

The Human Spleen in Malaria: Filter or Shelter?

Benoît Henry, Camille Roussel, Mario Carucci, Valentine Brousse, Papa Alioune Ndour, Pierre Buffet

▶ To cite this version:

Benoît Henry, Camille Roussel, Mario Carucci, Valentine Brousse, Papa Alioune Ndour, et al.. The Human Spleen in Malaria: Filter or Shelter?. Trends in Parasitology, 2020, 36, pp.435 - 446. 10.1016/j.pt.2020.03.001 . hal-03490391

HAL Id: hal-03490391

https://hal.science/hal-03490391

Submitted on 22 Aug 2022

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



The human spleen in malaria: filter or shelter?

- 3 Benoît Henry^{1,2,3,4}, Camille Roussel^{1,2,3}, Mario Carucci^{1,2,3}, Valentine Brousse^{1,2,3}, Papa
- 4 Alioune Ndour^{1,2,3}, Pierre Buffet^{1,2,3,4*}
- ¹Université de Paris, Biologie Intégrée du Globule Rouge, UMR_S1134, BIGR, INSERM, F-75015
- 6 Paris, France

1

2

- 7 ²Institut National de la Transfusion Sanguine, 6, rue Alexandre Cabanel, 75015 Paris, France
- 8 ³Laboratoire d'Excellence Gr-Ex, 24, boulevard du Montparnasse, 75015 Paris, France
- 9 ⁴APHP, Hôpital Necker Enfants Malades, Service des maladies infectieuses et tropicales, Centre
- d'Infectiologie Necker-Pasteur, Institut Imagine, 149, rue de Sèvres, 75015 Paris, France
- *Correspondence: <u>pabuffet@gmail.com</u> (P. Buffet)
- 13 **Keywords**: malaria; spleen; *Plasmodium*; gametocytes; hyper-reactive malarial
- splenomegaly; artemisinin.

16 Abstract

12

15

- 17 The human spleen is an immune sentinel and controls red blood cell (RBC) quality. By
- mechanically retaining subsets of infected RBC, the spleen may reduce the pace at which the
- 19 parasite biomass increases before the adaptive immune response operates. Conversely, the
- 20 spleen may contribute to malaria pathogenesis, particularly anemia that is associated with
- 21 splenomegaly. Large spleens may also shelter parasites in chronic carriers. Upon treatment
- 22 with artemisinins, the spleen clears circulating parasites by pitting and releases "once-
- 23 infected" RBC in circulation. This triggers post-artesunate delayed hemolysis and explains the
- long post-treatment positivity of HRP2-based dipsticks. Importantly, splenic retention of RBC
- also applies to gametocytes, the clearance of which may be enhanced by stiffening them with
- 26 drugs, a potential way to block malaria transmission.

Major importance of red blood cells-spleen interactions in the pathogenesis of human

malaria

27

28

29

30

31

32

33

34

35

36

37

38

39

40

41

42

43

44

45

46

47

48

49

50

Malaria is the most prevalent severe parasitic disease affecting humans. The causal microorganism, *Plasmodium* spp., develops in hepatocytes then in red blood cells (RBC), but only the asexual multiplication in RBC causes clinical manifestations. Invasion of RBC by Plasmodium triggers major alterations in this host cell. P. falciparum, the human-infecting *Plasmodium* specie that has been studied in greatest details, expresses parasitic proteins at the RBC surface, and modifies its deformability and adhesiveness. Adhesion of parasitized RBC to endothelial cells (and to other infected or uninfected RBC) is central in malaria pathogenesis but parasite-induced alterations of RBC deformability have also been associated with the severity of malaria [1-3]. The white pulp is a major operator of the humoral immune response, especially to circulating

White and red pulp structures (See Glossary) have specific functions in the human spleen.

antigens. The red pulp exerts a unique and subtle control on the surface integrity and

biomechanical properties of RBC. To be left in circulation, RBC must be fit enough to cross a

very specific structure of red pulp sinuses, the inter-endothelial slit (IES; Figure 1). Older

RBC, or RBC modified by innate or acquired conditions are eventually retained in the splenic

red pulp and processed by red pulp macrophages (RPM) [4].

The major modifications of RBC biomechanical properties induced by *Plasmodium* infection, point to the spleen as an important player in the pathogenesis of human malaria. The proportion of subjects with splenomegaly (the splenic index) has long been a defining marker of malaria endemicity. Although knowledge has progressed slowly due to the risk associated with splenic puncture or biopsy and limited access to imaging in endemic areas, significant advances have been made on the role of the spleen in malaria over the last few decades, especially regarding the innate control of infection and transmission [4,5] (Box 1). Ex vivo perfusion of human spleens [6] and the development of biomimetic tools [7–9] have generated new insights. The role of cell-mediated immunity has been extensively studied in murine models of malaria, and in malaria-infected or exposed humans through the analysis of circulating cell populations. However, the aforementioned constraints have hampered progress in the understanding of intra-splenic immune responses to malaria in humans. If the important role of platelets in malaria pathogenesis is likely [10], contribution of intra-splenic platelets, which represent a third of the total human pool, remains largely unexplored. In addition, substantial anatomic differences exist between human and murine spleens, especially regarding the red pulp which is devoid of bona fide IES in mice [11] (although mechanical filtration exists), and between human and murine *Plasmodium* species. Whether mouse models of malaria are relevant to explore the pathogenesis of human malaria is still a matter of controversy [12]. Unlike the spleen of humans and rats, the mouse spleen is devoid of sinuses. The mechanical sensing of RBC thus depends on holes in venules rather than on inter-endothelial slits in sinus walls [13]. It is unclear whether these markedly different microanatomic structures display a close or loose functional convergence, and whether the mouse spleen can pit RBC containing undeformable bodies, a major parasite clearance process in humans (please see section "Role of the spleen in parasite clearance following antimalarial therapy"). Whether the splenic function is preserved in humanized mice infected with P. falciparum is also questionable as stiff immature gametocyte and mature asexual stages – which should be filtered out by the spleen – are observed in the circulation of these mice [14]. We have therefore limited this review to observations in humans.

73

74

51

52

53

54

55

56

57

58

59

60

61

62

63

64

65

66

67

68

69

70

71

72

Functional anatomy of the human spleen

General features

The spleen is an oval retroperitoneal organ of 10 to 12 cm in its major axis, weighing 100 to 200 grams in adults, surrounded by a collagenous, poorly extensible capsule. It is considered poorly contractile in humans compared to diving or fast-running other animal species (seals, whales, dogs, horses) which harbor large "storage spleens". Splenic parenchyma is composed of 2 specific tissues, the red pulp and the white pulp. The **white pulp** is made of immune cells, organized in periarterial lymphoid sheaths containing mostly T cells, and lymphoid follicles containing mostly B cells. Follicles are surrounded by the marginal and perifollicular zones. The red pulp accounts for 75% of splenic volume and is composed of a unique association of **cords** and **venous sinuses**. Cords are circulatory structures that contain fibroblasts and, very predominantly, specific macrophages (RPM). Venous sinuses which collect blood from the cords, are made of endothelial cells, which are parallel to the blood flow, and are surrounded by a helicoid basal fiber (i.e., highly discontinuous) which leaves room for narrow spaces between endothelial cells (0.2 to 2 μ m), named inter-endothelial slits [13]. Rare plasma cells, mastocytes and other white blood cells are also observed in the red pulp.

Microcirculation in the human spleen

Overall, about 5% of arterial flow enters the spleen through the splenic artery, and is divided into 2 paths: approximately 85% of splenic flow enters a fast, closed circulation, which navigates around white pulp structures and follows a specific microcirculation from arterioles to venules delimited by specific MAdCAM1-positive cells in the perifollicular zone; 15% engage into the open and slow circulation in the red pulp, through the cords, then cross the unique structure of IES, to join venous sinuses upstream from venules. Crossing the IES is the

most stringent biomechanical challenge for circulating RBC. RBC altered by heat [15], stiffening chemicals [16], genetic defects [17], possibly pretransfusion storage [18], and *P. falciparum* [19,20] are retained in the cords, where RPM can process them.

Variable definitions of the human marginal zone

In rodents, a marginal zone (MZ), containing fibroblasts, dendritic cells, lymphocytes and specific macrophages ("metallophilic macrophages" and "MZ macrophages") has been described in the surroundings of lymphoid follicles, and is separated from the white pulp by the marginal sinus [13]. Such a structure is not present in humans, although an important population of B cells is found in this area, the MZ B cells, which have a major role in the early, T-independent antibody response to circulating antigens. Hence, the "human equivalent of MZ" is sometimes renamed peri-follicular zone, or as recently proposed, superficial zone [11].

Malaria in splenectomized subjects: a more severe disease?

From 1931 through 2019, 133 malaria attacks were reported in splenectomized patients, among which 69 were due to *P. falciparum*, 26 to *P. vivax*, 6 to *P. knowlesi*, and 3 to *P. malariae*. Infection was mixed in 1 case and in 28 cases, *Plasmodium* species was not reported [4] (supplemental Table S1). Only 2 cohorts assessing the risk of malaria in splenectomized subjects (33 in Malawi, 11 in Indonesia) have been prospectively followed and compared to controls. In both cohorts, acute malaria attacks were significantly more frequent in splenectomized patients [21,22]. In this population, severe attacks are also more frequent, mature forms of *P. falciparum* - which are normally sequestered by adherence to the

endothelium - are observed in circulation, and post-treatment parasite clearance is delayed [23,24]. However, the over-incidence of severe forms and mortality has not been confirmed by prospective controlled studies and its high frequency in published case reports or series may either reflect a genuine over-susceptibility or result from a publication bias. An increased risk of acute *P. vivax* attacks after splenectomy has been recently reported, and outweighs the risk of *P. falciparum* attacks (hazard ratio 7.7 vs 2.3) [22], without evidence for an increased risk of severe forms in this immune population. One case of acute malaria due to *P. knowlesi* has been described in an asplenic patient [25], and in another case a prolonged parasite clearance has been reported [26]. In summary, the increased risk of malaria has been robustly described in immune or semi-immune subjects, and is strongly suspected in non-immune travelers [27]. Non-immune splenectomized hosts should be very cautious when travelling to malaria-endemic areas. If these patients must travel to endemic areas, strict compliance to antimalarial prophylactic measures is warranted.

The spleen in acute malaria

Protective role

The human spleen senses subtle alterations in RBC deformability. This decreased deformability of *Plasmodium*-infected RBC (asexual stages), first observed with *P. knowlesi* [3], then with *P. falciparum* [1] is intense in RBC infected by mature forms, but already present, albeit moderately, at the ring stage [1,7,19,20,28]. Mechanisms include alterations of surface-to-volume ratio [20], and interactions between parasite proteins and the RBC cytoskeleton [29,30] which induce membrane rigidification. Importantly, **ektacytometry** - a reference technique to quantify RBC deformability [31] - and microfluidics have also

demonstrated that during acute malaria attacks, even non-infected RBC are less deformable than normal control RBC [2,28,32].

As evidenced by *ex vivo* perfusions of human spleens with *P. falciparum*-infected RBC, the spleen retains a proportion (approximatively half) of ring-infected RBC, in addition to the expected, almost complete, retention of schizont-infected RBC [19]. The mechanical retention of schizont-infected RBC is expected to display a small pathogenic impact *in vivo*, as the majority of mature forms cytoadhere in other organs, which prevents them from being filtered by the spleen. Retention of rings which affects a predominantly circulating component of the parasite biomass, takes place in the red pulp. This mechanism, which is difficult to explore directly in human subjects, is expected to reduce the circulating parasitic load, thereby also reducing the parasite biomass amenable to sequester in other organs [19].

The increased clearance of all RBC (infected and uninfected) by the spleen after acute malaria has been demonstrated through the autotransfusion of radiolabeled RBC. Thai patients with acute malaria had indeed an enhanced clearance of heat-stiffened RBC when splenomegaly was present [15] that also affected labeled normal RBC in a similar post-malarial context [33]. This enhanced splenic clearance of abnormal and normal RBC, which last several weeks, may contribute to the partial control of parasite loads. When comparing cerebral malaria with other severe forms (anemia, respiratory distress, acute renal and/or liver failure, shock), post-mortem studies also showed that intra-splenic **erythrophagocytosis** was more frequent in the latter [34]. Splenomegaly is also less frequent in cerebral malaria than in severe malarial anemia, as demonstrated by field studies performed in Uganda and Sudan, and including a total of 1108 children [35–37]. In one of these studies, splenomegaly was also associated with a survival advantage [37]. The human spleen can thus retain part of the parasite biomass, which may partially control the course of infection, potentially reducing the risk of cerebral malaria. This would, however, come at the expense of an increased retention and clearance of

non-infected RBC, leading to splenomegaly and anemia (Figure 1). Exploration of the innate retention of (dead or live) rings in the controlled human infection model [38] would be the most direct and relevant way to assess whether this potentially powerful protective process indeed exists, in a context of limited splenic activation at early steps of human infection, when splenomegaly is not yet present [15].

175

176

177

178

179

180

181

182

183

184

185

186

187

188

189

190

191

192

193

170

171

172

173

174

Detrimental role

The pathogenesis of acute malarial anemia is complex and only partially understood. It is associated with intra- and extra-vascular hemolysis, dyserythropoiesis, and the clearance of larger proportions of uninfected than infected RBC [39]. Several observations point to a splenic contribution to the pathogenesis of acute malarial anemia. Congestion of the red pulp is a key post-mortem feature in fatal malaria [40,41]. Decreased RBC deformability, which triggers intrasplenic retention ex-vivo (see above), correlates with hemoglobin nadir [32]. As stated above, splenomegaly is more prevalent in patients with severe malarial anemia than in other severe forms, and splenic clearance of RBC is increased after malarial attacks. Not least, erythrophagocytosis is enhanced in acute malaria and alterations of infected RBC render them prone to phagocytosis [42]. The phagocytic function of monocytes is also activated in malaria [43]. That erythrophagocytosis in malaria predominantly takes place in the macrophage-rich spleen remains to be directly demonstrated in humans. Very rarely, splenic-specific complications of malarial infection occur: pathologic splenic rupture, intra-splenic hematoma and splenic infarction. These complications have received little attention so far, and their precise mechanisms remain elusive [44]. In summary, multiple observations suggest a protective role for the spleen in acute malaria, even if definitive evidence from controlled studies is lacking. The innate retention of rings can prevent the rise

of peripheral parasite load thereby decreasing the risk of cytoadherence and sequestration of parasitized RBC in vital organs [45]. The potential down-side of this phenomenon is a splenic contribution to malarial anemia.

Role of the spleen in parasite clearance following antimalarial therapy

The pitting process

In patients recently cured from malaria by artemisinin derivatives, a proportion of circulating RBC display parasite proteins at the internal layer of their plasma membrane [46]. These "once-infected" RBC do not contain parasites [23,46,47] and their presence results from a spleen-specific physiological process called **pitting**, whereby an intra-erythrocytic body is extracted from the RBC without cell lysis [48]. Pitting clears physiological intra-erythrocytic items such as Howell-Jolly or Pappenheimer bodies. Malaria-related pitting can be quantified in the peripheral blood of patients through the double-staining of RBC with a nucleic acid marker and an antibody against parasite proteins like RESA (Ring-infected Erythrocyte Surface Antigen). Pitted RBC are parasite-negative and parasite membrane protein-positive. Pitting is not observed in splenectomized subjects [23], and has been replicated in human spleens perfused *ex vivo* with parasitized RBC pre-exposed to artemisinin derivatives [6]. Pitting does not occur (or to a very low level) with RBC containing live parasites, and has been so far explored only in *P. falciparum* infections.

Contribution of pitting to parasite clearance

In non-immune travelers treated with artemisinins, pitting is a major mechanism of parasite clearance [49]. In artemisinin-treated children living in malaria-endemic Mali, clearance

mechanisms may be more complex and influenced by the immune status of the host. Pitting appears to be an important contributor to clearance in infants, who are expected to have low anti-parasite humoral immunity; however in older children, immunity correlates with an even faster parasite clearance, likely through immune-mediated phagocytosis, and possibly through splenic retention without pitting [50]. Pitting is marked in patients treated with intravenous artesunate or oral artemisinin derivatives, and much lower after treatment with quinine, mefloquine or atoyaquone-proguanil [51].

Contribution of pitting to post-artesunate hemolysis

In Japanese then in German travelers treated with artesunate for hyperparasitemic severe malaria [52,53], acute hemolysis was observed 1 to 2 weeks after therapy. These hemolytic episodes were moderately intense and affected 5-27% of travelers [54,55]. Transfusion was required in 5 to 50% of cases [56] and no conventional cause of hemolysis was found. Initial observations in Southeast Asia had shown that once-infected RBC have a shorter lifespan than normal RBC [47]. In travelers, these hemolytic episodes (named **Post-Artesunate Delayed Hemolysis**, PADH) and the disappearance of once-infected RBC from the circulation were simultaneous. PADH is thus likely triggered by the clearance of once-infected RBC [57]. The peak concentration of once-infected RBC quantified by flow cytometry, 2-7 days after treatment initiation, is predictive of PADH [57,58]. Predicting PADH in resource-limited settings is important, as weekly follow-up of all patients recovering from severe malaria and preparation of transfusion are logistically demanding. Like RESA, the parasitic Histidin-Rich Protein 2 (HRP-2) is left as an imprint at the membrane of pitted RBC [58]. HRP-2 is the antigen used in many immunochromatographic malaria **rapid diagnostic tests**. The persistent positivity of the diagnostic dipstick, performed on diluted whole blood 3 to 7 days after

initiation of artesunate accurately predicted PADH [58]. Whether PADH is a significant problem in endemic areas has been a subject of debate; a prospective study following 217 Congolese children with acute malaria, treated with quinine or IV artesunate, and followed until 42 days after treatment only showed PADH in less than 1% of cases [59]. These patients, however, had parasitemia below the conventional 10% threshold for hyperparasitemia and had uncomplicated malaria. Loosely defined delayed hemolysis has been observed in 7% of African children treated with artesunate for severe malaria [60]. Further prospective studies in African children are ongoing. Interestingly, this approach also explains why HRP2-based dipsticks remain positive days to weeks after treatment of a malaria-attack with artemisinins. HRP2 persists in circulation not as soluble protein in plasma but as a cytoskeleton-associated protein in once-infected RBC.

The spleen in chronic parasite carriage.

A splenic contribution to chronic malarial anemia?

The mechanisms of malarial anemia are influenced by the infecting *Plasmodium* species, age and transmission intensity, amongst others factors. Excellent reviews have dissected these complex parameters [39,61]. Briefly, most important contributors are intra- and extra-vascular hemolysis, erythrophagocytosis, dyserythropoiesis (favored by frequent comorbidities such as iron or vitamin B12 deficiency, or malnutrition), some degree of bone marrow insufficiency, and possibly splenic clearance. The role of the spleen in chronic parasite carriage is complex and very difficult to explore. One study carried in Ghana [62] found a relationship between chronic parasite carriage, anemia and splenomegaly, as observed in multiple other populations and settings. This may reflect any combination of long-lasting RBC trapping in the spleen, enhanced erythrophagocytosis or increased immune reactivity.

Parasite biomass in the spleen

Recently, comprehensive pathological and molecular examinations of spleens (and concomitantly of peripheral blood) in mostly untreated patients undergoing splenectomy in a highly *P. falciparum* and *P. vivax*-endemic area of Indonesia have shown that, compared to peripheral blood, non-phagocytosed parasitized RBC concentrate into the spleen by several orders of magnitude [63]. Concentration of parasitized RBC was greatest in the splenic red pulp of *P. vivax*-infected patients. Red pulp congestion by infected and uninfected RBC was proportional to spleen weight. These major observations not only bring some substance to the protective role of the spleen filter in parasite clearance but also suggest that the spleen may be a parasite shelter, possibly supporting a cryptic intrasplenic parasitic cycle [4,64].

Hyper-reactive malarial splenomegaly

This condition, initially named tropical splenomegaly syndrome, then more precisely described in 1957, has been progressively deciphered over the last decades, the name hyper-reactive malarial splenomegaly (HMS) being proposed in 1983 [65]. The currently accepted diagnostic criteria for HMS associate marked splenomegaly, total plasma IgM above 2 times the local standard deviation, clinical response after antimalarials, and polyclonal lymphocytic response [66]. HMS is believed to account for an important proportion (40%) of massive splenomegaly in malaria-endemic areas [67], but recent and precise epidemiological data regarding its prevalence in general population are scarce. In the Gambia, 3 decades ago, the prevalence of HMS was estimated at 0.16% [68]. HMS has essentially been described in subjects chronically exposed to the parasite and is very rare in travelers. Typically, patients present with a gross and constant splenomegaly, sometimes clinically patent (abdominal

discomfort) and inconstantly associated with hepatomegaly. Total IgM are markedly elevated, with a polyclonal pattern. This elevation usually precedes splenomegaly. Cytopenia of variable importance can be present, usually attributed to hypersplenism. Positive diagnosis of malaria is difficult in HMS, as parasite load is usually very low, and only detected by molecular methods. Biological features of autoimmunity (cryoglobulinemia, rheumatoid factor, anti-nuclear antibodies) have been associated with HMS. Recently, a relatively specific pattern of anti-nuclear antibodies has been associated with chronic malaria [69].

289

290

291

292

293

294

295

296

297

298

299

300

301

302

303

304

305

306

307

308

309

310

311

312

313

A genetic component of HMS is suspected, based on familial studies [70], but its precise basis remains unknown (although certainly not Mendelian). Pathologic examination of the spleen in HMS has been rarely reported; published cases described essentially red pulp congestion and erythrophagocytosis, with only brief descriptions of the white pulp [71–73]. The pathogenesis of HMS is poorly understood, the most commonly accepted hypothesis involving a defect in regulatory T cells, leading to a B cell expansion after malaria infection, production of immune complexes which would be eventually phagocytosed by splenic macrophages. If this hypothesis is in line with splenomegaly and the enhanced immune response observed in HMS, it however falls short in providing an explanation for anemia and lower parasitic load. The evolution of HMS in endemic areas remains largely unknown. Historical data pointing a very important mortality (>50%) might be biased by the imprecision regarding the causes of death [74]. Scarce data have been collected in travelers, showing that regression of hyper IgM and splenomegaly are slow, over several months [75]. Travelers or expatriates affected by HMS exhibit a tendency to suffer from similar episodes of HMS upon re-exposure to the parasite, consistent with an innate pathogenic component and genetic susceptibility [76]. A major issue is the risk of evolution towards a low-grade marginal-zone splenic lymphoma (MZSL). This association, suspected 50 years ago has been essentially observed in West Africa. MZSL shares many features with HMS. In one study, B cell receptor clonality was

assessed in Ghanaian patients with HMS separated according to their response to antimalarials: clonality was absent in responders, constant in non-responders and observed in 2 out of 13 partial responders [77]. These findings, along with others, led to the hypothesis that splenic lymphoma represents an ultimate evolution of untreated HMS [78], but this risk has not been precisely quantified.

Antimalarial treatment of HMS relied on prolonged courses of primaquine, proguanil, mefloquine or chloroquine. In travelers not returning to malaria-endemic areas, a short regimen is as effective as prolonged treatments [76]. The value of such short courses remains unknown in endemic areas and the usual policy is to administer an intermittent prophylaxis to HMS patients persistently exposed to the parasite [79]. Splenectomy may prove dangerous in HMS patients, as it carries a risk of post-operative acute malaria [64,80,81] and exposes to the long-term risks associated with splenectomy [82].

The Fulani enigma

Fulani subjects, present across Sahelian Africa (from Southern Sudan to Eastern Senegal) used to be nomad pastoralists. In Mali and Burkina Faso, compared to other sympatric subjects, Fulani exposed to *Plasmodium* display a distinguishable phenotype, with frequent splenomegaly, enhanced anti-plasmodial humoral response, lower body temperature, lower hemoglobin and lower parasitic load. For these reasons, Fulani have been considered as protected from malaria. This phenotype is however neither complete nor constant (Supplemental Table S2), the most constant feature being palpable splenomegaly. The determinants of the "Fulani phenotype" have been mostly studied from an immunological standpoint, showing, in Fulani, an activated state of monocytes [83], enhanced proinflammatory cytokines production [84], strong response of dendritic cells to toll-like

receptors agonists [85], and a defect in regulatory T cells [86]. The genetic basis of this phenotype remains unclear, as the conventional RBC polymorphisms associated with malaria protection have not been found to be more frequent in Fulani [87]. The Fulani phenotype is in many aspects reminiscent of HMS, and HMS is over-prevalent in Fulani [68]. Comprehensive, multidisciplinary field studies are underway to decipher the determinants of this phenotype and deconvolute the relative contributions of RBC-spleen interactions and genetics [88].

Role of the spleen in the transmission of *P. falciparum*

Gametocytes, initial sexual forms of *Plasmodium* in RBC, are generated in a small proportion (0.1 to 5%) at each asexual replication stage. They progressively mature from stage I to IV (immature gametocytes) to stage V (mature forms) over 14 days, under the influence of external factors (reviewed in [89]) and the transcription factor PfAP2-G. The deformability of gametocytes is markedly reduced from stages I to IV, then drastically improves [90]. Immature stages accumulate in the spleen and in extra vascular spaces of the bone marrow [91,92] and are generally absent from the peripheral circulation, unlike mature stages. Circulation of mature gametocytes is a prerequisite for the parasite to be ingested by *Anopheles* and to maintain the parasite transmission cycle. The deformability of immature gametocytes is regulated in part by the parasite protein STEVOR, kinase A and AMPc [93]. A pharmacological approach aiming at artificially stiffening mature gametocytes has been developed, with the aim to induce their mechanical retention in the spleen, which would subsequently prevent their circulation and make them unavailable to *Anopheles*, hence blocking transmission. This approach has benefited from the technological improvements of

microsphiltration (Box 1). A large, high-throughput screening campaign to discover stiffening compounds is ongoing [8].

363

364

365

366

367

368

369

370

371

372

373

374

375

376

377

378

379

380

361

362

Concluding remarks

The spleen has been extensively investigated, in animal models and later in humans, from an immunological perspective. The exploration of how the spleen innately filters normal and altered RBC and eventually eliminates them (predominantly in the red pulp) has received attention from researchers half a century ago. New tools have enabled progress in this field over the last decade. A better understanding of human splenic physiology has shed light on the dual role "Dr. Jekyll and Mr. Hyde-like" of the spleen in human malaria (Table 1, Key Table). The spleen limits the increase in parasite biomass, therefore reducing sequestration and microvascular dysfunction in major target organs like the brain, but concomitantly likely contributes to malarial anemia. Similarly, the spleen-related production of once-infected RBC after treatment with artemisinins reduces RBC loss during therapy but sometimes induces delayed and clinically significant hemolysis. Current and further lines of research (see Outstanding Questions) will investigate the promising field of induction of splenic retention of Plasmodium gametocytes, a potentially important contributor to malaria elimination attempts. Not least, new exploratory tools (Box 1) will be of importance to investigate the role of cellular immunity antimalarial defense, a process essentially studied through in vitro studies and animal models so far.

381

382

Acknowledgements

The authors wish to thank Dr Geneviève Milon for her longstanding support and pertinent scientific guidance; Dr Steven Kho & prof. Nicholas Anstey for their comments on this manuscript and fruitful collaboration.

386

387

References

- Cranston, H.A. *et al.* (1984) Plasmodium falciparum maturation abolishes physiologic red cell deformability. *Science* 223, 400–403
- Dondorp, A.M. *et al.* (1997) Prognostic significance of reduced red blood cell deformability in severe falciparum malaria. *Am. J. Trop. Med. Hyg.* 57, 507–511
- 392 3 Miller, L.H. *et al.* (1971) Alteration in the rheologic properties of Plasmodium knowlesi--infected 393 red cells. A possible mechanism for capillary obstruction. *J. Clin. Invest.* 50, 1451–1455
- Buffet, P.A. *et al.* (2011) The pathogenesis of Plasmodium falciparum malaria in humans: insights from splenic physiology. *Blood* 117, 381–392
- 396 5 del Portillo, H. et al. (2012) The role of the spleen in malaria. Cell. Microbiol. 14, 343–355
- Buffet, P.A. *et al.* (2006) Ex vivo perfusion of human spleens maintains clearing and processing functions. *Blood* 107, 3745–3752
- Deplaine, G. *et al.* (2011) The sensing of poorly deformable red blood cells by the human spleen can be mimicked in vitro. *Blood* 117, e88-95
- 401 8 Duez, J. *et al.* (2018) High-throughput microsphiltration to assess red blood cell deformability 402 and screen for malaria transmission-blocking drugs. *Nat Protoc* 13, 1362–1376
- 403 9 Rigat-Brugarolas, L. *et al.* (2014) A functional microengineered model of the human splenon-on-a-chip. *Lab Chip* 14, 1715–24
- 405 10 Kho, S. *et al.* (2018) Platelets kill circulating parasites of all major Plasmodium species in human 406 malaria. *Blood* 132, 1332-1344
- 407 11 Steiniger, B.S. (2015) Human spleen microanatomy: why mice do not suffice. *Immunology* 145, 408 334–346
- 409 12 White, N. *et al.* (2010) The murine cerebral malaria phenomenon. *Trends in Parasitology* 26, 410 11–15
- 411 13 Bowdler, A.J. (2010) *The Complete Spleen: Structure, Function, and Clinical Disorders.*, Second 412 Edition. Humana Press.
- Duffier, Y. *et al.* (2016) A humanized mouse model for sequestration of Plasmodium falciparum sexual stages and in vivo evaluation of gametocytidal drugs. *Sci Rep* 6, 35025
- Looareesuwan, S. *et al.* (1987) Dynamic alteration in splenic function during acute falciparum malaria. *N. Engl. J. Med.* 317, 675–679
- Safeukui, I. *et al.* (2012) Quantitative assessment of sensing and sequestration of spherocytic erythrocytes by the human spleen. *Blood* 120, 424–430
- 419 Li, H. *et al.* (2018) Mechanics of diseased red blood cells in human spleen and consequences for hereditary blood disorders. *Proc. Natl. Acad. Sci. U.S.A.* 115, 9574–9579
- 421 18 Roussel, C. *et al.* (2019), Clairance post-transfusionnelle des globules rouges stockés: analyse 422 dans un modèle de rate humaine isolée-perfusée., in *Transfusion Clinique et Biologique*, pp. 423 S91–S106; https://doi.org/10.1016/j.tracli.2019.06.221
- Safeukui, I. *et al.* (2008) Retention of Plasmodium falciparum ring-infected erythrocytes in the slow, open microcirculation of the human spleen. *Blood* 112, 2520–2528

- Safeukui, I. *et al.* (2013) Surface area loss and increased sphericity account for the splenic
 entrapment of subpopulations of Plasmodium falciparum ring-infected erythrocytes. *PLoS One* 8, e60150
- Bach, O. *et al.* (2005) Falciparum malaria after splenectomy: a prospective controlled study of 33 previously splenectomized Malawian adults. *Trans. R. Soc. Trop. Med. Hyg.* 99, 861–867
- 431 22 Kho, S. *et al.* (2019) High risk of Plasmodium vivax malaria following splenectomy in Papua, 432 Indonesia. *Clin. Infect. Dis.* 68, 51–60
- Chotivanich, K. et al. (2002) Central Role of the Spleen in Malaria Parasite Clearance. J. Infect.
 Dis. 185, 1538–1541
- 435 24 Luethy, P.M. *et al.* (2018) Diagnostic challenges of prolonged post-treatment clearance of Plasmodium nucleic acids in a pre-transplant autosplenectomized patient with sickle cell disease. *Malar. J.* 17, 23
- 438 25 Boo, Y.L. *et al.* (2016) A case of severe Plasmodium knowlesi in a splenectomized patient.
 439 *Parasitol. Int.* 65, 55–57
- 440 26 Barber, B.E. *et al.* (2013) A Prospective Comparative Study of Knowlesi, Falciparum, and Vivax
 441 Malaria in Sabah, Malaysia: High Proportion With Severe Disease From Plasmodium Knowlesi
 442 and Plasmodium Vivax But No Mortality With Early Referral and Artesunate Therapy. *Clin Infect*443 *Dis* 56, 383–397
- Toovey, S. and Jamieson, A. (2003) Pretravel health advice for asplenic individuals. *J Travel Med* 10, 362; author reply 362-363
- 446 28 Herricks, T. *et al.* (2009) Deformability limits of Plasmodium falciparum-infected red blood cells.
 447 *Cell. Microbiol.* 11, 1340–1353
- Diez-Silva, M. *et al.* (2012) Pf155/RESA protein influences the dynamic microcirculatory behavior of ring-stage Plasmodium falciparum infected red blood cells. *Sci Rep* 2, 614
- Sisquella, X. *et al.* (2017) Plasmodium falciparum ligand binding to erythrocytes induce alterations in deformability essential for invasion. *Elife* 6:e21083
- 452 31 Bessis, M. *et al.* (1980) Automated ektacytometry: a new method of measuring red cell 453 deformability and red cell indices. *Blood Cells* 6, 315–327
- Dondorp, A.M. *et al.* (1999) Red blood cell deformability as a predictor of anemia in severe falciparum malaria. *Am. J. Trop. Med. Hyg.* 60, 733–737
- Looareesuwan, S. *et al.* (1987) Reduced erythrocyte survival following clearance of malarial parasitaemia in Thai patients. *Br. J. Haematol.* 67, 473–478
- 458 34 Prommano, O. *et al.* (2005) A quantitative ultrastructural study of the liver and the spleen in fatal falciparum malaria. *Southeast Asian J. Trop. Med. Public Health* 36, 1359–1370
- 460 35 Cserti-Gazdewich, C.M. *et al.* (2013) Inter-relationships of cardinal features and outcomes of
 461 symptomatic pediatric Plasmodium falciparum MALARIA in 1,933 children in Kampala, Uganda.
 462 *Am. J. Trop. Med. Hyg.* 88, 747–756
- 463 36 Giha, H.A. *et al.* (2009) Severe malaria in an unstable setting: clinical and laboratory correlates
 464 of cerebral malaria and severe malarial anemia and a paradigm for a simplified severity scoring.
 465 *Eur. J. Clin. Microbiol. Infect. Dis.* 28, 661–665
- Kotlyar, S. *et al.* (2014) Spleen volume and clinical disease manifestations of severe Plasmodium falciparum malaria in African children. *Trans. R. Soc. Trop. Med. Hyg.* 108, 283–289
- 468 38 Collins, K.A. *et al.* (2018) A controlled human malaria infection model enabling evaluation of transmission-blocking interventions. *J. Clin. Invest.* 128, 1551–1562
- 470 39 Douglas, N.M. et al. (2012) The anaemia of Plasmodium vivax malaria. Malar. J. 11, 135
- 471 40 Harnagel, E.E. and Rhudy, F.V. (1954) Falciparum malaria; report of a fatal case and autopsy findings. *Calif Med* 81, 89–91
- 473 41 Muehlethaler, K. et al. (2005) Fulminant cerebral malaria in a Swiss patient. *Infection* 33, 33–35
- Waitumbi, J.N. *et al.* (2000) Red cell surface changes and erythrophagocytosis in children with severe plasmodium falciparum anemia. *Blood* 95, 1481–1486
- 476 43 Fendel, R. *et al.* (2010) Hemolysis is associated with low reticulocyte production index and 477 predicts blood transfusion in severe malarial anemia. *PLoS One* 5, e10038

- 478 44 Imbert, P. *et al.* (2009) Pathological rupture of the spleen in malaria: analysis of 55 cases (1958-479 2008). *Travel Med Infect Dis* 7, 147–159
- 480 45 Buffet, P.A. *et al.* (2009) Retention of erythrocytes in the spleen: a double-edged process in human malaria. *Curr. Opin. Hematol.* 16, 157–164
- 482 46 Angus, B.J. *et al.* (1997) In vivo removal of malaria parasites from red blood cells without their destruction in acute falciparum malaria. *Blood* 90, 2037–2040
- 484 47 Newton, P.N. *et al.* (2001) A comparison of the in vivo kinetics of Plasmodium falciparum ring-485 infected erythrocyte surface antigen-positive and -negative erythrocytes. *Blood* 98, 450–457
- 486 48 Schnitzer, B. *et al.* (1972) Pitting function of the spleen in malaria: ultrastructural observations.
 487 *Science* 177, 175–177
- 488 49 White, N. (2017) Malaria parasite clearance. *Malar. J.* 16, 88
- Ndour, P.A. *et al.* (2015) Plasmodium falciparum clearance is rapid and pitting independent in immune Malian children treated with artesunate for malaria. *J. Infect. Dis.* 211, 290–297
- Wojnarski, M. *et al.* (2019) Plasmodium falciparum Clearance Is Pitting-Dependent With
 Artemisinin-Based Drugs but Pitting-Independent With Atovaquone-Proguanil or Mefloquine. *J. Infect. Dis.* 220, 535-539
- 494 52 Itoda, I. *et al.* (2002) Severe falciparum malaria with prolonged hemolytic anemia after successful treatment with intravenous artesunate. *Kansenshogaku Zasshi.* 76, 600–3
- Zoller, T. *et al.* (2011) Intravenous artesunate for severe malaria in travelers, Europe. *Emerg. Infect. Dis.* 17, 771–777
- Jauréguiberry, S. *et al.* (2015) Delayed-Onset Hemolytic Anemia in Patients with Travel Associated Severe Malaria Treated with Artesunate, France, 2011–2013. *Emerg. Infect. Dis.* 21,
 804–812
- 501 55 Kurth, F. *et al.* (2017) Severe malaria in Europe: an 8-year multi-centre observational study.
 502 *Malar J* 16, 57
- 503 56 Roussel, C. *et al.* (2017) Artesunate to treat severe malaria in travellers: review of efficacy and safety and practical implications. *J Travel Med* 24
- Jauréguiberry, S. *et al.* (2014) Postartesunate delayed hemolysis is a predictable event related to the lifesaving effect of artemisinins. *Blood* 124, 167–175
- 507 58 Ndour, P.A. *et al.* (2017) Measuring the Plasmodium falciparum HRP2 protein in blood from 508 artesunate-treated malaria patients predicts post-artesunate delayed hemolysis. *Sci Transl Med* 509 9
- 510 59 Fanello, C. *et al.* (2017) Post-treatment haemolysis in African children with hyperparasitaemic 511 falciparum malaria; a randomized comparison of artesunate and quinine. *BMC Infect Dis* 17, 512 575
- Rolling, T. *et al.* (2014) Delayed hemolysis after treatment with parenteral artesunate in African children with severe malaria--a double-center prospective study. *J. Infect. Dis.* 209, 1921–1928
- 515 61 White, N.J. (2018) Anaemia and malaria. Malaria Journal 17, 371
- 516 62 Crookston, B.T. *et al.* (2010) Exploring the relationship between chronic undernutrition and asymptomatic malaria in Ghanaian children. *Malar. J.* 9, 39
- Kho, S. *et al.* (2019), A hidden biomass of non-phagocytosed asexual and sexual stages of
 Plasmodium vivax and P. falciparum in the human spleen., presented at the 7th International
 Conference on Plasmodium vivax Research, Institut Pasteur, Paris, France
 (www.vivax2019.conferences-pasteur.org/programme)
- 522 64 Bachmann, A. *et al.* (2009) Absence of erythrocyte sequestration and lack of multicopy gene 523 family expression in Plasmodium falciparum from a splenectomized malaria patient. *PLoS One* 524 4, e7459
- 525 65 Bryceson, A. et al. (1983) Malaria and splenomegaly. Trans. R. Soc. Trop. Med. Hyg. 77, 879
- 526 66 Bates, I. and Bedu-Addo, G. (1997) Review of diagnostic criteria of hyper-reactive malarial 527 splenomegaly. *Lancet* 349, 1178
- 528 67 Bedu-Addo, G. and Bates, I. (2002) Causes of massive tropical splenomegaly in Ghana. *Lancet* 360, 449–454

- 530 68 Greenwood, B.M. *et al.* (1987) Ethnic differences in the prevalence of splenomegaly and malaria in The Gambia. *Ann Trop Med Parasitol* 81, 345–354
- Hommel, B. *et al.* (2014) Chronic malaria revealed by a new fluorescence pattern on the antinuclear autoantibodies test. *PLoS One* 9, e88548
- Martin-Peprah, R. *et al.* (2006) Investigation of familial segregation of hyperreactive malarial splenomegaly in Kumasi, Ghana. *Trans. R. Soc. Trop. Med. Hyg.* 100, 68–73
- 536 71 Ackerman, L. (1996) Hyperreactive malarial syndrome. J Am Board Fam Pract 9, 356–359
- 537 72 Chim, C.S. *et al.* (2004) Concurrent hyperreactive malarial splenomegaly and quartan malarial nephropathy Plasmodium malariae revisited. *Haematologica* 89, ECR21
- 539 73 Pryor, D.S. (1967) Splenectomy in tropical splenomegaly. *Br Med J* 3, 825–828
- 74 Crane, G.G. *et al.* (1972) Tropical splenomegaly syndrome in New Guinea. I. Natural history.
 74 Trans. R. Soc. Trop. Med. Hyg. 66, 724–732
- Puente, S. *et al.* (2001) Hyperreactive malarial splenomegaly in Europeans: report of five cases.
 J Travel Med 8, 322–324
- Van den Ende, J. *et al.* (2000) Hyperreactive malaria in expatriates returning from sub-Saharan Africa. *Trop. Med. Int. Health* 5, 607–611
- 546 77 Bates, I. *et al.* (1991) Use of immunoglobulin gene rearrangements to show clonal lymphoproliferation in hyper-reactive malarial splenomegaly. *Lancet* 337, 505–507
- 548 78 Bates, I. and Bedu-Addo, G. (1997) Chronic malaria and splenic lymphoma: clues to understanding lymphoma evolution. *Leukemia* 11, 2162–2167
- 550 79 Bisoffi, Z. *et al.* (2016) Chronic malaria and hyper-reactive malarial splenomegaly: a retrospective study on the largest series observed in a non-endemic country. *Malar J* 15, 230
- Bidegain, F. *et al.* (2005) Acute Plasmodium falciparum malaria following splenectomy for suspected lymphoma in 2 patients. *Clin. Infect. Dis.* 40, e97-100
- 554 81 Camprubi, D. *et al.* (2019) Post-splenectomy acute glomerulonephritis due to a chronic 555 infection with Plasmodium falciparum and malariae. *Journal of Travel Medicine* 26, 1–2
- Kristinsson, S.Y. *et al.* (2014) Long-term risks after splenectomy among 8,149 cancer-free
 American veterans: a cohort study with up to 27 years follow-up. *Haematologica* 99, 392–398
- Quin, J.E. *et al.* (2017) Major transcriptional changes observed in the Fulani, an ethnic group less susceptible to malaria. *Elife* 6, e29156
- 560 84 McCall, M.B.B. *et al.* (2010) Early interferon-gamma response against Plasmodium falciparum 561 correlates with interethnic differences in susceptibility to parasitemia between sympatric Fulani 562 and Dogon in Mali. *J. Infect. Dis.* 201, 142–152
- Arama, C. *et al.* (2011) Interethnic differences in antigen-presenting cell activation and TLR responses in Malian children during Plasmodium falciparum malaria. *PLoS One* 6, e18319
- Torcia, M.G. *et al.* (2008) Functional deficit of T regulatory cells in Fulani, an ethnic group with low susceptibility to Plasmodium falciparum malaria. *Proc. Natl. Acad. Sci. U.S.A.* 105, 646–651
- Modiano, D. *et al.* (2001) The lower susceptibility to Plasmodium falciparum malaria of Fulani of Burkina Faso (west Africa) is associated with low frequencies of classic malaria-resistance genes. *Trans. R. Soc. Trop. Med. Hyg.* 95, 149–152
- Henry, B. *et al.* (2017), Exploration of interethnic susceptibility to malaria in Benin. Abstract 17-LB-4411-ASTMH., presented at the 66th meeting of the American Society of Tropical Medicine and Hygiene, Baltimore, MD (https://doi.org/10.4269/ajtmh.abstract2017)
- 573 89 Meibalan, E. and Marti, M. (2017) Biology of Malaria Transmission. *Cold Spring Harb Perspect* 574 *Med* 7, a025452
- 575 90 Tibúrcio, M. *et al.* (2012) A switch in infected erythrocyte deformability at the maturation and blood circulation of Plasmodium falciparum transmission stages. *Blood* 119, e172–e180
- 577 91 Farfour, E. *et al.* (2012) The extravascular compartment of the bone marrow: a niche for Plasmodium falciparum gametocyte maturation? *Malar. J.* 11, 285
- 579 92 Joice, R. *et al.* (2014) Plasmodium falciparum transmission stages accumulate in the human 580 bone marrow. *Sci Transl Med* 6, 244re5

581	93	dani, G. et al. (2015) cAMP-Signalling Regulates Gametocyte-Infected Erythrocyte					
582	0.4	Deformability Required for Malaria Parasite Transmission. <i>PLoS Pathog</i> 11, e1004815					
583 584	94	Omar, A. and Freeman, S. (2016) Contrast-enhanced ultrasound of the spleen. <i>Ultrasound</i> 24, 41–49					
585	95	de Porto, A. et al. (2010) Assessment of splenic function. Eur. J. Clin. Microbiol. Infect. Dis. 29,					
586	33	1465–73					
587	96	Di Sabatino, A. et al. (2011) Post-splenectomy and hyposplenic states. The Lancet 378, 86–97					
588	97	Cameron, P.U. et al. (2011) Splenectomy Associated Changes in IgM Memory B Cells in an Adult					
589		Spleen Registry Cohort. PLOS One 6, e23164					
590	98	Lammers, A. et al. (2012) Hyposplenism: Comparison of different methods for determining					
591 592	99	splenic function. <i>Am. J. Hematol.</i> 87, 484–489 Duez, J. <i>et al.</i> (2015) Splenic retention of Plasmodium falciparum gametocytes to block the					
593	33	transmission of malaria. <i>Antimicrob. Agents Chemother</i> . 59, 4206–4214					
594	100	Picot, J. <i>et al.</i> (2015) A biomimetic microfluidic chip to study the circulation and mechanical					
595		retention of red blood cells in the spleen. Am. J. Hematol. 90, 339–345					
596							
597							
598	Glossary						
599	Cor	ds : Circulatory structures specific of the splenic red pulp devoid of endothelial lining. In					
600	the	the cords, circulation of red blood cells is slow, favoring close and prolonged interactions					
601	With	7th splenic macrophages which are very abundant in this unique environment. CD34+-					
602	posi	positive conventional arterioles traverse the red pulp but are not considered a component of					
603	the s	the slow and open pathway of the red pulp.					
604	Ekt	acytometry: Method to measure RBC deformability. Gradually increasing shear stress is					
605	applied to a solution of RBC diluted in a viscous, iso-osmolar medium. Measurement of RBC						
606	diffraction pattern enables the calculation of the elongation index, a correlate for RBC						
607	defo	rmability.					
608	Erythrophagocytosis: Phagocytosis of generally altered RBC by effector cells, a process						
609	takiı	ing place predominantly in the splenic red pulp and operated by red pulp macrophages.					
610	Hyp	per-reactive malarial splenomegaly (HMS): Rare form of chronic malaria, occurring in					
611	persistently exposed subjects, defined by a gross splenomegaly, elevated total IgM, high titers						
612	of anti-Plasmodium antibody with a polyclonal immune response pattern, and clinical						
613	response to antimalarials.						
614	Inte	Inter-endothelial slits (IES): Narrow spaces (0.2 to 2 microns) between adjacent elongated					

endothelial cells of sinuses of the splenic red pulp. Venous sinuses do not have a basal

membrane but a highly discontinuous helicoid basal fiber. RBC must cross inter-endothelial 616 617 slits as they navigate from the cords back to the general circulation, which challenges their deformability. Inter-endothelial slits are 2 - 10 times narrower than the smallest capillaries. 618 **Pitting**: Spleen–specific mechanism by which an intraerythrocytic body is extracted from the 619 RBC. The RBC is not lysed by the pitting process and returns to the circulation. Pitting plays 620 621 a major role in parasite clearance following therapy with artemisinins (but not following treatment with other antimalarial agents). 622 Post-artesunate delayed hemolysis: Acute hemolytic anemia, occurring 1 to 2 weeks after 623 624 treatment with artemisinin derivatives for severe, generally hyper-parasitemic malaria. This adverse event is triggered by the relatively synchronous clearance of pitted RBC. 625 626 Rapid diagnostic tests: Immunochromatographic tests for the diagnosis of *Plasmodium* parasitemia. These tests are performed on capillary blood collected by finger prick and detect 627 628 the presence of plasmodial antigens, either pan-specific (such as lactico-deshydrogenase), or species-specific (such as the histidine-rich protein-2). 629 Red pulp: Part of splenic parenchyma involved in the control of RBC biomechanical and 630 surface properties, erythrophagocytosis and reaction to circulating antigens. It includes cords 631 632 and venous sinuses. 633 Venous sinuses: Circulatory structures collecting blood downstream from the splenic cords and from the closed and fast perifollicular circulation of the spleen. The endothelial lining of 634 635 venous sinuses is made of parallel, elongated cells lying on discontinuous basal fibers. White pulp: Part of splenic parenchyma involved in immune responses, especially against 636

639

640

641

642

643

637

638

Box 1. Available tools to assess the human spleen

and lymphoid nodules.

Spleen palpation is the first and the only available assessment in most malaria-endemic settings. Spleen size is best quantified using Hackett's classification. The prevalence of splenomegaly correlates positively with malaria endemicity but lacks specificity.

circulating antigens and encapsulated bacteria. It comprises periarteriolar lymphoid sheaths

- Splenomegaly is frequent in children with malarial anemia in endemic areas, but rare in
- travelers with acute attacks.
- 646 Medical imaging assesses the size and structure of the spleen, and visualizes splenic
- abnormalities. Ultrasound coupled with Doppler is the most widely available technique.
- 648 Splenic contrast-enhanced ultrasonography using microbubbles [94] fine-tunes splenic
- evaluation and enables the relative quantification of flow in the fast and slow circulations.
- 650 Scintigraphy using radiolabeled tracers provides a functional assessment: sulphur colloids and
- sensitized RBC assess macrophage function while heat-stiffened RBC assess the mechanical
- 652 filtering function [95]. Splenic function can be inferred from the clearance kinetics of labelled
- RBC, and from the intensity of radioactivity in the splenic area compared to the hepatic or
- 654 cardiac area.
- 655 RBC-related markers of splenic function. When the filtering and pitting functions are
- 656 impaired, RBC containing Howell-Jolly bodies (small Giemsa-positive spheres) appear in
- circulation and can be observed on peripheral blood smears). Pocked RBC contain small
- 658 intracytoplasmic vesicles, visible using differential interference contrast microscopy. Their
- proportion in the peripheral blood increases above 2-3% in cases of chronic splenic
- dysfunction. This accurate marker [96] requires a specific equipment and trained personnel.
- 661 Circulating populations of immune cells, especially IgM memory B cells, have been linked to
- splenic function. This subpopulation is significantly reduced in splenectomized subjects
- 663 [97,98].
- 664 Ex vivo perfusion of human spleens retrieved following left pancreatosplenectomy preserves
- the filtering and phagocytic functions during a few hours [6]. This approach has uncovered
- the innate, mechanical retention of a proportion of ring-infected RBC [19], a process later
- 667 confirmed through filtration in vitro [7,20] and which may contribute to the intense
- accumulation of infected RBC in the spleen of chronic carriers.
- Some *Biomimetic tools* attempt to reproduce splenic function; microsphiltration evaluates the
- ability of RBC to squeeze through narrow slits between metallic beads, and accurately reflects
- 671 mechanical retention of RBC in the human spleen [7,8,99]. The initial experimental set-up
- [7] has been adapted to 96 or 384-wells microplates, enabling the parallel analysis of
- 673 hundreds of samples [8]. Spleen-mimetic microfluidics chips have been recently developed
- [9,100]. A stage-dependent retention in when chips were infused with infected, labeled RBC.

Table 1, Key Table. Dual role of the spleen in human malaria.

	Acute attacks	In treated patients	Chronic carriage	Transmission
Protective role	Retention of infected RBC	Clearance of infected RBC (pitting)	Limitation of the circulating parasitic load?	Retention of gametocytes
				Reservoir?
			- Contribution to	(contribution to
			chronic anemia?	chronic
Detrimental	- Rupture - Contribution to anemia	Post Artesunate Delayed Hemolysis	- Hyper-Reactive	asymptomatic
role			Malarial	carriage)
			Splenomegaly	
			-Splenic lymphoma?	

Figure 1. Interactions between parasitized and unparasitized RBC, macrophages, endothelial cells in the splenic red pulp, across different physiologic and pathologic conditions. (A): schematic view of the splenic microanatomy. PALS: periarteriolar lymphoid sheath. (B): Representation of the endothelial lining between splenic cords and venous sinus lumens. Normal RBC must squeeze through inter-endothelial slits (IES) to join venous circulation. Co: cords of Billroth. SL: sinus lumen. (C): during acute attacks of malaria, splenomegaly can be observed, and very rarely splenic rupture can occur. RBC infected with mature forms are retained in the spleen and processed by RPM. A proportion of ring-infected RBC crosses IES. Some uninfected RBC become also stiffer and might also be retained. (D): in the case of malarial infection in a splenectomized host, parasitemia is higher and mature forms, mostly sequestered in peripheral tissues and sometimes retained in the spleen, appear in the peripheral circulation. (E): during chronic parasite carriage, splenomegaly can inconstantly be observed and in rare cases, evolves towards hyper-reactive malarial splenomegaly. Infected RBC might be retained in a similar way as they are in acute attacks, as well as part of uninfected RBC. An intense intrasplenic retention of infected RBC might lead to a fully intrasplenic replication cycle of the parasite. (F): during acute malaria attacks treated with artemisinin derivatives, the pitting process occurs in the spleen: dead parasite remnants are expelled from the RBC without lysing it. "Once-infected" RBC harbor parasite proteins (green dotted line) at the inner side of their membrane, allowing them to be identified in the peripheral blood. In some cases, 1 to 2 weeks after acute attacks, synchronous hemolysis of once-infected RBC is responsible for post-artesunate delayed hemolysis (PADH). (G): in subjects carrying gametocytes, retention of immature forms (stages I to IV) occurs in the spleen due to their low deformability. After evolution towards mature stages, RBC infected with gametocytes are able to cross IES and therefore circulate in peripheral blood.

678

679

680

681

682

683

684

685

686

687

688

689

690

691

692

693

694

695

696

697

698

699

700

701

702

