



**HAL**  
open science

## **Freshwater ecoacoustics: Listening to the ecological status of multi-stressed lowland waters**

Gea H. van Der Lee, Camille Desjonquères, Jérôme Sueur, Michiel H.S. Kraak, Piet F.M. Verdonschot

### ► **To cite this version:**

Gea H. van Der Lee, Camille Desjonquères, Jérôme Sueur, Michiel H.S. Kraak, Piet F.M. Verdonschot. Freshwater ecoacoustics: Listening to the ecological status of multi-stressed lowland waters. *Ecological Indicators*, 2020, 113, pp.106252 -. <10.1016/j.ecolind.2020.106252>. <hal-03489973>

**HAL Id: hal-03489973**

**<https://hal.science/hal-03489973v1>**

Submitted on 20 May 2022

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Distributed under a Creative Commons CC BY-NC 4.0 - Attribution - Non-commercial use - International License

## Freshwater ecoacoustics: listening to the ecological status of multi-stressed lowland waters

5 Gea H. van der Lee<sup>1,\*</sup>, Camille Desjonquères<sup>2,3,\*</sup>, Jérôme Sueur<sup>3</sup>, Michiel H. S. Kraak<sup>1</sup>, Piet F. M. Verdonshot<sup>1,4</sup>

<sup>1</sup> Institute for Biodiversity and Ecosystem Dynamics (IBED-FAME), University of Amsterdam, P.O. Box 94240, 1090 GE Amsterdam, The Netherlands

10 <sup>2</sup> Terrestrial Ecology Group (TEG), Departamento de Ecología, Universidad Autónoma de Madrid, C-211, Madrid, Spain.

<sup>3</sup> Institut Systématique Evolution Biodiversité (ISYEB), Muséum National d'Histoire Naturelle, CNRS, Sorbonne Université, EPHE, 57 Rue Cuvier, CP 50, 75005, Paris, France

15 <sup>4</sup> Wageningen Environmental Research, Wageningen UR, P.O. Box 47, 6700 AA Wageningen, The Netherlands

\* These two authors contributed equally to this manuscript.

Corresponding author:

Camille Desjonquères, [cdesjonqu@gmail.com](mailto:cdesjonqu@gmail.com)

20 Departamento de Ecología, Room C211, Universidad Autónoma de Madrid, Ciudad Universitaria de Cantoblanco, 28049 Madrid

## **Abstract**

A major challenge in water quality assessment is to identify suitable indicators to monitor and assess the effects of anthropogenic stressors on the ecological status of freshwater ecosystems.

Passive acoustic monitoring is a novel approach that could potentially be used to detect invertebrate  
25 species and ecological processes such as dissolved oxygen dynamics in freshwater environments.  
The aim of the present study was to evaluate to what extent sounds can be used for water quality  
assessment. We performed a field study to relate acoustic indices to the intensity of several  
stressors, the invertebrate community composition and the dissolved oxygen dynamics in 20  
temperate lowland streams and drainage ditches impacted to a varying degree by agricultural  
30 activities and discharges from waste water treatment plants. Our results showed that the recorded  
acoustic patterns were primarily associated with the fluctuation in dissolved oxygen saturation,  
while specific frequency bands could be related to the sound-producing invertebrate community.  
We observed that acoustic indices do not allow to detect the adverse effects of anthropogenic  
stressors on the invertebrate community composition, presumably due to the prevalence of  
35 Heteroptera which are relatively insensitive to stressors, but make a lot of sounds. A strong relation  
between acoustic indices and oxygen fluctuation indicate that passive acoustic monitoring may be  
used to estimate metabolism in water bodies. We suggest that the next step in freshwater  
ecoacoustics will be to precisely characterise each source of sound emitted during the processes of  
primary production, respiration and re-aeration, in order to distinguish these parameters. This may  
40 overcome some of the challenges encountered in the estimation of metabolism from diel dissolved  
oxygen curves.

*Keywords: Water quality assessment, Invertebrates, Dissolved Oxygen Dynamics, Passive Acoustic  
Monitoring*

## 45 **Introduction**

Freshwater ecosystems are commonly impacted by various anthropogenic activities, including agricultural activities and municipal wastewater treatment plant (WWTP) discharges (Burdon et al., 2019; Ormerod et al., 2010). These activities can result in a combination of stressors, such as excess nutrients, increased water temperatures, pesticides, pharmaceuticals, personal care products and a  
50 suite of other contaminants, which can cause substantial changes in the community composition and ecological processes of the impacted water bodies (Allan, 2004; Karr, 1999; Palmer and Febria, 2012). A major challenge in protecting and restoring these ecosystems is to identify cost-effective indicators to monitor, assess and evaluate the effects of these stressors on the ecological status of freshwater ecosystems (Bonada et al., 2006; Friberg et al., 2011).

55 The majority of ecological indicators used in water quality assessments are based on point-in-time measurements of the community structure of various organism groups (Boulton, 1999; Resh, 2008). Communities, especially of macroinvertebrates, are relatively diverse and can therefore be used to assess a variety of anthropogenic stressors (e.g. Clapcott et al., 2012). These structural-based indicators, however, do not capture the dynamic properties of freshwater  
60 ecosystems (Palmer and Febria, 2012). It has been argued that repeated measurements of ecological processes could be helpful to capture system dynamics employing functional-based indicators (Bunn, 1995). To this purpose, diel change in dissolved oxygen (DO) concentrations have been measured with loggers in the open channel to calculate ecosystem metabolism, where DO  
65 concentrations are associated with photosynthesis during the day and respiration at night (Odum, 1956; Young et al., 2008). Yet, these structural and functional based indicators require different sampling techniques. Thus the development of a sampling method that can monitor and assess structural and functional indicators with a single technique could improve the efficiency of water quality assessments.

A potential approach to combine structural and functional water quality assessment is to use

70 passive acoustic monitoring, a method proposed by ecoacoustics that samples ambient sounds to  
tackle ecological questions (Farina and Gage, 2017; Sueur and Farina, 2015). This non-invasive and  
continuous monitoring method is emerging to address ecological questions in many ecosystems  
(Desjonquères et al., 2020b; Gibb et al., 2019; Linke et al., 2018; Sugai et al., 2019a, 2019b). Some  
terrestrial studies have already managed to link acoustic patterns to the degrees of human influence  
75 (Burivalova et al., 2018; Buxton et al., 2018a), however, only a limited number of studies have  
applied ecoacoustic monitoring in freshwater environments (but see Desjonquères et al., 2015;  
Karaconstantis et al., 2020; Linke et al., 2020).

Environmental sounds can emanate not only from sound producing species, whose presence  
and activity depends on local ecological conditions, but also from ecological processes and  
80 anthropogenic activities (Linke et al., 2018). In freshwater environments, there are four main taxa  
known for producing sounds: amphibians, crustaceans, fish and insects (Desjonquères et al., 2020a).  
In temperate zones, most of the sounds recorded in freshwater environments appear to be linked to  
insects belonging to the insect orders Heteroptera, Coleoptera, Trichoptera and Odonata,  
Heteroptera and more specifically the family Corixidae include the majority of **soniferous** species  
85 (Aiken, 1985; Desjonquères, 2016; Desjonquères et al., 2018). Sounds linked to ecological  
processes are likely due to a combination of primary production through plant photosynthesis as  
well as the decomposition of organic matter by microbial activity (Linke et al., 2018). Although all  
these species and processes can potentially be detected and monitored in freshwater environments,  
the knowledge on species- and process-specific sounds is still emerging, making it unclear to what  
90 extent sounds can be used as a monitoring tool in water quality assessment strategies.

One of the main challenges of acoustic monitoring is to extract ecologically meaningful  
attributes of sounds and to link these sounds to ecosystem variables, such as community  
composition and ecological processes (Linke et al., 2018; Sueur and Farina, 2015). To address this  
challenge, acoustic indices, which are the equivalent of ecological diversity indices based on sound

95 rather than on the number of species sampled, have been developed (Sueur et al., 2014). Such  
acoustic indices compute specific features of the sound spectrum (frequency representation) or  
waveform (temporal representation), which are thought to represent meaningful information about  
the ecosystem (Gage et al., 2017).

The aim of the present study was, therefore, to evaluate the potential of passive acoustic  
100 monitoring in freshwater environments by relating acoustic patterns to ecosystem structure and  
function in water bodies along a gradient of anthropogenic stress. We hypothesized that different  
acoustic indices relate to different structural and functional aspects of freshwater ecosystems. To  
test this hypothesis, we conducted a field study in 20 temperate lowland streams and drainage  
ditches throughout the Netherlands. First, we tested to what extent the invertebrate community  
105 reflected the intensity of the measured stressors including dissolved oxygen dynamics to confirm  
their use in water quality assessment. Then, we assessed if the sound-producing invertebrate  
community also reflected the same relationship to these stressors and dissolved oxygen dynamics.  
Finally, to evaluate the potential of passive acoustic monitoring in freshwater ecosystems, we  
correlated the acoustic indices to the measured stressors, the invertebrate community composition,  
110 the sound-producing invertebrate community composition and the dissolved oxygen dynamics.

## **Material and methods**

### *Study outline*

The 20 study sites were surrounded by varying degrees of agricultural land use in the riparian zone  
115 and WWTP effluent (Supplementary material 1). The sites had a comparable width (mean  $\pm$  sd =  
 $4.5 \pm 1.8$  m), depth ( $0.8 \pm 0.2$  m) and flow velocity ( $5.3 \pm 6.8$  cm/s). At each site, we measured  
nutrient concentrations, water temperature and proxies for pesticides, pharmaceuticals and personal  
care products to estimate the intensity of the stress originating from the agricultural activities and  
the WWTP discharges. As structural and functional indicators, we determined invertebrate

120 community composition and dissolved oxygen dynamics, respectively. At all study sites, we  
recorded the underwater sounds from which we calculated acoustic indices. The study was  
conducted between August 20<sup>th</sup> and October 23<sup>th</sup> 2018. We chose this period, as the dissolved  
oxygen dynamics are most distinct during this period (Van der Lee et al., 2018) and it is likely to be  
within the reproductive period of several aquatic insect species (Jansson, 1974). All of this is  
125 expected to result in the most distinct sound patterns between sites. Stressors, dissolved oxygen  
dynamics and acoustics data was collected during the first six weeks of the sampling period, while  
invertebrate samples were collected during the last two weeks of the sampling period to avoid  
disturbing the acoustic sampling.

### 130 *Stressors*

Nutrient concentrations were measured by collecting a weekly surface water grab sample at each  
site for six weeks. The samples were filtered over a 1.2 µm filter and analysed for total dissolved  
nitrogen (TDN) and orthophosphate (PO<sub>4</sub>-P) on a continuous flow analyser (SAN++ system, Skalar  
Analytical B.V., Breda, The Netherlands). The mean nutrient concentrations over the six weeks  
135 were calculated for further analysis.

Water temperature (°C) was measured every ten minutes for six weeks with HOBO®  
Temperature/Light loggers UA-002-64 (Onset Computer Corporation, Bourne, MA, USA). At each  
study site, the loggers were placed in the mid-channel, 15 cm under the water surface. The mean  
water temperature over the six weeks was calculated for further analysis.

140 Proxies for pesticides, pharmaceuticals and personal care products were derived from  
bioassays subjected to passive sampler extracts. To this end, the polar organic chemical integrative  
samplers (POCIS) containing 200 mg of Oasis hydrophilic-lipophilic balance sorbent were applied  
as passive samplers (Waters, MA, USA; Alvarez et al., 2004). At each site, four POCIS were  
deployed for six weeks in the middle of the water column to absorb polar compounds from the

145 surface water. After field exposure, the POCIS extracts were prepared and pooled for further  
analysis (details in Van der Lee et al., Under review). POCIS extracts were subjected to three in  
vitro chemical activated luciferase gene expression (CALUX®) bioassays at the BioDetection  
Systems laboratories (Amsterdam, the Netherlands), including the Estrogen receptor (ER $\alpha$ ), the  
androgen receptor antagonism (anti-AR) and the progesterone receptor antagonism (anti-PR)  
150 CALUX assays. The activity of the extracts were expressed as bio-analytical equivalents of the  
corresponding reference compounds and divided by the effect-based trigger (EBT) value of each  
assay to obtain a measure of the ecotoxicological risk caused by the bioactive compounds present at  
each site (Brion et al., 2019; Escher et al., 2018; Van der Lee et al., Under review). We considered  
ER $\alpha$  risk as a proxy for the presence of pharmaceuticals and personal care products and the mean of  
155 anti-AR and the anti-PR risks as a proxy for the presence of pesticides in the surface waters  
(Pieterse et al., 2015; Väitalo et al., 2016; Van der Lee et al., Under review). Each stressor variable  
was scaled to a standard deviation of one and centred at its mean for further analysis.

#### *Invertebrate community composition*

160 Six invertebrate samples were collected at each site on a single occasion. Three subsamples were  
taken with a pond net (1mm mesh size, 25 cm width) that was swept over a length of 0.5 m of  
submerged vegetation, while the other three subsamples were taken with the same net swept over  
the top layer of the sediment. The samples were stored overnight at 4 °C with oxygen supply,  
washed over 1 mm and 250  $\mu$ m sieves, sorted alive and preserved in 70% ethanol until  
165 identification. Overall, a total of 33298 individuals belonging to 106 invertebrate taxa were  
collected. Invertebrates were identified to the genus level with a few exceptions, specifically  
Oligochaeta (order), Hydracarina (order) and Diptera (family). Corixidae were, if possible, further  
identified to the species level, as they are the family with the highest number of sound-producing  
species (Aiken, 1985). In total, 26 of the identified taxa are known for producing sounds (Table 1).

170 The sum of taxon abundance for the six replicate invertebrate samples per site was  $\log_{10}(x+1)$   
transformed before further analysis to minimize the effect of high density taxa.

### *Dissolved oxygen dynamics*

Dissolved oxygen (DO) concentrations (mg/L) were measured with optical HOB0® Dissolved  
175 Oxygen loggers U26-001, protected by the antifouling protective guard U26-GUARD-2 (Onset  
Computer Corporation, Bourne, MA, USA). Following Van der Lee et al., (2018) measurements  
were taken every ten minutes for six consecutive days in the mid-channel, 15 cm under the water  
surface. This was repeated three times at each site during the six week period, rotating weekly  
between the sites. Percent DO saturation was calculated from the DO concentrations and  
180 temperature, assuming 0 ‰ salinity and 1 atm barometric pressure, using DOTABLES developed  
by the U.S. Geological Survey (2011) for further analysis (Van der Lee et al., 2018). The mean DO  
saturation was calculated per 10 minute time step over the 18 measurement days. The DO dynamics  
were represented by the mean DO saturation, while the fluctuation was calculated as the maximum  
values minus the minimum values.

185

### *Acoustic sampling*

The underwater sounds were monitored with nine autonomous recording platforms consisting of a  
HTI-96 hydrophone (flat frequency response between 20 Hz and 40 kHz, High Tech Inc., Long  
Beach, MS, USA) connected with a 20 m cable to one channel of an autonomous SM2 audio  
190 recorder (Wildlife Acoustics, Maynard, MA, USA). A single SM2 recorder connected to a  
hydrophone was set up at each site to record uncompressed audio files (in the wav format) at a 44.1  
kHz sampling frequency and a 16 bit digitization depth. The hydrophones were placed next to the  
dissolved oxygen sensors 15 cm below the water surface, with their piezoelectric element directed  
downward toward the sediment. The recording schedule was set to one minute every ten minutes,

195 24 hours a day for six consecutive days. This was repeated two times at each site during the six week period, rotating between the sites. There was a two to three week interval between the first and second recording week.

To ensure the quality of the recordings used for the analysis, two randomly selected recordings per day were systematically examined by listening to the recording and inspecting the spectrogram. Due to one malfunctioning hydrophone, the data obtained with this hydrophone (5652 recordings) was excluded from our analyses. Recordings with high levels of anthropogenic noise were identified in one site and this data was also excluded (1792 recordings). We were not able to identify and remove occasional anthropogenic noise, however we expect it to have a negligible effect on our results. As rain can introduce unwanted noise and bias acoustic analyses, we also removed recordings collected when it was raining. Rain periods were assessed using the closest  
205 Royal Netherlands Meteorological Institute (KNMI) meteorological station (Supplementary material 1), resulting in the removal of 2879 recordings. This way, we obtained a final dataset of 26989 recordings with an average of  $1350 \pm 575$  (mean  $\pm$  SD) one-minute recordings for each site (min: 209; max: 2095).

210

### *Acoustic analyses*

The systematic examination of the sound recordings also allowed us to identify the specific frequency bands in which the main acoustic patterns occurred. This way, we identified three frequency bands which appeared to delimit most types of sounds: 0-2 kHz, 2-7 kHz and 7-22.05  
215 kHz (Figure 1). For each of these three bands, we computed three acoustic indices: the Acoustic Complexity Index (*ACI*), the Spectral Entropy (*H<sub>f</sub>*), and the amplitude measured as the sum of raw amplitude from the spectrogram (*Amp*). These three indices were chosen because they are indicative of different aspects of the soundscape (Buxton et al., 2018b; Towsey et al., 2018) and they have been used previously in some of the first ecoacoustic studies in freshwater environments

220 (Desjonquères et al., 2015; Karaconstantis et al., 2020; Linke and Deretic, 2020). *ACI* is a measure that calculates the average difference of spectral amplitude between time windows (Pieretti et al., 2011). *Hf* is analogous to the Shannon entropy index from community ecology. Instead of species probability of presence, *Hf* uses the amplitude of each frequency bin in the mean spectrum (Sueur et al., 2008). This index thus yields a measure of the evenness of the probability mass function.

225 As sound is variable and dynamic over time it was recorded over multiple full days, which raises the issue of temporal autocorrelation and pseudo-replication (Desjonquères et al., 2020a). So far most research in ecoacoustics did not take into account such considerations, even though it may have dramatic impacts on the statistics and subsequent interpretation of the results. In this study, we proposed an innovative solution to this problem by employing Functional Data Analysis (FDA),  
230 which allows to represent a set of temporal data points as a single continuous mathematical function (Ramsay et al., 2009). Here, we used this method to describe and model the daily variations in each acoustic index at each site. To represent the smooth variation of the acoustic indices over a day, we calculated the mean values per ten minute time step resulting in 144 mean values per index per site. To obtain a representation of the daily variation for each site, we used 140 spline bases of order four  
235 and a smoothing parameter of  $10^4$ . This specific smoothing parameter was chosen to optimise the trade-off between degrees of freedom and generalised cross validation (Ramsay et al., 2009). The modes of variation between the sites were then displayed in a Functional Principal Component Analysis (FPCA) for each acoustic index (Ramsay et al., 2009). This way, we obtained the coordinates of each site in a functional FPCA space. In such a FPCA, each axis represents the  
240 maximum variance between the sites in the shape of the splines. The scores for the first FPCA axis (FPC1) explained more than 67% of all acoustic indices and was used in subsequence analyses.

### *Statistical analysis*

To assess whether the sound-producing invertebrate community represents the same indication of

245 the water quality as the entire community, we tested to what extent the separate unconstrained ordination (PCA) of the invertebrate taxa, both of the entire community and of only the sound-producing community, related to the stressors and dissolved oxygen dynamics. Then, to evaluate the potential of passive acoustic monitoring, we correlated the FPC1 of each acoustic index to a separate unconstrained ordination (PCA) of the measured stressors, the entire invertebrate  
250 community composition, the sound-producing invertebrate community composition and the dissolved oxygen dynamics. Significant relations between each acoustic index and each PCA was tested using a 999 permutation process. Significant vectors ( $p < 0.05$ ) for acoustic indices were plotted on the ordination to show the correlation with the ordination configuration.

All analyses were performed in R (R Core Team, 2015; v. 3.6.0) using the *seewave* package  
255 to compute the acoustic indices (Sueur et al., 2018; v. 2.1.3), the *fda* package to compute the FPCA analysis (Ramsay et al., 2014; v. 2.4.8) and the package *vegan* to compute the PCA and to fit the acoustics indices on the ordinations (Oksanen et al., 2013; v. 2.5-6).

## Results

260 The acoustic recordings contained various sound types associated with different processes including sound production by invertebrates, ticking and bubble sounds, wind and anthropogenic noise (Figure 1, Supplementary material 2-5). These sound types were in most cases confined to specific frequency bands (0-2, 2-7 and 7-22.05 kHz). The daily variation of acoustic indices depended on the index, the site and the frequency band considered (Figure 2). Some of the indices, such as  $Hf_{0-2}$   
265 and  $Amp_{0-2}$ , had similar temporal patterns with a maximum or minimum in the afternoon (12:00-18:00), but showed strong differences in the magnitude of the daily peaks. While others, like  $Amp_{2-7}$ , showed less differences in the magnitude of daily variation but higher differences in temporal patterns. Finally other indices, such as  $Hf_{2-7}$ ,  $Hf_{7-22}$  or  $Amp_{2-7}$  differed both in magnitude of daily variations and in temporal patterns, with in some sites a secondary peak of activity at night.

270 In terms of the stressors, the entire invertebrate community showed significant correlations with orthophosphate concentrations, the proxies for pesticides and pharmaceuticals and the fluctuation in dissolved oxygen saturation (Figure 3a, Table 2). The sound-producing invertebrate community only correlated significantly to the mean water temperature (Figure 3b, Table 2).

In terms of the acoustic indices, the intensity of the stressors only correlated significantly to 275  $Hf_{0-2}$  (Figure 4a, Table 3). Similarly to the stressors, the entire invertebrate community also correlated significantly with this acoustic index, as well as to  $Amp_{0-2}$  (Figure 4b, Table 3). The sound-producing invertebrate community composition correlated significantly with  $ACI$  and  $Hf$  in the frequency band 2-7 kHz as well as  $Amp_{7-22.05}$  and  $ACI_{0-2}$ . The loading scores for  $Amp_{7-22.05}$  and  $ACI_{0-2}$  were in the same direction as the loading score of *Micronecta* on PC2 (Figure 4c, Table 3). 280 The dissolved oxygen saturation correlated significantly with all acoustic indices, except for  $ACI_{0-2}$  and  $Amp_{2-7}$  (Figure 4d, Table 3). All arrows were mainly associated with the y-axis, representing fluctuations in dissolved oxygen. The strongest correlation was found with  $Amp_{0-2}$  ( $R^2 = 0.52$ ,  $p = 0.001$ ).

## 285 Discussion

The present study tested the relationship between acoustic patterns and the intensity of various anthropogenic stressors, invertebrate community composition and dissolved oxygen dynamics in temperate lowland streams and drainage ditches under a gradient of anthropogenic stress from agricultural activities and WWTP discharges. Our results showed that the acoustic patterns were 290 primarily associated with the composition of the sound-producing invertebrate community and the fluctuation in dissolved oxygen saturation. Below we discuss to what extent these results corroborate the utility of passive acoustic monitoring in water quality assessment.

The sound-producing invertebrate community composition correlated to two acoustic indices in the frequency band 2-7 kHz ( $ACI$  and  $Hf$ ), while the presence of *Micronecta* appeared to

295 be associated with the high frequency band (*Amp*<sub>7-22.05</sub>). This is coherent with the findings from  
previous studies showing that sounds produced by soniferous aquatic insects are concentrated  
within the 5-6.5 kHz range (Aiken, 1982), while *Micronecta* is known to generate a high-pitch  
sound with a dominant frequency in the 7-12 kHz band (Desjonquères et al., 2020b; Sueur et al.,  
2011). Indeed a wide diversity of invertebrates produce underwater sounds (Aiken, 1985;  
300 Desjonquères, 2016; Desjonquères et al., 2020a) and the family Corixidae is considered to be one of  
the main emitters of sounds in (shallow) freshwater environments (Desjonquères et al., 2018).  
Moreover, the daily pattern of these indices showed a distinct second peak at night, corroborating  
previous findings that Heteroptera chorus peak at 2 a.m. (Linke et al., 2020).

The acoustic indices were, however, not related to the intensity of the measured stressors.  
305 While the entire invertebrate community composition indicated several of the anthropogenic  
stressors, including nutrients, pollutants and dissolved oxygen fluctuation, the sound-producing taxa  
only related significantly to temperature. This corroborates the common use of invertebrate  
community composition in water quality assessment, as they are a diverse group reflecting a wide  
range of sensitivity to anthropogenic stressors (Resh and Rosenberg, 1993). The lack of a  
310 relationship between the measured stressors and the acoustic indices may be due to the fact that the  
majority of sound-producing invertebrate taxa occurring in these lowland waters belong to the order  
of Heteroptera, which are generally considered moderately tolerant to chemical stressors, such as  
nutrient loading and organic toxicants (Lock et al., 2013; Von der Ohe and Liess, 2004). Previous  
studies rather related their presence to the habitat structure of water bodies, such as vegetation  
315 coverage (Dias-Silva et al., 2010; Olosutean and Ilie, 2013), which was not included in our study.  
This was confirmed by Desjonquères et al., (2018) who showed that when vegetation density was  
included as part of the studied gradient, the acoustic community could indeed be related to the  
entire invertebrate community. So, even though the acoustic indices could represent the sound-  
producing invertebrate community, they may have limited value as indicator in water quality

320 assessment, as they did not represent the impact of different anthropogenic stressors.

Almost all acoustic indices (7 out of 9) were associated with the daily fluctuation in dissolved oxygen saturation and many of them showed a peak in the afternoon, which is a pattern typically observed for dissolved oxygen saturation in these water bodies. Similarly, Felisberto et al. (2015) observed that acoustic patterns in low (0.4-0.8 kHz) and medium (1.5-3.5 kHz) frequency bands followed the same diel cycle as measured by dissolved oxygen loggers in a marine environment. Interestingly, these frequency bands are similar to the ones for which high correlations values were observed in this study. Dissolved oxygen saturation in the water column is affected by 1) the release of oxygen by photosynthetic primary producers during the day, 2) the uptake of oxygen through respiration by all organisms and 3) the exchange of oxygen with the air (i.e. re-aeration) (Odum, 1956). Previous studies have shown that each of these processes produces sounds. Specifically, Kratochvil and Pollirer (2017) reported that an aquatic plant, *Elodea canadensis*, emits short sound pulses with a wide frequency band as it produces and releases oxygen bubbles in the water. Freeman et al. (2018) observed similar results for marine macroalgae and argued that these sounds may thus be used as an indicator for photosynthetic activity. The respiration by microorganisms decomposing organic matter (both aerobically and anaerobically) is also suspected to produce ticking sounds, as gas bubbles are formed and expelled (Felisberto et al., 2015; Linke et al., 2018). Lastly, Morse et al. (2007) were able to relate sounds at the water-air interface to re-aeration rates.

Our findings, along with these studies, indicate that acoustic indices could be used to estimate metabolism in water bodies, which may subsequently be used in water quality assessment. Passive acoustic monitoring may even overcome certain challenges encountered in the estimation of metabolism from diel dissolved oxygen curves, such as the possibility to split up different components if different processes emit different acoustic patterns, the ability to estimate re-aeration rates and the inclusion of anaerobic respiration (Staehr et al., 2012). Future research should focus on

345 the selection of suitable frequency bands or the detection of specific acoustic patterns that are not  
sensitive to confounding factors, affecting the acoustic patterns emitted by metabolism-related  
processes, such as the sounds of invertebrates, surface agitation due to wind and the influence of  
water movement on bubble formation and retention (Felisberto et al., 2015; Freeman et al., 2018).  
This may be achieved with a combination of laboratory and field studies identifying the acoustic  
350 patterns emitted by specific sources of underwater sounds.

In conclusion, the presently employed acoustics indices allowed the detection of sound-  
producing invertebrate taxa as well as the fluctuation in dissolved oxygen saturation. In terms of  
water quality assessment, the acoustic indices poorly indicated the intensity of the anthropogenic  
stressors compared to the traditional method which samples invertebrate communities with a net,  
355 presumably due to the dominance of the relatively insensitive Heteroptera in the sound-producing  
community. In contrast, the strong relation between acoustic indices and oxygen fluctuation  
indicated that passive acoustic monitoring may be used to estimate metabolism in these water  
bodies. The knowledge of these sounds is still emerging, we therefore suggest that the next step in  
freshwater ecoacoustics is to precisely characterise the sounds individually emitted by  
360 photosynthesis, respiration and re-aeration, so these processes can be distinguished. This would  
greatly enhance the potential of ecoacoustics as a monitoring tool in freshwater environments.

### **Author contributions**

GHvdL, CD and PFMV conceived the ideas and designed methodology; GHvdL conducted the  
365 field work, collecting the data and identifying invertebrate samples; CD and PFMV participated in  
the field work; CD and GHvdL analysed the data and led the redaction of the manuscript. All  
authors contributed critically to the drafts and gave final approval for publication.

### **Funding**

370 This research was part of the Smart monitoring project (443.324), funded by the foundation for applied water research STOWA.

### Acknowledgement

We thank Milo de Baat for preparing and deploying the POCIS and providing us with the CALUX  
375 assay data; Milo de Baat, Nienke Wieringa and Eline Reus for assistance in the field; Ralf Verdonschot, Dorine Dekkers, Mandy Velthuis and Annieke Borst for assistance with sorting invertebrate samples; Dorine Dekkers for checking the invertebrate identification; Peter Serne for analyzing the nutrient concentrations; all water boards that were involved in selecting suitable sites.

### 380 References

- Aiken, R.B., 1985. Sound production by aquatic insects. *Biological Reviews* 60, 163–211.
- Aiken, R.B., 1982. Shallow-water propagation of frequencies in aquatic insect sounds. *Canadian Journal of Zoology* 60, 3459–3461. <https://doi.org/10.1139/z82-436>
- Allan, J.D., 2004. Landscapes and riverscapes: the influence of land use on stream ecosystems. *Annu. Rev. Ecol. Evol. Syst.* 35, 257–284.
- Alvarez, D.A., Petty, J.D., Huckins, J.N., Jones-Lepp, T.L., Getting, D.T., Goddard, J.P., Manahan, S.E., 2004. Development of a passive, in situ, integrative sampler for hydrophilic organic contaminants in aquatic environments. *Environmental Toxicology and Chemistry: An International Journal* 23, 1640–1648.
- Bonada, N., Prat, N., Resh, V.H., Statzner, B., 2006. Developments in aquatic insect biomonitoring: a comparative analysis of recent approaches. *Annu. Rev. Entomol.* 51, 495–523.
- Boulton, A.J., 1999. An overview of river health assessment: philosophies, practice, problems and prognosis. *Freshwater Biology* 41, 469–479.
- Brion, F., De Gussem, V., Buchinger, S., Hollert, H., Carere, M., Porcher, J.-M., Piccini, B., Féray, C., Dulio, V., Könemann, S., 2019. Monitoring estrogenic activities of waste and surface waters using a novel in vivo zebrafish embryonic (EASZY) assay: Comparison with in vitro cell-based assays and determination of effect-based trigger values. *Environment international* 130, 104896.
- Bunn, S.E., 1995. Biological monitoring of water quality in Australia: workshop summary and future directions. *Australian Journal of Ecology* 20, 220–227.
- Burdon, F.J., Munz, N.A., Reyes, M., Focks, A., Joss, A., Räsänen, K., Altermatt, F., Eggen, R.I.L., Stamm, C., 2019. Agriculture versus wastewater pollution as drivers of macroinvertebrate community structure in streams. *Science of the Total Environment* 659, 1256–1265.
- Burivalova, Z., Towsey, M., Boucher, T., Truskinger, A., Apelis, C., Roe, P., Game, E.T., 2018. Using soundscapes to detect variable degrees of human influence on tropical forests in Papua New Guinea: Tropical-Forest Soundscapes. *Conservation Biology* 32, 205–215. <https://doi.org/10.1111/cobi.12968>

- Buxton, R.T., Lendrum, P.E., Crooks, K.R., Wittemyer, G., 2018a. Pairing camera traps and acoustic recorders to monitor the ecological impact of human disturbance. *Global Ecology and Conservation* 16, e00493. <https://doi.org/10.1016/j.gecco.2018.e00493>
- Buxton, R.T., McKenna, M.F., Clapp, M., Meyer, E., Stabenau, E., Angeloni, L.M., Crooks, K., Wittemyer, G., 2018b. Efficacy of extracting indices from large-scale acoustic recordings to monitor biodiversity: Acoustical Monitoring. *Conservation Biology* 32, 1174–1184. <https://doi.org/10.1111/cobi.13119>
- Clapcott, J.E., Collier, K.J., Death, R.G., Goodwin, E.O., Harding, J.S., Kelly, D., Leathwick, J.R., Young, R.G., 2012. Quantifying relationships between land-use gradients and structural and functional indicators of stream ecological integrity. *Freshwater Biology* 57, 74–90.
- Desjonquères, C., 2016. Acoustic diversity and ecology of freshwater environments: exploration in temperate environments (PhD Thesis). Museum national d’histoire naturelle-MNHN PARIS.
- Desjonquères, C., Gifford, T., Linke, S., 2020a. Passive acoustic monitoring as a potential tool to survey animal and ecosystem processes in freshwater environments. *Freshw Biol* 65, 7–19. <https://doi.org/10.1111/fwb.13356>
- Desjonquères, C., Rybak, F., Castella, E., Llusia, D., Sueur, J., 2018. Acoustic communities reflects lateral hydrological connectivity in riverine floodplain similarly to macroinvertebrate communities. *Scientific Reports* 8. <https://doi.org/10.1038/s41598-018-31798-4>
- Desjonquères, C., Rybak, F., Depraetere, M., Gasc, A., Le Viol, I., Pavoine, S., Sueur, J., 2015. First description of underwater acoustic diversity in three temperate ponds. *PeerJ* 3, e1393. <https://doi.org/10.7717/peerj.1393>
- Desjonquères, C., Rybak, F., Ulloa, J.S., Kempf, A., Bar Hen, A., Sueur, J., 2020b. Monitoring the acoustic activity of an aquatic insect population in relation to temperature, vegetation and noise. *Freshw Biol* 65, 107–116. <https://doi.org/10.1111/fwb.13171>
- Dias-Silva, K., Cabette, H.S., Juen, L., De Marco Jr, P., 2010. The influence of habitat integrity and physical-chemical water variables on the structure of aquatic and semi-aquatic Heteroptera. *Zoologia* 27.
- Escher, B.I., Aït-Aïssa, S., Behnisch, P.A., Brack, W., Brion, F., Brouwer, A., Buchinger, S., Crawford, S.E., Du Pasquier, D., Hamers, T., 2018. Effect-based trigger values for in vitro and in vivo bioassays performed on surface water extracts supporting the environmental quality standards (EQS) of the European Water Framework Directive. *Science of The Total Environment* 628, 748–765.
- Farina, A., Gage, S.H., 2017. *Ecoacoustics: The ecological role of sounds*. John Wiley & Sons.
- Felisberto, P., Jesus, S.M., Zabel, F., Santos, R., Silva, J., Gobert, S., Beer, S., Björk, M., Mazzuca, S., Procaccini, G., Runcie, J.W., Champenois, W., Borges, A.V., 2015. Acoustic monitoring of O<sub>2</sub> production of a seagrass meadow. *Journal of Experimental Marine Biology and Ecology* 464, 75–87. <https://doi.org/10.1016/j.jembe.2014.12.013>
- Freeman, S.E., Freeman, L.A., Giorli, G., Haas, A.F., 2018. Photosynthesis by marine algae produces sound, contributing to the daytime soundscape on coral reefs. *PloS one* 13, e0201766.
- Friberg, N., Bonada, N., Bradley, D.C., Dunbar, M.J., Edwards, F.K., Grey, J., Hayes, R.B., Hildrew, A.G., Lamouroux, N., Trimmer, M., 2011. Biomonitoring of human impacts in freshwater ecosystems: the good, the bad and the ugly, in: *Advances in Ecological Research*. Elsevier, pp. 1–68.
- Gage, S.H., Towsey, M., Kasten, E.P., 2017. Analytical methods in ecoacoustics. *Ecoacoustics. The Ecological Role of Sounds*, edited by Farina A. and Gage SH Oxford, UK: Wiley 273–296.
- Gibb, R., Browning, E., Glover-Kapfer, P., Jones, K.E., 2019. Emerging opportunities and challenges for passive acoustics in ecological assessment and monitoring. *Methods in Ecology and Evolution* 10, 169–185. <https://doi.org/10.1111/2041-210X.13101>

- Jansson, A., 1974. Annual periodicity of male stridulation in the genus *Cenocorixa* (Hemiptera, Corixidae). *Freshwater Biology* 4, 93–98. <https://doi.org/10.1111/j.1365-2427.1974.tb00941.x>
- Karaconstantis, C., Desjonquères, C., Gifford, T., Linke, S., 2020. micro- and macro-variation in a chain of waterholes. *Freshwater Biology* 65, 96–106.
- Karr, J.R., 1999. Defining and measuring river health. *Freshwater biology* 41, 221–234.
- Kratochvil, H.G., Pollirer, M., 2017. Acoustic effects during photosynthesis of aquatic plants enable new research opportunities. *Sci Rep* 7, 44526. <https://doi.org/10.1038/srep44526>
- Linke, S., Decker, E., Gifford, T., Desjonquères, C., 2020. Diurnal variation in freshwater ecoacoustics: Implications for site-level sampling design. *Freshw Biol* 65, 86–95. <https://doi.org/10.1111/fwb.13227>
- Linke, S., Deretic, J., 2020. Ecoacoustics can detect ecosystem responses to environmental water allocations. *Freshw Biol* 65, 133–141. <https://doi.org/10.1111/fwb.13249>
- Linke, S., Gifford, T., Desjonquères, C., Tonolla, D., Aubin, T., Barclay, L., Karaconstantis, C., Kennard, M.J., Rybak, F., Sueur, J., 2018. Freshwater ecoacoustics as a tool for continuous ecosystem monitoring. *Frontiers in Ecology and the Environment* 16, 231–238. <https://doi.org/10.1002/fee.1779>
- Lock, K., Adriaens, T., Van De Meutter, F., Goethals, P., 2013. Effect of water quality on waterbugs (Hemiptera: Gerromorpha & Nepomorpha) in Flanders (Belgium): results from a large-scale field survey, in: *Annales de Limnologie-International Journal of Limnology*. EDP Sciences, pp. 121–128.
- Morse, N., Bowden, W.B., Hackman, A., Pruden, C., Steiner, E., Berger, E., 2007. Using sound pressure to estimate reaeration in streams. *Journal of the North American Benthological Society* 26, 28–37.
- Odum, H.T., 1956. Primary Production in Flowing Waters 1. *Limnology and oceanography* 1, 102–117.
- Oksanen, J., Blanchet, F.G., Kindt, R., Legendre, P., Minchin, P.R., O’Hara, R.B., Simpson, G.L., Solymos, P., Stevens, M.H.H., Wagner, H., 2013. Package ‘vegan.’ *Community ecology package, version 2*.
- Olosutean, H., Ilie, D.M., 2013. Are semi-aquatic bugs (Heteroptera: Nepomorpha) indicators of hydrological stability of permanent ponds? *Aquatic insects* 35, 105–118.
- Ormerod, S.J., Dobson, M., Hildrew, A.G., Townsend, C., 2010. Multiple stressors in freshwater ecosystems. *Freshwater Biology* 55, 1–4.
- Palmer, M.A., Febria, C.M., 2012. The heartbeat of ecosystems. *Science* 336, 1393–1394.
- Pieretti, N., Farina, A., Morri, D., 2011. A new methodology to infer the singing activity of an avian community: The Acoustic Complexity Index (ACI). *Ecological Indicators* 11, 868–873. <https://doi.org/10.1016/j.ecolind.2010.11.005>
- Pieterse, B., Rijk, I.J.C., Simon, E., van Vugt-Lussenburg, B.M.A., Fokke, B.F.H., Van der Wijk, M., Besselink, H., Weber, R., van der Burg, B., 2015. Effect-based assessment of persistent organic pollutant and pesticide dumpsites using mammalian CALUX reporter cell lines. *Environmental Science and Pollution Research* 22, 14442–14454.
- R Core Team, 2015. *R: A Language and Environment for Statistical Computing*. R Foundation for Statistical Computing, Vienna, Austria.
- Ramsay, J., Hooker, G., Graves, S., 2009. *Functional Data Analysis with R and MATLAB*. Springer New York, New York, NY.
- Ramsay, J.O., Wickham, H., Graves, S., Hooker, G., 2014. *fda: Functional Data Analysis*.
- Resh, V.H., 2008. Which group is best? Attributes of different biological assemblages used in freshwater biomonitoring programs. *Environmental Monitoring and Assessment* 138, 131–138.
- Resh, V.H., Rosenberg, D.M., 1993. *Freshwater biomonitoring and benthic macroinvertebrates*.

Chapman & Hall New York, NY, USA:

- Staehr, P.A., Testa, J.M., Kemp, W.M., Cole, J.J., Sand-Jensen, K., Smith, S.V., 2012. The metabolism of aquatic ecosystems: history, applications, and future challenges. *Aquatic Sciences* 74, 15–29.
- Sueur, J., Aubin, T., Simonis, C., Lellouch, L., Brown, E.C., Depaetere, M., Desjonqueres, C., Fabianek, F., Gasc, A., Kasten, E., 2018. Package ‘seewave.’
- Sueur, J., Farina, A., 2015. Ecoacoustics: the ecological investigation and interpretation of environmental sound. *Biosemiotics* 8, 493–502. <https://doi.org/10.1007/s12304-015-9248-x>
- Sueur, J., Farina, A., Gasc, A., Pieretti, N., Pavoine, S., 2014. Acoustic Indices for Biodiversity Assessment and Landscape Investigation. *Acta Acustica united with Acustica* 100, 772–781. <https://doi.org/10.3813/AAA.918757>
- Sueur, J., Mackie, D., Windmill, J.F.C., 2011. So Small, So Loud: Extremely High Sound Pressure Level from a Pygmy Aquatic Insect (Corixidae, Micronectinae). *PLoS ONE* 6, e21089. <https://doi.org/10.1371/journal.pone.0021089>
- Sueur, J., Pavoine, S., Hamerlynck, O., Duvail, S., 2008. Rapid Acoustic Survey for Biodiversity Appraisal. *PLoS ONE* 3, e4065. <https://doi.org/10.1371/journal.pone.0004065>
- Sugai, L.S.M., Desjonquères, C., Silva, T.S.F., Llusia, D., 2019a. A roadmap for survey designs in terrestrial acoustic monitoring. *Remote Sens Ecol Conserv* rse2.131. <https://doi.org/10.1002/rse2.131>
- Sugai, L.S.M., Silva, T.S.F., Ribeiro, J.W., Llusia, D., 2019b. Terrestrial Passive Acoustic Monitoring: Review and Perspectives. *BioScience* 69, 15–25. <https://doi.org/10.1093/biosci/biy147>
- Towsey, M., Znidersic, E., Broken-Brow, J., Indraswari, K., Watson, D.M., Phillips, Y., Truskinger, A., Roe, P., 2018. Long-duration, false-colour spectrograms for detecting species in large audio data-sets. *J. Ecoacoust.* 2, IUSWUI. <https://doi.org/10.22261/JEA.IUSWUI>
- Välitalo, P., Perkola, N., Seiler, T.-B., Sillanpää, M., Kuckelkorn, J., Mikola, A., Hollert, H., Schultz, E., 2016. Estrogenic activity in Finnish municipal wastewater effluents. *Water research* 88, 740–749.
- Van der Lee, G.H., De Baat, M.L., Wieringa, N., Kraak, M.H.S., Verdonschot, R.C.M., Verdonschot, P.F.M., Under review. Structural and functional assessment of multi-stressed lowland waters. *Freshwater Science*.
- Van der Lee, G.H., Verdonschot, R.C., Kraak, M.H., Verdonschot, P.F., 2018. Dissolved oxygen dynamics in drainage ditches along a eutrophication gradient. *limnologica* 72, 28–31.
- Von der Ohe, P.C., Liess, M., 2004. Relative sensitivity distribution of aquatic invertebrates to organic and metal compounds. *Environmental Toxicology and Chemistry: An International Journal* 23, 150–156.
- Young, R.G., Matthaei, C.D., Townsend, C.R., 2008. Organic matter breakdown and ecosystem metabolism: functional indicators for assessing river ecosystem health. *Journal of the North American Benthological Society* 27, 605–625.

## Tables

385 Table 1: Invertebrate genera collected in this study that produce sound. Sound production in freshwater invertebrates was reviewed by Desjonquères (2016).

<b>Order</b>	<b>Family</b>	<b>Genus</b>
Trichoptera	Hydropsychidae	<i>Hydropsyche</i>
Coleoptera	Dytiscidae	<i>Cybister, Dytiscus</i>
	Haliplidae	<i>Haliplus</i>
	Hydrophilidae	<i>Anacaena, Enochrus, Helophorus</i>
Heteroptera	Corixidae	<i>Callicorixa, Corixa, Cymatia, Hesperocorixa, Micronecta, Paracorixa, Sigara</i>
	Nepidae	<i>Nepa, Ranatra</i>
	Pleidae	<i>Plea</i>
	Naucoridae	<i>Ilyocoris</i>
Decapoda	-	-

Table 2: Relations between the stressors and dissolved oxygen dynamics and the ordination of the invertebrate communities (all taxa and only sound-producing taxa), as squared correlation coefficient ( $R^2$ ) and significance ( $p$ ) permutation-tested using 999 randomizations ( $N = 20$ ).

	All invertebrate taxa		Sound-producing invertebrate taxa	
	$R^2$	$p$	$R^2$	$p$
Total dissolved nitrogen	0.08	0.541	0.204	0.148
Orthophosphate	0.31	0.038	0.003	0.982
Temperature	0.11	0.403	0.431	0.006
Proxy for pesticides	0.29	0.049	0.025	0.782
Proxy for pharmaceuticals	0.46	0.010	0.132	0.260
Mean dissolved oxygen	0.07	0.551	0.010	0.938
Fluctuation dissolved oxygen	0.40	0.011	0.087	0.482

Table 3: Relations between the acoustic indices and the ordination of different environmental variables as squared correlation coefficient ( $R^2$ ) and significance ( $p$ ) permutation-tested using 999 randomizations ( $N = 20$ ).

		Stressors		All invertebrate taxa		Sound-producing invertebrate taxa		Dissolved oxygen saturation	
		$R^2$	$p$	$R^2$	$p$	$R^2$	$p$	$R^2$	$p$
ACI	0-2	0.08	0.509	0.07	0.558	0.35	0.025	0.31	0.044
	2-7	0.11	0.346	0.20	0.143	0.41	0.031	0.37	0.017
	7-22.05	0.05	0.588	0.07	0.532	0.00	0.969	0.06	0.656
Hf	0-2	0.30	0.048	0.54	0.004	0.15	0.247	0.47	0.009
	2-7	0.11	0.326	0.16	0.236	0.38	0.019	0.44	0.006
	7-22.05	0.18	0.164	0.06	0.557	0.24	0.096	0.46	0.004
amp	0-2	0.21	0.108	0.40	0.015	0.13	0.279	0.52	0.001
	2-7	0.02	0.815	0.21	0.127	0.09	0.425	0.20	0.149
	7-22.05	0.08	0.509	0.07	0.558	0.35	0.025	0.31	0.044

## Figure legends

400 Figure 1: Example of the main acoustic patterns in the recordings with the three frequency bands indicated that delimit most types of sounds: 0-2 kHz, 2-7 kHz and 7-22.05 kHz. (a) Recording from site 2 at 1:50 pm on August 25<sup>th</sup> showing the sounds of gusts of wind and continuous low frequency sounds potentially corresponding to photosynthesis (0-2 kHz), ticking and bubble sounds (2-7 kHz and 7-22.05 kHz); (b) Recording from site 8 at 6:00 am on August 24<sup>th</sup> showing the sounds of gusts  
405 of wind and continuous low frequency sounds potentially corresponding to photosynthesis (0-2 kHz), Corixidae sounds (2-7 kHz) and *Micronecta* sounds (7-22.05 kHz); (c) Recording from site 12 at 5:40 pm on August 28<sup>th</sup> showing ticking sounds (7-22.05 kHz); (d) Recording from site 8 at 14:40 pm on September 16<sup>th</sup> showing anthropogenic noise (2-7 kHz). Information on the sites in Supplementary material 1. The selected sound recordings can be found in Supplementary material  
410 2-5.

Figure 2: FDA representation of the smooth variation of the acoustic indices over a single day. Each curve shows the FDA representation for a frequency band of an acoustic index in a site. The curves were obtained using a spline basis and a smoothing parameter. Notation:  $ACI_{0-2}$  corresponds to the  
415  $ACI$  index over the 0-2 kHz frequency band.

Figure 3: Relations between the stressors, dissolved oxygen dynamics and the ordination of a) entire invertebrate community composition and b) sound-producing invertebrate community composition. Only significant vectors ( $p < 0.05$ ) are plotted (details in Table 2). Taxa shown are prioritized on  
420 abundance.

Figure 4: Relations between the acoustic indices and the ordination of different environmental

variables, including a) stressors, b) dissolved oxygen dynamics, c) entire invertebrate community composition and d) sound-producing invertebrate community composition. Only significant vectors  
425 (p<0.05) are plotted (details in Table 3). Taxa shown are prioritized on abundance.







