, H. Dickensheets, R.P. Donnelly, IFN-lambdas mediate antiviral
 protection through a distinct class II cytokine receptor complex, Nat. Immunol. 4 (2003)
 69–77. https://doi.org/10.1038/ni875.
- [70] E.K. Miller, J.Z. Hernandez, V. Wimmenauer, B.E. Shepherd, D. Hijano, R. Libster,
 M.E. Serra, N. Bhat, J.P. Batalle, Y. Mohamed, A. Reynaldi, A. Rodriguez, M. Otello, N.
 Pisapia, J. Bugna, M. Bellabarba, D. Kraft, S. Coviello, F.M. Ferolla, A. Chen, S.J.
 London, G.K. Siberry, J.V. Williams, F.P. Polack, A mechanistic role for type III IFN-λ1
 in asthma exacerbations mediated by human rhinoviruses, Am. J. Respir. Crit. Care
 Med. 185 (2012) 508–516. https://doi.org/10.1164/rccm.201108-1462OC.
- [71] M. Contoli, S.D. Message, V. Laza-Stanca, M.R. Edwards, P.A.B. Wark, N.W. Bartlett,
 T. Kebadze, P. Mallia, L.A. Stanciu, H.L. Parker, L. Slater, A. Lewis-Antes, O.M. Kon,
 S.T. Holgate, D.E. Davies, S.V. Kotenko, A. Papi, S.L. Johnston, Role of deficient type
 III interferon-lambda production in asthma exacerbations, Nat. Med. 12 (2006) 1023–
 1026. https://doi.org/10.1038/nm1462.
- 1183 [72] T. Boonpiyathad, Z.C. Sözener, P. Satitsuksanoa, C.A. Akdis, Immunologic
 1184 mechanisms in asthma, Semin. Immunol. 46 (2019) 101333.
 1185 https://doi.org/10.1016/j.smim.2019.101333.
- 1186
 [73]
 P.D. Mitchell, P.M. O'Byrne, Epithelial-Derived Cytokines in Asthma, Chest. 151 (2017)

 1187
 1338–1344. https://doi.org/10.1016/j.chest.2016.10.042.
- 1188 [74] H. Gao, S. Ying, Y. Dai, Pathological Roles of Neutrophil-Mediated Inflammation in
 1189 Asthma and Its Potential for Therapy as a Target, J Immunol Res. 2017 (2017)
 1190 3743048. https://doi.org/10.1155/2017/3743048.
- 1191 [75] J. Travers, M.E. Rothenberg, Eosinophils in mucosal immune responses, Mucosal 1192 Immunol. 8 (2015) 464–475. https://doi.org/10.1038/mi.2015.2.
- In J. Chesné, F. Braza, G. Mahay, S. Brouard, M. Aronica, A. Magnan, IL-17 in severe asthma. Where do we stand?, Am. J. Respir. Crit. Care Med. 190 (2014) 1094–1101.
 https://doi.org/10.1164/rccm.201405-0859PP.
- 1196 [77] A. Bellini, M.A. Marini, L. Bianchetti, M. Barczyk, M. Schmidt, S. Mattoli, Interleukin (IL)4, IL-13, and IL-17A differentially affect the profibrotic and proinflammatory functions of
 fibrocytes from asthmatic patients, Mucosal Immunol. 5 (2012) 140–149.
 https://doi.org/10.1038/mi.2011.60.
- [78] X.-H. Yuan, Y.-M. Li, Y.-Y. Shen, J. Yang, Y. Jin, Clinical and Th1/Th2 immune
 response features of hospitalized children with human rhinovirus infection, J. Med.
 Virol. 92 (2020) 26–33. https://doi.org/10.1002/jmv.25587.
- [79] H.-C. Lee, M.B. Headley, Y.-M. Loo, A. Berlin, M. Gale, J.S. Debley, N.W. Lukacs, S.F.
 Ziegler, Thymic stromal lymphopoietin is induced by respiratory syncytial virus-infected airway epithelial cells and promotes a type 2 response to infection, J. Allergy Clin.
 Immunol. 130 (2012) 1187-1196.e5. https://doi.org/10.1016/j.jaci.2012.07.031.
- [80] P. Haag, H. Sharma, M. Rauh, T. Zimmermann, T. Vuorinen, N.G. Papadopoulos, S.T.
 Weiss, S. Finotto, Soluble ST2 regulation by rhinovirus and 25(OH)-vitamin D3 in the
 blood of asthmatic children, Clin. Exp. Immunol. 193 (2018) 207–220.
 https://doi.org/10.1111/cei.13135.
- [81] D.J. Jackson, H. Makrinioti, B.M.J. Rana, B.W.H. Shamji, M.-B. Trujillo-Torralbo, J.
 Footitt, null Jerico Del-Rosario, A.G. Telcian, A. Nikonova, J. Zhu, J. Aniscenko, L.
 Gogsadze, E. Bakhsoliani, S. Traub, J. Dhariwal, J. Porter, D. Hunt, T. Hunt, T. Hunt,
 L.A. Stanciu, M. Khaitov, N.W. Bartlett, M.R. Edwards, O.M. Kon, P. Mallia, N.G.
- Papadopoulos, C.A. Akdis, J. Westwick, M.J. Edwards, D.J. Cousins, R.P. Walton, S.L.
 Johnston, IL-33-dependent type 2 inflammation during rhinovirus-induced asthma

- 1217 exacerbations in vivo, Am. J. Respir. Crit. Care Med. 190 (2014) 1373–1382.
 1218 https://doi.org/10.1164/rccm.201406-1039OC.
- [82] L.M. Jurak, Y. Xi, M. Landgraf, M.L. Carroll, L. Murray, J.W. Upham, Interleukin 33
 Selectively Augments Rhinovirus-Induced Type 2 Immune Responses in Asthmatic but not Healthy People, Front Immunol. 9 (2018) 1895.
- 1222 https://doi.org/10.3389/fimmu.2018.01895.
- [83] L. Qin, C. Hu, J. Feng, Q. Xia, Activation of lymphocytes induced by bronchial epithelial
 cells with prolonged RSV infection, PLoS ONE. 6 (2011) e27113.
 https://doi.org/10.1371/journal.pone.0027113.
- [84] T. Marichal, D. Bedoret, C. Mesnil, M. Pichavant, S. Goriely, F. Trottein, D. Cataldo, M.
 Goldman, P. Lekeux, F. Bureau, C.J. Desmet, Interferon response factor 3 is essential
 for house dust mite-induced airway allergy, J. Allergy Clin. Immunol. 126 (2010) 836844.e13. https://doi.org/10.1016/j.jaci.2010.06.009.
- [85] J. Bizzintino, W.-M. Lee, I.A. Laing, F. Vang, T. Pappas, G. Zhang, A.C. Martin, S.-K.
 Khoo, D.W. Cox, G.C. Geelhoed, P.C. McMinn, J. Goldblatt, J.E. Gern, P.N. Le Souëf,
 Association between human rhinovirus C and severity of acute asthma in children, Eur.
 Respir. J. 37 (2011) 1037–1042. https://doi.org/10.1183/09031936.00092410.
- [86] Y.A. Bochkov, K. Watters, S. Ashraf, T.F. Griggs, M.K. Devries, D.J. Jackson, A.C.
 Palmenberg, J.E. Gern, Cadherin-related family member 3, a childhood asthma
 susceptibility gene product, mediates rhinovirus C binding and replication, Proc. Natl.
 Acad. Sci. U.S.A. 112 (2015) 5485–5490. https://doi.org/10.1073/pnas.1421178112.
- [87] K. Nakagome, Y.A. Bochkov, S. Ashraf, R.A. Brockman-Schneider, M.D. Evans, T.R.
 Pasic, J.E. Gern, Effects of rhinovirus species on viral replication and cytokine
 production, J. Allergy Clin. Immunol. 134 (2014) 332–341.
 https://doi.org/10.1016/j.jaci.2014.01.029
- 1241 https://doi.org/10.1016/j.jaci.2014.01.029.
- [88] C.T. Sikazwe, I.A. Laing, A. Imrie, D.W. Smith, Nasal Cytokine Profiles of Patients
 Hospitalised with Respiratory Wheeze Associated with Rhinovirus C, Viruses. 11
 (2019). https://doi.org/10.3390/v11111038.
- [89] X. Lu, E. Schneider, S. Jain, A.M. Bramley, W. Hymas, C. Stockmann, K. Ampofo, S.R.
 Arnold, D.J. Williams, W.H. Self, A. Patel, J.D. Chappell, C.G. Grijalva, E.J. Anderson,
 R.G. Wunderink, J.A. McCullers, K.M. Edwards, A.T. Pavia, D.D. Erdman, Rhinovirus
 Viremia in Patients Hospitalized With Community-Acquired Pneumonia, J. Infect. Dis.
 216 (2017) 1104–1111. https://doi.org/10.1093/infdis/jix455.
- [90] N. Principi, A. Zampiero, M. Gambino, A. Scala, L. Senatore, M. Lelii, B. Ascolese, C.
 Pelucchi, S. Esposito, Prospective evaluation of rhinovirus infection in healthy young children, J. Clin. Virol. 66 (2015) 83–89. https://doi.org/10.1016/j.jcv.2015.03.013.
- [91] M.J. Loeffelholz, R. Trujillo, R.B. Pyles, A.L. Miller, P. Alvarez-Fernandez, D.L. Pong, T.
 Chonmaitree, Duration of rhinovirus shedding in the upper respiratory tract in the first
 year of life, Pediatrics. 134 (2014) 1144–1150. https://doi.org/10.1542/peds.2014-2132.
- I256 [92] J. da Silva, C. Hilzendeger, C. Moermans, F. Schleich, M. Henket, T. Kebadze, P.
 Mallia, M.R. Edwards, S.L. Johnston, R. Louis, Raised interferon-β, type 3 interferon
 and interferon-stimulated genes evidence of innate immune activation in neutrophilic
 asthma, Clin. Exp. Allergy. 47 (2017) 313–323. https://doi.org/10.1111/cea.12809.
- 1260 [93] K.M. Wylie, The Virome of the Human Respiratory Tract, Clin. Chest Med. 38 (2017) 1261 11–19. https://doi.org/10.1016/j.ccm.2016.11.001.
- [94] Y. Wang, N. Zhu, Y. Li, R. Lu, H. Wang, G. Liu, X. Zou, Z. Xie, W. Tan, Metagenomic
 analysis of viral genetic diversity in respiratory samples from children with severe acute
 respiratory infection in China, Clin. Microbiol. Infect. 22 (2016) 458.e1–9.
 https://doi.org/10.1016/j.cmi.2016.01.006.
- [95] G. Freer, F. Maggi, M. Pifferi, M.E. Di Cicco, D.G. Peroni, M. Pistello, The Virome and Its Major Component, Anellovirus, a Convoluted System Molding Human Immune Defenses and Possibly Affecting the Development of Asthma and Respiratory Diseases in Childhood, Front Microbiol. 9 (2018) 686. https://doi.org/10.3389/fmicb.2018.00686.
- 1270 [96] M. Pifferi, F. Maggi, E. Andreoli, L. Lanini, E.D. Marco, C. Fornai, M.L. Vatteroni, M.
- 1271 Pistello, V. Ragazzo, P. Macchia, A. Boner, M. Bendinelli, Associations between nasal

- 1272 torquetenovirus load and spirometric indices in children with asthma, J. Infect. Dis. 192 1273 (2005) 1141–1148. https://doi.org/10.1086/444389.
- [97] D.L. Dinwiddie, J.L. Denson, J.L. Kennedy, Role of the Airway Microbiome in
 Respiratory Infections and Asthma in Children, Pediatr Allergy Immunol Pulmonol. 31
 (2018) 236–240. https://doi.org/10.1089/ped.2018.0958.
- 1277 [98] L.T. Stiemsma, K.B. Michels, The Role of the Microbiome in the Developmental Origins 1278 of Health and Disease, Pediatrics. 141 (2018). https://doi.org/10.1542/peds.2017-2437.
- [99] J.M. Larsen, S. Brix, A.H. Thysen, S. Birch, M.A. Rasmussen, H. Bisgaard, Children
 with asthma by school age display aberrant immune responses to pathogenic airway
 bacteria as infants, J. Allergy Clin. Immunol. 133 (2014) 1008–1013.
 https://doi.org/10.1016/i.jaci.2014.01.010.
- [100]M.-C. Arrieta, L.T. Stiemsma, P.A. Dimitriu, L. Thorson, S. Russell, S. Yurist-Doutsch,
 B. Kuzeljevic, M.J. Gold, H.M. Britton, D.L. Lefebvre, P. Subbarao, P. Mandhane, A.
 Becker, K.M. McNagny, M.R. Sears, T. Kollmann, CHILD Study Investigators, W.W.
 Mohn, S.E. Turvey, B.B. Finlay, Early infancy microbial and metabolic alterations affect
 risk of childhood asthma, Sci Transl Med. 7 (2015) 307ra152.
 https://doi.org/10.1126/scitranslmed.aab2271.
- [101]K. Hufnagl, I. Pali-Schöll, F. Roth-Walter, E. Jensen-Jarolim, Dysbiosis of the gut and
 lung microbiome has a role in asthma, Semin Immunopathol. 42 (2020) 75–93.
 https://doi.org/10.1007/s00281-019-00775-y.
- [102]K. Loewen, B. Monchka, S.M. Mahmud, G. 't Jong, M.B. Azad, Prenatal antibiotic
 exposure and childhood asthma: a population-based study, Eur. Respir. J. 52 (2018).
 https://doi.org/10.1183/13993003.02070-2017.
- [103] L. Hoskin-Parr, A. Teyhan, A. Blocker, A.J.W. Henderson, Antibiotic exposure in the
 first two years of life and development of asthma and other allergic diseases by 7.5 yr:
 a dose-dependent relationship, Pediatr Allergy Immunol. 24 (2013) 762–771.
 https://doi.org/10.1111/pai.12153.
- [104] D.M. Patrick, H. Sbihi, D.L.Y. Dai, A. Al Mamun, D. Rasali, C. Rose, F. Marra, R.C.T.
 Boutin, C. Petersen, L.T. Stiemsma, G.L. Winsor, F.S.L. Brinkman, A.L. Kozyrskyj, M.B.
 Azad, A.B. Becker, P.J. Mandhane, T.J. Moraes, M.R. Sears, P. Subbarao, B.B. Finlay,
 S.E. Turvey, Decreasing antibiotic use, the gut microbiota, and asthma incidence in
 children: evidence from population-based and prospective cohort studies, Lancet
 Respir Med. (2020). https://doi.org/10.1016/S2213-2600(20)30052-7.
- [105]W.A.A. de Steenhuijsen Piters, S. Heinonen, R. Hasrat, E. Bunsow, B. Smith, M.-C.
 Suarez-Arrabal, D. Chaussabel, D.M. Cohen, E.A.M. Sanders, O. Ramilo, D. Bogaert,
 A. Mejias, Nasopharyngeal Microbiota, Host Transcriptome, and Disease Severity in
 Children with Respiratory Syncytial Virus Infection, Am. J. Respir. Crit. Care Med. 194
 (2016) 1104–1115. https://doi.org/10.1164/rccm.201602-0220OC.
- [106] J.M. Mansbach, K. Hasegawa, D.M. Henke, N.J. Ajami, J.F. Petrosino, C.A. Shaw, P.A.
 Piedra, A.F. Sullivan, J.A. Espinola, C.A. Camargo, Respiratory syncytial virus and
 rhinovirus severe bronchiolitis are associated with distinct nasopharyngeal microbiota,
 J. Allergy Clin. Immunol. 137 (2016) 1909-1913.e4.
- 1314 https://doi.org/10.1016/j.jaci.2016.01.036.
- [107]K.M. Kloepfer, V.K. Sarsani, V. Poroyko, W.M. Lee, T.E. Pappas, T. Kang, K.A.
 Grindle, Y.A. Bochkov, S.C. Janga, R.F. Lemanske, J.E. Gern, Community-acquired
 rhinovirus infection is associated with changes in the airway microbiome, J. Allergy
 Clin. Immunol. 140 (2017) 312-315.e8. https://doi.org/10.1016/j.jaci.2017.01.038.
- [108]M.J. Giuffrida, N. Valero, J. Mosquera, A. Duran, F. Arocha, B. Chacín, L.M. Espina, J.
 Gotera, J. Bermudez, A. Mavarez, M. Alvarez-Mon, Increased Systemic
 Cytokine/Chemokine Expression in Asthmatic and Non-asthmatic Patients with
 Bacterial, Viral or Mixed Lung Infection, Scand. J. Immunol. 85 (2017) 280–290.
 https://doi.org/10.1111/sji.12532.
- [109]Y. Kama, M. Kato, Y. Yamada, T. Koike, K. Suzuki, M. Enseki, H. Tabata, K. Hirai, H.
 Mochizuki, The Suppressive Role of Streptococcus pneumoniae Colonization in Acute

- 1326 Exacerbations of Childhood Bronchial Asthma, Int. Arch. Allergy Immunol. 181 (2020) 1327 191–199. https://doi.org/10.1159/000504541.
- [110]S.M. Teo, H.H.F. Tang, D. Mok, L.M. Judd, S.C. Watts, K. Pham, B.J. Holt, M. Kusel,
 M. Serralha, N. Troy, Y.A. Bochkov, K. Grindle, R.F. Lemanske, S.L. Johnston, J.E.
 Gern, P.D. Sly, P.G. Holt, K.E. Holt, M. Inouye, Airway Microbiota Dynamics Uncover a
 Critical Window for Interplay of Pathogenic Bacteria and Allergy in Childhood
 Respiratory Disease, Cell Host Microbe. 24 (2018) 341-352.e5.
- 1333 https://doi.org/10.1016/j.chom.2018.08.005.
- [111] J.M. Mansbach, P.N. Luna, C.A. Shaw, K. Hasegawa, J.F. Petrosino, P.A. Piedra, A.F.
 Sullivan, J.A. Espinola, C.J. Stewart, C.A. Camargo, Increased Moraxella and
 Streptococcus species abundance after severe bronchiolitis is associated with
 recurrent wheezing, J. Allergy Clin. Immunol. 145 (2020) 518-527.e8.
 https://doi.org/10.1016/j.jaci.2019.10.034.
- [112]L. Cuthbertson, S.W.C. Oo, M.J. Cox, S.-K. Khoo, D.W. Cox, G. Chidlow, K. Franks, F.
 Prastanti, M.L. Borland, J.E. Gern, D.W. Smith, J.A. Bizzintino, I.A. Laing, P.N. Le
 Souëf, M.F. Moffatt, W.O.C. Cookson, Viral respiratory infections and the
 oropharyngeal bacterial microbiota in acutely wheezing children, PLoS ONE. 14 (2019)
 e0223990. https://doi.org/10.1371/journal.pone.0223990.
- [113]Y. Zhou, D. Jackson, L.B. Bacharier, D. Mauger, H. Boushey, M. Castro, J. Durack, Y.
 Huang, R.F. Lemanske, G.A. Storch, G.M. Weinstock, K. Wylie, R. Covar, A.M.
 Fitzpatrick, W. Phipatanakul, R.G. Robison, A. Beigelman, The upper-airway microbiota
 and loss of asthma control among asthmatic children, Nat Commun. 10 (2019) 5714.
 https://doi.org/10.1038/s41467-019-13698-x.
- [114]M.İ. Abdel-Aziz, P. Brinkman, S.J.H. Vijverberg, A.H. Neerincx, J.H. Riley, S. Bates, S.
 Hashimoto, N.Z. Kermani, K.F. Chung, R. Djukanovic, S.-E. Dahlén, I.M. Adcock, P.H.
 Howarth, P.J. Sterk, A.D. Kraneveld, A.H. Maitland-van der Zee, U-BIOPRED Study
 Group, Sputum microbiome profiles identify severe asthma phenotypes of relative
 stability at 12-18 months, J. Allergy Clin. Immunol. (2020).
 https://doi.org/10.1016/j.jaci.2020.04.018.
- [115]G. Lezmi, J. de Blic, Assessment of airway inflammation and remodeling in children
 with severe asthma: The next challenge, Pediatr. Pulmonol. 53 (2018) 1171–1173.
 https://doi.org/10.1002/ppul.24051.
- [116]G. Lezmi, A. Deschildre, R. Abou Taam, M. Fayon, S. Blanchon, F. Troussier, P.
 Mallinger, B. Mahut, P. Gosset, J. de Blic, Remodelling and inflammation in
 preschoolers with severe recurrent wheeze and asthma outcome at school age, Clin.
 Exp. Allergy. 48 (2018) 806–813. https://doi.org/10.1111/cea.13143.
- 1362 [117]A. Ray, J.K. Kolls, Neutrophilic Inflammation in Asthma and Association with Disease 1363 Severity, Trends Immunol. 38 (2017) 942–954. https://doi.org/10.1016/j.it.2017.07.003.
- [118]C.J. Bossley, L. Fleming, A. Gupta, N. Regamey, J. Frith, T. Oates, L. Tsartsali, C.M.
 Lloyd, A. Bush, S. Saglani, Pediatric severe asthma is characterized by eosinophilia
 and remodeling without T(H)2 cytokines, J. Allergy Clin. Immunol. 129 (2012) 974982.e13. https://doi.org/10.1016/j.jaci.2012.01.059.
- [119]L. Fleming, L. Tsartsali, N. Wilson, N. Regamey, A. Bush, Sputum inflammatory
 phenotypes are not stable in children with asthma, Thorax. 67 (2012) 675–681.
 https://doi.org/10.1136/thoraxjnl-2011-201064.
- [120] J.R. Grunwell, S.T. Stephenson, R. Tirouvanziam, L.A.S. Brown, M.R. Brown, A.M.
 Fitzpatrick, Children with Neutrophil-Predominant Severe Asthma Have
 Proinflammatory Neutrophils With Enhanced Survival and Impaired Clearance, J
 Allergy Clin Immunol Pract. 7 (2019) 516-525.e6.
 https://doi.org/10.1016/i.jaip.2018.08.024
- 1375 https://doi.org/10.1016/j.jaip.2018.08.024.
- [121]S.L. Taylor, L.E.X. Leong, J.M. Choo, S. Wesselingh, I.A. Yang, J.W. Upham, P.N.
 Reynolds, S. Hodge, A.L. James, C. Jenkins, M.J. Peters, M. Baraket, G.B. Marks,
 P.G. Gibson, J.L. Simpson, G.B. Rogers, Inflammatory phenotypes in patients with
 severe asthma are associated with distinct airway microbiology, J. Allergy Clin.
 Immunol. 141 (2018) 94-103.e15. https://doi.org/10.1016/j.jaci.2017.03.044.

- [122]J. Zhu, S.D. Message, Y. Qiu, P. Mallia, T. Kebadze, M. Contoli, C.K. Ward, E.S.
 Barnathan, M.A. Mascelli, O.M. Kon, A. Papi, L.A. Stanciu, P.K. Jeffery, S.L. Johnston,
 Airway inflammation and illness severity in response to experimental rhinovirus
 infection in asthma, Chest. 145 (2014) 1219–1229. https://doi.org/10.1378/chest.131567.
- [123] T. Guiddir, P. Saint-Pierre, E. Purenne-Denis, N. Lambert, Y. Laoudi, R. Couderc, R.
 Gouvis-Echraghi, F. Amat, J. Just, Neutrophilic Steroid-Refractory Recurrent Wheeze
 and Eosinophilic Steroid-Refractory Asthma in Children, J Allergy Clin Immunol Pract. 5
 (2017) 1351-1361.e2. https://doi.org/10.1016/j.jaip.2017.02.003.
- [124]K. Nakagome, M. Nagata, Involvement and Possible Role of Eosinophils in Asthma
 Exacerbation, Front Immunol. 9 (2018) 2220.
- 1392 https://doi.org/10.3389/fimmu.2018.02220.
- [125]L. Hatchwell, A. Collison, J. Girkin, K. Parsons, J. Li, J. Zhang, S. Phipps, D. Knight,
 N.W. Bartlett, S.L. Johnston, P.S. Foster, P.A.B. Wark, J. Mattes, Toll-like receptor 7
 governs interferon and inflammatory responses to rhinovirus and is suppressed by IL-5induced lung eosinophilia, Thorax. 70 (2015) 854–861.
 https://doi.org/10.1136/thoraxinl-2014-205465.
- [126] L. Fleming, N. Wilson, N. Regamey, A. Bush, Use of sputum eosinophil counts to guide
 management in children with severe asthma, Thorax. 67 (2012) 193–198.
- 1400 https://doi.org/10.1136/thx.2010.156836.
- [127]W.G. Teague, Blood Eosinophilia May Not Adequately Estimate Lung Fluid
 Eosinophilia in Childhood Asthma, J Allergy Clin Immunol Pract. 7 (2019) 2497–2498.
 https://doi.org/10.1016/j.jaip.2019.05.008.
- [128] P. Nagakumar, F. Puttur, L.G. Gregory, L. Denney, L. Fleming, A. Bush, C.M. Lloyd, S.
 Saglani, Pulmonary type-2 innate lymphoid cells in paediatric severe asthma:
 phenotype and response to steroids, Eur. Respir. J. 54 (2019).
 https://doi.org/10.1183/13993003.01809-2018.
- [129]S. Saglani, A. Bush, W. Carroll, S. Cunningham, L. Fleming, E. Gaillard, A. Gupta, C.
 Murray, P. Nagakumar, J. Paton, G. Roberts, P. Seddon, I. Sinha, Biologics for
 paediatric severe asthma: trick or TREAT?, Lancet Respir Med. 7 (2019) 294–296.
 https://doi.org/10.1016/S2213-2600(19)30045-1.
- [130] J. Just, R. Gouvis-Echraghi, R. Couderc, N. Guillemot-Lambert, P. Saint-Pierre, Novel
 severe wheezy young children phenotypes: boys atopic multiple-trigger and girls
 nonatopic uncontrolled wheeze, J. Allergy Clin. Immunol. 130 (2012) 103-110.e8.
 https://doi.org/10.1016/j.jaci.2012.02.041.
- [131]L.B. Bacharier, A. Beigelman, A. Calatroni, D.J. Jackson, P.J. Gergen, G.T. O'Connor,
 M. Kattan, R.A. Wood, M.T. Sandel, S.V. Lynch, K.E. Fujimura, D.W. Fadrosh, C.A.
 Santee, H. Boushey, C.M. Visness, J.E. Gern, NIAID sponsored Inner-City Asthma
 Consortium, Longitudinal Phenotypes of Respiratory Health in a High-Risk Urban Birth
 Cohort, Am. J. Respir. Crit. Care Med. 199 (2019) 71–82.
- 1421 https://doi.org/10.1164/rccm.201801-0190OC.
- [132] L.S. Subrata, J. Bizzintino, E. Mamessier, A. Bosco, K.L. McKenna, M.E. Wikström, J.
 Goldblatt, P.D. Sly, B.J. Hales, W.R. Thomas, I.A. Laing, P.N. LeSouëf, P.G. Holt,
 Interactions between Innate Antiviral and Atopic Immunoinflammatory Pathways
 Precipitate and Sustain Asthma Exacerbations in Children, The Journal of Immunology.
 183 (2009) 2793–2800. https://doi.org/10.4049/jimmunol.0900695.
- [133] A. Custovic, H.-J. Sonntag, I.E. Buchan, D. Belgrave, A. Simpson, M.C.F. Prosperi,
 Evolution pathways of IgE responses to grass and mite allergens throughout childhood,
 J. Allergy Clin. Immunol. 136 (2015) 1645-1652.e8.
 https://doi.org/10.1016/j.jppi.2015.02.041
- 1430 https://doi.org/10.1016/j.jaci.2015.03.041.
- [134]S. Illi, E. von Mutius, S. Lau, B. Niggemann, C. Grüber, U. Wahn, Multicentre Allergy
 Study (MAS) group, Perennial allergen sensitisation early in life and chronic asthma in
 children: a birth cohort study, Lancet. 368 (2006) 763–770.
- 1434 https://doi.org/10.1016/S0140-6736(06)69286-6.

- [135]S. Gabet, J. Just, R. Couderc, J. Bousquet, N. Seta, I. Momas, Early polysensitization
 is associated with allergic multimorbidity in PARIS birth cohort infants, Pediatr Allergy
 Immunol. 27 (2016) 831–837. https://doi.org/10.1111/pai.12622.
- [136] A.J. Hose, M. Depner, S. Illi, S. Lau, T. Keil, U. Wahn, O. Fuchs, P.I. Pfefferle, E.
 Schmaußer-Hechfellner, J. Genuneit, R. Lauener, A.M. Karvonen, C. Roduit, J.-C.
 Dalphin, J. Riedler, J. Pekkanen, E. von Mutius, M.J. Ege, MAS, PASTURE study
 groups, Latent class analysis reveals clinically relevant atopy phenotypes in 2 birth
 cohorts, J. Allergy Clin. Immunol. 139 (2017) 1935-1945.e12.
- 1443 https://doi.org/10.1016/j.jaci.2016.08.046.
- [137]P.G. Holt, D. Mok, D. Panda, L. Renn, G. Fabozzi, N.H. deKlerk, M.M.H. Kusel, M.
 Serralha, E.M. Hollams, B.J. Holt, P.D. Sly, R.L. Rabin, Developmental regulation of
 type 1 and type 3 interferon production and risk for infant infections and asthma
 development, J. Allergy Clin. Immunol. 143 (2019) 1176-1182.e5.
 https://doi.org/10.1016/j.jaci.2018.08.035.
- [138]A. Custovic, D. Belgrave, L. Lin, E. Bakhsoliani, A.G. Telcian, R. Solari, C.S. Murray,
 R.P. Walton, J. Curtin, M.R. Edwards, A. Simpson, M. Rattray, S.L. Johnston, Cytokine
 Responses to Rhinovirus and Development of Asthma, Allergic Sensitization, and
 Respiratory Infections during Childhood, Am. J. Respir. Crit. Care Med. 197 (2018)
 1265–1274. https://doi.org/10.1164/rccm.201708-1762OC.
- [139]B.M. Donovan, L. Bastarache, K.N. Turi, M.M. Zutter, T.V. Hartert, The current state of
 omics technologies in the clinical management of asthma and allergic diseases, Ann.
 Allergy Asthma Immunol. 123 (2019) 550–557.
 https://doi.org/10.1016/i.anai.2019.08.460.
- [140]N. Farzan, S.J. Vijverberg, M. Kabesch, P.J. Sterk, A.H. Maitland-van der Zee, The use
 of pharmacogenomics, epigenomics, and transcriptomics to improve childhood asthma
 management: Where do we stand?, Pediatr. Pulmonol. 53 (2018) 836–845.
 https://doi.org/10.1002/ppul.23976.
- [141]R. Hall, I.P. Hall, I. Sayers, Genetic risk factors for the development of pulmonary
 disease identified by genome-wide association, Respirology. 24 (2019) 204–214.
 https://doi.org/10.1111/resp.13436.
- [142] M.F. Moffatt, I.G. Gut, F. Demenais, D.P. Strachan, E. Bouzigon, S. Heath, E. von
 Mutius, M. Farrall, M. Lathrop, W.O.C.M. Cookson, GABRIEL Consortium, A largescale, consortium-based genomewide association study of asthma, N. Engl. J. Med.
 363 (2010) 1211–1221. https://doi.org/10.1056/NEJMoa0906312.
- [143]M.C. Altman, M.A. Gill, E. Whalen, D.C. Babineau, B. Shao, A.H. Liu, B. Jepson, R.S.
 Gruchalla, G.T. O'Connor, J.A. Pongracic, C.M. Kercsmar, G.K. Khurana Hershey,
 E.M. Zoratti, C.C. Johnson, S.J. Teach, M. Kattan, L.B. Bacharier, A. Beigelman, S.M.
 Sigelman, S. Presnell, J.E. Gern, P.J. Gergen, L.M. Wheatley, A. Togias, W.W. Busse,
 Jackson, Transcriptome networks identify mechanisms of viral and nonviral
 asthma exacerbations in children, Nat. Immunol. 20 (2019) 637–651.
- 1475 https://doi.org/10.1038/s41590-019-0347-8.
- [144] A. Bosco, S. Ehteshami, S. Panyala, F.D. Martinez, Interferon regulatory factor 7 is a
 major hub connecting interferon-mediated responses in virus-induced asthma
 exacerbations in vivo, J. Allergy Clin. Immunol. 129 (2012) 88–94.
 https://doi.org/10.1016/i.jaci.2011.10.038
- 1479 https://doi.org/10.1016/j.jaci.2011.10.038.
- [145]T. Aoki, Y. Matsumoto, K. Hirata, K. Ochiai, M. Okada, K. Ichikawa, M. Shibasaki, T.
 Arinami, R. Sumazaki, E. Noguchi, Expression profiling of genes related to asthma
 exacerbations, Clin. Exp. Allergy. 39 (2009) 213–221. https://doi.org/10.1111/j.13652222.2008.03186.x.
- [146]H. Persson, A.T. Kwon, J.A. Ramilowski, G. Silberberg, C. Söderhäll, C. OrsmarkPietras, B. Nordlund, J.R. Konradsen, M.J.L. de Hoon, E. Melén, Y. Hayashizaki, G.
 Hedlin, J. Kere, C.O. Daub, Transcriptome analysis of controlled and therapy-resistant
 childhood asthma reveals distinct gene expression profiles, J. Allergy Clin. Immunol.
 136 (2015) 638–648. https://doi.org/10.1016/j.jaci.2015.02.026.

- [147] A.M. Fitzpatrick, Y. Park, L.A.S. Brown, D.P. Jones, Children with severe asthma have
 unique oxidative stress-associated metabolomic profiles, J. Allergy Clin. Immunol. 133
 (2014) 258-261.e1–8. https://doi.org/10.1016/j.jaci.2013.10.012.
- [148] C.-Y. Chiu, M.-L. Cheng, M.-H. Chiang, Y.-L. Kuo, M.-H. Tsai, C.-C. Chiu, G. Lin, Gut
 microbial-derived butyrate is inversely associated with IgE responses to allergens in
 childhood asthma, Pediatr Allergy Immunol. 30 (2019) 689–697.
 https://doi.org/10.1111/pai.13096.
- [149] K.E. Duffy, R.J. Lamb, L.R. San Mateo, J.L. Jordan, G. Canziani, M. Brigham-Burke, J.
 Korteweg, M. Cunningham, H.S. Beck, J. Carton, J. Giles-Komar, C. Duchala, R.T.
 Sarisky, M.L. Mbow, Down modulation of human TLR3 function by a monoclonal
 antibody, Cell. Immunol. 248 (2007) 103–114.
- 1500 https://doi.org/10.1016/j.cellimm.2007.10.002.
- [150] P.E. Silkoff, S. Flavin, R. Gordon, M.J. Loza, P.J. Sterk, R. Lutter, Z. Diamant, R.B.
 Turner, B.J. Lipworth, D. Proud, D. Singh, A. Eich, V. Backer, J.E. Gern, C. Herzmann,
 S.A. Halperin, T.T. Mensinga, A.M. Del Vecchio, P. Branigan, L. San Mateo, F.
 Baribaud, E.S. Barnathan, S.L. Johnston, Toll-like receptor 3 blockade in rhinovirusinduced experimental asthma exacerbations: A randomized controlled study, J. Allergy
 Clin. Immunol. 141 (2018) 1220–1230. https://doi.org/10.1016/j.jaci.2017.06.027.
- [151] J.P. Wong, M.E. Christopher, S. Viswanathan, N. Karpoff, X. Dai, D. Das, L.Q. Sun, M.
 Wang, A.M. Salazar, Activation of toll-like receptor signaling pathway for protection against influenza virus infection, Vaccine. 27 (2009) 3481–3483. https://doi.org/10.1016/j.vaccine.2009.01.048.
- [152]X. Tian, F. Xu, W.Y. Lung, C. Meyerson, A.A. Ghaffari, G. Cheng, J.C. Deng, Poly I:C
 enhances susceptibility to secondary pulmonary infections by gram-positive bacteria,
 PLoS ONE. 7 (2012) e41879. https://doi.org/10.1371/journal.pone.0041879.
- [153]D. Torres, A. Dieudonné, B. Ryffel, E. Vilain, M. Si-Tahar, M. Pichavant, P. Lassalle, F.
 Trottein, P. Gosset, Double-stranded RNA exacerbates pulmonary allergic reaction
 through TLR3: implication of airway epithelium and dendritic cells, J. Immunol. 185
 (2010) 451–459. https://doi.org/10.4049/jimmunol.0902833.
- [154] E.M. Abrams, H.H. Raissy, Emerging Therapies in the Treatment of Early Childhood
 Wheeze, Pediatr Allergy Immunol Pulmonol. 32 (2019) 78–80.
 https://doi.org/10.1089/ped.2019.1043.
- [155]M. Cazzola, S. Anapurapu, C.P. Page, Polyvalent mechanical bacterial lysate for the
 prevention of recurrent respiratory infections: a meta-analysis, Pulm Pharmacol Ther.
 25 (2012) 62–68. https://doi.org/10.1016/j.pupt.2011.11.002.
- [156] M.D. Gutiérrez-Tarango, A. Berber, Safety and Efficacy of Two Courses of OM-85 BV
 in the Prevention of Respiratory Tract Infections in Children During 12 Months, CHEST.
 119 (2001) 1742–1748. https://doi.org/10.1378/chest.119.6.1742.
- [157] U.B. Schaad, OM-85 BV, an immunostimulant in pediatric recurrent respiratory tract
 infections: a systematic review, World J Pediatr. 6 (2010) 5–12.
 https://doi.org/10.1007/s12519-010-0001-x.
- [158] J.P. Collet, T. Ducruet, M.S. Kramer, J. Haggerty, D. Floret, J.J. Chomel, F. Durr,
 Stimulation of nonspecific immunity to reduce the risk of recurrent infections in children
 attending day-care centers. The Epicrèche Research Group, Pediatr. Infect. Dis. J. 12
 (1993) 648–652. https://doi.org/10.1097/00006454-199308000-00005.
- 1534 [159] J. Yin, B. Xu, X. Zeng, K. Shen, Broncho-Vaxom in pediatric recurrent respiratory tract
 1535 infections: A systematic review and meta-analysis, Int. Immunopharmacol. 54 (2018)
 1536 198–209. https://doi.org/10.1016/j.intimp.2017.10.032.
- [160] R. Ricci, C. Palmero, G. Bazurro, A.M. Riccio, V. Garelli, E. Di Marco, C. Cirillo, F.
 Braido, G.W. Canonica, G. Melioli, The administration of a polyvalent mechanical
 bacterial lysate in elderly patients with COPD results in serological signs of an efficient
 immune response associated with a reduced number of acute episodes, Pulm
 Pharmacol Ther. 27 (2014) 109–113. https://doi.org/10.1016/j.pupt.2013.05.006.
- [161]A. Emeryk, M. Bartkowiak-Emeryk, Z. Raus, F. Braido, G. Ferlazzo, G. Melioli,
 Mechanical bacterial lysate administration prevents exacerbation in allergic asthmatic

- 1544 children-The EOLIA study, Pediatr Allergy Immunol. 29 (2018) 394–401.
- 1545 https://doi.org/10.1111/pai.12894.
- [162] C.H. Razi, K. Harmanci, A. Abacı, O. Özdemir, S. Hızlı, R. Renda, F. Keskin, The
 immunostimulant OM-85 BV prevents wheezing attacks in preschool children, J. Allergy
 Clin. Immunol. 126 (2010) 763–769. https://doi.org/10.1016/j.jaci.2010.07.038.
- [163]Oral Bacterial Extract for the Prevention of Wheezing Lower Respiratory Tract Illness Full Text View ClinicalTrials.gov, (n.d.).
- 1551 https://clinicaltrials.gov/ct2/show/NCT02148796 (accessed February 9, 2020).
- 1552 [164]M. Huber, H. Mossmann, W.G. Bessler, Th1-orientated immunological properties of the 1553 bacterial extract OM-85-BV, Eur. J. Med. Res. 10 (2005) 209–217.
- [165]C. Zelle-Rieser, R. Ramoner, G. Bartsch, M. Thurnher, A clinically approved oral
 vaccine against pneumotropic bacteria induces the terminal maturation of CD83+
 immunostimulatory dendritic cells, Immunol. Lett. 76 (2001) 63–67.
 https://doi.org/10.1016/s0165-2478(00)00326-6.
- 1558 [166]J. Mauël, Stimulation of Immunoprotective Mechanisms by OM-85 BV, RES. 61 (1994) 1559 8–15. https://doi.org/10.1159/000196372.
- [167] F.H. Sousa, V. Casanova, F. Findlay, C. Stevens, P. Svoboda, J. Pohl, L. Proudfoot,
 P.G. Barlow, Cathelicidins display conserved direct antiviral activity towards rhinovirus,
 Peptides. 95 (2017) 76–83. https://doi.org/10.1016/j.peptides.2017.07.013.
- [168]Y. Li, X. Chu, C. Liu, W. Huang, Y. Yao, Y. Xia, P. Sun, Q. Long, X. Feng, K. Li, X.
 Yang, H. Bai, W. Sun, Y. Ma, Exogenous murine antimicrobial peptide CRAMP
 significantly exacerbates Ovalbumin-induced airway inflammation but ameliorates
 oxazolone-induced intestinal colitis in BALB/c mice, Hum Vaccin Immunother. 14
 (2018) 146–158. https://doi.org/10.1080/21645515.2017.1386823.
- [169] A.G. Telcian, M.T. Zdrenghea, M.R. Edwards, V. Laza-Stanca, P. Mallia, S.L.
 Johnston, L.A. Stanciu, Vitamin D increases the antiviral activity of bronchial epithelial
 cells in vitro, Antiviral Res. 137 (2017) 93–101.
 https://doi.org/10.1016/j.antiviral.2016.11.004.
- [170] C.L. Greiller, R. Suri, D.A. Jolliffe, T. Kebadze, A.G. Hirsman, C.J. Griffiths, S.L.
 Johnston, A.R. Martineau, Vitamin D attenuates rhinovirus-induced expression of intercellular adhesion molecule-1 (ICAM-1) and platelet-activating factor receptor
 (PAFR) in respiratory epithelial cells, J. Steroid Biochem. Mol. Biol. 187 (2019) 152– 159. https://doi.org/10.1016/j.jsbmb.2018.11.013.
- [171]A. Schögler, R.J. Muster, E. Kieninger, C. Casaulta, C. Tapparel, A. Jung, A. Moeller,
 T. Geiser, N. Regamey, M.P. Alves, Vitamin D represses rhinovirus replication in cystic
 fibrosis cells by inducing LL-37, Eur. Respir. J. 47 (2016) 520–530.
 https://doi.org/10.1183/13993003.00665-2015.
- [172] A.R. Martineau, D.A. Jolliffe, R.L. Hooper, L. Greenberg, J.F. Aloia, P. Bergman, G.
 Dubnov-Raz, S. Esposito, D. Ganmaa, A.A. Ginde, E.C. Goodall, C.C. Grant, C.J.
 Griffiths, W. Janssens, I. Laaksi, S. Manaseki-Holland, D. Mauger, D.R. Murdoch, R.
 Neale, J.R. Rees, S. Simpson, I. Stelmach, G.T. Kumar, M. Urashima, C.A. Camargo,
 Vitamin D supplementation to prevent acute respiratory tract infections: systematic
 review and meta-analysis of individual participant data, BMJ. 356 (2017) i6583.
 https://doi.org/10.1136/bmj.i6583.
- [173]M.S. Sandhu, T.B. Casale, The role of vitamin D in asthma, Ann. Allergy Asthma
 Immunol. 105 (2010) 191–199; quiz 200–202, 217.
 https://doi.org/10.1016/j.anai.2010.01.013.
- [174]T.-T. Wang, F.P. Nestel, V. Bourdeau, Y. Nagai, Q. Wang, J. Liao, L. Tavera-Mendoza,
 R. Lin, J.W. Hanrahan, S. Mader, J.H. White, J.H. Hanrahan, Cutting edge: 1,25dihydroxyvitamin D3 is a direct inducer of antimicrobial peptide gene expression, J.
 Immunol. 173 (2004) 2909–2912. https://doi.org/10.4049/jimmunol.173.5.2909.
- [175] V. Kim, R.M. Abreu, D.M. Nakagawa, R.M. Baldassare, F.J. Carrilho, S.K. Ono,
 Pegylated interferon alfa for chronic hepatitis B: systematic review and meta-analysis,
- 1597 J. Viral Hepat. 23 (2016) 154–169. https://doi.org/10.1111/jvh.12418.

- [176]K.J. Lavender, K. Gibbert, K.E. Peterson, E. Van Dis, S. Francois, T. Woods, R.J.
 Messer, A. Gawanbacht, J.A. Müller, J. Münch, K. Phillips, B. Race, M.S. Harper, K.
 Guo, E.J. Lee, M. Trilling, H. Hengel, J. Piehler, J. Verheyen, C.C. Wilson, M.L.
 Santiago, K.J. Hasenkrug, U. Dittmer, Interferon Alpha Subtype-Specific Suppression
 of HIV-1 Infection In Vivo, J. Virol. 90 (2016) 6001–6013.
 https://doi.org/10.1128/JVI.00451-16.
- [177]T.P. Trang, M. Whalen, A. Hilts-Horeczko, S.B. Doernberg, C. Liu, Comparative
 effectiveness of aerosolized versus oral ribavirin for the treatment of respiratory
 syncytial virus infections: A single-center retrospective cohort study and review of the
 literature, Transpl Infect Dis. 20 (2018) e12844. https://doi.org/10.1111/tid.12844.
- [178]S.A. Grim, G.E. Reid, N.M. Clark, Update in the treatment of non-influenza respiratory
 virus infection in solid organ transplant recipients, Expert Opin Pharmacother. 18
 (2017) 767–779. https://doi.org/10.1080/14656566.2017.1322063.
- [179]O. Ruuskanen, M. Waris, L. Kainulainen, Treatment of persistent rhinovirus infection
 with pegylated interferon α2a and ribavirin in patients with hypogammaglobulinemia,
 Clin. Infect. Dis. 58 (2014) 1784–1786. https://doi.org/10.1093/cid/ciu169.
- 1614 [180]S.S.Y. Wong, K.-Y. Yuen, The management of coronavirus infections with particular 1615 reference to SARS, J. Antimicrob. Chemother. 62 (2008) 437–441.
- 1616 https://doi.org/10.1093/jac/dkn243.
- [181]T.M. Becker, S.R. Durrani, Y.A. Bochkov, M.K. Devries, V. Rajamanickam, D.J.
 Jackson, Effect of exogenous interferons on rhinovirus replication and airway
 inflammatory responses, Ann. Allergy Asthma Immunol. 111 (2013) 397–401.
 https://doi.org/10.1016/j.anai.2013.07.029.
- [182] R. Djukanović, T. Harrison, S.L. Johnston, F. Gabbay, P. Wark, N.C. Thomson, R.
 Niven, D. Singh, H.K. Reddel, D.E. Davies, R. Marsden, C. Boxall, S. Dudley, V.
 Plagnol, S.T. Holgate, P. Monk, INTERCIA Study Group, The effect of inhaled IFN-β on
 worsening of asthma symptoms caused by viral infections. A randomized trial, Am. J.
 Respir. Crit. Care Med. 190 (2014) 145–154. https://doi.org/10.1164/rccm.20131222350C.
- [183]C. McCrae, M. Olsson, M. Aurell, C. Lundin, J. Paraskos, A. Cavallin, M. Kjerrulf, K.
 Karlsson, R. Marsden, A. Malmgren, P. Gustafson, T. Harrison, On-Demand Inhaled
 Interferon-Beta 1a for the Prevention of Severa Asthma Exacerbations: Results of the
 INEXAS Phase 2a Study, in: D12. IMMUNOTHERAPY IN LUNG DISEASE, American
 Thoracic Society, 2018: pp. A6165–A6165. https://doi.org/10.1164/ajrccmconference.2018.197.1_MeetingAbstracts.A6165.
- [184] A Study in Asthma Patients to Evaluate Efficacy, Safety and Tolerability of 14 Days
 Once Daily Inhaled Interferon Beta-1a After the Onset of Symptoms of an Upper
 Respiratory Tract Infection Full Text View ClinicalTrials.gov, (n.d.).
 https://clinicaltrials.gov/ct2/show/NCT02491684 (accessed February 4, 2020).
- 1636 Intps://cinicatinals.gov/ci2/snow/Ner/02491064 (accessed rebidary 4, 2020).
 1637 [185]H.J. Thibaut, A.M. De Palma, J. Neyts, Combating enterovirus replication: state-of-theart on antiviral research, Biochem. Pharmacol. 83 (2012) 185–192.
 1639 https://doi.org/10.1016/j.bcp.2011.08.016.
- [186]Effects of Pleconaril Nasal Spray on Common Cold Symptoms and Asthma
 Exacerbations Following Rhinovirus Exposure (Study P04295) Study Results ClinicalTrials.gov, (n.d.). https://clinicaltrials.gov/ct2/show/results/NCT00394914
 (accessed May 3, 2020).
- [187]D.P. Potaczek, S.D. Unger, N. Zhang, S. Taka, S. Michel, N. Akdağ, F. Lan, M. Helfer,
 C. Hudemann, M. Eickmann, C. Skevaki, S. Megremis, A. Sadewasser, B. Alashkar
 Alhamwe, F. Alhamdan, M. Akdis, M.R. Edwards, S.L. Johnston, C.A. Akdis, S. Becker,
 C. Bachert, N.G. Papadopoulos, H. Garn, H. Renz, Development and characterization
 of DNAzyme candidates demonstrating significant efficiency against human
 rhinoviruses, J. Allergy Clin. Immunol. 143 (2019) 1403–1415.
- 1650 https://doi.org/10.1016/j.jaci.2018.07.026.
- [188] A. Bush, Azithromycin is the answer in paediatric respiratory medicine, but what was
 the question?, Paediatr Respir Rev. (2019). https://doi.org/10.1016/j.prrv.2019.07.002.

- [189] F.M. de Benedictis, I. Carloni, R. Guidi, Question 4: Is there a role for antibiotics in
 infantile wheeze?, Paediatr Respir Rev. (2019).
- 1655 https://doi.org/10.1016/j.prrv.2019.11.001.
- [190] P. Li, T. Zhang, Y. Xiao, L. Tian, B. Cui, G. Ji, Y.-Y. Liu, F. Zhang, Timing for the
 second fecal microbiota transplantation to maintain the long-term benefit from the first
 treatment for Crohn's disease, Appl. Microbiol. Biotechnol. 103 (2019) 349–360.
 https://doi.org/10.1007/s00253-018-9447-x.
- 1660 [191]H.T. Wang, Š. Anvari, K. Anagnostou, The Role of Probiotics in Preventing Allergic 1661 Disease, Children (Basel). 6 (2019). https://doi.org/10.3390/children6020024.
- [192]C. Cuello-Garcia, A. Fiocchi, R. Pawankar, J.J. Yepes-Nuñez, G.P. Morgano, Y.
 Zhang, A. Agarwal, S. Gandhi, L. Terracciano, H.J. Schünemann, J.L. Brozek,
 Prebiotics for the prevention of allergies: A systematic review and meta-analysis of
 randomized controlled trials, Clin. Exp. Allergy. 47 (2017) 1468–1477.
 https://doi.org/10.1111/cea.13042.
- [193]A.N. Thorburn, C.I. McKenzie, S. Shen, D. Stanley, L. Macia, L.J. Mason, L.K. Roberts,
 C.H.Y. Wong, R. Shim, R. Robert, N. Chevalier, J.K. Tan, E. Mariño, R.J. Moore, L.
 Wong, M.J. McConville, D.L. Tull, L.G. Wood, V.E. Murphy, J. Mattes, P.G. Gibson,
 C.R. Mackay, Evidence that asthma is a developmental origin disease influenced by
 maternal diet and bacterial metabolites, Nat Commun. 6 (2015) 7320.
- 1672 https://doi.org/10.1038/ncomms8320.
- 1673 [194]T.J. Schuijt, J.M. Lankelma, B.P. Scicluna, F. de Sousa e Melo, J.J.T.H. Roelofs, J.D.
 1674 de Boer, A.J. Hoogendijk, R. de Beer, A. de Vos, C. Belzer, W.M. de Vos, T. van der
 1675 Poll, W.J. Wiersinga, The gut microbiota plays a protective role in the host defence
 1676 against pneumococcal pneumonia, Gut. 65 (2016) 575–583.
 1677 https://doi.org/10.1136/gutjnl-2015-309728.
- [195] V. Sencio, A. Barthelemy, L.P. Tavares, M.G. Machado, D. Soulard, C. Cuinat, C.M.
 Queiroz-Junior, M.-L. Noordine, S. Salomé-Desnoulez, L. Deryuter, B. Foligné, C.
 Wahl, B. Frisch, A.T. Vieira, C. Paget, G. Milligan, T. Ulven, I. Wolowczuk, C. Faveeuw,
 R. Le Goffic, M. Thomas, S. Ferreira, M.M. Teixeira, F. Trottein, Gut Dysbiosis during
 Influenza Contributes to Pulmonary Pneumococcal Superinfection through Altered
 Short-Chain Fatty Acid Production, Cell Rep. 30 (2020) 2934-2947.e6.
 https://doi.org/10.1016/j.celrep.2020.02.013.
- 1685 [196]R.L. Brown, R.P. Sequeira, T.B. Clarke, The microbiota protects against respiratory
 infection via GM-CSF signaling, Nat Commun. 8 (2017) 1512.
 https://doi.org/10.1038/s41467-017-01803-x.
- 1688 [197]N. Schwerk, F. Brinkmann, B. Soudah, M. Kabesch, G. Hansen, Wheeze in preschool 1689 age is associated with pulmonary bacterial infection and resolves after antibiotic 1690 therapy, PLoS ONE. 6 (2011) e27913. https://doi.org/10.1371/journal.pone.0027913.
- [198] P. Zimmermann, V.C. Ziesenitz, N. Curtis, N. Ritz, The Immunomodulatory Effects of
 Macrolides-A Systematic Review of the Underlying Mechanisms, Front Immunol. 9
 (2018) 302. https://doi.org/10.3389/fimmu.2018.00302.
- [199]L.B. Bacharier, T.W. Guilbert, D.T. Mauger, S. Boehmer, A. Beigelman, A.M. 1694 Fitzpatrick, D.J. Jackson, S.N. Baxi, M. Benson, C.-A.D. Burnham, M. Cabana, M. 1695 1696 Castro, J.F. Chmiel, R. Covar, M. Daines, J.M. Gaffin, D.A. Gentile, F. Holguin, E. Israel, H.W. Kelly, S.C. Lazarus, R.F. Lemanske, N. Ly, K. Meade, W. Morgan, J. Moy, 1697 T. Olin, S.P. Peters, W. Phipatanakul, J.A. Pongracic, H.H. Raissy, K. Ross, W.J. 1698 Sheehan, C. Sorkness, S.J. Szefler, W.G. Teague, S. Thyne, F.D. Martinez, Early 1699 Administration of Azithromycin and Prevention of Severe Lower Respiratory Tract 1700 Illnesses in Preschool Children With a History of Such Illnesses: A Randomized Clinical 1701 Trial, JAMA. 314 (2015) 2034–2044. https://doi.org/10.1001/jama.2015.13896. 1702
- 1703 [200] J. Stokholm, B.L. Chawes, N.H. Vissing, E. Bjarnadóttir, T.M. Pedersen, R.K. Vinding,
 1704 A.-M.M. Schoos, H.M. Wolsk, S. Thorsteinsdóttir, H.W. Hallas, L. Arianto, S. Schjørring,
 1705 K.A. Krogfelt, T.K. Fischer, C.B. Pipper, K. Bønnelykke, H. Bisgaard, Azithromycin for
 1706 episodes with asthma-like symptoms in young children aged 1-3 years: a randomised,

- double-blind, placebo-controlled trial, Lancet Respir Med. 4 (2016) 19–26.
- 1708 https://doi.org/10.1016/S2213-2600(15)00500-7.
- [201] P.J. Mandhane, P. Paredes Zambrano de Silbernagel, Y.N. Aung, J. Williamson, B.E.
 Lee, S. Spier, M. Noseworthy, W.R. Craig, D.W. Johnson, Treatment of preschool
 children presenting to the emergency department with wheeze with azithromycin: A
 placebo-controlled randomized trial, PLoS ONE. 12 (2017) e0182411.
- 1713 https://doi.org/10.1371/journal.pone.0182411.
- [202]M.A. Pincheira, L.B. Bacharier, J.A. Castro-Rodriguez, Efficacy of Macrolides on Acute
 Asthma or Wheezing Exacerbations in Children with Recurrent Wheezing: A
 Systematic Review and Meta-analysis, Paediatr Drugs. (2020).
- 1717 https://doi.org/10.1007/s40272-019-00371-5.
- [203]G.G. Brusselle, C. Vanderstichele, P. Jordens, R. Deman, H. Slabbynck, V. Ringoet, G.
 Verleden, I.K. Demedts, K. Verhamme, A. Delporte, B. Demeyere, G. Claeys, J.
 Boelens, E. Padalko, J. Verschakelen, G. Van Maele, E. Deschepper, G.F.P. Joos,
 Azithromycin for prevention of exacerbations in severe asthma (AZISAST): a
 multicentre randomised double-blind placebo-controlled trial, Thorax. 68 (2013) 322–
 329. https://doi.org/10.1136/thoraxinl-2012-202698.
- [204] P.G. Gibson, I.A. Yang, J.W. Upham, P.N. Reynolds, S. Hodge, A.L. James, C.
 Jenkins, M.J. Peters, G.B. Marks, M. Baraket, H. Powell, S.L. Taylor, L.E.X. Leong,
 G.B. Rogers, J.L. Simpson, Effect of azithromycin on asthma exacerbations and quality
 of life in adults with persistent uncontrolled asthma (AMAZES): a randomised, doubleblind, placebo-controlled trial, Lancet. 390 (2017) 659–668.
- 1729 https://doi.org/10.1016/S0140-6736(17)31281-3.
- [205] D.L. Hahn, M. Grasmick, S. Hetzel, S. Yale, AZMATICS (AZithroMycin-Asthma Trial In Community Settings) Study Group, Azithromycin for bronchial asthma in adults: an effectiveness trial, J Am Board Fam Med. 25 (2012) 442–459.
 https://doi.org/10.2122/jabfm.2012.04.110209
- 1733 https://doi.org/10.3122/jabfm.2012.04.110309.
- [206]S.A. Hiles, V.M. McDonald, M. Guilhermino, G.G. Brusselle, P.G. Gibson, Does
 maintenance azithromycin reduce asthma exacerbations? An individual participant data
 meta-analysis, Eur. Respir. J. 54 (2019). https://doi.org/10.1183/13993003.013812019.
- [207] J.L. Simpson, H. Powell, M.J. Boyle, R.J. Scott, P.G. Gibson, Clarithromycin targets
 neutrophilic airway inflammation in refractory asthma, Am. J. Respir. Crit. Care Med.
 1770 177 (2008) 148–155. https://doi.org/10.1164/rccm.200707-1134OC.
- [208]M. Yamaya, K. Nomura, K. Arakawa, M. Sugawara, X. Deng, N. Lusamba Kalonji, H.
 Nishimura, M. Yamada, R. Nagatomi, T. Kawase, Clarithromycin decreases rhinovirus
 replication and cytokine production in nasal epithelial cells from subjects with bronchial
 asthma: effects on IL-6, IL-8 and IL-33, Arch. Pharm. Res. (2017).
 https://doi.org/10.1007/s12272-017-0950-x.
- [209] V. Gielen, S.L. Johnston, M.R. Edwards, Azithromycin induces anti-viral responses in
 bronchial epithelial cells, Eur. Respir. J. 36 (2010) 646–654.
 https://doi.org/10.1183/09031936.00095809.
- [210] W.W. Busse, W.J. Morgan, P.J. Gergen, H.E. Mitchell, J.E. Gern, A.H. Liu, R.S.
 [210] W.W. Busse, W.J. Morgan, P.J. Gergen, H.E. Mitchell, J.E. Gern, A.H. Liu, R.S.
 [210] W.W. Busse, W.J. Morgan, P.J. Gergen, H.E. Mitchell, J.E. Gern, A.H. Liu, R.S.
 [210] W.W. Busse, W.J. Morgan, P.J. Gergen, H.E. Mitchell, J.E. Gern, A.H. Liu, R.S.
 [210] Gruchalla, M. Kattan, S.J. Teach, J.A. Pongracic, J.F. Chmiel, S.F. Steinbach, A.
 [210] Calatroni, A. Togias, K.M. Thompson, S.J. Szefler, C.A. Sorkness, Randomized trial of
 [210] omalizumab (anti-IgE) for asthma in inner-city children, N. Engl. J. Med. 364 (2011)
 [210] 1005–1015. https://doi.org/10.1056/NEJMoa1009705.
- [211]A. Deschildre, C. Marguet, J. Salleron, I. Pin, J.-L. Rittié, J. Derelle, R.A. Taam, M.
 Fayon, J. Brouard, J.C. Dubus, D. Siret, L. Weiss, G. Pouessel, L. Beghin, J. Just, Addon omalizumab in children with severe allergic asthma: a 1-year real life survey, Eur.
 Respir. J. 42 (2013) 1224–1233. https://doi.org/10.1183/09031936.00149812.
- [212]A. Esquivel, W.W. Busse, A. Calatroni, A.G. Togias, K.G. Grindle, Y.A. Bochkov, R.S.
 Gruchalla, M. Kattan, C.M. Kercsmar, G. Khurana Hershey, H. Kim, P. Lebeau, A.H.
 Liu, S.J. Szefler, S.J. Teach, J.B. West, J. Wildfire, J.A. Pongracic, J.E. Gern, Effects of
 Omalizumab on Rhinovirus Infections, Illnesses, and Exacerbations of Asthma, Am. J.

- 1762
 Respir. Crit. Care Med. 196 (2017) 985–992. https://doi.org/10.1164/rccm.201701

 1763
 0120OC.
- [213] D.B. Kantor, M.C. McDonald, N. Stenquist, B.J. Schultz, C.D. Smallwood, K.A. Nelson,
 W. Phipatanakul, J.N. Hirschhorn, Omalizumab Is Associated with Reduced Acute
 Severity of Rhinovirus-triggered Asthma Exacerbation, Am. J. Respir. Crit. Care Med.
 194 (2016) 1552–1555. https://doi.org/10.1164/rccm.201606-1145LE.
- [214] P.W. Heymann, T.A. Platts-Mills, J.A. Woodfolk, L. Borish, D.D. Murphy, H.T. Carper,
 M.R. Conaway, J.W. Steinke, L. Muehling, T.W. Gerald, J.L. Kennedy, A.-M. Irani, M.D.
 McGraw, S.V. Early, L.M. Wheatley, A.P. Adams, R.B. Turner, Understanding the
 Asthmatic Response to an Experimental Rhinovirus Infection: Exploring the Effects of
 Blocking IgE, J. Allergy Clin. Immunol. (2020).
- 1773 https://doi.org/10.1016/j.jaci.2020.01.035.
- [215]S.J. Teach, M.A. Gill, A. Togias, C.A. Sorkness, S.J. Arbes, A. Calatroni, J.J. Wildfire,
 P.J. Gergen, R.T. Cohen, J.A. Pongracic, C.M. Kercsmar, G.K. Khurana Hershey, R.S.
 Gruchalla, A.H. Liu, E.M. Zoratti, M. Kattan, K.A. Grindle, J.E. Gern, W.W. Busse, S.J.
 Szefler, Preseasonal treatment with either omalizumab or an inhaled corticosteroid
 boost to prevent fall asthma exacerbations, J. Allergy Clin. Immunol. 136 (2015) 1476–
 1485. https://doi.org/10.1016/j.jaci.2015.09.008.
- [216]M.A. Gill, A.H. Liu, A. Calatroni, R.Z. Krouse, B. Shao, A. Schiltz, J.E. Gern, A. Togias,
 W.W. Busse, Enhanced plasmacytoid dendritic cell antiviral responses after
 omalizumab, J. Allergy Clin. Immunol. 141 (2018) 1735-1743.e9.
 https://doi.org/10.1016/j.jaci.2017.07.035.
- 1784 [217] J. Just, A. Deschildre, S. Lejeune, F. Amat, New perspectives of childhood asthma
 1785 treatment with biologics, Pediatr Allergy Immunol. 30 (2019) 159–171.
 1786 https://doi.org/10.1111/pai.13007.
- [218] A. Gupta, M. Ikeda, B. Geng, J. Azmi, R.G. Price, E.S. Bradford, S.W. Yancey, J.
 Steinfeld, Long-term safety and pharmacodynamics of mepolizumab in children with
 severe asthma with an eosinophilic phenotype, J. Allergy Clin. Immunol. 144 (2019)
 1336-1342.e7. https://doi.org/10.1016/j.jaci.2019.08.005.
- 1791 [219]Y.S. Sabogal Piñeros, S.M. Bal, M.A. van de Pol, B.S. Dierdorp, T. Dekker, A. Dijkhuis, 1792 P. Brinkman, K.F. van der Sluijs, A.H. Zwinderman, C.J. Majoor, P.I. Bonta, L.
- 1793 Ravanetti, P.J. Sterk, R. Lutter, Anti-IL-5 in Mild Asthma Alters Rhinovirus-induced
- 1794 Macrophage, B-Cell, and Neutrophil Responses (MATERIAL). A Placebo-controlled,
- 1795 Double-Blind Study, Am. J. Respir. Crit. Care Med. 199 (2019) 508–517.
- 1796 https://doi.org/10.1164/rccm.201803-0461OC.
- 1797





partment	Study	Design and age-group	Main results	
In vitro and ex-vivo				
AEC	Edwards M, et al. 2013 [45]	RV-A16 Children: <u>Severe therapy resistant asthma (STRA)</u> 11y (9- 15y) vs healthy controls	In STRA: lower basal levels of TLR3, reduced RIG-I, MDA5 levels after RV stimulation; lower basal levels of TLR3, reduced RIG-I, MDA5 levels after RV stimulation No association with age, IgE levels, allergen reactivity, BAL/sputum neutrophils and eosinophils, lun function	
AEC	Parsons KS, et al. 2014 [46]	RV-B1 Adults: <u>asthma</u> / Healthy controls	In asthma: no impairment of expression of MDA5, TLR3 and reduced release of CXCL-10	
AEC	Contoli M, et al. 2015 [59]	RV-A16 Pre-treatment with IL-4 and IL-13 prior to infection Adults: atopic rhinitis / non-atopic healthy	In atopic cells: impaired immune response to RV-16 in the presence of IL-4 and IL-13, through inhit of TLR3 expression and signaling (IRF3),	
AEC	Moskwa S, et al. 2018 [61]	PIV-3 / RV-B1 Adults: atopic asthma / non-atopic asthma / healthy controls	<u>Atopic asthma:</u> After PIV3 infection: higher IFN-λ1, IFN- α, IFN- β and IRF7 expressions After RV1B infection: IFN- β higher than in <u>non-atopic asthmatics</u>	
АМ	Rupani H, et al. 2016 [47]	RV-A16 Pre-treatment with Imiquimod (TLR7 agonist) Adults: <u>severe asthma</u> / Healthy controls	In severe asthma: reduced TLR7 expression, inverse correlation with number of exacerbations Increased levels of 3 microRNAs (miR-150, miR-152, and miR-375) Restored TLR7 expression after blocking these microRNAs with anti-miRNA oligonucleotides	
BAL cells PBMC	Sykes A, et al. 2012 [48]	RV-A16 Adults: mild-to-moderate allergic asthma vs healthy controls	In asthma: no impairment of expression of PRR (TLR, RNA-helicases), their adaptor proteins, transcription factors downstream	
In vivo		-	·	
Sputum Serum PBMC	Deschildre A, et al. 2017 [49]	School-age children 8.9 y (6-16y) <u>Atopic asthma:</u> virus vs no virus Severe exacerbation (64% virus, 51% RV) / steady state (8 weeks later) (25% virus, 11% RV) 24% virus at both time	Virus at exacerbation: similar TLR3, RIGI-I and MDA-5 expression on blood monocytes and DC, similar levels of IFN- β , IFN- γ , IFN- λ 1 in sputum and plasma, higher airway IL-5 and eosinophils couthan patients with negative viral PCR Virus at both time: modification in PRR expression/function on PBMC, higher airway neutrophilic inflammation at steady state	
Bronchial	Zhu J, et al. 2019 [67]	<i>In vivo</i> inoculation - RV-A16 Young adults: <u>atopic asthma</u> (23 y \pm 1.4) vs non-atopic control subjects (27 y \pm 2.2)	In asthma: PRR expression (TLR3, MDA5, RIG-I) not deficient at baseline; and induced after RV bo epithelium and sub-epithelium	

	In vitro		
AEC	Wark PA, et al. 2005 [60]	RV-A16 Adults: <u>Asthma</u> vs healthy controls	In asthma: Impaired IFN- β , late virus release and late cell lysis, impaired apoptotic response
AEC	Edwards M, et al. 2013 [45]	RV-A16 Children: <u>Severe therapy resistant asthma (STRA)</u> 11y (9-15y), vs healthy controls	In STRA: IFN- β , IFN- λ 1, IFN- λ 2/3 mRNA levels lower and RV virus load higher No association with age, IgE levels, allergen reactivity, BAL/sputum neutrophils and eosinophils, lung function
AEC	Gielen V, et al. 2015 [57]RV-B1 / RV-A16Pre-treatment with IL-4 / IL-13 Children: STRA 11y (9-15y), vs healthy controls		In STRA: SOCS1 expression increased and related to IFN deficiency, increased viral replication Suppression of RV-induced IFN promoter activation in AEC by SOCS1, dependent on nuclear translocation
AEC	Parsons KS, et al. 2014 [46]	RV-B1 Adults: asthma / Healthy controls	In asthma: reduced release of IL-6, CXCL-8 and IFN- λ in response to RV, reduced release of CXCL-10
AEC	AEC Kicic A, et al. 2016 [58] RV-B1 / RV-B14 Children: <u>Asthma</u> 8.2 y (2.6-14.8) vs healthy children 8.4 y (3.2-15.6)		In asthma: greater virus proliferation and release, reduced apoptosis and wound repair (RV-B1), lower IFN-β, higher inflammatory cytokine production Addition of IFN-β: restored apoptosis, suppressed virus replication and improved repair of AEC from asthmatics, no effect on inflammatory cytokine production
AM	Rupani H, et al. 2016 [47]	RV-A16 Pre-treatment with Imiquimod (TLR7) Adults: <u>severe asthma</u> / Healthy controls	In severe asthma: reduced IFN responses Increased levels of 3 microRNAs (miR-150, miR-152, and miR-375): Blocking these microRNAs with anti-miRNA oligonucleotides increased IFN production
BAL cells	Sykes A, et al. 2012 [48]	RV-A16 Adults: mild-to-moderate allergic asthma vs healthy	In asthma: induction of type I IFN delayed and deficient, associated with airway hyperresponsiveness
Bronchial biopsy specimens	Baraldo S, et al. 2012 [56]	RV-A16 Children, 5y ±0.5: <u>asthma, atopy</u> / <u>Asthma, no atopy</u> / <u>Atopy, no asthma</u> / No atopy nor asthma (Controls)	In all groups vs controls: reduced type I and III IFN production, increased RV viral RNA IFN inversely correlated with airway eosinophils, IL-4, epithelial damage, total IgE (type III IFN)
Whole blood cultures	Bufe A, et al. NDV blood 2002 [62] Children: <u>Allergic asthma</u> / non-allergic asthma / allergic rhinitis / healthy control		In allergic asthma: lower virus-induced IFN-α in allergic asthma/ rhinitis: higher production of IFN-γ
PBMC	Gehlhar K, et al. 2006 [63]	RSV-1A / NDV Adults: <u>Allergic asthma</u> vs healthy controls	In allergic asthma: significant reduction of virus-induced IFN- α release, independent of virus used; no influence of medication (cortico-steroids)
РВМС	likura K, et al. 2011 [64]	RV-B14 <u>Wheeze</u> / healthy; Young children: 2-6y; Youth group: 7- 19 y; Adult group: ≥ 20y	Asthma (youth group): lower IFN-α production Asthma (youth group and adults): lower IL-6, TNF-α, IL-10 and sFasL productions Wheeze (young children): lower IL-10 associated with persistent wheeze at 2 y follow-up
PBMC	Sykes A, et al. 2012 [48]	RV-A16 Adults: mild-to-moderate allergic asthma vs healthy controls	In asthma: no impairement of type I IFN in PBMC, as opposed to BAL cells
PBMC	Durrani S, et al. 2012 [65]	RV-A16 Children 10-12 y: <u>allergic asthma</u> / <u>non-allergic asthma</u> / <u>airborne sensitization without asthma</u> / healthy controls.	Allergic asthma: lower IFN-responses after cross-linking FcɛRI (vs sensitization and vs controls); higher surface expression of FcɛRI on pDC and mDC Inverse relationship between total IgE and diminished IFN secretion, only when cross-linking of FcɛRI
РВМС	Simpson J, et al. 2016 [66]	RV-B1 Adults: <u>asthma</u> , 76% atopic. Sputum phenotypes: eosinophilic / neutrophilic / paucigranulocytic / granulocytic	In neutrophilic asthma: significantly less IFN- α than PBMC from eosinophilic and paucigranulocytic asthma after RV; IFN- α inversely correlated with serum IL-6, sputum IL-1 β Sputum neutrophils and dose of inhaled corticosteroids independent predictors of reduced IFN- α

In vivo				
Bronchial biopsies	Zhu J, et al. 2019 [67]	In vivo inoculation - RV-A16 Young adults: atopic asthma (23 y \pm 1.4) vs non-atopic control subjects (27 y \pm 2.3)	In asthma: IFNα/β deficiency in epithelium, at baseline, day 4 and week 6 post-inoculation; correlated with viral load, and clinical severity Sub-epithelium: lower frequencies of monocytes / macrophages expressing IFNα/β after RV infection in sub-epithelium; subepithelial neutrophils were the source of IFNα/β	
BAL monocyte and AM	Contoli M, et s al. 2006 [71]	<i>In vivo</i> inoculation - RV-A16 Adults: <u>severe asthma</u> (<i>ex vivo</i>) / mild asthma without steroids treatment (<i>ex vivo</i> and <i>in vivo</i>) vs healthy controls	In asthma / Ex vivo: Impaired induction and production of type III IFN-λ1 and IFN-λ2/3 in AEC and AM, impaired induction of IFN- λ upon LPS stimulation In asthma / in vivo: severity of symptoms, BAL virus load, airway inflammation, reduction of lung function inversely correlated with <i>ex vivo</i> production of IFN-λ	
Nasal washes	Miller EK, et al. 2012 [70]	Natural infection: 82% virus Children 5-18 y Upper respiratory infection; <u>Wheezing</u> / no-wheezing RV in 56% wheezing / 37% non-wheezing	Specific association between RV infection and asthma exacerbation, but no difference in virus titers, RV species and inflammatory or allergic molecules between RV+ wheezing and non-wheezing children In RV+ wheezing children: IFN- λ 1 level higher, increased with worsening symptoms	
Nasal washes	Kennedy J, et al. 2014 [29]	Natural infection: 74 children (4-18 y): <u>acute wheezing</u> (57% RV) / acute rhinitis (56% RV) / controls Inoculation (RVA-16): 24 young adults Asthma / healthy	Natural infection: No difference in viral loads and levels of IFN-λ1 between wheezing and acute rhinitis; In children with wheezing: lower levels of sICAM-1 Inoculation: No difference in viral load between groups, levels of sICAM-1 correlated with RV levels	
Nasal washes Induced sputum	Schwantes E, et al. 2014 [68]	Natural infection Adults: <u>asthma</u> vs healhy 17 exacerbations : 67% RV	In asthma, at exacerbation: sputum RNA levels of IFN- α 1, IFN- β 1 and IFN- γ correlated with exacerbation and the peak Asthma Index, early in the course of infection, higher levels of IL-13, IL-10	
Serum PBMC	Bergauer A, et al. 2017 [30]	<i>Ex vivo</i> inoculation of RV-1B on PBMC cultures <i>In vivo</i> natural infection Preschool children 4-6y <u>Asthma</u> at baseline (54% RV) / Asthma at exacerbation (100% RV) vs healthy controls	<i>Ex vivo</i> : induction of STAT1/STAT2 by RV in asthma / IRF9 in controls / IRF1 in both Natural infection (baseline): No increase of IFN- α expression in RV+ asthma patients compared with RV+ healthy controls; increase of IFN- λ protein with RV, both in asthma patients and controls Natural infection (exacerbation): reduced serum IFN- α at exacerbation compared with RV healthy children at baseline	
Sputum Serum PBMC	Deschildre A, et al. 2017 [49]	School-age children 8.9 y (6-16y) <u>Atopic asthma:</u> virus vs no virus Severe exacerbation (64% virus, 51% RV) / steady state (8 weeks later) (25% virus, 11% RV) / 24% virus at both time	Virus at exacerbation: similar levels of IFN- β , IFN- γ , IFN- λ 1 in sputum and plasma, higher airway IL-5 and eosinophils Virus at both time: lower levels of IFN- γ in plasma and sputum at exacerbation, higher airway neutrophilic inflammation at steady state	
Sputum Plasma	Lejeune S, et al. 2020 [8]	Preschool children 30 mo (1y-5y) <u>Recurrent wheeze / preschool asthma;</u> 47% atopy; 70% prone to exacerbation (≥ 2 exacerbations in the previous year) <u>Severe exacerbation</u> (94% virus, 73% RV) / steady state (8 weeks later) (67% virus, 50% RV)	Children prone to exacerbation: lower plasma concentrations of IFN-β and CXCL10 at exacerbation, and lower plasma levels of CXCL10 at steady state	

	In vitro			
Co-culture AEC and T cells	Qin L, et al. 2011 [83]	RSV-A2 AEC T cells extracted from adult PBMC (healthy volunteers)	Prolonged RSV infection in AEC induced T cells differentiation in Th2 and Th17 subsets that released IL-4, IFN- γ and IL-17 induced by supernatants from RSV-infected AEC	
AEC	Lee HC, et al. 2012 [79]	RSV-A AEC from healthy children and asthmatic children	Production of TSLP after infection of AEC with RSV, via activation of an innate signaling pathway that involved retinoic acid induced gene I, interferon promoter-stimulating factor 1, and nuclear factor-κB. Greater levels of TSLP after RSV infection in AEC from asthmatic children than from healthy children	
РВМС	Jurak L, et al. 2018 [82]	RV-A16 and IL-33 co-exposition Adults with <u>allergic asthma</u> vs healthy controls	In asthmatic cells: IL-5 and IL-13 release induced by RV, enhanced by IL-33, surface protein expression of ST2 induced by IL-33 Predominant source of IL-13 release: ILC2 In healthy cells: IL-33 had no effect on IL-5 and IL-13 production	
			In vivo	
BAL PBMC	Jackson D, et al. 2014 [81]	In vivo inoculation Ex vivo study: AEC, PBMC (T cells, ILC2) Adults: mild-to-moderate asthma vs non-atopic healthy volunteers	<i>In vivo</i> : IL-4, IL-5, IL-13 induced by RV in BAL in asthma Type 2 cytokines and IL-33 corelate with clinical outcomes and viral load <i>Ex vivo</i> : IL-5 and IL-13 production directly induced by IL-33 present in RV-infected AEC supernatants by human ILC2 and T cells	
PBMC Serum Nasal washes	Haag P, et al. 2018 [80]	Preschool children 4-6y Ex vivo study: RVB-1 inoculation In vivo: asthma (54% RV) / controls (45% RV) at baseline	<i>Ex vivo:</i> Induction of ILC2 genes in asthma following RV inoculation <i>In vivo:</i> Down regulatation of sST2 in asthma and controls by RV Up regulation of sST2 in RV- asthma patients with low levels of 25(OH)-VitD3 In asthma: direct correlation of serum sST2 with nasal IL-33	
Nasopharyn geal aspirates	Yuan XH, et al. 2020 [78]	Children aged less than 14y Bronchiolitis or pneumonia 19.7% RV, 35 bronchiolitis and 31 pneumonia cases	RV bronchiolitis: increased IL-4/IFN- γ and decreased TNF- α /IL-10 ratios, compared with RV pneumonia	
Sputum Plasma	Lejeune S, et al. 2020 [8]	Preschool children 30mo (1y-5y) <u>Recurrent wheeze / preschool asthma</u> ; 47% atopy; 70% prone to exacerbation (≥ 2 exacerbations in the previous year) Severe exacerbation (94% virus, 73% RV) / steady state (8 weeks later) (67% virus, 50% RV)	Children prone to exacerbation: lower plasma concentrations of IFN- γ , IL-5, TNF- α , IL-10, and lower levels of IFN- γ in sputum at exacerbation At steady state, lower plasma levels of IFN- γ	

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2 Table 1: Major studies (pediatric and adult population) assessing immune responses to viral infection during asthma exacerbation

3 Studies conducted in animal models, and/or without administration of viral particles (e.g. administration of TLR agonists, double-stranded RNAs) were not 4 included in this table.

AEC: Airway epithelial cells; AM: Alveolar macrophages; BAL: broncho-alveolar lavage; CCL: C-C motif ligand; CXCL: C-X-C motif ligand; FcɛRI: high-affinity IgE receptor; IgE: Immunoglobulin E;
 IFN: Interferon; IL: Interleukin; ILC2: type 2 innate lymphoid cells; IRF: interferon regulatory factor; mDC: myeloid dendritic cells; LPS: Lipopolysaccharides; MDA-5: melanoma differentiation-associated

7 protein 5; NDV: Newcastle disease virus; PBMC; peripheral blood mononuclear cells; pDC; plasmacytoid dendritic cells; PBMC; Peripheral blood mononuclear cells; PCR; Polymerase chain reaction;

, 8 9 PIV3: parainfluenza virus type 3; PRR: pattern recognition receptor; RIGI-I: retinoic-acid inducible gene I; RSV: Respiratory syncytial virus; RV: Rhinovirus; sFasL: Serum Soluble Fas Ligand; sICAM-

1: Soluble intercellular adhesion molecule-1; siRNA: small interfering RNA; SOCS1: Suppressor Of Cytokine Signaling 1; sST2: soluble suppression of tumorigenicity 2 (IL-33 receptor IL1RL1); STAT:

10 signal transducer and activator of transcription; STRA: severe therapy resistant asthma; Th2: T helper 2; TLR: Toll like receptor; TNF: Tumor necrosis factor.

Asthma development	Compartment	Study	Design and age-group	Main results
	Hypopharynx	Bisgaard H, et al. 2007 [32]	Hypopharyngeal samples were cultured from 321 neonates at 1 month of age	21% of children were colonized with <i>S. pneumoniae</i> , <i>M. catarrhalis</i> , <i>H. influenzae</i> , or a combination of these organisms. Colonization with one or more of these organisms, but not colonization with <i>S. aureus</i> , was significantly associated with persistent wheeze. Neonates colonized in the hypopharyngeal region with one or a combination of these organisms, were at increased risk for recurrent wheeze and asthma early in life.
	Gut microbiota	Arrieta MC, et al. 2015 [100]	319 children enrolled in the Canadian Healthy Infant Longitudinal Development (CHILD) Study	Children at risk of asthma exhibited transient gut microbial dysbiosis during the first 100 days of life Relative abundance of the bacterial genera <i>Lachnospira, Veillonella, Faecalibacterium</i> , and <i>Rothia</i> was significantly decreased in children at risk of asthma. This reduction in bacterial taxa was accompanied by reduced levels of fecal acetate. Inoculation of germ-free mice with these four bacterial taxa ameliorated airway inflammation in their adult progeny.
	Lung	Loewen K, et al. 2015 [102]	213,661 mother–child dyads with a median follow-up time of 9.3 years from birth Maternal antibiotic use was determined from records of oral antibiotic prescriptions	36.8% of children were exposed prenatally to antibiotics and 10.1% developed asthma. Prenatal antibiotic exposure was associated with an increased risk of asthma. Maternal antibiotic use during 9 months before pregnancy and 9 months postpartum were similarly associated with asthma.
		Hoskin-Parr L, et al. 2013 [103]	4,952 children from the Avon Longitudinal Study of Parents and Children (ALSPAC). Child antibiotic use and asthma, eczema and hay fever symptoms were reported	Children reported to have taken antibiotics during infancy (0-2 yr) were more likely to have asthma. The risk increased with greater numbers of antibiotic courses. The number of courses was also associated with a higher risk of eczema and hay fever but not atopy.
	Gut microbiota	Patrick DM, et al. 2020 [104]	2,644 children from the Canadian Healthy Infant Longitudinal Development (CHILD) prospective birth cohort and among them, 917 were analyzed for fecal microbiota	Reduction in asthma incidence over the study period was associated with decreasing antibiotic use in the first year of life. Asthma diagnosis in childhood was associated with infant antibiotic use (adjusted odds ratio [aOR] 2.15), with a significant dose-response; α-diversity of the gut microbiota was associated with a 32% reduced risk of asthma at age 5 years.
Acute asthma	Nasopharynx	Mansbach JM, et al. 2016 [106]	The MARC-35 prospective cohort study of 1,016 infants (age <1 year) hospitalized with bronchiolitis and followed for the development of recurrent wheezing	RSV infection was associated with high abundance of <i>Firmicutes</i> and <i>Streptococcus</i> and a low abundance of <i>Proteobacteria</i> and the genera <i>Haemophilus</i> and <i>Moraxella</i> . RV infection was associated with low <i>Streptococcus</i> and high <i>Haemophilus</i> and <i>Moraxella</i> . RSV/RV coinfections had intermediate abundances.
	Nasopharynx	De Steenhuijsen Piters WA, et al. 2016 [105]	106 children less than 2 years of age with RSV infection / 26 asymptomatic healthy control	5 nasopharyngeal microbiota clusters were identified, characterized by high levels of either <i>Haemophilus influenzae, Streptococcus, Corynebacterium, Moraxella,</i> or <i>Staphylococcus aureus.</i> RSV hospitalization were positively associated with <i>H. influenzae</i> and <i>Streptococcus</i> RSV-induced expression of IFN-related genes is independent of the microbiota cluster
	Nasopharynx	Kloepfer KM, et al. 2017 [107]	17 adult subjects collected nasal mucus samples on a weekly basis for 5 consecutive weeks	Asymptomatic RV infections were associated with a significant increase in the abundance of <i>Dolosigranulum</i> and <i>Corynebacterium</i> . RV infection preceded the increased odds of detection of <i>Streptococcus</i> and <i>Moraxella</i> by 1 week.
	Blood	Giuffrida LM, et al. 2017 [108]	14 asthmatic and 29 non-asthmatic adult patients with acute respiratory infection 10 healthy individuals	Higher blood concentrations of CCL2 and CCL5 in infected asthmatic patients than in non-asthmatic patients. Both asthmatic and non-asthmatics with pneumonia or bronchitis had higher concentrations of blood cytokines.

	Pharynx	Kama Y, et al. 2020 [109]	Pharyngeal samples of outpatients and/or in patients children with acute exacerbations of asthma (n= 111) (median age: 2.8/2.6, respectively)	The 3 major bacterial pathogens were <i>Streptococcus pneumoniae</i> (29.7%), <i>Moraxella catarrhalis</i> (11.7%), and <i>Haemophilus influenzae</i> (10.8%). Patients with <i>S. pneumoniae</i> colonization had significantly shorter wheezing episodes and reduced lung inflammation (including lower level of TNF- α)
Asthma natural history	Nasopharynx	Teo SM, et al. 2018 [110]	244 infants with acute respiratory tract illnesses (ARIs) followed through their first five years of life	Dominance with <i>Moraxella, Haemophilus</i> and <i>Streptococcus</i> cluster were positively associated with acute respiratory tract illness. This change frequently preceded the detection of viral pathogens and acute symptoms. These changes were associated with ensuing development of persistent wheeze in children developing early allergic sensitization
	Nasal microbiota	Zhou Y, et al. 2019 [113]	A 1 year longitudinal study in school-age children with mild-moderate persistent asthma treated with daily ICS (mean age 8.0 ± 1.8 years)	Children with nasal microbiota dominated by the commensal <i>Corynebacterium / Dolosigranulum</i> cluster at steady state experienced the lowest rates of exacerbation and disease progression. A switch towards the <i>Moraxella</i> - cluster was associated with highest risk of severe asthma exacerbation.
	Lung	Mansbach JM, et al. 2020 [111]	842 infants hospitalized for bronchiolitis with a follow -up at 3 years	Increased abundance of <i>Moraxella</i> or <i>Streptococcus</i> species 3 weeks after day 1 of hospitalization was associated with an increased risk of recurrent wheezing. Increased <i>Streptococcus</i> species abundance the summer after hospitalization was also associated with a greater risk of recurrent wheezing
	Oropharynx	Cuthbertson L, et al. 2019 [112]	Oropharyngeal swabs were collected from 109 children hospitalized for acute wheezing / 75 non-wheezing controls.	No significant difference in bacterial diversity between wheezers and healthy controls In wheezers, attendance at kindergarten or preschool was, associated with increased bacterial diversity.in contrast with rhinovirus (RV) infection
	Nasopharynx	<i>McCauley K. et al.</i> 2019 [35]	312 school-age asthmatic patients enrolled in a trial of omalizumab, nasal secretion samples collected after randomization	Nasal microbiotas dominated by <i>Moraxella</i> species were associated with increased exacerbation risk and eosinophil activation. <i>Staphylococcus</i> or <i>Corynebacterium</i> species–dominated microbiotas were associated with reduced respiratory illness and exacerbation events. <i>Streptococcus</i> species–dominated microbiota increased the risk of RV.
	Sputum	Abdel-Aziz MI. et al. 2020 [114]	Sputum samples were collected in 100 severe asthmatics of the U-BIOPRED adult patients cohort at baseline and after 12-18 months of follow-up	Two microbiome-driven clusters were identified, characterized by asthma onset, smoking status, treatment, lung spirometry results, percentage of neutrophils and macrophages in sputum. Patients of the most severe cluster displayed a commensal-deficient bacterial profile which was associated with worse asthma outcomes. Longitudinal clusters revealed high relative stability after 12-18 months in the severe asthmatics.
	Lung (BAL)	Robinson PFM, et al. 2019 [34]	Children: 35 wheezers categorized as MTW or EVW BAL obtained at steady state	There was no relationship between lower airway inflammation or infection and clinical preschool wheeze phenotypes. 2 groups identified: 1) a Moraxella species dysbiotic microbiota cluster that associated with airway neutrophilia and a mixed microbiota; 2) a mixed microbiota cluster with a macrophage- and lymphocyte-predominant inflammatory profile

Table 2: Major human studies (pediatric and adult population) assessing the impact of pathogenic bacteria and microbiota during the development, the attack and the natural history of pediatric asthma.

BAL: Broncho-alveolar lavage; EVW: Episodic Viral Wheeze, MTW: Multiple trigger Wheeze, RSV: Respiratory syncytial virus; RV: Rhinovirus

Graphical Abstract

- Characteristics of host-microbiota interactions in asthmatic children favor asthma attacks and may allow to define asthma endotypes.
- This knowledge offers promising perspectives to treat asthmatic children using precision medicine

