

Soil aggregation, ecosystem engineers and the C cycle

Patrick Lavelle, Alister Spain, Steven Fonte, Jose Camilo Bedano, Eric Blanchart, Victor Galindo, Michel Grimaldi, Juan Jose Jimenez, Elena Velasquez, Anne Zangerlé

▶ To cite this version:

Patrick Lavelle, Alister Spain, Steven Fonte, Jose Camilo Bedano, Eric Blanchart, et al.. Soil aggregation, ecosystem engineers and the C cycle. Acta Oecologica, 2020, 105, pp.103561 -. 10.1016/j.actao.2020.103561 . hal-03490669

HAL Id: hal-03490669 https://hal.science/hal-03490669v1

Submitted on 20 May 2022

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Soil aggregation, ecosystem engineers and the C cycle Patrick Lavelle¹, Alister Spain², Steven Fonte³, Jose Camilo Bedano⁴, Eric Blanchart⁵, Victor Galindo⁶, Michel Grimaldi⁷, Juan Jose Jimenez⁸, Elena Velasquez⁹, Anne Zangerlé¹⁰ 1. Université Paris Sorbonne, IEES-BIODIS, Paris, France 2. The University of Western Australia, Perth, Australia 3. Colorado State University, Fort Collins, USA 4. National University of Río Cuarto, Córdoba, Argentina 5. IRD, UMR Eco&Sols, Montpellier, France 6. Universidad del Valle, Cali, Colombia 7. Centre IRD, IEES, Centre IRD, Bondy, France 8. Instituto Pirenaico de Ecologia-CSIC, Jaca, Huesca, Spain 9. Universidad Nacional de Colombia, Palmira, Colombia 10. Ministry of Agriculture, Viticulture and Rural Development. ASTA, Luxemburg

HIGHLIGHTS

32 33

1. Invertebrates and roots are essential drivers of soil aggregation, but often overlooked.

35 36

2. Manual separation and NIR spectroscopy allow classifying macro-aggregates according to the physical or biogenic agents that produced them.

3738

39 3. A simple field method is proposed to measure in situ rates of production and losses in40 the different macro-aggregate pools.

41

42 4. Stocks, inputs and losses of C in aggregate pools characterize their contribution to C cycling and conservation.

44 45

46

5. This simple and robust approach can support the identification of management practices that best store and conserve C in soils

47 48

50

53

54

49

51 Abstract: Soil aggregation and its effects on soil C storage have been addressed in

52 thousands of research articles over the last 40 years. Research has been mostly focussed

on the resistance of aggregates to mechanical disruption and the role of organic matter in

- aggregate stabilization. On the other hand, relatively little attention has been paid to
- 55 identifying the microbial, plant root and macro-invertebrate actors and physical processes
- that continuously create and destroy aggregates. The sum and dynamics of these
- 57 processes determines the ability of soils to store and conserve C. Understanding the
- 58 interactions between aggregation dynamics and C transformations in soils therefore
- requires a precise identification of the agents that produced aggregates and knowledge of
- 60 the rates of formation and persistence in the pools thus identified.
- We propose to separate macro-aggregated components of different, physicogenic and
- 62 biogenic origins from non-macro-aggregated soil on a morphological basis, using a simple
- or physico-chemical agent which produced each
- 64 individual macro-aggregate can then be determined using Near Infrared Spectrometry
- 65 (NIRS). A general description of the distribution and quality of organic matter among the

different groups of macro-aggregates can be made. Simple soil re-aggregation or disaggregation test conducted in field conditions further measure the production of different macro-aggregates with time and their mean residence times in the studied soil. Respirometry measurements on each recognized category of macro-aggregates evaluate the respective C losses through respiration. The methods described here will allow the dominant pathways of C flow at a given site to be characterized and possible management options to increase C storage identified. We finally discuss the different assumptions made to build this simple model and offer ways to test the methodology under field conditions.

1. Introduction

While improved soil management offers one of many promising approaches for climate change mitigation, soil scientists and technicians are increasingly faced with the challenge of proposing and testing viable strategies to enhance C sequestration in soils (Altieri et al., 2015; Poeplau and Don, 2015; Paustian et al, 2016; Powlson et al., 2016; Smith et al., 2016; Bedano et al., 2019). As a major C reservoir, soil organic matter on Earth is estimated to be 1500 to 2400 Gt C, that is more than twice the mass of the C contained in the atmosphere (860 Gt). In addition, soils currently sequester an additional 3.2 ± 0.7 Gt C annually, equivalent to 29.4% of the annual emissions from fossil fuel burning. However, this positive contribution is offset each year by 1.5 \pm 0.7 Gt C emitted as a consequence of land use change and soil degradation (Le Quéré et al. 2018). We currently do not know whether soils will continue to store C as climate change accelerates, or lose it -as land degradation continues- and this represents a major uncertainty for climate change projections (Carvalhais et al. 2014). The international 4 per 1000 initiative proposes offsetting anthropogenic emissions by storing every year an extra 0.4 % in agricultural soils through adequate management options (Minasny et al., 2017). Such a challenge will require precise tools for implementation and monitoring at plot and farm scales. We argue that our

understanding of the mechanisms involved in the stabilization of C in soils and ways to harness our knowledge under field conditions need to be improved.

96 97

98

95

The Importance of physical protection for organic matter conservation

99 Three processes allow organic matter conservation in soils: physical protection in 100 aggregated structures, the association of transformed organic matter to mineral particles in 101 organo-mineral complexes and chemical recalcitrance (Six et al., 2004). We argue that 102 these processes are not independent and soil aggregation is expected to be a key process 103 in facilitating all three (Amezketa, 1999; Sollins et al., 1996; Feller and Beare, 1997; Six et 104 al., 2000a, 2002a and b; 2004; Von Lutzow et al., 2006). Physical protection is likely a first 105 step in conservation, that slows down the mineralization process by isolation of microbes 106 from their organic substrates and/or limits water and oxygen supplies; Kuzyakov et al., 107 2015; Negassa et al., 2015; Keiluweit et al., 2016). Suitable conditions are then created for 108 the other two processes to occur and a great part of SOM storage in soils may be in the 109 form of organo-mineral complexes (Cotrufo et al., 2019). Organic matter accumulation has 110 positive feed backs on in turn is widely acknowledged as an essential component for 111 aggregate formation and stabilization (Amezketa, 1999; Six et al., 2004; Abiven et al, 2009; 112 Fultz et al., 2013; Gumus and Seker, 2015; Zhu et al., 2017). However, a variable proportion of the leaf and root litter Carbon may have been 113 mineralized before these complexes are formed. It depends on the suite of initial 114 115 decomposition processes that associate physical processes like comminution, transfers in 116 the soil profile, inclusion in aggregate structures and chemical transformations associated 117 to digestion and humification (Lavelle and Spain, 2001). In determined conditions of climate, soil and plant cover, the stabilization of C in soils, in particulate (POM) or mineral 118 119 associated (MAOM) forms thus depends to a great extent on the efficiency of physical 120 protection of organic matter against decay at macro- or microsite scales (Parton et al., 121 1988, Lavelle, 2002; Jimenez and Lal., 2006; Kuzyakov et al., 2015). 122 The long admitted alternative option for accumulation in soils of C pools made recalcitrant 123 because of their chemical compositions is now strongly challenged (Lehmann et al., 2015).

Priming effects triggered by water-soluble organic compounds migrating down the soil profile or released by roots and invertebrate soil ecosystem engineers actually allow the mineralization of substrates reputed to be highly recalcitrant (Martin et al., 1992; Trigo et al., 1999; Fontaine et al., 2007; Lehmann et al., 2015).

There is a relative dearth of knowledge, however, of the diversity of ways aggregates are

There is a relative dearth of knowledge, however, of the diversity of ways aggregates are formed and the consequences this has on the amounts and quality of C concentrated and conserved inside these structures.

130131

132

129

What are soil aggregates??

Aggregates defined as "soil specific entities built from mineral and organic compounds with 133 134 stronger bonds between building blocks than with neighbouring particles"....."have a size, form and stability that is typical for individual soils depending on parent material and 135 136 texture, climate and vegetation, biological activity and management" (Yudina and 137 Kuzyakov, 2018). Their classification and evaluation are still largely determined by the 138 method used to view or isolate them. 139 Non-destructive viewing techniques require rather sophisticated approaches such as 140 tomography or the realization and image analysis of soil thin sections (Tracy et al., 2015; 141 Wang et al., 2016; Zhao et al., 2017; Scarciglia and Barca, 2017; Gutierrez-Castorena et al., 142 2018). Isolating techniques separate aggregates according to their resistance to physical 143 rupture following dry or wet sieving or slaking in water or other liquid substrates (Elliott, 144 1986; Le Bissonnais, 1996). Isolating may provide different results from viewing techniques 145 since the energy used to isolate may determine the size of elements separated (Ashman et 146 al., 2003; Kravchenko et al., 2018). 147 Separation of macroaggregates according to their morphology is a simple method that 148 associates viewing and isolating approaches (Topoliantz et al., 2000; Velasquez et al., 149 2007a). The major advantage is that it does not require sophisticated equipment and thus 150 can be applied by non-scientific operators, school children, students, technicians and 151 farmers to discover and quantify soil macro-aggregation.

Importance and diversity of macro-aggregates in natural soils

A wide range of possible mechanisms for aggregate formation exists, from the close association of mineral and/or organic particles in microaggregates, at microsite scales of a few tens of microns, to the production of highly organized long-lasting macro-aggregates made by soil ecosystem engineers such as ants, termites and earthworms (Lavelle et al., 1997; Tötsche et al., 2018, Zanella et al., 2018; Yudina and Kuzyakov, 2019). Aggregates frequently have a hierarchical organization and it is widely thought that microaggregates of a size < 250 µm, the most stable elements, are formed inside macroaggregates as part of their stabilization process (Six et al., 2014).

Many types of macro-aggregates of physicogenic or biogenic origin can be distinguished (Velasquez et al., 2007a; Zanella et al., 2018; Yudina and Kuzyakov, 2019. The simple visual separation of biogenic aggregates produced by roots and macroinvertebrate shows their importance in surface soil horizons: in natural soils of temperate (Pulleman et al., 2004) or tropical regions (Velasquez and Lavelle, 2019; Grimaldi et al., 2014), aggregates of biogenic origin, mainly produced by earthworm activities often represent 40 to 60% of the soil

Energy cost of forming aggregates

weight in the upper 15 cm of soil.

When considering the effects of soil aggregation on the C cycle, the release of C by respiratory activities during their construction is often overlooked. It may be an important element to consider when designing management options aimed at storing C in soils. For example, populations of the earthworm *Reginaldia omodeoi* in savannas of Ivory Coast ingest every year an estimated 850 Mg eq. dry soil and transform it into casts that are compact and highly resistant macroaggregates with a rather complex general structure (Lavelle, 1978; Blanchart,1993; Blanchart et al., 1997). The mechanical work required to burrow, ingest soil particles, take them through the gut and release them as casts is allowed by the assimilation of 9% of the C contained in the ingested soil, that is 1.2 Mg C ha⁻¹yr⁻¹ emitted as CO₂ through earthworm respiration (Lavelle, 1978). Energy cost derived

from C mineralization is expected to be lower for macro-aggregates formed by fungal entanglement of particles since hyphae likely grow in the connected porous space of the soil and do not spend energy moving particles. Data to address this point however, are lacking. The creation of physicogenic aggregates by processes like wetting/drying or freeze/thaw alternances apparently has no direct C cost.

186

187

181

182

183

184

185

Temporal stability and turnover time of aggregates

188 Aggregates experience dynamic processes of creation, stabilization, ageing, destabilization 189 and disruption (Marquez et al., 2019). A precise description of their spatio-temporal 190 dynamics and turnover is therefore an obligate step in evaluating the effect of aggregation 191 on C storage and conservation in soils. Assuming that C mineralization is either decreased 192 or stopped in aggregated structures, we need to know how much C is protected in these 193 structures, to what extent, in which forms and for how long time. 194 We actually know little of the temporal stability of aggregates of different origins and thus 195 how long these structures will conserve C before they collapse. Macro-aggregates formed 196 by entanglement of soil particles, by fine roots or mycorrhizal and other fungal hyphae, 197 generally have rather short life times, from a few days to a few months (Plante and Mc Gill, 198 2002; De Gryze et al., 2005, Segoli et al., 2013); they tend to disaggregate when the fungal 199 hyphae or root disappear and/or organic binding agents are mineralized. On the other hand, casts of the African endogeic species Reginaldia omodeoi can remain intact for long 200 201 periods of time - from 20 to 28 months - depending on soil texture and activity of 202 decompacting earthworm populations that feed on these structures and disperse them 203 (Blanchart et al., 1993b). Stabilized macro-aggregates are actually estimated to last up to 204 10 or 20 years if no disturbed (Lobe et al., 2011, Koesters et al., 2013; Marquez et al., 205 2019). 206 Many studies have monitored the build-up or decrease of aggregation as a result of 207 experimental conditions or changes in management options (Beare et al, 1994; Bronick et 208 al., 2005; Calonego and Rosolem, 2008). However, the direct measurement of aggregate 209 turnover under steady state conditions is difficult. It has only been achieved, to our

knowledge, using labelling with very specific rare earth or isotopic tracers (Plante and Mc Gill., 2002; De Gryze et al., 2005), tomography (Roose et al., 2016) or by using rather complex mechanistic simulation models (Lavelle and Meyer, 1983, Martin and Lavelle, 1992; Segoli et al., 2013; Marquez et al., 2019). These approaches can help with understanding aggregate turnover dynamics, but also have a number of limitations.

A field-based conceptual model and methodological approach

The rarity and diversity of attempts to describe aggregate life histories and dynamics is a clear indication that the problem is complex. Actually, no simple conceptual or methodological approach have been proposed so far. While much is known on the aggregation process, its diversity and importance, we argue that a few critical elements are still lacking to allow us to evaluate and accurately model the effect of soil aggregation dynamics on C conservation. This exists as a serious impediment to identify management practices and accurately predict their potential effect on soil C sequestration in a context of climate change.

This paper starts with a general bibliometric analysis. The aim was to identify knowledge gaps, that impede the implementation of a dynamic modelling of the soil aggregation process and its effect on C conservation in soils. The lack of consideration of the role of soil ecosystem engineers in general soil processes, is one such possible gap indicated by several authors (Lavelle, 2000; Bottinelli et al., 2015; Filser et al., 2016; Jouquet et al., 2016). We

protection into macro-aggregates.We finally discuss the coherence a

We finally discuss the coherence and feasibility of this approach in view of the assumptions made and identify research needs to improve it. Particular emphasis is set on providing concepts and methodologies accessible to non-specialized scientists, farmers and technicians.

then propose a simple conceptual model where the conservation of C in soil depends on its

2. Soil aggregation in the scientific literature

A search of the ISI Web of knowledge in November 2019, with the words 'soil 'and 'aggregat*' in title provided 2,692 papers, with a clear exponential increase occurring in the rates of publication on these and related topics over recent years. With the same keywords in topic, we obtained 77,427 articles. The large number of citations (>77,000), further illustrates the enormous interest in, and the importance of, this topic.

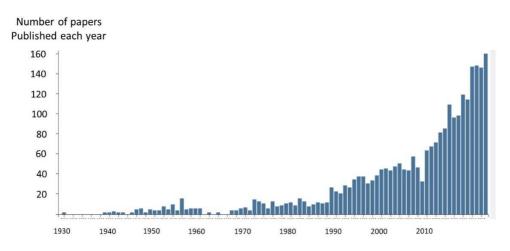


Figure 1: Number of papers published each year with words "soil" and "aggregate*" in title (source ISI Web) from 1900 to present.

The frequency of associated words in titles showed the major themes studied. **Organic matter** or **Carbon** was the most important topic associated with soil aggregates, treated in 757 (28.1%) papers. This shows the strong association perceived between the aggregation process and soil organic matter cycling. **Stability** was the second topic in importance (572 occurrences, 21.2%), a very important attribute of aggregates that measures their resistance to breakdown by physical stress, especially by water (301; 11.1%). **Soil management** (or till* or crop*) considered in 430 papers (16%) reflects the ongoing concern for physical degradation in managed soils and interest to identify options for reverting degradation. **Aggregate size** (418; 15.5%) is an important morphological attribute, considered an indicator of aggregate stability. **Water and erosion** (358; 6.7%) and **texture** (clay or text* or sand) (198; 7.4%) are studied for their important role in aggregate stabilization. In comparison, biological actors of aggregation comprised a low proportion of the papers. **Microbial** relationships with aggregation (microb* or microor* or bact* or

fung* or mycor*) were studied in only 307 (11.4%) papers, soil macroinvertebrates (earthworm* or lumbric* or termit* or ant* or formic* or macroinvert*) in 56 papers (2%), the same as roots (55; 2%). Finally, turnover of aggregates (7) or of organic matter in aggregates (9) have been very little considered in aggregate research.

3. An alternative approach to assess aggregate-associated C dynamics

In their seminal paper, Tisdall and Oades (1982) proposed a comprehensive conceptual framework to explain the origin, stabilization and dynamics of soil aggregation. They listed a number of key features that characterize the physical organization of aggregated structures and a diversity of mechanisms that provide stability against disruption. Such elements have been thoroughly described and discussed in later reviews (Elliott, 1986; Feller and Beare, 1997; Six et al., 2002a, 2014; Tötsche et al., 2018) and different elements have been introduced in recent modelling attempts (De Gryze et al., 2001, Marquez et al., 2019). Although this model has inspired a large number of studies, measuring the different compartments is still difficult and often requires rather high levels of expertise and equipment. Modelling the ageing and disruption dynamics of aggregates is even more difficult, partly because of the initial diversity of aggregate composition and structures that has not been considered so far in monitoring and modelling attempts. Stabilization or destabilization dynamics further depend on chemical, physical and biological processes associated with local soil conditions and the quality and quantity of organic material present in the aggregate.

3.1. General conceptual model

Our conceptual model is based on three simple assumptions:

1. Aggregate fractions should be separated according to their origin. This will prevent the mixing of very different aggregates into a single category based solely on their resistance to physical stress. We will recognize various pools of physicogenic or biogenic aggregates of root or invertebrate origin (Topoliantz et al., 2000; Velasquez et al., 2007a; Zanella et al., 2018). We expect each pool to have different dynamics

according to their initial diversity of compositions and structures (Hedde et al., 2005).

- 2. <u>Microaggregates are mainly formed within macroaggregates (Six et al., 2004)</u> and their dynamics are likely associated with the one of the larger structures that hosts them. We expect, however, that disaggregation of macro-aggregated structures will not go along with disaggregation of microaggregates and a pool of free microaggregates may be comprised in the non-macro-aggregated soil fraction (Tötsche et al., 2018).
- 3. Organic matter transformations are closely linked with aggregate dynamics. The incorporation of organic matter to the soil matrix is mostly a biological process associated with root growth, rhizodeposition, comminution, feeding and bioturbation processes. Flux of water-soluble organic matter that may transit through the porous space of the soil is expected to be negligible (Lavelle and Spain, 2001; Jimenez et al., 2006). Organic matter_is present in different forms and quantities in the different aggregated and non-macro-aggregated pools and associated to specific mineral components and porous spaces (Daniel et al., 1997; Capowiez et al., 2011; Arai et al., 2019). Mineralization rates are therefore likely determined by conditions that occur in aggregates.

In a given set of environmental - climate, soil and plant cover- conditions, macro-aggregates created by different processes and actors (see Fig. 2) accumulate in three compartments, which have different turnover times and mineralization rates and differently affect the overall C dynamics (Fig. 3).

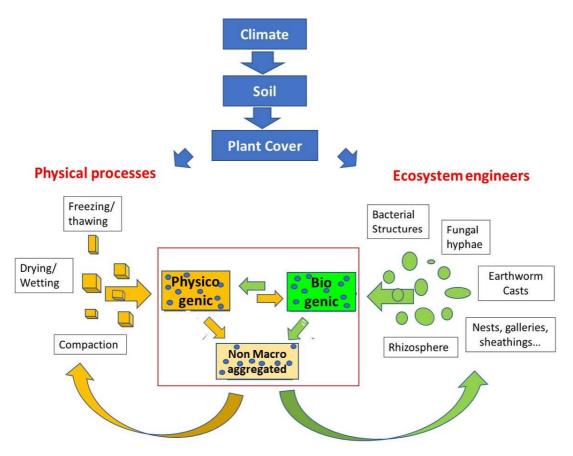
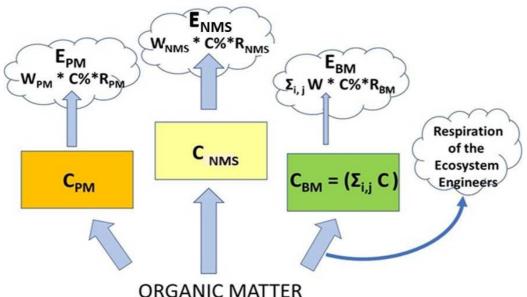


Figure 2: Conceptual framework for the diagnosis of soil macro-aggregation status and dynamics. Small circles in each category box symbolize microaggregates. Macroaggregates are classified in 3 categories, each of them having a definite number of sub categories indicated in boxes. Their dynamics involves creation from other aggregate pools, ageing and disruption that transfers their material to the non-macroaggregated pool, or a direct transformation into biogenic macroaggregates from the other 2 pools.



Fresh and transformed

323 324

Figure 3: Flow of organic matter and gas emissions through the aggregated and non-macroaggregated soil compartments.

325 326 327

328

Carbon in physical macro-aggregates (C_{PM}), non-macro-aggregated soil (C_{NMS}) and biogenic macro-aggregates (C_{BM}).

329 330 331 332 333

E_{PM}, E_{NMS} and E_{CBM} are gases released by respiratory activities from the C_{PM}, C_{NMS} and C_{BM} pools respectively. W: respective weights of the C_{PM} , C_{NMS} and C_{BM} pools: C%: % carbon contained in the respective pools; R_{PM}, R_{NMS} and R_{BM}: emission rate in %C of the different pools. Note that respiration of soil ecosystem engineers when they produce biogenic aggregates is an additional source of gas emissions.

334 335

Three major steps are necessary to run this general model of soil aggregation and associated C dynamics (Figure 2 and 3).

337 338

336

Diagnosis of the aggregation status of the soil that considers the origins and relative amounts of the different types of aggregates (Fig.2);

339 340 Assessing the production and turnover of aggregates by measuring the flows of materials among the different aggregated and non-aggregated fractions (Fig 2);

341 342 343

344

C cycling and aggregate dynamics: an analysis of the quality and quantity of C pools contained within each macro-aggregate or non-macro-aggregated fraction, their mineralization rates in the respective pools and the C losses and gains associated with their production, stabilization, ageing and disruption cycles (Fig 3).

3.2 Diagnosis of aggregation status

Separation and identification of the origins of aggregates is the first step in our approach.

Large macro-aggregated compartments

Size limits. There is a general agreement that a 250 μm size separates macro- from microaggregates. Since it was first proposed by Tisdall and Oades (1982), this limit has dominated the research although sub categories have been proposed within each class. This paper focuses on the large macroaggregates > 2 mm sub category that can be easily separated and identified.

Origin and identity. Manual fractionation allows the separation of soil into three main fractions (non-macro-aggregated, physicogenic and biogenic macro-aggregates) based on their sizes and general morphologies (Topoliantz et al., 2000; Pulleman et al., 2005; Velasquez et al., 2007a). This method has been applied successfully to describe macro-aggregation in a wide range of soils, with sandy to clayey textures, in temperate and tropical areas. An example is provided in Figure 4. A great proportion of soils worldwide, protected by permanent vegetation and possessing suitable moisture contents, comprise large proportions of up to 60% macro-aggregates of both physical and biogenic types (Velasquez et al., 2019).

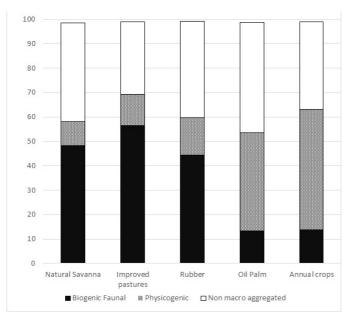


Figure 4: Relative proportions of biogenic, physicogenic macro-aggregates and non-macro-aggregated soil in different types of land use in the Eastern Plains of Colombia (Lavelle et al., 2014).

Physical aggregates are soil blocks produced by mechanical processes that create fissures in a continuous soil matrix (Boersma and Kooistra, 1994; Jongmans et al., 2001; Pulleman et al., 2005. They have angular blocky shapes with a dominance of sharp edges and plane surfaces. Particles within blocks are held together in a continuous matrix by organic and inorganic binding agents together with inter-particle binding due to Van der Waals forces (Hu et al., 2015). There is evidence that the same alternation of drying and rewetting events, that created these aggregates, may also activate the release of organic molecules, by microorganisms, that act as glues and consolidate these structures (Degens and Sparling, 1995; Cosentino et al., 2006). The addition of high molecular weight humic compounds may have similar effects (Piccolo et al., 1997; Yamaguchi et al., 2004). This set of aggregation processes when enhanced by physical compaction through use of tractors or cattle trampling in managed systems, tends to create very large clods and hard pans (Keen et al., 2013).

<u>Biogenic aggregates</u> are produced by organisms classified as ecosystem engineers for their ability to modify soil conditions through their physical activities (Blouin et al., 2013; Lavelle

et al., 2016). Ecosystem engineers produce a wide diversity of macro-aggregated structures that differ significantly in their morphologies, chemical and biochemical compositions (Decaëns et al., 2001; Mora et al., 2003; Velasquez et al., 2007b) (Figure 5). Specific mineral compositions and organic matter contents and natures gives them rather specific spectral signatures when illuminated with Near Infrared light (Hedde et al., 2005; Dominguez-Haydar, 2018).

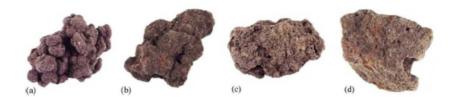


Figure 5: Different kinds of soil macro-aggregates: a. biogenic (fresh earthworm cast); b. biogenic (welded cast); c. intermediate (rounded aggregate); d. physicogenic (angular blocky aggregate) (Photo: Pulleman et al., 2005).

We recognize here four main categories which in turn comprise a large diversity of sub categories.

Fungal macro-aggregates are created when fungal hyphae entangle organic debris and mineral particles into stable aggregates that resist slaking and dispersion under the mechanical stresses applied in classical tests. They are usually not separated in soils with a permanent plant cover and are mostly seen in intensive cropping systems where communities of other ecosystem engineers are depleted.

Most of the macro-aggregation described in soils of cropped fields and laboratory tests is actually produced by this process (Tisdall and Oades, 1982; Plante and Mc Gill. 2002). Mycorrhizal fungi are considered major producers of this type of aggregation (Rillig and Mummey, 2006).

Earthworms_when present are major agents of soil bioturbation with widely different effects depending on their ecological strategies (Bouché, 1977; Lavelle and Spain, 2001). Epigeic earthworms live in the litter layers and are active composting agents that transform litter into largely organic pellets. Anecic species live in deep, subvertical galleries

surrounded by zones of compacted soil that may behave as stable aggregated structures. Their casts are usually relatively loose structures stabilized by high contents of 20 to 40% on average (Judas, 1992) of little-decomposed particulate organic matter. Endogeic earthworms live within the mineral soil horizons and feed on soil, with different concentrations of organic matter depending on their specific – oligohumic, mesohumic or polyhumic- adaptive strategies (Lavelle and Spain, 2001). They are the major producers of biogenic macro-aggregates in many soils. They typically ingest from 1 to 10 and sometimes up to 30 times their own weight of soil daily and total values of up to 1150 Mg dry soil ha⁻¹ have been measured, equivalent to a 10 cm thick soil layer (Lavelle, 1978). Their casts are either highly compacted structures, with bulk densities as high as 1.8 to 2.0 Mg m⁻³ (Blanchart et al., 1993), or loose structures, easily disaggregated, produced by the endogeic decompacting species (Blanchart et al., 1997).

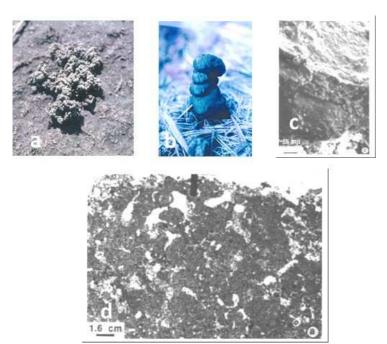


Figure 6: Loose (Chuniodrilus zielae) (a) and compact (Reginaldia omodeoi) (b) earthworm casts in African savanna (Lamto, Ivory Coast). (Photos P. Lavelle). Right: (c) Fracture of a surface cast of R. omodeoi observed with SEM. Observe the cortex made of fine particles that gives the surface of the cast a smooth and close aspect (Blanchart et al., 1993). (d) Thin section in the 0-10 cm layer of a soil from a humid savanna (Lamto, Ivory Coast) showing accumulation of globular casts mainly produced by the earthworm Reginaldia omodeoi (Blanchart et al., 1997).

436

The destruction and reorganization of aggregates may actually be a product of earthworm activity itself in some soils.

437 438

439

440

441

442

443

444

445

446

447

448

449

Social insect structures._Termites and ants may be active producers of soil macroaggregates. Their bioturbation of several Mg ha⁻¹ year⁻¹ is the result of a mixture of digging, building and casting activities (Lobry de Bruyn & Conacher 1990; Folgarait, 1998; Jouquet et al., 2011). Aggregates formed that way vary from very loose deposits and faeces to very compact tunnel-shaped aggregates or complex nest structures that may last years to decades and even more (Rajagopal et al., 1982; Bonell et al., 1986; Mermut et al., 1994;

Humphreys, 1994; Gorosito et al., 2005; Korb, 2011; Erens et al., 2015).

Maybe even more remarkable is the formation by humivorous termites of structures identified as pseudo-sand aggregates that end up comprising the entire volume of some tropical ferralsols (Wielemaker, 1984, Eschenbrenner, 1986; Balbino et al., 2002; Reatto et al., 2009; Millogo et al., 2011).

450

451

452

453

454

455

456

457

458

459

460

461

462

Root aggregates are structures that adhere to the roots. They are formed through five different processes: local compaction of soil, water regime alteration, rhizodeposition of gluing materials, decomposition of root material, and entanglement of soil particles by fine (0.2 to 1 mm in diameter) roots (Monroe and Kladivko, 1987; Miller and Jastrow, 1990; Morel et al., 1991; Materechera et al., 1992; Dorioz et al. 1993; Degens et al., 1994; Alami et al. 2000, Czarnes et al. 2000, Gale et al. 2000, Feeney et al. 2006; Demenois et al., 2018). An estimated 20% of plant photosynthesis is directly released at root tips with strong biological effects and induced aggregation (Miller and Jastrow, 1990; Lavelle and Spain, 2001). Macro-aggregation by root activities may be a rather fast process. In a five-week laboratory experiment, Trifolium pratense plants produced an average of 217.1 g and Plantago lanceolata, 142.1 g of macro-aggregates in pots that contained 800 g of dry soil moistened to field capacity (Zangerlé et al., 2011).

Mixed structures. Although some spectacular structures are produced by well-identified single actors in nature or laboratory experiments, aggregation seems frequently to be a cooperative process that involves different kinds of organisms, especially microorganisms. There is also evidence that fine roots often colonize fresh earthworm casts and thus possibly add their aggregative effects to those of the earthworm (Decaëns et al., 1999; Zangerlé et al., 2011; Fonte et al. 2012). Fungal hyphae likely participate in the consolidation of all sorts of other macroaggregates built by large ecosystem engineers.

471

472

473

474

475

476

477

478

479

480

481

482

483

484

485

486

487

488

489

490

491

492

464

465

466

467

468

469

470

Macro-aggregate specific identity and age

For a finer identification of physicogenic and biogenic macroinvertebrate or root macroaggregates, Near Infrared Spectrometry (NIRS) has proven to be a very efficient and practical tool (Hedde et al., 2005). NIRS is a non-destructive method that reflects the texture and amount and quality of organic matter contained in a soil sample. For each sample, NIRS provides a spectrum that can be divided into separate ranges of wave length longitudes (Velasquez et al., 2007b). Manually separated biogenic, physicogenic and nonmacro-aggregated aggregates exhibit significant differences (Dominguez-Haydar et al., 2018). The same method perfectly discriminated structures produced by different macro invertebrates (Hedde et al., 2005). Rather small samples of a few cg can be analysed separately. It is therefore possible to analyse individually all the macro-aggregates from a sample of a standard 10 x 10 x 10 cm size sample and group aggregates with similar spectral signatures into homogeneous categories. Using this method, Zangerlé et al. (2016) were able to separate eight categories of macro-aggregates from a forest soil in Luxemburg. They further identified the earthworm species that had produced the structure by comparing the spectral signatures with a bank of signatures established under laboratory conditions. In another experiment, the same authors observed changes in the spectral signature of casts as they aged (Zangerlé et al., 2014) showing potential of this method to identify different phases in the stabilization, ageing and disaggregation process of aggregates. Alternatively, enzymatic activities, microbial communities PLFA fingerprints or physical and chemical variables have been shown to discriminate among biogenic structures of different origins (Decaëns et al., 2001; Mora et al., 2003; Hedde et al., 2005;

Jouquet et al., 2013).

495

3.3. Assessing the production and turnover of aggregates

496 497

498

499

Our literature survey has confirmed that very few studies have been devoted to aggregate production and turnover in natural or managed systems.

500

501

502

503

504

505

506

507

508

509

510

511

512

513

514

515

516

517

518

519

520

521

Field reaggregation or disaggregation tests

A simple way to approach the dynamics of aggregation can be obtained by applying an insitu re-aggregation or disaggregation test as done by Blanchart (1992), Barros et al. (2001 and Gorosito (2007(). Blocks of soil from 10 x 10 x 10 cm to 25 x 25 x 30 cm depending on local conditions are taken in the field and a diagnosis of macro-aggregation is performed using the methodology proposed in section 3.1. Soil is then crushed and passed through a sieve with a 1 mm mesh size and taken back to fill the hole from which it had been excavated and reaggregation is monitored. The unit is covered with some litter taken from the surroundings and/or protected with a mesh to prevent the direct impact of rainfall that would create physicogenic macro-aggregates. Different sizes of mesh may be used to allow invertebrates and roots of different sizes to enter the experimental unit. Under these conditions, ecosystem engineers from the surrounding soil and natural physical processes will progressively form aggregates from the non-macro-aggregated soil. Experimental units are excavated at regular time intervals to follow the restoration of the macro-aggregated structure. In the experiment conducted by Blanchart (1992), the average percentage of macro-aggregates > 2mm was 12.9% when macroinvertebrates were not allowed to enter the unit, 49.9% when the mesh size allowed them to recolonize and 60.6% when the units had been inoculated with endogeic earthworms. The dynamics of disaggregation can also be observed putting the intact soil block, after fauna has been eliminated by a temporary drowning, in a net that will not allow roots and invertebrates to come in (Blanchart et al., 1997).

In some sites, macro-aggregate production can be measured using direct methodologies. Production of surface and subterranean casts by earthworms, root macro-aggregates and aggregated soil in the nest structures of termites and ants has been measured by ecologists in a range of laboratory and field situations (Lavelle, 1978; Lobry de Bruyn et al., 1990; Jouquet et al., 2013; Lavelle and Spain, 2001: Zangerlé et al., 2011) or simulated with mechanistic models (Martin and Lavelle, 1992). Data are still scarce because these measurements that require highly specific biological expertise are generally difficult and time consuming, especially when measuring structures produced in the soil matrix.

Spectral analysis of individual macro-aggregates

Macro-aggregates of biogenic root or invertebrate origin have a specific spectral signature that mostly reflect specific quality and concentrations of organic elements and texture (Hedde et al., 2005; Velasquez et al., 2007a; Zhang et al., 2009; Zangerlé et al., 2011). These specific signatures seem to progressively converge towards a common "bulk soil" signature as the structure ages. In a laboratory study conducted with the endogeic species *Aporrectodea caliginosa*, spectral signature of casts changed first rapidly, in the first 2 days after deposition and then slowly until day 45 to 60 when it was difficult to separate it from the bulk soil signature. This method allowed to separate casts aged less than 60 days and get an estimate of macro-aggregates of a given category during a 2-month period. (Zangerlé et al., 2016).

<u>Turnover time in steady state situations</u>

In a steady state situation, the amount of new structures created is expected to be compensated by the destruction of an equal amount. Based on reaggregation experiments conducted in natural environment, Blanchart (1992) was able to evaluate at 20 months and 28 months, in grass and shrub savannas respectively, the average residence time of casts of the earthworm *Reginaldia omodeoi* in a savanna of Ivory Coast. In this case, complementary studies have shown that macro-aggregated structures created by this

earthworm are eaten by another earthworm species from the Eudrilidae family that transform these highly compact casts into fragile erodible soil pellets (Blanchart et al., 1997). Very fast transformations of the soil structure have been observed with this method. In an experiment conducted in Amazonia, Barros et al, (2001) exchanged undisturbed soil blocks from a pasture and the natural forest to the other site. After one year, the heavily compacted soil of the pasture had recovered a macro-porosity close to that of the forest, while forest blocks inoculated in the pasture has almost attained the levels of compaction observed in this pasture. A similar experiment allowed Gorosito (200) to demonstrate that soil blocks from rice fields derived from a natural savanna were rapidly colonized, with similar abundances and diversities after one year and even higher values after 2 years.

Similar approaches developed with plants and other producers of biogenic aggregates might provide some estimates on the life duration of these structures. Further studies will then be required to identify the predominant disruption process and the fate of microaggregates that they contained.

4 C cycling and soil aggregation

Once recognized and measured the different aggregate pools in soil, the effect of soil aggregation dynamics on C cycling can be assessed using a simple conceptual framework (Figure 3). Assessing the effect of each recognized pool on soil C cycle first requires an evaluation of the energy cost (as mineralized C) of creating these macro-aggregates, if relevant. Then we need a precise knowledge of the amount and quality of the organic matter contained in this pool and an estimate of the persistence and general dynamics of the specific aggregate pool. In addition, one must have some ideas on the processes that lead to their disruption and transfer to the non-macro-aggregated pool or to another pool.

Soil C in aggregates and aggregation dynamics

Analysis of C contents separated among particulate organic matter (POM) and MAOM fraction associated to the soil mineral fraction will be applied to samples from the different recognized aggregate pools. This will allow measuring stocks of C associated to the

580 different pools. A number of studies has shown strong effects of plant cover and soil 581 management on these stocks and their temporal variations (Dexter et al., 1999: Conant et 582 al., 2001; Post et al., 2000; Martens, 2000; Zhang et al., 2013; 2016). 583 We hypothesize that transformations of particulate and mineral-associated organic matter 584 are linked to those of the aggregated or non-aggregated mineral pools that comprise them. 585 Dead fine and coarse root material and root exudates are transferred to the root biogenic 586 macro-aggregated structures (CBM roots) through rhizospheric activities. Organic debris of 587 root origin occasionally ingested by invertebrates will be further included in their faecal 588 pellets as earthworm casts or as organic macro-aggregates of rhizophagous coleoptera 589 larvae in other fractions of the CBM pool. Organic matter deposited in the leaf litter layer, 590 the other source of organic materials, is first transformed by comminution and natural 591 composting processes and progressively incorporated to the soil by digestion and 592 bioturbation processes associated with macroinvertebrate feeding activities and leaching of 593 dissolved organic matter. The decomposition rate of this organic matter is influenced by 594 the same processes that lead to its incorporation in the aggregates, gut transit or burial 595 below earthworm casts and insect deposits at the soil surface.

596

597

598

599

600

601

602

Energy cost of macro-aggregate production

As indicated before, the formation of certain biogenic macroaggregates has a significant cost in energy and C emission. This C loss may be evaluated by comparing C contents of the non-macroaggregated material originally used with the macroaggregates produced, or measuring the respiratory costs associated with the mechanical activities associated to their formation (Lavelle, 1978).

603604

605

606

607

<u>Carbon emission from the aggregated and non-macro-aggregated pools</u>

A large number of studies have compared the respiration activity of aggregates of different size fractions and in different conditions of soil management (Dexter et al., 1999; Ashman et al., 2003; Zhang et al., 2013; 2016). Carbon from each pool is mineralized at different

rates. For example, Martin (1991) showed that C mineralization proceeded at a much lower rate in casts of Reginaldia omodeoi than in the non-ingested soil sieved at 2mm. Digestion of the soil prior to cast release had decreased the C content of the soil by 9%, but mineralization inside the very compact cast had decreased microbial activity to such a low value that the non-digested soil and the cast had the same C contents after 100 days. With each day past this limit, the difference in C contents between macro-aggregated and nonmacro-aggregated soil would increase. Where casts may persist from 20 to 28 months, the ability of this species and type of biogenic aggregate to conserve C may be very important. The expected 'protection of C from mineralization however will likely depend on conditions created within the structure: aeration, temperature and moisture conditions, stoichiometric and other chemical environment conditions. The major water-soluble organic sources, root exudates, earthworm intestinal mucus or termite saliva that represent inputs of several Mg C per ha per year are mixed with the mineral soil and soon become part of the macro-aggregated soil fraction, in the forms of earthworm casts, root macro-aggregates or termite constructions. In the absence of invertebrate ecosystem engineers, leaf litter organic matter stays at the soil surface and the production of organo-mineral macro-aggregates is limited to root aggregate production. Water soluble organic matter leached from the litter layer may well circulate among macro-aggregated fractions as part of the soil drainage process and stimulate microbial decomposing activities creating priming effects (Fontaine et al., 2007). It should, however, not have a significant effect since it represents a rather limited flow as compared to other components of organic matter inputs (Lavelle and Spain, 2001; Jimenez and Lal, 2006). It is also expected to be rapidly flocculated in the presence of clay minerals (Toutain, 1981) or consumed upon entering the soil matrix and does not enter easily inside compact aggregates Carbon in the non-macro-aggregated soil pool (C_{NMS}) may have been released from disaggregated biogenic or physical aggregates or be generated by flocculation or condensation processes of water-soluble organic matter occurring in the interaggregate

608

609

610

611

612

613

614

615

616

617

618

619

620

621

622

623

624

625

626

627

628

629

630

631

632

633

634

space. Carbon of the physicogenic macro-aggregate pool (C_{PM}) may have been contained in the non-aggregated pool before it was transformed into physical aggregates.

5 Discussion

Our literature review confirmed that biological processes are severely undermined in aggregate studies was confirmed: only 11.5% of the papers considered microbial components, and 4% roots and invertebrate activities. This is clearly the result of restricted disciplinary focus where a holistic view is necessary (Lavelle et al., 2016; Briones, 2018). Five other important gaps, addressed below, appeared that are susceptible to hinder a global and practical understanding of processes at hand and ways to enhance their benefits in terms of ecosystem services. The approach that we propose to attain this goal is based on a number of assumptions, conceptual simplifications and the choice of technical options that often coincide with these gaps.

<u>Definition and classification of aggregates: The need for a comprehensive separation technique that combines viewing and isolating approaches.</u>

Soil macroaggregates may be seen in very different ways by scientists depending on the discipline they belong to. For soil ecologists, technicians and farmers, they often are visible and easily recognizable structures created by roots and invertebrate ecosystem engineers (Lavelle et al., 2006; Blouin et al., 2013; Jouquet et al., 2016). A severe drawback of using stress resistance of aggregates as a way to isolate them is that part of the resistance is provided by chemical and biological binding effects associated to the soil type, and not to the macro-aggregation process itself. On the other hand, a direct relationship between stabilization and conservation of C in soil conditions has not been proven to our knowledge. The manual separation technique that we propose is a combination of viewing and isolation techniques. This technique has proven very efficient at showing scientists, students and farmers the importance and vulnerability of the macro aggregation in soils. Synthetic indicators made with data provided with this method are always linked with

other physical, chemical or biological attributes of soil fertility and ecosystem services provisioning (Velasquez et al., 2007b; Lavelle et al., 2014; Grimaldi et al., 2014; Velasquez ad Lavelle, 2019). When the objective is to show the importance of the macro aggregation process to students or farmers and differential effects of soil management options, no specific training is required since trainees will separate structures the same way, whatever it is, in different soils and perceive differences. Efforts should be done however, to better standardize this technique (Jouquet et al., 2009). Clear recommendations should be made on how to properly disrupt the soil matrix according to natural breaking surfaces, in different conditions of soil texture and plant root densities. Separation of root and other biogenic aggregates may require specific actions depending on their sizes and structures. Soil texture - especially when clayey- or effects of mismanagement -when soils are highly compacted- may complicate this technique in some situations. An important issue is the size of the elements that have been identified as macro-aggregates. When felt important, smaller sieves could be used to isolate fractions down to 1, or even 0.25 mm. In this case, the observation of subsamples with a stereo microscope may provide the accuracy required by specific scientific studies (Topoliantz et al., 2000).

Macro vs. microaggregates. Can microaggregates be independent of macro-aggregates?

And to what extent?

The idea that microaggregates are formed inside macro-aggregates (Six et al. 2000) offers a welcome simplification in our approach. It is also in accordance with the self-organized soil model that proposes a hierarchical organization of soil, with successive functional units nested in each other as their size increases (Lavelle et al., 2016). While this view of soil organization is widely acknowledged and demonstrated by an important number of studies (Six et al., 2000; Bossuyt et al., 2004; Fonte et al., 2012), a few questions remain. First microaggregates are not a homogeneous category (Tötsche et al, 2018) although, there is some expectation that microaggregates have compositions derived from that of macroaggregates, although with generally less C and lower C:N ratio (Blanchart et al., 2000). Maybe more important is knowing what happens to microaggregates when

macroaggregates get disrupted. In the model of aggregate dynamics proposed by Marquez et al. (2018), microaggregates may be either disrupted when the macro-aggregate that comprise them is disrupted or be part of a transient fractions that is further reincorporated into macro-aggregates.

698

699

700

701

702

703

704

705

706

707

708

709

710

711

712

713

714

715

716

717

718

697

694

695

696

<u>Identify</u> aggregate origins to understand biological or other processes that determine their creation and further dynamics.

The very low proportion of articles that consider root or invertebrate generation of macroaggregates points at a very important gap. Specificities of composition, structure and microbial communities at species level of these macro-aggregates are very important determinants of their further stability and temporal dynamics (Hedde et al., 2005; Mora et al., 2003; Jouquet et al., 2013). NIRS has been showed to offer a cheap and efficient way to solve the problem. However, NIR spectral signatures have also proved to be unstable in ageing earthworm casts (Zangerlé et al., 2014). Changes in spectral signature probably reflect the rapid changes of microbial biomass and mineral N contents observed in freshly deposited earthworm casts (Lavelle et al., 1992). The Intriguing observation that casts aged >45 days have a signature similar to the bulk soil should be further investigated. Does this mean that the surrounding soil was actually mostly comprised of aged earthworm casts of the species considered? Or can we think of an alternative hypothesis? Fresh macroaggregates are hotspots of microbial activity where some microbial species are enhanced, not the whole soil community. Would the specific signature observed in freshly created structures reflect important transient biomass of these components? And signature observed later on an indication that these populations have decreased to their initial abundance in the bulk soil? Comparable observations made by Blackwood and Paul (2003) show the importance of this question.

719

720

721

Interactions among physicogenic and biogenic agents in soil macro-aggregation dynamics

In a finite soil volume, formation of new macro-aggregates in a steady state formation has to be done at expenses of other pools of aggregated or non-aggregated soil. Stable macro-aggregates produced by a given set of invertebrates or roots can be destroyed by other invertebrates that may use them as food, or roots and invertebrates that just need to make their way through the soil to grow or develop their porous domains. Organisms may cooperate in forming macro-aggregates. This is the case for example for earthworms and roots, although cooperation may well be specific (Zangerlé, et al., 2011). The equilibrium among ecosystem engineers with opposite compacting or decompacting effects may be a critical element when considering soil management options (Blanchart et al., 1997; Chauvel et al., 1999). Research on this topic is virtually absent.

C loss and conservation associated to aggregation

While soil aggregation is largely associated with C accumulation in soils, detailed underlying mechanisms are far from being completely understood. The option of our model, considering C cycling as a discrete process split among different aggregate pools of different origins with different histories probably deserves improved theoretical and experimental foundations. In surface soil layers where biological activity is concentrated, aggregation seems to be the best option for limiting C mineralization. A better understanding and classification of biological aggregation processes would probably help. The overall effects of these aggregation processes on C cycling might well differ along a continuum from short term fungal hyphae particle entanglement observed in poorly aggregated agricultural soils to potentially long-lived earthworm casts stabilized by a drying event or aggregates associated with social insect nests. In both extreme cases, respiration of the organisms responsible for the creation of aggregates have very different energy costs that should be incorporated in models. One can hypothesize that in steady state conditions, the cost of creating aggregates is compensated by their protection effect on soil organic matter (Lavelle and Spain, 2001; Lubbers et al., 2017). Disaggregation of soils

associated to intensive cropping practices may be explained by increased microbial mineralization in conditions where macro-aggregation by ecosystem engineers is deficient. In other cases, excessive activity of invasive ecosystem engineers, like European earthworm species in northern America forests, may well increase soil aggregation while soil C stocks decrease. Bioturbation activities then exceed the effect of physical protection in aggregates (Bohlen et al., 2004).

With soil depth, oxygen concentration naturally decreases and the mere rarefaction of biological activity is thought to favor C conservation. Mechanisms and conditions that allow migration of C in deep layers also require better understanding.

6 Conclusion

Synthesis of knowledges generated in different disciplinary fields led us to propose an alternative approach to evaluate the process of aggregate formation and its contribution to the conservation and accumulation of organic matter in soils. The methodology proposed is based on relatively simple and low-cost techniques that should be accessible to a large number of scientists, technicians and farmers. This condition is important to develop multifunctional agricultural practices that contribute to restore soil- based ecosystem services and especially soil critical contribution to the mitigation of global warming.

Explicitly considering biological mechanisms that determine soil aggregation will allow harnessing this process to ecosystem restoration practices and sustainable management options. In soils considered as self-organized systems, aggregates are constructed structures that have positive feedbacks on the communities that produced them (Lavelle et

al., 2016; Bedano et al., 2019). Aggregation is part of the "potential" of the ecosystem as defined by Holling (2000), a capital that varies along adaptive cycles and that determines ecosystem evolution. In situ soil reaggregation tests, that indirectly evaluate the ability of organisms to collectively organize and improve their habitat, might well appear as a simple way to measure such an important attribute. This evaluation of potential combined with an assessment of the connectance, mostly linked to local diversity and abundance of plant and

780 invertebrate ecosystem engineer communities, should allow progressing in the base line

781 evaluation of ecosystems before initiating restoration practices.

782

783

7 References

784 785 786

Abiven, S., S. Menasseri and C. Chenu. 2009. The effects of organic inputs over time on soil aggregate stability - A literature analysis. Soil Biology & Biochemistry 41. 1: 1-12.

787 788

- 789 Alami, Y., Achouak, W; Marol, C. and Heulin, T. 2000. Rhizosphere soil aggregation and plant growth promotion of sunflowers by an exopolysaccharide-producing Rhizobium sp. 790 791 strain isolated from sunflower roots. Appl. Env. Microb., 66, 3393-3398.
- 792 Altieri, M. A., C. I. Nicholls, A. Henao and M. A. Lana. 2015. Agroecology and the design of 793 climate change-resilient farming systems. Agronomy for Sustainable Development 35. 3: 794 869-890.

795

796 Amezketa, E. 1999. Soil aggregate stability: a review. Journal of Sustainable Agriculture 14. 797 83-151.

798

799 Ashman, M. R., P. D. Hallett and P. C. Brookes. 2003. Are the links between soil aggregate 800 size class, soil organic matter and respiration rate artefacts of the fractionation 801 procedure? Soil Biology & Biochemistry 35. 3: 435-444.

802

- 803 Bach, E. M., Hofmockel, K. S. 2 016. A time for every season: soil aggregate turnover 804 stimulates decomposition and reduces carbon loss in grasslands managed for bioenergy. 805 Glob. Ch. Biol. Bioener. 8, 3:588-99.
- 806 Balbino, L.C., Bruand, A., Brossard, M., Grimaldi, M., Hajnos, M., Guimaraes, M.F., 2002.
- 807 Changes in porosity and micro aggregation in clayey Ferralsols of the Brazilian Cerrado on
- clearing for pasture. Eur. J. Soil Sci., 53, 219-230. 808
- 809 Barois, I., Villemin, G., Lavelle, P., Toutain, F. 1993. Transformation of the soil structure
- 810 through Pontoscolex corethrurus (Oligochaeta) intestinal tract. Geoderma, 56, 57-66
- Barros, E., P. Curmi, V. Hallaire, A. Chauvel and P. Lavelle. 2001. The role of macrofauna in 811
- 812 the transformation and reversibility of soil structure of an oxisol in the process of forest to
- 813 pasture conversion. Geoderma 100, 1-2: 193-213.
- Bedano, J.C., Vaguero, F., Dominguez, A., Rodriguez, M.P., Wall, L., Lavelle, P. 814
- 815 Earthworms contribute to ecosystem process in no-till systems with high crop rotation
- 816 intensity in Argentina. Acta Oecologica 98. 14-24

- 818 Blackwood, C. B. and E. A. Paul. 2003. Eubacterial community structure and population size
- 819 within the soil light fraction, rhizosphere and heavy fraction of several agricultural
- **820** systems. Soil Biol. Biochem. 35, 1245-1255.
- 821 Blanchart, E., 1992. Restoration by earthworms (Megascolecidae) of the macro-aggregate
- 822 structure of a destructured savanna soil under field conditions. Soil Biol. Biochem., 24,
- **823** 12,1587-94.
- 824 Blanchart, E., Bruand, A., Lavelle. P. 1993. The physical structure of casts of Millsonia
- 825 anomala (Oligochaeta, Megascolecidae) in shrub savanna soils (Côte d'Ivoire). Geoderma,
- **826** 56 (1-4), 119-132.
- 827 Blanchart, E., P. Lavelle, E. Braudeau, Y. Le Bissonnais and C. Valentin. 1997. Regulation of
- 828 soil structure by geophagous earthworm activities in humid savannas of Cote d'Ivoire. Soil
- 829 Biol. & Biochem. 29. 3-4: 431-439.
- 830 Blanchart, E., Achouak, W., Albrecht, A., Barakat, M., Bellier, G., Cabidoche, Y.M., Hartmann,
- 831 C., Heulin, T., Larré-Larrouy, C., Laurent, J. Y., Mahieu, M., Thomas, F., Villemin, G., Watteau,
- 832 F. 2000 Déterminants biologiques de l'agrégation des vertisols des Petites Antilles.
- 833 Conséquences sur l'érodibilité des sols. Ét. Gest. Sols, 7: 309-328.
- 834 Blouin, M., Hodson, M. E., Delgado, E. A., Baker, G., Brussaard, L., Butt, K. R., Dai, J.,
- 835 Dendooven, L., Peres, G., Tondoh, J., Cluzeau, D., Brun, J.J. 2013. A review of earthworm
- impact on soil function and ecosystem services. Eur. J. Soil Sci., 64(2), 161-82.
- 837 Boersma, O. H., Kooistra, M. J., 1994. Differences in soil structure of silt loam Typic
- 838 Fluvaquents under various agricultural management practices. Agric. Ecosyst. Environ., 51,
- **839** 21-42.
- 840 Bohlen, P. J., S. Scheu, C. M. Hale, M. A. McLean, S. Migge, P. M. Groffman and D.
- 841 Parkinson. 2004. Non-native invasive earthworms as agents of change in northern
- temperate forests. Front. Ecol. & Envir. 2, 8: 427-435.
- 843 Bonell, M., Coventry, R. J., & J. A. Holt. 1986. Erosion of termite mounds under natural
- rainfall in semiarid tropical north-eastern Australia. Catena, 13, 11-28.
- 845 Bossuyt, H., Six, J., & P.F. Hendrix. 2004. Rapid incorporation of carbon from fresh residues
- 846 into newly formed stable microaggregates within earthworm casts. European Journal of
- 847 Soil Science, 55, 393-399.
- 848 Bossuyt, H., Six, J., & P.F. Hendrix. 2005. Protection of soil carbon by microaggregates
- within earthworm casts. Soil Biology and Biochemistry, 37, 251-258.
- 850 Bottinelli, N., Jouquet, P., Capowiez, Y., Podwojewski, P., Grimaldi, M., Peng, X., 2015. Why
- is the influence of soil macrofauna on soil structure only considered by soil ecologists? Soil
- **852** Till. Res. 146, 118-24.

- 853 Bouché, M. B. 1977. Stratégies lombriciennes. Soil Organism as Components of
- 854 Ecosystems. Ecol. Bull. (Stockholm), 25, 122-32.
- 855 Briones, M. J. I. 2018. The Serendipitous Value of Soil Fauna in Ecosystem Functioning: The
- 856 Unexplained Explained. Front. Envir. Sci. 6.
- 857 Bronick, C. J., Lal, R. 2005. Soil structure and management: a review. Geoderma. 124(1-
- **858** 2):3-22.
- 859 Cambardella, C. A.., Elliot, E. T. 1993. Methods for physical separation and characterization
- of soil organic matter fractions. Geoderma 56. 449-457.
- 861 Carvalhais, N., Forkel, M., Khomik, M., Bellarby, J., Jung, M., Migliavacca, M., Mu, M.,
- 862 Saatchi, S., Santoro, M., Thurner, M., Weber, U., Ahrens, B., Beer, C., Cescatti, A.,
- 863 Randerson, J.T., Reichstein, M., 2014. Global covariation of carbon turnover times with
- 864 climate in terrestrial ecosystems. Nature, 514, 7521, 213-217.
- 865 Castorena, E. V. G., M. D. Gutierrez-Castorena, T. G. Vargas, L. C. Bontemps, J. D. Martinez,
- 866 E. S. Mendez and C. A. O. Solorio. 2016. Micro mapping of microbial hotspots and biofilms
- from different crops using digital image mosaics of soil thin sections. Geoderma 279, 11-21.
- 868 Chauvel, A., M. Grimaldi, E. Barros, E. Blanchart, T. Desjardins, M. Sarrazin and P. Lavelle.
- 869 1999. Pasture damage by an Amazonian earthworm. Nature 398, 6722: 32-33.
- 870 Chenu, C., 1995. Extracellular polysaccharides: An interface between microorganisms and
- 871 soil constituents. Environmental impact of soil component interactions; natural and
- anthropogenic organics. 1, 217-33.
- 873 Chenu, C., and Stotzky, G., 2002. Interactions between microorganisms and soil particles: an
- 874 overview. Interactions between soil particles and microorganisms. Impact on the terrestrial
- ecosystems. P. M. Huang, J. M. Bollag and J. M. Senesi. Chichester, UK, John Willey and sons:
- **876** 3-40.

- 877 Conant, R. T., K. Paustian and E. T. Elliott. 2001. Grassland management and conversion into
- 878 grassland: Effects on soil carbon. Ecol. Appl. 11. 2: 343-355.
- 879 Cosentino, D., Chenu, C., Le Bissonnais, Y. 2006. Aggregate stability and microbial
- 880 community dynamics under drying-wetting cycles in a silt loam soil. Soil Biol. Biochem.
- **881** 38(8), 2053-62.
- 882 Cotrufo, M.F., Ranalli, M.G., Haddix, M.L. et al. Soil carbon storage informed by particulate
- 883 and mineral-associated organic matter. *Nat. Geosci.* 12, 989–994 (2019)
- **884** doi:10.1038/s41561-019-0484-6
- 886 Czarnes, S., Hallett, P.D., Bengough, A.G., Young, I.M., 2000. Root- and microbial-derived
- mucilages affect soil structure and water transport. Eur. J. Soil Sci., 51, 435-443.

- 888 Decaëns, T., Mariani, L., Lavelle, P., 1999. Soil surface macrofaunal communities associated
- with earthworm casts in grasslands of the Eastern Plains of Colombia. App. Soil. Ecol., 13(1),
- **890** 87-100.
- 891 Decaëns, T., 2000. Degradation dynamics of surface earthworm casts in grasslands of the
- eastern plains of Colombia. Biol. Fertil. Soils, 32:149-156.
- 893 Decaëns T, Galvis JH, Amezquita E., 2001. Properties of the structures created by ecological
- engineers at the soil surface of a Colombian savanna. C.R.A. Sciences, III, Life Sci. 324(5),
- **895** 465-478.
- 896 Degens, B.P., Sparling, G.P., Abbott, L.K., 1994. The contribution from hyphae, roots and
- 897 organic carbon constituents to the aggregation of a sandy loam under long-term clover
- 898 based and grass pastures. Eur. J. Soil Sci., 45, 459-468.
- 899 Degens, B. P., Sparling, G.P. 1995. Repeated wet-dry cycles do not accelerate the
- 900 mineralization of organic C involved in the macro aggregation of a sandy-loam soil. Plant &
- **901** Soil,175, 2:197-203.
- 902 De Gryze, S., Six, J., Brits, C., Merckx, R. 2005. A quantification of short-term macro-
- aggregate dynamics: influences of wheat residue input and texture. Soil Biol. Biochem., 37,
- **904** 55-66.
- 905 Demenois, J., F. Carriconde, P. Bonaventure, J. L. Maeght, A. Stoke and F. Rey. 2018. Impact
- 906 of plant root functional traits and associated mycorrhizas on the aggregate stability of a
- 907 tropical Ferralsol. Geoderma 312, 6-16.
- 908 Dexter, A. R., J. Arvidsson, E. A. Czyz, A. Trautner and B. Stenberg. 1999. Respiration rates of
- 909 soil aggregates in relation to tillage and straw-management practices in the field. Act. Agric.
- 910 Scand. Section B-Soil and Plant Science 49. 4: 193-200.
- 911 Dominguez-Haydar, Y., Castañeda, C., Rodriguez-Ochoa, R., Jimenez, J.J. 2018. Assessment
- 912 of soil fauna footprints at a rehabilitated coal mine using micromorphology and near
- 913 infrared spectroscopy (NIRS). Geoderma. 313, 135-145.
- 914 Dorioz, J.M., Robert, M., Chenu, C. 1993. The role of roots, fungi and bacteria on clay
- particle organization. An experimental approach. Geoderma, 56, 179-194.
- 916 Elliott, E.T., 1986. Aggregate structure and Carbon, Nitrogen, Phosphorus in native and
- 917 cultivated soils. Soil Sci. Soc. Am. J., 627-633.
- 918 Elliott, E. T., and Cambardella, C. A., 1991. Physical separation of soil organic matter. Agr
- **919** Ecosyst & Envir.34: 407-19.
- 920 Erens, H., Boudin, M., Mees, F., Mujinya, B. B., Baert, G., Van Strydonck, M., Boeckx, P., &
- 921 Van Ranst, E. 2015. The age of large termite mounds-radiocarbon dating of Macrotermes

- 922 falciger mounds of the Miombo woodland of Katanga, DR Congo. Palaeoclim. Palaeoeco.
- **923** 435, 265-271.
- 924 Eschenbrenner, V. 1986. Contribution des termites à la micro-agrégation des sols
- 925 tropicaux. Cah. ORSTOM, sér. Pédol. 22(4),397-408.
- 926 Feeney, D.S., Crawford, J.W., Daniell, T., Hallett, P.D., Nunan, N., Ritz, K., Rivers, M., Young,
- 927 I.M. 2006. Three-dimensional micro organization of the soil-root-microbe system. Microb.
- 928 Ecol., 52, 151-158.
- 929 Feller, C., Beare, M. H., 1997. Physical control of soil organic matter dynamics in the tropics.
- 930 Geoderma, 79(1-4), 69-116.
- 931 Filser, J., Faber, J.H., Tiunov, A.V., Brussaard, L., Frouz, J., De Deyn, G., Uvarov, A.V., Berg,
- 932 M.P., Lavelle, P., Loreau, M., Wall, D.H., Querner, P., Eijsackers, H., Jiménez, J.J. 2016. Soil
- 933 fauna: key to new carbon models. SOIL, 2(4), 565-582.
- Folgarait, P. J., 1998. Ant biodiversity and its relationship to ecosystem functioning: a review.
- 935 Biodiv. and Conserv., 7(9): 1221-1244.
- 936 Fonte, S.J., Quintero, C.D., Velasquez, E., Lavelle, P., 2012. Interactive effects of plants and
- 937 earthworms on the physical stabilization of soil organic matter in aggregates. Plant &
- 938 Soil.359, 1-2

- 939 Gale, W.J., Cambardella, C.A., Bailey, T.B. 2000. Root-derived carbon and the formation and
- 940 stabilization of aggregates. Soil Sc. Soc. Am. J., 64, 201-207.
- 941 Gorosito, N., Curmi, P., Hallaire, V., Folgarait, P., Lavelle, P. 2005. Morphological changes in
- 942 Camponotus punctulatus (Mayr) anthills of different ages. Geoderma, 132 (3-4), 249-260.
- 943 Gorosito, M. 2007. Camponotus punctulatus: su rol en la comunidad de hormigas,
- 944 colonización de nuevas áreas y causas de su explosión demográfica frente a un disturbio
- 945 agrícola. Universidad Quilmes, Bernal, Argentina.
- 947 Grimaldi, M., Oszwald, J., Doledec, S., Hurtado M. P., Miranda, I. Arnaud de Sartre, X., Assis,
- 948 W. S., Castaneda, E., Desjardins, T., Dubs, F., Guevara, E., Gond, V., Lima, T., Marichal, R.,
- 949 Michelotti, F., Mitja, D. N., Noronha C., Oliveira, M. N., Ramirez, B., Rodriguez, G., Sarrazin,
- 950 M., da Silva, M. L., Silva Costa, L. G., de Souza, S. L., Veiga I., Velasquez, E., P. Lavelle. 2014.
- 951 Ecosystem services of regulation and support in Amazonian pioneer fronts: searching for
- **952** landscape drivers. Landsc. Ecol., 29, 2: 311-328.
- 953 Gumus, I. and C. Seker. 2015. Influence of humic acid applications on modulus of rupture,
- 954 aggregate stability, electrical conductivity, carbon and nitrogen content of a crusting
- **955** problem soil. Solid Earth 6, 4: 1231-1236.

- 957 Gutierrez-Castorena, M. C., E. V. Gutierrez-Castorena, T. Gonzalez-Vargas, C. A. Ortiz-
- 958 Solorio, E. Suastegui-Mendez, L. Cajuste-Bontemps and M. N. Rodriguez-Mendoza. 2018.
- 959 Thematic micro-maps of soil components using high-resolution spatially referenced
- mosaics from whole soil thin sections and image analysis. Eur. J. Soil Sci. 69, 2: 217-231.

- 962 Hedde, M., Lavelle, P., Joffre, R., Jimenez, J. J., Decaëns, T. 2005. Specific functional
- 963 signature in soil macro-invertebrate biostructures. Funct. Ecol., 19(5), 785-93.
- 964 Holling, C. S. 2001. Understanding the complexity of economic, ecological, and social
- **965** systems. Ecosystems 4, 5: 390-405.
- 966 Humphreys, G.S., 1994. Bioturbation, biofabrics and the biomantle: an example from the
- 967 Sydney Basin. Micromorphology in Soil: Studies in management and Genesis (eds A.J.
- 968 Ringrose-Voase and G. Humphreys), Amsterdam, pp. 787-798.
- 969 Hu, F.N., Xu, C.Y., Li, H., Li, S., Yu, Z.H., Li, Y., et al. 2015. Particles interaction forces and
- 970 their effects on soil aggregates breakdown. Soil. Til. Res. 147:1-9.
- 971 Jimenez, J. J. and R. Lal. 2006. Mechanisms of C sequestration in soils of Latin America. Crit.
- **972** Rev. Plant Sci., 25. 4: 337-365.
- 973 Jongmans, A. G., Pulleman, M. M., Marinissen, J. C. Y. 2001. Soil structure and earthworm
- 974 activity in a marine silt loam under pasture versus arable land. Biol. Fertil. Soils. 33, 279-
- **975** 285.
- 976 Jouquet, P., Podwojewski, P., Bottinelli, N., Mathieu, J., Ricoy, M-, Orange, D., Tran, T.D.,
- 977 Valentin, C. 2008. Above-ground earthworm casts affect water runoff and soil erosion in
- **978** Northern Vietnam. Catena, 74, 1:13-21.
- 979 Jouquet, P., A. Zangerlé, C. Rumpel, D. Brunet, N. Bottinelli and T. T. Duc. 2009. Relevance
- and limitations of biogenic and physicogenic classification: a comparison of approaches for
- 981 differentiating the origin of soil aggregates. Eur. J. Soil Sci. 60, 6: 1117-1125.

- 983 Jouquet, P., Traore, S., Choosai, C., Hartmann, C., Bignell, D. 2011. Influence of termites on
- 984 ecosystem functioning. Ecosystem services provided by termites. Eur. J. Soil Biol., 47(4):
- **985** 215-22.
- 986 Jouquet, P., P.-A. Maron, V. Nowak and D. Toan Tran. 2013. Utilization of microbial
- 987 abundance and diversity as indicators of the origin of soil aggregates produced by
- 988 earthworms. Soil Biol. & Biochem. 57, 950-952.
- 989 Jouquet, P., Guilleux, N., Caner, L., Chintakunta, S., Ameline, M., Shanbhag, R. R. 2016.
- 990 Influence of soil pedological properties on termite mound stability. Geoderma, 262, 45-51.
- 991 Judas, M. 1992. Gut content analysis of earthworms (Lumbricidae) in a Beechwood. Soil
- 992 Biol. Biochem., 24, 12: 1413-17.

- 993 Keen, A., Hall, N., Soni, P., Gholkar, M. D., Cooper, S., Ferdous, J. 2013. A review of the
- 994 tractive performance of wheeled tractors and soil management in lowland intensive rice
- **995** production. J. Terramec. 50, 1:45-62.
- 996 Keiluweit, M., Nico, P. S., Kleber, M., Fendorf, S. 2016. Are oxygen limitations under
- 997 recognized regulators of organic carbon turnover in upland soils? Biogeochem.,127(2-
- **998** 3):157-71.
- 999 Keyes, S. D., Gillard, F., Soper, N., Mavrogordato, M. N., Sinclair, I. Roose, T. 2016. Mapping
- 1000 soil deformation around plant roots using in vivo 4D X-ray Computed Tomography and
- 1001 Digital Volume Correlation. J. Biomech. 49, 9: 1802-1811.
- 1002 Koesters, R., A. C. Preger, C. C. Du Preez and W. Amelung. 2013. Re-aggregation dynamics
- 1003 of degraded cropland soils with prolonged secondary pasture management in the South
- **1004** African Highveld. Geoderma 192, 173-181.
- 1005
- 1006 Korb, J. 2011 Termite mound architecture, from function to construction. Biology of
- 1007 Termites, a Modern Synthesis. Y. Roisin, D.E. Bignell, N. Lo. Dordrecht, Springer: 349-373.
- 1008 Kravchenko, A., W. Otten, P. Garnier, V. Pot and P. C. Baveye. 2019. Soil aggregates as
- 1009 biogeochemical reactors: Not a way forward in the research on soil-atmosphere exchange
- **1010** of greenhouse gases. Glob. Ch. Biol, 25, 7: 2205-2208
- 1011
- 1012 Kuzyakov, Y., Blagodatskaya, E., 2015. Microbial hotspots and hot moments in soil: Concept
- **1013** & review. Soil Biol. Biochem., 83:184-99.
- 1014 Lavelle, P., 1978. Les Vers de Terre de la savane de Lamto (Cote d'Ivoire). Peuplements,
- populations et fonctions dans l'écosystème. Publ. Lab. Zool., ENS, 18, 301pp
- 1016 Lavelle, P. & Meyer, J. A. 1983. Allez-les-Vers, a simulation model of dynamics and effect on
- 1017 soil of populations of *Millsonia anomala* (Oligochæta, Megascolecidæ). New Trends in Soil
- 1018 Biology. P. Lebrun, A. M. André, A. de Medts, C. Gregoire-Wibo and G. Wauthy. Louvain-la-
- **1019** Neuve, Dieu-Brichard : 503-519.
- 1020 Lavelle, P., G. Melendez, B. Pashanasi and R. Schaefer. 1992. Nitrogen mineralization and
- 1021 reorganization in casts of the geophagous tropical earthworm *Pontoscolex corethurus*
- 1022 (Glossoscolecidae). Biol. Fert. Soil14. 49-53.
- Lavelle, P., Bignell, D., Lepage, M., Wolters, V., Roger, P., Ineson, P., Heal, O. W., Dhillion, S.
- 1024 1997. Soil function in a changing world: the role of invertebrate ecosystem engineers. Eur. J.
- **1025** Soil Biol. 33, 4:159-93.
- 1026 Lavelle, P., Spain, A., 2001. Soil Ecology. Kluwer Pub., 654pp.
- 1027 Lavelle, P., 2000. Ecological challenges for soil science. Soil Sci., 165(1):73-86.

- **1028** Lavelle, P. 2002. Functional domains in soils. Ecol. Res.17, 4: 441-450.
- 1029 Lavelle, P., Rodriguez, N., Arguello, O., Bernal, J., Botero, C., Chaparro, P., Gomez, Y.,
- 1030 Gutierrez, A., Hurtado, M.P., Loaiza, S., Pullido, S.X., Rodriguez, E., Sanabria, C., Velasquez,
- 1031 E., Fonte, S.J., 2014. Soil ecosystem services and land use in the rapidly changing Orinoco
- 1032 River Basin of Colombia. Agr. Ecos. & Envir., 185:106-117.
- 1033 Lavelle, P., Spain, A., Blouin, M., Brown, G., Decaëns, T., Grimaldi, M., Jimenez, J.J., McKey,
- 1034 D., Mathieu, J., Velasquez, E., Zangerlé, A. 2016. Ecosystem Engineers in a Self-organized
- 1035 Soil: A Review of Concepts and Future Research Questions. Soil Sci., 181, 91-109.
- 1036 Le Bayon, R. C., Binet, F. 2001. Earthworm surface casts affect soil erosion by runoff water
- and phosphorus transfer in a temperate maize crop. Pedobiologia, 45, 5:430-442.
- 1038 Le Bissonnais, Y. 1996. Aggregate stability and measurement of soil crustability and
- erodibility: I. Theory and methodology. Eur. J. Soil Sci.47. 425-437.
- 1040
- 1041 Lehmann, J., Kleber, M., 2015. The contentious nature of soil organic matter. Nature.;528
- **1042** (7580):60-68.
- 1043 Le Quéré, C., Andrew, R. M., Friedlingstein, P., Sitch, S., Pongratz, J., Manning, A. C.,
- 1044 Korsbakken, J. I., Peters, G. P., Canadell, J. G., Jackson, R. B., Boden, T. A., Tans, P. P.,
- 1045 Andrews, O. D., Arora, V. K., Bakker, D. C. E., Barbero, L., Becker, M., Betts, R. A., Bopp, L.,
- 1046 Chevallier, F., Chini, L. P., Ciais, P., Cosca, C. E., Cross, J., Currie, K., Gasser, T., Harris, I.,
- 1047 Hauck, J., Haverd, V., Houghton, R. A., Hunt, C. W., Hurtt, G., Ilyina, T., Jain, A. K., Kato, E.,
- 1048 Kautz, M., Keeling, R. F., Klein Goldewijk, K., Körtzinger, A., Landschützer, P., Lefèvre, N.,
- 1049 Lenton, A., Lienert, S., Lima, I., Lombardozzi, D., Metzl, N., Millero, F., Monteiro, P. M. S.,
- 1050 Munro, D. R., Nabel, J. E. M. S., Nakaoka, S.-I., Nojiri, Y., Padin, X. A., Peregon, A., Pfeil, B.,
- 1051 Pierrot, D., Poulter, B., Rehder, G., Reimer, J., Rödenbeck, C., Schwinger, J., Séférian, R.,
- 1052 Skjelvan, I., Stocker, B. D., Tian, H., Tilbrook, B., Tubiello, F. N., van der Laan-Luijkx, I. T., van
- der Werf, G. R., van Heuven, S., Viovy, N., Vuichard, N., Walker, A. P., Watson, A. J.,
- 1054 Wiltshire, A. J., Zaehle, S., Zhu, D. 2018. Global Carbon Budget 2017, Earth Syst. Sci. Data,
- 1055 10, 405-448, https://doi.org/10.5194/essd-10-405-2018.
- 1056 Lobe, I., A. Sandhage-Hofmann, S. Brodowski, C. C. du Preez and W. Amelung. 2011.
- 1057 Aggregate dynamics and associated soil organic matter contents as influenced by
- prolonged arable cropping in the South African Highveld. Geoderma 162. 3-4: 251-259.
- 1059 Lobry de Bruyn, L. A. & Conacher, A. J. 1990. The role of termites and ants in soil
- modification: A review. Aust. J. Soil Res. 28: 55-93.
- 1061 Lubbers, I., Pulleman, M., Van Groenigen, J. W. 2017. Can earthworms simultaneously
- 1062 enhance decomposition and stabilization of plant residue carbon? Soil Biol. Biochem.
- **1063** 105:12-24.

- 1064 Marquez, C. O., V. J. Garcia, R. C. Schultz and T. M. Isenhart. 2019. A conceptual framework
- to study soil aggregate dynamics. Eur. J. Soil Sci. 70. 3: 466-479.
- 1066 Martin A. 1991. Short- and long-term effects of the endogeic earthworm *Millsonia anomala*
- 1067 (Omodeo) (Megascolecidae, Oligochaeta) of tropical savannas, on soil organic matter. Biol.
- **1068** Fertil. Soils, 11. 234-238.
- 1069 Martin, S., and Lavelle, P., 1992. A simulation model of vertical movements of an earthworm
- 1070 population (Millsonia anomala Omodeo, Megascolecidae) in an African savanna (Lamto,
- 1071 Ivory Coast). Soil Biol. Biochem., 24, 1419-1424.
- 1072 Materechera, S.A., Dexter, A.R., Alston, A.M., 1992. Formation of aggregates by plant roots
- **1073** in homogenized soils. Plant & Soil. 142, 1, 69-79.
- 1074 Mermut, A R, Arshad, M A., & Arnaud, R J. 1984. Micro pedological study of termite
- mounds of three species of *Macrotermes* in Kenya. Soil Sci. Soc. Amer- J. 48. 613-620.
- 1076 Miller, R.M., & Jastrow, J.D. 1990. Hierarchy of root and mycorrhizal fungal interactions
- with soil aggregation. Soil Biol. Biochem., 22, 5, 579-584.
- 1078 Millogo, Y., Hajjaji, M. & Morel, J. C. 2011. Physical properties, microstructure and
- mineralogy of termite mound material considered as construction materials. App. Clay Sci.
- **1080** 52. 1-2: 160-164.
- 1081 Minasny, B., Malone, B. P., Mc Bratney, A. B., Angers, D. A, Arrouays, D., Chambers, A., et
- **1082** al., 2017. Soil carbon 4 per mille. Geoderma, 292, 59-86.
- 1083 Monroe, C.D., Kladivko, E.J., 1987. Aggregate stability of a silt loam soil as affected by roots
- 1084 of corn, soybeans and wheat. Communications in Soil Science and Plant Analysis 18, 10:
- **1085** 1077-1087.
- 1086 Mora P, Seuge C, Chotte JL, Rouland C. Physico-chemical typology of the biogenic
- structures of termites and earthworms: a comparative analysis. 2003. Biol. Fert. Soil., 37(4),
- **1088** 245-249.
- 1089 Morel, J.L., Habib, L., Plantureux, S., Guckert, A. 1991. Influence of maize root mucilage on
- **1090** soil aggregate stability. Plant & Soil, 136, 1, 111-119.
- 1091 Negassa, W. C, Guber, A. K, Kravchenko, A. N, Marsh, T. L, Hildebrandt, B., Rivers, M. L.
- 1092 2015. Properties of Soil Pore Space Regulate Pathways of Plant Residue Decomposition and
- 1093 Community Structure of Associated Bacteria. Plos One. 10(4).
- Nunan, N., Wu, K. J., Young, I. M., Crawford, J. W., Ritz, K., 2003. Spatial distribution of
- 1095 bacterial communities and their relationships with the micro-architecture of soil. FEMS
- **1096** Microb. Ecol. 44, 2, 203-15.

- 1097 Paul, E. A. 2016. The nature and dynamics of soil organic matter: Plant inputs, microbial
- transformations, and organic matter stabilization. Soil Biol. Biochem. 98:109-26.
- 1099 Paustian K., Lehmann, J., Ogle, S., Reay, D., Robertson, G. P., Smith, P. 2016. Climate-smart
- 1100 soils. Nature., 532(7597), 49-57.
- 1101 Peth, S., Nellesen, J., Fischer G., Horn R. 2010. Non-invasive 3D analysis of local soil
- 1102 deformation under mechanical and hydraulic stresses by mu CT and digital image
- **1103** correlation. Soil & Till. Res. 111, 1: 3-18.
- 1104 Piccolo, A., Pietramellara, G., Mbagwu, J. S. C., 1997. Use of humic substances as soil
- conditioners to increase aggregate stability. Geoderma, 75 (3-4), 267-77.
- 1106 Plante, A. F., Mc Gill, W. B. 2002. Soil aggregate dynamics and the retention of organic
- 1107 matter in laboratory-incubated soil with differing simulated tillage frequencies. Soil. Till.
- **1108** Res. 66, 79-92.
- 1109 Poeplau, C. and A. Don. 2015. Carbon sequestration in agricultural soils via cultivation of
- 1110 cover crops A meta-analysis. Agr. Ecos. & Envir. 200, 33-41.
- 1111
- 1112 Post, W. M. and K. C. Kwon. 2000. Soil carbon sequestration and land-use change:
- processes and potential. Glob. Ch. Biol. 6, 3: 317-327.
- 1114
- 1115 Powlson, D. S., C. M. Stirling, C. Thierfelder, R. P. White and M. L. Jat. 2016. Does
- 1116 conservation agriculture deliver climate change mitigation through soil carbon
- sequestration in tropical agro-ecosystems? Agr. Ecos. & Envir. 220, 164-174.
- 1118 Puget, P., Angers, D.A., Chenu, C., 1999. Nature of carbohydrates associated with water-
- 1119 stable aggregates of two cultivated soils. Soil Biol. Biochem., 31(1), 55-63.
- 1120 Pulleman, M. M. and J. C. Y. Marinissen. 2004. Physical protection of mineralizable C in
- **1121** aggregates from long-term pasture and arable soil. Geoderma 120, 3-4: 273-282.
- 1122
- 1123 Pulleman, M. M., J. Six, A. Uyl, J. C. Y. Marinissen and A. G. Jongmans. 2005. Earthworms
- 1124 and management affect organic matter incorporation and microaggregate formation in
- agricultural soils. Appl. Soil. Ecol. 29, 1: 1-15.
- 1126 Rajagopal, D., Sathyanarayana, T., & Veeresh, G. K. 1982. Physical and chemical properties of
- termite mound and surrounding soils of Karnataka. J. Biol. Ecol. Soil. 2, 18-31.
- 1128 Reatto, A., Bruand, A., Martins, E. D., Muller, F., da Silva E. M., de Carvalho, O. A., Brossard,
- 1129 M., Richard, G. 2009. Development and origin of the microgranular structure in latosols of
- 1130 the Brazilian Central Plateau: Significance of texture, mineralogy, and biological activity.
- **1131** Catena 76. 2: 122-134.

- 1132 Reid, J.B., Goss, M.J., 1981. Effect of living roots of different plant species on the aggregate
- stability of two arable soils. J. Soil Sci., 32, 4, 521-541.
- 1134 Rieu, M., & Sposito, G., 1991. Fractal fragmentation, soil porosity, and soil-water properties
- 1135 .1. Theory. Soil Sci. Soc. Amer. J., 55, 5, 1231-1238.
- 1136 Rillig, M. C., Mummey, D. L. 2006. Mycorrhizas and soil structure. New Phytol. 171(1), 41-
- **1137** 53.
- 1138 Roose, T., Keyes, ., S. D., Daly, K. R., Carminati, A., Otten, W., Vetterlein D., Peth, S. 2016.
- 1139 Challenges in imaging and predictive modeling of rhizosphere processes. Plant & Soil 407.
- **1140** 1-2: 9-38.
- 1141 Scarciglia, F. and D. Barca. 2017. A powerful tool for assessing distribution and fate of
- potentially toxic metals (PTMs) in soils: integration of laser ablation spectrometry (LA-ICP-
- 1143 MS) on thin sections with soil micromorphology and geochemistry. Environmental Science
- **1144** and Pollution Research 24. 10: 9776-9790.
- 1145
- 1146 Segoli, M., De Gryze, S., Dou, F., Lee, J., Post, W. M., Denef, K., Six, J. 2013. AggModel: A soil
- organic matter model with measurable pools for use in incubation studies. Ecol. Model.
- **1148** 263: 1-9.
- 1149 Shipitalo, M. J., & R. Pots. 1988. Factors influencing the dispersibility of clay in worm casts.
- 1150 Soil Science Society of America Journal, 52, 764-769.
- 1151 Six, J., Elliott, E.T., Paustian, K. 2000a. Soil macro-aggregate turnover and microaggregate
- 1152 formation: a mechanism for C sequestration under no-tillage agriculture. Soil Biol.
- 1153 Biochem., 32, 14, 2099-2103.
- 1154 Six, J., Paustian, K., Elliott, E.T., Combrink, C. 2000b. Soil structure and organic matter: I.
- 1155 Distribution of aggregate-size classes and aggregate-associated carbon. Soil Sci. Soc. Am. J.,
- **1156** 64, 2, 681-689.
- 1157 Six, J., Conant, R.T., Paul, E.A., Paustian, K., 2002a. Stabilization mechanisms of soil organic
- matter: Implications for C-saturation of soils. Plant & Soil, 241, 155-176.
- 1159 Six, J., Feller, C., Denef, K., Ogle, S.M., Sa, J.C.D., Albrecht, A. 2002b. Soil organic matter,
- biota and aggregation in temperate and tropical soils Effects of no-tillage. Agronomie, 22,
- **1161** 7-8, 755-775.
- 1162 Six, J., H. Bossuyt, S. De Gryze and K. Denef. 2004. A history of research on the link between
- 1163 (micro)aggregates, soil biota, and soil organic matter dynamics. Soil & Tillage Research 79.
- **1164** 1: 7-31.
- 1165 Six, J. and Paustian, K. 2014. Aggregate-associated soil organic matter as an ecosystem
- property and a measurement tool. Soil Biol. & Biochem. 68: A4-A9.

- 1167 Smith, P., House, J. I., Bustamante, M., Sobocka, J., Harper, R., Pan, G. X., et al. 2016. Global
- change pressures on soils from land use and management. Glob. Ch. Biol. 22(3):1008-28.
- 1169 Sollins, P., P. Homann and B. A. Caldwell. 1996. Stabilization and destabilization of soil
- organic matter: Mechanisms and controls. Geoderma 74, 1-2: 65-105.
- 1171 Tisdall, J.M., Oades, J.M., 1982. Organic matter and water-stable aggregates in soils. J. Soil
- **1172** Sci., 33, 2, 141-163.
- 1173 Topoliantz, S., Ponge, J.F., Viaux, P. 2000. Earthworm and enchytraeid activity under
- 1174 different arable farming systems, as exemplified by biogenic structures. Plant & Soil, 225, 1-
- **1175** 2, 39-51.
- 1176 Tötsche, K. U., Amelung, W., Gerzabek, M. H., Guggenberger, G., Klump, E., Knief, C.,
- 1177 Lehndorff, E., Mikutta, R., Peth, S. Prechtel, A. Ray, N. Kogel-Knabner, I., 2018. Micro-
- aggregates in soils. J. Plant Nutr. Soil Sci., 181(1):104-36.
- 1179 Toutain F. 1981. Les humus forestiers : structure et mode de fonctionnement. R.F.F. XXXIII.
- **1180** 449-477.
- 1181 Tracy, S. R., K. R. Daly, C. J. Sturrock, N. M. J. Crout, S. J. Mooney and T. Roose. 2015. Three-
- dimensional quantification of soil hydraulic properties using X-ray Computed Tomography
- and image-based modeling. Water Resources Research 51. 2: 1006-1022.
- 1184 Velasquez, E., Lavelle, P., Andrade, M., 2007b. GISQ, a multifunctional indicator of soil
- **1185** quality. Soil Biol. Biochem., 39, 12, 3066-3080.
- 1186 Velasquez, E., Pelosi, C., Brunet, D., Grimaldi, M., Martins, M., Rendeiro, A. Barrios, E, & P.
- 1187 Lavelle. 2007a This ped is my ped: visual separation and near infrared spectra allow
- determination of the origins of soil macro-aggregates. Pedobiologia, 51, 75-87.
- 1189 Velasquez, E. and P. Lavelle. 2019. Soil macrofauna as an indicator for evaluating soil-based
- 1190 ecosystem services in agricultural landscapes. Acta Oecologica-International J. of Ecol., 100.
- 1191 von Lutzow, M., I. Kogel-Knabner, K. Ekschmitt, E. Matzner, G. Guggenberger, B. Marschner
- 1192 and H. Flessa. 2006. Stabilization of organic matter in temperate soils: mechanisms and
- their relevance under different soil conditions a review. Eur. J. Soil Sci.57. 4: 426-445.
- 1194 Wang, J. Y., Z. Q. Xiong and Y. Kuzyakov. 2016. Biochar stability in soil: meta-analysis of
- decomposition and priming effects. Glob. Ch. Biol. Bioenergy 8, 3: 512-523.
- 1196 Wielemaker, W. G. 1984. Soil formation by termites. PhD thesis. Wageningen University.
- 1197 Wollenberg, E., Richards, M., Smith, P., Havlik, P., Obersteiner, M., Tubiello, F. Herold N, M.,
- 1198 Gerber, P. Carter S., Reisinger A., van Vuuren D. P., Dickie A., Neufeldt, H. Sander B. O.,
- 1199 Wassmann, R. Sommer R., Amonette, J. E. Falcucci A., Herrero. M, Opio, C. Roman-Cuesta,
- 1200 R. M. Stehfest, E. Westhoek H. Ortiz-Monasterio, I, Sapkota, T. Rufino, M. C. Thornton P. K.,

- 1201 Verchot, L. West, P. C. Soussana, J. F. Baedeker, T. Sadler, M. Vermeulen S., Campbell B. M.
- 1202 2016. Reducing emissions from agriculture to meet the 2 degrees C target. Glob. Ch. Biol.
- **1203** 22, 12:3859-64.
- 1204 Yamaguchi, T., Takei, T., Yazawa, Y., Wong, M. T. F., Gilkes, R. J., Swift, R. S. 2004. Effect of
- 1205 humic acid, sodium, and calcium additions on the formation of water-stable aggregates in
- 1206 Western Australian wheatbelt soils. Aust. J. Soil Res. 2004;42(4):435-9.
- 1207 Young, I. M., Crawford, J. W. 2004. Interactions and self-organisation in the soil-microbe
- 1208 complex. Science. 304, 1634-1637.
- 1209 Yudina, A. and Y. Kuzyakov. 2019. Saving the face of soil aggregates. Glob. Ch. Biol25. 11:
- **1210** 3574-3577.
- 1211 Zangerlé, A., Pando, A., & P. Lavelle, 2011. Do earthworms and roots cooperate to build soil
- macro-aggregates? A microcosm experiment. Geoderma, 167, 303-309.
- 1213 Zangerlé, A., Hissler, C., Blouin, M and Lavelle, P. 2014. Near Infrared spectroscopy (NIRS)
- to estimate earthworm cast age. Soil Biol. & Biochem., 70, 47-53.
- 1215 Zangerlé A., Hissler, C. McKey, D. & P. Lavelle. 2016. Using near infrared reflectance
- spectroscopy (NIRS) to identify the contribution of earthworms to soil macro-aggregation in
- **1217** field conditions. Applied Soil Ecology, 104, 138-147.
- 1218 Zhao, D., M. X. Xu, G. B. Liu, X. Yao, D. F. Tuo, R. R. Zhang, T. Q. Xiao and G. Y. Peng. 2017.
- 1219 Quantification of soil aggregate microstructure on abandoned cropland during vegetative
- succession using synchrotron radiation-based micro-computed tomography. Soil & Til. Res.
- **1221** 165, 239-246.
- 1222 Zhang, S., Q. Li, Y. Lu, X. Zhang and W. Liang. 2013. Contributions of soil biota to C
- 1223 sequestration varied with aggregate fractions under different tillage systems. Soil Biol. &
- **1224** Biochem. 62, 147-156.
- 1225 Zhang, S., J. Li, X. Yang and B. Sun. 2016. Long-term effects of soil management regimes on
- carbon contents and respiration rates of aggregate size fractions. Soil Use & Manag. 32, 4:
- **1227** 525-534.
- 1228 Zhu, G. Y., Z. P. Shangguan and L. Deng. 2017. Soil aggregate stability and aggregate-
- 1229 associated carbon and nitrogen in natural restoration grassland and Chinese red pine
- 1230 plantation on the Loess Plateau. Catena 149, 253-260.