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Ca²⁺ signalling is critical for pancreatic stellate cell's pathophysiology : from fibrosis to cancer hallmarks

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Abstract :

Considerated Known as the key effector of pancreatic fibrosis and pancreatic cancer's stroma desmoplastic stromal reaction development, activated pancreatic stellate cells (aPSCs) are attracting more and more the attention of pancreatic research. Since a few years, a crucial role of Ca²⁺ signal in PSC's activation processes has been emerged. Although most of the work on PSCs and Ca²⁺ signal has been carried on aPSCs, because once seated in culture, they become activated within 2-5 days, there are also some studies focusing on the role of Ca²⁺ in quiescent pancreatic stellate cells (qPSCs). In this mini-review, we reported all findings on Ca²⁺ signal in qPSCs and aPSCs, in relation with to pancreatic fibrosis and cancer promotion.

Key words : calcium signalling, pancreatic stellate cells, ion channels, fibrosis, cancer

Introduction

Pancreatic stellate cell's research is receiving, since several years, an increased attention due to their crucial role of PSCs in the development of pancreatic fibrosis (PF) and cancer (PC) [1]. Initially characterized as resident quiescent vitamin A storing cells of the exocrine pancreas, they undergo transformations during pancreatic injury in order to become activated [2]. The aPSCs are responsible for the extracellular matrix (ECM) deposition, leading to the creation of a dense fibrotic stroma, presented during PF and PC [2,3]. Despite, the well studied hitherto mechanisms participating in PSC's activation, still much has to be learned, especially regarding to the processes triggering their transition from the quiescent to the activated state.

 Ca^{2+} acts as an universal intracellular second messenger implicated in the regulation of myriad biological processes, including protein secretion, gene expression, cell proliferation, migration, contraction and cell death, as well as cell differentiation [4,5]. Several studies have arlready elucidated the importance of Ca^{2+} signals in PSCs pathophysiology, suggesting its role in the modulation of PSC's phenotype transition. Here, we summarized what is already known about Ca^{2+} signalling in qPSCs and aPSCs, focusing on extracellular agent²s and ion channel's mediated- Ca^{2+} signals, in the PF and PC context.

How important pancreatic stellate cells are for pancreatic pathophysiology ?

The role of PSCs in the healthy and diseased pancreas has been underestimated for a long time. In the normal healthy pancreas, PSCs represent only 4-7% of the pancreas' total cell number, existing in a so-called quiescent state [6]. Under physiological conditions, qPSCs are characterized by their ability to store retinoid-containing cytoplasmic lipid droplets, having periacinar space location but also founded in perivascular and periductal regions [2,6]. The presence of retinoids participates in their quiescent state maintenance by inhibiting α -SMA expression [7]. Even though to be in an inactive state, qPSCs play a crucial role in maintaining pancreatic tissue architecture by modulating the ECM turnover though ECM protein degradation and synthesis [8]. Furthermore, the expression of toll-like receptors (TLRs 2-5) in rat qPSCs, suggests that they are also implicated in innate immunity [9]. However, it is still unknown whether human qPSCs possess TLRs as well; since they are hardly accessibles.

Nevertheless, in response to pancreatic injury or inflammation, qPSCs undergo morphological and functional transformations to become activated. This transition is highlighted by the

acquisition of a myofibroblast-like phenotype. This latter is characterized by an increased expression of α -SMA and excessive ECM component's production and secretion, especially type I collagen synthesis, as well as intensive proliferative, migratory, and contractile abilities. All these properties define aPSCs as the major actor involved in the development of PF, resulteding from the presence of ECM large protein amount [10]. PF constitute is the main feature of three pancreatic diseases : acute (AP), and chronic pancreatitis (CP) and as well as pancreatic ductal adenocarcinoma (PDAC), and is caused by the exacerbated activation of PSCs, inducing an increased deposition and reduced degradation of ECM [11,12]. Nonetheless, during pancreatic damage, a variety of extracellular stimuli including cytokines (e.g : interleukin [IL]-1, IL-6, IL-8 and tumor necrosis factor-α), and growth factors (e.g.: platelet-derived growth factor [PDGF] and transforming growth factor β [TGF β]) trigger PSC'activation [13-16]. Alcohol metabolites [17], trypsin, thrombin, fatty and bile acids [18*,19*,20*,21**], endothelin-1 (ET-1) [22], angiotensin II (ATII) [23] and hypoxic conditions [24] participate in PSC's activation as well. In turn, aPSCs are capable of an autocrine stimulation by secreting cytokines and growth factors, in order to maintain their activated state [10,13,14,25]. Moreover, aPSCs are in permanent dialogue with their neighbor cells, involving pancreatic acinar cells (PACs) during pancreatitis [18*] and pancreatic cancer cells (PCCs) during PDAC [26]. PSC's interplay with PACs and PCCs strengthens the poor prognosis of pancreatitis and PDAC, respectively. Throughout AP, aPSCs are responsible for the perpetual PACs necrosis leading to the remplacement of the pancreatic cellular component by non-cellular fibrotic tissue, caused by the necrotic amplification loop between PACs and aPSCs [27]. This phenomenon conducts to extended AP resulting in CP, that significantly increases the risk of PDAC development [18*,19*,20*,28,29]. Furthermore, in PDAC, and in hepatocellular carcinoma, activated stellate cells have the particularity of constituting the chief source of Cancer-Associated Fibroblasts (CAFs). During PDAC, the pancreatic CAFs (pCAF) or aPSCs are liable for more than 80% of the pancreatic tumor mass, representing the main component of the dense fibrotic stroma leading to the desmoplastic reaction, tightly surrounding the PCCs [30]. aPSCs enhance PDAC spreading by raising PCCs proliferation, migration et and invasion so as to form early distant metastases. In this line, aPSCs not only provide the ideal microenvironment for pancreatic tumor development and progression, but they also feed the tumor by the paracrine stimulation of PCCs through cytokines and growth factors secretion. Since recently, few studies have identified the existence of more than one distinct aPSC/pCAF subtypes, underlining their inter- and intra-tumor heterogeneity. This latter is defined from the cell surface marker expression, cell signaling, cytokine production, aPSC-PCC interactions, and gene expression, which differs between each subtype [31]. Among others, Öhlund et al.'s study, is of important interest, proposing two main aPSC subpopulations according to their spatial location. One subpopulation that interacts directly with the PCCs, characterized by high α -SMA expression, called myofibroblastic CAFs, and one located more distantly from the PCCs, defined by high IL-6, IL-11 and leukemia inhibitory factor expression and low α -SMA expression, called inflammatory CAFs [32]. Neuzillet et al. highlighted the importance of such aPSC heterogeneity by identifying four more pCAF subpopulations, classified depending on their : α-SMA and vimentin expression, ECM production, proliferation rate, immunogenic profile and role in PDAC prognosis [33]. All these recent studies brought a new feature of aPSC functioning, showing that according to the subtype, they can promote more or less PDAC progression. In addition, they restrict oxygen availability maintaining the ideal hypoxic conditions for the tumor, as well as they limit inflammatory infiltration and delivery of chemotherapeutic agents increasing PDAC chemoresistance. Contrary to PACs, there is a positive amplification loop between aPSCs and PCC in order to stimulate PDAC propagation [34-38]. It seems that aPSCs are the key link between these three pancreatic diseases and therefore understanding and limiting PSC's activation may be the key of for their mediacal treatment.

Distinguished Ca²⁺ signalling between quiescent and activated PSCs

Impaired Ca^{2+} homeostasis is one of the critical events triggering pancreatic failure, leading to AP, CP, or PDAC. However, the involvement of Ca^{2+} signalling in PSC's pathophysiology remains relatively little explored hitherto.Won *et al.* [21**] were the first to investigate the Ca^{2+} signature in mouse PSCs from healthy pancreas as well as from and a model of chronic pancreatitis, showing evidence of distinguished intracellular Ca^{2+} signaling between qPSCs and aPSCs.

Believed for a longtime to existe in a « quiescent » state under physiological conditions, qPSCs are far away from being really « inactive», since they can generate Ca^{2+} signals in response to extracellular stimuli. Won *et al.* **[21**]** nicely demonstrated that qPSCs can produce Ca^{2+} signals in response to the muscarinic agonist carbachol, ATII, ATP, and bradykinin (BK). This latter finding was supported by Gryshchenko *et al.* **[19*]**, showing that, in fact, qPSCs can respond to BK concentrations (100 pM), close to the ones present during AP (140 pM), through the activation of BK type 2 receptors. Indeed, Once linked to its receptor, BK generates a biphasic Ca^{2+} response, in qPSCs, represented by an initial large cytosolic Ca^{2+} transient signal followed by a sustained Ca^{2+} plateau. The Ca^{2+} transient signal

is caused by Ca²⁺ release from the endoplasmic reticulum (ER) stores, after phospholipaese C (PLC) and inositol triosphosphate receptor (InsP3R) activation, followed by a sustained while the Ca^{2+} plateau is induced by Ca^{2+} entry through Ca^{2+} release-activated Ca^{2+} (CRAC) channels. BK-elicited Ca²⁺ signals in qPSCs may play a key role in PSC transition transformation into the from the quiescent to activated state, since once activated, they respond to a much higher concentration of BK, involving BK type 1 receptor, which is a hallmark of inflammatory diseases, such as pancreatitis [18*,19*]. Moreover, BK-mediated Ca^{2+} signalling changes in aPSCs, inducing only a single cytosolic Ca^{2+} transient signal. Furthermore, qPSCs but also aPSCs as well, are capable of producing sustained toxic cytosolic Ca²⁺ signals in response to bile acids (BAs), such as sodium cholate and taurocholate, activating intracellular InsP3R-Ca²⁺ release and thus Ca²⁺ entry via CRAC channels [20*]. In turn, BK and BAs-induced cytosolic Ca²⁺ as well as and hydrogen peroxide-mediated Ca²⁺, activates the Ca²⁺/calmodulin-dependent enzyme nitric oxide synthase in order to produce a diffusible nitric oxide-mediated inflammatory response, acting on the PAC neighbor cells. This phenomenon triggers an amplification loop by promoting PAC's death and increasing PSC-stimulating factor's secretion from dying PACs and therefore aggravate AP, rising the risk to transform into CP [39*]. Another exeample of different Ca²⁺ signalling toolkit between qPSCs and aPSCs, is their responsiveness to ATP. Indeed, aPSCs are approximately threefold more sensitives to ATP than qPSCs. Won et al. [21**] established that ATP-mediated Ca²⁺ signals are initiated near to the nucleus and propagated into it, increasing nucleoplasmic Ca^{2+} concentrations in aPSCs, which stimulates their proliferation during CP. In accordance with these data, some other authors reported that ATP and but also UTP and UDP-induced intracellular Ca^{2+} signals are mediated through several types of purinergic receptors P2X and P2Y in aPSCs. Hennigs et al. [40] showed demonstrated that $P2X_4$ and $P2Y_2$ (shown to be upregulated in CP) are the most abundantly expressed isoforms in aPSCs. Interestingly, Haanes et al. [41] completed these findings, by reporting the importance of ATP-induced Ca^{2+} signals mediated by P2X₇, shown to be overexpresed and playing an important role in aPSCs proliferation, in CP. The authors concluded that depending on ATP concentration, P2X7 can drive PSC's proliferation (at low ATP concentration) or death (at high ATP doses). These could explain, in part, the high sensitivity of aPSCs to ATP compared to qPSCs, so they can maintain their activated state by promoting their $\frac{1}{2}$ proliferation in a Ca²⁺-dependent manner, leading to pancreatic fibrosis development. Giannuzzo et al. [42] highlighted the significance of P2X7 also in collagen's deposition during the desmoplastic reaction in PDAC. Nevertheless, contrary to qPSCs, only

aPSCs produce prominent nuclear Ca²⁺ signals in response to protease-activated receptors 1 driven by thrombin and PDGF, stimulating PSC proliferation [21**]. Furthermore, ET-1 and ATII, have been reported to be implicated in fibrosis progression by increasing Ca²⁺- dependent contractili¹/₁ty and proliferation of aPSCs. Moreover, only aPSCs can evoke intracellular Ca²⁺ signals in response to ET-1 through ET_A receptor [22], while both qPSCs and aPSCs respond to ATII *via* AT₁ receptor [23]. In contrast, some other agents, like melatonin [43,44] or the pesticide dimethoate [45], impair PSC's viability even though they promote Ca²⁺ mobilization from the ER.

Ca²⁺ signaling and ion channels in aPSCs

 Ca^{2+} entry is controlled by a large number of Ca^{2+} -permeable ion channels localized at the plasma membrane and PSCs have been shown to also express a number of Ca^{2+} channels.

Ion channels and pancreatitis

Zhang *et al.* [**46**^{**}] were the first to report the presence of transient receptor potential (TRP) channels, *et* particularly of TRPV4 in aPSCs from rat model of CP. The authors found demonstrated an upregulated TRPV4 expression in CP conditions compared to control, which is in concordance with the observed increased and sustained intracellular Ca²⁺ mobilization. Their study revealed that TRPV4-mediated Ca²⁺ mobilization is enhanced by tumor necrosis factor- α stimulation, showing the importance of cytokines in Ca²⁺ signalling modulation in the aPSCs activation. A Ffew years after, Gryshchenko *et al* [**19**^{*}], pharmacologically identified the presence of CRAC channels, suggesting a CRAC-mediated Ca²⁺ entry in both qPSCs and aPSCs. Their findings were clearly supported recently by Waldron *et al.* [**47**^{**}], who highlighted the expression of Orai1, Orai2, and Stim1 proteins, the main constituent of CRAC channels, in aPSCs of AP model. Using a CRAC channel inhibitor, they observed an inhibition of TLR-mediated fibro-inflammatory signals, leading to a decrease expression, among others, of IL-6, TGF_{β1}, type I collagen, and α-SMA, proposing a crucial role of CRAC-mediated Ca²⁺ entry in cytokine expression from aPSCs in AP.

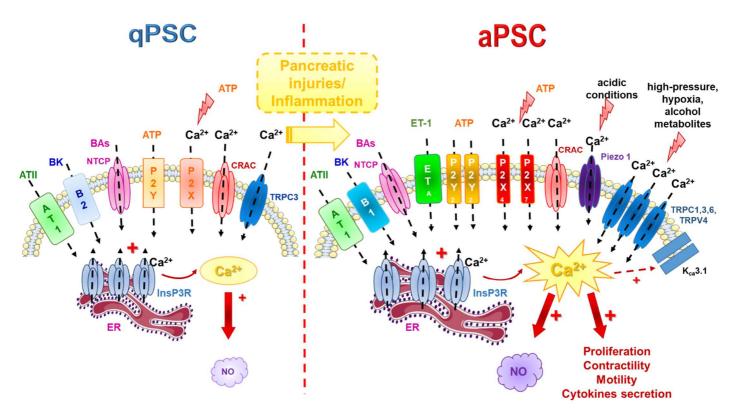
Ion channels and PDAC

Several reports have been focusing on the role of PSC-expressed ion channels in the context of PDAC as well. Fels *et al.* **[48**]** firstly established a role of TRPC1 ion channel in the activation of PSCs, under high-pressure application, which is one of the PDAC microenvironment features and one of PSC's activator stimuli, thereby perpetuating fibrosis

formation. They demonstrated that pressureion induces TRPC1 activation allowing Ca²⁺ influx into PSCs in order to regulate their migration and enhance their activated behaviour. Similar to pressure, it has been shown by Nielsen *et al.* [49**] that hypoxia, another hallmark of PDAC microenvironment, known to activate PSCs, induces an activation of TRPC6 leading to an increase of Ca²⁺ influx, which promotes aPSC's migration and promigratory factor's secretion by aPSCs. The report of Storck et al. [50**] brought evidence of cooperative regulation of PSC's migration, mediated by TRPC3 (shown to be upregulated in the PDAC stroma) and the calcium-activated potassium channel K_{Ca}3.1, under PDGF stimulation, known to activate PSCs. The authors intelligently demonstrated that Ca²⁺ entry through TRPC3 is involved into neighboring K_{Ca}3.1 channel's activation, resulting in increased PDGF-stimulated migration of PSCs. These data confirm, once more, the importance of cytokines in the regulation of PSC's Ca^{2+} signalling. To complete The key role of PDAC mircroenvironment on PSC's ion channels-mediated Ca²⁺ influx in the perpetuation of cancer progression, was also documented by Kuntze et al. [51**]. Their work determined the impact of Piezo1-mediated Ca²⁺ influx on PSC's physiopathophysiology, under acidification conditions, also characteristic of PDAC microenvironment. It appears that Indeed Piezo1 activity is modified reduced upon acidification, causing leading to Ca²⁺ influx decrease, The reduction of Piezo 1 activity would be probably due to the protonation-caused inhibition, in order to protect aPSCs from cell death and promote their migration into wellperfused invasive tumor regions where Piezo1 can be fully functionnal. Therefore, under physiological pH, the increased Piezo1 mediated-Ca²⁺ influx enhances PSC's migration as well as and myosin light chain isoform 9 phosphorylation, leading to aPSC's cytoskeletal architecture modification, which, in turn, increases aPSC's motility.

Conclusion

To summarize, pancreatic injury or inflammation induces a disequilibrium in PSC's composition, leading to an increased number of aPSCs, due to the transition of qPSCs to aPSCs. Taken together, it seems that in both qPSCs and aPSCs, Ca^{2+} signalling is initiated through the canonical pathway, involving InsP3R-mediated Ca^{2+} release from the ER, under diverse extracellular stimuli. Moreover, in aPSCs, ion channels-mediated Ca^{2+} influx apperars to be a key effector for the development and progression of PF and PDAC, as summearized in the conclusion schemae (**Figure 1**). And what if managing PSC's overloaded intracellular Ca^{2+} , by a combination of two or more ion channel inhibitors, is the answer for controlling PSC's transition and so PF and PDAC? Hence, controlling the transition of qPSCs to aPSCs



will limit pancreatic fibrosis formation and so PF and PDAC promotion. May the answer to this challenge is laying on the management of PSC's intracellular Ca^{2+} by using a combination of two or more ion channel inhibitors.

Figure 1 : Concluding model that distinguish the Ca²⁺ signature between quiescent (qPSCs) and activated PSCs (aPSCs). In response to pancreatic injury or inflammation qPSC undergo transformations to become activated. During this transition, the cell surface expression and function of several receptors and ion channels are modified. qPSCs are characterized by bradykinin type 2 receptor activation, while in aPSCs, bradykinin activates type 1 receptor. Moreover, aPSCs are defined by an overexpression of P2Y₂ receptor, P2X₄ and P2X₇ channels, as well as TRPC3 and TRPV4 channels. However, both qPSCs and aPSCs express AT1 receptors, NTCP transporters and CRAC channels but only aPSCs express ET_A receptor.

ATII = angiotensin II, AT1 = angiotensin II type 1 receptor, BK = bradykinin, B1 = bradykinin type 1 receptor, B2 = bradykinin type 2 receptor, BAs = bile acids, CRAC = Ca^{2+} release-activated Ca^{2+} channels, ER = endoplasmic reticulum, ET-1 = endothelin-1, ET_A = endothelin type A receptor, InsP3R = inositol trisphosphate receptor, NO = nitric oxide, NTCP = sodium-taurocholate cotransporting polypeptide, P2Y = purinergic receptors, P2X = purinergic channels, TRPC = transient receptor potential-canonical channels, TRPV = transient receptor potential-vanilloid channels

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CONFLICT OF INTERESTS STATEMENT

The authors declare no conflicts of interest

References

- 1. Apte M, Pirola R, Wilson J: Pancreatic stellate cell: physiologic role, role in fibrosis and cancer. *Curr Opin Gastroenterol* 2015, **31**:416–423.
- 2. Bachem MG, Schneider E, Gross H, Weidenbach H, Schmid RM, Menke A, Siech M, Beger H, Grünert A, Adler G: Identification, culture, and characterization of pancreatic stellate cells in rats and humans. *Gastroenterology* 1998, **115**:421–432.
- 3. Apte MV, Park S, Phillips PA, Santucci N, Goldstein D, Kumar RK, Ramm GA, Buchler M, Friess H, McCarroll JA, et al.: **Desmoplastic reaction in pancreatic cancer: role of pancreatic stellate cells**. *Pancreas* 2004, **29**:179–187.
- 4. Berridge MJ, Bootman MD, Lipp P: Calcium a life and death signal. *Nature* 1998, **395**:645–648.
- 5. Petersen OH, Michalak M, Verkhratsky A: Calcium signalling: Past, present and future. *Cell Calcium* 2005, **38**:161–169.
- Apte MV, Haber PS, Applegate TL, Norton ID, McCaughan GW, Korsten MA, Pirola RC, Wilson JS: Periacinar stellate shaped cells in rat pancreas: identification, isolation, and culture. *Gut* 1998, 43:128–133.
- 7. McCarroll JA, Phillips PA, Santucci N, Pirola RC, Wilson JS, Apte MV: Vitamin A inhibits pancreatic stellate cell activation: implications for treatment of pancreatic fibrosis. *Gut* 2006, **55**:79–89.
- 8. Phillips PA, Yang L, Shulkes A, Vonlaufen A, Poljak A, Bustamante S, Warren A, Xu Z, Guilhaus M, Pirola R, et al.: Pancreatic stellate cells produce acetylcholine and may play a role in pancreatic exocrine secretion. *Proc Natl Acad Sci U S A* 2010, **107**:17397–17402.
- 9. Masamune A, Kikuta K, Watanabe T, Satoh K, Satoh A, Shimosegawa T: Pancreatic stellate cells express Toll-like receptors. J Gastroenterol 2008, **43**:352–362.
- 10. Ferdek PE, Jakubowska MA: Biology of pancreatic stellate cells—more than just pancreatic cancer. *Pflugers Arch* 2017, **469**:1039–1050.
- 11. Jaster R, Emmrich J: Crucial role of fibrogenesis in pancreatic diseases. Best Pract Res Clin Gastroenterol 2008, 22:17–29.
- 12. Apte MV, Pirola RC, Wilson JS: Pancreatic stellate cells: a starring role in normal and diseased pancreas. *Front Physiol* 2012, **3**.

- 13. Apte M, Haber P, Darby S, Rodgers S, McCaughan G, Korsten M, Pirola R, Wilson J: Pancreatic stellate cells are activated by proinflammatory cytokines: implications for pancreatic fibrogenesis. *Gut* 1999, **44**:534–541.
- Schneider E, Schmid-Kotsas A, Zhao J, Weidenbach H, Schmid RM, Menke A, Adler G, Waltenberger J, Grünert A, Bachem MG: Identification of mediators stimulating proliferation and matrix synthesis of rat pancreatic stellate cells. *Am J Physiol-Cell Physiol* 2001, 281:C532– C543.
- 15. Vogelmann R, Ruf D, Wagner M, Adler G, Menke A: Effects of fibrogenic mediators on the development of pancreatic fibrosis in a TGF-β1 transgenic mouse model. *Am J Physiol-Gastrointest Liver Physiol* 2001, **280**:G164–G172.
- 16. Shimada M, Andoh A, Hata K, Tasaki K, Araki Y, Fujiyama Y, Bamba T: **IL-6 secretion by human** pancreatic periacinar myofibroblasts in response to inflammatory mediators. *J Immunol Baltim Md 1950* 2002, **168**:861–868.
- 17. Apte MV, Phillips PA, Fahmy RG, Darby SJ, Rodgers SC, McCaughan GW, Korsten MA, Pirola RC, Naidoo D, Wilson JS: **Does alcohol directly stimulate pancreatic fibrogenesis? Studies with rat pancreatic stellate cells**. *Gastroenterology* 2000, **118**:780–794.
- Gryshchenko O, Gerasimenko JV, Peng S, Gerasimenko OV, Petersen OH: Calcium signalling in
 the acinar environment of the exocrine pancreas: physiology and pathophysiology. *J Physiol* 2018, 596:2663–2678.

In this work, the authors highlight the change to bradykinin responsiveness between quiescent and activated pancreatic stellate cells, in acute pancreatitis models.

19. Gryshchenko O, Gerasimenko JV, Gerasimenko OV, Petersen OH: Ca²⁺ signals mediated by
 bradykinin type 2 receptors in normal pancreatic stellate cells can be inhibited by specific Ca²⁺ channel blockade. J Physiol 2016, 594:281–293.

Bradykinin type 2 receptor mediates bradykinin-induced Ca²⁺ signals leading to Ca²⁺ release from endoplasmic reticulum stores and so Ca²⁺ release-activated Ca²⁺ channel's activation, in normal pancreatic stellate cells.

- 20. Ferdek PE, Jakubowska MA, Gerasimenko JV, Gerasimenko OV, Petersen OH: Bile acids induce
- * necrosis in pancreatic stellate cells dependent on calcium entry and sodium-driven bile uptake. *J Physiol* 2016, **594**:6147–6164.

This report shows that sodium cholate and taurocholate bile acid salts entry induces Ca²⁺ release through the inositol triphosphate receptor from the endoplasmic reticulum stores, in quiescent and activated pancreatic stellate cells.

- 21. Won JH, Zhang Y, Ji B, Logsdon CD, Yule DI: Phenotypic changes in mouse pancreatic stellate cell
- ** Ca²⁺ signaling events following activation in culture and in a disease model of pancreatitis. Mol Biol Cell 2011, 22:421–436.

Quiescent and activated pancreatic stellate cells are characterized by a different Ca²⁺ signature. In activated pancreatic stellate cells, nuclear Ca²⁺ signals promote cell proliferation.

22. Klonowski-Stumpe H, Reinehr R, Fischer R, Warskulat U, Lüthen R, Häussinger D: **Production and Effects of Endothelin-1 in Rat Pancreatic Stellate Cells**. *Pancreas* 2003, **27**:67–74.

- 23. Reinehr R, Zoller S, Klonowski-Stumpe H, Kordes C, Häussinger D: Effects of Angiotensin II on Rat Pancreatic Stellate Cells. *Pancreas* 2004, **28**:129–137.
- 24. Masamune A, Kikuta K, Watanabe T, Satoh K, Hirota M, Shimosegawa T: **Hypoxia stimulates** pancreatic stellate cells to induce fibrosis and angiogenesis in pancreatic cancer. *Am J Physiol Gastrointest Liver Physiol* 2008, **295**:G709-717.
- 25. Luttenberger T, Schmid-Kotsas A, Menke A, Siech M, Beger H, Adler G, Grünert A, Bachem MG: Platelet-derived growth factors stimulate proliferation and extracellular matrix synthesis of pancreatic stellate cells: implications in pathogenesis of pancreas fibrosis. *Lab Investig J Tech Methods Pathol* 2000, **80**:47–55.
- 26. Allam A, Thomsen AR, Gothwal M, Saha D, Maurer J, Brunner TB: **Pancreatic stellate cells in pancreatic cancer: In focus**. *Pancreatol Off J Int Assoc Pancreatol IAP AI* 2017, **17**:514–522.
- 27. Hegyi P: Necrotic amplification loop in acute pancreatitis: pancreatic stellate cells and nitric oxide are important players in the development of the disease. *J Physiol* 2018, **596**:2679–2680.
- 28. Gerasimenko JV, Peng S, Tsugorka T, Gerasimenko OV: **Ca2+ signalling underlying pancreatitis**. *Cell Calcium* 2018, **70**:95–101.
- 29. Gerasimenko JV, Gryshchenko O, Ferdek PE, Stapleton E, Hébert TOG, Bychkova S, Peng S, Begg M, Gerasimenko OV, Petersen OH: Ca2+ release-activated Ca2+ channel blockade as a potential tool in antipancreatitis therapy. *Proc Natl Acad Sci U S A* 2013, **110**:13186–13191.
- 30. Norton J, Foster D, Chinta M, Titan A, Longaker M: Pancreatic Cancer Associated Fibroblasts (CAF): Under-Explored Target for Pancreatic Cancer Treatment. *Cancers* 2020, **12**.
- 31. Pereira BA, Vennin C, Papanicolaou M, Chambers CR, Herrmann D, Morton JP, Cox TR, Timpson P: **CAF Subpopulations: A New Reservoir of Stromal Targets in Pancreatic Cancer**. *Trends Cancer* 2019, **5**:724–741.
- 32. Öhlund D, Handly-Santana A, Biffi G, Elyada E, Almeida AS, Ponz-Sarvise M, Corbo V, Oni TE, Hearn SA, Lee EJ, et al.: **Distinct populations of inflammatory fibroblasts and myofibroblasts in pancreatic cancer**. *J Exp Med* 2017, **214**:579–596.
- 33. Neuzillet C, Tijeras-Raballand A, Ragulan C, Cros J, Patil Y, Martinet M, Erkan M, Kleeff J, Wilson J, Apte M, et al.: Inter- and intra-tumoural heterogeneity in cancer-associated fibroblasts of human pancreatic ductal adenocarcinoma. *J Pathol* 2019, **248**:51–65.
- 34. Habisch H, Zhou S, Siech M, Bachem MG: Interaction of stellate cells with pancreatic carcinoma cells. *Cancers* 2010, **2**:1661–1682.
- 35. Vonlaufen A, Joshi S, Qu C, Phillips PA, Xu Z, Parker NR, Toi CS, Pirola RC, Wilson JS, Goldstein D, et al.: **Pancreatic stellate cells: partners in crime with pancreatic cancer cells**. *Cancer Res* 2008, **68**:2085–2093.
- 36. Sousa CM, Biancur DE, Wang X, Halbrook CJ, Sherman MH, Zhang L, Kremer D, Hwang RF, Witkiewicz AK, Ying H, et al.: Pancreatic stellate cells support tumour metabolism through autophagic alanine secretion. *Nature* 2016, **536**:479–483.
- 37. Schneiderhan W, Diaz F, Fundel M, Zhou S, Siech M, Hasel C, Möller P, Gschwend JE, Seufferlein T, Gress T, et al.: **Pancreatic stellate cells are an important source of MMP-2 in human**

pancreatic cancer and accelerate tumor progression in a murine xenograft model and CAM assay. *J Cell Sci* 2007, **120**:512–519.

- 38. Hwang RF, Moore T, Arumugam T, Ramachandran V, Amos KD, Rivera A, Ji B, Evans DB, Logsdon CD: Cancer-associated stromal fibroblasts promote pancreatic tumor progression. *Cancer Res* 2008, **68**:918–926.
- 39. Jakubowska MA, Ferdek PE, Gerasimenko OV, Gerasimenko JV, Petersen OH: Nitric oxide signals
 are interlinked with calcium signals in normal pancreatic stellate cells upon oxidative stress and inflammation. Open Biol 2016, 6.

Bradykinin mediated-Ca²⁺ signals, sodium cholate and taurocholate-induced Ca²⁺ signals stimulate nitric oxide generation in quiescent and activated pancreatic stellate cells.

- 40. Hennigs J, Seiz O, Spiro J, Berna M, Baumann H, Klose H, Pace A: **Molecular Basis of P2-Receptor-Mediated Calcium Signaling in Activated Pancreatic Stellate Cells**. *Pancreas* 2011, **40**:740–746.
- 41. Haanes KA, Schwab A, Novak I: The P2X7 Receptor Supports Both Life and Death in Fibrogenic Pancreatic Stellate Cells. *PLoS ONE* 2012, **7**.
- 42. Giannuzzo A, Saccomano M, Napp J, Ellegaard M, Alves F, Novak I: **Targeting of the P2X7** receptor in pancreatic cancer and stellate cells. *Int J Cancer* 2016, **139**:2540–2552.
- Santofimia-Castaño P, Garcia-Sanchez L, Ruy DC, Sanchez-Correa B, Fernandez-Bermejo M, Tarazona R, Salido GM, Gonzalez A: Melatonin induces calcium mobilization and influences cell proliferation independently of MT1/MT2 receptor activation in rat pancreatic stellate cells. *Cell Biol Toxicol* 2015, **31**:95–110.
- 44. Estaras M, Moreno N, Santofimia-Castaño P, Martinez-Morcillo S, Roncero V, Blanco G, Lopez D, Fernandez-Bermejo M, Mateos JM, Iovanna JL, et al.: **Melatonin induces reactive oxygen species generation and changes in glutathione levels and reduces viability in human pancreatic stellate cells**. *J Physiol Biochem* 2019, **75**:185–197.
- 45. Martínez-Morcillo S, Pérez-López M, Soler-Rodríguez F, González A: **The organophosphorus** pesticide dimethoate decreases cell viability and induces changes in different biochemical parameters of rat pancreatic stellate cells. *Toxicol In Vitro* 2019, **54**:89–97.
- 46. Zhang LP, Ma F, Abshire SM, Westlund KN: Prolonged high fat/alcohol exposure increases
- **** TRPV4** and its functional responses in pancreatic stellate cells. *Am J Physiol Regul Integr Comp Physiol* 2013, **304**:R702–R711.

TRPV4 expression and TRPV4-mediated Ca²⁺ mobilization increase within pancreatic stellate cells during chronic pancreatitis. The latter is stimulated by the TNF- α cytokine in activated pancreatic stellate cells.

47. Waldron RT, Chen Y, Pham H, Go A, Su H-Y, Hu C, Wen L, Husain SZ, Sugar CA, Roos J, et al.: The

** Orai Ca2+ channel inhibitor CM4620 targets both parenchymal and immune cells to reduce inflammation in experimental acute pancreatitis. *J Physiol* 2019, **597**:3085–3105.

This is the first study identifying Orai1, Orai2 and STIM1 expression in mouse activated pancreatic stellate cells. The authors demonstrate pharmacologically that Orai-Ca²⁺ channels regulate fibro-inflammatory gene expression in mouse activated pancreatic stellate cells.

48. Fels B, Nielsen N, Schwab A: Role of TRPC1 channels in pressure-mediated activation of murine
 ** pancreatic stellate cells. *Eur Biophys J* 2016, 45:657–670.

High pressure stimulates TRPC1-mediated Ca²⁺ influx in order to regulate pancreatic stellate cell's migration, within pancreatic ductal adenocarcinoma microenvironment.

- 49. Nielsen N, Kondratska K, Ruck T, Hild B, Kovalenko I, Schimmelpfennig S, Welzig J, Sargin S,
- ** Lindemann O, Christian S, et al.: **TRPC6 channels modulate the response of pancreatic stellate** cells to hypoxia. *Pflüg Arch Eur J Physiol* 2017, **469**:1567–1577.

Pancreatic stellate cell's migration and promigratory factor's secretion is modulated by TRPC6-induced Ca²⁺ influx under hypoxic conditions.

- 50. Storck H, Hild B, Schimmelpfennig S, Sargin S, Nielsen N, Zaccagnino A, Budde T, Novak I, Kalthoff
- ** H, Schwab A: Ion channels in control of pancreatic stellate cell migration. Oncotarget 2016, 8:769–784.

This work reveals a functionnal cooperation between TRPC3 and K_{Ca} 3.1 channels in order to regulate pancreatic stellate cell's migration in pancreatic ductal adenocarcinoma stoma.

- 51. Kuntze A, Goetsch O, Fels B, Najder K, Unger A, Wilhelmi M, Sargin S, Schimmelpfennig S,
- ** Neumann I, Schwab A, et al.: Protonation of Piezo1 Impairs Cell-Matrix Interactions of Pancreatic Stellate Cells. Front Physiol 2020, 11.

Piezo1 mechanosensitive channel participate in the modulation of pancreatic stellate cell's migration through the Ca²⁺ influx. However Piezo1-mediated Ca²⁺ influx is modified under acidic conditions, in the pancreatic tumor stroma.