

# Should we reconsider cefazolin for treating staphylococcal meningitis? A retrospective analysis of cefazolin and cloxacillin cerebrospinal fluid levels in patients treated for staphylococcal meningitis

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1	Research note V2: CLM-19-16649
2	Title: Should we reconsider cefazolin for treating staphylococcal meningitis? A retrospective
3	analysis of cefazolin and cloxacillin CSF levels in patients treated for staphylococcal meningitis
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27	Abstract- 246/250 words
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#### 29 Abstract- 225/250 words

*Objectives:* The main objective of the study was to assess the meningeal penetration of cefazolin and
 cloxacillin in patients treated for methicillin-susceptible staphylococcal meningitis.

*Methods:* We retrospectively identified patients treated for *Staphylococcus* meningitis with measurements of cefazolin or cloxacillin concentrations in cerebrospinal fluid (CSF) using a liquidchromatography coupled with mass-spectrometry validated assay at the Nantes University Hospital between January 2009 and October 2019. *Staphylococcus* meningitis was defined by a compatible clinical presentation and a microbiological confirmation (positive CSF culture or positive specific polymerase chain reaction). Medical charts were retrospectively reviewed to collect microbiological, clinical data and to assess therapeutic success.

**Results:** Among the 17 included patients, 8 (47%) were treated with cefazolin and 9 (53%) with cloxacillin. Median daily dosages of cefazolin and cloxacillin were 8 (range 6-12) and 12 (range 10-13) grams respectively. Cefazolin and cloxacillin were mainly administered via continuous infusion. Eleven patients (65%) were males, median (IQR) age was 54 years (50;70), 14 (82%) had post-operative meningitis and 3 (18%) hematogenous meningitis. Median (IQR) antibiotic CSF concentrations were 2.8 (2.1;5.2) and 0.66 (0.5;0.9) mg/L for cefazolin and cloxacillin groups respectively. Cloxacillin was discontinued in 2 patients for therapeutic failure.

46 *Conclusions:* Patients with staphylococcal meningitis treated with high-dose continuous intravenous
47 infusion of cefazolin achieved therapeutic concentrations in CSF. Cefazolin appears to be a therapeutic
48 candidate which should be properly evaluated in this indication.

49

50 Keywords: Staphylococcus aureus, Staphylococcus epidermidis, meningitis, cefazolin, cloxacillin, CSF

51 antibiotic dosage, therapeutic drug monitoring

52

#### 53 Introduction

54 Staphylococcus spp. is a rare but challenging cause of meningitis which can occur following 55 hematogenous or direct meningeal inoculation, especially after neurosurgery. Anti-staphylococcal 56 penicillins are recommended for the treatment of methicillin-susceptible *Staphylococcus* meningitis [1], 57 even though their CSF penetration is a matter of debate [2]. Cefazolin is not recommended in this 58 indication because of a presumed poor CSF penetration [3].

The main objective of the herein study was to assess the CSF penetration of cefazolin and cloxacillin in patients treated for methicillin-susceptible staphylococcal meningitis. We present a series of patients treated for *Staphylococcus* meningitis either with high-dose cefazolin or cloxacillin and report the CSF and plasma concentrations of antibiotics as well as clinical outcomes.

#### 63 Materials and Methods

64 Patients with positive culture of CSF for Staphylococcus spp. (at least two samples in case of 65 Methicillin-susceptible coagulase-negative staphylococci (MSCNS)) or a positive polymerase chain 66 reaction (PCR) in CSF sample were retrospectively identified between January 2009 and October 2019 67 in 3 French hospitals. Among them, patients with CSF concentrations of cefazolin or cloxacillin 68 measured for therapeutic drug monitoring (TDM) were identified. Finally only patients with proven 69 staphylococcal meningitis (CSF parameters and clinical picture compatible with the diagnosis of 70 meningitis as defined by Tunkel and colleagues [1]) were selected. Medical charts were retrospectively 71 reviewed to collect clinical and microbiological data and outcomes. CSF parameters were reported at 72 the time of meningitis diagnosis and of cefazolin or cloxacillin concentration measurements in CSF. 73 Concomitant antibiotic plasma concentrations were collected when available. All CSF concentrations 74 were measured within 48h following CSF sampling and were systematically stored at -80°C if they were 75 not immediately performed. Antibiotic concentrations were measured in CSF and plasma using a liquid-76 chromatography coupled with mass-spectrometry validated assay [4]. Limit of quantitation was 0.5 77 mg/L for cloxacillin and cefazolin. A steady state plasmatic concentration (Css) was defined by a 78 concentration measured at least after 24 hours of anti-staphylococcal betalactams treatment.

Bacterial antibiotic susceptibilities, including oxacillin minimum inhibitory concentrations (MICs) were
determined using a Vitek 2 automated system (bioMérieux, France) and AST-P631 cards. Because

cefazolin is not included in AST-P631 cards, MICs of cefazolin were specifically determined by broth microdilution (BMD) antimicrobial susceptibility testing according to the European Committee on Antimicrobial Susceptibility Testing (EUCAST) guidelines. CSF concentrations were analyzed with respect to the latest epidemiological cut-off (ECOFF) values for cefazolin (2 mg/L) and cloxacillin (0.5 mg/L). The study received ethical approval from the University of Nantes Human Research Ethics Committee. Patients were informed of the study in accordance with French legal standards.

#### 87 **Results**

88 Study population

89 Seventeen patients met inclusion criteria during the study period (Figure S1). Patient characteristics are 90 reported in Table 1. Eleven (65%) were males and median (interquartile range [IQR]) age was 54 years 91 (50-70). Fourteen (84%) patients had post-operative meningitis and 3 (18%) hematogenous meningitis. 92 Eight (47%) patients were treated by cefazolin all with continuous intravenous infusion and nine (53%) 93 by cloxacillin mainly with continuous intravenous infusion (missing data for 3 patients). Daily dosages 94 and dosing times are detailed in Table 1 and Figure S2. Median MICs were 0.5 mg/L for cefazolin 95 (range 0.125-0.5) and oxacillin (range  $\leq 0.25$ -0.5) for patients treated by cefazolin and cloxacillin 96 respectively.

97 Pharmacokinetics-pharmacodynamics of cefazolin and cloxacillin in CSF

98 Twenty eight CSF concentrations were measured in the 17 patients: 14 in cefazolin group (range 1-4 99 per patient) and 14 in cloxacillin group (range 1-5 per patient). All CSF samples were drawn at least 24 100 hours after the initiation of antibiotic so steady state was achieved in CSF. CSF samples reported as 101 below the limit of quantitation were fixed at 0.5 mg/L for analysis (n=5 for cloxacillin and n=2 for 102 cefazolin). Median (IOR) CSF concentration was 2.8 mg/L (2.1;5.2) for cefazolin and 0.66 mg/L 103 (0.5;0.9) for cloxacillin. The median (IQR) CSF/plasma ratio was 4.3% (2.9;8.4) for cefazolin and 1.8 104 % (1.7;2.8) for cloxacillin. The median (IQR) CSF/MIC ratio was 7.38 (2.1;20.04) for cefazolin and 2.0 105 (1.34;3.4) for cloxacillin (Table 1). The median (IQR) CSF/ECOFF value ratio was 1.4 (1.05;2.6) for 106 cefazolin and 1.33 (1:1.805) for cloxacillin. CSF concentration of antibiotic was above ECOFF in 11 107 (79%) samples corresponding to 6 (75%) patients receiving cefazolin and 8 (57%) samples 108 corresponding to 6 (67%) patients receiving cloxacillin (Figure 1).

#### 109 Clinical outcomes

All patients were cured without any recurrences. In 2 patients treated with cloxacillin for external ventricular drain-associated *S. aureus* meningitis, CSF culture remained positive 2 days and 4 days after cloxacillin initiation respectively. They were considered by the physicians in charge as therapeutic failures leading to cloxacillin discontinuation and its replacement by another antibiotic. Of note in both cases the infected CSF drains had not been removed between diagnosis and antibiotic change.

#### 115 Discussion

In this study, cefazolin demonstrated levels of CSF penetration higher than previously expected in patients with staphylococcal meningitis and achieved therapeutic concentrations confirming preliminary results [5]. Cefazolin has become a first line option for the management of methicillin-susceptible *Staphylococcus* bacteraemia because of a good efficacy [6], favourable pharmacokinetics [4] and a good tolerability profile when compared with anti-staphylococcal penicillins [7]. Gelfand *et al* recently pointed out the lack of evidence regarding the treatment of CNS infections due to MSSA [8].

To the best of our knowledge the herein study reports the largest series of CSF measurements of cefazolin and cloxacillin in patients treated for staphylococcal meningitis. CSF penetration of cefazolin appeared as good as cloxacillin.

125 The increased CSF protein levels observed in our patients is suggestive of meningeal inflammation, a 126 major determinant of blood-brain barrier permeability and antibiotic penetration in CSF [9,10]. Of note, 127 six cefazolin CSF concentrations measured in 4 patients without meningitis were collected during study 128 period. Median (IQR) concentrations were 0.95 mg/L (0.5; 1.4) in CSF and 47.7 mg/L (42.3; 52.1) in 129 plasma with a median (IQR) CSF/plasma ratio of 2.1% (0.9;5.2). Knowing that meningeal inflammation 130 can vary a lot in nosocomial meningitis, we suggest performing systematic TDM in CSF when using 131 cefazolin in this indication. Naturally, this statement should also be applied to cloxacillin considering 132 the poor CSF penetration observed here, despite assigning the value of 0.5 mg/L to the assays below the 133 limit of quantification.

134 In general a concentration measured in CSF above ECOFF should be targeted throughout the treatment 135  $(100\% fT_{>MIC})$  [11]. The therapeutic failure of the 2 patients treated with cloxacillin who still had positive 136 CSF culture at least 2 days after antibiotic therapy is hard to ascertain because of the retention of the

138	the small number of patients included, especially those who received cefazolin or cloxacillin as a
139	monotherapy, and the variability of co-administered antibiotics limiting the comparison of clinical
140	outcomes between the two groups, (iii) single dosage in most patients yielding the data insufficient to
141	ascertain the stability of cefazolin concentrations in CSF.
142	In conclusion, continuous high-dose intravenous infusion of cefazolin achieved targeted concentrations
143	in CSF for a large majority of patients with staphylococcal meningitis. Based on our results this regimen
144	may be a suitable option in this indication. If chosen, we recommend associating cefazolin regimen with
145	a systematic therapeutic drug monitoring directly in CSF to check target attainment.
146	
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infected CSF drain in those patients. The limitations of our study include (i) its retrospective design, (ii)

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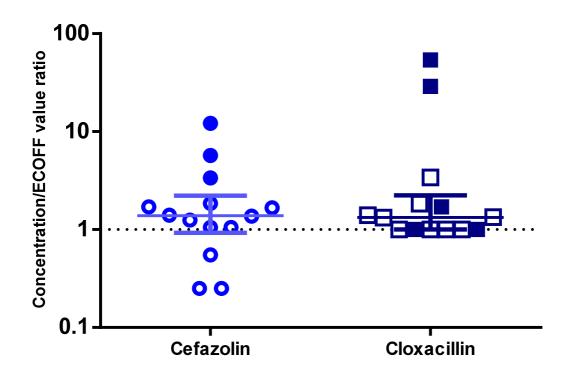
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#### 170 Authors' contributions

171 Paul Le Turnier (Conceptualization, Investigation, Data curation, Methodology, Writing - original 172 draft), Matthieu Grégoire (Conceptualization, Investigation, Formal analysis, Visualization, Writing -173 original draft), Guillaume Deslandes (Investigation), Karim Lakhal (Conceptualization, Investigation, 174 Writing - review and editing), Colin Deschanvres (Methodology, Investigation, Writing - review and 175 editing), Raphael Lecomte (Investigation, Writing - review and editing), Jean Philippe Talarmin 176 (Investigation), Vincent Dubée (Investigation), Ronan Bellouard (Investigation, Formal analysis), 177 David Boutoille (Supervision, Writing - review & editing), Anne Gaelle Leroy (Formal analysis, 178 Methodology, Writing - original draft), Benjamin Jean Gaborit (Conceptualization, Investigation, Data 179 curation, Methodology, Writing – original draft).

#### 180 **References**

- 181 [1] Tunkel AR, Hasbun R, Bhimraj A, Byers K, Kaplan SL, Scheld WM, et al. 2017 Infectious
  182 Diseases Society of America's Clinical Practice Guidelines for Healthcare-Associated
  183 Ventriculitis and Meningitis. Clin Infect Dis Off Publ Infect Dis Soc Am 2017;64:e34–65.
  184 https://doi.org/10.1093/cid/ciw861.
- [2] Chew R, Woods ML. Flucloxacillin does not achieve therapeutic cerebrospinal fluid levels
   against meticillin-sensitive Staphylococcus aureus in adults: A case report and review of the
   literature. Int J Antimicrob Agents 2016;47:229–31.
- 188 https://doi.org/10.1016/j.ijantimicag.2015.12.009.
- [3] Thys JP, Vanderkelen B, Klastersky J. Pharmacological Study of Cefazolin During Intermittent
   and Continuous Infusion: a Crossover Investigation in Humans. Antimicrob Agents Chemother
   191 1976;10:395–8. https://doi.org/10.1128/AAC.10.3.395.
- [4] Bellouard R, Deschanvres C, Deslandes G, Dailly É, Asseray N, Jolliet P, et al. Population
  Pharmacokinetic Study of Cefazolin Dosage Adaptation in Bacteremia and Infective
  Endocarditis Based on a Nomogram. Antimicrob Agents Chemother 2019;63.
  https://doi.org/10.1128/AAC.00806-19.
- 196 [5] Grégoire M, Gaborit B, Deschanvres C, Lecomte R, Deslandes G, Dailly É, et al. High-Dosage
  197 Cefazolin Achieves Sufficient Cerebrospinal Diffusion To Treat an External Ventricular
  198 Drainage-Related Staphylococcus aureus Ventriculitis. Antimicrob Agents Chemother 2019;63.
  199 https://doi.org/10.1128/AAC.01844-18.
- Weis S, Kesselmeier M, Davis JS, Morris AM, Lee S, Scherag A, et al. Cefazolin versus antistaphylococcal penicillins for the treatment of patients with Staphylococcus aureus bacteraemia.
  Clin Microbiol Infect Off Publ Eur Soc Clin Microbiol Infect Dis 2019;25:818–27. https://doi.org/10.1016/j.cmi.2019.03.010.
- [7] Loubet P, Burdet C, Vindrios W, Grall N, Wolff M, Yazdanpanah Y, et al. Cefazolin versus antistaphylococcal penicillins for treatment of methicillin-susceptible Staphylococcus aureus bacteraemia: a narrative review. Clin Microbiol Infect 2018;24:125–32. https://doi.org/10.1016/j.cmi.2017.07.003.
- [8] Gelfand MS, Cleveland KO. Re: "Cefazolin versus anti-staphylococcal penicillins for the treatment of patients with Staphylococcus aureus bacteremia" by Weis et al. Clin Microbiol
  Infect Off Publ Eur Soc Clin Microbiol Infect Dis 2019.
  https://doi.org/10.1016/j.cmi.2019.07.017.
- [9] Varatharaj A, Galea I. The blood-brain barrier in systemic inflammation. Brain Behav Immun 2017;60:1–12. https://doi.org/10.1016/j.bbi.2016.03.010.
- [10] Nau R, Sörgel F, Eiffert H. Penetration of drugs through the blood-cerebrospinal fluid/blood-brain barrier for treatment of central nervous system infections. Clin Microbiol Rev 2010;23:858–83. https://doi.org/10.1128/CMR.00007-10.
- [11] Mouton JW, Meletiadis J, Voss A, Turnidge J. Variation of MIC measurements: the contribution
   of strain and laboratory variability to measurement precision. J Antimicrob Chemother
- 219 2018;73:2374–9. https://doi.org/10.1093/jac/dky232.
- 220



**Figure 1: Cerebrospinal fluid concentrations (CSF) of cefazolin and cloxacillin in meningitis.** Concentrations are represented in function of EUCAST epidemiological cut-off (ECOFF) values. Median (Interquartile range) CSF concentration was 2.8 mg/L (2.1;5.2) for cefazolin and 0.66 mg/L (0.5;0.9) for cloxacillin. External ventricular drain are represented by circles (for cefazolin) and squares (for cloxacillin) and lumbar puncture by filled circles and squares.

	Cefazolin group	Cloxacillin group
~ · · · · · · · · · · · · · · · · · · ·	( <b>n=8</b> )	( <b>n=9</b> )
Study population characteristics		
Age, years, median (IQR)	58.4 (50.3;74)	54 (50;67)
Male gender, n (%)	6 (75)	5 (55)
BMI, Kg/m <sup>2</sup> , median (IQR)	25.9 (24.2;26.9)	22.3 (21;30.2)
Glomerular filtration (MDRD) on diagnosis, median (IQR) <sup>a</sup>	136 (115;146)	102 (93;114)
Mechanism of the meningitis, n (%)		
Post-operative meningitis	7 (88)	7 (78)
EVD associated meningitis	5 (62)	6 (66)
Hematogenous meningitis	1 (12)	2 (22)
Microbiological diagnosis of meningitis, n (%)		
S. aureus	4 (44)	6 (66)
S. epidermidis	3 (33)	3 (34)
S. lugdunensis	1 (11)	-
CSF parameters on diagnosis, median (IQR)		
White blood cells/mm <sup>3</sup>	440 (92;6022)	370 (181;1166)
Protein, g/L	1.52 (0.81;2.47)	1.76 (0.59;2.41)
Glucose mmol/L <sup>b</sup>	3.15 (0.17;4.75)	2.95 (1.42;4.1)
Lactate (mmol/L) <sup>c</sup>	4.5 (3.8;9.7)	5 (3.1;9.8)
CSF parameters in samples used for pharmacological		
CSF parameters in samples used for pharmacological measurements		
measurements	14	14
measurements CSF samples with antibiotic dosing, n		
measurements CSF samples with antibiotic dosing, n CSF sampling via EVD, n (%)	11 (79)	9 (64)
measurements CSF samples with antibiotic dosing, n CSF sampling via EVD, n (%) White blood cells/mm <sup>3</sup> , median (IQR) <sup>d</sup>	11 (79) 31.5 (6.75;68.5)	9 (64) 3 (2;21)
measurements CSF samples with antibiotic dosing, n CSF sampling via EVD, n (%) White blood cells/mm <sup>3</sup> , median (IQR) <sup>d</sup> Protein g/L, median (IQR) <sup>e</sup>	11 (79) 31.5 (6.75;68.5) 0.86 (0.67;1.27)	9 (64) 3 (2;21) 0.76 (0.54;1.13)
measurements CSF samples with antibiotic dosing, n CSF sampling via EVD, n (%) White blood cells/mm <sup>3</sup> , median (IQR) <sup>d</sup>	11 (79) 31.5 (6.75;68.5)	9 (64) 3 (2;21)
<ul> <li>measurements</li> <li>CSF samples with antibiotic dosing, n</li> <li>CSF sampling via EVD, n (%)</li> <li>White blood cells/mm<sup>3</sup>, median (IQR)<sup>d</sup></li> <li>Protein g/L, median (IQR)<sup>e</sup></li> <li>Glucose mmol/L, median (IQR)<sup>f</sup></li> <li>Lactate mmol/L, median (IQR)<sup>g</sup></li> </ul> Pharmacological results, median (IQR)	11 (79) 31.5 (6.75;68.5) 0.86 (0.67;1.27) 3.9 (2.27;4.75) 2.7 (2.1;3.3)	9 (64) 3 (2;21) 0.76 (0.54;1.13) 2.7 (1.8;3.5) 3.55 (3;4.1)
<ul> <li>measurements</li> <li>CSF samples with antibiotic dosing, n</li> <li>CSF sampling via EVD, n (%)</li> <li>White blood cells/mm<sup>3</sup>, median (IQR)<sup>d</sup></li> <li>Protein g/L, median (IQR)<sup>e</sup></li> <li>Glucose mmol/L, median (IQR)<sup>f</sup></li> <li>Lactate mmol/L, median (IQR)<sup>g</sup></li> </ul> Pharmacological results, median (IQR) CSF antibiotic concentration, mg/L	11 (79) 31.5 (6.75;68.5) 0.86 (0.67;1.27) 3.9 (2.27;4.75) 2.7 (2.1;3.3) 2.8 (2.1;5.2)	9 (64) 3 (2;21) 0.76 (0.54;1.13) 2.7 (1.8;3.5) 3.55 (3;4.1) 0.66 (0.5;0.9)
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<ul> <li>measurements</li> <li>CSF samples with antibiotic dosing, n</li> <li>CSF sampling via EVD, n (%)</li> <li>White blood cells/mm<sup>3</sup>, median (IQR)<sup>d</sup></li> <li>Protein g/L, median (IQR)<sup>e</sup></li> <li>Glucose mmol/L, median (IQR)<sup>f</sup></li> <li>Lactate mmol/L, median (IQR)<sup>g</sup></li> </ul> Pharmacological results, median (IQR) CSF antibiotic concentration, mg/L Plasma antibiotic concentration, mg/L Ratio concentration CSF/Plasma, % <sup>h</sup>	11 (79) 31.5 (6.75;68.5) 0.86 (0.67;1.27) 3.9 (2.27;4.75) 2.7 (2.1;3.3) 2.8 (2.1;5.2) 58.1 (55.5;70.4) 4.3 (2.9;8.4)	9 (64) 3 (2;21) 0.76 (0.54;1.13) 2.7 (1.8;3.5) 3.55 (3;4.1) 0.66 (0.5;0.9) 47.7 (46 ;51.1) 1.8 (1.7;2.8)
<ul> <li>measurements</li> <li>CSF samples with antibiotic dosing, n</li> <li>CSF sampling via EVD, n (%)</li> <li>White blood cells/mm<sup>3</sup>, median (IQR)<sup>d</sup></li> <li>Protein g/L, median (IQR)<sup>e</sup></li> <li>Glucose mmol/L, median (IQR)<sup>f</sup></li> <li>Lactate mmol/L, median (IQR)<sup>g</sup></li> </ul> Pharmacological results, median (IQR) <ul> <li>CSF antibiotic concentration, mg/L</li> <li>Plasma antibiotic concentration, mg/L</li> </ul>	11 (79) 31.5 (6.75;68.5) 0.86 (0.67;1.27) 3.9 (2.27;4.75) 2.7 (2.1;3.3) 2.8 (2.1;5.2) 58.1 (55.5;70.4)	9 (64) 3 (2;21) 0.76 (0.54;1.13) 2.7 (1.8;3.5) 3.55 (3;4.1) 0.66 (0.5;0.9) 47.7 (46 ;51.1)
measurements         CSF samples with antibiotic dosing, n         CSF sampling via EVD, n (%)         White blood cells/mm <sup>3</sup> , median (IQR) <sup>d</sup> Protein g/L, median (IQR) <sup>e</sup> Glucose mmol/L, median (IQR) <sup>f</sup> Lactate mmol/L, median (IQR) <sup>g</sup> Pharmacological results, median (IQR)         CSF antibiotic concentration, mg/L         Plasma antibiotic concentration, mg/L         Ratio concentration CSF/Plasma, % <sup>h</sup> CSF/MIC ratio	11 (79) 31.5 (6.75;68.5) 0.86 (0.67;1.27) 3.9 (2.27;4.75) 2.7 (2.1;3.3) 2.8 (2.1;5.2) 58.1 (55.5;70.4) 4.3 (2.9;8.4) 7.38 (2.1;20.04)	9 (64) 3 (2;21) 0.76 (0.54;1.13) 2.7 (1.8;3.5) 3.55 (3;4.1) 0.66 (0.5;0.9) 47.7 (46 ;51.1) 1.8 (1.7;2.8) 2.0 (1.34;3.4)
<ul> <li>measurements <ul> <li>CSF samples with antibiotic dosing, n</li> <li>CSF sampling via EVD, n (%)</li> <li>White blood cells/mm<sup>3</sup>, median (IQR)<sup>d</sup></li> <li>Protein g/L, median (IQR)<sup>e</sup></li> <li>Glucose mmol/L, median (IQR)<sup>f</sup></li> <li>Lactate mmol/L, median (IQR)</li> </ul> </li> <li>Pharmacological results, median (IQR)</li> <li>CSF antibiotic concentration, mg/L</li> <li>Plasma antibiotic concentration, mg/L</li> <li>Ratio concentration CSF/Plasma, %<sup>h</sup></li> <li>CSF/MIC ratio</li> </ul> <li>Characteristics of the antibotic therapy <ul> <li>Daily dosage, g [median (IQR) (min-max)]</li> </ul> </li>	11 (79) 31.5 (6.75;68.5) 0.86 (0.67;1.27) 3.9 (2.27;4.75) 2.7 (2.1;3.3) 2.8 (2.1;5.2) 58.1 (55.5;70.4) 4.3 (2.9;8.4) 7.38 (2.1;20.04) 8 (7.88;8) (6-12)	9 (64) 3 (2;21) 0.76 (0.54;1.13) 2.7 (1.8;3.5) 3.55 (3;4.1) 0.66 (0.5;0.9) 47.7 (46;51.1) 1.8 (1.7;2.8) 2.0 (1.34;3.4) 12 (12;12) (10-13)
<ul> <li>measurements <ul> <li>CSF samples with antibiotic dosing, n</li> <li>CSF sampling via EVD, n (%)</li> <li>White blood cells/mm<sup>3</sup>, median (IQR)<sup>d</sup></li> <li>Protein g/L, median (IQR)<sup>e</sup></li> <li>Glucose mmol/L, median (IQR)<sup>f</sup></li> <li>Lactate mmol/L, median (IQR)</li> </ul> </li> <li>Pharmacological results, median (IQR)</li> <li>CSF antibiotic concentration, mg/L</li> <li>Plasma antibiotic concentration, mg/L</li> <li>Ratio concentration CSF/Plasma, %<sup>h</sup></li> <li>CSF/MIC ratio</li> </ul> <li>Characteristics of the antibotic therapy <ul> <li>Daily dosage, g [median (IQR) (min-max)]</li> <li>Co administered antibiotic, n (%)</li> </ul> </li>	11 (79) 31.5 (6.75;68.5) 0.86 (0.67;1.27) 3.9 (2.27;4.75) 2.7 (2.1;3.3) 2.8 (2.1;5.2) 58.1 (55.5;70.4) 4.3 (2.9;8.4) 7.38 (2.1;20.04) 8 (7.88;8) (6-12) 6 (75)	9 (64) 3 (2;21) 0.76 (0.54;1.13) 2.7 (1.8;3.5) 3.55 (3;4.1) 0.66 (0.5;0.9) 47.7 (46;51.1) 1.8 (1.7;2.8) 2.0 (1.34;3.4) 12 (12;12) (10-13) 7 (77.8)
<ul> <li>measurements <ul> <li>CSF samples with antibiotic dosing, n</li> <li>CSF sampling via EVD, n (%)</li> <li>White blood cells/mm<sup>3</sup>, median (IQR)<sup>d</sup></li> <li>Protein g/L, median (IQR)<sup>e</sup></li> <li>Glucose mmol/L, median (IQR)<sup>f</sup></li> <li>Lactate mmol/L, median (IQR)</li> </ul> </li> <li>Pharmacological results, median (IQR)</li> <li>CSF antibiotic concentration, mg/L</li> <li>Plasma antibiotic concentration, mg/L</li> <li>Ratio concentration CSF/Plasma, %<sup>h</sup></li> <li>CSF/MIC ratio</li> </ul> <li>Characteristics of the antibotic therapy <ul> <li>Daily dosage, g [median (IQR) (min-max)]</li> <li>Co administered antibiotic, n (%)</li> <li>Levofloxacin</li> </ul> </li>	11 (79) 31.5 (6.75;68.5) 0.86 (0.67;1.27) 3.9 (2.27;4.75) 2.7 (2.1;3.3) 2.8 (2.1;5.2) 58.1 (55.5;70.4) 4.3 (2.9;8.4) 7.38 (2.1;20.04) 8 (7.88;8) (6-12) 6 (75) 4 (50)	9 (64) 3 (2;21) 0.76 (0.54;1.13) 2.7 (1.8;3.5) 3.55 (3;4.1) 0.66 (0.5;0.9) 47.7 (46;51.1) 1.8 (1.7;2.8) 2.0 (1.34;3.4) 12 (12;12) (10-13) 7 (77.8) 4 (44.4)
<ul> <li>measurements</li> <li>CSF samples with antibiotic dosing, n</li> <li>CSF sampling via EVD, n (%)</li> <li>White blood cells/mm<sup>3</sup>, median (IQR)<sup>d</sup></li> <li>Protein g/L, median (IQR)<sup>e</sup></li> <li>Glucose mmol/L, median (IQR)<sup>f</sup></li> <li>Lactate mmol/L, median (IQR)</li> <li>Pharmacological results, median (IQR)</li> <li>CSF antibiotic concentration, mg/L</li> <li>Plasma antibiotic concentration, mg/L</li> <li>Ratio concentration CSF/Plasma, %<sup>h</sup></li> <li>CSF/MIC ratio</li> </ul> Characteristics of the antibotic therapy <ul> <li>Daily dosage, g [median (IQR) (min-max)]</li> <li>Co administered antibiotic, n (%)</li> <li>Levofloxacin</li> <li>Clindamycin</li> </ul>	11 (79) 31.5 (6.75;68.5) 0.86 (0.67;1.27) 3.9 (2.27;4.75) 2.7 (2.1;3.3) 2.8 (2.1;5.2) 58.1 (55.5;70.4) 4.3 (2.9;8.4) 7.38 (2.1;20.04) 8 (7.88;8) (6-12) 6 (75)	9 (64) 3 (2;21) 0.76 (0.54;1.13) 2.7 (1.8;3.5) 3.55 (3;4.1) 0.66 (0.5;0.9) 47.7 (46;51.1) 1.8 (1.7;2.8) 2.0 (1.34;3.4) 12 (12;12) (10-13) 7 (77.8) 4 (44.4)
<ul> <li>measurements</li> <li>CSF samples with antibiotic dosing, n</li> <li>CSF sampling via EVD, n (%)</li> <li>White blood cells/mm<sup>3</sup>, median (IQR)<sup>d</sup></li> <li>Protein g/L, median (IQR)<sup>e</sup></li> <li>Glucose mmol/L, median (IQR)<sup>f</sup></li> <li>Lactate mmol/L, median (IQR)</li> <li>CSF antibiotic concentration, mg/L</li> <li>Plasma antibiotic concentration, mg/L</li> <li>Ratio concentration CSF/Plasma, %<sup>h</sup></li> <li>CSF/MIC ratio</li> </ul> Characteristics of the antibotic therapy <ul> <li>Daily dosage, g [median (IQR) (min-max)]</li> <li>Co administered antibiotic, n (%)</li> <li>Levofloxacin</li> <li>Clindamycin</li> <li>Rifampin</li> </ul>	11 (79) $31.5 (6.75;68.5)$ $0.86 (0.67;1.27)$ $3.9 (2.27;4.75)$ $2.7 (2.1;3.3)$ $2.8 (2.1;5.2)$ $58.1 (55.5;70.4)$ $4.3 (2.9;8.4)$ $7.38 (2.1;20.04)$ $8 (7.88;8) (6-12)$ $6 (75)$ $4 (50)$ $1 (12.5)$	9 (64) 3 (2;21) 0.76 (0.54;1.13) 2.7 (1.8;3.5) 3.55 (3;4.1) 0.66 (0.5;0.9) 47.7 (46;51.1) 1.8 (1.7;2.8) 2.0 (1.34;3.4) 12 (12;12) (10-13) 7 (77.8) 4 (44.4) - 2 (22.2)
<ul> <li>measurements</li> <li>CSF samples with antibiotic dosing, n</li> <li>CSF sampling via EVD, n (%)</li> <li>White blood cells/mm<sup>3</sup>, median (IQR)<sup>d</sup></li> <li>Protein g/L, median (IQR)<sup>e</sup></li> <li>Glucose mmol/L, median (IQR)<sup>f</sup></li> <li>Lactate mmol/L, median (IQR)</li> <li>CSF antibiotic concentration, mg/L</li> <li>Plasma antibiotic concentration, mg/L</li> <li>Ratio concentration CSF/Plasma, %<sup>h</sup></li> <li>CSF/MIC ratio</li> </ul> Characteristics of the antibotic therapy <ul> <li>Daily dosage, g [median (IQR) (min-max)]</li> <li>Co administered antibiotic, n (%)</li> <li>Levofloxacin</li> <li>Clindamycin</li> <li>Rifampin</li> <li>Other<sup>i</sup></li> </ul>	11 (79) 31.5 (6.75;68.5) 0.86 (0.67;1.27) 3.9 (2.27;4.75) 2.7 (2.1;3.3) 2.8 (2.1;5.2) 58.1 (55.5;70.4) 4.3 (2.9;8.4) 7.38 (2.1;20.04) 8 (7.88;8) (6-12) 6 (75) 4 (50)	9 (64) 3 (2;21) 0.76 (0.54;1.13) 2.7 (1.8;3.5) 3.55 (3;4.1) 0.66 (0.5;0.9) 47.7 (46;51.1) 1.8 (1.7;2.8) 2.0 (1.34;3.4) 12 (12;12) (10-13) 7 (77.8) 4 (44.4)
<ul> <li>measurements</li> <li>CSF samples with antibiotic dosing, n</li> <li>CSF sampling via EVD, n (%)</li> <li>White blood cells/mm<sup>3</sup>, median (IQR)<sup>d</sup></li> <li>Protein g/L, median (IQR)<sup>e</sup></li> <li>Glucose mmol/L, median (IQR)<sup>f</sup></li> <li>Lactate mmol/L, median (IQR)</li> <li>CSF antibiotic concentration, mg/L</li> <li>Plasma antibiotic concentration, mg/L</li> <li>Ratio concentration CSF/Plasma, %<sup>h</sup></li> <li>CSF/MIC ratio</li> </ul> Characteristics of the antibotic therapy <ul> <li>Daily dosage, g [median (IQR) (min-max)]</li> <li>Co administered antibiotic, n (%)</li> <li>Levofloxacin</li> <li>Clindamycin</li> <li>Rifampin</li> </ul>	11 (79) $31.5 (6.75;68.5)$ $0.86 (0.67;1.27)$ $3.9 (2.27;4.75)$ $2.7 (2.1;3.3)$ $2.8 (2.1;5.2)$ $58.1 (55.5;70.4)$ $4.3 (2.9;8.4)$ $7.38 (2.1;20.04)$ $8 (7.88;8) (6-12)$ $6 (75)$ $4 (50)$ $1 (12.5)$	9 (64) 3 (2;21) 0.76 (0.54;1.13) 2.7 (1.8;3.5) 3.55 (3;4.1) 0.66 (0.5;0.9) 47.7 (46;51.1) 1.8 (1.7;2.8) 2.0 (1.34;3.4) 12 (12;12) (10-13) 7 (77.8) 4 (44.4) - 2 (22.2)

 Table 1: Description of study population characteristics, CSF parameters, pharmacological data and outcomes.

BMI, body mass index; IQR, Interquartile range; MIC, Minimum Inhibitory Concentration; EVD, external ventricular drain; CSF, Cerebrospinal fluid

<sup>a</sup>missing data: cloxacillin group (n=1) <sup>b</sup>missing data: cloxacillin group (n=1) <sup>c</sup>missing data: cefazolin group (n=3), cloxacillin group (n=1) <sup>d</sup> missing data : cefazolin group (n=4), cloxacillin group (n=4) <sup>e</sup> missing data : cefazolin group (n=6), cloxacillin group (n=3) <sup>f</sup> missing data : cefazolin group (n=6), cloxacillin group (n=3) <sup>g</sup>missing data : cefazolin group (n=9), cloxacillin group (n=8) <sup>h</sup>missing data: cefazolin group (n=4), cloxacillin group (n=5) <sup>i</sup>Other antibiotics were linezolid (n=1 in the cefazolin group and n=1 in cloxacillin group), fosfomycin (n=1) in the cloxacillin group, acid fusidic (n=1) in the cefazolin group, gentamycin (n=1) in the cloxacillin group. <sup>j</sup>2 failures with positive culture above 48 h of antibiotic therapy and 1 underdosing in the cloxacillin group.