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Altered thymic CD4⁺ T-cell recovery after allogeneic hematopoietic stem cell transplantation is critical for Nocardiosis.

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Running title: Nocardiosis and thymic recovery

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Altered thymic CD4⁺ T-cell recovery after allogeneic hematopoietic stem cell transplantation is critical for Nocardiosis.

Abstract

- **Purpose of the study:** *Nocardia* affects immunocompromised human host exhibiting an altered cell-mediated immunity. Infectious risk after allogeneic hematopoietic cell transplantation (AHCT) is significantly correlated to the recovery status of donor-derived immune system, especially CD4⁺T-cells reconstitution and thymopoiesis. The purpose of this paper is to highlight a lack of cell-mediated immunity recovery for patients presenting a nocardiosis compared to a control cohort.
- Patients and methods: This is a case control retrospective monocentric study. We retrospectively analyzed a monocentric cohort of 15 cases of nocardiosis after AHCT and we explored the degree of patients' immunosuppression by phenotyping circulating lymphoid subpopulations, including NK cells, CD8+ T-cells, CD4+ T-cells and CD19+ B-cells. We focused on CD4⁺ T-cell subsets to appreciate thymic output, especially on naive CD4⁺ T-cells (NTE, CD45RA+/RO-CD4+ T-cells) and recent thymic emigrants (RTE, CD4+CD45RA+/RO-/CD31+). Infected patients were paired with a control cohort of patients with identical transplantation characteristics screened on hematological disease, AHCT conditioning, primary graft-versus-host disease (GHVD) prophylaxis, graft type, sex, age, and season at the AHCT and data concerning immunological reconstitution were compared.
 - **Results:** At onset of nocardiosis, circulating lymphocytes and CD4⁺ T-cells means count were respectively 730/μL and 162/μL. CD8⁺ T-cells, CD56⁺ NK cells and CD19⁺ B-cells means count were respectively 362/μL, 160/μL, 112/μL. CD4⁺ T-cells subpopulations, naïve CD4⁺ T-cells production was impaired with NTE and RTE means count at 26/μL and 11/μL respectively. Comparison between nocardiosis cohort and control cohort over time highlight significant lower cellular count for lymphocytes, CD4⁺ T-cells, NTE and RTE with p=0.001, p<0.001, p<0.001, p<0.001, p<0.001 respectively.
 - **Conclusion:** Immune recovery monitoring follow-up after AHCT is of particular importance to identify patients susceptible to develop Nocardiosis. Efficient microbiological investigations toward *Nocardia* such PCR should be used in case of compatible clinical presentation.

Key words: *Nocardia*, opportunistic infection, allogeneic hematopoietic stem cell transplant,
 immune recovery, thymic CD4⁺ T-cell.

Introduction

Nocardia is an ubiquitous group of environmental bacteria belonging to the suborder of aerobic Actinomycetes with *Rhodococus* within the family *Nocardiaceae*, close to the genus *Mycobacterium*. This is an aerobic gram-positive filamentous bacterium with hyphae-like branching. *Nocardia* are saprophytic organisms that can be found in soil, decomposing vegetation, organic matter from animal, as well as water and dust. Several heterogeneous species were described with numerous recent taxonomy revisions [1,2].

Nocardiosis is the name of *Nocardia spp* infection, and could affect immunocompromised human hosts [3]. This opportunistic pathogen affects particularly patients exhibiting an altered cell-mediated immunity [4]. Solid-organ and hematopoietic stem cell transplant recipients concerned one third of nocardiosis [2,5]. The clinical presentation is heterogeneous, non-specific and depends on infectious sites [4]. The most common infectious site is lung by respiratory tract infestation [6], nevertheless other localizations with disseminated presentation could be presented in blood, central nervous system (CNS) or lymph nodes [7]. The common radiological presentation does not have pathognomonic features as it shows lungs condensation with abscesses and nodules. CNS dissemination is also usually described with parenchymal abscesses [2,5,7,8]. As rare and late after allogeneic hematopoietic cell transplantation (AHCT), nocardiosis is potentially lethal with a survival rate reaching 84% (in retrospective studies) [9]. Gathering cohorts of immunosuppressed patients, incidence is estimated from 0.3 to 3.6% [2,9]. This incidence is likely under-estimated [5] as result of *Nocardia* identification challenge [10].

Despite improvement of supportive care during the last decades, infections are still the second cause of mortality after allogeneic hematopoietic stem cell transplantation (AHCT) [11,12]. The infectious risk after AHCT is significantly correlated to the delayed recovery of donor-derived immune system, especially CD4+ T-cells subset expansion [13–15]. Long-lasting steroids and immunosuppressive drugs use in AHCT impair immune recovery and are associated to nocardiosis presentation [2,9,10,16]. We report here a monocentric cohort of patients who underwent AHCT and presented a nocardiosis. The case control analysis depicts the defect of cell-mediated immunity recovery associated with this opportunistic infection.

Patients and methods

We described nocardiosis characteristics, management and outcome after AHCT in a hematopoietic cell transplant recipients' cohort treated at the University Hospital of Besançon. We analyzed immune reconstitution through circulating T-cells populations that were phenotyped before, at onset and after nocardiosis. The different circulating lymphoid populations available for analysis included: NK cells, CD8+ T-cells, CD4+ T-cells and CD19+ B-

- 74 cells. Our regular panel assessing immune reconstitution focused more precisely on naive
- 75 thymic emigrant CD4⁺ T-cells (NTE identified as CD45RA⁺/RO⁻) and recent thymic emigrants
- 76 CD4+ T-cells (RTE identified as CD45RA+/RO-/CD31+) to appreciate thymic output impact
- 77 after transplantation.

Patients

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- 79 Patients were selected from the local clinical data base between May 2001 and December
- 2017. We included all nocardiosis cases defined as "proven", "probable" or "possible" based
- on the similar invasive fungal infection (IFI) classification [17]. We defined "possible"
- 82 nocardiosis by the presence of clinical and radiological features (nodule, condensation,
- abscess), with or without dissemination [8], no alternative diagnosis and improvement after
- anti-*Nocardia* treatment in an immunocompromised host. "Probable" nocardia was defined by
- 85 identification of non-direct bacteriological criteria (direct exam, pathology, positive Nocardia
- PCR), and "proven" by positive *Nocardia* culture confirmation, as described recently [18] (table
- 1). Pathology arguments to bacterial infection or gram-positive bacillus were considered as
- 88 non-direct bacteriological criteria.
- 89 In purpose to strength our analysis, we selected a matched 2:1 control cohort based on the
- 90 following matching criteria: on hematological disease, AHCT conditioning (myeloablative, MAC
- or reduced intensity conditioning regimens, RIC), primary graft-versus-host disease (GHVD)
- 92 prophylaxis (drugs regimens), HLA matching (MRD, MUD, MMUD and haploidentical), sex,
- age, and season at the AHCT from retrospective data base. Immune reconstitution data over
- time were compared between these two groups during the first two years after AHCT.
- Data were collected retrospectively. This non-interventional study was conducted under the
- 96 French Medical Research Data Processing Advisory Committee (CCTIRS) and the French
- 97 Information Technology and Privacy Commission (CNIL) recommendations about anonymous
- 98 clinical data use. All patients sign a consent form permitting to use their clinicals data
- 99 anonymously respecting the Helsinski declaration.
- 100 We collected data concerning hematological disease, AHCT management, follow-up of
- immune recovery, and nocardiosis characteristics directly from the charts.

Statistic methods

- Statistical analyses and graphs were performed with Rstudio® Version 1.1.463. Continuous
- variables were summarized with means and standard deviation with interval of confidence at
- 105 95%. All observations over time were represented on stripcharts with identification of median
- value. Immune recovery was evaluated at least every three months during the first 2 years
- 107 (more frequently for frail patients with GVHD or recurrent infection) and compared with the

control population. Patients were censured at death time. Means of T-cell subset count were compared by Mann-Whitney test.

Conditioning and GVHD prophylaxis were adapted to hematological disease and comorbidity

AHCT management

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112 score, such as updated Sorror's score [19], as well as to our local policy and European Bone 113 Marrow Transplantation Group (EBMT) recommendations. During the first month after AHCT, all patients were hospitalized in High Efficiency Particulate 114 115 Air laminar flow chamber until neutrophilic recovery. An intravenous antibiotic therapy was administered empirically or adjusted to patient's environment, in case of fever [20]. Bacterial, 116 viral and fungal prophylaxis were administered as already described: long-term valaciclovir 117 [21], fluconazole or posaconazole during 6 months or more in case of GVHD [22,23], 118 trimethoprim-sulfamethoxazole (TMP-SMX 800mg/160mg) three time a week, or in case of 119 adverse event atovaquone or aerosol of pentamidine [24]. Intravenous polyvalent 120

immunoglobulins were administrated (400mg/kg) in case of immunoglobulin G <3g/L. Vaccines

were administered according to EBMT schedules [25] and adapted to the immune recovery.

Cytomegalovirus (CMV) PCR and fungal biomarkers (galactomannan (GM) completed by 123 124 specific Aspergillus PCRs) monitoring were performed once a week until immunosuppressive 125 therapy withdrawal [26]. In case of sepsis the following tests were systematically performed: 126 blood cultures, urine bacteriological analysis, GM, Aspergillus, Mucormycosis [27], Toxoplasma gondii, and CMV, C-reactive protein count, thoracic CT scan, and also 127 Pneumococcus and Legionella antigenuria in case of pulmonary infection. In case of lack of 128 microbiological documentation by non-invasive approach, *Nocardia* detection was included in 129 our pneumological investigational protocol for immunocompromised patients by bronchial-130 131 alveolar lavage (BAL). Nocardia detection was also realized in case of abscess after biopsy and association of clinical and radiological presentation compatible with nocardiosis [8]. In 132 case of nocardiosis diagnosis, a cerebral evaluation was systematically performed by magnetic 133

Specific microbiological detection of *Nocardia*

All bacteriological samples were initially observed by Gram direct examination. Then they were usually seeded in standard bacteria growth media with daily examination for growth. In this kind of regular growth media, *Nocardia* species grow slowly, until 2 to 3 weeks [10]. *Nocardia* colonies appeared chalky white in case of *hyphae* production.

resonance imaging (MRI) or by CT scan if MRI was not rapidly available.

Nocardia confirmation was centralized to the referent laboratory in Lyon (*Observatoire français* des nocardioses), and was identified by PCR as recently published [18]. Antimicrobial agent
 sensibility was tested in vitro on isolates.

Immune recovery assessment

- Lymphocytes counts were evaluated by XN-10 system automat (Sysmex, Japan). Absolute 144 lymphocytes, CD4+ T-cells, CD8+ T-cells, CD56+ NK cells and CD19+ B-cells counts were 145 determined by flow cytometry by TetraCXP1 method and fluorescence evaluation by FC500 146 147 cytometer (Beckman Coulter, France) [28]. Naïve CD4+ T-cells population was characterized by CD45RA+/CD45RO-/CD4+ labeling in immunofluorescence. CD31+ expression was 148 149 evaluated in order to identify RTE lymphocytes and subsequent thymic lymphocyte maturation 150 [29]. Our laboratory benchmark for general healthy population was 493-1666 CD4+ T-cells/µL, 224-1112 CD8+ T-cells/μL, 75-520 CD19+ B-cells/μL and 73-654 CD56+ NK cells/μL. 151 Quantitative immunoglobulin evaluation was also performed to quantitively assess the humoral 152 153 immune recovery.
- 154 Chimerism was performed on bone marrow or on blood by semi-quantitative PCR with unique 155 donor and recipient short tandem repeat markers (STR) amplification [30]. The quantification 156 was obtained by donor and recipient STR loci fluorescence GeneScan computer analyze 157 (Applied Biosystem, France).

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Results

Population

- Between May 2001 and December 2017, we highlighted 15 cases of nocardiosis in the 752
- AHCT realized in our hematological department, incidence rate of 1.99%.
- In the nocardiosis cohort one patient received 2 AHCT, acute myeloid leukemia (AML) was the
- most frequent hematological disease (61%), 10 patients (63%) were in complete response
- 165 (CR) at the transplantation time, 2 in partial response, 3 in progressive disease. Males were
- preponderant (sex ratio 4:1) (table 2). Myeloablative conditioning regimens (MAC) were used
- for 9 patients (60%) and the 6 others conditioning regimens (40%) were reduce intensity
- 168 conditionings (RIC). Ten patients (77%) received anti-thymoglobulin serum for GVHD
- prophylaxis during conditioning regimen (2.5 mg/kg for 1 patient, 5 mg/kg for 8 and 7.5 mg/kg
- 170 for one). Median CD34+ infused cells count was 2.69 x 10⁶/kg [IC 95%, 1.85-3.57].
- 171 Characteristics were similar to control cohort as report in table 2.

Nocardiosis presentation

Six nocardiosis occurred in patient receiving AHCT in spring whereas others were equally presented in summer, autumn and winter. Median duration time between AHCT and nocardiosis diagnosis was 218 days [IC95%, 118-317]. On the 15 nocardiosis, 7 (47%) were localized then 8 (53%) were disseminated (at least 2 infectious sites). Thirteen (87%) presented a pulmonary involvement, 8 (53%) were neurological nocardiosis with central

nervous system (CNS) involvement with 4 (24%) ocular involvement, 2 (12%) were skin nocardiosis and 1 was a bacteriema (table 3 - 4). Main radiological presentation was abscesses (11/15, 73%), with lungs nodules (7/13, 53%) and condensation (9/13, 69%) half time associated to abscesses (5/13, 38%), and CNS abscesses (8/8). As previously defined, 5 nocardiosis cases were proved, 6 were probable, 4 were possible (table 4). Ten (66%) were documented by PCR, 3 (20%) by cyto-histopathology, 5 (33%) by late culture with 1 (6%) on blood culture. Five patients had nocardia detection through at least 2 different methods. Four patients had missing documentation: 3 without possible laboratory *Nocardia* detection on samples and one which the sample was lost. Nevertheless they were highly suspected based on clinical and radiology presentation and because of the negative regular microbiological investigation regard and the favorable evolution under specific anti-*Nocardia* treatment.

- Two patients presented a nocardiosis recurrence: one after a second AHCT 3 years later and the other one 2 months after anti-microbial prophylaxis ending. For this patient, the secondary prophylaxis had been maintained Eight patients received TMP-SMX prophylaxis at nocardiosis diagnosis and relapse. Among them, 3 had gut GVHD. Four patients presented concomitant CMV reactivation and 4 other patients presented co-infection (2 *Mucormycosis* and 2 *Pneumocystis*). One of the patients with pneumocystosis did not followed the prescribed TMP-SMX prophylaxis. Median duration time of antimicrobial treatment was 180 days [IC95%, 130-229].
- At onset of nocardiosis infection, 11 patients (65%) received an imipenem-base induction antimicrobial therapy and 9 (53%) received a TMP-SMX-base consolidation antimicrobial therapy. Details were reported in table 3. All documented Nocardia were TMP-SMX sensitive.

Immunosuppression and GHVD

- In the nocardiosis cohort, GVHD prophylaxis was mostly performed by cyclosporin (CsA) associated to methotrexate for 8 patients (47%). Twelve patients (80%) presented an acute GVHD (aGVHD). All were skin aGVHD with 8 Glückberg's grade 1-2 and 4 more than grade 2. One presented a gut aGVHD (stage 1) and one a liver aGVHD (stage 1). The mean cumulative steroid dose administrated after onset of aGVHD was 25mg/kg/4weeks [IC95%, 15.5-34.9], compared to 19mg/kg/4weeks [IC95%, 8.5-28.6] in the control cohort but was not significative (p=0.20) (table 2).
- At nocardiosis diagnosis, 14 patients received immunosuppressive drugs allocated as follow: tacrolimus for 6 (40%), CsA for 4 (27%), high dose prednisone for 2 (13%), JAK2 inhibitor for 1 (6%), JAK2 inhibitor associated to CsA and mycophenolate mofetil, pegylated interferon and lenalidomide for 1. Only one patient had active skin aGVHD (stage 2) and 11 (73%) presented a cGVHD which were mostly moderate (41%) with skin lesions (77%). Thirteen patients

- 213 received steroid at nocardiosis diagnosis with a prednisone mean dose at 0.3 mg/kg/day
- 214 [IC95%, 0.19-0.58].
- 215 **Immune recovery** (figures 1 and 2 A-H)
- 216 At nocardiosis diagnosis immune recovery was deficient with a low cellular count for CD4⁺ T-
- cells. Subsets including CD8+ T-cells, CD56+ NK cells and CD19+ B-cells counts were in normal
- 218 ranges. However, immunoglobulin G mean level was low at 4.25 g/L. Means and standard
- 219 derivations with intervals of confidence were summarized in table 5. Concerning CD4+ T-cells
- subpopulations, memory/activated CD45RA⁻/RO⁺ T-cells was preponderant with mean count
- at 147/μL, naïve CD4+ T-cells production was impaired with naïve CD4+CD45RA+/RO- T-cells
- 222 (NTE) and CD4+CD45RA+/RO-/CD31+ cells (RTE) means count at 26/μL and 11/μL
- 223 respectively.
- Assessment of immune recovery over time after AHCT (figure 1) highlights a smaller individual
- variation and an lower cellular count of CD4+ T-cells, CD45RA-/RO+ T-cells, NTE, RTE, CD8+
- T-cells, and CD19⁺ B-cells in nocardiosis cohort compared to control cohort. CD56+ NK cells
- 227 distribution was more heterogeneous among both cohorts. Cell counts for the different subsets
- of immune cells are represented over time in figure 2. The case-control comparison of those
- values were evaluated by Mann-Whitney test (table 5) and highlighted significant lower cellular
- 230 count for lymphocytes (p=0.001), CD4+ T-cells (p<0.001), CD45RA-/RO+ T-cells (p=0.02), NTE
- 231 (p<0.001) and RTE (p<0.001), CD8+ T-cells (p=0.001) and CD19+ B-cells (p=0.001). CD56+
- NK cells cellular count was not different (p=0.33).
- 233 At the 7th month after AHCT, the median time of nocardiosis presentation, immune cellular
- recovery in the nocardiosis cohort was lower comparatively to the control cohort with circulating
- lymphocytes, CD4+ T-cells, CD8+ T-cells, CD56+ NK cells and CD19+ B-cells means count
- 236 (table 5). This relative deficient cellular count was persistent over time like that was presented
- 237 at 24 months.
- 238 Concerning CD4+CD45RA-/RO+ T-cells, NTE and RTE mean count, lower CD4+ T-cells in the
- 239 nocardiosis cohort compared to the control cohort result in a lower cellular count in those three
- subsets. That was persistent over time after AHCT in the nocardiosis cohort.

Outcomes

- 242 Five patients died after nocardiosis but only one death has been directly related to nocardiosis
- 243 16 days after diagnosis. The other 4 patients died of relapse of the hematological disease
- included one arising 32 days after nocardiosis diagnosis with lost of chimerism . No death was
- related to GVHD or other infection. Donor chimerism has been maintained in those who did
- 246 not relapse of their hematological disease.

Discussion

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Nocardiosis is a rare infection with polymorphic clinical presentation. Characteristics of our 249 250 rural high-risk population were similar to previous studies[5,31,32] including sex ratio and 251 incidence [33]. Indeed nocardiosis is more frequently diagnosed in dry, windy climates, facilitating aerosolization and dispersal in organism via respiratory routes [34,35] and in 252 agricultural workers [7]. The most common infectious site is lung by respiratory tract infestation 253 [6] as shown in our cohort (87%). The common radiological presentation with lungs 254 condensation with abscesses and nodules, and CNS abscesses were similar as already 255 described [2,5,7,8]. 256 Concerning nocardiosis diagnosis over time, we observed a higher incidence over few years. 257 Indeed in a previous epidemiologic evaluation between 2001 and 2012 [36] we shown 5 258 nocardiosis than we expanded with 5 nocardiosis between 2012 and 2016, and 5 more 259 260 between 2016 and 2017. Moreover recent use of alternative donor for AHCT (2 haploidenticals, 261 mismatch 9/10) and sequential conditioning (1 patient) inducing deeper immunosuppression [37] could explain higher incidence over time in our center. Outbreak was 262 263 already described by other teams [38] but this was not the case in our center. Indeed among the 5 nocardiosis between 2016 and 2017, 4 were documented by PCR with different Nocardia 264 species (N. shimofuensis/higoensis, farcinica, abscessus and nova), 1 was not documented. 265 In another hand, most samples were not routinely kept more than 1 week (48 hours for BAL, 5 266 days for blood cultures) which did not allowed Nocardia detection and also contributed to its 267 identification challenge [18]. Nocardia PCR permitted more quickly and higher number of 268 269 Nocardia detection. Also six nocardiosis were documented by PCR with specificity reported upon 74% [18]. The issues with that tool arise in case of chronic bronchopulmonary disease, 270 271 with colonization detection [18]. Widely use of PCR in our center could explain the higher number of nocardiosis. Three PCR were realized on BAL: 1 was associated to a gram-positive 272 bacillus on direct exam impossible to identified, and the 2 others were without any other 273 documentation. None presented a chronic bronchopulmonary disease. Colonization 274 275 hypothesis for the 2 Nocardia isolated in BAL was unlikely considering the severe neurological disseminated presentation for one, and a severe pulmonary infection with radiological 276 compatible presentation for the other. 277 278 Time between AHCT and nocardiosis diagnosis was quite different in our study (median time 218 days) compared to previous reports [39]. Usually nocardiosis was presented early (1st-2nd 279 month) and lately (1st-2nd year) after AHCT This particularity may be explained by the 280 proportion of RIC (40%) or alternative AHCT (20%) [37,40]. CsA and high dose steroid (more 281 than 1mg/kg) use, age at transplantation (mean 49 years-old) and CMV reactivation (26%) 282

were known to altered immune recovery, notably thymic function [41-43]. These

characteristics were identical compared to literature for current patients undergoing to AHCT [44]. Cumulative steroid dose >23mg/kg/4weeks was also described as an increased risk of infection [45] but we did not permit to observe statistical difference in steroid dose between our 2 cohorts. Half of the patients (8/15) received TMP-SMX prophylaxis at nocardiosis diagnosis. This corroborates that TMP-SMX 3 time a week prophylaxis was not sufficient for nocardiosis prophylaxis as describes in others studies [9,10,16,46,47], but may reduce nocardiosis mortality [48]. However TMP-SMX resistance is rare [49], and poor absorption could be frequent, especially in context of gut GVHD and/or polymedication. Atovaquone is known to be useless [48] for nocardia prophylaxis. Specific recommendations were published by *Lebeaux et al.* in 2014 about nocardiosis diagnosis and management after AHCT [50] reminding the necessity to use fast molecular diagnosis such PCR and to start a large empirical antimicrobial therapy as soon as Nocardia is suspected (secondary adjusted) that should be maintained over 6 to 12 months if diagnosis of nocardiosis is confirmed.

Immune recovery after AHCT is delayed and impaired, frequently described in the literature [37,51]. CD4+ T-cells recovery is delayed to several years after AHCT. Large use of immunosuppressive drugs at nocardiosis diagnosis (88%) delay CD4+ T-cells recovery. Moreover patients with nocardiosis seemed to present a low CD8+ T-cells and CD19+ B-cells recovery, with a significative lower cellular count, that could reflect a higher immunosuppression. Low B-cells count associated to hypogammaglobulinemia may need to consider optimization of immunoglobulin substitution strategy. As already reported, infections may arise despite an acceptable quantitative NK-cells and CD8⁺ T-cells recovery [51–53], which values were similar to our laboratory benchmarks suggesting a minor implication against Nocardia. Immune recovery against Nocardia is rather mediated by CD4+ T-cells [54]. Nevertheless in our Nocardia cohort normal absolute count CD4⁺ T-cells was not sufficient to protect against Nocardia. CD4+ T-cells recovery after AHCT initially was induced by peripheral clonal expansion of donor memory T cells expressing activation markers (CD45RA-/RO+). This initial response is usually followed by central naïve CD4+ T-cells expansion CD45RA+/RO-[55-57]. Activated memory T cells recovery in our cohort were shown to be expanded but also failed to prevent nocardiosis. NTE were practically undetectable before and during nocardiosis follow-up. Those descriptions suggest that *Nocardia* did not contribute to delay CD4⁺ T-cells recovery but was rather a consequence of cellular immune recovery delay.

As same as NTE, RTE count was significantly lower in nocardiosis cohort than control cohort and continued over time. Delay for NTE and RTE repopulation until "normal" level [51,52] was described as a key point that infection risk and mortality risk decreased after AHCT [29,57,58]. Peripheral naïve T-CD4+ cells RTE or not (CD31+ or not) [59] may have equal function but RTE presented a broader polyclonal TCR repertoire compared to non-thymic naïve T-CD4+ cells

[60]. TCR repertoire restriction with lack of thymic function may conduct to reduce anti infectious capacity. RTE recovery induces efficient protection against opportunist infection after AHCT [61]. Monitoring efficiency of thymopoiesis by quick tools may allow identifying high-risk patients for opportunistic infection and then personalized

The analysis of our cohort suggest that the risk of *Nocardia* infection after AHCT is potentially lethal and should be considered with a more intensive laboratory investigation strategy including systematic PCR detection. The assumption of such opportunistic infection is tricky regarding the non specific aspect of its clinical and radiological presentation. Diagnosis should be considered until effective immune cellular recovery, particularly for T-CD4+ cells. We highlight through nocardiosis that the lack of thymopoiesis with low RTE recovery may need a particular attention towards opportunistic infections and prophylaxis with TMP-SMX, even not completely reliable, must be prolonged if necessary. A larger multicentric cohort should be conducted to corroborate those results and to provide more robust recommendations.

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Conflict of interest

We have no Conflict of Interest.

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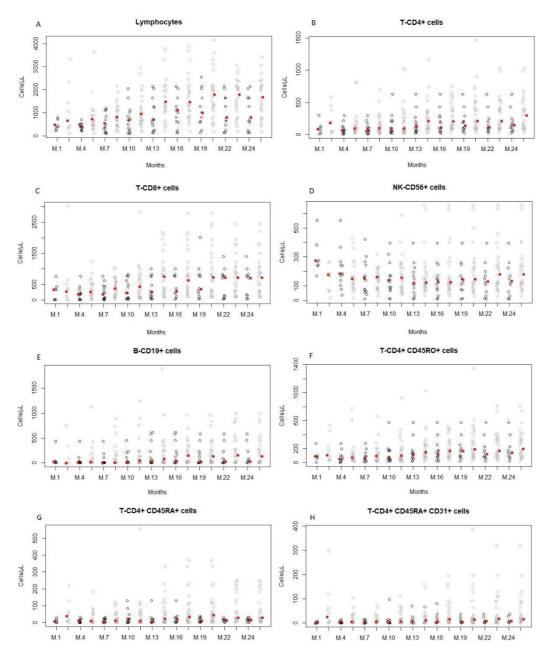


Fig 1 A-H: Immune recovery over time during 2 years after AHCT in stripchart. Nocardiosis cohort is in back, control cohort is in light grey. Median point is in red.

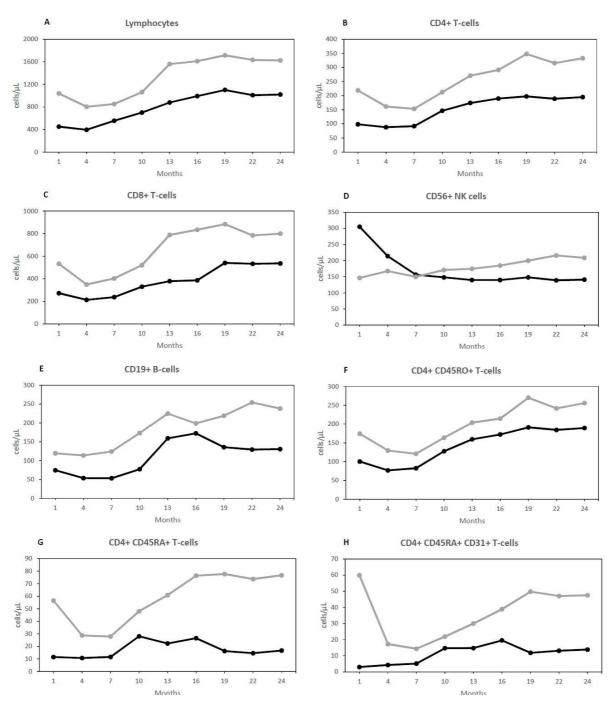


Fig 2 A-H: Immune recovery over time during 2 years after AHCT with means comparaison. Nocardiosis cohort is in back, control cohort is in grey.

(Table 1) Nocardiosis definitions					
Possible	Compatible host +clinic and radiologic criteria				
Probable	Possible + non-direct microbiologic criteria (direct exam, pathology, PCR)				
Proved	Previous criteria + positive culture				

(Table 2) Transplant characteristics								
Nocardiosis cohort Control cohort								
Mean age (years) (p=0.11)	49 [43-55]	43 [37-49]						
<u>Sex ratio</u>	4:1	4:1						
Hematological disease								
Acute myeloid leukemia	9 (61%)	18 (61%)						
Acute lymphoblastic leukemia	2 (13%)	4 (13%)						
Myelodysplasia	2 (13%)	4 (13%)						
Multiple myeloma	2 (13%)	4 (13%)						
<u>Conditioning</u>								
Myeloablative conditioning	9 (60%)	18 (60%)						
Reduce intensity conditioning	6 (40%)	12 (40%)						
HSCT type								
Bone marrow	8 (54%)	16 (54%)						
Peripheral blood stem cells	7 (46%)	14 (46%)						
<u>Donor type</u>								
Sibling	8 (54%)	16 (54%]						
Unrelated donor	4 (27%)	10 (33%)						
9/10	1 (6%)	-						
Haplo identical	2 (13%)	4 (13%)						
CMV donor/recipient status								
+/+	4 (27%)	11 (37%)						
+/-	3 (20%)	2 (6%)						
-/+	5 (33%)	9 (30%)						
-/-	3 (20%)	8 (27%)						
<u>Graft-versus-host disease</u>								
Acute grade	12 (80%)	21 (70%)						
Grade I-II	8 (53%)	15 (50%)						
Grade III-IV	4 (27%)	6 (20%)						
Chronic	11 (73%)	21 (70%)						
Mild	1 (6%)	10 (33%)						
Moderate	7 (47%)	7 (24%)						
Severe	3 (20%)	4 (13%)						
Steroid cumulative dose								
Mean during 1 st month (p=0.20)	25 mg/kg/4weeks	19 mg/kg/4weeks						
	[15.5-34.9]	[8.5-28.6]						

(Table 3) Nocardiosis characteristics (n=15)							
Prophylaxis (TMP-SMX)	8 (53%)	Antibiotic treatment (n=17)					
		Induction					
Infection status		Ceftriaxone	1 (6%)				
Localized	7 (47%)	Ceftriaxone + Linezolid	1 (6%)				
Disseminated	8 (53%)	IMP	5 (29%)				
		IMP + AMK	3 (18%)				
Infection site		IMP + TMP-SMX	2 (12%)				
Pulmonary	13 (87%)	IMP + Linezolid	1 (6%)				
CNS	8 (53%)	MERO + Linezolid	2 (12%)				
Ocular	4 (27%)	MERO + Linezolid + AMK	1 (6%)				
Cutaneous	2 (12%)	Cefuroxim + TMP-SMX	1 (6%)				
Bacteremia	1 (6%)						
		Consolidation					
<u>Documentation</u>		TMP-SMX	4 (24%)				
Cyto-histopathology	3 (20%)	TMP-SMX + Cefuroxim	3 (18%)				
PCR	10 (66%)	TMP-SMX + Ceftriaxone	1 (6%)				
Culture	5 (66%)	TMP-SMX + Cefotaxim	1 (6%)				
		Moxifloxacin	2 (12%)				
<u>Co-infection</u>		Ceftriaxone	2 (12%)				
CMV	4 (26%)	MERO + Linezolid	1 (6%)				
Pneumocystis	2 (13%)	Minocycline + Ciprofloxacin	1 (6%)				
Mucormycosis	2 (13%)	Dead before	2 (12%)				
		<u> </u>					
Mean steroid dose at diagnosis	0.30mg/	kg/day [0.19-0.58]					

IMP imipenem, MERO meropenem, AMK amikacin, TMP-SMX trimethoprim-sulfamethoxazole, CNS Central Nervous System

(Table 4)) Nocardiosis de	etails							
Patient	Prophylaxis	Infectious site	Radiology	Sample site	Pathology	Culture	PCR	Species	Status
#1	TMP-SMX	Lung	Chest CT scan : lung abscess	Lung biopsy	+	+	+	N. cyriacigeorgica	Proved
#2	None	Skin	Smooth tissue CT scan : abscess	Skin biopsy	-	+	+	N. farcinica	Proved
#3	TMP-SMX	CNS, Oc, Lung	Cerebral MRI : multiple abcesses + Chest CT scan : lungs nodules	Lung biopsy	indirect	+	+	N. farcinica	Proved
#4	TMP-SMX	CNS, Oc, Lung	Cerebral CT scan : multiple abcesses + Chest CT scan : lung nodule	BAL	-	not made	+	N. nova	Probable
#5	TMP-SMX	CNS, Oc, Lung	Cerebral CT scan : multiple abcesses + Chest CT scan : lung nodule	CSF, vitreous, BAL	indirect	-	-	•	Possible
#6	TMP-SMX	Lung, pleural	Chest CT scan : lung nodule + pleural condensation	Lung biopsy	+	not made	+	N. shimofuensis / higoensis	Probable
#7	Pentamidine	CNS, Oc, Lung	Cerebral CT scan: multiple abcesses + Chest CT scan: small lung nodule + pleural condensation	BAL	not made	-	+	N. nova	Probable
#8	Pentamidine	CNS, Lung	Cerebral MRI : 2 large abcesses + Chest CT scan : lung nodule	CSF, BAL, Abscess	+	+ (abscess)	+ (CSF)	N. nova	Proved
#9	TMP-SMX	CNS, Lung	Cerebral MRI : abcesses + Chest CT scan : lung abscess + lungs nodules	No-biopsy able, BAL not made	not made	not made	not made	-	Possible
#10	Atovaquone	Lung	Chest CT scan: lungs condensations	BAL not made Biopsy not made	not made	not made	not made	-	Possible
#11	TMP-SMX	CNS, Lung, Skin	Cerebral CT scan: abcesses + Body CT scan: lungs condensations + liver abscess	Blood culture	not made	+	not made	N. farcinica	Proved
#12	TMP-SMX	Lung	Chest CT scan: lungs condensations + lungs nodules	BAL	indirect	Few GPB, none identification able	+	N. abscessus	Probable
#13	None	Lung	Chest CT scan: lungs nodules, condensation, abcesses + pleural condensation	Pleural biopsy	indirect	-	+	N. farcinica	Probable
#14	None	CNS	Cerebral CT scan : compressive abcess	CSF	-	-	+	N. spp	Probable
#15	None	Lung	Chest CT scan: lung abcess +pleural condensations	BAL No-biopsy able	-	-	not made	-	Possible

ime points		At day +218 (median time of		At 24 months	after AHCT	Mann-	
	onset	nocardiosis presentation)		710 2 1 1110111111	Whitney test		
	Nocadiosis cohort	Nocadiosis cohort	Contrôle cohort	Nocadiosis cohort	Contrôle cohort	williney test	
Lymanhaaytaa	730/μL	554/μL	851/μL	1020/μL	1625/μL	p=0.001	
Lymphocytes	[394-1067]	[356-753]	[669-1032]	[635-1404]	[1330-1920]	ρ=0.001	
CD4+ T-cells	162/μL	92/μL	153/μL	195/μL	332/μL	n < 0, 001	
CD4+ 1-cells	[80-244]	[49-134]	[97-210]	[102-288]	[236-429]	p<0.001	
CD4+ CD45RO+ T-cells	147/μL	83/μL	121/μL	189/μL	256/μL	n=0.02	
	[72-223]	[43-122]	[72-170]	[102-277]	[183-328]	p=0.02	
CD4+ CD45RA+ T-cells	26/μL	12/μL	28/μL	17/μL	77/μL	p<0.001	
	[8-44]	[6-17]	[16-40]	[11-22]	[46-108]		
CD4+ CD45RA+ CD31+ T-cells	11/μL	5/μL	14/μL	14/μL	48/μL	p<0.001	
CD4+ CD43KA+ CD31+ 1-Cells	[0-23]	[2-8]	[8-21]	[7-21]	[23-73]	ρ<0.001	
CD8+ T-cells	362/μL	238/μL	403/μL	556/μL	800/μL	n=0.001	
	[125-600]	[112-363]	[300-506]	[297-775]	[570-1030]	p=0.001	
CD56+ NK cells	160/μL	156/μL	150/μL	141/μL	209/μL	n=0.22	
	[94-227]	[97-216]	[126-173]	[81-201]	[153-264]	p=0.33	
CD19 B-cells	112/μL	54/μL	124/μL	131/μL	238/μL	p=0.001	
CD13 P-CEII2	[12-214]	[8-115]	[42-206]	[27-234]	[137-339]		

immune reconstitution at time points with cells counts, interval of confidence at 95% and Mann Whitney test.